Estimation of contemporary effective population size in plant populations: limitations of genomic datasets

Roberta Gargiulo[1]*, Véronique Decroocq[2], Santiago C. González-Martínez[3], Ivan Paz-Vinas[4], Jean-Marc Aury[5], Isabelle Lesur Kupin[3], Christophe Plomion[3], Sylvain Schmitt[6], Ivan Scotti[7], Myriam Heuertz[3]

Royal Botanic Gardens, Kew, Richmond, Surrey, UK

INRAE, UMR 1332 BFP, Virologie, Villenave d'Ornon, France

INRAE, Univ. Bordeaux, Biogeco, Cestas, France

Department of Biology, Colorado State University, Fort Collins, Colorado, USA

Génomique Métabolique, Genoscope, Institut François Jacob, CEA, CNRS, Univ Evry, Université Paris-Saclay, Evry, France

AMAP, Univ. Montpellier, CIRAD, CNRS, INRAE, IRD, Montpellier, France

INRAE, URFM, Avignon, France

*corresponding author: r.gargiulo@kew.org; robertaxgargiulo@gmail.com

Abstract

Effective population size (N_e) is a pivotal evolutionary parameter with crucial implications in conservation practice and policy. Genetic methods to estimate N_e have been preferred over demographic methods because they rely on genetic data rather than time-consuming ecological monitoring. Methods based on linkage disequilibrium, in particular, have become popular in conservation as they require a single sampling and provide estimates that refer to recent generations. A recently developed software based on linkage disequilibrium, GONE, looks particularly promising to estimate contemporary and recent-historical N_e (up to 200 generations in the past). Genomic datasets from non-model species, especially plants, may present some constraints to the use of GONE, as linkage maps and reference genomes are seldom available, and SNPs genotyping is usually based on reduced-representation methods. In this study, we use empirical datasets from four plant species to explore the limitations of plant genomic datasets when estimating N_e using the algorithm implemented in GONE, in addition to exploring some typical biological limitations that may affect N_e estimation using the linkage disequilibrium method, such as the occurrence of population structure. We show how accuracy and precision of N_e estimates potentially change with the following factors: occurrence of missing data, limited number of SNPs/individuals sampled, and lack of information about the location of SNPs on chromosomes, with the latter producing a significant bias, previously unexplored with empirical data.

Keywords

conservation genomics, effective population size, GONE, linkage disequilibrium, plants

Introduction

Effective population size (N_e) is an evolutionary parameter introduced by Sewall Wright (Wright 1931)**,** which determines the rate of genetic change due to genetic drift and is therefore linked with inbreeding and loss of genetic variation in populations, including adaptive potential (Franklin 1980; Jamieson and Allendorf 2012; Waples 2022). The importance of contemporary effective population size in conservation biology is increasingly recognized and the concept implemented in conservation practice (Luikart et al. 2010; Frankham et al. 2014; Montes et al. 2016) and policy (Hoban et al. 2013; Graudal et al. 2014; Kershaw et al. 2022; O'Brien et al. 2022). For example, N_e has been included as a headline genetic indicator to support Goal A and Target 4 of the Kunming-Montreal Global Biodiversity Framework of the UN's Convention on Biological Diversity (CBD 2022), as the proportion of populations within species with $N_e > 500$, that are expected to have sufficient genetic diversity to adapt to environmental change (Jamieson and Allendorf 2012; Hoban et al. 2020).

Contemporary N_e can be estimated using demographic or genetic methods (Wright 1969; Luikart et al. 2010; Wang et al. 2016; Waples 2016; Felsenstein 2019). Demographic estimators require detailed ecological observations over time for the populations of interest (Wright 1969; Nunney 1993; Felsenstein 2019), which is not necessary for genetic estimators (Wang et al. 2016; Waples 2016). Methods that can provide N_e estimates based on a single sampling point in time (Wang 2016) have become particularly popular, especially in studies focused on species for which budget and time allocated are limited, elusive species that are difficult to track and monitor (Luikart et al. 2010), and species for which information about distribution is scarce. The current biodiversity crisis and the limited resources for conservation have recently fuelled the development and application of N_e estimators that rely on cost-effective, non-genetic proxy data across a wide range of species of conservation concern (Hoban et al. 2020, 2021a). Population census size, N_C, has been used to infer N_e when genetic N_e estimates are not available, relying on the ratio $N_e/N_C = 0.1$ (where N_C is the adult census size of a population) (Palstra and Fraser 2012; Frankham et al. 2014; Hoban et al. 2021b). This rule-of-thumb ratio is pragmatic for conservation (but see Fady and Bozzano 2021), as shown in application tests in different countries for different species of conservation concern (Thurfjell et al. 2022; Hoban et al. 2023). However, research needs to progress to better understand N_e estimation methods and potential deviations from the ratio $N_e/N_C = 0.1$, which are expected for example across populations within species or in species with life-history traits that favour individual persistence (Jamieson and Allendorf 2012; Hoban et al. 2020, 2021b; Frankham 2021; Laikre et al. 2021; Gargiulo et al. 2023). Current genetic estimators of contemporary N_e work well in small and isolated populations, which match many populations of conservation concern, but they are difficult to apply in species with a large and continuous distribution (Fady and Bozzano 2021; Santos-del-Blanco et al.

2022). In such species, genetic isolation by distance, overlapping generations, and difficulty to define representative sampling strategies can affect the accuracy of estimates of N_C, N_e and their ratio (Neel et al. 2013; Nunney 2016; Santos-del-Blanco et al. 2022). Plant species embody some of the features mentioned above, as they often have complex life-history traits (e.g., overlapping generations, long lifespans), reproductive systems (i.e., mixed clonal and sexual reproduction, mixed selfing and outcrossing strategies) and continuous distribution ranges (Petit and Hampe 2006; De Kort et al. 2021). Therefore, they are particularly interesting to help improve our understanding of N_e estimation methods.

Genetic drift generates associations between alleles at different loci, known as linkage disequilibrium (LD), at a rate inversely proportional to N_e (Waples et al. 2016). LD between loci can be used to obtain a robust estimate of contemporary N_e from genetic data at a single time point, and this explains the popularity of the LD method compared to the earlier developed two-sample temporal methods (for an early review, see Luikart et al. 2010) and the development of numerous tools for the estimation of LDN_e from genetic and genomic data (Do et al. 2014; Barbato et al. 2015; Wang et al. 2016; Santiago et al. 2020). The N_e estimates obtained with the LD method generally refer to a few generations back in time (Luikart et al. 2010; Do et al. 2014) and, depending on the genetic distances between loci, it is possible to obtain N_e at different times in the past (see the review on timescales of N_e estimates in Nadachowska-Brzyska et al. 2022). In particular, LD between closely linked loci can be used to estimate N_e over the historical past (Sved 1971; Hayes et al. 2003; Qanbari et al. 2010; Do et al. 2014; Barbato et al. 2015; Wang et al. 2016; Santiago et al. 2020), whereas loosely linked or unlinked loci can be used to estimate N_e in the recent past (Waples 2006; Waples and Do 2008; Sved et al. 2013; Wang et al. 2016; Qanbari 2019). However, as other methods to estimate N_e, the LD method is not devoid of biases and drawbacks, mostly relating to the assumption that the population is isolated, which is rarely satisfied (Hill 1981; England et al. 2010; Waples and England 2011), and to the occurrence of age-structure in populations (Nunney 1991; Yonezawa 1997; Waples and Do 2010; Robinson and Moyer 2013; Waples et al. 2014; Hössjer et al. 2016; Ryman et al. 2019).

