Configurable Full Subtractor and Adder by Lego-like Mixing and Matching of Engineered Bacteria

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Abstract

One of the long-term goals of synthetic bioengineering is to create configurable and programmable biological systems by just mixing and matching "LEGO"-like bio-modules. Here, we introduce a configurable and modular multi-cellular system where, from a small library of nine discrete engineered bacterial cells, a full subtractor and a full adder can be built on demand by just mixing and matching seven appropriate cell types in a culture. Here, each set of engineered bacteria was modelled as an ‘artificial neuro-synapse’ that, in a co-culture, formed a single layer artificial neural network (ANN) type architecture that worked as a biochemical full subtractor or full adder. The system is configurable with interchangeable cellular modules, whereby through simply interchanging two cell types in the subtractor culture, a full adder can be built and vice versa. This Lego-like mix and match system is mathematically predictive, and provide a flexible and scalable means to build complex cellular functions. This work may have significance in biocomputer technology development, multi-cellular synthetic biology, and cellular hardware for ANN.

Introduction

Performing computations with engineered living cells has enormous importance in biocomputer technology development at the micron scale [1-3], where microprocessor-based computers have limitations due to energy, cost, and technological constraints [3]. However, the success of biocomputing technology depends on expanding the horizons of implementing complex computational device functions in living cells. Such system designs [4-6] follow the hierarchical electronic design principle, where the component genetic logic gates are layered according to the electronic analogous design either in a single cell [7-9] or in a multicellular distributed format [10-12]. However, layering multiple logic gates in a single cell result in crosstalk among various parts and hinders building complex functionality [11, 12]. Multicellular distributed computing has been adopted to build more complex functions, where various engineered cells carry a part of the integrated circuits and are connected with other cells through engineered chemical communications [10-12]. As each engineered cell is physically insulated from the other engineered cells, this reduces the crosstalk. Such strategies were used to build integrated logic circuits [10] and maze-solving biocomputers[13] in bacteria and an adder [11] and analog-to-digital converter[12] in mammalian cells.

However, such systems are complicated to build, require multiple layers of various engineered cells, and depend on the availability of the orthogonal diffusible chemical communication channels. Further, they are nonconfigurable, mathematically unpredictable, and demonstrate a single function. One of the long-term goals of synthetic biology and bioengineering is to create configurable and programmable biological systems by just mixing and matching "Lego"-like bio modules to build multiple predictable biological functions [14].

Here we introduce a configurable and modular engineered multi-cellular system where, from a small library of nine discrete engineered bacterial cells, a full subtractor and a full adder can be built on demand by just mixing and matching seven appropriate cell types in a culture. The

system is configurable with interchangeable cellular modules, whereby by simply interchanging two cell types, a full subtractor culture can be reconfigured into that of a full adder and vice versa.

A full subtractor and a full adder form indispensable parts of modern ICs where they are found in the arithmetic logic units and the digital signal processing units to perform binary subtraction and binary addition respectively. A full subtractor is a function that performs subtraction of two bits, one is minuend and other is subtrahend, taking into account borrow of the previous adjacent lower minuend bit. The digit from which another digit is to be subtracted is known as minuend. Similarly, the number which is subtracted from the minuend is known as a subtrahend. It has three inputs, the minuend, subtrahend, and borrow-in from a previous full subtractor and two outputs, the difference and output borrow, respectively. The output borrow is connected as the input of a subsequent full subtractor. Full Adder is a function that adds three input bits and produces two outputs. The first two inputs get added along with a third input, that represents an input carry from a previous full adder. The outputs are the sum and a carry-out to the next adder. The output carry of a first adder is connected to the input of the second adder. Both the functions are complex to build with cells. No cellular subtractor has been realised yet. A mammalian multicellular full adder [11] required twenty-two component genetic logic circuits in nine different types of cells, which were layered according to the electronic circuit design and connected through engineered cellular communications.

To create two complex and challenging functions in an easy, configurable Lego-like format, we adapted the basic concepts of artificial neural network (ANN) architecture to represent engineered bacteria as artificial neuro-synapses or “bactoneurons” wherein a hetergenous population of such cells constitute a single layer ANN. Previously, we demonstrated [15,16] that such single-layer ANN-type architectures with engineered bacteria could generate multiple

computing functions like decoders, encoders, majority functions, Fredkin gates , and double Feynman gates.

Motivated by the success of this design concept, in this work, we built two single-layered Bactoneural-Networks (BNNs), each comprised of 7 bactoneruons formulated from a library of a just 9 bactoneuron types, which can implement two complex computing functions, a full subtractor and a full adder. The two BNN's differed from each other by 2 bactoneuron types and a simple re-configuration effected by interchanging these two, reprograms the entire bactoneural layer between two separate but equally complex computing functions. The reconfigurable full subtractor and adder made from engineered bacteria have several advantages: i) the system is configurable and the cells are interchangeable. It showed two independent functionalities by replacing just two bactoneurons: ii) the Lego-like wetware design is modular, simpler, and smaller, the system creates two complex and challenging functions in an easy format; iii) both the subtractor and adder were realised by 7 different cell populations arranged in a single layer, without the need of layering the cells by engineered chemical communications; iii) the behaviour of each module and the whole system can be described with mathematical equations of a single type, and each cellular type showed agreement between simulation and experiments; iv) robust rules for engineering such systems is in place; v) the design is general and can be built with any type of cells.

Results

Design of the configurable subtractor and adder in ANN type architecture

First, the subtractor and adder truth tables are mapped in a single-layer ANN-type architecture format (Fig. 1, Fig. S1) following a set of rules coherent with ANN [15]. In short, first we segregated each of the truth tables into its individual output channels and considered the relationship of that specific output with all the input combinations. Next, we grouped those combinations into smaller truth tables such that each input corresponding to a particular output

possess a weight with only one unique type of sign: ‘+," "-," or "0’. We now reduced the resultant truth table by ignoring any component with a weight value of "0’. If the output value 1 (true) in the smaller resultant truth table appeared only once, we terminated fragmenting the truth table. Otherwise, we kept fragmenting the truth table until the above condition appeared. The nine resultant truth tables represented the functional and mathematical characteristics of nine "artificial neuro-synapses" (Figure 1a), or unit bactoneurons, which need to be engineered as cellular devices. Together, the nine bactoneurons work as a configurable cellular system, such that when the corresponding seven unit bactoneurons are assembled into a single layered ANN-type architecture, both the subtractor and adder could be realised by interchanging two bactoneurons from a library of nine.

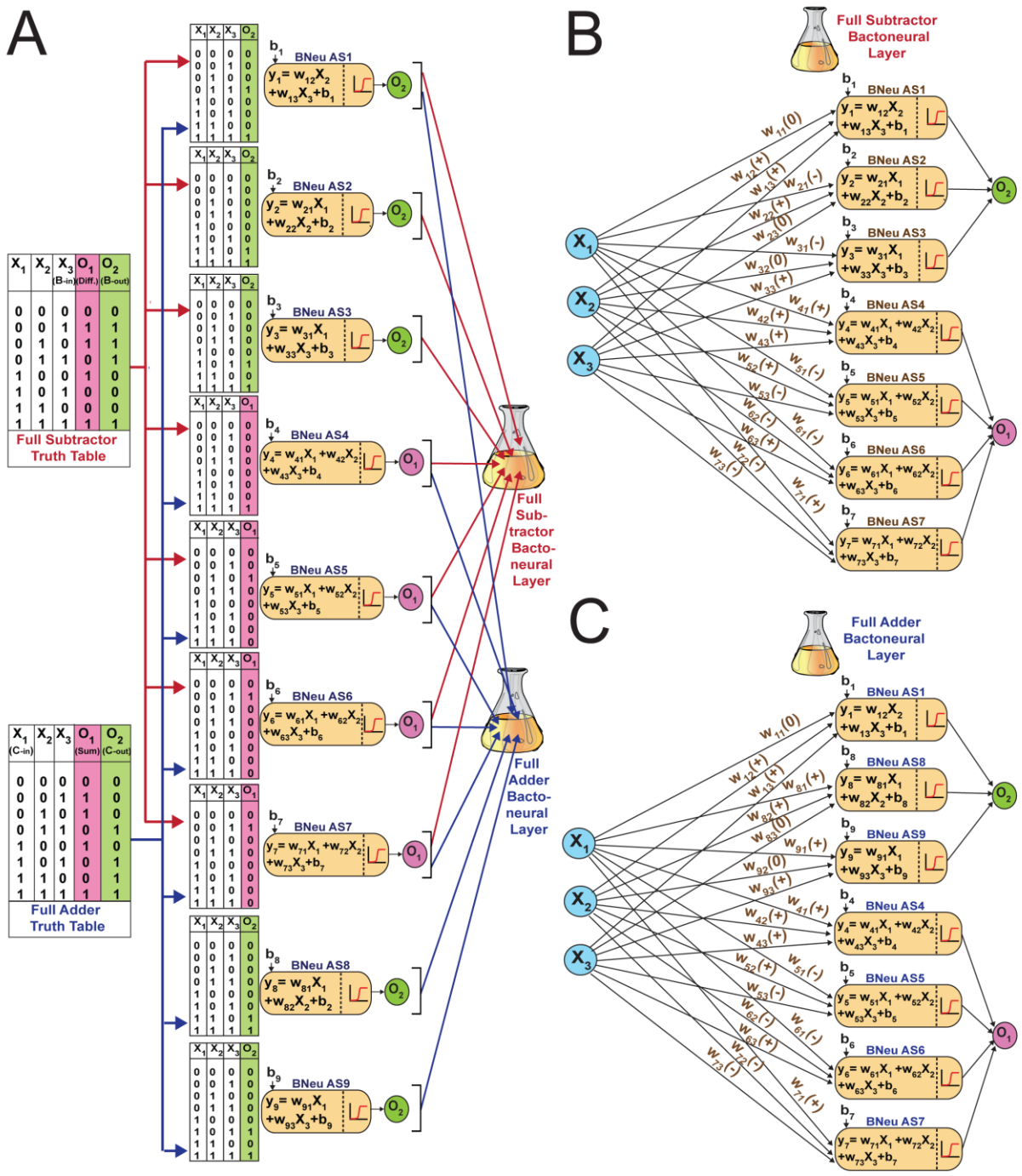

Figure 1: Fragmentation and mapping of the truth tables of Full Subtractor & Full Adder into configurable bactoneural layers. (A) The two functions Full Subtractor (above) and Full-adder (below) are first fragmented into smaller and simpler functions according to the rules of ANN and are represented by the vertical array of sub truth tables. The sub-functions generating out of full Subtractor are indicated through red connecting arrows and the ones generating out full adder are shown through blue connecting arrows. The corresponding output channels are shown in colour (O_1 is magenta and O_2 is green). These functions get mapped onto the Bactoneurons where the parameters of the summation function are implemented through and emanate from synthetic genetic circuits fabricated based on established design principles connecting ANN and transcriptional regulation. X_1, X_2 & X_3 represent the input channels, w_{ij} represents the weight of the jth input for the ith bactoneuron. b_i represents the bias of the ith

bactoneuron. The weighted summation function activates the activation function of the bactoneuron that respond through expression of fluorescent proteins that serve as output channels. Each bactoneuron is connected to any one output channel. Finally, populations of appropriate bactoneurons can be mixed and matched in a heterogenous culture to constitute bactoneural layers that can be configured according to the function desired. **(B)** Configuration of the bactoneural single layer network for performing full Subtractor function. The bactoneural layer composed 7 bactoneurons (AS-1-7) takes weighted inputs, combines them linearly in a summation function and responds through the output channels after activation through a sigmoidal function. **(C)** Configuration of the full adder bactoneural layer composed of Bactoneurons AS-1,4,5,6,7,8,9. The figure demonstrates how the re-configuration of the bactoneural layer by replacing just 2 bactoneurons out of seven, AS-2 & 3 with AS-8 & 9, reprograms the system from a full subtractor to a full adder.

Design of cellular devices and unit bactoneuron

The bactoneurons were constructed by building transcriptional synthetic genetic circuits in the bacteria (Fig. 2). The inputs X_1, X_2, and X_3 were replaced with three chemical inducers, N-acyl homoserine lactone (AHL) isopropyl b-D-1 thiogalactopyranoside (IPTG), and anhydrotetracycline (aTc) respectively. The outputs O_1 and O_2 were manifested with fluorescent proteins E2 crimson, and EGFP respectively (Figure 2). Synthetic genetic circuits inside bacteria process chemical inputs and produce output by expressing fluorescent proteins, such that the input-output relation mimics a conventional activation function[17] for an artificial neuro-synapse (eq 1). Thus, for a given bactoneuron j,

$$O_j=1(1+e-(X_1.w_{j1}+X_2.wj_2+ X_3.w_{j3}+ b_j)) \quad (1)$$

where O_j is the fluorescent protein output of the bactoneuron, X_i represents the input inducer concentration, w_{ji} represents the weight of the corresponding input X_i, and b_j represents the bias for the bactoneuron j. We designed the molecular-genetic circuits of all the bactoneurons based on the signs of the 'weight' parameter in activation function equations for the corresponding bactoneuron, (Fig. 2). The positive and negative weight values in an activation function of a bactoneuron respectively correspond to the activation and repression of output

reporter gene expression mediated by a specific inducer. Zero weight corresponds to an invariant relation.

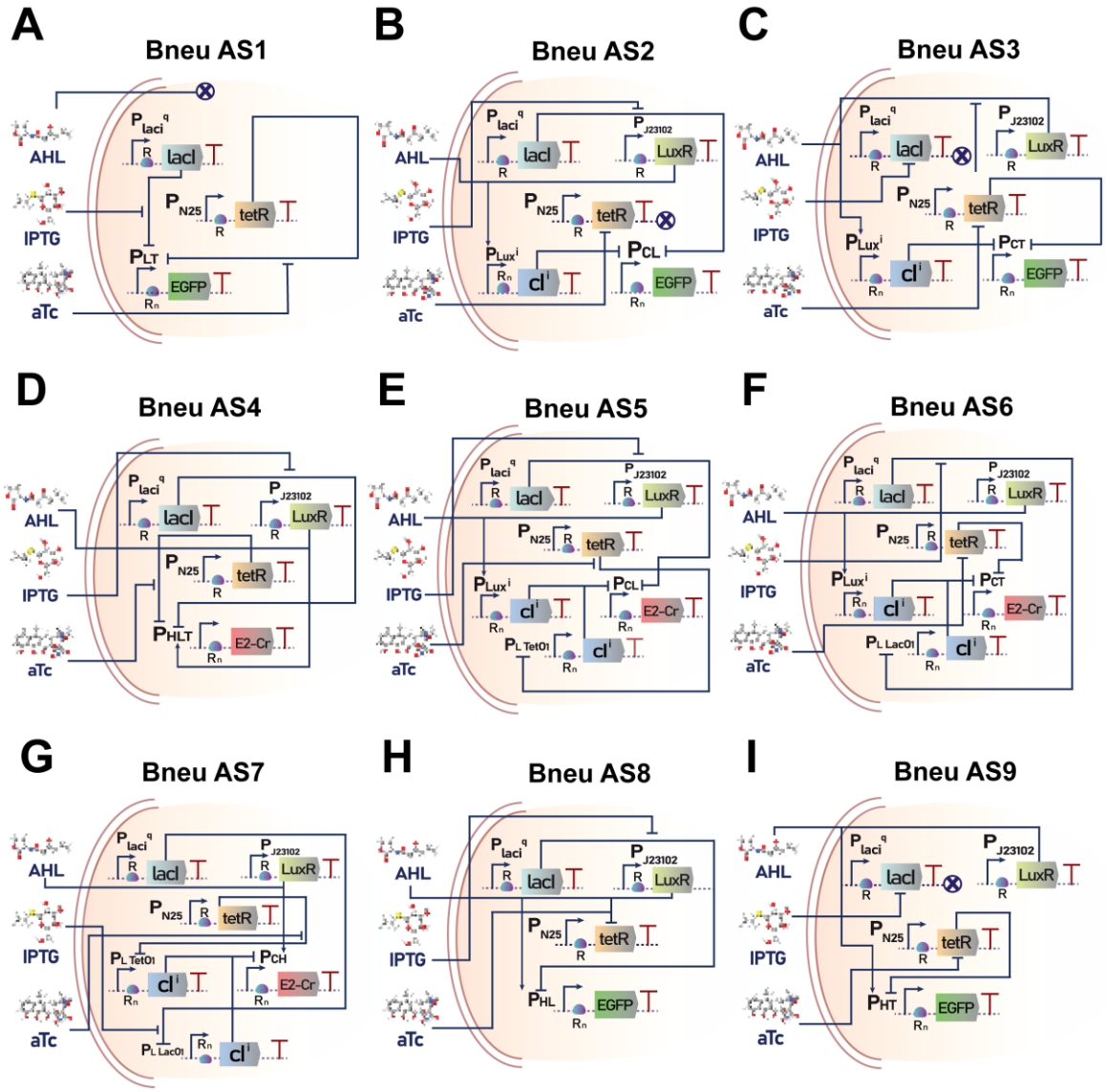

Figure 2: Generic maps of the synthetic genetic networks that encode corresponding specific functions to each bactoneuron type. (A) Bneu AS1, **(B)** Bneu AS2, **(C)** Bneu AS3, **(D)** Bneu AS4, **(E)** Bneu AS5, **(F)** Bneu AS6, **(G)** Bneu AS7, **(H)** Bneu AS8 and **(I)** Bneu AS9. The three abstract inputs X1, X2 & X3 are served by extracellularly provided AHL, IPTG & aTc respectively. Abstract outputs O1 and O2 are served by E2Crimson & EGFP respectively. In different combinations of the three inputs, they influence the transcriptional machinery of the synthetic genetic circuitry by interacting with transcription factors LuxR, LacI & TetR respectively by either activating the factor (LuxR) or deactivating it (LacI & tetR) through conformational changes. The sensitivity of the interaction of the input inducer with its cognate transcription factor was equated with the weight of the input and the process of interaction could be seen as the execution of the summation function. This interaction affects the transcription factor's binding affinity with its respective operator sites on the sequence of

various promoters within the genetic circuit in a non-linear fashion (activation by LuxR or repression by LacO1 or TetO2) thereby enabling the implementation of the activation functions. Weight and bias adjustment was mainly effected through iterative changes at different bioparts. R refers to RBSs that were remained constant and was not part of iterative updates. R_n refers to RBSs that were part for the iterative process where "n" corresponds to RBS number. cI^i refers to either of 2 variants of the lambda repressor proteins cI^w or cI^* which were interchanged as part of iterations. cI^w refers to the wild type version of cI whereas cI* refers to the frameshifted version[18]. $Plux^i$ refers to promoters $Plux^w$ or $Plux^*$ which were interchanged across iterations. We also iteratively modified the relative copy numbers of different gene expression cassettes by changing the origin of replication of the plasmids they were present in (not shown in figure). Gene expression cassettes that do not have any downstream effect on the output and are associated with "0" weight inputs are shown with their interaction lines terminating in a **"X"** sign.

