Unveiling the role of SRY in male-biased cancers: Insights into the molecular basis of sex disparities in high-grade glioma and melanoma

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ABSTRACT

Sex disparities have been observed in many tumor types affecting non-reproductive organs. Typically, the incidence and mortality rates of such cancers are higher in men. Although differences in lifestyle and environmental exposures are known contributors, knowledge of the molecular mechanisms driving sexual dimorphism in tumor development and therapy response remains limited. To address this question, we comprehensively studied the sex-determining region Y (SRY) gene, a male-specific gene that is critical in development. First, we screened 2,448 samples from 11 cancer types to identify those with a higher incidence in men and increased expression of SRY. In cases of high-grade glioma and melanoma, men with tumors exhibiting high SRY expression had a worse prognosis. Our results suggest that SRY target genes show altered expression when SRY is overexpressed. These gene sets are linked to cell growth, epithelial-mesenchymal transition, inflammation, and repression of tumor suppressor pathways (TP53 activation and activity of immune cells). In summary, we present the first comprehensive investigation of SRY expression and its association with clinical outcomes in men with high-grade glioma and melanoma. Our results shed light on the molecular basis for sex disparities and lay the foundation for investigation of various target genes and novel cancer treatments in men with high-grade glioma and melanoma.

Keywords: sexual dimorphism in cancer; high-grade glioma; melanoma; SRY; gene targets

INTRODUCTION

Sexual dimorphism refers not only to the differences between males and females in physiologic and behavioral characteristics, but also to disease susceptibility and treatment outcomes [1]. Many cancers exhibit sexual dimorphism in terms of incidence, prognosis, and treatment response [2]. For organs with non-reproductive functions, cancer incidence and mortality are usually higher in men [3,4]. While lung cancer, bladder cancer, liver hepatocarcinoma, esophageal cancer, and high-grade gliomas are more common in men, gallbladder, anal, and thyroid cancers are more common in women [3,4]. Overall, the underlying reasons for sexual dimorphism in cancer are poorly understood, in particular the genetic and hormonal contributions involved [4]. More knowledge is required for developing tailored prevention and treatment strategies that consider the unique biology and health needs of each gender.

Among the male-specific genetic factors potentially related to cancers that are more common in men, the male-specific region in chromosome Y has attracted interest. This region is 54 MB in size and contains roughly 80 protein-coding genes [5]. The sex-determining region Y (SRY) gene is a transcription factor that stands out among these genes. SRY is the crucial gene that initiates male sex determination in mammals, including humans [6,7]. Elucidating the role of SRY has greatly advanced our understanding of the molecular basis of sexual differentiation. SRY codes for a protein that regulates the expression of other genes (e.g., SOX9 and SF1) involved in the differentiation of the bipotential gonad into a male gonad, a primordial step in male sexual development [8]. In humans, mutations or abnormalities in SRY expression can lead to disorders of sexual development [9]. For example, individuals with XY chromosomes but a mutation in the SRY gene may develop as phenotypic females, while individuals with XX chromosomes but the SRY gene translocated to the X chromosome may develop as phenotypic males [10]. Moreover, growing evidence suggests that the SRY may be involved in the development of certain types of cancer, particularly those that are more common in men [11,12,13,14]. SRY expression levels have already been correlated with cancer progression and prognosis in patients with hepatocellular carcinoma [11,12] and in a few other tumors,

such as prostate cancer [13,14].

Skin cutaneous melanoma (SKCM) is a type of cancer that arises from the pigment-producing skin cells known as melanocytes. SKCM is one of the top 10 most common cancers [15], with exposure to ultraviolet radiation being its primary cause [16]. Additionally, SKCM is the most aggressive and deadly form of skin cancer, and is responsible for most skin cancer deaths [17]. Early detection of melanoma is associated with a favorable prognosis [18,19]. However, if the melanoma has metastasized, the five-year survival rate drops to around 27%. Melanoma treatment options include surgery, radiation therapy, chemotherapy, and newer targeted and immunotherapy treatments [20,21]. Melanoma incidence rates differ between genders, with a higher incidence (up to 50% more in some countries) observed in men [15].

High-grade gliomas (HGG), which include glioblastoma, anaplastic astrocytoma, and anaplastic oligodendroglioma, originate from the supportive neuroglial cells of the central nervous system [22]. HGG can cause cognitive impairment, personality changes, and seizure disorders, and treatment requires complex multidisciplinary care. Despite intensive efforts to develop new therapies for HGG, treatment options remain limited, and prognosis is still poor [23]. Moreover, HGG are more prevalent in men than in women, with a male-to-female ratio of approximately 3:2 [24]. The reasons for this difference in incidence are not fully understood, but related, in potential, to hormonal or genetic factors [25].

Here, we investigated the role of the SRY gene in tumors that are more common in men (in non-reproductive organs). We used a comprehensive strategy to assess SRY expression in bulk and single-cell datasets along with clinical and survival information, followed by a large-scale functional analysis in 11 cancer types. Our results identified two highly aggressive tumor types with higher incidence in men, high-grade glioma and melanoma, in which increased SRY expression was associated with a worse prognosis. Moreover, our results linked high SRY expression to specific oncogenic pathways.

RESULTS

SRY is differentially expressed in aggressive tumors that are more common in men

We employed an integrative approach to assess SRY expression levels and its potential regulatory impact on tumors with higher incidence in men. We used gene expression (bulk and single-cell) as well as clinical data to investigate SRY expression patterns and its association with patients' prognosis and oncogenic pathways in 2,448 tumor samples from 11 cancer types (Figure 1A) (see Methods for details). We started analyzing the gender distribution of cancer patients from The Cancer Genome Atlas (TCGA) and identified several tumor types that were at least 20% more frequent in men (Figure 1B and Supplementary Table S1). As expected, based on data from The United States and Europe, a higher number of cases was observed among men in TCGA for esophageal carcinoma, lung squamous cell carcinoma, bladder urothelial carcinoma, kidney carcinoma, liver hepatocellular carcinoma, skin cutaneous melanoma, and high-grade (III-IV) glioma [2,26].

