Pelvic spine reduction affects diet but not gill raker morphology in two polymorphic brook

stickleback (*Culaea inconstans*) populations

Running title: Brook stickleback diet and gill rakers

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Abstract

Pelvic spine polymorphism occurs in several species in the stickleback family

(Gasterosteidae). Given parallel selection driving similar phenotypic polymorphisms in multiple

stickleback species, we sought to determine the extent of parallelism in the ecological

consequences of pelvic spine reduction. Based on a metabarcoding analysis of brook stickleback

gut contents in two polymorphic populations, we found a shift towards a planktonic diet was

associated with pelvic spine reduction. These results contrast with those found in threespine

stickleback where pelvic spine reduction is associated with a shift towards a benthic diet. Hence,

we found non-parallel consequences of spine polymorphism across species. Furthermore, a

change in gill raker morphology has been consistently implicated in the change in diet in pelvic-

reduced threespine stickleback. But we found no evidence of any difference in gill raker

morphology associated with pelvic spine polymorphism in brook stickleback.

Introduction

The maintenance of polymorphisms in natural populations is a central concept in

evolutionary biology. Polymorphisms are of particular interest when they are maintained at a

frequency that is too high to be explained by mutation-selection balance (Ford 1945, Lande

1975). In cases where a polymorphism persists in a population at high frequency, the

maintenance of the polymorphism is typically explained by some form of balancing selection or

by heterosis (Dobzhansky 1971, Takahata et al. 1992, Rainey et al. 2000, Gigord et al. 2001,

Punzalan et al. 2005). These cases are of interest because they afford the opportunity to study the

genetic and ecological mechanisms that result in the balance of selective forces that maintain

diversity. Balancing selection may be driven by a single ecological mechanism, such as in the

case of pollinator-mediated negative frequency dependent selection on flower colour (e.g.

Gigord et al. 2001). Balancing selection may also be the result of the combined effects of

multiple ecological mechanisms (i.e. predation, competition, and parasitism). Balancing selection

maintaining a particular polymorphism has the potential to affect other traits via pleiotropy, and

the polymorphism may cause divergence in a suite of traits via plastic or evolutionary responses

to the ecological or behavioural consequences of the polymorphism. For example, polymorphism

in the length (or presence) of protective spines in freshwater populations of threespine

stickleback (*Gasterosteus acculeatus* Linnaeus, 1758) is partly driven by a balance of predation

pressures from fish and other predators, and the polymorphism is often associated with a suite of

divergent traits associated with diet, habitat, parasitism, and mate choice (Rennison et al. 2019b).

Studies of adaptive polymorphisms in threespine sticklebacks have provided foundational

contributions to our understanding of evolutionary processes (Schluter and McPhail 1992,

Reimchen 1994, Schluter 2000, Bolnick 2004, Colosimo et al. 2005, Chan et al. 2010, Hendry et

al. 2013, Schluter et al. 2022). The establishment of threespine stickleback as a model system in

evolutionary biology (Reid et al. 2021) has also increased interest in the study of adaptive

polymorphisms within other species in the stickleback family, Gasterosteidae (Blouw and Boyd

1992, Shapiro et al. 2009, Klepaker et al. 2013, Shikano et al. 2013, Lowey et al. 2020, Jeffries

et al. 2022, Willerth et al. 2022). For example, spine reduction or loss has evolved independently

in four stickleback genera that share a common ancestor approximately 27 million years ago:

Gasterosteus, *Pungitius*, *Apeltes*, and *Culaea* (Nelson 1969, Nelson 1977, Hagen and Blouw

1983, Blouw and Hagen 1984a, Chan *et al.* 2010, Varadharajan et al. 2019). The involvement of

predation by birds, fish, mammals, and insects as a selective agent on spines has been established

by many studies involving multiple stickleback species (Hoogland et al. 1957, Hall 1956,

Reisman and Cade 1967, Reimchen 1980, Reist 1980*a*, Reist 1980*b*, Reist 1983, Blouw and

Hagen 1984b, Foster et al. 1988, Reimchen 1992, Reimchen 1994, Vamosi 2002, Rundle et al.

2003, Vamosi and Schluter 2004, Marchinko 2008, Lescak and von Hippel 2011, Lescak et al.

2012, Miller et al. 2017). These studies typically take advantage of the existence of variation

among individuals within a population (or within sympatric or parapatric populations) to

determine which pelvic phenotype (e.g. spined or unspined) is favoured by a given predator. For

example, several studies have shown that gape-limited predators, such as trout, select for

increased spine length or against spine loss, whereas predators that are not gape-limited, such as

pike or aquatic insects, select for reduced spines or spine loss (Miller et al. 2017). Given that

multiple predators occur in an environment (e.g. a lake may contain insects, fish, and birds that

prey on stickleback), and that predators are not uniformly distributed within an environment

(Reimchen 1994), spine polymorphism within stickleback populations is likely maintained by a

balance of selection driven by multiple predators across multiple habitats or multiple points in

time.

The ecological correlates of spine polymorphism have been well-studied and

characterized in threespine stickleback (Rennison et al. 2019a, Rennison et al. 2019b). This

includes differences in predation between habitats, different habitats types, and different food

resources which presents the opportunity for additional adaptive or plastic responses. A noteable

example of pelvic spine divergence in some lakes is known to be associated with morphological

and ecological divergence between reproductively isolated spined limnetic and unspined benthic

threespine stickleback (Schluter and McPhail 1992, Hatfield and Schluter 1999, Schluter 1993,

Hatfield 1997, Boughman 2001, Peichel et al. 2001, Rundle and Schluter 2004). Even without

reproductive isolation, a well-established change associated with spine reduction in threespine

stickleback is a shift in diet away from planktonic food sources and towards more benthic food

items, which is associated with a change in gill raker morphology towards fewer, shorter, and

thicker gill rakers in benthic feeding fish (Reimchen 1980, McPhail 1984, Ridgeway and

McPhail 1984, Schluter 1993, Nagel and Schluter 1998, Rundle et al. 2003, Matthews et al.

2010). It is unclear, however, whether the reduction in spines that has occurred in other

stickleback species is associated with parallel ecological and morphological changes, such as

parallel changes in diet or gill raker morphology.

