Pollen thermotolerance of a widespread plant in response to climate warming: possible local adaptation of populations from different elevations

Karolína Hrubá[1,2], Paolo Biella[3] 3, Jan Klečka[1]

[1] Institute of Entomology, Biology Centre of the Czech Academy of Sciences, České Budějovice, Czech Republic
[2] Department of Zoology, Faculty of Science, University of South Bohemia, České Budějovice, Czech Republic
[3] ZooPlantLab, Department of Biotechnology and Biosciences, University of Milano-Bicocca, Milan, Italy

Corresponding Author:
Jan Klečka[1]
Institute of Entomology, Biology Centre of the Czech Academy of Sciences, Branišovská 31, České Budějovice, 37005, Czech Republic
Email address: jan.klecka@entu.cas.cz

Abstract

One of the most vulnerable phases in the plant life cycle is sexual reproduction, which depends on effective pollen transfer, but also on the thermotolerance of pollen grains. Pollen thermotolerance is temperature-dependent and may be reduced by increasing temperature associated with global warming. A growing body of research has focused on the effect of increased temperature on pollen thermotolerance in crops to understand the possible impact of temperature extremes on yield. Yet, little is known about the effects of temperature on pollen thermotolerance of wild plant species. To fill this gap, we selected *Lotus corniculatus* s.l. (Fabaceae), a species common to many European habitats and conducted laboratory experiments to test its pollen thermotolerance in response to artificial increase in temperature. To test for possible local adaptation of pollen thermal tolerance, we compared data from six lowland (389 - 451 m a.s.l.) and six highland (841 - 1030 m a.s.l.) populations. We observed pollen germination *in vitro* at 15, 25, 30, and 40°C. While lowland plants had stable germination rate at a wide range of temperatures between 15 and 30°C, with reduced germination rate observed only at extremely high temperatures (i.e., 40°C), the germination rate of highland plants was reduced already when the temperature reached 30°C, temperature commonly exceeded in the lowland during warm summers. This suggests that lowland populations of *L. corniculatus* may be locally adapted to higher temperature for pollen germination. On the other hand, pollen tube length decreased with increasing temperature in a similar way in lowland and highland plants. The overall average pollen germination rate significantly differed between lowland and highland populations, with highland populations displaying higher germination rate. On the other hand, the average pollen tube length was slightly smaller in highland populations. In conclusion, we found that pollen thermotolerance of *L. corniculatus* is reduced at high temperature and that the germination of pollen from plant populations growing at higher elevations is more sensitive to increased temperature, which suggests possible local adaptation of pollen thermotolerance.

Introduction

Climate change, in particular increasing temperature, represents a major threat to biodiversity and is already leading to changes in the geographic distribution of many species of plants and animals (Lenoir et al., 2008; Biella et al., 2017; Freeman et al., 2018; Fazlioglu, Wan & Chen, 2020). Globally, average air temperature is predicted to increase by as much as 2.4°C–4.8°C by the end of the 21st century in the high-emissions scenario SSP5-8.5 according to the Sixth Assessment Report of the IPCC (Lee et al., 2021). Understanding the effect of increasing temperature on the survival and reproduction of plants and animals and their capacity to adapt is thus an urgent task (Aitken et al., 2008; Alberto et al., 2013).

One of the most vulnerable phases in the plant life cycle is sexual reproduction. Successful reproduction in most plant species depends on effective pollination by animals

(Ollerton, Winfree & Tarrant, 2011), but also on the thermotolerance of pollen grains (He et al., 2017; Rosbakh et al., 2018), which can be highly variable in species growing in different environmental conditions (Steinacher & Wagner, 2012; Rosbakh & Poschlod, 2016). That pollen germination and pollen tube growth depend, among other factors, on temperature has been known for a long time (Brink, 1924; Das et al., 2014; Pham, Herrero & Hormaza, 2015; Shi et al., 2018; Hebbar et al., 2018). Other stages of the process of sexual reproduction are also temperature-dependent (Hedhly, Hormaza & Herrero, 2009; Zinn, Tunc-Ozdemir & Harper, 2010; Lohani, Singh & Bhalla, 2020), but pollen germination seem to be particularly sensitive to heat stress (Young, Wilen & Bonham-Smith, 2004; Zinn, Tunc-Ozdemir & Harper, 2010). Optimum temperature allows higher pollen germination success and faster pollen tube growth (Hedhly, Hormaza & Herrero, 2005), but when the temperature exceeds a species-specific optimum, pollen germination rapidly decreases and pollen tube growth stops (Lewis, 1942; Kakani et al., 2005; Zinn, Tunc-Ozdemir & Harper, 2010). Therefore, both pollen germination rate and pollen tube length can provide key information on the effects of thermal stress on the male fitness and reproduction in plants.

Global warming may thus represent a serious threat to pollen germination and the overall plant reproductive system (Hedhly, Hormaza & Herrero, 2005). Apart from increasing mean temperature, temperature extremes and heat waves are predicted to be more intense, more frequent, and to last longer (IPCC, 2021). However, the susceptibility of key stages of the sexual reproduction of plants to high temperature has been so far studied almost exclusively in crops or other commercially exploited plants (Wang et al., 2019; Bheemanahalli et al., 2019; Lovane et al., 2021; Lohani, Singh & Bhalla, 2022), but not in wild plant species, with the exception of a few studies on trees (Pigott & Huntley, 1980; Flores-Rentería et al., 2018) and forbs (McKee and Richards, 1998; Rosbakh & Poschlod, 2016).

Moreover, insights about the impact of global warming on plant populations can be gained from studies of local adaptation and acclimation to temperature along environmental, e.g. elevational gradients, to which plant species respond by with various trait changes (Aitken et al., 2008; Alberto et al., 2013; Wagner et al., 2016). Some studies indicate that plants in their native habitat exhibit higher fitness compared to plants from other populations introduced to that same location. Notably, plants from larger populations demonstrate more pronounced evidence of local adaptation (Leimu & Fischer, 2008). Additionally, Lortie & Hierro (2022) provided evidence about plant adaptation to local climate based on the analysis of seed germination, seedling growth, biomass, and other measures. Also, studies of crops confirmed intraspecific variation in pollen thermal tolerance by comparing the effects of temperature on pollen thermotolerance of different cultivars (Kakani et al., 2005; Walters & Isaacs, 2023). On the other hand, Flores-Rentería et al., (2018) tested pollen thermotolerance of *Pinus edulis* across different elevations, but their prediction that pollen from sites with higher air temperature is more tolerant

to high temperature was not confirmed. The possibility of local adaptation of pollen thermotolerance to temperature thus remains an open question.

