# Sparsity of higher-order interactions enables learning and prediction for microbiomes

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**Microbiome engineering offers the potential to leverage microbial communities to improve outcomes in human health, agriculture, and climate. To translate this potential into reality, it is crucial to reliably predict community composition and function. But a brute force approach to cataloguing community function is hindered by the combinatorial explosion in the number of ways we can combine microbial species. An alternative is to parametrize microbial community outcomes using simplified, mechanistic models, and then extrapolate these models beyond where we have sampled. But these approaches remain data-hungry, as well as requiring an *a priori* specification of what kinds of mechanism are included and which are omitted. Here, we resolve both issues by introducing a new, mechanism-agnostic approach to predicting microbial community compositions using limited data. The critical step is the discovery of a sparse representation of the community landscape. We then leverage this sparsity to predict community compositions, drawing from techniques in compressive sensing. We validate this approach on *in silico* community data, generated from a theoretical model. By sampling just $\sim 1\%$ of all possible communities, we accurately predict community compositions out of sample. We then demonstrate the real-world application of our approach by applying it to four experimental datasets, and showing that we can recover interpretable, accurate predictions from highly limited data.**

## Introduction

Our planet is host to a multitude of microbial communities, also known as microbiomes, which perform an enormous range of functions in shaping biogeochemical processes, agricultural productivity, and animal and human health [1–3]. In recent years, there have been concerted efforts to modify such communities in order to alter plant, animal, human, and environmental health for the better [4–14]. But the complexity and diversity of microbial communities represent a major challenge in making this process predictable, with controlled outcomes. Complex interactions, potentially involving multiple species, mean that the final composition and function of a microbiome may be very different from its initial composition. High diversity of distinct taxa means that there is a combinatorially-large number of distinct initial compositions. Putting these two issues together

makes a brute-force approach to cataloging potential microbiomes impossible: with as few as ten species, around 1000 experiments would be needed to survey the full range of possible outcomes. With 100 species, that number becomes $10^{30}$ [15].

Building mechanistic models of microbial interactions has the potential to alleviate this issue. If we can write down and parameterize a model that accurately represents interactions, but using only a limited amount of experimental data, we might be able to use such a model to predict outcomes beyond those experiments. A longstanding approach to modeling interactions has been to use pairwise interactions among species, with the strength of this pairwise dependence encoded in a community interaction matrix [16, 17]. But there is a recent realization that microbial communities may be infused with higher-order interactions, where the influence of one species on another is dependent on the presence or absence of a third, fourth, or fifth species [18–22]. With the potential for such higher-order interactions, fitting mechanistic models could just bring us back to a similar combinatorial problem to the one we started with[17, 18, 23, 24]. Finally, deep learning has been deployed to address this question[23, 25], at the expense of making results and predictions challenging to interpret.

Therefore, developing a method that requires only limited experimental data, is agnostic to any particular ecological model, and provides ecologically-interpretable, accurate results, would add an important tool to the microbial ecologist's toolbox. Here, we will address this need, introducing an approach that reveals a previously-hidden sparsity in both synthetic and experimental microbial communities. This sparsity means that the outcomes of microbial community assembly are highly constrained, in effect containing much less information than the full combinatorial set would suggest. We then leverage this sparsity, using algorithms from compressive sensing to accurately recover this sparse representation. Thus we predict the late-time outcomes for microbial community composition, from highly limited input data, in a way that is readily interpretable in terms of the sparsity of species interactions.

**Fig. 1. A sparse recovery algorithm can be used to predict microbial community end points A.** Given a species pool with $S$ members, there are $2^S - 1$ sub-communities, based on different choices of initial presence-absence of the members. This problem of exponential scale means that only a few sub-communities can be sampled in the laboratory. In this paper we focus on predicting end-point community compositions of unseen sub-communities, given limited data. **B** We found that the relative abundances of any species in the pool when stacked according to the sub-communities in which it was initially present is sparse in a weighted Walsh-Hadamard basis. This means that even though a chosen focal species (green,round) may have numerous, different steady-state abundances in different sub-communities, a weighted Walsh-Hadamard representation of this vector will have only a few components. The numerous, different possible steady state abundances are represented by the colors in the column corresponding to $\vec{N}_i(\vec{\sigma})$. Sparsity of the representation, $\vec{\beta}_i(\vec{\omega})$, is visualised by the number of transparent components, or boxes, which indicate a lot of the coefficients in this representation are insignificant. This sparsity may be leveraged by using a sparse recovery technique, called compressive sensing, which prescribes that a much smaller, generic sampling of sub-communities is required to efficiently predict steady-state abundances of the species in all other sub-communities.

## Modeling Approach

**The challenge of predicting microbial community composition.** We first explicitly state the problem to address: given a set of $S$ species to draw from, there are $2^S - 1$ possible combinations, or seed communities, that can be formed, based on the initial presence-absence of species. This initial condition naturally does not capture the full subsequent behavior of the community, which could be extremely complex. Here, we'll make a simplifying assumption that, from a given initial condition, all species will approach an equilibrium at late times, such that each species ends up with a relative abundance that only depends on the seed community composition. This excludes more general dynamics, including chaos, limit cycles, or priority effects. We justify this assumption by noting, as in [15], that this simple behavior is what has been frequently observed in small experimental communities. Furthermore, this behavior is predicted by many mechanistic models [15]. Still, even with this assumption, if there are any interactions among species at all, it is reasonable to assume that the abundance of a species at steady state will in general depend on which other species are present, and may therefore be quite different in different seed communities.

Mathematically, we will label each possible community one can obtain from a pool of $S$ species using a binary vector $\vec{\sigma}$, with ones (zeros) denoting the presence (absence) of each species in the initial community. There are $2^S$ distinct values of $\vec{\sigma}$ corresponding to the different species combinations possible, with one ecologically trivial case of all species being absent initially. The steady-state abundance of a species $\alpha$ in one of the sub-communities can then be written as $N_\alpha(\vec{\sigma})$, since the abundance by assumption depends on the presence of species in the seed community. Further, since species $\alpha$ is absent from half of the $2^S$ possible combinations, $N_\alpha(\vec{\sigma})$ can take up to $2^{S-1}$ non-zero, and potentially distinct values. Thus representing $N_\alpha(\vec{\sigma})$ in the $\vec{\sigma}$ basis requires up to $2^{S-1}$ non-zero coefficients, which we label $a_{\vec{\sigma}}$. For each species, we have thus defined a map between the $\vec{\sigma}$ space and the space of steady-state abundances. This formalism, of defining maps based on composition, is not new, and has recently been explored in many works, in the context of composition and community functions. These maps have been referred to as ecological landscapes. [15, 20, 26, 27] Here, we focus on the notion of ecological landscape, where the mapping of interest is between the initial composition of the community, and the final, steady-state abundance of each species. We thus consider as many ecological landscapes as there are species.

