New resources for environmental genomics in the wild Mediterranean grass *B. distachyon*

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ABSTRACT

Closely related to economically important crops, the grass *Brachypodium distachyon* has been originally established as a pivotal species for grass genomics but more recently flourished as a model for developmental biology. Grasses encompass more than 10,000 species and cover more than 40% of the world land area from tropical to temperate regions. Unlocking the sources of phenotypic variation in *B. distachyon* is hence of prime interest in fundamental and applied research in agronomy, ecology and evolution. We present here the *B. distachyon* diversity panel, which encompasses 332 fully sequenced accessions covering the whole species distribution from Spain to Iraq. By combining population genetics, niche modeling and landscape genomics, we suggest that *B. distachyon* recolonized Europe and the Middle East following the last glacial maximum. Consequently, the species faced new environmental conditions which led to clear associations between bioclimatic variables and genetic factors as well as footprints of positive selection in the genome. Altogether, this genomic resource offers a powerful alternative to *Arabidopsis thaliana* to investigate the genetic bases of adaptation and phenotypic plasticity in plants and more specifically in monocots.

Keywords: genetic diversity, local adaptation, landscape genomics, GEA, grass, *Brachypodium distachyon*

INTRODUCTION

In the face of accelerating climate change, understanding how plant populations adapt to new environmental conditions has sparked great interest (Lasky et al. 2023). Environmental constraints varying in space and time alter the local frequencies of adapted genotypes, leaving detectable signatures at selected genes. Identifying these signatures is valuable for understanding the processes underlying adaptation, and for providing candidate genes for functional studies. In this context, advances in genomics and DNA sequencing technology have revolutionized the field of landscape genomics by enabling high-resolution genome-wide analyses (for review Bourgeois and Warren 2021). For instance, genotype-environment association analyses (GEA) are powerful tools to identify alleles associated with ecologically relevant factors (Rellstab et al. 2015; Lasky et al. 2023). On the other hand, genome-wide scans of selection (Nielsen et al. 2005; Tang et al. 2007; Gautier 2015) have been largely used to identify genetic factors under selection without a prior neither on the phenotype under selection nor on the selective constraints acting on it. Those approaches, however, all rely on one fundamental aspect: the availability of genomic resources for many accessions occurring in contrasting habitats.

Despite the pressing need for new model species (Marks et al. 2023), research on local adaptation in plants is still dominated by *A. thaliana* (Provart et al. 2016; Woodward and Bartel 2018). The *A. thaliana* 1001 genomes project, combined with decades of functional studies, has indeed produced an unmatched wealth of knowledge about the processes of adaptation and evolution in plants (e.g. Hancock et al. 2011; Horton et al. 2012; Durvasula et al. 2017; Lee et al. 2017; Wu et al. 2017; Fulgione et al. 2018; Exposito-Alonso et al. 2019; Takou et al. 2019; Exposito-Alonso 2020; Wieters et al. 2021; Monroe et al. 2022). Even though the development of pan-genomes for crops (Montenegro et al. 2017; Wang et al. 2018; Haberer et al. 2020; Jayakodi et al. 2020; Walkowiak et al. 2020; Lovell et al. 2021) will undoubtedly help to understand stress resilience in plants of agronomical interest, they nonetheless remain of limited value to tackle the diversity of paths to adaptation found in natural systems. Developing alternative wild systems, especially in grasses which play a key role in ecosystem functioning (Groves 2000), is thus very timely.

Initially established as a model for crop genomics (International Brachypodium Initiative 2010), the grass species *Brachypodium distachyon* is now a pivotal system for developmental

biology (Hasterok et al. 2022; Raissig and Woods 2022) including the study of flowering time (Ream et al. 2014; Woods et al. 2014; Sharma et al. 2017; Woods et al. 2020; Bouché et al. 2022), stomata traits (McKown and Bergmann 2020; Nunes et al. 2020; Zhang et al. 2022; Slawinska et al. 2023) or cell-wall development (Coomey et al. 2020). In addition to harbouring a large mutant collection (Dalmais et al. 2013) and modern tools for mutagenesis (Hus et al. 2020), *B. distachyon* is a wild and widespread species with a small diploid genome (272 Mb). As such, it features functional and population genomics resources that are not combined in any other grass system for which a germplasm diversity panel has been established, such as rice or switchgrass (Wang et al. 2018; Lovell et al. 2021).

   B. distachyon occurs naturally in oligotrophic habitats around the Mediterranean rim (López-Alvarez et al. 2015; Catalán et al. 2016). The combination of a geographic mosaic and climatic stability allowed Mediterranean species to diversify at regional and local scales (Nieto Feliner 2014). In this context *B. distachyon* constitutes an excellent system to investigate the genetic bases of local adaptation in a species adapted to arid climates. Earlier works on this topic mostly focused on a small set of accessions originating from Turkey and Spain (Del'Acqua et al. 2014; Des Marais et al. 2017; Bourgeois et al. 2018, Wyler et al. 2018; Stritt et al. 2018; Skalska et al. 2020) and the genetic basis of trait variation is still largely characterized through QTL mapping and mutant screening in this species (e.g. Barbieri et al. 2012; Woods et al. 2017; 2020; Jiang et al. 2017). Yet, genome-wide sequencing data have been produced for about 260 accessions originating from Spain, France, Italy, Turkey, Lesser Caucasus and Iraq (Gordon et al. 2017; 2020; Skalska et al. 2020; Stritt et al. 2022). Those resources, however, were never analyzed at once (but see Stritt et al. (2022) for the analysis of 196 accessions) despite their great potential to unlock the source of genetic and phenotypic diversity in this species. In the current study, we filled up a last geographical gap by collecting and sequencing an additional set of *B. distachyon* accessions from Greece and Montenegro. We combined all available genomic data into a diversity panel encompassing 332 accessions and asked: i) What is the recent demographic history of *B. distachyon* populations ii) To what extent have environmental factors shaped genetic diversity in this species and, iii) Which genes have been selected by the environment. This resource is made available to the community to stimulate research and facilitate comparisons across established and emerging model species for plant adaptation.

RESULTS AND DISCUSSION

Population structure and demographic history of *B. distachyon*

We used publicly available sequencing data from 196 natural *B. distachyon* accessions originating from Spain, France, Italy, Turkey, and Iraq (Gordon et al. 2017; Skalska et al. 2020; Stritt et al. 2022) previously analyzed in Stritt et al. (2022). We also added 65 additional sequencing data produced by Gordon et al. (2020) for accessions from Spain, Turkey and the Lesser Caucasus. Finally, we collected and sequenced an additional set of 71 accessions from Montenegro, Greece and France. Altogether, we assembled a diversity panel that encompasses 332 accessions (Fig. 1a, Table S1) for which whole-genome sequencing data are available with a minimum coverage of 20X.

B. distachyon belongs to the Brachypodium species complex. The three species in this complex, *B. distachyon*, *B. stacei* and the allopolyploid *B. hybridum,* can be difficult to distinguish morphologically but are straightforward to identify through genotyping (Giraldo et al. 2012; Catalán et al. 2016). Following extensive fieldwork, all accessions collected in the past in Morocco, Afghanistan, Iran, Pakistan, Australia and USA have been genotyped as *B. hybridum* (Wilson et al. 2019; Stritt et al. 2022; Fig. S1 for the geographical distribution of the 2420 genotyped accessions). In contrast to *B. hybridum* and *B. stacei, B. distachyon* is extremely rare in Israel (Wilson et al. 2019) and in France only recorded in the South of the country (https://www.tela-botanica.org/bdtfx-nn-10075-synthese). The 332 *B. distachyon* accessions currently at hand (Fig. 1a, Table S1) are thus likely to cover the species range comprehensively.

We identified 10,227,760 high-confidence single nucleotide polymorphisms (SNPs) in the diversity panel and applied different filtering criteria according to the requirement of each analysis in the following sections, as described in Materials and Methods. While we merged datasets from different studies, a hierarchical clustering based on 75,000 random SNPs shows that samples do not cluster according to the study of origin (Fig. S2), indicating no large technical biases. Based on 196 samples, Stritt et al. (2022) found that *B. distachyon* accessions cluster in three main genetic lineages (A, B, C) that further split into five clades (A_East, A_Italia, B_East, B_West and C). Our current analysis based on 332 accessions did not reveal any additional discrete genetic clusters (Fig. 1a, b, c), even after expanding the sampling range. Indeed, the principal component analysis (PCA, Fig. 1b), sNMF (Fig. 1c and Fig. S3a,b)

and the Evanno method (Evanno et al. 2005; Fig. S3c) all suggest that our accessions group

into five main genetic clades.

