1   **Song evolution and diversification patterns are uncorrelated in honeyeaters**

4   Eleanor M. Hay[1], Steven L. Chown[1], Matthew D. McGee[1]

7   [1] School of Biological Sciences, Monash University, Melbourne, VIC 3800, Australia

10   *Corresponding author; email: eleanor.m.hay@outlook.com

ABSTRACT

Birdsongs are well-known acoustic signals and are thought to play a key role in sexual

selection. Despite the established role of birdsong in sexual selection and reproductive

isolation at microevolutionary scales, its macroevolutionary impacts on clade diversification

remain largely unexplored. Here, we test the hypothesis that shifts in song evolution

influence diversification using honeyeaters, a diverse clade of songbirds restricted to

Australasia. Using song data for 163 honeyeater species, we employ phylogenetic

comparative models of trait evolution to detect shifts in optima for song evolution and use

hidden state-dependent speciation and extinction models to assess the influence of these shifts

on diversification. Contrary to our expectations, we find no correlation between shifts in song

evolution and shifts in diversification rates, and models suggest other factors play a role.

Overall, shifts in song evolution do not significantly contribute to diversification in

honeyeaters, likely due to other ecological factors influencing diversification in this group

including range size and dispersal ability.

Keywords: sexual signals, speciation, extinction, macroevolution, RevBayes, trait evolution

INTRODUCTION

Determining what factors cause species diversification is a central goal of many evolutionary studies. The influence of morphological traits (Cooney & Thomas, 2021; Rabosky et al., 2013; Simões et al., 2020) and environmental factors such as ecological opportunity (Dumont et al., 2012; Mahler et al., 2010), on rates of speciation and diversification have been a major focus, but other factors including sexual signals, can also play a role (Maan & Seehausen, 2011; Ritchie, 2007; Wagner et al., 2012). Divergence and development of reproductive isolation are key initial phases in speciation events (Coyne & Orr, 2004; Hernández-Hernández et al., 2021; Jarvis, 2021). Sexual signals frequently emerge as the most divergent traits between closely related species and often experience strong selection (Ritchie, 2007; Servedio & Boughman, 2017). The evolution and divergence of sexual signals can outpace ecological divergence (Arnegard et al., 2010) and consequently it has been hypothesized that rapid changes in sexual signals could drive species diversification (Maan & Seehausen, 2011; Servedio & Boughman, 2017).

Vocalizations are in part sexual signals that have the potential to influence speciation. Acoustic signals are used by a wide range of organisms and are complex, behavioral traits that enable individuals of the same species to identify one another (Laidre & Johnstone, 2013). Acoustic signals have broad-scale functions in species recognition and fine-scale functions in mate choice, thus potentially influencing speciation (Catchpole & Slater, 2003; Wilkins et al., 2013). Divergence of acoustic signals has played a role in the early stages of speciation in a wide range of organisms including cicadas (Marshall et al., 2008), crickets (Gray & Cade, 2000; Tinghitella et al., 2018; Zuk et al., 2006), frogs (Boul et al., 2007), and birds (Grant & Grant, 2006). In the context of reproductive character displacement, however, the strength of selection on signals depends on range overlap, with sexual signals expected to be under greater divergence when species are sympatric rather than allopatric (Maan &

Seehausen, 2011). Therefore, at macroevolutionary scales (i.e., clade or family-level), the

strength of selection on acoustic signals is expected to vary between species and among

clades. It remains unclear whether the processes of divergence, assortative mating, and

selection of acoustic traits, which occur at the microevolutionary scale, can have

macroevolutionary consequences for clade diversification.

Birdsong is one of the best-known acoustic signals and is an important sexual signal.

Male birds sing to attract females: consequently, song can act as an important premating

isolation barrier and influence speciation (Price, 2008; Tobias et al., 2020). Indeed, it has

been long proposed that song evolution could explain the diversity of passerine birds

(Baptista & Trail, 1992; Fitzpatrick, 1988). The role of birdsong in speciation and

reproductive isolation has been studied at fine scales, and empirical evidence supports

theoretical expectations. Divergence of song has contributed to reproductive isolation in

warblers (Irwin et al., 2001), and is thought to have influenced the early stages of peripatric

speciation in antbirds in Amazonia (Seddon & Tobias, 2007). Additionally, long-term studies

on Darwin's finches have demonstrated that sexual selection on differences in vocalizations,

resulting from changes in beak shape morphology, have impacted mate choice and

contributed to reproductive isolation and speciation (Grant & Grant, 2006; Podos, 2001;

Podos & Nowicki, 2004). Although acoustic signals are commonly hypothesized to influence

diversification (Wilkins et al., 2013), most studies have focused on population or sister

species comparisons, and empirical studies testing the influence of signal divergence on

speciation at broader scales remain scarce (Mason et al., 2017). Where the relationship

between song and speciation has been investigated at macroevolutionary scales, bursts in

rates of speciation coincide with bursts in song evolution, however, no significant

correlations have been uncovered at the family-level (Mason et al., 2017), with the important

caveat that bursts in song evolution were examined via shifts in evolutionary rates measured

with Brownian motion models, rather than via more complex models that can incorporate

changes in optima position (Uyeda and Harmon 2014).

The relationship between diversification and birdsong has proven increasingly

difficult to disentangle, largely due to song itself being a complex trait, which in birds is

influenced by developmental mode and constrained by factors such as beak morphology,

body size, and the environment (Derryberry et al., 2018; Friedman et al., 2019; Friis et al.,

2022). The majority of macroevolutionary studies on birdsong have therefore focused on the

underlying drivers of song evolution. Like most sexual signals, acoustic signals are expected

to diverge between close relatives, especially if ranges overlap, potentially leading to a lack

of phylogenetic signal in the trait (Pfennig & Pfennig, 2010). However, song frequency is

understood to be phylogenetically conserved (Mikula et al., 2021), largely due to constraints

imposed by morphological traits, especially body size (Derryberry et al., 2018; Pearse et al.,

2018). Moreover, the developmental mode of song is thought to influence song evolution and

could complicate its relationship with speciation. Birdsong ranges from learned to innate, and

learning is understood to be a form of phenotypic plasticity which is influenced by cultural

transmission and can impact evolutionary outcomes (Pfennig et al., 2010). Learned songs are

typically associated with higher rates of evolution (Mason et al., 2017) and divergence

(Lachlan & Servedio, 2004) than innate songs, although conflicting evidence exists for this

(Freeman et al., 2017). Such complexity in the evolution of acoustic traits provides a

challenge for diversification approaches and can result in disparity when investigations are

conducted over varying phylogenetic and spatial scales.

