Ancient and ongoing hybridization in the *Oreochromis* cichlid fishes

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Abstract

Hybridization may enable adaptive diversification by generating unique genetic combinations when hybrid lineages are faced with ecological opportunity. Conversely, hybridization with exotic species may be detrimental to native biodiversity, by leading to homogenisation and the loss of important genetic material associated with local adaptation. Here we studied both ancient and contemporary hybridization in cichlid fishes of the genus *Oreochromis* (tilapia), which are among the most important fish for global aquaculture. We use whole genome resequencing of 575 individuals from 23 species, focussing on Tanzania, a natural hotspot of tilapia diversity, and a country where hybridization between exotic and native species in the natural environment has been previously reported. We reconstruct the first genome-scale phylogeny of the genus and reveal prevalent ancient gene flow across the *Oreochromis* phylogeny. This introgression has not led to large-scale adaptive radiation as seen in other cichlid lineages. We identify multiple cases of contemporary hybridization between native and introduced species in the wild, linked to the use of non-native species in aquaculture improvement and stocking for capture fisheries. Our study shows how ancient hybridization contributed to modern tilapia diversity, but is now a threat to both the genetic integrity of wild populations and the long-term prospects of the global tilapia aquaculture industry.

Introduction

Global aquaculture production has increased dramatically in recent decades, and is projected to take a key role in meeting the food demands of a growing human population, whilst alleviating nutritional poverty (Ahmed et al., 2019; Hall et al., 2017; Naylor et al., 2021). Tilapia of the genus *Oreochromis* (Cichlidae: Oreochromini) are a group of cichlid fish native to Africa and the Middle East, and are now the second largest group (by tonnage produced) of any aquaculture fish globally, following carp (Cyprinidae). *Oreochromis* production is dominated by a single species, the Nile tilapia, *Oreochromis niloticus*, which accounted for 4.5 out of the total 5.5 million tonnes of tilapia produced in 2020, the third most of any fish species globally. The remaining ~1 million tonnes are only accurately recorded to the genus level (FAO, 2022). The success of Nile tilapia has been attributed to high growth rates as well as tolerance of a wide range of environmental conditions, such as temperature, salinity, pH and low dissolved oxygen enable (El-Sayed & Fitzsimmons, 2023). In principle, the use of wild relatives of the Nile tilapia in breeding programmes could be used to further enhance production (Lind et al., 2012). However, despite this importance for food security, many of the 37 described *Oreochromis* species (Ford et al., 2019) are poorly known.

Although *Oreochromis* tilapia are widespread across Africa, subject to major capture fisheries, and key aquaculture species, considerably more attention has been given to the evolution of haplochromine cichlid systems (Joyce et al., 2011; Salzburger, 2018; Seehausen, 2006; Sturmbauer, 1998). This focus has been due to the propensity of haplochromines to form expansive lacustrine adaptive radiations, characterised by rapid speciation and extensive ecomorphological diversification (Malinsky et al., 2018; McGee et al., 2020). This occurred despite low levels of genetic diversity, a high proportion of which is shared between species due to incomplete lineage sorting and widespread interspecific introgression (Svardal et al., 2021). It has been proposed that extensive lacustrine radiations of haplochromine cichlids have been facilitated by the introgression arising from hybridization of ancestral lineages (Meier et al., 2017; Svardal et al., 2020). This may fuel

differentiation and diversification by supplying novel combinations of genetic variants

(Singhal et al., 2021), and sharing of ancient alleles that have already been filtered by

selection (Marques et al., 2019). By contrast, ongoing or recurring hybridization may

decrease differentiation and homogenise gene pools, therefore reducing the propensity of a

lineage to radiate.

In contrast to the haplochromines, extensive radiation has not taken place in the

Oreochromis (Seehausen, 2006). Most *Oreochromis* species occupy rivers and are allopatric

to apparent sister taxa (Ford et al., 2019; Trewavas, 1983)*;* the only known *Oreochromis*

lacustrine radiations are small and limited to Lake Malawi and Lake Natron (3 species each).

To date, comprehensive phylogenomic examinations of ancestral introgression occurring

during the evolution of *Oreochromis* are lacking, and therefore it is currently not clear what

role ancestral introgression has played in *Oreochromis* diversification*.*

Given the high productivity of some *Oreochromis* species in aquaculture and capture

fisheries, they have been widely introduced across tropical and subtropical freshwaters

globally (Canonico et al., 2005; Lévêque, 1995). In Africa, introductions to the natural

environment have resulted in widely documented contemporary hybridization (Blackwell et

al., 2020; Bradbeer et al., 2019; Champneys et al., 2021; Ciezarek et al., 2022; Shechonge

et al., 2018). The risk of hybridisation between native and exotic *Oreochromis* species is

likely to increase with the projected growth of aquaculture across Africa (Prabu et al., 2019).

Detailed analysis of the extent of hybridization between co-occurring tilapia species will

therefore be valuable for identifying species and populations of conservation concern.

This study focuses on the genomic implications on ancestral and contemporary

introgression in *Oreochromis*, focusing on wild populations in Tanzania. The country is a

hotspot for diversity of the genus with at least 21 species present, including a minimum of

three species that have been widely introduced outside their native range, namely *O.*

niloticus, *O. leucostictus* and *O. esculentus* (Shechonge et al., 2019). To reconstruct the

phylogenetic relationships between species, and to quantify ancestral and modern

hybridization, we present a genome-wide resequencing dataset of the genus *Oreochromis*.