In this study, we aimed to explore the limitations of plant genomic datasets when estimating contemporary N_e. We mostly focused on estimating N_e using the software GONE (Santiago et al. 2020), but we also provide N_e estimates obtained in NeEstimator (Do et al. 2014). These programmes provide recent historical and contemporary N_e estimates, respectively, using the LD method, though they differ mostly in the data requirement and timescales of estimates provided.

In particular, we explored the technical requirements of the software GONE by conducting power analyses aimed at testing how the number of SNPs, the proportion of missing data, the number of

individuals, the lack of information about the location of SNPs on chromosomes, and the occurrence of population structure could affect N_e estimation. The N_e estimates obtained in GONE were then compared to the ones obtained in NeEstimator and discussed in light of the biological and ecological features of the species.

Our findings help better understand the limitations and potentialities of genomic datasets when estimating LD-based, one-sample N_e, providing new insights on how to use current methods.

Methods

Datasets

We selected four datasets obtained with different high-throughput sequencing techniques from different plant taxa (*Symphonia globulifera* L.f. (Clusiaceae), *Mercurialis annua* L. (Euphorbiaceae), *Fagus sylvatica* L. (Fagaceae), *Prunus armeniaca* L. (Rosaceae)), to represent different botanical groups, ecosystems, generation times and reproductive strategies. Sampling strategies in the datasets encompassed different sample sizes for markers and individuals, and datasets featured distinct levels of population genetic structure (Table 1).

For boarwood, *S. globulifera* s.l., a widespread and predominantly outcrossing evergreen tree typical of mature rainforests in Africa and the Neotropics (Degen et al. 2004; Torroba-Balmori et al. 2017), we used the targeted sequence capture dataset described in Schmitt et al. (Schmitt et al. 2021). Three sympatric gene pools were identified in a lowland forest in French Guiana, likely corresponding to three biological species, described as *Symphonia* sp. 1, *Symphonia* sp. 2 and *Symphonia* sp. 3 (Schmitt et al. 2021). To avoid the influence of admixture on the estimation of N_e, we first divided the dataset in three subsets based on the analysis of genetic structure performed in (Schmitt et al. 2021), selecting only the individuals with a Q-value (cluster membership coefficient) $\geq 95\%$ to each of the three genetic clusters (Species 1, Species 2 and Species 3; Supplementary File 1). We then selected the 125 genomic scaffolds with the largest number of SNPs (see Table 1).

For the annual mercury, *M. annua*, an annual plant with great variety of mating systems (monoecious, dioecious, androdioecious) and ploidy levels (2x, 4x-12x) (Obbard et al. 2006b, a), typical of open or disturbed habitats in Europe and North Africa, we used the gene capture data set described in (González-Martínez et al. 2017), obtained from several diploid dioecious populations representative of three main gene pools in the species (total sample size = 40 individuals). We selected the 48

scaffolds with the largest number of SNPs and ran the analyses by considering separately each gene pool: (1) ancestral populations from Turkey and Greece, (2) range-front populations from northeastern Spain, or (3) range-front populations from northern France and the UK (see Table 1).

For the common beech, *F. sylvatica*, a deciduous predominantly outcrossing tree of European temperate forests (Merzeau et al. 1994), we analysed genomic scaffolds from a single, contiguous stand (plot N1; (Oddou-Muratorio et al. 2021) within a French population (Mt. Ventoux, southeastern France), in which population genetic structure is neither observed nor expected (Csilléry et al. 2014). Mapping of short-reads paired Illumina sequences was independently performed for each one of the 167 individuals of the population against the genome assembly (available at www.genoscope.cns.fr/plants) using bwa-mem2 2.0 (Li and Durbin 2009). SNPs were first called using GATK 3.8 (Van der Auwera and O'Connor 2020) using the following parameters: -nct 20 - variant_index_type LINEAR variant_index_parameter 128000. SNPs were also called using samtools v1.10 / bcftools v1.9 (Danecek et al. 2021) with default parameters. Following these two SNPs calling steps, we performed a three-steps filtering process: (i) only diallelic SNPs were kept, (ii) the minimum allele frequency (MAF, upper case used at the individual level), calculated on the basis of all the reads containing the SNP, was set to 30%, (iii) individual genotypes with sequencing depth less than 10 were recoded into « ./. » meaning that both alleles are missing. We then identified SNPs commonly found by GATK and samtools using the - diff flag of vcftools v0.1.15 with tabix-0.2.5 (Danecek et al. 2011). A nucleotide polymorphism was considered to be a SNP if at least one individual was found to be heterozygous at the position. On average, for each individual, 88.5% of the sequencing reads mapped properly onto the assembly. The final VCF contained 18,192,174 variants available at the Portail Data INRAe (doi:10.57745/FJRYI1).

We re-ordered the 406 genomic scaffolds available based on their number of SNPs, and selected 150 scaffolds with the largest number of SNPs. We tested different combinations of input subsets, with numbers of scaffolds ranging from 12 to 150 (provided that SNPs per scaffold < 1 million and total number of SNPs < 10 millions, see the requirements of GONE below), and numbers of individuals ranging from 5 to 167 (total sample size).

For the apricot, *P. armeniaca*, we estimated N_e using whole genome resequencing data (21× depth of coverage by ILLUMINA technology) for wild Central Asian, self-incompatible populations of the species (Groppi et al. 2021). Variant sites were mapped to the eight chromosomes of the species and ranged between 2.3 and 6.2 million per chromosome (total number of variant sites: 24 M). As these exceeded the total number allowed in GONE, we downsampled the number of SNPs prior to the analyses. We also analysed the datasets by considering the different gene pools recovered in Groppi et al. (2021)

(Supp. Fig. S20), namely the Southern (red cluster) and Northern (yellow cluster) gene pools, as obtained in fastStructure (Raj et al. 2014) (see next subsection).

Data analyses in GONE

Analyses for all species. We performed N_e estimation in the software GONE (Santiago et al. 2020). GONE generates contemporary or recent historical estimates of Ne (i.e. in the 100-200 most recent generations) using the LD method. GONE requires linkage information, ideally represented by SNPs mapped to chromosomes. Chromosome mapping is rarely available for non-model species, and in our case was only fully available for the apricot (*P. armeniaca*) dataset. In the absence of chromosome mapping information for the other species, we used the linkage information associated with genomic scaffolds. In terms of requirements, GONE accepts a maximum number of chromosomes of 200 and a maximum number of SNPs of 10 million, with a maximum number of SNPs per chromosome of 1 million, although the software uses up to 50,000 random SNPs per chromosome for the computations when the total number of SNP is larger. A complete workflow of the analyses carried out in GONE is available at https://github.com/Ralpina/Ne-plant-genomic-datasets (Gargiulo, 2023).

Influence of missing data on N_e estimation. The influence of missing data on N_e estimation in GONE was evaluated using the dataset from *F. sylvatica*. After keeping 67 individuals with less than 95% missing data, we permuted individuals (without replacement) to generate 150 datasets of 35 individuals, and estimated N_e in GONE for each dataset. Proportion of missing data per individual for each permuted dataset was calculated in vcftools v0.1.16 (Danecek et al. 2011) from an average of ~25% to 95%; results were plotted in R v4.2.2 (R Core Team 2019). In addition, we used the dataset of *P. armeniaca* to evaluate how N_e changed when manually introducing missing data. We selected all individuals from the Northern gene pool with a Q-value (cluster membership coefficient) $\geq$ 99% (77 individuals) to rule out the influence of admixture, and replaced some of the individual genotypes with missing values using a custom script (available at: https://github.com/Ralpina/Ne-plant-genomic-datasets). We generated four datasets, each with a proportion of missing data per individual of 20%, 40%, 60% and 80%, respectively, and then computed N_e in GONE for each dataset obtained.

