Integration of variant annotations using deep set networks boosts rare variant association genetics

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Abstract

Rare genetic variants can strongly predispose to disease, yet accounting for rare variants in genetic analyses is statistically challenging. While rich variant annotations hold the promise to enable well-powered rare variant association tests, methods integrating variant annotations in a data-driven manner are lacking. Here, we propose DeepRVAT, a set neural network-based approach to learn burden scores from rare variants, annotations and phenotypes. In contrast to existing methods, DeepRVAT yields a single, trait-agnostic, nonlinear gene impairment score, enabling both risk prediction and gene discovery in a unified framework. On 21 quantitative traits and whole-exome-sequencing data from UK Biobank, DeepRVAT offers substantial increases in gene discoveries and improved replication rates in held-out data. Moreover, we demonstrate that the integrative DeepRVAT gene impairment score greatly improves detection of individuals at high genetic risk. We show that pre-trained DeepRVAT scores generalize across traits, opening up the possibility to conduct highly computationally efficient rare variant tests.

Introduction

The recent arrival of population-scale whole-exome and whole-genome sequencing studies[1] vastly expands the potential to understand the genetic underpinnings of human traits. While genome-wide association studies (GWAS) on common variants have identified a compendium of trait-associated loci[2], mapping these largely noncoding variants to affected genes as well as addressing the typically subtle effect sizes remains challenging[3,4]. In contrast, rare variants can exhibit large effects[5], aiding the discovery of effector genes[6,7], the unraveling of molecular mechanisms underlying traits, and in turn, the identification of potent drug targets[8–10]. Of further medical relevance, modeling of rare variant effects has recently shown promise in the identification of individuals at high risk for disease and in deriving polygenic risk scores that generalize better across populations than those based only on common variants[11].

However, extending the GWAS strategy to rare variants must contend with large numbers of low-frequency variants, leading to low statistical power due to sparsity and an increased multiple testing burden. To compensate, rare variant association testing (RVAT) methods aggregate rare variants at the level of genomic regions, typically genes[12,13]. Furthermore, the presence or magnitude of impact on gene function for a given rare variant cannot typically be inferred directly. Therefore, RVAT methods leverage functional predictions of variant effect[12,14–16], such as conservation scores or variant effect predictions for splicing, gene expression or protein structure[17–19] to prioritize variants.

Burden testing, a common RVAT strategy, uses annotations to filter presumably uninformative variants and to weight them. These weights are then aggregated to a gene-level burden score and tested for association with discrete or quantitative traits[20–25].

Complementary to burden tests, variance component tests also use annotations for filtering and weighting variants, as part of a kernel function, and can account for both protective and deleterious variants[13,26,27]. Recently proposed RVAT methods based on variance component tests convincingly demonstrated the added value of incorporating a broad spectrum of annotations, either by performing a meta-analysis over different test types and annotations[28] or using specialized kernels tailored to different annotation types[29] (**Methods**; **Supp. Table 1**). However, like earlier methods, *ad hoc* variant filtering and weighting schemes remain integral components of the workflow. The few RVAT methods that use data-driven weights and integrate multiple annotations are, in practice, limited in the number of annotations and in the flexibility of scoring function that can be considered, and they are computationally demanding[30,31]. Finally, none of these methods lends itself to phenotype prediction, and they have not been considered for this task (**Methods**; **Supp. Table 1**).

To address these issues, we present DeepRVAT (Deep Rare Variant Association Testing). Our framework uses a deep set network for modeling traits by integration of rare variant annotations. The model handles variable numbers of rare variants per individual, leverages dozens of continuous or discrete annotations, and accounts for both additive and interaction effects. All model parameters are learned directly from the training data, minimizing the need for *ad hoc* modeling choices that characterize existing methods. The trained DeepRVAT model gives rise to a single, trait-agnostic gene burden scoring function, which can be used for different genetic analyses, including rare variant association tests and to refine polygenic risk prediction to account for rare variant effects. We offer user-friendly software that provides pre-trained DeepRVAT models for direct application on new data sets, as well as a pipeline for retraining the model, for example to incorporate other annotations.

