Genomic Diversity Illuminates the Species History and Environmental

Adaptation of *Drosophila suzukii*

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Abstract

Biological invasions carry substantial practical and scientific importance, and represent natural evolutionary experiments on contemporary timescales. Here, we investigated genomic diversity, invasion history, and environmental adaptation of the crop pest *Drosophila suzukii* using whole-genome sequencing data and environmental metadata for 29 population samples from its native and invasive range. Our analysis of genome-wide variation revealed maximal diversity in a population from eastern China, with other populations largely containing a subset of the genetic diversity present in this sample, consistent with an ancestral or refugial species range encompassing this region of Asia. Our analyses of genomic diversity and population history recapitulated some previously suggested dynamics of genetic structure, invasion bottlenecks, and admixture events. However, we suggest that clearer inferences of the geographic origins of worldwide invasions will require denser geographic sampling of genetic variation, particularly in Asia. Using a genome-environment association approach, we detected genetic signals of local adaptation associated with nine distinct environmental factors related to altitude, wind speed, precipitation, temperature, and human land use. We detected some unique functional signatures for each environmental variable, such as a prevalence of cuticular genes associated with annual precipitation. We also inferred biological commonalities in the adaptation to diverse selective pressures, particularly in terms of the apparent contribution of nervous system evolution to enriched processes (ranging from neuron development to circadian behavior) and/or top genes associated with all nine environmental variables. Our findings provide insights into the invasion history of *D. suzukii* and depict a finer-scale adaptive landscape underlying this species' invasion success.

Introduction

One of the main goals of ecological and evolutionary genomics is to understand how organisms evolve in response to novel environments. Biological invasions, while often ecologically and economically damaging, represent unique opportunities to build our understanding of local adaptation, as natural experiments that expose introduced species to new biotic and abiotic factors on contemporary time scales (Lee 2002; Prentis et al. 2008; Colautti and Lau 2016). Invasive species can exhibit rapid phenotypic and genetic changes during the invasion process, driven by various evolutionary mechanisms such as selection, drift, mutation, and gene flow (Colautti and Lau 2016; Hodgins et al. 2018). These changes can result in the adaptive evolution of invasive populations to the novel environments they encounter (Colautti and Barrett 2013). Although there have been emerging studies on the evolutionary biology of invasive species in recent years, the source and nature of the genetic variation underlying such adaptation are still not well characterized (Reznick et al. 2019; Welles and Dlugosch 2019).

Drosophila suzukii, also known as spotted wing *Drosophila*, is a promising model for studying adaptive evolution during invasions. *Drosophila suzukii* is a highly polyphagous vinegar fly that originated from Asia. It first expanded to Hawaii in 1980, and has recently invaded North America and Europe, and subsequently Réunion Island (Indian Ocean) and South America since the late 2000s (Asplen et al. 2015). *Drosophila suzukii* differs from other *Drosophila* species in its unique ability to oviposit on both unripe and ripe fruits, using its serrated ovipositor to pierce the skin of soft-skinned fruits. This has allowed it to exploit a novel ecological niche and avoid competition with other vinegar flies that typically feed on overripe and rotting fruits (Cini et al. 2012; Atallah et al. 2014), causing severe economic losses to fruit crops (Knapp et al. 2021). It also exhibits remarkable phenotypic plasticity and genetic diversity, which may facilitate its

adaptation to different climatic conditions and host plants (Gibert et al. 2019; Little et al. 2020;

Olazcuaga et al. 2020). To obtain a comprehensive evolutionary genetic understanding of the

invasion history of *D. suzukii*, we need to understand both the demography of this species

(including the history and structure of populations from Asia and introduced regions) and the

genetic basis and ecological drivers of adaptive evolutionary changes that have allowed this

species to occupy diverse worldwide environments.

Multiple genetic studies have investigated the demographic history of invasive

populations of *D. suzukii*. Adrion et al. (2014) analyzed six X-linked gene fragments from

populations in the continental US, Hawaii, Japan, and Spain. They detected differentiation

signals between European, Asian, and US populations, but not among US populations, and

suggested independent invasions into Europe and the continental US. Using data from 25

microsatellite markers, Fraimout et al. (2017) further characterized invasion bottlenecks and

inferred more comprehensive worldwide invasion scenarios of *D. suzukii*, using an approximate

Bayesian computation random forest (ABC-RF) approach. The resulting model, which was

constrained by the observed chronology of the species' invasion, indicated that some invading

populations had multiple genetic sources. A more recent population genomic study that focused

on samples from the US, as well as single sites in Brazil, Ireland, Italy, Japan, and Korea, used

autosomal single nucleotide polymorphisms (SNPs) to infer population structure and admixture

(Lewald et al. 2021). Their model mostly agrees with the above microsatellite analysis, except for

an admixture event from the western US back to Asia. Collectively, these studies suggest some

emerging consensus about the invasion history of *D. suzukii*, along with some lingering

uncertainty about admixture during this process. However, the limited study of Asian *D. suzukii*

samples has left the history of the species within this continent less clear, along with the precise

geographic origins of invading populations.

In contrast, the genetic basis of environmental adaptation in *D. suzukii* is still largely

unexplored. Among the few relevant studies is that of Olazcuaga et al. (2020), which used

genomic sequencing data from 22 worldwide populations to search for SNPs with greater

frequency differences between Asian (China, Japan) and non-Asian populations than are

observed at most loci due to founder event bottlenecks, in hopes of identifying genetic variants

that may underlie the invasion success of introduced populations. A subsequent study examining

this same data set also found a small number of transposon insertions with strong frequency

differences between American/European and Asian populations (Mérel et al. 2021). Another

study identified F_{ST} outliers among Hawaiian *D. suzukii* populations (Koch et al. 2020). Apart

from these limited comparisons, the genetic changes that may have helped *D. suzukii* to adapt to

diverse worldwide environments are entirely unknown.

With the increasing availability of genomic resources from non-model species, genotype-

environment association (GEA), also known as environmental association analysis (EAA), is

becoming a widely-used approach to understand the potential relationship between specific

environmental factors and adaptive genetic variation (Rellstab et al. 2015). GEA is also useful in

identifying subtle changes in allele frequencies that are difficult to detect with outlier tests based

on traditional population genomic approaches, especially when the number of studied

populations is relatively large, and there is high gene flow counteracting patterns of local

adaptation (Kawecki and Ebert 2004). The capability of GEA to identify adaptive genetic

changes and environmental drivers of local adaptation has been demonstrated with whole-

genome pool-seq data from *Drosophila melanogaster* (Bogaerts-Márquez et al. 2021). Therefore,

GEA could be helpful in understanding the genotype-environment relationships underlying the

invasion success of *D. suzukii.*

In the present study, we perform population genomic analyses on whole-genome pool-seq

data of 29 population samples from native and invasive ranges to investigate the genomic

diversity, invasion history and environmental adaptation of *D. suzukii*. We investigate the

geographic pattern of genetic diversity to obtain clues regarding the history of this species within

Asia. We investigate population structure and perform demographic inference, assessing

agreement with published models of invasion history. Finally, we test the association between

SNP frequencies and nine environmental variables across sampling locations, identifying both

specific and shared functional signatures of adaptation to these diverse selective pressures.

Results

Genomic Diversity Reaches its Maximum in Eastern China

We investigated the genomic polymorphism of 29 *D. suzukii* populations derived from

Asia (n=8), Europe (n = 11), and North and South America (n = 10), and Asia (n = 8),

encompassing both newly reported and previously published samples (Figure 1A; Table S1;

Olazcuaga et al. 2020). Whole-genome sequences were obtained from 29 pooled samples

consisting of 50-212 female and male individuals. The depth of mapped reads after quality

control ranged from 23X to 66X among population samples, with an average of 45X (Table S1).

We first estimated nucleotide diversity (π_S) across synonymous SNPs to investigate the

effects of rapid invasions on neutral genetic diversity. The genome-wide π_S ranged from a

minimum of 0.041 (autosomes) and 0.019 (X chromosome) in Hawaii, US (US-Haw) to a

maximum of 0.059 (autosomes) and 0.045 (X chromosome) in Ningbo, China (CN-Nin). The

acute drop in π_S of introduced European and American populations relative to that of the native

Chinese and Japanese populations reflects previously reported founder event bottlenecks (Figure

1B, diagonal; Figure S1, S2). The observed patterns were also recapitulated with SNPs at all

types of sites (Figure S3). Lower π_S and more contrasting among-population π_S differences in X

chromosome reflect effectively prolonged bottlenecks due to their lower effective population size

(Pool & Nielsen, 2007; Figure S1). We also observed a greater loss of rare alleles in the invasive

populations as a typical consequence of bottlenecks (Figure S4). These founder events also

increased genetic differentiation as measured by F_{ST}, especially between continents (Figure 1B;

Figure S3), and particularly for the X chromosome (Figure S2).

Previous studies have hypothesized a relatively wide native range of *D. suzukii* in East and

Southeast Asia (Adrion et al. 2014; Fraimout et al. 2017). We examined genetic variation within

and among Asian populations to search for clues regarding the history of *D. suzukii* within its

native continent. The Ningbo population in eastern China had the highest π_S − 15.6 % greater

than any other population for the autosomes, and 30.6 % greater for the X chromosome (Figure

1B; Figure S1). Notably, D_{XY} values involving the Ningbo population did not meaningfully

exceed the magnitude of Ningbo's π, indicating that all other investigated locations primarily

have a subset of the genetic diversity present in Ningbo. These findings are consistent with the

other sampled populations descending in relatively recent population genetic time from a

Ningbo-like ancestor, and eastern China being within or closer to the ancestral range of the

species.

Population Structure and Invasion History of *D. suzukii*

Principal component analysis (PCA) of allele frequencies was next performed to offer

insights into the population structure of *D. suzukii*. The top three principal components (PC 1-3)

explained 57.44% (autosomes, Figure 1C) and 72.35% (X chromosome, Figure S2B) of the

variance among populations. In both autosomes and X, three-dimensional PCA and matrices of

both D_{XY} and window F_{ST} recapitulated both continuous and hierarchical geographic structure.

These results together showed the expected clustering of populations into four distinct ranges

(East Asia, Hawaii, Americas, and Europe; Figure 1B, 1C; Figure S2). Much of the observed

population differentiation is most likely due to founder event bottlenecks and admixture during

worldwide expansion (Fraimout et al. 2017), whereas migration following population

establishment would need to be overwhelmingly high to have significant impacts given the very

brief time scale of the global invasion. The two more northerly populations from the western

US, Oregon (US-Sok) and central California (US-Wat), show an affinity with the Hawaiian

population, which aligns with the suggestion that these populations received a genetic

contribution from Hawaii in addition to East Asia (Fraimout et al. 2017), while populations from

southern California and the central and eastern US show less evidence of such admixture. These

findings are also reflected by the clustering patterns in F_{ST} -based neighbor-joining trees (Figure

S5).

Figure 1. *Drosophila suzukii* populations show maximal diversity in eastern China and continent-level genetic

structure. (A) The geographic locations of the studied 29 natural populations are depicted as dots. In addition to the

22 populations sampled by Olazcuaga et al. (2020), populations newly sampled at independent locations are circled

in black. Populations newly sampled at nearby locations are circled and center-dashed in black, with the number of

total population samples in brackets. The year of the first recorded occurrence in each geographic range (colored

grey in the map) is given in brackets in the color legend. Further information about each sample is presented in

Table S1. (B) Population differentiation in allele frequencies (F_{ST}; lower triangle), between-population sequence

distances (D_{XY}; higher triangle), and within-population nucleotide diversity (π_S; diagonal) across autosomal

synonymous SNPs are displayed as a heatmap. Population names are colored by their geographic region. (C)

Autosomal genetic structure is shown by three-dimensional principal components analysis (PCA) based on allele

frequencies of the two most frequent alleles across all populations. Each dot represents a population. Labeled are

Hawaii and western coastal US populations. See the X chromosome version of (B) and (C) in Figure S2.

To provide a complementary understanding of the invasion history from genome-wide

information, we used TreeMix (Pickrell and Pritchard 2012) to build maximum-likelihood trees

with admixture graphs based on allele counts of SNPs from autosomes and X chromosome,

separately (Figure 2; Figure S7). We rooted the tree along the branch leading to the Ningbo

population (CN-Nin), since the above diversity analysis suggested that it could reflect the most

basal lineage among the investigated populations. We also sequentially added a range of

migration events (0 - 20) to trees to capture gene flow and parsimoniously improve the model fit,

and found that tree models with 11 mixture events explain most of the variance in relatedness

between populations (autosomes: 99.6%, X chromosome: 99.9%), while remaining interpretable

for both autosomes and X chromosome, with greater numbers of migration events resulting in

diminishing model improvements (Figure 2; Figure S6).