To assess the contribution of SRY expression levels in these types of tumors, we first examined differences between tumors and normal tissues. Increased SRY expression was observed in high-grade and low-grade gliomas, lung squamous cell carcinoma, and pancreatic adenocarcinoma, while skin cutaneous melanoma and esophageal cancer exhibited SRY underexpression (p-value < 0.001; Wilcoxon test) (Figure 1C).

Figure 1. Identifying SRY differential expression in cancer types that are more prevalent among men. A) Schematic view of the approach used to investigate SRY gene expression in male cancer patients. **B)** Tumors that are more common in men (based on the number of samples) in the TCGA cohort. **C)** Comparison of SRY expression levels (TPM) in tumors from male patients from TCGA and healthy tissues from GTEx (Wilcoxon tests; **** p-value ≤ 0.0001, *** p-value ≤ 0.001). Abbreviations: ESCA, esophageal carcinoma; LUSC, lung squamous cell carcinoma; BLCA, bladder urothelial carcinoma; KIRP, kidney renal papillary cell carcinoma; LIHC, liver hepatocellular carcinoma; KIRC, kidney renal clear cell carcinoma; STAD, stomach adenocarcinoma; SKCM, skin cutaneous melanoma; HGG, high-grade glioma (grades III-IV); PAAD, pancreatic adenocarcinoma; and LGG, low-grade glioma (grade II).

SRY is preferentially expressed in malignant glioma cells, and its high expression is associated with worse prognosis in men with HGG

We investigated the prognostic value of SRY in the six cancer types with higher frequency in men (Figure 1C). We used log-rank tests to compare progression-free survival rates of 1,068 patients stratified in two groups based on their SRY expression levels: high (transcripts per million [TPM] ≥ 0.1) or low (TPM < 0.1). Considering all tumors, only HGG and SKCM showed significant associations (log rank test p-values < 0.05) between SRY expression and patients' prognoses (Supplementary Figure S1 and Figure 2A). In both types of cancer, high SRY expression was associated with a worse prognosis.

Deeper investigation of SRY in HGG patients from the TCGA (Supplementary Table S2) determined that 50% of patients with high SRY expression had recurrence in up to 300 days, while patients with low SRY expression had later recurrence (over 1,500 days) (Figure 2A). Next, we performed multivariate Cox regression to estimate the associations between SRY expression and progression-free survival in HGG together with relevant clinical information. High SRY expression emerged as a pivotal factor in disease recurrence (hazard ratio of 1.79 [95% CI, 1.18 to 2.72]; p-value = 0.006), Figure 2B).

To confirm the prognostic value of SRY in an independent set of gliomas, we used gene expression and clinical information from 238 men with HGG in the Chinese Glioma Genome Atlas (CGGA) (Supplementary Table S3). In agreement with our findings in the TCGA cohort, high expression of SRY (TPM > 0.1) was associated with a worse prognosis (Figure 2C). While 50% of HGG patients with a high SRY expression died within 500 days, patients with low SRY expression had survival times of ~1,000 days (Figure 2C). To further investigate the relationship between SRY expression and glioma aggressiveness, we assessed SRY expression in low-grade gliomas (LGG), which are less aggressive and have a better prognosis than HGG. SRY expression was significantly lower in LGG (and even lower in normal cortex) compared to HGG (p-value < 0.001; Wilcoxon rank test) (Figure 2D). This result is consistent with our previous findings showing that gliomas with higher SRY expression are more aggressive (Figure 2A-C).

Finally, we used single-cell gene expression data to assess SRY expression in cells from HGG samples (Supplementary Figures S2-S3). SRY was exclusively expressed in malignant tumor cells and not in other cell types such as macrophages, oligodendrocytes, stromal cells, or T-cells in the tumor microenvironment (Figure 2E). Taken together, our results indicate that SRY is predominantly expressed in malignant HGG cells and that its overexpression is significantly associated with a worse prognosis for patients with HGG.

Figure 2. SRY is preferentially expressed in malignant glioma cells and its overexpression is associated with a worse prognosis in men with HGG. A) Progression-free survival in the TCGA cohort of men with HGG and high (TPM ≥ 0.1) or low (TPM < 0.1) SRY expression. **B)** Multivariate Cox regression analysis considering several variables among men with HGG: SRY expression, age (median: 52 years), gain of chromosome 7 and loss of chromosome 10, TERT expression status and four glioma histologies: GBM, glioblastoma; A, astrocytoma; OA, oligoastrocytoma; and OD, oligodendroglioma. **C)** Overall survival for CGGA cohort of men with HGG and high (TPM ≥ 0.1) or low (TPM < 0.1) SRY expression. **D)** Expression levels of SRY in healthy (frontal) cortex samples from male individuals (GTEx cohort) and in tumor samples

from men with LGG or HGG from the TCGA and CGGA cohorts (Wilcoxon tests; **** p-value ≤ 0.0001, *** p-value ≤ 0.001, ** p-value ≤ 0.01); **E)** Results of single-cell RNA sequencing from seven HGG samples from two studies (acessions GSE141946 and GSE131928) showing SRY expression in cancer cells (malignant; in green), but not in other cells identified.

SRY target genes are associated with essential cancer aggressiveness pathways

Previous reports have shown that SRY regulates genes associated with cancer development and progression [11,27]. To evaluate SRY's downstream impact on tumor progression, we first selected its target genes identified in chromatin immunoprecipitation (ChIP-Chip) experiments [28] (Supplementary Table S4). Differential expression profiling between tumors with high (TPM ≥ 0.1) or low (TMP < 0.1) SRY expression (see Methods) identified 310 SRY target genes (Figure 3A; Supplementary Table S5). Next, pathway enrichment analysis of this set of SRY targets identified three enriched pathways: *epithelial mesenchymal transition* (associated with metastatic expansion, generation of tumor cells with stem cell properties, and resistance to cancer treatment); *proteoglycans in cancer* (associated with cancer cell adhesion and migration); and *NF Kappa B signaling* (associated with proliferation, cell survival, growth, and angiogenesis), among others (Figure 3B).

