Repeated passive visual experience modulates spontaneous and novelty-evoked neural activity.

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Abstract

Familiarity creates subjective memory of repeated passive innocuous experiences, reduces neural and behavioral responsiveness to those experiences, and enhances novelty detection. The neural correlates of the internal model of familiarity and the cellular mechanisms of enhanced novelty detection following multi-day repeated passive experience remain to be better understood. Using the mouse visual cortex as a model system, we test how the repeated passive experience of an orientation-grating stimulus for multiple days alters spontaneous, and non-familiar stimuli evoked neural activity in neurons tuned to familiar or non-familiar stimuli. We found that familiarity elicits stimulus competition such that stimulus selectivity reduces in neurons tuned to the familiar stimulus, whereas it increases in those tuned to non-familiar stimuli. Consistently, neurons tuned to non-familiar stimuli dominate local functional connectivity. Furthermore, responsiveness to natural images, which consists of familiar and non-familiar orientations, increases subtly in neurons that exhibit stimulus competition. We also show the similarity between familiar grating stimulus-evoked and spontaneous activity increases, indicative of an internal model of altered experience.

Introduction

Long-term repeated experience of an innocuous stimulus leads to behavioral habituation and a subjective memory of the stimulus. Habituation enables organisms to respond to novel or behaviorally relevant stimuli^{1,2}. Repeated exposure to an orientation-grating visual stimulus not associated with reward or punishment over multiple days selectively reduces behavioral exploration of that orientation in mice^{3,4}. However, at the level of neural activity, the effect of the multi-day repeated experience of orientation grating stimuli has yielded contradictory findings. Studies have found a familiar stimulus-selective increase in local field potential and altered oscillations⁵⁻⁹. Single-unit recordings have found an increase in peak responsiveness and a decrease in average responsiveness to familiar grating stimuli^{3,10}. Calcium imaging studies have yielded mixed results with no change¹¹, increased¹², or decreased average responsiveness¹³⁻¹⁵ to familiar grating stimuli. Our recent study found that familiarity with a grating stimulus reduces average neural responsiveness to that stimulus, consistent with some of the earlier studies⁴. While the reduction in average responsiveness to the familiar stimulus could explain behavioral habituation, how repeated passive non-natural stimulus experience over multiple days alters spontaneous activity and enhance novelty detection at the cellular level is unclear.

Functionally distinct inputs, including those tuned to different orientations, are intermingled on dendrites of visual cortical neurons¹⁶⁻¹⁹. Therefore, reduced responsiveness to the familiar stimulus could trigger competition and enhance responsiveness to novel stimuli. Consistently, short-term visual adaptation to an oriented grating or Gabor stimulus shifts the tuning curve toward or away from the adapted orientation depending on various stimulus features and enhances novelty detection²⁰⁻²⁹. Similarly, the multi-day repeated passive experience of orientation grating stimulus has been shown to increase orientation selectivity¹⁰, whereas other

studies found a lack of orientation tuning curve shift¹³⁻¹⁵. Whether multi-day passive experience enhances long-term (>12 hours) responsiveness to non-familiar stimuli and whether it involves competition between familiar and non-familiar stimuli remains unclear.

Expectations or subjective memory of the familiar stimulus may arise from the stored internal model of the familiar stimulus. Spatiotemporal characteristics of spontaneous activity, which occurs without stimulus, resemble evoked activity and may serve as an internal model of an animal's sensory environment³⁰⁻⁴². However, recent studies in mice and zebrafish show that spontaneous and visually evoked activity are dissimilar, further diverging over development ⁴³⁻⁴⁵. If spontaneous activity serves as an internal model of sensory experience, the repeated passive sensory experience should alter the spontaneous activity structure to resemble the activity evoked by this familiar experience in adulthood.

We show that in layer 2/3 of the mouse visual cortex, familiarity with a grating stimulus enables competitive plasticity between repeatedly experienced (familiar) and once experienced (non-familiar) orientations leading to higher responsiveness to non-familiar stimuli than when these stimuli were novel. Altered responsiveness to familiar and non-familiar stimuli changes stimulus selectivity differentially dependent on the stimulus preference of neurons. Interestingly, the fraction of neurons activated by natural images increased subtly, and the neurons that became more responsive to natural images exhibited higher stimulus competition than those that became less responsive. Furthermore, spontaneous and familiar stimulus-evoked activity patterns became more similar and shared the same neural space. Based on these observations, we speculate that superficial visual cortical neurons store an internal model of altered experience through changes in spontaneous activity and detect deviation from familiarity through changes in stimulus selectivity elicited by stimulus competition.

Results

Repeated passive exposure to specific orientation reduces neural responsiveness selectively to the familiar stimulus.

We imaged spontaneous and visually evoked calcium transients in the awake head and body-restrained GCaMP6s transgenic mice habituated to the imaging apparatus (Fig. 1A, B). Visual stimuli consisted of eight trials of sinusoidal phase-reversing orientation gratings and a set of ten natural images (Fig. 1B). The phase-reversing orientations do not drift, thus 0°, 45°, 90°, and 135° are the same as 180°, 225°, 270°, and 315° (hereafter referred to as 0°,45°,90°, and 135°, respectively). For the next eight days, head-fixed mice passively experienced two sessions of 60 seconds of the gray screen followed by five blocks of 100 seconds of 45° phase reversing grating stimulus with 30 seconds of the gray screen between blocks. The two sessions were separated by ~1-2 hours. We refer to the passive repeated exposure to the non-natural grating stimulus for eight days as an "altered experience." The next day after the altered experience the same neurons were imaged, similar to the first imaging session (Fig. 1B-F). 1375 neurons from ten mice (7 males and 3 females) were identified from both sessions combined and ranked based on the quality of soma in the anatomical and correlation maps generated by Suite2p⁴⁶. 561 neurons with clearly visible somata in both sessions were included for further analyses.

To measure visually evoked calcium transients, we compared the matched neurons across imaging sessions pre (S1) and post (S2) altered experience. We found that the trial-averaged mean dF/F0 over the 3-second stimulus period from all neurons matched between sessions selectively reduced to 45° (familiar) but not to other grating or natural image stimuli (non-familiar) (Fig. 1G). Familiar stimulus-specific reduction in population response indicates that plasticity is not an artifact associated with the animal's behavioral state.

Since the population's mean dF/F0 to all grating and natural image stimuli was close to 4%, we classified neurons with greater or less than 4% dF/F0 as high and low-responsive neurons, respectively. Interestingly, the extent of change in neural activity (dF/F0 on S2 – dF/F0 on S1 for each neuron (ΔdF/F0)) following the altered experience depends on the initial activity level on S1. Highly responsive neurons were more dynamic, with large increases and decreases from their initial dF/F0 compared to low-responsive neurons (Fig. 1H). In contrast to non-familiar stimuli, highly responsive neurons to the familiar 45° stimulus mostly showed a reduction in dF/F0 (Fig. 1H).

Familiarity alters the tuning preference of low but not high-responsive neurons

Due to the intermingling of synapses with multiple orientation preferences, even in neurons selectively tuned to one of the orientations, a reduction in responsiveness to familiar orientation is likely to increase the responsiveness to non-familiar orientations. Such competition underlies short-term adaptation to orientation grating responsiveness²⁹. However, whether long-term (>12 hours) changes to preferred orientation also happen following multi-day grating stimulus exposure remains unclear. First, we classified neurons as responsive to a stimulus if the trial averaged mean dF/F0 is greater than two standard deviations above the mean trial averaged gray screen period preceding the stimulus. We next identified neurons that were classified as responsive and had a good tuning curve fit in both sessions (286/561 neurons; Fig. 2A). We binned these well-fit neurons by their preferred orientation into four groups centered on the four stimuli (0° - 22.5° and >157.5°-180° as 0°, >22.5°- 67.5° as 45°, >67.5°-112.5° as 90°, and >112.5°-157.5 as 135° tuned neurons). We found a modest reduction in neurons whose preferred orientation was closer to the trained orientation (~27, 23, 26, and 24% of neurons were tuned to 0°,45°,90°, and 135°, respectively, in S1 but changed to 31, 15, 28, and 26% for these

orientations; Fig. 2B, top). However, when restricting this comparison to highly responsive neurons, the proportion of neurons tuned to different orientations did not differ (Fig. 2B, middle). In contrast, the distribution of tuning preferences differed for neurons that remained low responsive in both sessions (31, 23, 24, and 23% for 0°,45°,90°, and 135°, respectively in S1 and 36, 11, 30, and 22% for these orientations in S2; Fig. 2B, bottom). These results indicate that altered experience shifts the tuning preference of low but not highly responsive neurons. We next compared the fraction of neurons that retained their original tuning preference after the altered experience. We found that more neurons retained their stimulus preference if the stimulus was not made familiar. 73% of neurons maintained their tuning preference within their bin if the stimulus did not become familiar, compared to 56% if it became familiar (Fig. 2C, D, middle). Consistent with our observation that the distribution of tuning preferences did not change for highly responsive neurons, we found that a comparable fraction of highly responsive neurons retained their original stimulus preference regardless of familiarity (82% of neurons preferring non-familiar and 71% of neurons preferring familiar stimulus retained their tuning preference). In contrast, 38% of low-responsive neurons retained their preferred orientation when the stimulus became familiar, whereas 62% retained it when it remained non-familiar (Fig. 2C, D, bottom). These results show that familiarity elicits competition between stimuli, so non-familiar stimulus responsiveness becomes more dominant in low-responsive neurons.