The transcriptional genetic circuits (Fig 2) were built based on a set of synthetic promoters on which transcription factors LuxR, LacR, TetR, and Ci [18] bind to either individually or simultaneously to hinder and/or activate the transcriptions. The transcription factors, promoters, and their interactions formed various feed-forward, feedback, and combination mechanisms based on the bactoneuron circuit designs (Fig. 2). The mode of transcription was controlled by the interactions of the transcription factors with appropriate input chemicals (AHL, IPTG, and aTc) through allosteric regulation. The quantitative relation between inputs and outputs within a proper parameter range should demonstrate appropriate bactoneuron behaviour.

Construction, characterization, and optimization of bactoneurons

Among the nine bactoneurons required to configure a subtractor or adder, the basic form for five two-input bactoneurons (BNeu AS1, AS2, AS3, AS8, and AS9) were available from a bactoneuron library we made earlier [15,16]. The rest of the four bactoneurons (BNeu AS4, AS5, AS6, and AS7) are three input systems that were built from scratch. We first constructed the initial set of molecular constructs according to the design (Fig. 2) and inserted them into DH5αZ1 strains [19] of *E.coli*. Next, we measured fluorescence protein expression at '0' and

‘1’ (saturated) inducer concentrations (Figs. S2, S3, S4, and S5). These experiments gave us fold changes between the highest output signal (value "1") (F.C.) and the highest leakage, (where the leakage was defined as the basal level expression of the reporter fluorescent proteins under the input conditions, where output expression should be zero) total leakage from all possible inducer (0,1) combinations (Σ leakage, Σ L.), and variation in the highest signals among various bactoneurons (S.V). Further, we performed dose response experiments as a function of various inducer concentrations (Figs S2-S5) by varying one inducer concentration while keeping the others as constants (saturated or ‘0’ concentration, as the case may be). The dose response behaviours were fitted with the activation function equation (equation 1), and the weight and bias parameters were estimated from the fitted equations (Figs. S2-S5). The initial constructs for AS5, AS6, and AS7 showed suboptimal behaviour. They (AS5A, AS6A, and AS7A) generally had i) low fold changes (F.C.) between the highest output signal (value "1") and the highest leakage, ii) high total leakage (Σ L.) , iii) low weight values, and iv) high variability of the signals at ON states among various bactoneurons (S.V.) (Fig 3C). We defined/fixed a set of optimum values arbitrarily or based on previous observations [13,15] (Fig. 3A). The minimum fold change between the highest leakage and the ON state was set at 7. The maximum total leakage was set at 0.2 and 0.4 for 2-input and 3-input bactoneurons, respectively. The weight values define the sharpness of the transition from the OFF state to the ON state as a function of input. The minimum weight values were set as 8.5, 9.5, and 10.5 for input AHL, IPTG, and aTc, respectively. Further, we measured the signal strength at ON states of all bactoneurons expressing a single fluorescent protein (EGFP or E2-Crimson) and fixed the maximum variation to be within 20%. Though bactoneuron AS4 satisfied the optimal set points in its first construct, the initial constructs for AS5, AS6, and AS7 failed. The measurements performed on all the two input bactoneurons taken from the library (BNeu AS1, AS2, AS3, AS8, and AS9) showed optimum behaviour.

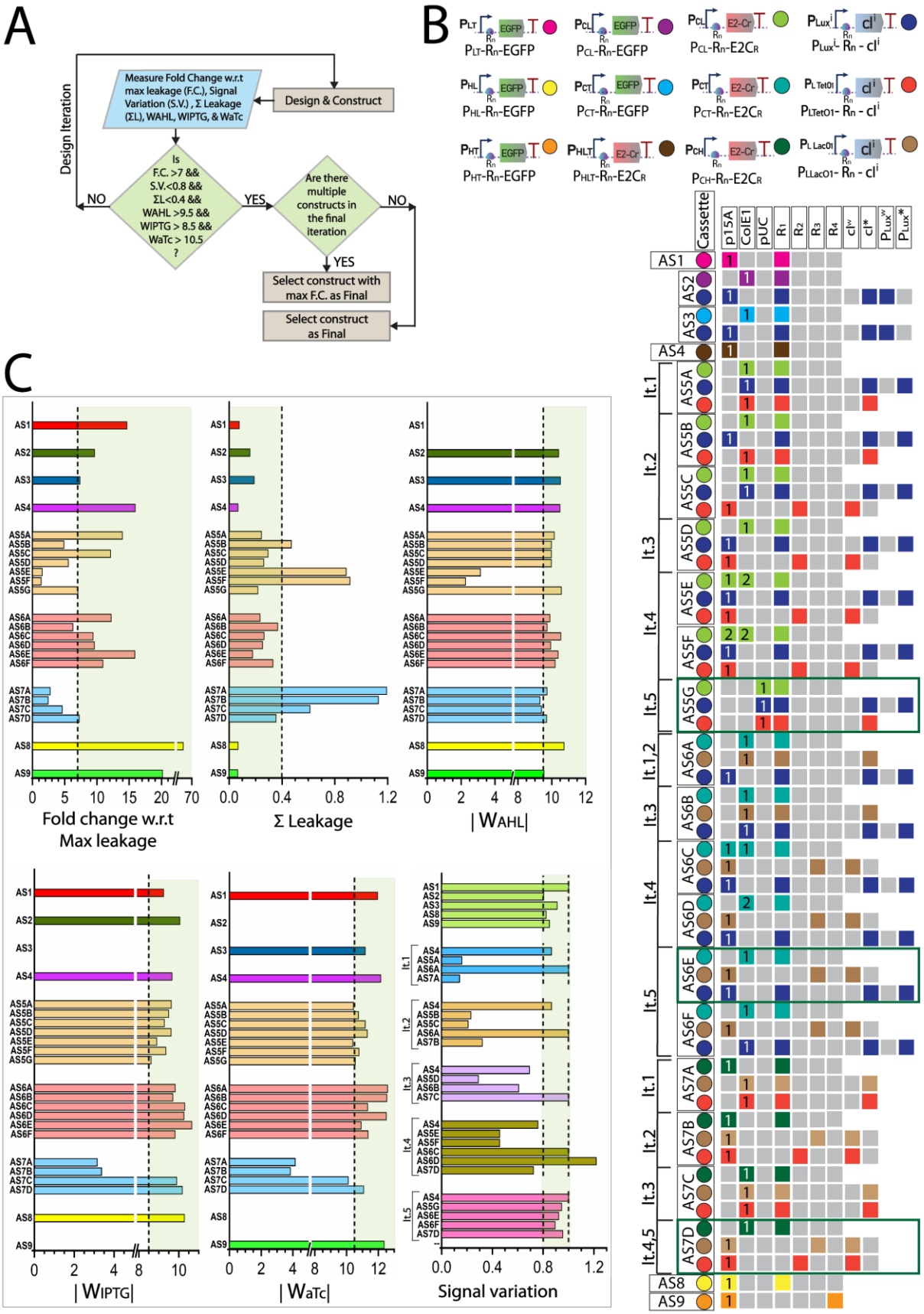
A
Design Iteration
Measure Fold Change w.r.t max leakage (F.C.), Signal Variation (S.V.), Σ Leakage (ΣL), WAHL, WIPTG, & WaTc
Design & Construct
Is F.C. >7 && S.V.<0.8 && ΣL<0.4 && WAHL >9.5 && WIPTG > 8.5 && WaTc > 10.5 ?
NO
YES
Are there multiple constructs in the final iteration
NO
YES
Select construct with max F.C. as Final
Select construct as Final
B
PLT-Rn-EGFP
PCL-Rn-EGFP
PCL-Rn-E2CR
PLux- Rn - cI
PHL-Rn-EGFP
PCT-Rn-EGFP
PCT-Rn-E2CR
PLTetO1- Rn - cI
PHT-Rn-EGFP
PHLT-Rn-E2CR
PCH-Rn-E2CR
PLLacO1- Rn - cI
Cassette
p15A
ColE1
pUC
R1
R2
R3
R4
cIw
cI*
PLuxw
PLux*
AS1
AS2
AS3
AS4
AS5A
AS5B
AS5C
AS5D
AS5E
AS5F
AS5G
AS6A
AS6B
AS6C
AS6D
AS6E
AS6F
AS7A
AS7B
AS7C
AS7D
AS8
AS9
It.1
It.2
It.3
It.4
It.5
It.1,2
It.4,5
C
Fold change w.r.t Max leakage
Σ Leakage
| WAHL|
| WIPTG|
| WaTc|
Signal variation

Figure 3: Details of the process of iterations to optimize Bactoneuron paramters. (A) Flowchart of the algorithm followed for bactoneural parameter optimization iterations. 6 principle parameters and their thresholds (shown) were set, which, when all satisfied, qualifies a bactoneuron contruct for incorporation in the configurable bactoneural layer**.** Iteration continues until such condition is achieved **(B)** The Biological design configuration space and details of the iteration steps and the bio-parts interchanged or modified to update the five parameters of each bactoneuron are illustrated. Biological circuit designs of each bactoneuron contruct involves one or more gene expression cassettes that were chosen as candidates for design configuration iterations. Such cassettes are grouped with colour-coded circles under the construct name bracket. Biological designs were iterated through updating either the relative copy number of a gene expression cassette with respect to others in the genetic circuit (high copy pUC or medium copy ColE1 or low copy p15A) and/or by modulation of the translation initiation rate by changing the strength of the RBS in the cassette (a library of 4 different RBSs of varying strengths[33] (refer to Supplementary table ST4) were used R_1, R_2, R_3 or R_4) and/or by modulation of the strength of repression by the Lambda repressor protein by changing between two variants of the protein cI^w or cI^* [18] and/or by modulating the promoter transcriptional strength of the AHL+LuxR activated promoter $Plux^i$ by changing between either of two variants $Plux^w$ & $Plux^*$. The gene expression cassette in question, where one or more of the modifications from the above described design space were effected in an iteration, is indicated through its colour coded circle. The specific bio-part configuration used for that gene expression cassette in an iteration is indicated through squares that glow with the same colour as of the cassette colour code. Squares coloured in grey indicates an unused configuration. For bactoneurons that went through multiple design iterations, the corresponding constructs generated at each iteration are grouped under its iteration number bracket denoted by "It.n" where n indicates iteration number. The constructs that were chosen as final at the final iteration step are highlighted in green boxes. Bactoneuron contructs under no iteration bracket denotes no design iteration performed for them. **(C)** Parameter values achieved by different bactoneuron constructs. Each bactoneuron construct were subjected to characterization and dose response experiments which yielded parameter values which were either measured or derived (see Methods). The set threshold and the acceptable region for each parameter are indicated by the dashed lines and the light green shaded area respectively. A bactoneuron construct gets selected as final when all 6 parameters fall within the acceptable region. Fold change w.r.t. max leakage (F.C.) is calculated by the fluorescence intensity at "ON" state of the construct divided by the Fluorescence signal at the highest leakage. Σ leakage (Σ L) is the summation of all the leakages in different induction conditions. The W_{AHL}, W_{IPTG} & W_{aTc} are derived from fitting the dose response data points against the log-Sigmoid function. Signal variation (S.V.) is drawn by scaling the fluorescence signal (@ "ON" state) of all bactoneurons in an iteration set (with the same output channel) by considering the highest signal of all bactoneurons as "1". The signal intensity from the strongest and weakest fluorescing bactoneuron must lie within 20% of each other. The parameter "Signal variation" (S.V.) is thus a comparative parameter across constructs in an iteration, while other parameters are intrinsic to the construct itself. Thus for S.V., all bactoneurons together must lie within the acceptable region, for the iteration set to be considered final. S.V. is thus shown in iteration groups.

We performed a detailed failure analysis on the disqualified bactoneuron constructs (Table S1). Now, keeping the general molecular design framework intact (Fig. 2), we optimised those suboptimal bactoneurons by modifying the genetic constructs, altering the interaction strength among the various transcription factors and promoters, and modulating their relative amounts in the cell. This had been done by changing plasmid copy numbers, using mutated forms of transcription factors, altering ribosome binding sites and promoter sequences, and redistributing the bioparts (Fig. 3B). Guided by the optimisation flow chart (Fig. 3A) and the failure analysis (Table S1), the optimisation was conducted in multiple iterations (Fig. 3B) until quantitative set points were reached (Fig. 3C). The schematics of genetic constructs and bioparts distribution among various plasmids in each iteration for each bactoneurons are shown in figures S6–S9.

Simulation and validation of optimized bactoneurons.

Based on the fitted parameters for each optimised bactoneurons, we performed computational simulations using parameterized activation functions. (Fig. 4). Further, we experimentally tested the behaviour of each optimised bactoneurons by simultaneously changing the concentrations of combinations of two inputs from among AHL, IPTG, and aTc and measuring the fluorescence at each data point. To reduce the experimental bias, in each case, a different set of inducer concentrations was selected compared to the dose response experiments (Table S2). These validation tests showed a close topological match between the simulated and experimental behaviour (Fig. 4).

