Effect of diffusivity of amyloid beta monomers on the formation of senile plaques

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Abstract

Alzheimer's disease (AD) presents a perplexing question: why does its development span decades, even though individual amyloid beta (Aβ) deposits (senile plaques) can form rapidly in as little as 24 hours, as recent publications suggest? This study investigates whether the formation of senile plaques can be limited by factors other than polymerization kinetics alone. Instead, their formation may be limited by the diffusion-driven supply of Aβ monomers, along with the rate at which the monomers are produced from amyloid precursor protein (APP), and the rate at which Aβ monomers undergo degradation. A mathematical model incorporating the nucleation and autocatalytic process (via the Finke-Watzky model), as well as Aβ monomer diffusion, was proposed. The obtained system of partial differential equations was solved numerically, and a simplified version was investigated analytically. The computational results predicted that it takes approximately 7 years for Aβ aggregates to reach a neurotoxic concentration of 50 μM. Additionally, a sensitivity analysis was performed to examine how the diffusivity of Aβ monomers and their production rate impact the concentration of Aβ aggregates.

Keywords: neuron; Alzheimer's disease; Finke−Watzky model, mathematical modeling; neurotoxicity; polymerization

Abbreviations

Aβ	amyloid beta
AD	Alzheimer's disease
CV	control volume
F-W	Finke-Watzky

1. Introduction

Alzheimer's disease (AD) is a serious neurodegenerative condition and remains the primary cause of dementia [1-3]. One prominent feature of AD is the spontaneous clustering of amyloid peptides into fibrillar aggregates, driven by a self-sustaining process known as secondary nucleation [4]. It is widely recognized that individuals with AD display a significant buildup of amyloid beta (Aβ) deposits, also known as senile plaques, in their brain tissue. Yet, there remains ongoing debate regarding whether these deposits actively contribute to the onset of the disease or simply signify its advancement [5-7]. Aβ monomers stem from the breakdown of amyloid precursor protein (APP), synthesized in neurons, by β- and γ-secretase enzymes [6,8]. The majority of Aβ monomers generated are released into the extracellular space, where they have the potential to aggregate [9].

Multiple studies have linked the propagation of amyloidogenic proteins to various neurodegenerative conditions like AD, Parkinson's disease, Amyotrophic Lateral Sclerosis, and Huntington's disease [10,11]. In ref. [12], the Finke-Watzky (F-W) two-step model was employed to fit previously published data on amyloid-beta (Aβ) aggregation, which were reported in refs. [13,14]. Typically, data is provided for two isoforms: Aβ40 and Aβ42, with Aβ42 being more prone to aggregation.

The buildup of senile plaques in individuals with AD develops gradually over many years. Interestingly, a study conducted in ref. [15] revealed that individual plaques can form quite rapidly, within approximately 24 hours. To understand this apparent paradox, this investigation explored the role of Aβ monomer diffusion in the process of Aβ aggregate formation. The proposed hypothesis states that the slow diffusion of Aβ, along with the limited rate of synthesis of Aβ monomers, can be contributing factors to the gradual formation of senile plaques. Thus, it is proposed that the rate of plaque formation is influenced not only by the kinetics of their growth but also by the rate at which the essential reactants (Aβ monomers) are produced and supplied through diffusion, and also by the rate at which Aβ monomers are destroyed by the proteolytic system.

Mathematical models were developed in [16-18] to understand the intraneuronal mechanisms driving the formation of amyloid-β plaques. Previous research primarily focused on simulating intracellular processes and neglected the extracellular diffusion of Aβ monomers. Given that Aβ monomers can diffuse very easily, it is important to incorporate this extracellular diffusion process. The current study builds upon recent studies reported in [19-21], broadening the scope of prior models by focusing on the diffusion of Aβ monomers in the extracellular space and its impact on the formation of Aβ aggregates.

2. Materials and models

2.1. Model equations

The F-W model simplifies the process into two steps, delineating the formation of particular aggregates through nucleation and autocatalysis reactions. It consists of two pseudo-elementary reaction steps: nucleation and autocatalysis, respectively. During the nucleation step, new aggregates form continuously, while in the autocatalysis step, these aggregates undergo rapid surface growth [12,22]. The two pseudo-elementary reaction steps involve the following:

$$A \xrightarrow{k_1} B\,, \tag{1}$$

$$A + B \xrightarrow{k_2} 2B\,, \tag{2}$$

where A stands for a monomeric protein, while B stands for an amyloid-converted protein. The kinetic constants k_1 and k_2 denote the rates of nucleation and autocatalytic growth, respectively [12]. The primary nucleation process, as described by Eq. (1), involves monomers only. Secondary nucleation, as outlined in Eq. (2), encompasses both monomers and pre-existing aggregates of the same peptide [4].