To fill this knowledge gap, we tested pollen thermotolerance of a common wild plant species, *Lotus corniculatus* s.l., by measuring pollen germination and pollen tube length in response to increasing temperature in 12 populations from two different elevations. We asked the following questions:

1. Does the pollen germination rate and pollen tube length decrease with increasing temperature? We hypothesised that pollen performance is limited by the temperature of 40 °C, because this temperature exceeds the maximum summer temperatures observed in the study region.
2. Does pollen from lowland populations have higher germination rate and pollen tube length at higher temperatures compared to pollen from highland populations? We hypothesized that pollen of plants from the lowland populations has higher termotolerance at higher temperatures because of local adaptation of lowland plants to warmer climate.

Materials & Methods

Study sites

The study was carried out in the southern part of the Czech Republic. We conducted our experiment at six lowland sites (389 – 451 m a.s.l.) in the surroundings of the city of České Budějovice and six highland sites (841 – 1030 m a.s.l.) in the nearby Šumava Mountains (Table1.). We did not measure climatic data in situ because it was not necessary to obtain exact measurements of microclimatic conditions for the purpose of our study. We only aimed to select a set of highland sites generally colder than selected lowland sites. Therefore, we obtained publicly available temperature data released by the Czech Hydrometeorological Institute (https://www.chmi.cz/) from the nearest meteorological stations located within several km from the sites at a similar elevation – one station in the lowland and one in the highland (Table 1). We also downloaded climate data for each site from the Worldclim database (https://www.worldclim.org/, Fick & Hijmans, 2017) based on exact GPS coordinates of the sites. We selected the average maximum temperature of the warmest months as the most biologically relevant among the climate variables in Worldclim because our sampling was done in the summer when the temperatures reach the maximum values. The temperature values for each site were extracted based on its GPS coordinates using the R package *raster* (Hijmans, 2021). The differences in maximum temperature between the lowland (mean 23.9°C) and highland sites (mean 19.6°C) were >4°C (Table 1.). This difference in elevation represents a good proxy for the effect of global warming, because it corresponds to predictions of future

global warming by 2.4°C–4.8°C by the end of the 21st century in the high-emissions scenario SSP5-8.5 according to the Sixth Assessment Report of the IPCC (Lee et al., 2021).

Study species

Our study species is a dicotyledonous forb *Lotus corniculatus* L. s.l. (Fabaceae). We selected this species because of its common occurrence across the Czech Republic spanning the entire climatic gradient from the warmest lowlands to the coldest mountain ranges (Chytrý et al., 2021). *L. corniculatus* has a long flowering period (April – August) and is strictly entomogamous; the most frequent pollinators are solitary bees and bumblebees (Chytrý et al., 2021). Pollen germination in *L. corniculatus* has been studied by Wagner et al. (2016), who tested its cold tolerance and found that pollen of *L. corniculatus* requires temperatures above 5°C for germination and reached >80% germination rate already at 10°C. We are not aware of published data on the effect of higher temperatures on pollen germination in this species.

Table 1. The list of study sites in the lowland and the highland with temperature details.
T_{max} *warmest month* represents the average highest air temperature of the warmest month at the study sites according to Worldclim, T_{max} *three days prior* represents the average of the highest temperature in the day of collection and two days before the collection. The elevation and GPS coordinates of the centre of each site are also provided.

Site ID	Site name	Elevation (m a.s.l.)	T_{max} warmest month (°C)	T_{max} 3 days prior (°C)	GPS coordinates
Lowland 1	Břehov	408	24.1	22.0	49.0193356N, 14.3288503E
Lowland 2	Haklovy Dvory	389	24.0	22.0	48.9990081N, 14.3891314E
Lowland 3	Křenovice	398	24.0	22.0	48.9899975N, 14.3616067E
Lowland 4	Branišov	406	23.9	25.0	48.9782317N, 14.4154694E
Lowland 5	Třebín	414	23.8	25.0	48.9665339N, 14.3844175E
Lowland 6	Kvítkovice	451	23.6	25.0	48.9584728N, 14.3290661E
Highland 1	Strážný CHKO	940	19.1	20.1	48.9290808N, 13.7092800E
Highland 2	Strážný	841	19.6	20.8	48.9174489N, 13.7183381E
Highland 3	Křišťanov	930	19.6	22.5	48.9078519N, 14.0228328E
Highland 4	Koryto	870	20.1	22.5	48.9315508N, 13.9978347E
Highland 5	Arnoštov	890	19.9	22.5	48.9061311N, 14.0014181E
Highland 6	Sněžná	1030	19.0	26.0	48.8961061N, 13.9393994E

Data collection

Before the collection of *L. corniculatus* plants, we prepared 1.5 litres of BK culture medium (Brewbaker & Kwack, 1963) modified for best performance in Fabaceae according to Tushabe & Rosbakh (2021) in the laboratory. We used 50g 10% 1L sucrose, 50 mg boric acid H_3BO_3, 150 mg calcium nitrate $Ca(NO_3)_2$, 100 mg magnesium sulphate heptahydrate $H_{14}MgO_{11}S$, 50 mg potassium nitrate KNO_3 for medium preparation. We made up the medium with 500 ml deionized water. We calibrated the medium to pH 5.5. Finally, we autoclaved the medium for 2 hours at 120°C. We did not add agar, thus the medium remained liquid, which allowed us easier manipulation.

We collected the plants during June – July 2022 in the field. In each of 12 sites, we collected 20 individuals between 10:00 and 16:00 hours, when we observed the highest activity of pollinators. We selected plants with fresh-looking flowers (without visible signs of senescence). Entire plant stems with flowers were cut and immediately placed into plastic zip-lock bags with a piece of wet paper towel and stored in polystyrene cooling boxes at ambient temperature to prevent their wilting during transport to the lab.

