Date: 16 Feb 2023

Title: The effect of ascertainment on penetrance estimates for rare variants: implications for establishing pathogenicity and for genetic counselling

Running title: Ascertainment effects on penetrance estimates

Authors: Andrew D. Paterson*[1,2], Sang-Cheol Seok[3], Veronica J. Vieland[3,4]

ORCID:

ADP: 0000-0002-9169-118X

SCS: 0000-0003-3955-9891

VJV: 0000-0002-3004-3840

Affiliations:

1. Program in Genetics and Genome Biology, The Hospital for Sick Children, Toronto, Ontario, M5G 1X8, Canada
2. Divisions of Epidemiology and Biostatistics, Dalla Lana School of Public Health, University of Toronto, Toronto, Ontario, Canada
3. Mathematical Medicine LLC, Chicago, IL
4. Departments of Pediatrics and Biostatistics (Emerita), The Ohio State University, Columbus, OH

***Correspondence:** andrew.paterson@sickkids.ca

Keywords:

Penetrance, ascertainment, Mendelian disease, multiplex family, rare variant, variant pathogenicity

Abstract:

Next-generation sequencing has led to an explosion of genetic findings for many rare diseases. However, most of the variants identified are very rare and were identified in small pedigrees, which creates challenges in terms of penetrance estimation and translation into genetic counselling in the setting of cascade testing. We use simulations to show that for a rare (dominant) disorder where a variant is identified in a small number of small pedigrees, the penetrance estimate can both have large uncertainty and be drastically inflated, due to underlying ascertainment bias. We have developed PenEst, an app that allows users to investigate the phenomenon across ranges of parameter settings. We also illustrate robust ascertainment corrections via the LOD score, and recommend a LOD-based approach to assessing pathogenicity of rare variants in the presence of reduced penetrance.

Next-generation sequencing has led to an explosion in the number of genetic findings for many rare diseases. For certain types of rare coding variants (e.g. missense, or protein truncating), if the variant is sufficiently rare and has bioinformatic predictions that are severe, current algorithms result in it being classified as pathogenic (1). However, the analysis of large-scale sequencing from cohorts, such as ExAC (2), gnomAD (3), and the UK Biobank (4), has shown that many such variants may often lack clinically significant impact. For example, ExAC estimated that individuals from population cohorts carried a mean of 53 variants previously thought to be sufficient causes of Mendelian diseases. Additionally, 88% of such variants had MAF>1%, implying that they are likely not sufficient causes. This may indicate that such variants are not causally related to disease, or perhaps, that they are causally related but with reduced penetrance.

Penetrance plays an important role in understanding disease pathology, in the appropriate classification of pathogenic variants, and perhaps above all in the context of genetic counseling. However, most of the variants reported to date have been very rare and identified in small sets of unrelated individuals (sometimes just one) or small pedigrees. Penetrance cannot be estimated from a single case, or a single parent-offspring trio presenting with a *de novo* mutation in the offspring. But even with multiple cases or families, determination of the penetrance can present challenges. Here we focus on one such challenge: ascertainment.

Typically a variant of interest is first identified in one individual with a given phenotype. Investigators may then sequence either additional relatives of the individual, or additional individuals or families presenting with the same or closely related phenotypes, with the goal of

bolstering the case for pathogenicity. Thus, ascertainment of individuals to be sequenced typically proceeds in stages. The precise ascertainment process used to enrol individuals and/or families is usually at least to some extent unsystematic, and may vary between families. Ascertainment is therefore challenging to model when attempting to estimate the penetrance of a variant.

One situation in which ascertainment can be easily handled is "single" ascertainment, in which the probability of an affected individual being ascertained is proportional to the number of affected individuals in the family (5). In fact, much of the literature on inferring pathogenicity or estimating penetrance tends to assume single ascertainment, e.g., (6), where ascertainment is addressed by conditioning on "the proband," a procedure which is strictly correct only under true single ascertainment. While it is true that the typical study ascertains families through one individual who may be designated as the single "proband", this does not ensure that the study meets the proportionality requirement of single ascertainment. This requirement would be violated, e.g., if families with four affected members were more than twice as likely to be recruited as families with just two; or, if the probability of a second sibling being ascertained were dependent on the ascertainment status of the first. And in general, if either (i) ascertainment is not truly single, or (ii) even if it is, if an appropriate ascertainment correction is not incorporated into the estimation method, then penetrance estimates will be biased. Here we consider the magnitude of that bias, across a range of plausible ascertainment models and varying amounts of available data.