To overcome such limitations, we test the relationship between song and

diversification using a monophyletic clade restricted to a particular region, recognizing the

power that focusing on a diverse single clade can bring (Marquet et al., 2004). Thus, we focus

on honeyeaters (Aves: Meliphagidae), a phenotypically diverse clade of birds (192 species;

Gill et al., 2020) that range throughout Australasia and are known for their distinctive

vocalizations (Higgins et al., 2008). Song in honeyeaters is learned and varies greatly among

species, with some species exhibiting simple and monotonal songs, whereas others have

complex and variable songs (Fig. 1). This variation in vocalizations, combined with the high

diversity and endemism of honeyeaters, makes this group an ideal model for studying the

evolution of vocalizations and their potential role in species diversification.

Specifically, we use honeyeaters as a model clade to test the hypothesis that shifts in

song evolution are correlated with shifts in diversification. We predict that if song is a major

driver of diversification, then shifts in song evolution toward evolutionary optima will

promote speciation and be correlated with shifts in diversification. Using a dataset of five

song variables for 163 species of honeyeater, we model song evolution and identify shifts in

song evolution, and then test whether these shifts are associated with significant increases in

rates of diversification.

METHODS

Song and phylogenetic data

We use a dataset of male honeyeater song traits from a previous study (Hay et al., 2024).

Containing data for 163 honeyeater species, the song dataset considers five relevant aspects

of song (Fig. 1; Table S1) which have commonly been explored in past song studies

(Derryberry et al., 2018; Mason & Burns, 2015; Mason et al., 2017): peak frequency

(frequency which the most sound energy was produced), maximum frequency (maximum

frequency of the highest pitched note in song), minimum frequency (minimum frequency of

the lowest pitched note in song), song bandwidth (difference between maximum and

minimum frequency), and song pace (number of notes/song duration). Song variables were

$\log_{10}$-transformed prior to statistical analysis to meet parametric assumptions of normality

and homogeneity of variance, and because logarithmic scales of sound frequency correspond

to how birds perceive and modulate sound (Cardoso, 2013).

To model trait evolution and estimate speciation rate we used a honeyeater phylogeny

from Hay et al. (2022), which contains 192 species of honeyeater. The phylogeny was

trimmed to represent the 163 honeyeater species for which we had song data and using the

'*drop.tip*' function from the 'ape' package (Paradis et al., 2004) in R (R Core Team, 2013).

Trait values are predicted to be more similar between closely related species, however sexual

signals are often expected to be the most divergent traits between species (Servedio &

Boughman, 2017). To test for how conserved song traits are, we used the '*phylosig*' function

from the R package 'phytools' (Revell, 2012), and estimated the phylogenetic signal of each

song variable using Pagel's lambda (Pagel, 1999) and a likelihood ratio test (Table S2).

Pagels lambda is a scaling parameter that ranges from 0 to 1. Using this measure, a lambda of

0 indicates no correlation between species, and a lambda of 1 indicates that the correlation of

trait values between species is equal to expectations of Brownian motion (Revell et al., 2008).

Analyses

A range of phylogenetic comparative methods, implemented through the program RevBayes

(Höhna et al., 2016) were used to test the relationship between song and diversification.

RevBayes enables Bayesian inference for phylogenetic comparative methods and uses a

probabilistic graphical model framework which is a powerful generic framework for

specifying and analyzing statistical models (Höhna et al., 2014; Höhna et al., 2016). First, to

explore patterns of song evolution in honeyeaters, we used relaxed Ornstein-Uhlenbeck (OU)

models of trait evolution to identify shifts in trait evolution of song traits in RevBayes.

Additionally, relaxed OU models were implemented in a multivariate framework using

RevBayes and verified using multivariate OU models using the R package 'PhylogeneticEM'

(Bastide et al., 2018). The influence of these shifts on diversification were then tested with

hidden state-dependent speciation and extinction models (Beaulieu & O'Meara, 2016) in

RevBayes (Höhna et al., 2016).

Relaxed OU models of trait evolution were used to model song evolution along the

honeyeater phylogeny. Relaxed OU models allow the optimal phenotype to vary over

branches of the phylogenetic tree to estimate tip-specific measures of OU evolution (Uyeda

and Harmon 2014) and to infer branch-specific evolutionary optima of song evolution in

honeyeaters. Here, we first assessed patterns of OU trait evolution in each of the five song

traits (peak frequency, maximum frequency, minimum frequency, bandwidth, and pace)

separately. Trait evolution is inherently a multivariate process (Revell & Harmon, 2022;

Walsh & Blows, 2009), thus we also implemented multivariate relaxed OU models. The

function '*dnPhyloOrnsteinUhlenbeckREML*' does not enable the inclusion of a correlation

matrix to account for covariation between multivariate traits. To overcome this limitation, we

performed a phylogenetic principal component analysis on the five song variables to ensure

no correlation between traits, using the function *'phyl.pca'* from the R package "phytools"

(Revell, 2012). We considered pPC1 to pPC3, which accounted for 97.6 % of the total

variation, in analysis (Table S3). Relaxed OU models were implemented in RevBayes using

reversible-jump Markov chain Monte Carlo and were executed through the CIPRES Science

Gateway v3.3 online server (Miller et al., 2010). The MCMC sampler was set for 4 runs of

50,000 generations with a burn-in of 10%. Tracer v1.7.1 (Rambaut et al., 2018) was used to

visualize log files and to ensure convergence. Because priors can influence performance of

OU models, particularly the number of expected shifts, all models were repeated with three

different priors on the number of shifts (15, 30, and 45). Model fit was assessed using

stepping-stone and path sampler analyses, and the best model was selected based on marginal

likelihood values. Branch-specific rates of relaxed OU evolution were visualized using the

'*plotTree*' function from the R package 'RevGadgets' (Tribble et al., 2021). Priors had little

effect on estimates, and all cases the preferred models had 45 shifts (Table S4; Fig. S1-S6).

Shifts in song evolution were determined based on tip theta values extracted from relaxed OU

models. Under an OU model, theta represents optimal values in which continuous characters

are assumed to evolve toward. Species were identified to have shifts in song evolution if their

tip theta value varied by more than one standard deviation from the mean. We also classified

species to have a shift if they fell within a clade in which all other species were associated

with shifts (Tables S5-S10).

As an additional measure to test for shifts in trait evolution, and to verify the results

from the multivariate relaxed OU models in RevBayes, we used the R package

'PhylogeneticEM' (Bastide et al., 2018). PhylogeneticEM simulates a scalar OU model of

evolution that allows for maximum likelihood estimation of shifts in OU trait evolution and

uses this approach to automatically detect position of shifts along a phylogeny, while

accounting for correlations among traits (Bastide et al., 2018). The function '*phyloEM*' was

used to test for OU shifts in multivariate trait evolution, using pPC1-pPC3, the same dataset

which was used for the multivariate OU model in RevBayes.