This pathway analysis also identified the *Estrogen response late* pathway as enriched. While estrogen is typically associated with female sex hormones, it is also produced in men, albeit at lower levels. Additionally, abnormal activation of the estrogen response late pathway has been associated with an increased risk of developing cancers in men [29]. On the other hand, the *P53 tumor suppressor pathway* was associated with downregulated target genes for SRY (Figure 3B).

To provide a comprehensive view of target genes differentially expressed in tumors with high SRY expression, we performed protein-protein interaction (PPI) analyses using these genes. The PPI network of downregulated SRY target genes preferentially contained proteins related to synthesis, release, and regulation of neurotransmitters (Figure 3C), essential functions of the nervous system commonly altered in HGG [30]. Furthermore, the majority of these genes (69.2%) have already

been associated with cancer, particularly demonstrating reduced expression (61%), and with validated or putative tumor suppression functions (Supplementary Table S6). These findings are thus in agreement with our observation of their downregulation in HGG samples.

The PPI network of overexpressed SRY target genes revealed proteins linked to extracellular matrix remodeling, cell adhesion, migration, angiogenesis, tumor invasion, and metastasis, such as FN1 [31], ICAM1 [32], COL1A1 [33], PLAU [34], and SERPINE1 [35] (Figure 3D). These functions are commonly associated with tumors that have a poor prognosis, in agreement with our previous results (Figure 2A-C) and our hypothesis that SRY expression is associated with tumor aggressiveness. Collectively, these results provide insights into the molecular mechanisms driving cancer aggressiveness, including activation of cell migration, tissue invasion, angiogenesis, and suppression of immune cell infiltration, which could explain why men with HGG and high SRY expression have a worse prognosis (with more advanced tumor progression and lower survival rates) than those with low SRY expression.

Figure 3. HGG samples with high expression of SRY show target genes associated with activation of cancer aggressiveness pathways and the suppression of tumor suppressor features. A) Differentially expressed SRY target genes between HGG samples showing higher (≥ 0.1 TPM) versus lower SRY expression (< 0.1 TPM). **B)** Functional enrichment analysis (KEGG pathways and MSigDB Hallmarks) of SRY target genes up- and down-regulated in HGG samples. **C)** Protein-protein interaction networks of SRY target genes that were down-regulated in samples with higher SRY expression. **D)** Protein-protein interaction networks of SRY target genes that were upregulated in samples with higher SRY expression.

High expression of SRY is associated with a worse prognosis in melanoma patients

High SRY expression (TPM ≥ 0.1) was also associated with a worse prognosis (log rank test p-value < 0.05) in samples from men with skin cutaneous melanoma (SKCM) in the TCGA (Supplementary Table S7). While half of those with high SRY expression had disease recurrence within 500 days, recurrence in half of the patients with lower

SRY expression persisted up to 1,000 days (Figure 4A). To more accurately assess other variables potentially associated with tumor progression, we performed multivariate Cox regression analysis considering SRY expression together with other clinical information from these patients (age and tumor stage). Only tumors with high expression of SRY (TPM ≥ 0.1) were significantly associated with length of survival (p-value < 0.03; hazard ratio of 2.77 [95% CI, 1.9 to 7.02]) (Figure 4B). Our data suggest that SRY was a major factor contributing to tumor recurrence in this cohort of men with melanoma.

We then profiled SRY expression in tumors from men with melanoma versus healthy skin samples. SRY was underexpressed in these tumors (p-value < 0.0001; Wilcoxon rank test) compared to healthy sun-exposed skin samples from the Genotype-Tissue Expression Project (GTEx) cohort (Figure 4C). Since human skin is composed of three layers made up of different cells (e.g., keratinocytes, melanocytes, Langerhans cells, and eccrine cells), we hypothesized that SRY may have distinct expression in these cell types. To test this hypothesis, we performed deconvolution of bulk RNA-seq expression data (healthy sun-exposed skin samples from GTEx) using CIBERSORTx [36], a tool that combines support vector regression with prior knowledge on scRNA-Seq expression profiles to estimate cell fractions. In agreement with our hypothesis, healthy skin keratinocytes and eccrine cells have robust SRY expression, while melanocytes have limited SRY expression (Figure 4D). Next, we measured SRY expression in an original single cell dataset [37]. Figure 4E shows that only keratinocytes express SRY in healthy skin. Altogether, these results indicate that SRY overexpression was associated with a poor prognosis for men with melanoma, similar to its role in HGG. Furthermore, the finding that SRY is also expressed in healthy skin, but not in melanocytes, suggests that its dysregulation in melanocytes may contribute to the development of melanomas.

Figure 4. SRY is associated with worse prognosis in men with melanoma and over-expressed in melanocytes only in tumors, but not in healthy skin. A) Progression-free survival of men with melanoma expressing SRY at high (TPM ≥ 0.1) or low (TPM < 0.1) levels in the TCGA cohort. **B)** Multivariate Cox regression of melanoma patients considering SRY expression together with clinical information about age (median: 63 years), and AJCC T (T1 - T4) and N (N0 - N3) stages. **C)** Expression levels of SRY in melanoma tumors from men in the TCGA cohort are underexpressed versus healthy sun-exposed skin samples (GTEx cohort) (Wilcoxon test; **** p-value ≤ 0.0001). **D)** Proportions of distinct cell types estimated from bulk RNA-seq data of 464 healthy skin samples (p-value < 0.05). **E)** In healthy skin samples from men in a scRNA-seq cohort (accession: GSE151091), SRY is expressed only in keratinocytes.

