Delayed postglacial colonization of *Betula* in Iceland and the circum North Atlantic

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DEDICATION

This study is dedicated to the memory and scientific legacy of Dr. Sarah E. Crump (1987-2022).

ABSTRACT

As the Arctic continues to warm, woody shrubs are expected to expand northward. This process, known as "shrubification", has important implications for regional biodiversity, food web structure, and high-latitude temperature amplification. While the future rate of shrubification remains poorly constrained, past records of plant immigration to newly deglaciated landscapes in the Arctic may serve as useful analogues. We provide one new postglacial Holocene sedimentary ancient DNA (*sed*aDNA) record of vascular plants from Iceland and place a second Iceland postglacial *sed*aDNA record on an improved geochronology; both show Salicaceae present shortly after deglaciation whereas Betulaceae first appears more than 1000 years later. We find a similar pattern of delayed Betulaceae colonization in eight previously published postglacial sedaDNA records from across the glaciated circum North Atlantic. In nearly all cases, we find that Salicaceae colonizes earlier than Betulaceae and that Betulaceae colonization is increasingly delayed for locations farther from glacial-age woody plant refugia. These trends in Salicaceae and Betulaceae colonization are consistent with the plant families' environmental tolerances, species diversity, reproductive strategies, seed sizes, and soil preferences. As these reconstructions capture the efficiency of postglacial vascular plant migration during a past period of high-latitude warming, a similarly slow response of some woody shrubs to current warming in glaciated regions, and possibly non-glaciated tundra, may delay Arctic shrubification and future changes in the structure of tundra ecosystems and temperature amplification.

INTRODUCTION

As the planet warms, the Arctic is warming at 2 to 4 times the global rate (Rantanen et al., 2022). This warming is impacting Arctic ecosystems through a process known as 'shrubification', or the northward expansion and increased height and density of deciduous woody vegetation (Tape et al., 2006; Myers-Smith et al., 2011; Elmendorf et al., 2012; Sweet et al., 2015). Arctic shrubification alters the structure of tundra ecosystems, its regional biodiversity, food web structure, and nutrient availability (Elmendorf et al., 2012; Fauchald et al., 2017; Collins et al., 2018). Arctic shrubification also has implications for the climate system by reducing surface albedo (Sturm et al., 2005) and increasing atmospheric water vapor (Pearson et al., 2013) – both positive feedbacks that amplify high-latitude warming (Thompson et al., 2022). However, large uncertainties remain regarding the rate of future shrubification, such that the magnitude of these positive feedbacks is poorly constrained in predictive earth system models. One way to reduce future climate uncertainties is through the analysis of well-constrained paleoenvironmental records dating to warmer-than-present periods in Earth's recent history (e.g., Tierney et al., 2020).

Traditionally, macrofossils and pollen have formed the backbone of Quaternary paleovegetation reconstructions (Birks, 2018). However, macrofossils are not consistently preserved in sedimentary records, and pollen production is restricted to seed-bearing taxa, which can be obscured by long-distant transport from sources 1000s of km away (Hyvärinen, 1970; Birks, 2003; Crump et al., 2019). For example, in Iceland, exotic tree pollen from taxa never reported as macrofossils since the Miocene and Pliocene (i.e., *Pinus, Quercus, Corylus, Alnus,* and *Ulmus*, Denk et al., 2011a, b) are found in Early Holocene sedimentary records following retreat of the Icelandic Ice Sheet (Caseldine et al., 2006; Eddudóttir et al., 2015). Hence, the first appearance of pollen need not reflect local colonization of a given taxon. Recent analytical advances in

sedimentary ancient DNA (*sed*aDNA) allow for more reliable vegetation reconstructions (e.g.,

Capo et al., 2021), due in part to their application in continuous lake sediment records that capture

colonization times following local deglaciation. Taxa identified by DNA in lake surface sediment

closely match those in catchment vegetation cover, confirming the local origin of DNA in lake

sediment and improved reliability for taxa presence compared to pollen (Sjögren et al., 2017; Alsos

et al., 2018). Past records of vascular plant *sed*aDNA can therefore provide new insight into

postglacial colonization patterns (Crump et al., 2019; Alsos et al., 2022) as well as the response of

vegetation to changes in temperature during prior warm periods (Clarke et al., 2020; Crump et al.,

2021; Huang et al., 2021).

In this study we present one new Icelandic record (Stóra Viðarvatn) and update the age

model of a second previously published record from Iceland (Torfdalsvatn, Alsos et al., 2021). For

these two lakes, we also compare existing pollen and *sed*aDNA datasets to highlight the ability of

*sed*aDNA in pinpointing the timing of each taxon's first appearance. We then compare *sed*aDNA

records from eight other published North Atlantic lakes that track local vegetation assemblages

since deglaciation, with particular attention to postglacial colonization patterns of two woody plant

families, Salicaceae and Betulaceae. Collectively, the North Atlantic vascular plant DNA data sets

demonstrate that Salicaceae colonized deglaciated lands quickly, whereas Betulaceae was delayed

by up to several thousand years. As these colonization patterns took place during a period of high

latitude warming, our datasets suggest that ongoing warming may also lead to different northward

expansion rates of woody shrubs in glaciated regions, and possibly tundra regions as well. As a

result, the rate of Arctic shrubification may be delayed, with important consequences for high-

latitude ecosystems and temperature amplification.

RESULTS AND INTERPRETATION

Stóra Viðarvatn, Iceland

Stóra Viðarvatn (#4, Table 1, Figure 1) is a large lake (surface area: 2.4 km^2, depth: 48 m) in northeast Iceland, with no pre-existing records of Holocene vegetation change. In winter 2020, we recovered sediment core 20SVID-02 from 17.4 m water depth near the center of the lake. The core contains 893 cm of sediment, collected in 1.5 m increments. The basal 20 cm are laminated, clay-rich sediment, transitioning to organic gyttja by 873 cm depth, which characterizes the remainder of the record, interbedded with tephra (volcanic ash) deposits (Figure S1). The chronology of the Early to Middle Holocene portion of the record is based on 5 tephra layers (see Materials and Methods), of which the lowest, at 886.5 cm depth, is identified as the Askja S tephra (10830 ± 57 BP, Bronk Ramsey et al., 2015), and the highest, at 334 cm depth, as Hekla 4 (4200 ± 42 BP, Dugmore et al., 1995). Because the sedimentation rate between the six tephra layers is essentially linear, we extrapolate that rate from the Askja S tephra layer 7 cm to the base of the sediment core at 893.5 cm depth, which provides a minimum deglaciation age of ~10950 BP (Figure S2A).