We provide a first phylogenomic analysis of the group and show that *Oreochromis* is typified

by a high degree of ancestral introgression. We also identify a species of hybrid origin, the

Lake Kiungululu tilapia, *O. chungruruensis*. We further find that anthropogenic activities have

resulted in widespread recent hybridization, highlighting a potential risk to the conservation

of species and their genetic diversity.

Materials and methods

Sampling and sequencing

We sampled and generated sequencing data for 433 individuals, and combined this with

previously published data from 142 individuals (Table S1) from across southern and eastern

Africa (Figure 1). Samples were from 65 locations, and 21 drainage basins across Tanzania,

as well as South Africa, Kenya, Angola, Malawi, Mozambique and Uganda. Individuals were

morphologically identified to species using information outlined in (Genner et al., 2018).

Individuals were classified as hybrids if they had characteristics intermediate between

different species. Our final dataset comprised of 25 species, including the outgroups

Maylandia zebra and *Sarotherodon galilaeus* and 23 out of the 37 species of *Oreochromis*

currently recognised, including *Oreochromis (Alcolapia) grahami* (Fricke et al., 2023). Fish

specimens were mostly obtained from local fishers or through experimental fishing surveys.

Live fish were euthanised with an overdose of anaesthetic (MS-222). Samples for

subsequent DNA extraction were collected by clipping off the right pectoral fin, which was

placed in a labelled vial of ethanol. Whole voucher specimens were retained, with each

individual pinned on a styrofoam board, photographed and preserved in formalin or ethanol.

Figure 1. Locations of the *Oreochromis* sampled across Africa (bottom left) and, within

Tanzania (right). Sample sites where contemporary hybrids were identified from genomic

data are highlighted in red. Key drainage basins are labelled. The bar chart (top left) shows

the total number of specimens (contemporary hybrids in red, non-hybrids in white) located

per drainage basin within Tanzania, for all basins with more than ten samples, or country

outside of Tanzania. All basins within Tanzania with fewer than ten specimens sampled are

grouped into "Tanzania Other". Abbreviation: SA - South Africa. Geographic coordinates of

sampling locations are shown in Table S1.

DNA was extracted from fin clips using a PureLink Genomic DNA extraction kit (Life

Technologies). Sequencing was conducted using either PCR-free library preparation with

Illumina HiSeq 2500 (250bp paired-end), automated KAPA DNA library preparation with

Illumina HiSeq 2500 (125bp paired-end), LITE library preparation with Illumina HiSeq 4000

(150bp paired-end) or LITE library preparation with NovaSeq S4 (150bp paired-end).

SNP calling

SNPs were called against previously published near chromosome-level assemblies of *M.*

zebra (GCF_000238955.4) and *O. niloticus* (GCF_001858045.2) (Conte et al., 2019). Raw

reads were checked for adapter sequence contamination using BBMerge, within BBMap

(v38.06) (Bushnell, 2014). Where necessary, reads were trimmed for adapters using fastp

(v0.20.0) (Chen et al., 2018), with quality-trimming disabled. Reads were then mapped

separately against both assemblies using the mem function in bwa (v0.30.0) (Li & Durbin,

2009), with raw mappings sorted by coordinate and having mate coordinates and insert size

fields added using samtools (v1.9) (Li et al., 2009). Joint genotyping was then carried out on

all 575 samples using bcftools (v1.10.2) (Li et al., 2009). First, bcftools mpileup was used

with a minimum base and mapping qualities of 30. Multi-allelic variant calling was then

carried out using bcftools call. Variants were filtered if they were within three base pairs of

any other variant, if variant quality score was less than 30, if depth at a given site (across all

samples) was less than 500 or greater than 9000, or if minor allele count was less than

three.

Backbone phylogeny reconstruction

To construct a species-level phylogeny, we identified a panel of 91 reference individuals that

were unlikely to be recent hybrids (Table S1). These individuals were identified using

morphology, evidence of monophyly in a full genome neighbour-joining tree of individuals not

classified morphologically as hybrid, and evidence of monophyly in a mitochondrial genome

neighbour-joining tree of those individuals (Supplementary Data 1). Using this reference

panel, separate phylogenetic trees were inferred for each of the *M. zebra* and *O. niloticus*

mapped datasets. Trees were constructed using both a maximum-likelihood SNP-based

concatenation tree, inferred using IQ-TREE (v1.6.12) (Nguyen et al., 2015), as well as a

multi-species coalescent summary-based tree, inferred using ASTRAL (v5.4.12) (Rabiee et

al., 2019; Zhang et al., 2018), based on trees inferred from recombination-free 10kb

windows across the genome with IQ-TREE (Supplementary Methods). Robinson-Foulds

(RF) distances (Robinson & Foulds, 1981) were calculated between each 10kb-window tree

and the inferred ASTRAL species tree using the ETE 3 python package (Huerta-Cepas et

al., 2016), in order to further quantify how widespread phylogenetic discordance was. For

species where the inferred phylogenetic trees raised taxonomic questions, such as a lack of

monophyly of all individuals assigned to a species, exploratory Principal Component

Analysis (PCA) was carried out on all individuals of the relevant species, but excluding those

inferred to be a contemporary hybrid (see below), using plink (v2.0.0) (Purcell et al., 2007).

