Genome wide assessment of genetic diversity and transcript variations in 17 accessions of the model diatom *Phaeodactylum tricornutum*

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Abstract

Diatoms, a prominent group of phytoplankton, have a significant impact on both the oceanic food chain and carbon sequestration, thereby playing a crucial role in regulating the climate. These highly diverse organisms exhibit a wide geographic distribution across various latitudes. In addition to their ecological significance, diatoms represent a vital source of bioactive compounds that are widely used in biotechnology applications. In the present study, we investigated the genetic and transcriptomic diversity of 17 accessions of the model diatom *Phaeodactylum tricornutum* including those sampled a century ago as well as more recently collected accessions. The analysis of the data reveals a higher genetic diversity and the emergence of novel clades, indicating an increasing diversity among the *P. tricornutum* species complex, compared to the previous study. Transcript analysis identified novel transcript including non-coding RNA and other categories of small RNA such as PiwiRNAs. Additionally, transcripts analysis using differential expression as well as Weighted Gene Correlation Network Analysis has provided evidence that the suppression or downregulation of genes cannot be solely attributed to loss of function mutations. This implies that other contributing factors, such as epigenetic modifications, may play a crucial role in regulating gene expression. Our study provides novel genetic resources, which are now accessible through the platform PhaeoEpiview (https://PhaeoEpiView.univ-nantes.fr), that offer both ease of use and advanced tools to further investigate microalgae biology and ecology, consequently enriching our current understanding of these organisms.

Introduction

Photosynthetic microalgae are important components of life in the oceans providing organic biomass and fueling a range of key biogeochemical processes. Diatoms in particular are widely recognized as one of the most significant phylum of phytoplankton, owing to their substantial contribution to primary productivity, carbon fixation, and biogeochemical cycling of essential nutrients such as nitrogen and silicon [1, 2]. In addition to their ecological importance, diatoms are a rich source of bioactive compounds with diverse applications in various industries, including nutraceuticals, nanotechnology, pharmaceuticals, and food and feed industries [3, 4]. In recent years, using model species in diatoms has dramatically increased our knowledge about the biology and ecology of these important organisms [5]. Notably, one species, the diatom *Phaeodactylum tricornutum* has proven to be a robust model for research, yielding a wealth of knowledge and advancing our understanding in this field of research.

P. tricornutum, a marine pennate diatom is commonly found in coastal waters, including tidal areas, estuaries, rock pools and shallow waters exposing the species to important fluctuations in light intensity and salinity. The diatom is a well-established model with a genome that has been fully assembled and well annotated as well as an expanding molecular tool box using the reference strain Pt1 8.6 [6-10]. Genome wide sequencing of ten accessions of *P. tricornutum* (Pt1 to Pt10) using Illumina, identified throughout the genome diverse variations, including single nucleotide and insertion deletion polymorphisms (SNPs, INDEls) and copy number variations [11]. This study provided important insights into the genetic diversity of the isolates clustering them into four distinct clades with a conserved genetic and functional makeup. Previous studies have revealed distinguishing features among different accessions. Pt4 displayed a low non-photochemical quenching (NPQ), Pt5 demonstrated higher adhesion, Pt6 exhibited substantial lipid accumulation, Pt8, Pt3 and Pt9 have different cell morphologies and Pt3 demonstrated increased tolerance to variations in salinity, among other traits [12-14].

The 10 sequenced accessions of *P. tricornutum* were mostly collected more than a century ago and have been preserved either as lab cultures or frozen stocks in culture collections for extended periods of time. As a consequence, their genetic composition may have been affected, potentially favoring genes that are adapted to lab conditions [15-18]. Seven more recent isolates were collected from the environment and sequenced, Pt11

(HongKong, China), Pt12, Pt13 (both from Bourneuf Bay, West Atlantic, France), Pt14 (Gulf of Salerno, Italy), Pt15 (East China sea), Pt16 (Helgoland, Atlantic Ocean, North Sea, Germany) and Pt17 (Banyuls Bay, Gulf of Lion, France) (Figure 1, Table S1).

DNA sequence polymorphism can be the primary cause in phenotypic variations but it remains only the first step in understanding how these polymorphisms can affect the phenotype. Variations in transcript levels are another proxy to better understand the contribution of genes to phenotypic variations. Differences in gene expression can be attributed to different DNA sequence polymorphisms including SNPs and INDELs that can nullify gene function or induce variations in splicing. It is important to ask whether the genes that show differences in expression are under selective pressure and whether there is a link between transcript level variations and DNA sequence polymorphisms. DNA sequence may not explain differences in gene expression, cases rather known to be the consequences of epigenetic factors including DNA methylation and post translational modifications of histones [19]. These aspects are overlooked in genetic diversity studies in microalgae which mostly consider only DNA sequence variations.

In the present study, we analysed the genetic diversity of 17 accessions of the model diatom *P. tricornutum* by examining both DNA sequences and transcript levels. These accessions were collected from various coastal regions of world seas and oceans including recently sampled accessions that were not included in the previous study that had sequenced accessions collected over a span of 100 years. Our findings indicate a higher genetic diversity that defined more distinct clades and long term balancing selection of genes in old and newly sampled accessions. However, no clear link between the year of sampling and genetic diversity could be established, rejecting the hypothesis of loss of heterozygoty in cultured strains. Furthermore, our study identified novel transcripts among which various non coding RNA species and provides insights into the regulation of genes mediated by genetic and transcript diversity. Our study offers easy and valuable access to these novel genetic resources, particularly focusing on a model species, through the PhaeoEpiView platform (https://PhaeoEpiView.univ-nantes.fr) [10]. This unprecedented accessibility provides a multitude of opportunities for exploring diverse ecological niches by leveraging the genetic diversity of this model organism, thereby expanding our understanding of the biology and ecology of microalgae.

Material and methods

Material used and growth conditions

A total of eighteen different accessions of *P. tricornutum* were acquired from the Provasoli-Guillard National Center for Culture of Marine Phytoplankton, Roscoff and Nantes culture collections (Table S1). All of the accessions were grown axenically using artificial sea water (EASW) [20] in batch cultures at 19°C, under 12/12 light dark period with a light intensity of 70 µmol photons m^{-2} s^{-1}.

Growth curves

The cultures were grown in 30 ml of EASW with an initial concentration of 10^5 cells / ml. Cell counts were measured using flow cytometry (CytoFLEX, Becman, USA) every 2 days for 20 days, with 1 ml of sample taken from each accession culture each time. After 7 days of culture, 1 ml from each of Pt11 to Pt17 were used for light microscopy to describe variant shapes and cell size using Axio inverted microscope (ZEISS, Germany). Photos were further analyzed using Zeiss software (ZEN 2.6).

Pulse amplitude modulated variable *chlorophyll a* measurements

Variable fluorescence measurements were carried out using an Imaging PAM fluorometer (Walz) with a blue measuring light (450 nm), controlled by the software ImagingWin v2.46i (Heinz Walz GmbH, Effeltrich, Germany). The actinic and saturating light were also blue and provided by fluorometer LED panel. The saturation pulse intensity was 6000 µmol photons m^{-2} s^{-1} for 0.8 s. Samples were dark-adapted for one hour before carrying out any measurements. For the construction of RLC (Rapid Light-response Curves) [21], the samples were exposed to nine incremental intensities of actinic light with an irradiance step duration of 30 s. The PAR (photosynthetically active radiation; 400 – 700 nm) steps used were: 0, 5, 19, 31, 37, 42, 47, 56, 75, 143, 280 and 519 µmol photons m^{-2} s^{-1}. The first point of the RLC corresponds to the dark-adapted state, yielding the minimum fluorescence yield (F_o) and the maximum fluorescence yield (F_m), allowing the calculation of the maximum PSII quantum efficiency (F_v/F_m) as $F_v/F_m = (F_m-F_o)/F_m$. The remaining light steps measured the fluorescence yield (F'), the maximum fluorescence yield (F_m') in the light-exposed state and the effective PSII quantum yield at each experimental light level (E) as $F_q'/F_m' = (F_m'- F')/F_m'$. Relative PSII electron transport rates were calculated as rETR(E) = F_q'/F_m'(E) x E and non-photochemical quenching as NPQ = $(F_m - F_m')/F_m'$. Maximum relative electron transport rates

141 ($rETR_{max}$) were estimated by fitting the RLCs with the photosynthesis-light response model of
142 [22] and maximum NPQ (NPQ_{max}) by fitting the NPQ-light response model of [23].

143 **DNA extraction and PCR protocol**

144 After 7 days of culture, cells were centrifuged at 4000 x g for 20 min and washed twice with
145 1XPBS. DNA was isolated using a CTAB protocol as described previously [24, 25]. A
146 volume of 1.5 mL of CTAB buffer (450 µL 10% CTAB, 420 µL of 5 M NaCl, 60 µL of 0.5 M
147 EDTA, 150 µL of 1 M Tris HCL) preheated to 65 °C was placed into a 2 ml plastic tube
148 together with diatom pellets and incubated for 1 hour at 65 °C, then DNA was isolated using
149 Chloroform Isoamyl (24/1) after centrifugation for 10 minutes (12000 × g). The upper phase
150 was removed and incubated with 3.2 µg of RNase A for 1 h at 37°C. DNA was isolated again
151 using Chloroform Isoamyl (24/1) after centrifugation (12000 × g) to remove protein and
152 RNA. Same volume of isopropanol and 8% volume of ice cold Ammonium Acetate 7.5M
153 were used for precipitation at -20°C overnight. Nucleic acids were recovered after
154 centrifugation (12000 × g) at 4°C and purified by absolute ethanol and 70% ethanol
155 respectively. DNA was measured using NanoDrop ND-1000 spectrophotometer (Thermo
156 Scientific, Wilmington, DE, USA). PCR amplification was carried out on Mastercycler®nexus
157 ×2 (eppendorf, Germany) in 20 µL total volume including 9.6 µL Go Taq, 50 ng DNA and 10
158 µM forward and reverse primers. The PCR program consisted of 95°C for 5 min, then 35
159 cycles of 95°C denaturation for 30s, annealing at appropriate annealing temperature (56°C-
160 62°C) for 30 s, 72°C extension for 30 s, and a final extension step at 72°C for 10 min. PCR
161 products were electrophoresed on 1% agarose gel, and the gel images were acquired using
162 EBOX CX5 System (VILBER BIO IMAGING, France). Primer sequences are listed in Table
163 S2.

