An adaptable, user-friendly score sheet to monitor welfare in experimental fish

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Abstract

Fish are increasingly used as experimental animals across research fields. Currently, a quarter of

all experimental animals used in Europe are fish. Less than 20% of these are standard model

species. Welfare assessments for experimental fish are in their infancy compared to rodents.

This can be attributed to the diversity of species used, the relative recency of fish as go-to model

for research, and challenges to assess welfare and pain in non-vocal underwater species. The

lack of guidelines and tools presents a challenge for researchers (particularly, for newcomers),

for ethics committees, and for implementing refinement measures.

Here, we present an adaptable, user-friendly score sheet for fish. The parameters contained in

the excel tool are based on a literature review, have been validated by expert interviews, and

evaluated by a fish pathologist. The tool allows to score individuals as well as groups, calculates

summary scores and visualizes trends. We provide the underlying literature, give use examples

and provide instructions on the adaptation and use of the score sheet.

We hope that this tool will empower researchers to include welfare assessment in their routines,

foster discussions on fish welfare parameters among scientists, facilitate interactions with ethics

committees, and most importantly, enable the refinement of fish experiments.

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Author contributions

IAK conceived the project, HS and IAK designed the analysis with input from MFG, MFG

collected the data and performed the analyses with support from AB and HS, HS and IAK

supervised and guided analyses, MFG wrote the manuscript draft, HS, IAK, AB and MFG

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Declaration of conflicting interests

The authors declare that there is no conflict of interest.

Research ethics

No animals were used for this research.

Data availability

All data used are available in the supplementary material.

Supplemental File 1: Literature search parameters

Supplemental File 2: List of retained and analysed literature, initial parameter list and

refinements, expert questions

Supplemental File 3: Collection of fish welfare parameters

Supplemental File 4: Score sheet

Introduction

A wide range of research areas use fish as experimental models. Topics include toxicological

tests [1], pharmacology [2], developmental biology [3,4], complex human diseases [5] and behaviour [6].

Fish are also often used to probe the response of aquatic organisms to the impact of global

change, ocean warming and ocean acidification [7]. Teleost fish share key genetic, anatomical and

physiological properties with humans [5,8] and therefore are valuable organisms to investigate

basic evolutionary concepts [9–12], stem cells and regeneration [13], the impact of chemicals and

pollutants on organismic health [14](12), or pharmacology [15].

In 2020, a quarter (27.4%) of all animals used for research in the EU were fish. Zebrafish

(*Danio rerio*) as the most widely known experimental fish model constituted 13% of all

experimental fish [15], but numerous other fish species beyond zebrafish are used in research [5,16],

such as medaka (*Oryzias latipes*) [17], fat head minnow (*Pimephales promelas*) [18], trout

(*Salmonidae*) [19], cichlids (*Cichlidae*) [20], sticklebacks (*Gasterosteidae*) [21], guppies (*Poecilia

reticulata*) [22], mollies (*Poecilia latipinna*) [23], or gobies (*Gobidae*) [24], to just name a few.

Evolutionarily, some of these species are more distantly related than humans to armadillos [25].

In accordance with widespread legal requirements to implement the 3Rs (Replace, Reduce,

Refine [26]) in animal experiments, the awareness and demand to objectively assess welfare in

animal experiments continues to increase, also in research using fish. The 3Rs postulate to

replace, if possible, animal models with non-living models, to reduce the number of animals

necessary to fulfil an experiment, and to refine the experiments to avoid or reduce negative

impacts, and they serve as foundation for laws and recommendations on animal welfare in

research[27]. Measures supporting the third R, refinement, are very much dependent on the

monitoring and documentation of animal welfare during husbandry and experimentation, which

is legally mandatory in many countries. The tool commonly used to measure welfare are score

sheets.

The aim of score sheets is to document the state of welfare and to detect alterations in welfare

between groups, individuals, or time-points in a semi-quantitative, semi-objective manner.