We focus here on sibship data. The impact of ascertainment for more complex pedigrees can be approximated by considering large sibship sizes. For simplicity, we assume all parents are

82 phenotypically and genotypically unknown; including parental information does not
83 substantively affect results. We assume a very rare variant of interest (VOI), and an autosomal
84 dominant disease D. Let a qualifying individual (QI) be anyone who is both heterozygous (HET)
85 for the VOI and also affected (AFF) with D. Let r be the number of QI sibs within a family, and
86 let t be the number of AFF sibs regardless of VOI genotype. We also assume that, regardless of
87 VOI status, an individual might develop D due to other factors, which might be genetic
88 (involving one or more VOIs at other loci or other variants within the same gene) and/or
89 environmental (e.g., due to infections). Let γ be the combined penetrance across all causes other
90 than the VOI under study. Since we assume the VOI is very rare, γ is effectively the population
91 prevalence of D.
92
93 In order to consider a range of plausible ascertainment scenarios, we employ the general family-
94 based k-model of ascertainment (7). In its simplest form, this model stipulates that the probability
95 that a family is ascertained is proportional to r^k, where k controls the model. For example, when
96 $k = 1$, the probability of ascertainment is strictly proportional to r: this is equivalent to classical
97 "single ascertainment". Similarly, when $k = 0$, so that every family with $r \geq 1$ is ascertained,
98 this model is equivalent to classical "complete" or "truncate" ascertainment. We generalize this
99 model in two ways. First, we assume that ascertainment requires $r \geq 1$, that is, every
100 ascertained family contains at least one QI, but we allow that there may be additional preferential
101 ascertainment of families based on t alone, that is, that investigators may preferentially ascertain
102 families with more affected individuals without knowing (or prior to knowing) the VOI status of
103 those additional individuals. Second, we allow that even an individual carrying the VOI may

develop disease due to any other independent causes at work in the general population. With these two extensions in mind, our ascertainment model becomes

P[sibship is ascertained | r, t] = $c(r^k + t); for\ r \geq 1$, and 0 otherwise

where c is a normalizing constant.

Let f be the attributable penetrance, or the penetrance due to the VOI for HET individuals. (Note that when $\gamma > 0$, β=P[AFF|HET] $= \gamma + f - \gamma f$. However, we focus here on estimation of f itself rather than β.) In what follows, we estimate f in two ways:

(ii) $\tilde{f}$ is obtained by counting the proportion of AFF individuals among all HET individuals in the data set, after dropping one QI individual per family, that is, applying the correction for single ascertainment;

(ii) $\tilde{f}^*$ is obtained by counting the proportion of AFF individuals among all HET individuals in the data set, that is, without applying any ascertainment correction.

$\tilde{f}^*$ is a naïve estimate, which would be correct if the families were not ascertained based on either phenotype or genotype. It is, however, clearly incorrect under any of our ascertainment models. Our interest in this estimate is to establish how biased it becomes under various ascertainment scenarios. $\tilde{f}$ by contrast, does apply the frequently employed single ascertainment correction, and again, our interest in $\tilde{f}$ is to establish how biased it will be under ascertainment scenarios other than single ascertainment. Expected values of $\tilde{f}$ and $\tilde{f}^*$ were obtained via simulation, by averaging each estimate's value across 1,000 replicates per generating condition, and standard errors were obtained by averaging the standard deviation of each estimate across those same 1,000 replicates. (While the expected values are easily calculated analytically, the standard errors

are not.) All simulations and calculations were done in MATLAB (2021.9.10.0.1739362 (R2021a), Natick, Massachusetts: The MathWorks Inc.).

Let s be the number of siblings in a family, and let N be the number of s-sized sibships in a dataset. Fig 1 shows results for true single ascertainment (k=1), for $s = 2$, as a function of sample size N. Here we assume that the true value of f=0.5. As can be seen, in this case, the mean of $\tilde{f}$ = 0.5, the generating value, as expected. But using $\tilde{f}^*$ the estimates are seriously upwardly biased in all data sets, regardless of N. Note that because each sibship contains at least one QI, by stipulation, the minimum value of $\tilde{f}^*$ is 0.50.