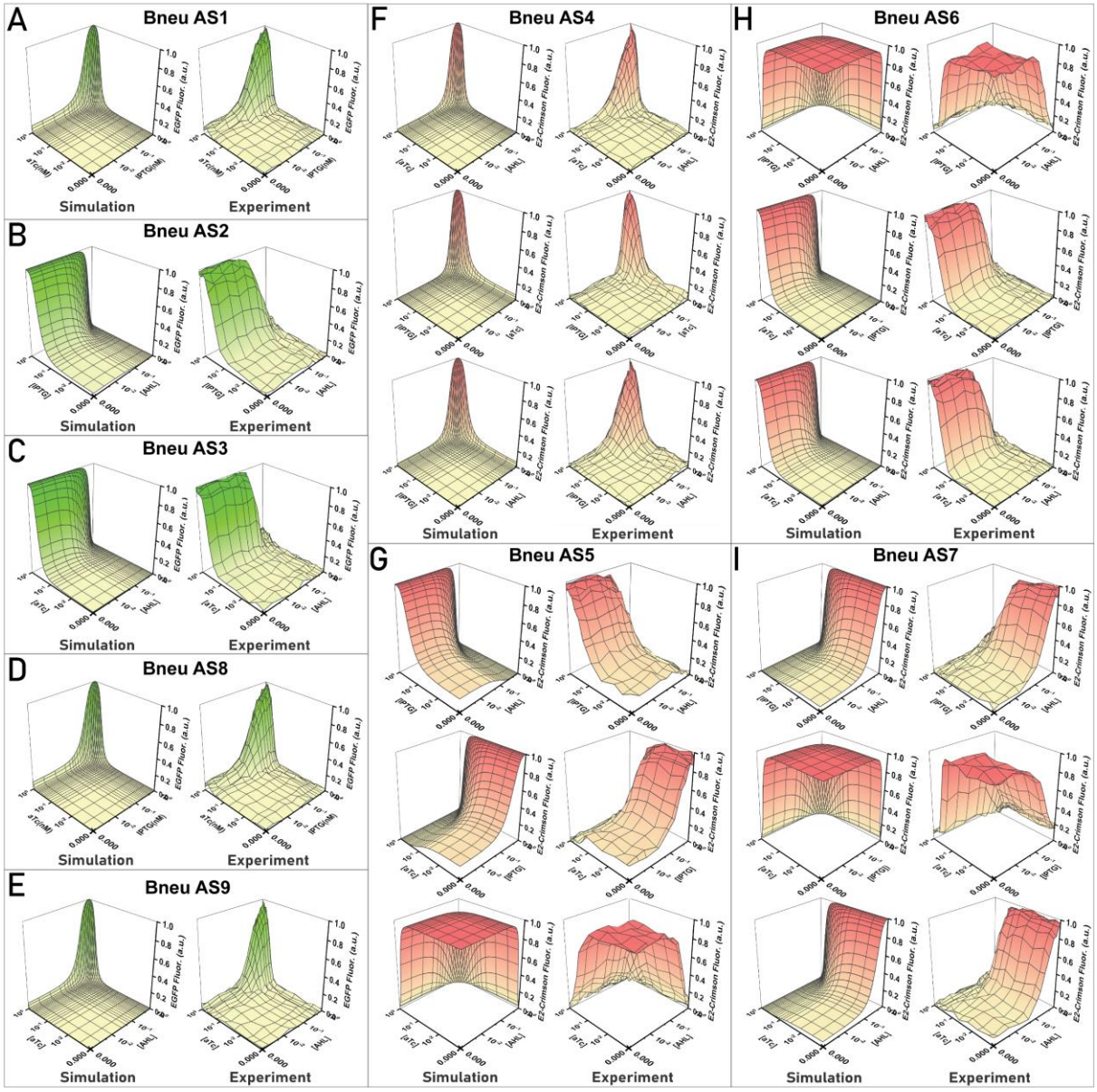

Figure 4: Simulations drawn from parameterized mathematical models of optimized Bactoneurons and its validation through validation experiments. Mathematical models were developed from parameters obtained from Dose response experiments (see Supplementary figures S2-5) and simulations were developed from these models by varying 2 inducers across 50x50 concentration points (while keeping the third inducer, if any, constant at "0" or "1" as the case may be) (see Methods) and their response in corresponding mathematical output signal intensity for each point. The simulations are shown towards the left of a panel for a bactoneuron. These simulations were validated by finding a good topological match with the validation experiments where the same corresponding set of inducers were varied across 16x16 concentration points in an actual experiment (that were different from those used for dose-response, see Supplementary table ST2) keeping the third, if any, constant, and plot with the corresponding reporter protein fluorescence. The validation experiments are shown towards the right of a panel for a bactoneuron. Panel **(A)** for Bneu AS1, **(B)** Bneu AS2, **(C)** Bneu AS3, **(D)** Bneu AS8, **(E)** Bneu AS9, **(F)** Bneu AS4, **(G)** Bneu AS5, **(H)** Bneu AS6 and **(I)** Bneu AS7. Bactoneurons connected to an EGFP output channel are represented in green shaded topology and those connected E2Crimson are shown in red shaded topology

Mix and match bactoneurons for a configurable adder and subtractor.

The nine optimised bactoneurons now worked as a configurable cellular system that could generate a full subtractor or a full adder. By mixing BNeu AS1, AS2, AS3, AS4, AS5, AS6 , and AS7 in a co-culture, we created a bactoneural layer that harboured the full subtractor function. Next, the subtractor co-culture was exposed to various combinations of input chemicals (AHL, IPTG, and aTc) according to the truth table, and the output fluorescence was seen with a fluorescence microscope. For the Full Subtractor, aTc served as the input Borrow (Bin), AHL served as the minuend and IPTG serves as the subtrahend. The E2-crimson expression (Output O_1) signified difference, and the EGFP expression (O_2) signified borrow-out. The results showed proper subtractor behaviour (Fig. 4A; Fig. S10A). Similarly, by just replacing BNeu AS2 and AS3 with BNeu AS8 and AS9 we could re-configure the bactoneural layer for a full adder behaviour, where aTc and IPTG served as the general inputs and AHL served as the input Carry (Cin); E2-Crimson expression signified sum and EGFP expression signified carry-out. The experimental results showed proper adder behaviour under fluorescence microscope. (Fig. 4B, Fig. S10B).

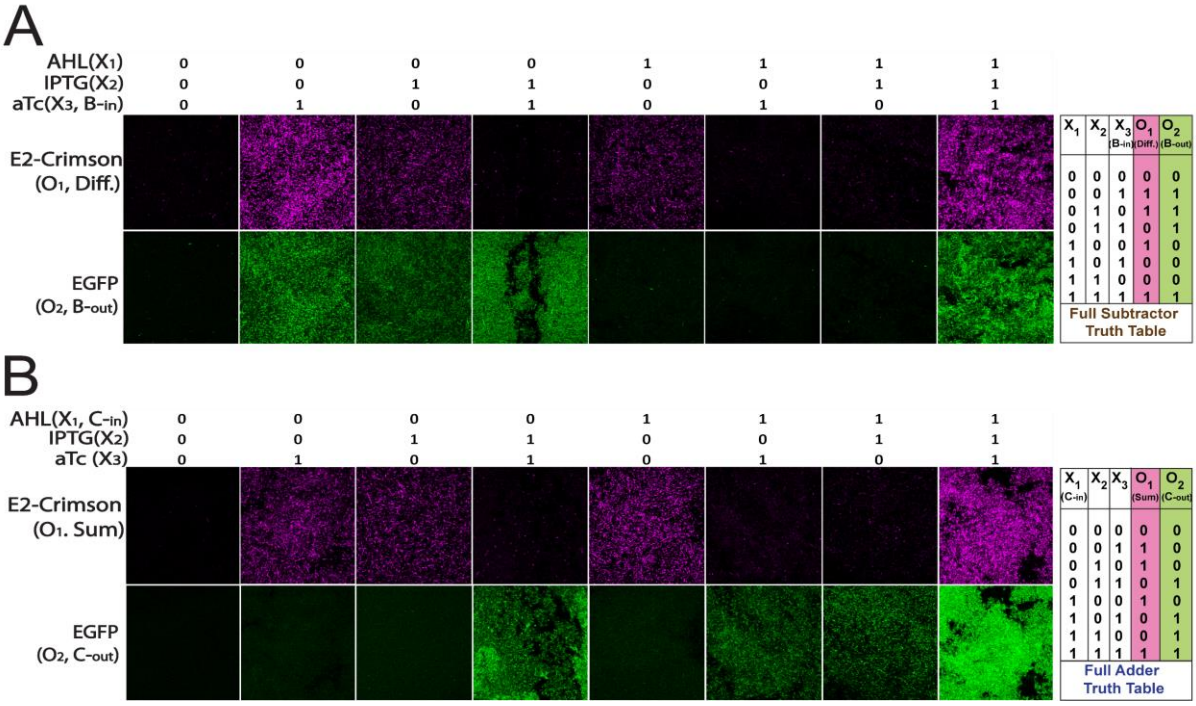

Figure 5: Fluorescence microscopy for A) Full Subtractor and B) Full Adder. Combinations of appropriate Bactoneuron types as required to constitute the corresponding Bactoneural layer were co-cultured for 16hours (with a sub-culture of 100 times dilution at the 10hour time point) with appropriate inducer combinations as per the respective truth table (shown to the right). The cultures were PBS washed and viewed as slides under relevant laser channels and emission filters (see methods). Heterogenous populations of E-coli cells forming the bactoneural layer can be seen in the fields under DIC/merged channels (see supplementary figure S10) where they respond in accordance to the input combinations. Each sub-population of bactoneurons in the bactoneural layer either gets activated (indicated by fluorescence from its respective output protein) or remains inactive (showing no fluorescence) based on the input inducer combinations. Logical adherence to the respective truth tables can be observed indicating proper operation of the Full Subtractor or Full Adder functions through the bactoneural layer.

Discussion

We demonstrated a configurable full subtractor and a full adder using engineered bacteria. The system is modular, whereby mixing and matching seven different bacterial populations can give rise to either a subtractor or an adder. Out of seven, five of the modular bacteria (BNeu AS1, AS4, AS5, AS6, and AS7) are common in both subtractor and adder, and by replacing only two bacterial types, it can switch between either a full subtractor or full adder. The

universal approximation theorem suggests that a feed-forward artificial neural network (ANN)-type architecture may approximate a wide variety of non-linear functions [17]. Thus, an ANN architecture might be a good strategy to build complex biocomputing functions. In this work, full subtractor and adder were created through a single-layer ANN-type architecture, where no layering of the cells with chemical communication was required. Thus, this design makes the construction of complex computing functions in cells easier. The absence of a second layer of cells with chemical communication might make the system respond faster to input signals.

Modularity and configurability are important hallmarks of engineering systems [20]. This helps to manage the complexity, perform parallel work, and accommodate future uncertainty. In this work, each engineered cell, which represented an artificial neuro-synapse within an ANN framework, worked as a modular and configurable system such that mixing and matching could generate two different functions. A few of the bactoneurons were taken from previous works, where they worked as modules to generate other functions. Interestingly, those 'bactoneurons' worked perfectly well within the configurable subtractor and adder system, suggesting the subtractor and adder possess a high degree of modularity and scalability.

Complex decision-making capability at the micron scale may have unprecedented applications in smart therapy [21-23], bioproduction [24] and bio-remediation [25]. Theoretical studies [26-28] predicted that to perform complex and large computations, multicellular distributed computing was necessary for its scalability, error tolerance and burden reduction per individual cell. However, often such experimental systems are fragile and unpredictive [3]. One of the hurdles in building functional multi-cellular systems is the unavailability of quantitative parameters, which are required for understanding as well as controlling the system [29-31]. In this work, each of the modular bactoneurons was quantitatively characterised and mathematically predictable. As the whole ANN architecture related to its modules (nodes)

through a mathematical framework, the behaviour of the ultimate function, subtractor or adder, was also predictable.

Though the work has been demonstrated with bacteria, the design of the ANN architecture and its individual 'bactoneurons' is abstract and general. Thus, it can be implemented with any type of cell, bacterial or mammalian.

Taken together, this 'lego-like'' mix and match modular and configurable cellular system provide flexible and scalable means to build complex biological functions with engineered cells. This work may have significance in biocomputer technology development, multi-cellular synthetic biology, and cellular hardware for ANN with potential industrial and therapeutic applications.

Materials and Methods

RBS design and promoters, genes, and plasmids.

Promoters, ribosome biding sites (RBS), genes & transcription terminators were all assembled using the Network Brick pipeline [32] into pZ expression systems [19] according to the corresponding genetic circuit designs. RBS with various strengths were designed and translation initiation rates for designed RBSs (Table S3) were calculated thermodynamically from RBS calculator [33]. Standard molecular biology protocols were used. Restriction enzymes, T4 DNA ligase, buffers & DNA ladders were from New England BioLabs. PCR reactions were performed using the KOD Hotstart DNA polymerase (Merck Millipore). Plasmid isolation, PCR purification & Gel extraction were done using QIAGEN kits. All promoter sequences, gene sequences and primer sequences are shown in table S4. Details of

the cellular devices and plasmids are shown in figures S6-S9. Primers were synthesized by Integrated DNA Technologies, Singapore. Oligos & gene Products were synthesized by Invitrogen GeneArt Gene Synthesis service, Thermo Fischer. All constructs & bio-parts were sequence verified by Eurofins Genomics India Pvt. Ltd., Bangalore, India.

Bacterial strains and culture conditions

For all cloning purposes *Escherichia coli* DH5α strain was used. For all experiments, DH5αZ1 strain [18] was used as the chassis organism. Chemically competent DH5αZ1 were co-transformed with sequence verified plasmid constructs as per design of ach of the bactoneurons and were plated onto LB-agar, Miller (Himedia) plates with appropriate antibiotics (ampicillin (Himedia) at 100 µg/ml, chloramphenicol (Himedia) at 34 µg/ml). For all liquid cultures, LB-broth, Miller (Himedia) was used with appropriate antibiotics. Appropriate chemical inducers were provided into liquid cultures (AHL ($C_{10}H_{15}NO_4$,) (Merck) 0 – 5000 nM, IPTG (Himedia) 0- 10,000 µM, aTc (Sigma) 0 -200 ng/ml. Incubation was done 37° C, ~ 280 rpm in New Brunswick Scientific, Innova 42 incubator shaker.

Testing of Bactoneuron constructs for adherence to respective truth-tables

Four distinct single colonies were picked from the plates and grown in liquid culture. For induction, overnight cultures were diluted (100 times) in fresh LB with appropriate inducer combinations as per the truth table (Fig. 1). This induction was given for a total of 16 h with a sub-culture of 1/100 dilution at the 10hr time-point. The ‘0’ and ‘1’ states signified zero and saturated inducer concentration.

Dose Response Experiments

3 or 4 single colony-overnight cultures for each bactoneurons were set up in in LB-Broth as before. For an experimental set, each of these cultures were subjected to 16hour induction (with

a sub-culture of 1/100 dilution into identical composition media at the 10hr time-point) in separate 3mL LB broth cultures with different inducer concentration points, where one inducer was varied across 10 or more intermediate concentration points (Table S2) between its "0" to "1" state (see scaling below) while the other inducer(s) was/were kept constant at "0" or "1" concentration as the case maybe. This was performed for each bactoneuron for each of the 3 or 2 inducers with non-negative weights.

Fluorescence Measurements, Normalization and Scaling

Post 16hr induction cells were washed and re-suspended in phosphate buffered saline (PBS, pH 7.4) for fluorescence measurements. Cells were re-suspended into PBS such that OD_{600} values were between 0.7-0.9. These cultures were then loaded onto a black flat-bottom 96-well plate (Greiner) & using a Synergy HTX Multi-Mode reader (Biotek Instruments, USA), OD_{600} & fluorescence was measured using appropriate combinations of excitation and emission filters. Cells were excited by a white light source that had been passed through an excitation filter of 485/20 nm band-pass for EGFP and 610/10nm band-pass for E2Crimson while emission was collected by 516/20 nm band-pass filter for EGFP at gain 60 & 610/10nm band-pass filter at gain 70 for E2Crimson. To collect the fluorescence and OD data at least 3 biological replicates had been considered for each condition. To normalize the raw fluorescence reading obtained, to the number cells in the well, the raw fluorescence values were divided by the respective OD_{600} values of each well. Auto-fluorescence was measured as average normalized fluorescence of the untransformed DH5αZ1 set (no plasmid set) and subtracted from the normalized fluorescence value of the experimental set. The resultant value was averaged across biological replicates to obtain the fluorescence intensity at an induction condition. The above process can be mathematically represented as follows:

$$Unscaled\ Normalized\ Fluorescence =$$

$$\left[\frac{(Absolute\ Flourescence\ value\ from\ experimental\ cell\ population)}{(OD\ of\ experimental\ cell\ population)}\right]-$$

$$\left[\frac{(Absolute\ Flourescence\ value\ from\ No\ plasmid\ cell\ population)}{(OD\ of\ No\ plasmid\ cell\ population)}\right]\ldots.\ \textbf{(2)}$$

These normalized fluorescence values were then scaled down between 0 and 1 by considering the normalized fluorescence value at the induction point of maximum expected/observed fluorescence to be 1 and by dividing all other fluorescence values by this fluorescence value.

$$Scaled\ Normalized\ Fluorescence=$$

$$\left[\frac{(Unscaled\ Normalized\ Fluorescence\ at\ any\ induction\ point)}{(Unscaled\ Normalized\ Fluorescence\ at\ induction\ point\ of\ maximum\ expected\ and\ oberevd\ Fluorescence)}\right]\ldots.\ \textbf{(3)}$$

Fitting & Scaling input concentration points

The scaled fluorescence values (y-axis) obtained were plotted against varying inducer concentration points (x-axis) onto a scatter plot on Origin2018 (OriginLab Corporation, USA). Using the in-built Orthogonal Distance Regression algorithm of the software, the scatter plot was fitted with a modified form of equation 1

$$\mathrm{Oj} = \frac{1}{1+e^{-(w_j n.X_j n+B)}}\ \ldots\ldots.\ \textbf{(4a)}$$

where, Y represents the normalized & scaled fluorescent outputs, W_{jn} represents the weight of the nth inducer input for the jth bactoneuron , X_{jn} represents scaled concentration for the nth input and $B = w_{jm}.x_{jm} + w_{jp}.x_{jp} + bj\ldots.$ **(4b)**

Where wj(m,p), xj(m,p) are weight and input for other two (m and p) inducers. B is a constant for an inducer specific dose response experiment for a specific bactoneuron. Inducer concentration points were scaled between 0 & 1 where “0” represents absence of the inducer

in the experimental set & “1” represents that concentration of the inducer where Oj just attains a value of 0.999 for inducers with positive weight sign or 0.001 for those with negative weight sign. All values of concentration above this point were designated as “1”. The bias bj value of the bactoneuron was derived by solving from different parameterized versions of equation 4b from different dose response experiments.

