Radial askew endodermal cell divisions reveal IRK functions in division orientation

R. M. Imtiaz Karim Rony[1,2], Roya Campos[1], Patricio Perez-Henriquez[1], and Jaimie M. Van Norman[1,2]*

[1]Departments of Botany and Plant Sciences and [2]Molecular, Cell, and Systems Biology, Center for Plant Cell Biology, Institute of Integrative Genome Biology, University of California, Riverside, USA

***Corresponding Author:**
Jaimie M. Van Norman, jaimie.vannorman@ucr.edu

KEYWORDS
Arabidopsis root, development, cell division, division plane orientation, endodermis, cell polarity

SHORT TITLE
IRK functions in cell division orientation

ABSTRACT

Oriented cell divisions establish plant tissue and organ patterning and produce different cell types; this is particularly true of the highly organized Arabidopsis root meristem. Mutant alleles of *INFLORESCENCE AND ROOT APICES RECEPTOR KINASE* (*IRK*) exhibit excess cell divisions in the root endodermis. IRK is a transmembrane receptor kinase that localizes to the outer polar domain of these cells, which suggests directional signal perception is necessary to repress endodermal cell division. Here, a detailed examination revealed many of the excess endodermal divisions in *irk* have division planes that specifically skew towards the outer lateral side, therefore we termed them 'radial askew' divisions. Expression of an IRK truncation, lacking the kinase domain, retains polar localization and rescues these radial askew divisions, but the roots exhibit excess periclinal endodermal divisions. Using markers of cell identity, we show that the daughters of radial askew divisions transition from endodermal to cortex identity similar to those of periclinal divisions. These results extend the requirement for IRK beyond repression of cell division activity to include cell division plane positioning. Based on its polarity, we propose that IRK at the outer lateral endodermal cell face participates in division plane positioning to ensure normal root ground tissue patterning.

INTRODUCTION

In multicellular eukaryotes, cell division produces diverse cell types and increases cell number for body plan elaboration and growth. Cell divisions are typically categorized as proliferative (symmetric) and formative (asymmetric) . Proliferative cell divisions are typically physically symmetrical, producing daughter cells of the same size and cell fate. In contrast, formative cell divisions can be physically symmetrical or asymmetrical but the daughter cells acquire distinct identities. Due to the cell wall, the relative position of individual plant cells is fixed in space, as such, previous division orientations are 'recorded' by the placement of the walls (Facette et al., 2018; Rasmussen and Bellinger, 2018). The orientation of plant cell divisions is informed by extracellular cues and intracellular polarized proteins and is crucial for plant cell fate determination and tissue organization (Rasmussen et al., 2011; Van Norman, 2016; Wallner, 2020; Hartman and Muroyama, 2023). Because the developmental trajectory of a daughter cell is influenced by positional information (van den Berg et al., 1995; Marhava et al., 2019), division plane orientation is a key developmental decision.

The orientation of plant cell divisions is defined by how the new cell wall is positioned relative to the surface of the organ (Livanos and Müller, 2019). Accordingly, periclinal cell

62 divisions are oriented parallel to the surface and anticlinal cell divisions are oriented
63 perpendicular to the surface. For anticlinal cell divisions, the division plane can be aligned with
64 the organ's longitudinal or transverse axis (Figure 1A, B); therefore, we distinguish between
65 longitudinal anticlinal and transverse anticlinal cell divisions. In the root, periclinal cell divisions
66 are typically formative as they generate additional layers of distinct cell types (De Smet and
67 Beeckman, 2011; Pillitteri et al., 2016). Whereas, anticlinal cell divisions are often proliferative,
68 generating more cells of the same type. Normal tissue and organ patterning require the specific
69 orientation of proliferative and formative cell divisions relative to the plant body axes. Yet our
70 understanding of how division plane orientation is precisely controlled and how it is coupled with
71 cell division frequency remains limited.
72
73 The highly stereotypical cellular organization of the *Arabidopsis thaliana* root makes it an
74 ideal system to investigate how the frequency and orientation of cell division contribute to
75 tissue/organ patterning. The root ground tissue (GT), is a good example of how a specific series
76 of oriented formative cell divisions give rise to distinct cell types, the cortex and endodermis.
77 First, the GT stem cell, the cortex/endodermal initial (CEI) (Figure 1A, B), undergoes a
78 formative, transverse anticlinal division to produce the CEI daughter cell (CEID). This daughter
79 cell then undergoes a formative, periclinal division to produce endodermis towards the inside
80 and cortex cells towards the periphery of the root (Figure 1B) (Dolan et al., 1993; Scheres and
81 Benfey, 1999)). As plants mature, endodermal cells can undergo another periclinal division to
82 produce another cortex layer called middle or secondary cortex (Paquette and Benfey, 2005;
83 Cui, 2016). After their formation, cortex and endodermal cells proliferate through symmetrical,
84 transverse anticlinal divisions, producing more cells in their respective longitudinal files.
85 Symmetrical, longitudinal anticlinal divisions (LADs) can also occur producing more GT cells in
86 the root's radial axis (Figure 1A), however, these divisions are rarely observed in young wild
87 type roots and, as a result, eight of each GT cell type, including the CEI, are most often
88 observed around the stele (Dolan et al., 1993; Scheres and Benfey, 1999). Deviations in cell
89 division planes lead to irregular daughter cell shapes allowing the consequences of misoriented
90 root cell divisions to be followed in time and space.
91
92 Strict regulation of GT cell division in the root's radial axis is disrupted in mutant alleles
93 of *IRK* (*INFLORESCENCE AND ROOT APICES RECEPTOR KINASE*), which exhibit excess
94 endodermal longitudinal anticlinal and periclinal cell divisions (Campos et al., 2020). These
95 excess cell divisions coincide with promoter activity of *CYCLIN D6;1* (Campos et al., 2020), a

specific D-type cyclin, which is associated with formative, but not proliferative, GT cell divisions (Sozzani et al., 2010). Yet, no increase in proliferative, transverse anticlinal divisions was observed in *irk-4*, suggesting IRK specifically represses cell divisions that produce more cells in the root's radial axis (Campos et al., 2020). We propose that IRK specifically represses an endodermal cell division program that widens the GT and that longitudinal anticlinal and periclinal endodermal cell divisions are developmentally regulated downstream of IRK.

