The genomic landscape of adaptation to a new host plant

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Abstract

Adaptation to novel ecological niches is known to be rapid. However, how ecological divergence

translates into reproductive isolation remains a consequential question in speciation research. It is

still unclear how coupling of ecological divergence loci and reproductive isolation loci occurs at

the genomic level, ultimately enabling the formation of persistent species. Here, we investigate the

genomic underpinning colonization of a new niche and formation of two host races in *Tephritis*

conura peacock flies that persist in sympatry. To uncover the genomic differences underlying host

plant adaptation, we take advantage of two independent sympatric zones, where host plant

specialists using the thistle species *Cirsium heterophyllum* and *C. oleraceum* co-occur, and address

what regions of the genome that diverge between the host races in a parallel fashion. Using

Population Branch Statistics, dxy and BayPass we identify 2996 and 3107 outlier regions

associated with host use among western and eastern Baltic populations, respectively, with 82%

overlap. The majority of outliers are located within putative inversions, adding to the growing

body of evidence that structural changes to the genome are important for adaptations to persist in

face of gene flow. Potentially, the high contents of repetitive elements could have given rise to the

inversions, but this remains to be tested. The outlier regions were enriched for genes involved in

e.g. metabolism and morphogenesis, potentially due to the selection for processing different

metabolites, and changes in size and ovipositor length respectively. In conclusion, this study

suggests that structural changes in the genome, and divergence in independent ecological functions

may facilitate the formation of persistent host races in face of gene flow.

Introduction

How biological variation and novel adaptation arise are fundamental and unresolved questions in our understanding of the origins of biodiversity. Ecological adaptation to novel environments and food sources are known to result in rapid phenotypic and genetic divergence (Bush 1969; Boag and Grant 1981; Feder et al. 1988; Filchak et al. 2000; Herrel et al. 2008; Nosil 2012), but to what extent this ecological adaptation leads to persistent speciation is contested (Schluter 2009; Anderson and Weir 2022; Anderson et al. 2023). Both the ecological context, including the consistency (Hendry 2009; Bolnick 2011) and multidimensionality of the divergent niches (Rice and Hostert 1993; Nosil and Sandoval 2008; Nosil et al. 2009; Chevin et al. 2014), and the genomic underpinnings of adaptation to those niches (Chevin et al. 2014) may influence the likelihood of divergent lineages becoming distinct species over time. Whether ecological adaptations and reproductive isolation are linked, at either the phenotypic or genomic scale, is important to the speciation process. Without reproductive isolation, ecological shifts can lead to collapses of formerly isolated population, resulting in increased gene flow and breakdown of the speciation process (Gow et al. 2006; Taylor et al. 2006; Lackey and Boughman 2017). Thus, a challenge for evolutionary biologists is to identify the genomic architecture that enables persistent species to form (Kulmuni et al. 2020).

One way to uncover the genomic architecture that enables speciation is to assess sequence differences that are robust to gene flow between populations in sympatry. Gene flow can be limited in certain regions of the genome by the presence of genes that are involved in reproductive isolation, and differences in these regions are likely contributing to the speciation process (Schluter

and Rieseberg 2022). However, interpreting which of the differences between existing species

caused speciation may be challenging, as additional genes involved in reproductive isolation may

have accumulated since the species diverged. Investigating the genomic differences between

recently diverged taxa and focusing on independent contact zones can help resolve this challenge

(Schluter and Rieseberg 2022). Regions that repeatedly differ among differentially-adapted taxa

are likely to reflect regions that have played a role in speciation (Bohutínská et al. 2021). Extensive

theoretical and empirical evidence has shown that genomic architecture coupling coadapted loci,

including inversions, is particularly important for persistence of differential adaptation in the face

of gene flow (Feder et al. 2003; Wellenreuther and Bernatchez 2018; Berdan et al. 2022; Schaal et

al. 2022). Yet, the extent to which linkage of ecological adaptation and reproductive isolation loci

is facilitated by pleiotropic effects of the same genes, coupling of genes through structural

variation, or co-inheritance of uncoupled genomic regions upheld by correlational selection is not

well understood.

Herbivorous insects and the plants they interact with are excellent models for studying the genomic

basis of speciation. These are two of the most speciose groups of eukaryotes (Mora et al. 2011;

Christenhusz and Byng 2016), and specialization of phytophagous insects and their hosts is

hypothesized to drive this divergence (Vidal and Murphy 2018). The specificity and

multidimensionality of the niches constituted by the host plants (Hardy and Otto 2014) and strong

dependency on host plants for reproduction and survival (e.g., assortative mating based on host

recognition; Bush 1969) may increase the chance of ecological speciation in herbivorous insects

(Hardy and Otto 2014). These features make phytophagous insects effective systems for

understanding the genomic basis of ecological adaptation. Changes in host repertoire, either

through host range expansion or host shifts, have previously been associated with chemosensory

genes underlying butterfly host choice (van Schooten et al. 2020), alter oral secretions in aphids

(Boulain et al. 2019; Shih et al. 2023), and code for enzymes in the digestive systems of both

butterflies and aphids (Nallu et al. 2018; Singh et al. 2020; Shih et al. 2023), and genes underlying

phenological shifts in *Rhagoletis* flies (Feder et al. 1988; Feder et al. 2003). Ecological divergence

resulting in reduced gene flow among sympatric or parapatric populations of herbivorous insects

causes in the formation of host races (Berlocher and Feder 2002). However, how the genetic

changes underlying ecological adaptation are coupled to or translated into reproductive isolation

at the genomic level, enabling co-existence and long term persistence of differentially adaptive

host races, remains a challenge to resolve. One possibility is that the rate at which differentially

adapted host races accumulate the genetic architecture enabling coupling or strong reproductive

isolation could determine the time required for stable co-existence. This is the case in birds, where

sister taxa with inversions co-exist at shorter divergence times (Hooper et al. 2019).

Here, we use the Tephritid fly *Tephritis conura* to uncover the genomic basis of ecological

divergence and reproductive isolation promoting stable coexistence of two differentially adapted

host races. These host races infest the ancestral host thistle *Cirsium heterophyllum* and the derived

host thistle *C. oleraceum*, respectively. They differ in phenology and host plant preference

(Romstock-Volkl 1997), and have host-specific larval performance and survival (Diegisser et al.

2008) resulting in relatively strong reproductive isolation (Seitz and Komma 1984; Romstock-

Volkl 1997; Diegisser et al. 2006a; Diegisser et al. 2006b). Moreover, mitochondrial networks and

allozyme markers suggest they are genetically differentiated (Diegisser et al. 2006a; Diegisser et

al. 2006b). Taken together, these findings suggest that there is strong divergent ecological selection

acting on these host races. This study system also exhibits independent contact zones where the

host races are found in sympatry both west and east of the Baltic Sea (Nilsson et al. 2022), enabling

us to determine the genomic regions that differ between host races in parallel, and hence are likely

to confer ecological adaptation. Specifically, we address which genomic regions are involved in

the host shift, whether structural variation is important, and to what extent these regions are

resistant to gene flow between the host races.

Methods

The study species *T. conura* (Loew, 1844; Diptera) is a true fruit fly in the family Tephritidae.

Most species in this family are specialists on one or a few plants (Aluja and Norrbom 2001) where

larvae feed on fruits, seeds, flowers, stems or plant roots (Christenson and Foote 1960; Headrick

and Goeden 1998). Within Tephritidae, several examples of host races occur in the genera *Eurosta*

(Craig et al. 1993), *Rhagoletis* (Bush 1969; Feder et al. 1998) and *Tephritis* (Diegisser et al. 2004;

Diegisser et al. 2006b). Within *T. conura*, host plant specialization has been documented in

populations in continental Europe, following a host shift from *C. heterophyllum* (referred to as

CH-flies from here on) to the derived host plant *C. oleraceum* (referred to as CO-flies) (Romstock-

Volkl 1997). The host races are nearly indistinguishable morphologically, with ovipositor length

being the only trait that enables classification in isolation (Diegisser et al. 2007; Nilsson et al.

2022). Sampling adult flies on their host plants or as larvae in infested buds also facilitates

discrimination.

In order to uncover the genomic regions involved in host plant adaptation in *T. conura*, we

assembled and annotated a reference genome and used whole genome resequencing of four

populations from each host race. For both host races, *T. conura* larvae inside infested *Cirsium* buds

of the respective host plants were collected during June and July 2018. We sampled infested buds

from eight populations, four for each host race, including sites in allopatric populations of *C.*

oleraceum in Germany and Lithuania, allopatric populations of *C. heterophyllum* in northern

Sweden and Finland, and sympatric populations of both host races in southern Sweden and Estonia

(Fig. 1; Table S1). This design enables us to identify the genomic regions that consistently differ

between host races, as the sympatric regions sampled on each side of the Baltic Sea constitute

independent contact zones. The pupae used for the reference genome was sampled from a CH-fly

individual from the CHST population (Fig. 1D).

Thistle buds containing *T. conura* larvae were stored individually in netted cups at 21°C in a

laboratory at the Department of Biology, Lund University, Sweden. Emerging adult flies were

provided with honey water (1:2 ratio) *ad libitum* smeared on top of the netting. Three days after

the first emergence, representing a sufficient interval for all flies to eclose, cups containing adult

flies were relocated to a climate chamber (7°C; 8:16 light:dark cycle). We considered all flies

emerging from a given bud as potential siblings and therefore sampled only a single adult male

from each thistle bud for genomic analysis to avoid including related individuals. Flies and the

pupae were euthanized through snap freezing and stored at -80°C.