High expression of SRY activates genes associated with melanoma progression and aggressiveness

Similar to the analyses of HGG tumors (Figure 3), we explored the differential expression of SRY target genes [28] (Supplementary Table S8). Four proto-oncogenes (epidermal growth factor receptor [EGFR], FLT, BCL11B and PAX5) were identified among overexpressed genes (Figure 5A). These genes have been implicated in

various cellular processes related to cancer, including cell proliferation, differentiation, and apoptosis. For example, EGFR is often upregulated in tumors, including melanoma, and its high expression contributes to tumor development and progression by promoting cell growth and survival, inhibiting apoptosis, and enhancing tumor invasion and metastasis [38,39]; FLT3 has been explored as a potential therapeutic target and showed promising results in melanoma [40,41].

Pathway enrichment analysis of target genes overexpressed in SKCM samples with higher SRY expression identified associations with key cancer pathways (Figure 4B). These included the KRAS signaling pathway, which is associated with cell proliferation, differentiation, and survival [42]; proteoglycans in cancer, which is associated with changes in the composition and organization of the extracellular matrix, thereby affecting the tumor microenvironment and tumor progression [43]; and, similar to HGG, the estrogen response (early and late) pathways. Moreover, two cellular pathways were shared between HGG and SKCM overexpressed SRY target genes: the estrogen response (late) and proteoglycans in cancer, suggesting their importance in tumors with high SRY expression.

In PPI analysis of target genes overexpressed in SKCM tumors with high SRY expression, EGFR emerged as the most highly connected protein (Figure 5C). Activation of the EGFR pathway leads to increased cell growth and tumor formation in many cancers, including melanoma. Other proteins related to melanoma progression and aggressiveness, such as WNT7B [44] , WNT5A [45], WNT11 [46] (Wnt pathway) and other individual proteins (e.g., SFN [47], COL1A1 [33], CLDN4 [48], PKP3 [49], and SDC1 [50]), were highly connected in the PPI analysis. Overall, these results indicate that SRY target genes in samples with high SRY expression are implicated in cellular pathways associated with melanoma progression and aggressive tumors, supporting high SRY expression as a contributor to poor prognosis among men with melanoma (Figure 5).

Figure 5. Several SRY targets are overexpressed in men with melanoma who had a worse prognosis. A) Overexpressed SRY target genes in SKCM samples with high expression of SRY (≥ 0.1 TPM). **B)** Functional enrichment analysis (KEGG pathways and MSigDB Hallmarks) of SRY target genes overexpressed in SKCM samples with high expression of SRY. **C)** Protein-protein interaction network of overexpressed SRY targets in SKCM patients with high expression of SRY.

DISCUSSION

Although sexual dimorphism has been observed in many cancer types [51–53], our understanding of these sex disparities at the molecular level is still very limited. Here, we integrated and analyzed a large amount of gene expression data (including single-cell expression) along with clinical and functional datasets to perform a comprehensive analysis of SRY gene expression in men with high-grade glioma or melanoma. Overall, high SRY expression was associated with a worse prognosis and linked to oncogenic pathways in men with these two cancers.

      High-grade glioma is more prevalent in men than women [2], but the reasons for this difference are not fully understood and likely involve hormonal, genetic, and

environmental factors [1]. In our investigation, we discovered that high (TPM ≥ 0.1) expression of SRY, a male-specific gene, was a poor prognostic factor among men with high-grade glioma. Moreover, SRY's target genes which were overexpressed in high-grade glioma patients with a worse prognosis, are involved in cellular pathways associated with tumorigenic processes such as extracellular matrix remodeling, cell adhesion, migration, angiogenesis, tumor invasion, and metastasis. Conversely, those with higher SRY expression also had underexpression of genes that, in normal conditions, facilitate infiltration and activity of immune cells within the tumor microenvironment. Thus, altogether these results indicate that SRY overexpression may activate key genes related to tumor aggressiveness and immune system modulation in high-grade glioma patients, and may be driving the worse prognosis in these patients.

Skin cutaneous melanoma (SKCM) is an increasingly common cancer and has a higher incidence rate in men than in women, with a lifetime risk of 1 in 26 for men and 1 in 41 for women [17]. Again, it is reasonable to suppose that this biased incidence is due to behavioral, genetic and hormonal factors. Our results show that men with melanoma and high SRY expression have a worse prognosis. As in high-grade glioma, tumors with a higher SRY expression showed overexpression of SRY target genes associated with cell proliferation, cell differentiation, and changes in the composition and organization of the extracellular matrix – all well-described cellular functions associated with melanomas aggressiveness. We found higher SRY expression in healthy skin than in melanoma, but this expression is limited to non-melanocyte cells, mostly keratinocytes. These findings suggest that SRY overexpression in melanoma (mainly melanocytes) may be a dysregulation that contributes to activation of genes involved in tumorigenesis, tumor progression and aggressiveness.

Among all cellular pathways enriched in men with high-grade glioma or melanoma and overexpression of SRY, only two were shared, the estrogen response (late) and the proteoglycans in cancer pathways. The estrogen response cellular signaling pathway is involved in the regulation of various physiologic processes, including cell proliferation and differentiation. While estrogens regulate many aspects of reproductive development and function in women, they also play a role in

men. In men, estrogen 17β-estradiol (E2) is a relevant hormone for hypothalamic-pituitary-testicular axis regulation, reproductive function, growth hormone-insulin-like growth factor-1 axis regulation, bone growth, skeletal health, glucose metabolism, brain development, and behavior [54,55]. Moreover, in men, estrogens are synthesized by aromatase (an enzyme that converts androgens to estrogens), which is expressed in several somatic tissues, especially in the brain [56]. In the male brain, aromatase is involved in several brain functions, including synaptic plasticity, neurogenesis, neuroprotection, and regulation of sexual behavior [56,57]. On the other hand, high levels of estrogen in men are linked to an increased risk of some cancers, including prostate cancer [29]. The fact that the SRY DNA-binding domain specifically recognizes proximal upstream elements in promoters of sex-specific genes encoding P450 aromatase suggests a connection between SRY and aromatase expression (and thus estrogen production) [58,59].

