1 **Title**
2
3 Meta-Research: understudied genes are lost in a leaky pipeline between genome-wide assays
4 and reporting of results
5
6 **Authors**
7

8 Reese AK Richardson[1,2], Heliodoro Tejedor Navarro[2,3], Luis A Nunes Amaral[2,3,4,5]*, Thomas
9 Stoeger[2,6,7]*
10

[1]Interdisciplinary Biological Sciences, Northwestern University

[2]Department of Chemical and Biological Engineering, Northwestern University

[3]Northwestern Institute on Complex Systems, Northwestern University

[4]Department of Physics and Astronomy, Northwestern University

[5]Department of Molecular Biosciences, Northwestern University

[6]The Potocsnak Longevity Institute, Northwestern University

[7]Simpson Querrey Lung Institute for Translational Science, Northwestern University

*Correspondence to:

L.A.N.A. (amaral@northwestern.edu), T.S. (thomas.stoeger@northwestern.edu)

23
24
25 **Abstract**
26

Present-day publications on human genes primarily feature genes that already appeared in
many publications prior to completion of the Human Genome Project in 2003. These patterns
persist despite the subsequent adoption of high-throughput technologies, which routinely
identify novel genes associated with biological processes and disease. Although several
hypotheses for bias in the selection of genes as research targets have been proposed, their
explanatory powers have not yet been compared. Our analysis suggests that understudied
genes are systematically abandoned in favor of better-studied genes between the completion
of -omics experiments and the reporting of results. Understudied genes are similarly
abandoned by studies that cite these -omics experiments. Conversely, we find that publications
on understudied genes may even accrue a greater number of citations. Among 45 biological
and experimental factors previously proposed to affect which genes are being studied, we find

that 35 are significantly associated with the choice of hit genes presented in titles and abstracts
of -omics studies. To promote the investigation of understudied genes we condense our
insights into a tool, *find my understudied genes* (FMUG), that allows scientists to engage with
potential bias during the selection of hits. We demonstrate the utility of FMUG through the
identification of genes that remain understudied in vertebrate aging. FMUG is developed in
Flutter and is available for download at fmug.amaral.northwestern.edu as a MacOS/Windows
app.
45
46 **Introduction**
47
Research into human genes concentrates on a subset of genes that were already frequently
investigated prior to the completion of the Human Genome Project in 2003[1-5]. This
concentration stems from historically acquired research patterns rather than present-day
experimental possibilities[6,7]. For most human diseases, these patterns lead to little correlation
between the volume of literature published on individual genes and the strength of supporting
evidence from genome-wide approaches[8-13]. For instance, we found that 44% of the genes
identified as promising Alzheimer's disease targets by the U.S. National Institutes of Health
(NIH) Accelerating Medicine Partnership for Alzheimer's Disease (AMP-AD) initiative have never
appeared in the title or abstract of any publication on Alzheimer's disease[13]. Furthermore,
when comparing gene-disease pairs, there is no correlation between the ranks of support by
transcriptomics and occurrence in annotation databases[9].
59
Although -omics technologies can provide insights on numerous genes across the genome at a
time and thus offer the promise to counter historically acquired research patterns[14-17], this
discrepancy has persisted[9,18-22] even as the popularity of -omics technologies has risen[5,23,24]. We
therefore sought to use bibliometric data to delineate where and why understudied human
protein-coding genes are abandoned as research targets following -omics experiments. In the
absence of any prior quantitative testing of existing hypotheses, it remains unclear whether
policies to promote the exploration of a greater set of disease-related genes should focus on
how experiments are conducted, how results are reported, or how these results are
subsequently received by other scientists.
69
70 **Data**
71
We considered 450 genome-wide association studies (GWAS, from studies indexed by the
NHGRI-EBI GWAS catalog[25]), 296 studies using affinity capture mass spectrometry (Aff-MS,
indexed by BioGRID[26]), 148 transcriptomic studies (indexed by the EBI Gene Expression Atlas,
EBI-GXA[27]), and 15 genome-wide screens using CRISPR (indexed by BioGRID Open Repository of
CRISPR Screens, BioGRID ORCS[26]) (see PRISMA diagrams in **Figures S1-S4**). We denote genes
that are found to have statistically significant changes in expression or associations with a
phenotype as 'hit' genes.
79
As a surrogate for a given gene having been investigated closer, we consider whether it was
reported in the title or abstract of a research article. We determined which genes were

mentioned in the title or abstract of articles using annotations from gene2pubmed[28] and
PubTator[29]. We used NIH iCite v32 for citations[30]. For determining which gene properties were
associated with selection as research targets, we synthesized quantitative measures from a
variety of authoritative sources (see **Methods**).
86
87 **Results**
88
89 *Understudied genes are abandoned at synthesis/writing stage*
90
We sought to identify at which point in the scientific process understudied genes are ignored as
research targets in investigations using -omics experiments (**Figure 1A**). To receive scholarly
attention, a gene must travel through a pipeline from biological reality to experimental results
to write-up of those results. These results must be extended by subsequent research by other
scholars. Understudied genes do not progress all the way through the pipeline, but it is unclear
where this leak primarily occurs. The first possibility is that seemingly understudied genes are,
in fact, not understudied as they would rarely be identified through experiments. Prior studies
have, however, shown that understudied genes are frequent hits in high-throughput
experiments[8,9,31], suggesting that this is not the case. The second possibility is that
understudied genes are frequently found as hits in high-throughput experiments but are not
investigated further by the authors. The final possibility is that subsequent studies do not
continue work on understudied genes revealed by the initial study.
103
104 Evaluating the first possibility, we found that understudied genes were frequently found as hits
105 in high-throughput experiments. (**Figure 1B**, **Figure S5**, and **Figure S6**). This demonstrates, in
106 line with earlier studies[8-13], that the lack of publications on some genes is not explained by
107 underlying biological experimental evidence.
108
109 Evaluating the second possibility, we found that hit genes that are highlighted in the title or
110 abstract are strongly over-represented among the 20% highest-studied genes in all biomedical
111 literature (**Figure 1B**). These trends are independent of significance threshold (**Figure S5**) and
112 (except for CRISPR screens) whether we considered the current scientific literature or literature
113 published before 2003, before any of these articles had been published (**Figure S6**).
114
Understudied genes are least frequently elevated to the title/abstract in transcriptomics
experiments and most frequently elevated to the title/abstract in CRISPR screens. GWAS
studies tend to return better-studied genes as hits; the median hit gene in GWAS studies was
more popular than 75% of genes. Hit genes promoted to the title/abstract in GWAS studies had
a median popularity greater 85% of all protein coding genes. This may explain the prior
observation that the total number of articles on individual genes partially correlates with the
total number of occurrences as a hit in GWAS studies[32].
122
123 Evaluating the final possibility, we found that the reception of -omics studies in later scientific
124 literature either reproduced authors' initial selection of highly studied genes or slightly
125 mitigated it. Jointly, the above findings reinforce that understudied genes become abandoned