Accessions from Spain and France share common ancestry and cluster together in the

B_West clade (n = 72). One accession from France (MonSV5), however, shares common

ancestry with the main Italian clade (A_Italia) while four accessions from Spain have a mixed

ancestry and cannot be assigned to any of the clades (Spa-Sou-GR6_4, Spa-Nor-S6D, Mon3,

Arn1). Accessions from eastern regions belong to two clades, one comprising Balkan and

Figure 1 - Sample distribution and population structure a) the map displays the origin of the 332 accessions used in the study. The tree represents the phylogeny of the five genetic clades and indicates the divergence estimates. The color code will apply to the rest of the study b) PCA based on 16.381 independent SNPs c) Structure plot displaying the shared ancestry among accessions. The black arrows point at the four admixed accessions found in Spain d) Population size evolution over time computed for the four derived genetic clades e) Tajima's D and pi per genetic clade computed in 5kb windows over the entire genome.

coastal Turkish individuals (A_East, n = 94), and the other including mainland Turkish and

Anatolian individuals (B_East, n = 73). In Italy, two genetic clades, the A_Italia (n = 66) and C,

co-occur. We also observed that all the accessions from Montenegro share ancestry with the

rest of the C clade, as does one individual from Greece (Meg7; Fig. 1a-c). Altogether, the C

clade comprises 27 accessions. Eleven accessions from the Lesser Caucasus have been

identified as belonging to two different genetic clusters, with three accessions belonging to

the A_East cluster and eight accessions belonging to the B_East cluster. Finally, three

accessions from coastal Turkey (2_14_15, 2_14_20, 2_20_16) clustered with the B_West, as

already reported in previous studies (Skalska et al. 2020; Stritt et al. 2022).

Using a multispecies coalescent approach, we estimated that the split between the

ancestral C lineage and the A/B lineages occurred 102 thousand years ago (kya, 90% highest

posterior density interval [HPDI] 50–170 ky), the split between the A and B lineages 45 kya

(90% HPDI 21–76 ky) and the split within the A and B lineages 13 kya (90% HPDI 5-23 ky) and

23 kya (90% HPDI 11-39ky) respectively (Fig. 1a). These estimates are in the same order of

magnitude as the ones we obtained with Relate, a method used for estimating genealogies

genome-wide. For this analysis, the ancestral C clade was used to polarize SNPs. Relative

cross-coalescence rates (CCR) between the remaining four genetic clades indicate that the

split (CCR < 0.5) between the A and B lineages occurred within the last 100,000 years ago

while the splits (CCR < 0.5) within the A and B lineages occurred within the last 5 kya (Fig. S4).

We also used Relate to compute effective population size (N_e) evolution over time. This

analysis revealed an abrupt decline in N_e across all clades around 30 kya, all followed by

population expansion in a very recent past (Fig. 1d). Congruent with the recent population

decline and expansion observed for each genetic clade, we found overall negative Tajima's D

values but still substantial remaining genetic diversity (pi) for all genetic clades (Fig. 1e).

Nonetheless, the B_East clade displays higher Tajima's D and genetic diversity levels than the

other genetic clades (Kruskall-Wallis test, all p-values < 5.2e-05) which could be explained by

its more stable effective population size before the most recent drop/expansion ca. 5 kya.

Note that all our dating methods rely on the use of a molecular clock and must thus be

interpreted with some caution. Our results nonetheless indicate major changes in the recent

demographic history of *B. distachyon.* Because these demographic changes accelerated

within the last 10,000 years, we speculated that the species experienced a shift of its

distribution following the Last Glacial Maximum (LGM, 22 kya) and more specifically during

the Holocene period (11.7 kya) which marks the beginning of deglaciation in Europe.

Ecological niche modeling and distribution of *B. distachyon* during the last glacial period

The climate, vegetation and landscape of Northern Eurasia (north of ca. 40°N and from 10°W

to 180°E) underwent massive changes during the last glacial period (Binney et al. 2017; Davis

et al. 2022). Under the cooler and dryer conditions faced during the LGM specifically, forests

retreated to glacial refugia in Spain, Italy or the Balkans (for review Feliner 2011; Nieto Feliner

2014) and land remained mostly covered by steppe and tundra (Binney et al. 2017). Following

the LGM, deglaciation led to the recolonization of Eurasia by woody plants and forests (Binney

et al. 2017) and hence to substantial changes in dominant biomes. These recent climatic

events have shaped the biogeography and genetic diversity of plants at large (Feliner 2011).

In *A. thaliana*, for instance, relict populations occupied post-glacial Eurasia first and were later

replaced by non-relict populations whose range expansions, accompanied by admixture,

largely shaped modern populations (Lee et al. 2017).

To test for a potential shift in the distribution of *B. distachyon* in the recent past, we

performed niche modeling analyses using Maxent climate niche models (Phillips et al. 2006).

Due to the high correlation among bioclimatic variables for current time (Fig. S5a), we

proceeded with a strategy based on selecting variables using an *a priori* understanding

(Burnham and Anderson, 2004) of the *B. distachyon* life cycle: minimum temperature

averaged from November to February (hereafter *tmin_Nov-Feb*) was chosen as potentially

important for the vernalization process; precipitation levels averaged from March to June

(hereafter *prec_March-June*), solar radiation levels averaged from March to June

(*srad_March-June*) and elevation were chosen as relevant for plant phenology during the

growing season.

We first fitted consecutive models for the whole species distribution with these four

variables under current environmental conditions (see Materials and Methods), before

projecting them over past conditions. Models with combinations of two variables had the

lowest AIC values generally, however, only the model using *prec_March-June* and *tmin_Nov-*

Feb showed Area under the Curve (AUC) values and predictive ability better than null models

(p-value < 0.05) and was consequently chosen as our final model (see Fig. S5b for the
distribution of these variables across genetic clades).

Based on this model and using paleoclimatic datasets, we projected the potential whole-
species distribution in Europe, North Africa, and the Middle East since the Last Glacial
Maximum (ca. 22 kya), as well as for the five genetic clades (Fig. S6). In contrast to the niche
modeling performed with the geographically restricted current bioclimatic variables, we
extended the analysis to a much wider geographic area in order to be able to detect putative
changes in the species distribution. Our results suggest that climatic conditions similar to
currently suitable conditions for *B. distachyon* extended southwards under LGM conditions
with Northern Africa, Levant countries as well as the Iberic and Arabian Peninsula providing
the most suitable habitats for the species (Fig. S6). During the LGM, the sea level was about
120–150 meters lower than at present (Yokoyama et al. 2000), which may have facilitated
migration between Southern Europe and North Africa (Ortiz et al. 2007). Furthermore, Levant
countries were mostly dry to subhumid habitats (Jennings et al. 2015). Hence, their
geographical proximity with modern accessions and habitat suitability makes Northern Africa,
Levant countries and the Iberic peninsula good candidates for *B. distachyon* glacial refugia.
While the high suitability in the Arabian Peninsula might be seen as an overfit of our models
at first, it is worth noting that annual rainfall levels under LGM conditions were much more
important than nowadays in this region, making part of the Arabian Peninsula a dry to
subhumid habitat (Jennings et al. 2015) likely suitable for *B. distachyon*.

We did not include information about soil as it is not projected for past conditions. For an
oligotroph species as *B. distachyon*, the fundamental niche we computed might therefore
differ from the realized one, and without modern samples available for all these four putative
glacial refugia one can only speculate about a migration scenario (Lee et al. 2017). Yet, a
natural conclusion is that *B. distachyon*, like most plants, operated a shift during the last
glaciation and recolonized Europe and the Middle East following deglaciation. Our results
contrast to some extent with those of López-Alvarez et al. (2015), who found that the
distribution of *B. distachyon* extended southwards without a complete shift. This early study
was based on the only samples collected at that time in Spain and Turkey. As it failed to
recover the presence of the species in Italy and the Balkans under current environmental

conditions, we argue that the projection to past conditions might be less reliable than the one
we presented here based on a more comprehensive sampling of the species.

