Long term trend and short-term dynamics of a willow ptarmigan population

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Abstract

Willow ptarmigan *Lagopus lagopus* is abundant in Arctic and tundra regions, but rapid climate warming has raised concerns about possible declines as has been observed in several bird species. In this study, we used a hierarchical state-space model to analyze data from a 139 km line transects in mid-Sweden over 48 years. Adult numbers and breeding success were analyzed separately, and we included covariates on vole abundance, numbers of snow-free days in autumn and spring, and the last day of frost in May. We assessed long-term trends in the adult population and estimated the effects of breeding success and weather variation on short-term changes.

The estimates of adult density did not show any trend for the period 1976 to 2023, and the dynamics were characterized by a strong direct negative density dependence indicating a stationary process. A number of possible mechanisms have been suggested for how a warmer climate affects willow ptarmigan population dynamics, but our results do not support the hypothesis that lack of snow in autumn and spring increases the vulnerability of willow ptarmigan to predation and leads to population decline. Breeding success is an important driver of changes in, but independent of, adult density. In addition to predation, we propose that climate conditions and emerging vegetation during egg formation and laying is important. We suggest that our results can be explained by a diverse predator assemblage that makes it difficult for the population to escape top-down control, resulting in short-term fluctuations at lower densities.

Keywords: Lagopus lagopus trend population dynamics

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Introduction

Negative trends and population declines have been reported for several bird species in the northern hemisphere, and climate change has been identified as the most important process behind the declines (Smith et al., 2020). Willow ptarmigan *Lagopus lagopus* is an abundant herbivore found in the Arctic and tundra regions (Storch, 2007). It has an extensive distribution from the open tundra to moorlands and subarctic forests at lower altitudes. Willow ptarmigan, with its white winter plumage, is adapted to harsh winter conditions, and it burrows in snow as protection from cold weather and predation (Andreev, 1991; Hoglund, 1980). Willow ptarmigan exhibits large annual fluctuations attributed to variation in breeding success (Myrberget, 1974; Steen and Erikstad, 1996). Adult mortality, nest loss, and chick mortality have mostly been attributed to predation (Erikstad et al., 1982; Marcström and Höglund, 1980; Smith and Willebrand, 1999; Steen et al., 1988). Population change show negative density–dependent growth (Pedersen et al., 2004), but the variation in breeding success is not related to adult abundance (Hörnell-Willebrand et al., 2006).

A recent review of the long-term population trends of willow ptarmigan identified negative trends in parts of its distribution (Fuglei et al., 2020), and several potential factors could affect adult survival and breeding success negatively (Henden et al., 2017). The ongoing changes in climate is expected to result in winters with shorter snow seasons, which could increase the vulnerability of the white ptarmigan on snow-free ground to birds of prey (Henden et al., 2020; Melin et al., 2020). On the other hand, air temperature after snow-melt is important for the timing of spring, with warmer conditions contributing to earlier phenology in vegetation communities (Kelsey et al., 2021). Early onset of spring with appearance of new nutritious vegetation positively affects nutritional status and breeding conditions (Brittas, 1988; Eriksen et al., 2023; Erikstad and Andersen, 1983; Marcström and Höglund, 1980).

Historically, willow ptarmigan has been an important game species and commodity in game markets (Hjeljord and Loe, 2022; Lloyd, 1867; Sotherton et al., 2009). Today, its value as a game for consumption has declined, and it is mostly a species of interest for sport hunting. However, the need for harvest management has been addressed in recent studies (Aanes et al., 2002; Pedersen et al., 2004; Smith and Willebrand, 1999). In response to these concerns, national schemes of line transects with pointing dogs and distance sampling (Thomas et al., 2010) have been developed to estimate densities in Sweden, Norway and Finland the last 15 - 25 years (Eriksson et al., 2006; Helle et al., 2016; Nilsen et al., 2024). Systematically collected field data over several decades provide important insights in both short- and long-term dynamics of ecological systems (Lindenmayer et al., 2012; Magurran et al., 2010). Time series of harvest statistics and catch data have provided valuable insights into population dynamics (Keith, 1963; Royama, 1992). However, long-term studies on counts and estimates of abundance are less common among wildlife species, but see (Jacobson et al., 2004; Ranta et al., 1995; Sæther et al., 2008).

The recent development of hierarchical state-space models in a Bayesian framework has improved the possibility of jointly analyzing several different data sources of wildlife time series and increased the possibility of identifying factors that are important for population change (de Valpine and Hastings, 2002; Kéry and Royle, 2016; Lee et al., 2015). A great advantage is that they can provide separate estimates of observation and process error, which otherwise tend to

bias estimates of model parameters (Aanes et al., 2002; Freckleton et al., 2006). These models
are well suited to iteratively using new data, updating short-term predictions, and supporting
management decisions (Bowler et al., 2020; Frost et al., 2022; Henden et al., 2020). The main
aim of this study was to investigate the drivers of short-term population dynamics of the adult
willow grouse and assess potential population trends over the last 50 years. To achieve this
we used a unique set of line-transects from the Lövhögen area in Sweden collected between
1963 and 2023 and developed a hierarchical state-space model to estimate changes in adult
population density and breeding success. We estimated the effects of vole abundance and spring
conditions on breeding success, and their effect on adult population change. The lack of snow in
autumn and spring was also used as predictor of adult change. The model includes parameters
for density dependence and a linear trend for adult change.

Material and methods

Study Area

The study area, Lövhögen 80 km^2, is located in the southern part of (62'N, 13'E) of the
Swedish alpine tundra, where the landscape is gradually transformed into continuous boreal
conifer forest. The altitude varies between 750 and 1 000 m.a.s.l. and is dominated by birch for-
est and open tundra (Brittas, 1988). The tree line is about 800 m.a.s.l., and about 20% is wooded
mainly with mountain birch *Betula* spp.. There are small scattered stands of Norway spruce *Picea*
abies and Scots pine *Pinus silvestris* above the tree line and a few scattered willow *Salix spp.*
thickets. Almost one-third of the area is peatland, characterized by sedge *Carex* spp., cloudberry
Rubus chamaemorus and cotton grass *Eriophorum* spp. About half of the study area is heathland,
dominated by dwarf shrubs such as heather *Calluna vulgaris*, bilberry *Vaccinium myrtillus*, crow
berry *Empetrum hermaphroditum* and dwarf-birch *Betula nana*. The climate is continental, with
temperatures averaging -10 C° in January and snow cover stay for about 6 months in winter.
The average temperature is +14 C° for July. There are few lakes, but frequent small creeks drain
the area into larger streams. The study area is privately owned by a single landowner, and the
harvest of willow grouse is highly restricted (Aanes et al., 2002).