Influence of number of SNPs on N_e estimation. The influence of the number of SNPs on N_e estimation in GONE was evaluated using the dataset of *P. armeniaca*. From the Northern gene pool, we first selected the individuals with a Q-value $\geq$ 99% to rule out the influence of admixture. We drew random subsets of variant sites (without replacement) including 40K, 80K, 150K, 300K, 500K, 3.5M, 7M, and 10M SNPs, respectively, and generated 50 replicates for each subset; we then estimated N_e in GONE

for each subset and obtained the 95% confidence intervals across the 50 replicate subsets with the same number of SNPs (using the *median* and *quantile* functions in R).

Influence of sample size on N_e estimation. We used the Northern gene pool of *P. armeniaca* to assess how N_e estimates changed depending on the number of samples considered and the uncertainty associated with individual sampling. We first downsampled the number of SNPs to 3.5M (to satisfy GONE requirements), and varied the sample sizes included in the analyses from 15 to 75 (i.e. ~the total number of individuals of the Northern gene pool with a Q-value ≥ 99%). For each sample size group, we generated 50 random subsets (without replacement within the subset) of individuals and estimated N_e in GONE for each random subset; we then estimated 95% confidence intervals across subsets with the same sample size (using the function *stat_summary(fun.data = median_hilow, fun.args = list(conf.int = 0.95)* in R).

Influence of population admixture on N_e estimation. We also evaluated how genetic structure within gene pools influenced N_e estimation in GONE for both the Southern and Northern gene pools of *P. armeniaca*. We first downsampled the number of SNPs to 3.5M to satisfy GONE requirements, as described above. We then divided the individuals of each gene pool into five non-overlapping classes based on individual Q-values (lower bounds of 70%, 80%, 90%, 95%, and 99%), resampled individuals (without replacement) in each Q-value class 50 times, standardising sample sizes to the sample size of the smallest Q-value class within a gene pool (i.e., 21 individuals as in the 99% Q-value class of the Southern gene pool and 77 individuals as in the 99% Q-value class of the Northern gene pool, see Supplementary Table S1 for original sample sizes). We then estimated N_e in GONE and obtained 95% confidence intervals across the 50 resampled datasets of the same Q-value class within a gene pool (using the R function *stat_summary* mentioned above). We also combined all individuals from the two gene pools (255 individuals), resampled 77 individuals 50 times without replacement, and estimated N_e in GONE and the related confidence intervals as explained above.

Effect of using genomic scaffolds rather than chromosomes. We evaluated the effect of using genomic scaffolds to estimate linkage groups when chromosome information is not available. Using the downsampled dataset of 3.5M SNPs from *P. armeniaca*, we selected from the Northern gene pool 45 random individuals with a Q-value ≥ 99%, to rule out the influence of admixture. For this dataset, five different chromosome maps were then created, progressively assigning SNPs to 8 (true value), 16, 32, 64 and 128 chromosomes (as if they were genomic scaffolds). We then estimated N_e in GONE using five corresponding chromosome map files and keeping the same ped (genotypes) file.

Data analyses in NeEstimator

We also used the LD method as implemented in the software NeEstimator v2 (Do et al. 2014) to estimate N_e in our datasets. NeEstimator assumes that SNPs are independently segregating (typically, SNPs at short physical distances, for example those in the same short genomic scaffolds or loci, are filtered previous to analysis, see below), and therefore it provides an N_e estimate based on the LD generated by random genetic drift, which reflects N_e in very recent generations (Waples et al. 2016). However, accuracy and precision will be both affected by (1) the assumption of independent segregation in genomic data sets, as SNPs are necessarily packed on a limited number of chromosomes and thus they provide non-independent information, and especially (2) the occurrence of overlapping pairs of loci, each locus appearing in multiple pairwise comparisons (i.e. two aspects of the issue known as pseudoreplication; (Purcell et al. 2007; Waples et al. 2016; 2022). Although the influence of this issue on bias and precision is difficult to address completely, some bias corrections have been proposed, for example applying a correction based on the genome size of the species being analysed (formula in Waples et al. 2016), using only one SNP per scaffold or thinning scaffolds based on discrete window sizes (Purcell et al. 2007). To explore the changes in precision, we also estimated N_e on datasets thinned by subsampling SNPs using windows of 10,000 positions or 5,000 positions, such that no two SNPs found at a distance smaller than 10,000 or 5,000, respectively, were included in the analyses (*-thin* option in vcftools).

As low-frequency alleles upwardly bias N_e, we followed the recommendations in (Waples and Do 2010) and used a threshold minimum allele frequency, P_{crit}, to filter rare alleles occurring at a frequency lower than 0.02 when sample sizes > 25, and $1/(2S) \leq P_{crit} \leq 1/S$ when sample sizes $\leq$ 25 (where S = sample size), and excluded singleton alleles (Waples and Do 2010). We also ran the analyses without applying a filter for rare alleles. Confidence intervals were obtained via jackknifing over samples (Do et al. 2014; Jones et al. 2016).

Results and Discussion

Our study explores the limitations associated with genomic datasets when estimating N_e using the LD method as implemented in the software GONE. The estimation of N_e in GONE failed for the three biological species of *S. globulifera*, as the software returned the error "too few SNPs" for each of the three species datasets. This was caused by the relatively small number of SNPs per scaffold (averaging ~250 SNPs) and, in turn, by the relatively short length of the scaffolds (length ranging from 5,421 to 931 positions). We then estimated N_e for the three species datasets using NeEstimator. N_e ranged

from 93 (CI: 39-Infinite) in Species 3, to 246 (CI: 199-320) in Species 2 and to 371 (CI: 318-441) in Species 1 (Table 2). These estimates may not reflect the population-wide true effective population size, but rather a quantity close to the neighbourhood size (Ns), i.e. the inverse of the probability of identity by descent of two uniting gametes, which is typical of sparsely sampled continuous populations of forest trees (see Neel et al. 2013; Nunney 2016; Santos-del-Blanco et al. 2022). Moreover, we observed that thinning datasets increased N_e estimates and generated broader CIs, ranging from 213 (CI: 38-Infinite) for Species 3, 773 (CI: 316-Infinite) in Species 2, and 1,382 (CI: 677-56,918) in Species 1 (Table 2). An increase in N_e estimates and their CIs is expected when reducing the number of closely linked SNPs, as thousands of physically linked loci with limited recombination are predicted to bias N_e downward while also narrowing CIs because of pseudoreplication (Waples et al. 2016; 2022). Removing the MAF filter on the full dataset produced the expected inflating effect of rare alleles on N_e (Waples and Do 2010); however, when considering the thinned dataset, including rare alleles reduced N_e estimates rather than increasing them. This suggests that systematically removing SNPs below a certain distance (when thinning), may introduce unexpected patterns of LD which result in N_e estimates that are difficult to predict and interpret.

