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2 **Title:** The effect of ascertainment on penetrance estimates for rare variants: implications for
3 establishing pathogenicity and for genetic counselling

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5 **Running title:** Ascertainment effects on penetrance estimates

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24 **Abstract:**

25 Next-generation sequencing has led to an explosion of genetic findings for many rare diseases.
26 However, most of the variants identified are very rare and were identified in small pedigrees,
27 which creates challenges in terms of penetrance estimation and translation into genetic
28 counselling in the setting of cascade testing. We use simulations to show that for a rare
29 (dominant) disorder where a variant is identified in a small number of small pedigrees, the
30 penetrance estimate can both have large uncertainty and be drastically inflated, due to underlying
31 ascertainment bias. We have developed PenEst, an app that allows users to investigate the
32 phenomenon across ranges of parameter settings. We also illustrate robust ascertainment
33 corrections via the LOD score, and recommend a LOD-based approach to assessing
34 pathogenicity of rare variants in the presence of reduced penetrance.

35 Next-generation sequencing has led to an explosion in the number of genetic findings for many
36 rare diseases. For certain types of rare coding variants (e.g. missense, or protein truncating), if
37 the variant is sufficiently rare and has bioinformatic predictions that are severe, current
38 algorithms result in it being classified as pathogenic (1). However, the analysis of large-scale
39 sequencing from cohorts, such as ExAC (2), gnomAD (3), and the UK Biobank (4), has shown
40 that many such variants may often lack clinically significant impact. For example, ExAC
41 estimated that individuals from population cohorts carried a mean of 53 variants previously
42 thought to be sufficient causes of Mendelian diseases. Additionally, 88% of such variants had
43 MAF>1%, implying that they are likely not sufficient causes. This may indicate that such
44 variants are not causally related to disease, or perhaps, that they are causally related but with
45 reduced penetrance.

46
47 Penetrance plays an important role in understanding disease pathology, in the appropriate
48 classification of pathogenic variants, and perhaps above all in the context of genetic counseling.
49 However, most of the variants reported to date have been very rare and identified in small sets of
50 unrelated individuals (sometimes just one) or small pedigrees. Penetrance cannot be estimated
51 from a single case, or a single parent-offspring trio presenting with a *de novo* mutation in the
52 offspring. But even with multiple cases or families, determination of the penetrance can present
53 challenges. Here we focus on one such challenge: ascertainment.

54
55 Typically a variant of interest is first identified in one individual with a given phenotype.
56 Investigators may then sequence either additional relatives of the individual, or additional
57 individuals or families presenting with the same or closely related phenotypes, with the goal of

58 bolstering the case for pathogenicity. Thus, ascertainment of individuals to be sequenced
59 typically proceeds in stages. The precise ascertainment process used to enrol individuals and/or
60 families is usually at least to some extent unsystematic, and may vary between families.
61 Ascertainment is therefore challenging to model when attempting to estimate the penetrance of a
62 variant.
63
64 One situation in which ascertainment can be easily handled is “single” ascertainment, in which
65 the probability of an affected individual being ascertained is proportional to the number of
66 affected individuals in the family (5). In fact, much of the literature on inferring pathogenicity or
67 estimating penetrance tends to assume single ascertainment, e.g., (6), where ascertainment is
68 addressed by conditioning on “the proband,” a procedure which is strictly correct only under true
69 single ascertainment. While it is true that the typical study ascertains families through one
70 individual who may be designated as the single “proband”, this does not ensure that the study
71 meets the proportionality requirement of single ascertainment. This requirement would be
72 violated, e.g., if families with four affected members were more than twice as likely to be
73 recruited as families with just two; or, if the probability of a second sibling being ascertained
74 were dependent on the ascertainment status of the first. And in general, if either (i) ascertainment
75 is not truly single, or (ii) even if it is, if an appropriate ascertainment correction is not
76 incorporated into the estimation method, then penetrance estimates will be biased. Here we
77 consider the magnitude of that bias, across a range of plausible ascertainment models and
78 varying amounts of available data.
79
80 We focus here on sibship data. The impact of ascertainment for more complex pedigrees can be
81 approximated by considering large sibship sizes. For simplicity, we assume all parents are

82 phenotypically and genotypically unknown; including parental information does not
83 substantively affect results. We assume a very rare variant of interest (VOI), and an autosomal
84 dominant disease D . Let a qualifying individual (QI) be anyone who is both heterozygous (HET)
85 for the VOI and also affected (AFF) with D . Let r be the number of QI sibs within a family, and
86 let t be the number of AFF sibs regardless of VOI genotype. We also assume that, regardless of
87 VOI status, an individual might develop D due to other factors, which might be genetic
88 (involving one or more VOIs at other loci or other variants within the same gene) and/or
89 environmental (e.g., due to infections). Let γ be the combined penetrance across all causes other
90 than the VOI under study. Since we assume the VOI is very rare, γ is effectively the population
91 prevalence of D .