126 between the completion of -omics experiments and the reporting of results, rather than being
127 abandoned by later research.
128
129 *Subsequent reception by other scientists does not penalize studies on understudied genes*
130
131 The abandonment of understudied genes could be driven by the valid concern of biomedical
132 researchers that focusing on less-investigated genes will yield articles with lower impact[17], as
133 observed around the turn of the millenium[33]. If this were the case, preemptively avoiding
134 understudied hits would be the rational decision for authors of -omics studies.
135
We thus decided to complement our preceding analysis by an analysis explicitly focused on
citation impact. Notably, we found that the concern of publications on understudied genes
receiving fewer citations does not hold for present-day research on human genes; in biomedical
literature at-large, articles focusing on less-investigated genes accumulate more citations, an
effect that has held consistently since 2001 (**Figure 2).** Important to human health, this also
holds when only considering disease-related fields (**Figure S7**).
142
To rule out that these macroscopic observations stem from us having aggregated over different
diseases, we separately analyzed 602 disease-related MeSH terms. We found 29 MeSH terms
with a statistically significant Spearman correlation using Benjamini-Hochberg FDR < 0.01 (**Table
S4**), of which 27 showed a negative association and only 2 a positive association. This result
suggests that is may actually be rational for most scientists to pursue studies focusing on
understudied genes, although most scientists specializing in a disease may also not receive
more citations when focusing on understudied genes.
150
Returning to our observation that understudied hits from high-throughput assays are not
promoted to the title and abstract of the resulting publication, we next tested if different
experimental approaches demonstrated distinct associations between gene popularity and
citations (**Figure S8**). Among 264 technique-related MeSH terms tested, there were 20 MeSH
terms with a statistically significant Spearman correlation using Benjamini-Hochberg FDR < 0.01
(**Table S5**), of which 16 showed a negative association and only 4 a positive association.
Notably, MeSH terms representing high-throughput techniques (e.g. D055106:Genome-Wide
Association Study and D020869:Gene Expression Profiling) showed no significant association.
This finding suggests that authors of high-throughput studies have little to gain or lose citation-
wise by highlighting understudied genes.
161
To summarize, our investigations detail the previously described separation between "large-
scale" and "small-scale" biological research[34-36]. Authors of high-throughput studies do not
highlight understudied genes in the title or abstract of their publications, the sections of the
publication most accessible to other scientists. While, overall, understudied genes (and high-
throughput assays themselves[5]) correlate with increased citation impact, for high-throughput
studies any potential gain in citations is either absent or too small to be significant. Thus, there
may not be any incentive for authors of high-throughput studies to highlight understudied
genes.

Identification of biological and experimental factors associated with selection of highlighted genes

To illuminate why understudied genes are abandoned between experimental results and the write-up of results, we performed a literature review to identify factors that have been proposed to limit studies of understudied genes (**Table S1**). These factors range from evolutionary factors (e.g., whether a gene only has homologs in primates), to chemical factors (e.g., gene length or hydrophobicity of protein product), to historical factors (e.g., whether a gene's sequence has previously been patented) to materialistic factors affecting experimental design (e.g., whether designed antibodies are robust for immunohistochemistry).

As any of these factors could plausibly affect gene selection within individual domains of biomedical research, we returned to the -omics data described above (**Figure 1**) and measured how much these factors align with the selective highlighting of hit genes in the title or abstract of GWAS, Aff-MS, transcriptomics, and CRISPR studies.

We identified 45 factors that relate to genes and found 35 (14 out of 23 binary factors and 21 out of 22 continuous factors) associated with selection in at least one assay type at $p < 0.001$ (**Figure 3, Table S2,** and **Table S3**). Across the four assay types, the most informative binary factor describes whether there is a plasmid available for a gene in the AddGene plasmid catalog. This might reflect that many different research groups produce reagents surrounding the genes that they actively study. The most informative continuous factor is the number of research articles about a gene, supporting the conclusion that gene popularity drives whether it is highlighted or not (**Figure 1**).

To better understand how all 45 factors are related, we performed a cluster analysis of the collected factors (**Figure 4, Figure S9,** and **Figure S10**). This clustering reflects the fact that many suggested factors influencing the abandonment of understudied genes are not independent. For instance, we find that the number of articles about a gene is heavily correlated with the number of annotations for that gene in all surveyed databases. In another case, gene length is heavily correlated with the number of GWAS annotations for a gene, as described before in terms of transcript length and single-nucleotide polymoprhisms[37].

Study limitations

Our study has several limitations. First, all analysis is subject to annotation errors in the various databases we employ. While these should be rare and not affect our overall findings, they may affect users who are interested in genes with discordant annotations. Second, we focus only on human genes. Different patterns of selection may exist for research on genes in other organisms. Third, our literature review also identified further factors that we could not test more directly because of absent access to fitting data. These are: experts' tendency to deepen their expertise[3], a perceived lack of accuracy of -omics studies[17,38], -omics serving research purposes beyond target gene identification[22], the absence of good protocols for mass

spectrometry[39], the electronic distribution and reading of research articles[40], rates of
reproducibility[17], career prospects of investigators[7,41], and the human tendency to fall back to
simplifying heuristics when making decisions under conditions with uncertainty[42]. Fourth, we
cannot resolve further which specific step between the conduct of an experiment and the
writing of a research article leads to the abandonment of hit genes. Finally, we interpret the
results of high-throughput experiments based on their representation in the NHGRI-EBI GWAS,
BioGRID, EBI-GXA and BioGRID ORCS databases. The authors of the original studies may have
processed their data differently, obtaining different results.

