Identification of domains in *Plasmodium falciparum* proteins of unknown function using DALI search on Alphafold predictions

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Abstract

Plasmodium falciparum, the causative agent of malaria, poses a significant global health challenge, yet much of its biology remains elusive. A third of the genes in the *P. falciparum* genome lack annotations regarding their function, impeding our understanding of the parasite's biology. In this study, we employ structure predictions and the DALI search algorithm to analyse proteins encoded by uncharacterized genes in the reference strain 3D7 of *P. falciparum*. By comparing Alphafold predictions to experimentally determined protein structures in the Protein Data Bank, we found similarities to known domains in 353 proteins of unknown function, shedding light on their potential functions. The lowest-scoring 5% of similarities were additionally validated using the size-independent TM-align algorithm, confirming the detected similarities in 88% of the cases. Notably, in over 70 *P. falciparum* proteins the presence of domains resembling heptatricopeptide repeats, which are typically involvement in RNA binding and processing, was detected. This suggests this family, which is important in transcription in mitochondria and apicoplasts, is much larger in *Plasmodium* parasites than previously thought. The results of this domain search provide a resource to the malaria research community that is expected to inform and enable experimental studies.

Introduction

Plasmodium falciparum, the malaria parasite responsible for over 600 000 deaths every year (World Health Organization, 2022), has a complex life cycle and its biology is still only partly understood. Of its 5268 predicted genes, 1407 remain without any annotation (release 62, January 2023 (Amos et al., 2022; Aurrecoechea et al., 2009)). Annotations of genes are most often based on sequence similarity to well characterized proteins in other organisms (Böhme et al., 2019). As part of an experimental gene-by-gene analysis of unknown proteins encoded on *P. falciparum* chromosome 3 we found that structural similarities of Alphafold structures matched well the experimental evidence of several proteins of unknown function (Kimmel et al., 2023). This indicates that many parasite proteins have a conserved evolutionary origin but evolved beyond recognition on the primary sequence level but not on the structural level. It can therefore be assumed that comparisons of the structural prediction of unknown proteins will yield useful information on proteins currently annotated as unknown. The availability of Alphafold structure predictions (Jumper et al., 2021) now enables the identification of domains based on geometric comparisons, independent of DNA and amino acid sequences. These similarities could provide information about gene products that so far lacked annotation. Here we apply this approach to all genes of unknown function in *P. falciparum*, specifically to the reference strain 3D7. Several sequence-independent algorithms exist to compare protein structures to each other based on geometry, including VAST (Gibrat et al., 1996), Dali (Holm, 2022), Foldseek (van Kempen et al., 2022), CE (Shindyalov & Bourne, 1998), the protein size-

independent scoring algorithm TM-align (Zhang, 2005) and others (reviewed in (Kufareva & Abagyan,
2011)). For this screen we chose the DALI search algorithm as it has several advantages. It is set up to
search the PDB and has an integrated graphical user (GUI) interface that also allows quick assessment
of PFAM-annotated domains in the aligned structures. Further it has a built-in GUI for visual 3D-
assessment of the alignment of hits to the query. We chose an open approach, where we screen all
proteins of unknown function for the presence of any domains, rather than searching for a
predefined set of domains or restricting the search to a small set of proteins of interest. This has the
advantage that discoveries can be made independent of hypotheses that are related to specific
processes in the parasite.
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Using this approach, we here report the identification of domains with similarities to experimentally
determined protein domains in 287 proteins encoded by genes of unknown function from *P.*
falciparum 3D7 by an open-ended search. In addition, we used a targeted approach to search for
proteins with similarity to the armadillo domain-contain ASA2 and ASA3 proteins which came to our
attention as a frequent similarity in the open-ended approach. In this targeted search we found folds
resembling heptatricopeptide repeats, which are typically involved in RNA-binding and -processing,
in 53 previously un-annotated proteins, indicating that this family is much larger than previously
though. We provide these results as a resource to the community to support experimental work.
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Results
Identification of domains in proteins of unknown function by open-ended domain search
In order to identify known domains in *P. falciparum* proteins of unknown function, all 1407 proteins
which were named protein of unknown function on PlasmoDB (The Plasmodium Genome Database
Collaborative, 2001) were selected for analysis. The Alphafold predictions of the proteins encoded by
these genes were compared to experimentally determined structures in the protein data bank (PDB)
(Burley et al., 2019) using the search algorithm DALI (Holm, 2022). Included in the search were
proteins that contained "globular" folds, thus excluding 476 structures which contained solely
disordered, extended and fibrous folds, as this is a requirement for successful DALI searches. The
resulting 931 hits were visually assessed to exclude structures that aligned only by two or fewer
helices or beta strands and this resulted in the identification of similarity to at least one annotated
domain in 287 of the 1407 unknown proteins (Figure 1A, Supplementary data 1).
77
The DALI search assigns a Z-score to each alignment which reflects its quality, where Z-scores of
below 2 indicate non-specific similarity. While all except one of the 287 structure alignments that
were considered specific and a good fit by the visual inspection had Z- scores of 5 or higher,
PF3D7_1336600 had a lower Z-score. However, the lower score in that case was due to its small size
as it still showed a very good alignment (also shown in Figure 2). The six highest-scoring similarities
had Z-scores of over 30 (Figure 1B). Of the proteins in which domains were identified, 254 contained
one newly identified domain, 23 contained two and five proteins contained 3 newly identified
domains (Figure 1C). As an example of a protein in which similarity to two domains was detected
PF3D7_1013300 was visualized (Figure S1). Among the identified domains, most (>200) domains
occurred only once, 27 occurred twice and 17 occurred three times or more (Figure 1D).
88
To validate these results, the lowest-scoring 5% of similarities found (based on DALI Z-score), were
assessed using a second algorithm. For this TM-align was used which is independent of protein size
(Zhang, 2005). This algorithm scores similarities between two protein structures on a scale from 0 to