The topology of our TreeMix histories generally agreed with previous inferences (Adrion

et al. 2014; Fraimout et al. 2017; Lewald et al. 2021) and with our above results, including the

presence of clear genetic groupings from China, Japan, the United States, and Europe. The tree

structure largely agreed between our autosomal and X chromosome trees, and showed general

consistency across models where differing numbers of migration events were estimated. One

exception concerned the differential placement of the American clade. The autosomal tree

grouped the American and European lineages, which split from an Asian lineage closer to the

sampled Japan populations than to China. The X chromosome tree instead placed the

American group within the sampled Japan populations. Both results differ from the model of

Fraimout et al. (2017), which utilizes source populations from China for both continental

invasions, although each of our trees does include a specific migration event from China, and the

published model did entail ancestry from Japan to the mainland US via Hawaii.

The placement of the Hawaii population in both our autosomal and X chromosome trees

departed from the observed chronology of the species' expansion and from the model estimated

by Fraimout et al. (2017), which was conditioned on observed invasion dates. In both of our

trees, Hawaii was placed as an offshoot of the mainland US group, albeit with considerable drift

(Figure 2), even though *D. suzukii* was observed in Hawaii about 30 years sooner than on the

mainland. Although our trees do entail admixture from Hawaii into southern California

populations, the previously proposed history of Hawaii also contributing higher levels of

admixture into the Oregon (US-Sok) and central California (US-Wat) populations (versus Hawaii

arising from a population similar to those two) is easier to reconcile with observed invasion

history. These results highlight the challenges in estimating population history from genetic data

alone, from a finite number of populations, and in the presence of substantial drift and admixture

along some lineages.

The autosomal and X chromosome trees also differed in their placement of the

population from Réunion (an Indian Ocean island that is part of France). The autosomal tree

placed Réunion within the American clade, but with substantial bidirectional migration between

this population and European samples. Whereas, the X chromosome tree placed Réunion

alongside a population from France, which is more consistent with the European origin estimated

by Fraimout et al. (2017).

The autosomal tree also indicated an admixture event from western US back to Japan

with an admixture proportion of 32% (Figure 2A), consistent with a previous inference of western

US/Asia admixture using a larger number of US populations (Figure 4 of Lewald et al. 2021).

Other instances of migration events from a more recent into an older population on a different

continent were also suggested from Réunion to Europe (from the autosomal tree discussed above)

and from Brazil to three separate French population samples based on the X chromosome tree

(Figure 2B). Whereas some "migration events" might actually reflect independent invasions into

the same continent, followed by secondary contact during the range expansion process, these

"reverse migrations" cannot be explained in the same way. It is not clear that neutral migration

between continents within just a few years should really be expected to displace any noticeable

portion of an established population's gene pool. Alternatively, the precise accuracy of the

TreeMix models may be limited by incomplete population sampling, or by violations of the

neutral assumptions of the method, such as adaptive introgression, as was argued for gene flow

back into the African ancestral range of *D. melanogaster* (Svedberg et al. 2021).

Figure 2. Population topology and migration events of *D. suzukii* suggested by TreeMix. Maximum likelihood

graphs were built by allele counts at (A) autosomal and (B) X-chromosomal SNPs, allowing 11 migration events.

Horizontal branch lengths are proportional to the amount of genetic drift that has occurred on the branch. The

scale bar shows ten times the average standard error of the entries in the sample covariance matrix, reflecting the

precision of branch lengths estimates. Population names are colored by their geographic region. The year of the

first recorded occurrence in each geographic range is given in brackets in the color legend. Migration arrows are

colored according to their weight, which approximates admixture proportions. The residual fit from these graphs is

presented in Figure S7.

Figure 3. Chromosomal distribution of genetic polymorphism in *D. suzukii* informs the ordering and orientations of

contigs, as well as levels of centromeric and telomeric repression. Window nucleotide diversity (π_w) values are

displayed across (A) the X chromosome and major autosomal arms (B) 2L, (C) 2R, (D) 3L, (E) 3R. Chromosome 4 is

not shown as it only contains 12 windows. Each window is a continuous genomic region that includes 125,000

analyzed sites. Each dot represents the average π_w across populations within their geographic range as colored.

Only populations from major continental ranges are shown for chromosomal patterns to be clear. Within each

chromosome arm, contigs are ordered by length. Separate contigs are indicated by grey or white shading.

Polymorphism, Divergence, and the Genomic Locations of *D. suzukii* Contigs

Genome-wide patterns of polymorphism and divergence are useful to reveal various

aspects of the evolutionary history of a species. The *D. suzukii* reference assembly (Paris et al.

2020) is somewhat fragmented, with two or more contigs assigned to each major chromosome

arm, but with unknown orientation with respect to each other. We note that in other examined

D. melanogaster group species, crossing-over rate and window nucleotide diversity (π_w) are reduced

in centromere- and telomere-proximal regions (*e.g.*, True et al. 1996). When we examined the

chromosomal distribution of π_w (Figure 3), we noticed that certain arrangements of contigs

would result in the expected patterns of reduced diversity at the ends of each arm, and relatively

smooth shifts in the diversity of large windows. For instance, the second longest contig in

chromosome arm 3L is likely to the left of the longest contig in a reverse orientation, potentially

with the smaller high diversity contig in between, (Figure 3D), although either of the low diversity

contig end could be centromeric or telomeric. Similarly in chromosome arm 3R, although the

assembly was more fragmented and the centromere- or telomere-proximal regions consist of

multiple contigs, the second and third longest contigs appeared to contain the transitions between

the high diversity mid-arm region and the centromeric/telomeric zones (Figure 3E). Therefore,

genome-wide polymorphisms could serve as useful information to aid the ordering and orienting

of contig-level genome assemblies (Nakabayashi and Morishita 2020), although a combination of

both systematic neighbor-matching approaches and manual correction would be needed, and

integration with other forms of evidence (*e.g.,* comparative genomic or experimental data) would

be preferable.

Genomic polymorphism levels could also be indirectly influenced by centromeric and

telomeric repression of crossing over, leading to a stronger influence of natural selection reducing

linked variation (Maynard Smith & Haigh, 1974; Charlesworth et al. 1993). From the

distribution of window-based nucleotide diversity (π_w) across chromosome arms, we noticed that

centromere- and telomere-proximal regions of reduced diversity showed similarly narrow

declines in π_w (Figure 3). These patterns are more similar to those in *D. simulans* than to *D.*

melanogaster – which has broader centromeric regions of low crossing over (Langley et al. 2012).

These results suggest a relatively weaker suppression of crossing over in the centromere-proximal

regions in *D. suzukii*. It also appears that regions of low nucleotide diversity (which probably

coincide with regions of low recombination) cover a relatively small fraction of the genome,

which should limit the potentially biasing effects of natural selection on demographic inferences

and facilitate the localization of selective sweeps (Schrider et al. 2016; Lange and Pool 2018).

We next leveraged our large pooled sequencing data set to improve inferences about

which contigs map to the X chromosome. Out of a total of 546 contigs, 313 were previously

assigned to autosomes and X chromosome through either direct mapping or comparing a

female-to-male read depth ratio (Paris et al. 2020). We added to these annotations by

implementing an approach based on correlations in sequencing depth of coverage across

population samples that included varying numbers of females and males (see Materials and

Methods). Based on this analysis, we assigned 170 contigs as autosomal or X-linked. Our

classification of previously-assigned contigs was 96% consistent with past inferences, but we

corrected four previous assignments that were based on female-to-male read depth ratio, whereas

our method did not assign eight previously-assigned contigs.

To reveal levels of selective constraint, we estimated the divergence between *D. suzukii*

and its close relative *D. biarmipes* (Ometto et al. 2013; Suvorov et al. 2022; Figure S8). Compared

to the divergence of *D. melanogaster* from *D. simulans*, *D. suzukii* shows a higher genome-wide

divergence from *D. biarmipes*. The intronic and intergenic regions also have a higher divergence

relative to that of nonsynonymous sites, suggesting that a lower proportion of these non-coding

regions are constrained in *D. suzukii* and *D. biarmipes* than in *D. melanogaster* and *D. simulans* (Begun

et al. 2007; Lange and Pool 2018). Non-coding divergence is potentially inflated by repetitive

elements (i.e., repeatome), which represent about half of the genome expansion of *D. suzukii*

compared to the *D. melanogaster* genome (Paris et al. 2020). This expansion of the repeatome

could reflect a lower long-term effective population size (N_e) in the *D. suzukii* lineage. Given that

the repeatome is predominantly contributed by transposable elements (TEs), that hypothesis

aligns with the negative correlation between TE content and N_e found among 22 worldwide *D.*

suzukii populations also sampled in this study (Mérel et al. 2021). However, the greater π of *D.*

suzukii than *D. melanogaster* (Figure S1; Lack et al. 2016a; Lewald et al. 2021) could instead

indicate a greater N_e for *D. suzukii* within the past $4N_e$ generations.

Environmental Adaptation Reveals the Genetic and Functional Basis of Invasion

Success

The worldwide expansion of *D. suzukii* has exposed this species to selection pressures from

varying local environmental conditions (Olazcuaga et al. 2020; Mérel et al. 2021). To identify

environmental factors that have contributed to adaptive genetic differentiation at various levels

and loci under positive selection, we performed a whole-genome scan using GEA analysis

between environmental and genetic differentiation (see Materials and Methods). Our selection of

environmental variables for GEA started with a preliminary set of 26 candidate variables that are

potentially relevant in the adaptation process of *D. suzukii* (Figure 4A), based on data availability

and prior knowledge of *Drosophila* ecology (Kellermann et al. 2012; Hamby et al. 2016; Bogaerts-

Márquez et al. 2021). Among common strategies of performing GEA with a large set of relevant

environmental variables, univariate association with all environmental variables could increase

the number of statistical tests, thus increasing the difficulty of controlling rates of false discovery.

On the other hand, including multiple variables might cause a multicollinearity issue (Rellstab et

al. 2015). We opted to retain nine of the least correlated environmental variables for univariate

tests, representing altitude, wind speed, as well as multiple aspects of temperature, precipitation,

and human land usage for GEA analysis (Figure 4B). Although the temperature of the coldest

quarter had a significant negative correlation with wind speed, we kept both variables for GEA,

as cold stress and wind-related factors are known to be potential drivers of local adaptation in

Drosophila (Bogaerts-Márquez et al. 2021). As indicated by their coefficients of variation (*CVs*), the

selected environmental variables had moderate ($15\% \leq CV < 30\%$) to high ($CV \geq 30\%$)

variability across our sampling locations (Table S3).

Figure 4. Identification of least-correlated environmental variables for genotype-environment association (GEA)

analysis in *D. suzukii*. (A) Pairwise correlations among a preliminary set of 26 environmental variables that are

potentially impactful on *D. suzukii*. (B) A final set of nine of the most relevant and least correlated environmental

variables that were chosen for GEA analysis. The Pearson correlation coefficients are colored from -1 (perfect

negative correlation) to 1 (perfect positive correlation). Significance correlations (p < 0.05) are indicated by

asterisks. See Table S3 for environmental values used to calculate correlation coefficients.

From 5,752,156 genome-wide SNPs with a minor allele frequency (MAF) higher than

5%, we identified an average of 3,033 (SD = 823.4) unique candidate variants that were

significantly (genome-wide $q < 0.05$) associated with each of the nine candidate environmental

variables (Table S4). These variants corresponded to an average of 3,345 overlapping or

neighboring genes (Table S4), suggesting that selection pressures from the tested environmental

variables (or correlated factors) have meaningfully contributed to adaptive genetic variation in *D.*

suzukii, even on the brief time scale of its worldwide expansion. Among all tested environmental

factors, mean temperature of the coldest quarter was associated with the greatest number of

putatively adaptive variants (4,250 SNPs). Two precipitation-related variables, annual

precipitation (4,141 SNPs) and precipitation seasonality (3,389 SNPs), have the next largest loci

count. The ratio of built area to vegetation (i.e., crops and forests) and the ratio of crops to

forests were associated with the fewest genetic variants (2,369 and 1,608 SNPs, respectively).