In brook stickleback (*Culaea inconstans* Kirtland, 1840), within-population pelvic spine

polymorphism does not involve reproductively isolated morphs (Lowey et al. 2020) and the

ecological and morphological changes associated with pelvic spine reduction are not necessarily

consistent among populations (Willerth et al. 2022). To date, however, no study has directly

addressed whether pelvic spine reduction is associated with a shift in diet in brook stickleback in

a parallel manner to the shift in diet in pelvic-reduced threespine stickleback. Hence, the extent

of ecological or phenotypic parallelism among stickleback species associated with the reduction

or loss of pelvic spines is unknown. Resolving this question about the extent of parallelism

among stickleback species will help address more general questions about the extent of

phenotypic parallelism across taxa (Bolnick et al. 2018, De Lisle and Bolnick 2020).

In this study, we sought to understand whether the outcome of balancing selection based

on similar selective forces (in this case, predation) in different species results in parallel or non-

parallel ecological and morphological changes. We addressed the hypothesis that pelvic spine

polymorphism is associated with a shift in diet in brook stickleback in a parallel manner to

threespine stickleback – with pelvic-reduced fish eating a more benthic diet. To address this

hypothesis, we used metabarcoding of gut contents to compare diet among spined and pelvic-

reduced brook stickleback in two polymorphic populations. Because a change in gill raker

morphology has been strongly implicated in diet shifts in spined and pelvic-reduced threespine

stickleback, we also investigated gill raker morphology associated with pelvic reduction.

Methods

Sample collection and study site

Brook stickleback were collected in Alberta, Canada, from Muir Lake (UTF-8 encoded

WGS84 latitude and longitude: 53.627659, -113.957524) and Shunda Lake (52.453899, -

116.146192) in the summer of 2017 and 2019 using unbaited minnow traps. To sample fish from

the littoral zone, where they were more likely to have a benthic diet, traps were set within

throwing distance from the shore (maximum 4m to 5m from shore) at a depth of 0.5m to 2m. To

sample fish from the limnetic zone, where they were more likely to have a planktonic diet, traps

were set at least 50m from shore and suspended from floats at a depth of 1-2m. All traps were

retrieved within one to twelve hours after being set. In each year, all brook stickleback were

retained until we had collected 40 (in 2017) or 30 (in 2019) spined individuals. Then, once we

had captured 30 or 40 spined fish, we retained only pelvic-reduced individuals until we had

achieved a balanced sample of spined and pelvic-reduced morphotypes (Table 1). Pelvic

morphology was assessed at the time of capture via close inspection of the pelvic region of each

fish with forceps, and sex was identified at the time of capture based on the presence of male

nuptial colouration or the presence of eggs. Each fish included in this study was euthanized by

immersion in an overdose mixture of lake water and eugenol. In 2019, the entire gut of each fish

was removed and preserved in 95% ethanol. The body of each fish was preserved in 70%

ethanol. As described in Willerth et al. (2022), we stained each fish with Alizarin red to observe

calcium-containing osteocytes more clearly (such as in pelvic bones, spines, branchial arches,

and gill rakers) and we confirmed pelvic morphology in the lab using magnified digital images.

The fish collected from Muir Lake and Shunda Lake were used in previous comparisons

of stable isotope signatures and morphology between spined and unspined brook stickleback

(Willerth et al. 2022). These lakes are well-suited for studies of brook stickleback pelvic spine

polymorphism because they contain relatively high abundances of spined and pelvic-reduced

morphs (Nelson 1977, Lowey et al. 2020). The pelvic-reduced morphs include unspined

individuals that completely lack spines or any vestige of a pelvic girdle, but also individuals with

vestigial (a.k.a. intermediate) pelvic structures that lack some pelvic element (e.g. one spine

missing, half the pelvic girdle missing, etc.; see Klapaker et al. 2013 for a detailed description of

variation in pelvic morphology across stickleback species). Recent surveys and stocking records

(Alberta Environment and Parks 2021) suggest that salmonids, such as brown trout (*Salmo trutta*

Linnaeus 1758) and rainbow trout [*Ocorhychus mykiss* (Walbaum 1792)], are the dominant

predators in both lakes, although we also observed other predators, such as loons [*Gavia immer*

(Brunnich 1764)], dragonfly nymphs (Gomphidae and other unidentified families), giant water

bugs [*Lethocerus americanus* (Leidy 1847)], and backswimmers (Notonectidae) in both lakes

(Willerth et al. 2022). Both lakes contain abundant Daphniid zooplankton, and littoral

macroinvertebrate sampling revealed Trichoptera larvae, amphipods, leeches, and Chironomidae

larvae (Willerth et al. 2022). Sampling permits were issued by the Government of Alberta, and

fish handling protocols were approved by the Animal Care Committee at Mount Royal

University (Animal Care Protocol ID 101029 and 101795).

Diet analysis

Fish guts collected from 95 fish in 2019 (Table 1) were shipped to the Canadian Centre

for DNA Barcoding at the University of Guelph for membrane-based DNA extraction and

amplicon sequencing (Ivanova et al. 2006). This metabarcoding approached used five PCR

primer sets designed to amplify a barcode region of the cytochrome c oxidase subunit I (COI)

gene in arthropods, mollusks, annelids, amphipods, and microalgae (details of COI amplification

can be found at http://ccdb.ca/site/wp-content/uploads/2016/09/CCDB_Amplification.pdf). A

second round of PCR amplification added Ion Torrent sequencing adapters and a unique

multiplex identifier (MID) sequence to the 5' end of amplicons for each sample. Amplified COI

fragments were then pooled and single-end sequenced on an Ion Torrent PGM sequencing

platform (Thermo Fisher, Waltham, MA, USA). The resulting sequence data was automatically

de-multiplexed by the PGM Torrent Browser (Thermo Fisher, Waltham, MA, USA). We

removed primer and adapter sequences from the de-multiplexed reads, and discarded reads that

consisted of only primer or adapter sequence using CUTADAPT v2.3 (Martin 2011). We used

DADA2 (Callahan et al. 2016) in R v4.2.2 (R Core Team 2022) to trim and filter reads, estimate

error rates for the filtered and trimmed amplicon dataset, combine identical reads into unique

sequences, and infer biologically meaningful (as opposed to spurious) amplified sequence

variants (ASVs). In the filtering and trimming step, we removed reads containing any unknown

basepairs, truncated reads to 200 bp, discarded reads less than 200 bp, truncated reads at the first

instance of a quality score less than 2, discarded reads that match the phiX genome, and

discarded reads with greater than 2 "expected errors" after truncation (where the number of

"expected errors" is calculated from the per-basepair quality scores for each read). In the

inference of ASVs, because Ion Torrent sequencing tends to make homopolymer errors and

produces higher numbers of indels (relative to Illumina sequencing), we applied a homopolymer

gap penalty of -1 and applied a more restrictive threshold for the number of insertions in one

sequence relative to another (BAND_SIZE = 32). We then created an ASV abundance table

(with the abundance of each ASV for each fish) and removed chimeric sequences using the

"consensus" method. We exported the list of ASVs across all samples to a fasta file and used the

JAMP pipeline in BOLDigger (Buchner & Leese 2020) to identify the best-fitting taxonomic

identification for each ASV.