However, one can only measure what one can define and quantify, and animal welfare remains

a tricky concept particularly for fish [28]. EU regulations, for example, contain no notion of

welfare [29]. National definitions of welfare vary (ranging from the five freedoms [30] to dignity- or

well-being-focused approaches [31]). While fish are treated as sentient by law [29], and despite key

initiatives [28,32–37]), some sources still express a need for debate on the matter [27,32,38]. Knowledge

on fishes´ emotions and behaviour changes is available on some species [8,28,39,40], but their

subjective pain perception is sometimes discussed controversially [32,41]. Consequently, while the

literature does discuss potential signs of distress in fish, evaluation tools for validated clinical

and behavioural indicators of wellbeing are hard to come by for fish other than zebrafish [42–44].

In addition to these conceptual challenges, fish score sheets need to account for the previously

mentioned diversity of species with distinct behaviours, morphology, physiology, and

environmental requirements [8], and the absence of characteristics well established for rodents,

such as vocalisations, body language, and spontaneous as well as induced behaviours such as

grooming [35,45,46], or for agricultural species [46]. While decipherable behaviours have been

identified in some fishes, many remain understudied or uninterpretable [40]. Fish are often housed

in considerably large groups, which on the one hand complicates individual welfare monitoring

and can result in specific welfare problems (e.g., hierarchy or territory issues (28)), but on the

other hand expands the set of observable and measurable behavioural indicators beyond

individual characteristics. If sourced from the wild, they come with a range of common

background conditions [47].

Finally, in addition to these system-inherent challenges, training opportunities taking fish

welfare into account are presently severely limited. Good welfare practices depend on skilled

staff [8,40], but courses for fish researchers, authorities and ethics committees are usually part of a

very broad education on "non-rodent species". Fish research publications typically do not

include score sheets [8,28], which limits open conversation about useful and implementable

quantitative or qualitative parameters for welfare evaluations. A potential ally in experimental

fish welfare is the field of aquaculture, where fish welfare translates to disease incidence,

growth rates and meat quality and thus to business revenue (e.g., MyFishCheck [48–51], but limited

mutual interactions impede the transfer of welfare related tools. Together, these challenges

result in a lack of resources, expertise and frameworks with regard to fish welfare scoring in

experimental research, which poses a serious challenge for researchers in the implementation of

the third R for fish, particularly when researchers are new to the organism.

This publication aims to provide fish researchers with a resource facilitating the task of

refinement. We provide a versatile, multispecies, modular, literature-based, and user-friendly

score sheet and thereby empower researchers to assess the welfare of their animals, implement

refinement strategies, and fulfil legal welfare monitoring requirements in an evidence-based

manner. To develop this tool, we first systematically listed and evaluated fish welfare

parameters. This entailed a scoping literature review and an exhaustive compilation of clinical

and behavioural parameters that could be used as indicators of the welfare state of fish. We also

conducted qualitative guided conversations with researchers and veterinarians working on

different fish species and experimental systems, and reduced the list of potential indicators to a

handful of individual and group parameters. Possibility of systematisation, non-intrusiveness,

and modularity of parameters (e.g. module "behaviour") were choice criteria. For each

parameter, we then defined possible, easy-to-identify deviations from the desirable state, and

defined mild versus severe deviations from the ideal state. We provide these parameters in the

context of a modular MS office-based scoring table that a) accommodates individuals as well as

groups of fish and b) can be adapted easily for the species, the experimental context or the

housing situation.

We are convinced that this score sheet will simplify welfare monitoring for fish researchers

worldwide, promote the use of pre-studies to optimize animal numbers and experimental

conditions, facilitate 3R implementation as well as licence application procedures, and serve as

useful starting point for discussions within and between labs regarding fish welfare.