Note too that even the correct estimate $\tilde{f}$ shows considerable sampling variability. For instance, with N=10, $\tilde{f}$ will be >70% or <30% in approximately 40% of all data sets when f=50%. This variability remains appreciable even for N=50.

For ascertainment models other than single, overall variability remains similar to what is shown in Fig 1, but even $\tilde{f}$ tends to be biased, with mean $\tilde{f}$ = 0.60, 0.50, 0.43 and 0.38 for k = 2, 1, 0 and −1, respectively. In all cases, the uncorrected $\tilde{f}^*$ will return even more biased estimates, with mean $\tilde{f}^*$ = 0.89, 0.88, 0.87 and 0.86, for k = 2, 1, 0 and −1, respectively.

Fig 2 shows the impact of the population prevalence γ on average penetrance estimates. Focusing first on single ascertainment (k=1) and $\tilde{f}$ =0.5, we can see that regardless of k, the expected value of $\tilde{f}$ is relatively independent of γ until γ becomes quite high. Note that for f = 0.5 and γ = 0.5, the actual probability that a VOI carrier is affected under our generating model is

150 0.5 + 0.5 – (0.5)(0.5) = 0.75, which is in line with the estimates returned by $\tilde{f}$. $\tilde{f}^*$ might be said
151 to be even more robust to γ, although this is because in this case $\tilde{f}^*$ is already close to the top of
152 the scale for γ=0. Moreover, $\tilde{f}^*$ appears not only robust to γ, but also to f itself, with estimates
153 >70% even for f=0.05, and >80% for f=0.05 when γ=0.5. These patterns repeat for different
154 values of k, with visible impact only on the magnitude of the bias for any given (f, γ)
155 combination. Ascertainment effects will be reduced as s increases. Users who are interested in
156 investigating penetrance estimates for other ascertainment models, other combinations of
157 parameter values or other sibship sizes are encouraged to download the PenEst app:
158 https://github.com/MathematicalMedicine/PenetranceEstimator.
159
160 In general, our simulations show that under unsystematic ascertainment schemes, or in cases
161 where appropriate ascertainment corrections are not included in the estimation procedure, there
162 is a high risk of over-estimating the penetrance of any given VOI. This finding is consonant with,
163 and may in large part explain, reports for specific variants. For example, multiple coding variants
164 in *PRNP* had been reported to cause rare dominant monogenic neurodegenerative disease, but
165 there was a 30-fold higher prevalence of variants previously suggested to be causal in this gene
166 in ExAC compared to the expected frequency calculated from the estimated prevalence of the
167 disorder (8). Specifically for three variants the lifetime risk of developing disease was <10%.
168 Similarly, GWAS array data from the UK Biobank were used to estimate pathogenicity,
169 penetrance, and expressivity of putative disease-causing rare variants (MAF<1%) that were
170 directly genotyped and had good quality (9). Focused on maturity-onset diabetes of the young
171 and developmental disorders, many specific variants were found for which the penetrance --
172 estimated either in families ascertained for the presence of the VOI or in disease cohorts -- was

173 much higher than that obtained from a population-based cohort. These observations have
174 implications for genetic counselling, including the recommendation of invasive screening
175 procedures and administration of preventative treatment.
176
177 Some approaches to the interpretation of rare coding variants assume either full or high
178 penetrance (10), for the sake of simplicity. Extensive criteria have been proposed to claim a
179 causal relationship between variants and disease, and authors have urged caution in presuming
180 full penetrance for pathogenic variants (11). But in practice, penetrance remains an important
181 factor in assessing pathogenicity. For instance, the ACMGG/AMP joint consensus
182 recommendations (1) warns against ignoring the possibility of reduced penetrance in establishing
183 segregation of a VOI with a phenotype, but also instructs that "lack of segregation…provides
184 strong evidence against pathogenicity." (p. 15) And in practice, many laboratories will rule out
185 candidate VOIs when they are found among unaffected relatives. Particularly in the absence of a
186 rigorous and accurate estimate of the actual penetrance, this complicates the use of segregation
187 information in assessments of pathogenicity.
188
189 We close by noting that there is one essentially "ascertainment assumption free" (12) method for
190 estimating the penetrance, viz., by conditioning on all of the phenotypic data. This is the
191 ascertainment correction implicit in the usual LOD score (13-15), and also the LOD score
192 allowing for linkage disequilibrium or LD-LOD (6, 16, 17), and in principle any program that
193 allows calculation of the LOD score will support this method. As in Thompson (6) the
194 calculation is done here assigning the VOI (which plays the role of the "marker") and the disease
195 allele the same (rare) frequency (we have used 0.001 in the simulations), assuming complete