We used PHYLOSEQ (McMurdie et al. 2013) to create a data object in R that allowed us

to associate the abundance of each taxon identified in each fish's gut with other sample data (i.e.

lake, sex, size, and pelvic phenotype). We removed any taxa identified as brook stickleback or

bacteria (the latter being more indicative of gut microbiome than diet). Because many taxa were

not identified to genus or species, we agglomerated taxa at the rank of family, while also

removing taxa that were not identified to the level of family, using the tax_glom function in

phyloseq. We analyzed individual-level variation in gut contents by performing an ordination

using non-metric multidimensional scaling (NMDS) and Bray-Curtis distances based on family-

level taxonomic assignment.

To test the hypothesis that brook stickleback pelvic spine reduction is associated with a

shift in diet, we used DESeq2 (Love et al. 2014) to conduct Wald tests of the Log_2 fold-

differences in the abundances of each eukaryote family in the diets of brook stickleback between

lakes and between pelvic phenotypes (with p-values corrected for multiple testing with the

Benjamin Hochberg method). Because we had previously found a significant difference in length

between fish collected in near-shore "littoral" traps and fish collected in "limnetic" traps

suspended >50m from shore (Willerth et al. 2022), which would lead to collinearity in statistical

models investigating the effects of fish length and trap location on diet, we looked for

correlations among predictor variables using generalised linear models with a Gaussian error

distribution (for length – a continuous numerical variable) or a binomial error distribution (for

factor variables – pelvic phenotype, sex, and trap location). Given the results of these tests

(details below in the results section) showing significant correlation between length and habitat

(but between no other predictor variables), and given the lack of any fish caught in our

"limnetic" traps in Shunda Lake, we interpreted length as a proxy for habitat, and removed

habitat (or "trap location") as a variable in subsequent analyses. In the comparison between

lakes, we specified a model that included lake, pelvic morphology, sex, and scaled total length

(i.e. scaled to mean = 0 and variance = 1). Because we expect diet might be affected by allometry

(Aguirre et al. 2008, Reimchen et al. 2016), we also included interactions with length (and all

other two way interactions) in our model. We subsequently conducted separate analyses for each

lake, wherein we specified models (one per lake) that included pelvic phenotype (spined or

pelvic-reduced), sex, scaled total length, and two-way interactions.

Gill rakers

Fish that had been bleached and stained with alizarin red for morphological analyses

(Willerth et al. 2022) were used to investigate variation in gill raker morphology associated with

different pelvic phenotypes. We removed the first branchial arches from both sides of each fish

with forceps and scissors, cleared away all the gill filaments, and photographed the gill rakers in

a plastic weighing boat using a Nikon SMZ-75T Greenough-type stereo microscope with a

mounted Imaging Source camera and NIS-Elements D Software (©2021 Nikon Corporation).

Following Kaeuffer et al. (2011), we counted the number of gill rakers on the left and right gill

arches, and we measured the length of the second through fourth gill raker from the epibranchial-

ceratobranchial joint on the ceratobranchial, which were consistently the three longest gill rakers

on each arch. We used generalized linear models in R with a gaussian error distribution to model

the effects of fish length, sex, year of capture, lake, and pelvic phenotype on variation in gill

raker length or number. As described above, we used length as a proxy for habitat to avoid

collinearity between the length and habitat variables. We set contrasts among factors using the

contr.sum function, and we used the CAR package (Fox and Weisberg 2019) to generate an

ANOVA table with type III sums of squares. We included two-way interactions in our initial

models, and then used a reverse-stepwise approach for model selection, removing any interaction

terms that were not significant ($p > 0.05$).

Results

A list of all taxa identified via metabarcoding in Muir Lake and Shunda Lake is available

in the supplementary material (Table S1 and Table S2). At the population level, brook

stickleback in Muir Lake and Shunda Lake had taxonomically diverse diets. We identified

thirteen Eukaryote phyla among the Amoebozoa, Archaeplastida, Chromalveolata, Fungi, and

Metazoa. The Metazoa was by far the most diverse taxon in both lakes where we identified six

different phyla, the most abundant and diverse of which were the arthropods. We identified six

arthropod classes: Arachnida, Branchiopoda, Copepoda, Insecta, Malacostraca, and Ostracoda.

Insecta was by far the most abundant and diverse class (among all phyla) in the diets of brook

stickleback in both lakes, with six orders and sixteen families. We focused on family-level

classifications because many taxa were not resolved to genus or species. Large individual-level

differences in diet among brook stickleback obscured any obvious group-level differences in diet

between lakes or between pelvic morphologies in an ordination plot (Figure 1). Prior to

statistically evaluating differences in diet between lakes and between pelvic phenotypes, we

evaluated correlations among predictor variables. Using a type II sums of squares ANOVA (i.e.

evaluating a main effect conditional on all other main effects without significant interaction

effects), we found a significant effect of trap location (limnetic or littoral) on fish length in Muir

Lake (df = 1, Wald's X^2 = 4.7997, p = 0.028), and a significant effect of fish length on trap

location in Muir Lake (df = 1, Wald's X^2 = 4.8788, p = 0.027). No other predictor variables were

correlated. Wald tests of the Log_2 fold-differences in the abundances of each eukaryote family in

the diets of brook stickleback between lakes suggested that there were significant differences in

the diets of brook stickleback in Muir Lake compared to Shunda Lake attributable to several taxa

(Figure 2).