Samples for *sed*aDNA analysis were taken from 75 equi-spaced depths throughout core 20SVID-02. Of these, 73 yielded amplifiable vascular plant DNA using the *trnL* P6 loop primer set. The two samples that failed (810 and 856.5 cm depth) were sampled within thick tephra layers with low organic content. Following data filtering (see Materials and Methods), the *trnL* dataset yields 15,738,794 total assigned reads, with an average of 209,851 reads per sample. qPCR cycle threshold (C_T) values, which reflect PCR efficiency and the quantity of suitable target sequences for amplification, average 32.1 across samples. While there is inter-sample variation in DNA reads and C_T values, linear regressions show nearly stable temporal trends (Figure S2), which indicates that the availability of DNA to PCR amplification is consistent throughout the

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record. Metabarcoding technical quality (MTQ) and analytical quality (MAQ) scores are below

suggested thresholds for low quality (0.75 and 0.1, respectively, Rijal et al., 2021) only in some

samples during the Early Holocene (Figure S2). However, given that the scores correlate with

species richness due to the latter values all falling below 30 (Figure S2), the low MTQ and MAQ

scores are likely an artifact of the requirement that the 10 best represented barcode sequences are

required for calculation (Rijal et al., 2021), and not an indication of low-quality DNA. As PCR

amplification results in sequence read abundances that may not reflect original relative abundances

in a sample (Nichols et al., 2018), we focus our discussion on taxa presence/absence, with a greater

number of replicates representing a higher likelihood of taxa presence. We identified 53 taxa

throughout Stóra Viðarvatn's lake sediment record. Of the key woody shrub taxa, Salicaceae is

present at the base of the core (892.5 cm depth, 10850 BP) and is present nearly continuously

through the core. Betulaceae first appears ~2300 years later (701.5 cm depth, 8560 BP), and has a

sporadic presence until the Middle Holocene at ~5500 BP (Figure 2A).

Torfdalsvatn, Iceland

Torfdalsvatn (#3, Table 1, Figure 1) is a small lake (surface area: 0.41 km^2, depth: 5.8 m) in north

Iceland from which sediment cores have been analyzed since the early 1990s to create

paleoecological records from pollen and macrofossils (Björck et al., 1992; Rundgren, 1995, 1998),

and *sed*aDNA (Alsos et al., 2021). The paleovegetation records derived from pollen/macrofossils

and *sed*aDNA were constructed from separate sediment cores that have independent age models

derived from radiocarbon and tephra layers of known age. To temporally compare the two

sediment records, radiocarbon ages need to be calibrated using the same calibration curve,

correlative tephra layer ages must agree, and age model construction requires a common method

(i.e., linear interpolation vs. Bayesian). For the pollen/macrofossil records (Rundgren 1995, 1998),

this required calibrating the radiocarbon ages using IntCal20 (Reimer et al., 2020) and application

of a Bayesian age modeling approach that informs sediment rate changes with prior information

(Blaauw and Christen, 2011) (see Geirsdóttir et al., 2020). For the *sed*aDNA record (Alsos et al.,

2021), we revised the tephra ages previously used to establish a more reliable Bayesian age model

(see Methods and Materials). In doing so, the *sed*aDNA and pollen records feature similar linear

sedimentation rates and are now synchronized by the G10ka Series tephra layers (Figure S2C).

The revised age model for the Early Holocene portion of the pollen/macrofossil record (Figure

S2B, Geirsdóttir et al., 2020) sets the timing of deglaciation, inferred from the onset of organic

sedimentation at 801 cm depth, to 11800 BP.

Differences on the order of centuries between the pollen/macrofossil and *sed*aDNA records

may be artifacts of chronological uncertainty. This is because although the revised age models for

the pollen/macrofossil and *sed*aDNA records feature common tephra layers, variable levels of

proxy data resolution between the two records increases uncertainty when comparing the timing

of inferred changes. Pollen of both Salicaceae (*Salix spp.*) and Betulaceae (*Betula spp.*) first appear

by ~11800 BP (Figure 3). The oldest *B. nana* macrofossil dates to ~9300 BP, similar to

macrofossils of *S. herbacea* (~9400 BP) and *S. phylicifolia* (~9300 BP) (Figure 3). The *trnL* primed

*sed*aDNA data indicate that Salicaceae did not colonize Torfdalsvatn's catchment until ~10300

BP, whereas Betulaceae arrived later at ~9500 BP (Figure 3).

Resolving Icelandic records of woody taxa colonization

Based on the original age model, Torfdalsvatn pollen records were interpreted to capture

ecological changes associated with the abrupt climate oscillations between the Bølling-Allerød

(14700 to 12900 BP) and Younger Dryas (12900 to 11700 BP) periods (Björck et al., 1992;

Rundgren, 1995). However, recent calibration of the radiocarbon ages in the Torfdalsvatn cores

from Björck et al. (1992) and Rundgren (1995) suggest that the organic-rich portion of the record

began less than 11800 years ago (Figure S2B, Geirsdóttir et al., 2020). While there is sediment at

deeper levels, it is described as silty clay (Björck et al., 1992) with sand and gravel (Rundgren,

1995). These characteristics suggest that the sediment likely originates from a rapidly deglaciating

environment, perhaps with lingering ice sheet meltwater in the catchment and a rapid sediment

accumulation rate that is difficult to constrain without secure age control. Therefore, we suggest

that the maximum age for the final retreat of the Icelandic Ice Sheet from Torfdalsvatn's

catchment, when organic-rich sedimentation began, is ~11800 BP, several thousand years younger

than previously proposed (Björck et al., 1992; Rundgren, 1995). We use the updated age models

to reconstruct the history of plant colonization in Iceland.