Familiarity differentially alters the tuning width of highly responsive neurons based on their tuning preference.

To test whether stimulus competition occurs in neurons that remain highly responsive to grating stimuli following altered experience, we compared the tuning width of these neurons (Fig. 3A). Decreased responsiveness to the familiar stimulus or increased responsiveness to non-familiar

stimulus would broaden and sharpen the tuning width of neurons tuned to familiar or the same non-familiar stimulus, respectively. We found that the median full width at half-maximum was ~ 59°, consistent with the 28-29° half-width at half-maximum reported previously⁴⁷. Altered experience differentially influenced the tuning width of neurons preferring familiar and non-familiar stimuli (Fig. 3B-E). Neurons tuned to 0° showed a non-significant left shift of tuning width distribution, and 135° (orthogonal to familiar stimulus) had no change (Fig. 3B, E). In contrast, 90° tuned neurons showed a significant left shift of frequency distribution and reduced average tuning width, whereas familiar stimulus tuned neurons showed a right shift (not significant) and a subtle increase in average tuning width (Fig. 3C, D).

To test whether the sharpening and broadening of the tuning curve is a result of stimulus competition, we compared the average ΔdF/F0 of these neurons. We found that the broadening of the tuning curve of familiar stimulus preferring neurons is solely due to a reduction in familiar stimulus responsiveness. In contrast, the sharpening of the tuning curve of non-familiar (90°) neurons is due to both a subtle increase in 90° response and a slight reduction in 45° response, indicative of stimulus competition (Fig. 3C, D). These results show that competition in highly responsive neurons primarily occurs in non-familiar stimulus preferring neurons.

Neurons tuned to non-familiar stimuli dominate functional connectivity

To assess whether stimulus competition elicited by altered experience changes the functional connectivity of neurons, we identified neurons with significant coactivity during the entire imaging period consisting of multiple visual stimuli (Fig. 4A). Two neurons are functionally connected if the number of their coactive imaging frames is >95% of the cumulative distribution of their coactivity obtained by 1000 random circular shifts of the imaging frames⁴⁸. Though there was a significant increase in the average number of functionally connected neurons (node

degrees) to 0° tuned neurons, we did not observe changes to the node degree of neurons tuned to other orientations, including the familiar orientation (Fig. 4B). This was surprising because we previously found a reduction in node degrees following familiarity⁴. However, in that study, functional connectivity was assessed within the familiar stimulus period whereas our current analysis encompasses imaging frames spanning multiple stimuli. Therefore, we could detect functional connectivity associated with non-familiar and familiar stimuli. To test whether functional connectivity is reorganized despite no change in average functional connectivity, we identified the tuning preferences of the functionally connected partners. We found that >30% of node degrees are between similarly tuned neurons. Altered experience enhances the fraction of node degrees with non-familiar stimuli tuned neurons (0° and 90°) for both familiar and non-familiar stimulus preferring neurons (Fig. 4C). 0° and 90° neurons show ~50% increase in their functional connectivity following repeated 45° passive experience. These results indicate that stimulus competition associated with familiarity alters functional connectivity favoring the dominance of non-familiar stimulus tuned neurons.

Responsiveness to natural images is altered following familiarity with a grating stimulus Changes to orientation grating stimulus responsiveness could influence neural responsiveness to natural images, which consist of multiple orientations. To test this possibility, we compared the fraction of neurons responsive to natural images before and after the altered experience with the 45° stimulus. We found that the fraction of non-responsive neurons reduced and transitioned to low responsiveness to the set of ten natural images tested (Fig. 5A-C). Interestingly, the neurons that became responsive (gain) to these natural images also showed increased responsiveness to non-familiar grating stimuli, consisting of cardinal orientations, and decreased responsiveness to the familiar grating stimulus. In contrast, neurons that became non-responsive (loss) after altered

experience or those that remained stably responsive had no or minor changes to non-familiar stimuli responsiveness (Fig. 5D). These results suggest that an increase in responsiveness to 0° and 90°, presumably due to stimulus competition associated with familiarity, alters responsiveness to natural image stimuli.

Spontaneous activity patterns reflect the altered experience

We next compared whether the spontaneous activity is modulated by altered experience in neurons that retained high responsiveness to stimuli (Fig. 6A). For this analysis, imaged neurons were independently matched between the spontaneous period and evoked activity images independently for pre- and post-altered experience imaging sessions. Spontaneous activity was quantified as the average dF/F0 obtained from every 3 seconds bin (to match the average stimulus duration) over the ~237 seconds of imaging in darkness. We found that the average dF/F0 of spontaneous activity did not change in familiar or non-familiar preferring neurons following the altered experience (Fig. 6B). Though the average magnitude of spontaneous activity may not be significantly different, the population response similarity with the stimulus-evoked response could change following an altered experience. We calculated the similarity ratio to assess changes to similarity (described in Methods) between spontaneous and stimulus-evoked activity in highly responsive neurons. An increase in the ratio above one indicates increased similarity between spontaneous and evoked activity following altered experience. The similarity between spontaneous and 0°, 45°, 90°, 135°, and gray screen evoked responses were 45%, 27%, 31%, 34%, and 38%, respectively on S1 and it changed to 43%, 38%, 27%, 36%, and 55% on S2 for the same stimuli (Fig. 6C). When restricted this analysis to neurons based on their tuning preferences, the similarity of spontaneous and preferred stimulusevoked activity increased by three-fold for familiar stimulus preferring neurons (27% to 84%)

similarity). Spontaneous activity and the preferred-stimulus evoked response were greater for horizontal orientation before the altered experience (0°; 63% similarity) but moderately reduced to 45% following the altered experience. The similarity of spontaneous activity and preferred stimulus evoked response in neurons tuned to 90° (38% on S1) and 135° (36% on S1) stayed roughly the same (Fig. 6D).

We also measured the overlap of stimulus-evoked and spontaneous activity for individual three second time bins. We projected the neural response of stimulus-evoked activity for individual stimulus cycles onto the observed spontaneous activity neural space (described in Methods; Fig. 6E). The overlap of spontaneous activity with each of the four grating stimuli evoked responses (obtained from all responsive neurons) was evenly distributed with a slight propensity for 0° on the first day of imaging. After the altered experience, however, the spontaneous neural activity increased its overlap with that of the familiar 45° stimulus with a corresponding decrease in overlap with the 0° and 90° stimuli (Fig. 6F). Thus, neural activity more closely mimicked the response pattern for the familiar grating pattern after repeated presentations, even when no stimuli were present.

Discussion

Consistent with a vast body of literature on habituation, we show that repeated orientation-grating stimulus experience reduces neural responsiveness to the familiar stimulus in the mouse visual cortex. Neurons in the visual cortex receive functionally diverse inputs, with neighboring synapses tuned to different orientations^{16,19}. Long-term reduction in responsiveness to one set of inputs evoked by familiarity is likely to elicit a compensatory increase in other inputs, perhaps to maintain neuronal activity homeostasis. Competition between stimuli is well established using different paradigms, such as monocular deprivation^{49,50}, cross-modal plasticity

⁵¹⁻⁵⁴, and sensory adaptation^{20,25,26}. In contrast, studies that found reduced responsiveness to the familiar stimulus following the multi-day passive experience of an orientation grating stimulus did not report altered tuning preference. Here, we show that responsiveness to non-familiar stimuli is higher than when these stimuli were novel in low-responsive neurons tuned to the familiar stimulus and high-responsive neurons tuned to a non-familiar stimulus, indicative of stimulus competition in these neurons. The greater vulnerability of lower responsive neurons to familiarity-evoked stimulus competition could be because weaker synapses are more likely to undergo potentiation or depression^{55,56}. Alternatively, subtle changes to inhibition that suppress familiar inputs and disinhibit non-familiar inputs could have a higher impact on lower responsive neurons. Depression and potentiation of familiar and non-familiar stimulus-responsive synapses in lower responsive neurons could lead to fewer of them retaining the familiar stimulus as the preferred stimulus. Similar plasticity mechanisms, albeit to a lesser extent, in highly responsive neurons may underlie subtle sharpening of tuning width of neurons tuned to non-familiar stimuli. Due to the slightly lower sampling of orientation tuning space in this study, we cannot rule out that the increased tuning width is due to a subtle shift in tuning preference.