The split among the three main genetic lineages of *B. distachyon* predates the LGM (Fig. 1a,
Fig. S4) and it is therefore likely that accessions from the A_East/A_Italia, B_East/B_West and
C genetic clades experienced different demographic histories. The geographic distribution of
the five genetic clades is more parsimoniously explained by independent expansions and is
sustained by the rapid increase of the effective population size for each clade within the last
5 ky (Fig. 1d). It has already been proposed that lineages and genetic clades may have used
different migration corridors to recolonize Europe and the Middle East (Stritt et al. 2022). An
East–West phylogeographical break, as often reported in Mediterranean species (Nieto
Feliner 2014), could explain that accessions from the B_West and B_East genetic clades group
together phylogenetically despite the current geographic gap (Fig. 1a). This scenario further
supports a North African corridor for accessions from the B lineage, as already speculated in
Stritt et al. (2022). Such a demographic scenario has been demonstrated in the herbaceous
perennial species *Erophaca baetica* for instance, where plants from the Iberic peninsula are
clearly derived from North African populations while geographically disconnected from the
Greek and Turkish ones (Casimiro-Soriguer et al. 2010; Nieto Feliner 2014). In contrast, we
previously observed a South-to-North gradient of declining N_e, genetic diversity, and shared
ancestry for A_Italia clade, which suggests a northwards expansion of this clade (Stritt et al.
2022).

Genetic clades occupy different ecological niches and display adaptive loci
The Mediterranean basin comprises a mosaic of habitats (Nieto Feliner 2014) and consistent
with recolonizations through different routes, the five genetic clades of *B. distachyon* occupy
nowadays different geographical areas and ecological niches (Fig. 2). To detect significant
differences among the ecological niche models (ENMs) of the five clades under current
conditions, we created pseudoreplicate niche comparisons and compared them with the
niche similarity score produced using the empirical data of the realized distribution (Fig. 2a-
e). Note that the three accessions from Turkey clustering with the B_West clade (2_14_15,
2_14_20, 2_20_16, Fig. 1a) were excluded from the analysis. According to our models, there
was no overlap between the niche suitability among the five clades. For the A_Italia vs C,

B_West vs A_East, B_West vs A_Italia comparisons, p-values ranged from 0.01 and 0.05. For

all the other comparisons, p-values were <0.01. We however acknowledge that the ENM of

the C clade must be interpreted cautiously due to the small sample size.

More specifically, the higher predicted niche suitability for the A_East clade was at the

coasts of Turkey, in mainland Greece, Italy, southern France, and parts of the Iberian

Peninsula (Fig. 2a). The A_Italia clade displays a high suitability in Italy, Iraq, the northwester

Iberian Peninsula as well as on the east coast of the Adriatic Sea (Fig. 2b). For the B_East clade,

the areas with the highest suitability scores were mainland and eastern Turkey, northern

Greece, coastal Bulgaria, as well as a part of central Spain (Fig. 2c), while the B_West clade

displayed higher suitability in northern Spain, southern France, parts of Italy and Greece, as

well as northwestern Turkey (Fig. 2d). Finally, the C clade shows high niche suitability in

southern Italy, the southern Iberian Peninsula, the Greek Aegean islands, and Iraq (Fig. 2e).

Figure 2 - Environmental Niche Modeling for current conditions using the masking region. The maps display environmental suitability for the five genetic clades a) A_East b) A_Italia c) B_East d) B_West e) C and f) at the species level.

264   We have already shown that adaptation at a regional scale led to specific footprints of

265   positive selection in the genome of accessions from the B_East and B_West genetic clades

266   (Bourgeois et al. 2018), but this previous analysis was somewhat limited by the number of

267   samples then sequenced (27 and 17 accessions respectively). As we found that *B. distachyon*

268   genetic clades now occupy different niches, we extended this initial study by testing for

genome-wide footprints of positive selection at a regional level, using the genetic clades as focal populations. To do so, we calculated the X^TX statistic, a measure comparable to single SNP Fst that accounts for the neutral genetic covariance across populations (Günther and Coop 2013) and detects highly differentiated alleles across several populations at once without SNP polarization into ancestral or derived alleles (Gautier 2015b). By selecting the top 0.1% X^TX outliers (Fig. 3a), we identified 1477 highly differentiated genes putatively under positive selection (Fig. 3a and c). The GO annotation of the X^TX gene set (Table S3) revealed a significant over-representation of genes involved in response to chemical (p-value = 2.01E-4) and metabolic process (p-value = 1.72E-04) including glutathione metabolic process (3.99E-06), a metabolic process involved in the control of reactive oxygen species (ROS) and hence stress response (Mittler et al. 2022) .

However, local adaptation is likely to occur at a much finer geographical scale (Gloss et al. 2022) and populations may each carry adaptations to their local climates (Lasky et al. 2023). Under the hypothesis that distinct lineages adapt to environmental gradients using the same traits (Lasky et al. 2023) through balancing selection, GEAs are powerful bottom-up tools to characterize the genetic basis of adaptation and identify which selective constraints might have shaped genetic diversity in a species. After excluding alleles displaying a minor allele frequency < 0.05, we tested for significant associations between the 2,867,335 remaining SNPs and 32 environmental variables related to precipitation levels, temperature, aridity index, solar radiation or elevation (Fig. 3b, Fig. S5a) with GEMMA while correcting for population structure (Zhou and Stephens 2012). We considered 5 kb windows significantly associated with a given environmental variable when they display at least two SNPs above the false discovery rate threshold (See Materials and Methods).

For each bioclimatic variable, we extracted genes located in significantly associated regions. Out of the 32,432 total genes already annotated in *B. distachyon* genome, 2,379 are significantly associated to at least one of the environmental variables (Fig. 3c, Fig. S7, Table S2). Annual mean precipitation (bio12, Fig. 3a), together with bioclimatic variables associated to conditions from March to June (precipitation or aridity), yielded the highest number of genes (Fig. 3c). On the other hand, only 32 and 21 genes are located in regions associated with elevation or annual mean temperature (bio1) respectively. Interestingly, even though the 32 bioclimatic variables we chose show some levels of correlation (Fig. 3b, Fig. S5a), we

only observed a partial overlap among the associated gene sets. For instance, while we identified 313 genes associated with precipitation levels in spring (*prec_March_June*) and 145 genes associated with precipitation levels in winter (*prec_Nov_Feb*), those two gene sets only share 113 genes indicating that genes playing a role in adaptation to climate may also be season-specific. The Gene Ontology (GO) annotation performed with the 2,379 genes associated with at least one bioclimatic variable (Table S3), however, did not reveal any significant process or molecular function.

We found little overlap between the GEAs and the scan of positive selection (Fig. 3c). This was expected given that we use the five genetic clades as focal populations for the X^TX analysis

Figure 3 - Genotype-environment association and X^TX analyses a) Manhattan plots displaying regions under positive selections (the dotted line indicates the 0.1% outlier threshold) and the association between genomic regions and annual precipitation levels bio12 (the dotted line indicates the FDR threshold) b) Correlations among top 7 bioclimatic variables (associated with more than 100 genes) c) Upset plot displaying the overlaps among gene sets associated with bioclimatic variables or positive selection. The top 7 variables are presented. Set size shows the number of genes in significant regions of each specific variable. Intersection size shows the number of candidate genes associated with a variable (single dot) or shared among variables (multiple dots linked).

and therefore putatively detect loci influencing adaptation at a larger geographical scale than

with the GEAs. We also estimated the average age of alleles per gene with GEVA (Albers and

McVean 2020). To avoid spurious age estimates, we only kept the 29,761 genes which harbor

at least five SNPs (referred hereafter as genome-wide level). We used the gene lists produced

by the GEA and the X^TX analyses only if they contained at least fifteen genes. Even though

statistical differences were observed among GEA and X^TX gene sets, age estimates indicate

that the large majority of alleles potentially involved in adaptation emerged 40 to 20 kya (Fig.

S7b) and are slightly older than the recolonization of Europe and the Middle East by *B.*

distachyon. In contrast to positive selection on *de novo* mutations (Barrett & Schluter, 2008),

selection from standing variation is predicted to promote faster evolution (Hermisson and

Pennings 2005) and this latter result suggests that the recycling of older alleles may have

played an important role in local adaptation in our system.