164 **RNA extraction**

165 A total of 300 ml exponentially growing cells were centrifuged at 4000 x g, 4°C for 20 min
166 and immediately re-suspended in 500 µL TRIzol®Reagent (Invitrogen, Thermo Fisher
167 Scientific, USA) and vortexed vigorously before being stored at - 80 °C. RNA was isolated
168 using Trizol reagent as described previously [26]. Purity and quantity of RNA were assessed
169 using NanoDrop ND-1000 spectrophotometer (Thermo Scientific, USA). To remove
170 genomic DNA, RNA samples were treated with Ambion™DNase I (Invitrogen, Thermo
171 Fisher Scientific, USA), according to manufacturer's instructions. RNA was quantified using
172 Qubit™ RNA BR Assay Kit, 500 assays (Invitrogen, Thermo Fisher Scientific, USA) and 1

173 µg of RNA was reverse transcribed using ImPron-II™ Reverse Transcription System
174 (Promega, USA) according to manufacturer's instructions.

175 **qPCR protocol**

176 Each Q-PCR reaction was carried out in a total volume of 10 µL containing 3 µM of each
177 primer, 1 µL cDNA, 1 µL of H_2O and 5 µL 2×SYBR Green Super mix. qPCR was performed
178 in 96-well plates in a CFX96 Touch Real-time PCR Detection System (Bio-Rad), following a
179 two temperature cycle protocol: an initial denaturation at 95°C for 3 min was followed by 39
180 cycles of 95°C denaturation for 5 seconds and 60°C annealing and extension for 20 seconds.
181 A melting curve was performed after the amplification cycles to verify specificity to the target
182 amplicon (temperature ramp from 65 to 95°C with 0.5 increments). Each sample was run in
183 triplicates. Primers sequence details are shown in Table S2.

184 **Bioinformatics analysis**

185 **Variant calling analysis**

186 Paired-end Illumina libraries from each ecotype (Pt1 to Pt17) were first trimmed using
187 Trimmomatic [27] with "ILLUMINACLIP:adapters.fa:2:30:10:2:keepBothReads
188 LEADING:20 TRAILING:20 SLIDINGWINDOW:4:20 MINLEN:40". Reads were then
189 mapped with BWA-mem2 [28] on *P. tricornutum* Phatr2 assembly (accession
190 GCA_000150955.2). Variant calling and filtering were performed with GATK package
191 version 4.2.2.0, following GATK best practices [29]. In short, HaplotypeCaller module was
192 run with a call confidence of 30, a sample ploidy of 2 and double precision was activated for
193 pair-HMM algorithm. Variants that were called were functionally annotated by snpEff [30]
194 with *P. tricornutum* database v5.0 and transposable elements annotation from [7], an
195 upstream/downstream region size set at 2kb and gene putative loss of function (LOF)
196 annotation activated. Only SNPs and insertions/deletions (INDELs) were retained from
197 annotated VCF files. Finally, GATK's VariantFiltration module was used on SNPs with the
198 following filters: "QD<2.0; QUAL<30; SOR>3.0; FS>60.0; MQ<40" and on INDELs with:
199 "QD<2.0; QUAL<30; FS>200.0". INDELs of size above 50bp were extracted with
200 SelectVariant module and some of the longest were validated by PCR.

201 **Fixation index computation**

202 Fixation index (Fst) was computed with ANGSD 0.939 [31] between all 17 accessions
203 possible pairs. First, in order to compare only regions where data were present for all samples,

the callable genome size was defined where read coverage on reference was no less than 10X across all accessions. Allele frequencies were computed for all ecotypes using ANGSD "-doSaf 1 -GL 2 -minMapQ 1 -minQ 20", then folded site frequency spectrum (2DSFS) was determined with "realSFS -maxIter 100 -fold 1" for all ecotypes combinations. Finally, we computed all pairwise Fst on the resulting indexes with "realSFS fst index -fold 1". All Fst values were gathered in a matrix and displayed as a heatmap using the R [32] package Pheatmap 1.0.12 (https://scicrunch.org/resolver/RRID:SCR_016418).

Population clustering

Callable SNPs and INDELs were analyzed with ADMIXTURE 1.3.0 [33] with a random seed of 12345679, cross-validation (CV) activated and a bootstrap value of 200. Numbers of ancestral populations (K-value) were tested from 1 to 17 and cross-validation error was plotted in order to select K leading to the lowest CV error. A PCA was then performed on Q-estimates for K=15 and estimated ancestral fractions were plotted with R.

CNV and gene loss analysis

For each ecotype (Pt1 to Pt17), raw number of mapped fragments from the Variant Calling Analysis BAM files were counted on each Phatr3 gene [7] using featureCounts [34] in unstranded paired-end mode and reads were assigned to all their overlapping features ("-O" option). Genes with no counts were deemed as possibly lost. Raw counts were then normalized for each gene following FPKM formula: FPKM_normalized_count = (gene_raw_count x 10^9) / (gene_length x total_sample_counts). Similar to previous work [11], binary logarithm Fold Change (log2FC) was calculated as the log2 ratio of normalized count for each gene to the average (mean) normalized count of all the genes per accession. Genes with a log2FC >= 2 were considered as showing putative Copy Number Variation (CNV) compared to the reference strain. Finally, lost genes and genes exhibiting CNV in only one accession were marked as ecotype-specific. Heatmap plots were made in R using Pheatmap 1.0.12 (https://scicrunch.org/resolver/RRID:SCR_016418) and UpsetR 1.4.0 [35] packages.

Genes with loss of function

After variant annotation by snpEff, we used an in-house script to find the total and specific number of genes affected by loss-of-function (LoF) mutations for each ecotype (Pt1-Pt17).

First, we selected genes with variants LoF alleles retained in the VCF annotation file. Then, we searched for the genes that are specific to each accession and considered the non-accession specific genes common if shared by two or more accessions.

Site Frequency Spectrum (SFS) analysis

A matrix of allele counts per ecotype was created from the variants called previously on the callable genome. Briefly, for each biallelic SNP, a value of 0, 1 or 2 was determined for each ecotype, depending on its ploidy (homozygous on reference allele, heterozygous reference/alternate alleles or homozygous on alternate allele, respectively). Moreover, functional annotation as described previously and the affected gene (if applicable) were added for each SNP. Folded SFS was then calculated for each functional category of SNPs (nonsense, non-synonymous, synonymous, intergenic) and the resulting data was plotted with R.

Searching for signatures of Balancing Selection (BS) on non-synonymous SNPs

One of the signature of balancing selection is the excess of nonsynonymous polymorphisms segregating at intermediate frequencies [36]. Genes with less than 10 synonymous (S) + non-synonymous (NS) SNPs were filtered out from the allele counts matrix (see "*Site Frequency Spectrum (SFS) analysis*"). The ratio non-synonymous versus synonymous diversity was estimated by Watterson's theta θ assuming twice as many non-synonymous than synonymous sites (θwNS/ θwS), defined as "(Number of NS/2) / (Number of S)" was calculated for each of the remaining 9267 genes. The 91 genes with an excess of non-synonymous SNPs (showing a θ ratio over 3) were extracted and further investigated. Finally, the same process was performed ecotype-wise, with a number of genes with a θ ratio > 3 ranging from 34 in Pt14 to 62 in Pt4.

Phylogeny of the 17 accessions

A matrix of genome-wide biallelic SNPs and INDELs allele counts per ecotype (0, 1 or 2 depending on the called ploidy of the variant, see "*Site Frequency Spectrum (SFS) analysis*") was computed for 640,454 variants found in the population Pt1 to Pt17. Canberra distance and average linkage functions were identified to produce the tree that represented best the matrix by the "find_dend()" method from R library "dendextend" 1.16.0 [37]. Then, an unrooted neighbourg-joining tree was built using "phangorn" R package 2.10.0 [38] on the Canberra distance matrix and colored according to the ecotypes clades.

Expression analysis

After filtering raw data with the removal of adapters and low quality reads, clean reads were aligned against the reference genome using HISAT2 2.0.5 [39]. Reads were assigned to each transcript using the FPKM metric which normalizes for differences in library size and gene length. In order to compare gene expression levels in different accessions, the graphical representation of the distribution of gene expression and FPKM levels in different samples has been performed using the ggplot2 R package (v3.4.0)[40].

Principal Component Analysis (PCA)

To elucidate the relationships between distinct accessions, we conducted Principal Component Analysis (PCA) on the gene expression values (FPKM) of all the samples. Specifically, we first computed the average FPKM value for two replicates of each sample, followed by a logarithmic transformation of this average value (log2(FPKM+1)). Ultimately, we represented the samples according to their expression levels. In our analysis, we evaluated how well the samples were represented in PCA using the cos2 (square cosine) metric as a measure of quality. A cos2 value closer to one indicates a stronger representation of the variable by the two displayed components.

Repeat detection in novel transcripts

Reads from both replicates of Pt1 to Pt17 RNAseq libraries were mapped with HISAT2 2.0.5 [39] using the default parameters and all the mapping information were combined. The reads were then assembled with StringTie 1.3.3b [41] and novel transcripts were kept. We screened these 656 novel transcripts for repeats with RepeatMasker (*RepeatMasker Open-4.0*. 2013-2015 <http://www.repeatmasker.org>) Galaxy Tool wrapper version 4.0.9 (slow settings with matrices for 43% GC content), using a manually curated database of 71 reference transposable elements (TE) of *P. tricornutum*. RepeatMasker was run on the public server at https://usegalaxy.org [42].

WGCNA network analysis

The network was obtained using Weighted Gene Correlation Network Analysis (WGCNA) R package (version 1.17) [43]. Before constructing the co-expression network, we filtered out genes having a row median less than 10 reads. The expression matrix was transformed with the vst (Variance Stabilizing Transformation) function from DESeq2 R package (version

1.32.0) [44]. The sequencing steps for the network construction for *P. tricornutum* accessions have been described previously [45].

For Network construction the WGCNA R package [43] was used to identify network modules from 36 RNA-Seq datasets representing expression data from 18 *P. tricornutum* accessions (two replicates per accession). First, the quality of the raw counting matrix was checked. A hierarchical clustering analysis based on the "average" method allowed us to identify the Pt1R2 as an outlier, so this sample was filtered out from further analysis.