To address the hypothesis that brook stickleback pelvic spine reduction (or loss) is

associated with a shift in diet, we evaluated the differential abundance of each taxon identified in

the stomachs of spined and pelvic-reduced brook stickleback in Muir Lake and Shunda Lake. In

Muir Lake, Succineidae (amber snails) were significantly more abundant in the diets of spined

fish (Figure 3). In Shunda Lake, Simuliidae (black flies) were significantly more abundant in the

diets of spined fish, whereas Baetidae (small "minnow" mayflies) and Pythiaceae (a family of

parasitic Oomycete water moulds) were significantly more abundant in the diets of pelvic-

reduced fish (Figure 3). Diet also differed significantly based on fish length, with larger fish

eating more Scirtidae (marsh beetles) in Muir Lake. In Shunda lake, larger fish ate more

Hyalellidae (amphipods) and Hydridae (cnidarian hydras) but less Baetidae, Pythiaceae,

Oecophoridae (a family of moths), Caenidae (squaregill mayflies), Sellaphoraceae (diatoms), and

Saprolegniaceae (a family of Oomycete freshwater moulds). There were no significant

differences in diet associated with sex in Muir Lake, whereas females ate more Pythiaceae in

Shunda Lake.

275   To investigate a potential change in gill raker morphology between spined and unspined

276 brook stickleback associated with a shift in diet, we compared gill raker morphology between

277 individuals with spined and reduced pelvic phenotypes in Muir Lake and Shunda Lake. We

278 conducted separate analyses where we looked for an effect on maximum gill raker length (i.e. the

279 length of the longest gill raker in each individual), mean gill raker length, the length of a specific

280 gill raker (e.g. the second gill raker from the epibranchial-ceratobranchial joint on the

281 ceratobranchial), the number of gill rakers on the left or right gill arch, or the mean number of

282 gill rakers on both arches. We also conducted analyses involving the effect of pelvic morphology

283 with three categories (i.e. spined, vestigial, and unspined) instead of two categories (i.e. spined

284 and pelvic-reduced). Whereas the effects of some factors varied in magnitude among these

285 analyses, no analysis revealed a significant effect of pelvic morphology on gill raker size or

286 number. For clarity, we report only the details of our analysis involving the effect of pelvic

287 reduction (with two categories of pelvic morphology) on maximum gill raker length (Table 2,

288 Figure 4) and on the mean number of gill rakers (Table 3, Figure 5). Gill raker length and

289 number did not differ significantly between spined and pelvic-reduced brook stickleback. Gill

290 raker length and number were significantly affected by fish size, with larger fish having more

291 and longer gill rakers. Brook stickleback in Muir Lake had more gill rakers and longer gill rakers

292 than those in Shunda Lake. Male brook stickleback had significantly longer gill rakers than

293 female brook stickleback, especially at larger sizes. The effect of fish size on gill raker length

294 was weaker for samples collected in 2019 than for samples collected in 2017 (see Tables 2 and 3

295 for details of statistical results).

297 **Discussion**

The pelvic spine polymorphism in brook stickleback populations is likely maintained by

a balance of selective forces from multiple predators. These predators have different habitat

preferences, and an analysis of stable isotope signatures revealed that spined and pelvic reduced

brook stickleback are associated with different habitat types (Willerth et al. 2022), suggesting

that predation may drive this pelvic spine polymorphism and therefore has ecological

consequences. In this study, we tested the hypothesis that spined and pelvic-reduced brook

stickleback have different diets. Consistent with our hypothesis, we found significant differences

in the abundance of some taxa in the diets of spined and pelvic-reduced brook stickleback.

Pelvic spine reduction in brook stickleback and threespine stickleback populations has

evolved in parallel based on similar selective forces. The extent to which we should expect

parallelism to extend beyond a particular phenotype of interest (in this case, pelvic spine

reduction), resulting in parallel ecological, behavioural, and morphological consequences, is

unknown. If parallelism extends beyond pelvic spine reduction in these stickleback species, we

expect that, like threespine stickleback, pelvic spine reduction in brook stickleback should be

associated with a more benthic diet.

Our observations were contrary to this expectation, which implies non-parallelism in the

ecological consequences of this adaptive phenotype. In Muir Lake, spined brook stickleback ate

more amber snails, which are land snails associated with lake shores (Lanoo and Bovbjerg 1985),

suggesting that spined, and not pelvic-reduced, brook stickleback have a more benthic diet in

Muir Lake. In Shunda Lake, spined fish ate more blackflies and less Baetid mayflies, which were

likely ingested as the aquatic larval phase of these insects (as opposed to the flying adults).

Blackfly larvae often form a large component of the benthic biomass in well-oxygenated flowing

waters, such as streams or waterfalls (Alder et al. 2004, Malmqvist et al. 2004). Baetid larvae are

typically good swimmers and may be found in moving or still water, often associated with

submerged vegetation (Ide 1931, Harper and Harper 1981, Giberson et al. 2007). Whereas we

did not collect brook stickleback from within the outlet stream of Shunda Lake (there is no

visible above-ground inlet stream), we did collect all our samples within 800m to the South and

West of the small dam at the mouth of the outlet. Brook stickleback have a high propensity for

dispersal (Nelson and Paetz 1992), suggesting that all the fish among our samples had the

opportunity to forage adjacent to the outlet stream of Shunda Lake where there is moving water

flowing out over the dam. Unfortunately, our sampling records are too coarse to allow an

assessment of the proportions of spined and pelvic-reduced fish as a function of proximity to the

outlet stream in Shunda Lake. Nonetheless, our diet results from Shunda Lake suggest that

spined fish tended to forage more in the benthos, likely adjacent to the outlet stream, and pelvic-

reduced fish tended to forage on free-swimming littoral prey. The finding that the parasitic fungi-

like oomycete, Pythiaceae, was at significantly higher abundance in the diets of pelvic-reduced

fish in Shunda Lake is difficult to interpret because the parasite is unlikely to have been a

standalone food item. Pythiaceae, known as water moulds, are parasites of plants and animals,

and may have been found in the guts of mayflies (or other herbivorous prey), or may have been

ingested as part of some other diet item, such as rotting vegetation. It is worthwhile to note that,

whereas there were no significant differences in the abundances of planktonic taxa (such as

Daphniids) between spined and pelvic-reduced fish, pelvic-reduced fish tended to eat more

Daphniids than spined fish in both lakes (Figure 3), which is consistent with the pelvic-reduced

fish having a less benthic-oriented diet (i.e. contrary to our original hypothesis).

