Integrating Biotic Interactions In Niche Analyses Unravels Patterns Of Community Composition in Clownfishes

Alberto García Jiménez[1], Antoine Guisan[2,3], Olivier Broennimann[2,3], Théo Gaboriau[1*] and Nicolas Salamin[1*]

[1*]Department of Computational Biology, University of Lausanne, Lausanne, Switzerland.
[2]Department of Ecology and Evolution,University of Lausanne, Lausanne, Switzerland.
[3]Institute of Earth Science Dynamics, University of Lausanne, Lausanne, Switzerland.
[*] indicates co-last authorship.

Contributing authors: agarcia26286@gmail.com;

Abstract

Biotic interactions shape the ecology of species and communities, yet their integration into ecological niche modeling methods remains challenging. Despite being a central topic of research for the past decade, the impact of biotic interactions on species distributions and community composition is often overlooked. Mutualistic systems offer ideal case studies for examining the effects of biotic interactions on species niches and community dynamics. This study presents a novel approach to incorporating mutualistic interactions into niche modeling, using the clownfish-sea anemone system. By adapting existing niche quantification frameworks, we developed a method to estimate the partial effects of known interactions and refine ecological niche estimates. This approach allows for a more comprehensive understanding of how mutualistic relationships influence species distributions and community assembly patterns. We also used mutualistic information to investigate the resource-use overlap, identifying patterns of competition within clownfish communities. Our results reveal significant deviations in niche estimates when biotic interactions are considered, particularly for specialist species. Host partitioning among clownfish species reduces resource-use overlap, facilitating coexistence in species-rich habitats and highlighting mutualism's role in promoting and maintaining diversity. We uncover complex dynamics in resource-use overlap among clownfish species, influenced by factors such as species richness, ecological niche overlap, and host specialization. Specialist-generalist interactions strike an optimal balance, supporting high species richness while minimizing competition. These insights enhance our understanding of clownfish biodiversity patterns, demonstrating how diverse mutualistic strategies contribute to diversity build-up and mitigate competitive exclusion in saturated communities. The analytical framework presented has broad applications beyond the clownfish-sea anemone system, potentially extending to a broader range of interactions. It enables a more comprehensive understanding of biodiversity maintenance in complex ecosystems and constitutes a valuable tool for conservation planning and ecosystem management.

Keywords: biotic interactions, clownfish, community composition, competition, mutualism, niche, species distribution, spatial ecology

Introduction

Diverse types of biotic interactions govern the interconnection between species in nature. Among them, mutualism, a relationship in which different species benefit from each other, has attracted the attention of biologists and the public at large. Mutualism plays a substantial role in evolutionary processes. It has impacted major evolutionary transitions like the origin of the eukaryotic cell or the colonization of land by symbiotic plants, and contributes to the increase of biodiversity (Bastolla *et al.* 2009) by enhancing the survival and success of interacting species (Benton 2009). It also has an important ecological impact by favouring ecosystem stability, and facilitating species dispersal and resilience (Hale *et al.* 2020; Le Roux *et al.* 2020), impacting species distributions (Pellissier *et al.* 2013; Schleuning *et al.* 2015; Marjakangas *et al.* 2020).

Mutualistic interactions present a gradient of intensity from full generalists to exclusive specialists, leading to diverse ecological and evolutionary dynamics (Bascompte & Jordano 2007; Sverdrup-Thygeson *et al.* 2017; Gracia-Lázaro *et al.* 2018). Generalist mutualism promotes ecosystem resilience through flexible responses to environmental changes (Maia *et al.* 2021). In contrast, specialist mutualism can drive co-evolution and lead to highly adapted species pairs (Cook & Rasplus 2003). Generalist-specialist transitions can foster diversification through ecological speciation (Lunau 2004), where reproductive isolation evolves as a by-product of adaptation to different ecological niches (Chomicki *et al.* 2019; Frachon *et al.* 2023). As species adapt to specific mutualistic partners, they may undergo niche partitioning, reducing niche overlap to avoid competitive exclusion (Salas-Lopez *et al.* 2022), allowing closely related species to coexist in the same ecosystem (Schoener 1974). Plant-pollinator systems exemplify this process, with related plant species evolving to attract different pollinators or flower at different times, reducing pollination competition (Van der

Niet *et al.* 2014). In the marine environment, the clownfish (Amphiprioninae) mutualism with host sea anemones might have triggered their rapid diversification through ecological speciation, a process known as adaptive radiation (Litsios *et al.* 2012).

The evolutionary success of this group of 30 reef fish species, is attributed to their unique mutualistic associations with sea anemones in the Indo-Pacific Ocean. The mutualism significantly enhances clownfish survival and reproductive success (Lubbock 1980; Fautin 1991), with each species developing specific associations with up to ten sea anemone species, resulting in both generalist and specialist behaviors. Despite their similar ecological characteristics (habitat, diet and social structure), clownfishes exhibit higher levels of coexistence in species-rich locations (Camp 2016; Elliott & Mariscal 2001), contrary to expectations of increased competition. We hypothesize that mutualistic interactions with sea anemones facilitate niche partitioning among clownfish species, reducing interspecific competition and enabling coexistence. This mechanism would provide ecological support for the adaptive radiation hypothesis, demonstrating mutualism' role in promoting clownfish diversity. However, conclusive evidence of niche partitioning among clownfish species is lacking, hampering the testing of adaptive radiation hypotheses and challenging our understanding of clownfish diversification.

Furthermore, the interplay between generalist and specialist strategies likely influences clownfish distributions and community assembly (Chesson 2020; Bartholomew *et al.* 2022; Wandrag *et al.* 2022). Communities dominated by specialist-specialist interactions are expected to show limitations in species richness due to increased competition for specific anemone hosts. Conversely, communities with more generalist interactions could support higher diversity by reducing direct competition and allowing for more efficient resource utilization. Investigating this interplay could reveal spatial patterns linked to species richness, elucidating clownfish ecological roles, and deepening our understanding of how mutualistic interactions shape community structure, species coexistence, and biodiversity patterns.

To test these hypotheses, we need to estimate the realized environmental niches of clownfish species, while accounting for the effect of explicit mutualistic associations. However, current approaches quantifying ecological niches and modeling species distributions involve techniques like principal component analysis (PCA; Broennimann *et al.* 2012) and niche-based spatial modelling of species distributions (SDMs; see Guisan *et al.* 2014, Valavi *et al.* 2021; Norberg *et al.* 2019), do not directly incorporate biotic interactions. This is despite the demonstrated improvement in SDM predictions that the integration of biotic interactions can bring (Wisz *et al.* 2013; Early & Keith 2019; Kass *et al.* 2020; Jenkins *et al.* 2020) and the recurrent calls to develop SDMs that can integrate these interactions (see Soberón 2010; Boulangeat *et al.* 2012; de Araujo *et al.* 2013; Leach *et al.* 2016; D'Amen *et al.* 2018; but see Konig *et al.* 2021). Consequently, the role of biotic interactions is frequently overlooked (Anderson

2017; Palacio & Girini 2018), which can lead to misinterpretations of ecological niches and species distributions (Zurell *et al.* 2020; Moullec *et al.* 2022). In our case, such misinterpretations would hinder our capacity to fully understand the significant role that mutualism plays in shaping the ecological niche of clownfishes.

Here we built on an existing environmental niche quantification approach and implemented a novel method to reduce the biotic uncertainty, estimate the partial effects of known interactions, and accordingly refine the estimate of the ecological niche. To capture the mutualistic dependence of clownfishes on sea anemones and understand their effects on clownfishes niche, we adapted the 'COUE' framework (Guisan *et al.* 2014) by defining new metrics describing the effect of biotic interactions on niche quantification (Fig. 1). We compared the impact of biotic interactions on niche quantification between generalist and specialist clownfishes, hypothesizing that sea anemone associations significantly affect clownfish niches and distributions, with a greater effect on specialists than on generalists. Furthermore, we anticipate that incorporating mutualistic interactions as a limited resource into the quantification of resource overlap will reflect current competition dynamics among clownfishes, as well as their relationship to species richness and community assembly patterns.