As our present analysis is correlative, it also is tempting to propose controlled trials where
published manuscripts on high-throughput studies randomly report hit genes in the abstract
even if not investigated further by the authors.

Discussion

Efforts to address the gaps in detailed knowledge about most genes have crystallized as
initiatives promoting the investigation of understudied sets of genes[16,43-47], an approach to gene
scholarship recently termed 'unknomics'[48]. We believe that enabling scientists to consciously
engage with bias in research target selection will enable more biomedical researchers to
participate in unknomics, to the potential benefit of their own research impact and towards the
advancement of our collective understanding of the entire human genome.

To achieve this goal, we combined all the above insights to create a tool we denoted *find my*
understudied genes (FMUG). Our literature review revealed several tools and resources aiming
to promote research of understudied genes by publicizing understudied genes[49-56] or by
providing information about hit genes[7,57-61]. However, we noted the absence of tools enabling
scientists to actively engage with factors that align with gene selection. Although such factors
are largely correlated when considering all genes (**Figure 4, Figure S9,** and **Figure S10**), some
cluster together and the influence of specific factors could vary across laboratories. For
instance, scientists could vary in their ability to perform proteomics, or ability to explore
orthologous genes in *C. elegans*, or ability to leverage human population data, or perform
standardized mouse assays.

Our tool makes selection bias explicit, while acknowledging that different laboratories vary in
their techniques and capabilities for follow-up research. Rather than telling scientists about the
existence of biases, FMUG aims to prompt scientists to make bias-aware informed decisions to
identify and potentially tackle important gaps in knowledge that they are well-suited to
address. For this reason, we believe that FMUG will not be of value only to scientists engaging
in high-throughput studies, but also by scientists wishing to mine existing datasets for hit genes
that they would be well-positioned to investigate further.

FMUG takes a list of genes from the user (ostensibly a hit list from a high-throughput -omics
experiment) and provides the kind of information that will allow a user to select genes for
further study.

Users can employ filters that reflect the factors identified in our literature review and
supported by our analysis. The default information provided to users consists of factors that are
representative of the identified clusters (**Figure 4**) and strongly associated with gene selection
in high-throughput experiments (**Figure 3, Table S2,** and **Table S3**). In extended options, users
can select any factor that demonstrated a significant association with the selection of genes.
For instance, a user may need to decide whether loss-of-function intolerant genes should be
considered for further research or not, or whether there should be robust evidence that a gene
is protein-coding. Some of these filters are context aware. For instance, a user may select genes
that have already been studied in the general biomedical literature but not yet within the
literature of their disease of interest.

To provide real-time feedback, users are, in parallel, presented the number of articles about
genes in their initial input list and the number of articles about genes that passed their filters.
Users can then export their filtered list of genes. In the interest of researcher privacy, FMUG
keeps all information local to the user's machine. Usage of FMUG is illustrated in **Figure 5A** and
demonstrated in **Movie S1**. FMUG is developed in Flutter and is available for download at
fmug.amaral.northwestern.edu as a MacOS/Windows app. For the development of custom
software and analytical code, we provide the data underlying FMUG at
github.com/amarallab/fmug_analysis.

To determine the practical usefulness of FMUG to scientists we used an early prototype of
FMUG to identify understudied genes associated with aging. One of these genes was Splicing
factor, proline- and glutamine-rich (*Sfpq*), which had not yet been investigated toward its role in
biological aging. We found *Sfpq* to be transcriptionally downregulated during murine aging.
Others had shown *Sfpq* to be required for the transcriptional elongation of long genes[62]. This
led us to hypothesize that during vertebrate aging, the transcripts of long genes become
downregulated in most tissues **(Figure 5B).** We found this hypothesis to be supported through a
multi-species analysis which we published in December 2022 in Nature Aging[63], with another
group publishing so in January 2023 in Nature Genetics[64], and a third group in iScience in March
2023[65].

Materials and Methods

Genes information

Homo sapiens gene information was downloaded from NCBI Gene on Aug 16, 2022
[ftp.ncbi.nlm.nih.gov/gene/DATA/GENE_INFO/All_Data.gene_info.gz]. Only genes with an
unambiguous mapping of Entrez ID to Ensembl ID were used (n = 36,035). Number of gene
synonyms, protein-coding status, and official gene symbol were derived from this dataset. A
gene symbol was considered undefined if the gene's entry for HGNC gene symbol was "-".