What purpose could stimulus competition serve? Habituation allows resources to be spent on detecting novel or behaviorally relevant stimuli. Models of habituation that involves synaptic depression or enhanced inhibition could explain a reduction in familiar stimulus responsiveness but do not explain how novel stimuli are preferentially detected and how a reduction in neural activity generates the knowledge of familiarity^{1,57}. Predictive coding-based models suggest that internal model of visual experience allow for the selective transmission of sensory information that deviates from the expectation⁵⁸⁻⁶¹. The internal model could serve as the knowledge of familiarity, and their inhibitory influence on familiar stimulus responsiveness may contribute to

novelty detection. Consistently, deviation from or memory of expected sensory experience has been shown to modulate neuronal activity in the visual cortex^{30,38,59,62-69}.

Similarly, spontaneous activity also influences stimulus-evoked activity^{70,71}. Our findings indicate that internal model of familiarity and enhancement of novel stimuli responsiveness may occur independently. Spontaneous activity structure more closely matches the observed response to familiar stimuli. Furthermore, the increase in similarity is more prominent in familiar stimulus preferring neurons. These results suggest that internal model or expectation of familiarity may arise from spontaneous activity patterns. The influence of spontaneous activity on evoked activity may contribute to familiarity^{11,72}. Our findings indicate that the suppression of familiar stimulus responsiveness occurs independent of changes in spontaneous activity. We show that the average amplitude of spontaneous activity is not altered by familiarity, and the change in the similarity of prestimulus gray screen elicited and spontaneous activity is not significant. The amplitude of familiar stimulus evoked activity, much larger than that of prestimulus gray screen activity, is reduced. Instead, we found that suppression and enhancement of familiar and nonfamiliar stimuli responsiveness go hand-in-hand, presumably driven by stimulus competition. However, further experiments are needed to establish the independence of spontaneous and evoked activity changes following familiarity. Our observations contrast with a recent study that showed increased spontaneous but no change in evoked activity in the visual cortex following multiple days of grating stimulus exposure in head-fixed but not body-restrained mice. Furthermore, the increase in spontaneous activity and behavioral habituation did not remain specific to the experienced orientation¹¹. It is unclear whether the differences in the results are due to locomotion, which is known to influence neuronal activity in the visual cortex significantly⁷³, or stimulus features (drifting vs. phase reversing gratings).

Stimulus competition is more evident in highly responsive neurons tuned to non-familiar stimuli and in familiar stimulus tuned lower responsive neurons that transition to high responsiveness. One interpretation of increased responsiveness to non-familiar stimuli in these neurons is that they reflect the detection of deviation from the expected familiar stimulus. In this view, error detection is built in the neurons by stimulus competition and occurs independently of the spontaneous activity representation of the expected or familiar stimulus (Fig. 7). Though not directly addressed in this study, we also speculate that stimulus competition could explain neural activity that persists when an expected stimulus is skipped^{63,74,75} if the direction of competition between familiar and non-familiar inputs is opposite in excitatory and specific class of inhibitory neurons (Fig. 7). This speculation is based on studies that show stronger familiar stimulus evoked activity in somatostatin expressing inhibitory neurons⁷⁶, which are also involved in detecting skipped expected stimulus⁷⁷. In this model (Fig. 7), reduced excitation and increased inhibition by the familiar stimulus will weaken neural activity, whereas increased excitation and reduced inhibition by novel stimuli will enhance it. Furthermore, skipping an expected stimulus will lead to disinhibition and elicit neural activity.

Beyond enabling novelty detection, stimulus competition may contribute to memory generalization or unintended learning. We found a subtle increase in the fraction of neurons responding to the same natural images after familiarity with a 45° grating stimulus. Natural images consist of multiple orientations with a predominance of cardinal angles⁷⁸, whose responsiveness was also increased in neurons activated by natural images post familiarity. We propose that non-learned stimuli with features that compete with or are similar to the familiar stimulus will lead to increased and decreased neuronal responsiveness.

Methods

Mice

All animal procedures are approved by the University of Kansas Institute of Animal Use and Care Committee and meet the NIH guidelines for the use and care of vertebrate animals.

C57BL/6J-Tg (Thy1-GCaMP6s) GP4.3Dkim/J (https://www.jax.org/strain/024275) were maintained as heterozygotes ⁷⁹. A maximum of five mice were housed in a standard cage but individually housed after the cranial window surgery. Mice were housed on a 12h-light/12h-dark cycle.

Cranial window

~ 4-month-old GCaMP6S mice received a cranial window on the right hemisphere over the visual cortex. A small scalp incision was made over the midline of the skull. A 5-mm diameter circle covering the visual cortex was scored using a biopsy punch. The skull was thinned along the scored circle with a fine drill using a sterile 0.5mm diameter round burr (Fine Science Tools). The bone flap was removed with fine forceps leaving behind the dura. A 5-mm diameter sterile circular glass coverslip (Harvard Apparatus) was positioned over the opening. Vetbond and cyanoacrylate glue was applied between the coverslip and bone to keep the coverslips in place. Metabond (C&B Metabond) was applied over the exposed skull. ~2-weeks after the surgery, a titanium head-post was affixed around the window to restrain mice during imaging. A black paint powder was applied over the cement, and a light-blocking cone was attached to the titanium headpost to block monitor light from reaching the photomultiplier tubes (PMTs) during imaging of visually evoked activity.

Widefield calcium imaging

Widefield calcium imaging was performed 14 days after cranial window surgeries to identify the visual cortex. Imaging was performed in a custom-built upright microscope with a 4X objective (Nikon). Awake mice, habituated for two days to the microscope, were positioned 20 cm in front of a high refresh rate monitor displaying a horizontal bar (1° of the visual field) drifting at 10 Hz. Images were collected using an sCMOS camera at 5Hz (1024 x 1024 pixels; Photometrics). GCaMP6 was excited by an LED (Lambda FLED, Sutter) filtered through a bandpass filter (470/40, 49002 Chroma), and the emission was filtered through a 525/50 bandpass filter. Reference vasculature was imaged with a 470 nm fiber-coupled LED powered by T-Cube LED drivers (Thorlabs). Images were downsized to 256x256 pixels, and magnitude maps, based on GCaMP6 fluorescence, were computed by extracting the Fourier component of fluorescence changes to matched stimulus frequency. The fractional change in fluorescence represents response magnitude, and the magnitude maps were thresholded at 30% of the peak-response amplitude. The visually responsive cortex was mapped by overlaying the magnitude maps over the 470nm reference image.

Two-photon imaging

Neurons within the mapped visual cortex (~100-150 µm below the dura) were imaged at 4.22 Hz, using a Sutter MOM multiphoton microscope, in head-fixed awake mice restrained in a body tube. The Ti: sapphire laser (MaiTai HP: Newport SpectraPhysics; 940 nm) was routed to the microscope using table optics. The power was adjusted (20-40mW) using a rotating half-wave plate and a polarizing beam splitter to avoid signal saturation. A pair of galvanometric mirrors scan the laser beams to the back aperture of the objective (Nikon 16X 0.8 NA). The emission signal was collected through the same objective, passed through a short pass filter to block infrared wavelengths, and routed to a GaASP PMT after passing through a 540/50 bandpass

filter. Image acquisition was controlled by Scanimage (Vidrio Technologies). The imaging field was a single Z frame of 336 x 336 um (256 x 256 pixels) consisting of 100 or more cells.