IRK encodes a transmembrane receptor kinase of the Arabidopsis RECEPTOR-LIKE KINASE (RLK) super family (Shiu and Bleecker, 2001; Shiu and Bleecker, 2003) and accumulates to the outer polar domain of the endodermal plasma membrane (PM). Although *IRK* is expressed in several root cell types, endodermal-specific expression of *IRK* is sufficient to rescue the cell division defects (Campos et al., 2020; Rodriguez-Furlan et al., 2022). Unexpectedly, IRK maintains its polar localization when the intracellular kinase domain is missing and this IRK truncation remains largely functional, rescuing the endodermal LADs but not the periclinal divisions in *irk-4* (Rodriguez-Furlan et al., 2022). Polar accumulation of IRK to the outer lateral endodermal cell face suggests it perceives an extracellular cue that originates peripheral to the endodermis and is required to maintain GT organization and cell number in the root's radial axis. However, the molecular details of IRK function remain unclear.

Here, we closely examined the excess endodermal divisions in *irk* mutants and identified a distinct cell division orientation defect. Unexpectedly in *irk-4,* a null allele, the majority of endodermal cell divisions previously characterized as periclinally oriented, instead, have outwardly skewed division planes. We defined these as 'radial askew' endodermal divisions, because these oblique divisions consistently produce small, abnormally shaped cells peripheral to the endodermis. In the weaker *irk-1* allele, roots have fewer radial askew endodermal cell divisions, but have excess longitudinal anticlinal and periclinal divisions. Therefore, while both *irk* alleles have excess GT cells in the radial axis, the orientation of the excess cell divisions is different. Additionally, the IRK intracellular truncation rescues the radially askew endodermal divisions in *irk-4*, but excess periclinal divisions are present, phenocopying *irk-1*. These results expand the function of IRK beyond repression of endodermal cell division activity showing it is also required for cell division plane orientation. Given its polar accumulation in the PM, IRK may participate in division plane orientation through physical involvement with cell division machinery or through perception of directional, positional information. We propose a model whereby the

129 presence of IRK at the outer polar domain occludes that endodermal cell face for division plane
130 selection or cell plate attachment.
131
132 **RESULTS**
133 **Many endodermal cell divisions in *irk-4* are abnormally oriented**
134 Close examination of endodermal cell division orientation in *irk-4* revealed that divisions
135 we had classified as periclinal were not actually oriented parallel to the root's surface. If a
136 periclinal division is considered in 2-dimensions (2D) in the longitudinal axis, the new cell plate
137 attaches to sites to opposite root/shootward endodermal faces within a cell file. However, in *irk-*
138 *4,* we observed abnormal divisions where cell plates attach to adjacent endodermal faces -
139 either the shoot- or rootward face and the outer lateral faces (Figure 1C-E). These abnormal,
140 oblique division planes resulted in new cell walls that often appeared periclinally oriented when
141 viewed only in longitudinal optical sections. However, in 3D, as observed in a series of
142 longitudinal and transverse optical sections (in confocal z-stacks (Campos and Van Norman,
143 2022)), key differences between true periclinal and these abnormal divisions become apparent.
144 Periclinal endodermal divisions produce daughter cells of nearly equal size, whereas the
145 abnormal divisions in *irk-4* result in unequal sized daughter cells with the smaller, prism-shaped
146 cell peripherally positioned. Intriguingly, we did not observe any abnormal divisions oriented
147 toward the inner lateral side of the endodermis, indicating division planes in *irk-4* are not
148 randomly misoriented. Given the specific orientation of these divisions, we termed them radial
149 askew endodermal divisions.
150
151 To assess the frequency of radial askew endodermal divisions, we examined a set of
152 confocal images (Rodriguez-Furlan et al., 2022) to specifically parse radial askew from periclinal
153 endodermal divisions. This revealed that 88.8% of the endodermal divisions in *irk-4* classified as
154 periclinal (n = 224 divisions) were actually radial askew divisions. Thus, there was no significant
155 difference in the number of periclinal divisions in *irk-4* compared to wild type (Wt) (Figure 1F). In
156 young Wt roots, periclinal divisions are somewhat rare and 5.1% of the divisions classified as
157 periclinal (n = 59 divisions) were found to be radial askew. In *irk-1*, a partial loss of function
158 allele, we found that 15.6% of divisions classified as periclinal (n = 154 divisions) are radial
159 askew oriented. Despite this, *irk-1* roots have significantly more periclinal divisions than Wt
160 (Figure 1F). These analyses revealed key phenotypic differences between the *irk* alleles: *irk-4*
161 has substantially more radial askew and longitudinal anticlinal divisions than Wt or *irk-1*,
162 whereas *irk-1* has significantly more periclinal and longitudinal anticlinal divisions that Wt.