Differences in taxa presence/absence between the *sed*aDNA and pollen records from

Torfdalsvatn likely reflect different mechanisms of transport for DNA and pollen. *Sed*aDNA for

both Salicaceae and Betulaceae from Torfdalsvatn shows a colonization delay relative to

deglaciation, whereas pollen from both taxa is present, albeit in low abundance, in the oldest

sediment samples (Figure 3). This is most likely explained by the local catchment origin of

*sed*aDNA (Sjögren et al., 2017) versus pollen, which can be transported by wind from 1000s of

km away (Caseldine et al., 2006; Eddudóttir et al., 2015). Therefore, the occurrence of low pollen

counts prior to the corresponding identification with *sed*aDNA (Alsos et al., 2021) likely reflects

long-distance transport from sources outside of Iceland rather than local establishment in the

catchment – an inference supported by a lack of *Salix* and *Betula* macrofossils during this interval

(Figure 3, Rundgren, 1998). While poor DNA preservation could explain the delay in *sed*aDNA,

other vascular plants are identified before Salicaceae and Betulaceae DNA appear, suggesting

adequate DNA preservation for PCR amplification (Alsos et al., 2021). Moreover, the

corresponding sediment is rich in clay (Rundgren, 1995), which preferentially binds with and

stabilizes DNA (Kanbar et al., 2020), making poor preservation unlikely. Therefore, we conclude

that the colonization of Salicaceae and Betulaceae were delayed by 1500 and 2300 years,

respectively, after local deglaciation of Torfdalsvatn's catchment.

Relative to Torfdalsvatn, the catchment of Stóra Viðarvatn deglaciated ~1000 years later

at ~10850 BP (Figure 2A). Based on our new *sed*aDNA record, Salicaceae colonized the catchment

rapidly, as it was identified in the lowermost sample at ~10850 BP, whereas Betulaceae does not

appear in the record until 8560 BP, more than 2000 years later. Considering C_T values are relatively

stable throughout the entire Holocene record (Figure S3), the delayed appearance of Betulaceae

does not likely result from preservation bias. While Stóra Viðarvatn does not have any independent

paleovegetation records, a pollen record from a nearby peat section Ytra-Áland in Thistilfjörður

(Karlsdóttir et al., 2014), ~12 km to the east, provides a useful cross-examination of woody shrub

colonization patterns. In the Ytra-Áland record, *Salix* pollen is found in high counts well above

the mean in the lowermost sample dated to 10400 BP, suggesting bona fide early colonization

(Figure 4). For *Betula*, there is limited to no pollen present until ~8000 BP, after which there is

sporadic presence as we observe with *sed*aDNA (Figure 4). The *Betula* pollen-limited period at

Ytra-Áland is followed by the first peak of *Betula* presence in the total land pollen during ~7700

to 6800 BP. According to size differentiation between the two *Betula* species in Iceland

(Karlsdóttir et al., 2007), the pollen recovered in this peak period is mostly from the dwarf birch

B. nana, but with a fair amount of pollen from the shrub-like *B. pubescens*, suggesting arctic-alpine

heath with stands of downy birch in the region (Karlsdóttir et al., 2014). Despite the coarser

resolution of Ytra-Áland's age model, which only has two tephra layer dates bounding the timing

of first appearance of these taxa (10400 BP and 7050 BP, Karlsdóttir et al., 2014), there is

reasonable agreement between lake and peat records for the timing of arrival and subsequent Early

Holocene establishment of Salicaceae and Betulaceae in northeast Iceland.

Based on the *sed*aDNA records from Torfdalsvatn and Stóra Viðarvatn, we find that as the

Icelandic Ice Sheet receded and exposed new land Salicaceae colonized the landscape. This

occurred 0 to 1700 years after inferred deglaciation, whereas Betulaceae colonization was further

delayed by a few thousand years (~2300 years in both cases). As Torfdalsvatn is the oldest dated

terrestrial site of deglaciation in Iceland, this implies that ~1700 years was required for Salicaceae

to establish a viable population on the island. By the time Stóra Viðarvatn deglaciated, it is likely

that other portions of north Iceland had also deglaciated allowing Salicaceae to more efficiently

colonize new regions of Iceland. The similar timing of Betulaceae colonization relative to

deglaciation in the two catchments (~2300 years) but different times of deglaciation (11800 and

10850 BP) suggests that the evolution of catchment conditions for Torfdalsvatn and Stóra

Viðarvatn may be additional factors controlling the local colonization of this taxon on Iceland. The

earlier presence of low concentrations of Salicaceae and Betulaceae pollen in Torfdalsvatn are

most likely derived from long distance wind dispersal from outside Iceland.

DISCUSSION

Postglacial *sed*aDNA records from the circum North Atlantic

Eight additional records of vascular plant *sed*aDNA from lake sediments located around the North

Atlantic that were glaciated during the Last Glacial Maximum are available to compare with the

Icelandic data (see Material and Methods for details). These include records from Baffin Island

(n=1, Crump et al., 2019), northern Greenland (n=1, Epp et al., 2015), Svalbard (n=2, Alsos et al.,

2016; Volstad et al., 2020), northern Norway (n=3, Rijal et al., 2021; Alsos et al., 2022), and

southern Norway (n=1, ter Schure et al., 2021) (Figure 1). All eight records have age control

derived from calibrated radiocarbon ages. Except for Baffin Island, we infer minimum ages of

deglaciation based on the age of the oldest sediment.

Baffin Island: Lake Qaupat (#1, Figure 1) is a small lake (surface area: 0.08 km^2, depth:

9.2 m) situated at 35 m asl on southern Baffin Island. Cosmogenic ^{10}Be exposure dating of Lake

Qaupat's impounding moraine constrains the timing of deglaciation to 9100 ± 700 BP (Crump et

al., 2019). Lake Qaupat and most of its catchment were below sea level until 7700 ± 300 BP, when

postglacial isostatic recovery raised the basin above the ocean and its catchment was available for

vascular plant colonization. Salicaceae is present in the record at shortly after isolation from the

ocean at 7400 BP, but no vascular plants are documented by *sed*aDNA in the older marine

sediment. The first appearance of Betulaceae *sed*aDNA occurs at 5900 BP, more than 1000 years

after Salicaceae and 2000 years after the lake and its catchment were above sea level (Crump et

al., 2019).