Data

Line transect survey

A permanent monitoring program based on line transect surveys using pointing dogs was
established in 1963 to estimate the abundance of adult willow ptarmigan. A total of 18 lines,
600 m apart, with a total distance of 139 km cover the entire study area. The start and end
points of each line were marked with permanent poles. These lines have remained unchanged
since the survey began. The perpendicular distance to the observed willow ptarmigans was used
to estimate the probability of detection. Initially, the effective strip width (ESW) was calculated
as the average of the observed distances within 200 m of the line. In the period A between
1963 and 2004, counts were made in late June. The perpendicular distance to all detected
adults were registered and, sex was determined by plumage and/or behavior. It was not possible
to determine the sex of some adults, and the observed ratio of males to females was used to
estimate the number of undetermined sex. In late June, females are usually incubating, and are
therefore very difficult to detect. Therefore, the estimated number of adult males was assumed
to correspond to a pair of two adults. Unfortunately, the distances between 1963 and 1975

were destroyed when the Boda wildlife research station burned down, but the estimated adult densities for that period are still available. The distances were lost in 1979, 1980, 1985 and 1986, and registered as missing values in the data. The model development was restricted to the period with available distances that starting in 1976. The original estimates of adult densities between 1963 and 1975 are presented in the appendix. In period B from 2004, the study area was included in the national monitoring program for willow ptarmigan in the tundra (Eriksson et al., 2006), and the counting was delayed to late July or early August, similar to other areas. In period B, it was possible to observe both adult and brood size in the field. It was not assumed that a brood was always accompanied by a pair because sex segregation among adults may occur already in August (Marcström and Höglund, 1980). The number of adult observations per year varied between 73 and 206, with a median of 129 during the study period. The survey was completed within a week, and most members of the monitoring team have remained for many years.

Brood size

Between 1963 and 1994, brood size was estimated from the age ratio of harvested willow ptarmigan. Harvest management in Lövhögen is aimed to sample the population and differs from many other areas. Hunting occur during 5 - 7 days after the season opens 25th of August, and a previous estimate of harvest rate was on average 0.144 but with large variation (s.d. = 0.106) (Aanes et al., 2002). Hunters are divided into 2-4 teams with several hunters and one professional dog handler. The area is divided into sections that are visited only once to ensure that the effort is equal in all parts of the area. In 1963 and 1971, the relationship between the age distribution of flushed willow ptarmigan in August was compared to the age ratio of harvested willow ptarmigan in early September. The difference in age ration was minor, 66% and 74% in counts and 63% and 70% in harvested birds (Marcström and Höglund, 1980). From 2004, when counts were made in early August, we used the number of observed chicks per two adults as the average brood size.

Vole numbers

Voles were snap-trapped in late June and later in early August during in the same time as the willow ptarmigan surveys, and they accumulated between 200 and 600 trap nights each year. The logarithm-transformed number of voles per 100 trap nights was used as a vole index (Hörn-feldt, 1978), data were standardized for each period and then combined. Three years lacked observations (1976, 1977 and 1990). The first initial two years of the study were replaced by the median of the complete time series. It was possible to replace the third missing observation in the series with the average for 1989 and 1991.

Meteorological data

Data on snow depth was obtained from the Swedish Meteorological and Hydrological Insti-tute for the Särna station (61.70, 13.13), 437 m.a.s.l. The station is about 28 km southwest of Lövhögen. Data were not available for May in 2000 due to the renovation of the station; there-fore, we used the average number of observations for each time and day in May in 1999 and 2001 as data for 2000.

The number of snow-free days was extracted for spring (April and May) and autumn (October and November). The number of snow-free days was standardized and included as a covariate

of adult change. We extracted the ambient temperatures for all days in May at 03:00 h during
the study. The last day with a temperature less than -2.2C° has been validated as a measure
of spring vegetation growth (Schwartz et al., 2006). We used as an index of annual variation in
availability of newly-growing and nutrient-rich plants. The dates were standardized and included
as covariate in the model for breeding success.

Analysis

Table 1 – Overview of the parameters estimated in the hierarchical models of adult density and breeding success. The breeding success model provide input to the model of adult change. For more information refer to the analysis section.

Model	Parameter
Population dynamics	Breeding success the previous year
	Snow-free days in spring the same year
	Snow-free days in autumn the previous year
	Density dependence
	Linear trend
Breeding success	Vole abundance
	Late frost

We developed a hierarchical state-space model in JAGS (Plummer et al., 2003) following
(Henden et al., 2020; Kéry and Royle, 2016). We used Markov Chain Monte Carlo (MCMC)
methods to estimate the posterior distribution of parameters and adult density estimates. The
model was run for 30 000 iterations with a burn-in of 8 000. The thinning was set to 1, and three
chains were used to evaluate convergence. The package MCMCvis (Youngflesh, 2018) was used
to evaluate the trace plots and posterior distribution of the model parameters. All parameters
had an R-hat value less than 1.01. The main model and submodel for breeding success were
validated by simulating replicate data and comparing predictions with both original and simulated
data (Schaub and Kéry, 2021). The Bayesian P-value was close to 0.5 for both the main and
breeding success model (0.49 and 0.41 respectively). Parameter estimates are presented with
a 95% credible interval (Cr. I), and the plot of the estimated time series of adult densities show
the 80% highest posterior density. All analyses and graphics were performed in R (R Core Team,
2024) and the JagsUI package was used as an interface to JAGS from R (Kellner, 2024).

Distance sampling and the probability of detection. Distance data were prepared for analysis using
a cutoff of 200 m and a bin width of 20 m. This resulted in 10 distance intervals, where each
observation was categorized into a specific distance interval (1-10). During the 48 years in which
distances were available we included 3531 distances less than 200 m. Thus, the line transects
covered an area of approximately 55 km^2.

The detection probability was computed in a multinomial model of cell probabilities, assum-
ing a half-normal detection function. See eq. 1 where x denotes the midpoint distance of the
distance class. All distances were pooled to estimate the detection probability, and we did not
add any covariates to the detection equation. However, we included separate intercepts of the
estimate of $log(\sigma_{A,B})$ to evaluate the potential effect of the different periods on the probability
of detection ($pcap_{A,B}$). Further information can be found in the model code in the Appendix, and
in chapter 9.7 in Kéry and Royle (2016).

(1)
$$g(x; \sigma) = exp(\frac{-x^2}{2\sigma^2})$$

Model of breeding success. We estimated the effects of vole abundance and spring phenology
on standardized brood size and used these estimates as an independent variable in the part of
the model that describes adult changes. This made it possible to estimate breeding success for
the period 1995 - 2003 when age ratio of harvested willow ptarmigan was not available due
to changes in harvest management. We used standardized data on average brood size, vole
abundance and late frost in May to estimate annual changes in breeding success. Data on brood
size were positively correlated with the vole index (0.324 (P = 0.044) but negatively correlated
with late frost (-0.322, P = 0.045). However, the vole index and late frost were not significantly
correlated (-0.132, P = 0.369). We assumed that the error terms were normally distributed, (eq.
2).Steen et al. (1988) present previous analysis on the effects of vole abundance and weather on
breeding success in our study area.

(2) $$\mu.chick_t = \beta_0 + \beta_{1vole} * voles_t + \beta_{2frost} * frost_t + \epsilon_t$$

Main model of adult change. We used two process models: one without covariates for the first
year and a second with covariates for subsequent years. In the section without covariates, the
initial density and observation error ($\mu_t = \beta 0_A$) are estimated (Sillett et al., 2012). The second
part include covariates and models population change in subsequent years (eq. 8) including
process variation (eq. 6, *PrOc*). The number of observations is assumed to follow a binomial
distribution based on the probability of observation (pcap) and the number of individuals (N) (eq.
3), which are assumed to show a Poisson distribution with mean λ (eq. 4). λ is the product of
density and the area covered by the line-transect surveys (eq. 5). The logarithm of the expected
density (μ_t) is assumed to be normally distributed (eq. 6). We then use a Gompertz-model to
estimate the change in adult densities ($log(D_t)$) in subsequent years. $\beta 0_{A,B}$ is the intercept for
the two different sampling periods, $\beta 1$ is the strength of density dependent and subtracted with
1 to scale the parameter such that 0 is lack of density dependence. $\beta 2$ is the parameter of the
linear trend, $\beta 3$ is the effect of variation in estimated breeding success, $\beta 4$ is the effect of snow-
free days in spring in the same year, and $\beta 5$ is the effect of snow-free days in autumn the previous
year.