1, where scores over 0.5 indicate that searched structures contain the queried fold. Of the 16
structures assessed 14 showed a TM-score over 0.5 (Figure 2), when compared to the top annotated
hit from the DALI search (which was an experimentally determined structure downloaded from the
PDB and cropped to the domain that showed similarity). The remaining two showed TM-scores over
0.4. Thus, the two algorithms show good agreement even among the lowest-scored similarities,
suggesting a low proportion of false positive similarities.
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Comparison with published data
To assess whether the observed similarities have the potential to predict the function of the *P.*
falciparum proteins of unknown function, we assess the literature for published experimental data.
Although all of the 287 proteins for which similarity was found are encoded by genes that are
annotated as of unknown function, some experimental data exists for eight of them (Table 1). The
cellular location of the proteins encoded by PF3D7_0307600, PF3D7_0313400 and PF3D7_0319900
was previously assessed by fusing the endogenous gene with the sequence encoding GFP in an
experimental screen of genes of unknown function from chromosome three in *P. falciparum* 3D7
parasites (Kimmel et al., 2023) (Table 1). PF3D7_0205600, PF3D7_1013300 and PF3D7_1329500
were analysed by the same approach (Schmidt et al., 2022) (Birnbaum et al., 2017; Khosh-Naucke et
al., 2018) while the *P. berghei* ortholog of PF3D7_1132400 was expressed as an endogenous HA-
fusion (Zuegge et al., 2001) and their cellular locations observed (Table 1). For PF3D7_0903600
functional data of the orthologue in *Toxoplasma gondii* recently became available (Dubois et al.,
2023).
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PF3D7_0205600 and PF3D7_1329500 are located in the nucleus, supporting that the identified
regions with similarity to CDC45-like and TIG domains, which are found in DNA replication proteins
and transcription factors, respectively, might serve the same functions. PF3D7_1013300 showed
similarity to both domains that make up nicastrins (Xie et al., 2014)and equally has a transmembrane
domain and a short C-terminal tail (Figure S1). In other organisms nicastrin is part of the gamma
secretase protein complex that proteolytically processes integral membrane proteins (Shah et al.,
2005). Because inhibitors against the human gamma secretase complex had no effect on *P.*
falciparum (Li et al., 2009) and homologs of the complex components were not found by sequence
similarity, the gamma secretase complex was thought to be absent in *Plasmodium* parasites. Yet, the
location of PF3D7_1013300 in the ER and the cell periphery would be consistent with nicastrin in
other systems where it gets modified in the Golgi before transport to its final destination at the
plasma membrane. Thus, the experimental data is compatible with the observed structural similarity
and suggests that at least the nicastrin component of the gamma secretase complex is present in
P. falciparum parasites.
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129   For PF3D7_0903600 a HOOK_N domain-like fold was detected, which typically occurs in dynein-
130   associated cargo adaptors. It's ortholog in *Toxoplasma gondii* has recently been shown to interact
131   with typical hook-interacting proteins and be involved in processes typical for hook proteins (Dubois
132   et al., 2023), giving credibility to the detected domain.
133
For PF3D7_0313400 (AAA domain), PF3D7_0319900 (UTRA domain) and PF3D7_1132400 (CRT-like
domain) the identified domains are not specific to distinct cellular locations but the experimentally
observed cellular location do not disagree with their potential function. As such the domain
predictions are possible based on the available experimental data. Solely for PF3D7_0307600, which

we found to harbour similarity to a Rad51 domain (typically involved in DNA repair), the GFP-fusion

protein was - contrary to our prediction - not found in the nucleus (Kimmel et al., 2023). This might

be due to a faulty prediction, a repurposing in *P. falciparum* or an altered cellular location due to the

GFP-fusion, which is possible given it is not essential (Kimmel et al., 2023; Schwach et al., 2015).

Table 1: Comparison of published data with predicted domains.

PlasmoDB ID	Newly identified domain and its function	Experimental data available for protein	Match of domain identification and experimental data
PF3D7_0205600	CDC45-like; DNA replication (Z-score 28.0)	GFP-fusion localizes to nucleus and cytoplasm (Birnbaum et al., 2017).	Predicted function **supported** by experimental data.
PF3D7_0307600	Rad51; DNA repair (Z-score 24.2)	GFP-fusion gives unknown localization pattern within the parasite (Kimmel et al., 2023).	Predicted function **not supported** by experimental data. Expected would be nuclear localization.
PF3D7_0313400	AAA; chaperone (Z-score 13.5)	GFP-fusion localizes to the parasite cytoplasm (Kimmel et al., 2023).	Predicted function **possible** based on experimental data.
PF3D7_0319900	UTRA; Ligand-binding, modulates transcription factor activity in response to small molecules (Z-score 6.7)	GFP-fusion gives unknown localization pattern in the parasite (Kimmel et al., 2023).	Predicted function **possible** based on experimental data.
PF3D7_0903600	HOOK_N; dynein-associated cargo adaptor proteins (Z-score 12.0)	*Toxoplasma gondii* homolog *Tg*HOOK regulates apical positioning and secretion of micronemes and contributes to egress, motility, host cell attachment, and invasion. Interacts with homologs of HOOK interaction partners TgFTS or TgHIP. (Dubois et al., 2023).	Predicted function **supported** by experimental data.
PF3D7_1013300	Ncstrn_small (small lobe) and Nicastrin (large lobe); Two domains making up nicastrin, a part of complex for intra-membrane proteolysis of integral membrane proteins. This protein is modified in Golgi or trans-Golgi network. (Z-score 20.4)	GFP-fusion localizes to the periphery of ring stages, trophozoites and schizonts and is visible in ER in trophozoites and schizonts. (Khosh-Naucke et al., 2018). Isolated from detergent-resistant membranes. HA-fusion gives "dotted labelling within the parasite cytoplasm" (Yam et al., 2013).	Predicted function **supported** by experimental data.
``PF3D7_1132400	CRT-like; chloroquine resistance transporter and homologues. Arabidopsis homologues involved in thiol transport from plastid to cytosol. (Z-score 33.8)	In *P. berghei* non-nuclear, non-apicoplast signal of HA-fusion protein. Predicted to have 9 to 10 TM domains. Western blot shows electrophoresis anomalies common to membrane proteins (Sayers et al., 2018).	Predicted function **possible** based on experimental data.
PF3D7_1329500	TIG; in cell surface receptors and in transcription factors for DNA binding	GFP-fusion localizes to nucleus in ring and trophozoite stage, in addition foci in the parasite periphery at K13 compartment in trophozoites stage (Schmidt et al., 2022).	Predicted function **supported** by experimental data.