Influence of missing data on N_e estimation

The effect of missing data on N_e estimation is evident from the results obtained when analysing the dataset of *F. sylvatica*, in both GONE and NeEstimator, and from the results obtained when analysing the dataset of *P. armeniaca* in which genotype data were manually excluded. For *F. sylvatica*, 35 individuals had a proportion of missing data < 50% (Fig. 1B). Increasing the proportion of missing data in the permuted datasets of 35 individuals produced acute increases in N_e estimates in GONE (see Fig. 1A); for instance, increasing the median proportion of missing data per individual from 25% to 35% produced N_e estimates increasing from 200 to 3 millions. Likewise, when missing data proportion per individual of *P. armeniaca* increased above 20%, we obtained N_e estimates that were > 350 times larger than those obtained from the original dataset (average missing data proportion per individual ~ 8%) (Fig. 2). N_e estimation in GONE failed when the datasets included a proportion of missing data exceeding 60% in each individual. This relationship between missing data and N_e estimates is consistent with what was previously found (e.g., Marandel et al. 2020), although the loss of accuracy in the N_e estimation is extreme and suggests that individuals with > 20% missing data should be removed from the dataset before estimating N_e in GONE. However, in the specific case of *F. sylvatica* and in other species with large effective population sizes, reducing the sample size (S) to a number << true N_e introduces a further bias in the N_e estimation using the LD method, in addition to the sampling error already expected because of the finite sample size (Peel et al. 2013).

In NeEstimator, even selecting the 35 individuals with < 50% missing data failed to produce N_e estimates in *F. sylvatica*, in contrast with a previous study showing that the method implemented in the software was relatively robust with a similar percentage of missing data (Nunziata and Weisrock 2018). As NeEstimator handles missing data using a weighted harmonic mean that assumes random and independent occurrence of missing data (Peel et al. 2013; Do et al. 2014), a higher and therefore non-random occurrence of missing data in the longest scaffolds of the beech dataset might have prevented the N_e calculation in NeEstimator.

Figure 1. In (A), ranked median N_e estimates in the most recent generation in 150 datasets of 35 individuals with different proportions of missing data (excluding individuals with a proportion of missing data > 0.95) of *F. sylvatica*; points represent median values and ranges represent standard deviations for the proportion of missing data per individual. Analyses based on the dataset with the twenty-seven genomic scaffolds with the largest number of SNPs (excluding the scaffolds with > 1 M SNPs). In (B), proportion of missing data per individual in the complete dataset of *F. sylvatica*.

Figure 2. Influence of missing data on N_e estimation in GONE. Missing genotypes were manually introduced in the dataset of *P. armeniaca*, generating pseudo-genotypes with an average proportion of missing data ranging from 20% to 80%. The analysis in GONE failed to produce an estimate for missing data proportions of 60% and 80%. The original dataset is shown for comparison (missing data = 8%). Note the different y-scales in the three facets.

Influence of number of SNPs on N_e estimation

The influence of the number of SNPs per chromosome was explored using the dataset from *P. armeniaca* (Northern gene pool), which was the only dataset with SNPs fully mapped to chromosomes. Increasing the number of SNPs per chromosome affected the point N_e estimates only slightly, and influenced the precision of the estimates more obviously, especially for a total number of above 300,000 SNPs, corresponding to an average of 10,000 SNPs per chromosome of *P. armeniaca* used by GONE (Fig. 3). Accuracy and precision of N_e estimates based on LD are expected to be affected by two types of pseudoreplication: (1) the non-independent information content provided by thousands of linked SNPs, and especially (2) the occurrence of overlapping pairs of loci, each locus appearing multiple times in pairwise comparisons (Waples et al. 2016, 2022). Therefore, the narrower confidence intervals we obtained when increasing the number of SNPs are partially due to the

inclusion of overlapping pairs of loci for the N_e estimation, which artificially increases the degrees of freedom that make CIs tight.

For practical purposes, our results show that adding more than 2,000 polymorphic SNPs per chromosome, with a large sample size (~75), does not improve the accuracy of the estimation, in line with what is shown in previous studies focusing on LDN_e (Marandel et al. 2020). (Santiago et al. 2020) noted that the accuracy of the estimation is proportional to sample size and to the square root of SNPs pairs, and therefore researchers might partially compensate for small sample sizes by increasing the number of SNPs. However, as the information content of a dataset depends on the amount of recombination and on the pedigree of the individuals included in the analyses, an estimation based on a small number of samples will not necessarily be representative of the entire population, especially if N_e is large (King et al. 2018; Santiago et al. 2020). Furthermore, the marginal benefit of increasing the number of SNPs beyond tens of thousands is counterbalanced by poor precision if CIs are generated using incorrect degrees of freedom, which is often the case with thousands of non-independent SNPs (Do et al. 2014; Jones et al. 2016; Moran et al. 2019; Luikart et al. 2021; Waples et al. 2022).

The estimates produced by NeEstimator from the datasets of *P. armeniaca* subsampled with two thinning windows (Table 2) revealed high consistency in point estimates and jackknife confidence intervals obtained, regardless of the thinning windows and the P_{crit} used, and despite the number of SNPs being halved in one of the two thinned datasets compared to the other. However, the estimates were lower than those obtained in GONE and based on at least ten times more SNPs. Considering that a downward rather than an upward bias is expected due to pseudoreplication when closely linked and overlapping pairs of loci are used, the higher N_e estimates in GONE can be explained with the higher power associated with many SNPs, and therefore an improved accuracy and precision when thinning datasets is offset by the reduced power associated with reducing the number of SNPs in the analysis (Waples et al. 2022).

Figure 3. N_e estimates obtained in GONE over the most recent generation for the Northern gene pool of *Prunus armeniaca* as a function of the number of SNPs. Points represent median values across 50 replicates; shaded area represents 95% confidence intervals across replicates. Note that GONE uses a maximum of 50,000 SNPs per chromosome, even if provided with a larger number (with 1 million per chromosome being the maximum number accepted); the number of SNPs in each of the eight subsets analysed ranged from 10^4 to 10^7, corresponding to a range of ~5,000 to ~20,000 polymorphic SNPs per chromosome used by GONE.

Influence of sample size on N_e estimation

We evaluated the influence of sample size using the Northern gene pool of *P. armeniaca*. Increasing sample sizes to over thirty samples led to more consistent N_e estimates and reduced the chances of obtaining N_e estimates only representative of a few individual pedigrees (Fig. 4), as previously observed when using the linkage disequilibrium method (Palstra and Ruzzante 2008; Waples and Do 2010; Tallmon et al. 2010; Antao et al. 2011; Waples et al. 2016; Nunziata and Weisrock 2018; Marandel et al. 2019; Santiago et al. 2020). Including in the N_e estimation a number of samples that is representative of the true N_e of the population is crucial in large populations, where the genetic drift signal in recent generations is weak (Palstra and Ruzzante 2008; Luikart et al. 2010; Do et al. 2014; Barbato et al. 2015; Wang et al. 2016; Santiago et al. 2020). On the contrary, small populations

experience more genetic drift, hence the LD method is particularly powerful in such populations. Estimates of N_e remain small in small populations even with larger sample sizes, hence the important conservation implication that small populations cannot be mistaken for large populations (Waples and Do 2010; Waples et al. 2016; Santiago et al. 2020). For the Northern gene pool of wild apricots, we obtained an N_e estimate < 2,000 when sample size was equal to 15, and progressively increasing up to a plateau of N_e ~4,000, for larger sample sizes. This confirms the expectation that a large sample size is needed to estimate a large N_e (Tallmon et al. 2010; Antao et al. 2011).