Visual stimulus to head restrained mice

Visual stimuli were delivered on a high refresh rate monitor placed 20 cm in front of the headrestrained animals covering 94° x 61° of the visual field. The software for generating visual stimuli was modified from a custom-written stimulus suite (a kind gift from Dr. Mark Bear's lab) written in Matlab (Mathworks) using the PsychToolbox extension (http://psychtoolbox.org). Mice were habituated to a gray screen by head-restraining them under the microscope for two days (30 minutes each day). On the first imaging session (S1), 10-15 minutes after head fixation, the GCaMP6 response was first imaged without visual stimulus (total darkness) to record spontaneous activity. ~2 minutes later, visually evoked activity imaging was performed. Visual stimuli consisted of 30 seconds of the gray screen followed by 8-cycles of 100% contrast, sinusoidal, phase reversing (2 Hz, 0.05 cycles /degree) grating stimuli of different orientations $(0^{\circ}, 45^{\circ}, 90^{\circ}, 135^{\circ} - 3 \text{ seconds each})$ and a set of ten natural images (0.3 second/image - 3 second/image)seconds per set) interspersed with 6 seconds of the gray screen. The order of stimuli was different in each cycle. Grayscale natural images were obtained from Berkeley Segmentation Dataset, contrast normalized, and resized to 1600 x 1068 pixels. Grating stimuli covered the entire monitor display value range between black and white. Gamma correction was performed to ensure the total luminance in the gray screen and grating stimuli were the same.

For the next eight days, head-restrained mice were exposed to two sessions of 60 seconds of the gray screen followed by five blocks of 100 seconds of phase-reversing grating stimulus with 30 seconds of the gray screen between blocks. The two sessions were separated by ~1-2 hours. On the following day (S2), spontaneous and evoked activity was imaged as before. We

tried to closely match the same field of view imaged in the first session for post-training imaging on the seventh day.

Calcium imaging analysis

Motion registration and ROI detection in the time-series images were performed using Suite2p RRID: SCR_016434 ⁴⁶. Tau and neuropil coefficient for spike deconvolution were set at 2.0 and 0.5, respectively. Suite2p generated ROIs were chosen as cells (cellular ROI) if the soma was visible in the mean or maximum projection image. All suite2p ROIs were matched between sessions. The following imaging sessions were matched for the analyses - S1-evoked and S2-evoked; S1-spontaneous and S1-evoked; S2-spontaneous and S2-evoked. If an ROI is present in one of the matching sessions but not the other, then a manual ROI was placed if the mean projection image showed the presence of a morphologically similar neuron at the same location.

Cellular fluorescence (F) was corrected for neuropil contamination, estimated as the ratio of blood vessel fluorescence to that of neuropil (Fneu). Neuropil-corrected fluorescence (Fcorr) was calculated as F - (0.5xFneu). dF/F0 is calculated as (Fcorr – F0)/F0, where F0 is defined as the mode of the Fcorr density distribution. Cellular ROIs that did not have at least one peak greater than 10% dF/F0 anywhere in the time series in at least one of the matched sessions were excluded. The 10% dF/F0 could lie anywhere in the time series (corresponding to the gray screen or the stimulus period) and is averaged out by the trial variation. We then manually examined all matched neurons and only selected ones with clearly visible soma in both sessions.

For each neuron, dF/F0 elicited by each stimulus was calculated as the mean dF/F0 of imaging frames corresponding to three seconds of the stimulus period and the preceding three seconds of the gray screen from the eight trials. The mean of trial-averaged dF/F0 of a neuron for each stimulus and the preceding gray screen was calculated as the mean of average dF/F0 during

the three-second stimulus and three-second gray screen periods, respectively. Neurons are considered active if the mean of trial-averaged dF/F0 during the stimulus period is greater than two standard deviations of the mean of the trial-averaged dF/F0 of the gray screen preceding it (p<0.05, paired t-test). Neurons are considered high or low responsive if they pass the criteria for active neurons and have a trial-averaged mean dF/F0 of >4 or <4, respectively, for at least one of the grating stimuli.

To obtain an orientation tuning curve, the trial-averaged area under the curve of dF/F0 during the 3 seconds of each grating stimulus was fit as a function of stimulus angle (ϕ) with a von Mises function (Eqn. 1) in neurons responsive to any grating stimuli.

$$f(\phi) = Ae^{K(\cos[2(\phi - \theta)] - 1)} + b$$
 Eqn. 1

The function is defined by four fit parameters: a preferred stimulus orientation that gives the maximum response (θ) , a tuning curve width (K), a response amplitude (A), and an intercept (b). Note, in our equation the angle difference $(\phi - \theta)$ is doubled to fit the 180° data to the standard 360° von Mises function. Fits were calculated with a maximum likelihood estimate of θ and K of using CircStat in Matlab and least-squares regression was then used to identify A and B. The fraction of explained variance, B^2 , was calculated, and the full width at half-maximum B^2 .

$$FWHM = \arccos\left[\frac{1}{K} \cdot \ln(\frac{e^K}{2} + \frac{e^{-K}}{2})\right]$$

Note, this equation is one half of the standard equation for FWHM of a von Mises function due to our range of angles from 0° to 180° . The amplitude (Amax, Eqn. 2) at the preferred stimulus angle (θ) and amplitude (Amin, Eqn. 3) at the opposite angle, 90° $\pi/2$ radians from preferred, were also calculated.

$$A_{max} = f(\phi = \theta)$$
 Eqn. 2

A_min =
$$f(\phi=\theta+90^{\circ}\pi/2)$$
 Eqn. 3

Only neurons with a tuning curve fit with R²>0.7 and were identified as visually responsive were used for analyses involving preferred orientations.

The fraction of neurons preferring an indicated orientation is calculated as the number of neurons whose preferred orientation is within 22.5° of the indicated bin divided by the number of neurons in all the bins. To calculate the fraction of neurons that retained the original or changed their preferred orientation, we binned (± 22.5° from the indicated orientation) neurons based on their S1 preferred orientation and calculated the number of matched neurons on S2 with the indicated preferred orientation divided by the total number of neurons in that group. To binarize as 45° and non-45° neurons, we pooled neurons tuned within 22.5° to 67.5° as 45° preferred neurons and the rest as non-45° neurons. The fraction of neurons responsive (active) for natural images was calculated as the number of neurons considered active for natural images divided by the total number of identified neurons.

Paired FWHM of matched highly responsive neurons on S1 and S2 evoked sessions was calculated as the mean FWHM of all matched highly responsive neurons that had the preferred orientation within the same bin on both sessions. $\Delta dF/F0$ was calculated as mean trial averaged dF/F0 on S2 – S1 for the same stimulus for each neuron.

Deconvolved spikes obtained from Suite2p for cellular ROIs were thresholded (> 2 SD from the mean) and binarized to assess functional connectivity. Briefly, neuron pairs are considered functionally connected if the number of their coactive frames exceeds 95% of the cumulative probability distribution generated by a 1000 random circular shift of their activity. The functional connectivity matrix generated above was used to determine the node degree - the

number of edges connected to each node (neuron) during the entire imaging period using MATLAB graph and degree functions.

To quantify spontaneous activity, the mean dF/F0 for each three seconds imaging period in darkness was first calculated for each neuron. The mean of 39 dF/F0 was obtained for each neuron. Neurons were matched with the evoked session on the same day to classify them in preferred orientation bins. Mean spontaneous dF/F0 was calculated for matched high responsive neurons tuned to the indicated orientation in the corresponding evoked session.

The cosine similarity between neural responsiveness to spontaneous and the indicated stimulus was calculated as similarity = X.Y/||X||||Y||, where X and Y are the average spontaneous dF/F0 and the mean of trial-averaged dF/F0 elicited by the indicated stimulus in highly responsive neurons. The same analysis was restricted to highly responsive neurons tuned to the indicated bin orientation to identify the similarity of responses based on tuning preference. Neurons matched between spontaneous and evoked imaging on S1 and S2 were used for this analysis.

We also measured how much spontaneous neural activity overlapped with the stimulus-driven response from the grating patterns for individual three second time windows. We defined the neural subspace of spontaneous activity by its covariance matrix of all three second windows of imaged activity without stimuli. The response for each individual stimulus averaged across the three seconds was then also defined by a vector in neural space. This vector of each stimulus response was multiplied by the spontaneous covariance matrix to estimate the amount of overlap between each response and the spontaneous activity. For this analysis of the population, we used all cells that were active with dF/F0 > 1 and tuned with an $R^2 > 0.7$. Overlap was calculated with the following equation 45,80 :

$$A = \frac{S_{Stim}^T C_{Spont} S_{Stim}}{\|S_{Stim}\|^2 \cdot \max(\sigma_{Spont})}$$

The neural response vector to each stimulus, S_{Stim} , was projected into the neural space that was observed during the spontaneous neural activity, defined by its covariance matrix, C_{Spont} . We normalized by dividing by the largest singular value of the covariance matrix, C_{Spont} . Each stimulus response was also normalized to a unit vector by dividing by the squared Euclidean norm of the S_{Stim} to compare relative responses across cells rather than global activity levels. Thus, the amount of overlap could range from 0 to 1, with a maximum overlap of 1 when with S_{Stim} perfectly aligns to the spontaneous neural dimension with the largest variance and 0 if S_{Stim} aligns with a neural dimension with no spontaneous activity. For comparison among the four different grating pattern stimuli, we calculated the percentage of spontaneous activity that overlapped with each individual grating out of the total overlap across the four different patterns within a cycle. A two-way ANOVA with S1 vs. S2 and animal as the two factors for the 8 stimuli cycles was used to test for significant differences in the overlap between the pre-and post-altered experience imaging sessions for the four grating stimuli and to generate 95% confidence intervals.