163 Because *irk-1* has many fewer radial askew endodermal divisions than *irk-4,* these divisions
164 appear to be a specific phenotype observed upon complete loss of IRK function. These
165 observations indicate that IRK operates not only to repress endodermal cell division activity but
166 also functions in division orientation.
167
168 **The IRK kinase domain is dispensable for its role in cell division orientation**
169 We previously showed that GFP fused to a truncated version of IRK missing its kinase
170 domain (IRKΔK-GFP) was polarly localized in the endodermis (Figure 1G) and, while it fully
171 rescued the *irk-4* endodermal longitudinal anticlinal division (LAD) phenotype, it did not rescue
172 the periclinal division phenotype (Rodriguez-Furlan et al., 2022). This is reminiscent of the *irk-1*
173 root phenotype. As *irk-1* contains a T-DNA insertion near the beginning of the region coding for
174 the kinase domain (Campos et al., 2020), we predict that production of a partially functional
175 truncation results in the milder *irk-1* cell division phenotype. With the detection of novel
176 phenotypic consequences of *irk* loss of function and differences between the *irk* alleles, we
177 investigated whether IRKΔK-GFP could rescue the radial askew cell division phenotype of *irk-4*.
178
179 We found that *irk-4* roots expressing the full-length IRK-GFP or IRKΔK-GFP showed
180 rescue of endodermal LAD and radial askew phenotypes (Figure 1G, H). However, the *irk-4*
181 IRKΔK-GFP roots had significantly more periclinal endodermal divisions than Wt or *irk-4* IRK-
182 GFP plants, making them phenotypically similar to *irk-1*. This indicates presence of IRKΔK at
183 the outer polar domain is sufficient to repress longitudinal anticlinal and radial askew
184 endodermal divisions in *irk-4,* but leads to excess periclinal divisions. Thus, although excess
185 endodermal divisions are present in *irk-4* IRKΔK-GFP roots, they are predominantly periclinally
186 oriented, which is more typical for endodermal formative divisions (Paquette and Benfey, 2005;
187 Cui, 2016). These observations suggest the IRK kinase domain is largely dispensable in
188 endodermal division plane orientation, but is involved in repression of endodermal cell division
189 activity.
190
191 **Radial askew endodermal cell divisions in *irk-4* coincide with *CYCD6;1* promoter activity**
192 In Wt roots, formative GT cell divisions are associated with promoter activity of a specific
193 D-type cyclin, *pCYCD6;1*, this includes divisions of the initial cells and periclinal endodermal
194 divisions, which generate middle cortex (Sozzani et al., 2010). In *irk* mutants, excess
195 endodermal LADs also coincide with *pCYCD6;1* activity (Campos et al., 2020). This putatively
196 links longitudinal anticlinal and periclinal endodermal divisions through some shared regulatory

197 mechanism(s) downstream of IRK function and suggests IRK specifically represses an
198 endodermal cell division program that operates in the root's radial axis.
199
200 With the addition of radial askew divisions to the population of excess endodermal
201 divisions in *irk-4*, we conducted a detailed examination of *pCYCD6;1:erGFP* expression in *irk*
202 and Wt roots grown on 0.2x MS, which aggravates *irk* root phenotypes (Campos and Van
203 Norman, 2022; Goff et al., 2023). Similar to our previous report, *pCYCD6;1* activity is
204 substantially increased in the *irk* endodermis compared to Wt. We quantified abnormal
205 endodermal cell divisions together with expression of *pCYCD6;1:erGFP* and mapped this
206 information in the root's longitudinal axis (Figures 2, 3, and S1). In Wt roots, endodermal LADs
207 occur much less frequently than in *irk* and the daughter cells are rarely associated with
208 *pCYCD6;1* activity (Figure 2A-D). Under these conditions, periclinal divisions occur in Wt at this
209 age (6 days post-stratification) and, as expected, often associate with *pCYCD6;1* activity (Figure
210 S1A-D). In *irk-1*, there are more endodermal LADs than in Wt with daughter cells frequently
211 showing *pCYCD6;1* activity. *irk-4* roots have endodermal LADs at nearly every position in the
212 longitudinal axis and, with rare exception, these daughter cells show *pCYCD6;1* activity (Figure
213 2C, D). Like in Wt, periclinal endodermal divisions in *irk* alleles most often associate with
214 *pCYCD6;1* activity (Figure S1C, D). These results reveal extensive excess endodermal LADs in
215 *irk* mutants and confirm their strong association with *pCYCD6;1* activity.
216
217 Radial askew endodermal cell divisions are also associated with *pCYCD6;1* activity
218 (Figure 3). In the rare instances when radial askew divisions are observed in the Wt
219 endodermis, they are associated with *pCYCD6;1* activity (Figure 3A-C). This association was
220 also observed in *irk* mutants, where there are numerous radial askew endodermal divisions
221 (Figure 3B, C). In Wt and *irk-1*, radial askew endodermal divisions begin to occur five or more
222 cells above the quiescent center (QC), however in *irk-4,* radial askew divisions occur much
223 more frequently and closer to the QC (Figure 3C). Like endodermal LADs in *irk*, the daughter
224 cells of radial askew divisions nearly always show *pCYCD6;1* activity. Altogether, our
225 endodermal cell division maps reveal extensive excess endodermal cell divisions in the *irk*
226 mutants, particularly *irk-4,* where the daughter cells of LADs and radial askew divisions
227 consistently show *pCYCD6;1* activity. The association of *pCYCD6;1* activity with each of the
228 excess endodermal cell division orientations in *irk* further demonstrates the requirement for IRK
229 in repression of cell division activity in the endodermis.
230