Material & Methods

Data Collection

Clownfishes inhabit the Indo-Pacific Ocean, from the East coast of Africa to Polynesia in the Pacific, and from the coast of Japan to the South of Australia, while sea anemones have a broader, worldwide distribution (Fautin & Allen 1992). We collected 1,636 occurrences of ten host sea anemone species (mean: 163.6; min: 68; max: 335) and 4,258 occurrences of 30 clownfish species (mean: 146.8; min: 2; max: 860) from RLS, GBIF, OBIS and Hexacoral databases (Atlas of Living Australia 2017; GBIF.org 2018; OBIS 2017; Fautin 2008 respectively). Datasets were filtered to remove duplicates and misplaced or misidentified occurrences. After filtering, two clownfish species (Amphiprion pacificus and thiellei) with fewer than five occurrences were excluded.

Environmental data from GMED (Basher *et al.* 2018) and Bio-Oracle (Tyberghein *et al.* 2012; Assis *et al.* 2018) were obtained at 0.083° resolution, representing approximately 9,2 km near the equator, covering physical, chemical, and biological factors. The 53 environmental variables collected were restricted to shallow reefs and the epipelagic zone above 50m depth, using the UNEP-WCMC warm water coral reef map (UNEP-WCMC 2018). After removing variables with excessive missing data and discarding highly correlated variables (Pearson's correlation > 0.8), we retained eight environmental variables: mean current velocity, mean salinity, mean temperature, mean nitrate concentration, nitrate concentration range, mean chlorophyll concentration, dissolved oxygen concentration range, and mean phytoplankton concentration.

Quantifying ecological niches

We constructed a global environmental space using the first two principal components of a scaled PCA based on the eight selected environmental variables, which explained 39% and 22% of the total variance, respectively. We used the *ecospat* R package (Broennimann *et al.* 2012) to estimate the occurrence density o for both clownfishes and sea anemones in a two-dimensional grid of 100 by 100 cells:

$$o = \frac{\delta(n)}{\max(n)} \qquad (1)$$

where $\delta(n)$ is the 2D kernel density estimation of the number of occurrences on the defined environmental space, and $\max(n)$ is the maximum number of occurrences in any grid cell of the environmental space. The occurrence density o ranges from 0 for environments where the species is not observed to 1 where it is most observed. It represents the species ecological niche (i.e. realized environmental niche) and can be used as a proxy of the probability of occurrence in a given environment (Drake & Richards 2018). .

Refining the ecological niche using mutualistic interactions

We compiled the species-specific mutualistic interactions between sea anemones and clownfishes from scientific literature (Fautin 1985, 1991; Godwin & Fautin 1992; Ollerton *et al.* 2007; Ricciardi *et al.* 2010; Litsios *et al.* 2012) and reputable online sources (https://amphiprionology.wordpress.com, www.fishbase.org, https://reeflifesurvey.com) to construct an association matrix $\mathcal{A}$ where each clownfish species s was represented by a vector of interactions (Fig. 1):

$$\mathcal{A}_s = \begin{bmatrix} \alpha_1 & \cdots & \alpha_n \end{bmatrix}, \alpha \in \{0, 1\}$$

with n being the number of sea anemone species (here 10 in total). Given the obligate nature of the clownfish-anemone mutualism, we assumed that environments unsuitable for host anemones would be unavailable to clownfish, resulting in α values of either 0 or 1 in our case (see Supplementary Material & Methods for potential extensions). Environmental availability for clownfish would thus depend on two factors: i) the association between a present host species and the clownfish, as defined by matrix $\mathcal{A}$, and ii) the occurrence density of sea anemone species in the environment. The host availability ω was estimated as

$$\omega = 1 - \prod_{k=1}^{n} (1 - o_k \alpha_k) \qquad (2)$$

where α_k is the association between a clownfish and its host anemone k (derived from the matrix $\mathcal{A}$), while $_k$ is the occurrence density of the host k. ω values range from 0 to 1 and represent the availability score across the two-dimensional environmental space where a suitable host is

present. Then, the refined occurrence density o' of the focal clownfish species given the mutualistic associations was

$$o' = \omega o \qquad (3)$$

where o is the occurrence density of the focal clownfish species, and ω is the host availability across the environmental space.

Given clownfishes' limited dispersal capacity (Jones *et al.* 2005; Almany *et al.* 2017) and local adaptation of both clownfishes (Huyghe & Kochzius 2015; Clark *et al.* 2021; Ducret *et al.* 2022) and sea anemones (Sachkova *et al.* 2020; Will *et al.* 2021; Prakash *et al.* 2021), we anticipated regional ecological variations. We divided the study area into 27 regions across 5 marine realms based on MEOW (Spalding *et al.* 2007; Fig. S1). We estimated the ecological niche o for both clownfish and sea anemones, and the mutualism-refined niche o' for clownfishes, at regional scales by assessing species niches within each marine region's environmental space subset. Main analyses were conducted at a global scale and without environmental variable selection to test the robustness of our findings across spatial scales and variable inclusion (Fig. S2 and S3). Additionally, we projected the occurrence densities of each estimated niche into the geographical space for spatial representation of the results (Supplementary Material & Methods). We also conducted sensitivity analyses to evaluate the effect of the association matrix in our framework (Fig. S4 and Supplementary Material & Methods).

Effect of mutualistic interactions on species niches

We compared the clownfishes niche estimates with and without accounting for the sea anemones hosts occurrences (ecological niche o vs. mutualism-refined niche o') to understand the effect of implementing explicit mutualistic interactions in niche estimations. We measured different characteristics of niche space by adapting the 'COUE' framework (Guisan *et al.* 2014). In particular, we quantified changes in species' realized niches by defining three categories of occupied environmental space, accounting for the influence of sea anemone hosts. With N representing the number of cells in the environmental grid, we defined 'Unavailable' as the proportion of environmental space not occupied by the hosts of the given clownfish species and thus not available for the clownfish, calculated as $N_{(o>0\ \&\ o'=0)}/N_{(o>0)}$, 'Used' as the proportion of environmental space occupied by the given clownfish species, calculated as $N_{(o>0\ \&\ o'>0)}/N_{(o>0)}$, and 'Unoccupied' as the proportion of environmental space inhabited by any host of the given clownfish species but not by the clownfish, calculated as $N_{(o'>0)}/N_{(\omega>0)}$. To evaluate how the generalist-specialist spectrum affects ecological niche estimates incorporating mutualistic data, we classified species based on host associations. Following Ollerton *et al.* (2007) and Litsios *et al.* (2014), we defined generalists as species interacting with three or more hosts, and specialists as those with fewer than three hosts.

Fig. 1: Scheme of the proposed framework to assess the effect of biotic interactions and whether differential mutualistic behaviors may produce biases in the ecological niche estimation. For each clownfish species, georeferenced occurrences were collected, and ecological niche was estimated from the environmental space created using the selected environmental variables. Additionally, georeferenced occurrences of all sea anemones species were collected to infer their ecological niches following the same procedure as for the clownfish. Hosts ecological niches were combined into a single multi-hosts 'niche' using the interaction vector following the provided formula to create an envelope of host availability (ω). Finally, we constrained the clownfish ecological niche (o) by the host availability (ω) to obtain a mutualism-refined ecological niche (o'). Comparisons between the host availability envelope and the estimated clownfish ecological niche (dashed lines) provided the UUU parameters, determining Unavailable environments (environmentally suitable but unavailable due to lack of host availability), Used environments as they were both suitable and available, and Unoccupied environments as those that were available but not suitable for the clownfish.

Effect of mutualistic interactions on species niche overlaps

We examined how clownfish mutualistic interactions influence niche overlap among clownfish species. Using Schoener's D, we calculated pairwise ecological niche overlap between species, both with and without considering host anemone associations. Ecological and mutualism-refined niche overlap inadequately represents actual competition in clownfish, which is primarily driven by host use. To estimate resource-use competition more accurately, we developed a multi-layered approach. We replicated the environmental space into layers corresponding to each host, projected species ecological niches onto these layers, and constrained them by host suitability (skipping equation 2 and taking each host's occurrence density as ω). We then averaged layer-specific overlap values to obtain a single overlap measure for each species pair (Fig. S5). This method accounts for the host-specific nature of clownfish competition and provides a more realistic estimate of resource-use overlap. By integrating host specificity and environmental preferences, it captures the interplay between habitat requirements and resource competition in the clownfish-sea anemone mutualism system. We compared overlap estimates (ecological niche, mutualism-refined niche, and resource-use) and examined differences based on species pair specialization levels (generalist-generalist, generalist-specialist, and specialist-specialist).