Differences in diet associated with pelvic spine reduction in threespine stickleback are

frequently accompanied by differences in gill raker morphology (McPhail 1984, Ridgeway and

McPhail 1984, Schluter 1993, Nagel and Schluter 1998, Rundle et al. 2003, Matthews et al.

2010). Longer, more slender, more numerous gill rakers are better for planktonic feeding,

whereas shorter, thicker, less numerous gill rakers are better for benthic feeding. In brook

stickleback, we did not observe a significant difference in gill raker morphology between spined

and pelvic-reduced fish. There are several possible reasons why the differences in diet between

spined and pelvic-reduced brook stickleback are not associated with differences in gill raker

morphology. First, aside from the non-significant shift to feeding on more Daphniids in pelvic-

reduced fish (as described in the previous paragraph), the diet differences associated with pelvic

reduction in brook stickleback do not necessarily involve a shift to planktonic feeding. Instead,

the shift in diet may reflect a shift from benthic feeding in spined fish (e.g. blackfly larvae or

snails) to littoral open-water (or, at least, non-benthic) feeding in pelvic-reduced fish (e.g. small

minnow mayfly larvae). Second, spined and pelvic-reduced brook stickleback are not

reproductively isolated, and each lake likely contains a panmictic population (Lowey et al.

2020). So, in the absence of pleiotropic effects of the spine polymorphism, divergence (or

polymorphism) in other traits that are not directly affected by balancing selection is unlikely to

evolve. Lastly, the diet differences between spined and pelvic-reduced brook stickleback may not

induce any developmental plasticity in gill raker morphology.

It is unclear whether the diet differences between spined and pelvic-reduced brook

stickleback are associated with a difference in habitat. It is possible that different pelvic morphs

use different resources in the same habitat. If this were the case, then maintenance of spine

polymorphism must depend on different predators in the same habitat exerting balancing

selection pressure simultaneously or in a temporally segregated manner. A fruitful avenue of

future research would likely be an investigation of temporally varying predation on brook

stickleback between seasons and across multiple generations.

Observations of parallel evolution have long been interpreted as evidence for the

involvement of natural selection as a mechanism of evolutionary change. Evolutionary biologists

are now trying to understand when, and when not, to expect the evolution of parallel phenotypes

to be associated with parallel ecological consequences (e.g. the same behavioural or habitat

divergence) and parallel biological pathways (e.g. the same genes or the same mutations). In the

case of threespine stickleback pelvic spine reduction, there seems to be parallelism among

populations in the genetic basis of spine reduction, and there appears to be parallelism in the

ecological consequences and correlates of spine reduction (Miller et al. 2015). Our comparison

with instances of pelvic spine reduction in brook stickleback suggests a reduced extent of

parallelism between species. Whether, and why, the extent of parallelism might generally be

lower in between-species versus between-population comparisons, despite very similar selective

scenarios (e.g. based on balancing or divergent selection due to predation in freshwater

environments), will undoubtedly be the focus of future research.

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Data Availability

Private URL for DRYAD archive during peer review (Clicking the link immediately launches a

download of the data files):

https://datadryad.org/stash/share/CJuLVp4HYMFaQMBDsCUnMMhK_jh4j4V-eSz9Qp6T21E

Literature Cited

Aguirre, W. E., K. E. Ellis, M. Kusenda, and M. A. Bell. 2008. Phenotypic variation and sexual

dimorphism in anadromous threespine stickleback: implications for postglacial adaptive

radiation. *Biological Journal of the Linnean Society*, 95:465-478.

Adler, P. H., D. C. Currie, D. M. Wood, R. M. Idema, and L. W. Zettler. 2004. *The black flies

(simuliidae) of North America*. Comstock Pub. Associates, New York City, NY.

Alberta Environment and Parks. (Accessed 2021, June 1). Fish and Wildlife Information

Management System. https://www.alberta.ca/fisheries-and-wildlife-management-

information-system-overview.aspx

Blouw, D. M., and G. J. Boyd. 1992. Inheritance of reduction, loss, and asymmetry of the pelvis

in Pungitius pungitius (ninespine stickleback). *Heredity*, 68(1), 33-42.

Blouw, D. M., and D. W. Hagen. 1984a. The adaptive significance of dorsal spine variation in

the fourspine stickleback, *Apeltes quadracus*, I. Geographic variation in spine number.

Canadian Journal of Zoology, 62(7):1329-1339.

Blouw, D. M., and D. W. Hagen. 1984b. The adaptive significance of dorsal spine variation in

the fourspine stickleback, *Apeltes quadracus*. III. Correlated traits and experimental

evidence on predation. *Heredity*, 53(2):371-382.

Bolnick, D. I. 2004. Can intraspecific competition drive disruptive selection? An experimental

test in natural populations of sticklebacks. *Evolution*, 58(3), 608-618.

Bolnick, D. I., R. D. Barrett, K. B. Oke, D. J. Rennison, and Y. E. Stuart. 2018. (Non) parallel

evolution. *Annual Review of Ecology, Evolution, and Systematics, 49*(1):303-330.

Boughman, J. W. 2001. Divergent sexual selection enhances reproductive isolation in

sticklebacks. *Nature*, 411(6840):944-948.

Buchner, D., and F. Leese. 2020. BOLDigger–a Python package to identify and organise

sequences with the Barcode of Life Data systems. *Metabarcoding and Metagenomics*,

4:e53535.

Callahan B. J., P. J. McMurdie, M. J. Rosen, A. W. Han, A. J. A. Johnson, and S. P. Holmes

(2016). DADA2: High-resolution sample inference from Illumina amplicon data. *Nature*

Methods, 13:581-583.

Chan, Y. F., M. E. Marks, F. C. Jones, G. Villarreal Jr, M. D. Shapiro, S. D. Brady, A. M.

Southwick, D. M. Absher, J. Grimwood, J. Schmutz, R. M. Myers, D. Petrov, B. Jónsson,

D. Schluter, M. A. Bell, and D. M. Kingsley. 2010. Adaptive evolution of pelvic

reduction in sticklebacks by recurrent deletion of a Pitx1 enhancer. *Science*,

327(5963):302–305.

