

## 1 **IgG4 serum levels are not elevated in cases of Post-COVID syndrome**

2 Jonas Abel<sup>1†</sup>, Annika J. Walter<sup>2†</sup>, Vivian Glück<sup>1†</sup>, Clara L. Magnus<sup>1†</sup>, Thomas Glück<sup>3</sup>, Philipp  
3 Schuster<sup>2</sup>, Stefan Blaas<sup>4</sup>, Ida Montanari<sup>5</sup>, Michael Koller<sup>6</sup>, Arno Mohr<sup>4</sup>, Thilo Hinterberger<sup>5</sup>,  
4 Bernd Salzberger<sup>7</sup>, Kerstin Renner<sup>8</sup>, Matthias Mack<sup>8</sup>, Robert Bals<sup>9</sup>, Tina Schmidt<sup>10</sup>, Verena  
5 Klemis<sup>10</sup>, Martina Sester<sup>10</sup>, Romina Kardashi<sup>11</sup>, Katja de With<sup>11</sup>, Thomas H. Loew<sup>5</sup>, Maximilian  
6 Malfertheiner<sup>4</sup>, Michael Pfeifer<sup>4</sup>, André Gessner<sup>1,2</sup>, Barbara Schmidt<sup>1,2</sup>, Daniel  
7 Schmalenberger<sup>4</sup>, and David Peterhoff<sup>1,2\*</sup> on behalf of the COVIDYS consortium

8 \* corresponding author: david.peterhoff@ur.de

9 † equal contribution

10 <sup>1</sup> Institute for Clinical Microbiology and Hygiene, University Hospital Regensburg, Regensburg,  
11 Germany

12 <sup>2</sup> Institute for Medical Microbiology and Hygiene, University of Regensburg, Regensburg, Germany

13 <sup>3</sup> Clinic Trostberg, Trostberg, Germany

14 <sup>4</sup> Department of Pneumology, Clinic Donaustauf, Donaustauf, Germany

15 <sup>5</sup> Department of Psychosomatic Medicine, University Hospital Regensburg, Regensburg, Germany

16 <sup>6</sup> Center for Clinical Studies, University Hospital Regensburg, Regensburg, Germany

17 <sup>7</sup> Department of Infection Control and Infectious Disease, University Hospital Regensburg,  
18 Regensburg, Germany

19 <sup>8</sup> Department of Nephrology, University Hospital Regensburg, Regensburg, Germany

20 <sup>9</sup> Department of Internal Medicine V, Pneumology and Intensive Care Medicine, Saarland University,  
21 Homburg/Saar, Germany

22 <sup>10</sup> Department of Transplant and Infection Immunology, Saarland University, Homburg/Saar, Germany

23 <sup>11</sup> Division of Infectious Diseases, University Hospital Carl Gustav Carus, Technische Universität  
24 Dresden, Dresden, Germany

25 **Abstract**

26 Recently, unexpectedly high virus-specific IgG4 levels were reported after more than two  
27 mRNA vaccinations. Class switch towards IgG4 occurs after long-term antigen exposure,  
28 downregulates immune responses and is associated with several autoimmune diseases.

29 Here, we examined differences in antigen-specific IgG subtypes in serum samples from 64  
30 Post-COVID patients and an equally sized cohort of convalescent controls.

31 In both cohorts, the relative amounts of spike protein-specific IgG subtypes were comparable.  
32 IgG1 was the most frequent, followed by IgG3, IgG2, and IgG4. A difference between cohorts  
33 was observed only for IgG2, which was significantly lower in the Post-COVID cohort. Further  
34 analysis of the reactive IgG4 revealed a small but significant difference for the spike protein  
35 receptor-binding domain but not for the spike ectodomain.

36 Since the total IgG4 levels are very low, we do not expect a biologically relevant role in Post-  
37 COVID syndrome. However, reduced virus-specific IgG2 levels could contribute to the  
38 persistence of SARS-CoV-2, causing chronic inflammation in the setting of Post-COVID  
39 syndrome.