Additionally, the effects of SRY expression on the increased expression of genes related to epithelial-mesenchymal transition (EMT) may also be associated with estrogens [60]. Estrogen-promoted signaling is related to induction of EMT in estrogen-responsive tissues [60,61]. In ovarian and prostate cancer, estradiol treatment induces estrogen receptor (ER)-α-dependent EMT, which is inhibited by receptor silencing [62]. Nevertheless, loss of ER-α expression in breast and endometrial cancer promotes morphologic changes, motility, improved invasion, and increased expression of EMT markers [63,64]. In human glioblastoma-derived cell lines, 17β-estradiol induced EMT via changes in cell morphology, expression of EMT markers, and cell migration and invasion assays [60].

While no relationships between SRY and proteoglycans in tumors or tumorigenesis have been reported, evidence from developmental diseases may suggest a potential connection. One such disease, campomelic dysplasia, is a congenital skeletal abnormality in humans which manifests with notable skeletal deformities along with distinct facial features, including macrocephaly, micrognathia, cleft palate, flat nasal bridge, and sex reversal [65]. The genetic basis of this condition involves mutations in the SRY-related gene SOX9 [66]. Specifically, SOX9 expression initiates during mesenchymal condensation preceding cartilage formation and remains within the perichondrium and chondrocytes. Thus, this

transcriptional activator likely plays a significant role in the establishment and maintenance of the chondrocytic phenotype by regulating cartilage-specific genes, including collagen types II and XI, aggrecan, and proteoglycans [67]. Consequently, SRY indirectly influences the expression of proteoglycans through its interaction with SOX9. Therefore, in conditions with high SRY expression, an upregulation of proteoglycans would be expected, reinforcing our findings in both glioblastoma and melanoma.

In light of our findings linking high SRY expression and estrogen-response genes in high-grade glioma and melanoma, we raise the question whether endocrine therapy, which involves drugs that block the estrogen receptor or reduce estrogen levels, could benefit these patients. While we have found limited evidence to support the use of endocrine therapy to treat high-grade glioma or melanoma, based on our data, clinical trials may be warranted, particularly in cases where other standard therapies have proven ineffective.

CONCLUSION

Taken together, our results provide insight into the biological mechanisms driving two cancers that are more common in men – high-grade glioma and melanoma – and suggest novel targets for putative therapeutic intervention. Specifically, we demonstrate that SRY overexpression predicts poor prognosis in high-grade glioma and melanoma, and we showed that this high SRY expression may activate target genes and cellular pathways involved in tumorigenesis, tumor progression, and immune system repression. Further studies are needed to explore the potential of these findings in developing new treatments for men with high-grade glioma or melanoma.

METHODS

RNA sequencing data

To determine cancers in The Cancer Genome Atlas - TCGA cohort that are more common in men (https://portal.gdc.cancer.gov), we obtained clinical data with gender information for 8,065 patients with 19 distinct types of solid primary tumors. Sex-specific tumor types and tumor types represented by fewer than 150 patients were not considered for analysis. We defined cancers more common in men as those with male to female ratios ≥ 1.2.

For the 11 tumor types identified as more common in men, we downloaded all available processed RNA-Seq data (raw gene counts and transcript per million [TPM] expression levels) from men: bladder urothelial carcinoma (BLCA, N=300), esophageal carcinoma (ESCA, N=158), high-grade glioma (HGG, N=248), kidney renal clear cell carcinoma (KIRC, N=345), kidney renal papillary cell carcinoma (KIRP, N=214), low-grade glioma (LGG, N=136), liver hepatocellular carcinoma (LIHC, N=250), lung squamous cell carcinoma (LUSC, N=371), pancreatic adenocarcinoma (PAAD, N=98), skin cutaneous melanoma (SKCM, N=61), and stomach adenocarcinoma (STAD, N=267). We evaluated the median TPM-normalized expression levels of SRY and considered for further analyses only tumors with a median TPM expression ≥ 0. These were: BLCA, ESCA, LGG, LUSC, HGG, PAAD and SKCM.

To compare SRY expression levels between these selected tumor samples and corresponding healthy tissue samples from men, we also obtained processed RNA-Seq data of 1,780 samples from The Genotype-Tissue Expression (GTEx Release V8) data portal (https://gtexportal.org): bladder (N=14), esophagus (N=363), brain (frontal) cortex (N=334), lung (N=395), pancreas (N=207), and skin (N=467).

To evaluate our results in gliomas in an independent cohort, we also obtained processed RNA-Seq data from men in the Chinese Glioma Genome Atlas - CGGA (http://www.cgga.org.cn), comprising 238 HGG and 131 LGG samples of primary tumors.

Survival analyses

To evaluate whether SRY expression impacts the prognosis of cancer patients, we obtained progression-free survival data from TCGA and curated by Liu *et al.* [68], and retrieved additional clinical information using the R package *TCGAbiolinks* (version 2.24.3). Overall survival and clinical data for the CGGA cohort were obtained directly from the CGGA data portal.

For each tumor type, we stratified patients into two groups based on SRY expression: patients with high (TPM ≥ 0.1) or low (TPM < 0.1) SRY expression. Survival analyses between these groups were then performed using log-rank tests; results were considered significant at a p-value < 0.05. Kaplan-Meier survival curves were created using the R packages *survival* (version 3.5.3) and *survminer* (version 0.4.9). For tumors with significant survival differences (p-value < 0.05), we next performed multivariate Cox regression analyses to evaluate the prognostic value of SRY in context with other relevant clinical variables. Only variables with < 5% of missing values were included. Forest plots were built using the R package *forestmodel* (version 0.6.2).