The PCAs on each subset of individuals used all biallelic SNPs found with a minor allele

count of at least 3 in this subset of individuals, and SNPs were pruned for linkage

disequilibrium ($r^2 > 0.6$) over 20kb windows using bcftools.

Assessment of ancestral introgression

Ancestral introgression between species was assessed using the panel of 91 reference

individuals, and both the *M. zebra* and *O. niloticus* mapping datasets. We used both SNP-

based D statistics (Green et al., 2010), as well as *f*-branch statistics (Malinsky et al., 2018),

calculated using Dsuite (v0.4) (Malinsky et al., 2021), and genome-wide 10kb window

phylogenetic-tree-based analyses. These tree-based analyses included the branch-length

tests (BLT) and discordant count tests (DCT), using the scripts provided by Suvorov et al.,

(2022), and chi-squared tests of discordant topology frequencies inferred using IQ-TREE

(Minh et al., 2020; Suvorov et al., 2022) (Supplementary Methods). To test whether

introgression was more likely between species sharing a drainage basin or between closely

or more distantly related species, for each species we identified the species counterpart with

the highest *f*-branch (i.e. the highest degree of gene flow). We then inferred whether these

species pairs were more likely to have shared modern drainage basins than expected by

chance using a randomised permutation test. Similarly, we identified for each species

whether their high *f*-branch was more or less phylogenetically related than the average of all

the species compared ($z <= -2$ for less phylogenetically distant, $z >= 2$ for more

phylogenetically distant). We then tested whether we observed more species with

significantly phylogenetically similar or dissimilar counterparts than expected by chance

using a permutation test.

Analysis of putative introgression events

Putative introgression events, identified by the *f*-branch analysis, were further investigated to

identify introgressed genomic regions. This was carried out using a novel statistic (Dwt;

Dweighted_topo), derived from topology weightings calculated from 'topology weighting by

iterative sampling of subtrees' (Twisst) (Martin & Van Belleghem, 2017), computed along the

22 linkage groups of the *M. zebra* reference assembly (Supplementary Methods). Dwt is

designed to highlight putatively introgressed regions. Dwt = 0 indicates regions where the

species tree (sptree) has the highest weighting. Dwt > 0 indicates where disc1 (the putative

introgression topology) is most heavily weighted, with 1 indicating it is fully supported (no

weighting for either of the other topology). Similarly, Dwt < 0 indicates where disc2 (the other

alternate topology) is most heavily weighted. $Dwt = \frac{disc1 - disc2}{disc1 + disc2} *$

$\frac{max(sptree,disc1,disc2) - sptree}{max(sptree,disc1,disc2) + sptree}$.

206   Putative introgression events where the Twisst analysis showed no clear excess of

either the species-tree topology or hybridization topology were further investigated to identify

signals of hybrid speciation. Following Olave et al. (2022), we used ADMIXTURE analyses

of the relevant species at both K=2 and K=3, as well as assessment of species-diagnostic

SNPs. Putative introgression dates were also estimated based on estimates of Dxy and π in

phylogenetically identified introgressed regions of the genome, calculated using

genomics_general (https://github.com/simonhmartin/genomics_general) (see Supplementary

Methods).

Contemporary hybridization

The 486 individuals that were not among the reference panel were tested for evidence of

recent hybridization. We carried out separate analyses in each sampled water drainage

basin within Tanzania. For each drainage basin, biallelic SNPs were extracted for each

individual sampled, alongside reference individuals for all the taxa recorded in the water

body by Shechonge et al. (2019). SNPs with a minor allele count of at least 3 were retained,

and SNPs were pruned for linkage disequilibrium (r^2>0.6) over 20kb windows using bcftools.

For each drainage basin, a supervised ADMIXTURE (Alexander et al., 2009) analysis was

carried out only involving the relevant species or populations, with a K value equal to the

number of locally recorded taxa for the ADMIXTURE analysis, and each reference species

assigned to their known group. Test individuals were assigned a hybrid status if they had

multiple ancestry components where the lower end of the standard error was at least 0.1.

This cut-off was used to identify up to second generation backcrosses. We further confirmed

that all of the reference individuals of the same species were tightly clustered using PCAs

performed in plink, and comprised a monophyletic group with no long branch lengths, in

phylogenetic trees inferred with IQ-TREE (v2.0) with automated model detection, 5

independent runs and 1,000 rapid bootstraps. These analyses were carried out with the

same SNP set as the ADMIXTURE runs, except it was further filtered for sites where at least

one individual had each of the homozygous reference and alternate alleles for the IQ-TREE

analysis. We further characterised contemporary hybridization using panels of species-

diagnostic SNPs (Supplementary Methods).

Results

Mapping, phylogenomic and ancestral introgression analyses were carried out against both

the *O. niloticus* and *M. zebra* reference assemblies separately. Results were similar and

conclusions identical (Supplementary Materials), therefore only the *M. zebra* reference

analyses are reported below.

Sampling, sequencing, read mapping and SNP calling of 23 species.