Finally, to test the influence of shifts in song evolution on diversification rates we

used hidden state-dependent speciation and extinction models (HiSSE; Beaulieu & O'Meara,

2016), implemented in RevBayes (Höhna et al., 2016). The HiSSE framework enables testing

of character dependent diversification and additionally models an unobserved hidden

character that potentially impacts species diversification rates. HiSSE models test the

influence of binary characters on speciation and extinction rates and can disentangle rate

variation due to the character of interest with the inclusion of a hidden state. HiSSE

represents a substantial improvement on previous state-dependent diversification models

(Maddison & FitzJohn, 2015; Rabosky & Goldberg, 2015). Because HiSSE is restricted to

testing the influence of binary characters on clade diversification, species associated with

shifts in song evolution were coded with a '1' and all other species were coded as '0' (Tables

S5-S10). This was done to test whether shifts in song evolution are associated with

differential diversification rates. Speciation and extinction models can be subject to bias

when species are missing (Mynard et al., 2023), therefore we implemented HiSSE models

across the full honeyeater phylogeny containing 192 species of honeyeater (Hay et al., 2022).

The 29 honeyeater species with no song data were treated as having unknown tip states and

were coded with '?' rather than '0' or '1'. The MCMC sampler was set for 4 runs of 3,000

generations and a burn-in of 10%. Tracer v1.7.1 (Rambaut et al., 2018) was used to visualize

output and determine convergence. The R package 'RevGadgets' (Tribble et al., 2021) was

used to visualize model output with the '*plotTrace*' function, and summary statistics were

calculated using *'summarizeTrace'* (Table S11).

RESULTS

Phylogenetic signal varied substantially among the five song variables. Values of Pagel's

lambda ranged from 0.31 to 0.92 and the significance values of the likelihood ratio tests were

all below $p < 0.001$ (Table S2). Song bandwidth had the lowest signal ($\lambda = 0.31$), followed by

maximum frequency ($\lambda = 0.58$), pace ($\lambda = 0.66$), peak frequency ($\lambda = 0.75$) and then

minimum frequency ($\lambda = 0.92$).

Univariate relaxed OU models revealed several patterns in song evolution. Peak

frequency and maximum frequency had very similar estimates for relaxed OU models of trait

evolution. Here, the clades *Myzomela + Sugomel* (including *Lichmera lombokia*) and

Philemon were associated with shifts in song evolution for peak and maximum frequency

(Fig. 2a and b; Tables S5-S6). Minimum frequency also contained similar estimates to peak

and maximum frequency but did not uncover a shift for *Philemon* (Fig. 2c; Table S7). Despite

this, we found little evidence that shifts in trait evolution influenced honeyeater

diversification rates for peak frequency (mean diversification$_{no shift}$ = 2.38, 95 % CI = [1.80,

3.15]; mean diversification$_{shift}$ = 3.42, 95 % CI = [2.29, 4.88]; Fig. 2a; Table S11), maximum

frequency (mean diversification$_{no shift}$ = 2.41, 95 % CI = [1.82, 3.20]; mean diversification$_{shift}$

= 3.39, 95 % CI = [2.28, 4.87]; Fig. 2b; Table S11), and minimum frequency (mean

diversification$_{no shift}$ = 2.47, 95 % CI = [1.87, 3.23]; mean diversification$_{shift}$ = 3.93, 95 % CI =

[2.34, 6.11]; Fig. 2c; Table S11). There was also no evidence that shifts in song bandwidth

evolution influenced rates of diversification (mean diversification$_{no shift}$ = 2.67, 95 % CI =

[2.04, 3.49]; mean diversification$_{shift}$ = 2.64, 95 % CI = [0.72, 4.82]; Fig. 2d; Table S11).

Interestingly, we found shifts in pace evolution to be associated with lower rates of

diversification (mean diversification$_{no shift}$ = 2.78, 95 % CI = [2.11, 3.75]; mean

diversification$_{shift}$ = -0.72, 95 % CI = [-6.91, 0.36; Fig. 2e; Table S11) and this was

statistically significant, however, this result could be due to the lower sample size for species

associated shifts in pace evolution (7 species).

The two methods of multivariate OU trait evolution (RevBayes and PhylogeneticEM)

uncovered relatively similar shifts in song evolution for honeyeaters. Species from the genera

Philemon, Myzomela, and *Meliarchus* were associated with shifts in OU trait evolution for

both approaches (Fig. 3a; Table S10; Fig. S7). The PhylogeneticEM method did reveal a shift

associated for the larger clade containing *Ptilotula*, *Anthochaera*, *Melidectes*, and relatives,

which was not recovered in the RevBayes relaxed multivariate OU model (Fig. 3a; Table

S10; Fig. S7). Both methods had similar estimates for diversification and speciation rates

(Fig. 3b and c; Table S11). Nevertheless, we found no evidence that shifts in song evolution

were associated with increased rates of diversification for both the RevBayes (mean

diversification$_{no\ shift}$ = 2.40, 95 % CI = [1.80, 3.24]; mean diversification$_{shift}$ = 3.49, 95 % CI

= [2.37, 5.04) and PhylogeneticEM approaches (mean diversification$_{no\ shift}$ = 2.18, 95 % CI =

[1.56, 3.05]; mean diversification$_{shift}$ = 3.31, 95 % CI = [2.38, 4.77]), and the 95 %

confidence intervals overlap substantially (Fig. 3b and c; Table S11).

DISCUSSION

In this study, we used honeyeaters to explore the macroevolutionary consequences of song

evolution, a sexually selected behavioral trait, which has commonly been hypothesized to

influence speciation in birds. We found no evidence that shifts in song evolution were

associated with increases in diversification rates. The significant overlap between confidence

intervals of states is present in all tests, except for song pace, and hidden state-dependent

speciation and extinction models suggest that there are other factors (hidden states) which

play a role. Ultimately, shifts in song evolution do not appear to contribute to diversification

in honeyeaters, possibly due to other factors including geographical processes and dispersal

to islands dominating speciation history in honeyeaters (Hay et al., 2022), and any apparent

influence of song is likely mediated by these factors and underlying variation in body size.

Most of the interspecific diversity of song frequency and pace in honeyeaters can be

explained by evolutionary history. Theory suggests that divergence is expected for sexually

selected traits between closely related species (Maan & Seehausen, 2011; Servedio &

Boughman, 2017). Thus, the level of phylogenetic signal uncovered for frequency and pace is

contrary to such theoretical expectations. This is unsurprising, however, given other studies

have found similar levels of phylogenetic signal for song traits in birds: peak frequency was

found to have a lambda of 0.87 for 5085 species of passerine birds (Mikula et al., 2021), and

a lambda of 0.74 for 1022 passerine species (Friis et al., 2022). Our results are consistent with these previous studies and confirm that peak frequency of song is phylogenetically conserved. This is likely driven by allometric constraints imposed by body size (Fletcher, 2004), which itself is a conserved trait and directly impacts the length of the vocal tract and the size of the syrinx (Bertelli & Tubaro, 2002; Suthers & Zollinger, 2008). Certainly, body size has been demonstrated to have a prominent influence on honeyeater song and is understood to structure variation in peak frequency and pace (Hay et al., 2024). Interestingly, minimum frequency was found to have the highest phylogenetic signal rather than peak frequency. It is unclear if this is the case in other clades as past studies have typically focused on peak frequency. Nonetheless, body size is understood to place a limit on the lowest frequency that a bird can effectively produce (Fletcher, 2004; Friis et al., 2021), so perhaps minimum frequency is particularly constrained by body size.