Simulation

All bactoneurons were modelled based on the activation function (equation 1). Parameterized activation function with numerical values of wj and bj were used for computational simulations for ach bactoneuron by generating 50x50 matrices of estimated output fluorescence values “Oj” where 2 inputs simultaneously varied across 50x50 scaled relative concentration points between “0” & “1” . The third inducer (if any) was kept at “0” or “1” as the case may be. We used a fixed set of scaled relative concentration points to draw all the simulations as listed in table S5.

Validatory experiments for verifying mathematical models & simulations

Verification of the mathematical models & simulations were done through validatory experiments subjecting an overnight culture of a single colony (n=1) of an appropriately transformed DH5αZ1 cell to 16hr induction as described previously, where 2 inducers were varied simultaneously across 16x16 concentration points (keeping the third inducer, if any, constant at “0” or “1” as the case may be) that were different from those that were used in the dose response experiments. The list of concentration points for validation experiments for different inducers & bactoneurons were are listed in table S2. Fluorescence readings were recorded, normalized, scaled and plot against the scaled concentration points of 2 inducers.

Confocal Microscopy

Bactoneurons were grown overnight. Based on the bactoneural configuration of full subtractor or adder ANN (Figure 1), inoculum of each bactoneuron cultures were equivalently mixed and the combined inoculum was added with 100 times dilution into fresh LB broths with appropriate inducer combinations as per truth table. These 10hr co-cultures were again sub-cultured with 100 times dilution for another 6hours and the final co-cultures were washed twice with PBS (pH 7.4). The final washed cell pellets were again re-suspended in fresh PBS, added onto glass slides, covered with coverslips and sealed with a sealer. These were observed under 60X water immersion objective in a Nikon AIR si confocal microscope supplied with the resonant scanner and coherent CUBE diode laser system. The sample fields were subjected to excitation of the fluorescent proteins by sequentially illuminating with 2 laser channels, with λ = 488 nm for EGFP, and λ= 561 nm for E2-Crimson. Fluorescence emissions were captured through a T-PMT with relevant emission filters of 525/50 nm band-pass for EGFP & 700/75 nm band-pass for E2-Crimson. Differential interference contrast (DIC) images were also acquired for each captured field. ImageJ Fiji software was used for processing of acquired images.

Author Contributions

D.B. and S.C. performed all the experiments. D.B. and S.B. designed the experiments, analyzed the data, and wrote the manuscript. S.B. conceived the study.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

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Supplementary Information

Configurable Full Subtractor and Adder by Lego-like Mixing and Matching of Engineered Bacteria

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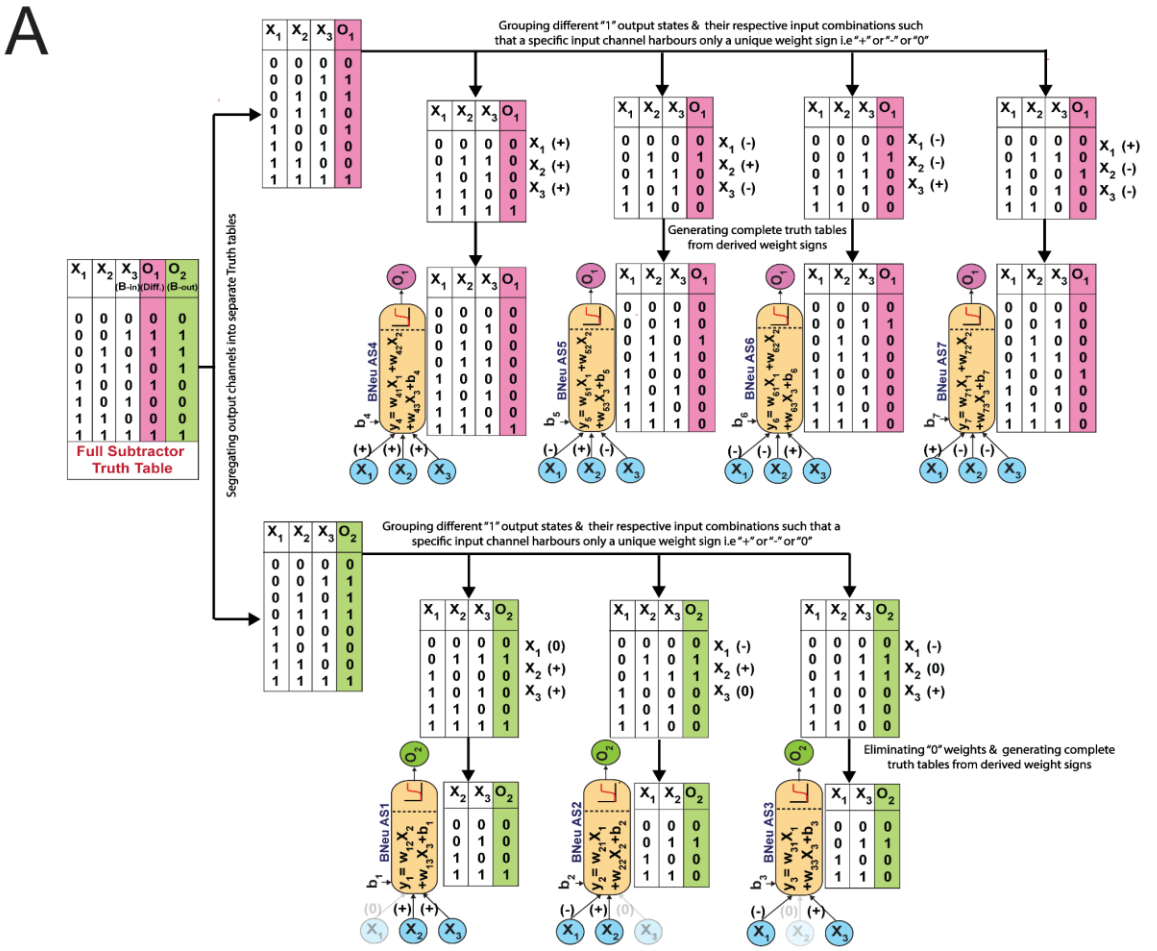
A
Full Subtractor Truth Table
Segregating output channels into separate Truth tables
Grouping different "1" output states & their respective input combinations such that a specific input channel harbours only a unique weight sign i.e "+" or "-" or "0"
Generating complete truth tables from derived weight signs
Grouping different "1" output states & their respective input combinations such that a specific input channel harbours only a unique weight sign i.e "+" or "-" or "0"
Eliminating "0" weights & generating complete truth tables from derived weight signs
BNeu AS1
BNeu AS2
BNeu AS3
BNeu AS4
BNeu AS5
BNeu AS6
BNeu AS7

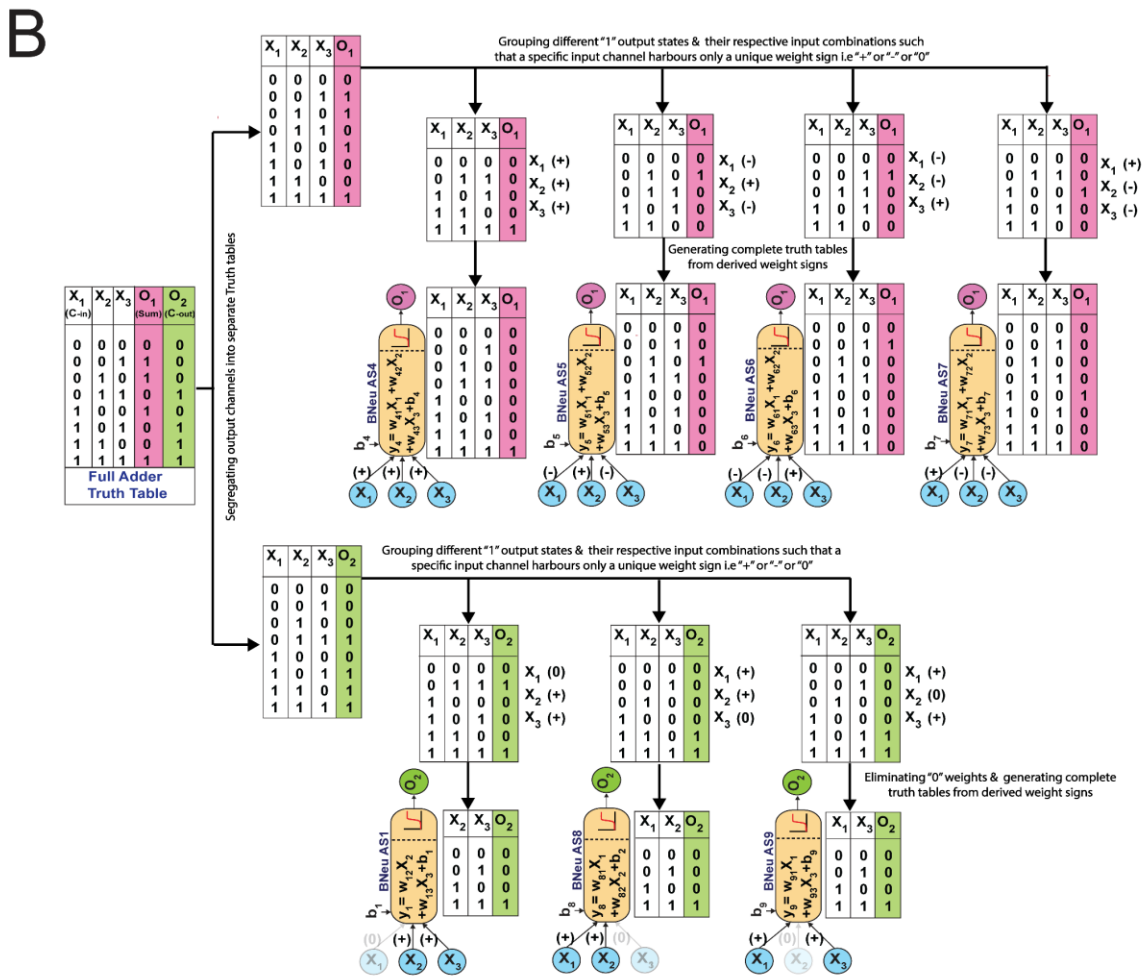
B
Full Adder Truth Table
Segregating output channels into separate Truth tables
Grouping different "1" output states & their respective input combinations such that a specific input channel harbours only a unique weight sign i.e "+" or "-" or "0"
Generating complete truth tables from derived weight signs
Grouping different "1" output states & their respective input combinations such that a specific input channel harbours only a unique weight sign i.e "+" or "-" or "0"
Eliminating "0" weights & generating complete truth tables from derived weight signs
BNeu AS1
BNeu AS8
BNeu AS9
BNeu AS4
BNeu AS5
BNeu AS6
BNeu AS7

Figure S1: Fragmentation of Truth tables through ANN design rules and derivation of Bactoneurons with corresponding weight signs for A) Full Subtractor B) Full Adder. "0" weight inputs, if any, of a bactoneuron have been faded in the Bactoneural schematic.

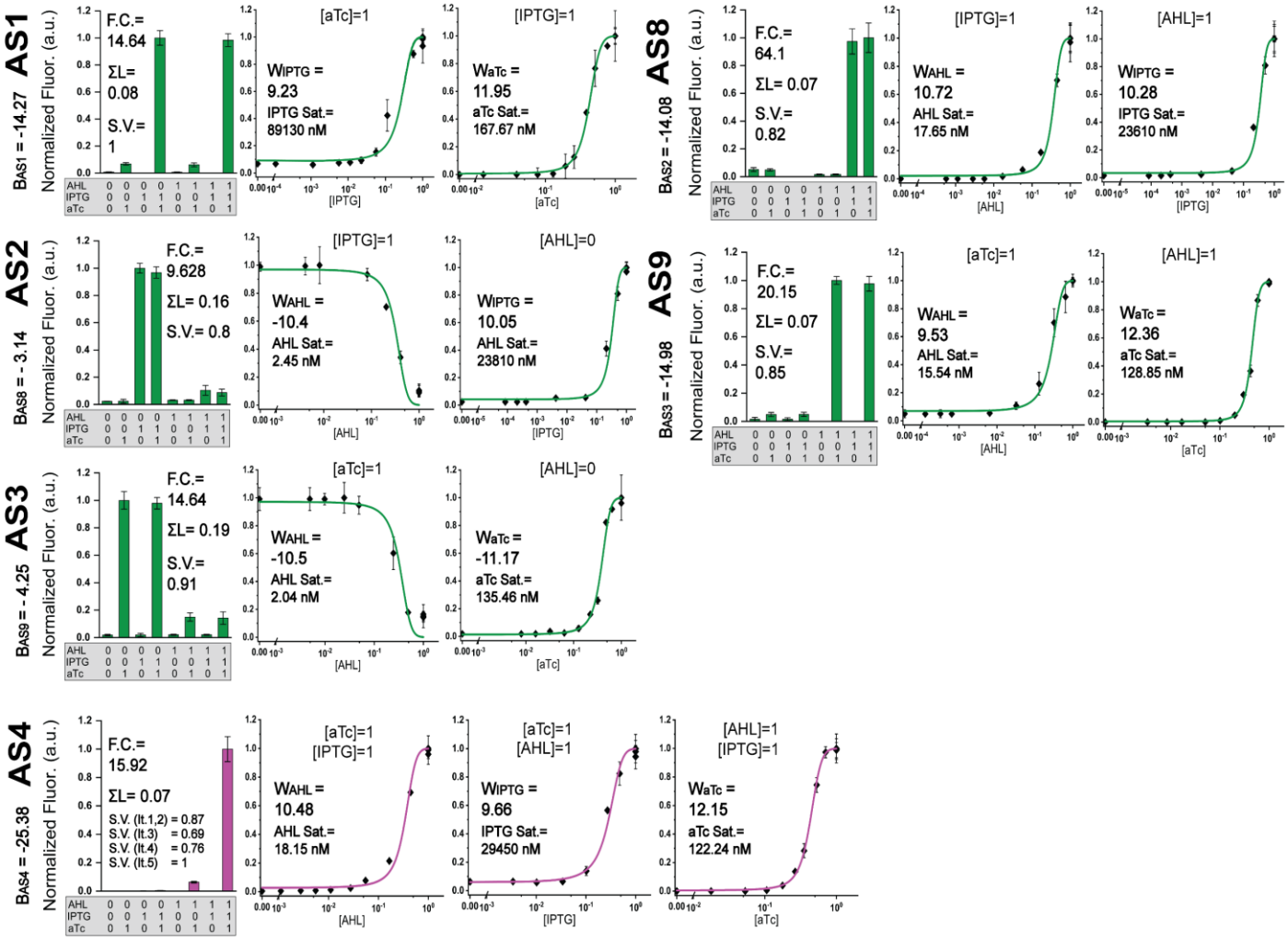

Figure S2: Details of characterization and Dose Responses experiments of Bactoneurons that went through no iteration and were selected as final. (AS1, AS2, AS3, AS4, AS8 & AS9). The experimental sets of each Bactoneuron type, yielded corresponding parameter values (shown in figure) of Fold Change (F.C.), Σ Leakage (ΣL), Signal variation (S.V.), Bias ($B_{AS\#}$), Weights ($W_{inducer}$) & Inducer Saturation points (Sat.) The fluorescence readings obtained from the dose response experiments were fitted with the Log-Sigmoid function (see Methods) to derive the Weight & Bias parameters

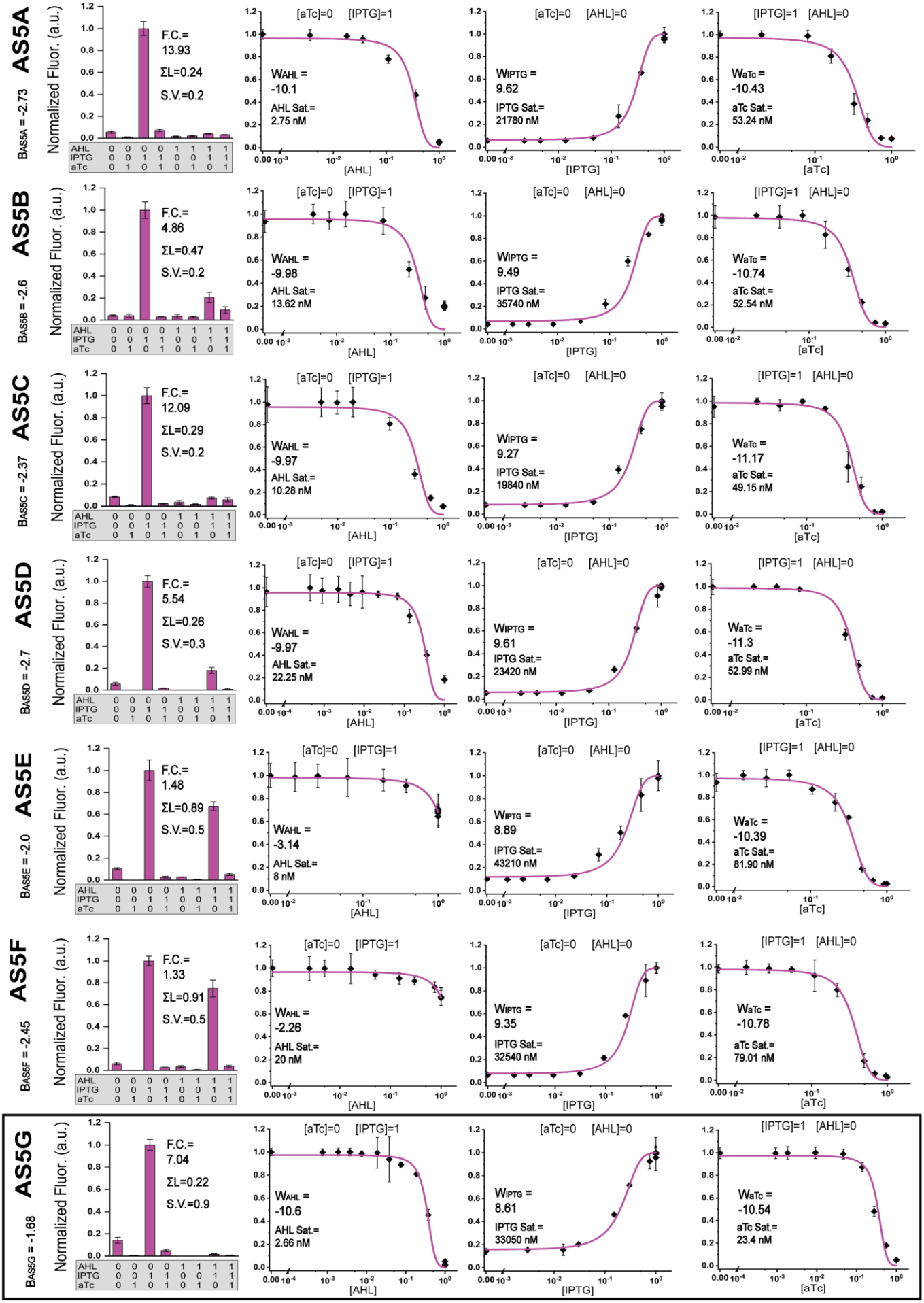

Figure S3: Details of characterization and Dose Responses experiments across iterations of Bactoneuron 5 (AS5A, AS5B, AS5C, AS5D, AS5E, AS5F & AS5G) yielding corresponding parameter values (shown in figure) of Fold Change

(F.C.), Σ Leakage (ΣL), Signal variation (S.V.), Bias ($B_{AS\#}$), Weights ($W_{inducer}$) & Inducer Saturation points (Sat.). The fluorescence readings obtained from the dose response experiments were fitted with the Log-Sigmoid function (see methods) to derive the Weight & Bias parameters. The final selected Construct is shown boxed.

