Opportunistic evidence of the impact of bacterial infections on social integration in vampire bats

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Abstract

Social integration can affect an individual's susceptibility to infectious disease.

Conversely, infectious disease can reduce an individual's social activity. Yet, it remains unclear

to what extent short-term infections can inhibit social integration and the formation of new

relationships. During a previous study on relationship formation, we captured 21 female

common vampire bats (*Desmodus rotundus*) from different wild populations and housed them

together in captivity. Upon introduction, we observed an unplanned outbreak of bacterial

infections that caused cutaneous lesions. After treating infected bats with an antibiotic, five bats

recovered, but four others suffered lasting injuries. Given that observations of how natural

infections alter relationship formation are rare, we analyzed how allogrooming rates changed

over time among familiar and new dyads consisting of the nine infected and 12 asymptomatic

bats. We found that (1) infected bats demonstrated reduced activity and social behavior, (2)

more severely infected bats gave and received less allogrooming than asymptomatic bats, (3)

the effect of infection was larger for new dyads relative to familiar dyads, and (4) this effect

decreased as infected bats recovered and new dyads became more familiar. These

opportunistic observations were consistent with the hypothesis that short-term infections can

impact the formation of new relationships.

53   Social integration, or the quantity and quality of an individual's social relationships, can

54 affect an individual's susceptibility to infection (e.g., Pavez-Fox et al., 2021; Rushmore et al.,

55 2017; Snyder-Mackler et al., 2016, 2020; Tung et al., 2012) and ability to tolerate or suppress

56 the adverse effects of parasites (Almberg et al., 2015; Jog et al., 2022; Loehle, 1995;

57 Stockmaier et al., 2023). Conversely, infectious disease can inhibit the social behaviors of

58 infected individuals (Hart, 1988; Lopes et al., 2021; Stockmaier et al., 2021) by diminishing their

59 motivation to interact with others (e.g., Fishkin & Winslow, 1997; Moreno et al., 2021;

60 Stockmaier et al., 2018), reducing their movement (e.g., in mice (*Mus musculus domesticus*,

61 Lopes et al., 2016) and mongoose (*Mungos mungo*, Fairbanks et al., 2014)), or causing others

62 to avoid them (Behringer et al., 2006; Boillat et al., 2015; Hawley et al., 2020; Kiesecker et al.,

63 1999; Lopes et al., 2022; Poirotte et al., 2017). These feedbacks between social integration and

64 infectious disease are complex in part because the effects of infection on social behavior vary

65 widely across pathogens, hosts, host sex, levels of infection severity, and types of social

66 relationships (reviewed in Hawley et al., 2020). In particular, stable social bonds may be more

67 important and should, therefore, be more resilient to immune challenges or changes in

68 individual behaviors compared to weak social relationships (Poirotte & Charpentier, 2020;

69 Seyfarth & Cheney, 2011; Stockmaier et al., 2020). For example, in vampire bats, sickness

70 behavior reduces individual activity and social grooming among non-kin, but it does not

71 decrease maternal grooming of offspring (Stockmaier et al., 2020). Similarly, mandrills avoid

72 grooming parasite-infected non-kin but continue to groom infected close maternal kin (Poirotte &

73 Charpentier, 2020). While these studies focus on the immediate effect of infections on social

74 behaviors, it remains unclear to what extent infections impact affiliative social bond formation.

75   New social bonds form when individuals join a new social community, which is also

76 when individuals are exposed to new pathogens. If exposure to new pathogens is more likely

77 when encountering new social partners, and if infectious disease causes sickness behavior that

78 limits individual activity and the motivation to interact, then acute infection during the earliest

stages of relationship formation might negatively affect rates of social integration. In a group

with both familiar and new social partners, we therefore predicted that the effect of infection on

social integration should be stronger in new, unfamiliar dyads (pairs) compared to familiar dyads

with well-established relationships. In new dyads of unfamiliar bats, infections should initially

reduce social interaction rates for infected bats; as the infected individuals recover and sickness

behaviors, such as lethargy and social withdrawal, resolve, interaction rates of infected and

asymptomatic bats should then converge over time. In familiar dyads, we expected this

interaction to be weaker because infections should have a smaller impact on how interaction

rates change over time in more established relationships.

To address this hypothesis, we report opportunistic evidence that an acute bacterial

infection reduced social integration in common vampire bats (*Desmodus rotundus*) in a captive

setting. Social integration is observable in this species as strangers transition from no-cost

clustering (huddling for warmth) to low-cost allogrooming (cleaning the fur or wings of other

bats) and to high-cost food sharing (regurgitations of ingested blood to unfed individuals, Carter

et al., 2020). Allogrooming is the best measure of relationship formation because it is a directed

investment of time and energy (unlike clustering) that also occurs frequently enough to track

over time through relationship formation (unlike food sharing). During a previous 4-month study,

we tracked changes in allogrooming among 21 female vampire bats before and after a week of

forced proximity in which small groups of three bats from three distant wild sites were housed

together in captivity for one week (Razik et al., 2022). Shortly after the 21 bats from different

sites were first introduced in captivity, we observed an unplanned outbreak of a bacterial

infection, which we treated with antibiotics (Razik et al., 2022). In this paper, we re-analyzed

published data on relationship formation by including new measures of infection severity to

estimate the social effect of these infections. Specifically, we tracked changes in allogrooming

over time among the nine infected female bats and the 12 asymptomatic female bats,

comparing dyads that were previously familiar or unfamiliar with each other.