Single-cell RNA sequencing data

To check SRY expression in single-cell RNA sequencing (scRNA-Seq) data obtained from patients with primary HGG, we obtained data from two previously published studies: [69] (GEO accession: GSE131928) and [70] (GEO accession: GSE141946). For the first dataset, SRY expression was directly assessed in 3,825 cells from 14 men. For the second dataset, scRNA-Seq data from 3 male individuals was reprocessed using the R package *Seurat* (version 4.3.0) [71]. Briefly, only cells with (a) at least 400 and at most 5,000 genes detected and (b) less than 25% mitochondrial reads were retained for further analysis (8,892 cells). Standard preprocessing steps, including normalization, scaling, and cell cycle phase scoring, were run without modifications. UMAP dimensionality reduction was performed with default parameters, and the Seurat function *FindAllMarkers* was used to perform differential gene expression with thresholds set to log2(fold-change) > 0.1 and minimum percent expressed > 0.1. The top 10 marker genes based on highest

log2(fold-changes) were selected for each cell type.

Moreover, we evaluated SRY expression in 2,981 skin cells of 8 healthy men using processed scRNA-Seq data obtained directly from a previous study [37] (GEO accession: GSE151091).

Gene expression analyses

Differences of SRY expression levels (TPM-normalized) between healthy and tumor samples, and between distinct glioma grades, were assessed using Wilcoxon rank tests (R package *ggpubr* version 0.5.0). Boxplots were generated using the R package *ggplot2* (version 3.4.0).

For men with HGG or SKCM, we performed differential gene expression analyses between individuals with high (TPM ≥ 0.1) or low (TPM < 0.1) SRY expression using DESeq2 [72]. Only genes with absolute log2(fold change) > 1 and a false discovery rate (FDR) adjusted p-value < 0.05 were considered as differentially expressed. Differentially expressed SRY target genes were identified based on previous ChIP-Chip experiments performed by Li and collaborators [28].

Functional and protein-protein network analyses

To elucidate the potential biological impact of SRY expression in men with HGG or SKCM, we performed functional enrichment analyses of differentially expressed SRY targets using the ShinyGO tool (v 0.77) [73]. As input reference datasets, we used KEGG pathways (Release 86.1, https://www.genome.jp/kegg) and cancer hallmarks from the Molecular Signatures Database - MSigDB (Release 6.1, http://www.gsea-msigdb.org/gsea/msigdb/).

Protein-protein networks of differentially expressed SRY target proteins were constructed using the STRING database (v 2023) [74] and Cytoscape (v 3.8.0, https://cytoscape.org). Only experimental and text-mining protein-protein interactions with scores > 0.4 were considered, and only proteins with at least one interaction were maintained.

Cell fraction imputation of healthy skin samples

To estimate the proportions of melanocytes, keratinocytes, eccrine, dendritic

cells, and T-cells in RNA-Seq samples obtained from GTEx (467 healthy skin samples), we used the CIBERSORTx tool [36]. This tool uses support vector regression combined with prior knowledge on scRNA-Seq expression profiles (reference datasets) to estimate the abundance of distinct cell subpopulations in bulk RNA-Seq samples. As input for the reference dataset, we provided the scRNA-Seq expression profile of marker genes used to cluster the five cell types in healthy skin samples from a previous study [37]. Imputation of cell fractions was run in CIBERSORTx with batch correction enabled (B-mode), disabled quantile normalization, relative run mode, and 1,000 permutations. Only samples with a p-value < 0.05 were considered for further analyses.

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REFERENCES

Clocchiatti A, Cora E, Zhang Y, *et al*. Sexual dimorphism in cancer. *Nat Rev Cancer* 2016; **16**: 330-339

Siegel RL, Miller KD, Wagle NS, *et al*. Cancer statistics, 2023. *CA Cancer J Clin* 2023; **73**: 17-48

Cook MB, Dawsey SM, Freedman ND, *et al*. Sex disparities in cancer incidence by period and age. *Cancer Epidemiol Biomarkers Prev* 2009; **18**: 1174-1182

Dorak MT, Karpuzoglu E. Gender differences in cancer susceptibility: an inadequately addressed issue. *Front Genet* 2012; **3**: 268

Skaletsky H, Kuroda-Kawaguchi T, Minx PJ, *et al*. The male-specific region of the human Y chromosome is a mosaic of discrete sequence classes. *Nature* 2003; **423**: 825-837

Kashimada K, Koopman P. Sry: the master switch in mammalian sex determination. *Development* 2010; **137**: 3921-3930

Hughes JF, Page DC. The history of the Y chromosome in man. *Nat Genet* 2016; **48**: 588-589

Mäkelä J-A, Koskenniemi JJ, Virtanen HE, *et al*. Testis Development. *Endocr Rev* 2019; **40**: 857-905

Lucas-Herald AK, Bashamboo A. Gonadal development. *Endocr Dev* 2014; **27**: 1-16

Ahmed SF, Alimusina M, Batista RL, *et al*. The Use of Genetics for Reaching a Diagnosis in XY DSD. *Sex Dev* 2022; **16**: 207-224

Liu C, Ren Y-F, Dong J, *et al*. Activation of SRY accounts for male-specific hepatocarcinogenesis: Implication in gender disparity of hepatocellular carcinoma. *Cancer Lett* 2017; **410**: 20-31

Xue T-C, Zhang L, Ren Z-G, *et al*. Sex-determination gene SRY potentially associates with poor prognosis but not sex bias in hepatocellular carcinoma. *Dig Dis Sci* 2015; **60**: 427-435

Murakami S, Ninomiya W, Sakamoto E, *et al*. SRY and OCT4 Are Required for the Acquisition of Cancer Stem Cell-Like Properties and Are Potential Differentiation Therapy Targets. *Stem Cells* 2015; **33**: 2652-2663

Correa RG, Krajewska M, Ware CF, *et al*. The NLR-related protein NWD1 is associated with prostate cancer and modulates androgen receptor signaling. *Oncotarget* 2014; **5**: 1666-1682