Greenland: Bliss Lake (#2, Figure 1) is a small lake (depth: 9.8 m) situated at 17 m asl on

the northern coastline of Greenland (Peary Land). Bliss Lake initially deglaciated 11000 BP (Olsen

et al., 2012) and the first appearance of Salicaceae *sed*aDNA is found in low read counts in

sediment dated to 10800 BP but only in high read counts beginning at 7400 BP (Epp et al., 2015).

As the lake was inundated by marine water between 10480 and 7220 BP, (Olsen et al., 2012), 7400

BP is interpreted to be the most reliable date for Salicaceae colonization in the catchment as sea

level regressed and isolated the lake from the ocean (Epp et al., 2015).

Svalbard: Jodavannet (#5, Figure 1) is a small lake (depth: 6.4 m) situated at 140 m asl on

the east coast of Wijdefjorden on northern Spitsbergen. Regional cosmogenic exposure dating

suggests the area began to deglaciate between 14600 and 13800 BP (Hormes et al., 2013), although

the base of Jodavannet's sediment record suggests a minimum timing of lake deglaciation by

11900 BP (Volstad et al., 2020). Salicaceae *sed*aDNA is first found at 9800 BP in Jodavannet,

whereas Betulaceae was not detected with *sed*aDNA metabarcoding (Volstad et al., 2020). Lake

Skartjørna (#6, Figure 1) is a small lake (surface area: 0.10 km^2, depth: 7.5 m) situated at 65 m asl

on the west coast of Spitsbergen. As sea level fell due to glacial isostatic adjustment, Lake

Skartjørna likely became isolated from the sea at ~13000 BP (Landvik et al., 1987), although the

base of the recovered sediment record, which provides a minimum deglaciation age, is dated to

8600 BP (Alsos et al., 2016). Salicaceae *sed*aDNA is identified in Lake Skartjørna's basal sample

at 8600 BP and Betulaceae is first identified at 7000 BP (Alsos et al., 2016).

Norway: Langfjordvannet (#7, Figure 1) is a small lake (surface area: 0.55 km^2, depth: 34.8

m) situated at 66 m asl on the coast of northern Norway (Rijal et al., 2021). Langfjordvannet

deglaciated by 16150 BP, Salicaceae is first identified in *sed*aDNA by 15500 BP, and Betulaceae

by 10000 BP (Alsos et al., 2022). Eaštorjávri South (#8, Figure 1) is a small lake (surface area:

0.06 km^2, depth: 5.4 m) situated at 260 m asl near the coast of northern Norway (Rijal et al., 2021).

Eaštorjávri South deglaciated by 11060 BP, with Salicaceae *sed*aDNA identified in the basal

sample at 11060 BP and consistently present above that level. Betulaceae is identified, albeit with

low read counts, in the basal sample as well, but not identified in the overlying sample. Since

Betulaceae *sed*aDNA is not present with substantial read counts until 9950 BP (Alsos et al., 2022),

we conservatively set this as Betulaceae's first appearance in Eaštorjávri South. Nordvivatnet (#9,

Figure 1) is a small lake (surface area: 0.05 km^2, depth: 13.3 m) situated at 82 m asl on the coastline

of northern Norway (Rijal et al., 2021). Nordvivatnet deglaciated by 12880 BP, with Salicaceae

*sed*aDNA identified in the basal sample. Betulaceae *sed*aDNA is identified in the basal sample as

well but below our cut-off for taxa presence (see Materials and Methods). The first sample with

high read counts indicative of Betulaceae presence is at 12080 BP (Alsos et al., 2022), which we

conservatively set as the timing of the taxa's first appearance in Nordvivatnet. Finally, Lake

Ljøgottjern (#10, Figure 1) is a small lake (surface area: 0.02 km^2, depth: 18 m) situated at 185 m

asl in southeastern Norway. Lake Ljøgottjern deglaciated by 9300 BP and both Salicaceae and

Betulaceae *sed*aDNA were first detected by 7900 BP (ter Schure et al., 2021).

Betulaceae colonization is delayed relative to Salicaceae in the circum North Atlantic

The timing of postglacial Salicaceae and Betulaceae colonization varies across the North Atlantic

regions, with first-appearance dates ranging from 15500 BP in Norway to 5900 BP on Baffin Island

(Figure 5). Salicaceae appears immediately after inferred deglaciation at 4 out of 10 sites, while

colonization dates for the other 6 locations range from 200 to 2100 years after deglaciation and

exhibit no clear spatio-temporal pattern (Figure 6). For Betulaceae, colonization times range from

800 to 6150 years after deglaciation, with sites closest to source refugia south of the ice sheets

(Figure 1) having generally shorter colonization delays (Figure 6), consistent with seedling

viability having a higher probability of success with shorter distance from source (Nathan, 2006).

The one exception is Langfjordvannet, in northern Norway (site #7, Figure 1, Alsos et al., 2022),

which deglaciated at 16150 BP (Figure 5). Considering that Betulaceae only arrives during the

Holocene Epoch (last 11700 years) across all North Atlantic sites, the anomalously long delay of

Betulaceae to Langfjordvannet suggests that this taxon may have required the Holocene's

relatively stable climate, as reflected by Greenland oxygen isotope records (Figure 5, Seierstad et

al., 2014), to successfully establish itself. Alternatively, the colonization delay of Betulaceae to

Langfjordvannet may reflect the slow development of stable soils, which are required for

Betulaceae establishment, due to the lake's steep catchment (Otterå, 2012). In contrast to

Betulaceae, Salicaceae appears to have been an efficient colonizer to Langfjordvannet during the

colder and less stable climate of the Late Glacial period and earliest Holocene (Figure 5, Seierstad

et al., 2014). Finally, except for Lake Ljøgottjern in southern Norway (site #10, Figure 1) where

Betulaceae and Salicaceae apparently colonize at about the same time (ter Schure et al., 2021), all

other sites show that Betulaceae colonization is delayed by 800 to 5500 years relative to Salicaceae

colonization dates (Figure 5).