CURRENT LIMITATIONS AND PERSPECTIVES

We have not considered here that plants may also adapt along gradients via distinct strategies

and traits, as shown in *A. thaliana*, where genetic factors causing local adaptation to elevation

vary across regions and populations (Yan et al. 2021; Gamba et al. 2022). Because we filtered

SNPs with more than two alleles, we may have generally excluded such genetic factors from

our analyses, and the subsequent GEAs might thus largely underestimate the number of

adaptive genes in our system. This might explain why we found so few genes associated with

elevation, while altitude is well known to influence key traits such as plant size (Moles et al.

2009), flowering time (Kooyers et al. 2015; Vidigal et al. 2016; Wadgymar et al. 2018) or

freezing tolerance (Zhen and Ungerer 2008). As we also observed substructure within genetic

clades (Fig. S3), within-genetic clades GEAs combined with common garden experiments

might be more pertinent to identify population-specific loci. Furthermore, most traits exhibit

a polygenic architecture, and detecting a large number of variants with subtle effects (for

review Yeaman 2022) can be challenging with classical GEA or genome-wide association

analyses (de Miguel et al. 2022). Hence, many crucial questions remain open to understand

the genetic architecture as well as the geographical scale of adaptation in *B. distachyon.* With

this diversity panel, the plant community is provided with a new means to unlock the source

of natural variation in a monocot adapted to arid climate, beyond what previous pan-genome

and QTL mapping studies permitted in this system.

MATERIALS AND METHODS

Sampling and genotyping

Seeds from 299 *Brachypodium* plants were collected in 2018 and 2019 from southern France,

Greece, and Montenegro. A representative subset of 110 plants was grown from seeds in

greenhouse conditions, and DNA was extracted from their leaves with a DNeasy plant kit from

Qiagen, following the manufacturer instructions. To confirm that only *B. distachyon*

individuals were selected for whole genome sequencing, plants were genotyped using the

microsatellite marker ALB165 (Giraldo et al. 2012) and by Sanger sequencing the GIGANTEA

gene (López-Alvarez et al. 2012). Out of the 88 accessions identified as *B. distachyon*, 71 were

selected for sequencing, while the remaining accessions were excluded from the rest of the

analysis. The libraries were prepared and sequenced using Illumina HiSeq2500 (150 bp PE) by

Novogene. Information about additional accessions collected and genotyped by genotyping-

by-sequencing (n = 1897) were obtained from Wilson et al. (2019) and displayed on a map

with QGIS (version 3.4.13) together with all the *B. distachyon* used in this study.

Sequencing, SNP calling, and filtering

Paired-end reads from the 71 newly sequenced accessions, together with the 65 *B. distachyon*

accessions sequenced by Gordon et al. (2020), were aligned to version 3.0 of the *B. distachyon*

Bd21 reference genome (https://phytozome-next.jgi.doe.gov) with bwa mem version 0.7.17-

r1188 (Li 2013). Aligned reads were converted to bam files and sorted with samtools version

1.7 (Li et al. 2009), while accessions with multiple lanes were merged using the same program.

Read duplicates were then removed with PICARD MarkDuplicates version 2.23.3, and variants

were called with GATK v. 4.1.2.0 (McKenna et al. 2010). The resulting gvcf files were combined

with the 196 gvcf files from the study of Stritt et al. (2022), leading to a dataset of 332

accessions. Subsequently, only SNPs with quality-by-depth higher than 8 and Phred-score

more than 20 were kept using VCFTOOLS version 0.1.15 (Danecek et al. 2011).

Population genetic structure and demographic history

The genetic structure of the samples was estimated by using principal component analysis and admixture analysis. To obtain independent SNPs, they were filtered for minor allele frequency > 0.05 and linkage disequilibrium < 0.4 using the function snpgdsLDpruning of the R package SNPRelate (Zheng et al. 2012). The principal components were calculated using the resulting 15,960 independent SNPs and the function snpgdsPCA of SNPRelate. On the same dataset, ancestry coefficients were estimated using the function sNMF of the R package LEA (Frichot and François 2015). Results for 2-10 K values for 20 replicates per K were retained, and the optimal K value was determined by calculating *ΔK* as described by (Evanno et al. 2005). The hierarchical cluster analysis was eventually performed using a set of 75,000 random SNPs with the R package pvclust version 2.2-0 (Suzuki and Shimodaira 2006).

We obtained population size history using Relate v1.1.7 (Speidel et al. 2019) while filtering for FS > 60, SOR > 3, MQ > 40, -5.0 < MQRankSum < 5.0, QD < 2, ReadPosRankSum < -4.0, INFO/DP > 16,000. We also excluded genotypes with a genotype quality GQ < 20, an individual depth DP < 8x, and excluded sites with more than 20% missing genotypes, but importantly did not filter for a maf. This resulted in 6,728,435 SNPs. Relate also requires alleles to be polarized into ancestral and derived alleles. To do so, we used the genotypes from the ancestral C clade (Stritt et al. 2022), assuming the most frequent allele in that group to be ancestral. We then assigned each allele in the four other clades as ancestral or derived. We assume that individuals are strongly inbred, each owning two copies of the same haplotype. We therefore kept only one haplotype per individual. We used the Relate package (Speidel et al. 2019) to infer genome-wide genealogies and coalescence rates across all individuals. To take into account local variation in recombination rates, we used a recombination map for *B. distachyon* obtained from Huo et al. (2011). To obtain demographic estimates of times and population sizes, we used a mutation rate of 7×10^{-9} substitutions/generation (Lynch et al. 2016) and a generation time of one year as *B. distachyon* is annual. We set the prior for the effective haploid population size to 75,000 individuals based on previous estimates (Stritt et al. 2022). We subsequently fitted a time-varying population size history and inferred genome-wide topologies using the EstimatePopulationSize.sh script provided with the Relate package.

For each annotated gene (https://phytozome-next.jgi.doe.gov), the nucleotide diversity (pi) was calculated using pixy version 1.2.7.beta1 in 5 kb windows (Korunes and Samuk 2021).

401 Tajima's D was derived from these pi values in R using theta(pi) and the number of segregating

402 sites.

Estimating divergence times

We used the multispecies coalescent approach implemented in bpp v.4.2.9 (Rannala and Yang 2003; Flouri et al. 2018) to estimate the age of the splits among genetic lineages and clades, as also described in Stritt et al. (2022). 28 accessions (Cm4, Ren22, Msa27, Lb13, BdTR7a, 2_14_13, 4_52_6, 1c_25_14, Luc1, ABR6, Tso18, Bd30-1, BdTR2B, BdTR3C, 1a_32_12, Bd21, Mca12, Cro24, Cb23, San12, Arm-Arm-2B, Geo-G30i2, Geo-G31i4, Alb-AL1A, Ko2, MonSV13, Myt1, Vyt1) were selected to represent the five genetic clades. 200 random genomic regions of 1 kb length and at least 100 kb apart were chosen. For each of the 28 accessions, the 200 sequences were obtained by calling consensus sequences from the respective bam file. Inverse gamma priors were set to (3, 0.014) for the root age τ and to (3, 0.002) for the population size parameter θ, which corresponds to a mean theta of 0.001 and a mean root age of one million years, assuming a constant mutation rate of 7×10^{-9} substitutions per site per generation. The rooted species tree was defined as (((A_East, A_Italia), (B_East, B_West)), C_Italia), as inferred by Stritt et al. (2022). The MCMC was run four times independently, each time with 408,000 iterations, including a burn-in of 8,000 iterations. Highest posterior density intervals (HPDI) were estimated with the R package HDInterval version 0.2.4 (Meredith and Kruschke 2022). Relative cross-coalescence extracted from the output of Relate were also used to estimate divergence times, with relative cross-coalescence < 0.5 considered as populations being fully separated.

Ecological niche modeling

Raster maps were downloaded for all bioclimatic variables from WorldClim 2.1 (Fick and Hijmans 2017) at a 30 sec resolution (about one kilometer). Global aridity index raster maps were obtained from https://cgiarcsi.community/data/global-aridity-and-pet-database/. We used the geolocations of the 332 accessions to extract bioclimatic variables at each locality with the R package raster 3.5-29. The 19 classical WorldClim variables (bio1 to bio19) and elevation were used unmodified. New variables were created from monthly values for precipitation, solar radiation, average temperature, maximum temperature, minimum

temperature, and aridity, by calculating the averages of four months of putative biological

interest (March to June, and November to February) (Table S1). Elevation data were

downloaded using Google Earth. Correlations among bioclimatic variables were plotted as a

bubble plot with the R package corrplot Version 0.92 (Wei and Simko 2021).