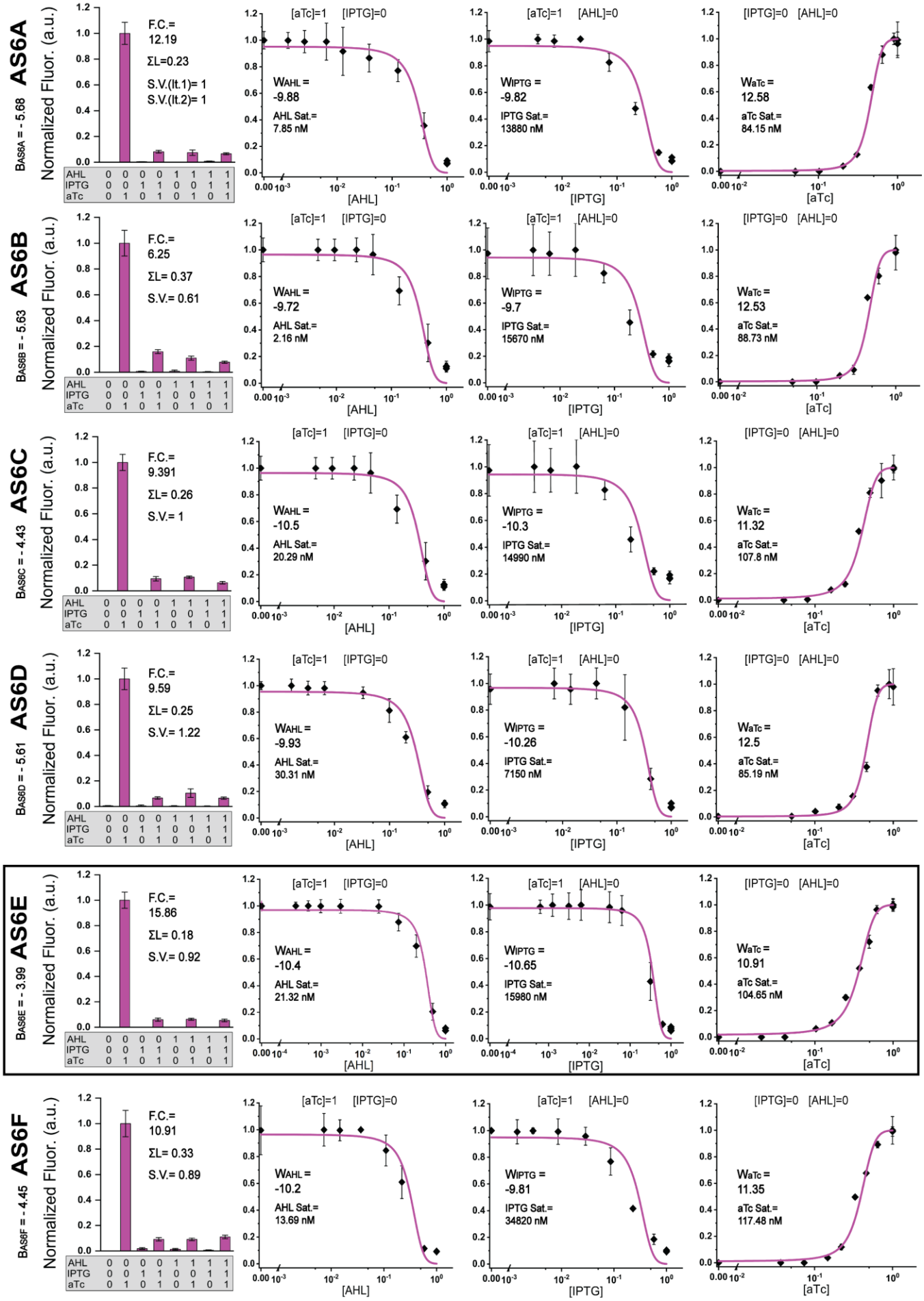

Figure S4: Details of characterization and Dose Responses experiments across iterations of Bactoneuron 6 (AS6A, AS6B, AS6C, AS6D, AS6E &AS6F) yielding corresponding parameter values (shown in figure) of Fold Change (F.C.), Σ Leakage (ΣL), Signal variation (S.V.), Bias ($B_{AS\#}$), Weights ($W_{Inducer}$) & Inducer Saturation points (Sat.). The fluorescence readings obtained from the dose response experiments were fitted with the Log-Sigmoid function (see Methods) to derive the Weight & Bias parameters. The final selected Construct is shown boxed.

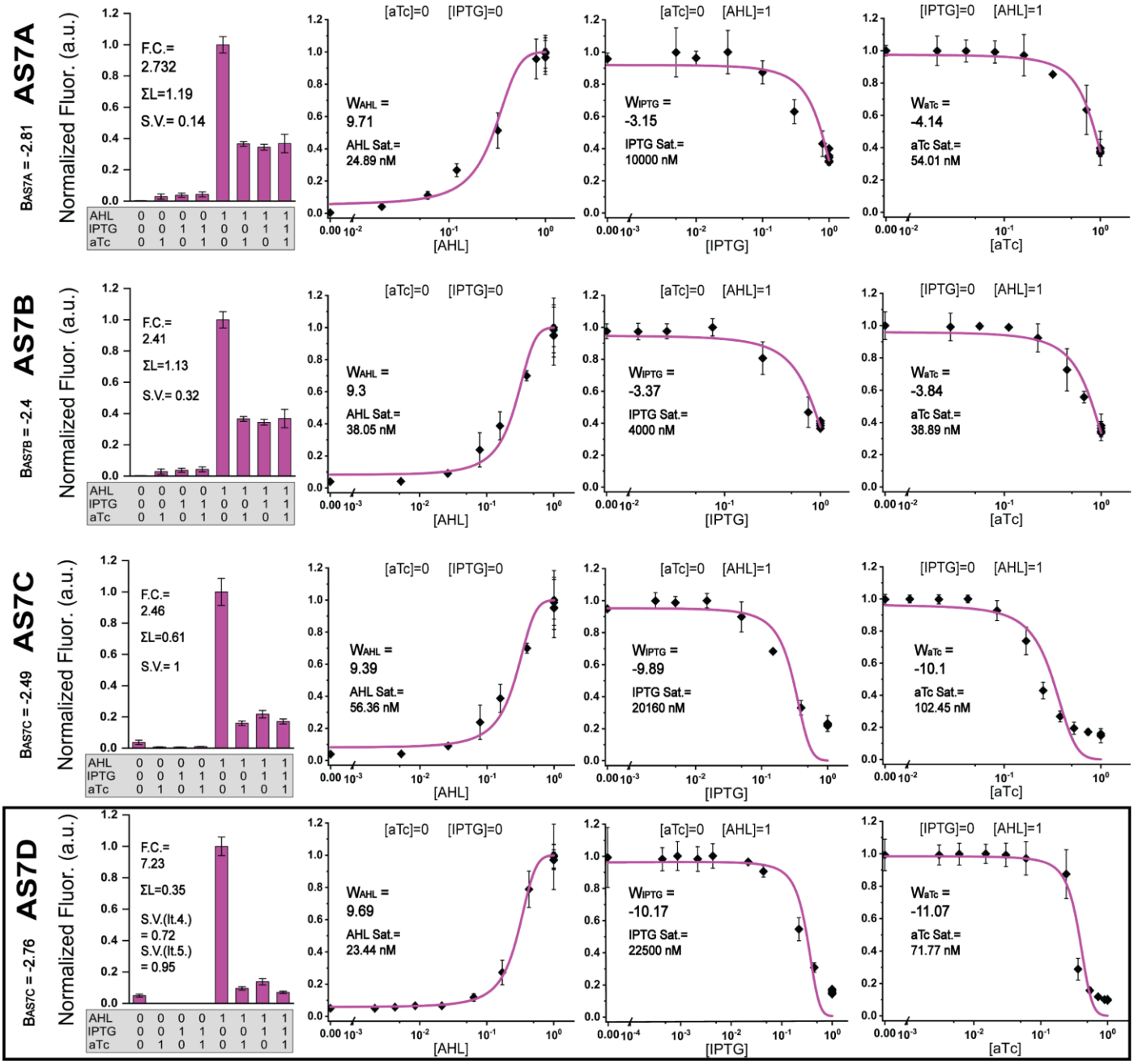

Figure S5: Details of characterization and Dose Responses experiments across iterations of Bactoneuron 7 (AS7A, AS7B, AS7C & AS7D) yielding corresponding parameter values (shown in figure) of Fold Change (F.C.), Σ Leakage (ΣL), Signal variation (S.V.), Bias ($B_{AS\#}$), Weights ($W_{Inducer}$) & Inducer Saturation points (Sat.). The fluorescence readings obtained from the dose response experiments were fitted with the Log-Sigmoid function (see Methods) to derive the Weight & Bias parameters. The final selected Construct is shown boxed.

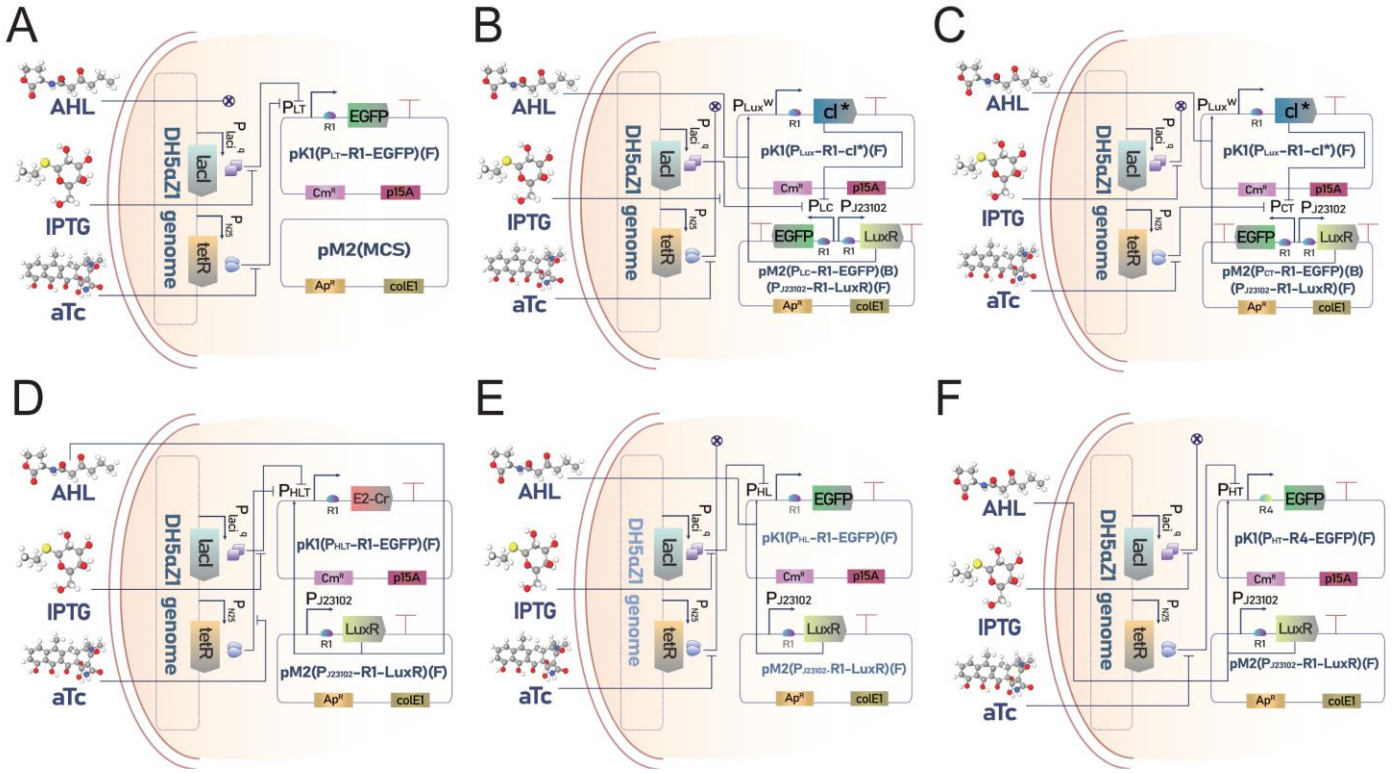

Figure S6: Detailed genetic circuit designs & plasmid maps of Bactoneurons that went through no iteration and were selected as final. (A) BNeu AS1, (B) BNeu AS2, (C) BNeu AS3, (D) BNeu AS4, (E) BNeu AS8, (F) BNeu AS9. Names of plasmids constructed & incorporated in the circuits are also shown within the plasmid schematic. MCS represents empty network brick cloning site. "F" & "B" denotes forward & reverse direction of cassette respectively.