To reveal the potential genetic and functional basis of invasion success under multiple

environmental challenges throughout the species' range, we first examined the functions of genes

linked to the top 10 environment-associated loci for each variable as ranked by association q-

value, and then by the g parameter estimating the sensitivity to environmental differentiation as

a tie-breaker. We found many of these genes have known functions that could facilitate

adaptation to the associated environmental factor (Table S5).

Among genes linked to altitude-associated loci, the second-ranked candidate *ab* is known

to control wing size in *Drosophila* (Simoes da Silva et al. 2019). Interestingly, wing size was found

to have increased in a highland Ethiopia *D. melanogaster* population, potentially assisting flight in

thin, cool air (Lack et al. 2016b). Ranked next to *ab* is *Gbs-70E*, which plays roles in glycogen

metabolism and the development of eggs inside the maternal ovary (Kerekes et al. 2014).

Another top gene, the lysine demethylase *Kdm2*, is upregulated in response to hypoxia (Batie et al.

2017).

With wind speed, the top first candidate *Ttc30* is an essential gene in the biogenesis of

sensory cilia, which are key to both chemosensory and mechanosensory functions in *Drosophila*

(Avidor-Reiss et al. 2004; Avidor-Reiss and Leroux 2015). Another top candidate, *Arr2*, is

involved in olfaction, hearing, and vision (Alloway and Dolph 1999; Elaine Merrill et al. 2005;

Senthilan et al. 2012). In light of the relevance of wind for insect flight, we also noted that a third

top candidate, *vn*, is a developmental gene named for its wing phenotype (Wang et al. 2000).

There is also some evidence for precipitation-related local adaptation. The top gene *mmy*

associated with precipitation seasonality (i.e., annual range of precipitation) was shown to

regulate chitin synthesis and cuticle production. Since precipitation is correlated with desiccation

resistance across the *Drosophila* phylogeny (Kellermann et al. 2012), *D. suzukii* may have

developed adaptive strategies of modifying chitin biosynthesis under conditions of desiccation

(Rezende et al. 2008; Clark et al. 2009), which was also implied in seasonal plasticity of natural

Drosophila populations (Shearer et al. 2016; Horváth et al. 2023). In addition, the gene *osy*

(*CG33970*) contributes to the formation of the outer cuticle layer and is expressed more highly in

D. suzukii than in *D. melanogaster* (Wang et al. 2020). Furthermore, two of the top genes associated

with annual precipitation (*Abd-B* and *bab1*) regulate cuticle pigmentation (Rogers et al. 2013),

which may or may not correlate with desiccation tolerance in *Drosophila* species (Wang et al.

2021). We also note that although environmental fitness effects on these testes-expressed genes

are not known, the same SNP near *CG17944* and *nxf4* was among the highest-scoring variants for

both annual precipitation and precipitation seasonality (variables that have a non-significantly

negative correlation between them; Figure 4).

Another important environmental barrier to invasion success is temperature. For the

mean temperature of the coldest quarter, a top gene was *Ac78C*, which has roles in circadian

regulation and taste (Ueno and Kidokoro 2008; Duvall and Taghert 2013). With the mean

temperature of the warmest quarter, the top genes *crp*, *Mrtf*, and *Ubx* help control the

development of trachea (Han et al. 2004; Guha and Kornberg 2005; Wong et al. 2015), which

may be important in limiting water loss in hot environments (Gibbs et al. 2003).

For the ratio of built to vegetated area, a different variant near the cuticle-related gene *osy*

(which was also indicated above for precipitation seasonality) was detected. Another top outlier

was the nervous system gene *trv*, which is involved in thermosensitivity (Honjo et al. 2016). For

the relative levels of crop and forest cover, the first-ranked variant was near *Mtk*, which encodes

an antifungal and antibacterial peptide (Levashina et al. 1995), and we note that mushrooms

(which are more available in forest) have been proposed as overwintering food sources for *D.*

suzukii (Wallingford et al. 2018), and the evolution of immune genes has been found to differ

strongly between mushroom-feeding and human commensal *Drosophila* species (Hill et al. 2019).

With regard to the differential light environments entailed by forest versus farm habitats, we note

that the next highest gene, *CadN2*, helps connect photoreceptor neurons to their targets (Prakash

et al. 2005).

Beyond the top candidate genes that have related functions to specific types of

environmental changes, we also found a wide range of nervous system genes associated with

multiple environmental factors. For instance, among the top five altitude-associated loci, three

have known functions in the nervous system of *Drosophila*, including the first-ranked gene *Cmpy*,

which enables proper growth control at neuromuscular junctions (James and Broihier 2011), *ab*,

which regulates dendritic complexity (Li et al. 2004; Sugimura et al. 2004), and *not*, which is

essential for stabilizing synaptic homeostasis within glia (Wang et al. 2020). Such genes were also

linked to top-10 loci associated with wind speed (*dpr6*), precipitation (*Msp300*, *Tusp*, *5-HT2A*),

temperature (*ATP6AP2, CG13579, D*), and land use variables (*Bsg*, *Mp*, *velo*). Different variants

associated with *scrt*, a regulator of neuronal cell fate, were among the top results for both mean

diurnal range and the ratio of crop to forest cover.

Next, we examined environment-specific adaptation on a more comprehensive basis

through a gene ontology (GO) enrichment analysis of the top 500 genes associated with each

environmental variable (Figure 5). As correlates of temperature in the coldest quarter, cAMP

metabolic process was the top enriched category, followed by two other related purine

metabolism groupings. We note that cAMP is important in circadian regulation (*e.g.*, Palacios-

Muñoz & Ewer, 2018), which is known to play an important role in *Drosophila* environmental

adaptation (*e.g.,* Helfrich-Förster et al. 2020), as also implicated by the presence of "entrainment

of circadian clock" on our top GO category list for altitude. More broadly, purine metabolism

was inferred as a strategy of cold acclimation in *D. suzukii* (Enriquez and Colinet 2019). For

diurnal temperature range, the top category was "regulation of growth", and we note that some

drosophilids have evolved to have larger body sizes in more challenging thermal environments

(Gilchrist and Partridge 1999; Calboli et al. 2003; Lack, Yassin, et al. 2016).

With precipitation, we identified "chitin metabolic process" as the top GO term

associated with annual precipitation, as well as "chitin-binding" with precipitation seasonality.

Together with the chitin synthesis genes we described above for precipitation, adaptation to the

overall intensity and seasonal variation of precipitation by modifying cuticular chitin may be

implied. For crop to forest ratio, the category "antimicrobial humoral response" included the top

gene *Mtk* listed above.

GO classes
- Biological process
- Molecular function
- Cellular component

	GO description	p-value	# genes
Altitude	nucleus localization	1.35e-03	7
	R7 cell fate commitment	1.89e-03	7
	R7 cell differentiation	2.44e-03	12
	sensory organ precursor cell fate determination	2.51e-03	6
	synaptic vesicle	2.67e-03	8
	intracellular protein-containing complex	2.76e-03	24
	endomembrane system	4.40e-03	53
	entrainment of circadian clock	4.99e-03	5
	translation initiation factor activity	5.85e-03	5
	centrosome duplication	6.85e-03	5
Ann. precip.	chitin metabolic process	9.70e-04	5
	transferase complex	1.84e-03	24
	I band	3.19e-03	8
	nucleoplasm	3.36e-03	24
	cellular protein-containing complex assembly	3.39e-03	23
	single fertilization	3.63e-03	5
	supramolecular fiber organization	3.84e-03	24
	cytoplasm	4.18e-03	145
	hydrolase activity, acting on glycosyl bonds	4.24e-03	8
	actin cytoskeleton organization	4.32e-03	27
Ann. mean wind speed	centrosome	2.40e-03	11
	ligand-gated sodium channel activity	2.61e-03	5
	cellular protein catabolic process	5.34e-03	19
	spindle	6.26e-03	9
	triglyceride lipase activity	7.07e-03	6
	nitrogen compound metabolic process	9.00e-03	167
	primary metabolic process	1.08e-02	175
	protein ubiquitination	1.19e-02	16
	organic substance metabolic process	1.26e-02	182
	actin cytoskeleton reorganization	1.39e-02	5
Mean temp. coldest qtr.	cAMP metabolic process	5.00e-04	6
	purine nucleotide metabolic process	1.35e-03	16
	oxidoreductase activity, acting on the aldehyde or oxo group of donors	6.18e-03	6
	sevenless signaling pathway	9.82e-03	5
	positive regulation of JNK cascade	1.03e-02	6
	positive regulation of cell projection organization	1.44e-02	8
	identical protein binding	1.54e-02	15
	cation transport	1.71e-02	32
	organic hydroxy compound metabolic process	2.35e-02	12
	locomotion	3.01e-02	60
Ratio crop to forest	cell-substrate junction	3.90e-04	9
	cell surface	1.15e-03	14
	extrinsic component of cytoplasmic side of plasma membrane	2.46e-03	5
	signaling receptor binding	5.99e-03	24
	positive regulation of filopodium assembly	7.48e-03	5
	cell-matrix adhesion	9.53e-03	5
	antimicrobial humoral response	1.05e-02	10
	cortical actin cytoskeleton organization	1.16e-02	6
	germ-band shortening	1.67e-02	5
	cell fate determination	1.85e-02	16

	GO description	p-value	# genes
Precip. Seasonality	response to sucrose	2.45e-03	5
	signaling receptor activity	5.59e-03	48
	detection of chemical stimulus involved in sensory perception	8.82e-03	12
	exopeptidase activity	9.20e-03	9
	chitin binding	1.45e-02	8
	positive regulation of response to external stimulus	1.65e-02	11
	hindgut development	1.68e-02	10
	gastrulation involving germ band extension	1.81e-02	6
	positive regulation of response to biotic stimulus	2.01e-02	10
	histone binding	2.20e-02	6
Mean diurnal temp. range	regulation of growth	1.70e-04	33
	regulation of cellular component organization	1.74e-03	49
	cell growth	2.39e-03	18
	R3/R4 cell differentiation	2.52e-03	7
	dorsal/ventral pattern formation, imaginal disc	3.20e-03	11
	embryo development	3.22e-03	48
	post-embryonic animal organ development	3.92e-03	46
	regulation of developmental process	4.26e-03	56
	compound eye morphogenesis	4.81e-03	33
	generation of neurons	5.56e-03	79
Mean temp. warmest qtr.	extracellular matrix structural constituent	1.00e-03	7
	spindle pole	1.19e-03	6
	regulation of neurogenesis	1.64e-03	18
	centrosome	2.73e-03	11
	regulation of dendrite development	3.16e-03	9
	regulation of neuron projection development	3.26e-03	14
	cellular macromolecule biosynthetic process	3.73e-03	95
	sex differentiation	4.97e-03	17
	pattern specification process	6.56e-03	47
	brain development	7.22e-03	17
Ratio built to vegetation	appendage morphogenesis	3.88e-03	38
	adenylate cyclase-inhibiting G protein-coupled receptor signaling pathway	5.49e-03	6
	positive regulation of synaptic transmission	5.64e-03	5
	carbohydrate biosynthetic process	5.93e-03	6
	anterior Malpighian tubule development	5.97e-03	5
	wing disc morphogenesis	6.41e-03	32
	cleavage furrow	8.33e-03	5
	neuroblast differentiation	9.56e-03	11
	late endosome	1.02e-02	8
	carbohydrate derivative biosynthetic process	1.20e-02	17

Figure 5. GO enrichment analysis of candidate genes from the gene-environment association analysis of *D. suzukii*.

The top 10 GO categories enriched by the top 500 genes associated with each environmental variable are shown in

each panel (labelled on the left), with permutation p-values and the number of associated genes in each GO

category. Descriptions of GO categories are colored by their GO class (see legend at top right). Only GO

categories including more than five associated genes are listed here. For a full list of enriched GO categories, see

Table S6.