196 linkage disequilibrium between the two (D′ = 1), and also assuming 0 recombination between the
197 marker and the disease allele. Free parameters in the model are then the three penetrances; in our
198 calculations we also include the admixture parameter α of Smith (18), representing the
199 probability that any given family is of the "linked" type, which adds robustness when phenocopy
200 levels are high. Maximizing the LD-LOD over the free parameters gives us the LD-MOD, which
201 occurs at the maximum likelihood estimate (m.l.e.) of $\hat{f}$ of f (12-15).
202
203 Fig. 3 shows results corresponding to the simulations in Fig 1A and Fig 2A, C. As can be seen, $\hat{f}$
204 behaves very much like $\tilde{f}$ when k = 1 (Fig 3A), but it retains almost complete robustness to
205 ascertainment, and also to γ at least until γ is quite large (Fig 3B). (As with $\tilde{f}$, as γ gets very
206 large, $\hat{f}$ covers both cases due to the VOI and also cases among variant carriers due to other
207 causes.) Comparing Fig 3A with Fig 1A, $\hat{f}$ shows slightly greater sampling variability than $\tilde{f}$;
208 this is due to the inherent ascertainment correction built in to $\hat{f}$. The slight but systematic over-
209 or under-estimation of f seen in Fig 3B is due to the small sample size; as N increases $\hat{f} \rightarrow f$
210 (results not shown). However, in small samples the upward bias can be appreciable particularly
211 when f is small; e.g., when $f = 0.05$ ($\gamma = 0$), for N = 20, the expected value of $\hat{f} = 0.165$.
212
213 Note, however, that while maximizing the LD-LOD is a highly ascertainment-robust method for
214 estimating f, the LD-MOD itself is not a good statistic for representing the strength of evidence
215 for co-segregation, because it is not additionally conditioned on ascertainment through the VOI.
216 However, once we ascertain so as to require the VOI to be present in the family, there is no
217 remaining LD information in the sibship, since LD information is conveyed entirely by the
218 marker allele frequencies in the parents. Therefore, we recommend using the ordinary (linkage

219 equilibrium) LOD, or LE-LOD, for assessing strength of evidence for co-segregation. Because
220 maximizing the LE-LOD itself will not return true m.l.e.s of *f* under the LD model, we
221 recommend evaluating the LE-LOD at the maximizing model obtained from the LD-MOD, for a
222 statistic we annotate as LE-LOD(max). (This maximization procedure is not inherently
223 inflationary; see Supplemental Results (A). Thompson et al. (6) proposed a form of Bayes factor
224 for assessing evidence for co-segregation of the VOI with disease; see the Supplemental Results
225 (B) for some comparisons between their Bayes factor and LE-LOD(max).) Fig 3(D) shows the
226 distribution of the LE-MOD(max), for the same data shown in Fig 1. (Here parents are treated as
227 genotypically known but phenotypically unknown.) As expected, based on just a few 2-child
228 sibships, evidence of co-segregation of the VOI with disease is quite weak. It requires at least N
229 = 30 2-child families before there is a reasonable chance of obtaining a substantial LE-
230 LOD(max).
231
232 **Declaration of interests**
233 The authors declare no competing interests
234 **Acknowledgements:**
235 This work was supported in part by Mathematical Medicine LLC, with special thanks to Jo
236 Valentine-Cooper for creation of the PenEst app.
237
238 **Web resources**
239 PenEst app : https://github.com/MathematicalMedicine/PenetranceEstimator/
240
241 **Data and code availability**
242 https://github.com/MathematicalMedicine/PenetranceEstimator/

243 **References**

244 1. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for
245 the interpretation of sequence variants: a joint consensus recommendation of the American College of
246 Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med.
247 2015;17(5):405-24.

248 2. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, et al. Analysis of protein-
249 coding genetic variation in 60,706 humans. Nature. 2016;536(7616):285-91.