Statistical analysis

Statistical tests were performed using Prism 9 or MATLAB. No statistical methods were used to predetermine sample sizes. The sample sizes are comparable to previous literature. Test for normal distribution was done with the Kolmogorov-Smirnov test. p < 0.05 was considered statistically significant. Sample sizes are reported in figure legends. Samples were individual mice or neurons , or stimulus cycles (indicated in legends). For obtaining 95% confidence intervals for similarity ratios, 1000 similarity values between two stimuli were obtained by bootstrapping with a replacement for the S1 and S2 data set, and the S2-S1 ratio was calculated.

The bootstrapped ratios were sorted in ascending order, and 50th and 950th values were used as the lower and upper bound of the 95% confidence interval. Similarly, 100th and 900th values were taken as the lower and upper bound of a 90% confidence interval. Statistical procedures are two-sided and are listed in figure legends.

Availability of materials and data

• The datasets generated during and/or analysed during the current study are available from the corresponding author on request.

References

- 1 Cooke, S. F. & Ramaswami, M. in *The Cognitive Neurosciences, 6th Edn, eds D. Poeppel, G. Mangun, and M. Gazzaniga, (Cambridge: MIT Press)* 197-206 (2020).
- Rankin, C. H. *et al.* Habituation revisited: an updated and revised description of the behavioral characteristics of habituation. *Neurobiol Learn Mem* **92**, 135-138, doi:10.1016/j.nlm.2008.09.012 (2009).
- Cooke, S. F., Komorowski, R. W., Kaplan, E. S., Gavornik, J. P. & Bear, M. F. Visual recognition memory, manifested as long-term habituation, requires synaptic plasticity in V1. *Nat Neurosci* **18**, 262-271, doi:10.1038/nn.3920 (2015).
- 4 Niraula, S. *et al.* Excitation-inhibition imbalance disrupts visual familiarity in amyloid and non-pathology conditions. *Cell Rep* **42**, 111946, doi:10.1016/j.celrep.2022.111946 (2023).
- Montgomery, D. P., Hayden, D. J., Chaloner, F. A., Cooke, S. F. & Bear, M. F. Stimulus-Selective Response Plasticity in Primary Visual Cortex: Progress and Puzzles. *Front Neural Circuits* **15**, 815554, doi:10.3389/fncir.2021.815554 (2021).
- Montey, K. L., Eaton, N. C. & Quinlan, E. M. Repetitive visual stimulation enhances recovery from severe amblyopia. *Learn Mem* **20**, 311-317, doi:10.1101/lm.030361.113 (2013).
- 7 Kissinger, S. T., Pak, A., Tang, Y., Masmanidis, S. C. & Chubykin, A. A. Oscillatory Encoding of Visual Stimulus Familiarity. *J Neurosci* **38**, 6223-6240, doi:10.1523/JNEUROSCI.3646-17.2018 (2018).
- Frenkel, M. Y. *et al.* Instructive effect of visual experience in mouse visual cortex. *Neuron* **51**, 339-349, doi:10.1016/j.neuron.2006.06.026 (2006).
- 9 Aton, S. J., Suresh, A., Broussard, C. & Frank, M. G. Sleep promotes cortical response potentiation following visual experience. *Sleep* **37**, 1163-1170, doi:10.5665/sleep.3830 (2014).
- Gao, M., Lim, S. & Chubykin, A. A. Visual Familiarity Induced 5-Hz Oscillations and Improved Orientation and Direction Selectivities in V1. *J Neurosci* **41**, 2656-2667, doi:10.1523/JNEUROSCI.1337-20.2021 (2021).
- Miller, J. K., Miller, B. R., O'Neil, D. A. & Yuste, R. An increase in spontaneous activity mediates visual habituation. *Cell Rep* **39**, 110751, doi:10.1016/j.celrep.2022.110751 (2022).
- 12 Kaneko, M., Fu, Y. & Stryker, M. P. Locomotion Induces Stimulus-Specific Response Enhancement in Adult Visual Cortex. *J Neurosci* **37**, 3532-3543, doi:10.1523/JNEUROSCI.3760-16.2017 (2017).
- Makino, H. & Komiyama, T. Learning enhances the relative impact of top-down processing in the visual cortex. *Nat Neurosci* **18**, 1116-1122, doi:10.1038/nn.4061 (2015).
- 14 Kim, T., Chaloner, F. A., Cooke, S. F., Harnett, M. T. & Bear, M. F. Opposing Somatic and Dendritic Expression of Stimulus-Selective Response Plasticity in Mouse Primary Visual Cortex. *Front Cell Neurosci* **13**, 555, doi:10.3389/fncel.2019.00555 (2019).
- Henschke, J. U. *et al.* Reward Association Enhances Stimulus-Specific Representations in Primary Visual Cortex. *Curr Biol* **30**, 1866-1880 e1865, doi:10.1016/j.cub.2020.03.018 (2020).
- Wilson, D. E., Whitney, D. E., Scholl, B. & Fitzpatrick, D. Orientation selectivity and the functional clustering of synaptic inputs in primary visual cortex. *Nat Neurosci* **19**, 1003-1009, doi:10.1038/nn.4323 (2016).
- Jia, H., Rochefort, N. L., Chen, X. & Konnerth, A. Dendritic organization of sensory input to cortical neurons in vivo. *Nature* **464**, 1307-1312, doi:10.1038/nature08947 (2010).
- lacaruso, M. F., Gasler, I. T. & Hofer, S. B. Synaptic organization of visual space in primary visual cortex. *Nature* **547**, 449-452, doi:10.1038/nature23019 (2017).
- 19 Chen, T. W. *et al.* Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature* **499**, 295-300, doi:10.1038/nature12354 (2013).