As broader evidence for a shared (or biologically similar) underlying genetic basis of

adaptation to multiple environmental factors, we examined the overlap of the most significant

genes and most enriched GO categories between different environmental variables. We found

the top candidate genes to be mostly associated with both temperature and precipitation. The

gene sets showed relatively greater overlap among climatic variables (including altitude), whereas

the two land usage variables had less overlap with climatic variables or with each other (Figure

6A). Since the patterns of shared genes cannot be fully explained by correlations between

environmental values (Figure 4B), at least some of the genes may have been responding to

multiple selective pressures. The overall proportions of shared GO categories were lower than

those of shared genes, indicating that the shared genes do not necessarily lead shared functional

categories between environmental variables. Relatively higher GO term sharing was observed

between altitude and either wind speed or precipitation, and between diurnal temperature range

and temperature of the warmest quarter (Figure 6B). Based on the shared genes and GO terms

observed, it is possible that during the rapid range expansion of *D. suzukii*, pleiotropy may have

facilitated local adaptation to multiple selective pressures (Hämälä et al. 2020; Kinsler et al.

2020).

Consistent with our gene-based analyses of universal adaptive function, we found three of

the top shared biological processes clearly related to nervous system functions, including the

topmost "synaptic transmission, glutamatergic" (shared by altitude, wind speed, diurnal

temperature range, and temperature of the warmest quarter), "regulation of neurogenesis"

(altitude, diurnal temperature range, temperature of the warmest quarter, and ratio of built area

to vegetation), and "central nervous system development" (precipitation seasonality, diurnal

temperature range, and temperature of the warmest quarter) (Figure 6C). Further, as mentioned

above, "cAMP metabolic process" (shared by temperature of the coldest quarter, precipitation

seasonality and ratio of built area to vegetation) could entail neurologically modulated changes in

circadian behavior. Each of these traits was associated with at least three environmental

variables, which suggests a multifaceted role for nervous system evolution in facilitating the

invasion success of *D. suzukii* under multiple environmental challenges.

Figure 6. Overlapping genes and GO categories among environmental factors reveal the shared genetic and

functional basis of environmental adaptation in *D. suzukii*. The numbers and proportions of shared (A) environment-

associated genes and (B) enriched GO categories among environmental factors are shown in heatmaps. Here, joint

proportion represents the fraction of the genes or GO terms associated with either of two environmental variables

that are associated with both variables. (C) Top GO categories of each type are depicted as bubbles. Bubbles are

colored by the negative logarithm of the combined p-value of enrichment across all environmental variables, and are

scaled by the number of enriched genes. The number of environmental variables that enrich a given GO category is

indicated by the top horizontal axis.

Discussion

We performed population genomic analyses of 29 population samples of *D. suzukii* to

investigate the genomic diversity, invasion history, and environmental adaptation of this highly

invasive species across its worldwide distribution. Our data shows a genetic grouping of these

populations into four primary geographic regions: eastern Asia (containing the native range),

Hawaii, the Americas, and Europe. We estimated the highest genetic diversity in eastern China

(Ningbo), suggesting this region could be within or near the ancestral range of *D. suzukii*. We

confirmed moderate founder event bottlenecks in introduced populations, and our analysis

suggested that admixture during the expansion of *D. suzukii* populations might be more complex

than previously estimated, but that further sampling in Asia is needed to address the precise

origins of introduced populations. We then identified the environmental drivers for local

adaptation of *D. suzukii*, including altitude, wind speed, precipitation, temperature, and human

land usage, where temperature of the coldest quarter shows the clearest evidence of

environmental adaptation. Our results suggested extensive local adaptation in response to

specific environmental challenges. We also revealed appreciable sharing of genes and functional

pathways underlying invasion success across multiple environmental pressures, which were most

obvious with nervous system genes.

Insights and mysteries regarding the history of *D. suzukii*

Surprisingly, the relative levels of diversity of *D. suzukii* populations have been the subject

of conflicting findings. Here, we detected the highest overall genetic diversity for *D. suzukii* in

eastern Asia, especially in the eastern China Ningbo population (Figure 1B; Figure S1). Our

results contrast with some prior findings, such as higher diversity in several North American

populations relative to a Japanese population (Adrion et al. 2014), the species' highest diversity

being in north-central China (Fraimout et al. 2017), or alternatively in Japan (Gautier et al.

2022). These conflicts may reflect challenges in accurate estimation of genetic diversity,

especially when the number of investigated loci is limited (Adrion et al. 2014; Fraimout et al.

2017), or when rare alleles are filtered by a relatively strict allele count threshold (Gautier et al.

2022), given that rare variants are especially likely to be lost during bottlenecks. In this study, we

accounted for such biases by applying stringent filtering by sequencing read base quality and

mapping quality, by including putative rare alleles in genome-wide analyses, and by focusing on

synonymous diversity which has a relatively higher ratio of signal to noise (more true variation

relative to sequencing errors).

Beyond the eastern China population's maximal diversity, we found that no pairwise D_{XY}

values meaningfully exceeded Ningbo π, implying that two Ningbo *D. suzukii* genomes are as

different from each other as any pair of genomes worldwide (paralleling a result for a Zambia

population of the human commensal *D. melanogaster*; Pool et al. 2012). Those findings are

consistent with all sampled *D. suzukii* populations originating from a Ningbo-like ancestor in

recent population genetic time. This inference could conceivably imply that *D. suzukii* only

expanded across portions of its current eastern Asian range due to its relationship with human

agriculture. Alternatively, in light of the apparently large effective population size of *D. suzukii*,

the plausible time frame of divergence among Asian populations could extend back toward the

last glacial maximum (LGM). We note that Ningbo is near the historical northern limit of

temperate woodlands during the LGM (Ray and Adams 2001). If this region is indeed part of a

limited overlap between the continental habitats of this temperate species during glacial and

interglacial periods, then this stability of occupation could help explain the Ningbo population's

elevated genetic diversity, whereas other eastern Asian populations' somewhat lesser diversity

might stem from mild founder event bottlenecks during post-glacial range expansions.

Further sampling and analysis of Asian *D. suzukii* populations is urgently needed to

advance our understanding of the history of this species. Even the current distribution of *D.*

suzukii in eastern Asia is not entirely clear. According to TaxoDros

(https://www.taxodros.uzh.ch/), *D. suzukii* has been sampled from China, Japan, North and

South Korea, Burma, Cambodia, Thailand, and Taiwan. It is unclear whether the lack of

records from neighboring countries such as Laos and Vietnam, where the species is predicted to

thrive (Santos et al. 2017), reflects a genuine absence or insufficient sampling effort.

Understanding the history of *D. suzukii* in Asia will require the analysis of genomic diversity from

dozens of population samples from across its Asian range, including from countries beyond

China, Japan, and Korea, where all population genetic study to date has focused.

Geographically denser population genomic data from Asian *D. suzukii* would also

facilitate clearer conclusions regarding the origins of lineages that invaded regions such as

Hawaii, the mainland US, and Europe. Above, we noted some contrasts between our TreeMix

results, which placed mainland US and Europe populations closer to sampled Japan populations

than to China, and the model of Fraimout et al. (2017), which primarily drew upon China source

populations for these invasions. However, our autosomal TreeMix results suggested that neither

of these invasive lineages falls within the clades of sampled populations from either China or

Japan (and our X chromosome analysis echoes this finding for Europe). Our results are

compatible with the possible origins of continental *D. suzukii* invasions from one or more Asian

populations that are somewhat genetically differentiated from any of those analyzed here. At

present, we suggest that the geographic origin of these economically significant introductions

remains an open question.

Our TreeMix analysis also suggested that worldwide patterns of admixture may be more

complex than suggested by previously inferred scenarios (Adrion et al. 2014; Fraimout et al.

2017; Gautier et al. 2022). However, caution is required to interpret admixture events that were

inferred solely from this analysis, and there may be several reasons why some inferences differ

between ours and prior analyses. First, as indicated above, we are still working with a very

incomplete panel of populations, especially from Asia, and methods may differ in terms of how

they respond to the absence of historically important populations such as the sources of the

invasive lineages. Second, whereas the study of Fraimout et al. (2017) restricted its consideration

to founder events compatible with the observed chronology of species expansion, our TreeMix

results represent an investigation of population history through genomic data alone, without

assuming that each worldwide invasion was detected rapidly upon its occurrence. In species with

complex population structure, multiple histories could lead to identical covariance matrices, thus

allowing for several different invasion histories compatible with the data (Pickrell and Pritchard

2012). Finally, as mentioned by Adrion et al. (2014) and Fraimout et al. (2017), the additional

migration events predicted in our analysis could indicate that genome-wide data is more sensitive

for discriminating among complex invasion scenarios and population structures than a limited

number of loci.

As indicated above, our TreeMix results also differed somewhat between autosomal and

X-linked loci. To some extent, demographic history is expected to differ between these loci,

including due to different levels of bottleneck severity at distinct effective population sizes, and

the unbalanced sex ratio of this species, especially when the multiple mating is present or sex

ratios depart from null expectations (Pool and Nielsen 2009; Tait et al. 2018; Weißinger et al.

2019; Manko et al. 2021). However, it is not clear that tree topology or gene flow presence

should differ substantially on such grounds. In addition to concerns listed above such as model

identifiability and missing populations, the consistency of autosomal versus X-linked inferences

could also be influenced by natural selection, particularly if it influences a longer stretch of the

genome with reduced recombination, such as a centromeric or telomeric region, or a

polymorphic inversion.

A key limitation of our pool-seq data is that it cannot provide haplotype information. In

light of the challenges in performing demographic inference from pooled sequencing data, there

would be considerable benefit in high quality individual sequencing of multiple wild-caught flies

from each of a broad array of populations (particularly in Asia), in order to facilitate quantitative

inferences regarding population history. Such data, especially if combined with efficient model

selection methods (*e.g.,* ABC-RF, (Pudlo et al. 2016; Fraimout et al. 2017), could provide a more

confident and precise demographic model which could both inform species history and serve as

null hypothesis for the detection of natural selection.

Environmental Drivers of Adaptation in *D. suzukii*

Here, we presented a GEA analysis that investigated the most geographically and

genetically diverse set of *D. suzukii* populations and the most comprehensive set of environmental

factors to date, which enabled unprecedented power to capture even minor adaptive genetic

differentiation in response to distinct environmental challenges during the species' rapid

invasions. In addition to our identification of climatic factors including temperature,

precipitation-related variables and wind speed as the most frequently correlated with putatively

local adaptative variants (consistent with previous GEA analysis in *D. melanogaster*, *e.g.*, Bogaerts-

Márquez et al. 2021), we also for the first time identified large numbers of genome-wide variants

associated with altitude and human land usage-related variables (Table S4), which were not

included in most GEA studies despite their potential significance to local adaptation. In

particular, the detected associations with ratios of developed land to vegetation and of cropland

to forests highlights the ecological impacts of urbanization and agriculture on natural populations

of insects. Since the prior selection of environmental variables is critical for successful GEA

analyses, we also provided an instructive example for correlation-based selection to identify the

most relevant and least redundant environmental factors (Rellstab et al. 2015).

Nervous System Evolution is Ubiquitous in Environmental Adaptation of

Drosophila

In *D. suzukii*, we found nervous system and related sensory and behavior annotations

associated with top genes for all nine environmental variables studied. Concordantly, we found

that GO categories related to the nervous system were among the most shared across

environmental variables (Figure 6C). In *D. melanogaster*, related GO categories like 'neuron

development', 'nervous system development' and 'eye development' were also enriched among

genes associated with environmental variation among natural populations within North America

or Europe, and across seasons within Europe (Bogaerts-Márquez et al. 2021). GO categories

associated with the nervous system have also shown evidence of positive selection in various

genome scans of *D. melanogaster* (Langley et al. 2012; Pool et al. 2012; Pool 2015), including a

study of parallel evolution in cold-adapted populations (Pool et al. 2017). Given the

morphological evidence of neuron-muscular junction evolution across the entire *Drosophila*

phylogeny (Campbell and Ganetzky 2012), we therefore propose a broad adaptive importance of

the nervous system in *Drosophila* species and potentially other insects. Such evolutionary

processes may have either maintained ancestral neural functions in novel challenging

environments, or created novel phenotypes that better fit the new optima arising from complex

combinations of environmental factors.

Cautions with Interpretations of Association Results and Future Directions

While we have generated intriguing hypotheses about gene functions that may underlie

the environmental adaptation of *D. suzukii*, it is difficult to distinguish between correlated

environmental selective pressures that may have driven the detected associations, including not

only the 17 environmental factors that were excluded in the process of variable reduction, but

also correlated biotic or abiotic factors not represented in global databases. As an intrinsic

limitation of GEA analysis that cannot be accounted for by applying stricter thresholds,

associations observed with a particular environmental factor might stem from adaptation to other

covarying factors (Rellstab et al. 2015). For example, the two tracheal branching genes *crp* and

Mrtf (Han et al. 2004; Wong et al. 2015) associated with mean temperature of the warmest

quarter could represent adaptations to reduce water loss under conditions of elevated water

vapor pressure (Telonis-Scott et al. 2012), which is closely related to humidity and has a

significant positive correlation with mean temperature of the warmest quarter (Figure 4B).