We used ecological niche models (ENMs) to estimate the climatic niches of the five

genomic clades of *B. distachyon* and to investigate the degree of climatic niche overlap in

both current and past conditions. As inputs for the models, we used the geographical

coordinates of sequenced individuals and a set of environmental variables that were selected

based both on biological relevance and on minimum collinearity between them: minimum

temperature averaged from November to February, precipitation levels averaged from March

to June, solar radiation levels averaged from March to June and elevation. The ENMs were

produced with the MaxEnt software version 3.4.4 (Phillips et al. 2006), which uses machine

learning to predict potential geographic distributions of species across different landscapes

(Elith et al. 2006). The strength of association between the current distribution of different

genomic clades and present climate variables was tested following the methods used by

Williams et al. (2015) and Skalska et al. (2020). In brief, ENMs were considered to have

identified relationships between the distribution of *B. distachyon* and the environmental

predictors that were stronger than expected by chance given the spatial patterns in the data

if the Area under the Curve (AUC) values of the real model were in the top five of the 100

models (99 null and 1 real), as described by Beale et al. (2008).

The ENMs were visualized using QGIS version 3.4.13-Madeira and maps were created to

show the predicted suitable habitats for each clade. ENM modeling was restricted to the

relevant study area by using a prepared mask (seen in Fig. 2) which was informed by very

broad estimates of the extent of the sampling efforts (Fig. S1) and field observations. This

included mainland Spain, Portugal, Turkey, Greece, Slovenia, Croatia, Bosnia, Montenegro,

Albania, North Macedonia, Bulgaria, Georgia, Armenia and Azerbaijan. In France, *B.*

distachyon is only found in the South (https://www.tela-botanica.org/bdtfx-nn-10075-

synthese) and we only included the Occitanie and Provence-Alpes-Côte d'Azur administrative

regions within the mask, rather than the whole country. For the same reason, we only

included the Italian administrative regions to the south of Torino, Milan and the west of the

Po valley. Sardinia, Corsica and the Balearic Islands were excluded. The dataset also included

three sample points in Northern Iran and Iraq. No suitable administrative region could be
found to effectively delineate these areas. We therefore created a buffer (1 decimal degree
radians) around the sampling points.

To study past clade distributions, we projected current climatic niches onto paleoclimate
maps using variables with a resolution of 2.5 arc-min for the Last Glacial Maximum (LGM)
obtained from WorldClim1.4 (Fick and Hijmans 2017). The atmospheric general circulation
models used in this study are the Community Climate System Model version 4 (CCSM4) (Gent
et al. 2011). Past Maxent model projections were 'clamped' which restricts them to the range
of climatic conditions that are seen in the current climate conditions (where the model was
trained). This is recommended to avoid spurious model extrapolations when projecting into
novel environments (Elith et al. 2010). Clamped areas can be seen coloured grey in Fig. S6.

The program ENMTools version 1.3 (Warren et al. 2010; Warren et al. 2021) was used to
test whether there are statistically significant ecological differences between the clades,
based on the ENMs produced by Maxent. Niche similarity was first calculated between each
clade combination for the empirical data. 99 pseudoreplicate niche comparisons between
each clade were then produced by pooled and randomized occurrence points of the empirical
data. The overlap between ENMs generated from the empirical data for each clade
comparison was then compared to the null distribution obtained using the pseudoreplicates,
using Schoener's D and the I statistic (Warren et al. 2010). We concluded that ENMs produced
by two clades are significantly different if the empirical data had values smaller than the 95
of the pseudoreplicate values.

Genotype-environment association studies
We used GEMMA 0.98.5 (Zhou and Stephens 2012) to fit a linear mixed model and test for
association between SNPs and the 32 bioclimatic variables described above, while correcting
for population structure. A centered relatedness matrix was first produced with the option -
gk 1. Association tests were performed using the option -maf 0.05 to exclude SNPs with minor
allele frequency with values less than 0.05, as power is lacking for detecting associations using
rare alleles (Marees et al. 2018).

We applied a False Discovery Rate (FDR, Benjamin and Hochberg 1995) threshold to control
for the expected false positives rates among the rejected null hypotheses. To further reduce

false positives, a sliding window approach was implemented across the whole genome, with window size of 5 kb and 2.5 kb overlap, using the R package rehh version 3.2.2 (Gautier et al. 2012). The selected windows contained at least two SNPs above the FDR threshold. We then used the BEDTOOLS version 2.26.0 intersect option (Quinlan and Hall 2010) to extract genes located in or overlapping with significant regions based on the v.3.2 annotation of the *B. distachyon* (https://phytozome-next.jgi.doe.gov).

Scans of selection

The $X^T X$ analysis was performed with BayPass using the five genetic clades as focal populations (Gautier 2015). We generated the input file by using vcftools --count to calculate the allele frequency of each SNP present in our vcf (no filtering on minimum allele frequency). We then ran Baypass on our actual dataset with the following parameters: -pilotlength 500 -npilot 15 -burnin 2500 -nthreads 6. Top 0.1% outliers were chosen as a threshold of significance, as more conservative than the threshold calculated with a pseudo-observed dataset simulated with the co-variance matrix (Gautier 2015). We also used a sliding window approach, with window size of 5 kb and 2.5 kb overlap, using the R package rehh (Gautier et al. 2012).

Age estimates and gene ontology enrichment

The age of each single SNP was computed with GEVA (Albers and McVean 2020) to estimate the average SNP age for each annotated gene (https://phytozome-next.jgi.doe.gov). All private SNPs to the combined A and B lineages were polarized using the ancestral C lineage using custom R scripts. GEVA was run on the five main scaffolds (corresponding to the five chromosomes) using the genetic map produced by (Huo et al. 2011) and the polarized SNP dataset. The average age of genes associated with the 32 bioclimatic variables described above or outliers in the $X^T X$ analysis was compared to the average age of genes at the genome wide level with Wilcoxon test. We only used the gene lists produced by the GEA and the $X^T X$ analyses if they contained at least fifteen genes. Gene Ontology search was performed with PANTHER v17.0 (http://www.pantherdb.org) using the built-in database for *B. distachyon.*

DATA AVAILABILITY

Seeds will be distributed through GRIN but can in the meanwhile be obtained in small quantities upon request. Illumina paired-end sequencing data generated for this project are archived on the European Nucleotide Archive, project number PRJEB61986. Archive numbers for the reads produced by Gordon et al. (2018, 2020), Skalska et al. (2020) and Stritt et al. (2022) are available in the respective publications. Scripts are available at https://github.com/nminad/env_genomics.

AUTHOR CONTRIBUTION

NM conceived the study, performed the niche modeling, population genetics and GEA analyses, wrote the manuscript. RH and YB performed age estimates, population size evolution and statistics. CS, DC and MT collected samples. HW performed the niche modeling analysis. ACR conceived the study and wrote the manuscript. All authors have read and agreed to the published version of the manuscript.

ACKNOWLEDMENTS

We are extremely grateful to the Swiss National Science foundation (project 31003A_182785) and the Research Priority Program Evolution in action from the University of Zurich for their generous funding. We would like to thank Chris Thomas and Léa Frachon for valuable discussions on niche modelling and GEA respectively.

SUPPLEMENTARY INFORMATION

7 figures and 3 tables are available in the supplementary materials.

REFERENCES

Albers PK, McVean G. 2020. Dating genomic variants and shared ancestry in population scale sequencing data. PLoS Biol. 18:1–26.

Anderson D, Burnham K. 2004. Model selection and multi-model inference. Second. NY: Springer-Verlag, 63(2020), p.10.

Barbieri M, Marcel TC, Niks RE, Francia E, Pasquariello M, Mazzamurro V, Garvin DF. 2012. QTLs for resistance to the false brome rust *Puccinia brachypodii* in the model grass *Brachypodium distachyon*. 55:152–163.

Beale CM, Lennon JJ, Gimona, A, 2008. Opening the climate envelope reveals no macroscale associations with climate in European birds. Proceedings of the National Academy of Sciences 105(39), pp.14908-14912.

Benjamin Y, Hochberg Y. 1995. Controlling the False Discovery Rate: A Practical and Powerful Approach to Multiple Testing. Stat. Methodol. 57:289–300.

Binney H, Edwards M, Macias-fauria M, Lozhkin A, Anderson P, Kaplan JO, Andreev A, Bezrukova E, Blyakharchuk T, Jankovska V, et al. 2017. Vegetation of Eurasia from the last glacial maximum to present: Key biogeographic patterns. Quat. Sci. Rev. 157:80–97.