231 **Peripheral daughter cells of radial askew endodermal divisions often express a marker of**
232 **cortex identity**
233 Endodermal periclinal, longitudinal anticlinal, and radial askew cell divisions all result in
234 an increased number of GT cells in the radial axis of *irk* roots. LADs produce more endodermal
235 cells around the stele and periclinal divisions produce a third GT layer, the middle cortex. We
236 expect that the daughters of endodermal LADs will maintain endodermal identity, while those of
237 periclinal divisions will transition to cortex identity (Figures 4, S2, and S3). As endodermal radial
238 askew cell divisions produce peripherally located, prism-shaped cells of unknown fate, we
239 examined reporter gene expression for endodermal and cortex cell identity (Figure 4). We first
240 examined *SCARECROW* promoter activity (*pSCR:erGFP*), which is expressed in endodermal
241 cells throughout the root (Di Laurenzio et al., 1996; Wysocka-Diller et al., 2000). In Wt, 43% (3
242 of 7) of peripheral daughters of radial askew divisions show *pSCR* activity. In *irk-1* and *irk-4*,
243 94% (25 of 26) and 60% (176 of 293) of these daughter cells show *pSCR* activity (Figure 4A-C),
244 respectively. Thus, regardless of genotype, not all peripheral daughters of radial askew
245 endodermal divisions show *pSCR* activity, implying they do not maintain endodermal identity.
246
247 Next, we examined activity of the *CORTEX2* promoter driving nuclear localized-yellow
248 fluorescent protein (*pCO2:nlsYFP*), which is expressed in cortex cells in the root meristem
249 (Heidstra et al., 2004; Paquette and Benfey, 2005). In Wt roots, cells immediately peripheral to
250 the endodermis have cortex identity (Figure 1B), whether those cells are derived from periclinal
251 CEID or endodermal divisions. Radial askew divisions were very infrequent in Wt and *irk-1*
252 expressing *pCO2:nlsYFP*. The single peripheral radial askew daughter in Wt did not express
253 *pCO2* and only one of four did in *irk-1.* In *irk-4,* ~21% (46 of 216) of the peripheral daughters of
254 radial askew divisions exhibited *pCO2* activity (Figure 4D-F). Comparison of the activity of
255 *pSCR* and *pCO2,* particularly in *irk-4*, suggests that peripheral daughters of endodermal radial
256 askew divisions gradually acquire cortex cell fate similar to what is observed for the peripheral
257 daughters of endodermal periclinal divisions.
258
259 To more directly investigate this, we examined *irk-4* roots simultaneously expressing
260 *pSCR* and *pCO2* reporters. Using linear unmixing, the daughters of endodermal longitudinal
261 anticlinal, periclinal, and radial askew divisions were examined (Figure 5A-D). The daughter
262 cells of endodermal LADs exclusively showed *pSCR* expression as expected, whereas the
263 daughter cells of periclinal and radial askew divisions showed expression of either or both of
264 these markers (Figure 5B). This suggests the daughter cells of radial askew and periclinal

265 divisions progress along a similar developmental trajectory and supports the idea that radial
266 askew divisions are formative divisions. Moreover, our observations suggest that *pCO2* activity
267 is progressively turned on in more distal peripheral daughters in files of these prism-shaped
268 cells (Figure 5C). Lastly, peripheral radial askew daughter cells more frequently show
269 simultaneous *pSCR* and *pCO2* activity, suggesting acquisition of cortex cell identity progresses
270 more slowly, than in the peripheral daughters of periclinal divisions (Figure 5D). Overall, we find
271 that peripheral radial askew daughters acquire cortex identity.
272
273 **Radial askew and periclinal endodermal divisions often co-occur in the root's**
274 **longitudinal axis**
275 Typically root cell divisions bisect cells strictly parallel or perpendicular to the root
276 surface. When considered in 2D in the transverse axis, periclinal endodermal divisions occur
277 when a new cell plate attaches to the circumferential endodermal faces and LADs occur when
278 the new cell plate attaches to the outer and inner endodermal faces. In each division, the pair of
279 resulting daughter cells of each division are similarly sized, but periclinal divisions are formative
280 and produce a new cell layer with a distinct cell fate (middle cortex, Figure 1A-C). In contrast,
281 the daughters of endodermal LADs remain endodermal. Radial askew endodermal divisions are
282 distinct, with a division plane oblique to the root surface that creates daughter cells of different
283 sizes, with the smaller prism-shaped cell consistently positioned peripherally. When considered
284 in 2D in the transverse axis, these divisions occur when a new cell plate attaches to the outer
285 lateral endodermal face and an adjacent circumferential endodermal face. Given the positions of
286 cell plate attachment (Figure 1C), radial askew divisions could simply be misoriented
287 longitudinal anticlinal or periclinal endodermal cell divisions. To address this we determined
288 whether endodermal cell division orientations could be spatially correlated.
289
290 Within individual cell files, we located endodermal cells that had undergone radial askew
291 division and assessed the number of instances in which the adjacent shootward and/or rootward
292 cell(s) had undergone a periclinal or longitudinal anticlinal division or had not divided at all
293 (immediately adjacent radial askew divisions were not counted, Figure 5E). In Wt, the rare radial
294 askew endodermal divisions (2 instances) had shootward and/or rootward neighbors that had
295 divided periclinally or not at all (Figure 5F). In *irk-1,* radial askew endodermal divisions were
296 adjacent to a periclinal division >60% of the time. Otherwise, the neighbor had not divided at all.
297 In *irk-4*, radial askew divisions were most often adjacent to endodermal cells that had not
298 divided and to periclinally divided cells just over 30% of the time. Unexpectedly, radial askew