We validate the model using simulations before applying it to 21 quantitative traits and whole-exome-sequencing data from 167,245 UK Biobank (UKBB) individuals, demonstrating a larger number of discoveries compared to existing methods. Additionally, we demonstrate improved replication of DeepRVAT associations in held-out individuals from UKBB. Finally, we combine DeepRVAT gene impairment scores with polygenic risk models, which yields improved prediction accuracy for extreme value phenotypes in UKBB traits.

Results

A deep set network-based RVAT framework

DeepRVAT is an end-to-end model (**Fig. 1a**) that first accounts for nonlinear effects from rare variants on gene function (gene impairment module) to then model variation in one or multiple traits as linear functions of the estimated gene impairment scores (phenotype

Figure 1 | DeepRVAT model overview and downstream use cases.
(**a**) DeepRVAT predicts phenotype from genotype by modeling gene impairment from individual-level rare variants and annotations (Gene impairment module, **b**), and additive trait-specific gene effects.

(**b**) DeepRVAT gene impairment module. Gene impairment scores are estimated from personalized rare variant profiles for each individual and gene combined with an arbitrary number of variant annotations as input. Variants are fed through an embedding network, φ, to compute a variant embedding, followed by permutation-invariant aggregation to yield a gene embedding. A second network, ρ, estimates the final gene impairment score. All parameters of the gene impairment module are shared across genes and traits.

(**c,d**) Downstream analyses based on the DeepRVAT impairment score.

(**c**) Gene discovery. The trained DeepRVAT impairment module can be used as a scoring function to define association tests for gene discovery. This improves both the number of discoveries, as well as their replication rate in held-out data, compared to existing methods.

(**d**) Polygenic risk prediction incorporating rare variants. The DeepRVAT gene impairment module can be used as input to train predictive models for phenotype from genotype, for example, to improve risk stratification based on polygenic risk scores (PRS) with common variants. Created with BioRender.com.

module). The gene impairment module (**Fig. 1b**) outputs a gene- and trait-agnostic gene impairment score estimating the combined effect of rare variants, thereby allowing the model to generalize to new traits and genes. Technically, a deep set neural network[32] architecture is employed to aggregate the effects from multiple discrete and continuous annotations for an arbitrary number of rare variants. This architecture captures additive and nonlinear interactions and does not rely on *a priori* assumptions about the relevance of individual annotations, such as common assumptions about the relationship between allele frequency and variant effects[20,22] (**Methods**).

For each trait used in model training, a set of informative seed genes that are associated with the given trait are required. The DeepRVAT software offers an integrated workflow that employs conventional rare-variant association tests to identify seed genes (**Methods**). Next, the gene impairment module is trained end-to-end together with the phenotype modules on the seed genes. Once trained, the DeepRVAT gene impairment module can be applied to genes not used during training to test for genetic associations using the established principles of conventional burden tests (**Fig. 1c**). The estimated gene impairment scores from the trained DeepRVAT module can also be used to predict phenotype from genotype, thereby providing a flexible way to derive polygenic risk scores[11,33] that account for rare variant effects (**Fig. 1d**). The computational complexity of training DeepRVAT scales linearly with the number of individuals, while computational demands of association testing with DeepRVAT are one to two orders of magnitude lower than existing approaches for incorporating multiple annotations[28,29] (**Supp. Fig. 1.1**). This enables applications to phenome-wide association studies on biobank-scale datasets.

Model validation using simulated data

To validate DeepRVAT for gene discovery, we considered a semi-synthetic dataset derived from whole exome sequencing (WES) from 167,245 caucasian UK Biobank individuals (November 2020, 200k WES release[34]; **Methods**). We annotated all 12,704,497 WES variants using minor allele frequency (MAF), VEP[35] consequences, missense variant impact scores (SIFT[14] and PolyPhen2[16]), and omnibus statistical deleteriousness scores (CADD[15] and ConDel[36]), as well as predicted annotations for effects on protein structure (PrimateAI[18]), and aberrant splicing (AbSplice[17]). To create synthetic phenotypic data, we first simulated impairment scores for genes designated to be causally associated to the trait. To this end, we stochastically assigned each variant an effect based on its annotations (**Supp. Fig. 2.1, Methods**), followed by aggregation for each gene. Finally, the individual phenotype values were simulated as a linear function of the gene impairment scores, covariates mimicking age and sex, and independent Gaussian noise.