Bouché F, Woods DP, Linden J, Li W, Mayer KS, Amasino RM, Périlleux C. 2022. EARLY FLOWERING 3 and Photoperiod Sensing in *Brachypodium distachyon*. Front. Plant Sci. 12:1–14.

Bourgeois Y, Stritt C, Walser JC, Gordon SP, Vogel JP, Roulin AC. 2018. Genome-wide scans of selection highlight the impact of biotic and abiotic constraints in natural populations of the model grass *Brachypodium distachyon*. Plant J. 96:438–451.

Casimiro-Soriguer R, Talavera M, Balao F, Terrab A, Herrera J, Talavera S. 2010. Phylogeny and genetic structure of *Erophaca* (Leguminosae), a East-West Mediterranean disjunct genus from the Tertiary. Mol. Phylogenet. Evol.56:441–450.

Catalán P, López-Álvarez D, Bellosta C, Villar L. (2016). Updated taxonomic descriptions, iconography, and habitat preferences of Brachypodium distachyon, B. stacei, and B. hybridum (Poaceae). Anales Del Jardín Botánico De Madrid, 73(1), e028.

Coomey JH, Sibout R, Hazen SP. 2020. Grass secondary cell walls*, Brachypodium distachyon* as a model for discovery. New Phytol. 227:1649–1667.

Dalmais M, Antelme S, Ho-Yue-Kuang S, Wang Y, Darracq O, d'Yvoire MB, Cézard L, Légée F, Blondet E, Oria N, et al. 2013. A TILLING Platform for Functional Genomics in *Brachypodium distachyon*. PLoS One 8.

Danecek P, Auton A, Abecasis G, Albers CA, Banks E, Depristo MA, Handsaker RE, Lunter G, Marth GT, Sherry ST, et al. 2011. The variant call format and VCFtools. Bioinformatics 27:2156–2158.

Davis BAS, Fasel M, Kaplan JO, Russo E, Burke A. 2022. The climate and vegetation of Europe, North Africa and the Middle East during the Last Glacial Maximum (21, 000 years BP) based on pollen data. Preprint: https://doi.org/10.5194/cp-2022-59.

Del'Acqua MD, Zuccolo A, Tuna M, Gianfranceschi L, Pè ME. 2014. Targeting environmental

adaptation in the monocot model *Brachypodium distachyon*: a multi-faceted approach.
BMC Genomics 15:801.

Durvasula A, Fulgione A, Gutaker RM, Alacakaptan SI, Flood PJ, Neto C, Tsuchimatsu T,
Burbano HA, Picó FX, Alonso-Blanco C, et al. 2017. African genomes illuminate the early
history and transition to selfing in *Arabidopsis thaliana*. Proc. Natl. Acad. Sci. 114:5213–
5218.

Elith J, Graham C, P. Anderson R, Dudík M, Ferrier S, Guisan A, J. Hijmans R, Huettmann F, R.
Leathwick J, Lehmann A, et al. 2006. Novel methods improve prediction of species'
distributions from occurrence data. Ecography. 29:129–151.

Elith J., Phillips SJ, Hastie T, Dudík M, Chee YE and Yates CJ. 2011. A statistical explanation of
MaxEnt for ecologists. Diversity and distributions, 17:43-57.

Exposito-Alonso M. 2020. Seasonal timing adaptation across the geographic range of
Arabidopsis thaliana. Proc. Natl. Acad. Sci. U. S. A. 117:9665–9667.

Evanno G, Regnaut S, Goudet J. 2005. Detecting the number of clusters of individuals using
the software STRUCTURE: A simulation study. Mol. Ecol. 14:2611–2620.

Exposito-Alonso M, Exposito-Alonso M, Gómez Rodríguez R, Barragán C, Capovilla G, Chae E,
Devos J, Dogan ES, Friedemann C, Gross C, et al. 2019. Natural selection on the
Arabidopsis thaliana genome in present and future climates. Nature 573:126–129.

Feliner GN. 2011. Southern European glacial refugia: A tale of tales. Taxon 60:365–372.

Feliner GN. 2014. Patterns and processes in plant phylogeography in the Mediterranean
Basin. A review. Perspect. Plant Ecol. Evol. Syst.16:265–278.

Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global
land areas. Int. J. Climatol. 37:4302–4315.

Flouri T, Jiao X, Rannala B, Yang Z. 2018. Species tree inference with *BPP* using genomic
sequences and the multispecies coalescent. Mol. Biol. Evol. 35:2585–2593.

Frichot E, François O. 2015. LEA: An *R* package for landscape and ecological association
studies. Methods Ecol. Evol. 6:925–929.

Fulgione A, Koornneef M, Roux F, Hermisson J, Hancock AM. 2018. Madeiran *Arabidopsis
thaliana* reveals ancient long-range colonization and clarifies demography in Eurasia.
Mol. Biol. Evol. 35:564–574.

Gamba D, Lorts C, Haile A, Sahay S, Lopez L, Xia T, Kulesza E, Elango D, Kerby J, Yifru M, et al.
2022. The genomics and physiology of abiotic stressors associated with global elevation
gradients in *Arabidopsis thaliana*. bioRxiv https://doi.org/10.1101/2022.03.22.485410.

Gautier M. 2015. Genome-Wide Scan for Adaptive Divergence and Association with
Population-Specific Covariates. Genetics 201:1555–1579.

Gautier M, Vitalis R, Baillarguet D, Cedex F-M. 2012. rehh : an R package to detect footprints
of selection in genome-wide SNP data from haplotype structure. Bioinformatics
28:1176–1177.

Gent PR, Danabasoglu G, Donner LJ, Holland MM, Hunke EC, Jayne SR, Lawrence DM, Neale
RB, Rasch PJ, Vertenstein M, et al. 2011. The community climate system model version

4. J. Clim. 24:4973–4991.

Giraldo P, Rodríguez-Quijano M, Vázquez JF, Carrillo JM, Benavente E. 2012. Validation of microsatellite markers for cytotype discrimination in the model grass *Brachypodium distachyon*. Genome 55:523–527.

Gloss AD, Vergnol A, Morton TC, Laurin PJ, Roux F, Bergelson J. 2022. Genome-wide association mapping within a local *Arabidopsis thaliana* population more fully reveals the genetic architecture for defensive metabolite diversity. Philos. Trans. R. Soc. B Biol. Sci. 377.

Gordon SP, Contreras-Moreira B, Levy JJ, Djamei A, Czedik-Eysenberg A, Tartaglio VS, Session A, Martin J, Cartwright A, Katz A, et al. 2020. Gradual polyploid genome evolution revealed by pan-genomic analysis of *Brachypodium hybridum* and its diploid progenitors. Nat. Commun.11:1–16.

Gordon SP, Contreras-Moreira B, Woods DP, Des Marais DL, Burgess D, Shu S, Stritt C, Roulin AC, Schackwitz W, Tyler L, et al. 2017. Extensive gene content variation in the *Brachypodium distachyon* pan-genome correlates with population structure. Nat. Commun.8.

Groves RH. 2000. Temperate grasslands of the southern hemisphere. In Grasses Systematics and Evolution. Jacobs, S. W. L. and Everett, J., eds. Melbourne: CSIRO Publishing, pp. 3781–3791.

Günther, T. & Coop, G. Robust Identification of Local Adaptation from Allele Frequencies. Genetics 195, 205–220 (2013).

Haberer G, Kamal N, Bauer E, Gundlach H, Fischer I, Seidel MA, Spannagl M, Marcon C, Ruban A, Urbany C, et al. 2020. European maize genomes highlight intraspecies variation in repeat and gene content. Nat. Genet.52:950–957.

Hancock AM, Brachi B, Faure N, Horton MW, Jarymowycz LB, Sperone FG, Toomajian C, Roux F, Bergelson J. 2011. Adaptation to climate across the *Arabidopsis thaliana* genome. Science 334:83–86.

Hasterok R, Catalan P, Hazen SP, Roulin AC, Vogel JP, Wang K, Mur LAJ. 2022. Brachypodium: 20 years as a grass biology model system; the way forward? Trends Plant Sci.27:1002–1016.

Hermisson J, Pennings PS. 2005. Soft Sweeps: Molecular Population Genetics of Adaptation From Standing Genetic Variation. 169:2335–2352.