92
93 In order to consider a range of plausible ascertainment scenarios, we employ the general family-
94 based k -model of ascertainment (7). In its simplest form, this model stipulates that the probability
95 that a family is ascertained is proportional to r^k , where k controls the model. For example, when
96 $k = 1$, the probability of ascertainment is strictly proportional to r : this is equivalent to classical
97 “single ascertainment”. Similarly, when $k = 0$, so that every family with $r \geq 1$ is ascertained, this
98 model is equivalent to classical “complete” or “truncate” ascertainment. We generalize this
99 model in two ways. First, we assume that ascertainment requires $r \geq 1$, that is, every ascertained
100 family contains at least one QI, but we allow that there may be additional preferential
101 ascertainment of families based on t alone, that is, that investigators may preferentially ascertain
102 families with more affected individuals without knowing (or prior to knowing) the VOI status of
103 those additional individuals. Second, we allow that even an individual carrying the VOI may

104 develop disease due to any other independent causes at work in the general population. With
105 these two extensions in mind, our ascertainment model becomes

$$106 \quad P[\text{sibship is ascertained} \mid r, t] = c(r^k + t); \text{ for } r \geq 1, \text{ and } 0 \text{ otherwise}$$

107 where c is a normalizing constant.

108

109 Let f be the attributable penetrance, or the penetrance due to the VOI for HET individuals. (Note
110 that when $\gamma > 0$, $\beta = P[\text{AFF}|\text{HET}] = \gamma + f - \gamma f$. However, we focus here on estimation of f itself
111 rather than β .) In what follows, we estimate f in two ways:

112 (i) \tilde{f} is obtained by counting the proportion of AFF individuals among all HET
113 individuals in the data set, after dropping one QI individual per family, that is,
114 applying the correction for single ascertainment;

115 (ii) \tilde{f}^* is obtained by counting the proportion of AFF individuals among all HET
116 individuals in the data set, that is, without applying any ascertainment correction.

117

118 \tilde{f}^* is a naïve estimate, which would be correct if the families were not ascertained based on
119 either phenotype or genotype. It is, however, clearly incorrect under any of our ascertainment
120 models. Our interest in this estimate is to establish how biased it becomes under various
121 ascertainment scenarios. \tilde{f} by contrast, does apply the frequently employed single ascertainment
122 correction, and again, our interest in \tilde{f} is to establish how biased it will be under ascertainment
123 scenarios other than single ascertainment. Expected values of \tilde{f} and \tilde{f}^* were obtained via
124 simulation, by averaging each estimate's value across 1,000 replicates per generating condition,
125 and standard errors were obtained by averaging the standard deviation of each estimate across
126 those same 1,000 replicates. (While the expected values are easily calculated analytically, the

127 standard errors are not.) All simulations and calculations were done in MATLAB
128 (2021.9.10.0.1739362 (R2021a), Natick, Massachusetts: The MathWorks Inc.).

129
130 Let s be the number of siblings in a family, and let N be the number of s -sized sibships in a
131 dataset. Fig 1 shows results for true single ascertainment ($k=1$), for $s = 2$, as a function of sample
132 size N . Here we assume that the true value of $f=0.5$. As can be seen, in this case, the mean of $\tilde{f} =$
133 0.5 , the generating value, as expected. But using \tilde{f}^* the estimates are seriously upwardly biased
134 in all data sets, regardless of N . Note that because each sibship contains at least one QI, by
135 stipulation, the minimum value of \tilde{f}^* is 0.50 .

136
137 Note too that even the correct estimate \tilde{f} shows considerable sampling variability. For instance,
138 with $N=10$, \tilde{f} will be $>70\%$ or $<30\%$ in approximately 40% of all data sets when $f=50\%$. This
139 variability remains appreciable even for $N=50$.

140
141 For ascertainment models other than single, overall variability remains similar to what is shown
142 in Fig 1, but even \tilde{f} tends to be biased, with mean $\tilde{f} = 0.60, 0.50, 0.43$ and 0.38 for $k = 2, 1, 0$
143 and -1 , respectively. In all cases, the uncorrected \tilde{f}^* will return even more biased estimates, with
144 mean $\tilde{f}^* = 0.89, 0.88, 0.87$ and 0.86 , for $k = 2, 1, 0$ and -1 , respectively.