This study employs the F-W model to simulate the conversion of monomers, whose concentration is C_A, into aggregates, whose concentration is represented by C_B. These aggregates include different forms of Aβ oligomers, protofibrils, and fibrils [23]. The buildup of Aβ aggregates results in the formation of amyloid plaques. The simplified nature of the F-W model means it cannot distinguish between different types and sizes of aggregates. Typically, the F-W model is used to study how an initial concentration of monomers transforms into aggregates. In this study, a novel extension of the F-W model is proposed to address scenarios where monomers are continuously introduced through diffusion from the left-hand side of the control volume (CV), represented by the position x=0 (Fig. 1). At the boundary x=0, which could represent locations such as the cellular membrane [24,25], Aβ monomers are produced through the cleavage of amyloid precursor protein (APP). The boundary at x=L represents an extracellular location. Thus, the model proposed in this paper is

$$\frac{\partial C_A}{\partial t} = D_A \frac{\partial^2 C_A}{\partial x^2} - k_1 C_A - k_2 C_A C_B - C_A \frac{\ln(2)}{T_{1/2,A}}\,, \tag{3}$$

$$\frac{\partial C_B}{\partial t} = k_1 C_A + k_2 C_A C_B\,. \tag{4}$$

The initial conditions are outlined below:

At $t=0$: $C_A=0$, $C_B=0$. (5a,b)

The boundary conditions are outlined below:

At $x=0$: $-D_A \frac{\partial C_A}{\partial x} = q_{A,0}$. (6)

At $x=L$: $\frac{\partial C_A}{\partial x} = 0$. (7)

Eqs. (6) and (7) assume the release of Aβ monomers from the boundary situated at x=0. The boundary at x=L is modeled as a symmetric boundary (L represents the half distance between senile plaques). Therefore, there is no flux of Aβ monomers through this boundary.

The model utilizes independent variables detailed in Table 1, while Table 2 gives a summary of the dependent variables. Table 3 gives an overview of the parameters used in the model.

Table 1. Independent variables utilized in the model.

Symbol	Definition	Units
t	Time	s
x	Coordinate measuring the distance from the CV's face from which Aβ monomers are supplied	μm

Table 2. Dependent variables utilized in the model.

Symbol	Definition	Units
C_A	Concentration of Aβ monomers	μM
C_B	Concentration of Aβ aggregates	μM

Table 3. Parameters utilized in the model and the discussion of results.

Symbol	Definition	Units	Value or range	Reference or estimation method	Value(s) used in computations

$C_{A,0}$	Aβ monomer concentration	μM	0.25×10^{-3} - 0.425×10^{-3} [a]	[26]	
$C_{B,tox}$	Neurotoxic Aβ oligomer concentration	μM	50 [b]	[27]	
D_A	Diffusivity of Aβ monomers	$\mu m^2/s$	62.3 [c]	[26]	62.3
k_1	Rate constant for the first step of the F-W model, describing Aβ aggregate nucleation	s^{-1}	2.22×10^{-9} [d]	[13,12]	2.22×10^{-9}
k_2	Rate constant for the second step of the F-W model, describing autocatalytic growth of Aβ aggregates	$\mu M^{-1}\ s^{-1}$	9.44×10^{-6} [d]	[13,12]	9.44×10^{-6}
L	Half-distance separating senile plaques	μm	50	Estimated from Fig. 1a in [28]	50
$q_{A,0}$	Flux of Aβ monomers from the left-hand side boundary of the CV with a volume L^3	$\mu M\ \mu m\ s^{-1}$	1.1×10^{-5} [e]	[27]	1.1×10^{-5}
$T_{1/2,A}$	Half-life of Aβ monomers	s	4.61×10^{3} [f]	[26]	4.61×10^{3}, 4.61×10^{9} (to represent $T_{1/2,A}\to\infty$)

[a] The parameter ranges from 250 pM to 425 pM, as reported in [26]. Although this parameter is not directly utilized in the model, it is included in the table due to its significance in discussing the predicted concentration of Aβ monomers.

[b] The value corresponds to the Aβ42 isoform [27]. Although this parameter is not directly utilized in the model, it is included in the table because it is relevant for discussing the model's predictions concerning the time it takes for Aβ aggregates to reach neurotoxic concentrations.

[c] The Aβ monomer diffusivity value is 0.623×10^{-6} cm^2/s [26]. This value is smaller than the one reported in [29] (1.43×10^{-6} cm^2/s) because the estimate from [26] accounts for tortuosity. Tortuosity considers the reduction of diffusion in brain tissue due to barriers like cells and the extracellular matrix.

[d] The values reported in [12] were obtained by fitting data from [13], resulting in values of $k_1 = 8\times10^{-6}$ h^{-1}, $k_2 = 3.4\times10^{-2}$ μM^{-1} h^{-1} [12].

[e] Based on [27], the human brain synthesizes Aβ42 monomers at a rate of 0.33×10^{-6} μmol s^{-1}. Considering the brain's approximate volume to be 1.5×10^{15} μm^3, the production rate per unit volume is estimated as 2.2×10^{-22} μmol μm^{-3} s^{-1}. In a small cube with a side length of L, which represents the CV in Fig. 1, the Aβ42 production rate is 2.75×10^{-17} μmol s^{-1}. Aβ monomers are produced at the lipid membrane [30]. Assuming all Aβ42 is released from the cube's left-hand side face with an area of L^2, the release rate per unit area is estimated as 1.1×10^{-20} μmol μm^{-2} s^{-1} $=1.1\times10^{-5}$ μM μm s^{-1}.

[f] As per ref. [26], the value is 1.28 hours.

The dimensionless version of Eqs. (3)-(7) can be found in section S1.1 of the Supplementary Materials. The Aβ aggregation process is affected by three dimensionless parameters: (i) the diffusivity of Aβ monomers, $D_A^* = D_A / \left(L^2 k_1\right)$; (ii) the production rate of Aβ monomers, $q_{A,0}^* = q_{A,0} k_2 / \left(k_1^2 L\right)$; and (iii) the half-life of Aβ monomers, $T_{1/2,A}^* = T_{1/2,A} k_1$.