An average of 41.3 million paired-end reads were sequenced for the 433 individuals (range

10.4 - 106 million) (Figure 1). Mapped reads from all 575 individuals (including the previously

published data for 142 individuals) had average depth of 6.3 (range 1.7-18.6), and average

properly paired mapping of 75% (range 45-89%). In total 55,949,298 filtered SNPs were

called. Full mitochondrial genomes of at least 10,000bp were successfully assembled for

437 out of the 450 individuals not morphologically identified as hybrid, with an average

length of 16,541bp (range 11,048-17,126bp).

Phylogenetic inference

A total of 1,692,136 non-coding SNPs were used for the maximum-likelihood phylogenetic

inference with 15,310 recombination-free 10kb windows used for ASTRAL. Most of the

taxonomic species produced monophyletic groups, and therefore collapsed to a single node.

Those that were not monophyletic were instead collapsed to the population level (Figure 2a).

There were some inconsistencies between the ASTRAL and maximum-likelihood trees,

particularly with regard to the phylogenetic placement of *O. tanganicae, O. placidus placidus*

and *O. korogwe* (Figure S2-5). The ASTRAL results also suggest *O. hunteri* to be a

monophyletic species, whereas the ML trees do not (Figure S2-5). All following analyses are

reported using the ASTRAL species tree, given that unlike the maximum-likelihood tree it

accounts for the likely widespread incomplete lineage sorting in the dataset. This is indicated

by the high levels of phylogenetic discordance shown by high Robinson-Foulds (RF)

distances between each 10kb window tree and the species tree. No 10kb window tree had a

RF distance from the species tree of less than 0.08 (Figure S1).

Figure 2. a) Phylogenetic tree inferred using ASTRAL from the *M. zebra* mapping dataset,

with conspecific individuals collapsed. Branch lengths are in coalescent units. All nodes had

posterior probability of 1.0. b) Phylogenetic tree inferred using ASTRAL from the *M. zebra*

mapping dataset, without species collapsed, highlighting only the individuals from the Lake

Malawi "Chambo" assemblage of *O. squamipinnis* and *O. karongae*. c) PCA analyses of the

major taxonomic questions raised by this phylogenetic tree, showing the "Chambo" group

(left), *O. rukwaensis* being distinct from *O.* "mtera" (middle), and subspecies of *O. placidus*

and *O. shiranus* not supporting current classification (right).

There were instances of taxonomic species where individuals from different

populations did not form monophyletic groups (Figure 2a, Figure S2), and PCA analyses

further confirmed a lack of clustering of taxonomically identified species (Figure 2c). *O.*

rukwaensis individuals from the Rukwa drainage basin were resolved as a clade but were

phylogenetically distinct from individuals collected from the Mtera Reservoir, which had

provisionally been identified as *O. rukwaensis* based on morphology (Shechonge et al.,

2019). Here, we refer to this population as *O. "mtera"*. We found no evidence of a taxonomic

split between *O. shiranus* and *O. placidus*, and instead *O. shiranus shiranus* was resolved

as sister to *O. placidus placidus*, whereas *O. shiranus chilwae* is sister to *O. placidus*

rovumae. Similarly, the analysed "Chambo" species, *O. squamipinnis* and *O. karongae*, are

not reciprocally monophyletic in our anlayses (Figure 2b,c).

Widespread ancestral introgression

There was consistently a high degree of ancestral introgression across the *Oreochromis*

phylogeny evident in our analyses. Using the genome-wide phylogenetic trees constructed

from 10kb windows, 44,030 out of the 113,564 trios tested with the blt and dct tests showed

significant deviations from expected values under incomplete lineage sorting alone,

consistent with introgression. Similarly, the frequencies of 10kb window-tree discordant

topologies indicated introgression at 42/89 nodes in the ASTRAL tree. For the SNP-based D

statistics, 2,658 out of the 2,925 tested trios showed significantly higher levels of allele

sharing between non-sister taxa than expected, with Dp values indicating a range of

introgression proportions ranging from 0-0.45 (Table S2; for this analysis, individuals were

pooled into populations or species, hence the lower number of tested trios than the blt and

dct analyses).

F-branch statistics were calculated between each branch in the tree and each tip,

revealing a few branches which appeared to exhibit relatively large amounts of gene flow

(Figure 3a). Notably high *f*-branch values were recorded associated with two of the

taxonomic uncertainties highlighted above; namely the *O. shiranus- O. placidus* group

(especially involving *O. shiranus chilwae*) and between *O. rukwaensis* and *O. "mtera"*

(Figure 3a).

We identified the counterpart for each species which had the highest *f*-branch value.

These species pairs occupied the same drainage basin more often than expected by chance

(*p*=0.0002). This was the case for seven species (compared to simulated range of 0-8

species): *O. leucostictus* / *O. niloticus*, *O. girigan* / *O. korogwe* Mlingano, *O. karomo* / *O.*

tanganicae, "Chambo" / *O. chungrurensis*, *O. jipe* / *O. girigan*, *O. chungruruensis* /

"Chambo", and *O. shiranus shiranus* / *O. chungruruensis*. Three out of these seven species

pairs were related to a single putative hybrid speciation event of *O. chungruruensis* (see

results below). After removing these species from the analysis, shared drainage basin less

strongly predicted the *f*-branch values (*p*=0.03; 4 species, simulated range 0-7). Similarly,

the species pairs with evidence of introgression were more likely to be more closely related

than the mean phylogenetic distance between each species (Z-score <= -2) than expected

by chance (*p*=0.0004). This was the case for four species pairs (compared to simulated

range 0-4 species): *O. jipe* with *O. girigan*, *O. amphimelas* with *O. esculentus*, *O. rukwaensis*

with *O.* "mtera" and *O. malagarasi* with *O. variabilis*.