Colosimo, P. F., K. E. Hosemann, S. Balabhadra, G. Villarreal Jr, M. Dickson, J. Grimwood, J.

Schmutz, R. M. Myers, D. Schluter, and D. M. Kingsley. 2005. Widespread parallel

evolution in sticklebacks by repeated fixation of ectodysplasin alleles. *Science*,

307(5717):1928-1933.

De Lisle, S. P., and D. I. Bolnick. 2020. A multivariate view of parallel evolution. *Evolution*,

74(7):1466-1481.

Dobzhansky, T. 1971. *Genetics of the Evolutionary Process*. Columbia University Press, New

York, NY.

Ford, E. B. 1945. Polymorphism. *Biological Reviews* 20:73–88.

Foster, S.A., V.B. Garcia, and M.Y. Town. 1988. Cannibalism as the cause of an ontogenetic

shift in habitat use by fry of the threespine stickleback. *Oecologia*, 74(4):577-585.

Fox, J., and S. Weisberg. 2019. An {R} Companion to Applied Regression, Third Edition.

Thousand Oaks CA: Sage. URL:

https://socialsciences.mcmaster.ca/jfox/Books/Companion/

Giberson, D. J., S. K. Burian, amd M. Shouldice. 2007. Life history of the northern mayfly

Baetis bundyae in Rankin Inlet, Nunavut, Canada, with updates to the list of mayflies of

Nunavut. *The Canadian Entomologist*, 139(5):628-642.

Gigord, L. D., M. R. Macnair, and A. Smithson. 2001. Negative frequency-dependent selection

maintains a dramatic flower color polymorphism in the rewardless orchid *Dactylorhiza*

sambucina (L.) Soò. *Proceedings of the National Academy of Sciences of the United*

States of America, 98(11):6253-6255.

Hagen, D. W., and D. M. Blouw. 1983. Heritability of dorsal spines in the fourspine stickleback

(*Apeltes quadracus*). *Heredity*, 50(3):275-281.

Hall, M. F. 1956. A comparative study of the reproductive behaviour of the sticklebacks

(Gasterosteidae). Ph.D. Thesis. St. Hugh's College, Oxford. 370 p.

Harper, F., and P. P. Harper. 1981. Northern Canadian mayflies (Insecta: Ephemeroptera),

records and descriptions. *Canadian Journal of Zoology*, 59:1784–1789.

Hatfield, T. 1997. Genetic divergence in adaptive characters between sympatric species of

stickleback. *The American Naturalist*, 149(6):1009-1029.

Hatfield, T., and D. Schluter. 1999. Ecological speciation in sticklebacks:

environment☐dependent hybrid fitness. *Evolution*, *53*(3):866-873.

458 Hendry, A. P., C. L. Peichel, B. Matthews, J. W. Boughman, and P. Nosil. 2013. Stickleback

459   research: the now and the next. *Evolutionary Ecology Research*, *15*(2):111-141.

460 Hoogland, R., D. Morris, and N. Tinbergen. 1957. The spines of sticklebacks (*Gasterosteus* and

461   *Pygosteus*) as means of defence against predators (*Perca* and *Esox*). *Behaviour*, 10:205-

462   236.

463 Ide, F. P. 1937. Descriptions of eastern North American species of baetine mayflies with

464   particular reference to nymphal stages. *The Canadian Entomologist*, 69:219–231.

465 Ivanova N. V., J. R. deWaard, and P. D. N. Hebert. 2006. An inexpensive, automation-friendly

466   protocol for recovering high-quality DNA. *Molecular Ecology Notes*, 6:998-1002.

467 Jeffries, D. L., J. A. Mee, and C. L. Peichel. 2022. Identification of a candidate sex determination

468   gene in Culaea inconstans suggests convergent recruitment of an Amh duplicate in two

469   lineages of stickleback. *Journal of Evolutionary Biology*, 35(12):1683-1695.

470 Kaeuffer, R., C. L. Peichel, D. I. Bolnick, and A. P. Hendry. 2012. Parallel and nonparallel

471   aspects of ecological, phenotypic, and genetic divergence across replicate population

472   pairs of lake and stream stickleback. *Evolution*, *66*(2):402-418.

473 Klepaker, T., K. Østbye, and M. A. Bell. 2013. Regressive evolution of the pelvic complex in

474   stickleback fishes: a study of convergent evolution. *Evolutionary Ecology Research,*

475   *15*(4):413-435.

476 Lande, R. 1975. The maintenance of genetic variability by mutation in a polygenic character

477   with linked loci. *Genetics Research*, *26*(3):221-235.

478 Lannoo, M. J., and R. V. Bovbjerg. 1985. Distribution, dispersion, and behavioral ecology of the

479   land snail *Oxyloma retusa* (Succineidae). *Proceedings of the Iowa Academy of Science*,

480   92(2):67-69.

Lescak, E. A., and F. A. von Hippel. 2011. Selective predation of threespine stickleback by

rainbow trout. *Ecology of Freshwater Fish*, 20(2):308-314.

Lescak, E. A., F. A. von Hippel, B. K. Lohman, and M. L. Sherbick. 2012. Predation of

threespines stickleback by dragonfly naiads. *Ecology of Freshwater Fish*, 21(4):581-587.

Love, M. I., W. Huber, and S. Anders. 2014. Moderated estimation of fold change and dispersion

for RNA-seq data with DESeq2. *Genome Biology*, 15(12):550.

Lowey, J., Q. Cheng, S. M. Rogers, and J. A. Mee. 2020. Persistence of pelvic spine

polymorphism in a panmictic population of brook stickleback (*Culaea inconstans*) in

Alberta, Canada. *Canadian Journal of Zoology*, 98(10):643-649.

Malmqvist, B., P. H. Adler, K. Kuusela, R. W. Merritt, and R. S. Wotton. 2004. Black flies in the

boreal biome, key organisms in both terrestrial and aquatic environments: a review.

Ecoscience, 11(2):187-200.

Marchinko, K. B. 2008. Predation's role in repeated phenotypic and genetic divergence of armor

in threespine stickleback. *Evolution*, 63(1):127-138.

Martin, M. 2011. Cutadapt removes adapter sequences from high-throughput sequencing reads.

EMBnet.Journal, 17(1):10-12.