If C_A remains nearly constant along x, it suggests that the diffusion of Aβ monomers is very fast. One supporting rationale for this assumption is that if soluble Aβ diffuses slower than the rate of aggregation, growth will occur primarily at the tips of the aggregate, leading to a branched, tree-like structure. However, this contradicts experimental observations [31]. With this assumption, the CV depicted in Fig. 1 can be regarded as a lumped capacitance body and Eqs. (3) and (4) can be recast as follows:

$$\frac{dC_A}{dt} = -k_1 C_A - k_2 C_A C_B - C_A \frac{\ln(2)}{T_{1/2,A}} + \frac{q_{A,0}}{L}, \tag{8}$$

$$\frac{dC_B}{dt} = k_1 C_A + k_2 C_A C_B . \tag{9}$$

The dimensionless versions of Eqs. (8) and (9) are given in section S1.2 of the Supplementary Materials. When Eqs. (8) and (9) are added, the following result is obtained:

$$\frac{dC_A}{dt} + \frac{dC_B}{dt} = -C_A \frac{\ln(2)}{T_{1/2,A}} + \frac{q_{A,0}}{L} . \tag{10}$$

For $T_{1/2,A} \to \infty$, integrating Eq. (10) with respect to time and using the initial condition from Eq. (5a,b) gives the following result:

$$C_A + C_B = \frac{q_{A,0}}{L} t . \tag{11}$$

The increase in $C_A + C_B$ over time is due to the flux of Aβ monomers through the left-hand side boundary of the CV. By eliminating C_A from Eq. (4) using Eq. (11), the following equation is obtained:

$$\frac{dC_B}{dt} = k_1 \left(\frac{q_{A,0}}{L} t - C_B \right) + k_2 \left(\frac{q_{A,0}}{L} t - C_B \right) C_B . \tag{12}$$

Eq. (12) is similar to the one studied in ref. [32]. To find the exact solution of Eq. (12) with the initial condition given by Eq. (5b), the DSolve function followed by the FullSimplify function in Mathematica 13.3 (Wolfram Research, Champaign, IL) was employed. The resulting solution is as follows:

$$C_B = \frac{k_1}{k_2} \times \left(-1 + \frac{\left(k_2 q_{A,0}\right)^{1/2} \exp\left[k_1 t + \frac{k_2 q_{A,0}}{2L} t^2 \right]}{\left(k_2 q_{A,0}\right)^{1/2} - (2L)^{1/2} k_1 \left(F\left(\frac{k_1 L^{1/2}}{\left(2 k_2 q_{A,0}\right)^{1/2}} \right) \right) - \exp\left[k_1 t + \frac{k_2 q_{A,0}}{2L} t^2 \right] F\left[\frac{k_1 L + k_2 q_{A,0} t}{\left(2 k_2 q_{A,0} L\right)^{1/2}} \right]} \right) . \tag{13}$$

In this equation, $F(x)$ is Dawson's integral:

$$F(x) = \frac{1}{2}\sqrt{\pi}\, e^{-x^2} erfi(x), \tag{14}$$

where $erfi(x)$ is the imaginary error function.

If $t \to 0$, Eq. (13) implies that $C_B \to \frac{k_1 q_{A,0} t^2}{2L}$, indicating that at small *t*, the concentration of Aβ aggregates is directly proportional to the kinetic constant that describes the nucleation of Aβ aggregates. C_A can then be found using Eq. (11) as:

$$C_A = \frac{q_{A,0}}{L} t - C_B . \tag{15}$$

The exact solution provided by Eqs. (13) and (15) is quite cumbersome. A more elegant approximate solution, also valid for the case of $T_{1/2,A} \to \infty$, is

$$C_A = \frac{\frac{q_{A,0}}{L}}{k_1 + k_2 \frac{q_{A,0}}{L} t}, \tag{16}$$

$$C_B = \frac{q_{A,0}}{L} t . \tag{17}$$

Note that as $t \to \infty$, Eq. (16) predicts that $C_A \to 0$. The dimensionless forms of Eqs. (16) and (17) are given in section S1.3 in Supplementary Materials.

2.2. Sensitivity analysis

One of the primary aims of this research is to examine the sensitivity of Aβ aggregate concentration to variations in the diffusivity of Aβ monomers. The study also explored how changes in the rate of Aβ monomer production, represented by $q_{A,0}$, affect the concentration of Aβ aggregates.

This was accomplished by calculating the local sensitivity coefficients, which represent the first-order partial derivatives of the observable (the concentration of Aβ aggregates) with respect to the diffusivity of Aβ monomers or the flux of Aβ monomers entering the CV through the left-hand side boundary, following methodologies described in [33-36]. The sensitivity coefficient of C_B to, for instance, D_A, was calculated as follows:

$$\frac{\partial C_B}{\partial D_A} \approx \left. \frac{C_B\left(D_A + \Delta D_A\right) - C_B\left(D_A\right)}{\Delta D_A} \right|_{\text{other parmeters kept constant}}, \tag{18}$$

where $\Delta D_A = 10^{-3} D_A$ represents the step size. Calculations were conducted using various step sizes to assess the sensitivity coefficients' independence of the step size.

The non-dimensionalized relative sensitivity coefficient was calculated following the methods outlined in [34,37], as demonstrated below (for example):

$$S_{D_A}^{C_B} = \frac{D_A}{C_B} \frac{\partial C_B}{\partial D_A}. \tag{19}$$

S3. Numerical solution

Eqs. (3) and (4) form a system of two partial differential equations (PDEs). They were solved subject to initial conditions given by Eq. (5) and boundary conditions given by Eqs. (6) and (7) using a well-validated MATLAB PDEPE solver (MATLAB R2020b, MathWorks, Natick, MA, USA).

