Splicing factor SRSF1 is essential for homing of precursor spermatogonial stem cells in mice

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Abstract

Spermatogonial stem cells (SSCs) are essential for continuous spermatogenesis and male fertility. The underlying mechanisms of alternative splicing (AS) in mouse SSCs are still largely unclear. We demonstrated that SRSF1 is essential for gene expression and splicing in mouse SSCs. Crosslinking immunoprecipitation and sequencing (CLIP-seq) data revealed that spermatogonia-related genes (e.g., *Plzf*, *Id4*, *Setdb1, Stra8*, *Tial1*/*Tiar*, *Bcas2*, *Ddx5*, *Srsf10*, *Uhrf1*, and *Bud31*) were bound by SRSF1 in the mouse testes. Specific deletion of *Srsf1* in mouse germ cells impairs homing of precursor SSCs leading to male infertility. Whole-mount staining data showed the absence of germ cells in the testes of adult conditional knockout (cKO) mice, which indicates Sertoli cell-only syndrome (SCOS) in cKO mice. The expression of spermatogonia-related genes (*Gfra1*, *Pou5f1*, *Plzf*, *Dnd1*, *Stra8*, and *Taf4b*) was significantly reduced in the testes of cKO mice. Moreover, multiomics analysis suggests that SRSF1 may affect survival of spermatogonia by directly binding and regulating *Tial1*/*Tiar* expression through AS. In addition, immunoprecipitation mass spectrometry (IP-MS) and co-immunoprecipitation (Co-IP) data showed that SRSF1 interacts with RNA splicing-related proteins (SART1, RBM15, and SRSF10). Collectively, our data reveal the critical role of SRSF1 in spermatogonia survival, which may provide a framework to elucidate the molecular mechanisms of the posttranscriptional network underlying homing of precursor SSCs.

Keywords

SRSF1, alternative splicing, infertility, Sertoli cell-only syndrome, precursor spermatogonial stem cells, homing of precursor SSCs

Introduction

SCOS, also known as del Castillo syndrome or germ cell aplasia, is one of the most common causes of severe non-obstructive azoospermia (NOA) (Wang et al., 2023). SCOS is the presence of only Sertoli cells in the testicular tubules of the testes, with no germ cells present (Juul et al., 2014; Wang *et al.*, 2023). It is well known that abnormal self-renewal and differentiation of SSCs lead to SCOS (Kanatsu-Shinohara and Shinohara, 2013; La and Hobbs, 2019). In mice, gonocytes begin homing at 0 to 3 days postpartum (dpp) and then develop into SSCs at 4 to 6 dpp for continuous self-renewal and differentiation (Lee and Shinohara, 2011; McLean et al., 2003; Tan and Wilkinson, 2020). The mechanisms regulating homing of precursor SSCs are hence crucial for forming SSC pools and establishing niches (Oatley and Brinster, 2012). Spermatogonia migrate to form two distinct subtypes in mice. The first subtype develops into precursor SSCs that provide an SSC population for adult spermatogenesis, whereas the second subtype transitions directly to differentiated spermatogonia that contribute to the first round of spermatogenesis but do not self-renew (Kluin and Derooij, 1981; Law et al., 2019). Therefore, homing of precursor SSCs to establish niches is essential for SSC self-renewal and differentiation.

Many transcription factors (e.g., FOXO1, PLZF, POU5F1, TAF4B, CHD4, BCL6B, BRACHYURY, ETV5, ID4, LHX1, POU3F1, DMRT1, NGN3, SOHLH1, SOHLH2, SOX3, and STAT3) promote SSC self-renewal and differentiation (Cafe et al., 2021; Song and Wilkinson, 2014). However, the molecular mechanisms of the posttranscriptional network underlying homing of precursor SSCs are not sufficiently clear. Previous studies have identified the key RNA-binding proteins DND1 and DDX5 in SSCs with a unique and dominant role in posttranscriptional regulation (Legrand et al., 2019; Yamaji et al., 2017). Surprisingly, recent studies have found that the RNA-binding proteins SRSF10, UHRF1, BUD31, and BCAS2 regulate alternative splicing (AS) in mouse spermatogonia (Liu et al., 2022; Liu et al., 2017; Qin et al., 2023; Zhou et al., 2022). It is well known that testes are rich in AS events (Mazin et al., 2021; Venables, 2002). Thus, understanding the mechanisms of AS in human reproduction can provide new insights into clinical diagnosis. However, the underlying mechanisms of how AS functions in homing of precursor SSCs are still largely unclear.

Serine/arginine-rich splicing factor 1 (SRSF1; previously SF2/ASF) is a widely studied and important splicing factor involved in cancer progression, heart development, and thymus development (Du et al., 2021; Katsuyama et al., 2019; Katsuyama and Moulton, 2021; Liu et al., 2021; Lv et al., 2021; Qi et al., 2021; Xu et al., 2005). Our previous work has shown that SRSF1 deficiency impairs primordial follicle

formation during meiotic prophase I and leads to primary ovarian insufficiency (POI) (Sun et al., 2023b). However, the underlying mechanisms by which SRSF1 regulates pre-mRNA splicing in mouse SSCs remain unknown. A mouse model with *Srsf1* conditional deletion can effectively address this uncertainty. This study showed that specific deletion of *Srsf1* in mouse germ cells leads to NOA by impairing homing of mouse precursor SSCs. We further verified that SRSF1 directly binds and regulates *Tial1*/*Tiar* expression via AS, which may be critical for homing of mouse precursor SSCs.

Results

SRSF1 may have a vital role in posttranscriptional regulation in the testes

To investigate the role of SRSF1 in spermatogenesis, the dynamic localization of SRSF1 in the testis was evaluated. Fascinatingly, the results of SRSF1 and γH2AX co-staining revealed that SRSF1 was expressed during spermatogenesis (Figure 1A and S1). RT–qPCR and Western blotting results showed that the expression of SRSF1 fluctuated during the developmental stages of the testes (Figure 1B and 1C). Concurrently, the results of SRSF1 and PLZF co-staining revealed that SRSF1 was highly expressed in the nuclei of spermatogonia (Figure 1D). To further explore the function of SRSF1 in regulating SSC self-renewal and differentiation, CLIP-seq was performed in adult mouse testes (Sun et al., 2023a). GO enrichment analyses of the SRSF1 peak-containing genes revealed that spermatogenesis-related genes were regulated by SRSF1 (Figure 2A and Table S1). In combination with previous studies, we found that spermatogonia-related genes (*Plzf*, *Id4*, *Setdb1, Stra8*, *Tial1*/*Tiar*, *Bcas2*, *Ddx5*, *Srsf10*, *Uhrf1*, and *Bud31*) were bound by SRSF1. To provide in-depth insight into the binding of spermatogonia-associated genes, the SRSF1-binding peaks of the gene transcripts were shown by using IGV (Figure 2B). The co-staining results showed localization and expression of the spermatogonia-related proteins in mouse testes (Figure 2C). Together, these results suggested that SRSF1 may have a vital role in posttranscriptional regulation in the testes, particularly during spermatogonial development.

SRSF1 deficiency leads to SCOS

To define the specific involvement of SRSF1 in spermatogonia, we studied the physiological roles of SRSF1 in vivo using a mouse model. Considering that global *Srsf1* knockout is lethal in mice (Xu *et al.*, 2005), we used a conditional allele of *Srsf1* (*Srsf1*^{Fl}) in which exons 2, 3, and 4 of *Srsf1* are flanked by two *loxP* sites (Figure 3A). By crossing *Srsf1*^{Fl} and *Vasa*-Cre mice, we obtained *Vasa*-Cre *Srsf1*$^{Fl/del}$ mice with *Srsf1* deletion in germ cells (Figure 3A and 3B). We verified the absence of the SRSF1 protein

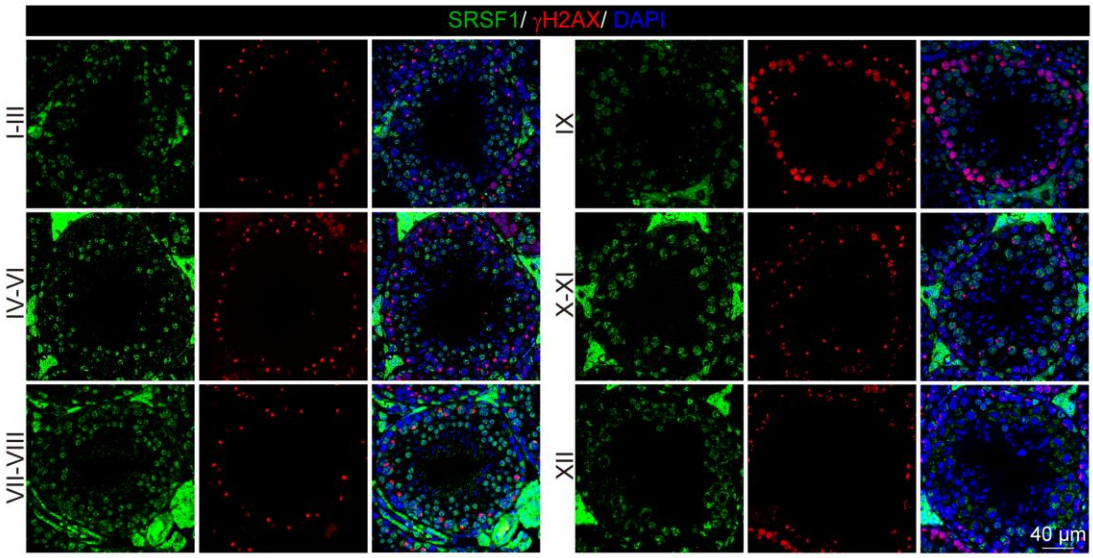


Figure S1 Dynamic localization of SRSF1 during spermatogenesis.

Co-immunostaining was performed using SRSF1 and γH2AX antibodies in adult mouse testes. DNA was stained with DAPI. Scale bar, 40 μm.