Matthews, B., K. B. Marchinko, D. I. Bolnick, and A. Mazumder. 2010. Specialization of trophic

position and habitat use by sticklebacks in an adaptive radiation. *Ecology*, *91*(4):1025-

1034.

McMurdie, P. J., and S. Holmes. 2013. phyloseq: an R package for reproducible interactive

analysis and graphics of microbiome census data. *PloS One*, *8*(4):e61217.

McPhail, J. D. 1984. Ecology and evolution of sympatric sticklebacks (Gasterosteus):

morphological and genetic evidence for a species pair in Enos Lake, British Columbia.

Canadian Journal of Zoology, *62*(7):1402-1408.

Miller, S. E., D. Metcalf, and D. Schluter. 2015. Intraguild predation leads to genetically based

character shifts in the threespine stickleback. *Evolution*, *69*(12):3194-3203.

Miller, S.E., M. Barrueto, and D. Schluter. 2017. A comparative analysis of experimental

selection on the stickleback pelvis. *Journal of Evolutionary Biology*, 30(6):1165-1176.

Nagel, L., and D. Schluter. 1998. Body size, natural selection, and speciation in sticklebacks.

Evolution, *52*(1):209-218.

Nelson, J. S. 1969. Geographic variation in the brook stickleback, *Culaea inconstans*, and notes

on nomenclature and distribution. *Journal of the Fisheries Board of Canada*, 26(9):2431-

2447.

Nelson, J. S. 1977. Evidence of a genetic basis for absence of the pelvic skeleton in brook

stickleback, *Culaea inconstans*, and notes on the geographical distribution and origin of

the loss. *Journal of the Fisheries Board of Canada*, 34(9):1314-1320.

Nelson, J. S., and M. J. Paetz. 1992. *The fishes of Alberta* (pp. 319–323). The University of

Alberta Press, Edmonton, AB, Canada.

Peichel, C. L., K. S. Nereng, K. A. Ohgi, B. L. Cole, P. F. Colosimo, C. A. Buerkle, D. Schluter,

and D. M. Kingsley. 2001. The genetic architecture of divergence between threespine

stickleback species. *Nature*, *414*(6866):901-905.

Punzalan, D., F.H. Rodd, and K. A. Hughes. 2005. Perceptual processes and the maintenance of

polymorphism through frequency-dependent predation. *Evolutionary Ecology*, 19(3):303-

320.

R Core Team. 2022. R: A language and environment for statistical computing. R Foundation for
Statistical Computing, Vienna, Austria. URL https://www.R-project.org/.

Rainey, P. B., A. Buckling, R. Kassen, and M. Travisano. 2000. The emergence and maintenance
of diversity: insights from experimental bacterial populations. *Trends in Ecology and*
Evolution, 15(6):243-247.

Reid, K., M. A. Bell, and K. R. Veeramah. 2021. Threespine stickleback: a model system for
evolutionary genomics. *Annual Review of Genomics and Human Genetics*, *22*:357-383.

Reimchen, T. E. 1980. Spine deficiency and polymorphism in a population of *Gasterosteus*
aculeatus: an adaptation to predators? *Canadian Journal of Zoology*, 58(7): 1232-1244.

Reimchen. T. E. 1992. Injuries on stickleback from attacks by a toothed predator
(*Onchorhynchus*) and implications for the evolution of lateral plates. *Evolution*,
46(4):1224-1230.

Reimchen, T. E. 1994. Predators and morphological evolution in threespine stickleback. *in* The
evolutionary biology of the threespine stickleback. *Edited by* Bell, M.A., and S.A. Foster.
Oxford University Press, New York, NY.

Reisman, H. M., and T. J. Cade. 1967. Physiological and behavioral aspects of reproduction in
the brook stickleback, *Culaea inconstans*. *American Midland Naturalist*, 77:275-295.

Reimchen, T. E., D. Steeves, and C. A. Bergstrom. 2016. Sex matters for defence and trophic
traits of threespine stickleback. *Evolutionary Ecology Research*, 17(4):459-485.

Reist, J. D. 1980a. Selective predation upon pelvic phenotypes of brook stickleback, *Culaea*
inconstans, by northern pike, *Esox lucius*. *Canadian Journal of Zoology*, 58(7):1245-
1252.

Reist, J. D. 1980b. Predation upon pelvic phenotypes of brook stickleback, *Culaea inconstans*,

by selected invertebrates. *Canadian Journal of Zoology*, 58(7):1253-1258.

Reist, J. D. 1983. Behavioural variation in pelvic phenotypes of brook stickleback, *Culaea*

inconstans, in response to predation by northern pike, *Esox lucius*. *Environmental*

Biology of Fishes, 8(3-4):255-267.

Rennison, D. J., S. M. Rudman, and D. Schluter. 2019a. Parallel changes in gut microbiome

composition and function during colonization, local adaptation and ecological speciation.

Proceedings of the Royal Society B, 286(1916):20191911.

Rennison, D. J., Y. E. Stuart, D. I. Bolnick, and C. L. Peichel. 2019b. Ecological factors and

morphological traits are associated with repeated genomic differentiation between lake

and stream stickleback. *Philosophical Transactions of the Royal Society*

B, *374*(1777):20180241.

Ridgway, M. S., and J. D. McPhail. 1984. Ecology and evolution of sympatric sticklebacks

(*Gasterosteus*): mate choice and reproductive isolation in the Enos Lake species pair.

Canadian Journal of Zoology, *62*(9):1813-1818.

Rundle, H. D., S. M. Vamosi, and D. Schluter. 2003. Experimental test of predation's effect on

divergent selection during character displacement in sticklebacks. *Proceedings of the*

National Academy of Sciences, 100(25):14943-14948.

Rundle, H. D., and D. Schluter. 2004. Natural selection and ecological speciation in sticklebacks.

Adaptive Speciation, *19*(3):192-209.

Schluter, D. 1993. Adaptive radiation in sticklebacks: size, shape, and habitat use efficiency.

Ecology, 74(3):699-709.

Schluter, D. 2000. *The Ecology of Adaptive Radiation*. Oxford University Press, Oxford.

Schluter, D., and J. D. McPhail. 1992. Ecological character displacement and speciation in

sticklebacks. *The American Naturalist*, 140(1):85-108.