Genome assembly and improvement

DNA from a single *T. conura* pupa harvested from a *C. heterophyllum* bud using was extracted by

SciLifeLabs, Solna, Sweden following their in-house protocol for PacBio sequencing. Briefly, the

DNA was sequenced using PacBio long read technology and sequences were error-corrected using

SMRT Link analysis. Sequencing resulted in 4,173,684 reads with a total of 74.3Gbp and an N50

read length of 17.8 kb. PacBio sequences were assembled with hifiasm 0.7-dirty-r255 (Cheng et

al. 2021), which was run using default parameters, followed by purge_dups (v. 1.2.5; (v. 1.2.5;

Guan et al. 2020) to identify and remove heterozygous duplication (e.g. haplotigs) in the resulting

assemblies.

The reference assembly was scanned with Kraken2 to identify bacterial, viral and fungal

sequences, and contigs with a contamination proportion of greater than 10% were removed (150

contigs; 47,038,732 bp). Due to the abundance of short contigs in our assembly, we used a close

relative, *Rhagoletis pomonella*, to achieve a chromosome-level perspective. We aligned the *T.*

conura contigs to the *R. pomonella* assembly (GCF_013731165.1_Rhpom_1.0; accessed 22 Nov

2022; length = 1223 Mbp, scaffolds = 32,060, scaffold N50 = 72Mbp) using minimap2 (asm20;

Li et al. 2021) within the RagTag wrapper (v. 2.1.0; Alonge et al. 2022). RagTag ordered *T. conura*

contigs based on *R. pomonella* scaffolds. This order was used in downstream analyses to cluster

T. conura contigs within hypothetical linkage groups. Putative sex-linked contigs were identified

as those with an excess of hits (at least one hit per million bp, approx. top 5% of contigs) when

blasted (tblastn; v.2.11.0; Camacho et al. 2009) with a set of *Drosophila melanogaster* X-linked

proteins. This resulted in 102 putative X-enriched contigs, 68.6% of which were clustered on two

RagTag linkage groups: NW_023458556.1_RagTag (n = 56) and NW_023458567.1_RagTag (n

= 14; Fig. S1). The genome was sequenced from a pupa of unknown sex, but coverage of both

long and short reads mapped across these contigs suggest that the pupa was male, as *Tephritis* is

one of the only fly genera in which females are heterogametic (Vicoso and Bachtrog 2015).

We used RepeatModeler (v. 2.0.3; Flynn et al. 2020) and RepeatMasker (v. 4.1.2; Smit et al. 2015)

to identify and soft mask repetitive elements in the *T. conura* reference assembly. For

RepeatModeler, we specified the 'rmblast' engine and ran the LTR structural pipeline. We ran the

-gccalc option in RepeatMasker to account for variable GC content. To put the *T. conura* results

in context we accessed repeat summaries for nine Tephritinae species with published genomes

(Sproul et al. 2022) While Sproul et al. (2022) performed additional repeat screens, we used the

output from their first round of masking, in which they used RepeatModeler2.0 (search engine

"ncbi") and RepeatMasker4.1.0 to generate custom repeat libraries.

Gene prediction and annotation

We annotated the assembly using the MAKER pipeline (3.01.03; Cantarel et al. 2008). We

generated expressed sequence predictions using RNAseq reads from flash-frozen flies from

various life stages, sexes and populations (Table S2). Extractions were performed using Sigma

Aldrich's Plant RNA kit, which we have found performs best with these flies. After sample quality

testing, Illumina TruSeq Stranded mRNA libraries were prepared by SciLifeLab (Stockholm,

Sweden, https://ngisweden.scilifelab.se) and sequenced using NovaSeq6000. We used two tools

to generate EST predictions from this RNAseq evidence. First, we mapped the reads to the soft-

masked reference genome using the splice aware mapper HISAT2 (v. 2.2.1; Kim et al. 2019).

Mapped reads were sorted and indexed with SAMtools (v. 1.14; Danecek et al. 2021). Transcripts

were predicted and merged into a single annotation with StringTie (v. 2.1.4; Kovaka et al. 2019).

We also generated a *de novo* transcriptome prediction with Trinity (v. 2.11.0; Grabherr et al. 2011).

Protein evidence was downloaded from Uniprot database (UniProt Consortium 2021; accessed

June 2021) under the taxonomy"Acalyptratae 99 [43741]" and only reviewed homologs were used.

Protein and EST predictions were synthesized into a final annotation with 27588 transcripts. The

final annotation was assessed using BUSCO and the diptera_odb10 database, which found 84.6%

completeness (82.9% single copy, 1.7% duplicated), 3.7% fragmented and 11.7% missing. We

functionally annotated the predicted genes with eggNOG emmaper (v. 2.1.5; Huerta-Cepas et al.

2018) using default settings. Functional annotations were returned for 14812 transcripts, including

gene ontology (GO) terms for 9894 of these.

DNA extraction and sequencing

We extracted DNA for whole genome sequencing from entire flies using Qiagen DNeasy Blood

and Tissue Kit (Qiagen Corp., Valencia, CA), following the standard protocol with some small

modifications (Supplementary Methods 1). Frozen flies were briefly thawed and ground with

plastic pestles in the tissue lysis buffer. Samples were incubated with proteinase K (3.5h, 56°C),

and then RNAse was added and the pellet eluted using a 56μl of EB buffer. Library preparation

and whole genome resequencing were performed by SciLifeLab (NGI-Sweden, Solna,Sweden)

using Illumina TruSeq PCR-free prep kit with an insert size of 350bp. Sequencing of 2x150 bp

paired-end reads was carried out on 8 Illumina HiSeqX lanes.

Mapping, variant calling and allele frequencies

We aligned the whole genome sequencing data of all individuals to the reference genome using

bwa mem (v.7.17-r1188; Li and Durbin 2009), then merged, sorted and indexed using samtools

(v. 1.14; Li et al. 2009). PCR duplicates were identified and removed using MarkDuplicates in

Picard tools (v2.10.3; Broad Institute). Genome-wide mean coverage was calculated with samtools

(v 1.9; Fig. S2).

To account for low coverage, we utilized genotype likelihood based methods for analyses when

possible. To accommodate both genotype likelihood and genotype call based analyses, we

produced both (1) a Beagle file containing genotype likelihoods for the 96 *T. conura* individuals

and (2) a VCF containing genotype calls for the 96 individuals as well as three outgroup individuals

using ANGSD (v. 0.940; Korneliussen et al. 2014). The filters and steps to produce these two files

are described in Supplementary Methods 2. The final Beagle file and VCF contained 18,722,827

and 53,722,613 SNPs, respectively.

Per population, we generated site allele frequency indices (SAF) and inferred minor allele

frequencies (MAFs) using ANGSD, relying on the same filters as described for the Beagle file,

excluding those used for SNP calling (Supplementary Methods 2). We produced a consensus

FASTA from the mapped reads of one of the outgroup individuals (P18705_115, *Tephritis*

kogardtauica) using ANGSD and polarized the SAF files to this sequence (-anc outgroup.fa). As

this FASTA is produced from mapped reads of an outgroup individual, many positions did not

have sufficient coverage to be called in the FASTA. As such, the SAFs only estimated frequencies

at positions that both passed filters set during SAF production and had a non-ambiguous base call

in the outgroup FASTA file. We produced unfolded 1D and 2D site frequency spectra (SFS) for

each population and population pair using the SAF files in realSFS (Nielsen et al. 2012).

Population clustering

We investigated the genetic clustering of the individuals using principal component analysis

performed in PCAngsd (v. 0.982; Skotte et al. 2013) for exploratory and illustration purposes, and

a formal clustering analysis for 1-10 clusters (K) as implemented in NGSAdmix (v. 33; Skotte et

al. 2013). To prepare the input files for these tools, we estimated pairwise linkage disequilibrium

between all SNPs within 100 kb of each other using ngsLD (Fox et al. 2019). We then used the

included prune_ngsLD.py script to produce a list of linkage pruned SNPs, excluding edges from

the initial graph between sites with a distance greater than 50 kb and with an r^2 of less than 0.1 (--

max_dist 50000 --min_weight 0.1). We then subset our Beagle file to these linkage pruned sites

and used it as input for both PCAngsd and NGSAdmix. For the clustering analysis, we performed

between 10-100 replicate optimizations for each K with a maximum of 4000 iterations per

replicate. Replicates were halted when the replicates converged or 100 replicates had completed,

considering replicates as converging when the three highest likelihood replicates came within 2

log-likelihood units of each other, as in Pečnerová et al. (2021). We selected the best K as the

highest level of K achieving convergence.

Demographic history

Demographic history was estimated for each population using MSMC2 (v. 2.1.2; Schiffels and

Durbin 2014). We generated the individual VCF and mask bed file by using bcftools (v. 1.14; Li

2011) and *bamCaller.py* script from msmc-tools (https://github.com/stschiff/msmc-tools;

Accessed on 2023-02-09). We also generated a reference genome mask file to eliminate sites with

extremely low or high coverage, defined as 0.5x and 2x mean coverage respectively. We used

Samtools (v1.16) to compute the average sample coverage and generate a BED file per-sample.