The ideal approach to overcoming this combinatorial challenge would be if there were a more efficient representation of steady-state species abundances using fewer coefficients. In the field of signal processing, we would say that this is possible for a given signal if there is a representation that compresses it. We aren't guaranteed that such a representation exists for microbiomes, but below we'll show that viewed in the right basis, microbiomes are highly compressible.

**Species abundances can be represented using only a few coefficients in a novel basis.** The question of mapping initial community composition to species abundances at late times is related to the problem of mapping genotypes to fitness in evolutionary genetics, where species interactions are analogous to epistasis among distinct genetic mutations. This connection is made more concrete by considering a combinatorially complete fitness landscape of $S$ bi-allelic loci. For each of the $2^S$ genotypes, there exists a scalar value of interest, the fitness. The landscape can be represented as a vector $f$, with length, $2^S$. Each element is a fitness value, and corresponds to a genotype ordered by an $S$ bit binary number whose digits 1 and 0, respectively, signal the presence or absence of the mutation at the corresponding loci [28, 29]. We propound, that in the case of microbiomes and under the assumptions we outlined in the previous section, the genotypes are the distinct sub-communities that are possible based on combining species simply based on presence-absence, and the fitness vectors are analogous to the steady-state abundances, one vector for each species. The role of 0s and 1s in the binary string is changed, a mutation (the digit 1) means that a species is initially present.

Here, we draw upon recent progress in the field of evolutionary genetics to inspire a new approach, applicable to microbiomes. Specifically, some empirical fitness landscapes are sparse when represented in a type of high-dimensional Fourier basis, called the Walsh-Hadamard basis. [29–34]. Moreover, in the field of signal processing, it has been noted that most natural images and many time-series signals are stored efficiently in the same kind of basis [35]. Discovery of these sparse representations, or sparse coding, has then been used to learn entire datasets from sampling only limited data using algorithms from the field of compressive sensing [30, 34, 36].

These breakthroughs in other fields motivated us to consider the representation of species abundances in various sub-communities in a similarly-transformed perspective, which we denote as $\vec{\omega}$. Specifically, we used a weighted Walsh-Hadamard basis (see Methods) [34, 37, 38]. Up to multiplication by a diagonal matrix, this transform is the well-known Walsh-Hadamard (WH) transform, an analog of Fourier transforms in this discrete basis. It has several useful properties as discussed in our SI, including orthogonality [28, 29, 39]. Using the weighted Walsh-Hadamard transform, we get an alternative representation of species abundances $b(\vec{\omega})$ defined by

$$b(\vec{\omega}) = \Omega a(\vec{\sigma})$$

where $\Omega$ is the transformation matrix defined as $\Omega = VH$, $H$ is the Hadamard matrix and $V$ is a diagonal

weighting matrix, both described in Methods. Multiplication by the Hadamard matrix $H$ transforms the representation into an orthogonal basis, while multiplication by the weighting matrix $V$ implements an ecologically motivated assumption that down-weighs higher-order interactions. This is based on the assumption that while higher-order interactions are allowed, lower-order interactions are more likely, and the weighing encoded in $V$ biases us to infer these interactions.

To test whether microbiomes could plausibly be sparse in this basis, we first generated abundance data from simulated microbial communities using microbial consumer-resource (MiCRM) models (see Methods and SI) [40, 41]. The advantage of starting with simulated communities is that we are able to comprehensively generate all possible species combinations, and thus test rigorously to what extent sparsity does or does not hold in the $\vec{\omega}$ basis. In particular, from a pool of 16 species, we simulated all possible 65,536 species combinations, obtaining the late-time, steady-state abundances arising from each initial species combination. We repeated this for 10 different species pools to test the robustness of our results.

Figure 2B shows the coefficients in the $\vec{\omega}$ basis, $b_{\vec{\omega}}$, calculated from the steady-state abundances of a typical species from this simulated pool. A small fraction of the 32,768 ($= 2^{16-1}$) coefficients are significantly larger than the others and using just the 50 largest coefficients to estimate the observed abundances in each community, we explain 99% of the observed variance (see Figure 2C). Figure 2D shows the goodness-of-fit as we change the number of coefficients used to fit species abundance. The goodness of fit is measured as a prediction score [34, 42],

$$\text{PS} = \frac{1}{2 - R^2}, \qquad (1)$$

where $R^2$ is the coefficient of determination. The prediction score always lies between 0 and 1. For a model that could predict only the mean abundance (over all sub-communities in which it was initially present) regardless of the input sub-community, this prediction score would be 0.5 (corresponding to $R^2 = 0$). A score of 1 indicates perfect prediction. A high value of the goodness-of-fit measure indicates that we are able to capture most of the variation in species abundance using a small number of coefficients, reinforcing that this representation of species abundances is highly sparse in this basis. This trend is consistent for all species across all species pools we simulated (see Figure 2E).

## Results

**Compressive sensing predicts simulated community compositions from limited data.** In simulated consumer-resource models, we have established that there is a sparse representation of species abundances. But we obtained this sparse representation from a full

knowledge of all species abundances $N(\vec{\sigma})$, from all initial species combinations. Even if sparsity does also hold in real microbiomes, we will need a way to identify the largest, most significant coefficients in the transformed basis from only limited experimental data. Compressive sensing (CS) provides this method. Assuming that a sparse representation *does* exist for a given signal, compressive sensing will recover the signal in its entirety from only limited, generic sampling [30, 34, 36]. To predict community end-points from limited abundance data, we now apply compressive sensing in this ecological context. Specifically, we minimize the sum of number of non-zero coefficients and the squared deviation between limited, observed data and the best-fit in the $\vec{\omega}$ basis. This is implemented using the Basis Pursuit Denoising (BPDN) algorithm [34, 44, 45] (see Methods for a detailed description of the algorithm).