| | (Z-score 8.1) | | |

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Proteins containing Armadillo-like domains
It stood out that many unknown *P. falciparum* proteins showed similarity to Mitochondrial ATP
synthase subunit ASA2 and Mitochondrial F1F0 ATP synthase associated 32 kDa protein ASA3. These
two proteins are part of the Polytomella F-ATP synthase complex for which structures of several
states were determined by single-particle cryo-electron microscopy (Murphy et al., 2019). As
according to Interpro no domains are annotated in the ASA2 and ASA3 proteins, no equivalent
domains in the structurally similar *P. falciparum* proteins could be annotated by our approach. To
annotate this large group of *P. falciparum* proteins, we therefore sought to identify the shared
domain of ASA2 and ASA3. Using their structures from PDB (PDB 6rd4), we performed a DALI search
against all PDB structures. The top hits for either protein were ASA2 and ASA3, suggesting that both
were very similar to each other. Further both were similar to proteins containing armadillo domains
(PF00514), for example in catenin delta (PDB 3L6X (Ishiyama et al., 2010)), plakoglobin (PDB 3IFQ
(Choi et al., 2009)), plakophilin (PDB 1XM9 (Choi & Weis, 2005)) and several other proteins,
suggesting that both ASA2 and ASA3 contain domains, which are made up of several armadillo
repeats.
160
We then performed DALI searches of these armadillo domains from ASA2 (residues 1-326) and ASA3
(full length) against all Alphafold-predicted structures of *P. falciparum* in the database. 1158 genes
were found to encode proteins that contains domains similar to ASA2, ASA3 or both. As the Z-score is
dependent on protein size, a suitable cut-off had to be determined for the common domain of ASA2
and ASA3. Spot checks of hits across different Z-scores were performed and showed that hits with a
Z-score of 6.5 or higher showed good similarity to ASA2 or ASA3, with alignment of more than three
armadillo repeats which each consist of one helix pair. To avoid false positives, an even more
stringent Z-score cut-off of 7.0 was chosen which resulted in 121 *P. falciparum* (in 3D7) protein hits
(Figure 3A, Supplementary data 2). All of them were found in both searches (ASA2 and ASA3). The
hits included 18 heptatricopeptide repeat proteins, 13 RAP (**R**NA-binding domain abundant in
Apicomplexans) proteins - both families are thought to be RNA-binding (Hillebrand et al., 2018; Hollin
et al., 2021), - 3 that contain heptatricopeptide repeats and a RAP domain, 3 proteins that are named
armadillo-domain containing proteins, 72 proteins of unknown function and 12 proteins with other
annotations (Figure 3B). In 6 of these, there were also new domains identified in the open-ended
DALI search shown in Figure 1, including DUF559, Importin-beta, Exportin 1-like, Armadillo domain
and Atypical Armadillo domain. Both DUF559 domains (in PF3D7_0104600 and PF3D7_1207900) did
not overlap with the identified armadillo domain, while the remaining four covered the same
residues for which similarity to the ASA2-ASA3-armadillo domain was found (Figure S2).
179
The amino acid sequences of these 121 proteins were aligned using Clustal Omega (Sievers & Higgins,
2018). The sequences of the proteins with annotation did not cluster according to their PlasmoDB
annotations and generally showed little amino acid sequence similarity. We therefore decided to
cluster them by structure. However, the 121 *P. falciparum* hits with similarity to ASA2 and ASA3 were
too many to do this in one batch, thus first the Alphafold predictions of only those with an
annotation in PlasmoDB were clustered using the DALI all-against-all algorithm. Two distinct clusters
were observed and named cluster 1 and cluster 2 (Figure 3C). Cluster 1 was found to contain all
heptatricopeptide repeat proteins, all RAP proteins and ARM2 (Figure 2C-F). All of these proteins are
known or predicted to be RNA-binding proteins (Hillebrand et al., 2018; Hollin et al., 2021; Tang et

al., 2019), while all proteins in cluster 2 are protein-binding proteins, some of which can also bind
RNA (Frankel & Knoll, 2009; Fritz et al., 2009; Geiger et al., 2020; Henrici et al., 2020; Jacot et al.,
2016; Jakel, 1998; Kumar et al., 2023; Xu et al., 2002). Three structures were randomly chosen to
represent each cluster (Figure 3C-E) and aligned to batches of the 72 unknown *P. falciparum* proteins
that harboured similarity to ASA2 and ASA3 (Figure S3). This assigned 53 new proteins to cluster 1
and one new protein to cluster 2, while 18 proteins could not be assigned to either cluster (Figure
3F).
196
Of the 20 proteins previously known to contain RAP domains, 5 were known to also contain
heptatricopeptide repeats. Of the remaining 15 RAP proteins, 13 showed up in our search, suggesting
that these also contain heptatricopeptide repeats. The DUF559 domains identified in two newly
identified heptatricopeptide repeat-like proteins also show some similarity to RAP domains. Thus,
the co-occurrence of RAP domains with heptatricopeptide repeats in the same protein seems to be a
common combination.
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Discussion

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In this study we used the DALI algorithm to search for similarities of the available Alphafold-predicted
structures of all unknown *P. falciparum* (3D7) proteins to known domains in experimentally
determined structures. This similarity search reduced the number of proteins without any annotation
from 1407 to 1054, which is a 25% reduction. We expect the resulting set of several hundred
identified domains to be a useful resource to the malaria research community that will help
streamline further annotation and characterization efforts, and expected to aid the understanding of
the molecular mechanisms of malaria parasites. It provides information on the potential function of
these *P. falciparum* protein inferred from the structural similarity to known proteins in other
organisms. Additionally, in combination with functional and interactome data these domain searches
can aid and substantiate the assignment of proteins to protein groups or complexes which together
serve a specific function as e.g. recently done for proteins in mitosis (Kimmel et al., 2023, Brusini et
al., JCB).
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Open-ended approaches have the advantage that they can discover unexpected components and
pathways. The search in this study gives hints to the unexpected presence of nicastrin, which is part
of a functionality thought to be absent in *Plasmodium* parasites. This approach can also provide hints
for proteins that were expected to be present but had so far not been identified. One example from
our dataset is PF3D7_0404300 which shows similarity to Ran-binding proteins and could be a missing
component in transport in and out of the nucleus.
224
Unexpectedly, we discovered a large group of proteins in the parasite with very similar domains,
including all known heptatricopeptide repeat and RAP proteins as well as 53 unknown proteins.
Proteins containing heptatricopeptide repeats are known to bind and process polycistronic precursor
RNAs into mRNAs and rRNAs for the mitochondrial ribosomes and to stabilize mRNAs to prevent
decay (Hillebrand et al., 2018). Heptatricopeptide repeats proteins contain 37 amino acid long
repeats, while proteins from the related families of pentatricopepide repeat proteins and
octatricopeptide repeats proteins contain repeats which are 35 and 38 amino acids in length. Hepta-,
penta- and octatricopeptide repeat proteins perform comparable RNA binding and processing
functions, yet their distribution among different organism groups, like plants, apicomplexans and
animals, varies (Hillebrand et al., 2018). In humans only 6 heptatricopeptide repeat proteins have