We merged the VCF and mask files (msmc-tools, *generate_multihetsep.py* script) and used the

resulting file as input for MSMC2. To scale the result, we used a mutation rate of 3.46×10^{-9}

mutations per site per generation, which is derived from estimates in *Drosophila melanogaster*

(Keightley et al. 2009) and a mean generational interval of 1 year (Romstock-Volkl 1997). Because

this software is sensitive to read coverage, neutrality, and repetitive content, we compared the

MSMC2 output for three sets of samples (the full set of samples, the individuals with highest

coverage in each of the eight populations, and the individuals with coverage closest to the global

mean in each of the eight populations) at three levels of VCF filtering (the VCF output from

bcftools + *bamCaller.py* script described above; the VCF excluding all repetitive content, and the

VCF excluding all outlier windows identified by population branch structure analysis described

below).

Population differentiation and divergence

To measure population differentiation, *Fst* was calculated for each site, first by generating the site

frequency spectrum (2D-SFS) for each population pair, based on which we calculated per-site *Fst*

using a Bhatia estimator (Bhatia et al. 2013) as implemented in ANGSD (realSFS *Fst*). Average

pairwise *Fst* was estimated genome-wide and for nonoverlapping 10kb windows (realSFS stats2).

We used population branch statistics (PBS) to assess differentiation between sympatric

populations of the two host races when corrected for divergence between populations within host

races. This accounts for heterogeneity in background divergence between populations. We

calculated PBS for each site (realSFS stats2) for four population triads, where each sympatric

population was the target in one comparison. Triads focused on populations where host plants are

sympatric, for which we expect selection favoring host specificity to be strongest, and were

composed of a sympatric population specializing on host plant A compared against the sympatric

and allopatric populations specializing on host B in either the east or west transects. Thus, PBS_{CHES}

quantifies the differentiation between sympatric CH-flies (CHES) and sympatric CO-flies

(COES), corrected for divergence from allopatric CO-flies (COLI) populations (Fig. S3

Supplementary Methods 3). PBS was estimated for nonoverlapping 10kb windows. Outlier

windows in each of these triads were identified as those with PBS values greater than four standard

deviations away from the genome-wide mean (Salmón et al. 2021; Montejo-Kovacevich et al.

2022).

Genome-wide association analysis with BayPass

Genome-wide scans for association with ecotype were performed with BayPass (v. 2.3; Gautier

2015). We converted the filtered VCF to allele frequencies for each population with bcftools. The

dataset was further filtered to account for linkage by subsampling every 20th variant across each

contig into 20 unique sets of SNPs. The data were analyzed first under the BayPass core model

using default option, then under the auxiliary model using a Markov Chain Monte Carlo (MCMC)

algorithm and ecotype as a covariate. Three independent runs (using the option -seed) were

performed for each of the 20 SNP sets. Support for association with ecotype was evaluated using

the mean Bayes Factor of these three runs. Results from the 20 independent runs were combined

into a single dataset for all sites, and reported in deciban units, with 10db corresponding to 10:1

odds, 20db to 100:1 odds, etc.).

Outlier analysis and gene ontology term enrichment

To identify genomic regions associated with host use we identified outliers for three different

metrics, PBS and dxy that were assessed independently for the western and eastern populations

respectively, and BayPass that was estimated based on all eight populations. Windows with PBS

and dxy that exceeded the genome-wide mean by four standard deviations or had a Bayes Factor

(in decibans) of 20 or higher were classified as outliers. As sex chromosomes and autosomes differ

in effective population size and sex chromosomes evolve faster, we performed all outlier analyses

on three different data sets, one data set including the entire genome, one including only putatively

X-enriched contigs, and one including only autosomal contigs. This approach allowed us to

identify enriched functions that were associated with either X-enriched or autosomal contigs.

For each of these outlier sets, GO-enrichment analyses were performed using TopGO (Alexa and

Rahnenfuhrer 2022), with a minimum node size of 5. Significant enrichment was tested with one-

sided Fisher's exact tests corrected with the parent-child algorithm (Grossmann et al. 2007). GO

terms were considered significantly enriched with a p-value < 0.01. We also performed GO-

analyses on the outliers that occurred in two of the three outlier sets in the west or in the east to

capture the outliers that are most consistently diverged.

Nucleotide diversity and divergence from neutrality

We evaluated nucleotide diversity (π) and divergence from neutrality, as characterized by Tajima's

D, to understand genome-wide and local differences among populations. ANGSD was used to

calculate pairwise nucleotide diversity for each site (thetaD, realSFS saf2theta). These were used

to estimate Tajima's D and pairwise differences across the entire genome and over 10Kb

nonoverlapping windows (thetaStat do_stat). To calculate π, pairwise differences were divided by

the total number of sites used in each window. For both metrics, we set a minimum coverage

fraction of 0.4 for our window-based analysis, meaning we required genotypes for at least 4kb of

the 10kb in each window. Windows below this threshold were excluded from downstream

analyses.

Selection statistics

To identify SNPs associated with host races and different biogeographic scenarios we used Selscan

(v. 1.3; Szpiech and Hernandez 2014) to calculate the cross-population extended haplotype

homozygosity (XP-EHH). XP-EHH is a cross-population statistic that test for differential local

adaptation and calculates the extended haplotype homozygosity (EHH) for two populations

separately, integrated over recombination distance and compared in order to identify loci under

selection in one of the two populations. To prepare the data for XP-EHH analysis, SNPs unpruned

for linkage disequilibrium were phased with SHAPEIT2 v 2.r837 (Delaneau et al. 2013) using

default MCMC parameters (7 burn-in MCMC iterations, 8 pruning iterations, and 20 main

iterations), conditioning states for haplotype estimation (K = 100), and window size set at 0.5Mb

as recommended for whole genome sequence data. We also used an effective population size (Ne)

of 494, based on estimates calculated with SNeP (v. 1.1; Barbato et al. 2015). Here, we used the

mean across populations, as all estimates were very similar. XP-EHH was calculated in windows

of size 100 kb in each direction from core SNPs, allowing decay curves to extend up to 1 Mb from

the core, and SNPs with MAF < 0.05 were excluded from consideration as a core SNP. As we

lacked a fine-scale genetic map for *T. conura*, we assumed a constant recombination rate of 1.73

cM/Mb as found for the related species *Bactrocera cucurbitae*. Scores were then normalized

within contigs with the norm version of selscan version 1.3.0 program.

The XP-EHH method identifies genomic regions that underwent a selective sweep in one

population but remained variable in the second population. We calculated XP-EHH for six

population pairs to test for differences in selection between host races in sympatry and allopatry,

and to test for differences between western and eastern transects. For each pair, we identified

putative locality-specific sweeps by selecting those SNPs above the 99th percentile of the genome-

wide distribution. Genes under selection were detected by comparing the SNPs detected by the

XP-EHH test against a gene annotation for *T.conura* with bedtools v 2.29.2 window 2 kb upstream

and downstream these genes (Quinlan and Hall 2010).

Introgression statistics

We investigated if there is evidence of introgression between populations by estimating genome-

wide Patterson's D and f_4-ratios (Patterson et al. 2012) for all possible population trios, as

implemented in Dsuite's DTrios function (Malinsky et al. 2021). To assess variation in

introgression across the genome in sympatric populations, we investigated f_{dM} in windows across

the genome for eight focal trios, as implemented in the script ABBABABAwindows.py

(https://github.com/simonhmartin/genomics_general). We selected this statistic as it is designed to

detect introgression in localized regions of the genome and is suitable for detecting introgression

between both P1 & P3, as well as P2 & P3. Briefly, for each of the four sympatric populations as

P2, we estimated f_{dM} in non-overlapping 10 kb windows, assigning the closest allopatric population

of the same host plant to P1, with P3 being either the closest allopatric or sympatric population of

the alternate host plant. For all analyses we utilized a *Tephritis hyoscyami* individual

(P18705_127) as the outgroup.

Results

Our final assembly of the *T. conura* genome was 1.99 Gbp in 2713 contigs (N50 = 1.75 Mbp).

Benchmarking with Dipteran single copy orthologs found 97.0% completeness with low

duplication (2.4%), making it not only the longest but among the most complete Tephritidae

genomes available to date (Fig. S4). It was also the most repeat rich genome by more than 20%,

driven by an expansion of long terminal repeats that make up over 22% of the genomic content

(Fig. S5; Table S5). This history of genomic expansion and high repetitive content, which increase

the opportunity for both structural change and chromatin state differences, suggests that *T. conura*

has the potential for extensive and rapid genomic divergence driven by structural variants.

Genomically divergent host races

We generated whole genome sequence data from eight continental *T. conura* populations

specializing on either *C. oleraceum* and *C. heterophyllum*. Populations were located along two

parallel transects west and east of the Baltic Sea, covering regions where host plants can be found

in either allopatry or sympatry (Fig. 1D). We found considerable genomic divergence, with *C.*

heterophyllum (CH) and *C. oleraceum* (CO) host races representing distinct lineages. The two host

races separated in a formal clustering analysis implemented in Admixture (Fig. 1E) and

consistently, in a genome-wide exploratory analysis of variants, the first PC axis, explaining 4.7%

of the variance, separated individuals by host race (Fig. 1F). Within the derived CO host race,

individuals cluster by geography, with western and eastern populations separating along the second

PC axis and in the clustering analysis (Fig. 1F). The western sympatric CH population (CHSK)

appears to be the most distinct lineage, separating from both CO individuals and the remaining CH

individuals along PC1, forming a separate cluster under the best supported model in the clustering

analysis (K=3; Table S4). Individuals from the remaining populations were grouped largely by

ecotype, with some evidence of shared ancestry (Fig. S6). These results were supported by

pairwise genome-wide *Fst* and dxy comparisons, which showed the largest differentiation and

divergence occurred between populations of different host races (Fig. S7).