We found no evidence that shifts in song evolution were associated with higher rates of diversification in honeyeaters. Macroevolutionary influences of song evolution on speciation has been explored in 581 species of tanagers and ovenbirds and shifts in song were found to be correlated with shifts in speciation, but this was not significant when each family was considered separately (Mason et al., 2017). This is consistent with the findings of this study, which is restricted to the family-level, and could imply that macroevolutionary signals of song on diversification emerge only when very different clades of birds are examined simultaneously, as is the case for other taxa too (Rabosky & Huang, 2016; Rabosky et al., 2013). Additionally, the song dataset used in this study was missing data for 29 honeyeater species. As is often the case with missing data, species inhabiting the most remote areas were data deficient. This skewed our dataset towards common species and especially those found in Australia and New Zealand. Areas such as New Guinea, which has high honeyeater species richness (Marki et al., 2019), and islands throughout the Indo-Pacific, which contain endemic

species, were lacking in data. Given previous work highlighting island settings for speciation

in the group (Hay et al., 2022), this potential effect of our dataset should be considered a

caveat for the study.

We did find some evidence that shifts in song pace evolution influenced rates of

diversification and shifts in song pace were correlated with lower rates of diversification.

However, only seven species of honeyeater were associated with shifts in pace evolution, and

this low sample size has likely influenced this outcome. Given that the number of species

associated with shifts (7 species) is far lower than the number of missing species included in

the model (29 species), we recommend caution when interpreting this result.

Nonetheless, our results indicate that shifts in song evolution are not major drivers of

diversification in honeyeaters. Recent studies have found that faster rates of song evolution

do not result in higher speciation rates in New World passerine birds (Freeman et al., 2022),

and ecological and demographic factors may provide a better explanation for speciation

(Freeman et al., 2022; Harvey et al., 2019; Singhal et al., 2018). State-dependent speciation

and extinction models used in the present study indicated hidden states are associated with

differential rates diversification. Thus, other factors not considered here are likely

contributing to patterns of diversification. Honeyeaters have a dynamic evolutionary history:

Australia is the center of honeyeater diversity (Driskell & Christidis, 2004; Norman et al.,

2007; Oliveros et al., 2019), however, dispersal to islands throughout the Indo-Pacific, and

vicariant speciation due to connection and disconnection of New Guinea and the changing

biomes of Australia have played key roles in the diversification of this group (Andersen et al.,

2014; Hay et al., 2022; Marki et al., 2017; Toon et al., 2010). Moreover, past studies have

found range size and dispersal ability to significantly contribute to speciation in honeyeaters

when applying state-dependent speciation and extinction models (Hay et al., 2022).

Allopatric speciation is the predominant mode of speciation in honeyeaters and could explain

the lack of influence of shifts in song evolution on diversification because selection on signal

divergence is expected to be stronger for sympatry (Pfennig & Pfennig, 2010). Songs are

thought to diverge slowly in allopatry, and only rapidly increase on secondary contact,

contributing to reinforcement (Lachlan & Servedio, 2004; Servedio & Boughman, 2017).

Thus, contributions of song to speciation likely depends on the mode of speciation, so

although song could be aiding microevolutionary processes such as reproductive isolation,

other factors including range size and dispersal ability have contributed more to

diversification (Hay et al., 2022).

Across all models of trait evolution, shifts in song evolution are notable in clades of

Myzomela + Sugomel (including *Lichmera lombokia*) and *Philemon*. These clades have

previously been identified to have increased rates of speciation (Hay et al., 2022) and are also

the most speciose genera in honeyeaters (Gill et al., 2020). However, past studies have shown

that small range sizes resulting from dispersal to islands is a primary driver of speciation in

these clades (Hay et al., 2022), and the (non-significant) slight increase in diversification

rates observed with these shifts in song evolution could be due to these underlying factors. It

is also important to consider the influence of body size on these patterns because honeyeater

song is under strong allometric constraints (Hay et al., 2024), and these clades show great

differences in body size: all species of *Myzomela* weigh between 6 to 16 g, and species of

Philemon range from 50 g to 194 g, in which the majority of species are over 100 g

(Dunning, 2008; Tobias et al., 2022). Body size evolution has been correlated with speciation

and species diversity across different vertebrate clades (Rabosky & Adams, 2012; Rabosky et

al., 2013; Ricklefs, 2004), and has contributed to speciation in honeyeaters (Brennan &

Keogh, 2018; Marki et al., 2019). Therefore, it is possible that the shifts in song evolution

could be due to shifts in body size evolution, and underlying variation in body size could be

contributing to both the observed patterns of song evolution and diversification in this study.

Understanding the influence of reproductive isolation on diversification has been

recognized as an urgent question in evolutionary biology (Matute & Cooper, 2021),

especially because the evolution of reproductive isolation often defines speciation (Coyne &

Orr, 2004). Here, we contribute to understanding of this question by testing the hypothesis

that shifts in song evolution have influenced diversification in honeyeaters. We uncovered

several shifts in song evolution but found no evidence that these shifts were associated with

significant increases in diversification rates. Models also indicated that other factors (hidden

states) contribute to differential rates of diversification, implying that diversification of

honeyeaters is better explained by other factors.

Acknowledgements

We thank Leo Joseph and Margaret Cawsey at ANWC for assistance with accessing

recordings and acknowledge receipt of media from The Macaulay Library at the Cornell Lab

of Ornithology. EMH was supported by an Australian Government Research Training

Program (RTP) Scholarship, a Monash Graduate Excellence Scholarship (MGES), and a

Faculty of Science Dean's Postgraduate Research Scholarship.