- Kohn, A. Visual adaptation: physiology, mechanisms, and functional benefits. *J Neurophysiol* **97**, 3155-3164, doi:10.1152/jn.00086.2007 (2007).
- 21 King, J. L. & Crowder, N. A. Adaptation to stimulus orientation in mouse primary visual cortex. *Eur J Neurosci* **47**, 346-357, doi:10.1111/ejn.13830 (2018).
- Gutnisky, D. A. & Dragoi, V. Adaptive coding of visual information in neural populations. *Nature* **452**, 220-224, doi:10.1038/nature06563 (2008).
- Jeyabalaratnam, J. *et al.* Adaptation shifts preferred orientation of tuning curve in the mouse visual cortex. *PLoS One* **8**, e64294, doi:10.1371/journal.pone.0064294 (2013).
- Dragoi, V., Sharma, J. & Sur, M. Adaptation-induced plasticity of orientation tuning in adult visual cortex. *Neuron* **28**, 287-298, doi:10.1016/s0896-6273(00)00103-3 (2000).
- Solomon, S. G. & Kohn, A. Moving sensory adaptation beyond suppressive effects in single neurons. *Curr Biol* **24**, R1012-1022, doi:10.1016/j.cub.2014.09.001 (2014).
- Webster, M. A. Visual Adaptation. *Annu Rev Vis Sci* **1**, 547-567, doi:10.1146/annurev-vision-082114-035509 (2015).
- Jin, M. & Glickfeld, L. L. Magnitude, time course, and specificity of rapid adaptation across mouse visual areas. *J Neurophysiol* **124**, 245-258, doi:10.1152/jn.00758.2019 (2020).
- Homann, J., Koay, S. A., Chen, K. S., Tank, D. W. & Berry, M. J., 2nd. Novel stimuli evoke excess activity in the mouse primary visual cortex. *Proc Natl Acad Sci U S A* **119**, doi:10.1073/pnas.2108882119 (2022).
- Bharmauria, V., Ouelhazi, A., Lussiez, R. & Molotchnikoff, S. Adaptation-induced plasticity in the sensory cortex. *J Neurophysiol* **128**, 946-962, doi:10.1152/jn.00114.2022 (2022).
- 30 Xu, S., Jiang, W., Poo, M. M. & Dan, Y. Activity recall in a visual cortical ensemble. *Nat Neurosci* **15**, 449-455, S441-442, doi:10.1038/nn.3036 (2012).
- Tsodyks, M., Kenet, T., Grinvald, A. & Arieli, A. Linking spontaneous activity of single cortical neurons and the underlying functional architecture. *Science* **286**, 1943-1946, doi:10.1126/science.286.5446.1943 (1999).
- Sakata, S. & Harris, K. D. Laminar structure of spontaneous and sensory-evoked population activity in auditory cortex. *Neuron* **64**, 404-418, doi:10.1016/j.neuron.2009.09.020 (2009).
- Ringach, D. L. Spontaneous and driven cortical activity: implications for computation. *Curr Opin Neurobiol* **19**, 439-444, doi:10.1016/j.conb.2009.07.005 (2009).
- Miller, J. E., Ayzenshtat, I., Carrillo-Reid, L. & Yuste, R. Visual stimuli recruit intrinsically generated cortical ensembles. *Proc Natl Acad Sci U S A* **111**, E4053-4061, doi:10.1073/pnas.1406077111 (2014).
- Kok, P., Mostert, P. & de Lange, F. P. Prior expectations induce prestimulus sensory templates. *Proc Natl Acad Sci U S A* **114**, 10473-10478, doi:10.1073/pnas.1705652114 (2017).
- Kenet, T., Bibitchkov, D., Tsodyks, M., Grinvald, A. & Arieli, A. Spontaneously emerging cortical representations of visual attributes. *Nature* **425**, 954-956, doi:10.1038/nature02078 (2003).
- Jermakowicz, W. J., Chen, X., Khaytin, I., Bonds, A. B. & Casagrande, V. A. Relationship between spontaneous and evoked spike-time correlations in primate visual cortex. *J Neurophysiol* **101**, 2279-2289, doi:10.1152/jn.91207.2008 (2009).
- Han, F., Caporale, N. & Dan, Y. Reverberation of recent visual experience in spontaneous cortical waves. *Neuron* **60**, 321-327, doi:10.1016/j.neuron.2008.08.026 (2008).
- Berkes, P., Orban, G., Lengyel, M. & Fiser, J. Spontaneous cortical activity reveals hallmarks of an optimal internal model of the environment. *Science* **331**, 83-87, doi:10.1126/science.1195870 (2011).
- Ferezou, I. & Deneux, T. Review: How do spontaneous and sensory-evoked activities interact? *Neurophotonics* **4**, 031221, doi:10.1117/1.NPh.4.3.031221 (2017).

- 41 Fiser, J., Chiu, C. & Weliky, M. Small modulation of ongoing cortical dynamics by sensory input during natural vision. *Nature* **431**, 573-578, doi:10.1038/nature02907 (2004).
- 42 Yao, H., Shi, L., Han, F., Gao, H. & Dan, Y. Rapid learning in cortical coding of visual scenes. *Nat Neurosci* **10**, 772-778, doi:10.1038/nn1895 (2007).
- 43 Stringer, C. *et al.* Spontaneous behaviors drive multidimensional, brainwide activity. *Science* **364**, 255, doi:10.1126/science.aav7893 (2019).
- Avitan, L. & Stringer, C. Not so spontaneous: Multi-dimensional representations of behaviors and context in sensory areas. *Neuron* **110**, 3064-3075, doi:10.1016/j.neuron.2022.06.019 (2022).
- Avitan, L. *et al.* Spontaneous and evoked activity patterns diverge over development. *Elife* **10**, doi:10.7554/eLife.61942 (2021).
- Pachitariu, M. *et al.* Suite2p: beyond 10,000 neurons with standard two-photon microscopy. *bioRxiv* (2017).
- 47 Niell, C. M. & Stryker, M. P. Highly selective receptive fields in mouse visual cortex. *J Neurosci* **28**, 7520-7536, doi:10.1523/JNEUROSCI.0623-08.2008 (2008).
- Perez-Ortega, J., Alejandre-Garcia, T. & Yuste, R. Long-term stability of cortical ensembles. *Elife* **10**, doi:10.7554/eLife.64449 (2021).
- 49 Hensch, T. K. Critical period mechanisms in developing visual cortex. *Curr Top Dev Biol* **69**, 215-237, doi:10.1016/S0070-2153(05)69008-4 (2005).
- Espinosa, J. S. & Stryker, M. P. Development and plasticity of the primary visual cortex. *Neuron* **75**, 230-249, doi:10.1016/j.neuron.2012.06.009 (2012).
- Bavelier, D. & Neville, H. J. Cross-modal plasticity: where and how? *Nat Rev Neurosci* **3**, 443-452, doi:10.1038/nrn848 (2002).
- Mao, Y. T., Hua, T. M. & Pallas, S. L. Competition and convergence between auditory and cross-modal visual inputs to primary auditory cortical areas. *J Neurophysiol* **105**, 1558-1573, doi:10.1152/jn.00407.2010 (2011).
- Lee, H. K. & Whitt, J. L. Cross-modal synaptic plasticity in adult primary sensory cortices. *Curr Opin Neurobiol* **35**, 119-126, doi:10.1016/j.conb.2015.08.002 (2015).
- Yu, L., Cuppini, C., Xu, J., Rowland, B. A. & Stein, B. E. Cross-Modal Competition: The Default Computation for Multisensory Processing. *J Neurosci* **39**, 1374-1385, doi:10.1523/JNEUROSCI.1806-18.2018 (2019).
- Ramiro-Cortes, Y., Hobbiss, A. F. & Israely, I. Synaptic competition in structural plasticity and cognitive function. *Philos Trans R Soc Lond B Biol Sci* **369**, 20130157, doi:10.1098/rstb.2013.0157 (2014).
- Oh, W. C., Hill, T. C. & Zito, K. Synapse-specific and size-dependent mechanisms of spine structural plasticity accompanying synaptic weakening. *Proc Natl Acad Sci U S A* **110**, E305-312, doi:10.1073/pnas.1214705110 (2013).
- Ramaswami, M. Network plasticity in adaptive filtering and behavioral habituation. *Neuron* **82**, 1216-1229, doi:10.1016/j.neuron.2014.04.035 (2014).
- Keller, G. B. & Mrsic-Flogel, T. D. Predictive Processing: A Canonical Cortical Computation. *Neuron* **100**, 424-435, doi:10.1016/j.neuron.2018.10.003 (2018).
- 59 de Lange, F. P., Heilbron, M. & Kok, P. How Do Expectations Shape Perception? *Trends Cogn Sci* **22**, 764-779, doi:10.1016/j.tics.2018.06.002 (2018).
- Aitchison, L. & Lengyel, M. With or without you: predictive coding and Bayesian inference in the brain. *Curr Opin Neurobiol* **46**, 219-227, doi:10.1016/j.conb.2017.08.010 (2017).
- Padamsey, Z. & Rochefort, N. L. Defying Expectations: How Neurons Compute Prediction Errors in Visual Cortex. *Neuron* **108**, 1016-1019, doi:10.1016/j.neuron.2020.12.005 (2020).
- Keller, G. B., Bonhoeffer, T. & Hubener, M. Sensorimotor mismatch signals in primary visual cortex of the behaving mouse. *Neuron* **74**, 809-815, doi:10.1016/j.neuron.2012.03.040 (2012).