Methods

Ethics Statement

109   This work was reviewed and approved by the Smithsonian Tropical Research Institute

110   Animal Care and Use Committee (STRI ACUC no. 2015-0501-2022) and the Panamanian

111   Ministry of the Environment (no. SEX/A-67-2019).

Study subjects

For an experiment testing the effects of forced proximity on relationship formation (Razik

et al., 2022), we captured eight or nine common vampire bats from each of three sites in

Panama that were 120-350 km apart (Figure 1). Upon capture, the 26 bats (23 adult females,

two juvenile males, and one juvenile female) appeared healthy with no pathological lesions. We

housed the bats together in an outdoor flight cage (2.1 × 1.7 × 2.3 m) in Gamboa, Panama, for

approximately four months. Although we caught 23 females, two recaptured females entered

the captive colony several weeks after the colony was created, were not included in all phases

of our experiment, and were therefore not included in this analysis. We therefore analyzed the

allogrooming relationships among 252 new and 126 familiar dyads of 21 adult females. We

define 'dyads' as unique directed combinations of a groomer and receiver. We use the label

'new' for dyads introduced in captivity and 'familiar' for dyads captured from the same site

(hollow tree or cave). Details of the study colony can be found in (Razik et al., 2022; Razik et al.,

2021).

Behavior sampling

We used three infrared surveillance cameras to sample social interactions among bats

individually marked with unique combinations of forearm bands (Razik et al., 2022). Over 114

days, we sampled 682 h of interactions, recording all visible allogrooming and mouth-licking

interactions (≥ 5 seconds in duration) as well as any aggression. For each interaction, we noted

the actor, receiver, and duration. We used allogrooming to assess social integration because

allogrooming was 94% of the 17,592 observed social interactions, while mouth-licking and

aggression were only 6% and 0.3% of interactions, respectively. For each directed edge, we

summed the total seconds of allogrooming from the groomer to the receiver, and we summed

the total sampled seconds that each dyad had the opportunity to interact. For further details on

behavior sampling, see (Razik et al., 2022).

Bacterial infection and treatment

Two weeks after the 26 wild-caught bats were first introduced to one another in the flight

cage, 10 female bats began to show progressive symptoms of a cutaneous infection (blisters or

open lesions with pus, as well as inflamed tissue). Symptoms progressed for approximately two

weeks without signs of recovery. These infections were unexpected and, to our knowledge,

have not previously been described in vampire bat colonies. The 10 infected bats varied in the

severity of their symptoms. The most severe symptoms occurred in bats from a hollow tree near

La Chorrera, Panama. The most severely infected bat developed large, pus-filled lesions on its

back and had extreme inflammation around the wrists that led tissue to break apart and expose

bone; we euthanized this bat before it received antibiotic treatment and did not include it in our

analysis of social integration. Four other bats developed severe symptoms with lesions and

inflammation on one or both wrists, leading to tissue scarring and the inability to sustain flight.

One bat from the same roost experienced only minor inflammation and lesions on both wrists

but healed without permanent tissue scarring and continued to fly. Three bats from a cave near

Lake Bayano, Panama, were also infected. Two bats developed pus-filled abscesses: on the

lower abdomen and upper thigh for one bat, on the lower back for the other bat, and the third

bat developed only minor inflammation on 1-2 toes. Finally, one bat from a hollow tree near

Tolé, Panama, had a large, round lesion on its knee, but this was evident at the start of the

study and did not change over time. This bat displayed no other signs of infection. With most

cases having been observed in bats from either La Chorrera or Lake Bayano, it seems the bats

from Tolé were most resistant to the pathogen.

Swab and tissue samples from skin lesions were cultured to isolate bacterial pathogens,

which indicated the presence of *Bacillus sp.* and *Staphylococcus aureus*. There are several

Bacillus species (e.g., *Bacillus anthracis, Bacillus cereus,* and *Bacillus pumilus*) that may infect

the skin or colonize wounds, but these infections are uncommon and often present with classic

symptoms (e.g., black eschars and necrosis; Cromartie et al., 1947; Esmkhani & Shams, 2022;

Tena et al., 2007; Veysseyre et al., 2015) that we did not observe in our colony. Alternatively,

several species within the *S. aureus* complex, including *Staphylococcus schweitzeri* and

Staphylococcus argenteus, among others (Becker et al., 2019; Chew et al., 2021; Tong et al.,

2015), typically infect the skin and cause symptoms consistent with what we observed. *S.*

aureus complex has also been isolated from the feces of Old-World fruit bats (Fountain et al.,

2022; Held et al., 2017; Monecke et al., 2022; Olatimehin et al., 2018) and the common vampire

bat (Chaverri, 2006; Moreno et al., 1975).

A Kirby-Bauer antibiogram indicated the susceptibility of the isolated bacteria to the

antibiotic Enrofloxacin. Given the urgency and results of the antibiogram, we immediately used

Enrofloxacin antibiotic to treat the infected female bats (described above) over the course of

seven days via daily subcutaneous injections, but we did not perform additional testing to

identify the bacterial species or strain. After the antibiotic treatment of the remaining nine

infected bats (one was euthanized, see above), a subset of five bats recovered without injury

(henceforth "treated"), but the other four bats suffered lasting physical impairments that

rendered them unable to fly for the remainder of the study (henceforth "injured"). We refer to the

remaining 12 bats in our analyses as "asymptomatic." For a detailed timeline of events, see

Figure 1, and for details on experimental bats, see Table S1.

Figure 1

Detailed timeline of events.

Testing for sickness behavior

To confirm that bacterial infections reduced individual activity and social behavior, i.e.,

'sickness behavior', we compared the activity of nine infected, symptomatic bats versus 12

asymptomatic bats (all adult females) during a sample period of 1 week before the antibiotic

treatment (pre-treatment week) and again during 1 week at the end of the study (post-treatment

week) (Figure 1). An observer scored videos using focal sampling for the presence or absence

of three behavioral states: moving, (all movement, e.g., changing location in the cage),

allogrooming (including mouth-licking), and self-grooming, following definitions in (Stockmaier et

al., 2018). The observer could not be entirely blind to the condition of bats in the videos because

lesions were visible, but the analyzed behavioral categories were not subjective. For one hour

(starting one hour after sunset), an observer noted at 30 sec intervals whether a focal female

was engaged in any of the behaviors of interest. Therefore, we sampled each of the 21 bats for

14 days, resulting in 1680 samples per bat (120 per day).