Genes in title/abstract of primary research articles
303
Homo sapiens gene information was downloaded from NCBI Gene on Aug 16, 2022
[ftp.ncbi.nlm.nih.gov/gene/DATA/GENE_INFO/Mammalia/Homo_sapiens.gene_info.gz].
gene2pubmed was download from NCBI Gene on Aug 16, 2022
[ftp.ncbi.nlm.nih.gov/gene/DATA/gene2pubmed.gz][28]. PubTator gene annotations were
downloaded from NIH-NLM on July 12, 2022
[https://ftp.ncbi.nlm.nih.gov/pub/lu/PubTatorCentral/][28,66]. PubMed was downloaded on Dec
17, 2021 [https://ftp.ncbi.nlm.nih.gov/pubmed/baseline/].
311
312 Only using PMIDs annotated as primary research articles, a human gene was considered as
313 mentioned in the title/abstract of the publication if gene was annotated as being in the
314 title/abstract by PubTator and the article appeared in gene2pubmed.
315
316 *CRISPR articles*
317
BioGRID ORCS[26] v1.1.6 was downloaded on April 25, 2022
[https://downloads.thebiogrid.org/BioGRID-ORCS/Release-Archive/BIOGRID-ORCS-1.1.6/]. Any
genome-wide CRISPR knockout screens in human with an associated PubMed ID in which hit
genes were mentioned in the title or abstract was considered (n = 15). 9,268 unique genes were
found as hits. Of these, 18 (0.19%) were elevated to titles/abstracts in the reporting articles and
19 (0.21%) were elevated to titles/abstracts in citing articles. A full list of PubMed IDs is
available in Supplementary File 2.
325
326 *Transcriptomics articles*
327
EBI-GXA[27] release 36 was downloaded on Sep 15, 2020
[https://web.archive.org/web/20201022184159/https://www.ebi.ac.uk/gxa/download]. This is
the most recent release of EBI-GXA available as a bulk download. Any transcriptomics
comparisons with an associated PubMed ID in which hit genes were mentioned in the title or
abstract was considered (n= 148). Analysis was restricted to protein-coding genes (some
screens featured non-protein-coding genes, but this was not common to all analyses). DE was
called at Benjamini-Hochberg FDR $q < 0.05$. 18,295 unique genes were found as hits. Of these,
161 (0.88%) were elevated to titles/abstracts in the reporting articles and 692 (3.78%) were
elevated to titles/abstracts in citing articles. A full list of PubMed IDs is available in
Supplementary File 2.
338
339 *Affinity capture – mass spectrometry articles*
340
BioGRID[26] v3.5.186 was downloaded on April 25, 2022
[https://downloads.thebiogrid.org/BioGRID/Release-Archive/BIOGRID-3.5.186/]. Any
interactions involving a human gene as the prey protein with an experimental evidence code of
'Affinity Capture-MS' labeled as 'High-Throughput' that had an associated PubMed ID in which
hit genes were mentioned in the title or abstract was considered (n= 296). Prey proteins in

these interactions were considered hits. 7,919 unique genes were found as hits. Of these, 311
(3.93%) were elevated to titles/abstracts in reporting articles and 407 (5.14%) were elevated to
titles/abstracts in citing articles. A full list of PubMed IDs is available in Supplementary File 2.
349
350 *GWAS articles*
351
The NHGRI-EBI GWAS catalog[25] (associations and studies) was download on Aug 17, 2022
[https://www.ebi.ac.uk/gwas/docs/file-downloads]. Any GWAS screens with an associated
PubMed ID in which hit genes were mentioned in the title or abstract was considered (n= 450).
Only SNPs occurring within a gene were considered hits. 1,043 unique genes were found as hits.
Of these, 413 (39.6%) were elevated to titles/abstracts in reporting articles and 319 (30.6%)
were elevated to titles/abstracts in citing articles. A full list of PubMed IDs is available in
Supplementary File 2.
359
360 *Citing articles*
361
362 NIH iCite v32 was downloaded on Aug 25, 2022[30]
363 [https://nih.figshare.com/collections/iCite_Database_Snapshots_NIH_Open_Citation_Collectio
364 n_/4586573/32].
365
366 *Functional annotations*
367
368 Mapping of genes to Gene Ontology / Protein Interaction Database / WikiPathways / Reactome
369 / Kyoto Encyclopedia of Genes and Genomes / Human Phenotype Ontology / BioCarta
370 categories was derived from MSigDB v7.5 Entrez ID .gmt files, downloaded on Apr 12, 2022
371 [http://www.gsea-msigdb.org/gsea/downloads_archive.jsp].
372
373 *Between-species homology*
374
375 Homologene Build 68 was used to determine interspecies homology
376 [ftp.ncbi.nih.gov/pub/HomoloGene/build68/]. Human = taxid:9606, mouse = taxid:10090, rat =
377 taxid:10116, c. elegans = taxid:6239, d. melanogaster = taxid:7227, yeast = taxid:559292,
378 zebrafish = taxid:7955.
379
380 *Primate specificity*
381
382 Human genes were considered primate-specific if the only other members of their homology
383 group belonged to primate genomes. Primate taxonomy ids were downloaded from NCBI
384 Taxonomy on Sep 20, 2022
385 [https://www.ncbi.nlm.nih.gov/taxonomy/?term=txid9443[Subtree]].
386
387 *Number of publications in model organisms*
388

389   Gene information was downloaded from NCBI Gene on Aug 16, 2022
390   [ftp.ncbi.nlm.nih.gov/gene/DATA/GENE_INFO/All_Data.gene_info.gz].
391
392   Only using PMIDs annotated as primary research articles, genes was considered as mentioned
393   in the title/abstract of the publication if gene was annotated as being in the title/abstract by
394   PubTator and the article appeared in gene2pubmed.
395
396   Genes in model organisms were mapped to human genes and the number of articles on those
397   mapping to human genes were counted. If a model organism's gene had homology to human
398   but no associated publications, the number of publications was resolved to zero. Otherwise,
399   counts were listed as NA.
400
401   *Mouse phenotype hits*
402
403   International Mouse Phenotyping Consortium data release 17.0 was downloaded on Aug 18,
404   2022 [https://www.mousephenotype.org/data/release]. Mouse genes were matched to human
405   genes with Homologene.
406
407   *Gene Expression Atlas (EBI-GXA)*
408
409   EBI-GXA release 36 was downloaded on Sep 15, 2020
410   [https://web.archive.org/web/20201022184159/https://www.ebi.ac.uk/gxa/download]. This is
411   the most recent release of EBI-GXA available as a bulk download. For probability of DE, only
412   RNA-seq comparisons were considered and DE was called at Benjamini-Hochberg $q < 0.05$.
413
414   *Global RNA expression*
415
416   RNA consensus tissue gene data from HPA release 21.1 was downloaded on Sep 20, 2022
417   [https://www.proteinatlas.org/about/download]. Global RNA expression was estimated by
418   taking the median expression (nTPM) across tissues for each gene and the proportion of tissues
419   with detectable (≥1 nTPM) expression for each gene.
420
421   *Expression in HeLa cells*
422
423   RNA cell line gene data from HPA release 21.1 was downloaded on Sep 20, 2022
424   [https://www.proteinatlas.org/about/download]. Expression is in nTPM.
425
426   *Previous patent activity*
427
Genes with patent activity were defined from Table S1 of Rosenfeld and Mason, 2013[67]. Genes
were mapped with their HGNC symbol. This analysis aligned sequences in patents to the human
genome to estimate patent coverage of human coding sequences. Although this does not
necessarily reflect whether the mapped genes were claimed directly by the patent holder, as

noted by others[68], this analysis remains the most comprehensive available for determining
patent coverage of the human genome.