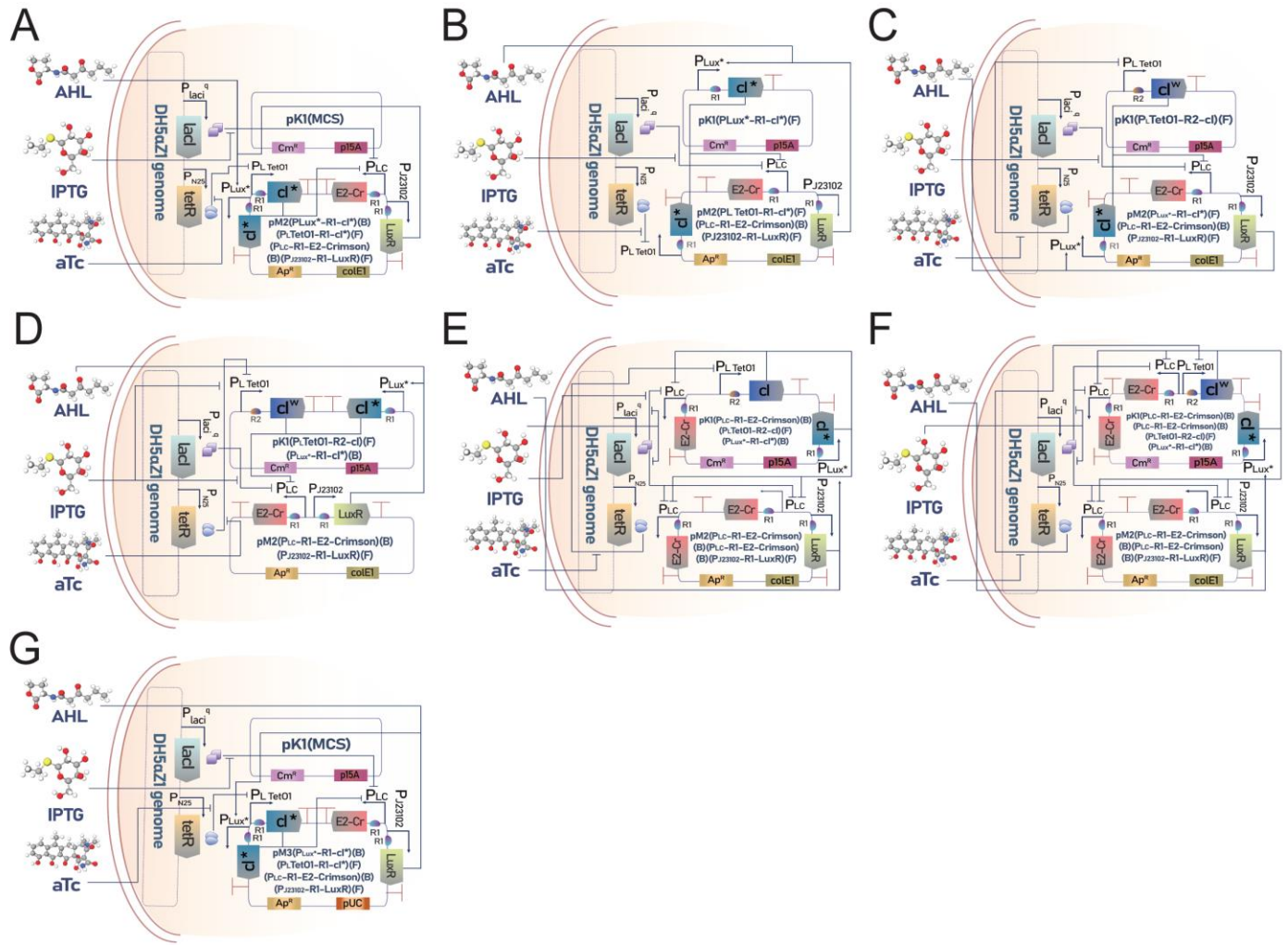

Figure S7: Detailed genetic circuit designs & plasmid maps of Bactoneuron 5 across iterations (A) BNeu AS5A, (B) BNeu AS5B, (C) BNeu AS5C, (D) BNeu AS5D, (E) BNeu AS5E, (F) BNeu AS5F & (G) BNeu AS5G. Names of

plasmids constructed & incorporated in the circuits are also shown within the plasmid schematic**.** MCS represents empty network brick cloning site. “F” & “B” denotes forward & reverse direction of cassette respectively.

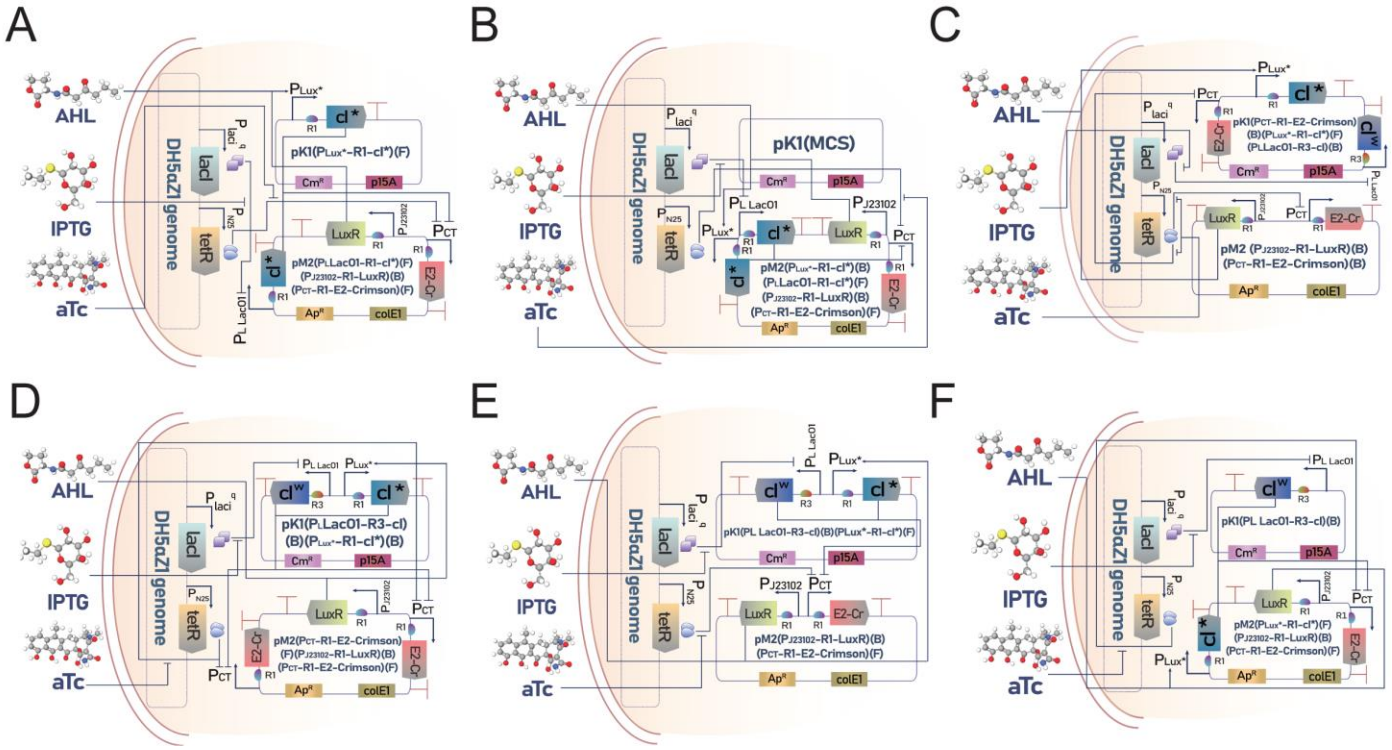

Figure S8: Detailed genetic circuit designs & plasmid maps of Bactoneuron 6 across iterations (A) BNeu AS6A, (B) BNeu AS6B, (C) BNeu AS6C, (D) BNeu AS6D, (E) BNeu AS6E & (F) BNeu AS6F. Names of plasmids constructed & incorporated in the circuits are also shown within the plasmid schematic**.** MCS represents empty network brick cloning site. “F” & “B” denotes forward & reverse direction of cassette respectively.

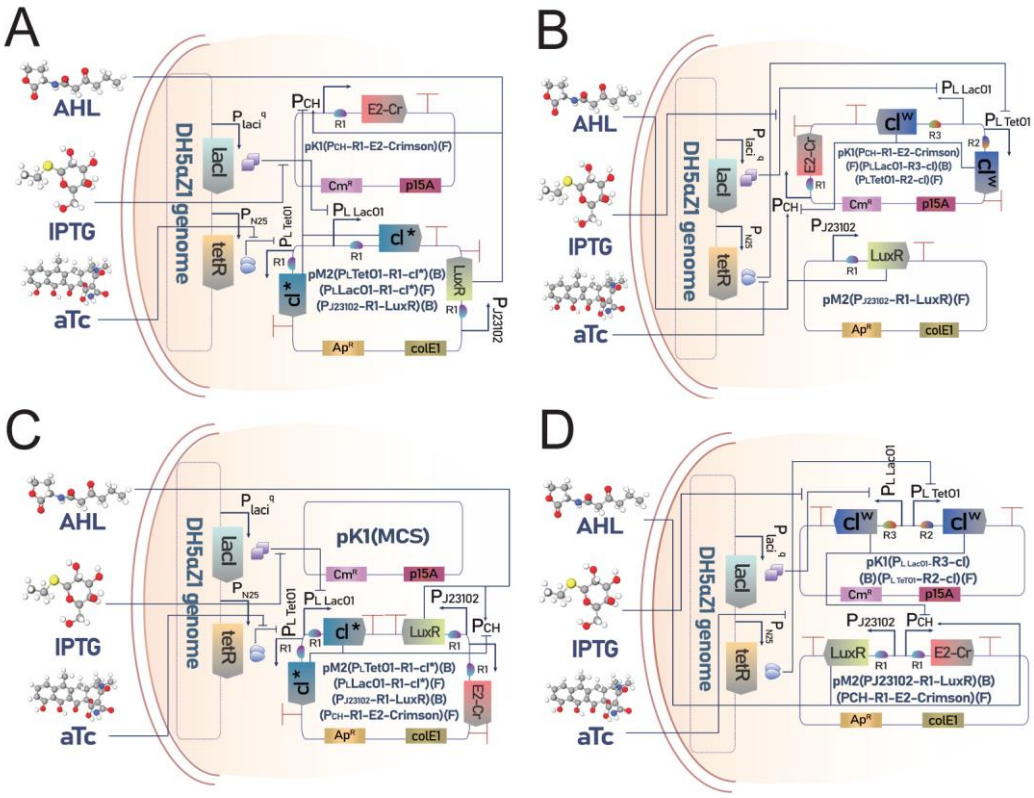

Figure S9: Detailed genetic circuit designs & plasmid maps of Bactoneuron 7 across iterations (A) BNeu AS7A, (B) BNeu AS7B, (C) BNeu AS7C & (D) BNeu AS7D. Names of plasmids constructed & incorporated in the circuits

are also shown within the plasmid schematic. MCS represents empty network brick cloning site. “F” & “B” denotes forward & reverse direction of cassette respectively.

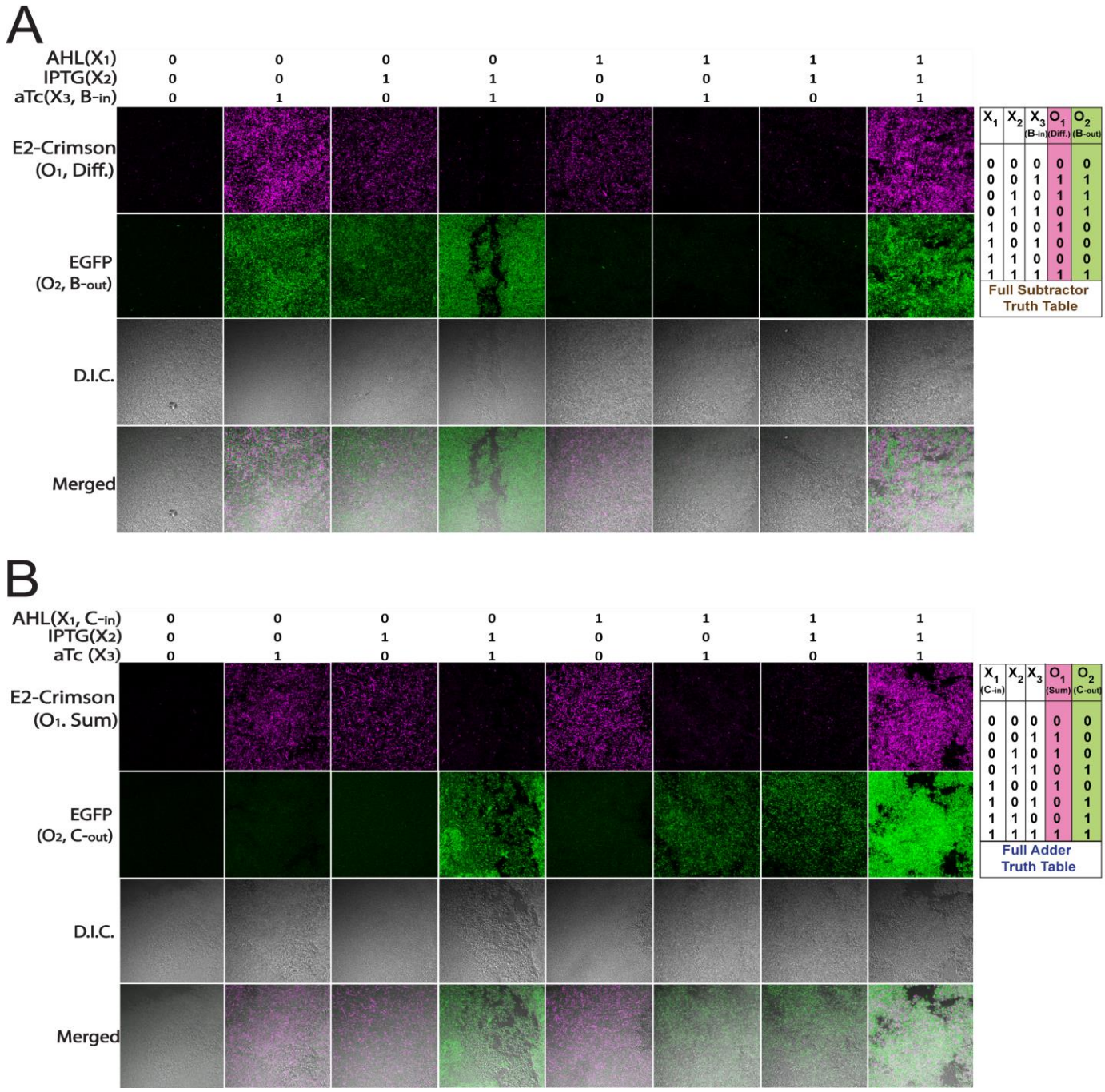

Figure S10: **Fluorescence microscopy images with corresponding differential interference contrast (DIC) and merged channels for A) Full Subtractor, B) Full Adder.** Combinations of appropriate Bactoneuron types as was required to constitute the corresponding Bactoneural layer were co-cultured for 16hours (with a sub-culture of 100 times dilution at the 10hour time point) with appropriate inducer combinations. The cultures were PBS washed and viewed as slides under relevant laser channels and emission filters. Heterogenous populations of E-coli cells forming the bactoneural layer can be seen in the fields under DIC/merged channels where they respond in accordance to the input combinations. Each sub-population of bactoneurons in the bactoneural layer either gets activated (indicated by gets fluorescence from its respective output protein) or remains inactive (showing no fluorescence) based on the input inducer combinations