To test if infection predicted the frequency of each behavioral state, we fit Bayesian

multilevel binomial regressions in R (brms package, Bürkner, 2017) and obtained median

posterior parameter estimates and 90% highest posterior density intervals (HPDI), i.e., the most

likely range of parameter values given the model. We used a 90% interval to prevent readers

from interpreting HPDIs as 95% confidence intervals or denoting "statistical significance." For

each behavioral state, model predictors were infection status (asymptomatic vs. infected),

treatment period (pre- vs. post-treatment), the interaction between infection status and period,

and bat identity as a random intercept. The response variable was the proportion of

observations doing a behavior (allogrooming, self-grooming, and moving) out of the total

number of observations of the bat on camera. To evaluate interactions between infection and

treatment period, we estimated marginal means using the *emmeans* package (Lenth &

Piaskowski, 2025). Across models, all parameters had above 1000 effective samples and an R-

hat equal to 1.00 with four chains, indicating successful convergence. Since the effects of acute

bacterial infection on vampire bat behavior have not been observed, we used conservative

default priors across all effects. However, to put our effect sizes in context, we compared them

with effect sizes from past experimental immune challenges, explained below.

Comparison of results to previous experimentally induced sickness behavior

We present our results alongside reference effect sizes from two past experiments on

the behavioral responses of female vampire bats to a simulated bacterial infection (Stockmaier

et al., 2018, 2020). In both experiments, behaviors of vampire bats were observed after being

injected with either lipopolysaccharide (LPS) or saline. Injections of LPS mimic bacterial

infections and cause physiological (e.g., increased white blood cell counts) and behavioral (e.g.,

lethargy, reduced grooming) symptoms in vampire bats (see Stockmaier et al., 2018). In the first

study (Stockmaier et al., 2018), vampire bats were observed in groups of 2-4 bats within small

cages. LPS injection reduced the odds of allogrooming by a factor of 3.73 (binomial mixed effect

model, Odds Ratio, 95% confidence interval (CI) = 2.25–6.50) and reduced the odds of self-

grooming by a factor of 16 (Odds Ratio, 95% CI = 12–27). In the second study (Stockmaier et

al., 2020), vampire bats were fasted to induce food sharing and observed in a larger group (36

bats) in a flight cage. Under these conditions, LPS injection reduced the odds of allogrooming

by a factor of 1.89 (Odds Ratio, 95% CI = 1.84–1.93). This smaller effect on allogrooming could

be due to differences in group size, cage size, the hunger state of the injected bats, or some

combination of those factors. We present reference effects from both studies because the

present study shares some features of both; the bats were unfasted like the first study, but held

in a large group in the same flight cage as the second study.

Effects of infection on symmetry of allogrooming given and received

For the bats that we studied, we expected that any sickness effects on allogrooming

given would cause at least some effects on allogrooming received because allogrooming is

highly reciprocal, and most allogrooming occurs bidirectionally (Carter & Wilkinson, 2013; Imran

Razik et al., 2022; Wilkinson, 1986). To assess whether infection reduced or removed the

symmetry of directed allogrooming, we compared the proportion of hourly allogrooming rates

that were bidirectional for dyads with at least one infected bat versus dyads with two

asymptomatic bats.

Effects of infection and familiarity on dyadic allogrooming rates over time

We assessed how allogrooming changed over time among familiar and new dyads of

bats that were either asymptomatic, infected, or injured. To do this, we fit Bayesian social

relation models in R (STRAND package, Ross et al., 2024). We used an aggregated binomial

outcome to model the proportion of total seconds of observation time with or without

allogrooming for each directed groomer-receiver dyad. Four dyadic covariates included whether

the dyad had been forced into proximity (co-housed in a small cage) during a previous

experiment (Razik et al., 2022, true/false), as well as the main effects and interaction between

familiarity (true/false) and infection severity (a sum of both individuals' infection severity scores:

0 = asymptomatic, 1 = infected, 2 = injured). This measure of infection severity assumes that

these states have linearly increasing and additive effects, but this simplifying assumption is

consistent with comparisons of allogrooming for infected bats versus all others (Fig. 2) and for

injured bats versus all others (Fig. S1). To explore how these effects change through time, we

binned the data by week (15 social networks across 16 weeks, excluding one week when bats

were not sampled altogether), fit the same model described above to each time bin, and

extracted posterior parameter estimates with 90% HPDIs. Model convergence diagnostics can

be found in the online data repository (47).

We predicted a three-way interaction between familiarity, time, and infection severity.

Specifically, we expected that (1) infections would have a greater negative impact on

allogrooming rates in new dyads relative to familiar dyads, and (2) the effect of infection on the

slope of dyadic allogrooming rates over time would be greater for new dyads than for familiar

dyads. Over time, both infection and familiarity effects should converge towards zero because

by the end of the study, new dyads are no longer completely new, and infected bats are no

longer infected.

270   **Results**

Confirming sickness behavior

Consistent with sickness behavior and recovery, infected bats spent less time moving

and allogrooming than asymptomatic bats before antibiotic treatment. These differences faded

to zero over time (Figure 2A, Figure 1C). Before antibiotic treatment, infected bats spent 2.6% of

their time allogrooming, 90% credibility intervals = [1.8, 3.6%], whereas asymptomatic bats

spent 8.4% [6.1, 11.1%] of their time allogrooming. Infected bats spent 12.8% [9.3, 16.9%] of

their time moving, compared to 32.0% [25.9, 38.7%] in asymptomatic bats. After recovery, the

previously infected bats no longer showed differences from the asymptomatic bats in

allogrooming or movement (Figure 2A, Figure 2C). In contrast with effects on allogrooming,

infections did not clearly reduce self-grooming (Figure 2B).