Methods

Scoping review

To gather the published state of the art regarding fish welfare assessments, we conducted a

scoping review [52] on welfare indicators for juvenile and adult fish. The main research question

was "Which welfare indicators and score sheets are currently used for fish?". PubMed and Web

of Science were queried with 40 combinations of the terms "fish", "rating", "welfare",

"indicator", "score board", "score card", "monitoring", "assessment", "humane endpoint",

"perch", "trout", and "zebrafish" (Supplemental File 1). When a search combination received

too many hits to screen and select, the search term was refined. The relevance of papers was

then assessed by screening title and the abstract. Papers were retained if these suggested the

paper could contain information about fish welfare. Retained papers were then screened for

welfare indicators and retained if they mentioned such parameters. Reference sections of

retained publications were additionally screened for publications that did not come up in the

original search and were subjected to the same screening process. 84 documents were directly

retrieved from the databases in round 1. An additional 15 relevant documents were identified

from snowballing the references of the round 1 documents. From the 99 documents containing

welfare parameters that were retained (Supplemental File 2), a list of welfare parameters was

compiled. In total, the literature search yielded 68 potential parameters (Supplemental File 2).

Expert interviews

To understand practical considerations and the diversity of needs and implementation situations,

ten fish facilities in Switzerland housing zebrafish, perch, trout, stickleback, and cichlids for

research on RNA, epigenetics, genetics and genomics, development, health and infection

biology, and behaviour were visited. The persons in charge of running the facility were

qualitatively interviewed in semi-structured interviews with an interview guide. Questions

included fish species, husbandry parameters, types of experiments done, current approaches to

measure welfare (tool, criteria, frequency, group or individual level, time, responsible person,

treatment of transgenic lines, reasons to not use a score sheet if none was used, source of the

tool), wishes and thoughts towards score sheets (what would make it easy to use, digital or

analog), and considerations concerning the collected parameters (relevance of parameters,

additional suggestions, species-specifics, how to group parameters, potential endpoints,

relevance of a total integrating score). Answers were recorded with pen and paper and later

transcribed into an excel sheet. Also, 4 additional conversations were conducted with experts on

animal welfare to understand what they would expect from or look for in a scoresheet.

Questions are included in supplemental file 2. The responses, except for a summary and

individual quotes, are not shared to maintain participant confidentiality.

Parameters

The 68 parameters identified in the literature were narrowed down to a meaningful, feasible set

of 29 parameters in a first round based by co-author HS, a diagnostic and research veterinarian

and fish pathologist with 20+ years of experience in fish health and fish welfare, and further

refined to 21 parameters for individuals and 10 parameters for groups based on the expert

interviews and on discussions within the team (who feature experience working with zebrafish,

trout, perch, and gobies, and rodents). Specifically, the following steps were taken: a) non-

invasively observable parameters were selected (for example, blood cortisol levels as stress

indicators require invasive measurements), b) related parameters were grouped (for example,

swimming equilibrium [53] and buoyancy [54] were summarised to navigation problems), c) a

verbose explanation for possible states was developed (for example, "normal for the species",

"mild weight loss/gain", and "severe weight loss/gain" for the parameter "nutrition"), d)

considerations that could affect scoring were collected (e.g., recent feeding may cause non-

problematic abdominal swelling), and e) observable states were linked to numeric scores with 0

= ideal state, 1 = not ideal, 2 = observe, act if possible, and 3 = act immediately, potentially

terminate (Table 1, see Supplemental File 3 for additional literature and species information).

Results

Expert Interviews

Expert interviews confirmed the idea that fish welfare assessments and their use in refinement

are an area of development with unsatisfactory options for fish researchers. A frequent practice

was to copy other researcher's score sheets and to satisfy administrative requests ("Once the

canton of … requested more parameters, so we just added one of them"). Opinions on welfare

were often based on years of experience working with fish, rather than on explicitly scorable,

model specific, and objective criteria. Only 3 of the 10 facilities indicate using a score sheet as

welfare tool. Two facilities stated not using a score sheet because it was not compulsory in their

region. Usually, more than 1 person was in charge of assessing fish health and state at the same

time. Some facilities stated that certain types of physical/morphological aberrations or

behavioral signs were considered as induced by standard lab conditions and were therefore

currently not considered in their score sheets as welfare issues.