Spatial patterns of clownfish interspecific niche overlap

We examined the relationship between species richness and both ecological niche and resource-use overlap, as well as their geographical patterns. Species richness was defined as the number of species predicted at a location, while overlap intensity was calculated as the average pairwise overlaps of all species at a site. Using spatial generalized linear mixed models (GLMMs) with the *spaMM* R package (Rousset & Ferdy 2014), we assessed the effect of species richness on both overlap estimates (ecological niche and resource-use), accounting for spatial autocorrelation (Fig. S6, S7 and Table S1). We then analyzed subsets of ecological niche and resource-use overlap for generalist-generalist, specialist-generalist, and specialist-specialist interactions, estimating species numbers for each interaction type. Finally, we conducted spatial GLMMs for

each subset to determine if niche overlap patterns varied by interaction type.

Results

Effect of mutualistic interactions on species niches

Incorporating mutualistic information significantly altered ecological niche estimates in 60% of the 108 regional niche subsets (Table S2). Across all clownfish species and regions, an average of 66% of a species ecological niche remained used after integrating mutualistic information ($\overline{X}_{Used} = 0.664 \pm 0.242$). Approximately one-sixth was unavailable due to unsuitable host environments ($\overline{X}_{Unavailable} = 0.179 \pm 0.249$), while a similar proportion was suitable for hosts but outside the clownfish ecological niche ($\overline{X}_{Unoccupied} = 0.178 \pm 0.193$). UUU proportions however, exhibited considerable variability among species (Fig. 2) and across regions (Fig. S8).

Fig. 2: Stacked bar plot showing the averaged UUU proportions per clownfish species among regions. Colours represent the different UUU parameters specified on the legend on top. White vertical lines represent the standard deviation across regions.

Specialists and generalists showed significant variations in UUU proportions (Fig. 3 and Table S3). Specialists had higher Unavailable ($H = 23.061, df = 1, p < 0.001$) and lower Used ($H = 17.129, df = 1, p < 0.001$) proportions in their ecological niches compared to generalists,

while no significant difference was detected for Unoccupied proportions ($H = 2.1735, df = 1, p = 0.14$). The stronger effects on specialist clownfish species Unavailable and Used proportions were evident through higher niche dissimilarity ($H = 39.987, df = 1, p < 0.001$), greater centroid shift ($H = 20.681, df = 1, p < 0.001$), and larger environmental shift ($H = 17.704, df = 1, p < 0.001$) when compared to generalists (Fig. S9). These UUU patterns were consistent at the spatial level (Fig. S10)

Fig. 3: Comparisons between generalists and specialists on the proportions of Unavailable (a), Used (b), and Unoccupied (c) proportions of the niche, adapted from the 'COUE' framework (Guisan *et al.* 2014). Violin plots show the distribution of the data. Statistical significance is represented following the legend: no significant (n.s.); $p < 0.05$ (*); $p < 0.01$ (**); $p < 0.001$ (***); $p < 0.0001$ (****).

Fig. 4: Pairwise species comparison of between ecological and mutualism-refined niche overlaps (a), and ecological niche and resource-use overlaps (b). Colored lines in the main plots (left) represent individual pairwise comparisons across categories, with blue lines indicating a decrease in overlap intensity and red lines showing an increase. Boxplots illustrate the overall intensity of niche overlap at each niche level, while violin plots show the distribution of pairwise overlaps for each category. Histograms on the right display the distribution of differences between pairwise overlaps, where blue bars indicate decreased overlap and red bars indicate increased overlap compared to ecological niche overlap. Statistical significance is represented following the legend: no significant (n.s.); $p < 0.05$ (*); $p < 0.01$ (**); $p < 0.001$ (***); $p < 0.0001$ (****).

Effect of mutualistic interactions on species niche overlaps

Clownfish ecological niche overlap, measured by Schoener's D statistic, was high (median $D_{ecological} = 0.741; IQR = 0.199$), indicating highly similar environmental suitability. Accounting for host associations slightly but significant ($V = 14,723; df = 1; p = 0.015$) increased niche overlap (median $D_{mutualism-refined} = 0.762; IQR = 0.220$) compared to ecological niche overlap. However, resource-use overlap was substantially lower (median $D_{resource-use} = 0.232; IQR = 0.304$) and significantly different from ecological and mutualism-refined niche overlap ($V = 35,327; df = 1; p < 0.001$ and $V = 33,650; df = 1; p < 0.001$, respectively; Fig. 4).

$D_{ecological}$ showed no significant differences among specialist-specialist, specialist-generalist, generalist-generalist interactions ($H = 4.193; df = 2; p = 0.122$). However, significant differences emerged after refining the ecological niche with mutualistic information ($H = 6.7509; df = 1; p = 0.034$), with stronger differences in resource-use overlap ($H = 71.623; df = 1; p < 0.001$). In specialist-specialist pairs, 15 comparisons showed zero resource-use overlap due to not shared hosts, while the remaining 13 pairs exhibited high overlap (median $D_{resource-use} = 0.456; IQR = 0.374$). Overlap decreased significantly in generalist-generalist (median $D_{resource-use} = 0.351; IQR = 0.212$) and generalist-specialist interactions (median $D_{resource-use} =$

$0.201; IQR = 0.172$), with the latter, being the most common among co-occurring clownfishes and exhibiting the lowest resource-use overlap (Fig. S11).

Geographical patterns of niche overlap

The Eastern and Western Coral Triangle regions showed the highest species richness and number of potential interactions (Fig 5a). High environmental suitability overlap was found in Pacific, Western and Central Indian Oceans regions for both ecological and mutualism-refined niches projections (Fig 5b; ecological niche projections shown). Resource-use overlap was more pronounced in the Tropical North-western Pacific and Somali/Arabian sea (Fig 5c). Generally, resource-use overlap was lower than environmental suitability overlap for within regions (Fig 5d), with the largest disparities in Temperate and Tropical Pacific regions, Northeast Australian Shelf, Coral Triangle, and Western Indian Ocean.

Resource-use overlap correlated positively with ecological overlaps across regions ($r = 0.800, p < 0.001$) and negatively with species richness ($r = -0.287, p < 0.001$). GLMMs revealed species richness negatively associated with resource-use overlap ($\beta = -0.013; 95\%CI[-0.015, -0.017]$), while environmental suitability overlap ($\beta = 0.668; 95\%CI[0.646, 0.689]$) showed a positive association. Their interaction had a negative impact on resource-use overlap ($\beta =$

0.015; 95%$CI[-0.011, 0.019]$; Fig. S12 and Table S4). Analyzing interaction subsets (generalist-generalist, specialist-generalist, and specialist-specialist), resource-use overlap among generalist-generalist interactions showed positively associated with generalist species richness ($\beta = 0.013$; 95%$CI[0.009, 0.017]$) and environmental suitability overlap ($\beta = 0.415$; 95%$CI[0.400, 0.430]$). Specialist-generalist and specialist-specialist interactions exhibited similar effects as the overall model, with varying magnitudes of association between environmental suitability and resource-use overlap (Table S4).

Discussion

Our study introduces a novel approach to incorporate mutualistic interactions in the estimation of ecological niches. Using clownfishes as a case study, we explored how species-specific mutualistic interactions with sea anemones influence the ecological niche. This allowed us to assess niche overlaps among clownfishes based on resource use, shedding light on competition patterns within clownfish communities. We then evaluated the role of mutualistic behaviour and host-partitioning in resource competing communities.

Effect of mutualistic interactions in clownfish species niches

Our study revealed significant misalignment between clownfish and host anemone ecological niches (Fig 2). Despite their obligate mutualism, we found only about 60% overlap between clownfish and host niches, contrary to our expectation of nearly-identical environmental suitability. This discrepancy exceeds that observed in non-dependent interactions of other organisms (e.g. Arumoogum *et al.* 2023). While some mismatch could be attributed to data limitations and methodological constraints, the extent of disagreement was unexpected given the nature of their relationship.

Comparing ecological niches (based on environmental factors) with mutualism-refined niches revealed deviations due to the presence or absence of suitable host anemones. These disparities can introduce potential biases into spatial models, especially for specialists species heavily constrained by biotic interactions (Meineri *et al.* 2012). Our approach incorporates explicit information about mutualistic interactions, refining ecological niche estimations and enabling detailed examination of environmental suitability discrepancies between clownfish and their host anemones.