Figure 3. a) Heatmap of the *f*-branch values calculated between each node (y axis) and tip

(x axis) of the *M. zebra* mapping ASTRAL tree. Boxes indicating the introgression from

"Chambo" into *O. chungruruensis* are highlighted in blue, and between *O. leucostictus* and

O. niloticus are highlighted in red. Boxes indicating introgression amongst the clades

associated with taxonomic uncertainties are highlighted in black. b) Normalised Dwt statistics

across the linkage groups showing regions with a stronger weighting for the introgression

tree (left of the solid line), where *O. leucostictus* and *O. niloticus* are sister (left; red), and

where "Chambo" and *O. chungruruensis* are sister (right; blue). Dots represent stretches of

the genome >=50kb where the four Twisst analyses consistently have Dwt = 1.0 showing

introgression between *O. leucostictus* and *O. niloticus* (red), *O. squamipinnis* and *O.*

chungruruensis (blue), and overlap between both comparisons (black). Abbreviations: On -

O. niloticus, Ol - *O. leucostictus*, On_s - *O. spilurus* + *O. aureus*; Ol_s - *O. variabilis* + *O.*

esculentus + *A. grahami,* Ch - "Chambo", Oc - *O. chungrurensis*, Ch_s - *O. rukwaensis* + *O.*

variabilis + *O. macrochir*, Oc_s - *O. placidus rovumae* + *O. korogwe* Mlingano + *O.*

mossambicus).

Based on the results of *f*-branch analyses and Dp values, we focused on ancestral

introgression between *O. niloticus* and *O. leucostictus* and between *O. chungruruensis* and

the "Chambo" group. These were the species pairs indicating the highest proportions of

introgression according to Dp (Table S2), as well as highest *f*-branch values between non-

sister or closely related branches (Figure S6). Four analyses were carried out to assess

gene flow in each case (Supplementary Materials). All analysis from the Dweighted_topo

statistics show generally similar putatively introgressed regions between the different

analyses, although some species combinations suggested more than others (Figure S7). In

both cases, very little of the genome had a higher weighting for the other alternate topology

(non-species or introgression tree), as would have been indicated by Dwt < 0 (Figure 3b). In

total, 52Mb of the genome was consistently found to be introgressed between *O.*

leucostictus and *O. niloticus,* and 4Mb introgressed between *O. chungruensis* and *"*Chambo"

(Figure 3b), from 761Mb in the *M. zebra* reference genome linkage groups. There was

significantly more of the genome introgressed in both comparisons (816Kb, with regions

across all 22 linkage groups; Figure 3b) than would be expected randomly (permutation test

p=0.0002; Figure S8).

The nature of *O. chungruruensis* introgression with "Chambo" and *O. shiranus*

shiranus was investigated by ADMIXTURE analysis assuming two populations (K=2; cross-

validation score = 0.48), with individuals of "Chambo", *O. chungruruensis* and *O. shiranus*

shiranus from the Lake Malawi basin (*O. placidus* and *O. shiranus chilwae* were not

recorded here so was not used). This suggested that 47-48% of the *O. chungruruensis*

genome had affinity with "Chambo", and 52-53% with *O. shiranus shianus*. When assuming

three populations (K=3; cross-validation score =0.58), *O. chungruruensis* formed its own

group (Figure 4). Diagnostic SNPs were also identified for "Chambo", the *O. shiranus - O.*

placidus group and *O. chungruruensis* (Supplementary Results). Note that diagnostic SNPs

could not be detected for *O. shiranus shiranus* alone, and so the monophyletic *O. shiranus-*

O. placidus group as a whole was used. These showed similar frequencies of SNPs fixed for

either the reference or the homozygous alternate alleles of both "Chambo" and *O. shiranus -*

O. placidus, with around half of diagnostic SNPs heterozygous (Figure 5b). Across the 22

linkage groups, 4,300 windows (each of length 200 SNPs) were found where all haplotypes

O. chungruruensis and *O. shiranus- O. placidus* were sister taxa, compared to 2,247

windows where *O. chungruruensis* and "Chambo" were sister. Based on values of π and

Dxy within these windows, we inferred introgression times of 21,520 (CI 16,374-47,075)

years ago between *O. shiranus/ O. placidus* and *O. chungruruensis* and 13,565 (CI 10,321-

29,673) years ago between "Chambo" and *O. chungruruensis*.

Figure 4. a) ADMIXTURE analysis of all *O. shiranus- O. placidus*, "Chambo" and *O. chungruruensis* at *K*=2 and *K*=3. b) The proportion of species diagnostic SNPs which are either homozygous for the alternate allele (Hom.), heterozygous (Het.), or homozygous for the reference allele (ref.) in the three *O. chungruruensis* individuals (abbreviation *shi-plac - O. shiranus + O. placidus*). * indicates genetically identified *O. shiranus shiranus* x "Chambo" hybrids in Lake Itamba.