(3) $$y_t \sim dbin(pcap_{A,B}, N_t)$$

(4) $$N_t \sim dpois(\lambda_t)$$

(5) $$\lambda_t = D_t * area_t$$

(6) $$log(D_t) \sim dnorm(\mu_t, PrOc)$$

(7) $$\mu_t = \beta 0_{A,B} + \beta 1 * \mu_{t-1} + \beta 2 * (t-1) +$$
$$\beta 3 * \mu.chick_{t-1} + \beta 4 * SnwSpr_t + \beta 5 * SnwAut_{t-1}$$

188   Following Royama (1992) we compared the first and last 20-years of the estimated adult
189 densities. We compared the mean (t-test) and variance (F-test) of the two periods, and examined
190 the autocorrelation plots for significant coefficients using the Bartlett band.

Figure 1 – The estimated dynamics of a) adult densities and b) breeding success during 1976 - 2023. The vertical lines show the 80% highest posterior density interval. The smooth stippled lines are produced by the loess function.

Results

Adult dynamics

Adult density was on average 5.4 adults per km^{-2}, with an interquartile range of 4.9 to 6.4 (Fig. 1a). The coefficient ($\beta2$) for the linear trend of logarithmic adult density was small and overlapped with zero, (0.007, 95%Cr.I. -0.007 to 0.021) (Fig. 2a). The mean logarithmic density for the first and second 20th year periods was 1.773 and 1.649 respectively (t = 1.700, df = 37.284, P = 0.097), and similar ratio of variance (F = 0.584, numerator df = 19, denominator df = 20, P = 0.246). In addition, the autocorrelation plots did not show any significant coefficient at time lags 1,2, ...10, indicating a stationary process.

The parameter for direct density dependence was -1.204 (*95%Cr.I.*, -1.472 to -0.871), which indicates a strong negative density dependence. The parameter for estimated breeding success

Figure 2 – Parameter values and 95% credible intervals for the explanatory variables of annual change in the a) adult density and b) breeding success.

was clearly positive, 0.439 (*95%Cr.I.*, 0.160 to 1.049). The parameters for snow free days in the previous autumn and current spring was positive but non-significant, 0.049 (*95%Cr.I.*, -0.048 to 0.144) and 0.023 (*95%Cr.I.*, -0.075 to 0.122), respectively. See figure 2a.

The errors for the first year (sdperctau) and subsequent years (sdproctau) were estimated to 0.595 (*95%Cr.I.* 0.077 to 0.980) and 0.212 (*95%Cr.I.*, 0.159 to 0.280) respectively. The intercepts for periods A and B of the second process model were 2.126 and 1.611, indicating a higher adult density in the first period but the 95% credible intervals greatly overlapped (1.551 - 2.675 and 0.966 - 2.359 respectively).

Breeding success, voles and late frost in May

Breeding was positively affected by vole abundance, (0.238 with *95%Cr.I.*, 0.045 to 0.478), but negatively affected by late frost in May (-0.337 with *95%Cr.I.*, -0.563 to -0.118; Fig. 2b). The adult density estimates did not correlate with breeding success, (-0.060, t = -0.409, df = 46, P = 0.684). The coefficients of the partial autocorrelation function at lags 1 and 3 were positive and 95% confidence intervals (Conf.I.) did not overlap zero, 0.426 (95%Conf.I. 0.143-0.709) and 0.382 (95%Conf.I. 0.099-0.283), respectively.

Changes in snow-free days and last frost in May

There was a large variation in the number of snow-free days in autumn and spring, and a significant linear increase trend over time, (slope=0.040 $R^2 = 0.274$, Std. Err. = 0.006, t= 6.093, P<0.0001). See figure 3a. From 1976 to about 1995, the last frost in May tended to occur increasingly later, followed by a period when the date of the last frost decreased to earlier dates again (Fig. 3b). The values of last frost in May was not correlated with the snow-free days in spring or autumn (r<0.011, P>0.94).

Detectability

The probability of detecting willow ptarmigan within the ±200 m strip around the line transect in period A and B was 0.441 (*95% Cr.I.*, 0.423 - 0.459) and 0.432 (*95% Cr.I.*, 0.412 - 0.453)

Figure 3 – a) The increase in snow-free days in autumn (October and November) and subsequent spring (April and May). The regression slope is positive and statistically significant, 0.040 (Std. Err. = 0.006 t= 6.093, P<0.0001). b) Last day in May with temperature below -2.2C°. The smooth stippled lines are produced by the loess function.

respectively. The average of these two estimates (0.436) corresponds to an effective strip width of 87.2 m, which is substantially longer than the 63.8 m that was used previously in the period 1963 - 1975. (See figure 4 in appendix.)

Discussion

The estimates of adult density did not show any trend for the period 1976 to 2023, and the dynamics were characterized a strong direct negative density dependence. We propose that the functional response by the predator guild in the region is the most likely process that explains these results. In addition, large-scale natal dispersal and recruitment of juvenile females can also contribute to stabilizing the population (Hörnell-Willebrand et al., 2014; Pulliam and Danielson, 1991). Earlier studies have shown that predation on young and adult willow ptarmigan is high during autumn, both in areas open and closed to harvest (Eriksen et al., 2025; Israelsen et al., 2020; Smith and Willebrand, 1999). However, our results do not support the hypothesis that lack of snow in autumn and spring increases vulnerability to predation and population decline, as suggested by Melin et al. (2020). In fact, recent studies have shown that in northern areas, more precipitation during winter with still below freezing temperatures leads to more snow in early spring, which might counteract earlier snowmelt from increase spring temperature (Bjerke et al., 2025). While this is one of the longest existing time series on willow ptarmigan density in Scandinavia, there is some uncertainties about the trend assessment since 1) the timing of the yearly monitoring was change during the course of the study and 2) the initial years of this time series (1963-1975) could not be included in the distance sampling model due to missing data, while this period appears to show the most extreme fluctuations and the highest adult estimates in the original time series. In fact a population decline during the late 60's with short-term fluctuations at lower densities seems to be in line with the population decline in willow ptarmigan in southeastern Norway during the last 150 years (Hjeljord and Loe, 2022). Similar declines and changed fluctuation patterns were reported in capercaillie (*Tetrao urogallus* and black grouse

Lyrurus tetrix during the 60's (Jahren et al., 2016). Further, the climate data (snow and temperature) were retrieved from the weather station closest to the study area, which was located at a considerably lower elevation (437 m.a.s.l compared to the ptarmigan monitoring at approximately 750 – 1100 m.a.s.l), potentially representing different climatic conditions relative to the study area. Despite these challenges, we believe this analysis provide valuable insight from a unique long term time-series about the population dynamics of willow ptarmigan, and supports many of the conclusions on willow ptarmigan biology from earlier studies.