Spatial projections of the UUU parameters showed 'Used' areas (suitable for both clownfish and host anemones) as predominant in species distributions (Fig. S13 and S14), indicating a strong agreement in environmental suitability. 'Unavailable' (suitable for clownfish only) and 'Unoccupied' (suitable for hosts only) areas represent less common environments, highlighting suitability mismatches between mutualistic partners (Fig. S15). The strong agreement between clownfish and host anemones ecological niches in predominant environments suggests an ecologically stable clownfish-sea anemone interaction,

Fig. 5: a) Estimated number of clownfish species occurring per location. b) Averaged ecological niche overlap among all co-occurring species per location. c) Average resource-use overlap among of all co-occurring species per location. d) Difference between ecological niche overlap and resource-use overlap computed as $D_{resource-use} - D_{ecological}$. Negative values represent higher ecological niche overlap than resource-use overlap and positive values represent higher resource-use niche overlap than ecological niche overlap.

likely maximizing habitat occupancy in a convergent manner. This pattern may indicate co-evolution, with both partners adapting to similar environmental conditions, driven by their symbiotic benefits.

Regions with high 'Unavailable' proportions due to host absence may indicate ecologically unstable or fluctuating populations, potentially representing sink populations. These areas show inconsistent observations of

both host anemones and clownfish. Such pattern could indicate non-self-sustaining populations relying on immigration from stable source populations. However, these sink populations might foster new ecological adaptations (Peniston *et al.* 2019). In source-sink dynamics, sink habitats can play crucial roles in adaptation by harboring increased genetic variation and experience reduced competition, allowing for survival of novel phenotypes (Holt, 1996; Kawecki, 2008). Temporal variation in harsh sink environments can facilitate niche evolution, with favorable periods allowing population growth and fixation of beneficial mutations (Lenormand, 2002; Holt *et al.* 2003). In clownfish-anemone systems, populations in low host availability areas might evolve wider environmental tolerances, utilize alternative hosts, or enhance dispersal capabilities. Long-term population genetic studies are needed to fully understand adaptive processes of clownfishes in marginal habitats.

'Unoccupied' environments could facilitate niche expansion for clownfish species (Bruno *et al.* 2003; Bulleri *et al.* 2016). Sea anemones may create favorable microenvironments (Arossa *et al.* 2021) enabling clownfish larvae to settle in otherwise harsh conditions. These areas could foster new environmental adaptations and driving range expansions (Chen *et al.* 2018; Álvarez *et al.* 2020). This host-mediated niche expansion might have facilitated the clownfish clade's expansion into the West Indian Ocean around 5 million years ago (Litsios *et al.* 2014), with long-established sea anemones (Titus *et al.*, 2019) providing an ecological opportunity for adapted clownfish. This hypothesis highlights the interplay between biotic interactions and abiotic factors in shaping species distributions and evolution. It also suggests sea anemones act as niche constructors, modifying environments for their symbionts, similar to mycorrhizal fungi altering soil for plants or corals building reef habitats.

Consistent with previous research on mutualistic networks (Bascompte & Jordano, 2007), generalist clownfish species showed higher niche overlap with host anemones compared to specialists. This suggests generalist niches are less constrained by biotic interactions, while specialist niches are strongly shaped by the specificity of their mutualistic relationship (Fig. 3). Specialist exhibited larger proportions of unavailable niches, likely due to their tight association with fewer hosts. This makes niche estimations incorporating biotic interactions more likely to differ from environmental-only models (Bateman *et al.* 2012), particularly for specialists linked to rare or patchily distributed hosts. Specialist niche models without accounting for biotic constrains may be prone to overfitting, potentially leading to inaccurate predictions, oversimplified understanding of complex ecological dynamics and misguided conservation efforts (Dormann *et al.* 2018). This is particularly crucial given as specialist species are often more vulnerable to climate change and extinction risks. Conversely, areas of overlapping predicted niches for clownfish and host anemones enhance model robustness, increasing confidence in distribution estimates and validating our understanding of their mutualism and ecological requirements.

While our niche estimations are considered reliable, potential biases stemming from imbalance occurrence data or incorrect biotic associations may exist. UUU parameters can identify areas needing better sampling or reveal misidentified associations, as seen in recent observations of *A. latezonatus*, *A. chagosensis* and *A. sebae* (Gaboriau *et al.* 2024). These species showed associations with more sea anemone species than previously known, potentially explaining high levels of unavailable niche proportions. *A. sebae*'s distribution, spanning from the Central Indian Ocean to the Eastern Coral Triangle, shows decreasing Unavailability proportions from west to east (100% in Central Indian Ocean Islands, 52% in Andaman, and 14% in Western Coral Triangle). This gradient suggests its western range could be facilitated by novel ecological interactions, indicating a shift in ecological structuring based on different host associations across its distribution. Similar patterns at smaller scales are observed for *A. chagosesis* (40% Unavailability in Central Indian Ocean Islands) and *A. latezonatus* (14% in East Central Australian Shelf and 100% in Lord Howe Island). These analyses provide a valuable tool for conservation strategies by identifying areas or populations requiring increased monitoring efforts.

Effect of mutualistic interactions on species niche overlaps

Our models provide insights into clownfish interspecific competition dynamics. Competition relies on physical interactions between species, making niche and distribution inferences crucial (Godsoe *et al.* 2015). Our approach enhances clownfish distribution estimations by considering their nested presence within hosts and allows studying interactions while accounting for resource partitioning due to varying host use.

Ecological niche overlaps were high, regardless of mutualistic associations and host use. As such, two clownfish in the same region would have similar environmental suitability despite inhabiting different sea anemones. Tropical reef fishes have narrow environmental niches, with limited tolerance to variations in temperature, salinity, pH, and oxygen levels (Brandl *et al.* 2020; Munday *et al.* 2008). Many species show reduced fitness when conditions deviate from their optimal range (Johansen *et al.* 2014; Rummer *et al.* 2013). Damselfishes exhibit decreased maximum oxygen uptake and aerobic scope at temperatures above 31°C (Habary *et al.* 2017). This specialization makes reef fish sensitive to environmental changes and vulnerable to climate impacts (Munday *et al.* 2008; Brandl *et al.* 2020). Resource-use overlap among clownfish species varied based on shared host anemones and mutualistic behaviours. Species sharing multiple host anemones exhibited greater overlap compared to those sharing fewer hosts. Generalist clownfish, associating with a wider range of anemone hosts, demonstrated higher resource-use overlap than specialists, restricted to fewer host species.

Our analyses revealed an inverse relationship between resource-use overlap and ecological niche overlap among

clownfishes. While sea anemone associations are critical for clownfish survival and reproduction (Lubbock 1980; Fautin 1991), factors driving species-specific host preferences remain unclear. We suggest that current mosaic of host preferences arose from long-term competitive dynamics aimed at minimizing resource-use overlaps, .facilitating species coexistence in optimal environments with high ecological niche overlap. This competition avoidance through resource partitioning is observed in various mutualistic systems, such as yucca-yucca moths, acacia-ant, plant-pollinator, and ant-myrmecophyte mutualisms (e.g., Addicott 1998; Palmer *et al.* 2003; Lee *et al.* 2010; Jeavons *et al.* 2020). In clownfishes, host resource-use overlap likely drives interspecific competition, as these species share fundamental ecological characteristics such as trophic position, reproductive behavior, phenology, and social structure. Host use and mutualistic behavior are primary differentiating factors, influencing coloration patterns and species recognition (Gaboriau *et al.* 2024). This suggests host use is a strong predictor of potential competitive dynamics among clownfish species.

While other ecological factors may influence local competition dynamics and the scale and resolution of the niche estimations could limit detection of biotic interactions (e.g. Pearson & Dawson 2003, Araujo & Rozenfeld 2014 and Fontoura *et al.* 2020), our analysis reveals potential effects of mutualistic interactions on clownfish coexistence and competitive dynamics at the regional scale. Our study reveals that generalist-specialist interactions are the most prevalent, with 67 unique interactions, particularly in species-rich environments. These interactions exhibit the lowest resource-use overlap, suggesting they are favoured in saturated communities, reducing competition and enabling species coexistence. Such interactions align with observations in plant-animal mutualistic systems, where generalist-specialist interactions contribute to biodiversity maintenance (Bascompte & Jordano 2007).

Geographical patterns of resource-use overlap

We observed distinct coexistence patterns among clownfish species across regions (Fig. S16 and Table S5, each linked to contrasting levels of ecological niche and resource-use overlap. These patterns reflect the complex interplay between host specialization and environmental adaptation in shaping distributions, aligning with the observed clownfish evolutionary dynamics (Litsios *et al.* 2014), suggesting a trade-off between host and environmental specialization.