With regard to a score sheet, isolated parameters were generally deemed insufficient to conclude

on the welfare state of individuals and groups. Also, behavioral signs of impairment were

considered more relevant than physical signs, which may be linked to the current lack of

understanding regarding the relevance of e.g., injury and related pain for fish wellbeing. The

relevance of an integrative total score was generally supported as reasonable given the mutual

impacts of parameters (e.g., damaged fins might lead to altered swimming behavior). A request

was to include potential measures to alleviate the identified issues in a score sheet, since

euthanasia of impacted fish would often create issues for experimental design. A digital version

was preferred, and a tablet-compatible app was suggested by several participants.

Score sheet

The score sheet developed based on the review and the interviews is available as Supplemental

File 4. It contains 21 potentially relevant parameters for individuals, and 10 parameters for

groups. These 21 parameters are based on literature from 17 distinct fish species from 11

taxonomic families. They are classified into the categories body condition, skin, fins, eyes,

gills/opercula, infection, swimming behavior, feeding behavior for individuals, and 2 additional

categories (group behavior and mortality) for groups.

For individually housed or marked fish, for example in cichlid behaviour experiments [6], the

status for a certain parameter is assessed verbosely by choosing from a drop-down menu (Figure

1). For example, for fin integrity, researchers can choose between "fin in healthy condition",

"slightly eroded / slightly missing", "moderate fin loss" or "severe fin loss / missing fin". The

sheet then automatically calculates a numerical score. For fish housed in groups, which is for

example usually the case for zebrafish, the estimated percentage of fish in the tank with certain

issues is recorded (Figure 1). For example, 80% of fish may be scored, for a particular

parameter, to display "no abnormalities / lesions", 10% with "slight abnormalities / lesions", 5%

with "moderate abnormalities / lesions" and 5% with "severe abnormalities / lesions". A

weighted average is then calculated for the tank based on formulas already implemented in the

score sheet. If percentages do not sum up to 100 due to entry errors, the respective cells in the

excel sheet turn orange.

In the score sheet for individually housed or marked fish, skipping the assessment of a

parameter in the individual score sheet results in a default score of 1 to avoid the false

assumption of good welfare based on non-scoring. If parameters cannot be scored because it is

either not feasible or not relevant regarding the circumstances, we recommend adapting the

score sheet and deleting the parameter (s) (Supplemental File 4 features instructions on how to

do that).

The score sheet tool provides a total score, a median, mean and maximum score for each

assessment. Both these integrative and the individual scores are color coded from green to red

on a relative scale for easy visual overview. 7 assessments (e.g., for one week) are designed to

fit on an A4 printout for easy collection and recording of past assessments. In addition, a trend

tab allows the tracking and visualization of integrative and individual scores across time. To

visualize trends, users need to copy-paste values (without formulas) from areas outlined with

heavy borders from the score sheet tab to the trends tab. Up to 31 data points (e.g., for one

month) can be accommodated within the current dimensions of the trend tab.

Use scenarios

Below, we use three hypothetical common research scenarios, to demonstrate how the score

sheet can be used to monitor welfare or refine experimental conditions. These scenarios are 1) a

pre-trial to identify useful welfare scoring parameters for a genetically modified zebrafish strain

with morphological impairments, 2) a pretrial to define a suitable acclimatization period of

wild-caught individuals before an experiment, and 3) the monitoring of fish welfare during an

infection trial to identify the appropriate termination point.

In scenario 1 (Figure 2), a strain of genetically modified zebrafish with developmental

morphological defects in spine and eyes, and slightly elevated mortality, is to be included in a

chemical exposure experiment (similar to e.g. [55]. The researchers are confronted with the task to

identify welfare parameters that are suited, in this impaired strain, to detect any additional

impairment caused by the experiment. To start with, they score a control tank over a month

using the tool for fish that are housed in groups. Scores relating to anatomy (morphology and

eye scores) are constitutively high due to the developmental impairment. Swimming behavior is

also impacted by the strains' defects, resulting in overall high behavioral scores. Skin

parameters and feeding behavior fluctuate slightly during the observation time. Mortality scores