250 3. Karczewski KJ, Francioli LC, Tiao G, Cummings BB, Alföldi J, Wang Q, et al. The mutational
251 constraint spectrum quantified from variation in 141,456 humans. Nature. 2020;581(7809):434-43.

252 4. Backman JD, Li AH, Marcketta A, Sun D, Mbatchou J, Kessler MD, et al. Exome sequencing
253 and analysis of 454,787 UK Biobank participants. Nature. 2021;599(7886):628-34.

254 5. Hodge SE, Vieland VJ. The essence of single ascertainment. Genetics. 1996;144(3):1215-23.

255 6. Thompson D, Easton DF, Goldgar DE. A full-likelihood method for the evaluation of causality of
256 sequence variants from family data. Am J Hum Genet. 2003;73(3):652-5.

257 7. Ewens WJ, Shute NC. The limits of ascertainment. Ann Hum Genet. 1986;50(4):399-402.

258 8. Minikel EV, Vallabh SM, Lek M, Estrada K, Samocha KE, Sathirapongsasuti JF, et al.
259 Quantifying prion disease penetrance using large population control cohorts. Sci Transl Med.
260 2016;8(322):322ra9.

261 9. Wright CF, West B, Tuke M, Jones SE, Patel K, Laver TW, et al. Assessing the Pathogenicity,
262 Penetrance, and Expressivity of Putative Disease-Causing Variants in a Population Setting. Am J Hum
263 Genet. 2019;104(2):275-86.

264 10. Jarvik GP, Browning BL. Consideration of Cosegregation in the Pathogenicity Classification of
265 Genomic Variants. Am J Hum Genet. 2016;98(6):1077-81.

266 11. MacArthur DG, Manolio TA, Dimmock DP, Rehm HL, Shendure J, Abecasis GR, et al.
267 Guidelines for investigating causality of sequence variants in human disease. Nature.
268 2014;508(7497):469-76.

269 12. Ewens WJ, Shute NC. A resolution of the ascertainment sampling problem. I. Theory. Theor
270 Popul Biol. 1986;30(3):388-412.
271 13. Greenberg DA. Inferring mode of inheritance by comparison of lod scores. Am J Med Genet.
272 1989;34(4):480-6.
273 14. Elston RC. Man bites dog? The validity of maximizing lod scores to determine mode of
274 inheritance. Am J Med Genet. 1989;34(4):487-8.
275 15. Vieland VJ, Hodge SE. The problem of ascertainment for linkage analysis. Am J Hum Genet.
276 1996;58(5):1072-84.
277 16. Slager SL, Huang J, Vieland VJ. Power comparisons between the TDT and two likelihood-based
278 methods. Genet Epidemiol. 2001;20(2):192-209.
279 17. Petersen GM, Parmigiani G, Thomas D. Missense mutations in disease genes: a Bayesian
280 approach to evaluate causality. Am J Hum Genet. 1998;62(6):1516-24.
281 18. Smith CA. Testing for heterogeneity of recombination fraction values in human genetics. Ann
282 Hum Genet. 1963;27:175-82.
283 19. Vieland VJ, Huang Y, Seok SC, Burian J, Catalyurek U, O'Connell J, et al. KELVIN: a software
284 package for rigorous measurement of statistical evidence in human genetics. Hum Hered. 2011;72(4):276-
285 88.

286 **Figure 1.** Swarm plots showing sampling distributions of penetrance estimates as a function of
287 number of families N.
288 Distributions of (A) $\tilde{f}$ and (B) $\tilde{f}^*$ are shown for simulations of 1000 replicates, with true
289 penetrance f=0.5. The number of sibs per family, s=2; phenocopy rate, γ=0. Users interested in
290 varying the parameters can use the PenEst app.
291
292 **Figure 2.** Expected values of penetrance estimates as a function of population prevalence γ and
293 ascertainment parameter k.
294 Top row: expected values of (A) $\tilde{f}$ and (B) $\tilde{f}^*$ when the true penetrance f=0.5. Bottom row:
295 expected values of (C) $\tilde{f}$ and (D) $\tilde{f}^*$ when f=0.2 (lower line sets) or f=0.8 (upper line sets). The
296 number of sibs per family, s=2. Users interested in varying the parameters can use the PenEst
297 app.
298
299 **Figure 3.** (A) Swarm plots showing sampling distributions of $\hat{f}$, as obtained from maximizing
300 the LD-LOD, as a function of number of families N; (B) Expected values of $\hat{f}$ as a function of
301 population prevalence γ and ascertainment parameter k, for f = 0.2, 0.5 and 0.8, reading from
302 bottom to top of the plot, respectively; (C) Swarm plots showing the distribution of LE-
303 LOD(max) as a function of N. Data are the same as used to generate Figures 1 and 2,
304 respectively. All calculations were done using KELVIN (19).