299 divisions in *irk-4* were very rarely flanked by LADs (Figure 5F). Given that endodermal radial
300 askew divisions are very rare in Wt, their co-occurrence with periclinal division is notable.
301 Likewise in *irk-4*, it is striking that radial askew divisions are usually adjacent to endodermal
302 cells that have not divided or have undergone a periclinal division, given that endodermal LADs
303 in *irk-4* are extremely numerous. These results suggest that, regardless of genotype,
304 endodermal radial askew divisions are most often adjacent to cells that have divided periclinally
305 or not divided at all. Given that endodermal radial askew daughter cells are peripherally located,
306 transition to cortex identity, and their neighbors are more likely to have undergone a periclinal
307 division, we propose radial askew divisions are abnormal GT formative divisions.
308
309 **DISCUSSION**
310 We have identified excess and obliquely skewed endodermal cell divisions in *irk*
311 mutants, a novel aspect of the mutant phenotype that reveals IRK functions to repress
312 endodermal cell division activity and is required for cell division orientation. Typically,
313 endodermal cell divisions bisect the cell into similarly sized daughters, however, the divisions
314 we define as radial askew have oblique division planes creating small prism-shaped cells. Of
315 particular surprise is the invariable positioning of these smaller daughter cells towards the outer
316 lateral side - nearer the cortex. If there were random misorientation of endodermal divisions in
317 *irk*, we would expect that approximately half would be oriented towards the inner lateral side,
318 such that the smaller daughter cells formed nearer the pericycle. Because this was not
319 observed in any of our experiments, there is no support for random misorientation of
320 endodermal cell division in *irk*; instead, these divisions have a specific orientation. This is
321 consistent with a hypothesis where positional information orients cell division planes away from
322 the lateral endodermal cell faces in Wt, but in *irk* mutants, information is missing and division
323 planes frequently contact the outer lateral face.
324
325 The specific orientations of the excess divisions among *irk* mutants suggest endodermal
326 division activity and orientation are linked and tightly controlled. The role of IRK in this control is
327 particularly compelling when the spatial relationship between the *irk* cell division orientation
328 defects and IRK polar accumulation are considered (Figure 6A, B). In the endodermis of *irk-1* or
329 *irk-4 IRKΔK-GFP* roots a portion of IRK is present at the outer lateral domain of the PM and
330 there are fewer radial askew and LADs, but more periclinal divisions than in *irk-4*. In those
331 genotypes, excess endodermal cell divisions are present but the division planes contact the
332 outer lateral endodermal face infrequently. Whereas in *irk-4*, the putative null allele, excess

333 endodermal divisions are very frequently oriented such that the division plane contacts the outer
334 lateral face (Figure 6C). Therefore, the presence of IRK or IRKΔK at the outer lateral
335 endodermal face has a negative, potentially repellent, influence on division plane positioning.
336
337 To instruct cell division plane orientation, a polar-localized protein would need to
338 influence positioning of the cell division machinery. In plants, this includes the mitotic spindle
339 and two plant-specific cytoskeletal structures, the preprophase band, which marks the future
340 division site, and the phragmoplast, which assembles and guides cell plate formation
341 (Rasmussen et al., 2013; Smertenko et al., 2017). Many studies have investigated the
342 mechanisms underlying how a new cell plate is attracted to the marked division site. However,
343 our findings are consistent with a mechanism that repels cell plates from particular sites. We
344 predict dueling attractive and repulsive cues would increase the precision and robustness of the
345 cell division orientation process. For instance, in the *ton recruiting motif* (*trm) 6,7,8* triple mutant
346 the preprophase band is not detectable, yet the defects in root cell division orientation and
347 stomata patterning are modest (Schaefer et al., 2017). This hints that other mechanisms exist to
348 position division planes and there is increasing evidence that polarized proteins can directly
349 and/or indirectly position division planes via linkage with the cytoskeleton. For example, during
350 stomatal development, BASL/BRX family polarity domain depletes cortical microtubules at
351 particular positions ensuring the preprophase band forms outside the site marked by this
352 polarity domain (Muroyama et al., 2020; Muroyama et al., 2023). Identification of the molecular
353 links between IRK and the mechanics of cell division orientation and/or cytokinesis are key
354 avenues of future investigation.
355
356 We found radial askew endodermal divisions often spatially correlate with periclinal
357 divisions and their peripheral daughters progressively acquire cortex identity, suggesting that
358 they are formative in nature. Formative plant cell divisions often must traverse a longer path
359 than proliferative divisions, which typically occur along the shortest path that divides the cell
360 equally (Besson and Dumais, 2011; Facette et al., 2018; Rasmussen and Bellinger, 2018);
361 indeed, the path of an endodermal periclinal division is the longest in this cell type. Therefore, a
362 simple conclusion is that radial askew endodermal divisions are just misoriented periclinal
363 divisions, this would suggest IRK is needed to maintain division orientation parallel to the outer
364 lateral face during 'long path' endodermal formative divisions. However, contemplating IRK
365 function only in the context of maintaining the orientation of formative, periclinal endodermal

366 divisions due to the long division path fails to account for the excess endodermal LADs, which
367 can also be considered 'long path' divisions and are a key attribute of the *irk* phenotype.
368
369 Moreover, radial askew divisions are often flanked by endodermal cells that have not
370 divided at all. Comparing the instances of radial askew endodermal divisions in *irk-4* flanked by
371 no division, an LAD, or a periclinal division to the average number of radial askew divisions per
372 root, reveals that radial askew divisions occur in consecutive stretches along endodermal cell
373 files. If they were simply misoriented periclinal divisions, we might expect them to be more
374 interspersed with properly oriented periclinal divisions. If they are not misoriented periclinal
375 divisions - could radial askew divisions be a distinctly oriented formative endodermal cell
376 division? Two details hint at this, first, in addition to always being at the periphery of the
377 endodermis, the daughters of radial askew endodermal divisions very often occur at the position
378 where two cortex cells circumferentially meet (e.g. Figure 1C and 5A). Second, besides the
379 endodermal cell division defects, the stele area of *irk* roots, particularly *irk-4,* is substantially
380 wider than Wt due to increases in cell size (Campos et al., 2020; Goff et al., 2023). This likely
381 increases mechanical force on the peripheral cell layers and cell division orientations are altered
382 in response to changes in mechanical tension (Shapiro et al., 2015; Louveaux et al., 2016;
383 Marhava et al., 2019). Thus, radial askew endodermal divisions could be a distinct and specific
384 division orientation that generates precisely placed cells poised to invade the cortex layer in
385 response to widening of internal root cell types.
386
387 We show extensive activity of *pCYCD6;1* in *irk* roots and, consistent with its reported
388 activity in Wt, it is associated with formative GT divisions (Sozzani et al., 2010). However,
389 *pCYCD6;1* activity also strongly associates with endodermal LADs, which are proliferative
390 divisions. It is possible that excess *pCYCD6;1* activity in *irk* mutants simply reflects the high
391 number of endodermal cell divisions occurring at any one time and a general defect in
392 spatiotemporal control of cell cycle activity. Persistent expression of cell cycle genes after
393 cytokinesis could explain the excess endodermal cell divisions observed in *irk* mutants.
394 However, there is no increase in (proliferative) transverse anticlinal endodermal divisions that
395 would lengthen the *irk* root meristem (Campos et al., 2020), nor are these divisions associated
396 with *pCYCD6;1* activity. Because all of the excess divisions in *irk* are oriented such that they
397 broaden the root's radial axis and associated with *pCYCD6;1* activity, we propose that IRK
398 represses a GT cell division program specific to the root's radial axis with CYCD6;1 downstream
399 of IRK in this pathway.