Figure 2

Infections reduced allogrooming and moving, but not self-grooming in vampire bats.

Note. Figure panels show the effects of infections on allogrooming (A), self-grooming (B), and

moving (C). Left-hand panels show median posterior parameter estimates (points) with 90%

highest posterior density intervals (HPDI). Vertical dashed lines show previous effect sizes

estimated from two LPS-induced immune challenge experiments (purple, (Stockmaier et al.,

2018); orange, (Stockmaier et al., 2020)). Right-hand panels show proportions of time for each

behavior (jittered data) for infected bats (red triangles) and asymptomatic bats (black squares)

during the outbreak (pre-treatment) and after bats recovered (post-treatment). Points with error

bars are predicted mean proportions with 90% credible intervals, marginalizing over random

effects.

Effects of infection on symmetry of allogrooming given and received

We found no evidence that infection reduced or removed allogrooming symmetry.

Dyadic allogrooming was bidirectional within the hour in 84% of cases with at least one infected

bat and in 81% of cases with two asymptomatic bats. The observation that allogrooming with an

infected bat was not more likely to be unidirectional than allogrooming among asymptomatic

bats confirms our assumption that any effect of infection on allogrooming given would also

impact allogrooming received.

Effects of infection and familiarity on dyadic allogrooming rates over time

The interaction between familiarity and infection severity changed over time (Figure 3).

Prior to antibiotic treatment (weeks 1 – 6), estimates of the interaction between familiarity and

infection severity were largely negative showing that the effect of infection was greater for new

dyads than for familiar dyads (Figure 3). As expected, this difference then faded during the post-

treatment period (weeks 8 – 16) as introduced bats became more familiar and as the infected

bats recovered (Figure 3). In familiar dyads, infection did not clearly impact dyadic allogrooming

rates over time, whereas in new dyads, there was a clear negative impact of infection severity

on allogrooming rates that diminished with time (Figure 4).

Figure 3.

The effect of infection on allogrooming initially differs between new and familiar dyads, but this

difference faded with time as bats recovered and became familiar.

Note. Y-axis shows the interaction between familiarity and infection severity as predictors of

allogrooming rate for each week (x-axis). Points with error bars show median posterior

parameter estimates with 90% highest posterior density intervals.

Figure 4.

Evidence that infections disproportionately reduced allogrooming in new dyads (developing

social relationships) relative to familiar dyads (existing social relationships).

Note. Y-axis shows the log-transformed (log(x + 1)) mean daily allogrooming rates among all

familiar dyads (left) and new dyads (right) across weeks. Yellow points and solid slopes indicate

dyads with an infection severity of 0-1 (e.g., two asymptomatic bats, or one infected bat),

whereas dark red triangles and dashed slopes indicate dyads with an infection severity of 2-4

(e.g., two infected bats, two injured bats). Shaded regions indicate 95% CI of the slope.

328   **Discussion**

We documented opportunistic evidence that acute symptoms of bacterial infection can

limit the development of allogrooming relationships in vampire bats. Relative to asymptomatic

bats, the initially infected bats spent less time moving and allogrooming. This sickness behavior,

which lasted approximately 2-3 weeks, led to changes in social integration, whereby infections

had a greater impact on the formation of new allogrooming relationships than on allogrooming in

existing relationships. Allogrooming rates were therefore influenced by a three-way interaction

between familiarity, infection severity, and time. Infections had a more negative impact on

allogrooming rates in new dyads relative to familiar dyads, such that the slope of allogrooming

over time was greatest for new dyads with infected and/or injured bats. Familiar dyads and new

dyads without infected bats were less impacted by infections. As predicted, the effects of

infection and familiarity effects, as well as their interaction, converged towards zero as new

dyads became familiar and infected individuals recovered over three months. This convergence

suggests that sickness effects were not entirely driven by injuries that reduced movement.

Because we did not randomly and experimentally infect the bats, a possible alternative

explanation for our findings is the reverse causal path to what we suggested above: perhaps

bats with worse social integration (or worse social competence) had weaker immune systems

and, therefore, suffered worse infections. However, this hypothesis cannot easily explain why

the gap in social integration faded over time after antibiotic treatment. If the infected bats were

already less social or less healthy for reasons besides the infection, then we would expect their

social integration to be worse throughout the study, not just during the outbreak. Also, if social

factors (like dominance or social competence) caused infection severity, then infection rates

should be similar within each wild population. Instead, infection rate and severity were

associated with wild population, suggesting that infections were a cause rather than a

consequence of how bats formed new relationships.

The sickness behavior we observed was similar in some respects to the responses of

vampire bats to simulated infections from injections of lipopolysaccharide, LPS (Stockmaier et

al., 2018, 2020). Using immune challenges like LPS allows for controlled experiments on the

connection between the inflammatory and the behavioral response to infection (Alciatore et al.,

2021; Fishkin & Winslow, 1997; Lopes et al., 2012, 2016; Love et al., 2023; Moreno et al., 2021;

Stockmaier et al., 2018, 2020; Willette et al., 2007), but naturally infecting pathogens can have

different, pathogen-specific effects. For instance, LPS-injected vampire bats significantly

reduced their self-grooming. Still, the naturally infected bats in our study did not, possibly

because the bacterial pathogens caused skin lesions that encouraged self-grooming even when

overall activity was reduced. Another pathogen that infects the skin of bats, the fungus

Pseudogymnoascus destructans (the cause of 'white-nose syndrome') also causes increased

self-grooming (Brownlee-Bouboulis & Reeder, 2013).