After confirming statistical calibration (**Fig. 2a, Supp. Fig. 2.2**), we compared the power of DeepRVAT to conventional burden testing as well as variance component testing with SKAT[26] analogously to the approach of Karczewski et al.[22], i.e. considering either predicted loss of function (pLOF) or missense variants and with weighting based on minor allele frequency (MAF; **Methods**). Existing methods rely on a predefined MAF cutoff. To

investigate the potential impact of the misspecification of the MAF cutoff, we varied the frequencies of the simulated causal variants (**Fig. 2b**, top to bottom, **Methods**). For each simulation setting, we applied the considered methods for various cutoff values of the MAF frequency (<0.01%, <0.1%, <1%; **Fig. 2b**, left to right). Whereas conventional tests were greatly affected by the alignment of the simulated MAF distribution of causal variants and the applied MAF filter, DeepRVAT was consistently well powered, including in settings for which additional non-causal variants were incorporated in the MAF cutoff. Collectively, these results support that DeepRVAT is able to estimate an appropriate weighting for variants from the data, including implicit prioritization of variants by MAF.

Similarly, we considered altering the proportion of rare variants that are selected to contribute to gene impairment effects (**Supp. Fig. 2.1, Supp. Fig. 2.3a**) and a rank-based evaluation criterion (**Supp. Fig. 2.3b**). Across these settings, DeepRVAT was consistently better powered than alternative methods. In sum, the benchmark using simulated data demonstrates that DeepRVAT yields results that are robust to a range of key parameters, including the MAF spectrum and the proportion of causal variants.

Figure 2 | Method assessment using simulated data. Considered were DeepRVAT using complete annotations, conventional burden tests and SKAT, each based on either pLOF or missense variants, and the combination of these four methods (Burden/SKAT combined).
(**a**) Assessment of calibration for DeepRVAT and conventional methods. Shown are Q-Q plots of the expected versus the observed *p*-value distribution. Lambda denotes the genomic inflation factor. Genes simulated to be causal in red; non-causal genes in black.
(**b**) Power comparison for alternative MAF distributions of simulated causal variants (rows)

as well as MAF filters applied to determine input variants for each of the considered tests (columns). Shown is average power (bar height) and standard deviation (error bars) across 10 simulation runs. Results on the diagonal correspond to the setting for which the MAF filter of the RVAT tests are aligned to the simulation setting; others correspond to varying degrees of MAF filter misspecification. Stacked bar plots on the right for each row denote the relative contribution of variants in different frequency bins for the respective simulation settings (averaged across all simulation runs).

DeepRVAT recovers a larger number of gene-trait associations using rare variants in UK Biobank

Next, we applied DeepRVAT to rare variants (MAF < 0.1%) from whole-exome data from the UK Biobank, considering the same 167,245 caucasian individuals as used in the simulation study. We used a total of 33 variant annotations (**Supp. Table 5; Supp. Fig. 3.1**), expanding the set considered in the simulation study by variant effect predictions for epigenetic markers in cell types from ENCODE[37] and the Roadmap Epigenomics[38] projects (using principal components of DeepSEA[19] predictions; **Supp. Fig. 3.2; Methods**), as well as binding to 6 RNA-binding proteins (selected predictions from DeepRiPe[39]; **Methods**). We considered a total of 21 quantitative traits, comprising standing height and 20 blood biomarkers (**Supp. Table 3; Methods**). We again compared DeepRVAT to burden tests and SKAT, either based on pLOF or missense annotations. Additionally, we considered STAAR[28] using the same set of annotations as employed in DeepRVAT, and the tests designed by Monti et al[29] – which comprise both collapsing and variance-component tests – using the same annotations and kernels as in the original work, which substantially overlap with the annotations we have considered for DeepRVAT (**Supp. Table 5**). Both methods employ multiple tests specific to different types of annotations.