In contrast to earlier suggestions of the derived host race arising before the onset of the last glacial

maximum c. 18000 years ago (Diegisser et al. 2006b) , an MSMC2 analysis suggests host races

diverged over one million years ago (Fig. 1G). These results broadly confirm findings from

clustering analyses, including western and eastern clustering of the CO host race. In particular, the

highly divergent CHSK population again appears as a distinct lineage, perhaps having diverged

from the other populations much earlier. However, similar patterns can arise when populations

have been affected by recent bottlenecks, and we have been unable to disentangle these alternative

explanations at this time. Finally, we found that our MSMC2 analyses were highly sensitive to

both genomic content (e.g., nonneutral sites, repetitive content) and coverage (Fig. S8)

Genomic regions underlying host plant specialization

The genomic regions that separated the host races were highly consistent between the western and

eastern transect for each of the tests we performed (PBS, dxy and BayPass Fig. 2A-B, C-D and E

respectively; Fig. S9-S12). We found consistently higher differentiation and divergence for

nonoverlapping 10kb windows located on contigs enriched for X-linked *D. melanogaster*

orthologs (X-enriched, putatively sex-linked) than the remainder of the genome. Both west and

east of the Baltic, population branch statistics (PBS) for sympatric populations recover many

outliers exceeding the mean by more than 4SD, both on putatively sex-linked and autosomal

contigs and in eastern and western host race comparisons (Fig. 2A-B; Fig. S10). Many of these

outliers clustered into distinct peaks, often affecting a majority of the 10kb windows within a given

contig. There was a remarkably high degree of overlap between western and eastern PBS outliers.

Fewer and less clustered outlier regions were identified using dxy, but dxy outliers were also highly

consistent across the two host race comparisons (Fig. 2C-D; Fig. S11-S12). BayPass, which

identified sites associated with host use among all eight populations, recapitulated many of the

divergent peaks found using the PBS analyses (Fig. 2E). However, using the conservative

recommended cutoff of BF > 20, only 60% of the BayPass outlier windows overlapped PBS

outliers in the west and east transects, and no BayPass outliers overlapped dxy outliers. We found

surprisingly low overlap among PBS, dxy and BayPass outliers within the eastern and western

host race comparison. As a result, we identified 'host race outlier' windows as those with support

from at least one statistic within each transect.

Host race outlier windows were more abundant on X-enriched contigs than those falling on

putative autosomes, and a greater proportion overlapped annotated genes (Fig. S13).

Gene ontology (GO) analyses recovered enrichment of early morphological development (e.g.,

GO:0048598 embryonic morphogenesis; GO:0001704 formation of primary germ layer) and

growth and mitosis (e.g., GO:1901722 regulation of cell proliferation, GO:0051782 negative

regulation of cell division; Fig. 3). RNA surveillance and molecular transport were also enriched

in both the west and the east, and these were similarly supported by enriched cellular components

(GO:0005921-gap junction; GO:0000701-condensed nuclear chromosome) and molecular

functions (GO:0070034 telomerase RNA binding; GO:0015078 proton transmembrane transporter

activity; GO:0005342-organic acid transmembrane transporter activity). A minority of outliers

were located on putative autosomes, and as a result the genes overlapping these windows in both

the west and east were enriched for few GO terms. However, appendage morphogenesis

(GO:0035107) was enriched among both X-enriched and autosomal outliers (Fig. 3), which may

support our hypothesis that nonphysical linkage underlies some host-specific adaptations in *T.*

conura. When we limited outlier windows to those with support from two or more statistics (i.e.,

PBS and Dxy or PBS and BayPass), we found that very few of the same biological process terms

were enriched compared to outliers supported by one or more statistic (Fig. 2F; Fig. S14). Rather,

a large number of terms involved in neurological processes were enriched, suggesting highly

differentiated and divergent regions may be involved in cue processing, related to interaction with

host plants, or communication, related to interaction with conspecifics (Fig. S14).

Selection for host race divergence

To understand the selective pressures acting on the regions of the genome that differentiate the

host races, we tested for evidence for selective sweeps, and whether these are stronger in the

derived CO host race. To do this, we assessed Tajima's D, nucleotide diversity (π) and extended

haplotype homozygosity. When considering all windows across the genome, Tajima's D was

similar among populations (Fig. S15-16). The exception was CHSK, which had a much higher

Tajima's D, supporting the conclusion from Admixture and MSCM analyses that this population

has experienced a recent bottleneck. In contrast to results for all windows, Tajima's D in outlier

windows is lower in CO populations than in CH populations, which is consistent with these regions

having experienced strong positive or purifying selection. Similarly, π within outlier windows is

also lower for CO populations (Fig. S17-19), suggesting selection has disproportionately decreased

nucleotide diversity in these regions in the CO population.

Across the genome, we found low Dxy, π, Tajima's D associated with repeat-dense regions of the

genome, even after imposing a coverage fraction filter that excluded windows missing genotypes

for 60% or more of the bases. However, we were able to confirm that this association is unlikely

to be driving the evidence for selective sweeps described above. We found that there was no

relationship between the proportion of repetitive content in outlier windows and estimates of Dxy

(Fig. S20) , π (Fig. S21), or Tajima's D (Fig. S22), suggesting these results are robust to the effects

of repetitive content.

Extended haplotype statistics reveal considerably stronger selection among the top selected SNPs

in CO populations when contrasted with CH populations. With a cutoff of the top 1% of SNPs, we

we recover a similar number of SNPs under selection in all the populations (Range: 253525 –

279839, Table S5). In all comparisons between CH and CO population pairs, the mean normalized

XP-EHH statistic of selected SNPs was higher in the CO host race. Meanwhile, in comparisons

within host race, there was evidence of stronger selection in sympatric populations in the western

transect (CHSK and COSK). We identified selected genes as those overlapping selected SNPs. We

found more genes overlapped selected SNPs in the CH host race (Table S5). This contrasting

pattern between SNPs and genes was due to the fact that the higher number of SNPs under selection

in CO populations affected a reduced number of annotated genes compared to CH populations

(Table S5). Furthermore, these genes were more likely to overlap with genes containing at least

one highly differentiated outlier window (as in Fig 3F; 4) in CO populations than in CH

populations (Fig. S23), again supporting that selected SNPs fall in a more concentrated, more

differentiated set of genes. This pattern is strongest in the COSK population, where in the number

of SNPs in annotated genes is an order of magnitude higher than in the other populations (Table

S5) and almost doubles the number of SNPs in annotated genes in CHSK.

Regions permeable to introgression

We find evidence consistent with introgression between the host races in both the eastern and

western sympatric regions. Using D-statistics, we found introgression between host races was

higher in the eastern than the western transect, consistent with this being an older contact zone for

these host races (Fig. S24, Table S6) We also found greater evidence of introgression into

sympatric populations of the alternate host race than into more distant allopatric populations.

Introgression was not evenly distributed across the genome, with outlier windows showing distinct

patterns of introgression compared to nonoutliers (Fig. 5). We used f_{dM} statistics to compare

introgression in 10kb windows across the genome. This statistic is expected to be positive when

introgression occurred between P3 and P2 and negative values if it occurred between P3 and P1.

After filtering, non-outlier windows had a much wider spread of f_{dM} values than outlier windows.

Similar to genome-wide results, f_{dM} in nonoutlier regions were consistently higher in trees testing

introgression into sympatric populations of the other host race than in trees testing introgression

into allopatric populations. In contrast, outlier regions appeared more resistant to introgression.

Across most triads, f_{dM} in outlier regions was lower, and in western transects significantly more

negative, than in nonoutliers. Negative values are consistent with greater introgression between

the more geographically distant populations, which may reflect historical gene flow. These

patterns suggest outlier regions have increased resistance to introgression between sympatric

populations. The exception to this result occurred in triads comparing introgression from CH

populations (either CHFI or CHES) into the two CO populations in the east. In both cases, outliers

had higher f_{dM} than nonoutliers.

Discussion

We find strong and consistent genomic divergence between the two *T. conura* host races in both

allopatry and sympatry. These results were highly parallel between contact zones both west and

east of the Baltic Sea. Putatively sex-linked contigs consistently show high divergence between

the host races. Furthermore, these contigs form discrete clusters when aligned to contiguous

regions in the genome of the closely related species *R. pomonella*. The recent and extensive

expansions of long terminal repeats, and discrete nature of the divergent genomic areas could

suggest a role for repeat-facilitated sex-linked inversions in separating the two host races of *T.*

conura. Potentially, these regions could couple the multifarious ecological traits that are associated

with host plant, including phenology (Romstock-Volkl 1997), host preference (Diegisser et al.

2008), morphology including ovipositor length (Diegisser et al. 2007; Nilsson et al. 2022), and

physiology including digestion of plant defense chemicals (Diegisser et al. 2008) and thus facilitate

coexistence between the host races. Recent research has shown that inversions are important for

maintaining both discrete morphs within species (Jones et al. 2012; Küpper et al. 2016; Tuttle et

al. 2016; Faria et al. 2019a), facilitate speciation (Wellenreuther and Bernatchez 2018; Faria et al.