indicate that strain mortality is 10% higher than usually observed in the wildtype. Based on

these documented and quantified observations, the researchers come to the following

conclusions for their main exposure experiment: they will pay particular attention to skin and

feeding parameters, since these parameters display variation already without treatment, and may

therefore be sensitive to stressors. Also, they take the 10% mortality rate into account during

sample size and power calculations and start the experiment with 10% more animals than

required to achieve the desired power. They design an experiment-specific score sheet where

they record anatomy and swimming behavior but – given that these parameters are high to begin

with, reducing their sensitivity – flag them as potentially unhelpful. They also use the data to

justify animal numbers in their animal experimentation permit application.

In scenario 2 (Figure 3), individuals from a wild fish species are caught and brought to the lab

for a behavior experiment (choice between two compartments, e.g. [56]) before being released

again. The researchers are confronted with the task to determine a suitable acclimation period

before the experiment: not too long to avoid unnecessary long captivity, not too short to avoid

confounding impacts of the stress during capture on the experiment. They catch several fish for

a pre-trial and assess their welfare individually for a month. Figure 3 shows data from one

representative individual. During the entire observation period, no signs of external damage,

lesion or infection are observed. However, a diverse range behavioral abnormalities (e.g.,

stereotypical swimming and startle responses to movement and sounds) are observed between

d1 and d13. These reactions decline in intensity and disappear in week 3. From d19 onwards, no

behavioral abnormalities are observed. Based on the pretrial, the researchers therefore conclude

(and have data to show) that an 18 day acclimation period is both required and sufficient for the

planned behavioral experiments, and choose their setup accordingly to ensure the reliability and

validity of their data.

In scenario 3 (Figure 4), researchers want to perform a drug screen on zebrafish infected with a

certain pathogen (similar to e.g. [57]) to find compounds that are active against said pathogen. The

researchers are confronted with the task of determining an experimental duration that is long

enough to show differences between control and treatment, and short enough to avoid

unnecessary suffering. They also need to determine a termination score (i.e. a cutoff where a

sick animal is euthanized and removed from the experiment to avoid excessive suffering). A

group of 10 fish is therefore used for a 4-week pretrial under control conditions (pathogen

infection without drug) and scored with the group tool. The score sheet reveals that first skin

defects are observed 14 days after pathogen exposure, and continuously increase in severity. On

day 23 after pathogen exposure, swimming issues (loss of buoyancy) are starting to be observed

and increase in prevalence until the end of the pretrial. Based on this pretrial, the researchers

determine to run the drug screen for 25 days, since meaningful differences between control and

treatment should be observable by this timepoint without causing unnecessary suffering of

experimental animals. They also decide that they will terminate any tank x compound

combination where the total score exceeds 11 two days in a row, since this score should not be

reached in the control within 25 days and would indicate that the compound causes harm.

Use and adaptation of the score sheet

Scoring 21 parameters on multiple fish every day is clearly not a feasible task. We want to

emphasize that a key step in the use of this (and any other) score sheet is the adaptation phase.

This phase includes a) test scoring of the parameters, followed by b) a choice of parameters that

are truly useful to assess welfare in the given species, situation, and experiment, and c) creating

an experiment-specific score sheet by deleting non-informative rows from the score sheet,

adding parameters, or adapting the weighting.

We strongly encourage a rigorous and information-driven reduction of the score sheet to

achieve maximum information and maximum compliance with minimal effort. The score sheet

file contains instructions to help with the adaptation and implementation phase (Supplemental

File 4, tab "instructions"). The adaptation phase encompasses 7 steps: "Adapt to setting", "Get

familiar", "Generate baseline", "Adapt criteria and weights", "Termination criteria". We hope to

discourage uncritical use of the score sheet with these instructions, since this would most likely

lead to frustration and non-compliance. In addition, the score sheet file contains instructions for

use after adaptation. The use phase is based on three steps: "Data entry", "Documentation",

"Statistics". The aim is to encourage sustainable long-term preservation strategies and support

good data management practices, e.g. for reporting to ethics committees.