400

401 In developmental biology, cell divisions are parsed into two groups: formative and
402 proliferative. Formative divisions are considered to require specialized cues to initiate and
403 properly orient; in plants, this often occurs in the long axis of the cell and creates an additional
404 cell layer with a distinct identity. In contrast, proliferative plant cell divisions tend to occur along
405 the shortest path to create equal sized daughter cells of the same identity. It is straightforward to
406 suggest that distinct mechanisms regulate proliferative and formative divisions. However, this is
407 not very satisfying when considering the endodermal divisions linked to IRK and CYCD6;1.
408 While neither IRK nor CYCD6;1 have known roles in proliferative cell divisions in the root's
409 longitudinal axis (Sozzani et al., 2010; Campos et al., 2020), they are linked to proliferative
410 divisions in the radial axis. This may mean that distinctly oriented proliferative divisions are also
411 differentially regulated from each other, in addition to the differential regulation between
412 proliferative and formative divisions. However, a more parsimonious explanation may be that
413 root cells separately regulate divisions that occur in different orientations. While this may seem
414 more complex than the currently accepted formative vs. proliferative division paradigm, it may
415 be more aligned with the underlying logic of plant body plan elaboration in respect to its organ
416 axes. Regardless, our study of IRK indicates that a distinct pathway operates to repress cell
417 divisions that specifically broaden the radial axis of the root GT - whether those divisions are
418 proliferative or formative.
419

420 **MATERIALS & METHODS**

421 **Plant material and growth conditions**

422 Seeds were surface sterilized with chlorine gas and plated on MS agar media. The
423 media contained 0.2X Murashige and Skoog salts (Caisson labs), 0.5g/L MES, 1% Sucrose,
424 and 1% agar (Difco) and was at pH 5.7. Plates were sealed with parafilm and placed at 4˚C for
425 24-72 hours for stratification. After stratification, plates were placed vertically in a Percival
426 incubator and grown under 16 hrs light/8hrs dark at a constant temperature of 22˚C. For each
427 experiment, the genotypes being analyzed were grown side-by-side on the same plate.
428 *Arabidopsis thaliana* ecotype Col-0 was used as the Wild type (Wt). The genotypes *irk-1*, *irk-4*,
429 *irk-4 pIRK:IRKΔK:GFP*, and *irk-4 pIRK:IRK:GFP* were obtained as previously described
430 (Campos et al., 2020; Rodriguez-Furlan et al., 2022). The cell type-specific reporters
431 *pCO2:nlsYFP (Paquette and Benfey, 2005)*, *pSCR:erGFP* (Di Laurenzio et al., 1996; Wysocka-
432 Diller et al., 2000), and *pCYCD6;1::GUS:GFP* (Sozzani et al., 2010) were received from the
433 Benfey lab and were crossed the *irk* mutants as previously described (Campos et al., 2020).

434

435 **Confocal microscopy and image analysis**

436 Roots were imaged at day 6 post-stratification (dps) and stained with ~10 μM propidium
437 iodide (PI) solubilized in water for 1-2 minutes; then, imaged on a Leica SP8 upright microscope
438 and imaging system housed in the Van Norman lab. Fluorescent protein (FP) signals were
439 captured using the following settings: PI (excitation 536 nm, emission 585-660 nm) and
440 GFP/YFP (excitation 488 nm, emission 492-530 nm). Z-stacks were acquired and analyzed with
441 the orthogonal sectioning tool of the LASX software. The total number of endodermal divisions
442 per root were counted from the QC to QC + 120μm or 20 cells as previously described (Campos
443 and Van Norman, 2022). Representative images of roots expressing the *pCYCD6;1:GUS:GFP*,
444 *pSCR:erGFP,* and *pCO2:nls:YFP* reporters were acquired from a transverse optical section at
445 the 10th endodermal cell (E10) shootward of the QC.

446 Signal separation GFP/YFP was performed by linear unmixing of a spectral image
447 (Zimmermann et al., 2014), acquired with a inverted Zeiss 880 microscope with 32 channel
448 spectral detector, objective 40X/1.2 W Korr FCS M27, MBS 488/561 and exciting with both 561
449 nm and 488 nm. Signal was collected simultaneously in 18 channels from 463 to 624 nm in ~9
450 nm bands. GFP/YFP and PI signal was subsequently separated using Zeiss ZEN software linear
451 unmixing tools.

452 For the images used to quantify endodermal cell divisions and generate the endodermal
453 cell division map (see below), Z-stacks were acquired from each root at 512×512 pixels with 1
454 μm between optical sections. All three genotypes expressing a single reporter were grown on
455 the same plate, 14-15 plants per genotype were imaged, and ≥10 plants per genotype were
456 analyzed in detail. For each root, the optical sections were manually analyzed from the QC
457 shootward 20 endodermal cells (E1-20), such that >160 endodermal cells per root were
458 examined for excess endodermal cell divisions.