Therefore, expanded characterization of the relationships between genotype, phenotype,

and fitness in this species is needed to further clarify the functional and phenotypic

interpretations associated with certain environmental factors and genes. Experimental

validations that leverage RNA interference (Boutros and Ahringer 2008) and/or transgenic

overexpression (Prelich 2012) to modify the expression of associated genes, and/or genome

editing techniques (Stern 2014; Turner 2014; Shalem et al. 2015) to target putatively adaptive

variants would also bring a more solid understanding about the invasive biology of this species in

distinct environments. Such functional studies could be complemented by population

experiments under controlled laboratory environments or field conditions (*e.g.,* Behrman et al.

2015; Rudman et al. 2022), in order to more clearly demonstrate the connections between

specific selective pressures and alleles or traits of interest.

Broader Impacts and Significance

Our work integrates genetic and environmental data to improve the reconstruction of the

invasion genomics of a crop pest carrying significant economic costs (Knapp et al. 2021), which

will hopefully inspire future studies on developing diverse pest control methods given the

adaptive and neutral genetic differentiation among *D. suzukii* populations. Understanding the

extent of local adaptation and its potential environmental drivers will also help predict the spread

and future distributions of invasive species (Colautti and Lau 2016). More broadly, the enhanced

understanding of how organisms may adapt to geographical, climatic and artificial selective

pressures from this study is also of value in assessing the susceptibility of natural populations to

climate change (Kellermann et al. 2012) and human activities (Barange et al. 2010).

Our analysis of genome-wide diversity has uncovered novel patterns (such as the maximal

diversity of an eastern China population), while reinforcing other prior findings and adding new

perspectives on the invasive history of *D. suzukii*. As suggested above, our findings indicate a

strong need for the sequencing and analysis of individual genomes from a larger number of

worldwide populations, especially from additional regions of eastern and southern Asia. Such

efforts could lead to a vastly improved understanding of both the evolution of *D. suzukii* within its

native continent and the geographic origins of worldwide invasions.

Materials and Methods

Fly Collection, DNA Preparation, and Pooled Sequencing

Fly samples from 29 populations were used, seven of which were sequenced for the

present study. The fly samples sequenced in this study were collected from wild *D. suzukii*

populations in two states of the USA, two provinces of Japan, and three European countries

(Figure 1; Table S1). Pooled whole adult flies (n = 100 ~ 183) from each population (Table S1)

were used for DNA extraction as previously described (Langley et al. 2011).　Library

preparations were conducted at the Next Generation Sequencing Core of University of

Wisconsin Madison Biotechnology Center (https://dnaseq.biotech.wisc.edu), where pair-end

(PE) reads at the length of 150bp were then generated for each of seven pooled DNA samples on

an Illumina NovaSeq 6000.

Pool-sequenced reads of 22 additional *D. suzukii* population samples, including from

Europe, the Americas, and Asia, were obtained from public data provided by Olazcuaga et al.

(2020) at EBI's SRA (Figure 1; Table S1). Taken together, we formed a comprehensive dataset

of 29 populations sampled from native and invasive ranges of *D. suzukii*.

Quality Control, Alignment, and Variant Calls from Pool-seq Data

To maximize the quality of our analyzed data, we built a high-throughput assembly and

quality control pipeline *poolWGS2SNP* with optimized performance, stringent filtering,

compatibility with large numbers of genomic contigs, and customized functions to call high-

confidence single-nucleotide variants from pool-sequenced data in *D. suzukii* (Figure S9), in part

by utilizing resources from the DrosEU bioinformatics pipeline (Kapun et al. 2020).

As an initial quality control of raw PE reads, adapters were removed, and the 3' end of

reads with base quality < 20 were trimmed using *fastp* (Chen et al. 2018). Further trimming was

performed using a self-developed python program *filter_PE_length_mem.py* (see Data Availability),

where any pair of forward and reverse reads with less than a total of 150 bases with base quality

(BQ) ≥ 20, as well as any individual reads with less than 25 bases with BQ ≥ 20 were discarded.

The trimmed and qualified reads were then mapped against the recently released near-

chromosome level *D. suzukii* genome assembly Dsuz-WT3_v2.0 that covers autosomes and the X

chromosome (Paris et al. 2020) using *bwa mem* (Li 2013). Reads with a mapping quality below 20

were then removed using *Samtools* (Li et al. 2009). We used Picard's *SortSam* to sort BAM files,

and used Picard's *MarkDuplicates* to mark PCR duplicates to avoid false variant calls

(http://broadinstitute.github.io/picard). Indel identification and realignment around indels were

performed using GATK's *RealignerTargetCreator* and *IndelRealigner* (Auwera and O'Connor 2020).

Finally, alignments in BAM format were checked for formatting errors using Picard's

ValidateSamFile. Summary statistics for quality checking of BAM files were generated using *bamdst*

(https://github.com/shiquan/bamdst).

To call SNPs, we merged the quality-checked BAM files of all population samples into

one file using Samtools *mpileup*, only retaining alignments with mapping quality no less than 20

and sites with base quality no less than 20. Variant calling was then performed on the mpileup

file using the heuristic SNP caller *PoolSNP* (Kapun et al. 2020). We used a nominally low value

for the parameter miss-frac (0.001) to require for each population sample individually, that depth

of coverage at a given site be 12 or greater (min-cov = 12), and that this site not be in the top 1%

of sites genome-wide for depth of coverage (max-cov = 0.99; calculated separately for each

population and for autosomal and X-linked contigs), in order to filter sites subject to copy

number variation. In the initial data set used for analysis of genome-wide diversity, we avoided

potential biases from allele frequency filters by using min-count = 1 and min-freq = 0. We

termed the resulting high-quality sites as 'analyzed sites' for brevity.

Identifying Autosomal and X-linked Contigs

We chose to perform all population genomic analyses and whole-genome scans separately

for SNPs from autosomes and the X chromosome for the following reasons: 1) autosomal and X-

linked variants have different allelic sample sizes as samples were obtained from both male and

female flies; 2) autosomes and the X chromosome could reflect different demographic histories

and outcomes of natural selection, *e.g.*, the lower effective population size of the X chromosome

than autosomes could lead to a higher impact of bottlenecks and selection on genomic diversity;

3) unbalanced sex ratios and male-biased dispersal could further differentiate autosomal versus X

chromosome variation (Clemente et al. 2018; Olazcuaga et al. 2020).

Since the assembly of *D. suzukii* reference genome is still at the contig level, chromosomal

identities of each contig are needed to perform separate analyses. However, 497 contigs that

represent ~43% of the assembly length have not been unambiguously mapped onto chromosome

arms of the *D. melanogaster* dm6 genome assembly. Although 264 of the unplaced contigs had

been assigned to autosomes and the X chromosome based on a female-to-male read depth ratio,

233 contigs that represent ~5% of the genome remained unassigned due to the lack of statistical

power (Paris et al. 2020).

Given our interest in accurately analyzing a larger proportion of the euchromatic

genome, we identified ~70% of these 233 unassigned contigs as autosomal and X-linked based

on the correlation between the mean read depth of each contig (among population samples) and

that across unambiguously aligned autosomal or X-linked contigs. We chose Spearman's rank

correlation instead of the Pearson correlation, as the distribution of depth data failed the

assumption of bivariate normality. A contig that has mean depth significantly correlated with

that of either known autosomal or X-linked contigs was assigned to the chromosome with a

higher correlation coefficient. Our method completely confirmed all prior mapping-based

assignments and had a ~96% consistency with the previous assignment based on female-to-male

read depth ratios. Inconsistent assignments for four contigs were corrected according to our

method (Table S2). The eight previously-assigned contigs that could not be assigned using our

method, as well as other unassigned contigs using all methods (totalling ~2.7 mb), were excluded

from downstream GEA analyses, because the assignment information is needed for estimating

effective sample size that were used to correct allele count data as an input to GEA.

Annotating Genomic Features and Estimating Divergence

To explore genomic diversity at synonymous sites and selective constraint for other site

types in *D. suzukii*, we classified the reference genome into nine exclusive categories of site

degeneracy and function (Lange and Pool 2018), including: non-degenerate (i.e.,

nonsynonymous) sites, two-, three-, and four-fold degenerate (i.e., synonymous) sites, 3' and 5'

UTRs, RNA-coding genes, introns, and intergenic regions. From input files including the

eukaryotic codon table, the published genome sequence and GFF3 annotation obtained at NCBI

RefSeq, we generated a letter-coded annotation (in FASTA format) mirroring both strands of the

whole genome sequence of *D. suzukii* and a coordinate-based annotation (in BED format) that

combines adjacent sites of the same category into a single row. Degeneracy was determined

based on the standard codon table. 5' UTRs were defined as regions between the start of the

first exon and the start of the first coding sequence (CDS), while 3' UTRs were defined as regions

between the end of the last exon and the end of the last CDS. In cases of overlapping genes and

alternative splicing that raise annotation conflicts, we followed an annotation priority in the

category order listed above.

We then estimated the divergence between *D. suzukii* and its close relative *D. biarmipes* in

each of these categories (Suvorov et al. 2022). We obtained results of multiple sequence

alignment between the current reference genomes of *D. suzukii* and *D. biarmipes* (Paris et al. 2020).

For each site category of *D. suzukii*, the unpolarized divergence was estimated as the number of

substitutions over the total number of sites within aligned blocks of reference genome sequences.

Estimating Nucleotide Diversity, F_{ST} D_{XY}

To compare genome-wide polymorphism among populations, we estimated nucleotide

diversity (π) across SNPs at four-fold degenerate sites (π_S) in addition to that at all categories of

sites (π_A), as π_S estimation is relatively less affected by sequencing errors than nucleotide diversity

estimated from other site categories (due to a higher ratio of real variation to errors). To

calculate π for each population sample, we adopted an unbiased estimator of nucleotide diversity

($\hat{\theta}_\Pi$) based on heterozygosity (Π), which has been optimized for pool-seq data (Ferretti et al.

2013). Numerically,

$\quad \pi_S = \hat{\theta}_\Pi = \frac{n_c}{n_c-1}\frac{\Pi}{L} = \frac{n_c}{n_c-1}\frac{1}{L}\sum_l \frac{2}{n_r(l)(n_r(l)-1)} m_l(n_r(l) - m_l)$ (1)

$\qquad$ Here, L represents the total number of genome-wide analyzed sites. Of a given

population sample, $n_r(l)$ represents the read depth of the top two alleles at the lth site (i.e., SNP)

and m_l represents the minor allele count. n_c as a normalization factor represents the haploid

sample size for either autosomes or X chromosome in a pool (Table S1). Strictly speaking, n_c as

a normalization factor should represent equally contributing chromosomes in a pool.

Nevertheless, for our data it is sufficient to use haploid sample size for either autosomes or X

chromosome to approximate n_c in the above equation, as the estimation of $\hat{\theta}_\Pi$ is not substantially

affected by the precise value of n_c when the number of individuals in the pool is large. The

above formula is a simplified version for SNP data, based on equation 3 in Ferretti et al. (2013).

$\qquad$ To examine patterns of polymorphism across chromosome arms, we also estimated

window nucleotide diversity (π_W) for all polymorphic sites. Each window was defined as a

continuous genomic region that includes 125,000 analyzed sites (Figure 3). Since chromosomal

identity was required in this analysis, we only took windows from 32 major contigs that contain

at least one full-size window and were unambiguously mappable to a chromosome arm of the *D.*

melanogaster dm6 genome assembly. Although such contigs only make up 57% of the *D. suzukii*

genome assembly, they contain a relatively larger proportion of all identified SNPs (72%), and

thus are still representative of genome-wide polymorphism.