MATLAB's ODE45 solver was used to solve Eq. (12) with initial condition (5b) (for large values of *t*, using MATLAB's solver works better than the analytical solution given by Eq. (13)). To ensure accurate results, the error tolerance parameters, RelTol and AbsTol, were set to 1e-10.

3. Results

Figs. 2 and 3 depict the scenario where the half-life of Aβ monomers is assumed to be infinite. In Fig. 2a, the concentration of Aβ monomers is approximately 0.465×10^{-3} μM, a value nearly within the range reported in ref. [26] (Table 3): 0.25×10^{-3} to 0.425×10^{-3} μM. The concentrations of monomers, C_A, and aggregates, C_B, gradually decrease from *x*=0 to *x*=*L*. The concentrations decrease because monomers diffuse from the boundary with the highest concentration (*x*=0) towards the boundary with a symmetric (zero flux) condition (*x*=*L*), where both Aβ monomer and aggregate concentrations reach their lowest levels (see Figs. 2a and 2b). In this scenario, the concentration of aggregates reaches the neurotoxic level of 50 μM (Fig. 2b) within the timescale of $t = 7.23$ years.

Initially, at the right-hand side boundary of the CV, $x = L$, the concentration of Aβ monomers is zero, as indicated by the initial condition in Eq. (5a). Subsequently, it rapidly increases, followed by a decline, and

then stabilizes at a low level due to the conversion of monomers into aggregates (Fig. 3a). The concentration of aggregates steadily increases in a linear fashion owing to the continuous supply of reactants (monomers) through the left-hand side boundary of the CV at x=0 (Fig. 3b). When the diffusivity of the monomers is decreased, the concentration of aggregates exhibits a slightly slower increase, as illustrated by the red solid curve in Fig. 3b corresponding to $D_A = 6.23$ μm²/s.

To gain a deeper understanding of how the concentrations of Aβ monomers and aggregates evolve over time, a simplified lumped capacitance model given by Eqs. (8) and (9) was developed. In Fig. S1, presented in the Supplementary Materials, the numerical solution of Eqs. (8) and (9) for the case of $T_{1/2,A} \to \infty$ was compared with the approximate analytical solution given by Eqs. (16) and (17). Remarkably, the approximate analytical solution of the lumped capacitance model shows excellent agreement with the numerical solution. As time progresses, the concentration of Aβ monomers decreases proportionally to $\frac{1}{t}$ (Eq. (16)). This is consistent with the F-W model, which disregards any potential equilibrium state between the monomers and aggregates, and predicts that the aggregation continues until all monomers are converted into aggregates [38,39]. The concentration of Aβ aggregates increases linearly with time (Eq. (17)). This is due to the reactants (Aβ monomers) entering the CV (Fig. 1) at a constant rate of $q_{A,0}L^2$; all reactants are subsequently converted into Aβ aggregates.

For a finite half-life of Aβ monomers, $T_{1/2,A} = 4.61\times10^3$ s (Figs. 4 and 5), the concentration of Aβ aggregates is reduced significantly compared to the scenario with an infinite half-life, decreasing from approximately 50 μM to around 5.25×10^{-3} μM (compare Fig. 4b with Fig. 2b). The significant drop in aggregate concentration occurs because, with a shorter half-life, a considerable portion of the monomers are broken down by the proteolytic system instead of forming aggregates. This finding has important implications for AD. Numerous studies have associated AD with the malfunctioning of the proteolytic machinery, leading to the inability to regulate Aβ concentrations effectively and degrade misfolded Aβ [40-42]. The study's findings suggest that Aβ aggregation can still occur even when Aβ monomer concentrations are relatively low, provided that the protein clearance mechanism fails to consistently degrade newly generated normal monomers. Preventing Aβ aggregation requires more than just breaking down faulty Aβ. Conversely, a well-functioning proteolytic system capable of degrading Aβ monomers can protect the brain from Aβ aggregates.

Interestingly, achieving this protection does not require the ability to degrade aggregates (notably, Eq. (4) lacks a degradation term).

When simulating the scenario with a finite half-life of Aβ monomers, which corresponds to a well-functioning Aβ monomer clearance system, the initial peak in the concentration of Aβ monomers is absent (compare Fig. 5a with Fig. 3a). Furthermore, the increase in the concentration of Aβ aggregates, C_B, does not follow a linear trend and occurs much slower, ultimately reaching a significantly lower value after 7 years (compare Fig. 5b with Fig. 3b).

When the diffusivity of Aβ monomers, D_A, is increased, the concentration of Aβ monomers, C_A, initially exhibits an increase. However, once D_A exceeds 30 μm²/s, C_A reaches a plateau and remains constant even with further increases in D_A (Fig. 6a). A similar trend is observed for the concentration of Aβ aggregates, C_B (Fig. 6b). This suggests that when the diffusivity of Aβ monomers exceeds 30 μm²/s, the diffusion of Aβ monomers no longer presents a limiting factor for Aβ aggregation.

Interestingly, in Fig. 6a, the concentration of Aβ monomers, C_A, for the scenario with an infinitely large half-life is higher than the concentration for the case with a finite half-life of Aβ monomers. It is important to note that the values of C_A shown in Fig. 6a are not steady-state concentrations of monomers; the steady-state monomer concentration is zero (as indicated by Eq. (16)). Fig. 6a is computed for a specific time interval (*t* is changing between 0 and 7.23 years), and if time is increased further, the monomer concentrations for both infinite and finite half-life scenarios will eventually reduce to zero. In Fig. 6b, it is evident that the concentration of aggregates is larger for the scenario with an infinite half-life of monomers, which is consistent with expectations.