The experiments were conducted in the laboratory using fresh pollen during the day of collection. We released the pollen from mature anthers of five flowers per individual using entomological forceps. The pollen was placed to an Eppendorf tube with liquid BK culture medium (described above). Afterwards, we put the Eppendorf tubes in plastic racks to incubators with four different temperature levels (15, 25, 30, and 40°C). These temperature levels were designed based on other pollen germination studies (Kakani et al., 2005; Kaushal et al., 2016; Mesihovic et al., 2016; Walters & Isaacs, 2023) and our own pilot experiments to cover temperatures likely within and above the optimum temperature range. In the lowland parts of the study region, summer temperatures measured 2 m above ground occasionally exceed 30°C, but never reach 40°C according to measurements of the Czech Hydrometeorological Institute (https://www.chmi.cz/), although the temperatures of microhabitats close to the ground where the flowers are located may be higher (Scherrer & Koerner, 2009). Five replicates for each combination of site and temperature level were incubated for 20 hours in the darkness. After the incubation, the samples were placed to the freezer (-20 °C), which allows long-term storage of the samples without damage to the pollen grains (Du et al., 2019). No damage to pollen grains was apparent after taking the samples out of the freezer after approximately two months. We processed the samples (counting of pollen grains and measurements of pollen tube length) during the autumn of the same year.

In the day of pollen processing, samples were left to defrost for 15 minutes at room temperature. Using a pipette, we removed three drops (1 drop = 40 µl) of the medium. We mixed the sample content with pipette suction. Medium with pollen was retrieved with clipped pipette tips with increased diameter to avoid possible damage to the germinated pollen tubes. We put these three drops on one slide and covered them by three cover slides. We put the slides with the

medium under the microscope with an attached camera (Canon EOS 77D). We observed the slides under 160x magnification and took three photos at random locations within each drop; i.e., nine photos per sample in total.

Photos obtained by a camera attached to the microscope, examples obtained under different temperatures in Figure 1, were processed in the Fiji distribution of ImageJ software (Schindelin et al., 2012). We visually counted all pollen grains in each photo and distinguished germinated pollen grains (those with a pollen tube at least the length of the pollen grain diameter). We calculated germination rate as the proportion of germinated grains divided by all pollen grains in a sample (pooled numbers from the nine photos per sample). We also measured the pollen tube length of germinated pollen grains by drawing a line along the full length of each pollen tube and measuring its length in ImageJ. Pollen tubes were measured only if they were longer than the pollen grain diameter. Burst pollen grains were not counted as germinated pollen. In total, we counted 25,427 pollen grains, mean number of pollen grains per photo was 14.5.

Figure 1. Example of photos of pollen grains. Pollen grains incubated at the temperatures of 15°C (A.), 25°C (B.), 30°C (C.), and 40°C (D.).

Data analysis

Data from the experiment were analyzed using a generalized linear mixed model (GLMM) with the glmer function implemented in the lme4 package (Bates et al., 2015) in R version 4.2.2 (R Core Team, 2022). Temperature data from Worldclim for focal sites were extracted in R using the library raster (Hijmans, 2021).

Temperature (a factor with four levels), elevation (a factor with two levels), and their interaction were used as predictors in the analyses. Pollen germination rate (proportion of pollen grains which germinated per sample) and pollen tube length were used as response variables in two separate models (Table 2.). For pollen germination data, we used binomial error distribution and included the number of pollen grains per sample as weights. For pollen tube length data, we used gamma error distribution with log link function. We performed model diagnostics to ensure that the model assumptions were satisfied. The site of plant origin was included as a random effect in both cases. We used likelihood ratio tests to evaluate the statistical significance of the predictors. In addition, we conducted post-hoc multiple comparison tests to compare pollen germination rate and pollen tube length between all combinations of temperature levels, separately for lowland and highland plants, using the function glht in the multcomp library for R (Hothorn, Bretz & Westfall, 2008).(Hothorn, Bretz & Westfall, 2008).

Results

We observed a statistically significant effect of the interaction of the temperature of incubation and the elevation of plant origin on pollen germination rate (χ^2 = 41.82, P = 4.376*10^{-09}, Table 2). Lowland plants had stable germination rate in the temperature range from 15 to 30°C, followed by a strong reduction at 40°C (Figure 2.). While on average between 47and 50% of pollen grains germinated at 15-30°C, pollen germination rate significantly dropped to 36% at 40°C (P<1*10^{-05} based on post-hoc tests comparing pollen germination rate at 40°C compared to the lower temperatures). On the other hand, the germination rate of pollen from highland plants significantly decreased already as the temperature reached 30°C, from 57% at 15°C and 53% at 25°C to 46% at 30°C and 45% at 40°C (P<0.001 based on post-hoc tests comparing pollen germination rates at 30°C or 40°C compared to 15°C or 25°C). In addition, the overall average pollen germination rate significantly differed between lowland and highland sites; the proportion of pollen grains which germinated was on average 8% higher in plants from the highland (z = 2.70, P = 0.0069, Figure 2.).

Figure 2. Pollen germination of plants from the lowland and the highland at four temperature levels. Mean values and SE estimated by a generalised linear mixed effects model are plotted (A. and B.). The fixed effect of temperature is displayed. Pollen germination rate is expressed as the proportion of pollen grains which germinated. The letters above the points show results of multiple comparison tests between all combinations of temperature levels, calculated separately for data on lowland and highland plants. Differences in pollen germination at temperature levels marked by a different letter were statistically significant (p<0.001 in all cases). The comparison of relative values of pollen germination rate in relation to the maximum is shown in C.

Analysis of pollen tube lengths also showed a statistically significant effect of the interaction of the incubation temperature and the elevation of plant origin (χ^2 = 17.12, P = 0.0007, Table 2). However, the differences between lowland and highland sites were less pronounced than in the case of pollen germination rate (Figure 3.). Pollen tube length decreased with increasing temperature in a similar way in lowland and highland populations (Figure 3.). The average pollen tube length was slightly shorter, on average by 68 μm, in plants from the highland compared to the lowland (t = -3.709, P = 0.0002, Figure 3.).

Figure 3. Pollen tube length of plants from the lowland and the highland at four temperature levels. Mean values and SE estimated by a generalised linear mixed effects model are plotted (A. and B.). The fixed effect of temperature is displayed. The letters above the points show results of multiple comparison tests between all combinations of temperature levels, calculated separately for data on lowland and highland plants. Differences in pollen tube length at temperature levels marked by a different letter were statistically significant (p<0.001 in all cases). The comparison of relative values of pollen germination rate in relation to the maximum is shown in C.