Breeding success is an important driver of changes in, but independent of, adult density. Our findings agree with numerous previous studies that have concluded that predation on nests and chicks follows fluctuations in voles and lemmings (Steen and Haugvold, 2009; Steen et al., 1988). Willow ptarmigan is well adapted to winters with deep snow, and the birch forest provides an ample source of twigs and catkins (Brittas, 1988; Hoglund, 1980). Small herbivores in northern areas feed regularly, emphasizing energy conservation rather than acquiring energy reserves (Thomas, 1987). Extremely cold weather and high precipitation during the first days after hatching can be detrimental (Bowler et al., 2020; Erikstad and Spidsø, 1982; Marcström and Höglund, 1980), but Steen et al. (1988) showed that the variation in weather after hatching was poorly correlated with breeding success. In stead, we propose that climate conditions during egg formation and laying are more important. Females generally form eggs when plants begin to grow in spring (Moss, 1997), and late spring is associated with delayed egg laying (Wiebe and Martin, 1995). In our study area, the variation in hatching date was on average 12 days (17 - 29 June) (Marcström and Höglund, 1980), and Brittas (1988) showed that the digestibility of spring food in willow ptarmigan crops was positively correlated with both female body condition during egg laying and subsequent breeding success. This is in accordance with the maternal hypothesis, which states that female body condition during egg laying and incubation affects the survival of chicks and subsequent recruitment (Blomqvist et al., 1997; Coslovsky and Richner, 2011). The negative effect of late frost in May reduces the availability of newly-growing and nutrient-rich plants, such as shoots of cotton grass *Eriophorum* spp. We believe it is important to explore the potential for vole abundance in early summer and the date for the last frost in May to provide an early indication of the average brood size in August.

Despite large annual variations, the long-term average onset of spring began earlier in our study area (Jin et al., 2019). However, we did not observe any effect of the increase in snow-free days in autumn and spring, but there are a number of possible mechanisms by which a warmer climate could affect willow ptarmigan population dynamics. For example, short periods of rain in winter can form an icy crust on both snow and birch twigs, which may force willow ptarmigans to leave an area (Hoglund, 1980). Another mechanism would be changes in the vegetation community (Bråthen et al., 2024; Tuomi et al., 2024).

The characteristics of the willow ptarmigan population dynamics are likely to vary depending on the food-web structure. The structure of trophic levels becomes simpler in the north compared to further south (Callaghan et al., 2004b), where the tundra becomes fragmented and surrounded by continuous boreal forests. At lower latitudes, animal diversity increases with various herbivores and predators (Callaghan et al., 2004a). Patterns of population dynamics depend on trophic interactions and time lags (Turchin and Taylor, 1992), and we expect willow ptarmigan populations to show increased densities and amplitudes along a geographical gradient from south to north similar to vole populations in Scandinavia (Erlinge, 1987; Soininen et

al., 2025; Turchin and Hanski, 1997) and snowshoe hares (*Lepus americanus*) in North America (Krebs, 2010). Our study area is in the southern range of the tundra, and we suggest that a diverse predator assemblage makes it difficult for the population to escape a top-down control. Further north, with fewer predator species, it may be more common to escape top-down control that might result in higher amplitudes and densities of willow ptarmigan populations.

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Conflict of interest disclosure

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Figure 4 – The original time series based on a strip width estimate of twice the average distances recorded. Note that distance measurements between 1963 to 1975 was lost in a fire.

314   **Original estimates of adult densities, 1963 - 2003**

Script and codes

```
model{

# Priors

for(i in 1:2){
alpha0[i] ~ dunif(-10,10)  # Intercept for sigma in half-.normal detection function
}

# Different intercept for the two different line transect periods, late June or early August

for(i in 1:2){
b0[i] ~ dunif(-10, 10)
}

b3 ~ dunif(-10, 10)    # Parameter for the effect of brood size
b1 ~ dunif(-1,1)       # Parameter for density dependence
b2 ~ dunif(-10,10)     # Parameter for the trend parameter (i.e. Year)
b4 ~ dunif(-10,10)     # Parameter for snow-free days in spring
b5 ~ dunif(-10,10)     # Parameter for snow-free days in spring (t-1)

## Precision (T=1) and process variation (T>1). Expressed as the standard deviation

PrEc <- 1 / (sdprectau * sdprectau)
sdprectau ~ dunif(0, 1)
PrOc <- 1 /(sdproctau * sdproctau)
sdproctau ~ dunif(0,1)

# Priors for estimating brood size (chicks per pair)

b0_chick ~ dunif(-10, 10)  # Intercept for brood size dependent on vole
b1_vole   ~ dunif(-10, 10)    # Parameter for the effect of vole abundance
b2_frost   ~ dunif(-10, 10)    # Parameter for the effect of vole abundance
chick.err <-  1 / (sdchick * sdchick)
sdchick ~ dunif(0, 1)

#############################################
## Distance sampling observation model for observations (binned)
## All lines are combined into one, same lines were surveyed all years

# Distance data:
# Construct cell probabilities for nD multinomial cells

for(i in 1:nobs){
    dclass[i] ~ dcat(fc[perII[i],]+ 0.01)  # 0.01 must be added for the model to converge
}
## End multinomial cells

for(s in 1:2){  # Two time periods of line transect surveys

for(g in 1:nD){    # Distance classes

# Half-normal detection function
  log(p[s,g]) <- -midpt[g] * midpt[g] / (2 *sigma[s]*sigma[s])
  pi[s,g] <- delta / B
  f[s,g] <- p[s,g] * pi[s,g]
  fc[s,g] <- f[s,g] / pcap[s]
 }   ## End detection

pcap[s] <- sum(f[s,]) # Pr(detection): sum of rectangular areas

log(sigma[s]) <- alpha0[s]# Linear detection probability model
}

## Likelihood
## Process model:

# Initial density model for Year 1
      y[1] ~ dbin(pcap[1], N[1])
```

```
386        N[1] ~ dpois(lambda[1])
387        lambda[1] <-  D[1] * area[1]
388        logD[1] ~ dnorm(mu[1], PrEc)
389        mu[1] <- b0[1]
390        D[1] <- exp(logD[1])
391
392 # Population dynamics model for subsequent years:
393     for (t in 2:T){
394        y[t] ~ dbin(pcap[Period[t]], N[t])
395        N[t] ~ dpois(lambda[t])
396        lambda[t] <- D[t] * area[t]
397        logD[t] ~ dnorm( mu[t] , PrOc )
398        mu[t] <- b0[Period[t]] + b1 * mu[t-1]  + b2 * (t-1) + b3 * mu_chick[t-1] + b4 * SnwSpr[t] + b5 *
399     SnwAut[t-1]
400        D[t] <- exp(logD[t])
401        }
402
403        bdd <- b1-1 # Adjusting parameter for density dependence for comparison with other parameters
404
405 # Add posterior predictive checks for main model
406 # Simulated observed counts based on the detection probability
407        for (t in 1:T){
408        y_Sim[t] ~ dbin(pcap[Period[t]], N[t])
409        y_Est[t] <- N[t]*pcap[Period[t]]
410        res1[t] <- y_Est[t] - y[t] # Comapre residuals with predictions
411       D_obs[t] <- pow(y[t] - y_Est[t], 2) / y_Est[t]
412       D_rep[t] <- pow(y_Sim[t] - y_Est[t], 2) / y_Est[t]
413        }
414
415        fit1 <- sum(D_obs)
416        fit2 <- sum(D_rep)
417
418 ## Model of brood size with vole index and frost in May as the predictors
419
420   for(i in 1:T){
421        # regression model on chick, given voles
422        chick[i] ~ dnorm(mu_chick[i], chick.err )
423        mu_chick[i] <- b0_chick + b1_vole * voles[i] + b2_frost * Frost[i]
424
425 # Add posterior predictive checks for chick model
426
427        y_Sim_chick[i] ~ dnorm(mu_chick[i], chick.err )
428        y_Est_chick[i] <- mu_chick[i]
429        res1_chick[i] <- y_Est_chick[i] - chick[i]
430        sq_res1_chick[i] <- pow(res1_chick[i],2)
431        res2_chick[i] <- y_Sim_chick[i] - mu_chick[i]
432        sq_res2_chick[i] <- pow(res2_chick[i],2)
433      } # End chick estimation
434
435        fit1_chick <- sum(sq_res1_chick[])
436        fit2_chick <- sum(sq_res2_chick[])
437
438 } # End model
```

```
441 library(tidyverse)
442 library(here)
443 library(jagsUI)
444 library(MCMCvis)
445 library(data.table)
446 data <- "DATA"
447 model <- "SCRIPT"
448 script2 <- "ADD_SCRIPT"
449 add_dat <- "ADD_DATA"
450 latex <- "LATEX/figures"
451
452 dat <- readRDS(here(data, "LovDatFinal2025.rds"))
453
454 # inits
455 inits <-Nst_init <- dat2$y + 1
456 inits <- function(){list(N=Nst_init)}
457
```

```
458  ni <- 30000   ;    nb <- 8000    ;   nt=1  ;    nc <- 3
459
460  para.check  <- c("y_Est", "fit1", "fit2", "res1", "mu_chick", "res1_chick", "fit1_chick", "fit2_chick")
461  para.names <- c("b0","bdd", "b2", "b3", "b4", "b5" , "pcap", "sdprectau", "sdproctau", "b0_chick", "b1_vole
462       ", "btCHICK" , "b2_frost")
463  para.pop <-c("D", "mu_chick")
464  para<-c(para.check, para.names, para.pop)
465
466  mod_maj <- jags (dat2, inits, para.check, here(model, "WillowGrouseModelFinalMaj14.jags"), n.thin=nt, n.
467       chains=nc, n.burnin=nb, n.iter=ni, parallel=TRUE)
468
469  print(mod_maj, dec=3)
470  any(unlist(mod_maj$Rhat)  > 1.05)
471
472  ## EVALUATE MODEL
473
474  # Discrepancy paramters. See main part of model.
475  FIT1   <- mod_maj$sims.list$fit1
476  FIT2   <- mod_maj$sims.list$fit2
477  plot(FIT2, FIT1)
478  abline(0,1)
479  mean(FIT2<FIT1) # Bayesian p value
480
481
482  # Discrepancy paramters. See chick part of model.
483  FIT1_chick  <- mod_maj$sims.list$fit1_chick
484  FIT2_chick  <- mod_maj$sims.list$fit2_chick
485  plot(FIT1_chick, FIT2_chick)
486  abline(0,1)
487  mean(FIT2_chick>FIT1_chick) # Bayesian p value
488
489  ############
490
491      Data and scripts are available online.
492
```