Horton MW, Hancock AM, Huang YS, Toomajian C, Atwell S, Auton A, Muliyati NW, Platt A, Sperone FG, Vilhjálmsson BJ, et al. 2012. Genome-wide patterns of genetic variation in worldwide *Arabidopsis thaliana* accessions from the RegMap panel. Nat. Genet.44:212–216.

Huo N, Garvin DF, You FM, Luo SMM, Gu YQ, Lazo GR, Philip J. 2011. Comparison of a high-density genetic linkage map to genome features in the model *grass Brachypodium distachyon*. Theor. Appl. Genet. 123:455–464.

Hus K, Betekhtin A, Pinski A, Rojek-Jelonek M, Grzebelus E, Nibau C, Gao M, Jaeger KE, Jenkins G, Doonan JH, et al. 2020. A CRISPR/Cas9-Based Mutagenesis Protocol for *Brachypodium*

distachyon and Its Allopolyploid Relative, *Brachypodium hybridum*. Front. Plant Sci. 11:1–
21.

International Brachypodium Initiative. 2010. Genome sequencing and analysis of the model
grass *Brachypodium distachyon*. Nature 463:763–768.

Jayakodi M, Padmarasu S, Haberer G, Bonthala VS, Gundlach H, Monat C, Lux T, Kamal N, Lang
D, Himmelbach A, et al. 2020. The barley pan-genome reveals the hidden legacy of
mutation breeding. Nature 588:284–289.

Jennings RP, Singarayer J, Stone EJ, Krebs-Kanzow U, Khon V, Nisancioglu KH, Pfeiffer M,
Zhang X, Parker A, Parton A, et al. 2015. The greening of Arabia: Multiple opportunities
for human occupation of the Arabian Peninsula during the Late Pleistocene inferred from
an ensemble of climate model simulations. Quat. Int.382:181–199.

Jiang Y, Wang X, Yu X, Zhao X, Luo N, Pei Z. 2017. Quantitative Trait Loci Associated with
Drought Tolerance in *Brachypodium distachyon*. Front. Plant Sci. 8:1–11.

Kooyers NJ, Greenlee AB, Colicchio JM, Oh M, Blackman BK. 2015. Replicate altitudinal clines
reveal that evolutionary flexibility underlies adaptation to drought stress in annual
Mimulus guttatus. New Phytol. 206:152–165.

Korunes KL, Samuk K. 2021. pixy: Unbiased estimation of nucleotide diversity and divergence
in the presence of missing data. Mol. Ecol. Resour. 21:1359–1368.

Lasky JR, Josephs EB, Morris GP. 2023. Genotype-environment associations to reveal the
molecular basis of environmental adaptation. Plant Cell 35:125–138.

Lee CR, Svardal H, Farlow A, Exposito-Alonso M, Ding W, Novikova P, Alonso-Blanco C, Weigel
D, Nordborg M. 2017. On the post-glacial spread of human commensal *Arabidopsis
thaliana*. Nat. Commun. 8:1–12.

Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM.
arXiv:1303.3997

Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R.
2009. The Sequence Alignment/Map format and SAMtools. Bioinformatics 25:2078–
2079.

López-Alvarez D, López-Herranz ML, Betekhtin A, Catalán P. 2012. A DNA barcoding method
to discriminate between the model plant *Brachypodium distachyon* and its close
relatives *B. stacei* and *B. hybridum* (Poaceae). PLoS One 7:e51058.

López-Álvarez D, Manzaneda AJ, Rey PJ, Giraldo P, Benavente E, Allainguillaume J, Mur L,
Caicedo AL, Hazen SP, Breiman A, et al. 2015. Environmental niche variation and
evolutionary diversification of the *Brachypodium distachyon* grass complex species in
their native circum-Mediterranean range. Am. J. Bot. 102:1073–1088.

Lovell JT, MacQueen AH, Mamidi S, Bonnette J, Jenkins J, Napier JD, Sreedasyam A, Healey A,
Session A, Shu S, et al. 2021. Genomic mechanisms of climate adaptation in polyploid
bioenergy switchgrass. Nature 590:438–444.

Lynch M, Ackerman MS, Gout JF, Long H, Sung W, Thomas WK, Foster PL. 2016. Genetic drift,
selection and the evolution of the mutation rate. Nat. Rev. Genet. 17:704–714.

Des Marais DL, Lasky JR, Verslues PE, Chang TZ, Juenger TE. 2017. Interactive effects of water

limitation and elevated temperature on the physiology, development and fitness of diverse accessions of *Brachypodium distachyon*. New Phytol. 214:132–144.

Marees, AT, de Kluiver, H, Stringer, S, et al. A tutorial on conducting genome-wide association studies: Quality control and statistical analysis. Int J Methods Psychiatr Res. 2018; 27:e1608. https://doi.org/10.1002/mpr.1608

Marks RA, Amézquita EJ, Percival S, Rougon-Cardoso A, Chibici-Revneanu C, Tebele SM, Farrant JM, Chitwood DH, VanBuren R. 2023. A critical analysis of plant science literature reveals ongoing inequities. Proc. Natl. Acad. Sci.120:10 e2217564120.

McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M, DePristo MA. 2010. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data*.* Genome Res*,* 20:1297-1303

McKown KH, Bergmann DC. 2020. Stomatal development in the grasses: lessons from models and crops (and crop models). New Phytol. 227:1636–1648.

Meredith M, Kruschke J. 2022. HDInterval. HDInterval: Highest (Posterior) Density Intervals. R package. https://CRAN.R-project.org/package=HDInterval

de Miguel M, Rodríguez-Quilón I, Heuertz M, Hurel A, Grivet D, Jaramillo-Correa JP, Vendramin GG, Plomion C, Majada J, Alía R, et al. 2022. Polygenic adaptation and negative selection across traits, years and environments in a long-lived plant species (*Pinus pinaster* Ait., Pinaceae). Mol. Ecol. 31:2089–2105.

Mittler R, Zandalinas SI, Fichman Y, Van Breusegem F. 2022. Reactive oxygen species signaling in plant stress responses. Nat. Rev. Mol. Cell Biol. 23:663–679.

Moles AT, Warton DI, Warman L, Swenson NG, Laffan SW, Zanne AE, Pitman A, Hemmings FA, Leishman MR. 2009. Global patterns in plant height. J. Ecol. 97:923–932.

Monroe JG, Srikant T, Carbonell-Bejerano P, Becker C, Lensink M, Exposito-Alonso M, Klein M, Hildebrandt J, Neumann M, Kliebenstein D, et al. 2022. Mutation bias reflects natural selection in *Arabidopsis thaliana*. Nature 602:101–105.

Montenegro JD, Golicz AA, Bayer PE, Hurgobin B, Lee H, Batley J, Edwards D. 2017. The pangenome of hexaploid bread wheat. Plant J. 90:1007–1013.

Nielsen R, Williamson S, Kim Y, Hubisz MJ, Clark AG, Bustamante C. 2005. Genomic scans for selective sweeps using SNP data. Genome Research 15:1566–1575.

Nunes TDG, Zhang D, Raissig MT. 2020. Form, development and function of grass stomata. Plant J. 101:780–799.

Nunes TDG, Berg LS, Slawinska MW, Zhang D, Redt L, Sibout R, Vogel J, Laudencia-Chingcuano D, Jesenofsky B, Lindner H, Raissig MT. 2023. Report Regulation of hair cell and stomatal size by a hair cell-specific peroxidase in the *grass Brachypodium distachyon*. Curr. Biol.33:1–11.

Ortiz MÁ, Tremetsberger K, Talavera S, Stuessy T, García-Castaño JL. 2007. Population structure of *Hypochaeris salzmanniana* DC. (Asteraceae), an endemic species to the Atlantic coast on both sides of the Strait of Gibraltar, in relation to Quaternary sea level changes. Mol. Ecol. 16:541–552.

Phillips SJ, Anderson RP, Schapire RE. 2006. Maximum entropy modeling of species
geographic distributions. Ecol. Modell. 190:231–259.

Provart NJ, Alonso J, Assmann SM, Bergmann D, Brady SM, Brkljacic J, Browse J, Chapple C,
Colot V, Cutler S, et al. 2016. 50 years of Arabidopsis research: Highlights and future
directions. New Phytol. 209:921–944.

Quinlan AR, Hall IM. 2010. BEDTools: a flexible suite of utilities for comparing genomic
features. Bioinformatics 26:841–842.