been detected while around 70 were predicted in plants and dinoflagellates. In plants this is in
addition to 450 pentatricopepide repeat proteins. In addition to the 18 previously described
heptatricopeptide repeat proteins in *Plasmodium*, we here identified many more potential members
of this group. If all 54 unknown proteins in cluster 1 of our clustering of the ASA2 and ASA3 hits
indeed are such RNA-binding proteins, this would place the number of heptatricopeptide repeat-like
RNA-binding proteins in *Plasmodium* (72) in a similar range as in plants. Interestingly, while some the
P. falciparum proteins with similarity to heptatricopeptide repeat proteins contain insertions that
break the 37-amino acid repeat motif that gives this domain its name, the geometry of the 37-amino
acid repeat seems conserved.

Another interesting aspect is that we found heptatricopeptide repeat-like folds in a large proportion
of proteins containing RAP and RAP-like DUF559 domains. RAP domains consist of a restriction
endonuclease-like fold and are found in RNA-binding proteins (Hollin et al., 2021). Thus,
heptatricopeptide repeats and RAP domains seem to be a common combination, suggesting that the
two domains might act in concert.

While we believe this search to provide a valuable and useful resource, there are a number of
limitations. The first inherent limitation is that this study used predicted structures of *P. falciparum*
proteins rather than experimentally determined structures, which should not be viewed with the
same confidence as experimentally determined structures. This circumstance has been discussed
extensively following the introduction of Alphafold (Jones & Thornton, 2022; Subramaniam &
Kleywegt, 2022; Terwilliger et al., 2022).

A second limitation was that the search presented here was not exhaustive. To the predicted
structures, we applied DALI, the only search platform with which it was feasible to manually analyse
several hundred protein structures in a reasonable time. This throughput is largely possible because
of the built-in alignment and domain annotation viewers. The DALI algorithm scores search results
based on length and quality of the structure alignment of the whole query structure.
As a result, this algorithm favoured the discovery of one domain of highest confidence, while further
domains in the same protein were only discovered if they incidentally had similar scores as the first
domain or if they occurred in the same experimentally determined structure that the query aligned
to. This is one reason, why the search presented in this study was not exhaustive. Other search
algorithms could circumvent this drawback, for example the VAST algorithm first detects potential
domains in the query structure and then performs searches for each of them individually in addition
to the search with the complete query structure. Generally, the use of other search algorithms, such
as VAST (Gibrat et al., 1996), Foldseek (van Kempen et al., 2022) and PDBeFold (Krissinel & Henrick,
2004), might result in the discovery of different domain similarities due to their different
implementations and ways of scoring similarity. This search being not exhaustive is an important
limitation that should be kept in mind when using these findings. Nevertheless, some of the
identified domains already give clear functional indications and a protein of interest can then be
analysed in more detail using other algorithms in which case further domains might be identified.

A third limitation of our approach is that domains that were not annotated in experimentally
determined structures could not be found. Searching against predicted structures of well-studied and
annotated proteins might therefore expand the number of detected domains. Of course, with the
caveat that predicted structures harbour some uncertainty.

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Finally, the identified domains were not all computationally and none were experimentally validated
in this study. Computational validation could be achieved by re-assessing the detected similarities
using a second algorithm. This could for instance be the cealign command in Pymol (Figure 1B)
followed by confirming or rejecting annotations based on a cut-off score, as we have done previously
(Schmidt et al., 2022). It could also be done with the TM-align algorithm which has a pre-defined cut-
off (Zhang, 2005) and was here used to validate the lowest 5% of the hits from this work. This
confirmed 14 of 16 hits, overall giving a good credibility for the detected similarities, particularly as it
can be assumed that the hits with higher scores are more reliable. Experimental validation is the gold
standard for any structure prediction (Sanderson & Rayner, 2017). The assessment of available data
in published literature on eight proteins annotated here, suggests that the majority of similarities
found might align with the function that can be inferred from the domain similarities. However, a
case-by-case validation is the only way to know whether the domains were identified correctly and
whether these domains serve the same function in *P. falciparum* as in other organisms. We did not
embark on a systematic experimental validation as this would take a considerable amount of time
and we believe that rapidly providing these search results to the wider community will be more
beneficial.
298
299   In conclusion, the results presented here are an example for the power of sequence-independent
300   structure comparison approaches for reducing the number of genes lacking annotation and
301   information on their potential biological function. Here applied to *P. falciparum*, it sheds light on the
302   biology of the malaria parasite and provides starting points for future research which will lead to a
303   better understanding of the biology of this pathogen.
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306   **Methods**
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Open-ended search
Lists of genes were downloaded from PlasmoDB v61 based on genomic location. All genes which
contained the word "unknown" in the gene name, for example "conserved Plasmodium protein,
unknown function", were included in the analysis. Pseudogenes were excluded.
Available Alphafold (alphafold.ebi.ac.uk, accessed December 2022-Mai 2023) (Jumper et al., 2021)
structures were visually assessed for a globular and compact fold which is a prerequisite for a
successful DALI search (Holm, 2022). The suitable Alphafold structures were submitted to a DALI PDB
search as available in the web application (http://ekhidna2.biocenter.helsinki.fi/dali/, accessed
December 2022-Mai 2023). The matches against PDB90 were analysed. The hits with the highest Z-
scores were visually assessed for suitable alignment (excluded alignment of just 2 or fewer α-helices
or β-strands) in the DALI 3D visualization tool. Of the top seven to ten hits with good alignment, the
Pfam 35.0 (Mistry et al., 2021) domain annotations were viewed in the DALI PFAM tool. A domain
annotation for a region of a PDB hit that aligned to the query Alphafold structure was considered a
reliable hit if there were no conflicting domain annotation. Domain annotations were considered
conflicting if other proteins with differently annotated domains aligned to the same amino acid
residues and had a Z-score within 1.0 of the protein harbouring the domain of interest.
324
Validation with TM-align