145
146 Fig 2 shows the impact of the population prevalence γ on average penetrance estimates.
147 Focusing first on single ascertainment ($k=1$) and $\tilde{f}=0.5$, we can see that regardless of k , the
148 expected value of \tilde{f} is relatively independent of γ until γ becomes quite high. Note that for $f =$
149 0.5 and $\gamma = 0.5$, the actual probability that a VOI carrier is affected under our generating model

150 is $0.5 + 0.5 - (0.5)(0.5) = 0.75$, which is in line with the estimates returned by \tilde{f} . \tilde{f}^* might be
151 said to be even more robust to γ , although this is because in this case \tilde{f}^* is already close to the
152 top of the scale for $\gamma=0$. Moreover, \tilde{f}^* appears not only robust to γ , but also to f itself, with
153 estimates $>70\%$ even for $f=0.05$, and $>80\%$ for $f=0.05$ when $\gamma=0.5$. These patterns repeat for
154 different values of k , with visible impact only on the magnitude of the bias for any given (f, γ)
155 combination. Ascertainment effects will be reduced as s increases. Users who are interested in
156 investigating penetrance estimates for other ascertainment models, other combinations of
157 parameter values or other sibship sizes are encouraged to download the PenEst app:
158 <https://github.com/MathematicalMedicine/PenetranceEstimator>.

159
160 In general, our simulations show that under unsystematic ascertainment schemes, or in cases
161 where appropriate ascertainment corrections are not included in the estimation procedure, there
162 is a high risk of over-estimating the penetrance of any given VOI. This finding is consonant with,
163 and may in large part explain, reports for specific variants. For example, multiple coding variants
164 in *PRNP* had been reported to cause rare dominant monogenic neurodegenerative disease, but
165 there was a 30-fold higher prevalence of variants previously suggested to be causal in this gene
166 in ExAC compared to the expected frequency calculated from the estimated prevalence of the
167 disorder (8). Specifically for three variants the lifetime risk of developing disease was $<10\%$.
168 Similarly, GWAS array data from the UK Biobank were used to estimate pathogenicity,
169 penetrance, and expressivity of putative disease-causing rare variants ($MAF < 1\%$) that were
170 directly genotyped and had good quality (9). Focused on maturity-onset diabetes of the young
171 and developmental disorders, many specific variants were found for which the penetrance --
172 estimated either in families ascertained for the presence of the VOI or in disease cohorts -- was

173 much higher than that obtained from a population-based cohort. These observations have
174 implications for genetic counselling, including the recommendation of invasive screening
175 procedures and administration of preventative treatment.

176
177 Some approaches to the interpretation of rare coding variants assume either full or high
178 penetrance (10), for the sake of simplicity. Extensive criteria have been proposed to claim a
179 causal relationship between variants and disease, and authors have urged caution in presuming
180 full penetrance for pathogenic variants (11). But in practice, penetrance remains an important
181 factor in assessing pathogenicity. For instance, the ACMGG/AMP joint consensus
182 recommendations (1) warns against ignoring the possibility of reduced penetrance in establishing
183 segregation of a VOI with a phenotype, but also instructs that “lack of segregation...provides
184 strong evidence against pathogenicity.” (p. 15) And in practice, many laboratories will rule out
185 candidate VOIs when they are found among unaffected relatives. Particularly in the absence of a
186 rigorous and accurate estimate of the actual penetrance, this complicates the use of segregation
187 information in assessments of pathogenicity.

188
189 We close by noting that there is one essentially “ascertainment assumption free” (12) method for
190 estimating the penetrance, viz., by conditioning on all of the phenotypic data. This is the
191 ascertainment correction implicit in the usual LOD score (13-15), and also the LOD score
192 allowing for linkage disequilibrium or LD-LOD (6, 16, 17), and in principle any program that
193 allows calculation of the LOD score will support this method. As in Thompson (6) the
194 calculation is done here assigning the VOI (which plays the role of the “marker”) and the disease
195 allele the same (rare) frequency (we have used 0.001 in the simulations), assuming complete

196 linkage disequilibrium between the two ($D' = 1$), and also assuming 0 recombination between the
197 marker and the disease allele. Free parameters in the model are then the three penetrances; in our
198 calculations we also include the admixture parameter α of Smith (18), representing the
199 probability that any given family is of the “linked” type, which adds robustness when phenocopy
200 levels are high. Maximizing the LD-LOD over the free parameters gives us the LD-MOD, which
201 occurs at the maximum likelihood estimate (m.l.e.) of \hat{f} of f (12-15).