$\qquad$ To estimate genome-wide pairwise F_{ST} between populations, we adopted an unbiased

multi-loci estimator known as Reynolds' estimator of the coancestry coefficient, which accounts

for unequal sample sizes among populations and is applicable for more than two alleles at a site

(Reynolds et al. 1983). We first heuristically partitioned the genome into windows that exceeded

a cross-sample average accumulated heterozygosity threshold of 100. The genome-wide F_{ST} was

then calculated as an average of window-F_{ST} $F_{ST}(w)$ weighted by the number of analyzed sites

within each window $(L(w))$, where $F_{ST}(w)$ was calculated as a weighted average of single-site

ratio estimators. Numerically,

$F_{ST} = \frac{1}{L}\sum_w L(w)\, F_{ST}(w)$ (2)

$F_{ST}(w) = (\sum_l a_l)/\sum_l (a_l + b_l)$ (3)

where

$a_l = \frac{1}{2}\sum_u (\tilde{p}_{1lu} - \tilde{p}_{2lu})^2 - \frac{(n_{1l}+n_{2l})(n_{1l}\tilde{\alpha}_{1l}+n_{2l}\tilde{\alpha}_{2l})}{4n_{1l}n_{2l}(n_{1l}+n_{2l}-1)}$ (4)

$a_l + b_l = \frac{1}{2}\sum_u (\tilde{p}_{1lu} - \tilde{p}_{2lu})^2 + \frac{(4n_{1l}n_{2l}-n_{1l}-n_{2l})(n_{1l}\tilde{\alpha}_{1l}+n_{2l}\tilde{\alpha}_{2l})}{4n_{1l}n_{2l}(n_{1l}+n_{2l}-1)}$ (5)

At the lth site in each population, $\tilde{p}_{1lu}$ and $\tilde{p}_{2lu}$ represent the frequency of the uth allele at the

lth site; $\tilde{\alpha}_{1l}$ and $\tilde{\alpha}_{2l}$ represent the heterozygosity; n_{1l} and n_{2l} represent the sample size. Unlike

the sequencing of individual genomes, pool-seq induces an uncertainty in the number of

individual alleles actually sequenced at a locus (i.e., effective sample size), and this uncertainty

decreases slowly even at high read depth (Ferretti et al. 2013). Since the sample size is an

important parameter for F_{ST} estimation, we took measures to obtain an estimate of the effective

sample size, n_{il}, at each given site (Ferretti et al. 2013). Numerically,

$n_{il} = \sum_j^{n_c} j P_c(j \mid n_r, n_c)$ (6)

where

$P_c(j \mid n_r, n_c) = \frac{n_c! S(n_r, j)}{(n_c-j)!\, n_c^{n_r}}$ (7)

Here, we explicitly estimated the probability of the number of j unique lineages sampled at a site

given n_r sampled reads and n_c equally contributing chromosomes in a pool, where $S(n_r, j)$ are

the Stirling numbers of the second kind, defined as the number of ways to partition n_r reads into

j non-empty sets (Ferretti et al. 2013). We then estimated n_{il} as the expected number of lineages

for each n_r and n_c. Ideally, n_c should be estimated as $2n_e$, where n_e is the effective pool size

representing the number of diploid individuals contributing the same amount of reads to a pool

(Gautier et al. 2013; Lange et al. 2022). Although we lack sample replicates to estimate n_e and

therefore used haploid sample size for n_c as an approximation, the probability estimation is still

reasonable given that our number of lineages for each pool is large (Table S1) (Ferretti et al.

2013).

Lastly, we estimated genome-wide pairwise D_{XY} as an absolute measure of population

differentiation that is independent of levels of within-population diversity. It was calculated as

pairwise differences per site between two populations, divided by L total analyzed sites (Nei 1987;

Hahn 2018). Numerically,

$D_{XY} = \frac{1}{L}\sum_l \sum_{ij} x_i y_j k_{ij}$ (8)

where x_i and y_j represent frequencies of the ith allele from population X and the jth allele from

population Y, and k_{ij} is either 1 or 0, depending on whether or not the alleles differ at the lth

site. Calculations in this section were all implemented with Python and Shell scripts (see Data

Availability).

Building Population Trees and Admixture Graphs

836   To infer the population structure assuming no migrations, we used MEGA X (Stecher et

837 al. 2020) to build neighbor-joining population trees (Figure S5) for autosomes and X

838 chromosome from Reynolds' distance, which is a transformation of pairwise $F_{ST} : D = -ln\,(1 -$

839 $F_{ST})$ (Reynolds et al. 1983). Since the position of the tree root is not identifiable through this

840 analysis, and the China-Ningbo population was identified through diversity and distance analyses

841 as a potentially basal lineage among the sampled populations, we rooted the tree along the

842 branch to Ningbo.

843   To further investigate population splits and mixtures, we used TreeMix (Pickrell and

844 Pritchard 2012) to infer population trees using allele counts that were corrected based on

845 effective sample size. The input SNPs were a subset of our total identified SNPs whose total

846 minor allele count across all samples was no less than ten, as it would allow us to complete this

847 computationally intensive analysis in a reasonable amount of time and avoid biases on tree

848 construction from false positive SNPs. We combined SNPs into blocks of 54 (-k 54) to account

849 for linkage disequilibrium across windows that have a median length of ~500 bp.

850   To determine the number of migration events (-m) that improved the model fit while

851 avoiding overfitting and losing interpretability, we ran TreeMix on our actual SNP data with no

852 migration and with -m from one to 20. Since the position of the root is only partially identifiable

853 through TreeMix inference, we rooted the tree at China-Ningbo, as suggested by Pickrell &

854 Pritchard (2012). We then estimated the fraction of the variance in relatedness between

855 populations that is accounted for by each model (f). We found that f began to plateau for both

856 the autosomal and X-linked models at m = 11. For the X chromosome model, we also

857 confirmed that all migration edges were statistically significant (p < 0.05) based on calculating

858 their p-values from the jackknife estimates of the migration weights and their standard error

(Pickrell and Pritchard 2012), whereas this analysis remained computationally intractable for the

autosomal case. We interpreted the migration weights *w* as an admixture proportion, as it was

correlated with ancestry fraction in previous simulations, although it tends to be a more

conservative approximation for high admixture proportions (Pickrell and Pritchard 2012).

Preparing Environmental Data

To generate environmental data for GEA, we selected a preliminary set of 26 candidate

environmental variables representing geographic, climatic and land cover-related factors (Figure

4A) that may be relevant in the adaptation process of *D. suzukii* based on prior knowledge

(Kellermann et al. 2012; Bogaerts-Márquez et al. 2021). With R packages 'raster' (v. 3.5.2) and

'SpatialPoints' (v. 1.4.6), we retrieved environmental data of high spatial resolution ($\sim$100 km^2)

in batch for the sampling locations of our 29 populations from online databases WorldClim (Fick

and Hijmans 2017) and Esri 2020 Land Cover (Karra et al. 2021). Annual mean values of

monthly climatic variables, including mean wind speed, solar radiation, and water vapor

pressure, were derived by averaging across 12 months of data.

Due to the large number of statistical tests that would result from running GEA on all the

environmental variables one by one, there is an increased difficulty in controlling rates of false

discovery. Additionally, including multiple highly correlated variables in a model would lead to

multicollinearity issues (Rellstab et al. 2015). To avoid these problems, we calculated a pairwise

Pearson correlation matrix from values of environmental factors across sampled locations (Figure

4A; Table S3), and then selected a subset of nine least correlated environmental variables for

one-by-one GEA analyses (Figure 4B). To avoid scale inconsistencies between estimated GEA

statistics, the environmental differentiation of each population was calculated as the absolute

difference between the environmental value of that population and the average across all

populations, standardized by the standard deviation (de Villemereuil and Gaggiotti 2015). This

standardized differentiation was then input to GEA (Table S3).

Environmental Association Analyses

To characterize the environmental adaptation of *D. suzukii*, we scanned the whole

genome for adaptive loci using the F_{ST}-based GEA method BayesScEnv (de Villemereuil and

Gaggiotti 2015). We chose this specific approach over other GEA methods mainly because it has

a lower false positive rate than other GEA approaches in the presence of hierarchical population

structure. It also allows for detecting patterns of allele frequency that are not linearly dependent

on environmental factors (Rellstab et al. 2015; de Villemereuil and Gaggiotti 2015).

For each environmental variable, the association analyses tested the relationship between

environmental and genetic differentiation among populations, for 5,752,156 genome-wide SNPs

with a MAF higher than 5%. To control for false positives, we chose stringent model parameters

expected to yield extremely conservative results, setting the prior probability of non-neutral

models as 0.02 (-pr_jump 0.02) and the prior probability of the competing environment-

unrelated locus-specific model as 0.9 (-pr_pref 0.9). These parameters correspond to assumptions

that genetic differentiation reflects the action of natural selection in just 2% of the genome, and

the focal environmental variable is only expected to be involved at 10% of the non-neutral loci.

To make this GEA analysis computationally feasible with our large SNP set, while still

analyzing all qualifying SNPs, we applied a split-run strategy: we subsampled SNPs across

concatenated sequences of contigs within the autosomes and the X chromosome separately, and

then ran subsamples with BayesScEnv in parallel. Since the null model of population structure is

estimated separately in each run, we subsampled non-adjacent SNPs at a fixed interval to limit

locus-specific biases in that estimation, where the length of the interval between jointly analyzed

SNPs was equal to the total number of subsamples. With a targeted subsample/interval size of

up to 10,000 SNPs, we divided the concatenated autosomal contigs into 490 subsamples (with

actual subsample sizes of 9982-9983 SNPs), and the concatenated X-linked contigs into 87

subsamples (with actual subsample sizes of 9893-9894 SNPs). Hence, the first autosomal

subsample contained SNP #1, SNP #491, and so on.

Convergence of each run was confirmed with the R package 'CODA'. Individual runs

were then merged across autosomes and X chromosome to calculate the genome-wide q-value

(q) of locally-estimated posterior error probability (PEP) across all sites, where we targeted a false

discovery rate (FDR) of 5% by setting the q threshold at 0.05 (Storey 2003; Muller et al. 2006).

For downstream analyses, to remove redundancy due to linkage disequilibrium, we pared down

closely linked candidate sites by maintaining the site with the lowest q within each 20 kb genomic

window. To assess the relative levels of support for associations between SNPs and a given

environmental variable, we ranked all candidate loci first by q and then by the estimated g

parameter as a tie-breaker, which measures the sensitivity of a locus to environmental

differentiation.

Identifying Candidate Genes

For each candidate SNP, the closest gene in each direction within a 200-exon flanking

region that overlapped with it was considered to be associated with that variant, in order to

encompass both potential coding and regulatory adaptation. To facilitate clear comparisons

among environmental variables with different numbers of significant variants, we focused on the

top 500 candidate genes that were linked to variants with the lowest significant q and highest g

within each environmental variable (Table S5).

GO Enrichment and Semantic Clustering

Gene ontology (GO) enrichment of the top 500 candidate genes associated with

candidate SNPs was performed via genomic permutation of outlier SNP positions (100,000,000

replicates), which accounts for the variability of gene length and the clustering of functionally

related genes, as described in previous work (Pool et al. 2017). For each GO category, a p-value

indicated the proportion of permutation replicates in which an equal or greater number of genes

was implicated.

We then prioritized the most informative and significant GO terms and removed

redundant terms that potentially share similar groups of genes by clustering GO terms based on

their semantic similarity and ranking representative terms of each cluster by their p-value

(Reijnders and Waterhouse 2021). For GO terms that were shared among associations with

multiple environmental variables, a combined p-value was calculated from the p-values of

independent enrichment tests using Fisher's method (Fisher 1938).

Supplementary Material

Supplementary figures S1−S9 and tables S1−S6 are available at Molecular Biology and

Evolution online.

Data Availability

All sequence data generated for this project are available from the NIH Short Read

Archive under project PRJNA973110, with specific sample information given in Table S1,

Supplementary Material online. All computational scripts created for this study have been

uploaded to https://github.com/Sfeng666/poolWGS2SNP and

https://github.com/Sfeng666/Dsuz_popgen_GEA (last accessed May 27, 2023).

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References

Adrion JR, Kousathanas A, Pascual M, Burrack HJ, Haddad NM, Bergland AO, Machado H, Sackton TB, Schlenke TA, Watada M, et al. 2014. *Drosophila suzukii*: the genetic footprint of a recent, worldwide invasion. *Mol Biol Evol* 31:3148–3163.

Alloway PG, Dolph PJ. 1999. A role for the light-dependent phosphorylation of visual arrestin. *Proc Natl Acad Sci USA* 96:6072–6077.

Asplen MK, Anfora G, Biondi A, Choi D-S, Chu D, Daane KM, Gibert P, Gutierrez AP, Hoelmer KA, Hutchison WD, et al. 2015. Invasion biology of spotted wing *Drosophila* (*Drosophila suzukii*): a global perspective and future priorities. *J Pest Sci* 88:469–494.