The 5% of domains with the lowest Z-score resulting from the open-ended DALI search were further
validated. For each of these, the PDB structure of the top hit of the DALI search was cropped to only
contain the residues annotated as the domain for which similarity was found. Where the top hit did
not contain this annotation, the highest-scoring PDB structure with this annotation was used. The
Alphafold structure was compared with this cropped PDB structure using TM-align (Zhang, 2005) as
available in the web application (https://seq2fun.dcmb.med.umich.edu//TM-align/, accessed
December 2022-Mai 2023). The TM-score based on the length of the cropped PDB structure was
assessed.
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ASA2/ASA3 search
All protein structures predicted for *P. falciparum* by AlphaFold (Jumper et al., 2021) were searched
using ASA2 residues 1-326 and full length ASA3 (chain 2 and 3 of PDB 6rd4 (Murphy et al., 2019))
using the DALI AF search (Holm, 2022). Alignments were visually assessed for at least 3 aligned
armadillo repeats (helix pairs), resulting in all proteins with a Z-score larger or equal to 7.0 to be
included for further analysis.
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342 **Visualization and further assessment**
343 Protein structures were analysed and visualized using PyMol 2.4.0 (Schrödinger, USA). Alignments
344 were performed using the PyMol command cealign. Figures were arranged in CorelDraw X6-8.
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References
Amos, B., Aurrecoechea, C., Barba, M., Barreto, A., Basenko, E. Y., Bażant, W., Belnap, R., Blevins, A.
S., Böhme, U., Brestelli, J., Brunk, B. P., Caddick, M., Callan, D., Campbell, L., Christensen, M. B.,
Christophides, G. K., Crouch, K., Davis, K., DeBarry, J., … Zheng, J. (2022). VEuPathDB: the
eukaryotic pathogen, vector and host bioinformatics resource center. *Nucleic Acids Research*,
50(D1), D898–D911. https://doi.org/10.1093/nar/gkab929

Aurrecoechea, C., Brestelli, J., Brunk, B. P., Dommer, J., Fischer, S., Gajria, B., Gao, X., Gingle, A.,
Grant, G., Harb, O. S., Heiges, M., Innamorato, F., Iodice, J., Kissinger, J. C., Kraemer, E., Li, W.,
Miller, J. A., Nayak, V., Pennington, C., … Wang, H. (2009). PlasmoDB: a functional genomic
database for malaria parasites. *Nucleic Acids Research*, *37*(Database), D539–D543.
https://doi.org/10.1093/nar/gkn814

Birnbaum, J., Flemming, S., Reichard, N., Soares, A. B., Mesén-ramírez, P., Jonscher, E., Bergmann, B.,
& Spielmann, T. (2017). A genetic system to study Plasmodium falciparum protein function.
Nature Methods, *14*(4), 450–456. https://doi.org/10.1038/nmeth.4223

Böhme, U., Otto, T. D., Sanders, M., Newbold, C. I., & Berriman, M. (2019). Progression of the
canonical reference malaria parasite genome from 2002–2019. *Wellcome Open Research*, *4*, 58.
https://doi.org/10.12688/wellcomeopenres.15194.2

Burley, S. K., Berman, H. M., Bhikadiya, C., Bi, C., Chen, L., Costanzo, L. Di, Christie, C., Duarte, J. M., Dutta, S., Feng, Z., Ghosh, S., Goodsell, D. S., Green, R. K., Guranovic, V., Guzenko, D., Hudson, B. P., Liang, Y., Lowe, R., Peisach, E., … Ioannidis, Y. E. (2019). Protein Data Bank: the single global archive for 3D macromolecular structure data. *Nucleic Acids Research*, *47*(D1), D520–D528. https://doi.org/10.1093/nar/gky949

Choi, H.-J., Gross, J. C., Pokutta, S., & Weis, W. I. (2009). Interactions of Plakoglobin and β-Catenin with Desmosomal Cadherins. *Journal of Biological Chemistry*, *284*(46), 31776–31788. https://doi.org/10.1074/jbc.M109.047928

Choi, H.-J., & Weis, W. I. (2005). Structure of the Armadillo Repeat Domain of Plakophilin 1. *Journal of Molecular Biology*, *346*(1), 367–376. https://doi.org/10.1016/j.jmb.2004.11.048

Dubois, D. J., Chehade, S., Marq, J.-B., Venugopal, K., Maco, B., Puig, A. T. I., Soldati-Favre, D., & Marion, S. (2023). *Toxoplasma gondii* HOOK-FTS-HIP Complex is Critical for Secretory Organelle Discharge during Motility, Invasion, and Egress. *MBio*. https://doi.org/10.1128/mbio.00458-23

Frankel, M. B., & Knoll, L. J. (2009). The Ins and Outs of Nuclear Trafficking: Unusual Aspects in Apicomplexan Parasites. *DNA and Cell Biology*, *28*(6), 277–284. https://doi.org/10.1089/dna.2009.0853

Fritz, J., Strehblow, A., Taschner, A., Schopoff, S., Pasierbek, P., & Jantsch, M. F. (2009). RNA-Regulated Interaction of Transportin-1 and Exportin-5 with the Double-Stranded RNA-Binding Domain Regulates Nucleocytoplasmic Shuttling of ADAR1. *Molecular and Cellular Biology*, *29*(6), 1487–1497. https://doi.org/10.1128/MCB.01519-08

Geiger, M., Brown, C., Wichers, J. S., Strauss, J., Lill, A., Thuenauer, R., Liffner, B., Wilcke, L., Lemcke, S., Heincke, D., Pazicky, S., Bachmann, A., Löw, C., Wilson, D. W., Filarsky, M., Burda, P.-C., Zhang, K., Junop, M., & Gilberger, T. W. (2020). Structural Insights Into PfARO and Characterization of its Interaction With PfAIP. *Journal of Molecular Biology*, *432*(4), 878–896. https://doi.org/10.1016/j.jmb.2019.12.024