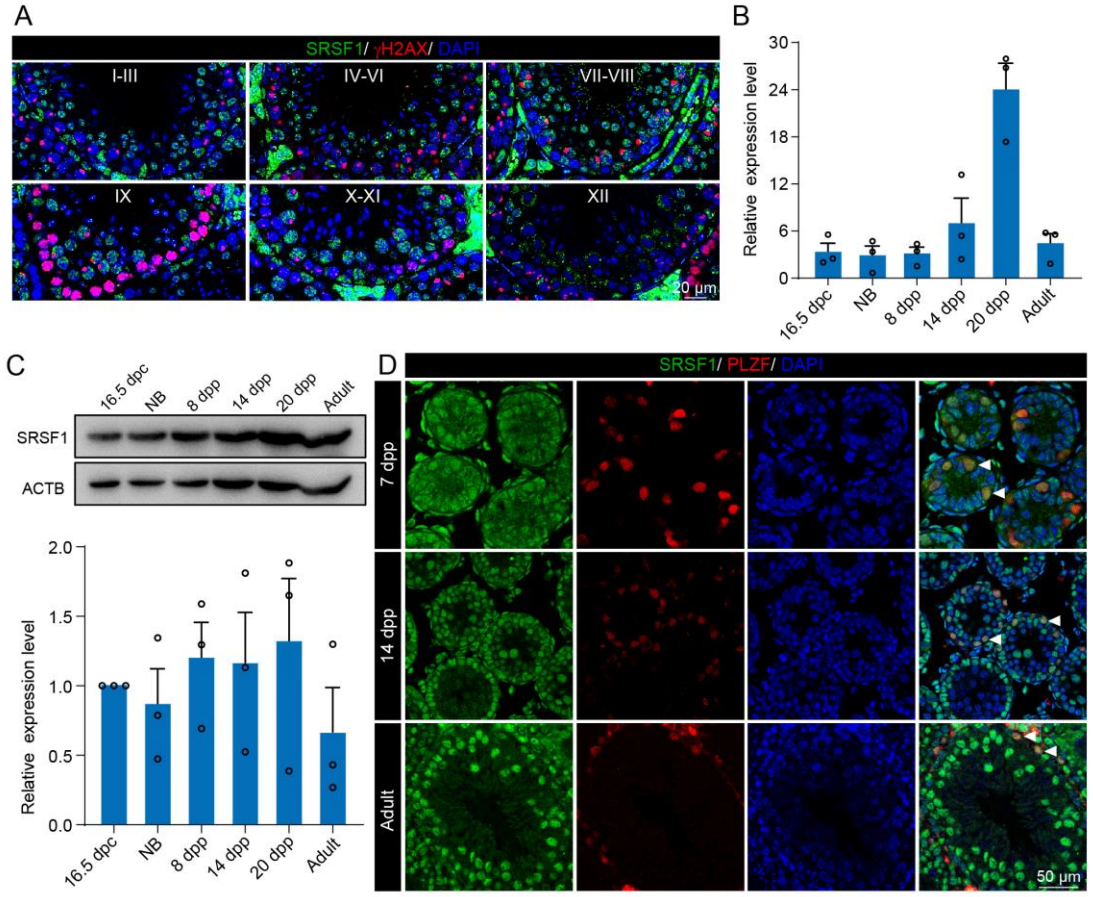


Figure 1 Expression and localization of SRSF1 in the testis of mice at different developmental stages.

(A) Dynamic localization of SRSF1 during spermatogenesis. Co-immunostaining was performed using SRSF1 and γH2AX antibodies from adult mouse testes. DNA was stained with DAPI. Scale bar, 20 μm.

(B) Expression of *Srsf1* in testes at different stages of development. The RT–qPCR data were normalized to *Gapdh*. *n*=3.

(C) Western blotting of SRSF1 expression in testes at different stages of development. ACTB served as a loading control. The value in 16.5 dpc testes were set as 1.0, and the relative values of testes in other developmental periods are indicated. n=3.

(D) Localization and expression of SRSF1 in spermatogonia. Co-immunostaining was performed using PLZF and SRSF1 antibodies in 7 dpp, 14 dpp, and adult mouse testes. DNA was stained with DAPI. Arrowheads, spermatogonia. Scale bar, 50 μm

in germ cells by co-immunofluorescence analyses with SRSF1 and PLZF antibodies (Figure 3C). Subsequently, the breeding experiment indicated that cKO mice had a standard mating capacity but that the absence of *Srsf1* led to complete infertility in males (Figure 3D). Histological examination of cKO epididymides revealed that sperm could not be found in the cauda epididymis (Figure 3E). Considering the limitations of sectioning, the cauda epididymal sperm count further validated this conclusion (Figure 3F). It was clear that spermatogenesis in the testes was severely impaired. Therefore, we focused our attention on the testes. The adult cKO mice were normal in size (Figure 3G). However, the sizes of cKO mouse testes were significantly reduced (Figure 3H). Histological examination of cKO testis sections showed that no germ cells could be visualized, and only a large number of Sertoli cells were observed in the testes of cKO mice (Figure 3I). Together, these results demonstrated that SRSF1 is critical for spermatogenesis and that its absence leads to SCOS.

Loss of SRSF1 impairs spermatogonia survival

To further confirm the absence of germ cells in the testes of cKO mice, PLZF and γH2AX co-staining was performed in adult mouse testes. These data suggested that SRSF1 deficiency impaired germ cell survival (Figure 4A). The results of VASA and TRA98 co-staining further confirmed this phenotype (Figure 4B). Considering the limitations of sectioning, we used whole-mount immunostaining to perform a comprehensive analysis and found that germ cells were indeed absent in the testes of adult cKO mice (Figure 4C). To dynamically analyse the loss of germ cells, we collected testes from 5 dpp, 7 dpp, and 14 dpp mice. Morphological results showed that the testes of 7 dpp and 14 dpp cKO mice were much smaller than those of Ctrl mice (Figure 5A). To determine the presence of germ cells in cKO testes,

VASA staining was performed in 5 dpp, 7 dpp, and 14 dpp Ctrl and cKO testes. The results showed that germ cells were still present in cKO mice but were significantly reduced in 7 dpp and 14 dpp cKO testes (Figure 5B). The quantifications of germ cells per tubule showed a significant reduction in the number of 7 dpp and 14dpp cKO testes, especially 14 dpp cKO testes (Figure 5B). In addition, TUNEL results showed that apoptosis significantly increased in cKO testes (Figure 5C). These data suggested that the absence of SRSF1 causes apoptosis in a large number of spermatogonia that are unable to survive.