It is intriguing that in the scenario with an infinite half-life of Aβ monomers, the concentration of Aβ monomers, C_A, appears to be nearly independent of the rate of Aβ monomer production, $q_{A,0}$, as indicated by the horizontal solid red line in Fig. 7a. To investigate this further, a simplified model based on the lumped capacitance approximation was developed; this model is given by Eqs. (8) and (9). This model assumes a lack of concentration variations, C_A and C_B, with respect to *x*; it accounts for concentrations changing with time but not with the distance from the *x*=0 plane. Fig. S1 explores a hypothetical scenario covering 70 years. Fig. S1a shows that C_A becomes small after a short transient and decreases to zero as time progresses. Since the lumped capacitance model assumes an infinitely large value of Aβ monomer diffusivity, this suggests that C_A is primarily determined by the difference between the production rate of Aβ monomers and their conversion rate into aggregates. The concentration of Aβ aggregates, C_B, exhibits a linear increase with time (Fig. S1b). This is attributed to a constant rate of Aβ

monomer production, $q_{A,0}$, which enables continuous conversion of Aβ monomers into aggregates. Similar trends can be observed in Fig. 3b, where the finite rate of diffusion of Aβ monomers is taken into account.

As depicted in Fig. 7b, for an infinite half-life of Aβ monomers the concentration of Aβ aggregates, C_B, at the boundary $x=L$ (50 μm) and $t = 7.23$ years, increases linearly with $q_{A,0}$ at large times. This behavior is attributed to the fact that $q_{A,0}$ represents the rate at which reactants are produced and subsequently converted into Aβ aggregates. The production rate, $q_{A,0}$, is assumed to be constant and independent of time. When the half-life of Aβ monomers is finite (4.61×10^3 s), an increase in $q_{A,0}$ results in a delayed increase in C_B, as long as $q_{A,0}$ remains small. However, once $q_{A,0}$ exceeds 0.4 μM·μm·s^{-1}, C_B begins to increase at the same rate as it does in the case with an infinite half-life of Aβ monomers (Fig. 7b).

The dimensionless sensitivity of the molar concentration of Aβ aggregates, C_B, to the diffusivity of Aβ monomers, D_A, remains consistently positive at all times t (Fig. 8a). This is because Aβ monomers serve as reactants in the F-W model, which describes Aβ aggregation (Eqs. (1) and (2)). An increase in the diffusivity of monomers enhances the supply of reactants (see Figs. S2 and S3 in the Supplementary Materials, showing the flux of Aβ monomers), resulting in a greater accumulation of aggregated Aβ.

The dimensionless sensitivity of the concentration of Aβ aggregates, C_B, to the rate of Aβ monomer production, $q_{A,0}$, which enters the model through the flux of Aβ monomers from the left-hand side boundary, is also positive at all times t (Fig. 8b). It is worth noting that in the scenario with an infinite half-life of Aβ monomers (represented by the red line in Fig. 8b), the sensitivity of C_B to $q_{A,0}$ quickly stabilizes at a constant value of 1. This behavior is attributed to the linear dependence of C_B on $q_{A,0}$ (as evident from the red solid line in Fig. 7b), indicating that C_B is directly influenced by the rate of production of the reactants (Aβ monomers). The value of 1 in the sensitivity calculation is a result of rescaling in Eq. (19) that is performed to ensure that the dimensionless sensitivity is independent of magnitudes of parameters.

4. Discussion, limitations of the model, and future directions

A popular hypothesis suggests that sporadic forms of AD are linked to impaired clearance of Aβ aggregates [43-45]. The study's findings suggest that preventing AD may not hinge solely on the

proteolytic system's capability to break down Aβ polymers. The results indicate that even if the proteolytic system only breaks down Aβ monomers, the aggregation of Aβ cannot advance due to the insufficient availability of reactants (monomers). This insight has significant implications for potential therapeutic approaches to AD.

The current model has limitations, primarily because the F-W model it utilizes does not account for the size variability of Aβ polymers. Future studies should focus on developing models that accurately depict the interaction between Aβ aggregates and tau tangles [8]. To tackle this issue, the model should incorporate cross-membrane diffusion processes, considering that Aβ aggregates primarily form in the extracellular space, while tau tangles are mainly intracellular. Moreover, future modeling should consider the activation of microglia in response to Aβ aggregation. Exploring the potential synergy between microglia-mediated degradation of Aβ aggregates and the neuroinflammatory response induced by microglia activation [46,23,15] is crucial.

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References

1. Hardy, J.: A hundred years of Alzheimer's disease research. Neuron **52,** 3-13 (2006)

2. Maqbool, M., Mobashir, M., Hoda, N.: Pivotal role of glycogen synthase kinase-3: A therapeutic target for Alzheimer's disease. European Journal of Medicinal Chemistry **107,** 63-81 (2016)

3. Hung, S., Fu, W.: Drug candidates in clinical trials for Alzheimer's disease. Journal of Biomedical Science **24,** 47 (2017)

4. Thacker, D., Barghouth, M., Bless, M., Zhang, E., Linse, S.: Direct observation of secondary nucleation along the fibril surface of the amyloid β 42 peptide. Proceedings of the National Academy of Sciences **120,** e2220664120 (2023)

5. Hardy, J., Selkoe, D.J.: The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. Science (New York, N.Y.) **297,** 353-356 (2002)

6. O'Brien, R.J., Wong, P.C.: Amyloid precursor protein processing and Alzheimer's disease. Annual Review of Neuroscience, Vol 34 **34,** 185-204 (2011)