Results

Phenotypic traits characterization

To assess phenotypic differences among the 17 accessions, we monitored their growth, cell morphology and photosynthetic features. Significant differences in growth rate were recorded at day 4 of the exponential phase (Fig.2a). In most of the cultures, cell growth rate dropped after day 11 and entered the stationary phase. Pt8, Pt3 and Pt10 showed faster growth rate and higher final concentrations than other accessions, with 1165×10^4, 1032×10^4 and 960×10^4 cells mL^{-1} respectively in the end of exponential phase ($p<0.05$). On the other hand, Pt4 and Pt9 showed slower growth rate than the others and the lowest final concentrations, with 419×10^4 and 559×10^4 cells mL^{-1} respectively.

Cell dimensions were measured for only 7 accessions (Pt11 to Pt17) and compared to the previously published Pt1 to Pt10 accessions [12]. Among all accessions, the previously measured Pt5 had the longest length (25-30 µm) and Pt14 cells were the shortest (10-15 µm). Pt12 were the thinnest cells with the lowest length/width ratio (9.9 ± 1.1) and Pt17 were the largest (4.6 ± 0.8) (Table S1, Fig. S1). Different morphotypes were observed in Pt16 (a mix of 75% fusiform and 25% of triradiate). As reported previously [11], we found few oval cells mixed with fusiform in Pt3 and Pt9 (7.3% and 7.6% respectively). Triradiate were reported in Pt8 [12] but we did not observe triradiate cells in our conditions. Triradiate morphotype was reported to be instable in this accession [46].

Measurements of photosynthetic parameters

330 To assess photosynthetic capacities of *P. tricornutum* accessions, we measured maximal
331 PSII quantum yield (F_v/F_m) and maximal relative electron transport rates (rETRmax). Among
332 all the accessions, Pt12 showed the highest F_v/F_m followed by Pt11 (p = 0.0047) and Pt13 (p =
333 0.027) (Fig. 2c). These three accessions also showed the best photosynthetic performances
334 based on rETRmax (Fig. 2b), while Pt4, Pt5, and Pt6 demonstrated the lowest values in
335 rETRmax and PSII quantum efficiency (Fig 2c). The English Channel accessions (Pt1, Pt2
336 and Pt3) and Eastern American accession (Pt5, Pt6 and Pt7) showed similar results.

337 To evaluate the response to excess light energy, we measured non-photochemical
338 quenching (NPQ). Pt9 and Pt17 showed the highest NPQ capacity, c. 8.2 and c. 9.2
339 respectively while Pt4 showed a lower NPQ capacity of around 2.5 (Fig 2d). These
340 observations suggest that Pt9 and Pt17 can tolerate environments with higher light intensity
341 compared to other accessions. This is consistent with their geographical distribution in
342 latitudes that receive greater amounts of solar radiation. Similarly, Pt4 NPQ reflects an
343 adaptation to its sampling location in higher latitudes, specifically the Norwegian Fjords.

344 **Variant calling analysis**

345 We performed variant calling analysis on previously published sequences of Pt1 to Pt10
346 [11] and the newly sequenced Pt11 to Pt17 accessions using the reference strain Pt1 8.6
347 genome sequences. All the accessions had a good sequence coverage allowing a confident
348 variant calling. We identified 731,357 single nucleotide polymorphisms (SNPs), 44,470
349 insertions (from 1 to 422 bp length) and 52,867 deletions (varying from 1 to 274 bp) (Fig. 3
350 A,B). Site frequency spectrum (SFS) which reflects the numbers of variants segregating at
351 different frequencies showed the expected excess of low frequency alleles (Fig. 3C). The
352 highest increase in low frequency SNPs was observed in non-sense polymorphisms followed
353 by non-synonymous polymorphisms as compared to intergenic and synonymous
354 polymorphisms. Most of the SNPs (59-63%) were found in genes while INDELS were mostly
355 detected in intergenic regions (Fig.3 D). Our analysis identified 22,4253 additional SNPs and
356 74,918 additional INDELs in Pt1 to Pt10 compared to our previous study [11].

357 Despite the higher number of discovered SNPs and INDELs, the overall trend of their
358 distribution among the 10 previously analyzed accessions remains the same. Across all the
359 accessions most of the SNPs were heterozygous (HetSNPs) with >95% in Pt1, Pt2, Pt3, Pt9,
360 Pt15 and Pt17 and 65% to 68% in Pt6, Pt7, Pt8 and Pt16 and the lowest proportion of

361 HetSNPs were found in Pt4, Pt5 and Pt10 to Pt14 (<49%). Across all the accessions, the
362 numbers of INDELs was similar except for Pt4, Pt6, Pt7 and Pt8 which showed the highest
363 number of INDELs (Fig. 3B, Table S3). Most INDELs were shared among the accessions. A
364 total of 14 INDELs were validated by PCR for randomly chosen loci (Fig. S2). To further
365 assess genetic diversity, we investigated copy number variations (CNVs) and gene loss (GL).
366 A total of 284 and 180 genes show CNV or GL respectively. Most of CNVs are shared and
367 eleven accessions out of 17 show specific CNVs with 40 genes in Pt10 followed by Pt4, Pt6,
368 Pt14 and Pt16 with 15, 15, 8 and 7 genes respectively (Fig. 3E, Table S4, Table S5).
369 Randomly chosen loci were validated by PCR for gene loss (Fig. S3).

370 To understand the functional impact of genetic variations among *P. tricornutum*
371 accessions, we examined the loss of function mutations (LoF) such as premature stop codons,
372 frameshifts and start loss. A total of 31,536 LoF was found, among which 588 were shared
373 across the accessions (Fig. 3F). Accession specific LoFs were mostly found in Pt4 (61), Pt14
374 (20) and Pt16 (17) (Table S6). LoF mutations were enriched in gene ontology (GO) categories
375 of molecular function type (Table S6) and in genes that belong to large gene families as
376 previously shown [11].

377 **Population structure and phylogeny of *P. tricornutum* accessions**

378 To examine the global population structure of *P. tricornutum* accessions, we used
379 pairwise fixation index (Fst), a measure of genetic differentiation revealing groups with low
380 Fst index (< 5%) (Figure 4A). To further estimate genetic relatedness among *P. tricornutum*
381 accessions, we used admixture proportion inference which allows the assignation of
382 individual genetic variations into clusters based on shared allele frequency patterns [33]. We
383 ran ADMIXTURE with various plausible values of *K* which is the number of source
384 populations and found a stable admixture pattern proportions with K=15, reflecting the
385 number of ancestral populations (Fig. S4, Table S7). Based on individual ancestry with
386 similarity of clusters between accessions, we distinguished 6 clades: Pt1, 2, 3, 9, 15 and 17 in
387 clade 1 with most of the clusters (up to 11) reflecting a clade with multiple ancestral
388 populations, Pt16 in clade 2, Pt4 in clade 3, Pt5, 10 and 11 in clade 4 with only 3 clusters, Pt6,
389 7 and 8 in clade 5 and Pt12, Pt13 and Pt14 in clade 6 (Figure 4B).

390 To confirm admixture analysis clades, we performed a Principle Component Analysis
391 (PCA) which revealed similar results reflecting common ancestry except for Pt14 which is far

392 from its admixture defined clade 6 (Fig. 4C). Of note, cluster composition proportions of Pt14
393 are different from Pt12 and Pt13. To further assess the segmentation among *P. tricornutum*
394 accessions, we used CNVs to run a hierarchical clustering and found that the 17 accessions
395 fall into 6 clusters supporting further both PCA and admixture analyses: Pt1, Pt2, Pt3, Pt9,
396 Pt15 and Pt17 in cluster 1, Pt16 in cluster 2, Pt4 in cluster 3, Pt6, Pt7 and Pt8 in cluster 4, Pt5,
397 Pt10 and Pt11 in cluster 5 and Pt12, Pt13 and Pt14 in cluster 6 (Fig. 4D). Phylogenetic
398 analysis at whole genome scale using genetic variations (SNPs and INDELs) of the 17
399 accessions supported further the clustering into six clades observed with Fst and PCA
400 analyses (Fig. 4E).

401 **Balancing and relaxed selection in Pt clades**

402 We calculated the ratio of nonsynonymous to synonymous nucleotide site diversity using
403 Watterson's estimate of theta (θwN/θwS) [47] as a measure of the efficiency of natural
404 selection. Ratios > 3 suggest that selective pressure maintains non-synonymous
405 polymorphisms, a signature for balancing selection (BS) which refers to selective processes
406 by which alleles are maintained in a population at frequencies larger than expected from
407 genetic drift alone [48], while θwN/θwS ratios < 1 refer to non-synonymous polymorphisms
408 being counter-selected pointing to a purifying selection. We identified 91 common genes
409 under BS and 2422 under purifying selection (Table S8). Most of the genes under BS are of
410 unknown function. However, those with known function are enriched in genes coding for
411 functions such as cell proliferation and growth, perception, transmembrane transport activity,
412 stress responses and adaptation to the environment.

413 **Identification of transcript level variations and co-expression network modules in *P.***
414 ***tricornutum* accessions**

415 Differences in gene expression is known to control inter and intra specific phenotypic
416 variations providing living organisms with abilities to colonize different ecological niches. To
417 identify differences in transcriptomes, RNAs from *P. tricornutum* accessions (Pt1 to Pt17)
418 were sequenced and mapped to the reference genome. Mapping of mRNAseq reads varied
419 from 41.27% to 94.75%. Pearson correlation coefficients between each of the two replicates
420 was mostly around 0.98 (Fig. S5). Interestingly, a total of 656 genes including 25 from the
421 chloroplast were novel. These genes are widely distributed over the genome among which
422 some were specific to each of the ecotypes and 385 genes were found to be common to all of
423 them (Table S9). They show an average length of 709.42 bp and are for nearly half of them

424 sense transcripts (334 genes) and antisense (322 genes). Except from few genes which were
425 annotated, most of these novel genes were of unknown function (Table S9). The majority of
426 novel genes were downregulated but their expression remains significant and cannot be
427 considered as part of a spurious phenomenon of background low level transcription.