Summarizing our methodology : our simulated communities are sparse in the $\vec{\omega}$ basis, and we hope compressive sensing allows us to recover these $\vec{\omega}$ coefficients from limited measurements of species abundances. We then use these 'best guess' $\vec{\omega}$ coefficients to compute species abundances outside of our original limited "training" data.

We tested this approach in our simulated communities, for which we have access to all data in practice, and where we have already shown that a sparse basis exists. Specifically, we restricted ourselves to observing/sampling $1\%$ or $327$ sub-communities of the 32,678 possible sub-communities with a given species present at the start. We also considered lower sampling percentages of $0.1\%$, $0.2\%$, and a higher percentage of $2\%$. With these limited "training" data, we used CS algorithms to calculate coefficients in the $\vec{\omega}$ basis (see Methods). Figure 3A shows that CS is able to infer the most significant coefficients accurately from limited data, and the accuracy increases as more data are obtained for training.

Next, we implement an exact inverse weighted WH transform on the best guess $\vec{\omega}$ to recover abundances from coefficients inferred by compressive sensing algorithms, and Figure 3B shows the resulting prediction score on out-of-sample data. We see that the algorithm's predictive power improves with more training data, and appears to asymptote at as little as 1% of all possible data included in the training set. We compare our method with two alternative approaches: an $l$-1 regularized regression on the $\vec{\sigma}$ basis that tries to estimate $N_\alpha(\vec{\sigma})$ directly, and random forest regression. (See Methods for detailed description of these algorithms) The $l$-1 regularized regression is adopted since the number of observations is smaller than the number of possible coefficients, i.e., the problem is under-determined. We found that compressive sensing outperforms both regularized regression and random forest regression, as can be seen from the median values of the predictions in Figure 3C. Finally, Figure 3D compares the prediction score of the three methods for each species, which is

**Fig. 2. Relative abundances of a synthetic community are sparse in the weighted Walsh-Hadamard ($\vec{\omega}$) basis A.** We simulated a pool of 16-species in silico using microbial consumer-resource models [40, 43]. Simulations started from all 65535 possible sub-communities corresponding to distinct species presence-absences ($\vec{\sigma}$) to obtain species abundances in the different steady-state communities. The relative abundance of a species, in the 32678 sub-communities in which it was initially present, is $N(\vec{\sigma})$. By using a weighted Walsh-Hadamard transform, species abundance can be represented in an orthogonal $\vec{\omega}$ basis. **B.** The coefficients used to represent abundances of a typical species in the 32678 sub-communities using the $\vec{\omega}$ basis are plotted sorted by absolute size. The vast majority of the 32678 coefficients are small; the inset plots the largest 50 coefficients. **C.** Using only 50 coefficients, we were able to explain most of the variation (99%) in species abundances. The color bar indicates the density of points in the hexplot. Panel **D.** demonstrates that a small number of $\vec{\omega}$ coefficients is sufficient to fit abundances almost perfectly. **E.** Box-plots of goodness-of-fit, aggregated for all 16 species in all 10 replicates, demonstrate that only a small number of $\vec{\omega}$ coefficients are required to explain most of the observed variation in abundances. We display medians in the text and by yellow lines, while mean statistics is denoted by white triangles. Coefficients in panel B are ordered by magnitude. The goodness-of-fit is calculated via Eq.1

a matched comparison at the species level of the three approaches, with training data held at 1%. This analysis reveals that some species are easier to predict, with a prediction score close to 1, while some others that are harder to predict. In the SI, we show that species which are easier to predict are dominated by low order interactions, and thus have a smoother abundance landscape. We can thus conclude that low-order interactions lead to a smoother, more predictable landscape. Despite this variation in performance at the species-level, compressive sensing outperforms regularized regression in almost all cases, and outperforms random forest regression in most cases.

Thus, compressive sensing provides a way to use the sparse basis for our simulated consumer-resource communities, along with limited observations of species abundances in different sub-communities, in order to predict species abundances out-of-sample.

**Compressive sensing predicts species abundances in experimental data.** Using simulated communities, we demonstrate that the challenge posed by an exponential number of sub-communities to the problem of ecolog-

ical predictions is massively reduced with compressive sensing. We now test the performance of CS as a predictive tool in a diverse range of real microbiomes, in the context of unknown community dynamics which likely depart from our idealized consumer-resource model, experimental noise, and incomplete data. We draw data from four published studies of microbial communities spanning *in vitro* and *in vivo* conditions, pool sizes from 5 to 16, and microbes from environments including the human gut, soil, and fruit flies [19, 20, 46, 47].

In the first study, Gould *et al.* assembled all combinations of 5 species *in vivo* in the gut of *Drosophila melanogaster*. The remaining three studies considered larger species pool sizes and hence assembled only a fraction of all possible species combinations *in vitro*. Sanchez-Gorostiaga *et al.* assembled 53 combinations of 6 starch-degrading soil microbes. Friedman *et al.* assembled 101 combinations of 8 soil microbes. From Clark *et al.*, we consider 187 combinations assembled of 16 gut microbes.

Figure 4 demonstrates the performance of compressive sensing, $l$-1 regularized regression, and random forests on predicting unseen communities using a k-fold