REFERENCES

Andersen, M. J., Naikatini, A., & Moyle, R. G. (2014). A molecular phylogeny of Pacific honeyeaters (Aves: Meliphagidae) reveals extensive paraphyly and an isolated Polynesian radiation. *Molecular Phylogenetics and Evolution, 71*, 308-315. https://doi.org/10.1016/j.ympev.2013.11.014

Arnegard, M. E., McIntyre, P. B., Harmon, L. J., Zelditch, M. L., Crampton, W. G., Davis, J. K., Sullivan, J. P., Lavoue, S., & Hopkins, C. D. (2010). Sexual signal evolution outpaces ecological divergence during electric fish species radiation. *The American Naturalist, 176*(3), 335-356. https://doi.org/10.1086/655221

Baptista, L. F., & Trail, P. W. (1992). The role of song in the evolution of passerine diversity. *Systematic Biology, 41*(2), 242-247. https://doi.org/10.2307/2992524

Bastide, P., Ané, C., Robin, S., & Mariadassou, M. (2018). Inference of adaptive shifts for multivariate correlated traits. *Systematic Biology, 67*(4), 662-680. https://doi.org/10.1093/sysbio/syy005

Beaulieu, J. M., & O'Meara, B. C. (2016). Detecting hidden diversification shifts in models of trait-dependent speciation and extinction. *Systematic Biology, 65*(4), 583-601. https://doi.org/10.1093/sysbio/syw022

Bertelli, S., & Tubaro, P. L. (2002). Body mass and habitat correlates of song structure in a primitive group of birds. *Biological Journal of the Linnean Society, 77*, 423-430. https://doi.org/10.1046/j.1095-8312.2002.00112.x

Boul, K. E., Funk, W. C., Darst, C. R., Cannatella, D. C., & Ryan, M. J. (2007). Sexual selection drives speciation in an Amazonian frog. *Proceedings of the Royal Society B: Biological Sciences, 274*(1608), 399-406. https://doi.org/10.1098/rspb.2006.3736

Brennan, I. G., & Keogh, J. S. (2018). Miocene biome turnover drove conservative body size

evolution across Australian vertebrates. *Proceedings of the Royal Society B:*

Biological Sciences, 285, 20181474. https://doi.org/10.1098/rspb.2018.1474

Cardoso, G. C. (2013). Using frequency ratios to study vocal communication. *Animal*

Behaviour, 85(6), 1529-1532. https://doi.org/10.1016/j.anbehav.2013.03.044

Catchpole, C. K., & Slater, P. J. B. (2003). *Bird Song: Biological Themes and Variations*.

Cambridge University Press.

Cooney, C. R., & Thomas, G. H. (2021). Heterogeneous relationships between rates of

speciation and body size evolution across vertebrate clades. *Nature Ecology and*

Evolution, 5(1), 101-110. https://doi.org/10.1038/s41559-020-01321-y

Coyne, J. A., & Orr, H. A. (2004). *Speciation*. Sinauer Associates.

Derryberry, E. P., Seddon, N., Derryberry, G. E., Claramunt, S., Seeholzer, G. F., Brumfield,

R. T., & Tobias, J. A. (2018). Ecological drivers of song evolution in birds:

disentangling the effects of habitat and morphology. *Ecology and Evolution, 8*(3),

1890-1905. https://doi.org/10.1002/ece3.3760

Driskell, A. C., & Christidis, L. (2004). Phylogeny and evolution of the Australo-Papuan

honeyeaters (Passeriformes, Meliphagidae). *Molecular Phylogenetics and Evolution,*

31(3), 943-960. https://doi.org/10.1016/j.ympev.2003.10.017

Dumont, E. R., Davalos, L. M., Goldberg, A., Santana, S. E., Rex, K., & Voigt, C. C. (2012).

Morphological innovation, diversification and invasion of a new adaptive zone.

Proceedings of the Royal Society B: Biological Sciences, 279(1734), 1797-1805.

https://doi.org/10.1098/rspb.2011.2005

Dunning, J. B. (2008). *CRC Handbook of Avian Body Masses*. Taylor & Francis Group, LLC.

Fitzpatrick, J. W. (1988). Why so many passerine birds? A response to Raikow. *Systematic*

Zoology, 37(1), 71-76. https://doi.org/10.2307/2413195

Fletcher, N. H. (2004). A simple frequency-scaling rule for animal communication. *The*

Journal of the Acoustical Society of America, 115(5 Pt 1), 2334-2338.

https://doi.org/10.1121/1.1694997

Freeman, B. G., Montgomery, G. A., & Schluter, D. (2017). Evolution and plasticity:

divergence of song discrimination is faster in birds with innate song than in song

learners in Neotropical passerine birds. *Evolution, 71*(9), 2230-2242.

https://doi.org/10.1111/evo.13311

Freeman, B. G., Rolland, J., Montgomery, G. A., & Schluter, D. (2022). Faster evolution of a

premating reproductive barrier is not associated with faster speciation rates in New

World passerine birds. *Proceedings of the Royal Society B: Biological Sciences,*

289(1966), 20211514. https://doi.org/10.1098/rspb.2021.1514

Friedman, N. R., Miller, E. T., Ball, J. R., Kasuga, H., Remes, V., & Economo, E. P. (2019).

Evolution of a multifunctional trait: shared effects of foraging ecology and

thermoregulation on beak morphology, with consequences for song evolution.

Proceedings of the Royal Society B: Biological Sciences, 286(1917), 20192474.

https://doi.org/10.1098/rspb.2019.2474

Friis, J. I., Sabino, J., Santos, P., Dabelsteen, T., & Cardoso, G. C. (2021). The allometry of

sound frequency bandwidth in songbirds. *The American Naturalist, 197*(5), 607-614.

https://doi.org/10.1086/713708

Friis, J. I., Sabino, J., Santos, P., Dabelsteen, T., Cardoso, G. C., & Jennions, M. D. (2022).

Ecological adaptation and birdsong: how body and bill sizes affect passerine sound

frequencies. *Behavioral Ecology, 33*(4), 798-806.

https://doi.org/10.1093/beheco/arac042

Gill, F., Donsker, D., & Rasmussen, P. (2020). *IOC World Bird List* Version 10.2).

https://doi.org/10.14344/IOC.ML.10.2.

Grant, P. R., & Grant, B. R. (2006). Evolution of character displacement in Darwin's finches.

Science, 313, 224-226. https://doi.org/10.1126/science.1128374

Gray, D. A., & Cade, W. H. (2000). Sexual selection and speciation in field crickets.

Proceedings of the National Academy of Sciences of the USA, 97, 14449-14454.

https://doi.org/10.1073/pnas.97.26.14449

Harvey, M. G., Singhal, S., & Rabosky, D. L. (2019). Beyond reproductive isolation:

demographic controls on the speciation process. *Annual Review of Ecology,*

Evolution, and Systematics, 50(1), 75-95. https://doi.org/10.1146/annurev-ecolsys-

110218-024701

Hay, E. M., McGee, M. D., & Chown, S. L. (2022). Geographic range size and speciation in

honeyeaters. *BMC Ecology and Evolution, 22*, 1-14. https://doi.org/10.1186/s12862-

022-02041-6

Hay, E. M., McGee, M. D., White, C. R., & Chown, S. L. (2024). Body size shapes song in

honeyeaters. *Proceedings of the Royal Society B: Biological Sciences*. https://doi.org/

10.1098/rspb.2024.0339

Hernández-Hernández, T., Miller, E. C., Roman-Palacios, C., & Wiens, J. J. (2021).