459

460 **Generation of the endodermal cell division maps**

461 To generate the endodermal cell division maps for each genotype, we examined
462 promoter activity and the presence of periclinal, radial askew, and longitudinal anticlinal
463 divisions in each optical transverse section from just above the QC shootward to endodermal
464 cell position 20 (E20). If a division was present, we documented whether the daughter cell was
465 positive or negative for the reporter. For plants expressing *pCYCD6;1:erGFP*, we examined
466 endodermal LADs, periclinal, and radial askew divisions. For plants expressing *pSCR:erGFP*
467 and *pCO2:nls:YFP*, we only examined the endodermal periclinal and radial askew divisions, as

468 the daughters of endodermal LADs remain endodermal cells. The total number of these
469 divisions was quantified per root and then counted as GFP/YFP positive (+) or GFP/YFP
470 negative (-). We note that the excess periclinal endodermal division phenotype in *irk-1* appears
471 slightly repressed in the reporter backgrounds (compare Figure 1F to S1B, S2B, and S3B),
472 however, this is likely due to general variability in the number of periclinal divisions in any
473 genotype and relatively low number of roots (n = ~40 vs. ~10, respectively) examined per
474 replicate.
475 The endodermal cell division map shows whether periclinal, radial askew, and/or
476 longitudinal anticlinal divisions were present at a given position (E1-E20). This information is
477 compressed as the map does not include the absolute number of divisions or map divisions in
478 the root's radial axis. Specifically, if a box in the division map is a solid color, then all the
479 daughter cells for a given endodermal division that is present expressed the reporter - whether
480 there was a single division or several. If some, but not all, of the daughter cells expressed the
481 reporter, then the box in the map has a gradient fill. Finally, if none of the daughter cells
482 expressed the reporter the box is unfilled (white). Endodermal division orientations are
483 represented by different shapes in each box in the endodermal cell division maps: LADs are
484 indicated by a black arrowhead on the right, internal side; periclinal divisions are indicated by a
485 black arrowhead on the lower, internal side, and radial askew divisions are indicated by a
486 curved line in the lower-right corner. If none of these endodermal cell divisions were detected,
487 the box is empty.
488
489 **Assessing the spatial correlation between endodermal division orientations**
490 To determine if endodermal radial askew divisions were correlated with periclinal or
491 longitudinal anticlinal division, the endodermal cell division pattern along the root's longitudinal
492 axis was examined in each genotype expressing *pCYCD6;1:GUS:GFP*. In endodermal cell files
493 from the QC upwards, transverse optical sections were examined for a radial askew division; then
494 the adjacent shootward and rootward cells were examined for an abnormal division or no division.
495 If the adjacent cell also had a radial askew division, we proceeded to look until an endodermal
496 cell that had a periclinal or longitudinal anticlinal division or no division was found up to E20 for
497 all cell files.

498 **Figure generation**

499 Confocal images were exported from Leica software (LASX) as TIF files, which were
500 cropped and resized in Adobe Photoshop. Graphs were generated with PRISM8 (GraphPad

software https://www.graphpad.com/, San Diego, USA). Schematics were created in Adobe Illustrator. The figures containing confocal images, graphs, and schematics were assembled in Adobe Illustrator.

ACKNOWLEDGEMENTS

We thank former members of the Van Norman lab, including Dr. Jason Goff, Dr. Jessica N. Toth, and Dr. Cecilia Rodriguez Furlan, and Dr. Carolyn Rasmussen (UC, Riverside (UCR)) for discussions of and feedback on this manuscript while it was in preparation. We also thank Dr. Patricia Springer (UCR) for her insightful suggestion for how to display the endodermal division map. We appreciate access to the Zeiss 880 confocal microscope housed in the Institute of Integrative Genome Biology Microscopy Core Facility (UCR). This work was supported by funds from UCR, including Initial Complement (IC) and Academic Senate funds, USDA-NIFA-CA-R-BPS-5156-H, and NSF CAREER #1751385 awarded to J.M.V.N.

AUTHOR CONTRIBUTIONS

Conceptualization: RC, JMVN, and RMIKR; Cell division methodology: RC; Endodermal division & FP mapping: RMIKR and RC; Dual FP examination: PPH; Resources: JMVN and RC.; Writing - Original Draft and Review and Editing: JMVN, RC, RMIKR, and PPH; Visualization: JMV, RMIKR, RC, and PPH; Supervision: JMVN; Funding Acquisition: JMVN.

REFERENCES

van den Berg C, Willemsen V, Hage W, Weisbeek P, Scheres B (1995) Cell fate in the Arabidopsis root meristem determined by directional signalling. Nature **378**: 62–65

Besson S, Dumais J (2011) Universal rule for the symmetric division of plant cells. Proc Natl Acad Sci U S A **108**: 6294–6299

Campos R, Goff J, Rodriguez-Furlan C, Van Norman JM (2020) The Arabidopsis Receptor Kinase IRK Is Polarized and Represses Specific Cell Divisions in Roots. Dev Cell **52**: 183–195.e4

Campos R, Van Norman JM (2022) Confocal Analysis of Arabidopsis Root Cell Divisions in 3D: A Focus on the Endodermis. Methods Mol Biol **2382**: 181–207

Cui H (2016) Middle Cortex Formation in the Root: An Emerging Picture of Integrated Regulatory Mechanisms. Mol Plant **9**: 771–773

De Smet I, Beeckman T (2011) Asymmetric cell division in land plants and algae: the driving force for differentiation. Nat Rev Mol Cell Biol **12**: 177–188

535 **Di Laurenzio L, Wysocka-Diller J, Malamy JE, Pysh L, Helariutta Y, Freshour G, Hahn MG,**
536 **Feldmann KA, Benfey PN** (1996) The SCARECROW gene regulates an asymmetric cell
537 division that is essential for generating the radial organization of the Arabidopsis root. Cell
538 **86**: 423–433

539 **Dolan L, Janmaat K, Willemsen V, Linstead P, Poethig S, Roberts K, Scheres B** (1993)
540 Cellular organisation of the Arabidopsis thaliana root. Development **119**: 71–84