7. Selkoe, D.J., Hardy, J.: The amyloid hypothesis of Alzheimer's disease at 25years. EMBO Molecular Medicine **8,** 595-608 (2016)

8. Hampel, H., Hardy, J., Blennow, K., Chen, C., Perry, G., Kim, S.H., Villemagne, V.L., Aisen, P., Vendruscolo, M., Iwatsubo, T., Masters, C.L., Cho, M., Lannfelt, L., Cummings, J.L., Vergallo, A.: The amyloid-β pathway in Alzheimer’s disease. Molecular Psychiatry **26,** 5481-5503 (2021)

9. Rahman, M.M., Lendel, C.: Extracellular protein components of amyloid plaques and their roles in Alzheimer’s disease pathology. Molecular Neurodegeneration **16,** 59 (2021)

10. Jucker, M., Walker, L.C.: Self-propagation of pathogenic protein aggregates in neurodegenerative diseases. Nature **501,** 45-51 (2013)

11. Klimova, N., Makarava, N., Baskakov, I.V.: The diversity and relationship of prion protein self-replicating states. Virus Research **207,** 113-119 (2015)

12. Morris, A.M., Watzky, M.A., Agar, J.N., Finke, R.G.: Fitting neurological protein aggregation kinetic data via a 2-step, minimal/"Ockham's razor" model: The Finke-Watzky mechanism of nucleation followed by autocatalytic surface growth. Biochemistry **47,** 2413-2427 (2008)

13. Bieschke, J., Zhang, Q., Powers, E.T., Lerner, R.A., Kelly, J.W.: Oxidative metabolites accelerate Alzheimer's amyloidogenesis by a two-step mechanism, eliminating the requirement for nucleation. Biochemistry **44,** 4977-4983 (2005)

14. Vestergaard, M., Kerman, K., Saito, M., Nagatani, N., Takamura, Y., Tamiya, E.: A rapid label-free electrochemical detection and kinetic study of Alzheimer's amyloid beta aggregation. Journal of the American Chemical Society **127,** 11892-11893 (2005)

15. Meyer-Luehmann, M., Spires-Jones, T.L., Prada, C., Garcia-Alloza, M., de Calignon, A., Rozkalne, A., Koenigsknecht-Talboo, J., Holtzman, D.M., Bacskai, B.J., Hyman, B.T.: Rapid appearance and local toxicity of amyloid-beta plaques in a mouse model of Alzheimer's disease. Nature **451,** 720-724 (2008)

16. Kuznetsov, I.A., Kuznetsov, A.V.: How the formation of amyloid plaques and neurofibrillary tangles may be related – A mathematical modelling study. Proceedings of the Royal Society A **474,** 20170777 (2018)

17. Kuznetsov, I.A., Kuznetsov, A.V.: Simulating the effect of formation of amyloid plaques on aggregation of tau protein. Proceedings of the Royal Society A-Mathematical Physical and Engineering Sciences **474,** 20180511 (2018)

18. Torok, J., Maia, P.D., Verma, P., Mezias, C., Raj, A.: Emergence of directional bias in tau deposition from axonal transport dynamics. PLOS Computational Biology **17,** e1009258 (2021)

19. Kuznetsov, A.V.: Numerical and analytical simulation of the growth of amyloid-β plaques. ASME Journal of Biomedical Engineering **146,** 061004 (2024)

20. Kuznetsov, A.V.: Numerical modeling of senile plaque development under conditions of limited diffusivity of amyloid-β monomers. bioRxiv 2024.01.09.574935 (2024)

21. Andrey, V.K.: The growth rate of senile plaques is determined by the competition between the rate of deposition of free aβ aggregates into plaques and the autocatalytic production of free aβ aggregates. bioRxiv 2024.04.06.588435 (2024)

22. Iashchishyn, I.A., Sulskis, D., Nguyen Ngoc, M., Smirnovas, V., Morozova-Roche, L.A.: Finke-Watzky two-step nucleation-autocatalysis model of S100A9 amyloid formation: Protein misfolding as "nucleation" event. ACS Chemical Neuroscience **8,** 2152-2158 (2017)

23. Chen, G., Xu, T., Yan, Y., Zhou, Y., Jiang, Y., Melcher, K., Xu, H.E.: Amyloid beta: Structure, biology and structure-based therapeutic development. Acta Pharmacologica Sinica **38,** 1205-1235 (2017)

24. Haass, C., Kaether, C., Thinakaran, G., Sisodia, S.: Trafficking and proteolytic processing of APP. Cold Spring Harbor Perspectives in Medicine **2,** a006270 (2012)

25. Masters, C.L., Selkoe, D.J.: Biochemistry of amyloid β-protein and amyloid deposits in Alzheimer disease. Cold Spring Harbor Perspectives in Medicine **2,** a006262 (2012)

26. Waters, J.: The concentration of soluble extracellular amyloid-beta protein in acute brain slices from CRND8 mice. PLoS One **5,** e15709 (2010)

27. Raskatov, J.A.: What is the “Relevant” amyloid β42 concentration? ChemBioChem **20,** 1725-1726 (2019)

28. Querol-Vilaseca, M., Colom-Cadena, M., Pegueroles, J., Nuñez-Llaves, R., Luque-Cabecerans, J., Muñoz-Llahuna, L., Andilla, J., Belbin, O., Spires-Jones, T., Gelpi, E., Clarimon, J., Loza-Alvarez, P., Fortea, J., Lleó, A.: Nanoscale structure of amyloid-β plaques in Alzheimer’s disease. Scientific Reports **9,** 5181 (2019)