Speciation across the Tree of Life. *Biological Reviews, 96*(4), 1205-1242.

https://doi.org/10.1111/brv.12698

Higgins, P. J., Ford, H. A., & Christidis, L. (2008). Family Meliphagidae. In J. del Hoyo, A.

Elliott, & D. A. Christie (Eds.), *Handbook of the Birds of the World: Penduline-tits to*

shrikes (Vol. 13, pp. 498–691). Lynx Edicions.

Höhna, S., Heath, T. A., Boussau, B., Landis, M. J., Ronquist, F., & Huelsenbeck, J. P.

(2014). Probabilistic graphical model representation in phylogenetics. *Systematic*

Biology, 63(5), 753-771. https://doi.org/10.1093/sysbio/syu039

Höhna, S., Landis, M. J., Heath, T. A., Boussau, B., Lartillot, N., Moore, B. R., Huelsenbeck,

J. P., & Ronquist, F. (2016). RevBayes: Bayesian phylogenetic inference using

graphical models and an interactive model-specification language. *Systematic Biology,*

65(4), 726-736. https://doi.org/10.1093/sysbio/syw021

Irwin, D. E., Bensch, S., & Price, T. D. (2001). Speciation in a ring. *Nature, 409*, 333-337.

https://doi.org/10.1038/35053059

Jarvis, E. D. (2021). At the beginning of speciation. *Science, 371*(6536), 1312.

https://doi.org/10.1126/science.abg5454

Lachlan, R. F., & Servedio, M. R. (2004). Song learning accelerates allopatric speciation.

Evolution, 58(9), 2049-2063. https://doi.org/10.1111/j.0014-3820.2004.tb00489.x

Laidre, M. E., & Johnstone, R. A. (2013). Animal signals. *Current Biology, 23*(18), R829-

833. https://doi.org/10.1016/j.cub.2013.07.070

Maan, M. E., & Seehausen, O. (2011). Ecology, sexual selection and speciation. *Ecology*

Letters, 14(6), 591-602. https://doi.org/10.1111/j.1461-0248.2011.01606.x

Maddison, W. P., & FitzJohn, R. G. (2015). The unsolved challenge to phylogenetic

correlation tests for categorical characters. *Systematic Biology, 64*(1), 127-136.

https://doi.org/10.1093/sysbio/syu070

Mahler, D. L., Revell, L. J., Glor, R. E., & Losos, J. B. (2010). Ecological opportunity and

the rate of morphological evolution in the diversification of Greater Antillean anoles.

Evolution, 64(9), 2731-2745. https://doi.org/10.1111/j.1558-5646.2010.01026.x

Marki, P. Z., Jonsson, K. A., Irestedt, M., Nguyen, J. M. T., Rahbek, C., & Fjeldsa, J. (2017).

Supermatrix phylogeny and biogeography of the Australasian Meliphagides radiation

(Aves: Passeriformes). *Molecular Phylogenetics and Evolution, 107*, 516-529.

https://doi.org/10.1016/j.ympev.2016.12.021

Marki, P. Z., Kennedy, J. D., Cooney, C. R., Rahbek, C., & Fjeldsa, J. (2019). Adaptive

radiation and the evolution of nectarivory in a large songbird clade. *Evolution, 73*,

1226-1240. https://doi.org/10.1111/evo.13734

Marquet, P. A., Fernández, M., Navarette, A. A., & Valdovinos, C. (2004). Diversity

emerging: towards a deconstruction of biodiversity patterns. In L. M. & H. L. (Eds.),

Frontiers of biogeography: New directions in the geography of nature (pp. 191-209).

Sinauer Associates.

Marshall, D. C., Slon, K., Cooley, J. R., Hill, K. B. R., & Simon, C. (2008). Steady Plio-

Pleistocene diversification and a 2-million-year sympatry threshold in a New Zealand

cicada radiation. *Molecular Phylogenetics and Evolution, 48*(3), 1054-1066.

https://doi.org/10.1016/j.ympev.2008.05.007

Mason, N. A., & Burns, K. J. (2015). The effect of habitat and body size on the evolution of

vocal displays in Thraupidae (tanagers), the largest family of songbirds. *Biological*

Journal of the Linnean Society, 114, 538-551. https://doi.org/10.1111/bij.12455

Mason, N. A., Burns, K. J., Tobias, J. A., Claramunt, S., Seddon, N., & Derryberry, E. P.

(2017). Song evolution, speciation, and vocal learning in passerine birds. *Evolution,*

71(3), 786-796. https://doi.org/10.1111/evo.13159

Matute, D. R., & Cooper, B. S. (2021). Comparative studies on speciation: 30 years since

Coyne and Orr. *Evolution, 75*(4), 764-778. https://doi.org/10.1111/evo.14181

Mikula, P., Valcu, M., Brumm, H., Bulla, M., Forstmeier, W., Petruskova, T., Kempenaers, B.,

& Albrecht, T. (2021). A global analysis of song frequency in passerines provides no

support for the acoustic adaptation hypothesis but suggests a role for sexual selection.

Ecology Letters, 24(3), 477-486. https://doi.org/10.1111/ele.13662

Miller, M. A., Pfeiffer, W., & Schwartz, T. (2010). Creating the CIPRES Science Gateway for

inference of large phylogenetic trees. 2010 Gateway Computing Environments

workshop (GCE), New Orleans, LA.

Mynard, P., Algar, A., Lancaster, L., Bocedi, G., Fahri, F., Gubry-Rangin, C.,

Lupiyaningdyah, P., Nangoy, M., Osborne, O., Papadopulos, A., Sudiana, I. M.,

Juliandi, B., Travis, J., & Herrera-Alsina, L. (2023). Impact of phylogenetic tree

completeness and misspecification of sampling fractions on trait dependent

diversification models. *Systematic Biology*. https://doi.org/10.1093/sysbio/syad001

Norman, J. A., Rheindt, F. E., Rowe, D. L., & Christidis, L. (2007). Speciation dynamics in

the Australo-Papuan *Meliphaga* honeyeaters. *Molecular Phylogenetics and Evolution,*

42(1), 80-91. https://doi.org/10.1016/j.ympev.2006.05.032

Oliveros, C. H., Field, D. J., Ksepka, D. T., Barker, F. K., Aleixo, A., Andersen, M. J.,

Alstrom, P., Benz, B. W., Braun, E. L., Braun, M. J., Bravo, G. A., Brumfield, R. T.,

Chesser, R. T., Claramunt, S., Cracraft, J., Cuervo, A. M., Derryberry, E. P., Glenn, T.

C., Harvey, M. G., Hosner, P. A., Joseph, L., Kimball, R. T., Mack, A. L., Miskelly, C.