428 The analysis of novel transcripts using RepeatMasker revealed that 12.26% of them
429 contained repetitive elements, primarily Copia LTR_retrotransposon of class I and rare
430 MuDR2 Terminal Inverted Repeats of class II (Table S10). Additionally, approximately
431 0.48% of the transcripts contained simple repeats. Using a coding potential assessment tool,
432 we identified several non-coding RNA with sizes varying between 202 bp and 8379 bp (Table
433 S11). Furthermore, we detected several other RNA types, including sRNA, miRNA, tRNA,
434 snoRNA, antisense and piwiRNAs (Table S11).

435 Then, we looked at differentially expressed genes (DEG) in our standard growth
436 conditions using RNA-Seq data across the 17 accessions. The reference strain Pt1 8.6, used
437 for comparative gene expression was derived from Pt1, which displayed the lowest number of
438 DEG (1308 genes) as expected (Figure 5 A). In contrast, Pt7 showed the highest number of
439 DEG (5086 genes). The remaining ecotypes showed variable numbers of DEG, with Pt5
440 exhibiting the lowest number at 1890 genes. On average, most ecotypes had approximately
441 4000 DEG. With the exception of Pt1, which exhibited a bias towards upregulation (twice as
442 many upregulated genes as downregulated genes), the other ecotypes displayed a balanced
443 ratio of upregulated and downregulated genes. A substantial fraction of DEGs, regardless of
444 their upregulation or downregulation status are found to be specific to one or multiple
445 ecotypes suggesting an ecotype-dependent gene expression pattern that likely underlies
446 ecotype specific trait (Figure 5B, C, Table S12, S13). On the other hand, only a minor subset
447 of DEGs displaying upregulation or downregulation are shared across all the ecotypes. We
448 conducted a principal component analysis using the average replicates expression to evaluate
449 whether the ecotypes exhibited comparable expression profiles. Our analysis distinguished
450 five clusters that partially aligned with the clades defined in this study, implying a correlation
451 to some extent between genetic diversity and expression patterns (Figure 5D).

452 To explore the relationship between phenotypic traits, specifically photosynthesis and
453 transcripts, we closely examined the NPQ response and the genes related to its regulation in
454 response to light. Indeed, the NPQ capacity depends on transthylakoidal proton gradient, but
455 also on antenna proteins called Light-Harvesting Complex Protein X (LHCX) and on the

Diatoxanthin Xanthophyll cycle [49], equivalent to the Zeaxanthin cycle in land plants [50]. Among the LHCX genes, only LCHX1 shows strong expression in all ecotypes, confirming its constitutive role in low light, while LHCX2 and 3 are involved in high light and LHCX4 expression increases in the dark [51]. Similarly, all genes involved in the Diatoxanthin Xanthophyll cycle show negligible expression in our low light culture conditions.

To understand the underlying nature of the conserved transcriptomic responses, we analyzed the enrichment of GO terms for both upregulated and downregulated DEGs (Figure 6BC). Additionally, we performed GO enrichment analysis on genes that were specifically upregulated or downregulated in a single accession as well as per clade, where applicable. Only few GO terms emerged from the analysis of accession specific DEGs, namely photosynthesis GO related terms (light harvesting, protein chromophore linkage) in Pt8, lipid metabolic processes and translation in Pt3 for downregulated genes while upregulated genes were enriched in ribosome biogenesis and rRNA processing in Pt7, protein transport in Pt16 and glucose metabolism in Pt11 (Figure 6BC, Table S12, S13).

The WGCNA package was used to construct gene co-expression network of transcripts from an expression matrix of ~432,000 transcripts derived from 36 RNA-seq samples, with 2 replicates collected from the 18 accessions including the reference strain Pt1 8.6. This approach yielded in 33 distinct co-expressed modules (labeled by different colors) with dark slate blue and plum2 modules containing each the smallest number of genes (106) and green yellow with the largest number of genes (1599) (Figure 6A, Table S14). These modules were constituted by genes demonstrating analogous expression profiles, which may or may not be consistent among different clades suggesting the existence of additional factors besides genetic polymorphisms that could modulate transcript levels. (Figure S6). The modules were further categorized into six distinct clusters, each characterized by a group of genes exhibiting comparable expression patterns thus implying their involvement in shared pathways (Fig. 6A, Table S15). Based on the GO annotation analysis, we identified significant functional enrichments in different groups. Group I displayed a substantial increase in oxidoreductase activity, while Group II showed an enrichment in calcium ion binding activity. Group III was characterized by an enrichment in chlorophyll binding and light harvesting, whereas Group IV was associated with RNA processing and translation. Group V showed a significant enrichment in photosynthesis and cell redox homeostasis, while Group VI exhibited an enrichment in cell division and DNA binding activity (Table S15).

Discussion

The present study was designed to comprehensively characterize the phenotypic, genetic, and transcriptomic diversity among seventeen distinct *P. tricornutum* accessions, which were collected from various locations across the world's oceans and included more recently sampled accessions compared to those examined in prior studies [11, 12]. Growth dynamics, cell morphology, and photosynthetic traits were monitored, and significant inter-accession differences were recorded. Notably, the Pt4 and Pt9 strains exhibited a distinct growth pattern, implying a probable correlation to their respective sampling locations. More specifically, Pt4, sampled from the Norwegian fjords, seems to be adapted to a lower light intensity regime than that employed in our study, while Pt9, a tropical strain, showed a slower growth at 19°C compared to the presumed higher temperature in its geographical location. Moreover, the evaluation of photosynthetic abilities across various accessions corroborated the association with the sampling sites. Pt4 displayed the smallest $rETR_{max}$ and PSII maximum quantum efficiency, while Pt12, Pt13, and Pt14 collected from the Mediterranean Sea and Atlantic side demonstrated higher photosynthetic performance, as evidenced by their important F_v/F_m and $rETR_{max}$ values. Measuring cell dimensions in the recently acquired accessions and their comparison with previously characterized ones revealed notable variations. Specifically, Pt12, Pt14, and Pt17 exhibited significant deviations, with Pt12 having the shortest cell length, Pt14 displaying the smallest length-to-width ratio, and Pt17 showing the largest cells when compared to the remaining accessions. Additionally, our investigation identified Pt16 as a new accession with a combination of triradiate and fusiform morphotypes. These disparities in cell sizes may confer an advantageous trait in terms of enhanced gliding capabilities and improved photosynthetic efficiency [52].

Variants calling analysis showed a larger number of SNPs and INDELs than previously reported in Pt1 to Pt10. This is due to the gapped alignment mode used for SNPs and INDELs calling which performs better than ungapped mapping [53]. Improvements made to HaplotypeCaller's algorithm since 2018 were also likely playing a role in the gain of sensitivity we noticed. Most of the SNPs were located in coding regions, while INDELs were mostly found in intergenic regions as a consequence of their highly deleterious effects within coding regions. Most of the SNPs were found to be heterozygous, indicating that *P. tricornutum* has a high level of heterozygosity. Our previous study has demonstrated a substantial level of heterozygosity in *P. tricornutum* populations. The recent sampling of genetically distinct accessions has reaffirmed the persistence of this trait, despite not being

522 related to the previously identified highly heterozygous accessions Pt1, Pt2, and Pt3, thereby
523 supporting the reliability of the heterozygosity measure. This high level of heterozygosity is
524 intriguing in *P. triconrutum* considering that the species is not known to reproduce sexually
525 suggesting an advantage of heterozygous alleles or the detrimental homozygous alleles that
526 get selected against as reported in inbred population of clonal honey bees, *Apis mellifera*
527 *capensis* which retained after 20 years of inbreeding high heterozygosity throughout its
528 genome due to selection against homozygotes [54]. Similar examples of heterozygosity
529 advantage through its maintenance at high proportions of the genome were reported in other
530 species, isolated wolf populations and a hermaphrodite worm after several generations of
531 selfing [55, 56]. An alternative explanation for the observed high heterozygosity could be due
532 to the mutations that occurred in the ancestral lineage of Pt1, Pt2, Pt3, Pt9, Pt15, which was
533 revealed through admixture analysis, indicating that these accessions share a common
534 ancestry and are closely related to Pt17, which also exhibits high heterozygosity. In contrast,
535 accessions with lower heterozygosity display a distinct ancestry pattern.

536 The SFS analysis provided compelling evidence consistent with the predictions of the
537 nearly neutral theory of evolution, revealing an excess of low frequency alleles. This
538 prevalence of lower frequency alleles in non-sense and non-synonymous polymorphisms can
539 be attributed to the effects of purifying selection, which acts against deleterious mutations.
540 Consequently, our investigation aimed to examine whether there was an elevated occurrence
541 of non-synonymous mutations in the Pt1 8.6 reference strain, in contrast to both the original
542 Pt1 accession and the closely related Pt2 strain within the same clade. This analysis sought to
543 determine if the Pt1 8.6 strain had undergone a process of "domestication." However,
544 unexpectedly, we did not observe differences in non-synonymous mutations. Instead, we
545 observed a remarkable predominance of LOF mutations in the reference strain Pt1 8.6,
546 suggesting an adaptation to laboratory culture conditions facilitated through LOF mediated
547 mechanisms. The majority of these LOF mutations resulted in the repression or reduced
548 expression of targeted genes. However, a substantial number of genes showed moderate to
549 high expression levels, implying that these LOF mutations may function as an evolutionary
550 mechanism for generating new functional genes, serving as an adaptive response to culture
551 conditions [57-59]. It is important to note that not all LOF mutations lead to complete
552 functional knockout. For instance, LOF mutations at the 5' or 3' regions of genes may not
553 entirely abolish their functions, and truncated proteins resulting from such mutations could act
554 as dominant-negative factors [60, 61]. Interestingly, about 10.75% (331 out of 3078 genes)
555 LOF showed moderate to high expression. Some specific examples of these genes with

known functions include: (i) a mitochondrial enzyme known as glutamate dehydrogenase (Phatr3 J13951), which has been reported to play a crucial role in carbon and nitrogen metabolism as well as energy supply under abiotic stresses in *Arabidopsis* and the red alga *Pyropia haitanensis* [62-65] ; (ii) an LCH15 protein (Phatr3 J48882), which functions as a chlorophyll binding protein and potentially contributes to the adaptation to different light conditions in laboratory cultures and (iii) a heat shock transcription factor (Phatr3 J49594) which similarly may contribute to the adaptation to lab culture temperatures.