29. Bora, R.P., Prabhakara, R.: Translational, rotational and internal dynamics of amyloid beta-peptides (A beta 40 and A beta 42) from molecular dynamics simulations. Journal of Chemical Physics **131,** 155103 (2009)

30. Dawkins, E., Derks, R.J.E., Schifferer, M., Trambauer, J., Winkler, E., Simons, M., Paquet, D., Giera, M., Kamp, F., Steiner, H.: Membrane lipid remodeling modulates γ-secretase processivity. Journal of Biological Chemistry **299,** (2023)

31. Cruz, L., Urbanc, B., Buldyrev, S.V., Christie, R., Gómez-Isla, T., Havlin, S., McNamara, M., Stanley, H.E., Hyman, B.T.: Aggregation and disaggregation of senile plaques in Alzheimer disease. Proceedings of the National Academy of Sciences of the United States of America **94,** 7612-7616 (1997)

32. Kuznetsov, I.A., Kuznetsov, A.V.: An analytical solution simulating growth of Lewy bodies. Mathematical Medicine and Biology: A Journal of the IMA **39,** 299-312 (2022)

33. J.V. Beck, K.J. Arnold, Parameter Estimation in Science and Engineering, Wiley, New York (1977)

34. Zadeh, K.S., Montas, H.J.: A class of exact solutions for biomacromolecule diffusion-reaction in live cells. Journal of Theoretical Biology **264,** 914-933 (2010)

35. Zi, Z.: Sensitivity analysis approaches applied to systems biology models. IET Systems Biology **5,** 336-346 (2011)

36. Kuznetsov, I.A., Kuznetsov, A.V.: Investigating sensitivity coefficients characterizing the response of a model of tau protein transport in an axon to model parameters. Computer Methods in Biomechanics and Biomedical Engineering **22,** 71-83 (2019)

37. Kacser, H., Burns, J.A., Fell, D.A.: The control of flux. Biochemical Society Transactions **23,** 341-366 (1995)

38. Martin, J.D.: Particle size is a primary determinant for sigmoidal kinetics of nanoparticle formation: A "disproof" of the Finke-Watzky (F-W) nanoparticle nucleation and growth mechanism. Chemistry of Materials **32,** 3651-3656 (2020)

39. Finke, R.G., Watzky, M.A., Whitehead, C.B.: Response to “Particle size is a primary determinant for sigmoidal kinetics of nanoparticle formation: A “Disproof” of the Finke–Watzky (F-W) nanoparticle nucleation and growth mechanism”. Chemistry of Materials **32,** 3657-3672 (2020)

40. Wang, D., Dickson, D.W., Malter, J.S.: Beta-amyloid degradation and Alzheimer's disease. Journal of Biomedicine & Biotechnology **2006,** 58406 (2006)

41. Baranello, R.J., Bharani, K.L., Padmaraju, V., Chopra, N., Lahiri, D.K., Greig, N.H., Pappolla, M.A., Sambamurti, K.: Amyloid-beta protein clearance and degradation (ABCD) pathways and their role in Alzheimer's disease. Current Alzheimer Research **12,** 32-46 (2015)

42. Kim, D.E., Priefer, R.: Therapeutic potential of direct clearance of the amyloid-β in Alzheimer's disease. Brain Sciences **10,** (2020)

43. Selkoe, D.J.: Clearing the brain's amyloid cobwebs. Neuron **32,** 177-180 (2001)

44. Tanzi, R.E., Moir, R.D., Wagner, S.L.: Clearance of Alzheimer's abeta peptide: The many roads to perdition. Neuron **43,** 605-608 (2004)

45. Saido, T., Leissring, M.A.: Proteolytic degradation of amyloid β-protein. Cold Spring Harbor Perspectives in Medicine **2,** a006379 (2012)

46. Hansen, D.V., Hanson, J.E., Sheng, M.: Microglia in Alzheimer's disease. The Journal of Cell Biology **217,** 459-472 (2018)

Figure captions

Fig. 1. A CV with a width of $L = 50$ μm, used to simulate the diffusion of Aβ monomers from the boundary at *x*=0 and the accumulation of Aβ aggregates. The boundary at *x*=*L* is assumed to be symmetric, indicating that no Aβ monomers pass through it.

Fig. 2. Concentration of Aβ monomers, C_A (a), and concentration of Aβ aggregates, C_B (b), plotted against the distance from the surface releasing Aβ monomers (e.g., cell membrane). These computational results are obtained for $t = 2.28 \times 10^8 \text{ s} = 7.23 \text{ years}$ and pertain to the scenario assuming an infinite half-life of Aβ monomers.

Fig. 3. (a) Concentration of Aβ monomers, C_A, plotted against time (in years). (b) The concentration of Aβ aggregates, C_B, also plotted against time (in years). The computational results are presented at the right-hand side boundary of the CV, $x = L$. This scenario assumes an infinite half-life of Aβ monomers.

Fig. 4. (a) The concentration of Aβ monomers, C_A, as a function of the distance from the surface releasing Aβ monomers (such as the cell membrane). (b) The concentration of Aβ aggregates, C_B, as a function of the distance from the same surface. The computational results are displayed at $t = 2.28 \times 10^8 \text{ s} = 7.23 \text{ years}$. This figure illustrates a scenario where Aβ monomers have a finite half-life ($T_{1/2,A} = 4.61 \times 10^3$ s).