2019b) and enable coexistence with sister taxa after a shorter period of time (Hooper and Price

2017). Our findings suggest that inversions may be important for coupling (Butlin and Smadja

2018) of ecologically co-adapted traits also in *T. conura*. However, we also found that discrete

autosomal regions show elevated PBS, *Fst* and Dxy, and were associated with host race divergence

in BayPass analyses. This supports that, in addition to physical coupling facilitated by inversions,

nonphysical coupling of host race-associated loci may be contributing to ecological divergence

and reproductive isolation in peacock flies.

Consistent with expectations under strong divergent ecological selection, the divergent areas are

highly consistent across the eastern and western sympatric zones, both for PBS and dxy.

Introgression statistics suggest low levels of gene flow across many, but not all, regions of the

genome in the sympatric areas. Regions of the genome not associated with host use should

exchange freely, and the presence of discrete genomic regions that remain divergent between host

races in sympatry suggests they contain divergent adaptations in the two host races. Moreover,

genomic regions important for host adaptation are expected to diverge in parallel in the sympatric

zones on both sides of the Baltic, as parallelism in response to a similar environment is a hallmark

of selection (Johannesson 2001; Stuart et al. 2017). Hence, the genomic regions identified in our

analyses are highly likely to be important for host race specific ecological adaptation.

Interestingly, there is also an overlap in functions of the genes enriched in outlier sets based on

PBS, dxy, BayPass or the intersect. Functions that frequently were enriched include metabolic

biological processes, neural signaling, and morphogenesis. The divergence in genes associated

with metabolic processes is expected as the host races suffer strong extrinsic inviability as larvae

when feeding on the wrong host plants (Diegisser et al. 2008). While the exact function of the

genes contributing to the enrichment of morphogenesis among diverged genes are not known, it

would be interesting to examine the expression of these genes during ovipositor formation, as the

relative length of the ovipositor is the most defining morphological difference among the host

races (Diegisser et al. 2007; Nilsson et al. 2022). There are also size differences between the host

races that potentially could be affected by these genes. Interpretation of neural signaling remains

speculative, but examining the expression of these genes in the brain when exposed to different

host plants would uncover if the divergence in neural signaling could be of importance in host

plant preference.

Tephritis conura has parallels with the co-existence of different host races of *Rhagoletis* infesting

hawthorne and apple, respectively (Feder et al. 1988; Filchak et al. 2000; Feder et al. 2003), the

parapatric ecomorphs of *Littorina* snails adapted to either wave-exposure or crab predation

(Johannesson and Johannesson 1996; Hollander et al. 2005; Johannesson et al. 2010; Faria et al.

2019a; Perini et al. 2020), and *Timema* walking sticks adapted to be cryptic on different host plants

(Nosil et al. 2002; Nosil 2007; Nosil and Sandoval 2008; Villoutreix et al. 2020). Each involve

strong selection pressures that maintain ecologically divergent population in sympatry despite

incomplete reproductive isolation. Rhagoletis flies are isolated by phenological difference and host

preference (Feder et al. 2003), but the genomic differences that underlie the differently adapted

ecomorphs is polygenic and can rapidly be recovered through selection on standing genetic

variation within the ancestral hawthorn host race (Egan et al. 2015). The genomic basis conferring

mimicry on different host plants in Timema walking consists of few loci of large effects, and the

background genomic divergence is lower (Villoutreix et al. 2020). The genomic basis of the

Littorina ecotypes is similar to that in *T. conura* in that variants within inversions contribute to

approximately half of the ecotype divergence (Koch et al. 2022). Compared to those systems, the

study races in *T. conura* have relatively high levels of genetic divergence and strong reproductive

isolation, arising from the combination of preference, phenology and performance in the

alternative niche (Diegisser et al. 2006a; Diegisser et al. 2006b, 2007; Diegisser et al. 2008;

Nilsson et al. 2022). Potentially, the adaptation is more multifarious, or the adaptation may have

taken place during a longer period of time. In contrast to previous divergence time estimates based

on mitochondria, MSMC analyses suggest that host race divergence may be old. Potentially, the

host races could be older than 500 kya as there is no convergence between the host races in the

MSMC plots even at this time scale. Further exploration of the dependency of the MSMC inference

of which fractions of the genome are included would be necessary to draw any firm conclusions

though. Given the morphological similarity of the species (Nilsson et al. 2022), it may seem

surprising that divergence times exceed previous estimates of just before the last glacial maximum.

However, our new estimates are consistent with the high level of genomic divergence and limited

introgression on the putatively sex-linked contigs observed in this study. Furthermore, we believe

this timeframe is also in accordance with strong reproductive isolation in this system, evidenced

by phenological differences (Romstock-Volkl 1997), oviposition preference and larval mortality

on the new host plant (Diegisser et al. 2008). Hence, *T. conura* provides a window into a later

stage of ecological speciation continuum (Stankowski and Ravinet 2021) with the potential to offer

interesting insights into the development of reproductive barriers following ecological

specialization.

Interestingly, the scaffolds that were enriched for *Drosophila melanogaster* X-linked proteins

showed on average higher divergence. Several different factors, including the lower effective

population sizes of sex chromosomes (Charlesworth et al. 2018), their potential lower rates of

recombination (Charlesworth 2017) and the faster Z (Mank et al. 2009) are expected to contribute

to this pattern, but differ depending on the evolutionary history of the sex chromosomes. These

processes could complicate the interpretation of genomic regions or genes important for ecological

adaptation on these contigs. In addition, while *Tephritis* flies are known to have ZW sex

determination (shown for *T. californica*; Vicoso and Bachtrog 2015), the origins of this switch to

female heterogamety are not known. Whether the Z-chromosome was converted directly from the

ancestral X-chromosome, or originated from an autosome while the ancestral X-chromosome lost

its sex determination role remains to be discovered, limiting our understanding of the evolutionary

dynamics of X-enriched contigs in *T. conura.* An additional intriguing possibility is that lower

levels of gene flow could result in the typically more divergent sex chromosomes with a large role

in reproductive isolation representing the ancestral species tree in face of introgression (Fontaine

et al. 2015). Future comparisons of coverage across the genomes of males and females and gene

expression data from reproductive tissues can shed further light on the sex chromosome in *T.*

conura and facilitate interpreting its importance for host race formation. The GO-terms for

putatively sex-linked scaffolds rarely overlapped with those enriched for outliers on the autosomes.

Long-read assemblies of the derived CO host race would be an interesting future venue to gain

further insights into the structural variation underlying host race formation and persistence in *T.*

conura. Moreover, the potential for transposable elements to contribute to reproductive isolation

and/or serve as a source for novel variation through altering regulation of gene expression and

remodeling chromosomes, has recently been recognized (Serrato-Capuchina and Matute 2018).

To which extent recent TE releases has contributed to the potential to adapt to novel niches, and

to reproductively isolate the host races of *T. conura* is an interesting venue for future research that

could help shed light on the role of TE releases in ecological speciation.

In a rapidly changing climate, understanding the ability to adapt to a novel niche is more important

than ever, as it has critical implications for conservation and our power to predict potential host

range expansions and new pest species following northward shifts of inset distributions. *Tephritis*

conura is an exemplar ongoing ecological speciation of two host races and future research on this

study system will offer unique insights into the changes in the genome necessary for rapid

colonization of novel niches.

Data availability

All code used will be deposited on github upon acceptance. The reference genome will be available

from NCBI and all resequencing data will be available as bam-files at NCBI/ENA upon manuscript

acceptance.

Author contributions

J. O. performed field work, extracted DNA and RNA, calculated recent effective population sizes

with SNeP and performed extended haplotype statistics analyses. K. J. N. performed field work,

performed bioinformatic analyses including PCAngsd, NGSadmix, *Fst*, dxy, pi and Tajima's D

with support from Z N and RAS, plotted figures and prepared the supplement. R. S. improved the

reference genome with analyses of microbial content and alignment to the *Rhagoletis pomonella*

genome, performed PBS analyses, BayPass analyses, GO-analyses and helped with other analyses

and plotting and wrote a first draft of the methods. Z. J. N. paved the pipeline used to generate *Fst*,

dxy, pi and Tajima's D as well as PCAngsd and NGSadmix. C. Y. generated alignments and the

maker annotation. Y. H. performed MSMC analyses. J. A. L. helped analyze *Fst*, dxy, pi and

Tajima's D. H. V. extracted DNA for the first long-read assembly. A. R. Conceived of, designed

and funded the study, performed field work, outlined the bioinformatics analyzes to be performed

and supported the students performing these, interpreted data, took a lead on the writing and wrote

the first draft of the introduction, results and discussion.

Acknowledgements

This work was funded by a fellowship from the Wenner-Gren foundations, a Swedish Research Council starting grant, and additional grants from the Crafoord foundation, Erik Philip Sörensens Stiftelse and Carl Tryggers Stiftelse covering sequencing to A.R. The authors acknowledge support from the National Genomics Infrastructure in Stockholm funded by Science for Life Laboratory, the Knut and Alice Wallenberg Foundation and the Swedish Research Council, and SNIC/Uppsala Multidisciplinary Center for Advanced Computational Science for assistance with massively parallel sequencing and access to the UPPMAX computational infrastructure. We thank Jes Johanneson for support when starting the work on this study system, and Emma Kärrnäs and Mathilde Schnuriger for assistance during field work.