Discussion

Until now, the effect of increasing temperature on plant sexual reproduction has been studied mostly in crops because of concerns about the effects of climate change on food production (Hedhly, Hormaza & Herrero, 2009; Zinn, Tunc-Ozdemir & Harper, 2010) but little attention has been paid to the effects of increasing temperature on sexual reproduction of wild plants (Rosbakh et al., 2018). Although species with frequent vegetative reproduction could persist with non-viable pollen, relying on vegetative reproduction leads to the loss of genetic variability and decreases the ability to adapt to unpredictable changes in the environment (Hedhly, Hormaza & Herrero, 2009; Eckert et al., 2010). Hence, research on the direct effects of increasing temperature on sexual reproduction of wild plants is needed to evaluate the threat posed by global warming for plant diversity.

Table 2. The results of generalized linear mixed effects models (GLMM) testing the dependence of pollen germination rate and pollen tube length on different temperature levels and elevation. Parameter estimates, their standard errors (SE), z or t values, and P values are provided for fixed effects of the elevation (lowland x highland), temperature (4 levels), and their interaction. Values of parameter estimates are provided at the scale of the linear predictors. Logit link function was used in the binomial GLMM testing the effect of elevation and temperature on pollen germination rate and log link function was used in the GLMM testing the effect of the predictors on pollen tube length. P values <0.05 are highlighted in bold.

Predictors (fixed effects)	Pollen germination rate				Pollen tube length			
	Estimate	SE	z	P	Estimate	SE	t	P
Intercept	-0.018	0.098	-0.19	0.8524	-1.305	0.071	-18.38	**< 2*10^{-16}**
Temperature 25	-0.053	0.065	-0.82	0.4118	-0.136	0.039	-3.49	**0.0005**
Temperature 30	-0.081	0.063	-1.30	0.1936	-0.425	0.038	-11.18	**< 2*10^{-16}**
Temperature 40	-0.539	0.06	-8.92	**< 2*10^{-16}**	-0.546	0.039	-14.17	**< 2*10^{-16}**
Elevation Highland	0.307	0.114	2.70	**0.0069**	-0.289	0.078	-3.71	**0.0002**
Temperature 25 : Elevation Highland	-0.086	0.090	-0.96	0.3390	0.026	0.051	0.50	0.6154
Temperature 30 : Elevation Highland	-0.369	0.088	-4.19	**2.9*10^{-5}**	0.182	0.052	3.51	**0.0004**
Temperature 40 : Elevation Highland	0.079	0.082	0.96	0.3388	0.093	0.050	1.88	0.0600

The effects of high temperature on pollen thermotolerance

Pollen germination rate and pollen tube length of *Lotus corniculatus* significantly decreased at high temperature. The pollen germination rate was reduced at 40°C in the lowland population and also at 30°C in the highland population, compared to lower temperatures. Although very rarely studied in wild plant species, a negative effect of high temperature on pollen germination rate has been observed in many species of crops (Zinn, Tunc-Ozdemir & Harper, 2010; Kaushal et al., 2016). For example, pollen germination rate of rice (*Oryza sativa*) significantly decreased when exposed to temperature higher than 32 – 35 °C in a study of Rang et al. (2011) and 38°C in a study by Shi et al. (2018). Also, almond (*Prunus dulcis)* had reduced pollen performance at 35°C, which inhibited pollen germination (Sorkheh et al., 2011), similarly to spring wheat (*Triticum aestivum* L.) exposed to 34°C (Bheemanahalli et al., 2019). Optimum temperature for pollen germination in cotton (*Gossypium hirsutum*) was approximately 30°C with a sharp decrease at higher as well as lower temperatures (Kakani et al., 2005). Among crops from

temperate regions, pollen germination rate of the northern highbush blueberry (*Vaccinium corymbosum*) was reduced at temperatures exceeding 35°C and almost completely prevented at 40° (Walters & Isaacs, 2023).

Pollen tube length was also significantly reduced with increasing temperature, but in this case, the reduction was more gradual. Previous studies of crops also provide examples of a negative effect of high temperatures on pollen tube growth. For instance, Longan (*Dimocarpus longan*), a subtropical fruit tree, showed decreased pollen tube length at 40 °C (Pham, Herrero & Hormaza, 2015; Pham et al., 2015). Also. Hebbar et al. (2018) found out that some cultivars of coconut (*Cocos nucifera*) have shorter pollen tubes at temperatures around 40 °C. Decreasing pollen tube length with increasing temperature was also observed in *Pistacia* spp (Acar & Kakani, 2010), in cotton (*Gossypium hirsutum*) at temperatures above the optimum temperature of 30°C (Kakani et al., 2005), and in the northern highbush blueberry (*Vaccinium corymbosum*) also at temperatures exceeding 30°C (Walters & Isaacs, 2023).

Signs of local adaptation

As hypothesized, we observed differences in the effect of increasing temperature on the pollen germination rate of lowland and highland populations of *L. corniculatus*. While pollen germination rate was stable at the incubation temperature of 15-30°C in the lowland plants and a reduction of pollen germination rate was observed only at 40°C, pollen germination rate of the highland plants was reduced already when the temperature of incubation reached 30°C. We suggest that this is due to local adaptation of the lowland plant populations to higher temperatures. The difference in the elevation of the lowland and highland sites was over 400 m and climate data show that the differences in long-term average temperatures between the lowland and highland sites exceed 4°C (Table 1). Plants in the lowland populations thus experience substantially higher temperatures during their flowering period and this provides selection pressure to adapt to higher temperatures.

Variation of plant traits along environmental gradients, which suggests local adaptation of plant populations to varying conditions has been observed in many species (Aitken et al., 2008; Alberto et al., 2013; Wagner et al., 2016). A large amount of indirect evidence comes from numerous common garden experiments which showed that various plant traits and different measures of plant performance depend on the origin of populations along the environmental gradient. Direct evidence from reciprocal transplant experiments is mixed, but some studies showed that plants at their native site had higher fitness than plants from other populations introduced to that site, with plants from larger populations showing stronger evidence for local adaptation (Leimu & Fischer, 2008). A recent meta-analysis of seed and seedling transplant experiments provided a strong overall support for plant adaptation to local climate based on the analysis of seed germination, seedling growth, biomass, and other measures (Lortie & Hierro, 2022). However, the only reciprocal transplant study on *Lotus corniculatus* we are aware of found no evidence for adaptation to climate based on comparisons of total biomass and fecundity

of plants reciprocally transplanted among three sites (Macel et al., 2007). Despite the large body of research on local adaptation, we are not aware of previous studies focusing on adaptation of pollen thermotolerance to increasing temperature, with the exception of a study of *Pinus edulis* along an elevational gradient (Flores-Rentería et al., 2018). However, that study did not confirm the hypothesis that pollen from plants growing at lower elevations is more tolerant to higher temperatures.