**Fig. 3. Sparsity in the $\bar{\omega}$ coefficients implies abundances can be practically recovered from limited data in simulated communities A.** The accuracy of predicting the 4 most significant $\omega$ coefficients by compressive sensing (CS) increases as the size of training data is increased. The largest coefficients, which explain most of the variance in abundance, are more easily predicted than smaller coefficients from limited data. The percentages in the parenthesis next to the coefficient symbol and index denote the percentage of the variance explained by each coefficient taken individually. This means, for example, that the first $\omega$ term explains $52\%$ of the total variance. **B.** The ability of compressive sensing to predict species abundances in new sub-communities when trained on a small fraction of all possible sub-communities is shown using the Prediction Score (Eq.1). Note that 1% corresponds to 327 sub-communities, which would requires 4 96-well plates. The median score, shown by the yellow line is annotated. The number of sub-communities sampled at each training level is indicated within parentheses. **C.** The prediction score of two alternative methods, $L1$ regularized regression and random forest regression is shown for comparison. We controlled for data so that all the methods were trained on the same 327 (1%) communities. Box plots denote the performance across the 160 species (10 replicates of pools with 16 species). A model that outputs only the mean abundances of a species in all sub-communities in which the species was initially present, regardless of the input sub-communities, would get a prediction score of $0.5$. We see that compressive sensing does objectively better than such a model (prediction score shown using red dashed line) while outperforming random forest regression and l1 regularized regression. **D.** Comparing the performance of CS, $L1$ regularised regression, and random forest on a species-by-species basis, we find that some species are easier to predict than others. Nevertheless, CS outperforms both regressions for most species. Permutation test p-value for the difference in the performance at $1\%$ sampling between compressive sensing and random forest regression is statistically significant at $10^{-4}$ and that between compressive sensing and $L1$ regularized regression is $5 \times 10^{-5}$. All results are averages over 5 runs.

cross-validation procedure (see Methods). We used a k-fold cross-validation procedure due to the small number of data points and report the prediction score on the stacked validation sets. Further, keeping the bias-variance trade-off [48] in mind, we set $k = 3$ for most experimental communities. A complete description is given in Methods. Figure 4 B-E show the performance of compressed sensing, alongside the alternative methods, on the individual species. As in simulations, we find that some species are easier to predict than others. Compressive sensing outperforms regression in the majority of the cases (22 out of 35). Thus, our compressive

sensing approach enables more accurate predictions of microbial community abundances.

## Discussion

The search for ways to optimize microbial community function is a critical goal in microbiome engineering. However, an exhaustive search through all possible ways to combine microbial taxa is impractical, and fitting mechanistic models can still require large amounts of data, while also imposing assumptions about community dynamics that may be violated for real micro-

**Fig. 4. Performance of compressive sensing on 4 real datasets.** We tested the performance of compressive sensing on 4 real datasets, ranging from 5-16 species, and in both in-vitro and in-vivo contexts. (Methods) **A** At a community level, compressive sensing outperforms both random forest regression and l1 regularized regression. **B** For the in-vivo gut fly dataset from [19], compressive sensing does better in predicting a majority of the species. Permutation test p-values are .09 and .06 for comparison against random forest regression and l1-regularized regression. **C** For the in-vitro 6-species soil community, we find that compressive sensing outperforms random forest marginally (permutation test p-value : 0.1) but does better than l1-regularized regression (p-value : 0.015). This dataset, overall, is harder to predict. **D** Compressive sensing outperforms both random forest regression and $L1$-regularized regression (pvalues : $4 \times 10^{-3}$ and $8 \times 10^{-3}$) in the 8-species in-vitro soil community assembled by Friedman et al.[46]. **E.** For the 16 species community assembled by Clark et al. [47], we find that compressive sensing outperforms other methods reaching statistically significant values in a permutation test. (p-value : $9 \times 10^{-3}$ for comparison with random forest regression, and $1.52 \times 10^{-5}$ for $L1$-regularized regression.) We report all results as averages of 5 runs. The error bars in panels B,C,D, and E indicate standard deviation from the mean across 5 runs. Species names on the x-axis are compiled from the data as provided by authors.

biomes. Here, we establish the method of compressive sensing as a new, model-agnostic, predictive tool, applicable in situations where experimenters have access to limited data. The success of our approach relies on the assumption that species abundances are sparse in a novel basis, which we demonstrate explicitly for simulated consumer-resource models.

We use this method in silico, in-vivo, and in-vitro. We also show that our method outperforms $l$-1 regularization and a popular machine learning method, random forest regression. This formalism does not require time-series data ; this omission also makes it easier to work with data obtained from high-throughput laboratory techniques. Further, it performs well with relative abundances, which is in line with how abundances are widely observed and calculated in this field, though in principle the approach will also work well on absolute abundances, as shown in our Supplementary Information.

Recently, there has been work on applying deep-

learning models to predict community structures [23, 25].These models perform well, but their performance can be obscured by large numbers of hyper-parameters and a difficulty in interpretability. On the contrary, this method is naturally endowed with interpretation in terms of the sparseness of higher-order interactions, and our compressive sensing algorithms requires only one tunable hyper-parameter. In cases where compressive sensing predicts species abundances accurately, it seems reasonable to conclude that interactions are sparse, and that low-order interactions dominate. This gives insight into the nature of interactions in real communities.

While our approach works well in the simulated and experimental data we applied it to, this method hinges on the assumption that there is a unique steady-state set of community abundances, rather than the potential for multiple equilibria, or more complex late-time dynamics. There is therefore scope to consider further generalizations in cases where experimental data suggests that these outcomes are possible. However, for cases where

experiment suggests a unique map from initial composition to late-time abundances, our approach paves a new way to reliably engineer microbial communities.

## Methods

**Predictions on a 5-species gut community.** Gould *et al.* [19] assembled all possible communities from a 5-species pool of bacteria that are known to colonize fly guts in both wild and laboratory conditions. All the 32 sub-communities (termed treatments) were assembled in germ-free flies, each with 48 replicate experiments. They reported colony-forming unit (CFU) counts for each replicate and each treatment. For each replicate, we first computed the relative abundance of each species in that community and then took the average of these relative abundances of each condition (i.e each sub-community type).

**Predictions on a 6-species starch degrading community.** Sanchez-Gorostiaga *et al.* [20] studied assemblages by considering a pool of $6$ amylolytic soil bacterial species. Out of the 63 possible communities, they reported abundance data (in colony-forming units) for 53 communities. We dropped two communities that were inconsistent with labeling, and worked with 51 data points. For each community and each replicate, we calculated the total biomass of the community, and divided the abundance of each species to find the relative abundance of each species in the community. If multiple replicate experiments were reported, we first computed the relative abundance of a species in each replicate and then averaged this across replicates to find the mean relative abundance.