Across all 21 traits, DeepRVAT identified 572 gene-trait associations (FDR < 5%; **Supp. Table 6**), which corresponds to a 127% increase in gene-trait associations compared to the most competitive baseline, combining burden and SKAT (252), a 29% increase compared to Monti et al. (444) and an 11% increase compared to STAAR (517) (**Fig. 3a**). At the level of individual traits, DeepRVAT was the top performer among all 8 methods in number of discoveries for 13 out of the 21 traits (**Fig. 3c**). DeepRVAT also retained calibration when applying the model to permuted phenotype data (**Supp. Fig. 3.3**).

Next, we evaluated the validity of the discoveries by assessing their replication in at least one of two studies on the full UK Biobank WES cohort (N=454,787 and 394,841 individuals)[20,22], which employed analysis strategies that resemble the baseline methods we have considered. Notably, across a wide range of nominal significance ranks, the replication rate of DeepRVAT exceeded the replication of discoveries of alternative methods (**Fig. 3b**). This suggests that not only does DeepRVAT have an improved capacity to detect subtle associations compared to existing methods, but it is also less susceptible to spurious ones.

Figure 3 | Application of DeepRVAT to gene discovery using WES and 21 phenotypes from 167,245 UK Biobank individuals. Considered are DeepRVAT, conventional burden or SKAT tests – each using either pLOF or missense annotations – their combination (Burden/SKAT combined), STAAR, and the approach from Monti et al.

(**a**) Cumulative number of gene-trait associations discovered by alternative methods across 21 traits (FDR<5%).

(**b**) Replication of the cumulative discoveries across all traits by alternative methods as in **a,** in a larger cohort (cohort supersets; UK Biobank full release[20,22], **Supp. Table 7**). Shown is, for each method, the total number of gene-trait associations that were also discovered in the larger cohort (according to the methodology of the respective studies) as a function of the rank of their nominal significance. Circles indicate the rank position that corresponds to FDR<5% (as in **a**). The gray line corresponds to a replication rate of one.

(**c**) Number of significant gene-trait associations from **a** broken down by trait.

We also considered DeepRVAT with a reduced set of annotations (MAF, pLOF and missense status), as well as a simplified model architecture with a fully linear gene impairment module. These models yielded lower performance than the full DeepRVAT model when considering discoveries and replication (**Supp. Fig. 3.4**), but still significantly outperformed baseline models. This indicates that the data-driven scoring function, the ability to consider a larger number of annotations, and the capacity to capture nonlinear effects all jointly contribute to the overall performance. This is further supported by an analysis of the relevance of different annotations in DeepRVAT (**Supp. Fig. 3.5**), which suggests that non-canonical annotations such as DeepSEA predictions add value even for coding variants.

DeepRVAT improves phenotype prediction by integrating rare variants

An important advantage of DeepRVAT over approaches that combine different annotations by aggregating across multiple tests such as STAAR and Monti et al. is that it provides phenotype predictions. To assess the added value of DeepRVAT for predicting phenotypes, we trained regression models that combine polygenic risk scores (PRS) derived from common variants[33] with gene impairment scores from DeepRVAT. For comparison, we trained analogous models in which the gene impairment was based on single-variant annotations as in conventional gene burden tests. All models were evaluated using 5-fold cross-validation across individuals. For each fold, conventional RVAT methods (*Burden/SKAT combined*; previous section) or DeepRVAT were employed for gene discovery (FDR < 0.05) followed by training of the phenotype prediction model on the same training set (**Methods**). All models were evaluated on the held-out validation set.