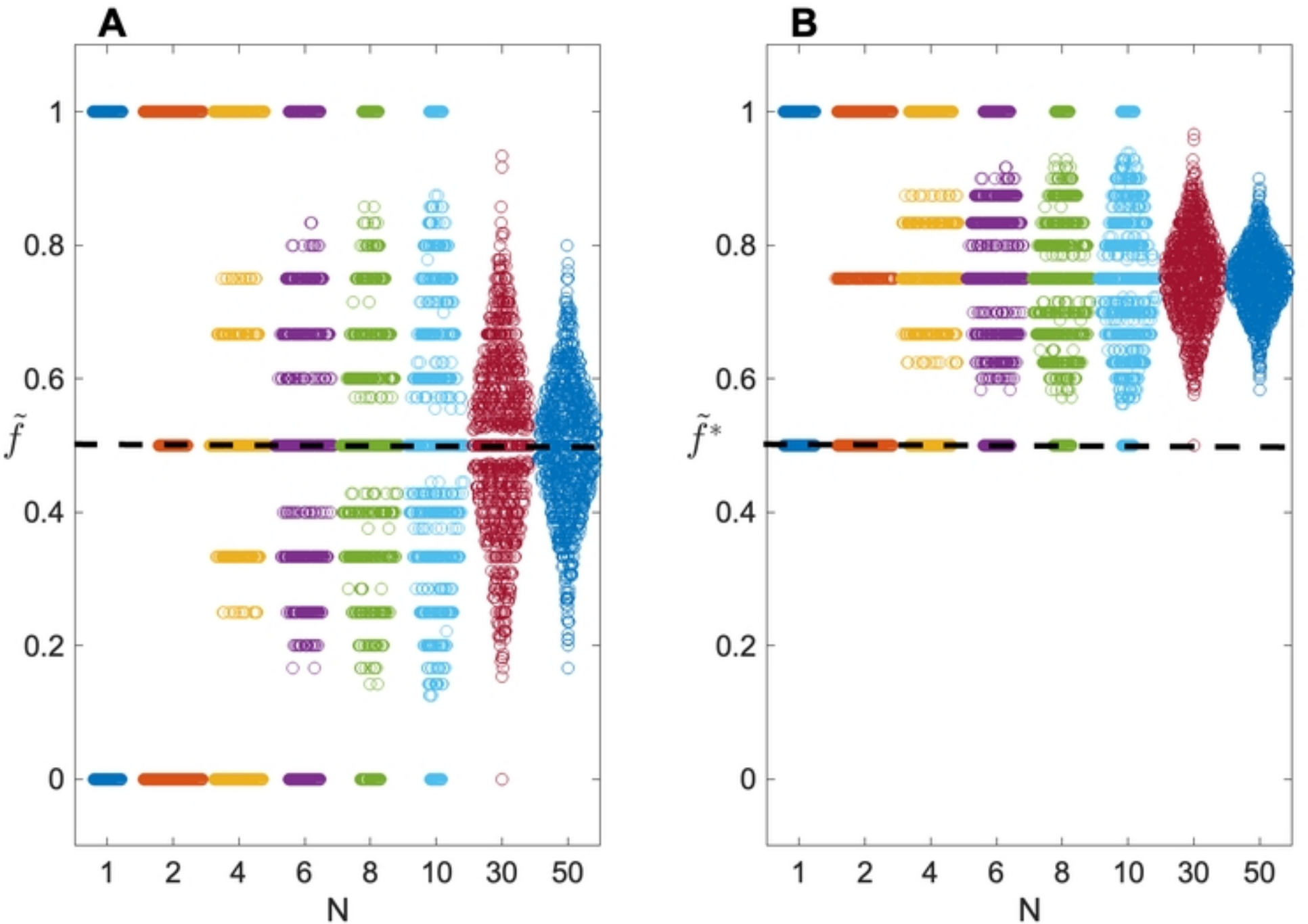

Figure 1

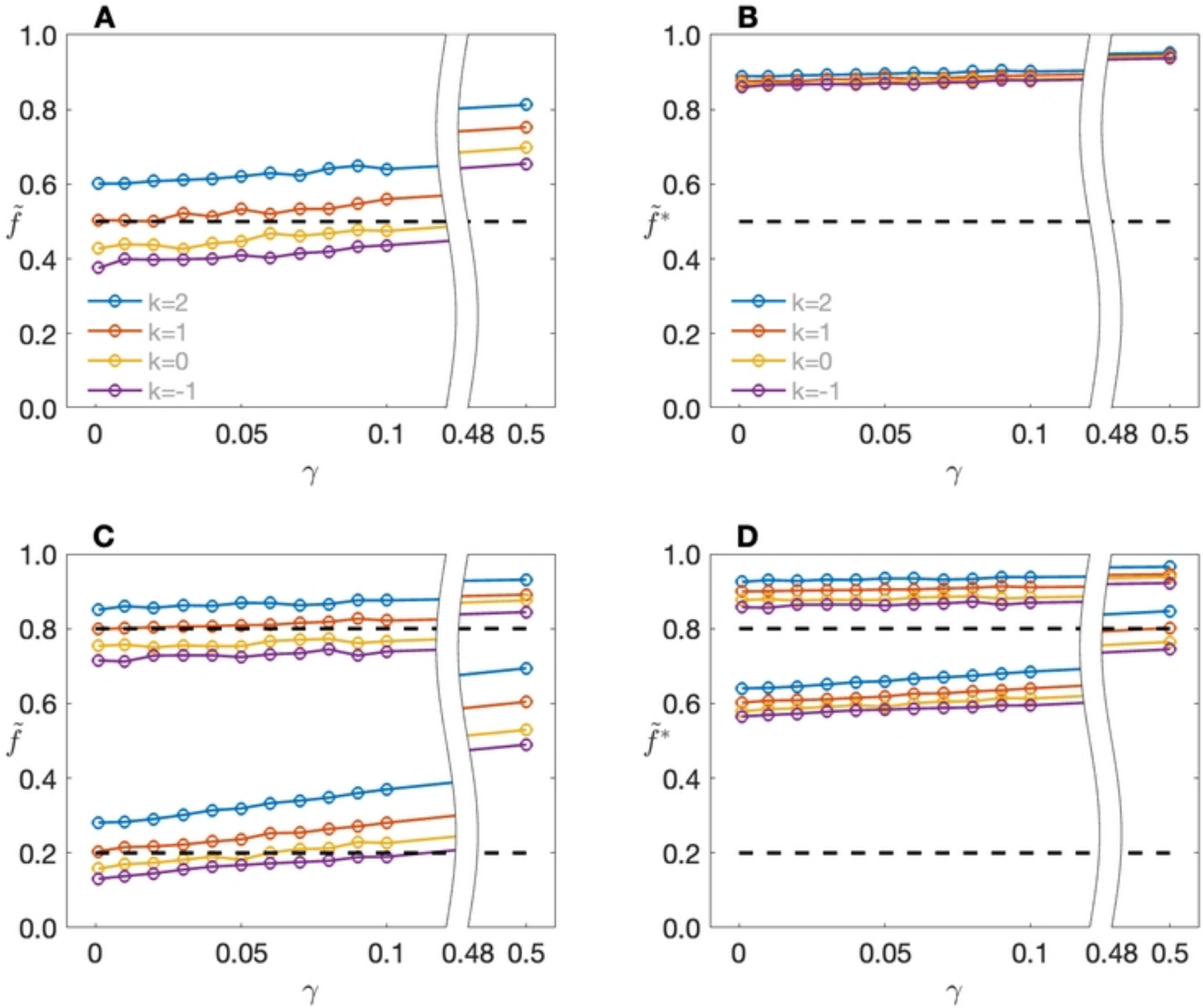
A
B
C
D
1.0
0.8
0.6
0.4
0.2
0.0
$\tilde{f}$
$\tilde{f}^*$
k=2
k=1
k=0
k=-1
0
0.05
0.1
0.48
0.5
γ