Based on the design of our experiment, we could not obtain a precise estimate of the optimum and maximum temperature for pollen germination of *L. corniculatus*. However, our choice of temperature levels reflected the local climate. Specifically, we observed that pollen germination rate of highland plants was significantly reduced at 30°C compared to 15 and 25°C, while lowland plants had stable pollen germination rate at 15, 25, and 30°C and pollen germination rate was significantly reduced only at 40°C. Data from nearby meteorological stations show that while maximum daily temperatures in the lowland (ca. 400 m a.s.l.) regularly exceed 30°C, they very rarely do so in the highland (ca. 900 m a.s.l.). For example, in 2022, 30°C was exceeded on 15 days at a station in České Budějovice (elevation 381 m a.s.l.) near the lowland sites, but never at a station in Nicov (elevation 935 m a.s.l.) near the highland sites, where the maximum recorded temperature was 29.0°C. This is reflected in the different effects of increasing temperature on the germination rate of pollen grains of plants from the highland compared to the lowland populations.

Possible acclimation and confounding factors

A caveat in our interpretation of the differences between the lowland and highland populations is that pollen thermotolerance may be affected also by the temperature the plants experienced in the field prior to the collection of pollen; i.e., there may be an effect of acclimation leading to acquired tolerance of high temperature (Sung et al., 2003). Due to acclimation, plants are able to express heat stress-responsive genes (Müller & Rieu, 2016) and also photosynthesize more efficiently in new climate conditions (Yamori, Hikosaka & Way, 2014). A study by Firon et al. (2012) also demonstrated a positive effect of previous exposure of tomato plants to high non-lethal temperature on their pollen thermotolerance under subsequent heat stress.

Accounting for the effect of acclimation in plants collected in the field is not trivial. To evaluate whether acclimation might bias our results, we compared temperature data from the meteorological stations near the lowland and highland sites. We specifically looked up the maximum air temperature on the day of sample collection and two days before that and compared the average values at individual sites. The average maximum air temperature during this time period ranged from 22.0 to 25.0°C at the lowland sites and between 20.1 and 26.0°C at the highland sites (Table 1). Plants in the lowland and the highland thus experienced very similar temperatures prior to pollen collection and we argue that the difference in the effect of high temperature on pollen germination in the lowland and highland plans observed in the lab is thus

unlikely to be biased by the effect of differences in temperature experienced by the plants in the field.

Apart from the effect of temperature, we observed that the average pollen germination rate was higher in plants from the highland populations compared to the lowland populations, but the pollen tubes were slightly but statistically significantly longer in plants from the lowland populations. We do not have a good explanation for these differences, although we suspect that higher pollen germination rate in plants from the highland might be related either to differences in general plant condition possibly caused by higher water availability in the mountains (Chytrý, 2017), or by differences in air humidity, which is known to affect pollen thermotolerance (Aronne, 1999; Leech, Simpson & Whitehouse, 2002). The negative effect of drought on pollen germination was confirmed in multiple crops, e.g. in maize (*Zea mays* L.) (Bheemanahalli et al., 2022), soybean (*Glycine max*) (Poudel et al., 2023), and rice (*Oryza sativa*) (Rao et al., 2019). We did not measure precipitation at the sites, but data from a drought monitoring project by the Global Change Research Institute of the Czech Academy of Sciences (https://www.intersucho.cz/) show that the area where the highland sites were located consistently has higher soil water content than the lowland area, including during the time of our sampling. Another important factor could be whether the plant is exposed to direct sunshine most of the day or is located in the shadow (Corbet, 1990), which we did not record.

Interestingly, a previous experiment on pollen germination of *Pinus edulis* showed variable pollen germination across an elevation gradient, with the highest pollen germination rate at intermediate elevations where the plants are in their optimal conditions (Flores-Rentería et al., 2018). Also, Rosbakh & Poschlod (2016) observed variable pollen germination rate of multiple forb species across an elevational gradient. They confirmed that pollen of lowland species germinates and grows pollen tubes at relatively high temperatures compared to highland species which perform better at lower temperatures. In addition, McKee & Richards (1998) showed that increasing temperature may hamper seed set of several *Primula* species. On the other hand, low temperature can also negatively affect pollen performance as in the case of *Tilia cordata* where it prevents sexual reproduction at the northern distribution limit (Pigott & Huntley, 1980). Minimum temperature requirements for pollen germination and pollen tube grows are also species specific depending on their elevation of occurrence (Rosbakh & Poschlod, 2016) and time of the year when they are flowering (Wagner eta al. 2016).

Multiple studies demonstrated that different environmental drivers, such as temperature, water, and nutrient availability often have interactive effects on plant vegetative growth, floral traits, and interactions with pollinators (Hoover et al., 2012; Descamps et al., 2018; Akter & Klečka, 2022). Hence, understanding the effects of a broader set of local environmental conditions on pollen thermotolerance will certainly be an interesting avenue for future research.

Conclusions

Our experiment with pollen germination in lowland and highland populations of *Lotus corniculatus*, which were tested across four temperature levels, provides evidence that pollen germination is more sensitive to high temperatures in highland plants compared to lowland plants. This is likely the result of local adaptation to higher temperatures in lowland populations. In addition, we found a negative effect of increasing temperature on pollen tube length, which may further prevent fertilization at high temperatures. We conclude that more research on the temperature-dependence of pollen thermotolerance of wild plants is needed to understand the impact of climate change on plant reproduction. Our data show that increasing temperature may hamper sexual reproduction of wild plants, similarly to crops, but differences among populations suggest potential for adaptation.

Acknowledgements

We would like to thank Jenna Walters for discussions about pollen germination, Dagmar Hucková and Davide Panzeri for their help with the preparation of culture medium, and Pavel Duda for his help in the field. We would also like to thank Nicola Tommasi and Zdeněk Faltýnek Fric for useful comments about data analyses. Last but not least, we are grateful to Eva Kaštovská and Petr Čapek for providing their incubators for the experiment.