Gibrat, J.-F., Madej, T., & Bryant, S. H. (1996). Surprising similarities in structure comparison. *Current Opinion in Structural Biology*, *6*(3), 377–385. https://doi.org/10.1016/S0959-440X(96)80058-3

Henrici, R. C., Edwards, R. L., Zoltner, M., van Schalkwyk, D. A., Hart, M. N., Mohring, F., Moon, R. W., Nofal, S. D., Patel, A., Flueck, C., Baker, D. A., John, A. R. O., Field, M. C., & Sutherland, C. J. (2020). The plasmodium falciparum artemisinin susceptibility-associated ap-2 adaptin μ subunit is clathrin independent and essential for schizont maturation. *MBio*, *11*(1), 1–16. https://doi.org/10.1128/mBio.02918-19

Hillebrand, A., Matz, J. M., Almendinger, M., Müller, K., Matuschewski, K., & Schmitz-Linneweber, C. (2018). Identification of clustered organellar short (cos) RNAs and of a conserved family of organellar RNA-binding proteins, the heptatricopeptide repeat proteins, in the malaria parasite. *Nucleic Acids Research*, *46*(19), 10417–10431. https://doi.org/10.1093/nar/gky710

Hollin, T., Jaroszewski, L., Stajich, J. E., Godzik, A., & Le Roch, K. G. (2021). Identification and phylogenetic analysis of RNA binding domain abundant in apicomplexans or RAP proteins. *Microbial Genomics*, *7*(3). https://doi.org/10.1099/mgen.0.000541

Holm, L. (2022). Dali server: structural unification of protein families. *Nucleic Acids Research*, *50*(W1), W210–W215. https://doi.org/10.1093/nar/gkac387

Ishiyama, N., Lee, S.-H., Liu, S., Li, G.-Y., Smith, M. J., Reichardt, L. F., & Ikura, M. (2010). Dynamic and
Static Interactions between p120 Catenin and E-Cadherin Regulate the Stability of Cell-Cell
Adhesion. *Cell*, *141*(1), 117–128. https://doi.org/10.1016/j.cell.2010.01.017

Jacot, D., Tosetti, N., Pires, I., Stock, J., Graindorge, A., Hung, Y.-F., Han, H., Tewari, R., Kursula, I., &
Soldati-Favre, D. (2016). An Apicomplexan Actin-Binding Protein Serves as a Connector and Lipid
Sensor to Coordinate Motility and Invasion. *Cell Host & Microbe*, *20*(6), 731–743.
https://doi.org/10.1016/j.chom.2016.10.020

Jakel, S. (1998). Importin beta , transportin, RanBP5 and RanBP7 mediate nuclear import of
ribosomal proteins in mammalian cells. *The EMBO Journal*, *17*(15), 4491–4502.
https://doi.org/10.1093/emboj/17.15.4491

Jones, D. T., & Thornton, J. M. (2022). The impact of AlphaFold2 one year on. *Nature Methods*, *19*(1),
15–20. https://doi.org/10.1038/s41592-021-01365-3

Jumper, J., Evans, R., Pritzel, A., Green, T., Figurnov, M., Ronneberger, O., Tunyasuvunakool, K.,
Bates, R., Žídek, A., Potapenko, A., Bridgland, A., Meyer, C., Kohl, S. A. A., Ballard, A. J., Cowie,
A., Romera-Paredes, B., Nikolov, S., Jain, R., Adler, J., … Hassabis, D. (2021). Highly accurate
protein structure prediction with AlphaFold. *Nature*, *596*(7873), 583–589.
https://doi.org/10.1038/s41586-021-03819-2

Khosh-Naucke, M., Becker, J., Mesén-Ramírez, P., Kiani, P., Birnbaum, J., Fröhlke, U., Jonscher, E.,
Schlüter, H., & Spielmann, T. (2018). Identification of novel parasitophorous vacuole proteins in
P. falciparum parasites using BioID. *International Journal of Medical Microbiology*, *308*(1), 13–
24. https://doi.org/10.1016/j.ijmm.2017.07.007

Kimmel, J., Schmitt, M., Sinner, A., Jansen, P. W. T. C., Mainye, S., Ramón-Zamorano, G., Toenhake, C.
G., Wichers-Misterek, J. S., Cronshagen, J., Sabitzki, R., Mesén-Ramírez, P., Behrens, H. M.,
Bártfai, R., & Spielmann, T. (2023). Gene-by-gene screen of the unknown proteins encoded on
Plasmodium falciparum chromosome 3. *Cell Systems*, *14*(1), 9-23.e7.
https://doi.org/10.1016/j.cels.2022.12.001

Krissinel, E., & Henrick, K. (2004). Secondary-structure matching (SSM), a new tool for fast protein
structure alignment in three dimensions. *Acta Crystallographica Section D Biological
Crystallography*, *60*(12), 2256–2268. https://doi.org/10.1107/S0907444904026460

Kufareva, I., & Abagyan, R. (2011). *Methods of Protein Structure Comparison* (pp. 231–257).
https://doi.org/10.1007/978-1-61779-588-6_10

Kumar, A., Vadas, O., Dos Santos Pacheco, N., Zhang, X., Chao, K., Darvill, N., Rasmussen, H. Ø., Xu, Y.,
Lin, G. M.-H., Stylianou, F. A., Pedersen, J. S., Rouse, S. L., Morgan, M. L., Soldati-Favre, D., &
Matthews, S. (2023). Structural and regulatory insights into the glideosome-associated
connector from Toxoplasma gondii. *ELife*, *12*. https://doi.org/10.7554/eLife.86049

Li, X., Chen, H., Bahamontes-Rosa, N., Kun, J. F. J., Traore, B., Crompton, P. D., & Chishti, A. H. (2009).
Plasmodium falciparum signal peptide peptidase is a promising drug target against blood stage
malaria. *Biochemical and Biophysical Research Communications*, *380*(3), 454–459.
https://doi.org/10.1016/j.bbrc.2009.01.083