Several possibilities may contribute to the delay in Betulaceae colonization compared to

Salicaceae colonization. First, the northernmost Salicaceae species reach bioclimate subzone A (1-

3 °C July temperature) whereas the northernmost Betulaceae species rarely go beyond bioclimate

subzone D (8-9 °C July temperature) (Walker et al., 2005). The different environmental tolerances

may explain why Salicaceae generally arrives earlier when regional temperatures are still climbing

compared to the timing of Betulaceae colonization when warmth is more broadly established

(Figure 5). In support, Alsos et al. (2022) showed that at least for Fennoscandian *sed*aDNA

vascular plant records, temperature was one of the dominant controls on postglacial dispersal

patterns. Second, species diversity is greater for the Salicaceae family compared to Betulaceae

family in the North Atlantic, and conventional DNA metabarcoding techniques cannot identify

*sed*aDNA to the species level. Iceland, for example, has four native Salicaceae taxa, i.e., three

dwarf, cold-tolerant arctic species (*S. herbacea, S. lanata,* and *S. arctica*) and one shrub-like

species (*S. phylicifolia*), and only two Betulaceae (*B. nana* and *B. pubescens*) (Kristinsson, 2008).

The suite of Salicaceae taxa can collectively tolerate a wider range of environments than

Betulaceae, including disturbed, wet, and nutrient limited substrate characteristic of deglacial environments (e.g., *S. herbacea,* Beerling, 1998). Third, the modes of reproduction for Salicaceae and Betulaceae are different. Salicaceae is clonal, meaning that plants mostly reproduce asexually by vegetative spreading, which is advantageous when colonizing new landscapes (e.g., Beerling, 1998). Betulaceae propagates instead by seed and therefore requires a larger number of founder individuals for successful establishment (Atkinsson, 1992; de Groot et al., 1997). Moreover, Betulaceae seeds are generally heavier than Salicaceae seeds (dry mass, *B. nana* = 0.12 to 0.30 mg, de Groot et al., 1997; *S. herbacea* = 0.07 to 0.14 mg, Beerling, 1998). This mass difference would provide a higher probability of long-distance wind dispersal for Salicaceae compared to Betulaceae and establishment of the pioneer plants.

Finally, modern observations broadly replicate the records of delayed woody taxa colonization in the North Atlantic. Studies tracking the migration of Salicaceae and Betulaceae in the forefields of retreating glaciers and to new islands also show a rapid and faster colonization of Salicaceae compared to Betulaceae (Whittaker, 1993; Magnússon et al., 2009; Burga et al., 2010; Synan et al., 2021). As an example, following the formation of the volcanic island Surtsey in 1964 CE, Salicaceae (*S. herbaceae, S. lanata* and *S. phylicifolia*) colonized between 1995 and 1999 CE and Betulaceae has yet to arrive (Magnússon et al., 2009). Complimentary studies tracking soil and nutrient development at proglacial sites in the North Atlantic (Vilmundardóttir et al., 2014) suggest that Salicaceae is environmentally unconstrained (Whittaker, 1993; Glausen and Tanner, 2019), whereas Betulaceae requires more developed soil for its ultimate viability (Synan et al., 2021). Collectively, these studies support the environmental constraints known for each woody taxon and reinforce the hypothesis that Salicaceae is a more efficient pioneer than Betulaceae, particularly in newly deglaciated landscapes.

Environmental dispersal mechanisms

Coupling the timing of postglacial woody shrub colonization in the North Atlantic with genetic approaches provides a valuable opportunity to identify probable environmental dispersal mechanisms (e.g., wind, sea ice, driftwood, and birds) for long-distance transport. Previously, Alsos et al. (2015) compared amplified fragment length polymorphism (AFLP) data from living plants from Iceland, Greenland, Svalbard, the Faroe Islands, and Jan Mayen to AFLP data from the same species from potential source regions. Genetic distances suggest that *Betula* (*sp. nana* and *pubescens*) and *Salix herbacea* populations in most locations descend from ancestors in a western European Last Glacial Maximum (LGM) refugia (Alsos et al., 2009; Eidesen et al., 2015), and that each taxa underwent a broad-fronted and rapid migration to newly deglaciated regions (Alsos et al., 2015). *B. nana* on Svalbard likely descends from a northwest Russian source (Alsos et al., 2007, 2015), and populations on Baffin Island and western Greenland originated from North America (Alsos et al., 2009; Eidesen et al., 2015). Molecular analysis of Icelandic *Betula* based on chloroplast haplotypes indicates that *Betula* may have post-glacially immigrated to Iceland from northern Scandinavia (essentially *B. nana*) and northwestern Europe (*B. pubescens*) (Thórsson et al., 2010). However, the relative roles of wind, sea ice, driftwood, and birds as transport vectors remain an open question.

Sea ice has been suggested to be a dispersal mechanism for postglacial vascular plants in the Arctic (Alsos et al., 2016). While seasonal sea ice persisted throughout the Holocene around Svalbard (Pieńkowski et al., 2021) and northern Greenland (Syring et al., 2020), general ocean circulation patterns require source populations of vascular plants in Arctic Siberia, the dominant source region for sea ice export to the North Atlantic, for sea ice to be a viable vector. Considering

that *B. nana* on Svalbard likely descends from a Russian refugia (Alsos et al., 2007, 2015), sea ice

is a reasonable transport mechanism for this location. However, genetic data from Salicaceae (*S.*

herbacea) on Baffin Island, Greenland, and Svalbard suggest that the modern populations from

these locations descend from source populations in either mid-latitude North America (south of

the Laurentide Ice Sheet) or western Europe (Alsos et al., 2007, 2009), and neither region exported

sea ice in the Early Holocene. This leaves wind and birds as likely mechanisms for *S. herbacea*

transport to Baffin Island, Greenland, and Svalbard during the Holocene. North of Iceland, proxy

evidence shows that sea ice disappeared by ~11200 BP (Figure 2B, Xiao et al., 2017) when surface

ocean currents were established bringing warm Atlantic waters to the North Iceland Shelf (Figure

2C, Eiríksson et al., 2000; Sha et al., 2022), nearly 500 years before the colonization of the first

woody taxon on the island (Figure 2A). Thus, with the absence of sea ice around Iceland's coastline

during woody plant colonization, wind and/or birds remain the likeliest dispersal mechanisms for

Iceland, similar to other Arctic regions.