For *M. annua*, we attempted N_e estimation despite the small sample sizes using both GONE and NeEstimator. All point N_e estimates were generally small (< 50) and confidence intervals were narrow, with larger N_e values for the Core gene pool (Table 2). These estimates are much smaller than those provided by González-Martínez et al. (2017) for present-day, coalescent N_e using extensive demographic modelling in fastsimcoal2, which produced values of about ~500,000 for the Core gene pool and ~40,000-50,000 for the other gene pools. In *M. annua*, we did not observe substantial differences in the N_e estimates obtained from the thinned datasets (for example an increase in N_e estimates due to partially removing the influence of pseudoreplication, as observed for *S. globulifera*) and this suggests that the downward bias caused by a small sample size is potentially much stronger than the downward bias associated with pseudoreplication (Table 2).

Figure 4. Change in the N_e estimates as a function of sample size in *P. armeniaca* (Northern gene pool). Ranges represent 95% confidence intervals estimated by generating random subsets of individuals (without replacement) 50 times. The insert shows a magnified view of the N_e estimates in the main plot (y-axis limit = 5000).

Influence of admixture on N_e estimation

The impact of admixture on N_e estimation was explored using the dataset of *P. armeniaca*. Estimates of N_e in the most recent generation generally decreased when the Q-value of the individuals included in the analysis increased (Fig. 5A). The larger N_e estimates in the most recent generations (1-4) when including more admixed individuals are consistent with the upward bias predicted by Waples and England (Waples and England 2011) for a sampled subpopulation that does not include all potential parents ("drift LD"); with higher admixture proportions (Fig. 5A), the N_e estimated for each gene pool (subpopulation) using the LD method tends to approach the N_e of the metapopulation instead (Waples and England 2011). However, the N_e estimate we obtained when combining the two gene pools ("all" in Fig. 5A) is lower than the N_e estimate obtained when considering highly admixed individuals in the Northern gene pool (70% in the right panel of Fig. 5A). This is consistent with the expected downward

N_e bias associated with a "mixture LD", caused by the Wahlund effect associated with the occurrence of more than one gene pool ((Waples and England 2011); (Waples and England 2011; Neel et al. 2013; Nunney 2016; Santos-del-Blanco et al. 2022)). The Southern gene pool showed a contrasting trend; N_e estimates remain lower than those obtained when combining the two gene pools, possibly because the small sample size for this gene pool contributed less to any potential mixture LD than the LD signal from individuals of the Northern gene pool (Fig. 5A).

Over the last 25 generations (Fig. 5B), we obtained higher N_e estimates when individuals from the Southern gene pool with a Q-value ≥ 99% were included. For the Northern gene pool, on the contrary, we obtained a lower N_e estimate when individuals with a Q-value ≥ 99% were included. This certainly depends on their different demographic histories, as the Southern gene pool seems to have undergone a recent bottleneck, whereas the Northern gene pool has a more stable demographic trend. The recent population decline for the Southern gene pool may be explained by the Soviet era and the current land-use change in the Fergana valley (mainly Uzbekistan) where the native forests were partially replaced with crop species. Nevertheless, there are more factors to consider for the interpretation of these divergent trends. First, the sample size of the Southern gene pool is smaller than that of the Northern gene pool (only 21 individuals vs. 77 individuals drawn from each Q-value group). Second, Santiago et al. (Santiago et al. 2020) warn about a typical artefactual bottleneck observed in GONE and caused by population structure (in Figure 2F of (Santiago et al. 2020), considering a migration rate = 0.2%). As we observed a consistent trend regardless of the individual Q-value, and the drop in N_e is particularly evident with a Q-value = 99%, we interpret this N_e drop as a true bottleneck, with the caveat of reduced accuracy linked to a small sample size for the Southern gene pool.

Figure 5. Influence of population structure on GONE N_e estimates for the Northern and Southern gene pools of *P. armeniaca*. Q-values refer to the results of the fastStructure analysis performed in Groppi et al. (2021) (lower bounds of individual Q-value to the main genetic cluster). N_e was estimated over 50 datasets of resampled individuals (77 in each Q-value class in the Northern gene pool and 21 in each Q-value class in the Southern gene pool, reflecting differences in sample sizes); in (B), only median values of the N_e estimates across 50 datasets and in the last 25 generations are shown. N_e estimates obtained for the combined gene pools are also shown ("all" in A and "all inds" in B).

Effect of using genomic scaffolds rather than chromosomes

To evaluate the effect of using genomic scaffolds as a proxy for linkage groups when chromosome information is not available, we sorted SNPs from the *P. armeniaca* dataset into a progressively larger number of scaffolds or chromosomes assumed. This produced inconsistent N_e estimates across the

datasets with increasing number of chromosomes assumed, with N_e values progressively rising from around 3×10^3 for 8 chromosomes (true value) to $> 8 \times 10^5$ when the number of chromosomes assumed was equal to 128 (Fig. 6). The algorithm implemented in GONE is based on the assumption that LD among pairs of SNPs at different genetic distances provides differential information about N_e at different times in the past (Santiago et al. 2020). Loosely linked loci give information about N_e in recent generations, as their recombination rate is higher and rate of LD-decay slower than that of closely linked loci (Sved and Feldman 1973). Therefore, the behaviour of the N_e estimates observed in Fig. 6 can be explained by considering that assigning potentially linked SNPs to different chromosomes free to recombine causes their LD to decay more slowly, overestimating N_e. This finding suggests that the N_e estimates found in GONE for *M. annua* and *F. sylvatica* may be further biased since, for both datasets, scaffolds were used as a proxy for chromosomes (Table 1).

Figure 6. Estimates of N_e calculated on datasets in which the same set of SNPs is assigned to a progressively larger number of assumed chromosomes, where 8 is the true number of chromosomes for *P. armeniaca* (per haploid count); 45 individuals from the Northern gene pool were used for this analysis.

Practical recommendations when estimating contemporary N_e in GONE

In this study, we have considered some of the technical limitations when estimating N_e for plant genomic datasets, including: (i) the occurrence of missing data, (ii) the limited number of SNPs/individuals sampled, (iii) the lack of genetic/linkage maps and of information about how SNPs map to chromosomes when estimating N_e using the software GONE. In addition, we have explored some biological limitations that may affect N_e estimation using the LD method, such as the occurrence of population structure. Our empirical results corroborate some previous findings, for example about the importance of having large samples sizes (ideally > 30 per subpopulation), especially when populations are large, and highlight the following requirements that genomic datasets should satisfy:

- non-random missing data should not exceed 20% per individual. Missing data also affect how SNPs are represented across loci and individuals sampled and can generate non-random patterns whose effect on N_e estimation is difficult to predict;
- having a large number of SNPs (> tens of thousands) is potentially important to allow users to generate non-overlapping subsets of loci that reduce the influence of pseudoreplication on confidence intervals (Waples et al. 2022). However, increasing the number of SNPs beyond a few thousands per chromosome does not produce significant changes in N_e estimates, as we observed in wild apricots;
- most importantly, having SNPs fully mapped to chromosomes is essential to obtain reliable estimates when using the software GONE.

In addition, the bias on N_e estimates due to the occurrence of gene flow and admixture can significantly affect the performance of single-sample estimators, as previously described (e.g., Neel et al. 2013). This bias and potential other biases associated with (i) further sources population structure (i.e. overlapping generations, demographic fluctuations including bottlenecks, reproductive strategies causing variance in reproductive success, etc.) and (ii) further technical issues associated with sampling strategies and genomic datasets can add up and generate results that are misleading for conservation. Therefore, a careful consideration of the issues above is essential when designing and interpreting studies focused on the estimation of N_e and other related indicators for conservation.