Druggability

437   Druggable genes were identified from Table S1 of Finan et al., 2017[69]. Genes were mapped with
438   their Ensembl identifier.

Gene length

442   GenBank was downloaded in spring 2017 (genome version GRCh38.p10). Gene length is defined
443   here as the span of the longest transcript on the chromosome. This aligns with the model of
444   gene length used in Stoeger et al., 2018[7].

Solubility

448   SwissProt protein sequences and mapping tables to Entrez GeneIDs were downloaded from
449   Uniprot in spring 2017. Protein GRAVY score (ignoring Pyrrolysine and Selenocysteine) was
450   estimated with BioPython[70].

Loss of function intolerance

454   Data was obtained from Karczewski et al.[71]. pLI scores > 0.9 on main transcripts, as flagged by
455   authors, were considered as highly loss-of-function intolerant as described by Lek et al.[72].

Number of GWAS hits

459   EBI GWAS catalog[25] (associations and studies) was download on Aug 17, 2022
460   [https://www.ebi.ac.uk/gwas/docs/file-downloads]. Loci were mapped to the nearest gene.

Status as understudied protein

464   The Illuminating the Druggable Genome understudied protein list was downloaded on Sep 20,
465   2022
466   [https://github.com/druggablegenome/IDGTargets/blob/master/IDG_TargetList_CurrentVersio
467   n.json].

Human Protein Atlas

HPA release 21.1 was downloaded on Sep 20, 2022 [https://www.proteinatlas.org/search].
Evidence for a protein's existence, as determined by NeXtProt, HPA, or UniProt was resolved as
True if the respective evidence entry was annotated as "Evidence at protein level". Status as a
membrane protein was determined by whether the 'Protein class' column contained the string

'membrane protein'. Antibodies were considered available for each protein if the protein's
entry in the 'Antibody' column was not null.
477
478 *Availability of plasmids*
479
480 The AddGene plasmid catalog was downloaded on Aug 12, 2022
481 [https://www.addgene.org/browse/gene/gene-list-data/?_=1666368044314].
482
483 *Availability of compounds*
484
485 The catalog of gene targets was downloaded from ChEMBL on Sep 20, 2022
486 [https://www.ebi.ac.uk/chembl/g/#browse/targets]. UniProt IDs were converted to Entrez IDs
487 to identify which human genes were affected by any compound.
488
489 *Mendelian inheritance*
490
491 Autosomal dominant [https://hpo.jax.org/app/browse/term/HP:0000006] and autosomal
492 recessive [https://hpo.jax.org/app/browse/term/HP:0000007] inherited disease-gene
493 associations were downloaded from the Human Phenotype Ontology on Sep 20, 2022. Genes
494 were considered to have evidence of Mendelian inheritance if they appeared in these lists of
495 associations.
496
497 *Code*
498
499 Code for analysis is available at github.com/amarallab/fmug_analysis. Code for FMUG is
500 available at github.com/amarallab/fmug.
501

502
Figure 1: While understudied genes appear often as hits high-throughput -omics experiments,
they are seldom highlighted by authors. a, Conceptual diagram depicting possible points of
abandonment for understudied genes in studies using high-throughput -omics experiments. **b,**
Bibliometric data reveals that understudied genes are frequently hits in -omics experiments but
are not typically highlighted in the title/abstract of reporting articles, nor in the title/abstract or
articles citing reporting articles. We considered articles reporting on genome-wide CRISPR
screens (CRISPR, n=15 articles), transcriptomics (T-omics, n=148 articles), affinity capture –
mass spectrometry (Aff-MS, n=296 articles), and GWAS (n=450 articles). Numbers to the right of
each box plot indicate the percentile (in terms of number of articles about that gene) of all
genes exceeded by the median gene in each box plot. ** denotes $p < 0.01$ and *** denotes $p <$
0.001 by two-sided Mann-Whitney U test, comparing genes highlighted in title/abstract to
genes present in hit lists.
515

Figure 2: Articles focusing on less popular genes tend to accrue more citations. a, Density plot
shows correlation between articles per gene before 2015 and median citations to articles
published in 2015. Contours correspond to deciles in density. Solid red line shows locally
weighted scatterplot smoothing (LOWESS) regression. ρ is Spearman rank correlation and p the
significance values of the Spearman rank correlation as described by Kendall and Stuart[73]. **b,**
Spearman correlation of previous gene popularity (i.e. number of articles) to median citations
per year since 1990. Solid blue line indicates nominal Spearman correlation, shaded region
indicates bootstrapped 95% confidence interval (n=1,000). Only articles with a single gene in
the title/abstract are considered, excluding the 30.4% of gene-focused studies which feature
more than one gene in the title/abstract.

Figure 3: We evaluated which gene-related factors are associated with elevation to the
title/abstract of an article featuring a high-throughput experiment. **a)** Association between
factors with binary (True/False) identities and highlighting hits in title/abstract of reporting
articles. Values represent the odds ratio between hits in the collected articles and hits
mentioned in the title or abstract of collected articles (e.g. hits with a compound known to
affect gene activity are 5.114 times as likely to be mentioned in the title/abstract in an article

using transcriptomics). Collected articles are described in **Figure 1B** and **Figures S5** and **S6.** 95%
confidence interval of odds ratio is shown in parentheses. * = $p < 0.05$, ** = $p < 0.01$, and *** =
$p < 0.001$ by two-sided Fisher exact test. Results are shown numerically in **Table S2**. For
consistency between studies, hits were restricted to protein-coding genes. Thus, status as a
protein-coding gene could not be tested. †No genes without a defined HUGO symbol were
found as hits in GWAS or transcriptomics studies. **b)** Association with factors with continuous
identities and highlighting hits in title/abstract of reporting articles. Values represent F, the
common-language effect size (equivalent to AUROC, where ~0.5 indicates little effect, >0.5
indicates positive effect and <0.5 indicates negative effect) of being mentioned in the
titles/abstracts of the collected articles described in **Figure 1B** and **Figures S5** and **S6.** * = $p <$
0.05, ** = $p < 0.01$, and *** = $p < 0.001$ by two-sided Mann-Whitney U test. Results are shown
numerically in **Table S3**.
546
547

548
Figure 4: Clustermap showing collected factors across all human protein-coding genes. Factors
are shown along the x axis, with genes along the y axis. Eight factors, representing the default
factors we selected for FMUG, are shown (all factors are shown in **Figure S7** and **Figure S8**).
Binary factors are coded to 0 (purple) and 1 (white), while continuous factors are ranked from 0
to 1 with ties resolved to minimum rank. Clustering was performed with Ward's method for
hierarchical clustering[74].