Table S1: Failure Analysis

Iteration Number	Bacto-neuron construct, figure number for molecular design , and acceptance status	Performance Analysis	Troubleshooting undertaken
1	AS5A Figure S7(A) (Rejected)	Construct AS5A (& AS7A) had significantly lower fluorescence intensity at "ON" states compared to constructs AS4 & AS6A in the 1st iteration set with about -81.56% deviation from AS4. It had 0.16 times the fluorescence intensity of AS6A (construct with highest fluorescence in iteration set) aTc weight was also below the threshold of -10.5. It is to be noted that the repressor cI* expresses from $\mathbf{P_{LUX}}$* & $\mathbf{P_{LtetO1}}$ promoters both in ColE1 origin. Since from the behaviour of BNeu AS8 & AS9, it was indicated that promoter $\mathbf{P_{CL}}$ (used in AS5 & AS8) & $\mathbf{P_{CT}}$ (used in AS6 & AS9) are of comparable strength, we reckoned that possibly, a significant amount of cI* expressing from the combined leakage of promoters $\mathbf{P_{LtetO1}}$ & $\mathbf{P_{LUX}}$* is diminishing the expression of E2Crimson from $\mathbf{P_{CL}}$ at the [0,1,0] input state.	First, we fixed the Fluorescence intensity of BNeu AS4 at its "ON" state was set as a reference point to judge other Bactoneural constructs since AS4 had passed all the parameter thresholds & exhibited comparatively better parameter values than other E2Crimson expressing constructs in the 1st iteration.. We decided to bring the Fluorescence values of all E2Crimson expressing constructs to within +/-10% of that of AS4 OR a minimum of 0.8 times the highest Fluorescence observed in the iteration set. To achieve this, the cassette $\mathbf{P_{LUX}}$*-R1-cI* was shifted from ColE1 to p15A origin to reduce the amount of leakage levels of cI*. This gives rise to construct AS5B. Also, alternatively, the cassette $\mathbf{P_{LtetO1}}$-R1-cI* in colE1 was replaced by $\mathbf{P_{LtetO1}}$-R2-cI^{w} in p15A in construct AS5C to check if this reduces any effects of Lambda repressor leakages.
2	AS5B Figure S7(B) (Rejected)	AS5B showed very little reduction in variation in "ON" state Fluorescence intensity, (down to -73.4% from -81.56% of AS4) & therefore far from our acceptable threshold. It was also only 0.23 times the highest fluorescence observed in the iteration set (from AS6A), far less than our threshold of 0.8. Moreover, there was a loss of fold change which dropped below the threshold of 7 to 4.861 and the Σ Leakage too breached its acceptable upper limit of 0.4. This suggests that maybe the transfer of $\mathbf{P_{LUX}}$*-R1-cI* to p15A is not enough to raise fluorescence levels of BNeu AS5.	In AS5B & AS5C we alternatively shifted the expression of lambda repressor from $\mathbf{P_{LUX}}$* & $\mathbf{P_{LtetO1}}$ promoters from ColE1 to p15A origin plasmids. Both modifications exhibited slight reductions in variations of "ON" state Fluorescence intensity against AS4. However, neither singularly could reduce it up to our stipulated threshold. We reckoned that if diminished E2Crimson expression from PRL was due to excessive combined leakage of Lambda repressor from $\mathbf{P_{LUX}}$*& $\mathbf{P_{LtetO1}}$, in the [0,1,0] input state, perhaps if both PLUX* & $\mathbf{P_{LtetO1}}$ were transferred under p15A origin, fluorescence levels of BNeu AS5 could be raised further. Toward this, we transferred the $\mathbf{P_{LUX}}$*-R1-cI* cassette of AS5A to a p15A origin plasmid & replaced the $\mathbf{P_{LtetO1}}$-R1-cI* cassette in ColE1 in AS5A with a $\mathbf{P_{LtetO1}}$-R2-cI^{w} cassette in p15A. The two modifications together gave rise to Construct AS5D.
2	AS5C Figure S7(C) (Rejected)	For AS5C, we moved the Lambda repressor under $\mathbf{P_{LtetO1}}$ in AS5A from a colE1 plasmid to p15A with the speculation that reduced leakage of the protein from $\mathbf{P_{LtetO1}}$ will cause the Fluorescence intensity of BNeu AS5 to rise in [0,1,0] state. We also replaced the frameshifted variant cI* with the wild type cI^{w} and employed a weaker RBS R2. The weak RBS was employed in accordance with our previous work for tuning wild type Lambda repressor levels from this same promoter. AS5C was still 75.94% weaker in fluorescence than AS4 and 0.21 times the highest fluorescence However, fold change and other parameters remained above the respective thresholds. We speculated that the above modification alone was insufficient to meet our purpose.	
3	AS5D Figure S7(D) (Rejected)	The ratio of fluorescence intensity of AS5D compared to the AS7C, the construct with the highest fluorescence intensity in the iteration set was 0.29, which was a mild improvement. AS5D displayed quite a better reduction in the variation of Fluorescence intensity against AS4 compared to its predecessors. However, at -58.2% it was still below the required levels & fold change against max leakage was also below 7 at 5.54. This indicated that an increase in Fluorescence intensity could perhaps be achieved by increasing the amount of E2Crimson expression instead of simply reducing Lambda repressor leakage levels.	We noted that the absolute fluorescence intensity of AS5D was roughly 1/3rd of that of AS4 in their respective "ON" states. Since R1 was already a strong RBS, we tried to increase the levels of E2Crimson by adding additional "$\mathbf{P_{CL}}$ -R1-E2Crimson" cassettes to the circuit. For AS5E we added another $\mathbf{P_{CL}}$-R1-E2Crimson cassette upstream of the existing one in the AS5D ColE1 origin plasmid. Another cassette was added upstream of the $\mathbf{P_{LtetO1}}$-R2-cI^{w} cassette of AS5D in p15A. For AS5F, the modifications were similar but a 3rd additional cassette was added in the p15A origin plasmid.
4	AS5E Figure S7(E) (Rejected)	Both AS5E & AS5F displayed a further significant reduction in the variation of Fluorescence intensity against AS4, down to around -39% from -58.2% in AS5D. The ratio of fluorescence intensity against the highest fluorescent intensity of the iteration (observed in AS6C) set improved up to 0.46 However, this still was not within our threshold limits. On the bright side, it did strongly suggest that an increasing number of E2-Crimson expressing cassettes within the cell was the key to mitigating the problem of differences in Fluorescence intensity with AS4, and reduction in Lambda repressor leakage levels by reducing the relative numbers of its expressing promoters $\mathbf{P_{LtetO1}}$ & $\mathbf{P_{LUX}}$* was a less effective approach. In fact, in both AS5E & AS5F, the drastic drop in fold changes to 1.49 & 1.34 respectively, and the rise of Σ Leakage was majorly due to leakage of E2Crimson expression from the [1,1,0] input state suggesting inability of the $\mathbf{P_{LUX}}$*-R1-cI* cassette to repress the $\mathbf{P_{CL}}$ -R1-E2Crimson cassettes properly in this condition. This also resulted in a drastic decrease in the absolute values of AHL weight.	It was realized that while increasing the number of E2Crimson expressing cassette $\mathbf{P_{CL}}$ -R1-E2Crimson was effective in raising the strength of fluorescence, it is also required to proportionately raise the level of lambda repressor expressing cassettes to avoid compromising deterioration of Fold changes & Σ Leakage values. It was observed that among all the constructs of BNeu AS5 so far, the highest fold changes and least Σ Leakage values were obtained in AS5A where the E2crimson expressing cassette PRL-R1-E2Crimson & lambda repressor expressing cassettes, $\mathbf{P_{LUX}}$*-R1-cI* & $\mathbf{P_{LtetO1}}$-R1-cI* were in equal proportions in the same plasmid with ColE1 origin. We decided that if the entire content within the MCS of this plasmid could to be transferred to a higher copy pUC origin plasmid, we could raise the E2Crimson levels while also maintaining the relative proportion of Lambda repressors expressing cassettes within the cell. This modification gave rise to construct AS5G.
4	AS5F Figure S7(F) (Rejected)		

5	AS5G Figure S7(G) (Accepted)	**All parameters reached threshold values & the construct was accepted as final for Bactoneuron AS5.** ***Comment:*** *Highest leakage was obtained in the [0,0,0] input state probably due to relatively lower amounts of LacI & tetR within the cell expressing from the genome as compared to the high number of their binding sites present in high copy pUC origin plasmid.*	**NA**
1,2	AS6A Figure S7(G) (Accepted in Iteration 1, Rejected after Iteration 2)	AS6A passes all the thresholds for the parameters that we had set. Although, its E2Crimson Fluorescence intensity is not within +/-10% of that of AS4 it satisfies the alternative criteria of a minimum of 0.8 times fluorescence intensity of the highest Fluorescence observed in the iteration set by virtue of being of the highest intensity itself. However, in the subsequent iteration 2, low relative fluorescence from AS5B, AS5C & AS7B (0.23, 0.21 & 0.32 resp.) compelled us to reduce the fluorescence intensity of BNeu AS6 so that the differences in intensity could be minimized up to the threshold values. In the present design, one cI* expressing cassette, $\mathbf{P_{LLacO1}}$-R1-cI* is under ColE1 origin while the other cassette $\mathbf{P_{LUX}}$*****-R1-cI* is under p15A origin. We reckoned that if the later is shifted onto a ColE1 plasmid, its inherent leakage could be used to our advantage to reduce fluorescence intensity.	$\mathbf{P_{LUX}}$*****-R1-cI* cassette present under p15A origin in AS6A was shifted to the ColE1 origin plasmid with the speculation that the inherent leakage of $\mathbf{P_{LUX}}$***** could work to our advantage to diminish the amount of E2Crimson expressing from PRT-R1-E2Crimson cassette at the [0,1,0] input state. Other bio-parts were left as before. This modification generated construct AS6B.
3	AS6B Figure S8(B) (Rejected)	A reduction in fluorescence intensity was observed in AS6B compared to AS6A but the modification also resulted in an unacceptable drop in fold change to 6.25. From the characterization graphs, it was evident that there is considerable leakage whenever aTc is present in the system (max leakage in [0,1,1] input state) suggesting an inability of cI* in the cell to sufficiently suppress $\mathbf{P_{CT}}$ when it is activated by aTc. Moreover, an attempt to increase the Fluorescence of Construct AS7B from the previous iteration 2 (see below) resulted in a huge rise in fluorescence in AS7C in the present iteration.The relative fluorescence of AS6B was 0.61 times AS7C which was below 0.8. Also, AS6B's fluorescence was 12.27% below that of AS4 which too was below threshold. This demanded a reduction in fluorescence from BNeu AS7 while also a rise in that of AS6 seemed beneficial	The low fold change in AS6B could be due to the inability of the cI* to sufficiently suppress $\mathbf{P_{CT}}$ when it is activated by aTc. Given that maximum leakage was observed in [0,1,1] input state, a deficient repression especially by the $\mathbf{P_{LLacO1}}$-R1-cI* cassette was indicated. For this, we replaced the frameshifted version with the wild-type version cI^{w} under promoter $\mathbf{P_{LLacO1}}$, & a weak RBS R3 was used in place of R1 in accordance with our previous work [ref]. $\mathbf{P_{LUX}}$*****-R1-cI* was returned under p15A origin as was in AS6A given that it otherwise had good fold changes with this arrangement. To bring fluorescence levels nearer to AS7C, 1 & 2 additional cassette(s) of $\mathbf{P_{CT}}$ -R1-E2Crimson were added under p15A, upstream of $\mathbf{P_{LUX}}$*****-R1-cI* in addition to the above changes to generate AS6C & AS6D resp.
4	AS6C Figure S8(C) (Rejected)	Although all other parameters satisfied their respective thresholds, AS6C overshot the Fluorescence intensity values of AS4 by 31.91% which was unacceptable. At iteration 4, it assumed the highest relative Fluorescence (AS6D attained 1.22 times higher fluorescence than AS6C, but wasn't considered since it fared worse in terms of % variation of Fluorescence w.r.t AS4). From the good values of fold change & Σ Leakage, it was deduced that the arrangement of cassettes, $\mathbf{P_{LLacO1}}$-R3-cI^{w} & $\mathbf{P_{LUX}}$*****-R1-cI* with both under p15A origin could repress additional OR1 sites on the multiple $\mathbf{P_{CT}}$ -R1-E2Crimson cassettes well, at relevant induction conditions. So this arrangement was accepted for further iterations. The fluorescence intensity, however, was required to be reduced & simply removing the extra e2Crimson expressing cassettes from p15A origin seemed promising.	The extra $\mathbf{P_{CT}}$ -R1-E2Crimson cassette was removed from the p15A origin plasmid to tune down fluorescence intensity levels. Only the cassette under ColE1 origin was retained. Arrangements of all other bio-parts were not disturbed. This yielded the construct AS6E. We went ahead to create another construct of BNeu AS6 in iteration 5, where, keeping everything else same as AS6E (above), we shifted the $\mathbf{P_{LUX}}$*****-R1-cI* cassette to under ColE1 origin from p15A, hoping that the small leakages of E2Crimson at [1,0,1], [0,1,1] & [1,1,1] input states could be further reduced using the increased leakage of cI* from $\mathbf{P_{LUX}}$***** to our benefit. This generated construct AS6F.
4	AS6D Figure S8(C) (Rejected)	The Fluorescence Intensity of AS6D overshot that of AS4 by 60.58%, a value which was unacceptable & worse than AS6C. So AS6D wasn't considered further and was rejected.	**NA**
5	AS6E Figure S8(E) (Accepted)	**All parameters reached threshold values & the construct was accepted as final for Bactoneuron AS6.** ***Comment:*** *Although AS6F also yielded parameters all of which passed the corresponding threshold values, AS6E was considered final for further use because AS6E yielded comparatively better values overall.*	**NA**
5	AS6F Figure S8(E) (Acceptable but Not used)	AS6F yielded parameters all of which passed the corresponding threshold values. However, it wasn't used further because AS6E yielded comparatively better values overall.	**NA**

1	**AS7A** **Figure S9(A)** **(Rejected)**	The fluorescence level was just 0.14x of AS6A (which had highest fluorescence intensity in the iteration set) & 83.69% below that of AS4. The fold change & Σ Leakage values were far from acceptable. This could be happening due to overall low levels of E2Crimson expressing from the $\mathbf{P_{CH}}$ promoter in the "ON" state of the construct at [1,0,0] input state probably due to leakage of cI* from its expression cassettes. This initial design was based on previous work where $\mathbf{P_{CH}}$ promoter expressing E2Crimson from p15A & cI* expressing from either $\mathbf{P_{LTetO1}}$ or $\mathbf{P_{LLacO1}}$ from ColE1 showed good kinetics. However here there are 2 cI* expressing cassettes $\mathbf{P_{LLacO1}}$ -R1-cI* & $\mathbf{P_{LTetO1}}$-R1-cI* both in ColE1 while $\mathbf{P_{CH}}$ -R1-E2Crimson was in p15A. So small leakages of cI* from the 2 cassettes could add up and greatly affect E2Crimson levels.	To reach the threshold of Fluorescence intensity within +/- 10% of that of AS4 OR a minimum of 0.8 times the highest Fluorescence observed in the iteration set. we needed to raise the AS7A fluorescence levels in its "ON" state. For this, we tried to reduce the levels of Lambda repressor leakage by placing its expressing promoters $\mathbf{P_{LTetO1}}$ & $\mathbf{P_{LLacO1}}$ in p15A instead ColE1. We also replaced the frameshifted version of lambda repressor (cI*) with its wild-type i.e., cI^{w}. Moreover, we replaced the strong RBS R1 with two weak RBS's, R1 for promoter $\mathbf{P_{LTetO1}}$ & R2 for promoter $\mathbf{P_{LLacO1}}$. The weak RBSs were employed in accordance with our previous work for tuning cI levels expressing from these same promoters. $\mathbf{P_{CH}}$ -R1-E2Crimson remained in p15A. This led to the construct AS7B.
2	**AS7B** **Figure S9(B)** **(Rejected)**	Compared to AS7A, there was improvement seen in terms of Fluorescence intensity which was now 0.32x the fluorescence intensity of AS6A (which had the highest fluorescence intensity in the iteration set) and variation from AS4 decreased to -63.03%. However, this was still far from the thresholds. Additionally, the fold change and Σ Leakage values remained almost equally bad which may have affected the inducer weights that were unsatisfactory too. This indicated that lambda repressor expression & its leakage levels may not be the primary issue in context of relative Fluorescence intensity. Instead of modulating the copy number of lambda repressor expressing cassettes, a more effective way to improve fluorescence intensity levels and fold change could be by raising the copy number of $\mathbf{P_{CH}}$ -R1-E2Crimson.	We reverted back to the original design of AS7A where Lambda repressor expressed from 2 cassettes $\mathbf{P_{LLacO1}}$-R1-cI* & $\mathbf{P_{LTetO1}}$-R1-cI* in ColE1 origin plasmid and instead shifted the $\mathbf{P_{CH}}$ -R1-E2Crimson cassette from p15A to ColE1.
3	**AS7C** **Figure S9(C)** **(Rejected)**	A dramatic increase in fluorescence was observed with AS7C overshooting AS4 by 44.57%, much beyond our threshold. This made AS7C the highest fluorescing construct in iteration 3. Apart from still having subpar fold changes and Σ Leakage values, this high fluorescence made the relative fluorescence of other BNeu constructs fall below 0.8 (0.69 for AS4, 0.29 for AS5D & 0.61 for AS6B). To bring relative fluorescence values within threshold limits it was necessary to reduce Fluorescence intensity from AS7C and/or increasing the same in others. Besides, subpar fold change & Σ Leakage values suggested that cI*, the frame-shifted version of lambda repressor was unable to efficiently repress all OR1 sites on $\mathbf{P_{CH}}$ promoters at relevant induction states. Using the wild type cI^{w} may improve this situation.	We retained $\mathbf{P_{CH}}$ -R1-E2Crimson cassette in ColE1 but replaced the $\mathbf{P_{LLacO1}}$-R1-cI* & $\mathbf{P_{LTetO1}}$-R1-cI* under ColE1 origin plasmid with $\mathbf{P_{LLacO1}}$-R3-cI & $\mathbf{P_{LTetO1}}$-R2-cI under p15A origin. Here we used the wild type Lambda repressor cI^{w} & weak RBS's R2 & R3 were employed to tune cI levels with respective promoters in accordance with our previous work [ref].
4, 5	**AS7D** **Figure S9(D)** **(Accepted)**	**All parameters reached threshold values at iteration 5 & the construct was accepted as final at iteration 5 for Bactoneuron AS7.** ***Comment:*** *In iteration 4 AS6C was considered as the construct with the highest fluorescence relative to which AS7D had a fluorescence of 0.72, which is below our threshold of 0.8. However, in the next & final iteration 5, AS6C was rejected for a new construct for Bactoneuron AS6 (AS6E) and now the construct with the highest fluorescence was AS4 relative to which the fluorescence of AS7D was 0.95 which is well within acceptable values.*	**NA**

***Fluor. Intensity of construct @ "ON" state/ Max Fluor. Intensity observed in iteration set** (Max 20% variation of signal between "ON" states of the highest & lowest fluorescing constructs of the iteration

Table S2: Concentrations of inducers* used in different induction conditions for characterization, Dose response & Validation experiments of Bactoneurons