- Gavornik, J. P. & Bear, M. F. Learned spatiotemporal sequence recognition and prediction in primary visual cortex. *Nat Neurosci* **17**, 732-737, doi:10.1038/nn.3683 (2014).
- Fiser, A. *et al.* Experience-dependent spatial expectations in mouse visual cortex. *Nat Neurosci* **19**, 1658-1664, doi:10.1038/nn.4385 (2016).
- Egner, T., Monti, J. M. & Summerfield, C. Expectation and surprise determine neural population responses in the ventral visual stream. *J Neurosci* **30**, 16601-16608, doi:10.1523/JNEUROSCI.2770-10.2010 (2010).
- Aitken, F. *et al.* Prior expectations evoke stimulus-specific activity in the deep layers of the primary visual cortex. *PLoS Biol* **18**, e3001023, doi:10.1371/journal.pbio.3001023 (2020).
- Poort, J. *et al.* Learning Enhances Sensory and Multiple Non-sensory Representations in Primary Visual Cortex. *Neuron* **86**, 1478-1490, doi:10.1016/j.neuron.2015.05.037 (2015).
- 68 Price, B. H., Jensen, C. M., Khoudary, A. A. & Gavornik, J. P. Expectation violations produce error signals in mouse V1. *bioRxiv*, 2021.2012.2031.474652, doi:10.1101/2021.12.31.474652 (2022).
- Zmarz, P. & Keller, G. B. Mismatch Receptive Fields in Mouse Visual Cortex. *Neuron* **92**, 766-772, doi:10.1016/j.neuron.2016.09.057 (2016).
- Haider, B., Duque, A., Hasenstaub, A. R., Yu, Y. & McCormick, D. A. Enhancement of visual responsiveness by spontaneous local network activity in vivo. *J Neurophysiol* **97**, 4186-4202, doi:10.1152/jn.01114.2006 (2007).
- Scholvinck, M. L., Friston, K. J. & Rees, G. The influence of spontaneous activity on stimulus processing in primary visual cortex. *Neuroimage* **59**, 2700-2708, doi:10.1016/j.neuroimage.2011.10.066 (2012).
- Gutnisky, D. A., Beaman, C. B., Lew, S. E. & Dragoi, V. Spontaneous Fluctuations in Visual Cortical Responses Influence Population Coding Accuracy. *Cereb Cortex* **27**, 1409-1427, doi:10.1093/cercor/bhv312 (2017).
- Niell, C. M. & Stryker, M. P. Modulation of visual responses by behavioral state in mouse visual cortex. *Neuron* **65**, 472-479, doi:10.1016/j.neuron.2010.01.033 (2010).
- Ross, J. M. & Hamm, J. P. Cortical Microcircuit Mechanisms of Mismatch Negativity and Its Underlying Subcomponents. *Front Neural Circuits* **14**, 13, doi:10.3389/fncir.2020.00013 (2020).
- Carrillo-Reid, L., Miller, J. E., Hamm, J. P., Jackson, J. & Yuste, R. Endogenous sequential cortical activity evoked by visual stimuli. *J Neurosci* **35**, 8813-8828, doi:10.1523/JNEUROSCI.5214-14.2015 (2015).
- Hayden, D. J., Montgomery, D. P., Cooke, S. F. & Bear, M. F. Visual recognition is heralded by shifts in local field potential oscillations and inhibitory networks in primary visual cortex. *J Neurosci* **41**, 6257-6272, doi:10.1523/JNEUROSCI.0391-21.2021 (2021).
- Hamm, J. P. & Yuste, R. Somatostatin Interneurons Control a Key Component of Mismatch Negativity in Mouse Visual Cortex. *Cell Rep* **16**, 597-604, doi:10.1016/j.celrep.2016.06.037 (2016).
- 78 Dragoi, V., Turcu, C. M. & Sur, M. Stability of cortical responses and the statistics of natural scenes. *Neuron* **32**, 1181-1192, doi:10.1016/s0896-6273(01)00540-2 (2001).
- Dana, H. *et al.* Thy1-GCaMP6 transgenic mice for neuronal population imaging in vivo. *PLoS One* **9**, e108697, doi:10.1371/journal.pone.0108697 (2014).
- 80 Elsayed, G. F., Lara, A. H., Kaufman, M. T., Churchland, M. M. & Cunningham, J. P. Reorganization between preparatory and movement population responses in motor cortex. *Nat Commun* **7**, 13239, doi:10.1038/ncomms13239 (2016).

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Author contributions: S.N. and J.S. designed the experiments. W.L.H, A.G.R, and J.S. performed data analysis. A.G.R. and J.S designed data analysis. J.S. wrote the paper and supervised the research.

Conflict of interest: The authors declare no competing financial interests.

Figure legends

Figure 1. Repeated orientation-grating stimulus exposure reduces responsiveness **selectively to the familiar stimulus.** A. representation of a head and body restrained mouse placed under microscope objective viewing visual stimuli. **B.** Experimental timeline (top). Each square indicates a day, and h1 and h2 are habituation days when mice were exposed to a gray screen. Spontaneous and visually evoked calcium transients were imaged in sessions S1 and S2. Visual stimuli consisted of full-field phase reversing grating stimuli of different orientations and natural images (NI; bottom). On the intervening days, mice experienced only one orientation of phase reversing grating stimulus. C. Representative standard deviation projection images from S1 (left) and S2 (right); scale bar: 50 µm. A neuron circled in blue is zoomed on the right; scale bar: 10 µm. **D.** Raster plot of calcium transients (dF/F0) from identified neurons in the imaging field in C on S1 (left) and S2 (right). E. dF/F0 of calcium transients from the neuron circled in C on S1 (blue) and S2 (orange) during the entire imaging period encompassing gray screen, different grating, and natural images (represented as bars or NI on top) stimuli. Gray bars represent the stimulus duration. F. Trial-averaged dF/F0 (%) of the transients in E for each stimulus. GS and stim represent the gray screen and the indicated stimulus period, respectively. Arrow represents the start of the stimulus. G. Population average of the mean trial-averaged dF/F0 for the indicated stimulus on S1 and S2. ** p<0.01; paired t test; n = 10 mice (circles on the histogram). Data presented as mean \pm sem. **H.** Scatterplot representing the relation between dF/F0 on S1 and the change in dF/F0 (Δ dF/F0) following altered experience. The circles represent neurons (561 neurons). The dotted line represents the approximate mean dF/F0 on S1 (4%). X-axis – log scale.

Figure 2. Altered experience elicits stimulus competition in low responsive neurons A.

Representative orientation tuning curve fit (orange line) of a low responsive neuron that retained (top) or shifted (bottom) its preferred stimulus following altered experience. The green circles represent the mean of trial-averaged dF/F0 of 0° , 45° , 90° , and 135° grating stimuli. **B.** Fraction of all (top; 286 neurons), high (middle; 122 neurons), or low (bottom; 96 neurons) grating responsive neurons with good tuning curve fit in both imaging sessions in indicated bins separated by 45° before (S1) and after (S2) altered experience. **C.** Fraction of all (top), high (middle), or low (bottom) neurons tuned to indicated S1 orientation that retained their tuning within the same bin or changed (\pm 45° or \pm 90° relative to S1). **D.** Same as C except that 0° , 90° , and 135° (non- 45°) bins are pooled for all (top; n = 222 non- 45° and 64 45° neurons), high responsive (middle; n = 122 non- 45° and 35 45° neurons), and low (bottom; 100 non- 45° , 29 45° neurons) responsive neurons. Original and not original represents the fraction of neurons that retained and changed their S1 preference, respectively. # p = 0.05, ** p < 0.01 McNemar's test for correlated proportions.

Figure 3. Altered experience elicits stimulus competition in high-responsive neurons. A. Representative orientation tuning curve fit (orange line) of a highly responsive neuron whose tuning curve sharpened following altered experience. The green circles represent the mean of trial-averaged dF/F0 of 0°, 45°, 90°, and 135° grating stimuli. **B-E.** Left. Cumulative probability distribution (CPD) of full width at half maximum (FWHM) of matched neurons that retained high responsiveness and preferred orientation before (S1) and after (S2) altered experience. * p<0.05, KS test; n = 36, 23, 33, and 30 matched neurons that remained highly responsive and tuned to 0°, 45°, 90°, and 135°, respectively. Middle. Mean FWHM of the same neurons. * p<0.05, *** p<0.001, paired t tests. Black circles represent individual neuron values. (C and D)

Right. Change in mean trial-averaged dF/F0 (Δ dF/F0) following altered experience for the neurons tuned to 45° (C) or 90° (D). * p < 0.05, ** p < 0.01, *** p < 0.001, Friedman test followed by Dunn's post hoc multiple comparisons test. Data are presented as mean \pm SEM. **Figure 4. Neurons tuned to non-familiar stimuli dominate functional connectivity. A.** Representative raster plot of calcium transients (dF/F0) from 43 neurons (N1-N43) neurons of one mouse. A plot of three of those neurons (N14,16,18) is magnified below. N14 and N16 are functionally connected to each other but not N18. **B.** Average node degrees of active neurons tuned to 0°, 45°, 90°, and 135° before (S1) and after (S2) altered experience. Data are presented as box (25th to 75th percentile) and whisker (minimum and maximum values) plots, the median value indicated as a horizontal line. n = 174, 193 (0°), 136, 90 (45°), 135, 149 (90°), 116, 130 (135°) neurons. *** p < 0.001 Mann Whitney U test. **C.** Identified functionally connected neurons are categorized based on their tuning preference.