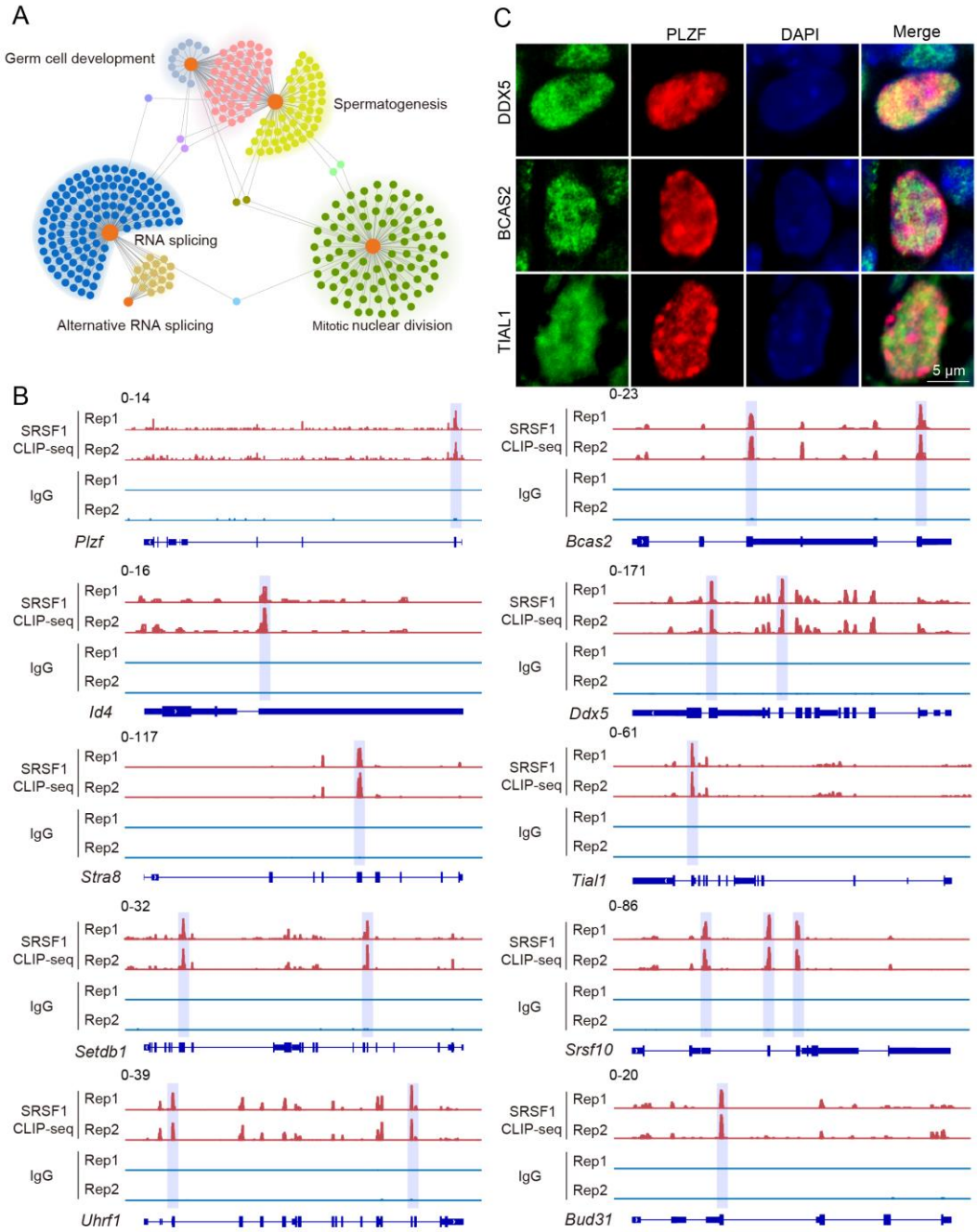


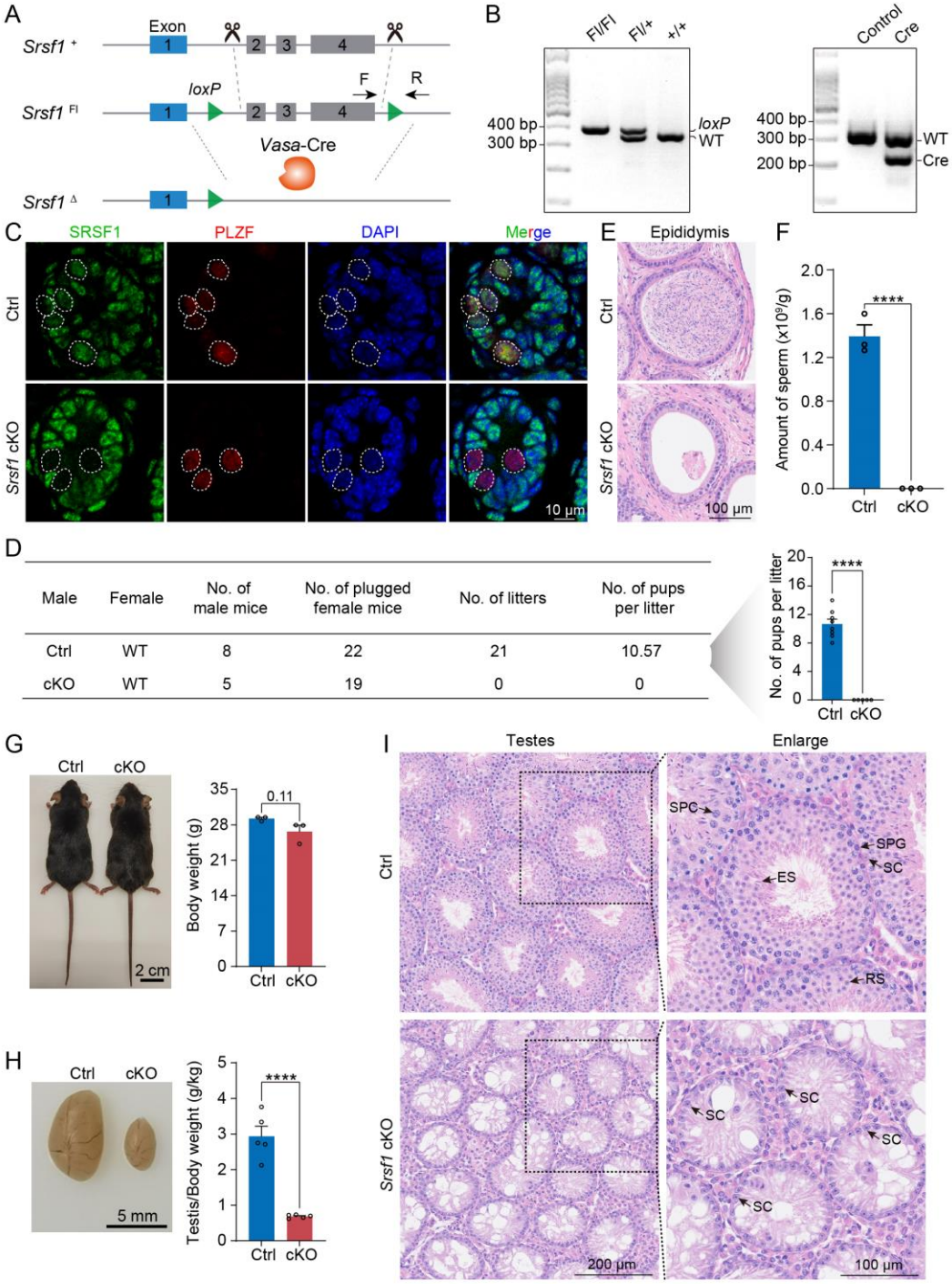
Figure 2 SRSF1-binding genes have an essential role in spermatogonia.

(A) Network showing GO enrichment analyses of SRSF1-binding genes.

167 (B) Representative genome browser views of spermatogonia-related gene transcripts bound by SRSF1. Higher peaks

168 are marked by a lavender area.

169 (C) Localization of the spermatogonia-related proteins in mouse testes. Scale bar, 5 μm.



170

171 **Figure 3 SRSF1 plays critical roles in spermatogenesis and male fertility.**

172 (A) *Vasa*-Cre mice were crossed with *Srsf1*$^{Fl/Fl}$ mice to generate *Srsf1* cKO mice. Cre-mediated deletion removed

173 exons 2, 3, and 4 of *Srsf1* and generated a null protein allele.

174 (B) Genotyping PCR was performed using *Vasa*-Cre and *Srsf1* primers.

175 (C) Co-immunostaining of SRSF1 and PLZF in 7 dpp Ctrl and cKO testes. DNA was stained with DAPI. Scale bar,
176 10 μm.
177 (D) Fertility test results showed a male infertility phenotype in the cKO mice (n= 5) compared to the Ctrl mice (n= 8).
178 The number of pups per litter was determined in the cKO (n= 5) and Ctrl (n= 8) mice.
179 (E) Haematoxylin-eosin-stained epididymis sections from adult Ctrl and cKO mice were obtained. Scale bar, 100
180 μm.
181 (F) Cauda epididymal sperm counting was performed. n=3.
182 (G) Normal body weight in cKO mice. The body sizes and weights of adult Ctrl and cKO mice are shown as the
183 mean ± SEM. n= 3.
184 (H) Testis atrophy in adult cKO mice. Testis sizes and weights/body of adult Ctrl and cKO mice are shown as the
185 mean ± SEM. n= 5.
186 (I) Haematoxylin-eosin-stained testis sections from adult Ctrl and cKO mice were obtained. Scale bar, left panel:
187 200 μm, right panel: 100 μm. SC, Sertoli cell; SPG, spermatogonium; SPC, spermatocyte; RS, round spermatid; ES,
188 elongated spermatid.
189 Unpaired Student's t-test determined significance; exact P value $P \geq 0.05$, ****$P < 0.0001$. The points and error bars
190 represent the mean ± SEM.
191
192 **Loss of SRSF1 impairs homing of precursor SSCs**
193 To further investigate the reason for the failure of spermatogonia to survive, we observed homing of
194 precursor SSCs in the testes of mice at 5 dpp. Interestingly, the results of VASA and SOX9 co-staining
195 showed that partial germ cells could not complete homing in 5 dpp cKO testes (Figure 6A and 6B). In
196 mice, starting at 3 dpp, cytoplasmic FOXO1 in some gonocytes begins to enter the nucleus (Goertz et al.,
197 2011). These cells further develop into prospermatogonia, which are expected to develop into
198 spermatogonial stem cells (Goertz *et al.*, 2011). Thus, immunohistochemical staining for FOXO1 was
199 performed on 5 dpp mouse testis sections (Figure 6C). Further, germ cell statistics of FOXO1 expression
200 in the nucleus showed a reduced number of prospermatogonia in cKO mice (Figure 6D). And germ cells
201 in which FOXO1 is expressed in the nucleus similarly undergo abnormal homing (Figure 6E). Thus, all
202 the above data indicated that SRSF1 has an essential role in the homing of precursor SSCs.
203 **SRSF1 is essential for gene expression in spermatogonia**
204 To investigate the molecular mechanisms of SRSF1 in spermatogonia, we isolated mRNA from 5 dpp
205 cKO and Ctrl mouse testes and performed RNA-seq (Figure S2). RNA-seq and RT–qPCR results showed
206 a significant reduction in the expression of *Srsf1* in 5 dpp cKO mouse testes (Figure 7A). Western blotting

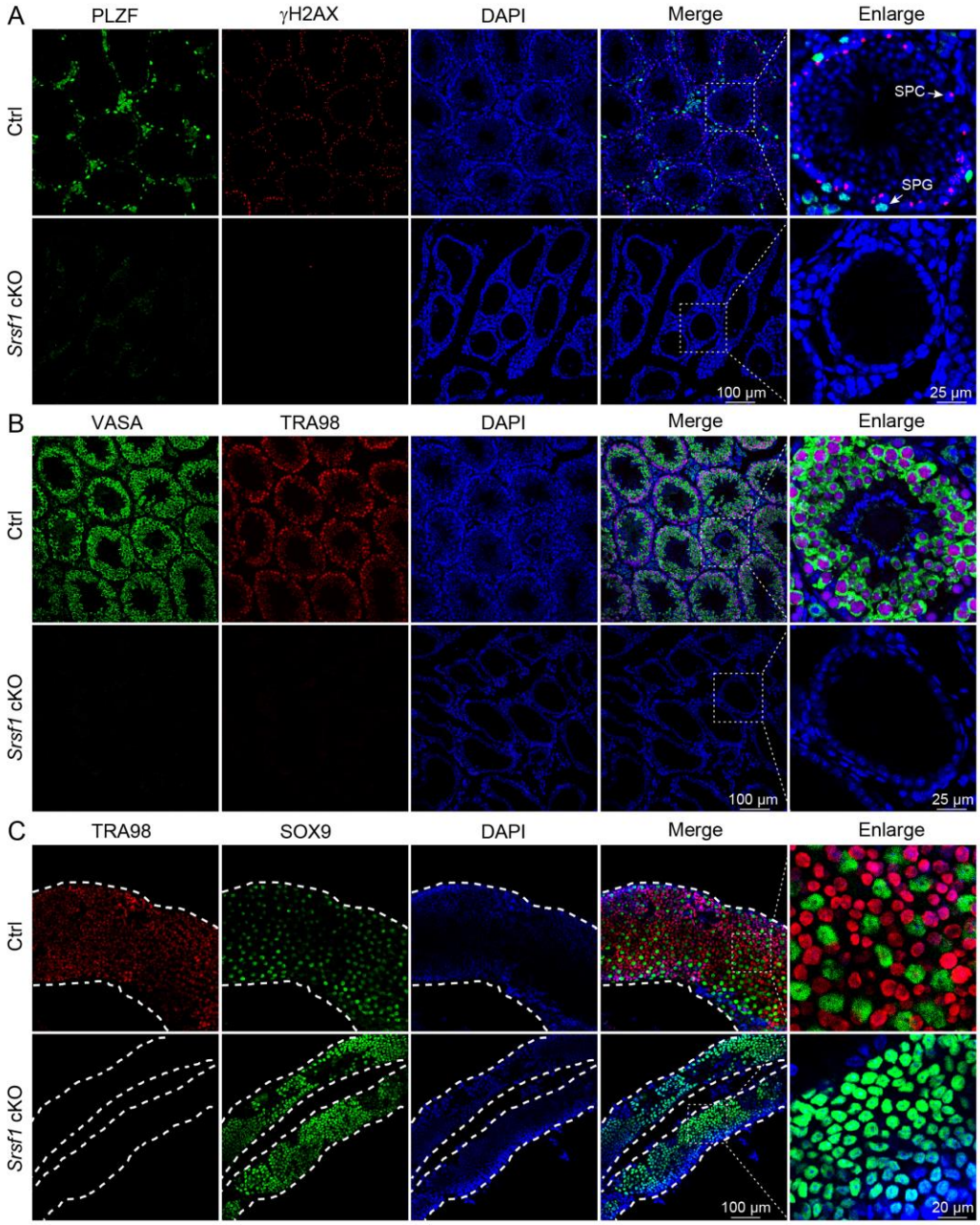


Figure 4 Loss of germ cells in adult cKO mouse testes.

(A) Co-immunostaining of PLZF and γH2AX in adult Ctrl and cKO testes. DNA was stained with DAPI. Scale bar, right panel: 25 μm, other panels: 100 μm.

(B) Co-immunostaining of VASA and TRA98 in adult Ctrl and cKO testes. DNA was stained with DAPI. Scale bar, right panel: 25 μm, other panels: 100 μm.