40

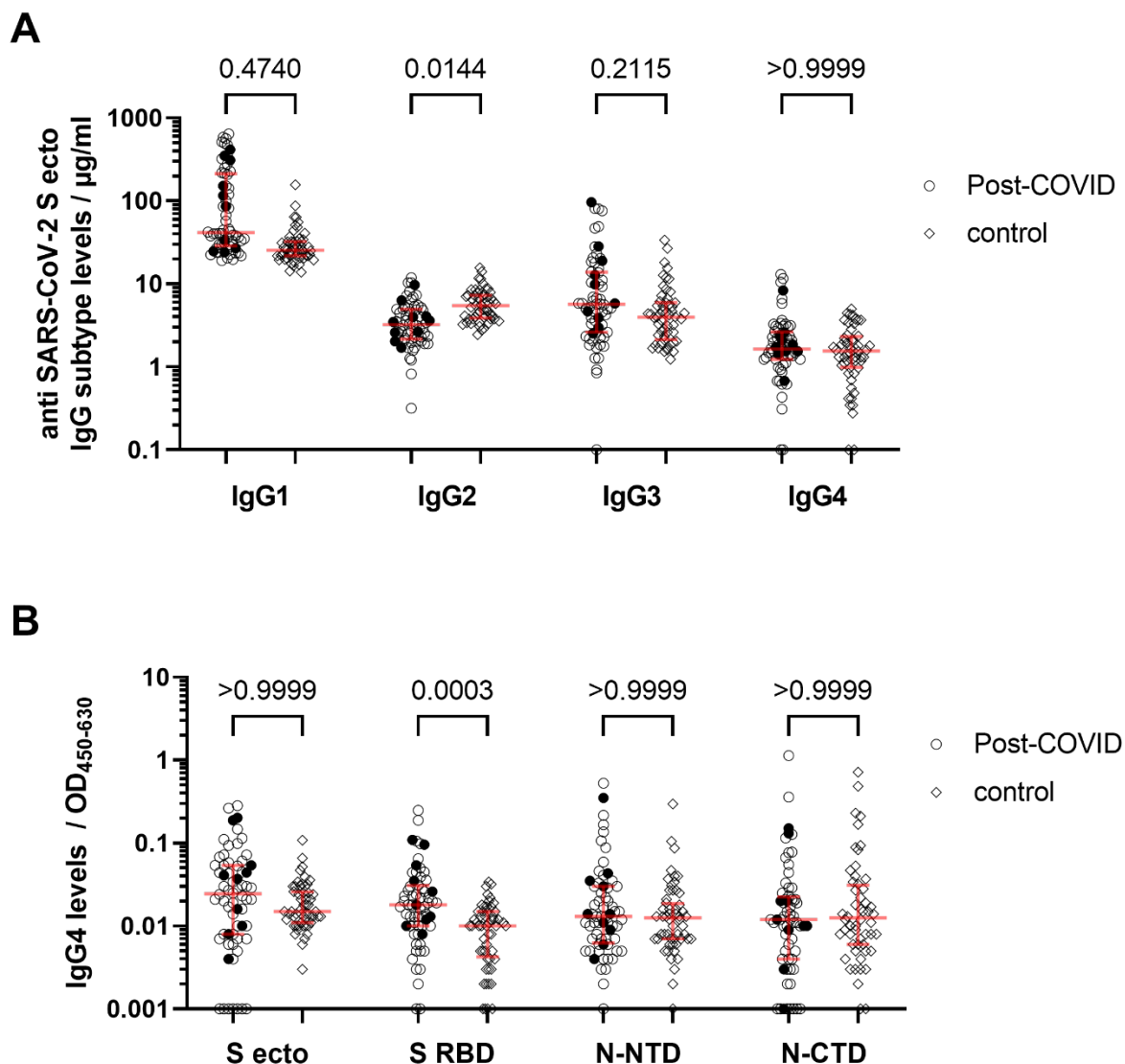
41 Numerous studies have recently described levels and durability of infection- or vaccination-  
42 induced SARS-CoV-2-specific serum antibodies as a correlate of protection. In addition to their  
43 virus-neutralizing effect through receptor competition and stabilization of the viral spike fusion  
44 machinery, the importance of their effector functions is currently debated (1). Effector functions  
45 are closely linked to the four antibody subtypes (IgG1, -2, -3, -4), i.e. sequence, structure and  
46 glycosylation of the antibodies' fragment crystallizable region (Fc) (2).