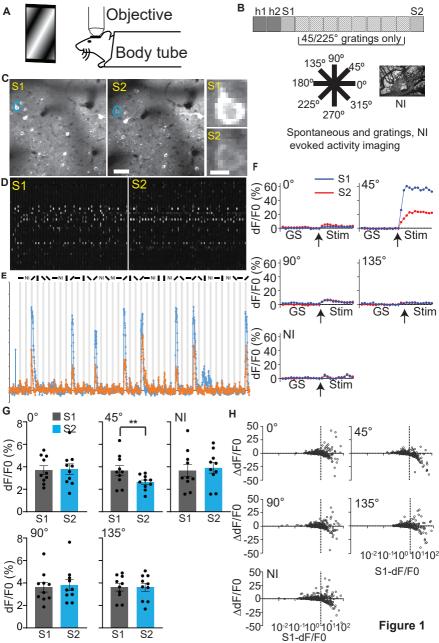
Figure 5. Increased fraction of neurons responsive to natural images following altered experience. A. Representative imaging fields on S1 (left) and S2 (right) from one mouse. Scale bar: 100 μ m. **B.** Population trial-averaged dF/F0 elicited by natural images from the neuron circled in blue in A. C. Fraction of high, low, and non-responsive neurons to natural images. Circles on the histogram represent average mouse values. n = 10 mice. # p = 0.05, * p < 0.05, paired t tests. **D.** Change in dF/F0 (S2 – S1 dF/F0) in response to 0, 45, 90, and 135° stimuli in neurons that became responsive (gain), unresponsive (loss), or remained active (stable) to natural images. * p < 0.05, ** p < 0.01, *** p < 0.001, Friedman test followed by Dunn's post hoc multiple comparisons test. Data are presented as mean \pm SEM.

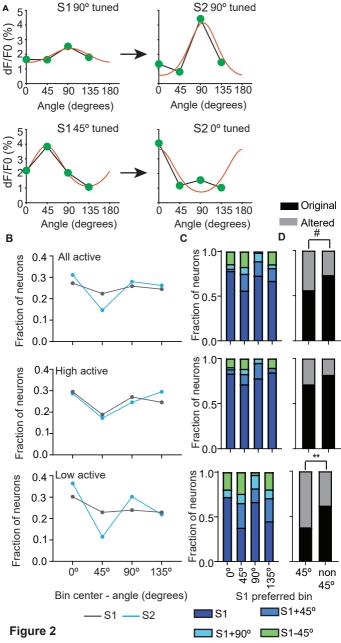
Figure 6. Spontaneous and familiar stimulus-driven activity patterns overlap. A. A representative standard deviation projection images from spontaneous and evoked imaging

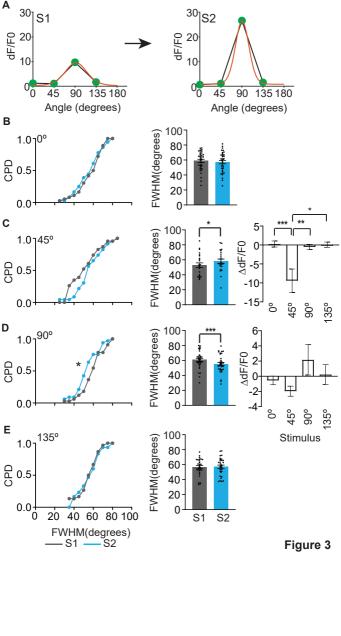
sessions (top; Scale bar: 100 µm) and corresponding dF/F0 raster plot (below). **B**. dF/F0 averaged over three seconds during spontaneous activity in highly responsive neurons tuned to the indicated orientation before (S1) and after the altered experience (S2). Data are presented as box (25th to 75th percentile) and whisker (minimum and maximum values) plots, with the median value indicated as a horizontal line. n = 50, 53, 50, 38 (S1) and 43, 19, 50, 40 (S2) neurons tuned to 0°, 45°, 90°, 135° bins, respectively from 9 mice. C. Fold-change in the similarity of responses to spontaneous activity and evoked activity elicited by indicated grating stimulus or gray screen (GS) in all neurons highly responsive to any grating stimulus. n = 191(S1) and 136 (S2) neurons from 9 mice. **D.** Fold change in the similarity of responses to spontaneous and evoked activity elicited by indicated stimulus in neurons tuned to that stimulus. $n = 50, 53, 50, \text{ and } 38 \text{ neurons (S1) and } 43, 19, 50, \text{ and } 40 \text{ neurons (S2) tuned to } 0^{\circ}, 45^{\circ}, 90^{\circ}, \text{ and } 40^{\circ}$ 135°, respectively from 9 mice. The dotted line indicates identical similarity after 45° experience. Black circles - the lower and upper bound of 95% confidence intervals (CI) obtained by bootstrapping with replacement. * null value (1.00) outside of 95% confidence interval. E. Schematic for calculating the overlap between evoked responses to stimuli and the spontaneous neural activity. The spontaneous neural activity is defined by the covariance matrix represented by the blue ellipsoid. Each trial of stimuli can then be represented as a vector defined as the response activity across all observed neurons. The calculated overlap between these stimuli responses and the spontaneous activity was normalized relative to the 1st principal component with maximum variance. F. Percentage of spontaneous neural activity that overlapped with the evoked responses for the four grating stimuli. The overlap for each of the stimuli for each time window was divided by the total overlap observed across all four stimuli in a cycle to show the relative percentages of spontaneous activity that mimicked each of the stimuli. The dotted line

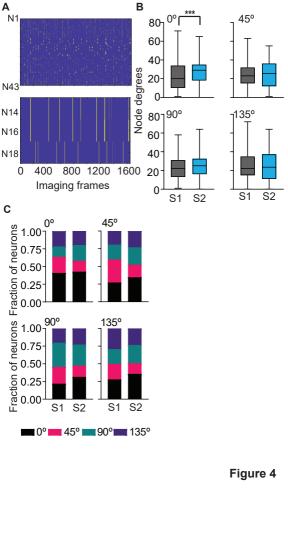
indicates an equal amount of overlap (25%) across the four stimuli. After the altered experience, neural activity during the spontaneous period significantly increased its overlap from S1 to S2 to that evoked with the familiar 45° stimulus and decreased for the 0° and 90° stimuli (*p<0.05, **p<0.01, ***p<0.001 two-way ANOVA, posthoc Tukey test). Data are presented as mean ± 95% confidence interval from the ANOVA analysis of eight repeated stimuli cycles for nine mice x two sessions.

Figure 7. A model for error detection enabled by stimulus competition. Repeated passive experience may weaken familiar or expected stimulus inputs and strengthen novel or unexpected stimuli inputs onto excitatory neurons. However, the direction of this change could be the opposite in certain classes of inhibitory neurons. Reduced excitation and enhanced inhibition elicited by familiar or expected stimulus will dampen neuronal activity; skipping the expected stimulus will lead to disinhibition and restore activity. In contrast, novel or unexpected stimuli will elicit higher neuronal activity. Double arrows and dashed arrows indicate the strengthening and weakening of synaptic inputs following passive exposure.

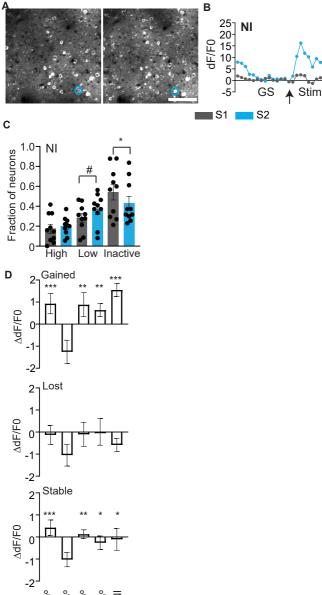








В



Stimulus

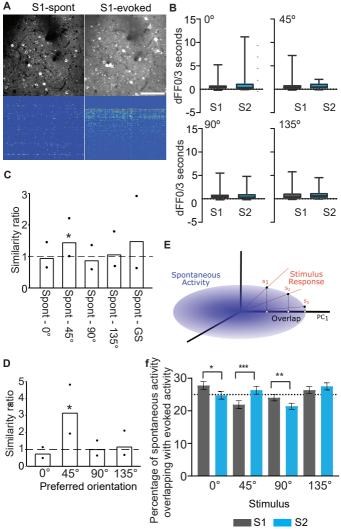


Figure 6

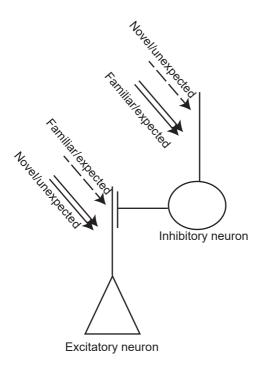


Figure 7