Our findings align with Camp *et al.* (2016), where approximately 25% of clownfishes inhabiting the Coral Triangle were found in interspecific cohabiting groups, where clownfish species richness often exceeded host availability. This region characterized by high species co-occurrence and significant differences between ecological and resource-use overlap, exhibits a common coexistence pattern involving a generalist and a specialist with low resource-use overlap, indicating opportunities for coexistence through niche partitioning (Polechová & Storch

2019). However, we identified two alternative coexistence scenarios in which resource-use overlap was high, suggesting on-going competition.

First, regions at distribution edges such as the Western Pacific, Somali/Arabian Peninsula, and Central and Western Indian Ocean, exhibited high resource-use overlap driven mainly by closely related species. These regions, characterized by more recent colonization events (Litsios *et al.* 2014), likely represent areas of ongoing adaptation to new environmental conditions. These species pairs share host anemones while adapting to different environmental conditions, indicating recent divergence and specialization in distinct ecological niches. Some of those interactions have been reported as never coexisting within the same location, suggesting fine-scale competitive exclusion not captured by our regional-scale analyses.

Second, we found distantly related species, like *A. frenatus* and *A. biaculeatus* or *A. chrysopterus* and *A. percula*, converging at distribution edges and sharing hosts despite evolving in different ecological niches, indicating secondary contact and convergence on common hosts (Gaboriau *et al.* 2024). In these cases, we observed divergence between species in other traits such as coloration, morphology, or positioning relative to the host anemone. This observation aligns with findings by Camp *et al.* (2016). Such trait divergence likely serves to reduce competition when species share host anemones, enabling cohabitation on the same host while maintaining species recognition to prevent hybridization.

Our study reveals complex resource-use overlap dynamics influenced by species richness, ecological overlap, and specialization levels. Generalist-generalist interactions show increased resource-use overlap as ecological overlap and generalist species number rise. Coexistence is most likely in communities with few generalists and moderate ecological overlap. However, generalists' flexibility in host switching may mitigate competition locally. Specialist-specialist interactions exhibit a different pattern, where a larger number of specialists allows reduced competition if ecological overlap is low. Specialist-generalist interactions strike an optimal balance, supporting high species richness and ecological overlap while maintaining low resource-use overlap. This balance likely promotes species coexistence and may explain the remarkable clownfish diversity in the Coral Triangle, the most environmentally competitive area. These findings underscore the importance of considering specialization levels and ecological factors in understanding biodiversity maintenance mechanisms in complex ecosystems like coral reefs.

Conclusion

Our study introduces a novel framework for assessing species interactions' impact on niche estimation, with significant implications for conservation strategies and understanding mutualistic networks. Applied to clownfishes, we revealed the crucial role of resource partitioning in regulating competition, illuminating the evolution of diverse clownfish-sea anemone associations. Our findings support

established hypotheses in mutualistic systems, demonstrating how diverse strategies promote ecosystem sustainability and mitigate negative effects in saturated communities. Competition avoidance through resource partitioning emerges as a central mechanism shaping clownfish communities across the Indo-Pacific, aligning with broader ecological principles. While our study highlights importance of resource partitioning, it also raises questions about its evolutionary origins versus dynamic responses to current host composition. This framework advances clownfish ecology understanding and provides a valuable tool for investigating mutualistic interactions broadly. It can be extended to account for various interaction types, contributing to a more comprehensive understanding of biodiversity maintenance in complex ecosystems.

Supplementary information

Acknowledgements. We thank Daniele Silvestro, Pablo Duchen and Thibault Latrille for their contribution to the discussions of this study.

Declarations

Funding.   Financial support for this research was provided by the University of Lausanne funds and the Swiss National Science Foundation (Grant Number: 310030_185223).

Conflict of interest statement.   The authors declare no competing interests in the publication of this work.

Data availability. All data sets used and produced, figures and R scripts can be retrieved from the DRYAD repository (https://doi.org/10.5061/dryad.2bvq83bv8).

Code availability.   The scripts used for the analyses will be deposited on Dryad upon acceptance of the manuscript, ensuring the reproducibility and accessibility of our research findings.

Author contribution.   AGJ, TG and NS designed the study, AGJ collected the data, developed the implementation, performed the modelling, carried out the analyses, and wrote the initial manuscript. TG and NS contributed to the development of the implementation, the interpretation of the analyses and the writing of the manuscript, with help of all authors.

References

[1] Addicott, J. F. (1998). Regulation of mutualism between yuccas and yucca moths: population level processes. *Oikos*, 119-129. https://doi.org/10.2307/3546474

[2] Almany G. R., Planes S., Thorrold S. R., Berumen M. L., Bode M., Saenz-Agudelo P., … Jones G. P. (2017). Larval fish dispersal in a coral-reef seascape. *Nature Ecology and Evolution*, *1*(6), 1–7. https://doi.org/10.1038/s41559-017-0148

[3] Álvarez I., Fernández I., Traoré A. et al. Genomic scan of selective sweeps in Djallonké (West African Dwarf) sheep shed light on adaptation to harsh environments. *Sci Rep* 10, 2824 (2020). https://doi.org/10.1038/s41598-020-59839-x

[4] Anderson R. P. (2017). When and how should biotic interactions be considered in models of species niches and distributions? *Journal of Biogeography*. https://doi.org/10.1111/jbi.12825

[5] Araujo, M. B., & Rozenfeld, A. (2014). The geographic scaling of biotic interactions. *Ecography*, 37(5), 406-415. doi:DOI https://doi.org/10.1111/j.1600-0587.2013.00643.x

[6] Araújo M.B., Anderson R.P., Márcia Barbosa A., Beale C.M., Dormann C.F., Early R., Garcia R.A., Guisan A., Maiorano L., Naimi B. and O'Hara R.B., (2019). Standards for distribution models in biodiversity assessments. *Science Advances*, 5(1), p.eaat4858.

[7] Arossa S., Barozzi A., Callegari M., Klein S. G., Parry A. J., Hung S-H., Steckbauer A., Aranda M., Daffonchio D, Duarte C. M. (2021). The Internal Microenvironment of the Symbiotic Jellyfish Cassiopea sp. From the Red Sea. *Frontiers in Marine Science* 8 2021. doi. https://doi.org/10.3389/fmars.2021.705915 ISSN.2296-7745.

[8] Assis J., Tyberghein L., Bosch S., Verbruggen H., Serrão E. A., & De Clerck O. (2018). Bio-ORACLE v2.0: Extending marine data layers for bioclimatic modelling. *Global Ecology and Biogeography*, *27*(3), 277–284. https://doi.org/10.1111/geb.12693

[9] Atlas of Living Australia (2017). Reef Life Survey - Survey Records. Occurrence dataset https://doi.org/10.15468/4v5twn accessed via GBIF.org on 2020-05-11.

[10] Bartholomew, D. C., Banin, L. F., Bittencourt, P. R. L., Suis, M. A. F., Mercado, L. M., Nilus, R., Burslem, D. F. R., & Rowland, L. (2022). Differential nutrient limitation and tree height control leaf physiology, supporting niche partitioning in tropical dipterocarp forests. *Functional Ecology*, 36, 2084–2103. https://doi.org/10.1111/1365-2435.14094

[11] Bascompte J., & Jordano P. (2007). Plant-Animal Mutualistic Networks: The Architecture of Biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, *38*(1), 567–593. https://doi.org/10.1146/annurev.ecolsys.38.091206.095818

[12] Basher Z., Bowden D. A., Costello M. J. (2018). Global Marine Environment Datasets (GMED). World Wide Web electronic publication. Version 2.0 (Rev.02.2018). Accessed at http://gmed.auckland.ac.nz in June 2019

[13] Bastolla U., Fortuna M., Pascual-García A. et al. (2009). The architecture of mutualistic networks minimizes competition and increases biodiversity. *Nature* 458, 1018–1020 (2009). https://doi.org/10.1038/nature07950

[14] Bateman B.L., VanDerWal J., Williams S.E. and Johnson C.N. (2012), Biotic interactions influence the projected distribution of a specialist mammal under climate change. *Diversity Distrib.*, 18: 861-872. https://doi.org/10.1111/j.1472-4642.2012.00922.x

[15] Boulangeat I., Gravel D. and Thuiller W. (2012), Accounting for dispersal and biotic interactions to disentangle the drivers of species distributions and their abundances. *Ecology Letters*, 15: 584-593. https://doi.org/10.1111/j.1461-0248.2012.01772.x

[16] Brandl S.J., Johansen J.L., Casey J.M. *et al*. Extreme environmental conditions reduce coral reef fish biodiversity and productivity. *Nat Commun* 11, 3832 (2020). https://doi.org/10.1038/s41467-020-17731-2