Discussion & Conclusion

This publication aims to provide a flexible framework to develop suited score sheets across a

number of species, experimental scenarios, and housing conditions. This is by no means the first

attempt to develop score sheets for experimental fish (see, for example, a zebrafish-specific pain

and distress score sheet in [58] Table 3). However, to our knowledge, it is the first attempt to

create an adaptable tool that is explicitly not supposed to be used as is, but rather to be molded

by the researcher to really fit their needs regarding welfare monitoring.

Applicability

The diversity of fish species kept for research purposes, and their housing both as single fish

and large groups is a challenge for welfare assessment. The presented work tackles this in two

ways. First, the developed score sheet remains adaptable. Parameters can be removed (which is

actually recommended), changed, or added (e.g., for highly specific behaviours, such as mouth

brooding or nest building). Second, the score sheet emphasizes the importance of relative

assessments. It does not provide absolute numerical cutoffs a priori but empowers users to

develop their own meaningful termination criteria based on the respective species, strain,

facility, and housing conditions.

The operationalization of termination criteria (endpoints where an experiment must be

terminated and the animals are euthanized to prevent further suffering) is one of the most

important aspects and purposes of score sheets. How to arrive at a combined measure of

maximally tolerable burden is one of the most important, but also most difficult and

controversial parts of welfare assessments. Importantly, termination criteria depend heavily on

the weighing of interests and the purpose of the experiment (which extent of suffering is

justifiable for the expected gain of knowledge). This score sheet should facilitate the definition

of termination criteria and facilitate conversations on termination criteria among fish

researchers.

Implications

This score sheet should benefitfish and fish researchers in five distinct ways. Firstly, the tool

aims to empower the individual researcher. Our interviews revealed a prevalence of subjective

assessments which may or may not reflect animal welfare criteria, a certain acceptance of

stereotypies as given, and a dependence on few knowledgeable care takers. All these aspects

could benefit from more objective scoring options. Secondly, the tool potentially fosters within-

group interactions and cross-facility communication regarding welfare and termination criteria.

This may improve collective welfare knowledge in the fish research community and inspire

initiatives and research towards better welfare assessment in fish. For example, our interviews

revealed that standard laboratory conditions are generally assumed to guarantee good welfare.

One way to assess this would be to conduct a project using the tool across facilities and housing

conditions. Thirdly, the tool may facilitate regulatory processes surrounding animal

experimentation. In Switzerland, score sheets are required to obtain animal experimentation

permits. This tool may thus support researchers in defining and submitting evidence-based

monitoring and stop-criteria. Fourthly, ethics committees are usually not familiar with welfare

of fish. This tool should facilitate the work of ethics committees to support well-planned and

well-monitored experimentation with fish, and the communication between researchers and

committees. A fifth aspect relates to teaching. Score sheets are valuable tools to discuss fish

needs, health, and welfare, and can be used to train junior researchers in accurate and attentive

evaluation of signs and behaviours. For example, a course could entail the application of a score

sheet to strains with known physical and behavioral impairments, or to a range of species.

Limitations

From the practical perspective, welfare monitoring can pose a challenge regarding the time

requirements and personnel resources of research groups . We strongly recommend prioritizing

the monitoring of few meaningful parameters at longer time intervals over more complex

monitoring plans which fail in the long-term. A key factor in welfare monitoring is compliance

and constancy, and these need to be given priority over perfection due to overburdening the

researchers or animal care takers. From the conceptual perspective, discussions on animal

welfare are often impacted by debates on fish nociception. We support the point of view that the

presence of protective responses and avoidance learning may be sufficient indicators of pain,

health, and welfare in fish, but agree that scoring of pain remains a challenge.

Recommendations

In summary, this manuscript provides an objective, adaptable welfare score sheet tool to the

wider fish research communityallowing monitoring of fish welfare in a semi-objective manner

Prospectively, it would be valuable to engage in cross-species and cross-field efforts to build on

the presented monitoring tool. Also the development of an adaptable welfare-monitoring smart-

phone app would be extremely practical for the daily monitoring. Also, visual tracking

applications [59] and sensors have great potential to facilitate fish welfare monitoring in the future.