Future outlook

As the planet continues to warm, glacier and ice sheet mass loss is accelerating (Hugonnet et al.,

2021), providing new territory for woody shrubs to colonize – the Greenland Ice Sheet currently

occupies 1.7 million km^2 (Morlighem et al., 2017) with >400,000 km^2 more covered by glaciers

elsewhere in the Arctic (RGI Consortium, 2017). Global warming will also allow woody plants to

migrate poleward across unglaciated Arctic tundra (e.g., Myers-Smith et al., 2011), which occupies

a substantially larger area compared to glaciated regions (5 billion km^2, Walker et al., 2005). In

this study, we demonstrate that Salicaceae can migrate into deglaciating North Atlantic habitats

rapidly whereas Betulaceae may be delayed by millennia. However, the ability of these taxa to

successfully follow the rapid pace of ongoing range shifts in non-glaciated tundra (e.g., Alaska

and Siberia) remains an open question (e.g., Davis and Shaw, 2001; Thuiller et al., 2009). This is

partially due to different baseline conditions of the tundra ecosystem compared to glacial regions

upon deglacial warming, which already contain established ecosystems and nutrient-rich soils

bound in the permafrost (e.g., Heijmans et al., 2022).

While the causes and rates of present-day climate change differ from the preceding

postglacial period (e.g., Tierney et al., 2020), the factors that we identify as potentially driving the

colonization patterns (i.e., environmental tolerance, modes of reproduction, and soil development)

are relevant in both scenarios. As a result, circum North Atlantic shrubification in deglaciating

regions may be prolonged – ultimately dampening the rate of change, at least temporarily, in

regional biodiversity and food web structure, as well as high-latitude temperature amplification.

An important next step for constraining future shrubification rates is developing detailed

paleoecological reconstructions of plant dynamics in multiple tundra regions (e.g., Clarke et al.,

2020; Huang et al., 2021), continuing these efforts in glaciated regions, and pairing plant histories

with independent local records of climate, such as temperature and precipitation. The latter

paleoclimate reconstructions will enable more detailed assessment of plant colonization lags

relative to climate (e.g., Crump et al., 2019). By pairing paleoecological and paleoenvironmental

records from formerly glaciated and tundra regions and integrating these with predictive models

(e.g., Braconnot et al., 2012), the community will be able to reduce uncertainties and better

quantify future shrubification rates, which are essential to inform ecological predictions for

policymakers.

MATERIALS AND METHODS

Lake sediment cores and age control

Stóra Viðarvatn (66.24°N, 15.84°W) is a large (2.4 km^2), deep lake (48 m) located at 151 m asl in northeast Iceland (Figure 1, Axford et al., 2007). In winter 2020, we recovered a composite 8.93 m long sediment core (20SVID-02, Fig. S1) from 17.4 m water depth using lake ice as a coring platform. The sediment was collected in 7 drives of ~150 cm each. The core sections were subsequently stored at 4 °C before opening for sediment subsampling. The chronology of 20SVID-02 is based on 5 marker tephra layers (volcanic ash) of known age: Askja S, G10ka Series, Kverkfjöll/Hekla, Kverkfjöll, and Hekla 4 tephra layers, with ages of 10830 ± 57 BP (Bronk Ramsey et al., 2015), 10400 to 9900 BP (Óladóttir et al., 2020), 6200 BP (Óladóttir et al., 2011), 5200 BP (Óladóttir et al., 2011) and 4200 ± 42 BP (Dugmore et al., 1995), respectively (see Supplemental Information). A Bayesian age model was generated using the R package rbacon and default settings (Blaauw and Christen, 2011) (Figure S2A).

Torfdalsvatn (66.06°N, 20.38°W) is a small (0.4 km^2), shallow (5.8 m) lake located at 52 m asl in north Iceland (Figure 1, Rundgren et al., 1997). Paleovegetation records from this lake are based on pollen, macrofossil assemblages (Björck et al., 1992; Rundgren, 1995, 1998; Rundgren and Ingólfsson, 1999), and *sed*aDNA (Alsos et al., 2021). Sediment cores from Torfdalsvatn were also recovered by a joint Colorado-Iceland team in 2004, 2008, 2012, and 2020. In modifying the lake sediment core chronologies, we use the recently calibrated age model (Figure S2B, Geirsdóttir et al., 2020) for the pollen/macrofossil records (Björck et al., 1992; Rundgren, 1995) that uses the IntCal20 calibration curve (Reimer et al., 2020). We note that the ~12000 BP Vedde Ash has been previously identified in this record based on major oxide composition (Björck et al., 1992). However, the age of the Vedde Ash is stratigraphically too old for the radiocarbon-based chronology, suggesting that this tephra layer likely represents ash from a separate and subsequent

volcanic eruption that produced the Vedde Ash, of which several possible correlations have been

identified in Iceland and mainland Europe (e.g., Pilcher et al., 2005; Kristjánsdóttir et al., 2017;

Matthews et al., 2011; Geirsdóttir et al., 2022). For the *sed*aDNA record, the Early Holocene

portion of lake sediment core (Alsos et al., 2021) features two age control points, the so-called

„Saksunarvatn Ash" (10267 ± 89 BP) and a graminoid radiocarbon age (6620 ± 120 BP, Alsos et

al., 2021). To better compare with the pollen/macrofossil record, we generated a new age model

for the *sed*aDNA record (Alsos et al., 2021), using the lowermost radiocarbon age and the upper

and lower limits of the main ~20 cm thick G10ka Series/Saksunarvatn tephra unit (10400 to 9900

BP) following the recent recognition of multiple associated tephra layers and dating of these layers

in Iceland (Harning et al., 2018, 2019; Óladóttir et al., 2020). The new Bayesian age model was

generated using the R package rbacon and default settings (Blaauw and Christen, 2011) with the

IntCal20 calibration curve (Reimer et al., 2020) (Figure S2C).