References

Aanes S, Engen S, SÆther BE, Willebrand T, Marcström V (2002). *Sustainable Harvesting Strategies of Willow Ptarmigan in a Fluctuating Environment. Ecological Applications* **12**, 281–290. `https://doi.org/10.1890/1051-0761(2002)012[0281:SHSOWP]2.0.CO;2`. URL: `http://doi.wiley.com/10.1890/1051-0761(2002)012%5B0281:SHSOWP%5D2.0.CO;2` (visited on 05/13/2022).

Andreev AV (1991). *Winter Adaptations in the Willow Ptarmigan. Arctic* **44**, 106–114. JSTOR: `40511070`. URL: `https://www.jstor.org/stable/40511070` (visited on 06/24/2024).

Bjerke JW, López-Blanco E, Tømmervik H, Striberny A, Davids C, Ólafsdóttir R, Karlsen SR, Sandström P, Turunen M, Rikkonen T, Arneberg MK, Siikavuopio S, Zinglersen K, Lynge-Pedersen K, Sandström S, Rautio P (2025). *Nordic Boreo-Arctic Lands under Rapid Climatic Change: A Review of Recent and Future Trends and Extreme Events. Earth-Science Reviews* **261**, 105012. `https://doi.org/10.1016/j.earscirev.2024.105012`. URL: `https://linkinghub.elsevier.com/retrieve/pii/S0012825224003404` (visited on 05/11/2025).

Blomqvist D, Johansson OC, Götmark F (1997). *Parental Quality and Egg Size Affect Chick Survival in a Precocial Bird, the Lapwing Vanellus Vanellus. Oecologia* **110**, 18–24. `https://doi.org/10.1007/s004420050128`. URL: `https://doi.org/10.1007/s004420050128` (visited on 09/13/2024).

Bowler DE, Kvasnes MAJ, Pedersen HC, Sandercock BK, Nilsen EB (2020). *Impacts of Predator-Mediated Interactions along a Climatic Gradient on the Population Dynamics of an Alpine Bird. Proceedings of the Royal Society B: Biological Sciences* **287**, 20202653. `https://doi.org/10.1098/rspb.2020.2653`. URL: `https://royalsocietypublishing.org/doi/10.1098/rspb.2020.2653` (visited on 09/06/2023).

Bråthen KA, Tuomi M, Kapfer J, Böhner H, Maliniemi T (2024). *Changing Species Dominance Patterns of Boreal-Arctic Heathlands: Evidence of Biotic Homogenization. Ecography* **2024**, e07116. `https://doi.org/10.1111/ecog.07116`. URL: `https://nsojournals.onlinelibrary.wiley.com/doi/10.1111/ecog.07116` (visited on 06/30/2025).

Brittas R (1988). *Nutrition and Reproduction of the Willow Grouse Lagopus Lagopus in Central Sweden. Ornis Scandinavica* **19**, 49. `https://doi.org/10.2307/3676527`. JSTOR: `3676527`. URL: `https://www.jstor.org/stable/3676527?origin=crossref` (visited on 05/05/2020).

Callaghan TV, Björn LO, Chernov Y, Chapin T, Christensen TR, Huntley B, Ims RA, Johansson M, Jolly D, Jonasson S, Matveyeva N, Panikov N, Oechel W, Shaver G, Elster J, Henttonen H, Laine K, Taulavuori K, Taulavuori E, Zöckler C (2004a). *Biodiversity, Distributions and Adaptations of Arctic Species in the Context of Environmental Change. AMBIO: A Journal of the Human Environment* **33**, 404–417. `https://doi.org/10.1579/0044-7447-33.7.404`. URL: `http://www.bioone.org/doi/abs/10.1579/0044-7447-33.7.404` (visited on 08/27/2024).

Callaghan TV, Björn LO, Chernov Y, Chapin T, Christensen TR, Huntley B, Ims RA, Johansson M, Jolly D, Jonasson S, Matveyeva N, Panikov N, Oechel W, Shaver G, Henttonen H (2004b). *Effects on the Structure of Arctic Ecosystems in the Short- and Long-term Perspectives. AMBIO: A Journal of the Human Environment* **33**, 436–447. `https://doi.org/10.1579/0044-7447-33.7.436`. URL: `http://www.bioone.org/doi/abs/10.1579/0044-7447-33.7.436` (visited on 08/27/2024).