Figures

Table 1. Parameters. Welfare parameters, explanations of these parameters, and signs to look

for. Includes scores in numbers and in verbose expressions. This table is translated into an

interactive scoring tool in Supplemental File 4.

Figure 1. Score sheets. A – Score sheet for individual animals. **B** – Score sheet for groups of

animals. These are screenshots taken from the interactive scoring tool provided as Supplemental

File 4.

Figure 2. Application Scenario 1. Example for a group scoring of an impaired mutant line of

zebrafish. View of the trends tab. The trends tab translates the scores into visual patterns, on the

one hand through score-dependent coloring of the cells, on the other hand through trend lines.

Each value in the top table corresponds to a datapoint in the trend lines (some pointed out by

circles and arrows). The summary scores, external signs, and group/behavioural signs are each

collated in a separate trend line figure.

Figure 3. Application Scenario 2. Example for an individual scoring of a wild-caught fish

during the acclimatization period. View of the trends tab. Both the score-dependent coloring on

the top and the trend lines at the bottom suggest that the fish is stressed until day 13 of the

acclimatization. No behavioural signs of stress are observed after day 19.

Figure 4. Application Scenario 3. Example for an individual scoring of an experimental fish

exposed to a compound. External signs (i.e. skin defects) are observed before the appearance of

behavioural signs and get progressively worse over the course of several weeks. Based on these

data, the end point of the planned experiment is determined after onset of signs, but before

welfare is too compromised.

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A - Score sheet for individual animals

Animal Exp. Permit														
ID of Tank / Individual														

Termination criteria:		Date	20230701	yyyymmdd		yyyymmdd		yyyymmdd		yyyymmdd		yyyymmdd		yyyymmdd
		Sored by	Jenni D.											
		Total score	24.0		21.0		21.0		21.0		21.0		21.0	21.0
		Total median	1.0		1.0		1.0		1.0		1.0		1.0	1.0
		Total mean	1.1		1.0		1.0		1.0		1.0		1.0	1.0
		Total max	3.0		1.0		1.0		1.0		1.0		1.0	1.0

External signs			18		13		13		13		13		13	13
Body condition	nutrition state	severe weight loss	3	na	1	na	1	na	1	na	1	na	1	na	1
	swelling, proliferative lesion	normal rounded abdomen	0	na	1	na	1	na	1	na	1	na	1	na	1
	spine	slight scoliosis	1	na	1	na	1	na	1	na	1	na	1	na	1
	egg-bound (female)	g-bound, egg-bound for longer per	3	na	1	na	1	na	1	na	1	na	1	na	1
	other deformities	no other deformation	0	na	1	na	1	na	1	na	1	na	1	na	1
Skin	scales	protruding scales	3	na	1	na	1	na	1	na	1	na	1	na	1
	skin lesions	small superficial lesions	1	na	1	na	1	na	1	na	1	na	1	na	1
	skin color	color normal for the strain	0	na	1	na	1	na	1	na	1	na	1	na	1
Fins	fin state	moderate tissue necrosis	2	na	1	na	1	na	1	na	1	na	1	na	1
Eyes	eye state	two sided damage	3	na	1	na	1	na	1	na	1	na	1	na	1
Gills/Opercula	gill state	normal operculum and gills	0	na	1	na	1	na	1	na	1	na	1	na	1
	ventilation	normal beat rate for the species	0	na	1	na	1	na	1	na	1	na	1	na	1
Infection	skin health / infection signs	few small infected areas	2	na	1	na	1	na	1	na	1	na	1	na	1