**Predictions on an 8-species soil microbiome community.** Friedman *et al.* [46] studied a community of 8 heterotrophic soil-dwelling bacteria, reporting their optical densities (ODs) after cross-checking actual cell counts using agar plating. They considered sub-communities which included all the 8 monocultures, all pairwise combinations, all three-species communities, all 8 leave-one-out communities, and the sub-community with all the species present. They combined species in different ratios (i.e other than $1:1$), and sampled the populations at various times, obtaining time series data. This difference in starting abundances however did not affect the steady state in most conditions and replicates. However, for $2$ sub-communities they found that the steady-state abundances within the replicates had a much larger variation than what could be accounted for by experimental noise, and one sub-community displayed bistability. We discarded these experimental conditions, and for all other data points proceeded to take the average of the final time (where we assume steady state has been attained) abundances across the replicates.

**Predictions on a subset of a 25-species synthetic human gut community.** Clark *et al.* [47] studied a community of 25 species that consisted of species spanning all major phyla in the human gut microbiome, and also are representative of the major metabolic functions in the gut. Of the $2^{25} - 1$ ecological communities possible, they sampled $\sim 600$. For these sub-communities, they reported read counts, and computed the relative abundances for each species using total read counts for each sub-community. They also found absolute abundances by multiplying the relative abundance with the $\mathrm{OD}_{600}$ measurement for each sample. As outlined in the Methods section of [47], we excluded sub-communities that were flagged as contaminated. In such a data-limited case, the performance of any algorithm will be difficult to test. Therefore, we considered subsets of 16 species with other 9 always being absent. With this, we had $\sim 0.2\%$ of the 16-species landscape sampled. We considered relative abundances in the data, after averaging over the number of replicate experiments for each sub-community. There were differing number of data points corresponding to each species, unlike the previous datasets. We therefore allowed the $k$ to vary in the cross-validation scheme. $k$ was set to 3, 5, or 7 depending on the size of the dataset for each species.

**Microbial consumer-resource models.** We considered a pool of 16 species, with a single externally supplied resource in a chemostat. Communities with cross-feeding that are supplied with a single resource externally have been studied in recent experiments [49–53]. This supplied resource is primarily consumed by only a few species. However, in the presence of cross-feeding, there are $20$ metabolites present in the community. Different species consume different resources at different rates, this information is encoded in the consumer matrix, $C$, with elements $c_{i\alpha}$. We choose a consumer matrix whose elements are sampled from a binary-gamma distribution [54]. We chose a binary-gamma distribution because this allows for positive uptake rates, but also allows us to control the sparsity of the consumer matrix. By considering all possible combinations of initial presence-absence of the species, we generated steady state abundances of all the species, by numerically solving the ODEs for the consumer-resource model (equations given in SI). Simulations reached steady state when the root mean square of the logarithmic growth rates rates of the species fell below a threshold, i.e., $\mathrm{RMS}\left(\frac{1}{N_i}\frac{dN_i}{dt}\right) < 10^{-2}$. Further, we verified that the extinct species could not have survived in the community by simulating a re-invasion attempt of the steady-state community.

Thus, for each species, we have $2^{16}$ abundance data points. Since each species is present in only half the combinations, the effective total data for each species is $2^{15}$. Further, we considered the relative abundance of a species in each combination. We calculated the

relative abundance by summing the biomass of each species in the sub-community and dividing this by the total biomass of that sub-community. Details of parameters used in the simulations are given in the SI.

**Representation of Abundances in the Walsh-Hadamard Basis.** We consider a combinatorially complete data-set of microbial abundances at steady state. Starting with a set of $S$ species, there are $2^S - 1$ of combining them, each assemblage characterised by the species initially present. We consider that the underlying population dynamics of these sub-communities, whatever they are, give rise to a unique steady state for each sub-community, i.e we do not consider cases of multi-stability. Borrowing from the language of genetics and epistasis, we consider a vector of abundances at steady state for each of the $S$ species. For concreteness, in this section, we consider a set of 3 species. The set of all the sub-communities, each with a different starting composition is enumerated as : $\vec{\sigma} = \{[000], [001], [010], [011], [100], [101], [110], [111]\}$. We include the ecologically trivial case of all species being initially absent. The element $[011]$, for example, denotes the sub-community with species 2 and 3 present at the start. For each of these unique conditions, we consider the steady-state abundance (say, cell counts) of each species. This allows for a mapping to be written down explicitly, between the decimal-ordered binary elements and the $S$ steady-state abundance vectors.

$$
\begin{bmatrix} 000 \\ 001 \\ 010 \\ 011 \\ 100 \\ 101 \\ 110 \\ 111 \end{bmatrix} \mapsto \begin{bmatrix} a_{000} \\ a_{001} \\ a_{010} \\ a_{011} \\ a_{100} \\ a_{101} \\ a_{110} \\ a_{111} \end{bmatrix}_0 , \begin{bmatrix} a_{000} \\ a_{001} \\ a_{010} \\ a_{011} \\ a_{100} \\ a_{101} \\ a_{110} \\ a_{111} \end{bmatrix}_1 , \begin{bmatrix} a_{000} \\ a_{001} \\ a_{010} \\ a_{011} \\ a_{100} \\ a_{101} \\ a_{110} \\ a_{111} \end{bmatrix}_2 \ldots \quad \textbf{(2)}
$$

Here the subscripts on the vectors denote the species indices. We can define a Walsh-Hadamard transform on this $2^S$ long vector of steady-state abundances of a species $\alpha$. In particular, we work with a weighted Walsh-Hadamard transform. The weighted transform is implemented by the matrix : $VH$ where the matrices $V$ and $H$ are generated by the recursion relations:

$$
H_{n+1} = \begin{bmatrix} H_n & H_n \\ H_n & -H_n \end{bmatrix} \quad \textbf{(3)}
$$

with $H_0 = 1$

$$
V_{n+1} = \begin{bmatrix} 0.5V_n & 0 \\ 0 & -V_n \end{bmatrix} \quad \textbf{(4)}
$$

with $V_0 = 1$.
$V$ is a diagonal weighting matrix that takes into account

the order of interactions, to account for averaging over different numbers of terms as a function of the order of interactions [37]. In our case, it serves to provide a way to bias inference from limited data towards lower-order interactions.