These observations are opportunistic and sparse (only 9 infected bats), but we believe

our results are valuable for two main reasons. First, effect size estimates for sickness behaviors

of vertebrates in response to naturally infecting pathogens are uncommon compared to

experiments that use immune challenges (but see Bouwman & Hawley, 2010; Brownlee-

Bouboulis & Reeder, 2013; Cárdenas-Canales et al., 2022; Fairbanks et al., 2015; Kollias et al.,

2004; Langager et al., 2023). Second, while several studies have quantified the effect of

immune challenges (infection or immunogenic antigens) on host social behavior (e.g., Fishkin &

Winslow, 1997; Lopes et al., 2016; Moreno et al., 2021; Stockmaier et al., 2018; Willette et al.,

2007), the effect of short-term infections on the formation and development of long-term social

bonds remains largely unexplored. Our results can inform future experiments on the effect of

infections on social integration and relationship formation.

Our findings also raise several questions. When might animals suppress their infection-

induced behavioral symptoms to continue social integration (Aubert et al., 1997; Langager et al.,

2023; Lopes, 2014; Lopes et al., 2012; Stockmaier et al., 2020; Weil et al., 2006; Willette et al.,

2007)? At what stages of relationship formation do sickness behaviors have the most

substantial effect on social interactions? What is the relationship between the duration or

intensity of sickness behavior and the effects on social integration? To address these questions

experimentally, researchers could use immune challenges at different doses (Dantzer et al.,

1998; Grigoleit et al., 2011; Willette et al., 2007), and/or administer them at varying stages of

relationship formation. Such standardized immune challenges may be better suited for isolating

the effects of sickness on social integration because they minimize confounding variation in

pathogen exposure or host vulnerability. In our case, differences in infection severity among

source populations suggested that susceptibility might have depended on factors such as

immune status or microbiome composition. For instance, site- or region-specific variation in the

skin microbiota (as shown in other bat species, see Avena et. al., 2016) could have influenced

colonization by opportunistic, pathogenic bacteria and the severity of resulting lesions, which, in

turn, could have affected the duration of sickness behavior and opportunities for social

integration.

In conclusion, we found opportunistic observational evidence for reduced social

integration in infected vampire bats after an outbreak of a naturally infecting pathogen. Just as

sickness behavior and other cues of infection can reduce social activity (reviewed in Hart, 1988;

Loehle, 1995; Lopes et al., 2021; Stockmaier et al., 2021) and mate attraction (reviewed in

Beltran-Bech & Richard, 2014), there may be similar effects on the formation of same-sex social

bonds. If there are critical periods of relationship formation, then infections during this time might

have lingering social consequences.

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GC; Resources: IR, GC; Software: IR, GC; Supervision: GC, SS; Validation: IR, GC, SS;

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References

Alciatore, G., Ugelvig, L. V., Frank, E., Bidaux, J., Gal, A., Schmitt, T., Kronauer, D. J., & Ulrich,

Y. (2021). Immune challenges increase network centrality in a queenless ant.

Proceedings of the Royal Society B, *288*(1958), 20211456.

https://doi.org/10.1098/rspb.2021.1456

Almberg, E. S., Cross, P. C., Dobson, A. P., Smith, D. W., Metz, M. C., Stahler, D. R., &

Hudson, P. J. (2015). Social living mitigates the costs of a chronic illness in a

cooperative carnivore. *Ecology Letters*, *18*(7), 660-667.

https://doi.org/10.1111/ele.12444

Aubert, A., Goodall, G., Dantzer, R., & Gheusi, G. (1997). Differential Effects of

Lipopolysaccharide on Pup Retrieving and Nest Building in Lactating Mice. *Brain,*

Behavior, and Immunity, *11*(2), 107-118. https://doi.org/10.1006/brbi.1997.0485

Avena, C.V., Parfrey, L.W., Leff, J.W., Archer, H.M., Frick W.F., Langwig, K.E., Kilpatrick, A.M.,

Powers, K.E., Foster, J.T., & McKenzie, V.J. (2016). Deconstructing the bat skin

microbiome: influences of the host and the environment. Frontiers in Microbiology, 7,

1753. https://doi.org/10.3389/fmicb.2016.01753

Becker, K., Schaumburg, F., Kearns, A., Larsen, A. R., Lindsay, J. A., Skov, R. L., & Westh, H.

(2019). Implications of identifying the recently defined members of the Staphylococcus

aureus complex *S. argenteus* and *S. schweitzeri*: A position paper of members of the

ESCMID Study Group for Staphylococci and Staphylococcal Diseases (ESGS). *Clinical*

Microbiology and Infection: The Official Publication of the European Society of Clinical

Microbiology and Infectious Diseases, *25*(9), 1064–1070.

https://doi.org/10.1016/j.cmi.2019.02.028

Behringer, D. C., Butler, M. J., & Shields, J. D. (2006). Ecology: Avoidance of disease by social

lobsters. *Nature*, *441*(7092), 421. https://doi.org/10.1038/441421a

Beltran-Bech, S., & Richard, F.-J. (2014). Impact of infection on mate choice. *Animal Behaviour*,

90, 159–170. https://doi.org/10.1016/j.anbehav.2014.01.026

Boillat, M., Challet, L., Rossier, D., Kan, C., Carleton, A., & Rodriguez, I. (2015). The

vomeronasal system mediates sick conspecific avoidance. *Current Biology*, *25*(2), 251-

255. https://doi.org/10.1016/j.cub.2014.11.061

Bouwman, K. M., & Hawley, D. M. (2010). Sickness behaviour acting as an evolutionary trap?

Male house finches preferentially feed near diseased conspecifics. *Biology Letters*, *6*(4),

462-465. https://doi.org/10.1098/rsbl.2010.0020

Brownlee-Bouboulis, S. A., & Reeder, D. M. (2013). White-nose syndrome-affected little brown

myotis (*Myotis lucifugus*) increase grooming and other active behaviors during arousals

from hibernation. *Journal of Wildlife Diseases*, *49*(4), 850-859.

https://doi.org/10.7589/2012-10-242

Bürkner, P.C. (2017). brms: An R Package for Bayesian Multilevel Models Using Stan. *Journal*

of Statistical Software, *80*, 1–28. https://doi.org/10.18637/jss.v080.i01

Cárdenas-Canales, E. M., Stockmaier, S., Cronin, E., Rocke, T. E., Osorio, J. E., & Carter, G.