References

Acar I, Kakani VG. 2010. The effects of temperature on in vitro pollen germination and pollen tube growth of *Pistacia* spp. Scientia Horticulturae 125:569–572. DOI: 10.1016/j.scienta.2010.04.040.

Aitken SN, Yeaman S, Holliday JA, Wang T, Curtis-McLane S. 2008. Adaptation, migration or extirpation: climate change outcomes for tree populations. Evolutionary Applications 1:95–111. DOI: 10.1111/j.1752-4571.2007.00013.x.

Akter A, Klečka J. 2022. Water stress and nitrogen supply affect floral traits and pollination of the white mustard, *Sinapis alba* (Brassicaceae). PeerJ 10:e13009. DOI: 10.7717/peerj.13009.

Alberto FJ, Aitken SN, Alía R, González-Martínez SC, Hänninen H, Kremer A, Lefèvre F, Lenormand T, Yeaman S, Whetten R, Savolainen O. 2013. Potential for evolutionary responses to climate change – evidence from tree populations. Global Change Biology 19:1645–1661. DOI: 10.1111/gcb.12181.

Aronne G. 1999. Effects of relative humidity and temperature stress on pollen viability of *Cistus incanus* and *Myrtus communis*. Grana 38:364–367. DOI: 10.1080/00173130050136154.

Bates D, Mächler M, Bolker B, Walker S. 2015. Fitting linear mixed-effects models using lme4. Journal of Statistical Software 67:1–48. DOI: 10.18637/jss.v067.i01.

Bheemanahalli R, Ramamoorthy P, Poudel S, Samiappan S, Wijewardane N, Reddy KR. 2022. Effects of drought and heat stresses during reproductive stage on pollen germination, yield, and leaf reflectance properties in maize (*Zea mays* L.). Plant Direct 6. DOI: 10.1002/pld3.434.

Bheemanahalli R, Sunoj VSJ, Saripalli G, Prasad PVV, Balyan HS, Gupta PK, Grant N, Gill KS, Jagadish SVK. 2019. Quantifying the impact of heat stress on pollen germination, seed set, and grain filling in spring wheat. Crop Science 59:684–696. DOI: 10.2135/cropsci2018.05.0292.

Biella P, Bogliani G, Cornalba M, Manino A, Neumayer J, Porporato M, Rasmont P, Milanesi P. 2017. Distribution patterns of the cold adapted bumblebee *Bombus alpinus* in the Alps and hints of an uphill shift (Insecta: Hymenoptera: Apidae). Journal of Insect Conservation 21:357–366. DOI: 10.1007/s10841-017-9983-1.

Brewbaker JL, Kwack BH. 1963. The essential role of calcium ion in pollen germination and pollen tube growth. American Journal of Botany 50:859–865. DOI: 10.1002/j.1537-2197.1963.tb06564.x.

Brink RA. 1924. The physiology of pollen I. The requirements for growth. American Journal of Botany 11:218–228. DOI: 10.1002/j.1537-2197.1924.tb05773.x.

Chytrý M. 2017. Physical geography of the Czech Republic. In: Chytrý M, Danihelka J, Kaplan Z, Pyšek P eds. Flora and Vegetation of the Czech Republic. Plant and Vegetation. Cham: Springer International Publishing, 1–23. DOI: 10.1007/978-3-319-63181-3_1.

Chytrý M, Danihelka J, Kaplan Z, Wild J, Holubová D, Novotný P, Řezníčková M, Rohn M, Dřevojan P, Grulich V, Klimešová J, Lepš J, Lososová Z, Pergl J, Sádlo J, Šmarda P, Štěpánková P, Tichý L, Axmanová I, Bartušková A, Blažek P, Chrtek J, Fischer FM, Guo W-Y, Herben T, Janovský Z, Konečná M, Kühn I, Moravcová L, Petřík P, Pierce S, Prach K, Prokešová H, Štech M, Těšitel J, Těšitelová T, Večeřa M, Zelený D, Pyšek P. 2021. Pladias Database of the Czech flora and vegetation. Preslia 93:1–87. DOI: 10.23855/preslia.2021.001.

Corbet SA. 1990. Pollination and the weather. Israel Journal of Botany 39:1–2:13–30. DOI: 10.1080/0021213X.1990.10677131.

Das S, Krishnan P, Nayak M, Ramakrishnan B. 2014. High temperature stress effects on pollens of rice (*Oryza sativa* L.) genotypes. Environmental and Experimental Botany 101:36–46. DOI: 10.1016/j.envexpbot.2014.01.004.

Descamps C, Quinet M, Baijot A, Jacquemart A-L. 2018. Temperature and water stress affect plant–pollinator interactions in *Borago officinalis* (Boraginaceae). Ecology and Evolution 8:3443–3456. DOI: 10.1002/ece3.3914.

Du G, Xu J, Gao C, Lu J, Li Q, Du J, Lv M, Sun X. 2019. Effect of low storage temperature on pollen viability of fifteen herbaceous peonies. Biotechnology Reports 21:e00309. DOI: 10.1016/j.btre.2019.e00309.

Eckert CG, Kalisz S, Geber MA, Sargent R, Elle E, Cheptou P-O, Goodwillie C, Johnston MO, Kelly JK, Moeller DA, Porcher E, Ree RH, Vallejo-Marín M, Winn AA. 2010. Plant mating systems in a changing world. Trends in Ecology & Evolution 25:35–43. DOI: 10.1016/j.tree.2009.06.013.

Fazlioglu F, Wan JSH, Chen L. 2020. Latitudinal shifts in mangrove species worldwide: evidence from historical occurrence records. Hydrobiologia 847:4111–4123. DOI: 10.1007/s10750-020-04403-x.

Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global land areas. International Journal of Climatology 37:4302–4315. DOI: 10.1002/joc.5086.

Firon N, Pressman E, Meir S, Khoury R, Altahan L. 2012. Ethylene is involved in maintaining tomato (*Solanum lycopersicum*) pollen quality under heat-stress conditions. AoB PLANTS 2012:pls024. DOI: 10.1093/aobpla/pls024.

Flores-Rentería L, Whipple AV, Benally GJ, Patterson A, Canyon B, Gehring CA. 2018. Higher temperature at lower elevation sites fails to promote acclimation or adaptation to heat stress during pollen germination. Frontiers in Plant Science 9:536. DOI: 10.3389/fpls.2018.00536.