DNA metabarcoding

We subsampled the Stóra Viðarvatn sediment core halves immediately upon splitting in a

dedicated clean lab facility with no PCR products in the Trace Metal Lab, University of Colorado

Boulder. All surfaces and tools were treated with 10% bleach and 70% ethanol before use.

Personnel wearing complete PPE took a total of 75 samples using clean metal spatulas. Samples

were then stored in sterile Falcon tubes at 4ºC until DNA extraction and metabarcoding. Sampling

resolution was based upon a preliminary age model with a sample estimated every 150 years.

We performed DNA extraction and metabarcoding library generation in a dedicated ancient

DNA laboratory at the Paleogenomics Lab, University of California Santa Cruz. While working

in this laboratory we followed commonly used practices for minimizing contamination of ancient

and degraded samples described previously (Fulton and Shapiro, 2019). We initially cleaned all

instruments and surfaces using 10% bleach and 70% ethanol before UV irradiation for at least 1

hour. From each homogenized sample of the core, we aliquoted two 50 mg subsamples and isolated

DNA following the column purification method described in Rohland et al. (2018) using Binding

Buffer D and eluting 50 µl volumes. We pooled extraction eluates derived from the same original

core sample prior to subsequent experiments and included one extraction control (containing no

sample) for every 12 samples processed to assess the presence of external contaminants or cross

contamination.

We performed quantitative PCR (qPCR) to determine the ideal dilution of extract to add to

the metabarcoding PCR, as well as the ideal number of PCR cycles. We added 2 µl of neat or

diluted extract to wells containing 1X Qiagen Multiplex master mix, as well as a final

concentration of 0.16 µm of the forward and reverse trnL primer and 0.12X SYBR Green I Dye.

Cycling conditions were as follows: a 15 min 95C activation, followed by 40 cycles of 94C for 30

sec, 57C for 30 sec, and 72C for 60 sec. We tested 1:10 and 1:100 dilutions of extract to water

alongside neat extract. We used cycle threshold (C_T) values to compare PCR efficiency between

dilutions and determined the ideal cycle number for the metabarcoding PCR as the number of

cycles at which the PCR curve reached the plateau phase.

We performed metabarcoding PCR on each extract using the best dilution as determined

by qPCR and versions of the chloroplast *trnL* primer set (Taberlet et al., 2007) that contained

attached truseq adapters (Table S2). We used five replicate PCRs for each primer set and for each

sample, with reaction conditions as follows: 2 µl of diluted or undiluted extract in 1X Qiagen

Multiplex Mix and a 0.16 µM concentration of each primer at a total volume of 25 µl.

Thermocycling began with 15 min at 95C, followed by a number of cycles (as determined by

qPCR) at 94C for 30 sec, 50-57C for 90 sec, 72C for 60 sec. We used one negative control PCR

(no sample) alongside every 16 samples to assess the possibility of contamination during the PCR

set up. Extraction controls also underwent metabarcoding. We cleaned reactions using SPRI select

beads and eluted them into a final volume of 20 µl.

We then used indexing PCR to attach the full truseq adapter and unique identifier. We

added 5 µl of cleaned PCR product from the metabarcoding PCR to 1X Kapa Hifi master mix and

dual-unique truseq style indices at a concentration of 0.4 µM. Thermocycling began with an

activation step at 98C for 3 min, followed by 8 cycles of 98C for 20 sec, 65C for 30 sec, 72C for

40 sec, and a final extension at 72C for 2 min. We cleaned reactions using SPRI select beads and

quantified DNA concentration using the Nanodrop 8000 Spectrophotometer. We pooled PCR

replicates from all samples in equimass ratios and sequenced the pools on an Illumina Nextseq 550

platform. Sequencing was performed using Illuminas v3 chemistry and a 150 cycle mid output

sequencing kit with paired end reads of 76 base pair length.

We processed raw fastq files and performed taxonomic assignment using the Anacapa

Toolkit (Curd et al., 2019) and the ArcBorBryo library, which contains 2280 sequences of Arctic

and boreal vascular plants and bryophytes (Sønstebø et al., 2010). Briefly, we removed adapter

sequences from the raw fastq data using cutadapt and trimmed low quality bases using FastX-

toolkit before reads were separated into paired and unpaired files (Hannon, 2010; Martin, 2011).

We then used dada2 for further filtering of chimeric sequences and for merging of paired reads

(Callahan et al., 2016.). We dereplicated identical sequences present in the data using dada2 and

generated ASV (amplicon sequence variant) tables and fastq files for each of the read types (paired

merged, paired unmerged, unpaired forward, and unpaired reverse). ASV's were assigned

taxonomy by globally aligning to a CRUX database generated from the ArcBorBryo library using

Bowtie2 before assignment with Anacapa's custom BLCA algorithm (Langmead and Salzberg,

2012). Outputted taxonomy site frequency tables containing taxonomic assignments and read

counts were subsequently analyzed using R. ASV sequences for Salicaceae and Betulaceae

identified in our samples are included in the supplement (Table S3).

To minimize the chance of false positives, we retained only sequences that: 1) were not

detected in negative controls, 2) had 100% match to sequences in the library, 3) had a minimum

of 10 reads per PCR replicate and occurred in a minimum of two of five PCR replicates, and 4)

had a minimum of 100 reads across all PCR replicates (Table S4). We verified non-native taxa

identified by this pipeline via comparison with the NCBI nucleotide database using BLAST

(http://www.ncbi.nlm.nih.gov/ blast/) and removed these due to the low likelihood of correct

taxonomic assignment. Finally, DNA quality scores were calculated following Rijal et al. (2021).