Fig. 1. Genomic divergence of *Tephritis conura* host races. Host races of *T. conura* specialize on *Cirsium heterophyllum* (CH; **A**) or *C. oleraceum* (CO; **B**) thistle buds. **(C)** A female *T. conura.* **(D)** Distribution and admixture proportions of populations sampled for this study. Purple represents regions of northern Europe where *C. heterophyllum* exists in allopatry, green where *C.*

677   *oleraceum* exists in allopatry, and striped where both host plants are found in sympatry. **(E)**
678   Admixture proportions estimated for K=3 (see Fig. S6A for K2-9). **(F)** Principal component (PC)
679   analysis of genetic differences among populations separates CH (purple) and CO (green)
680   individuals first PC axis (see Fig. S6B for a comparison of the second and third PC axes). **(G)**
681   MSMC analysis of demographic history.
682

Fig. 2. Highly divergent genomic windows between host races based on **(A,B)** population branch statistics (PBS), **(C,D)** dxy, and **(E)** BayPass analyses for association with host race. Statistics were evaluated over 10kb non-overlapping windows. PBS outliers were greater than four standard deviations above the mean. Dxy comparisons were between CHSK and COSK in the west, CHES and COES in the easta and outliers were also greater than four standard deviations above the mean. BayPass outliers had an average BayesFactor over 20 (in decibans). **(F)** Overlap between outlier sets for eastern and western transects.

Fig. 3. Functional enrichment of outlier genes. Gene set enrichment was performed using genes (+/- 2000bp) overlapping 10kb outlier regions identified using PBS, Dxy or BayPass in the western (blue) and eastern (coral) comparisons. BayPass outliers were included in both east and west, as this analysis was performed on all populations. Point size is scaled to reflect the number of genes annotated with that GO term (log10-transformed).

Fig. 4. Distribution of selected SNPs across the genome. Selection was estimated using extended haplotype statistics. Panels show top 1% of SNPs identified as under selection in each sympatric population when compared against the sympatric population of alternative host race (CHSK vs. COSK; CHES vs. COES). For all comparisons see Table S5.

Fig. 5. Introgression in outlier windows compared to genome-wide. Introgression was compared between triads of *T. conura* populations (trees below x-axis) using f_{dM}. Values close to zero indicate little to no introgression, negative values indicate greater relative introgression between the left-most population and P3, while positive values indicate greater relative introgression between the middle population and P3, where P3 is one of the two populations above each tree.

References

Alexa, A. and J. Rahnenfuhrer. 2022. topGO: enrichment analysis for gene ontology. R package version 2.48.0.

Alonge, M., L. Lebeigle, M. Kirsche, K. Jenike, S. Ou, S. Aganezov, X. Wang, Z. B. Lippman, M. C. Schatz, and S. Soyk. 2022. Automated assembly scaffolding using RagTag elevates a new tomato system for high-throughput genome editing. Genome Biology 23:258.

Aluja, M. and A. L. Norrbom. 2001. Fruit flies (*Tephritidae*): phylogeny and evolution of behaviour. CRC Press, Boca Raton.

Anderson, S. A. S., H. López-Fernández, and J. T. Weir. 2023. Ecology and the origin of nonephemeral species. Am. Nat. 0:000-000.

Anderson, S. A. S. and J. T. Weir. 2022. The role of divergent ecological adaptation during allopatric speciation in vertebrates. Science 378:1214-1218.

Barbato, M., P. Orozco-terWengel, M. Tapio, and M. W. Bruford. 2015. SNeP: a tool to estimate trends in recent effective population size trajectories using genome-wide SNP data. Front Genet 6:109.

Berdan, E. L., T. Flatt, G. M. Kozak, K. E. Lotterhos, and B. Wielstra, eds. 2022. Genomic architecture of supergenes: causes and evolutionary consequences [Theme Issue], Philosophical Transactions of the Royal Society B-Biological Sciences, 377:1856.

Berlocher, S. H. and J. L. Feder. 2002. Sympatric speciation in phytophagous insects: moving beyond controversy? Annu. Rev. Entomol. 47:773-815.

Bhatia, G., N. Patterson, S. Sankararaman, and A. L. Price. 2013. Estimating and interpreting FST: the impact of rare variants. Genome Res 23:1514-1521.

Boag, P. T. and P. R. Grant. 1981. Intense natural selection in a population of Darwin's finches

(*Geospizinae*) in the Galápagos. Science 214:82-85.

Bohutínská, M., J. Vlček, S. Yair, B. Laenen, V. Konečná, M. Fracassetti, T. Slotte, and F. Kolář.

2021. Genomic basis of parallel adaptation varies with divergence in *Arabidopsis* and its

relatives. Proc. Natl. Acad. Sci. 118:e2022713118.

Bolnick, D. I. 2011. Sympatric speciation in threespine stickleback: why not? International Journal

of Ecology 2011:e942847.

Boulain, H., F. Legeai, J. Jaquiéry, E. Guy, S. Morlière, J.-C. Simon, and A. Sugio. 2019.

Differential expression of candidate salivary effector genes in pea aphid biotypes with

distinct host plant specificity. Frontiers in Plant Science 10.

Broad Institute. Picard Tools, http://broadinstitute.github.io/picard/.

Bush, G. L. 1969. Sympatric host race formation and speciation in frugivorious flies of genus

Rhagoletis (*Diptera, Tephritidae*). Evolution 23:237-251.

Butlin, R. K. and C. M. Smadja. 2018. Coupling, reinforcement, and speciation. Am. Nat. 191:155-

172.

Camacho, C., G. Coulouris, V. Avagyan, N. Ma, J. Papadopoulos, K. Bealer, and T. L. Madden.

2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421.

Cantarel, B. L., I. Korf, S. M. Robb, G. Parra, E. Ross, B. Moore, C. Holt, A. Sánchez Alvarado,

and M. Yandell. 2008. MAKER: an easy-to-use annotation pipeline designed for emerging

model organism genomes. Genome Res 18:188-196.

Charlesworth, B., J. L. Campos, and B. C. Jackson. 2018. Faster-X evolution: theory and evidence

from *Drosophila*. Mol. Ecol. 27:3753-3771.

Charlesworth, D. 2017. Evolution of recombination rates between sex chromosomes.
Philosophical Transactions of the Royal Society B: Biological Sciences 372:20160456.

Cheng, H., G. T. Concepcion, X. Feng, H. Zhang, and H. Li. 2021. Haplotype-resolved de novo
assembly using phased assembly graphs with hifiasm. Nature Methods 18:170-175.

Chevin, L.-M., G. Decorzent, and T. Lenormand. 2014. Niche dimensionality and the genetics of
ecological speciation. Evolution 68:1244-1256.

Christenhusz, M. J. M. and J. W. Byng. 2016. The number of known plants species in the world
and its annual increase. Phytotaxa 261:201-217.

Christenson, L. D. and R. H. Foote. 1960. Biology of fruit flies. Annu. Rev. Entomol. 5:171-192.

Craig, T. P., J. K. Itami, W. G. Abrahamson, and J. D. Horner. 1993. Behavioral evidence for host-
race formation in *Eurosta solidaginis*. Evolution 47:1696-1710.

Danecek, P., J. K. Bonfield, J. Liddle, J. Marshall, V. Ohan, M. O. Pollard, A. Whitwham, T.
Keane, S. A. McCarthy, R. M. Davies, and H. Li. 2021. Twelve years of SAMtools and
BCFtools. Gigascience 10.

Delaneau, O., B. Howie, Anthony J. Cox, J.-F. Zagury, and J. Marchini. 2013. Haplotype
estimation using sequencing reads. The American Journal of Human Genetics 93:687-696.

Diegisser, T., J. Johannesen, C. Lehr, and A. Seitz. 2004. Genetic and morphological
differentiation in *Tephritis bardanae (Diptera: Tephritidae)*: evidence for host-race
formation. J. Evol. Biol. 17:83-93.

Diegisser, T., J. Johannesen, and A. Seitz. 2006a. The role of geographic setting on the
diversification process among *Tephritis conura (Tephritidae)* host races. Heredity 96:410-
418.

Diegisser, T., J. Johannesen, and A. Seitz. 2008. Performance of host-races of the fruit fly,

Tephritis conura on a derived host plant, the cabbage thistle *Cirsium oleraceum*:

Implications for the original host shift. J. Insect Sci. 8:1-6.

Diegisser, T., A. Seitz, and J. Johannesen. 2006b. Phylogeographic patterns of host-race evolution

in *Tephritis conura (Diptera: Tephritidae)*. Mol. Ecol. 15:681-694.

Diegisser, T., A. Seitz, and J. Johannesen. 2007. Morphological adaptation in host races of

Tephritis conura. Entomol. Exp. Appl. 122:155-164.

Egan, S. P., G. J. Ragland, L. Assour, T. H. Q. Powell, G. R. Hood, S. Emrich, P. Nosil, and J. L.

Feder. 2015. Experimental evidence of genome-wide impact of ecological selection during

early stages of speciation-with-gene-flow. Ecol. Lett. 18:817-825.

Faria, R., P. Chaube, H. E. Morales, T. Larsson, A. R. Lemmon, E. M. Lemmon, M. Rafajlović,

M. Panova, M. Ravinet, K. Johannesson, A. M. Westram, and R. K. Butlin. 2019a. Multiple

chromosomal rearrangements in a hybrid zone between *Littorina saxatilis* ecotypes. Mol.

Ecol. 28:1375-1393.

Faria, R., K. Johannesson, R. K. Butlin, and A. M. Westram. 2019b. Evolving inversions. Trends

Ecol. Evol. 34:239-248.