Saginala K, Barsouk A, Aluru JS, *et al*. Epidemiology of Melanoma. *Med Sci (Basel)* 2021; **9**

Lopes FCPS, Sleiman MG, Sebastian K, *et al*. UV Exposure and the Risk of Cutaneous Melanoma in Skin of Color: A Systematic Review. *JAMA Dermatol* 2021; **157**: 213-219

Hartman RI, Lin JY. Cutaneous Melanoma-A Review in Detection, Staging, and Management. *Hematol Oncol Clin North Am* 2019; **33**: 25-38

Petrie T, Samatham R, Witkowski AM, *et al*. Melanoma Early Detection: Big Data, Bigger Picture. *J Invest Dermatol* 2019; **139**: 25-30

Rabbie R, Ferguson P, Molina-Aguilar C, *et al*. Melanoma subtypes: genomic profiles, prognostic molecular markers and therapeutic possibilities. *J Pathol* 2019; **247**: 539-551

Babacan NA, Eroglu Z. Treatment Options for Advanced Melanoma After Anti-PD-1 Therapy. *Curr Oncol Rep* 2020; **22**: 38

Luke JJ, Flaherty KT, Ribas A, *et al*. Targeted agents and immunotherapies: optimizing outcomes in melanoma. *Nat Rev Clin Oncol* 2017; **14**: 463-482

Wen PY, Kesari S. Malignant gliomas in adults. *N Engl J Med* 2008; **359**: 492-507

Stupp R, Brada M, van den Bent MJ, *et al*. High-grade glioma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol* 2014; **25 Suppl 3**: iii93-iii101

Ostrom QT, Price M, Neff C, *et al*. CBTRUS Statistical Report: Primary Brain and Other Central Nervous System Tumors Diagnosed in the United States in 2015–2019. *Neuro Oncol* 2022; **24**: v1-v95

Yang W, Warrington NM, Taylor SJ, *et al*. Sex differences in GBM revealed by analysis of patient imaging, transcriptome, and survival data. *Sci Transl Med* 2019; **11**

Dyba T, Randi G, Bray F, *et al*. The European cancer burden in 2020: Incidence and mortality estimates for 40 countries and 25 major cancers. *Eur J Cancer* 2021; **157**: 308-347

Murakami S, Chishima S, Uemoto H, *et al*. The male-specific factor Sry harbors an

oncogenic function. *Oncogene* 2014; **33**: 2978-2986

Li Y, Zheng M, Lau Y-FC. The sex-determining factors SRY and SOX9 regulate similar target genes and promote testis cord formation during testicular differentiation. *Cell Rep* 2014; **8**: 723-733

Nelles JL, Hu W-Y, Prins GS. Estrogen action and prostate cancer. *Expert Rev Endocrinol Metab* 2011; **6**: 437-451

Guardia GDA, Correa BR, Araujo PR, *et al*. Proneural and mesenchymal glioma stem cells display major differences in splicing and lncRNA profiles. *NPJ Genom Med* 2020; **5**: 2

Bouvier C, Nihous H, Macagno N. [Soft tissue tumours with FN1 (Fibronectin 1) fusion gene]. *Ann Pathol* 2022; **42**: 242-248

Bui TM, Wiesolek HL, Sumagin R. ICAM-1: A master regulator of cellular responses in inflammation, injury resolution, and tumorigenesis. *J Leukoc Biol* 2020; **108**: 787-799

Verginadis II, Avgousti H, Monslow J, *et al*. A stromal Integrated Stress Response activates perivascular cancer-associated fibroblasts to drive angiogenesis and tumour progression. *Nat Cell Biol* 2022; **24**: 940-953

Kisling SG, Natarajan G, Pothuraju R, *et al*. Implications of prognosis-associated genes in pancreatic tumor metastasis: lessons from global studies in bioinformatics. *Cancer Metastasis Rev* 2021; **40**: 721-738

Simone TM, Higgins CE, Czekay R-P, *et al*. SERPINE1: A Molecular Switch in the Proliferation-Migration Dichotomy in Wound-'Activated' Keratinocytes. *Adv Wound Care* 2014; **3**: 281-290

Newman AM, Steen CB, Liu CL, *et al*. Determining cell type abundance and expression from bulk tissues with digital cytometry. *Nat Biotechnol* 2019; **37**: 773-782

Belote RL, Le D, Maynard A, *et al*. Human melanocyte development and melanoma dedifferentiation at single-cell resolution. *Nat Cell Biol* 2021; **23**: 1035-1047

Sigismund S, Avanzato D, Lanzetti L. Emerging functions of the EGFR in cancer. *Mol Oncol* 2018; **12**: 3-20

Pastwińska J, Karaś K, Karwaciak I, *et al*. Targeting EGFR in melanoma - The sea of possibilities to overcome drug resistance. *Biochim Biophys Acta Rev Cancer* 2022; **1877**: 188754

Esche C, Subbotin VM, Maliszewski C, *et al*. FLT3 ligand administration inhibits tumor growth in murine melanoma and lymphoma. *Cancer Res* 1998; **58**: 380-383

Cueto FJ, Sancho D. The Flt3L/Flt3 Axis in Dendritic Cell Biology and Cancer Immunotherapy. *Cancers* 2021; **13**

Kim HJ, Lee HN, Jeong MS, *et al*. Oncogenic KRAS: Signaling and Drug Resistance. *Cancers* 2021; **13**

Barkovskaya A, Buffone A Jr, Žídek M, *et al*. Proteoglycans as Mediators of Cancer Tissue Mechanics. *Front Cell Dev Biol* 2020; **8**: 569377

Chen S, Ding H, Wang K, *et al*. Inhibition of Wnt7b reduces the proliferation, invasion, and

migration of colorectal cancer cells. *Mol Biol Rep* 2023; **50**: 1415-1424

Bueno MLP, Saad STO, Roversi FM. WNT5A in tumor development and progression: A comprehensive review. *Biomed Pharmacother* 2022; **155**: 113599