[17] Broennimann O., Fitzpatrick M. C., Pearman P. B., Petitpierre B., Pellissier L., Yoccoz N. G., … Guisan, A. (2012). Measuring ecological niche overlap from occurrence and spatial environmental data. *Global Ecology and Biogeography*, *21*(4), 481–497. https://doi.org/10.1111/j.1466-8238.2011.00698.x

[18] Bruno J.F., Stachowicz J.J., Bertness M.D. (2003). Inclusion of facilitation into ecological theory. *Trends in Ecology & Evolution*, Volume 18, Issue 3, 2003, Pages 119-125, ISSN 0169-5347, https://doi.org/10.1016/S0169-5347(02)00045-9

[19] Bulleri F., Bruno J.F., Silliman B.R. and Stachowicz J.J. (2016), Facilitation and the niche: implications for coexistence, range shifts and ecosystem functioning. *Funct Ecol*, 30: 70-78. https://doi.org/10.1111/1365-2435.12528

[20] Camp E. F., Hobbs J.P. A., De Brauwer M., Dumbrell A. J. and Smith D. J. (2016). Cohabitation promotes high diversity of clownfishes in the Coral Triangle. *Proc. R. Soc. B*. 2832016027720160277. http://doi.org/10.1098/rspb.2016.0277

[21] Chen Y., Shenkar N., Ni P. et al. (2018). Rapid microevolution during recent range expansion to harsh environments. *BMC Evol Biol* 18, 187, 2018. https://doi.org/10.1186/s12862-018-1311-1

[22] Chesson, P. (2020). Species coexistence, Chapter 2. In: Theoretical Ecology: *Concepts and Applications (eds K.S. McCann & G. Gellner)*. Oxford University Press, Oxford, pp. 5–27.

[23] Clark, René D. and Aardema, Matthew L. and Andolfatto, Peter and Barber, Paul H. and Hattori, Akihisa and Hoey, Jennifer A. and Montes, Humberto R. and Pinsky, Malin L. (2021). Genomic signatures of spatially divergent selection at clownfish range margins. *Proc. R. Soc. B*. 288:20210407. https://doi.org/10.1098/rspb.2021.0407C

[24] Colwell, R. K., & Rangel, T. F. (2009). Hutchinson's duality: The once and future niche. *Proceedings of the National Academy of Sciences of the United States of America*, 106, 19651-19658. https://doi.org/10.1073/pnas.0901650106

[25] Cook, J. M., & Rasplus, J. Y. (2003). Mutualists with attitude: coevolving fig wasps and figs. *Trends in Ecology & Evolution*, 18(5), 241-248. 10.1016/S0169-5347(03)00062-4

[26] D'Amen M., Mod H. K., Gotelli N. J., & Guisan A. (2018). Disentangling biotic interactions, environmental filters, and dispersal limitation as drivers of species co-occurrence. *Ecography*. https://doi.org/10.1111/ecog.03148

[27] de Araújo C.B., Marcondes-Machado L.O. and Costa G.C. (2014). The importance of biotic interactions in species distribution models: a test of the Eltonian noise hypothesis using parrots. *J. Biogeogr.*, 41: 513-523. https://doi.org/10.1111/jbi.12234

[28] Dormann C.F., Bobrowski M., Dehling D.M., *et al*. Biotic interactions in species distribution modelling: 10 questions to guide interpretation and avoid false conclusions. *Global Ecol Biogeogr.* 2018; 27: 1004–1016. https://doi.org/10.1111/geb.12759

[29] Drake, J. M., and R. L. Richards. 2018. Estimating environmental suitability. *Ecosphere* 9(9):e02373. https://doi.org/10.1002/ ecs2.2373

[30] Ducret, H., Timm, J., Rodríguez-Moreno, M. et al. (2022). Strong genetic structure and limited connectivity among populations of Clark's Anemonefish (*Amphiprion clarkii*) in the centre of marine biodiversity. *Coral Reefs* 41, 599–609 (2022). https://doi.org/10.1007/s00338-021-02205-8

[31] Early R, Keith SA. (2018). Geographically variable biotic interactions and implications for species ranges. *Global Ecol Biogeogr.*; 28: 42– 53. https://doi.org/10.1111/geb.12861

[32] Elliott J.K. and Mariscal R.N. (2001). Coexistence of nine anemonefish species: differential host and habitat utilization, size and recruitment. *Mar. Biol*. 138:23–36. https://doi:10.1007/s002270000441

[33] Fautin D. (1985). Competition by anemone fishes for host actinians. *Proceedings of the 5th International Coral Reef Congress*, Vol. 5, pp. 373–377.

[34] Fautin D. G. (1991). The anemonefish symbiosis What is known and what is not. *Symbiosis*, *10*, 23–46.

[35] Fautin D.G. and G.R. Allen (1992). Field guide to anemonefishes and their host sea anemones. Western Australian Museum, Francis Street, Perth.

[36] Fautin D. G. and Buddemeier R. W. (2008). *Biogeoinformatics of the Hexacorals*
http://www.kgs.ku.edu/Hexacoral/

[37] Fontoura, L., Cantor, M., Longo, G.O., Bender, M.G., Bonaldo, R.M. and Floeter, S.R. (2020), The macroecology of reef fish agonistic behaviour. *Ecography*, 43: 1278-1290.
https://doi.org/10.1111/ecog.05079

[38] Frachon, L., Arrigo, L., Rusman, Q., Poveda, L., Qi, W., Scopece, G., & Schiestl, F. P. (2023). Putative signals of generalist plant species adaptation to local pollinator communities and abiotic factors. *Molecular Biology and Evolution*, 40(3), msad036.
https://doi.org/10.1093/molbev/msad036

[39] Franklin J. (2010). Mapping Species Distributions: Spatial Inference and Prediction (Ecology, Biodiversity and Conservation). *Cambridge University Press*.
https://doi:10.1017/CBO9780511810602

[40] Gaboriau, T., Marcionetti, A., Garcia Jimenez, A., Schmid, S., Fitzgerald, L. M., Micheli, B., Titus, B., & Salamin, N. (2024). Host-use drives convergent evolution in clownfish and disentangles the mystery of an iconic adaptive radiation. *bioRxiv*, 2024-07.
https://doi.org/10.1101/2024.07.08.602550

[41] *GBIF*.org (18 May 2018) GBIF Occurrence Download
https://doi.org/10.15468/dl.yohpbq

[42] Godsoe W., Jankowski J., Holt R. D., & Gravel D. (2017). Integrating Biogeography with Contemporary Niche Theory. *Trends in Ecology and Evolution*, *32*(7), 488–499.
https://doi.org/10.1016/j.tree.2017.03.008

[43] Godwin J. and Fautin D.G. (1992). Defense of Host Actinians by Anemonefishes. *Society, American*, (3), 902–908.

[44] Gracia-Lázaro C., Hernánde, L., Borge-Holthoefer J. et al. (2018). The joint influence of competition and mutualism on the biodiversity of mutualistic ecosystems. *Sci Rep* 8, 9253.
https://doi.org/10.1038/s41598-018-27498-8

[45] Guisan, A., Petitpierre B., Broennimann O., Daehler C., & Kueffer C. (2014). Unifying niche shift studies: insights from biological invasions. *Trends in Ecology & Evolution*, 29(5), 260–269.
https://doi.org/https://doi.org/10.1016/j.tree.2014.02.009

[46] Guisan A., Tingley R., Baumgartner J.B., Naujokaitis‐Lewis I., Sutcliffe P.R., Tulloch A.I., Regan T.J., Brotons L., McDonald‐Madden E., Mantyka‐Pringle C. and Martin T.G. (2013). Predicting species distributions for conservation decisions. *Ecology letters*, 16(12), pp.1424-1435.

[47] Guisan A., Thuiller W., & Zimmermann N. (2017). Habitat Suitability and Distribution Models: With Applications in R (Ecology, Biodiversity and Conservation). *Cambridge University Press*.
https://doi:10.1017/9781139028271

[48] Habary, A., Johansen, J. L., Nay, T. J., Steffensen, J. F., & Rummer, J. L. (2017). Adapt, move or die–how will tropical coral reef fishes cope with ocean warming?. *Global Change Biology*, 23(2), 566-577.
https://doi.org/10.1111/gcb.13488

[49] Hale K.R.S., Valdovinos F.S. & Martinez N.D. Mutualism increases diversity, stability, and function of multiplex networks that integrate pollinators into food webs.*Nat Commun* 11,2182 (2020).
https://doi.org/10.1038/s41467-020-15688-w

[50] Holt R.D., Gomulkiewicz R., and Barfield M. (2003). The phenomenology of niche evolution via quantitative traits in a 'black-hole' sinkProc. *R. Soc. Lond. B.*270215–224.
http://doi.org/10.1098/rspb.2002.2219

[51] Hutchinson, G. E. (1957). Concluding remarks. Cold Spring Harbor *Symposia on Quantitative Biology* 22: 415–427.