(C) Whole-mount co-immunostaining of TRA98 and SOX9 in adult Ctrl and cKO testes. DNA was stained with DAPI. White dashed lines, boundary of the tubule. Scale bar, right panel: 20 μm, other panels: 100 μm.

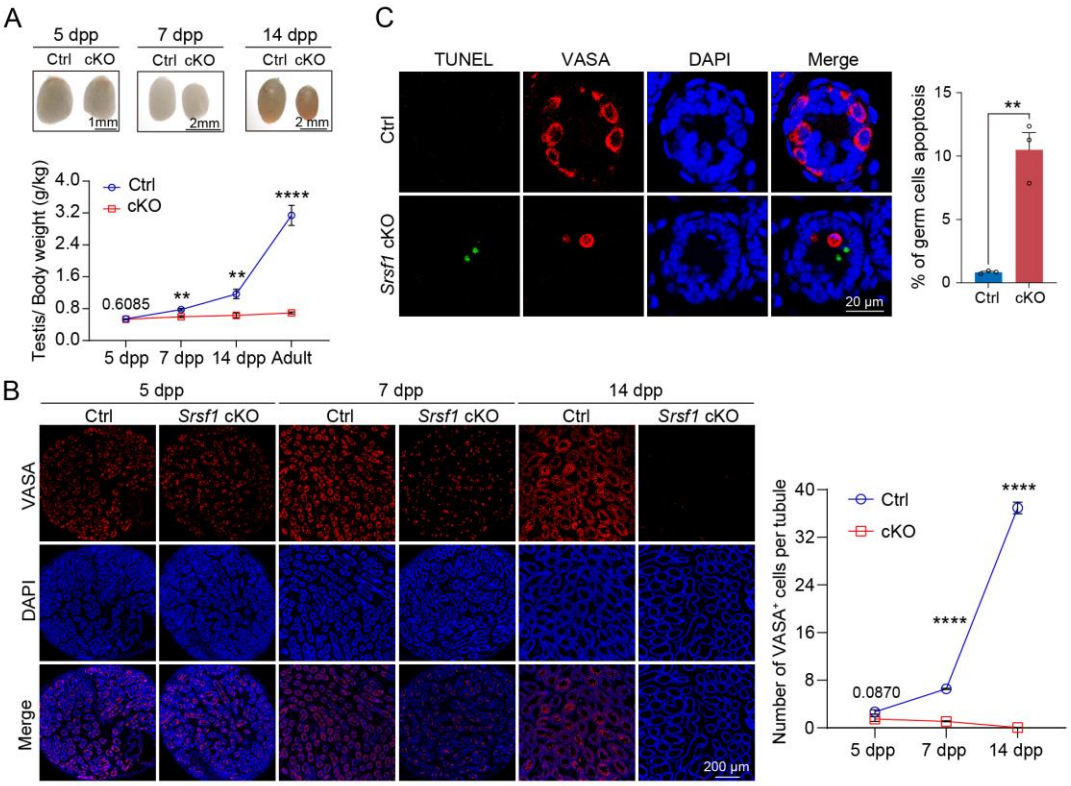


Figure 5 SRSF1 is required for spermatogonia survival.

(A) Testis sizes of 5 dpp, 7 dpp, and 14 dpp Ctrl and cKO mice are shown. The testis/body weight ratios (g/kg) of 5 dpp, 7 dpp, 14 dpp, and adult Ctrl and cKO mice are shown as the mean ± SEM. $n = 4$.

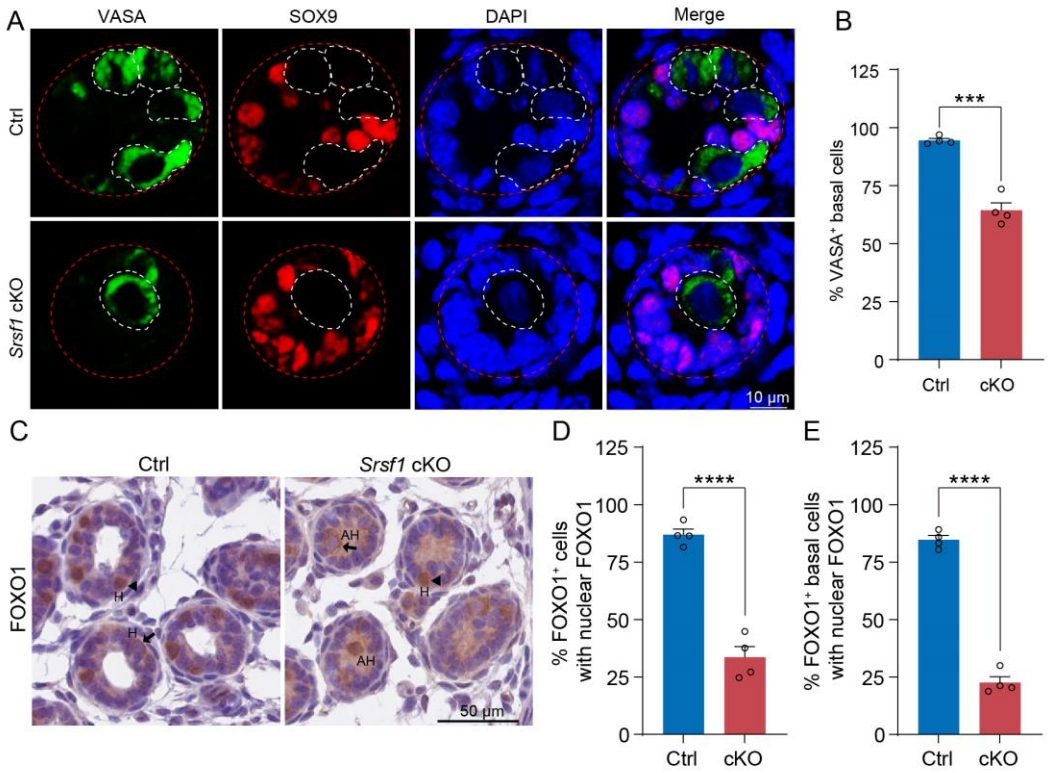
(B) Immunostaining of VASA in 5 dpp, 7 dpp, and 14 dpp Ctrl and cKO testes. DNA was stained with DAPI. Scale bar, 200 μm. Number of VASA-positive cells per tubule is the mean ± SEM. $n = 3$.

(C) TUNEL apoptosis assay was performed on sections from 7 dpp Ctrl and cKO testes. DNA was stained with DAPI. Scale bar, 20 μm. Percentage of germ cells apoptosis is the mean ± SEM. $n = 3$.

Unpaired Student's *t*-test determined significance; exact P value $P \geq 0.05$, $^{**}P < 0.01$, $^{****}P < 0.0001$. The points and error bars represent the mean ± SEM.

results showed that SRSF1 expression was significantly reduced in the testes of cKO mice at 5 dpp (Figure 7B). Hence, for Ctrl and cKO samples, the confidence level of the RNA-seq data was high. The volcano map and cluster heatmap showed 715 downregulated and 258 upregulated genes identified by RNA-seq data in 5 dpp cKO mouse testes (Figure 7C, 7D and Table S2). These gene GO enrichment analyses indicated abnormal germ cell development and cell cycle arrest in 5 dpp cKO mouse testes (Figure 7E). Surprisingly, the heatmap showed that spermatogonia associated gene (*Gfra1*, *Pou5f1*, *Plzf*, *Nanos3*, *Dnd1*, *Stra8*, and *Taf4b*) expression was significantly reduced in the testes of cKO mice at 5 dpp

234 (Figure 7F). Simultaneously, the tracks of differentially expressed genes are demonstrated by using IGV
235 (Figure 7G). Next, we validated the abnormal expression of spermatogonia associated genes
236 (downregulated: *Gfra1*, *Pou5f1*, *Plzf*, *Dnd1*, *Stra8*, and *Taf4b*; unchanged: *Nanos3*) by RT–qPCR (Figure
237 7H). Together, these data indicated that germ cell-specific deletion of *Srsf1* impairs the expression of
238 spermatogonia associated genes.



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240 **Figure 6 SRSF1 is required for homing of precursor SSCs.**

241 (A) Co-immunostaining of VASA and SOX9 in 5 dpp Ctrl and cKO testes. DNA was stained with DAPI. Scale bar,
242 10 μm. Red dashed circles, tubule. White dashed circles, germ cell.

243 (B) The percentage of VASA positive basal cells is shown as the mean ± SEM. $n = 4$.

244 (C) Immunohistochemical staining of FOXO1 in 5 dpp Ctrl and cKO testes. The nuclei were stained with
245 haematoxylin. Scale bar, 10 μm. Arrowheads, FOXO1 in the nucleus. Arrows, FOXO1 in the cytoplasm. H, homing.
246 AH, abnormal homing.

247 (D) The percentage of FOXO1 positive cells with nuclear FOXO1 is shown as the mean ± SEM. 913 Ctrl germ cells
248 and 592 cKO germ cells were obtained from 4 animals.

249 (E) The percentage of FOXO1 positive basal cells with nuclear FOXO1 is shown as the mean ± SEM. 913 Ctrl germ
250 cells and 592 cKO germ cells were obtained from 4 animals.

251 Unpaired Student's *t*-test determined significance; ***$P < 0.001$, ****$P < 0.0001$. The points and error bars represent
252 the mean ± SEM.