Contemporary hybridization ongoing across Tanzania

Eleven different drainage basins and three aquaculture establishments across Tanzania were tested for the presence of hybrids (Figure 1). A total of 76 hybrids (Table 1) were identified. These were identified from across seven drainage basins: Ruvu River (30/46 tested individuals identified as hybrid), Pangani River (15/51), Rufiji River (11/56), Wami River (6/109), Lake Malawi (2/12), Lake Rukwa (2/29) and Ruvuma River (1/3) and Lake Chala (1/1). Hybrids were also identified at two aquaculture establishments: the Songea hatchery (7/50) and Mbarali Farm (1/24). It should be stressed that these numbers should not be taken as representative, as sampling was non-random and biased towards individuals thought to be hybrids on the basis of morphology.

390 **Table 1.** Recent hybrids identified from the supervised ADMIXTURE analysis. See Table S1

391 for exact locations and ancestry components of each hybrid. * indicates the individual with

392 the "Bandia" phenotype with Lake Chala.

Species pair	Number of hybrids	Drainage basins recorded (number; locations)
O. niloticus x O. urolepis	27	Ruvu River (10; Mindu Reservoir), Rufiji (9; Kidatu), Pangani (7; Lake Jipe), Wami River (1; Kilosa)
O. leucostictus x O. urolepis	20	Ruvu River (17; Mindu), Wami River (3; Kilosa (2), Lake Nala (1))
O. niloticus x O. leucostictus x O. urolepis	5	Ruvu River (3; Mindu), Wami River (2; Kilosa)
O. niloticus x O. shiranus	5	Songea hatchery (5)
O. niloticus x O. "mtera"	3	Rufiji (2; Kidatu), Mbarali Farm (1)
O. shiranus x "Chambo"	2	Lake Malawi (2; Lake Itamba)
O. leucostictus x O. shiranus	2	Songea hatchery (2)
O. girigan x O. jipe	2	Pangani (2; Lake Jipe)
O. girigan x O. niloticus	2	Pangani (2; Lake Jipe)
O. girigan x O. jipe x O. niloticus	2	Pangani (2; Lake Jipe)
O. girigan x O. esculentus	2	Pangani (2; Lake Jipe)
O. niloticus x O. placidus rovumae	1	Ruvuma (1; Rovuma)
O. leucostictus x O. rukwaensis	1	Lake Rukwa (1; Lake Rukwa)
O. esculentus x O. leucostictus	1	Lake Rukwa (1; Lake Rukwa)
*O. rukwaensis x O. urolepis	1	Lake Chala (1; Lake Chala)

Most of the hybrids in our samples (68%) were between *O. urolepis, O. leucostictus* and *O.*

niloticus, with none directly between *O. leucostictus* and *O. niloticus*. Species diagnostic

SNPs, fixed in the confidently non-hybrid individuals of each of these three species, and with

a low frequency (<0.1) in other non-hybrid individuals, were identified for each, with numbers

ranging from 571 in *O. niloticus*, to 3,147 in *O. leucostictus* and 4,177 in *O. urolepis*. The

hybrids identified by ADMIXTURE between the three species were investigated for these

diagnostic SNPs, with results generally concordant with the ADMIXTURE ancestry

components (Figure 5). The individuals with ADMIXTURE ancestry components close to

50% had high levels of heterozygosity for both species, potentially indicating F1 hybrids.

When ancestry components rose above 50% for a species, it was generally associated with

more homozygous species diagnostic SNPs, suggesting backcrossing.

Figure 5. Species diagnostic SNP frequency for each individual inferred to be a hybrid

between the potential parent species of *O. leucostictus*, *O. niloticus* and *O. urolepis* from

ADMIXTURE analyses (ancestry components on far-right) (one per row). Diagnostic SNPs

shown for *O. urolepis* (left), *O. niloticus* (mid-left), *O. leucostictus* (mid-right).

Only 51/124 individuals morphologically suggested to be possible hybrids were

genetically confirmed, ranging across the Rufiji, Ruvu River, Wami River, Zanzibar, as well

as the Songea hatchery. 24 out of the remaining 321 test individuals were genetically, but

not morphologically identified as hybrids. These included individuals morphologically

assigned to *O. urolepis* from Mbarali Farm, the Ruvu River and the Wami River, *O. shiranus*

shiranus from Lake Itamba and Songea hatchey, *O. niloticus* from Lake Jipe*, O. girigan* from

Lake Jipe, *O. esculentus* from Lake Rukwa, *O. rukwaensis* from Lake Rukwa, *O. pangani*

from Lake Kalimau and *O. placidus rovumae* from the Muhuwesi River (a tributary of the

Ruvuma River).

The 'Bandia' individual, from a potentially hybrid population found in Lake Chala of

unknown origin, was identified as a probable *O. rukwaensis* x *O. urolepis* hybrid, according

to a panel of species diagnostic SNPs, as well as ADMIXTURE and Twisst analysis (see

Supplementary Results).