47 In this context, two independent publications recently described the finding of significantly  
48 increased IgG4 levels after more than two mRNA vaccinations, which was not seen in subjects  
49 vaccinated with adenoviral vectors (3,4). Antibodies of the IgG4 subtype occur after long-term  
50 antigen exposure, downregulating immune responses and inducing tolerance (2).  
51 Furthermore, IgG4 antibodies are associated with several autoimmune diseases (5). The  
52 authors suggested that mRNA-vaccine induced elevated IgG4-levels may be due to a  
53 prolonged availability of antigen in the germinal centers of the lymph nodes after mRNA  
54 vaccination or to a specific property of the mRNA vaccine. It is currently unknown whether  
55 IgG4 is also induced after SARS-CoV-2 infection in non-vaccinated individuals. Furthermore,  
56 the pathogenic potential of IgG4 in the context of SARS-CoV-2 vaccination and COVID-19 is  
57 unclear. This prompted us to investigate whether increased IgG4 levels also occur in patients  
58 suffering from Post-COVID syndrome. In these patients, prolonged circulation of viral antigen  
59 has been described (6). Increased class-switch of B-cells towards IgG4 might result in less Fc-  
60 mediated effector function and thus contribute to longer viral persistence and prolonged  
61 antigen circulation.

62 We examined this hypothesis in cohorts of 64 Post-COVID patients (20 males, 44 females;  
63 median age of 39.5 years, interquartile range [IQR] 30.0 to 50.3 years) and 64 COVID-19  
64 convalescent subjects (26 males, 38 females; median age of 37 years, IQR 30.8 to 48.0).  
65 Individuals in the two cohorts were infected in the early pandemic (March 2020 to May 2021  
66 for Post-COVID patients and February to May 2020 for the control group), when only Wuhan-  
67 like (D614G) and Alpha strain infections were present (7). Approval was obtained from the

68 ethical committee of the Faculty for Medicine, University of Regensburg, Germany (Ref.no. 20-  
69 1785-101 and 20-1896-101). Post-COVID and control cohorts were matched regarding sex,  
70 age and time interval from symptom onset of COVID-19 to serum donation, which took place  
71 at a median of seven months post symptom onset (IQR of 5.0 to 9.0 and 6.5 to 7 months,  
72 respectively). A total of 10 out of 64 Post-COVID patients (15.6%) were vaccinated at the time  
73 of serum sampling. The majority of Post-COVID patients suffered from fatigue (88.5%),  
74 dyspnea (81.0%), and cognitive and memory impairments (71.4%).

75 For detection of anti-Spike antibodies of the different IgG subtypes we used a SARS-CoV-2  
76 pre-fusion stabilized spike protein ectodomain (S ecto)-based ELISA (8). To be able to  
77 compare the different levels of IgG subtypes, we established four subtype variants of the  
78 monoclonal antibody CR3022 as standard for the respective subtype-specific ELISA (9).



79

80 **Figure 1: Serum IgG subtype levels in COVID-19 convalescent (n=64) and Post-COVID subjects**  
81 **(n=64).** Serum levels of vaccinated Post-COVID patients (n=10) are indicated by filled circles. Medians  
82 and IQRs are represented in red. P-values of the Kruskal-Wallis test adjusted by Dunn's *post-hoc* test  
83 are given for selected comparisons indicated by horizontal brackets. **(A)** Serum levels of IgG subtype 1-  
84 4 were quantified separately by comparison to individual subtype variants of the monoclonal antibody  
85 CR3022. The SARS-CoV-2 stabilized spike protein ectodomain (S ecto) was coated on the plate and  
86 binding of patient sera was detected by IgG subtype-specific conjugates. Serum concentration  
87 values  $\leq 0$  µg/ml were set to 0.1 µg/ml to allow for logarithmic scaling of the axis. **(B)** Antigen-specific  
88 IgG4 levels were determined after prolonged development of the ELISA for SARS-CoV-2's S ecto,  
89 receptor-binding domain (RBD), and nucleocapsid N- and C-terminal domain (NTD and CTD). OD<sub>450-600</sub>  
90 values  $\leq 0$  were set to 0.001 to allow for logarithmic scaling of the axis. For both analyses, P values  
91 changed only marginally when vaccinated Post-COVID patients were excluded.