Coslovsky M, Richner H (2011). *Predation Risk Affects Offspring Growth via Maternal Effects. Functional Ecology* **25**, 878–888. `https://doi.org/10.1111/j.1365-2435.2011.01834.x`. URL:

https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2435.2011.01834.x
(visited on 09/13/2024).

Valpine P, Hastings A (2002). *Fitting Population Models Incorporating Process Noise and Observa-*
tion Error. Ecological Monographs **72**, 57–76. https://doi.org/10.1890/0012-9615(2002)
072[0057:FPMIPN]2.0.CO;2. URL: https://onlinelibrary.wiley.com/doi/abs/
10.1890/0012-9615%282002%29072%5B0057%3AFPMIPN%5D2.0.CO%3B2 (visited on
03/27/2025).

Eriksen LF, Harald Ringsby T, Israelsen MF, Moa PF, Grøtan V, Nilsen EB (2025). *Climatic Variation*
Affects Seasonal Survival of an Alpine Bird Species. Wildlife Biology **n/a** (), e01386. https://doi.
org/10.1002/wlb3.01386. URL: https://onlinelibrary.wiley.com/doi/abs/10.1002/
wlb3.01386 (visited on 03/31/2025).

Eriksen LF, Ringsby TH, Pedersen HC, Nilsen EB (2023). *Climatic Forcing and Individual Hetero-*
geneity in a Resident Mountain Bird: Legacy Data Reveal Effects on Reproductive Strategies. Royal
Society Open Science **10**, 221427. https://doi.org/10.1098/rsos.221427. URL: https://
royalsocietypublishing.org/doi/full/10.1098/rsos.221427 (visited on 03/24/2025).

Eriksson T, Andersson J, Byström P, Hörnell-Willebrand M, Laitila T, Sandström C, Willebrand
T (2006). *Fish and Wildlife in the Swedish Mountain Region: Resources, Use and Management.*
International Journal of Biodiversity Science & Management **2**, 334–342. https://doi.org/
10.1080/17451590609618154. URL: https://www.tandfonline.com/doi/full/10.1080/
17451590609618154 (visited on 02/16/2021).

Erikstad KE, Andersen R (1983). *The Effect of Weather on Survival, Growth Rate and Feeding Time in*
Different Sized Willow Grouse Broods. Ornis Scandinavica (Scandinavian Journal of Ornithology)
14, 249–252. https://doi.org/10.2307/3676311. JSTOR: 3676311. URL: https://www.
jstor.org/stable/3676311 (visited on 06/23/2024).

Erikstad KE, Blom R, Myrberget S (1982). *Territorial Hooded Crows as Predators on Willow Ptarmi-*
gan Nests. The Journal of Wildlife Management **46**, 109–114. https://doi.org/10.2307/
3808413. JSTOR: 3808413. URL: https://www.jstor.org/stable/3808413 (visited on
03/21/2025).

Erikstad KE, Spidsø TK (1982). *The Influence of Weather on Food Intake, Insect Prey Selection and*
Feeding Behaviour in Willow Grouse Chicks in Northern Norway. Ornis Scandinavica (Scandina-
vian Journal of Ornithology) **13**, 176–182. https://doi.org/10.2307/3676295. JSTOR:
3676295. URL: https://www.jstor.org/stable/3676295 (visited on 03/27/2025).

Erlinge S (1987). *Predation and Noncyclicity in a Microtine Population in Southern Sweden. Oikos*
50, 347–352. https://doi.org/10.2307/3565495. JSTOR: 3565495. URL: https://www.
jstor.org/stable/3565495 (visited on 08/24/2024).

Freckleton RP, Watkinson AR, Green RE, Sutherland WJ (2006). *Census Error and the Detection of*
Density Dependence. Journal of Animal Ecology **75**, 837–851. https://doi.org/10.1111/j.
1365-2656.2006.01121.x. URL: https://onlinelibrary.wiley.com/doi/abs/10.1111/
j.1365-2656.2006.01121.x (visited on 05/25/2025).

Frost F, McCrea R, King R, Gimenez O, Zipkin E (2022). *Integrated Population Models: Achieving*
Their Potential. Journal of Statistical Theory and Practice **17**, 6. https://doi.org/10.1007/
s42519-022-00302-7. URL: https://doi.org/10.1007/s42519-022-00302-7 (visited on
09/18/2023).

Fuglei E, Henden JA, Callahan CT, Gilg O, Hansen J, Ims RA, Isaev AP, Lang J, McIntyre CL, Merizon RA, et al. (2020). *Circumpolar Status of Arctic Ptarmigan: Population Dynamics and Trends.* Ambio **49**, 749–761.

Helle P, Ikonen K, Kantola A (2016). *Wildlife Monitoring in Finland: Online Information for Game Administration, Hunters, and the Wider Public. Canadian Journal of Forest Research* **46**, 1491–1496. `https://doi.org/10.1139/cjfr-2015-0454`. URL: `https://cdnsciencepub.com/doi/full/10.1139/cjfr-2015-0454` (visited on 10/15/2023).

Henden JA, Ims RA, Yoccoz NG, Asbjørnsen EJ, Stien A, Mellard JP, Tveraa T, Marolla F, Jepsen JU (2020). *End-User Involvement to Improve Predictions and Management of Populations with Complex Dynamics and Multiple Drivers. Ecological Applications* **30**, e02120. `https://doi.org/10.1002/eap.2120`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1002/eap.2120` (visited on 09/14/2023).

Henden JA, Ims RA, Fuglei E, Pedersen ÅØ (2017). *Changed Arctic-alpine Food Web Interactions under Rapid Climate Warming: Implication for Ptarmigan Research. Wildlife Biology* **2017** (SP1), wlb.00240. `https://doi.org/10.2981/wlb.00240`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.2981/wlb.00240` (visited on 06/21/2024).

Hjeljord O, Loe LE (2022). *The Roles of Climate and Alternative Prey in Explaining 142 Years of Declining Willow Ptarmigan Hunting Yield. Wildlife Biology* **2022**, e01058. `https://doi.org/10.1002/wlb3.01058`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1002/wlb3.01058` (visited on 08/15/2024).

Hoglund NH (1980). *Studies on the Winter Ecology of the Willow Grouse Lagopus Lagopus Lagopus L. Swedish Wildlife Research*, 249–270.

Hörnell-Willebrand M, Marcström V, Brittas R, Willebrand T (2006). *Temporal and Spatial Correlation in Chick Production of Willow Grouse Lagopus Lagopus in Sweden and Norway. Wildlife Biology* **12**, 347–355. `https://doi.org/10.2981/0909-6396(2006)12[347:TASCIC]2.0.CO;2`. URL: `http://www.bioone.org/doi/abs/10.2981/0909-6396%282006%2912%5B347%3ATASCIC%5D2.0.CO%3B2` (visited on 02/16/2021).

Hörnell-Willebrand M, Willebrand T, Smith AA (2014). *Seasonal Movements and Dispersal Patterns: Implications for Recruitment and Management of Willow Ptarmigan (* Lagopus Lagopus *): Dispersal and Movements of Willow Ptarmigan. The Journal of Wildlife Management* **78**, 194–201. `https://doi.org/10.1002/jwmg.650`. URL: `http://doi.wiley.com/10.1002/jwmg.650` (visited on 05/05/2020).