Discussion

Here, we present evidence that the genus *Oreochromis*, a group of cichlid fish important for

global aquaculture, has undergone introgressive hybridization multiple times in the course of

its evolution, and is now experiencing large-scale contemporary hybridization resulting from

anthropogenic translocations. Almost all evidence of contemporary hybrids involved

populations introduced to boost aquaculture or fisheries production, threatening the

conservation of wild populations in Africa.

Phylogenomic resolution of Oreochromis and taxonomic questions

We inferred a first phylogenomic backbone tree for the clade, building on previous

work that was based on a handful of nuclear and mitochondrial markers (Ford et al., 2019).

In accordance with Ford et al., (2019), we found that the *Alcolapia* group, comprised of

species adapted to extreme soda lake environments (Trewavas, 1983), is nested within the

Oreochromis, forming a clade with *O. esculentus* and *O. amphimelas*. This supports the

conclusions of Ford et al. (2019) that *Alcolapia* is best considered as a subgenus within

Oreochromis. We consistently found *O. amphimelas,* to be sister to *A. grahami.* This would

suggest a single origin of tolerance of soda-lake conditions, which is present in *O.*

amphimelas, albeit less extreme than *Alcolapia* (Trewavas, 1983). However, unlike Ford et

al., (2019), our analyses suggests that the species previously placed in the subgenus

Nyasalapia ("Chambo", *O. rukwaensis*, *O. chungruruensis*, *O. variabilis*, *O. malagarasi*, *O.*

macrochir and *O. karomo*) (Trewavas, 1983) do form a monophyletic clade, with the

exception of *O. chungruruensis*, which has a hybrid origin between the "Chambo" and the *O.*

shiranus- O. placidus group.

Our study raises several taxonomic questions related to species classifications within

the *Oreochromis*. Two species of the subgenus *Nyaslapia*, *O. squamipinnis* and *O.*

karongae, which are morphologically similar and endemic to Lake Malawi, comprise a single

clade, but were not resolved as reciprocally monophyletic. Within the main lake, they have

been differentiated largely on the basis of male breeding dress: male *O. karongae* are

uniformly black (apart from white fin margins and pinkish genital papilla), while *O.*

squampinnis males have a contrasting blaze of colour, generally blue, on the top of the

head. In Lake Malawi itself, these traits are associated with other differences, including

breeding seasonality, microhabitat and trophic morphology (Trewavas, 1983), but

populations in crater lakes have not been thoroughly studied and have just been

provisionally identified on the basis of male colour. Our results do not support a phylogenetic

division based on male colour, particularly when the crater lake populations are considered.

Ongoing admixture between species or parallel evolution may explain the evolution of these

traits. The low number of specimens from Lake Malawi, as opposed to elsewhere in the

catchment, prevents us from drawing clear conclusions about its populations. We were

unable to sample the third *Nyasalapia* species from Lake Malawi, *O. lidole,* which has not

been seen alive since 2007 and may be extinct (Konings, 2018). Genomic data on this

species, potentially from museum specimens, would help with a taxonomic reassessment of

the group.

Our results suggest that the current taxonomic status of *O. shiranus* and *O. placidus*

needs revision. Each of these species is presently split into two geographically separated

subspecies (Trewavas, 1983). Specifically, *O. shiranus shiranus* is found in Lake Malawi and

the Upper-Middle Shire River, while *O. shiranus chilwae* is found in Lake Chilwa and Chiuta.

Meanwhile, *O. placidus placidus* is found in the Lower Shire and Lower Zambezi, while *O.*

placidus ruvumae is found in the Ruvuma system. Our results suggest that the current

groupings are incorrect and should be replaced by a Lake Malawi/ Shire/ Zambezi taxon and

a Chilwa/ Chiuta/ Ruvuma taxon, which more closely reflects historic and present river

system connectivity. There are few, if any, morphological features supporting the current

classification.

We found that the *Oreochromis* population from the Mtera Reservoir (on the Ruaha

section of the Rufiji system) did not cluster with *O. rukwaensis* sampled from the

neighbouring Lake Rukwa catchment. Instead, it was sister to a clade consisting of *O.*

rukwaensis, O. variabilis and *O. malagarasi.* This first genomic assessment of the population

is consistent with the Mtera population representing a candidate new species, which we refer

to as *O.* sp "mtera". There was some apparent introgression between *O. rukwaensis* and *O.*

"mtera", perhaps explaining the morphological similarity that led to previous assumptions of

their conspecific status (Genner et al., 2018).

Widespread ancestral introgression

We found that there was widespread, although uneven, introgression across the

Oreochromis phylogeny. The degree of introgression was only weakly predicted by

phylogenetic distance or whether species occupy the same drainage basin in the modern

day. This is, however difficult to test given the highly correlated nature and lack of

independence of introgression statistics, which are calculated on different subsets of four

taxon across a phylogenetic tree. We may therefore have lacked power to identify these

influences. We highlight two notable instances of introgression between relatively

phylogenetically divergent species, primarily between the "Chambo" and *O. shiranus* (likely

resulting in *O. chungruruensis*), and between *O. niloticus* and *O. leucostictus.* These latter

two species coexist in their indigenous habitat of the Albertine Rift lakes Edward, George

and Albert, and have been co-translocated to the same water bodies (e.g. Lake Victoria), yet

are rarely found to hybridize in the modern day (Ciezarek et al., 2022; Geletu & Zhao, 2022).