We first applied this strategy to a regression task to predict quantitative traits (**Fig. 4a**). Compared to models based on conventional RVAT burdens, the relative prediction improvement of DeepRVAT vs. common variant PRS was significantly larger across 21 traits (maximum improvement of 3.97% for LDL direct). However, the overall contribution of rare variants in general was modest (median improvement of 1.3% for DeepRVAT), which is consistent with the expectation that common variants explain most of the trait variation across the population. We hypothesized, however, that the importance of rare variants would be higher when applying the same modeling strategy to predicting the individuals in the bottom or top 1% of the phenotypic range (**Fig. 4b**). Indeed, the relative improvement for predicting the extreme percentiles was considerably larger (maximum improvement using DeepRVAT of 52.3% for predicting individuals with low LDL direct measurement), and we again found significant advantages of the DeepRVAT impairment score compared to alternative burden measures. Dissection analyses, combining alternative methods for gene discovery and burden score estimation, showed that the gene impairment score and the set of identified genes individually contribute to the added value of DeepRVAT (**Supp. Fig. 4.1**).

Figure 4 | Application of DeepRVAT and alternative gene burdens for phenotype prediction. Assessment of alternative rare variant gene burdens as features for phenotype prediction of 21 UK Biobank traits. Considered were polygenic risk scores derived from common variants, combined with either DeepRVAT or one of six alternative rare burden measures, with evaluation by 5-fold cross validation.

(**a**) Relative improvement of the prediction performance of a linear regression model that includes rare variant gene burdens versus a model based on common variant PRS only. Shown are relative differences in the coefficient of determination (R^2) for models that account for rare variants versus a common-variant PRS model.

(**b**) Analogous comparison as in **a**, however considering a logistic regression model to stratify individuals in the bottom or top 1% of the phenotypic distribution. Shown are relative differences in the area under the precision-recall curve (AUPRC) between a model that accounts for rare variants versus a common variant PRS model. Unless indicated otherwise ("ns"), the relative gains of including the DeepRVAT burden compared to alternative methods as in **a** and **b** are significant (p < 0.01, one-sided Wilcoxon test).

(**c**) Rank-based enrichment of individuals with extreme value phenotypes (top or bottom 1%) among individuals with strongly deviating predictions when using a model that combines PRS and a rare variant burden. Shown is, for each phenotype, the number of individuals with extreme-value phenotypes when ranking individuals by the magnitude of deviation using a model that accounts for rare variants versus a common-variant PRS model.

(**d**) Enrichment of outlier phenotype predictions (exceeding the 99% quantile) in individuals with extreme phenotypes. Shown is the enrichment of outlier individuals (defined as exceeding a certain Z-score cutoff, x-axis) for models that account for rare variants relative to the enrichment obtained from a common-variant PRS model.

(**e**) Evaluation of DeepRVAT for phenotype prediction on held-out phenotypes. Shown is a scatter plot of relative ΔR^2 value (computed as in **a**), considering a linear regression model

using DeepRVAT trained on all 21 phenotypes (x-axis) versus DeepRVAT when omitting the target phenotype upon training the gene impairment module (y-axis).
(**f**) Same analysis as in **e,** but comparing the relative ΔAUPRC for logistic regression models (as in **b**).

Next, we focused on the set of individuals with the strongest deviations between the rare variant model and common PRS predictor. Particularly with DeepRVAT, individuals showing the greatest deviation in predicted phenotypic values were more likely to exhibit extreme phenotype values (bottom or top 1%; **Fig. 4c**). Similarly, individuals with outlying phenotype predictions from the DeepRVAT model were more prominently enriched for extreme phenotype values compared to alternative rare variant predictors and a common variant PRS-only predictor (**Fig. 4d**). At the most extreme phenotype threshold (z-score ≥ 3.16), individuals were 35.7-fold more likely than the overall population to have a predicted phenotype in the 99.9th percentile, compared to 32.8-fold for the most competitive alternative rare variant predictor (PrimateAI) and 32.2-fold for the common variant-only model. These findings demonstrate the benefits of DeepRVAT gene impairment scores for phenotype prediction, particularly in identifying individuals with outlier phenotypes, compared to conventional PRS based on common variants and alternative rare variant models.

Finally, we tested the generalizability of DeepRVAT gene impairment scores across phenotypes not included in the training set (**Fig. 4e-f**). When comparing phenotype predictors derived from a gene impairment module trained on all 21 phenotypes to a model excluding the evaluated phenotype during impairment module training, we found no notable drop in model performance. This suggests that DeepRVAT impairment scores can be effectively applied to diverse phenotypes, demonstrating their robust applicability beyond the training dataset.