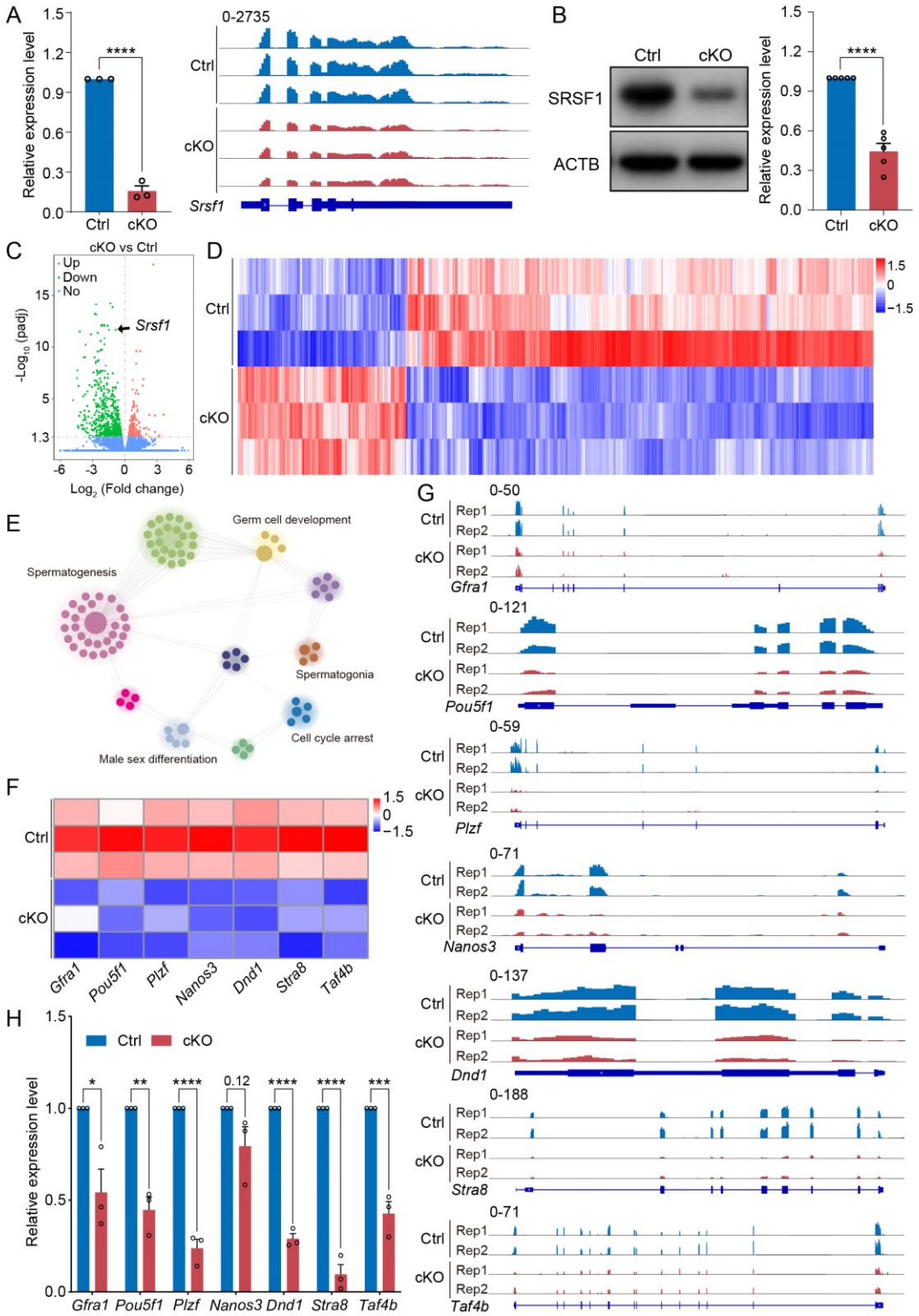


Figure 7 SRSF1 regulates the expression of spermatogonia-related genes.

(A) Expression of *Srsf1* in 5 dpp Ctrl and cKO mouse testes. The RT–qPCR data were normalized to *Gapdh*. *n*=5. The expression of *Srsf1* is shown as reading coverage in 5 dpp mouse testes.

(B) Western blotting of SRSF1 expression in 5 dpp mouse testes. ACTB served as a loading control. The value in

Ctrl testes was set as 1.0, and the relative values in cKO testes are indicated. n=5.

(C) Volcano map displaying the distribution of differentially expressed genes from RNA-seq data. The abscissa in the figure represents the gene fold change in 5 dpp cKO and Ctrl mouse testes. |log2FoldChange| ≥ 0. The ordinate indicates the significance of gene expression differences between 5 dpp cKO and Ctrl mouse testes. padj ≤ 0.05. Upregulated genes are shown as red dots, and downregulated genes are shown as green dots.

(D) Cluster heatmap of differentially expressed genes. The ordinate is the genotype, and the abscissa is the normalized FPKM (fragments per kilobase million) value of the differentially expressed gene. Red indicates a higher expression level, while blue indicates a lower expression level.

(E) Network showing GO enrichment analyses of differentially expressed genes.

(F) Heatmap of spermatogonia-related gene expression.

(G) The expression of spermatogonia-related genes is shown as read coverage.

(H) The expression of spermatogonia-related genes in 5 dpp cKO and Ctrl mouse testes. The RT–qPCR data were normalized to *Gapdh*. The value in the Ctrl group was set as 1.0, and the relative value in the cKO group is indicated. n=3. Unpaired Student's t-test determined significance; exact P value $P \geq 0.05$, *$P < 0.05$, **$P < 0.01$, ***$P < 0.001$, ****$P < 0.0001$. The points and error bars represent the mean ± SEM.

SRSF1 directly binds and regulates the expression and AS of *Tial1*/*Tiar*

Multiomics analyses were carried out in a subsequent study to identify the molecular mechanisms by which SRSF1 regulates spermatogonia survival. Considering that the CLIP-seq data were obtained from adult mouse testis, the set of genes bound by CLIP-seq was restricted to those expressed only in the 5 dpp mouse testis RNA-seq data. we found that 3543 of the 4824 genes bound by SRSF1 were expressed in testes at 5 dpp. Venn diagram data revealed that nine out of 715 down-regulated genes were bound by SRSF1 and underwent abnormal AS (Figure 8A). And one out of 258 upregulated genes was bound by SRSF1 and underwent abnormal AS (Figure 8A). Interestingly, we found that 39 unchanged genes were bound by SRSF1 and underwent abnormal AS (Figure 8A). The AS genes were subsequently investigated in 5 dpp cKO mouse testes using transcriptomic analyses. RNA-seq analyses showed that 162 AS events were significantly affected (FDR<0.05) in cKO mouse testes (Figure 8B, 8C and Table S3). Most of the 133 affected AS events (162) were classified as skipped exons (SEs), with ten AS events categorized as retained introns (RIs), thirteen as mutually exclusive exons (MXEs), four as alternative 5' splice sites (A5SSs), and two as alternative 3' splice sites (A3SSs) (Figure 8C). Additionally, the overall analysis of aberrant AS events showed that SRSF1 effectively inhibits the occurrence of SE and MXE events and promotes the occurrence of RI events (Figure 8C). Then, GO enrichment analyses of AS genes revealed

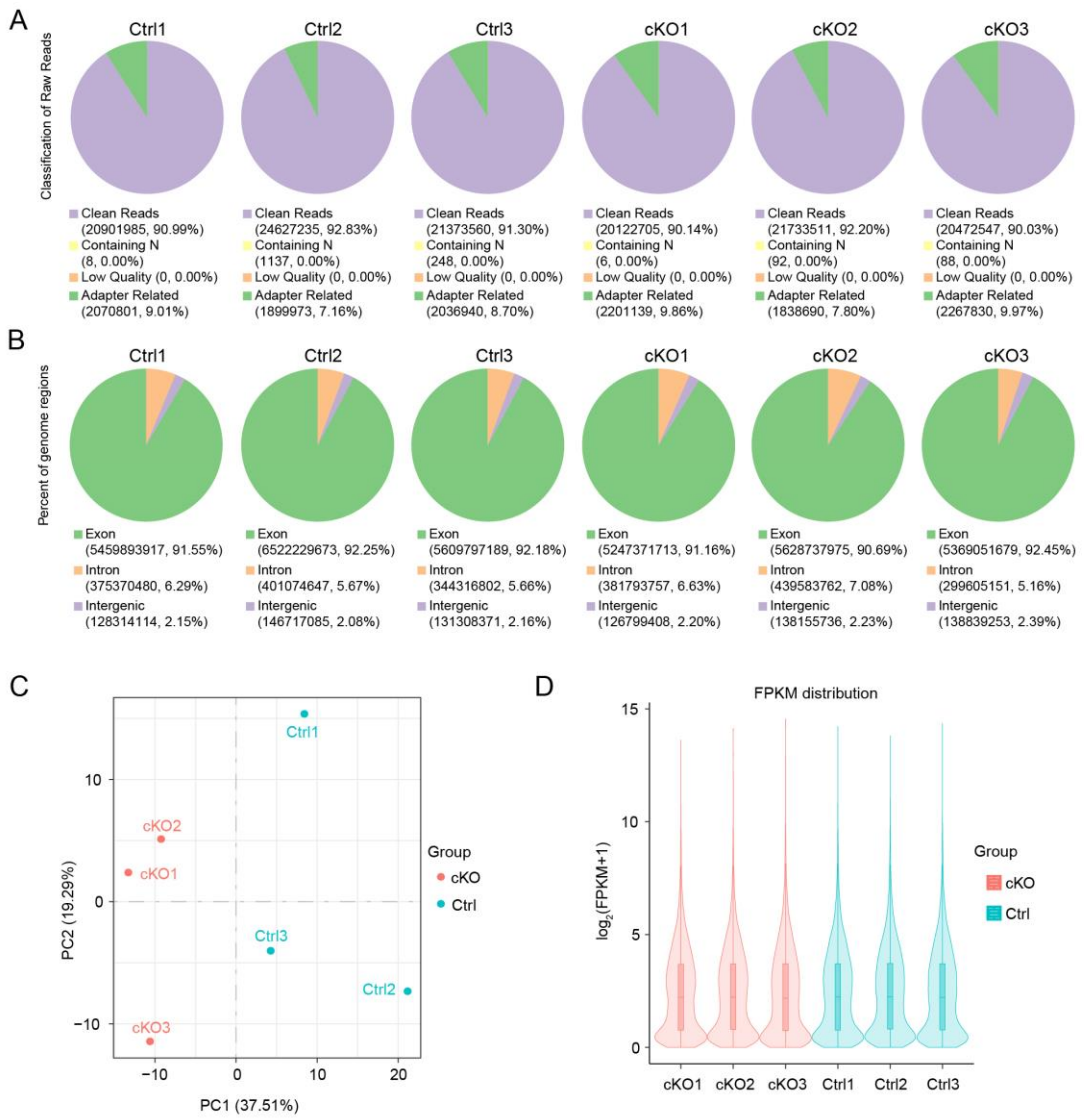


Figure S2 The libraries of RNA-seq are of good quality.

(A) Filtering of sequencing data per sample.

(B) Distribution of sequenced reads across genomic regions for all samples.

(C) Gene expression values (FPKM) of all samples were subjected to principal component analysis (PCA).