[52] Huyghe, F. and Kochzius, M. (2017). Highly restricted gene flow between disjunct populations of the skunk clownfish (*Amphiprion akallopisos*) in the Indian Ocean. *Mar Ecol*, 38: e12357.
https://doi.org/10.1111/maec.12357

[53] Jeavons, E., van Baaren, J., & Le Lann, C. (2020). Resource partitioning among a pollinator guild: A case study of monospecific flower crops under high honeybee pressure. *Acta Oecologica*, 104, 103527.
https://doi.org/10.1016/j.actao.2020.103527

[54] Jenkins D. A., Lecomte N., Andrews G., Yannic G., Schaefer J. A. (2020). Biotic interactions govern the distribution of coexisting ungulates in the Arctic Archipelago – A case for conservation planning, *Global Ecology and Conservation*, Volume 24, 2020, e01239, ISSN 2351-9894.
https://doi.org/10.1016/j.gecco.2020.e01239.

[55] Johansen, J. L., Messmer, V., Coker, D. J., Hoey, A. S., & Pratchett, M. S. (2014). Increasing ocean temperatures reduce activity patterns of a large commercially important coral reef fish. *Global Change Biology*, 20(4), 1067-1074.
https://doi.org/10.1111/gcb.12452

[56] Jones G. P., Planes S., & Thorrold S. R. (2005). Coral Reef Fish Larvae Settle Close to Home. *Current Biology*, *15*(14), 1314–1318.
https://doi.org/https://doi.org/10.1016/j.cub.2005.06.061

[57] Kass J.M., Anderson R.P., Espinosa-Lucas A., Juárez-Jaimes V., Martínez-Salas E., Botello F., Tavera G., Flores-Martínez, J.J. and Sánchez-Cordero, V. (2020), Biotic predictors with phenological information improve range estimates for migrating monarch butterflies in Mexico. *Ecography*, 43: 341-352. https://doi.org/10.1111/ecog.04886

[58] Kawecki, T. J. (2008). Adaptation to marginal habitats. *Annual review of ecology, evolution, and systematics*, 39(1), 321-342. https://doi.org/10.1146/annurev.ecolsys.38.091206.095622

[59] Konig C., Wuest R. O., Graham C. H., Karger D. N., Sattler T., Zimmermann N. E., and Zurell D. (2021). Scale dependency of joint species distribution models challenges interpretation of biotic interactions. *Journal of Biogeography* 48:1541-1551. https://doi.org/10.1111/jbi.14106

[60] Leach K., Montgomery W. I., & Reid N. (2016). Modelling the influence of biotic factors on species distribution patterns. *Ecological Modelling*. https://doi.org/10.1016/j.ecolmodel.2016.06.008

[61] Lee, C. T., & Inouye, B. D. (2010). Mutualism between consumers and their shared resource can promote competitive coexistence. *The American Naturalist*, 175(3), 277-288. https://doi.org/10.1086/650370

[62] Lenormand, T. (2002). Gene flow and the limits to natural selection. *Trends in ecology & evolution*, 17(4), 183-189. https://doi.org/10.1016/S0169-5347(02)02497-7

[63] Le Roux J.J. et al. (2020) Biotic Interactions as Mediators of Biological Invasions: Insights from South Africa. In: van Wilgen B., Measey J., Richardson D., Wilson J., Zengeya T. (eds) *Biological Invasions in South Africa. Invading Nature - Springer Series in Invasion Ecology*, vol 14. Springer, Cham. https://doi.org/10.1007/978-3-030-32394-3 14

[64] Lira-Noriega A., Soberón J., Miller C.P. (2013). Process-based and correlative modeling of desert mistletoe distribution: a multiscalar approach. *Ecosphere* 4(8): 1-23. https://doi.org/10.1890/ES13-00155.1

[65] Litsios G., Kostikova A., & Salamin N. (2014). Host specialist clownfishes are environmental niche generalists. *Proceedings of the Royal Society B: Biological Sciences*, 281(1795). https://doi.org/10.1098/rspb.2013.3220

[66] Litsios G., Pearman P.B., Lanterbecq D., Tolou N. and Salamin N. (2014), The radiation of the clownfishes has two geographical replicates. *J. Biogeogr.*, 41: 2140-2149. https://doi.org/10.1111/jbi.12370

[67] Litsios G., Sims C. A., Wüest R. O., Pearman P. B., Zimmermann N. E., & Salamin N. (2012). Mutualism with sea anemones triggered the adaptive radiation of clownfishes. *BMC Evolutionary Biology*, 12(1). https://doi.org/10.1186/1471-2148-12-212

[68] Lubbock R. (1980). Why are clownfishes not stung by sea anemones? *Proc. R. Soc. Lond. B.*20735–61. http://doi.org/10.1098/rspb.1980.0013

[69] Lunau, K. (2004). Adaptive radiation and coevolution—pollination biology case studies. *Organisms Diversity & Evolution*, 4(3), 207-224. https://doi.org/10.1016/j.ode.2004.02.002

[70] Marjakangas E-L., Abrego N., Grøtan V., *et al.* (2020). Fragmented tropical forests lose mutualistic plant–animal interactions. *Divers Distrib.* 2020; 26: 154–168. https://doi.org/10.1111/ddi.13010

[71] Mebs D. (2009). Chemical biology of the mutualistic relationships of sea anemones with fish and crustaceans. *Toxicon*. 2009 Dec 15;54(8):1071-4. doi: 10.1016/j.toxicon.2009.02.027. Epub 2009 Mar 5. PMID: 19268681.

[72] Meineri E., Skarpaas O., Vandvik V. (2012). Modeling alpine plant distributions at the landscape scale: Do biotic interactions matter? *Ecological Modelling*, Volume 231, 2012, Pages 1-10, ISSN 0304-3800, https://doi.org/10.1016/j.ecolmodel.2012.01.021

[73] Moullec F., Barrier N., Drira S., Guilhaumon F., Hattab T., Peck M. A., and Shin Y. J. (2022). Using species distribution models only may underestimate climate change impacts on future marine biodiversity. *Ecological Modelling* 464:11. https://doi.org/10.1016/j.ecolmodel.2021.109826.

[74] Munday, P. L., Jones, G. P., Pratchett, M. S., & Williams, A. J. (2008). Climate change and the future for coral reef fishes. *Fish and Fisheries*, 9(3), 261-285. https://doi.org/10.1111/j.1467-2979.2008.00281.x

[75] Norberg A., Abrego N., Blanchet F. G., Adler F. R., Anderson B. J., Anttila J., Araújo M. B., Dallas T., Dunson D., Elith J., Foster S. D., Fox R., Franklin J., Godsoe W., Guisan A., O'Hara B., Hill N. A., Holt R. D., Hui F. K. C., Husby M., Kålås J. A., Lehikoinen A., Luoto M., Mod H. K., Newell G., Renner I., Roslin T., Soininen J., Thuiller W., Vanhatalo J., Warton D., White M., Zimmermann N. E., Gravel D., and Ovaskainen O. (2019). A comprehensive evaluation of predictive performance of 33 species distribution models at species and community levels. *Ecological Monographs* 89(3):e01370. https://doi.org/10.1002/ecm.1370

[76] OBIS (2018). Ocean Biodiversity Information System Intergovernmental Oceanographic Commission

of UNESCO (2017)

[77] Ollerton J., McCollin D., Fautin D. G., & Allen G. R. (2007). Finding NEMO: Nestedness engendered by mutualistic organization in anemonefish and their hosts. *Proceedings of the Royal Society B: Biological Sciences*, *274*(1609), 591–598. https://doi.org/10.1098/rspb.2006.3758

[78] Palacio F.X. and Girini J.M. (2018), Biotic interactions in species distribution models enhance model performance and shed light on natural history of rare birds: a case study using the straight-billed reedhaunter *Limnoctites rectirostris*. *J Avian Biol*, 49: e01743. https://doi.org/10.1111/jav.01743

[79] Palmer T. M., Stanton M. L., and Young T. P. (2003). Competition and Coexistence: Exploring Mechanisms that Restrict and Maintain Diversity within Mutualist Guilds. *The American Naturalist* 162(S4): S63– 79. https://doi.org/10.1086/378682

[80] Pellissier L, Pinto-Figueroa E, Niculita-Hirzel H, Moora M, Villard L, Goudet J, Guex N, Pagni M, Xenarios I, Sanders I, Guisan A. (2013) Plant species distributions along environmental gradients: do belowground interactions with fungi matter? *Front Plant Sci*. 2013 Dec 10;4:500. doi: 10.3389/fpls.2013.00500. PMID: 24339830; PMCID: PMC3857535.