Acknowledgements and Funding information

This study was carried out within the short-term scientific mission "Estimating effective population size in genomic datasets: test of methods and assumptions", organised by Working Group 2 of the European Cost Action CA18134 "Genomic Biodiversity Knowledge for Resilient Ecosystems (G-BiKE)" . The work on *F. sylvatica* was supported by the Genoscope, the Commissariat à l'Énergie Atomique et aux Énergies Alternatives (CEA) and France Génomique (ANR-10-INBS-09–08). We are grateful to the genotoul bioinformatics platform Toulouse Occitanie (Bioinfo Genotoul, https://doi.org/10.15454/1.5572369328961167E12), to the Bordeaux Bioinformatics Center (CBiB), and to the Royal Botanic Gardens, Kew HPC (KewHPC) for providing computing and storage resources. We thank Enrique Santiago and Armando Caballero for their suggestions on how to interpret parameters and results using the software GONE. We thank Iris Bienbach, Alice Brambilla, Christine Grossen, Jo Howard-McCombe, and all the other members of the G-BiKE Working Group 2 chaired by Mike Bruford for the useful discussions about N_e estimation methods and strategies. IP-V is supported by the U.S. Geological Survey Powell Center for Synthesis and Analysis.

Data and code availability

The SNP matrices used in this study can be accessed at the following links: https://doi.org/10.5281/zenodo.4727831 (*Symphonia globulifera*), https://datadryad.org/stash/dataset/doi:10.5061/dryad.74631 (*Mercurialis annua*), https://doi.org/10.57745/FJRYI1 (*Fagus sylvatica*), https://doi.org/10.5281/zenodo.8124822 (*Prunus armeniaca*). The analyses carried out in this study and the related scripts are available at: https://github.com/Ralpina/Ne-plant-genomic-datasets (Gargiulo, 2023).

Table 1. Details of the different plant genomic datasets analysed in the present study.

Species name	Life-form	Reproductive system of populations analysed	Gene pools (#samples)	Data type	Average frequency of missing data per individual	#chromosomes/scaffolds/contigs analysed in GONE	Average #SNPs per scaffold or chromosome*	Total #SNPs **	Reference	Issues explored (affecting *Ne* estimation in GONE)
Symphonia globulifera L.f.	Perennial (tree)	Monoecious, mixed mating with predominant outcrossing (Degen et al. 2004)	Species 1 (228) Species 2 (107) Species 3 (30)	Targeted sequence capture	0.04	125 (contigs)	247	30,863	Schmitt et al. 2021	Minimum number of SNPs required
Mercurialis annua L.	Annual	Various mating systems, analyses based on dioecious populations; obligate outcrosser (González-Martínez et al. 2017)	Atlantic (12) Core (16) Mediterranean (12)	Targeted gene (exome) capture	0.01	48 (contigs)	670	32,151	González-Martínez et al. 2017	Influence of sample size
Fagus sylvatica L.	Perennial (tree)	Monoecious, predominant outcrossing (Merzeau et al. 1994)	Mt. Ventoux, France (167)	Whole genome sequencing	0.81 (with 27 scaffolds)	12-150 (scaffolds)	~470K (with 27 scaffolds)	~13 M (with 27 scaffolds)	See data availability section	Influence of missing data
Prunus armeniaca L.	Perennial (tree)	Monoecious, self-incompatible (Groppi et al. 2021)	Southern (56) Northern (199) (see Supplementary Table 1)	Whole genome sequencing	0.07	8 (chromosomes)	~3 M (440K)	~24 M (3.5 M in the subsampled dataset)	Groppi et al. 2021	Influence of number of SNPs, of sample size, of population structure, of using scaffolds

										instead of chromosomes

*in the map file, number of lines divided by number of scaffolds/chromosomes;

**number of lines in the map file

Table 2. Estimates of effective population sizes for each dataset analysed in NeEstimator and GONE. Estimates in bold represent the best possible N_e estimates obtained, considering biases and limitations of each dataset.

Species Gene pool (#samples)	Type of dataset	N_e in NeEstimator				N_e in GONE	
		#polymorphic loci analysed(1)	N_e using P-crit(2)	N_e excluding singletons	N_e with no MAF filtering	#polymorphic loci analysed (3)	N_e in the last generation (geometric mean) (4)
S. globulifera Species 1 (228)	Full	17,515	**371 (CI: 318-441)**	754 (CI: 623-949)	1,036 (CI: 841-1,340)	17,515	N/A
	Thinned	1,710	1,382 (CI: 677-56,918)	1,215 (CI: 674-4,997)	766 (CI: 478-1,756)		
Species 2 (107)	Full	14,906	**246 (CI: 199-320)**	380 (CI: 300-510)	547 (CI: 409-813)	14,906	N/A
	Thinned	1,199	773 (CI: 316-Infinite)	1,042 (CI: 414-Infinite)	564 (CI: 299-3,302)		
Species 3 (30)	Full	9,207	**93 (CI: 39-Infinite)**	86 (CI: 37-Infinite)	223 (CI: 65-Infinite)	9,207	N/A
	Thinned	546	213 (CI: 38-Infinite)	243 (CI: 46-Infinite)	150 (CI: 35-Infinite)		
M. annua Atlantic (12)	Full	17,854	**15 (CI: 7-58)**	15 (CI: 7-58)	22 (CI: 10-121)	17,854	40
	Thinned	250	16 (CI: 6.5-119)	16 (CI: 7-117)	24 (CI: 10-735)		

Core (16)	Full	27,874	**18.6 (CI: 10.2-46.2)**	18.6 (CI: 10.2-46.2)	34.7 (CI: 18.3-131.3)	27,874	123
	Thinned	404	21 (CI: 11-71)	21 (CI: 11-71)	44 (CI: 20-815)		
Mediterranean (12)	Full	18,032	**16 (CI: 10-32)**	16 (CI: 10-32)	26 (CI: 17-51)	18,032	103
	Thinned	233	12 (CI: 8-19)	12 (CI: 8-19)	20 (CI: 13-34)		
***F. sylvatica* (35)**	Full (12 or 27 scaffolds)	N/A	N/A	N/A	N/A	322,185 (12 scaffolds) 1,115,200 (27 scaffolds)	25 (12 scaffolds) 360 (27 scaffolds)
	Thinned (12 or 27 scaffolds)	20,432 or 55,160	No estimates produced	No estimates produced	No estimates produced		
P. armeniaca **Southern (21)**	Full 3.5 M SNPs subset	N/A	N/A	N/A	N/A	82,891	**184**
	Thinned 3.5 M SNPs subset - excluding sites at a distance < 10,000 positions	4,206	44 (CI: 33-64)	43 (CI: 32-62)	68 (CI: 51-99)		
	Thinned 3.5 M SNPs subset - excluding sites at a distance < 5,000 positions	8,378	44 (CI: 34-62)	41.5 (CI: 32-57)	66 (CI: 50-94)		
Northern (75)	Full 3.5 M SNPs subset	N/A	N/A	N/A	N/A	116,285	**3,526**

Thinned 3.5 M SNPs subset - excluding sites at a distance < 10,000 positions	5,838	315 (CI: 198-716)	352 (CI: 221-802)	447 (CI: 269-1,201)		
Thinned 3.5 M SNPs subset - excluding sites at a distance < 5,000 positions	11,468	305 (CI: 197-635)	344 (CI: 219-751)	427 (CI: 261-1,085)		

(1)polymorphic loci = total number of loci in NeEstimator minus number of non-polymorphic loci. Note that in NeEstimator SNPs=loci.