(D) The distribution of gene expression levels of different samples is demonstrated by violin plots as shown below. The horizontal coordinate of the graph is the name of the sample, and the vertical coordinate is $\log_2$(FPKM+1).

that four genes concerning germ cell development were altered in AS forms (Figure 8D). It has been shown that *Tial1*/*Tiar* affects the survival of primordial germ cells (Beck et al., 1998). Moreover, *Tial1*/*Tiar* is one of 39 unchanged genes that are bound by SRSF1 and undergo abnormal AS. Thus, multiomics analyses suggested that *Tial1*/*Tiar* were posttranscriptionally regulated by SRSF1. Next, we investigated the mechanism by which SRSF1 regulates the AS of *Tial1*/*Tiar*, RT–PCR results showed

that the pre-mRNA of *Tial1*/*Tiar* in 5 dpp cKO mouse testes exhibited abnormal AS (Figure 8E). We then visualized the different types of AS based on RNA-seq data by using IGV (Figure 8F and Figure S3). The results of RIP–qPCR showed that SRSF1 could bind to the pre-mRNA of *Tial1*/*Tiar* (Figure 8G). Interestingly, RNA-seq analyses showed that the FPKM of *Tial1*/*Tiar* was unchanged in 5 dpp cKO mouse testes (Figure 8H). RT–qPCR results showed that *Tial1*/*Tiar* transcript levels were not inhibited (Figure 8I). However, Western blotting showed that expression levels of TIAL1/TIAR isoform X2 were significantly suppressed (Figure 8J and Figure S3). In summary, the data indicate that SRSF1 is required for TIAL1/TIAR expression and splicing in spermatogonia survival.

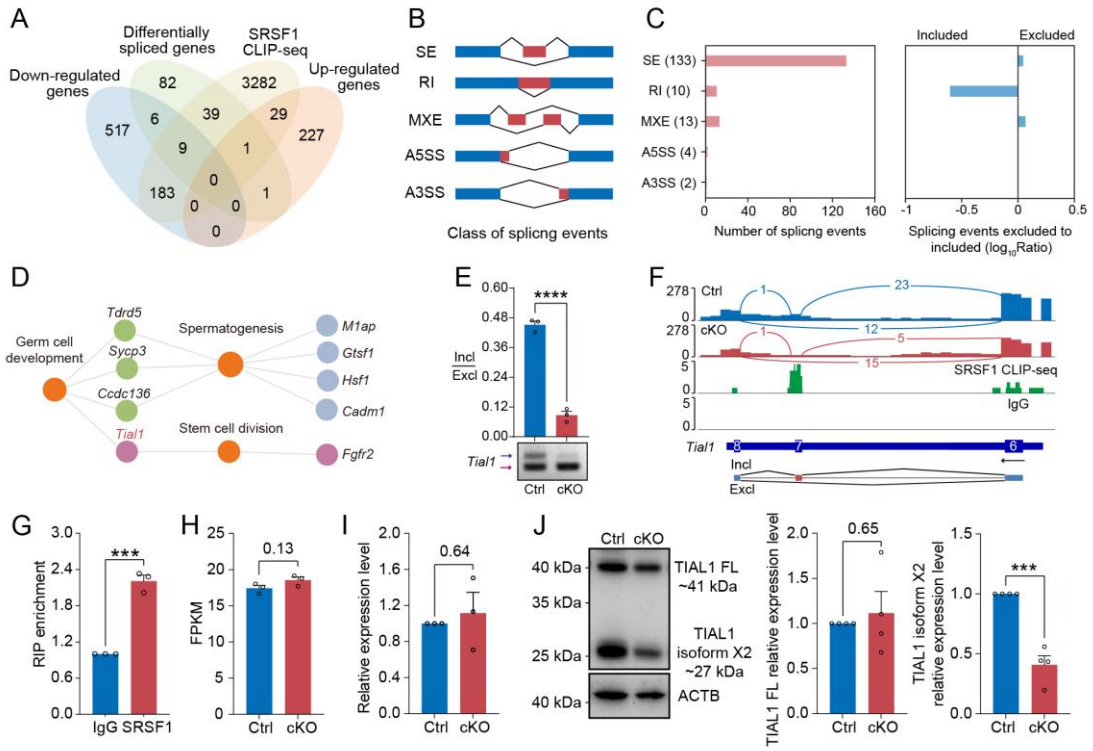


Figure 8 SRSF1 directly binds and regulates the expression and AS of *Tial1*/*Tiar*.

(A) Venn diagram showing the correlation among down-regulated, upregulated, alternatively spliced, and SRSF1-binding genes.

(B) Schematic diagram showing the classes of splicing events.

(C) Splicing events were analysed by number, exclusion, and inclusion.

(D) Network showing GO enrichment analyses of AS genes.

(E) The AS of *Tial1/Tiar* in 5 dpp Ctrl and cKO mouse testes was analysed by RT–PCR. *n*=3. The ratio of inclusion (Incl) to exclusion (Excl) is shown accordingly.

(F) Analyses of *Tial1/Tiar* expression and exon–exon junctions were performed.

(G) SRSF1 directly binds the pre-mRNA of *Tial1*/*Tiar* by RIP–qPCR in 5 dpp mouse testes. *n*=3.

(H) The FPKM of *Tial1*/*Tiar* in 5 dpp Ctrl and cKO mouse testes.

(I) The expression of *Tial1*/*Tiar* in 5 dpp Ctrl and cKO mouse testes. The RT–qPCR data were normalized to *Gapdh*. The value in the Ctrl group was set as 1.0, and the relative value in the cKO group is indicated. n=3.

(J) Western blotting of TIAL1/TIAR expression in 5 dpp mouse testes. ACTB served as a loading control. The FL/isoform X2 value of TIAL1/TIAR in the Ctrl group was set as 1.0, and the relative value in the cKO group is indicated. n=3. FL, full length.

Unpaired Student's *t*-test determined significance; exact P value $P \geq 0.05$, ***$P < 0.001$, ****$P < 0.0001$. The points and error bars represent the mean ± SEM.

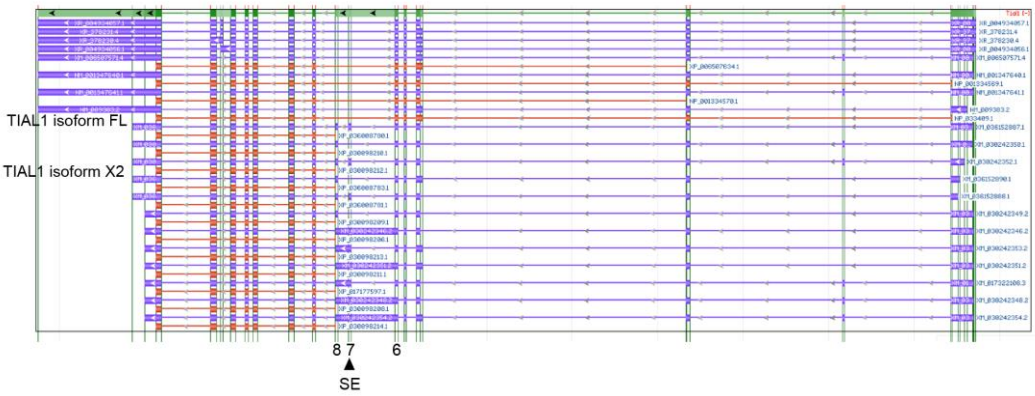
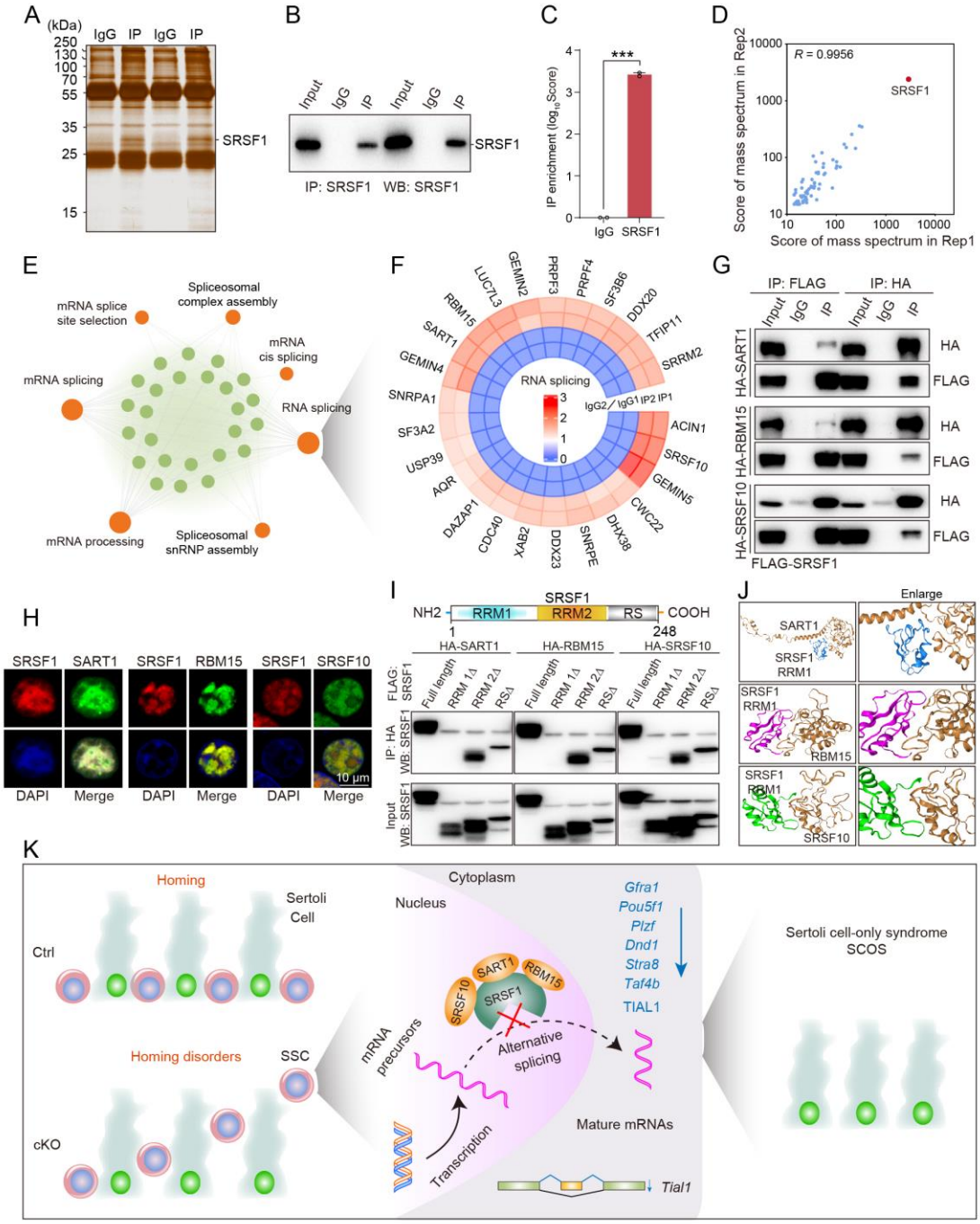


Figure S3 Transcription and translation information for *Tial1*.

Transcription and translation information for *Tial1* was obtained from genome data viewer.



333

334 **Figure 9 SRSF1 recruits AS-related proteins to modulate AS in testes.**

335 (A) Silver-stained gel of SRSF1 and control immunoprecipitates from 5 dpp mouse testis extracts.