G. (2022). Social effects of rabies infection in male vampire bats (*Desmodus rotundus*).

Biology Letters, *18*(9), 20220298. https://doi.org/10.1098/rsbl.2022.0298

Carter, G. G., Farine, D. R., Crisp, R. J., Vrtilek, J. K., Ripperger, S. P., & Page, R. A. (2020).

Development of New Food-Sharing Relationships in Vampire Bats. *Current Biology*,

30(7), 1275-1279. https://doi.org/10.1016/j.cub.2020.01.055

Carter, G. G., & Wilkinson, G. S. (2013). Food sharing in vampire bats: Reciprocal help predicts

donations more than relatedness or harassment. *Proceedings of the Royal Society B*,

280(1753), 20122573. https://doi.org/10.1098/rspb.2012.2573

Chaverri, G. (2006). Aerobic bacterial flora from the digestive tract of the common vampire bat,

Desmodus rotundus (Chiroptera: *Phyllostomidae*). *Revista De Biologia Tropical*, *54*(3),

717–724.

Chew, K. L., Octavia, S., Lai, D., Lin, R. T. P., & Teo, J. W. P. (2021). *Staphylococcus*

singaporensis sp. Nov., a new member of the *Staphylococcus aureus* complex, isolated

from human clinical specimens. *International Journal of Systematic and Evolutionary*

Microbiology, *71*(10). 005067. https://doi.org/10.1099/ijsem.0.005067

Cromartie, W. J., Bloom, W. L., & Watson, D. W. (1947). Studies on infection with *Bacillus*

anthracis; a histopathological study of skin lesions produced by *B. anthracis* in

susceptible and resistant animal species. *The Journal of Infectious Diseases*, *80*(1), 1–

13. https://doi.org/10.1093/infdis/80.1.1

Dantzer, R., Bluthé, R.-M., Gheusi, G., Cremona, S., Layé, S., Parnet, P., & Kelley, K. W.

(1998). Molecular basis of sickness behaviors. *Annals of the New York Academy of*

Sciences, *856*(1), 132–138. https://doi.org/10.1111/j.1749-6632.1998.tb08321.x

Esmkhani, M., & Shams, S. (2022). Cutaneous infection due to *Bacillus cereus*: A case report.

BMC Infectious Diseases, *22*(1), 393. https://doi.org/10.1186/s12879-022-07372-9

Fairbanks, B. M., Hawley, D. M., & Alexander, K. A. (2014). The impact of health status on

dispersal behavior in banded mongooses (*Mungos mungo*). *EcoHealth*, *11*(2), 258-262.

https://doi.org/10.1007/s10393-014-0912-4

Fairbanks, B. M., Hawley, D. M., & Alexander, K. A. (2015). No evidence for avoidance of visibly

diseased conspecifics in the highly social banded mongoose (*Mungos mungo*).

Behavioral Ecology and Sociobiology, *69*(3), 371-381. https://doi.org/10.1007/s00265-

014-1849-x

Fishkin, R. J., & Winslow, J. T. (1997). Endotoxin-induced reduction of social investigation by

mice: Interaction with amphetamine and anti-inflammatory drugs. *Psychopharmacology*,

132(4), 335-341. https://doi.org/10.1007/s002130050353

Fountain, K., Barbon, A., Gibbon, M. J., Lloyd, D. H., Loeffler, A., & Feil, E. J. (2022).

Staphylococcus aureus lineages associated with a free-ranging population of the fruit bat

Pteropus livingstonii retained over 25 years in captivity. *Scientific Reports*, *12*(1), 13457.

https://doi.org/10.1038/s41598-022-17835-3

Grigoleit, J.-S., Kullmann, J. S., Wolf, O. T., Hammes, F., Wegner, A., Jablonowski, S., Engler,

H., Gizewski, E., Oberbeck, R., & Schedlowski, M. (2011). Dose-dependent effects of

endotoxin on neurobehavioral functions in humans. *PloS One*, *6*(12), e28330.

https://doi.org/10.1371/journal.pone.0028330

Hart, B. L. (1988). Biological basis of the behavior of sick animals. *Neuroscience &*

Biobehavioral Reviews, *12*(2), 123-137. https://doi.org/10.1016/S0149-7634(88)80004-6

Hawley, D. M., Gibson, A. K., Townsend, A. K., Craft, M. E., & Stephenson, J. F. (2020).

    Bidirectional interactions between host social behaviour and parasites arise through

    ecological and evolutionary processes. *Parasitology*, 1–70.

    https://doi.org/10.1017/S0031182020002048

Held, J., Gmeiner, M., Mordmüller, B., Matsiégui, P.-B., Schaer, J., Eckerle, I., Weber, N.,

    Matuschewski, K., Bletz, S., & Schaumburg, F. (2017). Bats are rare reservoirs of

    Staphylococcus aureus complex in Gabon. *Infection, Genetics and Evolution: Journal of*

    Molecular Epidemiology and Evolutionary Genetics in Infectious Diseases, *47*, 118–120.

    https://doi.org/10.1016/j.meegid.2016.11.022

Jog, M. G., Sackett, M. E., Kisty, S. D., Hansen, J. A., & Stephenson, J. F. (2022). The

    behaviour of infected guppies depends on social context, parasite tolerance and host

    sex. *Animal Behaviour*, *187*, 97–104. https://doi.org/10.1016/j.anbehav.2022.03.001

Kiesecker, J. M., Skelly, D. K., Beard, K. H., & Preisser, E. (1999). Behavioral reduction of

    infection risk. *Proceedings of the National Academy of Sciences, 96*(16), 9165-9168.