			Concentration Points (nM)															
Sl. No.			1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
AS1	Dose Response Exp	AHL	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
		IPTG	0	10	100	5 x 10^2	10^3	$2x10^3$	$5x10^3$	10^4	$5x10^4$	10^5	$5x10^5$	10^6	$5x10^6$	10^7		
		aTc	0	2.16	6.48	12.96	21.6	32.4	43.2	64.81	86.42	129.62	216.04	302.456	432.08			
	Validation Exp.	AHL	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
		IPTG	0	200	600	1800	$4x10^3$	$8x10^3$	$15x10^3$	$2x10^4$	$3x10^4$	$4x10^4$	$5.5x10^4$	$7x10^4$	$8.9x10^4$	$3x10^5$	$9x10^5$	$8x10^6$
		aTc	0	0.4	0.86	3.24	9.72	25.92	38.89	51.85	69.13	90.74	112.34	138.26	167.67	194.44	280.85	388.87
AS2	Dose Response Exp	AHL	0	0.01	0.02	0.2	0.5	1	8	30	120	600	5000					
		IPTG	0	2	5	10	100	1000	5000	$12x10^3$	10^5	$6x10^5$	$15x10^5$	$4x10^6$	10^7			
		aTc	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	Validation Exp.	AHL	0	0.005	0.02	0.04	0.1	0.24	0.4	0.6	0.9	1.2	1.6	2.1	2.45	25	300	4000
		IPTG	0	50	150	500	1500	3600	5500	7500	10^4	$1.25x10^4$	$1.6x10^4$	$2x10^4$	$2.38x10^4$	$9x10^4$	$8x10^5$	$8x10^6$
		aTc	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
AS3	Dose Response Exp	AHL	0	0.01	0.02	0.05	0.1	0.5	1	3	8	30	120	600	5000			
		IPTG	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
		aTc	0	1.08	2.16	4.32	8.64	17.28	30.24	43.2	64.81	86.42	216.04	324.06	432.08			
	Validation Exp.	AHL	0	0.005	0.02	0.04	0.09	0.15	0.3	0.45	0.6	0.9	1.25	1.6	2.04	25	300	4000
		IPTG	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
		aTc	0	0.32	0.65	2.48	7.56	19.44	32.4	45.37	58.33	73.45	90.74	116.66	135.46	172.83	259.25	388.87
AS4	Dose Response Exp	AHL	0	0.02	0.05	0.1	0.2	0.5	1	3	8	30	120	1000	5000			
		IPTG	0	100	300	10^3	$3x10^3$	$8x10^3$	$14x10^3$	$5x10^4$	$2x10^5$	$8x10^5$	10^7					
		aTc	0	2.16	6.49	12.96	21.6	32.4	43.2	64.812	86.416	129.624	216.04	432.08				
	Validation Exp.	AHL	0	0.03	0.09	0.3	0.7	1.4	2.5	3.5	5.2	7	10	15	18.15	100	500	4000
		IPTG	0	50	140	400	900	2500	4500	7500	10^4	$1.5x10^4$	$2.1x10^4$	$2.5x10^4$	$2.945x10^4$	10^5	$9x10^5$	$8x10^6$
		aTc	0	0.11	0.43	1.73	5.4	10.8	19.44	30.2456	38.89	51.8496	69.1328	90.7368	122.2354	172.832	259.248	388.872
AS5A	Dose Response Exp.	AHL	0	0.01	0.05	0.1	0.3	1	8	30	120	5000						
		IPTG	0	50	100	300	1000	3000	8000	$5x10^4$	$2x10^5$	10^7						
		aTc	0	1.08	4.32	8.64	17.28	25.92	38.89	54.01	75.614	432.08						
AS5B	Dose Response Exp.	AHL	0	0.05	0.1	0.2	1	3	6	15	120	600	5000					
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$2x10^5$	$8x10^5$	10^7				
		aTc	0	1.08	2.16	4.32	8.64	17.28	25.92	38.89	54.01	172.832	432.08					
AS5C	Dose Response Exp.	AHL	0	0.05	0.1	0.2	1	3	6	15	45	120	600	5000				
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$8x10^5$	10^7					
		aTc	0	1.08	2.16	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.832	432.08				
AS5D	Dose Response Exp.	AHL	0	0.01	0.02	0.05	0.1	0.2	0.5	1.5	3	8	50	600	$5x10^3$			
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$2x10^5$	$8x10^5$	10^7					
		aTc	0	1.08	2.16	4.32	17.28	25.92	38.89	54.01	172.83	432.08						
AS5E	Dose Response Exp.	AHL	0	0.1	0.2	0.5	1.5	3	8	20	50	120	600	$5x10^3$				
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$2x10^5$	$8x10^5$	10^7					
		aTc	0	1.08	2.16	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.832	432.08				
AS5F	Dose Response Exp.	AHL	0	0.05	0.1	0.33	1	3	6	15	45	120	600	$5x10^3$				
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$8x10^5$	10^7						
		aTc	0	1.08	2.16	4.32	8.64	17.28	38.89	54.01	75.614	172.832	432.08					
AS5G	Dose Response Exp.	AHL	0	0.002	0.005	0.01	0.02	0.05	0.1	0.2	0.5	1	8	30	120	$5x10^3$		
		IPTG	0	100	500	10^3	$5x10^3$	10^4	$2.5x10^4$	$5x10^5$	10^6	$2x10^6$	$5x10^6$	10^7				
		aTc	0	0.02	0.04	0.21	1.08	3.24	6.48	12.9624	38.89	54.01	216.04	432.08				
	Validation Exp.	AHL	0	0.006	0.02	0.06	0.12	0.24	0.45	0.6	0.9	1.2	1.6	2	2.66	10	200	$4x10^3$
		IPTG	0	90	200	800	$2.4x10^3$	$4.5x10^3$	$6x10^3$	$9x10^3$	$14x10^3$	$18x10^3$	$22x10^3$	$28x10^3$	$33.05x10^3$	10^5	$9x10^5$	$8x10^6$
		aTc	0	0.03	0.11	0.43	1.29	3.02	4.54	6.91328	9.7218	11.66616	14.69072	17.28	23.397	51.85	172.83	388.87
AS6A	Dose Response Exp.	AHL	0	0.01	0.02	0.05	0.1	0.3	1	3	8	30	120	5000				
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$8x10^5$	10^7					
		aTc	0	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.83	432.08						
AS6B	Dose Response Exp.	AHL	0	0.01	0.02	0.05	0.1	0.3	1	3	30	120	$5x10^3$					
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$5x10^4$	$2x10^5$	$8x10^5$	107					
		aTc	0	4.32	8.64	17.28	25.92	38.89	54.01	172.83	432.08							
AS6C	Dose Response Exp.	AHL	0	0.1	0.2	0.5	1.5	3	8	20	50	120	600	$5x10^3$				
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$2x10^5$	$8x10^5$	10^7				
		aTc	0	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.83	432.08						
AS6D	Dose Response Exp.	AHL	0	0.05	0.1	0.2	1	3	6	15	45	120	600	5000				
		IPTG	0	50	100	300	10^3	$3x10^3$	$2x10^4$	$5x10^4$	2 $x10^5$	8 $x10^5$	10^7					
		aTc	0	4.32	8.64	17.28	25.92	38.89	54.01	75.614	432.08							
AS6E	Dose Response Exp.	AHL	0	0.005	0.01	0.02	0.06	0.5	1.5	4	10	25	60	150	400	10^3	$2x10^3$	$5x10^3$
		IPTG	0	10	20	50	100	500	10^3	$5x10^3$	10^4	$2.5x10^4$	$5x10^4$	10^5	$5x10^5$	10^6	10^7	
		aTc	0	2.16	4.32	10.8	17.28	25.92	38.89	51.8496	64.812	129.624	216.04	280.852	345.664	432.08		
	Validation Exp.	AHL	0	0.04	0.12	0.36	1	2	3.6	4.8	7.2	9.6	14	18	21.32	50	500	$4x10^3$
		IPTG	0	30	90	270	810	1500	2400	3600	4800	7200	$9x10^3$	$12x10^3$	$15.98x10^3$	$9x10^4$	$8x10^5$	$8x10^6$
		aTc	0	0.32	0.78	2.59	7.78	16.2	21.6	32.4	43.208	54.01	69.133	90.74	104.65	172.83	259.25	388.87
AS6F	Dose Response Exp.	AHL	0	0.1	0.2	0.5	1.5	3	8	20	50	120	$5x10^3$					
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$8x10^5$	10^7					
		aTc	0	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.83	432.08						

AS7A	Dose Response Exp.	AHL	0	0.5	1.5	3	8	20	50	120	600	5000						
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$2x10^5$	$8x10^5$	10^7				
		aTc	0	1.08	2.16	4.32	8.64	17.28	38.89	54.01	75.614	172.832	432.08					
AS7B	Dose Response Exp.	AHL	0	0.2	1	3	6	15	45	120	600	5000						
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$2x10^4$	$5x10^4$	$2x10^5$	$8x10^5$	10^7				
		aTc	0	1.08	2.16	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.832	432.08				
AS7C	Dose Response Exp.	AHL	0	0.5	1.5	3	8	20	50	120	600	$5x10^3$						
		IPTG	0	50	100	300	10^3	$3x10^3$	$8x10^3$	$5x10^4$	$2x10^5$	10^7						
		aTc	0	1.08	2.16	4.32	8.64	17.28	25.92	38.89	54.01	75.614	172.832	432.08				
AS7D	Dose Response Exp	AHL	0	0.05	0.1	0.2	0.5	1.5	4	10	60	150	400	10^3	$2x10^3$	$5x10^3$		
		IPTG	0	10	20	50	100	500	10^3	$5x10^3$	10^4	$5x10^4$	10^5	$5x10^5$	10^6	10^7		
		aTc	0	0.22	0.43	1.08	2.16	4.32	17.28	25.92	38.89	51.8496	64.812	129.624	216.04	432.08		
	Validation Exp.	AHL	0	0.04	0.12	0.36	1	2.5	3.6	5.4	8	12	16	20	23.44	80	500	4000
		IPTG	0	40	120	360	$1.2x10^3$	$2.4x10^3$	$4.8x10^3$	$7.2x10^3$	$9.6x10^3$	$12x10^3$	$15x10^3$	$18x10^3$	$22.5x10^3$	$9x10^4$	$8x10^5$	$8x10^6$
		aTc	0	0.13	0.39	1.17	3.24	6.05	11.67	15.55	21.604	32.4	43.208	56.17	71.77	108.02	194.44	388.87
AS8	Dose Response Exp	AHL	0	0.01	0.02	0.05	0.1	0.3	1	3	8	30	120	500	1500	5000		
		IPTG	0	2	5	10	100	10^3	$5x10^3$	$1.2x10^3$	$1x10^5$	$6x10^5$	$1.5x10^6$	$4x10^6$	10^7			
		aTc	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
	Validation Exp.	AHL	0	0.03	0.08	0.2	0.6	1.6	2.8	4	6	9	12	15	17.65	50	400	$4x10^3$
		IPTG	0	30	90	300	1200	3600	5500	7500	10^4	$1.25x10^4$	$1.6x10^4$	$2.1x10^4$	$2.361x10^4$	$9x10^4$	$8x10^5$	$8x10^6$
		aTc	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
AS9	Dose Response Exp	AHL	0	0.002	0.005	0.01	0.1	0.5	2	5	10	100	500	$2x10^3$	$5x10^3$			
		IPTG	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
		aTc	0	0.43	1.08	2.16	6.48	12.96	25.92	38.89	54.01	75.614	172.832	302.456	432.08			
	Validation Exp.	AHL	0	0.03	0.09	0.3	0.7	1.4	2.5	3.5	5.2	7	9	12	15.54	90	600	4000
		IPTG	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
		aTc	0	0.33	0.65	2.48	7.56	19.44	30.24	43.208	56.17	69.13	86.42	108.02	128.85	168.51	259.25	388.87

*concentration of AHL, IPTG and aTc in nM, µM, and ng/mL respectively

Table S3: Translation initiation rate calculated from RBS calculator [32]

RBS Name	Operating Promoter	Protein of Translational regulation	Translation Initiation rate (a.u.)	Sequence
R1	P_{LLacO1}/P_{LTetO1}	cI^w	40767	*GAATTC*ATTAAAGAGGAGAAA*GGTACC*
R2	P_{LLacO1}/P_{LTetO1}	cI^w	2091	*GAATTC*ATTTTTGAGGAGTTC*GGTACC*
R3	P_{LLacO1}/P_{LTetO1}	cI^w	518	*GAATTC*ATTCGGGAGGAGTGC*GGTACC*
R1	P_{HT}	EGFP	46	*GAATTC*ATTAAAGAGGAGAAA*GGTACC*
R4	P_{HT}	EGFP	10	*GAATTC*ATTGGAGAGGAGTCC*GGTACC*

Table S4: DNA sequences of Bio-parts & primers used in this study. lacO1, tetO2, Lux box, OR1 sites are coloured in orange, teal, pink & purple respectively. Transcription start site is shown in bold blue. -10 and -35 hexamers are underlined. Each promoter is flanked by XhoI and EcoRI restriction sites (marked in brown italics). RBS's are flanked by EcoRI & KpnI restriction sites (marked in brown italics). Plasmids generated in this study are detailed in Figure S6, S7, S8 & S9.

Part Name	Sequence (5' – 3')	Source
P_{LT}	*CTCGAG*TCCCTATCAGTGATAGAGATTGACATTGTGAGCGGATAACAAGATACTGAGCACATCCCTATCAGTGATAGAGAGATAATTGTGAGCGGATAACAATT*GAATTC*	Plasmid: pP_{IAA}7A1EGFP Ref [15]
P_{HT}	*CTCGAG*ACCTGTAGGATCGTACAGGTTTACGTCCCTATCAGTGATAGAGTATAGTCGAATAAATCCCTATCAGTGATAGAGA*GAATTC*	Plasmid: pP_{AAH} C3RBSHEGFP Ref [15]
P_{HL}	*CTCGAG*ACCTGTAGGATCGTACAGGTTTACGTTTGTGAGCGGATAACAATATAGTCGAATAAATTGTGAGCGGATAACAATT*GAATTC*	Plasmid: pP_{IAH} C3EGFP(R) Ref [15]
P_{HLT}	*CTCGAG*ACCTGTAGGATCGTACAGGTTTACGTTTGTGAGCGGATAACAATATAGTCGAATAAATCCCTATCAGTGATAGAGAGATGATTCCCTATCAGTGATAGAGAGATGATAATTGTGAGCGGATAACAATT*GAATTC*	Plasmid: pAR-$P_{luxATTA}$EGFP Ref [9]

P_{CL}	*CTCGAG*AATTGTGAGCGGATAACAATTGACATTGTGAGCGGATAACAAGATACTGAGCACATCTACCTCTGGCGGTGATAGATGATTACCTCTGGCGGTGATAGATGATTACCTCTGGCGGTGATA*GAATTC*	Plasmid: pP_{INA}7A2EGFP Ref [15]
P_{CT}	*CTCGAG*TACCTCTGGCGGTGATAGATTACCTCTGGCGGTGATATTGACATCCCTATCAGTGATAGAGATACTGAGCACATCCCTATCAGTGATAGAGA*GAATTC*	Plasmid: pP_{ANI}2A2EGFP Ref [15]
P_{CH}	*CTCGAG*ACCTGTAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTATAGTCGAATAAATACCTCTGGCGGTGATAGATGATTACCTCTGGCGGTGATAGATGATTACCTCTGGCGGTGATA*GAATTC*	Plasmid: pHC-1_EGFP Ref [16]
P_{lux*}	*CTCGAG*ACCTGTAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTACTTTCGAATAAA*GAATTC*	Plasmid: pX*C3EGFP Ref [15]
P_{lux}^{w}	*CTCGAG*ACCTGTAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTATAGTCGAATAAA*GAATTC*	Plasmid: pXC3cIfm Ref [15]
$P_{L\ TetO1}$	*CTCGAG*TCCCTATCAGTGATAGAGATTGACATCCCTATCAGTGATAGAGATACTGAGCACATCAGCAGGACGCACTGACC*GAATTC*	Ref [19]
$P_{L\ LacO1}$	*CTCGAG*AATTGTGAGCGGATAACAATTGACATTGTGAGCGGATAACAAGATACTGAGCACATCAGCAGGACGCACTGACC*GAATTC*	Ref [19]
P_{J23102}	*CTCGAG*TTGACAGCTAGCTCAGTCCTAGGTACTGTGCTAGC*GAATTC*	Plasmid: pEM-J23102-luxR Ref [9]
EGFP Forward sequencing Primer	CAAGGGCGAGGAGCTGTT	This Study
EGFP Reverse sequencing Primer	CCATGCCGAGAGTGATCC	This Study
E2Crimson Forward sequencing Primer	TGGATAGCACTGAGAACGTCAT	This Study
E2Crimson Reverse sequencing Primer	ACCACGGTGTAGTCCTCGTT	This Study
LuxR Forward sequencing Primer	ATGCCGACGACACATACAGA	This Study
LuxR Reverse sequencing Primer	TGATGCCTGGCTCTAGTAGTGA	This Study
cI^{i} Forward sequencing Primer	AAAGAAACCATTAACACAAGAGCA	This Study
cI^{i} Reverse sequencing Primer	TCAGCCAAACGTCTCTTCAG	This Study

Table S5: List of scaled relative concentration points to perform simulations of Bactoneurons

0	0.0012	0.003	0.006	0.01	0.02	0.03	0.04	0.05	0.06	0.07	0.08	0.09	0.1
0.12	0.14	0.16	0.18	0.2	0.22	0.24	0.26	0.28	0.3	0.33	0.36	0.39	0.42
0.45	0.48	0.51	0.54	0.57	0.6	0.64	0.68	0.72	0.76	0.8	0.84	0.88	0.92
0.96	1												