Feder, J. L., S. H. Berlocher, and S. B. Opp. 1998. Sympatric host-race formation and speciation

in *Rhagoletis* (*Diptera: Tephritidae*): a tale of two species for Charles D.

Feder, J. L., C. A. Chilcote, and G. L. Bush. 1988. Genetic differentiation between sympatric host

races of the apple maggot fly *Rhagoletis pomonella*. Nature 336:61-64.

Feder, J. L., J. B. Roethele, K. Filchak, J. Niedbalski, and J. Romero-Severson. 2003. Evidence

for inversion polymorphism related to sympatric host race formation in the apple maggot

fly, *Rhagoletis pomonella*. Genetics 163:939-953.

Filchak, K. E., J. B. Roethele, and J. L. Feder. 2000. Natural selection and sympatric divergence

in the apple maggot *Rhagoletis pomonella*. Nature 407:739-742.

Flynn, J. M., R. Hubley, C. Goubert, J. Rosen, A. G. Clark, C. Feschotte, and A. F. Smit. 2020.

RepeatModeler2 for automated genomic discovery of transposable element families. Proc

Natl Acad Sci U S A 117:9451-9457.

Fontaine, M. C., J. B. Pease, A. Steele, R. M. Waterhouse, D. E. Neafsey, I. V. Sharakhov, X.

Jiang, A. B. Hall, F. Catteruccia, E. Kakani, S. N. Mitchell, Y.-C. Wu, H. A. Smith, R. R.

Love, M. K. Lawniczak, M. A. Slotman, S. J. Emrich, M. W. Hahn, and N. J. Besansky.

2015. Extensive introgression in a malaria vector species complex revealed by

phylogenomics. Science 347:1258524.

Fox, E. A., A. E. Wright, M. Fumagalli, and F. G. Vieira. 2019. ngsLD: evaluating linkage

disequilibrium using genotype likelihoods. Bioinformatics 35:3855-3856.

Gautier, M. 2015. Genome-wide scan for adaptive divergence and association with population-

specific covariates. Genetics 201:1555-1579.

Gow, J. L., C. L. Peichel, and E. B. Taylor. 2006. Contrasting hybridization rates between

sympatric three-spined sticklebacks highlight the fragility of reproductive barriers between

evolutionarily young species. Mol Ecol 15:739-752.

Grabherr, M. G., B. J. Haas, M. Yassour, J. Z. Levin, D. A. Thompson, I. Amit, X. Adiconis, L.

Fan, R. Raychowdhury, Q. Zeng, Z. Chen, E. Mauceli, N. Hacohen, A. Gnirke, N. Rhind,

F. di Palma, B. W. Birren, C. Nusbaum, K. Lindblad-Toh, N. Friedman, and A. Regev.

2011. Full-length transcriptome assembly from RNA-Seq data without a reference genome.

Nature Biotechnology 29:644-652.

Grossmann, S., S. Bauer, P. N. Robinson, and M. Vingron. 2007. Improved detection of

overrepresentation of gene-ontology annotations with parent child analysis. Bioinformatics

23:3024-3031.

Guan, D., S. A. McCarthy, J. Wood, K. Howe, Y. Wang, and R. Durbin. 2020. Identifying and

removing haplotypic duplication in primary genome assemblies. Bioinformatics 36:2896-

2898.

Hardy, N. B. and S. P. Otto. 2014. Specialization and generalization in the diversification of

phytophagous insects: tests of the musical chairs and oscillation hypotheses. Proceedings

of the Royal Society B: Biological Sciences 281:20132960.

Headrick, D. H. and R. D. Goeden. 1998. The biology of nonfrugivorous tephritid fruit flies. Annu

Rev Entomol 43:217-241.

Hendry, A. P. 2009. Ecological speciation! Or the lack thereof? Can. J. Fish. Aquat. Sci. 66:1383-

1398.

Herrel, A., K. Huyghe, B. Vanhooydonck, T. Backeljau, K. Breugelmans, I. Grbac, R. Van

Damme, and D. J. Irschick. 2008. Rapid large-scale evolutionary divergence in

morphology and performance associated with exploitation of a different dietary resource.

Proc. Natl. Acad. Sci. 105:4792-4795.

Hollander, J., M. Lindegarth, and K. Johannesson. 2005. Local adaptation but not geographical

separation promotes assortative mating in a snail. Animal Behaviour 70:1209-1219.

Hooper, D. M., S. C. Griffith, and T. D. Price. 2019. Sex chromosome inversions enforce

reproductive isolation across an avian hybrid zone. Mol. Ecol. 28:1246-1262.

Hooper, D. M. and T. D. Price. 2017. Chromosomal inversion differences correlate with range

overlap in passerine birds. Nat. Ecol. Evol. 1:1526-1534.

Huerta-Cepas, J., D. Szklarczyk, D. Heller, A. Hernández-Plaza, S. K. Forslund, H. Cook, D. R.

Mende, I. Letunic, T. Rattei, Lars J. Jensen, C. von Mering, and P. Bork. 2018. eggNOG

5.0: a hierarchical, functionally and phylogenetically annotated orthology resource based

on 5090 organisms and 2502 viruses. Nucleic Acids Research 47:D309-D314.

Johannesson, B. and K. Johannesson. 1996. Population differences in behaviour and morphology

in the snail *Littorina saxatilis*: Phenotypic plasticity or genetic differentiation? Journal of

Zoology 240:475-493.

Johannesson, K. 2001. Parallel speciation: a key to sympatric divergence. Trends Ecol. Evol.

16:148-153.

Johannesson, K., M. Panova, P. Kemppainen, C. André, E. Rolán-Alvarez, and R. K. Butlin. 2010.

Repeated evolution of reproductive isolation in a marine snail: unveiling mechanisms of

speciation. Philos Trans R Soc Lond B Biol Sci 365:1735-1747.

Jones, F. C., M. G. Grabherr, Y. F. Chan, P. Russell, E. Mauceli, J. Johnson, R. Swofford, M.

Pirun, M. C. Zody, S. White, E. Birney, S. Searle, J. Schmutz, J. Grimwood, M. C. Dickson,

R. M. Myers, C. T. Miller, B. R. Summers, A. K. Knecht, S. D. Brady, H. Zhang, A. A.

Pollen, T. Howes, C. Amemiya, J. Baldwin, T. Bloom, D. B. Jaffe, R. Nicol, J. Wilkinson,

E. S. Lander, F. Di Palma, K. Lindblad-Toh, and D. M. Kingsley. 2012. The genomic basis

of adaptive evolution in threespine sticklebacks. Nature 484:55-61.

Keightley, P. D., U. Trivedi, M. Thomson, F. Oliver, S. Kumar, and M. L. Blaxter. 2009. Analysis

of the genome sequences of three *Drosophila melanogaster* spontaneous mutation

accumulation lines. Genome Res 19:1195-1201.

Kim, D., J. M. Paggi, C. Park, C. Bennett, and S. L. Salzberg. 2019. Graph-based genome

alignment and genotyping with HISAT2 and HISAT-genotype. Nat Biotechnol 37:907-

915.

Koch, E. L., M. Ravinet, A. M. Westram, K. Johannesson, and R. K. Butlin. 2022. Genetic

architecture of repeated phenotypic divergence in *Littorina saxatilis* ecotype evolution.

Evolution 76:2332-2346.

Korneliussen, T. S., A. Albrechtsen, and R. Nielsen. 2014. ANGSD: analysis of next generation

sequencing data. BMC Bioinformatics 15:356.

Kovaka, S., A. V. Zimin, G. M. Pertea, R. Razaghi, S. L. Salzberg, and M. Pertea. 2019.

Transcriptome assembly from long-read RNA-seq alignments with StringTie2. Genome

Biology 20:278.

Kulmuni, J., R. K. Butlin, K. Lucek, V. Savolainen, and A. M. Westram, eds. 2020. Towards the

completion of speciation: the evolution of reproductive isolation beyond the first barriers

[Theme issue], Philosophical Transactions of the Royal Society B-Biological Sciences,

375:1806.

Küpper, C., M. Stocks, J. E. Risse, N. dos Remedios, L. L. Farrell, S. B. McRae, T. C. Morgan, N.

Karlionova, P. Pinchuk, Y. I. Verkuil, A. S. Kitaysky, J. C. Wingfield, T. Piersma, K. Zeng,

J. Slate, M. Blaxter, D. B. Lank, and T. Burke. 2016. A supergene determines highly

divergent male reproductive morphs in the ruff. Nature Genetics 48:79-83.

Lackey, A. C. R. and J. W. Boughman. 2017. Evolution of reproductive isolation in stickleback

fish. Evolution 71:357-372.

Li, H. 2011. A statistical framework for SNP calling, mutation discovery, association mapping and

population genetical parameter estimation from sequencing data. Bioinformatics 27:2987-

2993.

Li, H. and R. Durbin. 2009. Fast and accurate short read alignment with Burrows–Wheeler

transform. Bioinformatics 25:1754-1760.

Li, H., B. Handsaker, A. Wysoker, T. Fennell, J. Ruan, N. Homer, G. Marth, G. Abecasis, R.

Durbin, and G. P. D. P. Subgroup. 2009. The sequence alignment/map format and

SAMtools. Bioinformatics 25:2078-2079.

Malinsky, M., M. Matschiner, and H. Svardal. 2021. Dsuite - fast D-statistics and related

admixture evidence from VCF files. Molecular Ecology Resources 21:584-595.