    https://doi.org/10.1073/pnas.96.16.9165

Kollias, G. V., Sydenstricker, K. V., Kollias, H. W., Ley, D. H., Hosseini, P. R., Connolly, V., &

    Dhondt, A. A. (2004). Experimental infection of house finches with *Mycoplasma*

    gallisepticum. *Journal of Wildlife Diseases*, *40*(1), 79-86. https://doi.org/10.7589/0090-

    3558-40.1.79

Langager, M. M., Adelman, J. S., & Hawley, D. M. (2023). Let's stick together: Infection

    enhances preferences for social grouping in a songbird species. *Ecology and Evolution*,

    13(10), e10627. https://doi.org/10.1002/ece3.10627

Lenth, R., & Piaskowski J. (2025). emmeans: Estimated Marginal means, aka Least-Squares

    Means. R package version 2.0.1, https://rvlenth.github.io/emmeans/

Loehle, C. (1995). Social barriers to pathogen transmission in wild animal populations. *Ecology*,

    76(2), 326-335. https://www.jstor.org/stable/1941192

Lopes, P. C. (2014). When is it socially acceptable to feel sick? *Proceedings of the Royal Society B*, *281*(1788), 20140218. https://doi.org/10.1098/rspb.2014.0218

Lopes, P. C., Adelman, J., Wingfield, J. C., & Bentley, G. E. (2012). Social context modulates sickness behavior. *Behavioral Ecology and Sociobiology*, *66*(10), 1421-1428. https://doi.org/10.1007/s00265-012-1397-1

Lopes, P. C., Block, P., & Konig, B. (2016). Infection-induced behavioural changes reduce connectivity and the potential for disease spread in wild mice contact networks. *Scientific Reports*, *6*, 31790. https://doi.org/10.1038/srep31790

Lopes, P. C., French, S. S., Woodhams, D. C., & Binning, S. A. (2021). Sickness behaviors across vertebrate taxa: Proximate and ultimate mechanisms. *Journal of Experimental Biology*, *224*(9), jeb225847. https://doi.org/10.1242/jeb.225847

Lopes, P. C., Woodhams, D. C., & Binning, S. A. (2022). Infection avoidance behaviors across vertebrate taxa: Patterns, processes and future directions. In V. Ezenwa, S. Altizer, R. Hall (Eds.), *Animal Behavior and Parasitism* (pp. 237-255). Oxford University Press.

Love, A. C., Anthony, A. C., Nash, A., Campos-Melara, A., Kodali, J., & DuRant, S. E. (2023). Simulated infection alters the behavior of pair bonded songbirds and their healthy neighbors. *Behavioral Ecology*, *34*(2), 251-260. https://doi.org/10.1093/beheco/arac120

Monecke, S., Schaumburg, F., Shittu, A. O., Schwarz, S., Mühldorfer, K., Brandt, C., Braun, S. D., Collatz, M., Diezel, C., Gawlik, D., Hanke, D., Hotzel, H., Müller, E., Reinicke, M., Feßler, A. T., & Ehricht, R. (2022). Description of Staphylococcal strains from Straw-Coloured Fruit Bat (*Eidolon helvum*) and Diamond Firetail (*Stagonopleura guttata*) and a review of their phylogenetic relationships to other *Staphylococci*. *Frontiers in Cellular and Infection Microbiology*, *12*, 878137. https://doi.org/10.3389/fcimb.2022.878137

Moreno, G., Lopes, C. A., Seabra, E., Pavan, C., & Corrêa, A. (1975). Bacteriological study of the intestinal flora of bats (*Desmodus rotundus*) (author's transl). *Arquivos Do Instituto Biologico*, *42*, 229–232.

Moreno, K. R., Weinberg, M., Harten, L., Salinas Ramos, V. B., Herrera M., L. G., Czirják, G. Á.,

& Yovel, Y. (2021). Sick bats stay home alone: fruit bats practice social distancing when

faced with an immunological challenge. *Annals of the New York Academy of Sciences*,

1505(1), 178-190. https://doi.org/10.1111/nyas.14600

Olatimehin, A., Shittu, A. O., Onwugamba, F. C., Mellmann, A., Becker, K., & Schaumburg, F.

(2018). *Staphylococcus aureus* complex in the Straw-Colored Fruit Bat (*Eidolon helvum*)

in Nigeria. *Frontiers in Microbiology*, *9*, 162. https://doi.org/10.3389/fmicb.2018.00162

Pavez-Fox, M. A., Negron-Del Valle, J. E., Thompson, I. J., Walker, C. S., Bauman, S. E.,

Gonzalez, O., Compo, N., Ruiz-Lambides, A., Martinez, M. I., Platt, M. L., Montague, M.

J., Higham, J. P., Snyder-Mackler, N., & Brent, L. J. N. (2021). Sociality predicts

individual variation in the immunity of free-ranging rhesus macaques. *Physiology &*

Behavior, *241*, 113560. https://doi.org/10.1016/j.physbeh.2021.113560

Poirotte, C., & Charpentier, M. J. E. (2020). Unconditional care from close maternal kin in the

face of parasites. *Biology Letters*, *16*(2), 20190869.

https://doi.org/10.1098/rsbl.2019.0869

Poirotte, C., Massol, F., Herbert, A., Willaume, E., Bomo, P. M., Kappeler, P. M., & Charpentier,

M. J. E. (2017). Mandrills use olfaction to socially avoid parasitized conspecifics.

Science Advances, *3*(4), e1601721. https://doi.org/10.1126/sciadv.1601721

Razik, I., Brown, B. K. G., Page, R. A., & Carter, G. G. (2021). Non-kin adoption in the common

vampire bat. *Royal Society Open Science*, *8*(2), 201927.