Freeman BG, Lee-Yaw JA, Sunday JM, Hargreaves AL. 2018. Expanding, shifting and shrinking: The impact of global warming on species' elevational distributions. Global Ecology and Biogeography 27:1268–1276. DOI: 10.1111/geb.12774.

He G, Hu F, Ming J, Liu C, Yuan S. 2017. Pollen viability and stigma receptivity in *Lilium* during anthesis. Euphytica 213:231. DOI: 10.1007/s10681-017-2019-9.

Hebbar KB, Rose HM, Nair AR, Kannan S, Niral V, Arivalagan M, Gupta A, Samsudeen K, Chandran KP, Chowdappa P, Vara Prasad PV. 2018. Differences in in vitro pollen germination and pollen tube growth of coconut (*Cocos nucifera* L.) cultivars in response to high temperature stress. Environmental and Experimental Botany 153:35–44. DOI: 10.1016/j.envexpbot.2018.04.014.

Hedhly A, Hormaza JI, Herrero M. 2005. The effect of temperature on pollen germination, pollen tube growth, and stigmatic receptivity in peach. Plant Biology 7:476–483. DOI: 10.1055/s-2005-865850.

Hedhly A, Hormaza JI, Herrero M. 2009. Global warming and sexual plant reproduction. Trends in Plant Science 14:30–36. DOI: 10.1016/j.tplants.2008.11.001.

Hijmans RJ. 2021. raster: Geographic data analysis and modeling.

Hoover SER, Ladley JJ, Shchepetkina AA, Tisch M, Gieseg SP, Tylianakis JM. 2012. Warming, CO_2, and nitrogen deposition interactively affect a plant-pollinator mutualism. Ecology Letters 15:227–234. DOI: 10.1111/j.1461-0248.2011.01729.x.

Hothorn T, Bretz F, Westfall P. 2008. Simultaneous inference in general parametric models. Biometrical Journal 50:346–363. DOI: 10.1002/bimj.200810425.

IPCC. 2021. Climate Change 2021: The Physical Science Basis. Contribution of working group i to the sixth assessment report of the intergovernmental panel on climate change. In Press. DOI: 10.1017/9781009157896.

Kakani VG, Reddy KR, Koti S, Wallace TP, Prasad PVV, Reddy VR, Zhao D. 2005. Differences in in vitro pollen germination and pollen tube growth of cotton cultivars in response to high temperature. Annals of Botany 96:59–67. DOI: 10.1093/aob/mci149.

Kaushal N, Bhandari K, Siddique KHM, Nayyar H. 2016. Food crops face rising temperatures: An overview of responses, adaptive mechanisms, and approaches to improve heat tolerance. Cogent Food & Agriculture 2. DOI: 10.1080/23311932.2015.1134380.

Lee J-Y, Marotzke J, Bala G, Cao L, Corti S, Dunne JP, Engelbrecht F, Fischer E, Fyfe JC, Jones C, Maycock A, Mutemi J, Ndiaye O, Panickal S, Zhou T, Milinski S, Yun K-S, Armour K, Bellouin N, Bethke I, Byrne MP, Cassou C, Chen D, Cherchi A, Christensen HM, Connors SL, Di Luca A, Drijfhout SS, Fletcher CG, Forster P, Garcia-Serrano J, Gillett NP, Kaufmann DS, Keller DP, Kravitz B, Li H, Liang Y, MacDougall AH, Malinina E, Menary M, Merryfield WJ, Min S-K, Nicholls ZRJ, Notz D, Pearson B, Priestley MDK, Quaas J, Ribes A, Ruane AC, Sallee J-B, Sanchez-Gomez E, Seneviratne SI, Slangen ABA, Smith C, Stuecker MF, Swaminathan R, Thorne PW, Tokarska KB, Toohey M, Turner A, Volpi D, Xiao C, Zappa G. 2021. Future global climate: scenario-based projections and near-term information. In: Masson-Delmotte V, Zhai P, Pirani A, Connors SL, Pean C, Berger S, Caud N, Chen Y, Goldfarb L, Gomis MI, Huang M, Leitzell K, Lonnoy E, Matthews JBR, Maycock TK, Waterfield T, Yelekçi O, Yu R, Zhou B eds. Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth : Assessment Report of the Intergovernmental Panel on Climate Change. Genf, Switzerland: IPCC, 1–195.

Leech L, Simpson DW, Whitehouse AB. 2002. Effect of temperature and relative humidity on pollen germination in four strawberry cultivars. Acta Horticulturae:261–263. DOI: 10.17660/ActaHortic.2002.567.53.

Leimu R, Fischer M. 2008. A meta-analysis of local adaptation in plants. PLOS ONE 3:e4010. DOI: 10.1371/journal.pone.0004010.

Lenoir J, Gégout JC, Marquet PA, de Ruffray P, Brisse H. 2008. A significant upward shift in plant species optimum elevation during the 20th century. Science 320:1768–1771. DOI: 10.1126/science.1156831.

Lewis D. 1942. The physiology of incompatibility in plants - I. The effect of temperature. Proceedings of the Royal Society of London. Series B - Biological Sciences 131:13–26. DOI: 10.1098/rspb.1942.0015.

Lohani N, Singh MB, Bhalla PL. 2020. High temperature susceptibility of sexual reproduction in crop plants. Journal of Experimental Botany 71:555–568. DOI: 10.1093/jxb/erz426.

Lohani N, Singh MB, Bhalla PL. 2022. Short-term heat stress during flowering results in a decline in canola seed productivity. Journal of Agronomy and Crop Science 208:486–496. DOI: 10.1111/jac.12534.

Lortie CJ, Hierro JL. 2022. A synthesis of local adaptation to climate through reciprocal common gardens. Journal of Ecology 110:1015–1021. DOI: 10.1111/1365-2745.13664.

Lovane M, Cirillo A, Izzo LG, Di Vaio C, Aronne G. 2021. High temperature and humidity affect pollen viability and longevity in *Olea europaea* L. Agronomy 12:1. DOI: 10.3390/agronomy12010001.

Macel M, Lawson CS, Mortimer SR, Šmilauerova M, Bischoff A, Crémieux L, Doležal J, Edwards AR, Lanta V, Bezemer TM, Van Der Putten WH, Igual JM, Rodriguez-Barrueco C, Müller-Schärer H, Steinger T. 2007. Climate vs. soil factors in local adaptation of two common plant species. Ecology 88:424–433. DOI: 10.1890/0012-9658(2007)88[424:CVSFIL]2.0.CO;2.