[81] Peniston J.H., Barfield M., and Holt R.D. (2019). Pulsed Immigration Events Can Facilitate Adaptation to Harsh Sink Environments. *The American Naturalist* 2019 194:3, 316-333. doi:10.1086/704608

[82] Pearson, R. G., & Dawson, T. E. (2003). Predicting the impacts of climate change on the distribution of species: are bioclimate envelope models useful?. *Global Ecology & Biogeography*, 12, 361-372.

[83] Polechová J. & Storch D. (2019). *Encyclopedia of Ecology (2nd Edition)*. Elsevier, Volume 3, p. 72-80. https://doi.org/10.1016/B978-0-12-409548-9.11113-3

[84] Prakash, S., Muthu, A., & Kumar, A. (2021). Population structure and reproductive performance in the sea anemone associated shrimp *Ancylocaris brevicarpalis* (Caridea: Palaemonidae). *Journal of the Marine Biological Association of the United Kingdom*, 101(1), 109-116. doi:10.1017/S002531542000137X

[85] R Core Team (2018). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Available online at https://www.R-project.org/

[86] Ricciardi F., Boyer, M., & Ollerton J. (2010). Assemblage and interaction structure of the anemonefish-anemone mutualism across the Manado region of Sulawesi, Indonesia. *Environmental Biology of Fishes*, *87*(4), 333–347. https://doi.org/10.1007/s10641-010-9606-0

[87] Rousset F. and Ferdy J-B. (2014). Testing environmental and genetic effects in the presence of spatial autocorrelation. *Ecography* 37(8): 781-790. http://dx.doi.org/10.1111/ecog.00566

[88] Rummer, J. L., Stecyk, J. A., Couturier, C. S., Watson, S. A., Nilsson, G. E., & Munday, P. L. (2013). Elevated CO_2 enhances aerobic scope of a coral reef fish. *Conservation Physiology*, 1(1), cot023. https://doi.org/10.1093/conphys/cot023

[89] Sachkova, M.Y., Macrander, J., Surm, J.M. et al. (2020). Some like it hot: population-specific adaptations in venom production to abiotic stressors in a widely distributed cnidarian. *BMC Biol 18*, 121 (2020). https://doi.org/10.1186/s12915-020-00855-8

[90] Salas-López, A., Violle, C., Munoz, F., Menzel, F., & Orivel, J. (2022). Effects of habitat and competition on niche partitioning and community structure in neotropical ants. *Frontiers in Ecology and Evolution*, 10, 863080. https://doi.org/10.3389/fevo.2022.863080

[91] Schlager S. (2017). "Morpho and Rvcg – Shape Analysis in R." In Zheng G, Li S, Szekely G (eds.), *Statistical Shape and Deformation Analysis*, 217–256. *Academic Press*. ISBN 9780128104934.

[92] Schleuning M., Fründ, J. and García, D. (2015), Predicting ecosystem functions from biodiversity and mutualistic networks: an extension of trait-based concepts to plant–animal interactions. *Ecography*, 38: 380-392. https://doi.org/10.1111/ecog.00983

[93] Schoener, T. W. (1974). Resource Partitioning in Ecological Communities: Research on how similar species divide resources helps reveal the natural regulation of species diversity. *Science*, 185(4145), 27-39. https://doi.org/10.1126/science.185.4145.27

[94] Soberón J. M. (2010). Niche and area of distribution modeling: A population ecology perspective. *Ecography*, *33*(1), 159–167. https://doi.org/10.1111/j.1600-0587.2009.06074.x

[95] Spalding M. D., Fox H. E., Allen G. R., Davidson N., Ferdaña Z. A., Finlayson M., Halpern B.S., Jorge M.A., Lombana A., Lourie S.A., Martin K.D., McManus E., Molnar J., Recchia C.A., Robertson J. (2007). Marine Ecoregions of the World: A Bioregionalization of Coastal and Shelf Areas. *BioScience*, *57*(7), 573–583. https://doi.org/10.1641/b570707

[96] Sverdrup-Thygeson A., Skarpaas, O., Blumentrath S., Birkemoe T., & Evju M. (2017). Habitat connectivity affects specialist species richness more than generalists in veteran trees. *Forest Ecology and Management*, *403*(1432), 96–102. https://doi.org/10.1016/j.foreco.2017.08.003

[97] Thioulouse J., Chessel D., Dolédec S., & Olivier J. M. (1997). ADE-4: A multivariate analysis and graphical display software. *Statistics and Computing*, *7*(1), 75–83. https://doi.org/10.1023/A:1018513530268

[98] Titus B. M., Benedict C., Laroche R., Gusmão L. C., van Deusen V., Chiodo T., Meyer C. P., Berumen M. L., Bartholomew A., Yanagi K., Reimer J. D., Fujii T., Daly M., & Rodríguez E. (2019). *Phylogenetic relationships among the clownfish-hosting sea anemones. Molecular Phylogenetics and Evolution*, 139. https://doi.org/10.1016/j.ympev.2019.106526

[99] Tyberghein L., Verbruggen H., Pauly K., Troupin C., Mineur F., & De Clerck O. (2012). Bio-ORACLE: A global environmental dataset for marine species distribution modelling. *Global Ecology and Biogeography*, *21*(2), 272–281. https://doi.org/10.1111/j.1466-8238.2011.00656.x

[100] UNEP-WCMC, WorldFish Centre, WRI, TNC (2021). Global distribution of warm-water coral reefs, compiled from multiple sources including the Millennium Coral Reef Mapping Project. Version 4.1. Includes contributions from IMaRS-USF and IRD (2005), IMaRS-USF (2005) and Spalding et al. (2001). *Cambridge (UK): UN Environment World Conservation Monitoring Centre*. https://doi.org/10.34892/t2wk-5t34

[101] Valavi R., Guillera-Arroita G., Lahoz-Monfort J. J., and Elith J. (2022). Predictive performance of presence-only species distribution models: a benchmark study with reproducible code. *Ecological Monographs* 92(1):e01486. https://doi.org/10.1002/ecm.1486

[102] Van der Niet, T., Peakall, R., & Johnson, S. D. (2014). Pollinator-driven ecological speciation in plants: new evidence and future perspectives. *Annals of Botany*, 113(2), 199-212. https://doi.org/10.1093/aob/mct290

[103] Wandrag, E.M., Catford, J.A. and Duncan, R.P. (2023), Niche partitioning overrides interspecific competition to determine plant species distributions along a nutrient gradient. *Oikos*, 2023: e08943. https://doi.org/10.1111/oik.08943

[104] Will H Ryan, Jaclyn Aida, Stacy A Krueger-Hadfield (2021). The Contribution of Clonality to Population Genetic Structure in the Sea Anemone, *Diadumene lineata*, Journal of Heredity, Volume 112, Issue 1, January 2021, Pages 122–139, https://doi.org/10.1093/jhered/esaa050

[105] Wisz M. S., Pottier J., Kissling W. D., Pellissier L., Lenoir J., Damgaard C. F., … Svenning J. C. (2013). The role of biotic interactions in shaping distributions and realised assemblages of species: Implications for species distribution modelling. *Biological Reviews*, *88*(1), 15–30. https://doi.org/10.1111/j.1469-185X.2012.00235.x

[106] Zurell D., Zimmermann N.E., Gross H., Baltensweiler A., Sattler T., Wüest R.O. (2020). Testing species assemblage predictions from stacked and joint species distribution models. *J Biogeogr.* 2020; 47: 101– 113. https://doi.org/10.1111/jbi.13608

Data

Clownfish
georeferenced
occurrences

Environmental
variables

Sea anemones
georeferenced
occurrences

Ecological niche

$$o = \delta(n)/\max(n)$$

Host-clownfish
interactions

**Mutualism-refined
niche**

$$o' = o\omega$$

Host availability

$$\omega_e = 1 - \prod_{k=1}^{n}(1 - o_{ek}\alpha_k)$$

UUU parameters

a

b

c