https://doi.org/10.1098/rsos.201927

Razik, I., Brown, B. K. G., & Carter, G. G. (2022). Forced proximity promotes the formation of

enduring cooperative relationships in vampire bats. *Biology Letters*, *18*(4), 20220056.

https://doi.org/10.1098/rsbl.2022.0056

Razik, I., Carter, G. G., Stockmaier, S., & Abou-Elias, M. (2023). *Data and R code for*

Opportunistic evidence of the impact of bacterial infections on social integration in

vampire bats [Dataset]. Link: https://github.com/imranrazik/infection_vampires

Ross, C. T., McElreath, R., & Redhead, D. (2024). Modelling animal network data in R using

STRAND. *Journal of Animal Ecology*, *93*(3), 254–266. https://doi.org/10.1111/1365-

2656.14021

Rushmore, J., Bisanzio D., Gillespie, T. R. (2017). Making new connections: insights from

primate-parasite networks. *Trends in Parasitology*, *33*(7), 547–560.

https://doi.org/10.1016/j.pt.2017.01.013

Seyfarth, R. M., & Cheney, D. L. (2011). The evolutionary origins of friendship. *Annual Review*

of Psychology, *63*(1), 153-177. https://doi.org/10.1146/annurev-psych-120710-100337

Snyder-Mackler, N., Burger, J. R., Gaydosh, L., Belsky, D. W., Noppert, G. A., Campos, F. A.,

Bartolomucci, A., Yang, Y. C., Aiello, A. E., O'Rand, A., Harris, K. M., Shively, C. A.,

Alberts, S. C., & Tung, J. (2020). Social determinants of health and survival in humans

and other animals. *Science*, *368*(6493), eaax9553.

https://doi.org/10.1126/science.aax9553

Snyder-Mackler, N., Sanz, J., Kohn, J. N., Brinkworth, J. F., Morrow, S., Shaver, A. O., Grenier,

J.-C., Pique-Regi, R., Johnson, Z. P., Wilson, M. E., Barreiro, L. B., & Tung, J. (2016).

Social status alters immune regulation and response to infection in macaques. *Science*,

354(6315), 1041-1045. https://doi.org/10.1126/science.aah3580

Stockmaier, S., Bolnick, D. I., Page, R. A., & Carter, G. G. (2018). An immune challenge

reduces social grooming in vampire bats. *Animal Behaviour*, *140*, 141–149.

https://doi.org/10.1016/j.anbehav.2018.04.021

Stockmaier, S., Bolnick, D. I., Page, R. A., & Carter, G. G. (2020). Sickness effects on social

interactions depend on the type of behaviour and relationship. *Journal of Animal*

Ecology, *89*(6), 1387-1394. https://doi.org/10.1111/1365-2656.13193

Stockmaier, S., Stroeymeyt, N., Shattuck, E. C., Hawley, D. M., Ancel Meyers, L., & Bolnick, D.

I. (2021). Infectious diseases and social distancing in nature. *Science*, *371*(6533),

eabc8881. https://doi.org/10.1126/science.abc8881

Stockmaier, S., Ulrich, Y., Albery, G. F., Cremer, S., & Lopes, P. C. (2023). Behavioural

defences against parasites across host social structures. *Functional Ecology*, *37*, 809-

820. https://doi.org/10.1111/1365-2435.14310

Tena, D., Martinez-Torres, J. A., Perez-Pomata, M. T., Sáez-Nieto, J. A., Rubio, V., & Bisquert,

J. (2007). Cutaneous infection due to *Bacillus pumilus*: Report of 3 cases. *Clinical*

Infectious Diseases: An Official Publication of the Infectious Diseases Society of

America, *44*(4), e40-42. https://doi.org/10.1086/511077

Tong, S. Y. C., Schaumburg, F., Ellington, M. J., Corander, J., Pichon, B., Leendertz, F.,

Bentley, S. D., Parkhill, J., Holt, D. C., Peters, G., & Giffard, P. M. (2015). Novel

staphylococcal species that form part of a *Staphylococcus aureus*-related complex: The

non-pigmented *Staphylococcus argenteus* sp. nov. and the non-human primate-

associated *Staphylococcus schweitzeri* sp. nov. *International Journal of Systematic and*

Evolutionary Microbiology, *65*(1), 15–22. https://doi.org/10.1099/ijs.0.062752-0

Tung, J., Barreiro, L. B., Johnson, Z. P., Hansen, K. D., Michopoulos, V., Toufexis, D., Michelini,

K., Wilson, M. E., & Gilad, Y. (2012). Social environment is associated with gene

regulatory variation in the rhesus macaque immune system. *Proceedings of the National*

Academy of Sciences, *109*(17), 6490-6495. https://doi.org/10.1073/pnas.1202734109

Veysseyre, F., Fourcade, C., Lavigne, J.-P., & Sotto, A. (2015). *Bacillus cereus* infection:

57 case patients and a literature review. *Medecine Et Maladies Infectieuses*, *45*(11–12),

436–440. https://doi.org/10.1016/j.medmal.2015.09.011

Weil, Z. M., Bowers, S. L., Dow, E. R., & Nelson, R. J. (2006). Maternal aggression persists

following lipopolysaccharide-induced activation of the immune system. *Physiology &*

Behavior, *87*(4), 694-699. https://doi.org/10.1016/j.physbeh.2006.01.005

Wilkinson, G. S. (1986). Social grooming in the common vampire bat, *Desmodus rotundus*.

Animal Behaviour, *34*(6), 1886-1889. http://dx.doi.org/10.1016/S0003-3472(86)80274-3

Willette, A. A., Lubach, G. R., & Coe, C. L. (2007). Environmental context differentially affects

behavioral, leukocyte, cortisol, and interleukin-6 responses to low doses of endotoxin in

the rhesus monkey. *Brain Behavior & Immunity*, *21*(6), 807-815.

https://doi.org/10.1016/j.bbi.2007.01.007