Figure 2

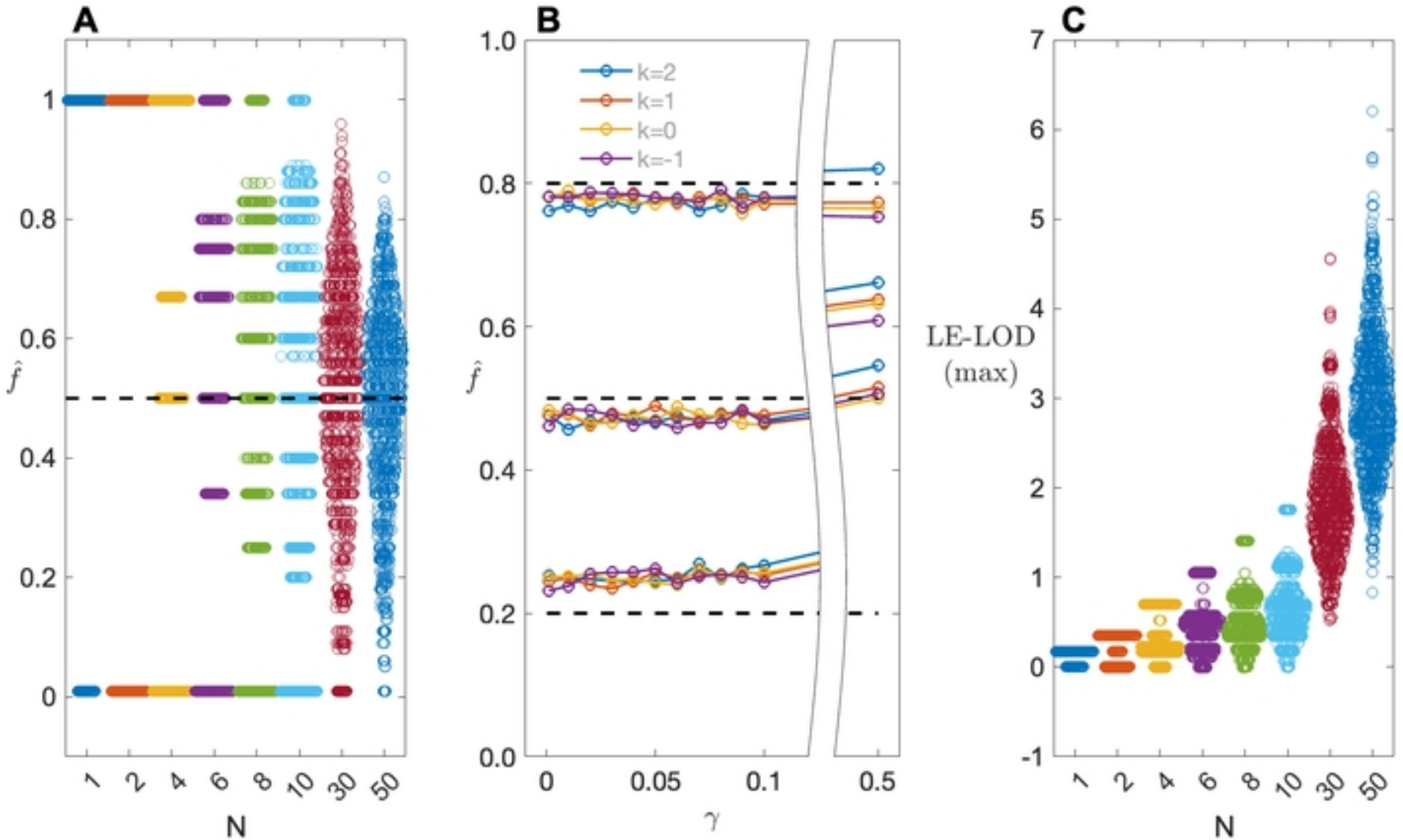
A
B
C
k=2
k=1
k=0
k=-1
$\hat{f}$
γ
N
LE-LOD
(max)
1
0.8
0.6
0.4
0.2
0
1.0
0.0
0.05
0.1
0.5
7
6
5
4
3
2
-1
1
2
4
6
8
10
30
50

Figure 3