Mank, J. E., K. Nam, and H. Ellegren. 2009. Faster-Z evolution is predominantly due to genetic

drift. Mol. Biol. Evol. 27:661-670.

Montejo-Kovacevich, G., J. I. Meier, C. N. Bacquet, I. A. Warren, Y. F. Chan, M. Kucka, C.

Salazar, N. Rueda-M, S. H. Montgomery, W. O. McMillan, K. M. Kozak, N. J. Nadeau, S.

H. Martin, and C. D. Jiggins. 2022. Repeated genetic adaptation to altitude in two tropical

butterflies. Nat. Commun. 13:4676.

Mora, C., D. P. Tittensor, S. Adl, A. G. B. Simpson, and B. Worm. 2011. How many species are

there on earth and in the ocean? PLoS. Biol. 9:8.

Nallu, S., J. A. Hill, K. Don, C. Sahagun, W. Zhang, C. Meslin, E. Snell-Rood, N. L. Clark, N. I.

Morehouse, J. Bergelson, C. W. Wheat, and M. R. Kronforst. 2018. The molecular genetic

basis of herbivory between butterflies and their host plants. Nat. Ecol. Evol. 2:1418-1427.

Nielsen, R., T. Korneliussen, A. Albrechtsen, Y. Li, and J. Wang. 2012. SNP calling, genotype

calling, and sample allele frequency estimation from new-generation sequencing data.

PLoS One 7:e37558.

Nilsson, K. J., J. Ortega, M. Friberg, and A. Runemark. 2022. Non-parallel morphological

divergence following colonization of a new host plant. Evol. Ecol. 36:859-877.

Nosil, P. 2007. Divergent host plant adaptation and reproductive isolation between ecotypes of

Timema cristinae walking sticks. Am. Nat. 169:151-162.

Nosil, P. 2012. Ecological speciation. Oxford University Press, Oxford.

Nosil, P., B. J. Crespi, and C. P. Sandoval. 2002. Host-plant adaptation drives the parallel evolution

of reproductive isolation. Nature 417:440-443.

Nosil, P., L. J. Harmon, and O. Seehausen. 2009. Ecological explanations for (incomplete)

speciation. Trends Ecol Evol 24:145-156.

Nosil, P. and C. P. Sandoval. 2008. Ecological niche dimensionality and the evolutionary

diversification of stick insects. PLoS One 3:e1907.

Patterson, N., P. Moorjani, Y. Luo, S. Mallick, N. Rohland, Y. Zhan, T. Genschoreck, T. Webster,

and D. Reich. 2012. Ancient admixture in human history. Genetics 192:1065-1093.

Pečnerová, P., G. Garcia-Erill, X. Liu, C. Nursyifa, R. K. Waples, C. G. Santander, L. Quinn, P.

Frandsen, J. Meisner, F. F. Stæger, M. S. Rasmussen, A. Brüniche-Olsen, C. Hviid Friis

Jørgensen, R. R. da Fonseca, H. R. Siegismund, A. Albrechtsen, R. Heller, I. Moltke, and

K. Hanghøj. 2021. High genetic diversity and low differentiation reflect the ecological

versatility of the African leopard. Curr Biol 31:1862-1871.e1865.

Perini, S., M. Rafajlovic, A. M. Westram, K. Johannesson, and R. K. Butlin. 2020. Assortative
mating, sexual selection, and their consequences for gene flow in *Littorina*. Evolution
74:1482-1497.

Quinlan, A. R. and I. M. Hall. 2010. BEDTools: a flexible suite of utilities for comparing genomic
features. Bioinformatics 26:841-842.

Rice, W. R. and E. E. Hostert. 1993. Laboratory experiments on speciation: what have we learned
in 40 years? Evolution 47:1637-1653.

Romstock-Volkl, M. 1997. Host race formation in *Tephritis conura*: determinants from three
trophic levels. Ecol. Stud. 130:21-38.

Salmón, P., A. Jacobs, D. Ahrén, C. Biard, N. J. Dingemanse, D. M. Dominoni, B. Helm, M.
Lundberg, J. C. Senar, P. Sprau, M. E. Visser, and C. Isaksson. 2021. Continent-wide
genomic signatures of adaptation to urbanisation in a songbird across Europe. Nat.
Commun. 12:2983.

Schaal, S. M., B. C. Haller, and K. E. Lotterhos. 2022. Inversion invasions: when the genetic basis
of local adaptation is concentrated within inversions in the face of gene flow. Philos Trans
R Soc Lond B Biol Sci 377:20210200.

Schiffels, S. and R. Durbin. 2014. Inferring human population size and separation history from
multiple genome sequences. Nature Genetics 46:919-925.

Schluter, D. 2009. Evidence for ecological speciation and its alternative. Science 323:737-741.

Schluter, D. and L. H. Rieseberg. 2022. Three problems in the genetics of speciation by selection.
Proc. Natl. Acad. Sci. 119:e2122153119.

Seitz, A. and M. Komma. 1984. Genetic polymorphism and its ecological background in tephritid

populations (*Diptera: Tephritidae*). Pp. 143-158 *in* K. Wöhrmann, and V. Loeschcke, eds.

Population Biology and Evolution. Springer Berlin Heidelberg, Berlin, Heidelberg.

Serrato-Capuchina, A. and D. R. Matute. 2018. The role of transposable elements in speciation.

Genes 9.

Shih, P.-Y., A. Sugio, and J.-C. Simon. 2023. Molecular mechanisms underlying host plant

specificity in aphids. Annu. Rev. Entomol. 68:431-450.

Singh, K. S., B. J. Troczka, A. Duarte, V. Balabanidou, N. Trissi, L. Z. Carabajal Paladino, P.

Nguyen, C. T. Zimmer, K. M. Papapostolou, E. Randall, B. Lueke, F. Marec, E. Mazzoni,

M. S. Williamson, A. Hayward, R. Nauen, J. Vontas, and C. Bass. 2020. The genetic

architecture of a host shift: an adaptive walk protected an aphid and its endosymbiont from

plant chemical defenses. Science Advances 6:eaba1070.

Skotte, L., T. S. Korneliussen, and A. Albrechtsen. 2013. Estimating individual admixture

proportions from next generation sequencing data. Genetics 195:693-702.

Smit, A. F., R. Hubley, and P. Green. 2015. RepeatMaster Open-4.0,

http://www.repeatmasker.org.

Sproul, J. S., S. Hotaling, J. Heckenhauer, A. Powell, A. M. Larracuente, J. L. Kelley, S. U. Pauls,

and P. B. Frandsen. 2022. Repetitive elements in the era of biodiversity genomics: insights

from 600+ insect genomes. bioRxiv:2022.2006.2002.494618.

Stankowski, S. and M. Ravinet. 2021. Defining the speciation continuum. Evolution 75:1256-

1273.

Stuart, Y. E., T. Veen, J. N. Weber, D. Hanson, M. Ravinet, B. K. Lohman, C. J. Thompson, T.

Tasneem, A. Doggett, R. Izen, N. Ahmed, R. D. H. Barrett, A. P. Hendry, C. L. Peichel,

and D. I. Bolnick. 2017. Contrasting effects of environment and genetics generate a

continuum of parallel evolution. Nat. Ecol. Evol. 1:0158.

Szpiech, Z. A. and R. D. Hernandez. 2014. selscan: an efficient multithreaded program to perform

EHH-based scans for positive selection. Mol Biol Evol 31:2824-2827.

Taylor, E. B., J. W. Boughman, M. Groenenboom, M. Sniatynski, D. Schluter, and J. L. Gow.

2006. Speciation in reverse: morphological and genetic evidence of the collapse of a three-

spined stickleback *(Gasterosteus aculeatus)* species pair. Mol. Ecol. 15:343-355.

Tuttle, Elaina M., Alan O. Bergland, Marisa L. Korody, Michael S. Brewer, Daniel J. Newhouse,

P. Minx, M. Stager, A. Betuel, Zachary A. Cheviron, Wesley C. Warren, Rusty A. Gonser,

and Christopher N. Balakrishnan. 2016. Divergence and functional degradation of a sex

chromosome-like supergene. Curr. Biol. 26:344-350.

UniProt Consortium. 2021. UniProt: the universal protein knowledgebase in 2021. Nucleic Acids

Res 49:D480-d489.

van Schooten, B., J. Meléndez-Rosa, S. M. Van Belleghem, C. D. Jiggins, J. D. Tan, W. O.

McMillan, and R. Papa. 2020. Divergence of chemosensing during the early stages of

speciation. Proc. Natl. Acad. Sci. 117:16438-16447.

Vicoso, B. and D. Bachtrog. 2015. Numerous transitions of sex chromosomes in Diptera. PLoS.

Biol. 13:e1002078.

Vidal, M. C. and S. M. Murphy. 2018. Bottom-up vs. top-down effects on terrestrial insect

herbivores: a meta-analysis. Ecol. Lett. 21:138-150.

Villoutreix, R., C. F. de Carvalho, V. Soria-Carrasco, D. Lindtke, M. De-la-Mora, M. Muschick,

J. L. Feder, T. L. Parchman, Z. Gompert, and P. Nosil. 2020. Large-scale mutation in the

evolution of a gene complex for cryptic coloration. Science 369:460-466.

Wellenreuther, M. and L. Bernatchez. 2018. Eco-evolutionary genomics of chromosomal

inversions. Trends Ecol. Evol. 33:427-440.