Admixture analysis, PCA, and hierarchical clustering all identified six clusters, which suggests that the samples had shared ancestry and similar geographical origins. However, not all accessions within the same cluster had shared geographical origins. The English Channel and East China Sea populations clustered together, indicating that *P. tricornutum* accessions may have been dispersed by various means, or that there were similar ecological niches across the sampling sites. Genome-wide phylogeny analysis confirmed six clades, with some accessions falling into previously identified clades and others forming two new clades, which suggests genetic divergence.

Several loci that are believed to be under balancing selection were found to have a high level of genetic diversity. Notably, genes coding for stress-inducible proteins (Phatr3_EG00471, Phatr3_J54019), outer membrane receptors (Phatr3_EG01193), and cell cycle genes (Phatr3_J34920, Phatr3_EG00817) displayed an excess of polymorphism, reflecting their role in protecting cells from stresses such as high temperatures, starvation, and infection, as well as in cell division and growth. Consistent with this, transmembrane protein have been previously observed to evolve faster than protein without a transmembrane domain in unicellular eukaryotes such as yeast [66] and *Ostreococcus* [67]. Interestingly, the genes identified under balancing selection were found in multiple clades and were specific to one or more accessions, suggesting that spatially varying selection forces may be related to local niches. It is expected that these same selection forces will apply to accessions from similar ecological niches or with a shared origin and sampling location.

Comparison of genes under BS between accessions sampled at divergent time points, namely 1910, 1930, 1956, 1989 and 2016 revealed several identical genes suggesting the persistence of long term balancing selection acting on these genomic regions (Figure 6D, Table S8). Notably, the majority of these genes were functionally associated with stress resistance and fundamental cellular processes, highlighting their potential significance in adaptation to various habitats. Long term balancing selection was found at genes involved in

589 diverse processes such as disease resistance, self-incompatibility, and heat stress providing
590 advantages and enhancing fitness in natural populations [68-71].
591
592 Profiling transcript levels in the 17 accessions identified novel genes in the assayed
593 growth conditions suggesting condition and accession specific genes that were not identified
594 in the numerous growth conditions previously reported [7]. Using expression profiling
595 analysis, we identified DEG that exhibited a clustering pattern among ecotypes similar to that
596 observed using DNA sequence variations. Interestingly, our analysis revealed that genes
597 carrying LOF mutations displayed a significant decrease in expression levels when compared
598 to their non-LOF counterparts (Figure 6E), implying the role of DNA sequence variations in
599 shaping gene expression patterns. Nonetheless, we also noticed a considerable number of
600 LOF mutations that did not result in downregulation. The observed LOF mutations with no
601 effect on gene expression is likely due to the robustness of the genome through gene
602 duplication and compensatory mechanisms allowing for the toleration of many LOF variants,
603 resulting in the majority of these mutations being silent and having little to no impact.
604 Multiple LOF mutations were observed across clades as well as within them, particularly
605 among accessions that exhibited extreme phenotypes (short cell size versus long ones, low
606 versus high photosynthetic performance). This suggests that there may be variations in genetic
607 backgrounds and/or epigenetic factors among these accessions. For instance, Pt12 and Pt14,
608 despite having vastly different cell morphologies (very long versus short cells), share 1273
609 LOF mutations. Similarly, Pt2 and Pt9, as well as Pt6 and Pt12, share 1479 and 1120 LOF
610 mutations respectively, but exhibit distinct photosynthetic performances.
611 Interestingly, we observed LOF mutations that led to upregulation of genes instead,
612 suggesting the potential existence of Gain of Function (GOF) mutations. These GOF
613 mutations known to occur mostly in unstructured regions may be attributed to the emergence
614 of transcription factor binding sites, miRNA binding sites, an RNA binding protein or new
615 functional domains [72, 73]. Since genes and their products do not operate in isolation but
616 rather in biological networks, these newly acquired domains may acquire functionality
617 through their interactions with other proteins. Additionally, compensatory mechanisms for
618 LOF or GOF may involve epigenetic processes that serve as a platform for interacting with
619 specific proteins or complexes. WGCNA analysis revealed several network modules that were
620 further merged into 6 clusters with similar expression patterns indicating co-regulated genes
621 and pathways. Both differential expression and WGCNA analysis corroborated the presence
622 of differences in transcript levels, which cannot be solely attributed to genetic variations. This

observation implies the involvement of other regulatory mechanisms, such as epigenetics, that are known to modulate gene transcription [6, 74, 75].

Conclusions

Our study provides a comprehensive assessment of the genetic and transcriptional diversity among 17 natural accessions of the model diatom *P. tricornutum*. Our investigation has uncovered novel clades, which are likely indicative of previously unexplored ecological niches. Moreover, we have identified new genes that expand the existing transcriptome repertoire of this species. By incorporating recently sampled accessions, we have further confirmed the high level of heterozygosity in *P. tricornutum* through population genetic analyses, suggesting that this characteristic arises from a heterozygous advantage. Our findings establish a crucial groundwork for future research that utilizes sequencing data from various *P. tricornutum* accessions which we made available via PhaeoEpiView platform (https://PhaeoEpiView.univ-nantes.fr) [10] for easy and comprehensive use. This will enhance our understanding of diatom biology, foster advancements in biotechnology applications, and optimize trait selection.

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Authors contribution

LT conceived and designed the study. TC performed and coordinated the bioinformatics analysis. FY conducted most of the experiments. AG extracted RNA and performed the PAM study. EM contributed to the bioinformatics analysis. BJ supervised the PAM study and assisted with data analysis. GP and LT supervised the genetic population study. OAM performed the WGCNA analysis. YW and UC assisted with DNA extraction. TC, FY, AG, GP and LT analysed and interpreted the data. LT supervised and coordinated the study. LT wrote the manuscript with input from TC, FY, AG, EM and OAM. All authors read and edited the manuscript.

Data availability

The data that support the findings of this study are openly available in BioProjects PRJNA430316 and PRJNA971163