Figure 5: We created FMUG to help researchers identify understudied genes among their genes of interest and characterize their tractability for future research. a, Diagram describing use of FMUG. **b,** An early prototype of FMUG led us to the hypothesis that transcript length negatively correlates with up-regulation during aging. First, we identified genes that strongly associate with age-dependent transcriptional change across multiple cohorts. We then performed a literature review for each of these genes to identify the most direct way the genes (or evolutionarily closely related genes or functionally closely related partner proteins) had been studied in aging. 64% had been functionally investigated in aging, 15% shown to change a measure of gene expression, 3% functionally investigated in a biological domain close to aging (such as senescence), and 5% shown to change a measure of gene expression in a biological

domain close to aging. For genes reported by others to change expression with age, we identified tissues in which transcripts of the genes change during aging. We computed 'feasibility scores' scientific strategies (GEM: G: strong genetic support, E: and experimental potential, M: homolog in invertebrate model organism) as described by Stoeger et al.[7] and total number of publications in MEDLINE. Splicing factor, proline- and glutamine-rich (*Sfpq*) had previously been demonstrated by Takeuchi et al. to be required for the transcriptional elongation of long genes[62]. When performing a data-driven analysis of factors that could possibly explain age-dependent changes of the entire transcriptome, we thus included gene and transcript lengths, and subsequently found them to be more informative than transcription factors or microRNAs[63].

Data Availability

All underlying data for figures are available at github.com/amarallab/fmug_analysis.

Acknowledgements

We thank Xiaojing Sui for testing FMUG and Northwestern Information Technology for technical assistance. RAKR was supported in part by the National Institutes of Health Training Grant (T32GM008449) through Northwestern University's Biotechnology Training Program. RAKR also acknowledges support from the Dr. John N. Nicholson fellowship from Northwestern University and Moderna Inc., "Identifying bias and improving reproducibility in RNA-seq computational pipelines". LANA was supported by NSF 1956338, NIH U19AI135964 and Simons Foundation DMS-1764421. TS was supported by NIH K99AG068544. We thank Alexander Misharin, Richard Morimoto, and Scott Budinger for feedback on an early prototype of FMUG which we used as part of our shared research into the biology of aging.

References

Hoffmann, R. & Valencia, A. Life cycles of successful genes. *Trends Genet.* **19**, 79-81, doi:10.1016/S0168-9525(02)00014-8 (2003).

Su, A. I. & Hogenesch, J. B. Power-law-like distributions in biomedical publications and research funding. *Genome Biol.* **8**, 404, doi:10.1186/gb-2007-8-4-404 (2007).

Edwards, A. M. *et al.* Too many roads not taken. *Nature* **470**, 163-165, doi:10.1038/470163a (2011).

Gillis, J. & Pavlidis, P. Assessing identity, redundancy and confounds in Gene Ontology annotations over time. *Bioinformatics* **29**, 476-482, doi:10.1093/bioinformatics/bts727 (2013).

Stoeger, T. & Nunes Amaral, L. A. The characteristics of early-stage research into human genes are substantially different from subsequent research. *PLoS Biol* **20**, e3001520, doi:10.1371/journal.pbio.3001520 (2022).

Grueneberg, D. A. *et al.* Kinase requirements in human cells: I. Comparing kinase requirements across various cell types. *Proc. Natl. Acad. Sci. U. S. A.* **105**, 16472-16477, doi:10.1073/pnas.0808019105 (2008).