McKee J, Richards AJ. 1998. The Effect of temperature on reproduction in five *Primula* species. Annals of Botany 82:359–374. DOI: 10.1006/anbo.1998.0697.

Mesihovic A, Iannacone R, Firon N, Fragkostefanakis S. 2016. Heat stress regimes for the investigation of pollen thermotolerance in crop plants. Plant Reproduction 29:93–105. DOI: 10.1007/s00497-016-0281-y.

Müller F, Rieu I. 2016. Acclimation to high temperature during pollen development. Plant Reproduction 29:107–118. DOI: 10.1007/s00497-016-0282-x.

Ollerton J, Winfree R, Tarrant S. 2011. How many flowering plants are pollinated by animals? Oikos 120:321–326. DOI: 10.1111/j.1600-0706.2010.18644.x.

Pham VT, Herrero M, Hormaza JI. 2015. Effect of temperature on pollen germination and pollen tube growth in longan (*Dimocarpus longan* Lour.). Scientia Horticulturae 197:470–475. DOI: 10.1016/j.scienta.2015.10.007.

Pigott CD, Huntley JP. 1980. Factors controlling the distribution of Tilia cordata at the northern limits of its geographical range. ii. history in north-west England. New Phytologist 84:145–164. DOI: 10.1111/j.1469-8137.1980.tb00757.x.

Poudel S, Vennam RR, Shrestha A, Reddy KR, Wijewardane NK, Reddy KN, Bheemanahalli R. 2023. Resilience of soybean cultivars to drought stress during flowering and early-seed setting stages. Scientific Reports 13:1277. DOI: 10.1038/s41598-023-28354-0.

R Core Team. 2022. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. https://www.R-project.org/.

Rang ZW, Jagadish SVK, Zhou QM, Craufurd PQ, Heuer S. 2011. Effect of high temperature and water stress on pollen germination and spikelet fertility in rice. Environmental and Experimental Botany 70:58–65. DOI: 10.1016/j.envexpbot.2010.08.009.

Rao G, Ashraf U, Kong L, Mo Z, Xiao L, Zhong K, Rasul F, Tang X. 2019. Low soil temperature and drought stress conditions at flowering stage affect physiology and pollen traits of rice. Journal of Integrative Agriculture 18:1859–1870. DOI: 10.1016/S2095-3119(18)62067-2.

Rosbakh S, Pacini E, Nepi M, Poschlod P. 2018. An unexplored side of regeneration niche: seed quantity and quality are determined by the effect of temperature on pollen performance. Frontiers in Plant Science 9:1036. DOI: 10.3389/fpls.2018.01036.

Rosbakh S, Poschlod P. 2016. Minimal temperature of pollen germination controls species distribution along a temperature gradient. Annals of Botany 117:1111–1120. DOI: 10.1093/aob/mcw041.

Scherrer D, Korner C. 2009. Infra-red thermometry of alpine landscapes challenges climatic warming projections: Thermometry of alpine landscapes. Global Change Biology:no-no. DOI: 10.1111/j.1365-2486.2009.02122.x.

Schindelin J, Arganda-Carreras I, Frise E, Kaynig V, Longair M, Pietzsch T, Preibisch S, Rueden C, Saalfeld S, Schmid B, Tinevez J-Y, White DJ, Hartenstein V, Eliceiri K, Tomancak P, Cardona A. 2012. Fiji: an open-source platform for biological-image analysis. Nature Methods 9:676–682. DOI: 10.1038/nmeth.2019.

Shi W, Li X, Schmidt RC, Struik PC, Yin X, Jagadish SVK. 2018. Pollen germination and in vivo fertilization in response to high-temperature during flowering in hybrid and inbred rice: Rice pollen & in vivo fertilization under heat. Plant, Cell & Environment 41:1287–1297. DOI: 10.1111/pce.13146.

Sorkheh K, Shiran B, Rouhi V, Khodambashi M, Wolukau JN, Ercisli S. 2011. Response of in vitro pollen germination and pollen tube growth of almond (*Prunus dulcis* Mill.) to temperature, polyamines and polyamine synthesis inhibitor. Biochemical Systematics and Ecology 39:749–757. DOI: 10.1016/j.bse.2011.06.015.

Steinacher G, Wagner J. 2012. Effect of temperature on the progamic phase in high-mountain plants: Temperature effects on the progamic phase in mountain plants. Plant Biology 14:295–305. DOI: 10.1111/j.1438-8677.2011.00498.x.

Sung D-Y, Kaplan F, Lee K-J, Guy CL. 2003. Acquired tolerance to temperature extremes. Trends in Plant Science 8:179–187. DOI: 10.1016/S1360-1385(03)00047-5.

Tushabe D, Rosbakh S. 2021. A compendium of in vitro germination media for pollen research. Frontiers in Plant Science 12.

Wagner J, Gastl E, Kogler M, Scheiber M. 2016. Cold tolerance of the male gametophyte during germination and tube growth depends on the flowering time. Plants 6:2. DOI: 10.3390/plants6010002.

Walters J, Isaacs R. 2023. Pollen germination and tube growth in northern highbush blueberry are inhibited by extreme heat. HortScience 58:635–642. DOI: 10.21273/HORTSCI17075-23.

Wang Y, Tao H, Tian B, Sheng D, Xu C, Zhou H, Huang S, Wang P. 2019. Flowering dynamics, pollen, and pistil contribution to grain yield in response to high temperature during maize flowering. Environmental and Experimental Botany 158:80–88. DOI: 10.1016/j.envexpbot.2018.11.007.

Yamori W, Hikosaka K, Way DA. 2014. Temperature response of photosynthesis in C3, C4, and CAM plants: temperature acclimation and temperature adaptation. Photosynthesis Research 119:101–117. DOI: 10.1007/s11120-013-9874-6.

Young LW, Wilen RW, Bonham-Smith PC. 2004. High temperature stress of *Brassica napus* during flowering reduces micro- and megagametophyte fertility, induces fruit abortion, and disrupts seed production. Journal of Experimental Botany 55:485–495. DOI: 10.1093/jxb/erh038.

Zinn KE, Tunc-Ozdemir M, Harper JF. 2010. Temperature stress and plant sexual reproduction: uncovering the weakest links. Journal of Experimental Botany 61:1959–1968. DOI: 10.1093/jxb/erq053.