Competing interests

None of the authors have any competing interests

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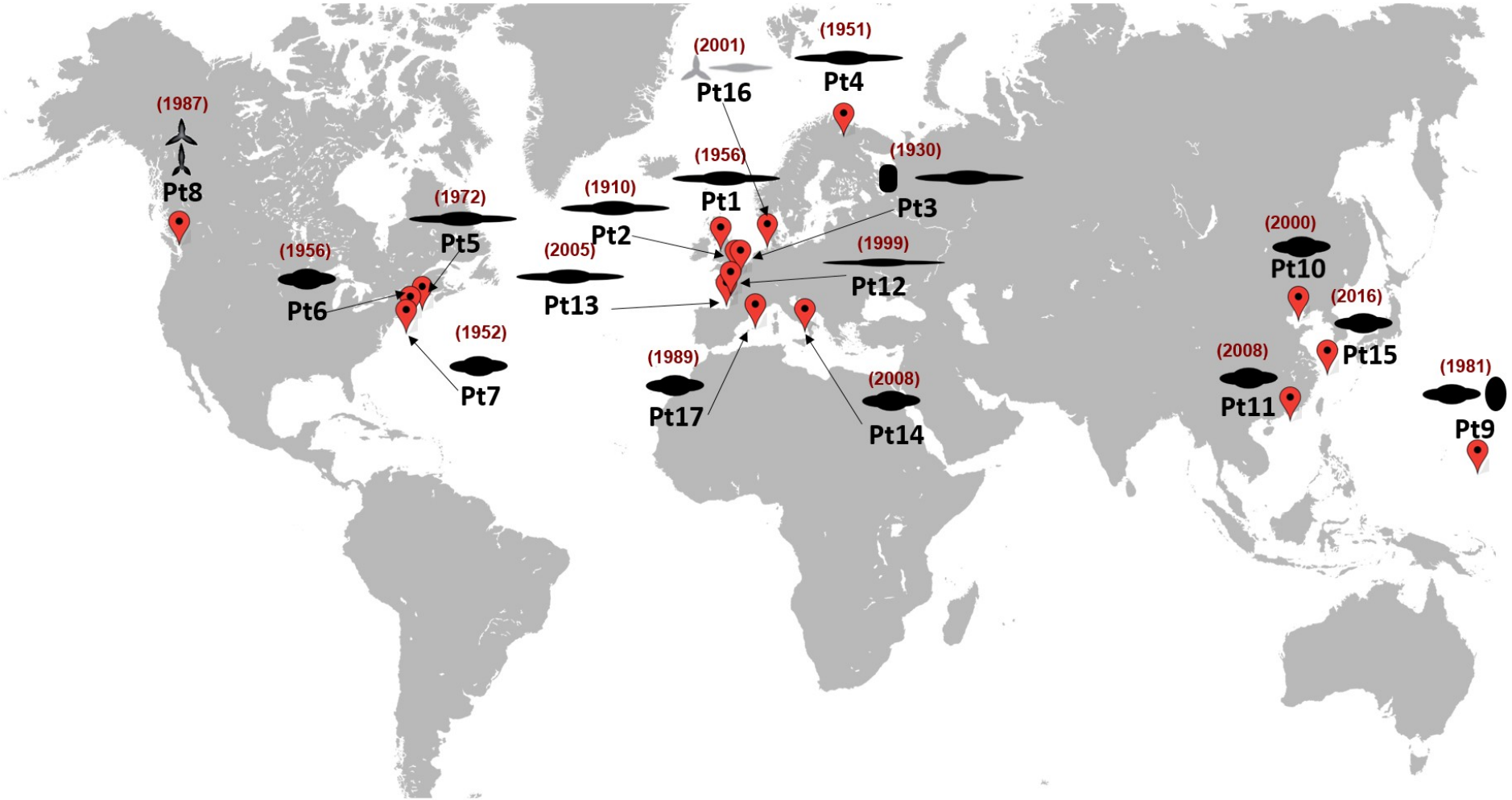
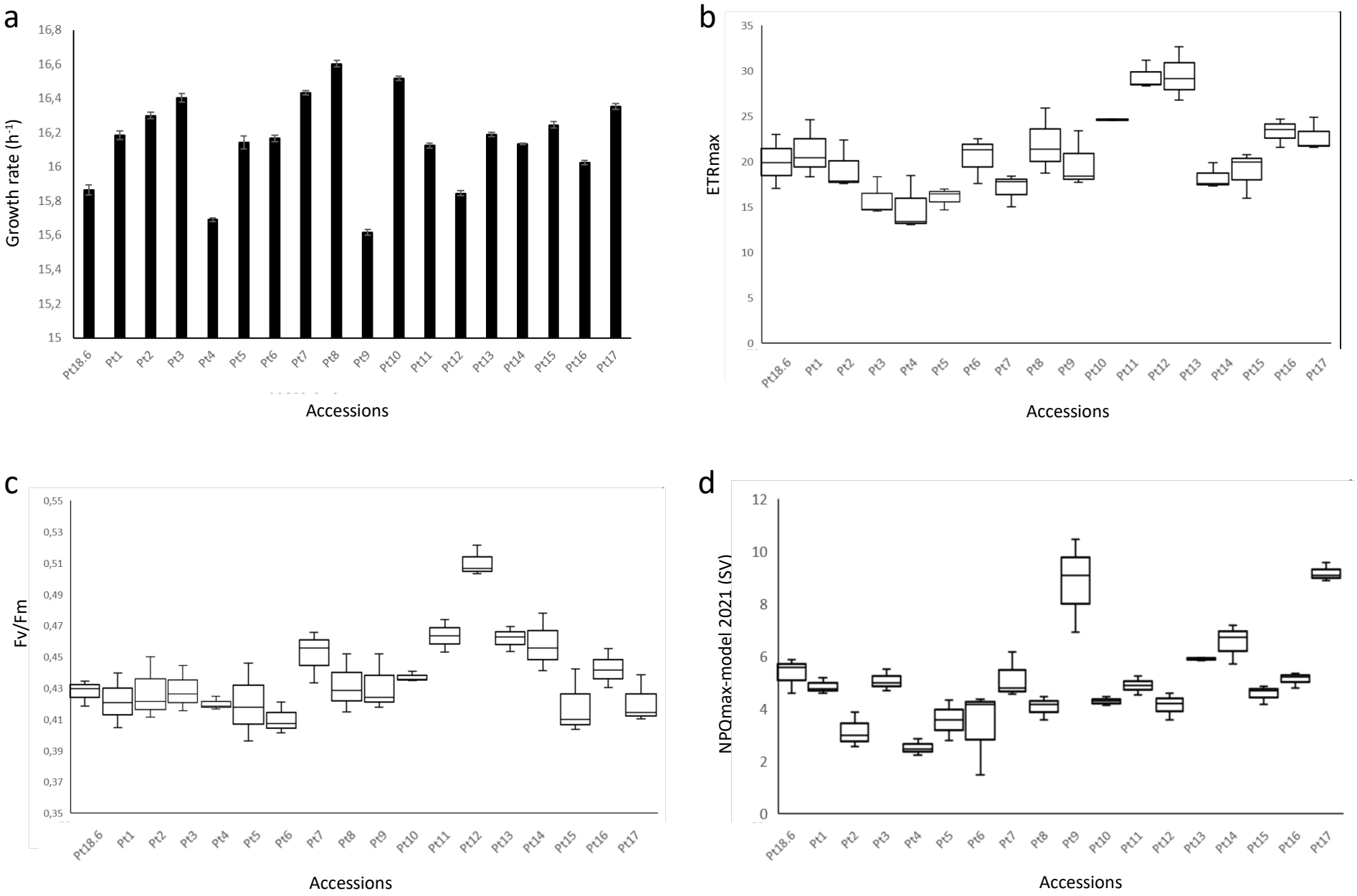
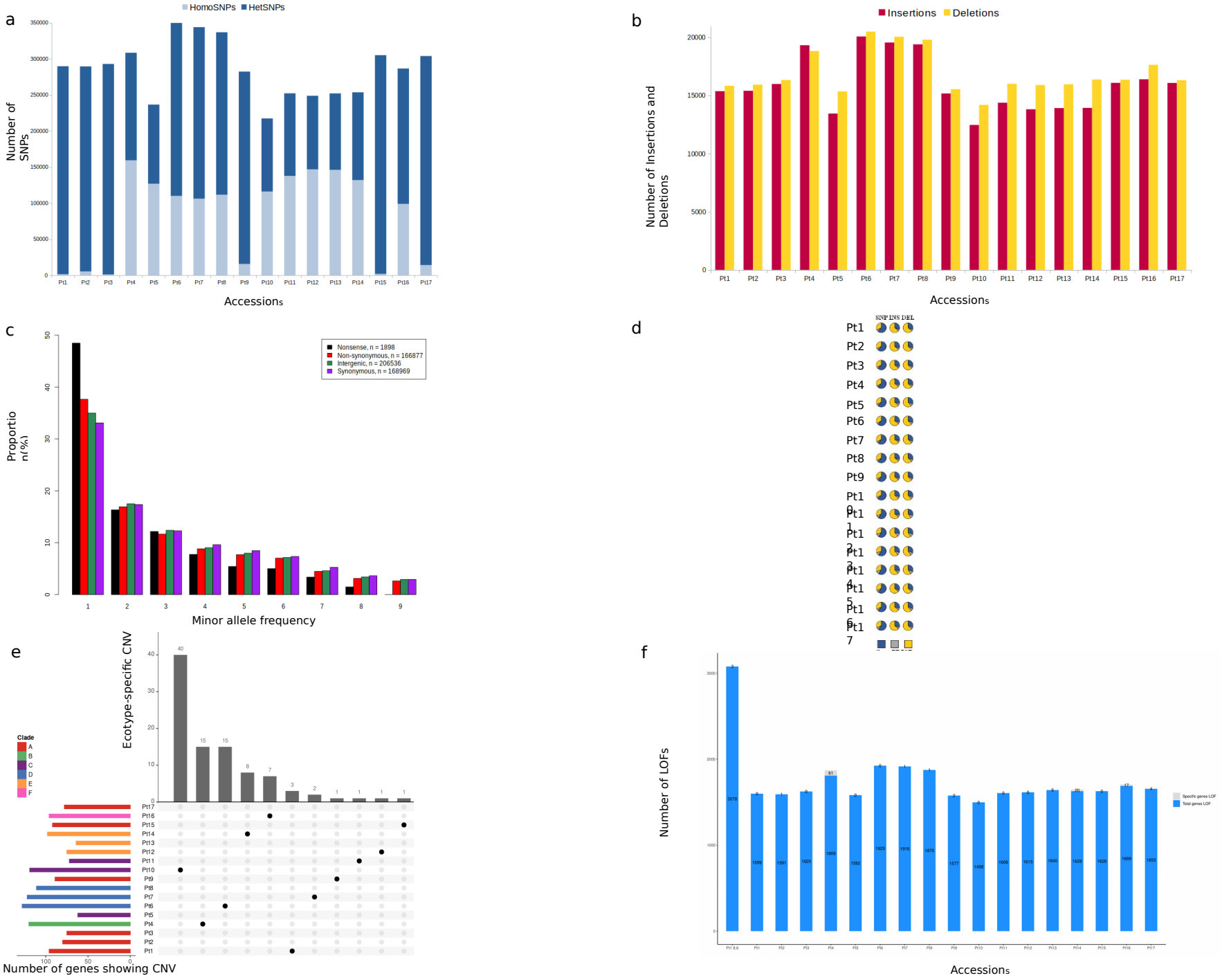
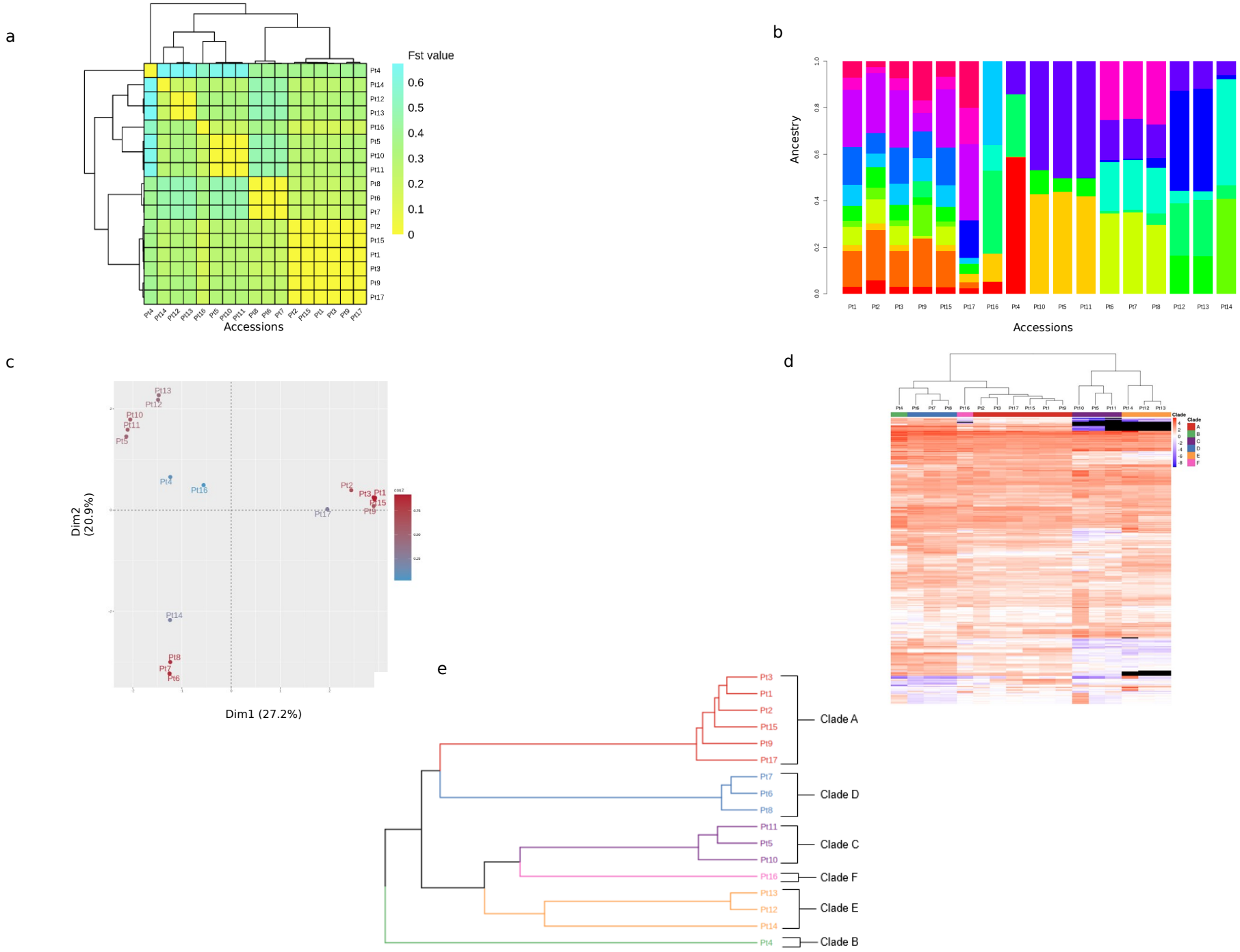
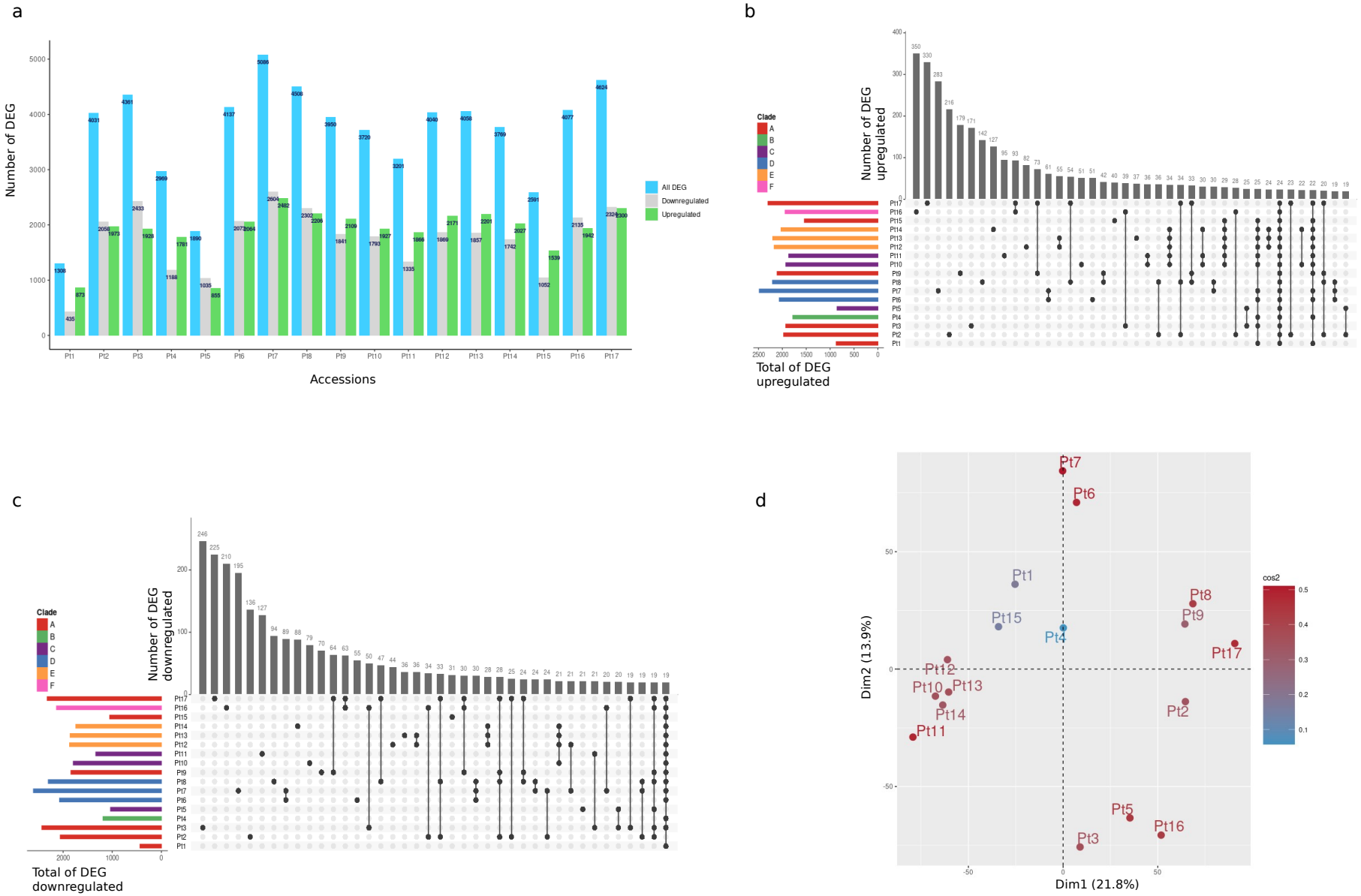
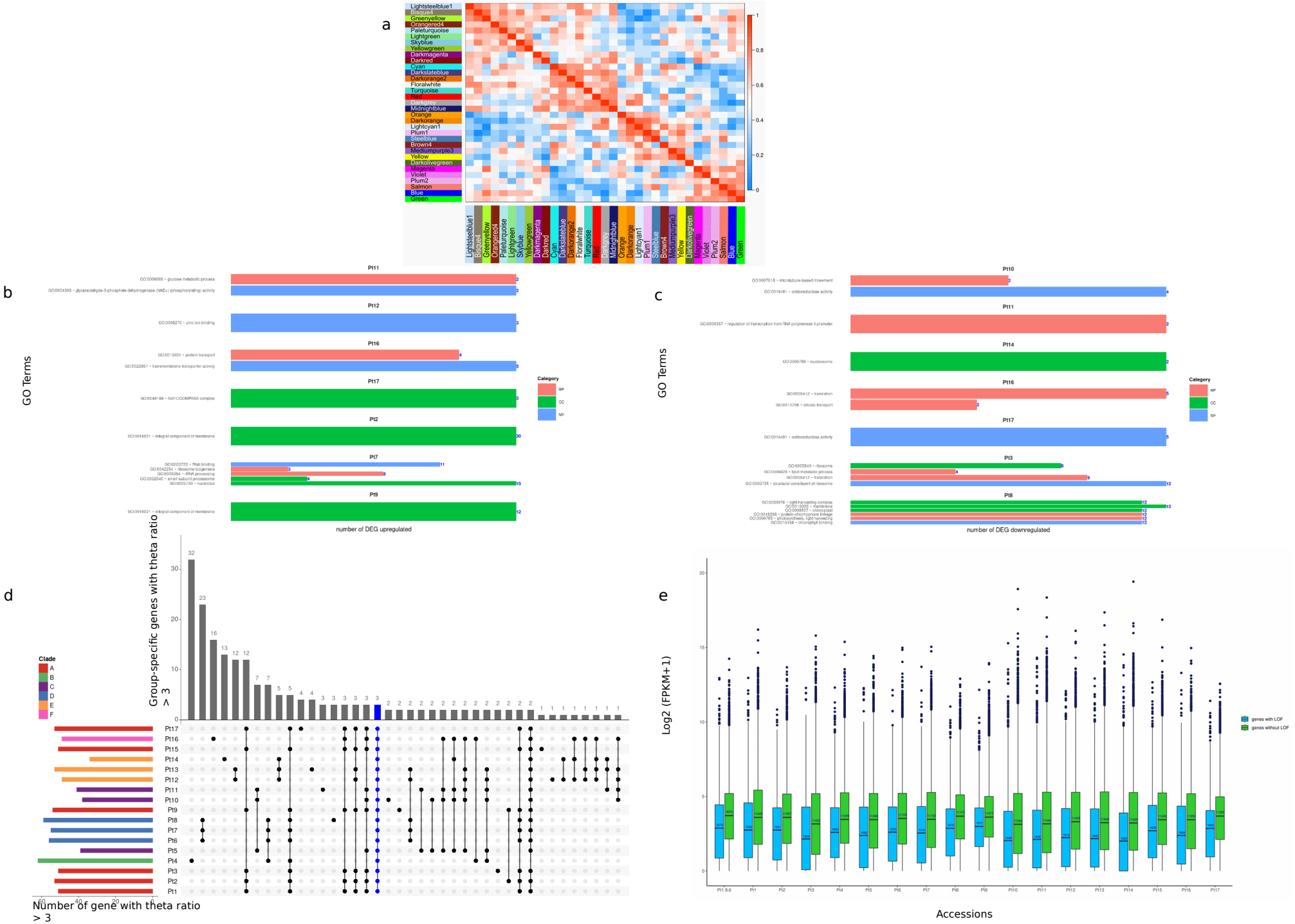