(2)as low-frequency alleles upwardly bias N_e, we followed the recommendations in Waples & Do (2010) and used P_{crit} to screen-out rare alleles = 0.02 when sample sizes > 25, and $1/(2S) \leq P_{crit} \leq 1/S$ when sample sizes ≤ 25 (where S = sample size), and excluded singleton alleles (Waples and Do 2010). For *S. globulifera*, P_{crit}=0.02; for *M. annua*, P_{crit}=0.05; For *P. armeniaca* - Southern gene pool, P_{crit}=0.03; Northern gene pool, P_{crit}=0.02.

(3) GONE only uses a subset of SNPs per chromosome (or scaffold), up to a maximum of 50,000 SNPs per chromosome (or scaffold),these are indicated in the OUTPUT_dataname file.

(4) no MAF filtering was applied in GONE (as recommended).

CIs represent jackknife confidence intervals. Thinned datasets were obtained by subsampling SNPs using windows of 10,000 positions (and also 5,000 positions for *F. sylvatica* and *P. armeniaca*); for *F. sylvatica* and *P. armeniaca*, we only estimated N_e in NeEstimator from thinned datasets, because of the difficulties in handling file conversion of large vcf datasets to the formats required in NeEstimator.

References

Antao T, Pérez-Figueroa A, Luikart G (2011) Early detection of population declines: high power of genetic monitoring using effective population size estimators. Evol Appl 4:144–154

Barbato M, Orozco-terWengel P, Tapio M, Bruford MW (2015) SNeP: a tool to estimate trends in recent effective population size trajectories using genome-wide SNP data. Front Genet 6:109

CBD (2022) Kunming-Montreal Global biodiversity framework. https://www.cbd.int/doc/decisions/cop-15/cop-15-dec-05-en.pdf

Csilléry K, Lalagüe H, Vendramin GG, et al (2014) Detecting short spatial scale local adaptation and epistatic selection in climate-related candidate genes in European beech (*Fagus sylvatica*) populations. Mol Ecol 23:4696–4708

Danecek P, Auton A, Abecasis G, et al (2011) The variant call format and VCFtools. Bioinformatics 27:2156–2158

Danecek P, Bonfield JK, Liddle J, et al (2021) Twelve years of SAMtools and BCFtools. Gigascience 10.: https://doi.org/10.1093/gigascience/giab008

Degen B, Bandou E, Caron H (2004) Limited pollen dispersal and biparental inbreeding in *Symphonia globulifera* in French Guiana. Heredity 93:585–591

De Kort H, Prunier JG, Ducatez S, et al (2021) Life history, climate and biogeography interactively affect worldwide genetic diversity of plant and animal populations. Nat Commun 12:516

Do C, Waples RS, Peel D, et al (2014) NeEstimator v2: re-implementation of software for the estimation of contemporary effective population size (N_e) from genetic data. Mol Ecol Resour 14:209–214

England PR, Luikart G, Waples RS (2010) Early detection of population fragmentation using linkage disequilibrium estimation of effective population size. Conserv Genet 11:2425–2430

Fady B, Bozzano M (2021) Effective population size does not make a practical indicator of genetic diversity in forest trees. Biol Conserv 253:108904

Felsenstein J (2019) Theoretical Evolutionary Genetics. University of Washington

Frankham R (2021) Improvements to proposed genetic indicator for CBD. Biological Conservation

Frankham R, Bradshaw CJA, Brook BW (2014) Genetics in conservation management: Revised recommendations for the 50/500 rules, Red List criteria and population viability analyses. Biol Conserv 170:56–63

Franklin IR (1980) Evolutionary change in small populations

Gargiulo R (2023) Ralpina/Ne-plant-genomic-datasets: Ne-plant-genomic-datasets (v1.0.0). Zenodo. https://doi.org/10.5281/zenodo.8134169

Gargiulo R, Waples RS, Grow AK, et al (2023) Effective population size in a partially clonal plant is not predicted by the number of genetic individuals. Evol Appl 16:750–766

González-Martínez SC, Ridout K, Pannell JR (2017) Range expansion compromises adaptive evolution in an outcrossing plant. Curr Biol 27:2544–2551.e4

Graudal L, Aravanopoulos F, Bennadji Z, et al (2014) Global to local genetic diversity indicators of evolutionary potential in tree species within and outside forests. For Ecol Manage 333:35–51

Groppi A, Liu S, Cornille A, et al (2021) Population genomics of apricots unravels domestication history and adaptive events. Nature Communications 12

Hayes BJ, Visscher PM, McPartlan HC, Goddard ME (2003) Novel multilocus measure of linkage disequilibrium to estimate past effective population size. Genome Res 13:635–643

Hill WG (1981) Estimation of effective population size from data on linkage disequilibrium. Genet Res 38:209–216

Hoban S, Bruford M, D'Urban Jackson J, et al (2020) Genetic diversity targets and indicators in the CBD post-2020 Global Biodiversity Framework must be improved. Biol Conserv 248:108654

Hoban S, Bruford MW, Funk WC, et al (2021a) Global Commitments to Conserving and Monitoring Genetic Diversity Are Now Necessary and Feasible. Bioscience 71:964–976

Hoban S, da Silva JM, Mastretta-Yanes A, et al (2023) Monitoring status and trends in genetic diversity for the Convention on Biological Diversity: An ongoing assessment of genetic indicators in nine countries. Conserv Lett. https://doi.org/10.1111/conl.12953

Hoban SM, Hauffe HC, Pérez-Espona S, et al (2013) Bringing genetic diversity to the forefront of conservation policy and management. Conserv Genet Resour 5:593–598

Hoban S, Paz-Vinas I, Aitken S, et al (2021b) Effective population size remains a suitable, pragmatic indicator of genetic diversity for all species, including forest trees. Biol Conserv 253:108906

Hössjer O, Laikre L, Ryman N (2016) Effective sizes and time to migration-drift equilibrium in geographically subdivided populations. Theor Popul Biol 112:139–156

Jamieson IG, Allendorf FW (2012) How does the 50/500 rule apply to MVPs? Trends Ecol Evol 27:578–584

Jones AT, Ovenden JR, Wang Y-G (2016) Improved confidence intervals for the linkage disequilibrium method for estimating effective population size. Heredity 117:217–223

Kershaw F, Bruford MW, Funk WC, et al (2022) The Coalition for Conservation Genetics: Working across organizations to build capacity and achieve change in policy and practice. Conservat Sci and Prac 4. https://doi.org/10.1111/csp2.12635

King L, Wakeley J, Carmi S (2018) A non-zero variance of Tajima's estimator for two sequences even for infinitely many unlinked loci. Theor Popul Biol 122:22–29

Laikre L, Hohenlohe PA, Allendorf FW, et al (2021) Authors' Reply to Letter to the Editor: Continued improvement to genetic diversity indicator for CBD. Conserv Genet 22:533–536

Li H, Durbin R (2009) Fast and accurate short read alignment with Burrows-Wheeler transform. Bioinformatics 25:1754–1760

Luikart G, Antao T, Hand BK, et al (2021) Detecting population declines via monitoring the effective number of breeders (N_b). Mol Ecol Resour 21:379–393

Luikart G, Ryman N, Tallmon DA, et al (2010) Estimation of census and effective population sizes: the increasing usefulness of DNA-based approaches. Conserv Genet 11:355–373

Marandel F, Charrier G, Lamy J-B, et al (2020) Estimating effective population size using RADseq: Effects of SNP selection and sample size. Ecol Evol 10:1929–1937

Marandel F, Lorance P, Berthelé O, et al (2019) Estimating effective population size of large marine populations, is it feasible? Fish Fish 20:189–198