541 **Facette MR, Rasmussen CG, Van Norman JM** (2018) A plane choice: coordinating timing and
542 orientation of cell division during plant development. Curr Opin Plant Biol **47**: 47–55

543 **Goff J, Imtiaz Karim Rony R, Ge Z, Hajný J, Rodriguez-Furlan C, Friml J, Van Norman JM**
544 (2023) PXC2, a polarized receptor kinase, functions to repress ground tissue cell divisions
545 and restrict stele size. bioRxiv 2021.02.11.429611

546 **Hartman KS, Muroyama A** (2023) Polarizing to the challenge: New insights into polarity-
547 mediated division orientation in plant development. Curr Opin Plant Biol **74**: 102383

548 **Heidstra R, Welch D, Scheres B** (2004) Mosaic analyses using marked activation and deletion
549 clones dissect Arabidopsis SCARECROW action in asymmetric cell division. Genes Dev
550 **18**: 1964–1969

551 **Livanos P, Müller S** (2019) Division Plane Establishment and Cytokinesis. Annu Rev Plant Biol
552 **70**: 239–267

553 **Louveaux M, Julien J-D, Mirabet V, Boudaoud A, Hamant O** (2016) Cell division plane
554 orientation based on tensile stress in Arabidopsis thaliana. Proc Natl Acad Sci U S A **113**:
555 E4294–303

556 **Marhava P, Hoermayer L, Yoshida S, Marhavý P, Benková E, Friml J** (2019) Re-activation of
557 Stem Cell Pathways for Pattern Restoration in Plant Wound Healing. Cell **177**: 957–
558 969.e13

559 **Muroyama A, Gong Y, Bergmann DC** (2020) Opposing, Polarity-Driven Nuclear Migrations
560 Underpin Asymmetric Divisions to Pattern Arabidopsis Stomata. Curr Biol. doi:
561 10.1016/j.cub.2020.08.100

562 **Muroyama A, Gong Y, Hartman KS, Bergmann DC** (2023) Cortical polarity ensures its own
563 asymmetric inheritance in the stomatal lineage to pattern the leaf surface. Science **381**: 54–
564 59

565 **Paquette AJ, Benfey PN** (2005) Maturation of the ground tissue of the root is regulated by
566 gibberellin and SCARECROW and requires SHORT-ROOT. Plant Physiol **138**: 636–640

567 **Pillitteri LJ, Guo X, Dong J** (2016) Asymmetric cell division in plants: mechanisms of symmetry
568 breaking and cell fate determination. Cell Mol Life Sci. doi: 10.1007/s00018-016-2290-2

569 **Rasmussen CG, Bellinger M** (2018) An overview of plant division-plane orientation. New
570 Phytol **219**: 505–512

571 **Rasmussen CG, Humphries JA, Smith LG** (2011) Determination of symmetric and
572 asymmetric division planes in plant cells. Annu Rev Plant Biol **62**: 387–409

Rasmussen CG, Wright AJ, Müller S (2013) The role of the cytoskeleton and associated proteins in determination of the plant cell division plane. Plant J **75**: 258–269

Rodriguez-Furlan C, Campos R, Toth JN, Van Norman JM (2022) Distinct mechanisms orchestrate the contra-polarity of IRK and KOIN, two LRR-receptor-kinases controlling root cell division. Nat Commun **13**: 235

Schaefer E, Belcram K, Uyttewaal M, Duroc Y, Goussot M, Legland D, Laruelle E, de Tauzia-Moreau M-L, Pastuglia M, Bouchez D (2017) The preprophase band of microtubules controls the robustness of division orientation in plants. Science **356**: 186–189

Scheres B, Benfey PN (1999) ASYMMETRIC CELL DIVISION IN PLANTS. Annu Rev Plant Physiol Plant Mol Biol **50**: 505–537

Shapiro BE, Tobin C, Mjolsness E, Meyerowitz EM (2015) Analysis of cell division patterns in the *Arabidopsis* shoot apical meristem. Proc Natl Acad Sci U S A **112**: 4815–4820

Shiu SH, Bleecker AB (2001) Receptor-like kinases from Arabidopsis form a monophyletic gene family related to animal receptor kinases. Proc Natl Acad Sci U S A **98**: 10763–10768

Shiu SH, Bleecker AB (2003) Expansion of the receptor-like kinase/Pelle gene family and receptor-like proteins in Arabidopsis. Plant Physiol **132**: 530–543

Smertenko A, Assaad F, Baluška F, Bezanilla M, Buschmann H, Drakakaki G, Hauser M-T, Janson M, Mineyuki Y, Moore I, et al (2017) Plant Cytokinesis: Terminology for Structures and Processes. Trends Cell Biol **27**: 885–894

Sozzani R, Cui H, Moreno-Risueno MA, Busch W, Van Norman JM, Vernoux T, Brady SM, Dewitte W, Murray JAH, Benfey PN (2010) Spatiotemporal regulation of cell-cycle genes by SHORTROOT links patterning and growth. Nature **466**: 128–132

Van Norman JM (2016) Asymmetry and cell polarity in root development. Dev Biol **419**: 165–174

Wallner E-S (2020) The value of asymmetry: How polarity proteins determine plant growth and morphology. J Exp Bot. doi: 10.1093/jxb/eraa329

Wysocka-Diller JW, Helariutta Y, Fukaki H, Malamy JE, Benfey PN (2000) Molecular analysis of SCARECROW function reveals a radial patterning mechanism common to root and shoot. Development **127**: 595–603

Zimmermann T, Marrison J, Hogg K, O'Toole P (2014) Clearing Up the Signal: Spectral Imaging and Linear Unmixing in Fluorescence Microscopy. *In* SW Paddock, ed, Confocal Microscopy: Methods and Protocols. Springer New York, New York, NY, pp 129–148