**Compressive Sensing and Sparse Recovery Algorithms.** Implementation of compressive sensing involves using optimization algorithms that find a sparse representation of the data using small subsets of observations. After a sparse representation has been found by the algorithm, the rest of the unseen data is computed by taking the inverse transform of the best-guess sparse representation. This inverse transform is the inverse of the matrix, $\Omega$, that is known/expected to sparsify the data. To find the sparse representations, we employed the basis pursuit denoising (BPDN) algorithm [44] which is an optimization problem posed as : $\underset{b}{\mathrm{argmin}}\left(\frac{1}{2}||Db - a||_2^2 + \lambda||b||_1\right)$. The idea is to find the $b$ vector that has the smallest $l_1$ norm while keeping the error from observed data as low as possible. To find the sparsest representation, the problem should actually minimize the $l_0$ norm instead of $l_1$ but this problem is non-convex and combinatorial [35], and $l_1$ norm minimization is a convex approximation to this. This algorithm has only one hyperparameter, $\lambda$, which can be tuned according to the expected sparsity of the representations. In real datasets, this value needs to be chosen carefully. We report all results in the main text based on the optimal hyperparameter. We used the SPORCO package [55] in Python to implement BPDN using an alternating direction method of multipliers (ADMM) algorithm [56].

**$l$-1 regularized regression.** For a species $\alpha$ with relative abundances $a(\vec{\sigma})$, we consider a representation

$$
\vec{g} = G_n a(\vec{\sigma})
$$

Here $G$ is a matrix defined recursively as :

$$
G_{n+1} = \begin{bmatrix} G_n & 0 \\ -G_n & G_n \end{bmatrix} \quad \textbf{(5)}
$$

with $G_0 = 1$ We note that, as elucidated in [34, 37], this transformation can be thought of as looking at the abundance landscape of a species as a local approximation around a single sub-community – the one with all species absent. This is akin to a Taylor expansion on the landscape, while the class of Walsh-Hadamard transforms corresponds to a Fourier transform [57] on the landscape, and is an approximation over the background of all sub-communities. We implement a similar BPDN algorithm as for compressive sensing with Walsh-Hadamard transform, where the algorithm tries to learn the $g$ coefficients.

**Bench-marking with Random Forest Regression.** Tree-based ensemble learners, like the random forest regressor and xgboost, are popular choices of supervised learning algorithms, especially when the predictors are not sure to be linearly related to the target variables. We implemented the random forest regressor as available in sci-kit learn [58].While there are arguments for using a multi-output regression instead of several single-output ones [59], we worked with fitting a random forest model to each species individually. For data control and an apples-to-apples comparison, we used the same data splits and number of folds as we did in compressive sensing. With random forest regression, there are multiple hyper-parameters to consider. To reduce the number of hyper-parameters to tune, given the small size of some of the experimental datasets, we used the number of estimators (trees) to be the sci-kit learn default (100) and the number of features for selection at every split to be $1/3$ of the number of species in each case.

## Bibliography

1. Falkowski, P. G., Fenchel, T. & Delong, E. F. The Microbial Engines That Drive Earth's Biogeochemical Cycles. *Science* **320**, 1034–1039 (2008). URL https://www.sciencemag.org/lookup/doi/10.1126/science.1153213.

2. Paerl, H. W. & Pinckney, J. L. A Mini-Review of Microbial Consortia: Their Roles in Aquatic Production and Biogeochemical Cycling. *Microbial Ecology* **31**, 225–247 (1996). URL https://www.jstor.org/stable/25152978. Publisher: Springer.

3. Flint, H. J., Scott, K. P., Duncan, S. H., Louis, P. & Forano, E. Microbial degradation of complex carbohydrates in the gut. *Gut Microbes* **3**, 289–306 (2012). URL http://www.tandfonline.com/doi/abs/10.4161/gmic.19897.

4. Brenner, K., You, L. & Arnold, F. H. Engineering microbial consortia: a new frontier in synthetic biology. *Trends in Biotechnology* **26**, 483–489 (2008). URL https://linkinghub.elsevier.com/retrieve/pii/S0167779908001716.

5. Olle, B. Medicines from microbiota. *Nature Biotechnology* **31**, 309–315 (2013). URL http://www.nature.com/articles/nbt.2548.

6. Tanoue, T. *et al.* A defined commensal consortium elicits CD8 T cells and anti-cancer immunity. *Nature* **565**, 600–605 (2019). URL http://www.nature.com/articles/s41586-019-0878-z.

7. Minty, J. J. *et al.* Design and characterization of synthetic fungal-bacterial consortia for direct production of isobutanol from cellulosic biomass. *Proceedings of the National Academy of Sciences* **110**, 14592–14597 (2013). URL http://www.pnas.org/lookup/doi/10.1073/pnas.1218447110.

8. Lindemann, S. R. *et al.* Engineering microbial consortia for controllable outputs. *The ISME Journal* **10**, 2077–2084 (2016). URL http://www.nature.com/articles/ismej201626.

9. Buffie, C. G. *et al.* Precision microbiome reconstitution restores bile acid mediated resistance to Clostridium difficile. *Nature* **517**, 205–208 (2015). URL http://www.nature.com/articles/nature13828.

10. Hu, J. *et al.* Design and composition of synthetic fungal-bacterial microbial consortia that improve lignocellulolytic enzyme activity. *Bioresource Technology* **227**, 247–255 (2017). URL https://linkinghub.elsevier.com/retrieve/pii/S096085241631728X.

11. Senne de Oliveira Lino, F., Bajic, D., Vila, J. C. C., Sánchez, A. & Sommer, M. O. A. Complex yeast–bacteria interactions affect the yield of industrial ethanol fermentation. *Nature Communications* **12**, 1498 (2021). URL https://www.nature.com/articles/s41467-021-21844-7. Number: 1 Publisher: Nature Publishing Group.

12. Weng, J.-K., Li, X., Bonawitz, N. D. & Chapple, C. Emerging strategies of lignin engineering and degradation for cellulosic biofuel production. *Current Opinion in Biotechnology* **19**, 166–172 (2008). URL https://www.sciencedirect.com/science/article/pii/S0958166908000268.

13. Piccardi, P., Vessman, B. & Mitri, S. Toxicity drives facilitation between 4 bacterial species. *Proceedings of the National Academy of Sciences* **116**, 15979–15984 (2019). URL https://www.pnas.org/doi/abs/10.1073/pnas.1906172116. Publisher: Proceedings of the National Academy of Sciences.