Mistry, J., Chuguransky, S., Williams, L., Qureshi, M., Salazar, G. A., Sonnhammer, E. L. L., Tosatto, S.
C. E., Paladin, L., Raj, S., Richardson, L. J., Finn, R. D., & Bateman, A. (2021). Pfam: The protein

families database in 2021. *Nucleic Acids Research*, *49*(D1), D412–D419.
https://doi.org/10.1093/nar/gkaa913

Murphy, B. J., Klusch, N., Langer, J., Mills, D. J., Yildiz, Ö., & Kühlbrandt, W. (2019). Rotary substrates
of mitochondrial ATP synthase reveal the basis of flexible F_1-F_o coupling. *Science*, *364*(6446).
https://doi.org/10.1126/science.aaw9128

Sanderson, T., & Rayner, J. C. (2017). PhenoPlasm: a database of disruption phenotypes for malaria
parasite genes. *Wellcome Open Research*, *2*, 45.
https://doi.org/10.12688/wellcomeopenres.11896.2

Sayers, C. P., Mollard, V., Buchanan, H. D., McFadden, G. I., & Goodman, C. D. (2018). A genetic
screen in rodent malaria parasites identifies five new apicoplast putative membrane
transporters, one of which is essential in human malaria parasites. *Cellular Microbiology*, *20*(1).
https://doi.org/10.1111/cmi.12789

Schmidt, S., Wichers-misterek, J. S., Behrens, H. M., Birnbaum, J., Henshall, I., Jonscher, E., Flemming,
S., & Spielmann, T. (2022). The Kelch13 compartment is a hub of highly divergent vesicle
trafficking proteins in malaria parasites. *BioRxiv [Preprint]*.
https://doi.org/https://doi.org/10.1101/2022.12.15.520209

Schwach, F., Bushell, E., Gomes, A. R., Anar, B., Girling, G., Herd, C., Rayner, J. C., & Billker, O. (2015).
PlasmoGEM, a database supporting a community resource for large-scale experimental genetics
in malaria parasites. *Nucleic Acids Research*, *43*(D1), D1176–D1182.
https://doi.org/10.1093/nar/gku1143

Shah, S., Lee, S.-F., Tabuchi, K., Hao, Y.-H., Yu, C., LaPlant, Q., Ball, H., Dann, C. E., Südhof, T., & Yu, G.
(2005). Nicastrin Functions as a γ-Secretase-Substrate Receptor. *Cell*, *122*(3), 435–447.
https://doi.org/10.1016/j.cell.2005.05.022

Shindyalov, I. N., & Bourne, P. E. (1998). Protein structure alignment by incremental combinatorial
extension (CE) of the optimal path. *Protein Engineering Design and Selection*, *11*(9), 739–747.
https://doi.org/10.1093/protein/11.9.739

Sievers, F., & Higgins, D. G. (2018). Clustal Omega for making accurate alignments of many protein
sequences. *Protein Science*, *27*(1), 135–145. https://doi.org/10.1002/pro.3290

Subramaniam, S., & Kleywegt, G. J. (2022). A paradigm shift in structural biology. *Nature Methods*,
19(1), 20–23. https://doi.org/10.1038/s41592-021-01361-7

Tang, Y., Meister, T. R., Walczak, M., Pulkoski-Gross, M. J., Hari, S. B., Sauer, R. T., Amberg-Johnson,
K., & Yeh, E. (2019). A mutagenesis screen for essential plastid biogenesis genes in human
malaria parasites. *PLOS Biology*, *17*(2), e3000136.
https://doi.org/10.1371/journal.pbio.3000136

Terwilliger, T., Liebschner, D., Croll, T. I., Williams, C. J., McCoy, A. J., Poon, B. K., Afonine, P. V.,
Oeffner, R. D., Richardson, J. S., Read, R. J., & Adams, P. D. (2022). AlphaFold predictions: great
hypotheses but no match for experiment. *BioRxiv*. https://doi.org/10.1101/2022.11.21.517405

The Plasmodium Genome Database Collaborative. (2001). PlasmoDB: An integrative database of the
Plasmodium falciparum genome. Tools for accessing and analyzing finished and unfinished
sequence data. *Nucleic Acids Research*, *29*(1), 66–69. https://doi.org/10.1093/nar/29.1.66

van Kempen, M., Kim, S. S., Tumescheit, C., Mirdita, M., Lee, J., Gilchrist, C. L. M., Söding, J., &
Steinegger, M. (2022). Fast and accurate protein structure search with Foldseek. *BioRxiv*.
https://www.biorxiv.org/content/10.1101/2022.02.07.479398v5

World Health Organization. (2022). *World malaria report 2022*.
https://www.who.int/publications/i/item/9789240064898

Xie, T., Yan, C., Zhou, R., Zhao, Y., Sun, L., Yang, G., Lu, P., Ma, D., & Shi, Y. (2014). Crystal structure of
the γ-secretase component nicastrin. *Proceedings of the National Academy of Sciences of the*
United States of America, *111*(37), 13349–13354. https://doi.org/10.1073/pnas.1414837111

Xu, X., Yamasaki, H., Feng, Z., & Aoki, T. (2002). Molecular cloning and characterization of
Plasmodium falciparum transportin. *Parasitology Research*, *88*(5), 391–394.
https://doi.org/10.1007/s00436-001-0578-z

Yam, X. Y., Birago, C., Fratini, F., Di Girolamo, F., Raggi, C., Sargiacomo, M., Bachi, A., Berry, L., Fall, G.,
Currà, C., Pizzi, E., Breton, C. B., & Ponzi, M. (2013). Proteomic Analysis of Detergent-resistant
Membrane Microdomains in Trophozoite Blood Stage of the Human Malaria Parasite
Plasmodium falciparum. *Molecular & Cellular Proteomics*, *12*(12), 3948–3961.
https://doi.org/10.1074/mcp.M113.029272

Zhang, Y. (2005). TM-align: a protein structure alignment algorithm based on the TM-score. *Nucleic*
Acids Research, *33*(7), 2302–2309. https://doi.org/10.1093/nar/gki524