7   Stoeger, T., Gerlach, M., Morimoto, R. I. & Nunes Amaral, L. A. Large-scale investigation
of the reasons why potentially important genes are ignored. *PLoS Biol* **16**, e2006643,
doi:10.1371/journal.pbio.2006643 (2018).
8   Riba, M. *et al.* Revealing the acute asthma ignorome: characterization and validation of
uninvestigated gene networks. *Sci Rep* **6**, 24647, doi:10.1038/srep24647 (2016).
9   Haynes, W. A., Tomczak, A. & Khatri, P. Gene annotation bias impedes biomedical
research. *Sci Rep* **8**, 1362, doi:10.1038/s41598-018-19333-x (2018).
10 Border, R. *et al.* No support for historical candidate gene or candidate gene-by-
interaction hypotheses for major depression across multiple large samples. *American
Journal of Psychiatry* **176**, 376-387 (2019).
11 Stoeger, T. & Nunes Amaral, L. A. COVID-19 research risks ignoring important host genes
due to pre-established research patterns. *Elife* **9**, doi:10.7554/eLife.61981 (2020).
12 Zhang, D. *et al.* Incomplete annotation has a disproportionate impact on our
understanding of Mendelian and complex neurogenetic disorders. *Science Advances* **6**,
eaay8299 (2020).
13 Byrne, J. A. *et al.* Protection of the human gene research literature from contract
cheating organizations known as research paper mills. *Nucleic Acids Research* **50**, 12058-
12070, doi:10.1093/nar/gkac1139 (2022).
14 Collins, F. S., Green, E. D., Guttmacher, A. E., Guyer, M. S. & Institute, U. S. N. H. G. R. A
vision for the future of genomics research. *Nature* **422**, 835-847,
doi:10.1038/nature01626 (2003).
15 Shendure, J., Findlay, G. M. & Snyder, M. W. Genomic Medicine-Progress, Pitfalls, and
Promise. *Cell* **177**, 45-57, doi:10.1016/j.cell.2019.02.003 (2019).
16 Lloyd, K. C. K. *et al.* The Deep Genome Project. *Genome Biol* **21**, 18, doi:10.1186/s13059-
020-1931-9 (2020).
17 Kustatscher, G. *et al.* Understudied proteins: opportunities and challenges for functional
proteomics. *Nat Methods* **19**, 774-779, doi:10.1038/s41592-022-01454-x (2022).
18 Rodriguez-Esteban, R. & Jiang, X. Differential gene expression in disease: a comparison
between high-throughput studies and the literature. *BMC Med Genomics* **10**, 59,
doi:10.1186/s12920-017-0293-y (2017).
19 Oprea, T. I. *et al.* Unexplored therapeutic opportunities in the human genome. *Nat Rev
Drug Discov* **17**, 317-332, doi:10.1038/nrd.2018.14 (2018).
20 Sinha, S., Eisenhaber, B., Jensen, L. J., Kalbuaji, B. & Eisenhaber, F. Darkness in the
Human Gene and Protein Function Space: Widely Modest or Absent Illumination by the
Life Science Literature and the Trend for Fewer Protein Function Discoveries Since 2000.
Proteomics **18**, e1800093, doi:10.1002/pmic.201800093 (2018).
21 Wood, V. *et al.* Hidden in plain sight: what remains to be discovered in the eukaryotic
proteome? *Open Biol* **9**, 180241, doi:10.1098/rsob.180241 (2019).
22 Donohue, C. & Love, A. Perspectives on the Human Genome Project and genomics.
Minnesota Studies in the Philosophy of Science **23** (in press).
23 Pena-Castillo, L. & Hughes, T. R. Why are there still over 1000 uncharacterized yeast
genes? *Genetics* **176**, 7-14, doi:10.1534/genetics.107.074468 (2007).
24 Ellens, K. W. *et al.* Confronting the catalytic dark matter encoded by sequenced
genomes. *Nucleic Acids Res* **45**, 11495-11514, doi:10.1093/nar/gkx937 (2017).

25   Buniello, A. *et al.* The NHGRI-EBI GWAS Catalog of published genome-wide association
studies, targeted arrays and summary statistics 2019. *Nucleic Acids Res* **47**, D1005-
D1012, doi:10.1093/nar/gky1120 (2019).
26   Oughtred, R. *et al.* The BioGRID database: A comprehensive biomedical resource of
curated protein, genetic, and chemical interactions. *Protein Science* **30**, 187-200 (2021).
27   Papatheodorou, I. *et al.* Expression Atlas: gene and protein expression across multiple
studies and organisms. *Nucleic acids research* **46**, D246-D251 (2018).
28   Maglott, D., Ostell, J., Pruitt, K. D. & Tatusova, T. Entrez Gene: gene-centered
information at NCBI. *Nucleic acids research* **35**, D26-D31 (2007).
29   Wei, C.-H., Allot, A., Leaman, R. & Lu, Z. PubTator central: automated concept
annotation for biomedical full text articles. *Nucleic acids research* **47**, W587-W593
(2019).
30   Hutchins, B. I., Santangelo, George. *iCite*, <https://doi.org/10.35092/yhjc.c.4586573 >
(2019).
31   Stoeger, T. & Nunes Amaral, L. A. COVID-19 research risks ignoring important host genes
due to pre-established research patterns. *Elife* **9**, e61981 (2020).
32   Stoeger, T., Gerlach, M., Morimoto, R. I. & Nunes Amaral, L. A. Large-scale investigation
of the reasons why potentially important genes are ignored. *PLoS biology* **16**, e2006643
(2018).
33   Pfeiffer, T. & Hoffmann, R. Temporal patterns of genes in scientific publications. *Proc.*
Natl. Acad. Sci. U. S. A. **104**, 12052-12056, doi:10.1073/pnas.0701315104 (2007).
34   Knorr Cetina, K. *Epistemic Cultures*. (Harvard University Press, 1999).
35   Alberts, B. M. Limits to growth: In biology, small science is good science. *Cell*
(Cambridge) **41**, 337-338 (1985).
36   Richardson, S. S. a. S., Hallam.   (Duke University Press, 2015).
37   Lopes, I., Altab, G., Raina, P. & De Magalhães, J. P. Gene size matters: an analysis of gene
length in the human genome. *Frontiers in Genetics* **12**, 559998 (2021).
38   Brown, L. A. & Peirson, S. N. Improving Reproducibility and Candidate Selection in
Transcriptomics Using Meta-analysis. *Journal of Experimental Neuroscience* **12**,
1179069518756296, doi:10.1177/1179069518756296 (2018).
39   Cesar-Razquin, A. *et al.* A Call for Systematic Research on Solute Carriers. *Cell* **162**, 478-
487, doi:10.1016/j.cell.2015.07.022 (2015).
40   Evans, J. A. Electronic publication and the narrowing of science and scholarship. *Science*
321, 395-399, doi:10.1126/science.1150473 (2008).
41   Alberts, B., Kirschner, M. W., Tilghman, S. & Varmus, H. Rescuing US biomedical
research from its systemic flaws. *Proc Natl Acad Sci U S A* **111**, 5773-5777,
doi:10.1073/pnas.1404402111 (2014).
42   Gilovich, T., Griffin, D. W. & Kahneman, D. *Heuristics and biases : the psychology of*
intuitive judgment. (Cambridge University Press, 2002).
43   Gerlt, J. A. *et al.* The Enzyme Function Initiative. *Biochemistry* **50**, 9950-9962,
doi:10.1021/bi201312u (2011).
44   Carter, A. J. *et al.* Target 2035: probing the human proteome. *Drug Discov Today* **24**,
2111-2115, doi:10.1016/j.drudis.2019.06.020 (2019).