14. Swenson, W., Wilson, D. S. & Elias, R. Artificial ecosystem selection. *Proceedings of the National Academy of Sciences* **97**, 9110–9114 (2000). URL http://www.pnas.org/cgi/doi/10.1073/pnas.150237597.

15. George, A. B. & Korolev, K. S. Ecological landscapes guide the assembly of optimal microbial communities. *PLOS Computational Biology* **19**, e1010570 (2023). URL https://journals.plos.org/ploscompbiol/article?id=10.1371/journal.pcbi.1010570. Publisher: Public Library of Science.

16. Maynard, D. S., Miller, Z. R. & Allesina, S. Predicting coexistence in experimental ecological communities. *Nature Ecology & Evolution* **4**, 91–100 (2020). URL http://www.nature.com/articles/s41559-019-1059-z.

17. Skwara, A., Lemos-Costa, P., Miller, Z. R.

& Allesina, S. Modelling ecological communities when composition is manipulated experimentally. *Methods in Ecology and Evolution* **14**, 696–707 (2023). URL https://onlinelibrary.wiley.com/doi/abs/10.1111/2041-210X.14028. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1111/2041-210X.14028.

18. Ansari, A. F., Reddy, Y. B. S., Raut, J. & Dixit, N. M. An efficient and scalable top-down method for predicting structures of microbial communities. *Nature Computational Science* **1**, 619–628 (2021). URL https://www.nature.com/articles/s43588-021-00131-x.

19. Gould, A. L. *et al.* Microbiome interactions shape host fitness. *Proceedings of the National Academy of Sciences* **115**, E11951–E11960 (2018). URL http://www.pnas.org/lookup/doi/10.1073/pnas.1809349115.

20. Sanchez-Gorostiaga, A., Bajić, D., Osborne, M. L., Poyatos, J. F. & Sanchez, A. High-order interactions distort the functional landscape of microbial consortia. *PLOS Biology* **17**, e3000550 (2019). URL https://dx.plos.org/10.1371/journal.pbio.3000550.

21. Bairey, E., Kelsic, E. D. & Kishony, R. High-order species interactions shape ecosystem diversity. *Nature Communications* **7**, 12285 (2016). URL https://www.nature.com/articles/ncomms12285. Number: 1 Publisher: Nature Publishing Group.

22. Morin, M. A., Morrison, A. J., Harms, M. J. & Dutton, R. J. Higher-order interactions shape microbial interactions as microbial community complexity increases. *Scientific Reports* **12**, 22640 (2022). URL https://www.nature.com/articles/s41598-022-25303-1. Number: 1 Publisher: Nature Publishing Group.

23. Baranwal, M. *et al.* Recurrent neural networks enable design of multifunctional synthetic human gut microbiome dynamics. *eLife* **11**, e73870 (2022). URL https://elifesciences.org/articles/73870.

24. Fisher, C. K. & Mehta, P. Identifying Keystone Species in the Human Gut Microbiome from Metagenomic Timeseries Using Sparse Linear Regression. *PLoS ONE* **9**, e102451 (2014). URL https://dx.plos.org/10.1371/journal.pone.0102451.

25. Michel-Mata, S., Wang, X.-W., Liu, Y.-Y. & Angulo, M. T. Predicting microbiome compositions from species assemblages through deep learning. *iMeta* **1**, e3 (2022). URL https://onlinelibrary.wiley.com/doi/abs/10.1002/imt2.3. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1002/imt2.3.

26. Sánchez, *et al.* Directed Evolution of Microbial Communities 19 (2021).

27. Sanchez, A. *et al.* The community-function landscape of microbial consortia. *Cell Systems* **14**, 122–134 (2023). URL https://www.sciencedirect.com/science/article/pii/S2405471222004999.

28. Weinreich, D. M., Lan, Y., Wylie, C. S. & Heckendorn, R. B. Should evolutionary geneticists worry about higher-order epistasis? *Current Opinion in Genetics & Development* **23**, 700–707 (2013). URL https://www.sciencedirect.com/science/article/pii/S0959437X13001421.

29. Weinreich, D. M., Lan, Y., Jaffe, J. & Heckendorn, R. B. The Influence of Higher-Order Epistasis on Biological Fitness Landscape Topography. *Journal of Statistical Physics* **172**, 208–225 (2018). URL http://link.springer.com/10.1007/s10955-018-1975-3.

30. Brookes, D. H., Aghazadeh, A. & Listgarten, J. On the sparsity of fitness functions and implications for learning. *Proceedings of the National Academy of Sciences* **119**, e2109649118 (2022). URL https://pnas.org/doi/full/10.1073/pnas.2109649118.

31. Ballal, A. *et al.* Sparse Epistatic Patterns in the Evolution of Terpene Synthases. *Molecular Biology and Evolution* **37**, 1907–1924 (2020). URL https://academic.oup.com/mbe/article/37/7/1907/5771371. Publisher: Oxford Academic.

32. Sailer, Z. R. & Harms, M. J. Detecting High-Order Epistasis in Nonlinear Genotype-Phenotype Maps. *Genetics* **205**, 1079–1088 (2017). URL https://academic.oup.com/genetics/article/205/3/1079/6066378. Publisher: Oxford Academic.

33. Yang, G. *et al.* Higher-order epistasis shapes the fitness landscape of a xenobiotic-degrading enzyme. *Nature Chemical Biology* **15**, 1120–1128 (2019). URL https://www.nature.com/articles/s41589-019-0386-3. Number: 11 Publisher: Nature Publishing Group.

34. Poelwijk, F. J., Socolich, M. & Ranganathan, R. Learning the pattern of epistasis linking genotype and phenotype in a protein. *Nature Communications* **10**, 4213 (2019). URL https://www.nature.com/articles/s41467-019-12130-8. Number: 1 Publisher: Nature Publishing Group.

35. Hastie, T., Tibshirani, R. & Wainwright, M. *Statistical Learning with Sparsity* (Routledge, Boca Raton, 2015), 1st edition edn.

36. Candes, E. & Wakin, M. An Introduction To Compressive Sampling. *IEEE Signal Processing Magazine* **25**, 21–30 (2008). URL http://ieeexplore.

ieee.org/document/4472240/.