Atallah J, Teixeira L, Salazar R, Zaragoza G, Kopp A. 2014. The making of a pest: the evolution of a fruit-penetrating ovipositor in *Drosophila suzukii* and related species. *Proc Roy Soc B* 281:20132840.

Auwera GAV der, O'Connor BD. 2020. Genomics in the cloud: using Docker, GATK, and WDL in Terra. O'Reilly Media.

Avidor-Reiss T, Leroux MR. 2015. Shared and distinct mechanisms of compartmentalized and cytosolic ciliogenesis. *Curr Biol* 25:R1143–R1150.

Avidor-Reiss T, Maer AM, Koundakjian E, Polyanovsky A, Keil T, Subramaniam S, Zuker CS. 2004. Decoding Cilia Function: Defining specialized genes required for compartmentalized cilia biogenesis. *Cell* 117:527–539.

Barange M, Cheung WWL, Merino G, Perry RI. 2010. Modelling the potential impacts of climate change and human activities on the sustainability of marine resources. *Curr Opin Environ Sustain* 2:326–333.

Batie M, Druker J, D'Ignazio L, Rocha S. 2017. KDM2 family members are regulated by HIF-1 in hypoxia. *Cells* 6:8.

Begun DJ, Holloway AK, Stevens K, Hillier LW, Poh Y-P, Hahn MW, Nista PM, Jones CD, Kern AD, Dewey CN, et al. 2007. Population genomics: whole-genome analysis of polymorphism and divergence in *Drosophila simulans*. *PLOS Biol* 5:e310.

Behrman EL, Watson SS, O'Brien KR, Heschel MS, Schmidt PS. 2015. Seasonal variation in life history traits in two *Drosophila* species. *J Evol Biol* 28:1691–1704.

Bogaerts-Márquez M, Guirao-Rico S, Gautier M, González J. 2021. Temperature, rainfall and wind variables underlie environmental adaptation in natural populations of *Drosophila melanogaster*. *Mol Ecol* 30:938–954.

Boutros M, Ahringer J. 2008. The art and design of genetic screens: RNA interference. *Nat Rev Genet* 9:554–566.

Calboli FCF, Gilchrist GW, Partridge L. 2003. Different cell size and cell number contribution in
two newly established and one ancient body size cline of *Drosophila subobscura*. *Evolution*
57:566–573.

Campbell M, Ganetzky B. 2012. Extensive morphological divergence and rapid evolution of the
larval neuromuscular junction in *Drosophila*. *Proc Natl Acad Sci USA* 109:E648–E655.

Charlesworth B, Morgan MT, Charlesworth D. 1993. The effect of deleterious mutations on
neutral molecular variation. *Genetics*. 34:1289-1303.

Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor.
Bioinformatics 34:i884–i890.

Cini A, Ioriatti C, Anfora G. 2012. A review of the invasion of *Drosophila suzukii* in Europe and a
draft research agenda for integrated pest management. *Bull Insectology* 65:149–160.

Clark MS, Thorne MA, Purać J, Burns G, Hillyard G, Popović ŽD, Grubor-Lajšić G, Worland
MR. 2009. Surviving the cold: molecular analyses of insect cryoprotective dehydration in
the Arctic springtail *Megaphorura arctica* (Tullberg). *BMC Genomics* 10:328.

Clemente F, Gautier M, Vitalis R. 2018. Inferring sex-specific demographic history from SNP
data. *PLOS Genet* 14:e1007191.

Colautti RI, Barrett SCH. 2013. Rapid adaptation to climate facilitates range expansion of an
invasive plant. *Science* 342:364–366.

Colautti RI, Lau JA. 2016. Contemporary evolution during invasion. In: Invasion Genetics. John
Wiley & Sons, Ltd. p. 101–121.

Duvall LB, Taghert PH. 2013. E and M circadian pacemaker neurons use different PDF
receptor signalosome components in drosophila. *J Biol Rhythms* 28:239–248.

Elaine Merrill C, Sherertz TM, Walker WB, Zwiebel LJ. 2005. Odorant-specific requirements
for arrestin function in *Drosophila* olfaction. *J Neurobiol* 63:15–28.

Enriquez T, Colinet H. 2019. Cold acclimation triggers lipidomic and metabolic adjustments in
the spotted wing drosophila *Drosophila suzukii* (Matsumara). *Am J Physiol Regul Integr Comp*
Physiol 316:R751–R763.

Ferretti L, Ramos-Onsins SE, Pérez-Enciso M. 2013. Population genomics from pool
sequencing. *Mol Ecol* 22:5561–5576.

Fick SE, Hijmans RJ. 2017. WorldClim 2: new 1-km spatial resolution climate surfaces for global
land areas. *Int J Climatol* 37:4302–4315.

Fisher RA. 1938. Statistical methods for research workers. Oliver and Boyd, Edinburgh.

Fraimout A, Debat V, Fellous S, Hufbauer RA, Foucaud J, Pudlo P, Marin J-M, Price DK,
1034   Cattel J, Chen X, et al. 2017. Deciphering the routes of invasion of *Drosophila suzukii* by
1035   means of ABC random forest. *Mol Biol Evol* 34:980–996.

1036   Gautier M, Foucaud J, Gharbi K, Cézard T, Galan M, Loiseau A, Thomson M, Pudlo P,
1037   Kerdelhué C, Estoup A. 2013. Estimation of population allele frequencies from next-
1038   generation sequencing data: pool-versus individual-based genotyping. *Mol Ecol* 22:3766–
1039   3779.

1040   Gautier M, Vitalis R, Flori L, Estoup A. 2022. f-Statistics estimation and admixture graph
1041   construction with Pool-Seq or allele count data using the R package poolfstat. *Mol Ecol*
1042   *Resour* 22:1394–1416.

1043   Gibbs AG, Fukuzato F, Matzkin LM. 2003. Evolution of water conservation mechanisms in
1044   Drosophila. *J Exp Biol* 206:1183–1192.

1045   Gibert P, Debat V, Ghalambor CK. 2019. Phenotypic plasticity, global change, and the speed of
1046   adaptive evolution. *Curr Op Insect Sci* 35:34–40.

1047   Gilchrist AS, Partridge L. 1999. A comparison of the genetic basis of wing size divergence in
1048   three parallel body size clines of *Drosophila melanogaster*. *Genetics* 153:1775–1787.

1049   Guha A, Kornberg TB. 2005. Tracheal branch repopulation precedes induction of the
1050   Drosophila dorsal air sac primordium. *Devel Biol* 287:192–200.

1051   Hahn MW. 2018. Molecular population genetics. Sunderland, MA: Sinauer Associates.

1052   Hämälä T, Gorton AJ, Moeller DA, Tiffin P. 2020. Pleiotropy facilitates local adaptation to
1053   distant optima in common ragweed (Ambrosia artemisiifolia). *PLOS Genet* 16:e1008707.

1054   Hamby KA, E. Bellamy D, Chiu JC, Lee JC, Walton VM, Wiman NG, York RM, Biondi A.
1055   2016. Biotic and abiotic factors impacting development, behavior, phenology, and
1056   reproductive biology of *Drosophila suzukii*. *J Pest Sci* 89:605–619.

1057   Han Z, Li X, Wu J, Olson EN. 2004. A myocardin-related transcription factor regulates activity
1058   of serum response factor in *Drosophila*. *Proc Natl Acad Sci USA* 101:12567–12572.

1059   Helfrich-Förster C, Bertolini E, Menegazzi P. 2020. Flies as models for circadian clock
1060   adaptation to environmental challenges. *Eur J NeuroSci* 51:166–181.

1061   Hill T, Koseva BS, Unckless RL. 2019. The genome of *Drosophila innubila* reveals lineage-specific
1062   patterns of selection in immune genes. *Mol Biol Evol* 36:1405–1417.

1063   Hodgins KA, Bock DG, Rieseberg LH. 2018. Trait evolution in invasive species. In: Annual
1064   Plant Reviews online. New York: John Wiley & Sons, Ltd. p. 459–496.

1065   Honjo K, Mauthner SE, Wang Y, Skene JHP, Tracey WD. 2016. Nociceptor-enriched genes
1066   required for normal thermal nociception. *Cell Rep* 16:295–303.

Horváth V, Guirao-Rico S, Salces-Ortiz J, Rech GE, Green L, Aprea E, Rodeghiero M, Anfora
G, González J. 2023. Gene expression differences consistent with water loss reduction
underlie desiccation tolerance of natural *Drosophila* populations. *BMC Biol* 21:35.

James RE, Broihier HT. 2011. Crimpy inhibits the BMP homolog Gbb in motoneurons to
enable proper growth control at the *Drosophila* neuromuscular junction. *Development*
138:3273–3286.

Kapun M, Barrón MG, Staubach F, Obbard DJ, Wiberg RAW, Vieira J, Goubert C, Rota-
Stabelli O, Kankare M, Bogaerts-Márquez M, et al. 2020. Genomic analysis of European
Drosophila melanogaster populations reveals longitudinal structure, continent-wide selection,
and previously unknown DNA viruses. *Mol Biol Evol* 37:2661–2678.

Karra K, Kontgis C, Statman-Weil Z, Mazzariello JC, Mathis M, Brumby SP. 2021. Global
land use / land cover with Sentinel 2 and deep learning. In: 2021 IEEE International
Geoscience and Remote Sensing Symposium IGARSS. p. 4704–4707.

Kawecki TJ, Ebert D. 2004. Conceptual issues in local adaptation. *Ecol Let* 7:1225–1241.

Kellermann V, Loeschcke V, Hoffmann AA, Kristensen TN, Fløjgaard C, David JR, Svenning J,
Overgaard J. 2012. Phylogenetic constraints in key functional traits behind species'
climate niches: patterns of desiccation and cold resistance across 95 *Drosophila* species.
Evolution 66:3377–3389.

Kerekes É, Kókai E, Páldy FS, Dombrádi V. 2014. Functional analysis of the glycogen binding
subunit CG9238/Gbs-70E of protein phosphatase 1 in *Drosophila melanogaster*. *Insect*
Biochem Mol Biol 49:70–79.

Kinsler G, Geiler-Samerotte K, Petrov DA. 2020. Fitness variation across subtle environmental
perturbations reveals local modularity and global pleiotropy of adaptation. *eLife* 9:e61271.

Knapp L, Mazzi D, Finger R. 2021. The economic impact of *Drosophila suzukii*: perceived costs
and revenue losses of Swiss cherry, plum and grape growers. *Pest Manag Sci* 77:978–1000.

Koch JB, Dupuis JR, Jardeleza M-K, Ouedraogo N, Geib SM, Follett PA, Price DK. 2020.
Population genomic and phenotype diversity of invasive *Drosophila suzukii* in Hawaiʻi. *Biol*
Invasions 22:1753–1770.

Lack JB, Lange JD, Tang AD, Corbett-Detig RB, Pool JE. 2016. A thousand fly genomes: an
expanded *Drosophila* Genome Nexus. *Mol Biol Evol* 33:3308–3313.

Lack JB, Yassin A, Sprengelmeyer QD, Johanning EJ, David JR, Pool JE. 2016. Life history
evolution and cellular mechanisms associated with increased size in high-altitude
Drosophila. *Ecol Evol* 6:5893–5906.

Lange JD, Bastide H, Lack JB, Pool JE. 2022. A population genomic assessment of three decades
of evolution in a natural *Drosophila* population. *Mol Biol Evol* 39:msab368.

Lange JD, Pool JE. 2018. Impacts of Recurrent Hitchhiking on Divergence and Demographic
Inference in *Drosophila*. *Genome Biol Evol* 10:1882–1891.

Langley CH, Stevens K, Cardeno C, Lee YCG, Schrider DR, Pool JE, Langley SA, Suarez C,
Corbett-Detig RB, Kolaczkowski B, et al. 2012. Genomic variation in natural populations
of *Drosophila melanogaster*. *Genetics* 192:533–598.

Lee CE. 2002. Evolutionary genetics of invasive species. *Trends Ecol Evol* 17:386–391.

Levashina EA, Ohresser S, Bulet P, Reichhart J-M, Hetru C, Hoffmann JA. 1995.
Metchnikowin, a novel immune-inducible proline-rich peptide from *Drosophila* with
antibacterial and antifungal properties. *Eur J Biochem* 233:694–700.