92 We found differences in the absolute levels of the four IgG subtypes for both cohorts  
93 **(Figure 1 A).** IgG1 is by far the most abundant subtype with median concentrations of  
94 41.4 µg/ml (IQR of 29.5 to 209.4 µg/ml) and 25.5 µg/ml (IQR of 21.9 to 32.5 µg/ml) for the

95 Post-COVID and control cohort. These serum concentrations match well with the  
96 concentrations described by Irrgang *et al.* after one immunization with the mRNA vaccine  
97 Comirnaty (median 17.3 µg/ml with IQR of 6.9 to 21.7 µg/ml). IgG2 and IgG3 levels were similar  
98 in both cohorts (medians of 3.2 µg/ml [IQR of 2.2 to 4.9 µg/ml] and 5.5 µg/ml [IQR of 3.9 to 7.3  
99 µg/ml] for IgG2 and 5.7 µg/ml [IQR of 2.6 to 13.7 µg/ml] and 4.0 µg/ml [IQR of 2.2 to 5.9 µg/ml]  
100 for IgG3 in the Post-COVID and control cohort, respectively). However, the levels in our study  
101 were higher than those detected by Irrgang *et al.* after one immunization with Comirnaty  
102 (medians 1.3 µg/ml and 1.0 µg/ml for IgG2 and IgG3 with IQRs of 0.9 to 2.0 µg/ml and 0.6 to  
103 2.1 µg/ml respectively). Levels of the IgG4 subtype were lowest in both cohorts, with a median  
104 of 1.6 µg/ml (IQR of 1.2 to 2.6 µg/ml for the Post-COVID and 1.0 to 2.3 µg/ml for the control  
105 cohort respectively), whereas no IgG4 was detectable with the assay described by Irrgang *et*  
106 *al.*. Only the difference in IgG2 between the two cohorts reached significance at an alpha level  
107 of 0.05. IgG2 has been linked to the recognition of polysaccharide structures of bacterial cell  
108 walls as well as human endogenous glycan epitopes (2,10). It is conceivable that less efficient  
109 recognition of SARS-CoV-2 S antigen glycans could contribute to prolonged antigen  
110 circulation.

111 To further investigate the impact of IgG4 antibodies in the Post-COVID context, we analyzed  
112 their reactivity against the S ecto protein, its receptor-binding domain (RBD) and the N- and C-  
113 terminal domain of the nucleocapsid-protein of SARS-CoV-2. In this assay we extended the  
114 development time of the ELISA from 4 min to 20 min to amplify the initially low signals. A small  
115 but significant difference was detected for the RBD (**Figure 1 B**), but not for the spike  
116 ectodomain and nucleocapsid proteins. Whether this difference between the cohorts reflects  
117 a pathomechanistic factor in the development of the Post-COVID syndrome is an open  
118 question. As the overall level of IgG4 is very low, we do not expect a biologically relevant role  
119 of minor differences. However, an additive effect in interaction with other factors or an impact  
120 on different subtypes of Post-COVID syndrome cannot be excluded. Furthermore, IgG4 is  
121 apparently not an appropriate biomarker for Post-COVID but may be predictive at an earlier

122 time point, e.g. during acute infection. We were unable to answer this question because serum  
123 samples from an early post-infection time point were not available for the Post-COVID cohort.

124 In summary, our analysis provides insights in the relative distribution of Spike-specific IgG  
125 subtypes in Post-COVID patients and convalescent subjects not vaccinated before infection,  
126 as well as in the differences between the two cohorts and antigen-dependent differences for  
127 IgG4. Based on our dataset we do not expect IgG4 to be a central parameter in the  
128 development of Post-COVID. Rather, low IgG2 levels could contribute to the prolonged  
129 presence of SARS-CoV-2 antigens driving chronic inflammation and thus persistent Post-  
130 COVID symptoms.