37. Poelwijk, F. J., Krishna, V. & Ranganathan, R. The Context-Dependence of Mutations: A Linkage of Formalisms. *PLOS Computational Biology* **12**, e1004771 (2016). URL https://dx.plos.org/10.1371/journal.pcbi.1004771.

38. Yitbarek, S., Guittar, J., Knutie, S. A. & Ogbunugafor, C. B. Deconstructing taxa x taxa x environment interactions in the microbiota: A theoretical examination. *bioRxiv* 647156 (2021). URL https://www.biorxiv.org/content/10.1101/647156v2. Publisher: Cold Spring Harbor Laboratory Section: New Results.

39. Doro, S. & Herman, M. A. On the Fourier transform of a quantitative trait: Implications for compressive sensing. *Journal of Theoretical Biology* **540**, 110985 (2022). URL https://linkinghub.elsevier.com/retrieve/pii/S0022519321004057.

40. Marsland, R., Cui, W., Goldford, J. & Mehta, P. The Community Simulator: A Python package for microbial ecology. *PLOS ONE* **15**, e0230430 (2020). URL https://dx.plos.org/10.1371/journal.pone.0230430.

41. Marsland, R., Cui, W. & Mehta, P. A minimal model for microbial biodiversity can reproduce experimentally observed ecological patterns. *Scientific Reports* **10**, 3308 (2020). URL https://www.nature.com/articles/s41598-020-60130-2. Number: 1 Publisher: Nature Publishing Group.

42. Nash, J. E. & Sutcliffe, J. V. River flow forecasting through conceptual models part I — A discussion of principles. *Journal of Hydrology* **10**, 282–290 (1970). URL https://www.sciencedirect.com/science/article/pii/0022169470902556.

43. Marsland, R. *et al.* Available energy fluxes drive a transition in the diversity, stability, and functional structure of microbial communities. *PLOS Computational Biology* **15**, e1006793 (2019). URL http://dx.plos.org/10.1371/journal.pcbi.1006793.

44. Chen, S. S., Donoho, D. L. & Saunders, M. A. Atomic Decomposition by Basis Pursuit. *SIAM Journal on Scientific Computing* (2006). URL https://epubs.siam.org/doi/10.1137/S1064827596304010. Publisher: Society for Industrial and Applied Mathematics.

45. Tibshirani, R. Regression Shrinkage and Selection Via the Lasso. *Journal of the Royal Statistical Society: Series B (Methodological)* **58**, 267–288 (1996). URL https://onlinelibrary.wiley.com/doi/abs/10.1111/j.2517-6161.1996.tb02080.x. _eprint: https://onlinelibrary.wiley.com/doi/pdf/10.1111/j.2517-6161.1996.tb02080.x.

46. Friedman, J., Higgins, L. M. & Gore, J. Community structure follows simple assembly rules in microbial microcosms. *Nature Ecology & Evolution* **1**, 0109 (2017). URL http://www.nature.com/articles/s41559-017-0109.

47. Clark, R. L. *et al.* Design of synthetic human gut microbiome assembly and butyrate production. *Nature Communications* **12**, 3254 (2021). URL http://www.nature.com/articles/s41467-021-22938-y.

48. Mehta, P. *et al.* A high-bias, low-variance introduction to Machine Learning for physicists. *Physics Reports* **810**, 1–124 (2019). URL https://www.sciencedirect.com/science/article/pii/S0370157319300766.

49. Goldford, J. E. *et al.* Emergent simplicity in microbial community assembly. *Science* **361**, 469–474 (2018). URL https://www.sciencemag.org/lookup/doi/10.1126/science.aat1168.

50. Gralka, M., Szabo, R., Stocker, R. & Cordero, O. X. Trophic Interactions and the Drivers of Microbial Community Assembly. *Current Biology* **30**, R1176–R1188 (2020). URL https://linkinghub.elsevier.com/retrieve/pii/S0960982220311611.

51. Estrela, S. *et al.* Functional attractors in microbial community assembly. *Cell Systems* S2405471221003793 (2021). URL https://linkinghub.elsevier.com/retrieve/pii/S2405471221003793.

52. Enke, T. N. *et al.* Modular Assembly of Polysaccharide-Degrading Marine Microbial Communities. *Current Biology* **29**, 1528–1535.e6 (2019). URL https://www.sciencedirect.com/science/article/pii/S0960982219303458.

53. Dal Bello, M., Lee, H., Goyal, A. & Gore, J. Resource–diversity relationships in bacterial communities reflect the network structure of microbial metabolism. *Nature Ecology & Evolution* **5**, 1424–1434 (2021). URL https://www.nature.com/articles/s41559-021-01535-8. Number: 10 Publisher: Nature Publishing Group.

54. Chang, C.-Y. *et al.* Engineering complex communities by directed evolution. *Nature Ecology & Evolution* **5**, 1011–1023 (2021). URL http://www.nature.com/articles/s41559-021-01457-5.

55. Wohlberg, B. SPORCO: A Python package for standard and convolutional sparse representations. *Proceedings of the 16th Python in Science Conference* 1–8 (2017). URL https://conference.scipy.org/proceedings/scipy2017/brendt_wohlberg.html. Conference Name: Proceedings of the 16th Python in Science Conference.

56. Boyd, S., Parikh, N. & Chu, E. *Distributed Optimization and Statistical Learning Via the Alternating Direction Method of Multipliers* (Now Publishers Inc, Hanover, MA, 2011).

57. Weinberger, E. D. Fourier and Taylor series on fitness landscapes. *Biol. Cybern.* **65**, 321–330 (1991). URL https://doi.org/10.1007/BF00216965.

58. Pedregosa, F. *et al.* Scikit-learn: Machine Learning in Python. *Journal of Machine Learning Research* **12**, 2825–2830 (2011). URL http://jmlr.org/papers/v12/pedregosa11a.html.

59. Schmid, L., Gerharz, A., Groll, A. & Pauly, M. Machine Learning for Multi-Output Regression: When should a holistic multivariate approach be preferred over separate univariate ones? (2022). URL http://arxiv.org/abs/2201.05340. ArXiv:2201.05340 [cs, stat].