Lewald KM, Abrieux A, Wilson DA, Lee Y, Conner WR, Andreazza F, Beers EH, Burrack HJ,
Daane KM, Diepenbrock L, et al. 2021. Population genomics of *Drosophila suzukii* reveal
longitudinal population structure and signals of migrations in and out of the continental
United States. *G3* 11:jkab343.

Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM,
http://arxiv.org/abs/1303.3997, last accessed May 31, 2023.

Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R,
1000 genome project data processing subgroup. 2009. The Sequence Alignment/Map
format and SAMtools. *Bioinformatics* 25:2078–2079.

Li W, Wang F, Menut L, Gao F-B. 2004. BTB/POZ-zinc finger protein abrupt suppresses
dendritic branching in a neuronal subtype-specific and dosage-dependent manner. *Neuron*
43:823–834.

Little CM, Chapman TW, Hillier NK. 2020. Plasticity is key to success of *Drosophila suzukii*
(Diptera: Drosophilidae) invasion. *J Insect Sci* 20:5.

Manko P, de Figueroa JMT, Oboňa J. 2021. Responses of the pest *Drosophila suzukii* population
to environmental variables in southern Spain (Granada, Andalusia) – results of a pilot
study. *Acta Mus Sil Sci Nat* 70:65–74.

Maynard Smith J, Haigh J. 1974. The hitch-hiking effect of a favourable gene. *Genet Res*. 23:23-
35.

Mérel V, Gibert P, Buch I, Rodriguez Rada V, Estoup A, Gautier M, Fablet M, Boulesteix M,
Vieira C. 2021. The worldwide invasion of *Drosophila suzukii* is accompanied by a large
increase of transposable element load and a small number of putatively adaptive
insertions. *Mol Biol Evol* 38:4252–4267.

Muller P, Parmigiani G, Rice K. 2006. FDR and Bayesian multiple comparisons rules. *JHU*
Biostatistics 115.

Nakabayashi R, Morishita S. 2020. HiC-Hiker: a probabilistic model to determine contig
orientation in chromosome-length scaffolds with Hi-C. *Bioinformatics* 36:3966–3974.

Nei M. 1987. Molecular evolutionary genetics. New York: Columbia University Press.

Olazcuaga L, Loiseau A, Parrinello H, Paris M, Fraimout A, Guedot C, Diepenbrock LM, Kenis
M, Zhang J, Chen X, et al. 2020. A whole-genome scan for association with invasion
success in the fruit fly *Drosophila suzukii* using contrasts of allele frequencies corrected for
population structure. *Mol Biol Evol* 37:2369–2385.

Ometto L, Cestaro A, Ramasamy S, Grassi A, Revadi S, Siozios S, Moretto M, Fontana P,
Varotto C, Pisani D, et al. 2013. Linking genomics and ecology to investigate the
complex evolution of an invasive *Drosophila* pest. *Genome Biol Evol* 5:745–757.

Palacios-Muñoz A, Ewer J. 2018. Calcium and cAMP directly modulate the speed of the
Drosophila circadian clock. *PLOS Genet* 14:e1007433.

Paris M, Boyer R, Jaenichen R, Wolf J, Karageorgi M, Green J, Cagnon M, Parinello H, Estoup
A, Gautier M, et al. 2020. Near-chromosome level genome assembly of the fruit pest
Drosophila suzukii using long-read sequencing. *Sci Rep* 10:11227.

Pickrell JK, Pritchard JK. 2012. Inference of population splits and mixtures from genome-wide
allele frequency data. *PLOS Genet* 8:e1002967.

Pool JE. 2015. The mosaic ancestry of the *Drosophila* Genetic Reference Panel and the *D.*
melanogaster reference genome reveals a network of epistatic fitness interactions. *Mol Biol*
Evol 32:3236–3251.

Pool JE, Braun DT, Lack JB. 2017. Parallel evolution of cold tolerance within *Drosophila*
melanogaster. *Mol Biol Evol* 34:349–360.

Pool JE, Corbett-Detig RB, Sugino RP, Stevens KA, Cardeno CM, Crepeau MW, Duchen P,
Emerson JJ, Saelao P, Begun DJ, et al. 2012. Population genomics of sub-Saharan
Drosophila melanogaster: African diversity and non-African admixture. *PLOS Genetics*
8:e1003080.

Pool JE, Nielsen R. 2007. Population size changes reshape genomic patterns of diversity. *Evolution*
61:3001–3006.

Pool JE, Nielsen R. 2009. Inference of historical changes in migration rate from the lengths of
migrant tracts. *Genetics* 181:711–719.

Prakash S, Caldwell JC, Eberl DF, Clandinin TR. 2005. *Drosophila* N-cadherin mediates an
attractive interaction between photoreceptor axons and their targets. *Nat Neurosci* 8:443–
450.

Prelich G. 2012. Gene overexpression: uses, mechanisms, and interpretation. *Genetics* 190:841–
854.

Prentis PJ, Wilson JRU, Dormontt EE, Richardson DM, Lowe AJ. 2008. Adaptive evolution in
invasive species. *Trends Plant Sci* 13:288–294.

Pudlo P, Marin J-M, Estoup A, Cornuet J-M, Gautier M, Robert CP. 2016. Reliable ABC model
choice via random forests. *Bioinformatics* 32:859–866.

Ray N, Adams JM. 2001. A GIS-based vegetation map of the world at the last glacial maximum
(25,000-15,000 BP). *Internet Archaeol* 11.

Reijnders MJMF, Waterhouse RM. 2021. Summary visualizations of Gene Ontology terms with
GO-Figure! *Front Bioinform* 1:6.

Rellstab C, Gugerli F, Eckert AJ, Hancock AM, Holderegger R. 2015. A practical guide to
environmental association analysis in landscape genomics. *Mol Ecol* 24:4348–4370.

Reynolds J, Weir BS, Cockerham CC. 1983. Estimation of the coancestry coefficient: basis for a
short-term genetic distance. *Genetics* 105:767–779.

Rezende GL, Martins AJ, Gentile C, Farnesi LC, Pelajo-Machado M, Peixoto AA, Valle D.
2008. Embryonic desiccation resistance in *Aedes aegypti*: presumptive role of the chitinized
Serosal Cuticle. *BMC Devel Biol* 8:82.

Reznick DN, Losos J, Travis J. 2019. From low to high gear: there has been a paradigm shift in
our understanding of evolution. *Ecol Let* 22:233–244.

Rogers WA, Salomone JR, Tacy DJ, Camino EM, Davis KA, Rebeiz M, Williams TM. 2013.
Recurrent modification of a conserved cis-regulatory element underlies fruit fly
pigmentation diversity. *PLOS Genet* 9:e1003740.

Rudman SM, Greenblum SI, Rajpurohit S, Betancourt NJ, Hanna J, Tilk S, Yokoyama T,
Petrov DA, Schmidt P. 2022. Direct observation of adaptive tracking on ecological time
scales in *Drosophila*. *Science* 375:eabj7484.

Santos LA dos, Mendes MF, Krüger AP, Blauth ML, Gottschalk MS, Garcia FRM. 2017.
Global potential distribution of *Drosophila suzukii* (Diptera, Drosophilidae). *PLOS ONE*
12:e0174318.

Schrider DR, Shanku AG, Kern AD. 2016. Effects of linked selective sweeps on demographic
inference and model selection. *Genetics* 204:1207–1223.

Senthilan PR, Piepenbrock D, Ovezmyradov G, Nadrowski B, Bechstedt S, Pauls S, Winkler M,
Möbius W, Howard J, Göpfert MC. 2012. *Drosophila* auditory organ genes and genetic
hearing defects. *Cell* 150:1042–1054.

Shalem O, Sanjana NE, Zhang F. 2015. High-throughput functional genomics using CRISPR–
Cas9. *Nat Rev Genet* 16:299–311.

Shearer PW, West JD, Walton VM, Brown PH, Svetec N, Chiu JC. 2016. Seasonal cues induce
phenotypic plasticity of *Drosophila suzukii* to enhance winter survival. *BMC Ecol* 16:11.

Simoes da Silva CJ, Sospedra I, Aparicio R, Busturia A. 2019. The microRNA-306/abrupt
regulatory axis controls wing and haltere growth in *Drosophila*. *Mech Dev* 158:103555.

Stecher G, Tamura K, Kumar S. 2020. Molecular Evolutionary Genetics Analysis (MEGA) for
macOS. *Mol Biol Evol* 37:1237–1239.

Stern DL. 2014. Identification of loci that cause phenotypic variation in diverse species with the
reciprocal hemizygosity test. *Trends Genet* 30:547–554.

Storey JD. 2003. The positive false discovery rate: A Bayesian interpretation and the q-value. *Ann*
Stat 31:2013–2035.

Sugimura K, Satoh D, Estes P, Crews S, Uemura T. 2004. Development of Morphological
Diversity of Dendrites in *Drosophila* by the BTB-Zinc Finger Protein Abrupt. *Neuron*
43:809–822.

Suvorov A, Kim BY, Wang J, Armstrong EE, Peede D, D'Agostino ERR, Price DK, Waddell PJ,
Lang M, Courtier-Orgogozo V, et al. 2022. Widespread introgression across a phylogeny
of 155 *Drosophila* genomes. *Curr Biol* 32:111-123.e5.

Svedberg J, Shchur V, Reinman S, Nielsen R, Corbett-Detig R. 2021. Inferring adaptive
introgression using hidden Markov models. *Mol Biol Evol* 38:2152–2165.

Tait G, Grassi A, Pfab F, Crava CM, Dalton DT, Magarey R, Ometto L, Vezzulli S, Rossi-
Stacconi MV, Gottardello A, et al. 2018. Large-scale spatial dynamics of *Drosophila suzukii*
in Trentino, Italy. *J Pest Sci* 91:1213–1224.

Telonis-Scott M, Gane M, DeGaris S, Sgrò CM, Hoffmann AA. 2012. High resolution mapping
of candidate alleles for desiccation resistance in *Drosophila melanogaster* under selection. *Mol*
Biol Evol 29:1335–1351.

True JR, Mercer JM, Laurie CC. 1996. Differences in crossover frequency and distribution
among three sibling species of *Drosophila*. *Genetics* 142:507–523.

Turner TL. 2014. Fine-mapping natural alleles: quantitative complementation to the rescue. *Mol*
Ecol 23:2377–2382.

Ueno K, Kidokoro Y. 2008. Adenylyl cyclase encoded by AC78C participates in sugar
perception in *Drosophila melanogaster*. *Eur J Neurosci* 28:1956–1966.

de Villemereuil P, Gaggiotti OE. 2015. A new F_{ST}-based method to uncover local adaptation
using environmental variables. *Methods in Ecology and Evolution* 6:1248–1258.

Wallingford AK, Rice KB, Leskey TC, Loeb GM. 2018. Overwintering behavior of *drosophila*
suzukii, and potential springtime diets for egg maturation. *Environ Entomol* 47:1266–1273.

Wang S-H, Simcox A, Campbell G. 2000. Dual role for *Drosophila* epidermal growth factor
receptor signaling in early wing disc development. *Genes Dev* 14:2271–2276.

Wang T, Morency DT, Harris N, Davis GW. 2020. Epigenetic signaling in glia controls
presynaptic homeostatic plasticity. *Neuron* 105:491-505.e3.

Wang Y, Farine J-P, Yang Y, Yang J, Tang W, Gehring N, Ferveur J-F, Moussian B. 2020.
Transcriptional control of quality differences in the lipid-based cuticle barrier in *Drosophila*
suzukii and *Drosophila melanogaster*. *Front Genet* 11:887.

Wang Y, Ferveur J-F, Moussian B. 2021. Eco-genetics of desiccation resistance in *Drosophila*. *Biol*
Rev 96:1421–1440.

Weißinger L, Schrieber K, Breuer M, Müller C. 2019. Influences of blackberry margins on
population dynamics of *Drosophila suzukii* and grape infestation in adjacent vineyards. *J*
Appl Entomol 143:802–812.

Welles SR, Dlugosch KM. 2019. Population genomics of colonization and invasion. In: Rajora
OP, editor. Population genomics: concepts, approaches and applications. Population
Genomics. Cham: Springer International Publishing. p. 655–683.

Wong MM-K, Liu M-F, Chiu SK. 2015. Cropped, *Drosophila* transcription factor AP-4, controls
tracheal terminal branching and cell growth. *BMC Devel Biol* 15:20.