Raissig MT, Woods DP. 2022. The wild grass *Brachypodium distachyon* as a developmental
model system. Current Topics in Developmental Biology 147: 33-71

Rannala B, Yang Z. 2003. Bayes estimation of species divergence times and ancestral
population sizes using DNA sequences from multiple loci. Genetics 164:1645–1656.

Ream TS, Woods DP, Schwartz CJ, Sanabria CP, Mahoy JA, Walters EM, Kaeppler HF, Amasino
RM. 2014. Interaction of photoperiod and vernalization determines flowering time of
Brachypodium distachyon. Plant Physiol. 164:694–709.

Rellstab C, Gugerli F, Eckert AJ, Hancock AM, Holderegger R. 2015. A practical guide to
environmental association analysis in landscape genomics. Mol. Ecol. 24:4348–4370.

Sharma N, Ruelens P, Mariëlla D, Maggen T, Dochy N, Torfs S, Kaufmann K, Rohde A, Geuten
K, Leuven KU, et al. 2017. A Flowering Locus C Homolog Is a Vernalization-Regulated
Repressor *in Brachypodium* and Is Cold Regulated in Wheat. Plant Phys. 173:1301–1315.

Skalska A, Stritt C, Wyler M, Williams Hefin W, Vickers M, Han J, Tuna M, Savas Tuna G, Susek
K, Swain M, et al. 2020. Genetic and Methylome Variation in Turkish *Brachypodium*
distachyon Accessions Differentiate Two Geographically Distinct Subpopulations. Int. J.
Mol. Sci. 21:6700.

Speidel L, Forest M, Shi S, Myers SR. 2019. A method for genome-wide genealogy estimation
for thousands of samples. Nat. Genet. 51:1321–1329.

Stritt C, Gimmi EL, Wyler M, Bakali AH, Skalska A, Hasterok R, Mur LAJ, Pecchioni N, Roulin AC.
2022. Migration without interbreeding: Evolutionary history of a highly selfing
Mediterranean grass inferred from whole genomes. Mol. Ecol. 31:70–85.

Stritt C, Gordon SP, Wicker T, Vogel JP, Roulin AC. 2018. Recent activity in expanding
populations and purifying selection have shaped transposable element landscapes
across natural accessions of the Mediterranean grass *Brachypodium distachyon*.
Genome Biol. Evol. 10:304–318.

Suzuki R, Shimodaira H. 2006. Pvclust: An R package for assessing the uncertainty in
hierarchical clustering. Bioinformatics 22:1540–1542.

Takou M, Wieters B, Kopriva S, Coupland G, Linstädter A, De Meaux J. 2019. Linking genes
with ecological strategies *in Arabidopsis thaliana*. J. Exp. Bot. 70:1141–1151.

Tang K, Thornton KR, Stoneking M. 2007. A New Approach for Using Genome Scans to Detect
Recent Positive Selection in the Human Genome. PLoS Biol. 5.

Vidigal DS, Marques ACSS, Willems LAJ, Buijs G, Méndez-Vigo B, Hilhorst HWM, Bentsink L,
Picó FX, Alonso-Blanco C. 2016. Altitudinal and climatic associations of seed dormancy
and flowering traits evidence adaptation of annual life cycle timing in *Arabidopsis*

thaliana. Plant Cell Environ. 39:1737–1748.

Wadgymar SM, Ogilvie JE, Inouye DW, Weis AE, Anderson JT. 2018. Phenological responses
793   to multiple environmental drivers under climate change: insights from a long-term
794   observational study and a manipulative field experiment. New Phytol. 218:517–529.

795   Walkowiak S, Gao L, Monat C, Haberer G, Kassa MT, Brinton J, Ramirez-Gonzalez RH, Kolodziej
796   MC, Delorean E, Thambugala D, et al. 2020. Multiple wheat genomes reveal global
797   variation in modern breeding. Nature 588:277–283.

798   Wang W, Mauleon R, Hu Z, Chebotarov D, Tai S, Wu Z, Li M, Zheng T, Fuentes RR, Zhang F, et
799   al. 2018. Genomic variation in 3,010 diverse accessions of Asian cultivated rice. Nature
800   557:43–49.

801   Warren DL, Glor RE, Turelli M. 2010. ENMTools: a toolbox for comparative studies of
802   environmental niche models. Ecography 33:607–611.

803   Warren DL, Matzke NJ, Cardillo M, Baumgartner JB, Beaumont LJ, Turelli M, Glor RE, Huron
804   NA, Simões M, Iglesias TL, et al. 2021. ENMTools 1.0: an R package for comparative
805   ecological biogeography. Ecography 44:504–511.

806   Wei T, Simko V. 2021. R package 'corrplot': Visualization of a Correlation Matrix. (Version
807   0.92), https://github.com/taiyun/corrplot.

808   Wieters B, Steige KA, He F, Koch EM, Ramos-Onsins SE, Gu H, Guo YL, Sunyaev S, de Meaux J.
809   2021. Polygenic adaptation of rosette growth in Arabidopsis thaliana. PLoS Genet. 17:1–
810   27.

811   Wyler M, Stritt C, Walser JC, Baroux C, Roulin AC. 2020. Impact of transposable elements on
812   methylation and gene expression across natural accessions of *Brachypodium distachyon.*
813   Genome Biol. Evol. 12:1994–2001.

814   Williams HW, Cross DE, Crump HL, Drost CJ, Thomas CJ. 2015. Climate suitability for European
815   ticks: Assessing species distribution models against null models and projection under
816   AR5 climate. Parasites and Vectors 8:1–15.

817   Wilson PB, Streich JC, Murray KD, Eichten SR, Cheng R, Aitken NC, Spokas K, Warthmann N,
818   Gordon SP, Vogel JP, et al. 2019. Global diversity of the Brachypodium species complex
819   as a resource for genome-wide association studies demonstrated for agronomic traits in
820   response to climate. Genetics 211:317–331.

821   Woods DP, Bednarek R, Bouché F, Gordon SP, Vogel JP, Garvin DF, Amasino RM. 2017. Genetic
822   architecture of flowering-time variation in *Brachypodium distachyon*. Plant Physiol.
823   173:269–279.

824   Woods DP, Dong Y, Bouché F, Mayer K, Varner L, Ream TS, Thrower N, Wilkerson C, Cartwright
825   A, Sibout R, et al. 2020. Mutations in the predicted DNA polymerase subunit POLD3
826   result in more rapid flowering of *Brachypodium distachyon*. New Phytol.227:1725-1735.

827   Woods DP, Ream TS, Minevich G, Hobert O, Amasino RM. 2014. PHYTOCHROME C is an
828   essential light receptor for photoperiodic flowering in the temperate grass,
829   *Brachypodium distachyon*. Genetics 198:397–408.

830   Woodward AW, Bartel B. 2018. Biology in bloom: A primer on the *Arabidopsis thaliana* model
831   system. Genetics 208:1337–1349.

Wu Q, Han T, Chen X, Chen J, Zou Y, Li Z, Xu Y. 2017. Long-term balancing selection contributes
to adaptation in *Arabidopsis* and its relatives. Genome Biol. 18:217.

Yan W, Wang B, Chan E, Mitchell-Olds T. 2021. Genetic architecture and adaptation of
flowering time among environments. New Phytol. 230:1214–1227.

Yeaman S. 2022. Evolution of polygenic traits under global vs local adaptation. Genetics
220:iyab134

Yokoyama Y, Lambeck K, De Deckker P, Johnston P, Fifield LK. 2000. Timing of the Last Glacial
Maximum from observed sea-level minima. Nature 406:713-716.

Zhang D, Abrash EB, Nunes TDG, Prados IH, Gil MXA, Jesenofsky B, Lindner H, Bergmann DC,
Raissig MT. 2022. Opposite polarity programs regulate asymmetric subsidiary cell
divisions in grasses. Elife11:e79913:2022.04.24.489281.

Zhen Y, Ungerer MC. 2008. Clinal variation in freezing tolerance among natural accessions of
Arabidopsis thaliana. New Phytol. 177:419–427.

Zheng X, Levine D, Shen J, Gogarten SM, Laurie C, Weir BS. 2012. A high-performance
computing toolset for relatedness and principal component analysis of SNP data.
Bioinformatics 28:3326–3328.

Zhou X, Stephens M. 2012. Genome-wide efficient mixed-model analysis for association
studies. Nat. Genet. 44:821–824.