45   Rodgers, G. *et al.* Glimmers in illuminating the druggable genome. *Nat Rev Drug Discov*
17, 301-302, doi:10.1038/nrd.2017.252 (2018).
46   Kustatscher, G. *et al.* An open invitation to the Understudied Proteins Initiative. *Nat*
Biotechnol **40**, 815-817, doi:10.1038/s41587-022-01316-z (2022).
47   *EUbOPEN*, <https://www.eubopen.org/> (
48   Rocha, J. J. *et al.* Functional unknomics: Systematic screening of conserved genes of
unknown function. *PLoS biology* **21**, e3002222 (2023).
49   Duek, P., Gateau, A., Bairoch, A. & Lane, L. Exploring the Uncharacterized Human
Proteome Using neXtProt. *J Proteome Res* **17**, 4211-4226,
doi:10.1021/acs.jproteome.8b00537 (2018).
50   Crow, M., Lim, N., Ballouz, S., Pavlidis, P. & Gillis, J. Predictability of human differential
gene expression. *Proc Natl Acad Sci U S A* **116**, 6491-6500,
doi:10.1073/pnas.1802973116 (2019).
51   Perdigao, N. & Rosa, A. Dark Proteome Database: Studies on Dark Proteins. *High*
Throughput **8**, doi:10.3390/ht8020008 (2019).
52   Essegian, D., Khurana, R., Stathias, V. & Schurer, S. C. The Clinical Kinase Index: A
Method to Prioritize Understudied Kinases as Drug Targets for the Treatment of Cancer.
Cell Rep Med **1**, 100128, doi:10.1016/j.xcrm.2020.100128 (2020).
53   Sheils, T. K. *et al.* TCRD and Pharos 2021: mining the human proteome for disease
biology. *Nucleic Acids Res* **49**, D1334-D1346, doi:10.1093/nar/gkaa993 (2021).
54   Rocha, J. *et al.* Functional unknomics: closing the knowledge gap to accelerate
biomedical research. *bioRxiv* (2022).
55   Higgins, D. P., Weisman, C. M., Lui, D. S., D'Agostino, F. A. & Walker, A. K. Defining
characteristics and conservation of poorly annotated genes in Caenorhabditis elegans
using WormCat 2.0. *Genetics* **221**, doi:10.1093/genetics/iyac085 (2022).
56   Wainberg, M. *et al.* A genome-wide atlas of co-essential modules assigns function to
uncharacterized genes. *Nat Genet* **53**, 638-649, doi:10.1038/s41588-021-00840-z (2021).
57   Rebhan, M., Chalifa-Caspi, V., Prilusky, J. & Lancet, D. GeneCards: a novel functional
genomics compendium with automated data mining and query reformulation support.
Bioinformatics **14**, 656-664, doi:10.1093/bioinformatics/14.8.656 (1998).
58   Tan, J. *et al.* ADAGE signature analysis: differential expression analysis with data-defined
gene sets. *Bmc Bioinformatics* **18**, 512, doi:10.1186/s12859-017-1905-4 (2017).
59   Kustatscher, G. *et al.* Co-regulation map of the human proteome enables identification
of protein functions. *Nat Biotechnol* **37**, 1361-1371, doi:10.1038/s41587-019-0298-5
(2019).
60   Wu, T. *et al.* clusterProfiler 4.0: A universal enrichment tool for interpreting omics data.
Innovation (Camb) **2**, 100141, doi:10.1016/j.xinn.2021.100141 (2021).
61   Jiang, J. *et al.* Systematic illumination of druggable genes in cancer genomes. *Cell Rep*
38, 110400, doi:10.1016/j.celrep.2022.110400 (2022).
62   Takeuchi, A. *et al.* Loss of Sfpq Causes Long-Gene Transcriptopathy in the Brain. *Cell Rep*
23, 1326-1341, doi:10.1016/j.celrep.2018.03.141 (2018).
63   Stoeger, T. *et al.* Aging is associated with a systemic length-associated transcriptome
imbalance. *Nature Aging* **2**, 1191-1206, doi:10.1038/s43587-022-00317-6 (2022).

64   Gyenis, A. *et al.* Genome-wide RNA polymerase stalling shapes the transcriptome during
741   aging. *Nat Genet* **55**, 268-279, doi:10.1038/s41588-022-01279-6 (2023).

742   65   Ibañez-Solé, O., Barrio, I. & Izeta, A. Age or lifestyle-induced accumulation of
743   genotoxicity is associated with a length-dependent decrease in gene expression.
744   *iScience*, 106368, doi:https://doi.org/10.1016/j.isci.2023.106368 (2023).

745   66   Wei, C. H., Allot, A., Leaman, R. & Lu, Z. PubTator central: automated concept
746   annotation for biomedical full text articles. *Nucleic Acids Res* **47**, W587-W593,
747   doi:10.1093/nar/gkz389 (2019).

748   67   Rosenfeld, J. A. & Mason, C. E. Pervasive sequence patents cover the entire human
749   genome. *Genome Med.* **5**, 27, doi:10.1186/gm431 (2013).

750   68   Tu, S. *et al.* Response to 'pervasive sequence patents cover the entire human genome'.
751   *Genome medicine* **6**, 1-3 (2014).

752   69   Finan, C. *et al.* The druggable genome and support for target identification and
753   validation in drug development. *Science translational medicine* **9**, eaag1166 (2017).

754   70   Cock, P. J. A. *et al.* Biopython: freely available Python tools for computational molecular
755   biology and bioinformatics. *Bioinformatics* **25**, 1422-1423,
756   doi:10.1093/bioinformatics/btp163 (2009).

757   71   Karczewski, K. J. *et al.* The mutational constraint spectrum quantified from variation in
758   141,456 humans. *Nature* **581**, 434-443, doi:10.1038/s41586-020-2308-7 (2020).

759   72   Lek, M. *et al.* Analysis of protein-coding genetic variation in 60,706 humans. *Nature* **536**,
760   285-291, doi:10.1038/nature19057 (2016).

761   73   Kendall, M. G. & Stuart, A. Inference and Relationship *The Advanced Theory of Statistics*
762   **2** (1973).

763   74   Ward Jr, J. H. Hierarchical grouping to optimize an objective function. *Journal of the*
764   *American statistical association* **58**, 236-244 (1963).

765