M., Peterson, A. T., Robbins, M. B., Sheldon, F. H., Silveira, L. F., Smith, B. T.,

White, N. D., Moyle, R. G., & Faircloth, B. C. (2019). Earth history and the passerine

superradiation. *Proceedings of the National Academy of Sciences of the USA, 116*(16),

7916-7925. https://doi.org/10.1073/pnas.1813206116

Pagel, M. (1999). Inferring the historical patterns of biological evolution. *Nature, 401*(6756),

877-884.

Paradis, E., Claude, J., & Strimmer, K. (2004). APE: analyses of phylogenetics and evolution

in R language. *Bioinformatics, 20*(2), 289-290.

https://doi.org/10.1093/bioinformatics/btg412

Pearse, W. D., Morales-Castilla, I., James, L. S., Farrell, M., Boivin, F., & Davies, T. J.

(2018). Global macroevolution and macroecology of passerine song. *Evolution, 72*(4),

944-960. https://doi.org/10.1111/evo.13450

Pfennig, D. W., & Pfennig, K. S. (2010). Character displacement and the origins of diversity.

The American Naturalist, 176, 26-44. https://doi.org/10.1086/657056

Pfennig, D. W., Wund, M. A., Snell-Rood, E. C., Cruickshank, T., Schlichting, C. D., &

Moczek, A. P. (2010). Phenotypic plasticity's impacts on diversification and

speciation. *Trends in Ecology & Evolution, 25*(8), 459-467.

https://doi.org/10.1016/j.tree.2010.05.006

Podos, J. (2001). Correlated evolution of morphology and vocal signal structure in Darwin's

fnches. *Nature, 409*, 185-188. https://doi.org/10.1038/35051570

Podos, J., & Nowicki, S. (2004). Beaks, adaptation, and vocal evolution in Darwin's finches.

Bioscience, 54, 501-510. https://doi.org/10.1641/0006-

3568(2004)054[0501:BAAVEI]2.0.CO;2

Price, T. (2008). *Speciation in Birds*. Roberts and Co.

R Core Team. (2013). R: a language and environment for statistical computing.

https://www.r-project.org/

Rabosky, D. L., & Adams, D. C. (2012). Rates of morphological evolution are correlated with

species richness in salamanders. *Evolution, 66*(6), 1807-1818.

https://doi.org/10.1111/j.1558-5646.2011.01557.x

Rabosky, D. L., & Goldberg, E. E. (2015). Model inadequacy and mistaken inferences of

trait-dependent speciation. *Systematic Biology, 64*(2), 340-355.

https://doi.org/10.1093/sysbio/syu131

Rabosky, D. L., & Huang, H. (2016). A robust semi-parametric test for detecting trait-

dependent diversification. *Systematic Biology, 65*(2), 181-193.

https://doi.org/10.1093/sysbio/syv066

Rabosky, D. L., Santini, F., Eastman, J., Smith, S. A., Sidlauskas, B., Chang, J., & Alfaro, M.

E. (2013). Rates of speciation and morphological evolution are correlated across the

largest vertebrate radiation. *Nature Communications, 4*, 1958.

https://doi.org/10.1038/ncomms2958

Rambaut, A., Drummond, A. J., Xie, D., Baele, G., & Suchard, M. A. (2018). Posterior

summarization in Bayesian phylogenetics using Tracer 1.7. *Systematic Biology, 67*(5),

901-904. https://doi.org/10.1093/sysbio/syy032

Revell, L. J. (2012). phytools: an R package for phylogenetic comparative biology (and other

things). *Methods in Ecology and Evolution, 3*(2), 217-223.

https://doi.org/10.1111/j.2041-210X.2011.00169.x

Revell, L. J., & Harmon, L. J. (2022). *Phylogenetic comparative methods in R*. Princeton

University Press.

Revell, L. J., Harmon, L. J., & Collar, D. C. (2008). Phylogenetic signal, evolutionary

process, and rate. *Systematic Biology, 57*(4), 591-601.

https://doi.org/10.1080/10635150802302427

Ricklefs, R. E. (2004). Cladogenesis and morphological diversification in passerine birds.

Nature, 430, 338-341. https://doi.org/10.1038/nature02755

Ritchie, M. G. (2007). Sexual selection and speciation. *Annual Review of Ecology, Evolution,*

and Systematics, 38, 79-102.

https://doi.org/10.1146/annurev.ecolsys.38.091206.095733

Seddon, N., & Tobias, J. A. (2007). Song divergence at the edge of Amazonia: an empirical

634   test of the peripatric speciation model. *Biological Journal of the Linnean Society, 90*,

635   173-188. https://doi.org/10.1111/j.1095-8312.2007.00753.x

636 Servedio, M. R., & Boughman, J. W. (2017). The role of sexual selection in local adaptation

637   and speciation. *Annual Review of Ecology, Evolution, and Systematics, 48*, 85-109.

638   https://doi.org/10.1146/annurev-ecolsys-110316-022905

639 Simões, T. R., Vernygora, O., Caldwell, M. W., & Pierce, S. E. (2020). Megaevolutionary

640   dynamics and the timing of evolutionary innovation in reptiles. *Nature*

641   *Communications, 11*(1), 3322. https://doi.org/10.1038/s41467-020-17190-9

642 Singhal, S., Huang, H., Grundler, M. R., Marchan-Rivadeneira, M. R., Holmes, I., Title, P. O.,

643   Donnellan, S. C., & Rabosky, D. L. (2018). Does population structure predict the rate

644   of speciation? a comparative test across australia's most diverse vertebrate radiation.

645   *The American Naturalist, 192*(4), 432-447. https://doi.org/10.1086/699515

646 Suthers, R. A., & Zollinger, S. A. (2008). From brain to song: the vocal organ and vocal tract.

647   In P. H. Zeigler & P. Marler (Eds.), *Neuroscience of Birdsong* (pp. 78-98). Cambridge

648   University Press.

649 Tinghitella, R. M., Broder, E. D., Gurule-Small, G. A., Hallagan, C. J., & Wilson, J. D.

650   (2018). Purring crickets: the evolution of a novel sexual signal. *The American*

651   *Naturalist, 192*(6), 773-782. https://doi.org/10.1086/700116

652 Tobias, J. A., Ottenburghs, J., & Pigot, A. L. (2020). Avian diversity: speciation,

653   macroevolution, and ecological function. *Annual Review of Ecology, Evolution, and*

654   *Systematics, 51*(1), 533-560. https://doi.org/10.1146/annurev-ecolsys-110218-025023

655 Tobias, J. A., Sheard, C., Pigot, A. L., Devenish, A. J. M., Yang, J., Sayol, F., Neate-Clegg,

656   M. H. C., Alioravainen, N., Weeks, T. L., Barber, R. A., Walkden, P. A., MacGregor,

657   H. E. A., Jones, S. E. I., Vincent, C., Phillips, A. G., Marples, N. M., Montano-

Centellas, F. A., Leandro-Silva, V., Claramunt, S., Darski, B., Freeman, B. G.,

Bregman, T. P., Cooney, C. R., Hughes, E. C., Capp, E. J. R., Varley, Z. K., Friedman,

N. R., Korntheuer, H., Corrales-Vargas, A., Trisos, C. H., Weeks, B. C., Hanz, D. M.,

Topfer, T., Bravo, G. A., Remes, V., Nowak, L., Carneiro, L. S., Moncada, R. A.,

Matysiokova, B., Baldassarre, D. T., Martinez-Salinas, A., Wolfe, J. D., Chapman, P.