Comments

Behavioural signs			6		8		8		8		8		8	8
Swimming	activity level	normal activity level for species	0	na	1	na	1	na	1	na	1	na	1	na	1
	breathing	normal breathing	0	na	1	na	1	na	1	na	1	na	1	na	1
	fin posture	fins are clamped	2	na	1	na	1	na	1	na	1	na	1	na	1
	navigation	porary loss of balance and orienta	2	na	1	na	1	na	1	na	1	na	1	na	1
	position in tank	ccupies tank as typical for specie	0	na	1	na	1	na	1	na	1	na	1	na	1
	scratching	no scratching, no jumping	0	na	1	na	1	na	1	na	1	na	1	na	1
	stereotypies	observation of stereotypies	2	na	1	na	1	na	1	na	1	na	1	na	1
Feeding	feeding	reacts normally to food	0	na	1	na	1	na	1	na	1	na	1	na	1

Comments

B - Score sheet for groups

	20230701	20230702	20230703	20230704	20230705	20230706	20230707	20230708	20230709	20230710	20230711	20230712	20230713	20230714	20230715	20230716	20230717	20230718	20230719	20230720	20230721	20230722	20230723	20230724	20230725	20230726	20230727	20230728	20230729	20230730	20230731
Total	13.00	10.00	10.00	10.00	8.00	10.00	8.00	2.00	6.00	4.00	2.00	4.00	4.00	0.00	0.00	0.00	2.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
median	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
mean	0.62	0.38	0.48	0.48	0.38	0.48	0.38	0.10	0.29	0.19	0.10	0.19	0.19	0.00	0.00	0.00	0.10	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
max	3.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	2.00	0.00	0.00	0.00	2.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
External signs	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
nutrition state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
spine	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
swelling, proliferative lesion	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
egg-bound (female)	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
other deformities	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
skin lesions	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
skin color	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
scales	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
fin state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
eye state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
gill state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
ventilation	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
skin health / infection signs	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Behavioural signs	13	10	10	10	8	10	8	2	6	4	2	4	4	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0
navigation	2	2	2	2	2	2	2	0	2	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
activity level	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
stereotypies	2	2	2	2	2	2	2	2	0	2	0	2	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
fin posture	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
position in tank	2	2	2	2	2	2	2	0	2	2	2	2	0	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0
breathing	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
scratching	2	2	2	2	2	2	2	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
feeding	2	2	2	2	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

d1-d13: diverse, strong behavioural stress indicators

d19 ff: no behavioural stress indicators

	20230701	20230702	20230703	20230704	20230705	20230706	20230707	20230708	20230709	20230710	20230711	20230712	20230713	20230714	20230715	20230716	20230717	20230718	20230719	20230720	20230721	20230722	20230723	20230724	20230725	20230726	20230727	20230728	20230729	20230730	20230731
Total	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.00	1.00	2.00	3.00	1.00	0.00	1.00	2.00	5.00	7.00	7.00	10.00	14.00	15.00	16.00	19.00	21.00	21.00
median	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.00	1.00	1.00	1.00
mean	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.05	0.05	0.10	0.14	0.05	0.00	0.05	0.10	0.24	0.33	0.33	0.48	0.67	0.71	0.76	0.90	1.00	1.00
max	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.00	1.00	1.00	1.00	1.00	0.00	1.00	1.00	2.00	2.00	2.00	2.00	2.00	3.00	3.00	3.00	3.00	3.00
External signs	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	2	3	1	0	1	2	5	5	5	9	11	11	11	13	13	14
nutrition state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1
spine	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
swelling, proliferative lesion	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
egg-bound (female)	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
other deformities	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
skin lesions	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	0	0	1	1	2	2	2	2	2	3	3	3	3	3
skin color	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
scales	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	1	0	1	2	1	1	2	2	2
fin state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	2	2	2	2	2	2
eye state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
gill state	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
ventilation	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	2	2	2	2	2	3
skin health / infection signs	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	1	1	2	2	2	2	2	3	3	3
Behavioural signs	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	2	1	3	4	5	6	8	7
navigation	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	1	1	1
activity level	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
stereotypies	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
fin posture	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	2	2	1
position in tank	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
breathing	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	2	2
scratching	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
feeding	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1

Chart annotations: experimental timeframe; first skin defects; first buoyancy issues.

Chart legend: Total; median; mean; max; External signs; Behavioural signs.