Discussion

We have introduced DeepRVAT, a data-driven deep learning model for association testing and phenotype prediction based on rare variants. Unlike existing methods, DeepRVAT infers the relevance of different annotations and their combination directly from data. In so doing, DeepRVAT eliminates the need for *post hoc* aggregation of test results derived from individual annotations using multiple testing schemes. DeepRVAT combines the benefits of a flexible deep neural network architecture with a calibrated statistical testing framework. It leverages the flexibility of deep neural networks to non-linearly integrate rare variant annotations while still offering a calibrated statistical framework for gene-phenotype association testing. DeepRVAT significantly outperformed state-of-the-art methods in gene discovery for 21 traits from the UK Biobank, leading to a substantial increase in performance and retrieving gene-trait associations with higher replication rates in held-out data. Moreover, DeepRVAT improved over polygenic risk scores based on common variants for phenotype prediction, particularly for extreme phenotypes, outperforming conventional burden scores based on single annotations. DeepRVAT is provided as a user-friendly

software package that supports both *de novo* training of gene impairment modules as well as application of pretrained ones, with dramatic improvements in computational efficiency over existing methods.

Although we found that DeepRVAT advances the state-of-the-art in two use cases central to genetics, the model is not free of limitations. First, a universal challenge of any method that considers annotations for rare variant aggregation is the question of how to choose the most relevant annotations. DeepRVAT is no exception, yet it can cope with a potentially very large number of annotations, pruning uninformative ones during the learning process. Second, we benchmarked DeepRVAT using exome sequencing data, but whole genome sequencing promises valuable insights into non-coding regions with numerous rare variants of uncertain impact. We hypothesize that the added value of DeepRVAT, which incorporates rich variant annotations, may be even larger in this context. Finally, while we showed that DeepRVAT is robust to various rare variant frequency cutoffs, it is still based on a dichotomy between rare and common variants. Recent reports have shown substantial overlap between GWAS loci and rare-variant associations[10,11,20]. These insights suggest the potential for future developments to jointly model and estimate rare and common variant effects in a unified framework.

The integration of rare variants will remain a major topic in quantitative genetics modeling. Among rare variant association methods, DeepRVAT belongs to the class of approaches that model gene impairment, a strategy that underpins high-impact variant filters in burden tests[20,22,23] or protein function impairment[25]. Our contribution to this family of approaches is to create a better optimized impairment score by allowing the integration of different annotations and learning a more flexible model directly from cohort data. Importantly, we found this gene impairment score to generalize well across traits. This transferability, combined with DeepRVAT's computational efficiency, is an important feature that will facilitate its application to new phenotypes, for example to conduct rapid rare-variant phenome-wide association studies (PheWAS), to perform rare variant associations in smaller cohorts from case-control studies, or to analyze cohorts from rare disease and cancer.

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Author contributions

B.C., E.H., J.G., & O.S. conceived the method. F.B. made additional conceptual contributions to the method. B.C., E.H., H.O., & F.H. prepared the data. B.C., E.H., H.O., K.M, M.M., F.M., & M.W. implemented the methods and analyzed the data. B.C., E.H., J.G., & O.S. interpreted the results and wrote the paper.

Competing interests

O.S. is a paid advisor of Insitro Inc. The remaining authors declare no competing interests.

Methods and data

A complete methods document with all relevant details is provided as *Supplementary methods*.

Supplementary Data

Supplementary methods: SupplementaryMethods.pdf
Supplementary figures and table captions: SupplementaryInformation.pdf
Supplementary tables: SupplementaryTables.xlsx

Code availability

The code to run DeepRVAT can be found here: https://github.com/PMBio/deeprvat/
The code to run all analysis done in this paper and re-generate the figures can be found here: https://github.com/PMBio/deeprvat-analysis/

Data availability

The genetic, phenotype, and covariate data are protected and are only available to researchers that have valid and approved research applications for these data within the UK Biobank (www.ukbiobank.ac.uk/).

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