M., Daly, B. G., Sorensen, M. C., Neu, A., Ford, M. A., Mayhew, R. J., Fabio Silveira,

L., Kelly, D. J., Annorbah, N. N. D., Pollock, H. S., Grabowska-Zhang, A. M.,

McEntee, J. P., Carlos, T. G. J., Meneses, C. G., Munoz, M. C., Powell, L. L., Jamie,

G. A., Matthews, T. J., Johnson, O., Brito, G. R. R., Zyskowski, K., Crates, R.,

Harvey, M. G., Jurado Zevallos, M., Hosner, P. A., Bradfer-Lawrence, T., Maley, J.

M., Stiles, F. G., Lima, H. S., Provost, K. L., Chibesa, M., Mashao, M., Howard, J. T.,

Mlamba, E., Chua, M. A. H., Li, B., Gomez, M. I., Garcia, N. C., Packert, M., Fuchs,

J., Ali, J. R., Derryberry, E. P., Carlson, M. L., Urriza, R. C., Brzeski, K. E.,

Prawiradilaga, D. M., Rayner, M. J., Miller, E. T., Bowie, R. C. K., Lafontaine, R. M.,

Scofield, R. P., Lou, Y., Somarathna, L., Lepage, D., Illif, M., Neuschulz, E. L.,

Templin, M., Dehling, D. M., Cooper, J. C., Pauwels, O. S. G., Analuddin, K.,

Fjeldsa, J., Seddon, N., Sweet, P. R., DeClerck, F. A. J., Naka, L. N., Brawn, J. D.,

Aleixo, A., Bohning-Gaese, K., Rahbek, C., Fritz, S. A., Thomas, G. H., &

Schleuning, M. (2022). AVONET: morphological, ecological and geographical data

for all birds. *Ecology Letters, 25*(3), 581-597. https://doi.org/10.1111/ele.13898

Toon, A., Hughes, J. M., & Joseph, L. (2010). Multilocus analysis of honeyeaters (Aves:

Meliphagidae) highlights spatio-temporal heterogeneity in the influence of

biogeographic barriers in the Australian monsoonal zone. *Molecular Ecology, 19*(14),

2980-2994. https://doi.org/10.1111/j.1365-294X.2010.04730.x

Tribble, C. M., Freyman, W. A., Landis, M. J., Lim, J. Y., Barido-Sottani, J., Kopperud, B. T.,

Höhna, S., & May, M. R. (2021). RevGadgets: an R package for visualizing Bayesian

phylogenetic analyses from RevBayes. *Methods in Ecology and Evolution, 13*(2),

314-323. https://doi.org/10.1111/2041-210x.13750

Uyeda, J. C., & Harmon, L. J. (2014). A novel Bayesian method for inferring and interpreting

the dynamics of adaptive landscapes from phylogenetic comparative data. *Systematic*

Biology, 63(6), 902-918. https://doi.org/10.1093/sysbio/syu057

Wagner, C. E., Harmon, L. J., & Seehausen, O. (2012). Ecological opportunity and sexual

selection together predict adaptive radiation. *Nature, 487*(7407), 366-369.

https://doi.org/10.1038/nature11144

Walsh, B., & Blows, M. W. (2009). Abundant genetic variation + strong selection =

multivariate genetic constraints: a geometric view of adaptation. *Annual Review of*

Ecology, Evolution, and Systematics, 40(1), 41-59.

https://doi.org/10.1146/annurev.ecolsys.110308.120232

Wilkins, M. R., Seddon, N., & Safran, R. J. (2013). Evolutionary divergence in acoustic

signals: causes and consequences. *Trends in Ecology & Evolution, 28*(3), 156-166.

https://doi.org/10.1016/j.tree.2012.10.002

Zuk, M., Rotenberry, J. T., & Tinghitella, R. M. (2006). Silent night: adaptive disappearance

of a sexual signal in a parasitized population of field crickets. *Biology Letters, 2*(4),

521-524. https://doi.org/10.1098/rsbl.2006.0539

 FIGURE CAPTIONS

Figure 1. Song spectrograms displaying frequency (kHz) over time (s) for five species of

honeyeater: a) New Zealand Bellbird (*Anthornis melanura*; xeno-canto accession number =

XC153005), b) Eastern Spinebill (*Acanthorhynchus tenuirostris;* xeno-canto accession

number = XC490800), c) Blue-faced Honeyeater (*Entomyzon cyanotis*; xeno-canto accession

number = XC439288), d) Noisy Friarbird (*Philemon corniculatus*; xeno-canto accession

number = XC287056), and e) Singing Honeyeater (*Gavicalis virescens*; xeno-canto accession

number = XC334251). The five song variables considered in this study are indicated with

annotations in a): peak frequency represents the frequency of maximum amplitude across the

entire song, maximum frequency is the highest pitched note in song, minimum frequency is

the lowest pitched note in song, bandwidth is the difference between maximum and minimum

frequency across the song, and pace is the number of notes over song duration. Birds are

illustrations by Eleanor Hay.

Figure 2. Results from relaxed OU and HiSSE models for each of the five song traits; peak

frequency (a), maximum frequency (b), minimum frequency (c), bandwidth (d) and pace (e),

Phylogenetic trees display branch specific theta values from relaxed OU models in

RevBayes. Dots at the tips of the trees indicate which species were identified as associated

with shifts in song evolution and were tested to influence speciation rates. Respective

diversification rates extracted from HiSSE models associated with the coding at the tips are

displayed in the density plots to the right of each phylogeny.

Figure 3. Honeyeater phylogenetic tree (a), the branches of the phylogeny display estimated

branch theta values from the multivariate relaxed OU model in RevBayes. Dots on the

branches of the phylogeny indicate shifts revealed by PhylogeneticEM. Dots at the tip of the

tree show the binary classifications for HiSSE models from the RevBayes and

PhylogeneticEM (labelled as PhyloEM) approaches. Density plots below the phylogeny show

the estimated diversification rates extracted from HiSSE models for shifts identified by

RevBayes (b) and PhylogeneticEM (c). Birds are illustrations by Eleanor Hay.

FIGURES

Figure 1.

Figure 2.

779   **Figure 3.**