(1987)
Pt8
(1956)
Pt6
(1972)
Pt5
(1952)
Pt7
(2005)
Pt13
(1910)
Pt2
(1956)
Pt1
(2001)
Pt16
(1951)
Pt4
(1930)
Pt3
(1999)
Pt12
(1989)
Pt17
(2008)
Pt14
(2000)
Pt10
(2016)
Pt15
(2008)
Pt11
(1981)
Pt9


a
Growth rate (h-1)
16,8
16,6
16,4
16,2
16
15,8
15,6
15,4
15,2
15
Pt18.6 Pt1 Pt2 Pt3 Pt4 Pt5 Pt6 Pt7 Pt8 Pt9 Pt10 Pt11 Pt12 Pt13 Pt14 Pt15 Pt16 Pt17
Accessions
b
ETRmax
35
30
25
20
15
10
5
0
Pt18.6 Pt1 Pt2 Pt3 Pt4 Pt5 Pt6 Pt7 Pt8 Pt9 Pt10 Pt11 Pt12 Pt13 Pt14 Pt15 Pt16 Pt17
Accessions
c
Fv/Fm
0,55
0,53
0,51
0,49
0,47
0,45
0,43
0,41
0,39
0,37
0,35
Pt18.6 Pt1 Pt2 Pt3 Pt4 Pt5 Pt6 Pt7 Pt8 Pt9 Pt10 Pt11 Pt12 Pt13 Pt14 Pt15 Pt16 Pt17
Accessions
d
NPQmax-model 2021 (SV)
12
10
8
6
4
2
0
Pt18.6 Pt1 Pt2 Pt3 Pt4 Pt5 Pt6 Pt7 Pt8 Pt9 Pt10 Pt11 Pt12 Pt13 Pt14 Pt15 Pt16 Pt17
Accessions