336 (B) IP experiment was performed in 5 dpp mouse testis extracts.

337 (C) IP of SRSF1 from IP-MS data.

338 (D) *Pearson's* correlation analysis showed the coefficient between the two samples for IP-MS data.

339 (E) Network showing GO enrichment analyses of SRSF1-binding proteins.

340 (F) Circular heatmap of AS-related proteins.

341 (G) Co-IP experiment for detecting the SRSF1 association between SART1, RBM15, and SRSF10 in 293T cells.

(H) SRSF1-mCherry cotransfected with SART1-eGFP, RBM15-eGFP, and SRSF10-eGFP 293T cells is shown. DNA was stained with DAPI. Scale bar, 10 μm.

(I) Co-IP experiment for detecting the SRSF1 truncated protein association between SART1, RBM15, and SRSF10 in 293T cells.

(J) A schematic diagram of protein interactions is shown.

(K) Schematic illustration of the molecular mechanisms by which SRSF1 regulates homing of mouse precursor SSCs.

SRSF1 recruits AS-related proteins to modulate AS in testes

To identify the interacting proteins for which SRSF1 exerts its AS role, we performed MS analyses of IP samples from 5 dpp mouse testis extracts. The silver-stained gel of SRSF1 and normal IgG showed several SRSF1-interacting proteins from 5 dpp mouse testis extracts (Figure 9A). The IP results indicated that SRSF1 was able to effectively IP the testis extracts of 5 dpp mice (Figure 9B). IP-MS data demonstrated the efficient enrichment of SRSF1 (Figure 9C and Table S4). These data showed that the two samples were highly reproducible, especially for SRSF1 (Figure 9D). Then, GO enrichment analyses of the IP proteins revealed that AS-related proteins could interact with SRSF1 (Figure 8E). A circular heatmap showed that SRSF1 could interact with AS-related proteins (e.g., SRSF10, SART1, RBM15, SRRM2, SF3B6, and SF3A2) (Figure 9F). The Co-IP results indicated that FLAG-SRSF1 interacted with HA-SART1, HA-RBM15 and HA-SRSF10 in 293T cells (Figure 9G). In addition, the fluorescence results showed complete co-localization of mCherry-SRSF1 with eGFP-SART1, eGFP-RBM15 and eGFP-SRSF10 in 293T cells (Figure 9H). Co-IP suggested that the RRM1 domain of SRSF1 interacted with HA-SART1, HA-RBM15 and HA-SRSF10 in 293T cells (Figure 9I). Determining the complex structures of these interactions is valuable, in which molecular docking has played an important role (Yan et al., 2017). HDOCK is a novel web server of our hybrid docking algorithm of template-based modelling and free docking (Yan *et al.*, 2017). The HDOCK analysis results depicted the RRM1 domain of SRSF1 with SRSF10, SART1, and RBM15 docking based on a hybrid strategy (Figure 9J). Together, the above data show that SRSF1 may interact with SRSF10, SART1, and RBM15 to regulate AS in 5 dpp mouse testes.

Discussion

Failure of spermatogonia survival led to SCOS

Disturbed spermatogenesis can cause SCOS and ultimately male sterility (Jiao et al., 2021). In recent

373 years, it has been reported that many spermatogonia-related gene deletions have disrupted SSC self-
374 renewal and differentiation in patient and mouse models (La and Hobbs, 2019; Tan and Wilkinson, 2020;
375 Wang et al., 2021). SCOS was observed in *Ddx5*, *Tial1*/*Tiar*, *Uhrf1*, *Pramef12*, *Dot1l*, and *Rad51* deletion
376 mouse models (Beck *et al.*, 1998; Legrand *et al.*, 2019; Lin et al., 2022; Qin et al., 2022; Wang et al.,
377 2019; Zhou *et al.*, 2022). Mouse models are still of great significance and reference for human SCOS
378 studies, and they will provide a better understanding of how SCOS occurs and develops over time.
379 Interestingly, our mouse model had SCOS (Figure 3D-I and Figure 4). The absence of germ cells
380 represents classical SCOS in adult mouse testes (Figure 4) (Wang *et al.*, 2023). In addition, we found
381 abnormal expression of spermatogonia-related genes (*Gfra1*, *Pou5f1*, *Plzf*, *Dnd1*, *Stra8*, and *Taf4b*) in
382 cKO mouse testes (Figure 7F-H). These differentially expressed genes regulate SSC self-renewal and
383 differentiation in mouse testes (Kanatsu-Shinohara and Shinohara, 2013; La and Hobbs, 2019; Tan and
384 Wilkinson, 2020). Thus, this provided an opportunity for us to better study the underlying molecular
385 mechanisms. These data indicate that SRSF1 deficiency impairs spermatogonia survival, leading to
386 SCOS in male mice.

387 **The formation of SSC pools and the establishment of niches are essential for spermatogenesis**

388 The earliest event in the development of the SSC population is the migration of prospermatogonia from
389 the centre of seminiferous cords where they have resided since sex determination of the embryonic gonad
390 to the basement membrane (Oatley and Brinster, 2012). In mice, this process is also known as homing of
391 precursor SSCs, which occurs in the initial 3 dpp and then develops into SSCs at 4 to 6 dpp for continuous
392 self-renewal and differentiation (Lee and Shinohara, 2011; McLean *et al.*, 2003; Oatley and Brinster,
393 2012; Tan and Wilkinson, 2020). Therefore, homing analysis of precursor SSCs was performed in 5 dpp
394 cKO mouse testes. Interestingly, the VASA and SOX9 co-staining results demonstrated that partial germ
395 cells could not complete homing in 5 dpp cKO testes (Figure 6A and 6B). Further, immunohistochemical
396 staining for FOXO1 and statistical results indicated that germ cells in which FOXO1 is expressed in the
397 nucleus similarly undergo abnormal homing (Figure 6C-E). Germ cells that do not migrate to the
398 basement membrane are unable to form SSC pools and establish niches (McLean *et al.*, 2003). These
399 SSCs that lose their ecological niche will cease to exist. In our data, TUNEL results showed that apoptosis
400 significantly increased in 7 dpp cKO mouse testes. At once, the quantifications of germ cells per tubule
401 showed a significant reduction in 7 dpp and 14 dpp cKO testes, especially 14 dpp cKO testes (Figure
402 5B). In conclusion, SRSF1 is crucial for the formation of SSC pools and the establishment of niches

through homing of precursor SSCs.

Abnormal AS impaired the survival of spermatogonia

AS is commonly found in mammals, especially in the brain and testes (Mazin *et al.*, 2021; Merkin et al., 2012; Wang et al., 2008). AS plays essential roles in the posttranscriptional regulation of gene expression during many developmental processes, such as SSC self-renewal and differentiation (Chen et al., 2018; Song et al., 2020). Recently, BUD31-mediated AS of *Cdk2* was shown to be required for SSC self-renewal and differentiation (Qin *et al.*, 2023). *Srsf10* depletion disturbed the AS of genes, including *Nasp*, *Bclaf1*, *Rif1*, *Dazl*, *Kit*, *Ret*, and *Sycp1* (Liu *et al.*, 2022). UHRF1 interacts with snRNAs and regulates AS of *Tle3* in mouse SSCs (Zhou *et al.*, 2022). Mettl3-mediated m6A regulates AS of *Sohlh1* and *Dazl* (Xu et al., 2017). We found that SRSF1 acts as an alternative RNA splicing regulator and directly interacts with *Tial1*/*Tiar* transcripts to regulate splicing events in spermatogonia (Figure 8E-G). Additionally, expression levels of TIAL1/TIAR isoform X2 were significantly suppressed (Figure 8J). Interestingly, *Tial1*/*Tiar* transcript levels were not inhibited (Figure 8H and 8I). These results suggested that SRSF1 explicitly regulates the expression of *Tial1*/*Tiar* via AS. Studies have reported that TIAL1/TIAR is essential for primordial germ cell development in mouse testes (Beck *et al.*, 1998). *Tial1/Tiar* deletion impairs germ cell survival leading to SCOS, consistent with our phenotype (Figure 3E-I and 4A-C) (Beck *et al.*, 1998). Taken together, SRSF1 may affect germ cell survival by directly binding and regulating *Tial1*/*Tiar* expression through AS.

We found that SRSF1 could interact with AS-related proteins (e.g., SRSF10, SART1, RBM15, SRRM2, SF3B6, and SF3A2) (Figure 9F). A recent study reported that SRSF10 deficiency impaired spermatogonia differentiation but did not affect homing of precursor SSCs (Liu *et al.*, 2022). However, our data showed that SRSF1 is essential for homing of mouse precursor SSCs. Therefore, this suggests that SRSF1 has a specific function in the homing of precursor SSCs independent of SRSF10.

SRSF1-mediated posttranscriptional regulation during homing of precursor SSCs provides new insights into the treatment of human reproductive diseases

Aberrant homing of precursor SSCs often lead to gametogenic failure, resulting in subfertility or infertility (Jiao *et al.*, 2021; Kanatsu-Shinohara and Shinohara, 2013; La and Hobbs, 2019; Song and Wilkinson, 2014). Loss-of-function mutations in humans and corresponding knockout/mutated mice have been extensively researched (Jiao *et al.*, 2021). However, AS-related posttranscriptional regulation during meiosis has not been well studied. In recent years, there have been reports that the RNA-binding

proteins SRSF10, UHRF1, BUD31, and BCAS2 regulate AS in mouse SSCs (Liu *et al.*, 2022; Liu *et al.*, 2017; Qin *et al.*, 2023; Zhou *et al.*, 2022). This study used a multiomics approach to perform in-depth analyses of SRSF1-mediated posttranscriptional regulatory mechanisms to enrich the field. It also provides new ideas and insights for clinical diagnosis and treatment.

In summary, this study demonstrates that SRSF1 plays a critical role in posttranscriptional regulation to implement homing of precursor SSCs (Figure 9K). Thus, the posttranscriptional regulation of SRSF1-mediated splicing is resolved during the formation of SSC pools and the establishment of niches.

Materials and Methods

Mouse strains

C57BL/6N and ICR mice were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd. *Srsf1*$^{Fl/Fl}$ mice were generated in the laboratory of Prof. Xiangdong Fu (University of California, San Diego, USA) and were kindly provided by Prof. Yuanchao Xue (Institute of Biophysics, Chinese Academy of Sciences, Beijing, China) (Xu *et al.*, 2005). *Vasa*-Cre mice were obtained from The Jackson Laboratory (Gallardo et al., 2007).To generate *Srsf1* cKO mice, *Vasa*-Cre mice were crossed with *Srsf1*$^{Fl/Fl}$ mice. The primers used for PCR to genotype *Srsf1*$^{Fl/Fl}$ and *Vasa*-Cre mice are shown in Table S5. All mice were bred and housed under specific pathogen-free conditions with a controlled temperature (22 ± 1°C) and exposed to a constant 12-hour light-dark cycle in the animal facilities of China Agricultural University. All experiments were conducted according to the guidelines and with the approval of the Institutional Animal Care and Use Committee of China Agricultural University (No. AW80401202-3-3).