Zuegge, J., Ralph, S., Schmuker, M., McFadden, G. I., & Schneider, G. (2001). Deciphering apicoplast
targeting signals – feature extraction from nuclear-encoded precursors of Plasmodium
falciparum apicoplast proteins. *Gene*, *280*(1–2), 19–26. https://doi.org/10.1016/S0378-
1119(01)00776-4

Figure 1

A

all proteins of
P. falciparum 3D7
5791

↓

name contains word
"unknown"
1407

↓ visual
assessment

AF-prediction contains
globular folds
931

↓ DALI search
against PDB

↓ exclude alignments to 2 or
fewer α-helices/β-strands

proteins with 1 or more
domains identified
288

B

Z-score 36.3
RMSD 3.4 Å over 330 aa
PF3D7_1112800 vs.
DUF885 (7Z5G)

Z-score 33.8
RMSD 3.5 Å over 152 aa
PF3D7_1132400 vs.
CRT-like (6UKJ)

Z-score 33.0
RMSD 3.9 Å over 304 aa
PF3D7_0716500 vs.
Amino_oxidase (2IVE)

Z-score 32.0
RMSD 3.8 Å over 272 aa
PF3D7_1026200 vs.
THUMP & UPF0020 (7Z5G)

Z-score 31.2
RMSD 2.7 Å over 152 aa
PF3D7_1351200 vs.
Glyco_hydro_35 (3W5G)

Z-score 31.2
RMSD 3.4 Å over 440 aa
PF3D7_1475200 vs.
DEAD, Helicase_C, Ha2 &
OB_NTP_bind (6I3P incl. RNA)

C

Number of proteins

257
25
5
1

Number of detected
domains per protein

D

# of times domain
was detected

C2, Wd40, Pentapeptide repeats, AB_hydrolase, RRM_1, ABM, Arm, xRRM, Methyltransferase, S1 RNA binding, Hsp20, Leucine Rich repeat, FKBP_C, Gal-lectin, DEAD

Most commonly detected domains

Figure 1: Identification of domains based on similarity of Alphafold prediction to PDB structures.
(A) Pipeline for detection of domains in proteins of unknown function. (B) Alignment of Alphafold
prediction (beige) to experimentally determined structured (blue) with domain annotation, with Z-
scores >30. Indicated are the Z-score resulting from the DALI search, the root mean square deviation
(RMSD) generated by Pymol cealign, the PlasmoDB gene accession number of the gene encoding the
P. falciparum protein, the name(s) of the domain(s) in the experimentally determined structure
according to Pfam and the PDB accession (blue) of the experimentally determined structure. (C)
Number of proteins in which one, two, three or four domains were detected by the open-ended
search. (D) Most commonly detected domains shown with number of times they were detected.
Domain names as they appear in Pfam.
527

Figure 2: TM-align assessment of lowest-scoring 5% of aligned domains. The Alphafold structure of the *P. falciparum* protein (beige) is aligned to the indicated residues of the PDB structure (blue), which corresponds to the annotated domain. Given are TM-score generated by TM-align, Z-score generated by DALI, RMSD generated by Pymol cealign, PlasmoDB gene ID of *P. falciparum* gene that encodes the respective protein, name of the identified domain of similarity, the PDB ID of the structure that contained the domain and the amino acid residues at which the domain was found. For 3FWB-A residues 18 to 92 are shown even though the annotated domain only including residues 24-52 because structural similarity extended to this region. TM-scores under 0.5 are shown in red.

Figure 3

Figure 3: Identification of ASA2/3-like armadillo domains in *P. falciparum* proteins using Alphafold

predictions. (A) Pipeline for detection of ASA2/3-like armadillo domains. (B) Annotations of *P.*

falciparum proteins which were found to be similar to ASA2 and ASA3. (C) Correspondence analysis

to cluster annotated *P. falciparum* proteins with similarity to ASA2 and ASA3 by structural similarity

to each other. Dots corresponding to proteins shown in (D) and (E) are labelled. Purple dots do not

belong to cluster 1 or 2. (D) Three representative structures selected from cluster 1 as defined in (C)

aligned to each other using cealign in Pymol. (E) Three representative structures selected from
cluster 2 as defined in (C). Structures shown in (D) and (E) were used to sort *P. falciparum* proteins of
unknown function with similarity to ASA2 and ASA3 into clusters 1 and 2. (F) Number of proteins of
different functions in cluster 1 and 2.
551

Supplementary Figure 1: Nicastrin domains detected in PF3D7_1013300. (A) Schematic representation of *Dictyostelium purpureum* nicastrin and PF3D7_1013300 (not to scale). Grey elements show corresponding domains. Z-score resulting from DALI score is indicated. Secondary structure elements of *D. purpureum* nicastrin that are not present in the crystal structure PDB 4R12 are shown with dashed lines. SP, predicted signal peptide; TM, transmembrane domain. (B) Crystal structure of *D. purpureum* nicastrin (PDB 4R12). Residues coloured as in (A). (C) Alphfold structure prediction of PF3D7_1013300. Residues coloured as in (A). (D) *D. purpureum* nicastrin (PDB 4R12, blue) and Alphfold structure prediction of PF3D7_1013300 (beige) aligned to each other using cealign in Pymol. RMSD generated by Pymol cealign is indicated.

565   Supplementary Figure 2

566
Supplementary Figure 2: Spatial overlap of ASA2 with hits identified in open-ended domain search.
Alphafold structure predictions of proteins for which domains were found in the open-ended search
and in the ASA2/ASA3-based search are shown (beige), and aligned to ASA2 residues 1-326 from PDB
6RK4 (red) and to the highest-scoring domain-annotated hit of the open-ended search (blue). For
PF3D7_1207900 only ASA2 residues 1-170 are shown for clarity. For PF3D7_0104600 and
PF3D7_1207900 the domains from the open-ended search (both DUF559, blue) and the armadillo
domain (red) align to different regions of the target protein. For the other four structures the
domains align to the same region of the target protein.

Supplementary Figure 3

Supplementary Figure 3: Structure based clustering of unknown proteins with ASA2/ASA3-like

armadillo domains by DALI all-against-all search. Proteins of known functions which are

representative of cluster 1 are labelled and highlighted in blue, those representative of cluster 2 are

labelled and highlighted in yellow. Proteins considered to belong to the same cluster as the

representative proteins are framed in the same color. (A) Batch 1. (B) Batch 2. (C) Batch 3.