131

132

### 133 **Author contributions:**

134 DP and BSc were responsible for the conception and design of the study. The Post-COVID  
135 cohort was assembled and samples generated by DS, AW, CLM, IM, AM, SB, and MP. VG,  
136 TG and DP contributed the control cohort samples. DP carried out the experimental design.  
137 JA and DP collected data. AW, CM, BSc and DP analysed and interpreted the data. Clinical  
138 symptoms of post-COVID patients were characterized by DS, TH, RB, RK, MMaI, TL, and KdW  
139 with valuable intellectual input by AG, PS, MK, KR, TS, VK, BSa, MMac, and MS. The  
140 manuscript was drafted by DP and revised by BSc. All authors approved the submitted version.

### 141 **Acknowledgments:**

142 We thank all study participants for their commitment and support. We acknowledge financial  
143 support through the BMBF Project COVIDYS (project no. 01EP2105A) and the pandemic  
144 responsiveness fund of The Bavarian Ministry of Science and Art.

### 145 **Conflicts of interest:**

146 All authors declare no conflicts of interest.

147 **References**

- 148 1. Zhang A, Stacey HD, D'Agostino MR, Tugg Y, Marzok A, Miller MS. Beyond  
149 neutralization: Fc-dependent antibody effector functions in SARS-CoV-2 infection. *Nat*  
150 *Rev Immunol.* 2022 Dec 19;1–16.
- 151 2. Vidarsson G, Dekkers G, Rispens T. IgG subclasses and allotypes: from structure to  
152 effector functions. *Front Immunol.* 2014;5:520.
- 153 3. Buhre JS, Pongracz T, Künsting I, Lixenfeld AS, Wang W, Nouta J, et al. mRNA  
154 vaccines against SARS-CoV-2 induce comparably low long-term IgG Fc galactosylation  
155 and sialylation levels but increasing long-term IgG4 responses compared to an  
156 adenovirus-based vaccine. *Front Immunol.* 2023 Jan 12;13:1020844.
- 157 4. Irrgang P, Gerling J, Kocher K, Lapuente D, Steininger P, Habenicht K, et al. Class  
158 switch towards non-inflammatory, spike-specific IgG4 antibodies after repeated SARS-  
159 CoV-2 mRNA vaccination. *Sci Immunol.* 2022 Dec 22;eade2798.
- 160 5. Mahajan VS, Mattoo H, Deshpande V, Pillai SS, Stone JH. IgG4-Related Disease. *Annu*  
161 *Rev Pathol Mech Dis.* 2014 Jan 24;9(1):315–47.
- 162 6. Swank Z, Senussi Y, Manickas-Hill Z, Yu XG, Li JZ, Alter G, et al. Persistent Circulating  
163 Severe Acute Respiratory Syndrome Coronavirus 2 Spike Is Associated With Post-acute  
164 Coronavirus Disease 2019 Sequelae. *Clinical Infectious Diseases.* 2023 Feb  
165 8;76(3):e487–90.
- 166 7. Glück V, Grobecker S, Tydykov L, Salzberger B, Glück T, Weidlich T, et al. SARS-CoV-  
167 2-directed antibodies persist for more than six months in a cohort with mild to moderate  
168 COVID-19. *Infection.* 2021 Aug;49(4):739–46.
- 169 8. Peterhoff D, Glück V, Vogel M, Schuster P, Schütz A, Neubert P, et al. A highly specific  
170 and sensitive serological assay detects SARS-CoV-2 antibody levels in COVID-19  
171 patients that correlate with neutralization. *Infection.* 2021 Feb;49(1):75–82.
- 172 9. Tian X, Li C, Huang A, Xia S, Lu S, Shi Z, et al. Potent binding of 2019 novel coronavirus  
173 spike protein by a SARS coronavirus-specific human monoclonal antibody. *Emerging*  
174 *Microbes & Infections.* 2020 Jan 1;9(1):382–5.
- 175 10. Schneider C, Smith DF, Cummings RD, Boligan KF, Hamilton RG, Bochner BS, et al.  
176 The human IgG anti-carbohydrate repertoire exhibits a universal architecture and  
177 contains specificity for microbial attachment sites. *Sci Transl Med.* 2015 Jan  
178 7;7(269):269ra1.

179