Hörnfeldt B (1978). *Synchronous Population Fluctuations in Voles, Small Game, Owls, and Tularemia in Northern Sweden. Oecologia*, 141–152.

Israelsen MF, Eriksen LF, Moa PF, Hagen BR, Nilsen EB (2020). *Survival and Cause-Specific Mortality of Harvested Willow Ptarmigan (Lagopus Lagopus) in Central Norway. Ecology and Evolution* **10**, 11144–11154. `https://doi.org/10.1002/ece3.6754`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1002/ece3.6754` (visited on 03/27/2025).

Jacobson AR, Provenzale A, Hardenberg A, Bassano B, Festa-Bianchet M (2004). *Climate Forcing and Density Dependence in a Mountain Ungulate Population. Ecology* **85**, 1598–1610. `https://doi.org/10.1890/02-0753`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1890/02-0753` (visited on 03/21/2025).

Jahren T, Storaas T, Willebrand T, Fossland Moa P, Hagen BR (2016). *Declining Reproductive Output in Capercaillie and Black Grouse – 16 Countries and 80 Years. Animal Biology* **66**, 363–400.

`https://doi.org/10.1163/15707563-00002514`. URL: `https://brill.com/view/`
`journals/ab/66/3-4/article-p363_11.xml` (visited on 05/05/2020).

Jin H, Jönsson AM, Olsson C, Lindström J, Jönsson P, Eklundh L (2019). *New Satellite-Based Esti-*
mates Show Significant Trends in Spring Phenology and Complex Sensitivities to Temperature and
Precipitation at Northern European Latitudes. International Journal of Biometeorology **63**, 763–
775. `https://doi.org/10.1007/s00484-019-01690-5`. URL: `http://link.springer.`
`com/10.1007/s00484-019-01690-5` (visited on 08/19/2024).

Keith LB (1963). *Wildlife's Ten-Year Cycle*. Madison: University of Wisconsin Press. 201 pp.

Kellner K (2024). *jagsUI: A Wrapper around 'rjags' to Streamline 'JAGS' Analyses*. manual. URL:
`https://CRAN.R-project.org/package=jagsUI`.

Kelsey KC, Pedersen SH, Leffler AJ, Sexton JO, Feng M, Welker JM (2021). *Winter Snow and*
Spring Temperature Have Differential Effects on Vegetation Phenology and Productivity across
Arctic Plant Communities. Global Change Biology **27**, 1572–1586. `https://doi.org/10.`
`1111/gcb.15505`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/gcb.`
`15505` (visited on 06/24/2024).

Kéry M, Royle JA (2016). *Applied Hierarchical Modeling in Ecology: Analysis of Distribution, Abun-*
dance and Species Richness in R and BUGS: Volume 1: Prelude and Static Models. Academic Press.

Krebs CJ (2010). *Of Lemmings and Snowshoe Hares: The Ecology of Northern Canada*. Proceedings
of the Royal Society B: Biological Sciences **278**, 481–489. `https://doi.org/10.1098/rspb.`
`2010.1992`. URL: `https://royalsocietypublishing.org/doi/full/10.1098/rspb.`
`2010.1992` (visited on 03/29/2025).

Lee AM, Bjørkvoll EM, Hansen BB, Albon SD, Stien A, Sæther BE, Engen S, Veiberg V, Loe LE,
Grøtan V (2015). *An Integrated Population Model for a Long-Lived Ungulate: More Efficient Data*
Use with Bayesian Methods. Oikos **124**, 806–816. `https://doi.org/10.1111/oik.01924`.
URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/oik.01924` (visited on
03/21/2025).

Lindenmayer DB, Likens GE, Andersen A, Bowman D, Bull CM, Burns E, Dickman CR, Hoffmann
AA, Keith DA, Liddell MJ, Lowe AJ, Metcalfe DJ, Phinn SR, Russell-Smith J, Thurgate N, Wardle
GM (2012). *Value of Long-Term Ecological Studies*. Austral Ecology **37**, 745–757. `https://doi.`
`org/10.1111/j.1442-9993.2011.02351.x`. URL: `https://onlinelibrary.wiley.com/`
`doi/abs/10.1111/j.1442-9993.2011.02351.x` (visited on 06/21/2024).

Lloyd L (1867). *The Game Birds and Wild Fowl of Sweden and Norway: With an Account of the Seals*
and Salt-water Fishes of Those Countries. F. Warne and Company. 734 pp.

Magurran AE, Baillie SR, Buckland ST, Dick JM, Elston DA, Scott EM, Smith RI, Somerfield PJ,
Watt AD (2010). *Long-Term Datasets in Biodiversity Research and Monitoring: Assessing Change*
in Ecological Communities through Time. Trends in Ecology & Evolution **25**, 574–582. `https:`
`//doi.org/10.1016/j.tree.2010.06.016`. URL: `https://linkinghub.elsevier.com/`
`retrieve/pii/S0169534710001552` (visited on 07/08/2024).

Marcström V, Höglund N (1980). *Factors Affecting Reproduction of Willow Grouse, Lagopus Lagopus,*
in Two Highland Areas of Sweden. Swedish Wildlife Research **11**, 285–314.

Melin M, Mehtätalo L, Helle P, Ikonen K, Packalen T (2020). *Decline of the Boreal Willow Grouse*
(Lagopus Lagopus) Has Been Accelerated by More Frequent Snow-Free Springs. Scientific Reports
10 (1), 6987. `https://doi.org/10.1038/s41598-020-63993-7`. URL: `https://www.`
`nature.com/articles/s41598-020-63993-7` (visited on 06/07/2023).

Moss R (1997). *Grouse and Ptarmigan Nutrition in the Wild and in Captivity*. *Proceedings of the Nutrition Society* **56**, 1137–1145. `https://doi.org/10.1079/PNS19970116`. URL: `https://www.cambridge.org/core/product/identifier/S0029665197000694/type/journal_article` (visited on 03/27/2025).

Myrberget S (1974). *Variations in the Production of the Willow Grouse Lagopus Lagopus (L.) in Norway, 1963-1972*. *Ornis Scandinavica (Scandinavian Journal of Ornithology)* **5**, 163–172. `https://doi.org/10.2307/3676059`. JSTOR: `3676059`. URL: `https://www.jstor.org/stable/3676059` (visited on 03/24/2025).

Nilsen EB, Vang R, Breisjøberget JI (2024). *Tetraonid Line Transect Surveys from Norway: Data from Statskog* (). `https://doi.org/10.15468/q2ehlk`. URL: `https://www.gbif.org/dataset/6a948a1c-7e23-4d99-b1c1-ec578d0d3159` (visited on 07/11/2024).

Pedersen HC, Steen H, Kastdalen L, Brøseth H, Ims RA, Svendsen W, Yoccoz NG (2004). *Weak Compensation of Harvest despite Strong Density–Dependent Growth in Willow Ptarmigan*. *Proceedings of the Royal Society of London. Series B: Biological Sciences* **271**, 381–385. `https://doi.org/10.1098/rspb.2003.2599`. URL: `https://royalsocietypublishing.org/doi/10.1098/rspb.2003.2599` (visited on 03/24/2025).