Fertility test

For 15 days, two 8-week-old ICR female mice were caged with one 8-week-old male control (Ctrl) or cKO mouse. The mice were kept individually after the appearance of the vaginal plug, and the dates were recorded. Male mice continue to be caged after two days. The number of pups from each female was recorded each day, and the date of parturition was recorded.

Immunostaining and histological analyses

Mouse testes were fixed with 4% paraformaldehyde (PFA, P6148-500G, Sigma–Aldrich) in PBS (pH 7.4) at 4°C overnight, dehydrated in graded ethanol solutions, vitrified with xylene, and embedded in paraffin. Testis sections were cut at a 5-μm thickness for immunostaining and histologic analyses. For

463 histological analyses, sections were dewaxed in xylene, rehydrated in a graded ethanol solution, and
464 stained with haematoxylin-eosin. After sealing the slides with neutral resin, a Ventana DP200 system was
465 used for imaging. For immunofluorescence analyses, antigen retrieval was performed by microwaving
466 the sections with sodium citrate buffer (pH 6.0). After blocking with 10% normal goat serum at room
467 temperature for 1 hour, the sections were incubated with primary antibodies (Table S6) in 5% normal
468 goat serum overnight at 4°C. After washing with PBS, the sections were incubated with secondary
469 antibodies (Table S6) at room temperature in the dark for 1 hour. The slides were mounted in an antifade
470 mounting medium with DAPI (P0131, Beyotime). Photographs were taken with a Nikon A1 laser
471 scanning confocal microscope and a Zeiss OPTOME fluorescence microscope. For
472 immunohistochemistry analyses, antigen retrieval was performed by microwaving the sections with
473 sodium citrate buffer (pH 6.0). 5 dpp testis sections were prepared as described in the instructions for
474 Immunohistochemistry Kit (PV-9001, ZSGB-BIO). After sealing the slides with neutral resin, a Ventana
475 DP200 system was used for imaging.

476 **Whole-mount immunostaining**

477 The testes were collected and dispersed with 5ml syringes. Blown-out tubules were fixed in PFA at 4°C
478 for 4 hours. The tubules were washed three times with PBS (pH 7.4) for 5 min each and stored at 4°C.
479 The tubules were permeated with 0.3% Triton X-100 for 1 h at 4°C. Then, whole-mount staining followed
480 the immunofluorescence protocol.

481 **TUNEL apoptosis analyses**

482 7 dpp testis sections were prepared as described in the instructions for the TUNEL Apoptosis Assay Kit
483 (C1088, Beyotime). Photographs were taken with a Nikon A1 laser scanning confocal microscope and a
484 Zeiss OPTOME fluorescence microscope. Apoptosis rate was derived from the number of TUNEL-
485 positive signals in tubules as a percentage of the total number of germ cells.

486 **RT–PCR and RT–qPCR**

487 Total RNA was extracted by using RNAiso Plus (9109, Takara), and the concentration was measured
488 with a Nano-300 ultramicro spectrophotometer (Allsheng). cDNA was obtained according to the
489 instructions of a TIANScript II RT kit (KR107, TIANGEN). The expression of transcripts of the target
490 gene was measured by using a LightCycle® 96 instrument (Roche) with Hieff UNICON SYBR green
491 master mix (11198ES08, Yeasen). AS analyses were performed on a RePure-A PCR instrument (BIO-
492 GENER). Primers were synthesized by Sangon Biotech (Table S5). The expression level of *Gapdh* or

493 *Actb* was used as the control, and this value was set as 1. Other samples' relative transcript expression
494 levels were obtained by comparing them with the control results.

495 **RNA-seq**

496 Total RNA was extracted from mouse testes according to the above protocol at 5 dpp. Briefly, mRNA
497 was purified from total RNA using poly-T oligo-attached magnetic beads. After fragmentation, we
498 established a transcriptome sequencing library and assessed library quality on an Agilent Bioanalyzer
499 2100 system. The clustering of the index-coded samples was performed on a cBot Cluster Generation
500 System using a TruSeq PE Cluster kit v3-cBot-HS (Illumina) according to the manufacturer's instructions.
501 After cluster generation, the library preparations were sequenced on the Illumina NovaSeq platform, and
502 150 bp paired-end reads were generated. After quality control, all downstream analyses were performed
503 on clean, high-quality data. The reference genome index was built, and paired-end clean reads were
504 aligned to the reference genome using HISAT2 software (version 2.0.5). FeatureCounts (version 1.5.0)
505 counted the reads mapped to each gene. Then, the fragments per kilobase million (FPKM) value of each
506 gene was calculated based on the length of the gene and the read count mapped to this gene. Differential
507 expression analyses of cKO/Ctrl mouse testes (three biological replicates per condition) were performed
508 using the DESeq2 R package (version 1.20.0). Genes with a padj $\leq$ 0.05 identified by DESeq2 were
509 considered differentially expressed.

510 **AS analyses**

511 rMATS software (version 3.2.5) was used to analyse the AS events in cKO mouse germ cells based on
512 RNA-seq data. Five types of AS events (SE, RI, MXE, A5SS, and A3SS) were revealed by rMATS
513 software. Our threshold for screening differentially significant AS events was a false discovery rate (FDR)
514 of less than 0.05. Splicing events with an FDR less than 0.05 and an inclusion-level difference with a
515 significance of at least 5% (0.05) were considered statistically significant. Integrative Genomics Viewer
516 (IGV, 2.16.0) software was used to visualize and confirm AS events based on RNA-seq data.

517 **Gene Ontology (GO) enrichment analyses**

518 The GO enrichment analyses of differentially expressed genes and AS genes were implemented with the
519 clusterProfiler R package (version 3.4.4), in which gene length bias was corrected. All expressed genes
520 (TPM sum of all samples >=1) are listed background. GRCm38/mm10 was used as a mouse reference
521 genome, and the Benjamini–Hochberg multiple methods was applied to adjust for multiple testing. GO

522 enrichment analyses with corrected *P* values of less than 0.05 were significantly enriched for
523 differentially expressed genes and AS genes.

524 **Western blotting**

525 Total protein was extracted with cell lysis buffer (P0013, Beyotime) containing PMSF (1:100, ST506,
526 Beyotime) and a protease inhibitor cocktail (1:100, P1005, Beyotime). Protein concentration was
527 determined with the BCA Protein Assay Kit (CW0014S, CWBiotech). The protein lysates were
528 electrophoretically separated on sodium dodecyl sulfate-polyacrylamide gels and electrically transferred
529 to polyvinylidene fluoride membranes (IPVH00010, Millipore). The membranes were blocked in 5%
530 skimmed milk for 1 hour and incubated with the primary antibodies (Table S6) for one night at 4°C. Then,
531 the membranes were incubated with secondary antibodies (Table S6) at room temperature for 1 hour. The
532 proteins were visualized using a Tanon 5200 chemiluminescence imaging system following incubation
533 with BeyoECL Plus (P0018S, Beyotime).

534 **IP, IP-MS, Co-IP**

535 Total protein was extracted with cell lysis buffer (P0013, Beyotime) containing PMSF (1:100, ST506,
536 Beyotime) and a protease inhibitor cocktail (1:100, P1005, Beyotime). After incubation on ice for 20 min,
537 the lysate was added to 20 μl of protein A agarose (P2051-2 ml, Beyotime) for pre-clearing at 4°C for 1
538 hour. Two micrograms of an SRSF1 antibody (sc-33652, Santa Cruz Biotechnology) and a normal mouse
539 IgG (sc-3879, Santa Cruz Biotechnology) were added to the lysate the mixture was incubated overnight
540 incubation at 4°C. The next day, 60 μl of protein A agarose was added to the lysate, which was then
541 incubated at 4°C for 4 hours. The agarose complexes containing antibodies and target proteins were
542 washed 3 times for 5 min at 4°C. IP and Co-IP were performed according to the above Western blotting
543 protocol. The complex was sent to the protein mass spectrometry laboratory for IP-MS analyses using a
544 Thermo Q-Exactive high-resolution mass spectrometer (Thermo Scientific, Waltham). Raw data from
545 the mass spectrometer were preprocessed with Mascot Distiller 2.4 for peak picking. The resulting peak
546 lists were searched against the uniport mouse database using Mascot 2.5 search engine.

547 **RNA immunoprecipitation (RIP) and RIP–qPCR**

548 As described previously (Gagliardi and Matarazzo, 2016), RIP was performed using 5 dpp mouse testes.
549 The testes were lysed in cell lysis buffer (P0013, Beyotime) containing PMSF (1:100, ST506, Beyotime),
550 a proteinase inhibitor cocktail (1:100, P1005, Beyotime), DTT (1:50, ST041-2 ml, Beyotime), and an
551 RNase inhibitor (1:20, R0102-10 kU, Beyotime). After incubation on ice for 20 min, the lysate was added

to 20 μl of protein A agarose (P2051-2 ml, Beyotime) for pre-clearing at 4°C for 1 hour. Two micrograms of an SRSF1 antibody (sc-33652, Santa Cruz Biotechnology) and a normal mouse IgG (sc-3879, Santa Cruz Biotechnology) were added to the lysate, which was then incubated overnight at 4°C. The next day, 60 μl of protein A agarose was added to the lysate, and the mixture was incubated at 4°C for 4 hours. The agarose complexes containing antibodies, target proteins, and RNA were washed 3 times for 5 min at 4°C and repeated. Protein-bound RNA was extracted using RNAiso Plus and a Direct-zol RNA MicroPrep Kit. RIP–qPCR was performed according to the above RT–qPCR protocol.

Statistical analyses

Pearson's correlation coefficients (R) were calculated by using the scores of the two samples for MS. GraphPad Prism software (version 9.0.0) was used for the statistical analyses, and the values and error bars represent the mean ± SEM. Significant differences between the two groups were analysed using Student's *t*-test. Statistical significance is indicated as follows: exact P value $P \geq 0.05$; $^{*}P < 0.05$; $^{**}P < 0.01$; $^{***}P < 0.001$; $^{****}P < 0.0001$).

Data availability

All data generated or analysed during this study are included in this published article, its supplementary information files and publicly available repositories. Mass spec data are included in supplementary information files (Table S7). The RNA-seq data were deposited in GEO (https://www.ncbi.nlm.nih.gov/geo/) under accession number GSE227575. The CLIP-seq data were deposited in GEO under accession number GSE227422 (Sun *et al.*, 2023a).

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