Published *sed*aDNA datasets

To place the Icelandic plant *sed*aDNA datasets in the context of the circum North Atlantic, we

compiled all existing lake *sed*aDNA records of vascular plants from this region that were formally

glaciated during the Last Glacial Maximum (Figure 1, Table 1). These include records from Baffin

Island (*n*=1, Crump et al., 2019), northern Greenland (*n*=1, Epp et al., 2015), Svalbard (*n*=2, Alsos

et al., 2016; Volstad et al., 2020), northern Norway (*n*=3, Rijal et al., 2021; Alsos et al., 2022), and

southern Norway (*n*=1, ter Schure et al., 2021). To ensure that all records were comparable, we

excluded 7 records from northern Norway: Sandfjorddalen, Horntjernet, Gauptjern, Jøkelvatnet,

Kuutsjärvi, Nesservatnet, Sierravannet (Rijal et al., 2021; Alsos et al., 2022). Nesservatnet and

Sierravannet contained only Late Holocene records and thus do not capture local postglacial

colonization patterns. A lack of supporting proxy records (e.g., carbon content) for Sandfjorddalen

and Horntjernet meant it was unclear whether these two records capture the complete postglacial

period. For Gauptjern, the inferred basal age (8776 BP) is inconsistent with the reconstructed

timing of deglaciation (~11000 BP, Hughes et al., 2016), which suggests the initial postglacial

interval may be missing. Finally, both Jøkelvatnet and Kuutsjärvi were impacted by glacial

meltwater during the Early Holocene when woody taxa are first identified (Wittmeier et al., 2015;

Bogren, 2019), and thus the inferred timing of plant colonization is probably confounded in this

unstable landscape by periodic pulses of terrestrial detritus. Unless otherwise specified, the timing

of deglaciation at each site is inferred from the basal age of the sediment record.

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AUTHOR CONTRIBUTIONS

GHM, ÁG and JS conceptualized funded the project; GHM, BS and ÁG supervised the project;

GHM, ÁG, DJH and JHR cored the lake; DJH and NA sampled the sediment; TT performed tephra

analyses and identification; SS performed DNA extraction and library generation; DJH performed

taxononmic assignment and processed the data, constructed age models, and compiled published

datasets; DJH wrote the paper with contributions and expertise from SS, KAJ, NA, TT, JS, ÁG,

BS and GHM.

COMPETING INTERESTS

The authors declare that they have no competing interests.

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995 **TABLES AND FIGURES**

997 Table 1: *Lake site information.*

Site #	Lake name	Region	Latitude (º)	Longitude (º)	Reference
1	Lake Qaupat	Baffin Island	63.68	-68.20	Crump et al. (2019)
2	Bliss Lake	Greenland	83.52	-28.35	Epp et al. (2015)
3	Torfdalsvatn	Iceland	66.06	-20.38	Alsos et al. (2021)
4	Stóra Viðarvatn	Iceland	66.24	-15.84	This study
5	Jodavannet	Svalbard	77.34	16.02	Volstad et al. (2020)
6	Lake Skartjørna	Svalbard	77.96	13.82	Alsos et al. (2016)
7	Langfjordvannet	Norway	70.15	20.54	Alsos et al. (2022)
8	Eaštorjávri South	Norway	70.43	27.33	Alsos et al. (2022)
9	Nordvivatnet	Norway	70.13	29.01	Alsos et al. (2022)
10	Lake Ljøgottjern	Norway	60.15	11.14	ter Schure et al. (2021)

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1002

1003 **Figure 1**: Overview map of the North Atlantic. Lake *sed*aDNA sites with vascular plant histories

1004 since local deglaciation shown in yellow (see Table 1 for site information). The extent of the Last

1005 Glacial Maximum (LGM) ice sheets is delineated in blue (Batchelor et al., 2019).

1006

Figure 2: Comparison of paleoclimate and lake *sed*aDNA records from Iceland. A) Presence/absence *sed*aDNA record from Stóra Viðarvatn (green) and Torfdalsvatn (red) for Betulaceae and Salicaceae, where light gray markers denote that no taxa was detected. For Stóra Viðarvatn *sed*aDNA, the bubble size is proportional to the number of PCR replicates, and vertical dark gray lines denote the timing of each lake's deglaciation. B) Biomarker-proxy record of sea ice from the North Iceland Shelf (inverted), where greater concentrations of IP_{25} correspond to more sea ice (Xiao et al., 2017). C) Diatom-based SST record from the North Iceland Shelf (Sha et al., 2022).

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Figure 3: Simplified paleovegetation records from Torfdalsvatn for two taxa: *Salix spp.* and *Betula spp.* Shown are pollen counts (bold red lines), where shaded regions indicate values above the mean (Rundgren, 1995), first occurrence of taxa macrofossils (black leaves, Rundgren, 1998), and DNA presence (red bubbles, Alsos et al., 2021).

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Figure 4: Simplified paleovegetation records from northeast Iceland for two taxa: *Salix spp.,* and *Betula spp.* Shown are Ytra-Áland pollen counts (bold green lines), where shaded regions indicate values above the mean (Karlsdóttir et al., 2014) and DNA presence from Stóra Viðarvatn (green bubbles, this study), where the bubble size is proportional to the number of PCR replicates.

1026

Figure 5: Timing of postglacial Salicaceae and Betulaceae colonization in the circum North Atlantic at 10 locations (see Figure 1 and Table 1). A) GISP2 δ^{18}O reflective of regional North Atlantic temperature variability (Seierstad et al., 2014), and B) lake *sed*aDNA records of vascular plants, where the brown boxes reflect the extent of sedimentary record and the red/blue circles denote the first appearance of Salicaceae/Betulaceae (Epp et al., 2015; Crump et al., 2019; Alsos et al., 2016, 2021, 2022; Volstad et al., 2020; Rijal et al., 2021; ter Schure et al., 2021; this study). Light red and blue circles denote the presence of a low number of taxa reads, which we conservatively do not interpret to reflect genuine presence (e.g., Epp et al., 2015). Grey circles denote the minimum timing of local deglaciation inferred from basal core ages, and in the case of Baffin Island, cosmogenic radionuclide dating of the lake's impounding moraine (Crump et al., 2019).

1039

1040 **Figure 6**: Colonization time after deglaciation (years) versus distance the lake is from the closest

1041 possible source south of the LGM ice sheet margin (km). For all circum North Atlantic sites,

1042 Salicaceae is shown in red and Betulaceae in blue. Data point for Langfjordvannet is open for

1043 Betulaceae.