a
HomoSNPs
HetSNPs
Number of SNPs
Accessions
b
Insertions
Deletions
Number of Insertions and Deletions
Accessions
c
Nonsense, n = 1898
Non-synonymous, n = 166877
Intergenic, n = 206536
Synonymous, n = 168969
Proportion (%)
Minor allele frequency
d
SNP INS DEL
e
Clade
A
B
C
D
E
F
Ecotype-specific CNV
Number of genes showing CNV
f
Number of LOFs
Accessions
Specific genes LOF
Total genes LOF


a
Fst value
0.6
0.5
0.4
0.3
0.2
0.1
0
Pt4 Pt14 Pt12 Pt13 Pt16 Pt5 Pt10 Pt11 Pt8 Pt6 Pt7 Pt2 Pt15 Pt1 Pt3 Pt9 Pt17
Accessions
b
Ancestry
Pt1 Pt2 Pt3 Pt9 Pt15 Pt17 Pt16 Pt4 Pt10 Pt5 Pt11 Pt6 Pt7 Pt8 Pt12 Pt13 Pt14
Accessions
c
Dim2 (20.9%)
Dim1 (27.2%)
Pt13 Pt12 Pt10 Pt11 Pt5 Pt4 Pt16 Pt2 Pt3 Pt1 Pt15 Pt9 Pt17 Pt14 Pt8 Pt7 Pt6
d
Clade
A
B
C
D
E
F
e
Pt3 Pt1 Pt2 Pt15 Pt9 Pt17
Clade A
Pt7 Pt6 Pt8
Clade D
Pt11 Pt5 Pt10
Clade C
Pt16
Clade F
Pt13 Pt12 Pt14
Clade E
Pt4
Clade B


a
Number of DEG
Accessions
All DEG
Downregulated
Upregulated
b
Number of DEG upregulated
Total of DEG upregulated
Clade
A
B
C
D
E
F
c
Number of DEG downregulated
Total of DEG downregulated
d
Dim1 (21.8%)
Dim2 (13.9%)
cos2


a
b
GO Terms
number of DEG upregulated
Category
BP
CC
MF
c
GO Terms
number of DEG downregulated
d
Group-specific genes with theta ratio > 3
Clade
A
B
C
D
E
F
Number of gene with theta ratio > 3
e
Log2 (FPKM+1)
Accessions
genes with LOF
genes without LOF

Legend

Figure 1. World map illustrating the sampling sites for the 17 *P. tricornutum* accessions analysed in the study with the year of sampling indicated in red.

Figure 2. Growth and photosynthetic features for the 17 accessions. **a** Growth rates in the 17 accessions, with error bars indicating standard deviations based on triplicate cultures. **b**. Mean maximum relative electron transport rate (rETRmax) for the 17 accessions. **c** The maximum PSII photochemical efficiency (Fv/Fm). **d** Non-photochemical quenching (NPQ) for the same 17 accessions.

Figure 3. Genetic diversity across *P. tricornutum* accessions. **a** Composition of heterozygous and homozygous SNPs, the stacked bar plot represents the number of SNPs discovered in Pt1 to Pt17 accessions, showing a contrasted proportion of homozyguous and heterozyguous SNPs. Homozyguous SNPs are displayed in light blue, heterozyguous SNPs are shown in dark blue. **b** The bar plot represents the number of insertions (red) and deletions (yellow) called in Pt1 to Pt17 accessions. **c** Folded site frequency spectrum SFS of 1898, 166877, 206536 and 168969 non-sense, non-synonymous, intergenic and synonymous SNPs, respectively. In order to obtain an unbiased SFS, only one accession was chosen to represent each group of genetically close accessions. These variants were thus called on Pt1-4-7-9-11-12-14-16-17. **d** Pie charts represent different proportion of SNPs and INDELs over all functional features of the genome; GENEs (blue), TEs (gray), IGRs (Intergenic Regions, represented in yellow). **e** The bar plots represent the total number of genes considered to exhibit CNV per accession (left, colored by clade) and those that are accession-specific (top, in grey). Only accessions having specific genes with CNV are shown in the matrix (center, in black). **f** The bar chart represents total and specific numbers of genes that are affected by loss-of-function (LoF) mutations for all ecotypes (Pt1-Pt17). The total number of genes (blue color) is the number of all non-duplicated genes on which a single variant (INDEL or SNP) was taken into account. The grey bars represent the number of genes unique to a specific accession and not present in the others.

Figure 4. Clustering of *P. tricornutum* accessions into clades. **a** The heat-map shows the genetic differentiation or association between all possible pairs of accessions. The colors indicate Fst values, which range from 0.02 to 0.4, with a color gradient from yellow to green, respectively. Values closer to 0 signify close genetic makeup and values closer to one indicate strong genetic structuring between the populations. **b** ADMIXTURE plot representing the ancestry genome fractions of Pt1 to Pt17 (in color) for K=15. **c** Principal component analysis (PCA) showing the distancebetween the seventeen accessions based on their shared genome structure. **d** The heatmap shows the log2 fold change (log2FC) of normalized reads counts between each reference gene and average of the read counts of all the reference genes per accession. FPKMs are used as normalized values, the log2 ratio of each gene FPKM over the mean FPKM of each accession being calculated. A blue to red color gradient in the heatmap represents low to high log2FC. The previously described clades A, B, C, D and the newly defined clades E and F are shown as colored annotations on the top. Genes having a null FPKM in a given accession (considered lost) are displayed in black. Only genes having a log2FC over 2 in at least one ecotype are plotted in this figure (222 genes) and are considered to exhibit Copy Number Variation (CNV). **e** Phylogenetic association of the 17 accessions based on genome-wide biallelic SNPs and Indels (640454 variants), built from a hierarchical clustering (canberra distance and average linkage functions).

Figure 5. Transcription levels variations in the 17 accessions. **a** The bar plots represent the number of differentially expressed genes (DEG) in the 17 accessions denoted on the Y axis. All DEGs are displayed in blue. Downregulated genes are shown in grey and upregulated in green. **b** The bar plots represent total number of upregulated genes in each accession (left, colored by clade) and those that are specific to a given group of accessions (top, in grey).. **c** The bar plots represent the total number of downregulated genes in each accession (left, colored by clade) and those that are specific to a given group of accessions (top, in grey). **d** Principal component analysis (PCA) showing the distancebetween the seventeen accessions based on gene expression values.

Figure 6. WGCNA and GO enrichment analysis. **a** Eigengene adjacency heatmap of the 33 merged modules of *P. tricornutum* accessions network. Each row and column in the heatmap corresponds to one module (represented by their color). The scale bar on the right represents the correlation strength ranging from 0 (blue) to 1 (red). **b** GO terms enrichment in each of Pt1, Pt7, Pt9, Pt11, Pt12, Pt16 and Pt17. In other accessions, no significant enrichment of GO terms was found. **c** The enrichment of GO terms was performed on the set of specific downregulated DEGs for each ecotype. Bar charts show which GO terms are represented and in which ecotype (Pt3, Pt8, Pt10, Pt11, Pt14, Pt16, Pt17). In other ecotypes, no significant GO enrichment terms were found. **d** The plot represents the total number of genes showing a signature for balancing selection (BS) per accessions (left, colored by clade) and those that are found only in a given group of accessions (top, in grey). Genes under BS in all accessions are shown in blue. **e** Distribution of gene expression levels in genes affected by loss-of-function mutations (blue) and unaffected genes (green) for each accession using FPKM values.

Supplementary figures

Figure S1. Light microscopy images of *P. tricornutum* accessions depicting cell morphology and size. **a** The proportions of oval morphology in Pt3. **b** The proportion of oval morphology in Pt9. **c** Cell size and morphology of Pt11 (c), Pt12 (d), Pt13 (e), Pt14 (f) Pt15 (g). **h** Cell size, morphology and proportion of tri-radiate morphology in Pt16. **i** Cell size and morphology in Pt17.

Figure S2. Gel images of PCR product illustrating the validation of INDELs observed in Pt11 to Pt17. **a** Insertion validation. **b** Deletion validation. The molecular weight marker is 1 kb plus DNA ladder (M), and the negative control is represented by (N).

Figure S3. Gel images of PCR product illustrating the validation of gene loss observed in Pt11 to Pt17. The molecular weight marker is 1 kb plus DNA ladder (M), and the negative control is represented by (N).

Figure S4. The plot shows the error of ADMIXTURE cross-validation process for K ranging from 1 to 17, from all accessions callable SNPs and INDELs. The lowest value (15) gives an indication of the ancestral populations number.

Figure S5. Inter-samples correlation heat map. The correlation coefficient is represented by the square of the Pearson correlation coefficient (R). The greater the value of R, the higher the degree of similarity between samples.

Figure S6. Heatmap visualization of gene expression and corresponding eigengenes across accessions in the 33 modules. The x-axis of both the heatmap and barplot displays the *P. tricornutum* accessions in the following order: Pt1R1, Pt2R1, Pt2R2, Pt3R1, Pt3R2 until Pt17R2. Each row of the heatmap corresponds to genes belonging to the module. Red color denotes overexpression and green underexpression.

Table S1. Sampling location, date and morphological features of the 17 accessions of *P. triorutum*

Table S2. List of primer sequences used in the study

Table S3. INDEL loci identified in Pt1 to Pt17 accessions

Table S4. Copy number variation (CNV) identified in Pt1 to Pt17 accessions

Table S5. Gene loss in Pt1 to Pt17 accessions

Table S6. Loss of function (LoF) loci in Pt1 to Pt17 accessions

Table S7. Table of the Fst distance between the different populations

Table S8. Genes under balancing selection identified in Pt1 to Pt17 accessions

Table S9. List of the novel genes identified in Pt1 to Pt17 accessions

Table S10. Repeats identified in novel transcripts

Table S11. Non coding RNA and other categories of RNA identified in novel transcripts

Table S12. List of accession specific genes up or down regulated and their GO

Table S13. List of genes up or down regulated and their GO in all accessions or per clade

Table S14. *P. tricornutum* accessions modules after merging and their annotation

Table S15. Module composition of identified clusters and their corresponding GO