Schluter, D., K. B. Marchinko, M. E. Arnegard, H. Zhang, S. D. Brady, F. C. Jones, M. A. Bell,

and D. M. Kingsley. 2021. Fitness maps to a large-effect locus in introduced stickleback

populations. *Proceedings of the National Academy of Sciences*, 118(3):e1914889118.

Shapiro, M. D., B. R. Summers, S. Balabhadra, J. T. Aldenhoven, A. L. Miller, C. B.

Cunningham, M. A. Bell, and D. M. Kingsley. 2009. The genetic architecture of skeletal

convergence and sex determination in ninespine sticklebacks. *Current Biology*,

19(13):1140-1145.

Shikano, T., V. N. Laine, G. Herczeg, J. Vilkki, and J. Merilä. 2013. Genetic architecture of

parallel pelvic reduction in ninespine sticklebacks. *G3: Genes, Genomes, Genetics*,

3(10):1833-1842.

Takahata, N., Y. Satta, and J. Klein. 1992. Polymorphism and balancing selection at major

histocompatibility complex loci. *Genetics* 130(4):925-938.

Vamosi, S. M. 2002. Predation sharpens the adaptive peaks: survival trade-offs in sympatric

sticklebacks. *Annales Zoologici Fennici*, 39(3):237-248.

Vamosi, S. M., and D. Schluter. 2004. Character shifts in the defensive armor of sympatric

sticklebacks. *Evolution*, 58(2):376-385.

Varadharajan, S., P. Rastas, A. Löytynoja, M. Matschiner, F. C. F. Calboli, B. Guo, A. J.

Nederbragt, K. S. Jakobsen, and J. Merilä. 2019. A high-quality assembly of the nine-

spined stickleback (*Pungitius pungitius*) genome. *Genome Biology and Evolution*,

11:3291–3308.

Willerth, K., E. Franks, and J. A. Mee. 2022. Parallel and non-parallel divergence within

polymorphic populations of brook stickleback, *Culaea inconstans* (Actinopterygii:

Gasterosteidae). *Biological Journal of the Linnean Society*, 135(1):159-173.

Table 1. Summary of samples analyzed in this study. Numbers in parentheses indicate the

samples used in the gut metabarcoding analysis.

	Pelvic Phenotype		
	Spined	Vestigial	Unspined
2017			
Muir Lake			
female	19	1	25
male	22	1	18
Shunda Lake			
female	20	22	6
male	25	8	3
2019			
Muir Lake			
female	3 (3)	0	7 (7)
male	25 (18)	1	27 (18)
Shunda Lake			
female	13 (13)	12 (10)	3 (3)
male	17 (12)	10 (9)	2 (2)

600 **Table 2.** ANOVA table for the generalized linear model of the effects of pelvic reduction (spined

601 or pelvic-reduced), lake (Muir Lake or Shunda Lake), sex, sampling year (2017 or 2018), and

602 body length (total length in mm) on maximum gill raker length in brook stickleback. Two-way

603 interaction terms that were not significant were not included in the final model shown here.

	Sum of Squares	df	F	p-value
Main effects:				
Pelvic reduction	5109	1	0.9238	0.337
Lake	112007	1	20.2544	< 0.001
Sex	84370	1	15.2569	< 0.001
Year	16909	1	3.0578	0.081
Body length	417938	1	75.5767	< 0.001
Interaction effects:				
Lake x body length	55977	1	10.1224	0.002
Sex x body length	109862	1	19.8665	< 0.001
Year x body length	28198	1	5.099	0.025
Residual	1548394	280		
Total	2378764			

606   **Table3.** ANOVA table for the generalized linear model of the effects of pelvic reduction (spined

607   or pelvic-reduced), lake (Muir Lake or Shunda Lake), sex, sampling year (2017 or 2018), and

608   body length (total length in mm) on mean number of gill rakers in brook stickleback. Interaction

609   terms were not significant and were not included in the final model shown here.

	Sum of Squares	df	F	p-value
Pelvic reduction	0.64	1	0.5474	0.460
Lake	5.65	1	4.8253	0.029
Sex	1.52	1	1.3004	0.255
Year	2.05	1	1.7528	0.187
Body length	29.06	1	24.8229	< 0.001
Residual	331.28	283		
Total	370.20			

Figure 1. Ordination plot of Non-metric Multidimensional Scaling axes based on sample-wise

615   Bray-Curtis distances in brook stickleback diet (with diet items classified to the level of family).

Figure 2. Log$_2$ fold change between Shunda Lake and Muir Lake in the abundance of eukaryotic

families in the diets of brook stickleback. Zero indicates no difference in abundance between

lakes, positive numbers indicate higher abundance in Shunda Lake, and negative numbers

indicate higher abundance in Muir Lake. Red symbols indicate a significant difference in

abundance with an adjusted p-value < 0.05 corrected for multiple testing. Families are arranged

(left to right on the x-axis) in order of highest to lowest fold change (in an absolute sense)

between lakes.

Figure 3. Log$_2$ fold change in the abundance of eukaryotic families in the diets of brook

stickleback in Muir Lake (top panel) and Shunda Lake (bottom). Points show the change in

abundance between spined and pelvic-reduced individuals, "S" symbols indicate change between

sexes, and "L" symbols indicate change attributed to fish length. A log$_2$ fold change of zero

indicates no difference in abundance, whereas positive values indicate higher abundance in

spined individuals, in males, or in longer individuals. Red symbols indicate a significant

633   difference in abundance with an adjusted p-value < 0.05 corrected for multiple testing. Two-way

634   interaction effects were included in the statistical model but, for clarity, are not included in this

635   visualization. Families are arranged (left to right on the x-axis) in order of highest to lowest fold

636   change (in an absolute sense) between pelvic morphotypes in each lake.

Figure 4. Comparison of maximum gill raker lengths between spined and pelvic-reduced brook

stickleback in Muir Lake and Shunda Lake. The horizontal span of each line indicates the range

of fish lengths for each group in the comparison (i.e. male spined, female spined, male reduced,

and female reduced). For this visualization, the effect of year of sampling is not depicted.

Figure 5. Comparison of mean number of gill rakers between spined and pelvic-reduced brook

stickleback in Muir Lake and Shunda Lake. The horizontal span of each line indicates the range

of fish lengths for each group in the comparison (i.e. male spined, female spined, male reduced,

and female reduced). For this visualization, the effect of year of sampling is not depicted.

649