Our results may be reflective of gene flow between the two species at early stages of the

speciation continuum (Stankowski & Ravinet, 2021).

We found that *O. chungruruensis* could have arisen as a result of hybrid speciation

between "Chambo" and either *O. shiranus shiranus* or the wider *O. shiranus/ O. placidus*

group. Interestingly, *O. chungruruensis* is currently classified as a member of the subgenus

Nyasalapia alongside *O. squamipinnis* and *O. karongae* (Trewavas, 1983), which was

supported by previous studies based on a handful of nuclear and mitochondrial markers

(Ford et al., 2019). *O. chungruruensis* is endemic to Lake Kiungululu, an isolated crater lake

within the northern sector of the Lake Malawi basin. Neither "Chambo", or *O. shiranus* have

been recorded in this lake, although "Chambo" and *O. shiranus shiranus* are found

throughout the Malawi basin and co-occur in several crater lakes in the northern sector

(Kingiri, Ilamba, Itamba and Ikapu). Indeed, our analysis finds two contemporary hybrids

between "Chambo" and *O. shiranus shiranus* in Lake Itamba. The age of Lake Kiungulu is

not known, although the nearby crater Lake Masoko has been estimated to be around

50,000 years old (Garcin et al., 2006), relatively soon before our estimated introgression

≤30,000 years ago. This may suggest a scenario where *O. chungruruensis* originated from

hybridization between ancestral populations of both within the lake.

Contemporary hybridization among Oreochromis species

Within Tanzania, there has been a long history of translocations of *Oreochromis* species for

capture fisheries, dating back to at least the 1950s, in addition to ongoing aquaculture

translocations, leading to the widespread colonisation by three species: *O. niloticus*, *O.*

leucostictus and *O. esculentus* (Shechonge et al., 2019). Interspecific hybridization has long

been recorded between *Oreochromis* species based on SNP panels, mitochondrial data and

microsatellites (Geletu & Zhao, 2022), although studies based on genome-wide data are

lacking. Our study has particularly demonstrated that hybridization between the invasive *O.*

niloticus, O. leucostictus and the native *O. urolepis*, previously recorded in Tanzania

(Ciezarek et al., 2022) is widespread. Although we found no evidence of hybrids between

the invasive *O. leucosticus* and *O. niloticus*, we did find five individuals which had ancestry

components of over 0.1 (10%) for both species as well as *O. urolepis*. This suggests that

there may be barriers preventing *O. leucostictus* and *O. niloticus* from directly hybridizing in

the modern day, despite the ancestral introgression we inferred between them. Hybridization

from either species with *O. urolepis* may however mediate gene flow between them.

Previous studies, based on mitochondrial data have recorded introgression directly between

the two species (Ndiwa et al., 2014), although Nyingi et al. (2009) found this introgression

signal was not present in the nuclear genome. Furthermore, it is not clear whether there was

thorough species sampling in these studies, raising the possibility of 'ghost' introgression

explaining these findings. Further work is necessary to determine whether there are genomic

or behavioural barriers reducing the likelihood of *O. leucostictus* and *O. niloticus* hybridizing

directly.

Interestingly, the Lake Chala Bandia individual appears to be a case of hybridization

between two species not native to the lake (*O. urolepis* and *O. rukwaensis*), plausibly in farm

ponds prior to its introduction to the lake. Previous studies on Bandia have suggested some

clustering with *O. urolepis*, but with a high degree of diversity in both mitochondrial DNA and

morphology (Dieleman et al., 2019; Moser et al., 2019). This is consistent with our finding of

hybrid origin. Our study is however, based on only a single Bandia individual, and so more

detailed population genomic analysis would be welcome.

Concluding remarks

Several caveats need to be considered when assessing our findings. Critically, our

study did not include every existing species or population of *Oreochromis* found across the

globe. These missing or 'ghost' taxa, in addition to extinct populations of *Oreochromis* may

have significantly misled analyses of ancestral introgression (Tricou et al., 2022a, 2022b).

Extensive sampling of tilapia at both the species and population levels across Africa and the

Middle East will be necessary to assess this, in addition to the development of bioinformatic

tools which can accurately infer phylogenetic networks with large datasets, including

genome-scale data and many individuals. Ghost taxa may have influenced the identification

of recent hybrids, particularly in water bodies where *Oreochromis* was less thoroughly

sampled. There is always the possibility for unsampled species to be responsible for the

observed signals.

Our results highlight the complex relationship between farmed animals and their wild

relatives. The diversity that exists within the wild may be used to enhance that of farmed

populations by selective breeding (Bentsen et al., 2017). However, this potentially important

diversity may also be lost due to unintentional hybridization with farm escapees. This

accidental hybridization may also in turn have a harmful effect on farm populations, by

potentially introducing alleles less beneficial for farming. This dynamic is particularly

pronounced in *Oreochromis*, and requires ongoing genomic monitoring and assessment, to

meet both food security and conservation outcomes.

Data availability

All raw-read data generated will be made available in the European Nucleotide Archive upon

publication. All SNP datasets and inferred phylogenetic trees will be made available in dryad

upon publication. All custom R and python scripts will be made available on github upon

publication.

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