Menck K, Heinrichs S, Wlochowitz D, *et al*. WNT11/ROR2 signaling is associated with tumor invasion and poor survival in breast cancer. *J Exp Clin Cancer Res* 2021; **40**: 395

Shiba-Ishii A. Significance of stratifin in early progression of lung adenocarcinoma and its potential therapeutic relevance. *Pathol Int* 2021; **71**: 655-665

Hao S, Yang C, Song P, *et al*. CLDN4 promotes growth of acute myeloid leukemia cells via regulating AKT and ERK1/2 signaling. *Biochem Biophys Res Commun* 2022; **619**: 137-143

Ruan S, Shi J, Wang M, *et al*. Analysis of Multiple Human Tumor Cases Reveals the Carcinogenic Effects of PKP3. *J Healthc Eng* 2021; **2021**: 9391104

Maeda T, Desouky J, Friedl A. Syndecan-1 expression by stromal fibroblasts promotes breast carcinoma growth in vivo and stimulates tumor angiogenesis. *Oncogene* 2006; **25**: 1408-1412

Suteau V, Munier M, Briet C, *et al*. Sex Bias in Differentiated Thyroid Cancer. *Int J Mol Sci* 2021; **22**

Costa AR, Lança de Oliveira M, Cruz I, *et al*. The Sex Bias of Cancer. *Trends Endocrinol Metab* 2020; **31**: 785-799

Colopi A, Fuda S, Santi S, *et al*. Impact of age and gender on glioblastoma onset, progression, and management. *Mech Ageing Dev* 2023; **211**: 111801

Mahboobifard F, Pourgholami MH, Jorjani M, *et al*. Estrogen as a key regulator of energy homeostasis and metabolic health. *Biomed Pharmacother* 2022; **156**: 113808

Russell N, Grossmann M. MECHANISMS IN ENDOCRINOLOGY: Estradiol as a male hormone. *Eur J Endocrinol* 2019; **181**: R23-R43

Cisternas CD, Garcia-Segura LM, Cambiasso MJ. Hormonal and genetic factors interact to control aromatase expression in the developing brain. *J Neuroendocrinol* 2018; **30**

Azcoitia I, Mendez P, Garcia-Segura LM. Aromatase in the Human Brain. *Androg Clin Res Ther* 2021; **2**: 189-202

Larney C, Bailey TL, Koopman P. Switching on sex: transcriptional regulation of the testis-determining gene Sry. *Development* 2014; **141**: 2195-2205

Haqq CM, King CY, Donahoe PK, *et al*. SRY recognizes conserved DNA sites in sex-specific promoters. *Proc Natl Acad Sci U S A* 1993; **90**: 1097-1101

Hernández-Vega AM, Del Moral-Morales A, Zamora-Sánchez CJ, *et al*. Estradiol Induces Epithelial to Mesenchymal Transition of Human Glioblastoma Cells. *Cells* 2020; **9**

Diaz-Ruano AB, Martinez-Alarcon N, Perán M, *et al*. Estradiol and Estrone Have Different Biological Functions to Induce NF-κB-Driven Inflammation, EMT and Stemness in ER+ Cancer Cells. *Int J Mol Sci* 2023; **24**

Di Zazzo E, Galasso G, Giovannelli P, *et al*. Estrogen Receptors in Epithelial-Mesenchymal

Transition of Prostate Cancer. *Cancers* 2019; **11**

Bouris P, Skandalis SS, Piperigkou Z, *et al.* Estrogen receptor alpha mediates epithelial to mesenchymal transition, expression of specific matrix effectors and functional properties of breast cancer cells. *Matrix Biol* 2015; **43**: 42-60

Liu Y, Zhao R, Chi S, *et al.* Is Upregulated by Estrogen and Promotes Epithelial-Mesenchymal Transition via p53 in Endometrial Cancer. *Mol Cancer Res* 2020; **18**: 204-215

Kayhan G, Calis P, Karcaaltincaba D, *et al.* Prenatal diagnosis of campomelic dysplasia due to deletion. *J Obstet Gynaecol* 2019; **39**: 1175-1176

Csukasi F, Duran I, Zhang W, *et al.* Dominant-negative SOX9 mutations in campomelic dysplasia. *Hum Mutat* 2019; **40**: 2344-2352

Haseeb A, Kc R, Angelozzi M, *et al.* SOX9 keeps growth plates and articular cartilage healthy by inhibiting chondrocyte dedifferentiation/osteoblastic redifferentiation. *Proc Natl Acad Sci U S A* 2021; **118**

Liu J, Lichtenberg T, Hoadley KA, *et al.* An Integrated TCGA Pan-Cancer Clinical Data Resource to Drive High-Quality Survival Outcome Analytics. *Cell* 2018; **173**: 400-416.e11

Neftel C, Laffy J, Filbin MG, *et al.* An Integrative Model of Cellular States, Plasticity, and Genetics for Glioblastoma. *Cell* 2019; **178**: 835-849.e21

Jacob F, Salinas RD, Zhang DY, *et al.* A Patient-Derived Glioblastoma Organoid Model and Biobank Recapitulates Inter- and Intra-tumoral Heterogeneity. *Cell* 2020; **180**: 188-204.e22

Hao Y, Hao S, Andersen-Nissen E, *et al.* Integrated analysis of multimodal single-cell data. *Cell* 2021; **184**: 3573-3587.e29

Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* 2014; **15**: 550

Ge SX, Jung D, Yao R. ShinyGO: a graphical enrichment tool for animals and plants. *Bioinformatics* December 2019

Szklarczyk D, Kirsch R, Koutrouli M, *et al.* The STRING database in 2023: protein-protein association networks and functional enrichment analyses for any sequenced genome of interest. *Nucleic Acids Res* 2023; **51**: D638-D646