Merzeau D, Comps B, Thiébaut B, Letouzey J (1994) Estimation of *Fagus sylvatica* L. mating system parameters in natural populations. Annales des Sciences Forestières 51:163–173

Montes I, Iriondo M, Manzano C, et al (2016) No loss of genetic diversity in the exploited and recently collapsed population of Bay of Biscay anchovy (*Engraulis encrasicolus*, L.). Mar Biol 163:98

Moran BM, Hench K, Waples RS, et al (2019) The evolution of microendemism in a reef fish (*Hypoplectrus maya*). Mol Ecol 28:2872–2885

Nadachowska-Brzyska K, Konczal M, Babik W (2022) Navigating the temporal continuum of effective population size. Methods Ecol Evol 13:22–41

Neel MC, McKelvey K, Ryman N, et al (2013) Estimation of effective population size in continuously distributed populations: there goes the neighborhood. Heredity 111:189–199

Nunney L (2016) The effect of neighborhood size on effective population size in theory and in practice. Heredity 117:224–232

Nunney L (1991) The influence of age structure and fecundity on effective population size. Proc Biol Sci 246:71–76

Nunney L (1993) The influence of mating system and overlapping generations on effective population size. Evolution 47:1329–1341

Nunziata SO, Weisrock DW (2018) Estimation of contemporary effective population size and population declines using RAD sequence data. Heredity 120:196–207

Obbard DJ, Harris SA, Buggs RJA, Pannell JR (2006a) Hybridization, polyploidy, and the evolution of sexual systems in *Mercurialis* (Euphorbiaceae). Evolution 60:1801–1815

Obbard DJ, Harris SA, Pannell JR (2006b) Sexual systems and population genetic structure in an annual plant: testing the metapopulation model. Am Nat 167:354–366

O'Brien D, Laikre L, Hoban S, et al (2022) Bringing together approaches to reporting on within species genetic diversity. J Appl Ecol 59:2227–2233

Oddou-Muratorio S, Gauzere J, Angeli N, et al (2021) Phenotypic and genotypic data of a European beech (*Fagus sylvatica* L.) progeny trial issued from three plots along an elevation gradient in Mont Ventoux, South-Eastern France. Ann For Sci 78. https://doi.org/10.1007/s13595-021-01105-9

Palstra FP, Fraser DJ (2012) Effective/census population size ratio estimation: a compendium and appraisal. Ecol Evol 2:2357–2365

Palstra FP, Ruzzante DE (2008) Genetic estimates of contemporary effective population size: what can they tell us about the importance of genetic stochasticity for wild population persistence? Mol Ecol 17:3428–3447

Peel D, Waples RS, Macbeth GM, et al (2013) Accounting for missing data in the estimation of contemporary genetic effective population size (N_e). Mol Ecol Resour 13:243–253

Petit RJ, Hampe A (2006) Some evolutionary consequences of being a tree. Annu Rev Ecol Evol Syst 37:187–214

Purcell S, Neale B, Todd-Brown K, et al (2007) PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet 81:559–575

Qanbari S (2019) On the extent of linkage disequilibrium in the genome of farm animals. Front Genet 10:1304

Qanbari S, Pimentel ECG, Tetens J, et al (2010) The pattern of linkage disequilibrium in German Holstein cattle. Anim Genet 41:346–356

Raj A, Stephens M, Pritchard JK (2014) fastSTRUCTURE: variational inference of population structure in large SNP data sets. Genetics 197:573–589

R Core Team (2019) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. Version 3.6.1. Citeseer. URL https://www.R-project.org/.

Robinson JD, Moyer GR (2013) Linkage disequilibrium and effective population size when generations overlap. Evol Appl 6:290–302

Ryman N, Laikre L, Hössjer O (2019) Do estimates of contemporary effective population size tell us what we want to know? Mol Ecol 28:1904–1918

Santiago E, Novo I, Pardiñas AF, et al (2020) Recent demographic history inferred by high-resolution analysis of linkage disequilibrium. Mol Biol Evol 37:3642–3653

Santos-del-Blanco L, Olsson S, Budde KB, et al (2022) On the feasibility of estimating contemporary effective population size (N_e) for genetic conservation and monitoring of forest trees. Biol Conserv 273:109704

Schmitt S, Tysklind N, Hérault B, Heuertz M (2021) Topography drives microgeographic adaptations of closely related species in two tropical tree species complexes. Mol Ecol 30:5080–5093

Sved JA (1971) Linkage disequilibrium and homozygosity of chromosome segments in finite populations. Theor Popul Biol 2:125–141

Sved JA, Cameron EC, Gilchrist AS (2013) Estimating effective population size from linkage disequilibrium between unlinked loci: theory and application to fruit fly outbreak populations. PLoS One 8:e69078

Sved JA, Feldman MW (1973) Correlation and probability methods for one and two loci. Theor Popul Biol 4:129–132

Tallmon DA, Gregovich D, Waples RS, et al (2010) When are genetic methods useful for estimating contemporary abundance and detecting population trends? Mol Ecol Resour 10:684–692

Thurfjell H, Laikre L, Ekblom R, et al (2022) Practical application of indicators for genetic diversity in CBD post-2020 global biodiversity framework implementation. Ecol Indic 142:109167

Torroba-Balmori P, Budde KB, Heer K, et al (2017) Altitudinal gradients, biogeographic history and microhabitat adaptation affect fine-scale spatial genetic structure in African and Neotropical

populations of an ancient tropical tree species. PLoS One 12:e0182515

Van der Auwera GA, O'Connor BD (2020) Genomics in the Cloud: Using Docker, GATK, and WDL in Terra. "O'Reilly Media, Inc."

Wang J (2016) A comparison of single-sample estimators of effective population sizes from genetic marker data. Mol Ecol 25:4692–4711

Wang J, Santiago E, Caballero A (2016) Prediction and estimation of effective population size. Heredity 117:193–206

Waples RK, Larson WA, Waples RS (2016) Estimating contemporary effective population size in non-model species using linkage disequilibrium across thousands of loci. Heredity 117:233–240

Waples RS (2022) What is N_e, anyway? J Hered 113:371–379

Waples RS (2016) Making sense of genetic estimates of effective population size. Mol. Ecol. 25:4689–4691

Waples RS (2006) A bias correction for estimates of effective population size based on linkage disequilibrium at unlinked gene loci. Conserv Genet 7:167–184

Waples RS, Antao T, Luikart G (2014) Effects of overlapping generations on linkage disequilibrium estimates of effective population size. Genetics 197:769–780

Waples RS, Do C (2008) ldne: a program for estimating effective population size from data on linkage disequilibrium. Mol Ecol Resour 8:753–756

Waples RS, Do C (2010) Linkage disequilibrium estimates of contemporary N_e using highly variable genetic markers: a largely untapped resource for applied conservation and evolution. Evol Appl 3:244–262

Waples RS, England PR (2011) Estimating contemporary effective population size on the basis of linkage disequilibrium in the face of migration. Genetics 189:633–644

Waples RS, Waples RK, Ward EJ (2022) Pseudoreplication in genomic-scale data sets. Mol Ecol Resour 22:503–518

Wright S (1931) Evolution in Mendelian Populations. Genetics 16:97–159

Wright S (1969) Evolution and the genetics of Populations Volume 2: The Theory of Gene Frequencies. The University of Chicago Press, Chicago

Yonezawa K (1997) Effective population size of plant species propagating with a mixed sexual and asexual reproduction system. Genet Res 70:251–258