202
203 Fig. 3 shows results corresponding to the simulations in Fig 1A and Fig 2A, C. As can be seen, \hat{f}
204 behaves very much like \tilde{f} when $k = 1$ (Fig 3A), but it retains almost complete robustness to
205 ascertainment, and also to γ at least until γ is quite large (Fig 3B). (As with \tilde{f} , as γ gets very
206 large, \hat{f} covers both cases due to the VOI and also cases among variant carriers due to other
207 causes.) Comparing Fig 3A with Fig 1A, \hat{f} shows slightly greater sampling variability than \tilde{f} ;
208 this is due to the inherent ascertainment correction built in to \hat{f} . The slight but systematic over-
209 or under-estimation of f seen in Fig 3B is due to the small sample size; as N increases $\hat{f} \rightarrow$
210 f (results not shown). However, in small samples the upward bias can be appreciable particularly
211 when f is small; e.g., when $f = 0.05$ ($\gamma = 0$), for $N = 20$, the expected value of $\hat{f} = 0.165$.

212
213 Note, however, that while maximizing the LD-LOD is a highly ascertainment-robust method for
214 estimating f , the LD-MOD itself is not a good statistic for representing the strength of evidence
215 for co-segregation, because it is not additionally conditioned on ascertainment through the VOI.
216 However, once we ascertain so as to require the VOI to be present in the family, there is no
217 remaining LD information in the sibship, since LD information is conveyed entirely by the
218 marker allele frequencies in the parents. Therefore, we recommend using the ordinary (linkage

219 equilibrium) LOD, or LE-LOD, for assessing strength of evidence for co-segregation. Because
220 maximizing the LE-LOD itself will not return true m.l.e.s of f under the LD model, we
221 recommend evaluating the LE-LOD at the maximizing model obtained from the LD-MOD, for a
222 statistic we annotate as LE-LOD(max). (This maximization procedure is not inherently
223 inflationary; see Supplemental Results (A). Thompson et al. (6) proposed a form of Bayes factor
224 for assessing evidence for co-segregation of the VOI with disease; see the Supplemental Results
225 (B) for some comparisons between their Bayes factor and LE-LOD(max).) Fig 3(D) shows the
226 distribution of the LE-MOD(max), for the same data shown in Fig 1. (Here parents are treated as
227 genotypically known but phenotypically unknown.) As expected, based on just a few 2-child
228 sibships, evidence of co-segregation of the VOI with disease is quite weak. It requires at least N
229 = 30 2-child families before there is a reasonable chance of obtaining a substantial LE-
230 LOD(max).

231

232 **Declaration of interests**

233 The authors declare no competing interests

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236 Valentine-Cooper for creation of the PenEst app.

237

238 **Web resources**

239 PenEst app : <https://github.com/MathematicalMedicine/PenetranceEstimator/>

240

241 **Data and code availability**

242 <https://github.com/MathematicalMedicine/PenetranceEstimator/>

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286 **Figure 1.** Swarm plots showing sampling distributions of penetrance estimates as a function of
287 number of families N .

288 Distributions of (A) \tilde{f} and (B) \tilde{f}^* are shown for simulations of 1000 replicates, with true
289 penetrance $f=0.5$. The number of sibs per family, $s=2$; phenocopy rate, $\gamma=0$. Users interested in
290 varying the parameters can use the PenEst app.

291

292 **Figure 2.** Expected values of penetrance estimates as a function of population prevalence γ and
293 ascertainment parameter k .

294 Top row: expected values of (A) \tilde{f} and (B) \tilde{f}^* when the true penetrance $f=0.5$. Bottom row:

295 expected values of (C) \tilde{f} and (D) \tilde{f}^* when $f=0.2$ (lower line sets) or $f=0.8$ (upper line sets). The

296 number of sibs per family, $s=2$. Users interested in varying the parameters can use the PenEst

297 app.

298

299 **Figure 3.** (A) Swarm plots showing sampling distributions of \hat{f} , as obtained from maximizing

300 the LD-LOD, as a function of number of families N ; (B) Expected values of \hat{f} as a function of

301 population prevalence γ and ascertainment parameter k , for $f = 0.2, 0.5$ and 0.8 , reading from

302 bottom to top of the plot, respectively; (C) Swarm plots showing the distribution of LE-

303 LOD(max) as a function of N . Data are the same as used to generate Figures 1 and 2,

304 respectively. All calculations were done using KELVIN (19).