Plummer M et al. (2003). *JAGS: A Program for Analysis of Bayesian Graphical Models Using Gibbs Sampling*. In: *Proceedings of the 3rd International Workshop on Distributed Statistical Computing*. Vol. 124. 125.10. Vienna, Austria., pp. 1–10.

Pulliam HR, Danielson BJ (1991). *Sources, Sinks, and Habitat Selection: A Landscape Perspective on Population Dynamics*. *The American Naturalist* **137**, S50–S66.

R Core Team (2024). *R: A Language and Environment for Statistical Computing*. manual. Vienna, Austria: R Foundation for Statistical Computing. URL: `https://www.R-project.org/`.

Ranta E, Lindström J, Lindén H (1995). *Synchrony in Tetraonid Population Dynamics*. *Journal of Animal Ecology* **64**, 767–776. `https://doi.org/10.2307/5855`. JSTOR: `5855`. URL: `https://www.jstor.org/stable/5855` (visited on 03/21/2025).

Royama T (1992). *Analytical Population Dynamics*. Vol. 10. Springer Science & Business Media.

Sæther BE, Lillegård M, Grøtan V, Drever MC, Engen S, Nudds TD, Podruzny KM (2008). *Geographical Gradients in the Population Dynamics of North American Prairie Ducks*. *Journal of Animal Ecology* **77**, 869–882. `https://doi.org/10.1111/j.1365-2656.2008.01424.x`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2656.2008.01424.x` (visited on 03/21/2025).

Schaub M, Kéry M (2021). *Integrated Population Models: Theory and Ecological Applications with R and JAGS*. Academic Press.

Schwartz MD, Ahas R, Aasa A (2006). *Onset of Spring Starting Earlier across the Northern Hemisphere*. *Global Change Biology* **12**, 343–351. `https://doi.org/10.1111/j.1365-2486.2005.01097.x`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2486.2005.01097.x` (visited on 06/19/2024).

Sillett TS, Chandler RB, Royle JA, Kéry M, Morrison SA (2012). *Hierarchical Distance-Sampling Models to Estimate Population Size and Habitat-Specific Abundance of an Island Endemic*. *Ecological Applications* **22**, 1997–2006. JSTOR: `41723110`. URL: `https://www.jstor.org/stable/41723110` (visited on 11/03/2023).

Smith A, Willebrand T (1999). *Mortality Causes and Survival Rates of Hunted and Unhunted Willow Grouse*. *The Journal of wildlife management*, 722–730.

Smith PA, McKinnon L, Meltofte H, Lanctot RB, Fox AD, Leafloor JO, Soloviev M, Franke A, Falk K, Golovatin M, Sokolov V, Sokolov A, Smith AC (2020). *Status and Trends of Tundra Birds across the Circumpolar Arctic*. Ambio **49**, 732–748. `https://doi.org/10.1007/s13280-019-01308-5`. URL: `https://doi.org/10.1007/s13280-019-01308-5` (visited on 06/21/2024).

Soininen EM, Magnusson M, Jepsen JU, Eide NE, Yoccoz NG, Angerbjörn A, Inge Breisjøberget J, Ecke F, Ehrich D, Framstad E, Henttonen H, Hörnfeldt B, Killengreen S, Olofsson J, Oksanen L, Oksanen T, Tveito OE, Ims RA (2025). *Macroecological Patterns of Rodent Population Dynamics Shaped by Bioclimatic Gradients*. Ecography **2025**, e07058. `https://doi.org/10.1111/ecog.07058`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/ecog.07058` (visited on 05/25/2025).

Sotherton N, Tapper S, Smith A (2009). *Hen Harriers and Red Grouse: Economic Aspects of Red Grouse Shooting and the Implications for Moorland Conservation*. Journal of Applied Ecology **46**, 955–960. `https://doi.org/10.1111/j.1365-2664.2009.01688.x`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2664.2009.01688.x` (visited on 03/23/2025).

Steen H, Erikstad KE (1996). *Sensitivity of Willow Grouse Lagopus Lagopus Population Dynamics to Variations in Demographic Parameters*. Wildlife biology **2**, 27–35.

Steen JB, Haugvold OA (2009). *Cause of Death in Willow Ptarmigan Lagopus l. Lagopus Chicks and the Effect of Intensive, Local Predator Control on Chick Production*. Wildlife Biology **15**, 53–59. `https://doi.org/10.2981/07-073`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.2981/07-073` (visited on 03/27/2025).

Steen JB, Steen H, Stenseth NC, Myrberget S, Marcström V (1988). *Microtine Density and Weather as Predictors of Chick Production in Willow Ptarmigan, Lagopus l. Lagopus*. Oikos **51**, 367–373. JSTOR: `3565320`. URL: `http://www.jstor.org/stable/3565320`.

Storch I (2007). *Conservation Status of Grouse Worldwide: An Update*. Wildlife Biology **13**, 5–12. `https://doi.org/10.2981/0909-6396(2007)13[5:CSOGWA]2.0.CO;2`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.2981/0909-6396%282007%2913%5B5%3ACSOGWA%5D2.0.CO%3B2` (visited on 06/21/2024).

Thomas L, Buckland ST, Rexstad EA, Laake JL, Strindberg S, Hedley SL, Bishop JR, Marques TA, Burnham KP (2010). *Distance Software: Design and Analysis of Distance Sampling Surveys for Estimating Population Size*. Journal of Applied Ecology **47**, 5–14. `https://doi.org/10.1111/j.1365-2664.2009.01737.x`. URL: `https://onlinelibrary.wiley.com/doi/abs/10.1111/j.1365-2664.2009.01737.x` (visited on 02/20/2023).

Thomas VG (1987). *Similar Winter Energy Strategies of Grouse, Hares and Rabbits in Northern Biomes*. Oikos **50**, 206–212. `https://doi.org/10.2307/3566002`. JSTOR: `3566002`. URL: `https://www.jstor.org/stable/3566002` (visited on 03/27/2025).

Tuomi MW, Utsi TA, Yoccoz NG, Armstrong CW, Gonzalez V, Hagen SB, Jónsdóttir IS, Pugnaire FI, Shea K, Wardle DA, Zielosko S, Bråthen KA (2024). *The Increase of an Allelopathic and Unpalatable Plant Undermines Reindeer Pasture Quality and Current Management in the Norwegian Tundra*. Communications Earth & Environment **5**, 414. `https://doi.org/10.1038/s43247-024-01451-2`. URL: `https://www.nature.com/articles/s43247-024-01451-2` (visited on 06/30/2025).

Turchin P, Hanski I (1997). *An Empirically Based Model for Latitudinal Gradient in Vole Population Dynamics*. The American Naturalist **149**, 842–874. `https://doi.org/10.1086/286027`.

URL: https://www.journals.uchicago.edu/doi/abs/10.1086/286027 (visited on
08/24/2024).

Turchin P, Taylor AD (1992). *Complex Dynamics in Ecological Time Series*. *Ecology* **73**, 289–305.

Wiebe KL, Martin K (1995). *Ecological and Physiological Effects on Egg Laying Intervals in Ptarmigan*.
The Condor **97**, 708–717. https://doi.org/10.2307/1369179. URL: https://academic.
oup.com/condor/article/97/3/708-717/5124439 (visited on 08/24/2024).

Youngflesh C (2018). *MCMCvis: Tools to Visualize, Manipulate, and Summarize MCMC Output*. *Jour-*
nal of Open Source Software **3**, 640. https://doi.org/10.21105/joss.00640.