Fig. 5. (a) Concentration of Aβ monomers, C_A, plotted against time (in years). (b) Similarly, the concentration of Aβ aggregates, C_B, depicted over time. The computational results are depicted at the right-hand side boundary of the CV, $x = L$. This scenario assumes a finite half-life of Aβ monomers ($T_{1/2,A} = 4.61 \times 10^3$ s).

Fig. 6. (a) Concentration of Aβ monomers, C_A, plotted against the diffusivity of Aβ monomers, D_A. (b) Concentration of Aβ aggregates, C_B, also plotted against the diffusivity of Aβ monomers, D_A. The computational results are shown at the right-hand side boundary of the CV, *x*=*L*, and $t = 2.28 \times 10^8 \text{ s} = 7.23 \text{ years}$.

Fig. 7. (a) The concentration of Aβ monomers, C_A, plotted against the flux of Aβ monomers from the left-hand side boundary of the CV, $q_{A,0}$. (b) The concentration of Aβ aggregates, C_B, also plotted against

the flux of Aβ monomers from the right-hand side boundary of the CV, $q_{A,0}$. The computational results are shown at the boundary $x=L$ and $t = 2.28\times10^{8}\ \text{s} = 7.23\ \text{years}$.

Fig. 8. (a) The dimensionless sensitivity of the molar concentration of Aβ aggregates, C_B, to the diffusivity of Aβ monomers, D_A, over time. (b) The dimensionless sensitivity of C_B to the flux of Aβ monomers from the left-hand side boundary of the CV, $q_{A,0}$, over time. The calculations were carried out for $\Delta D_A = 10^{-3} D_A$, and close results were obtained for $\Delta D_A = 10^{-2} D_A$. The computational findings are displayed at the right-hand side boundary of the CV, $x = L$.

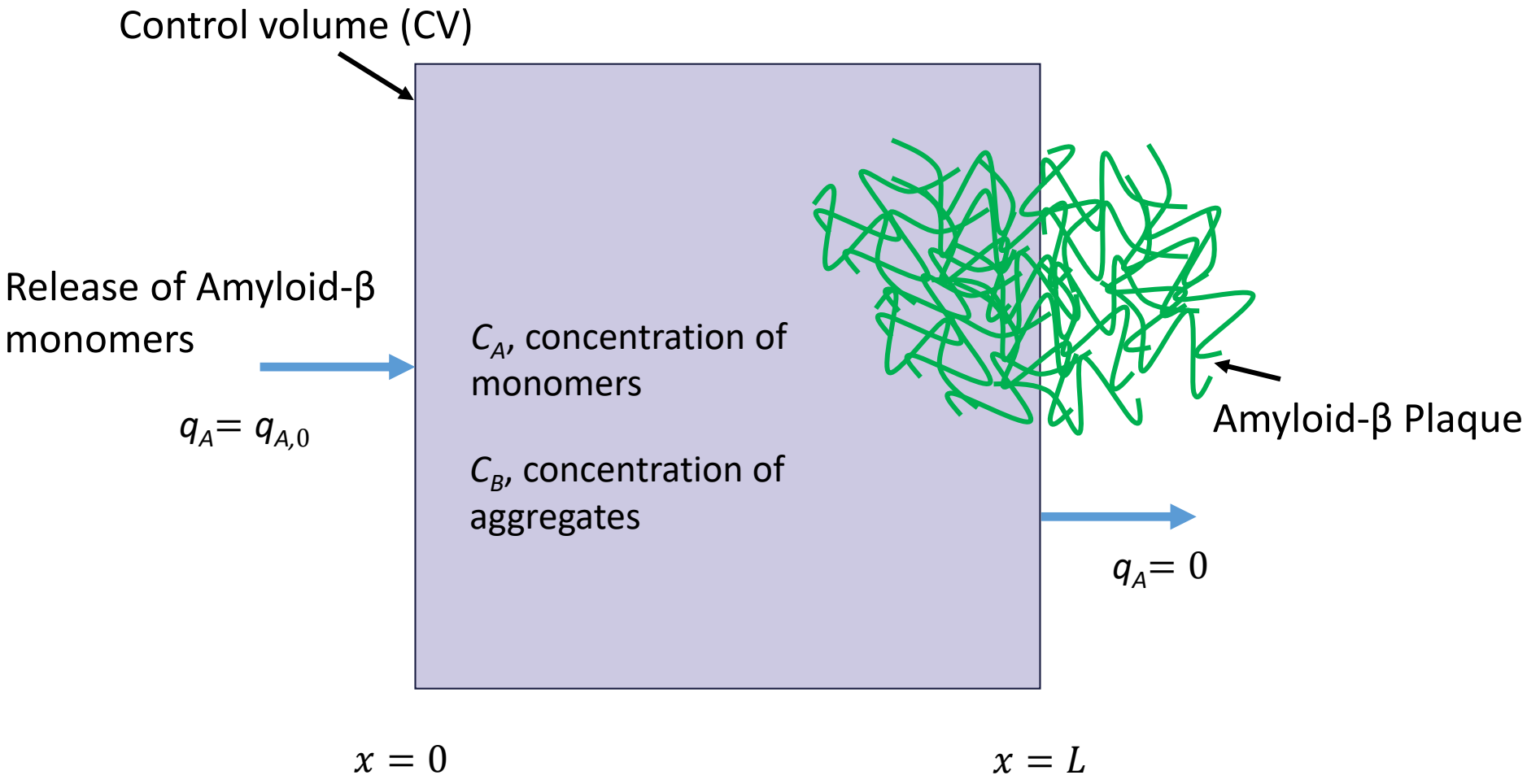
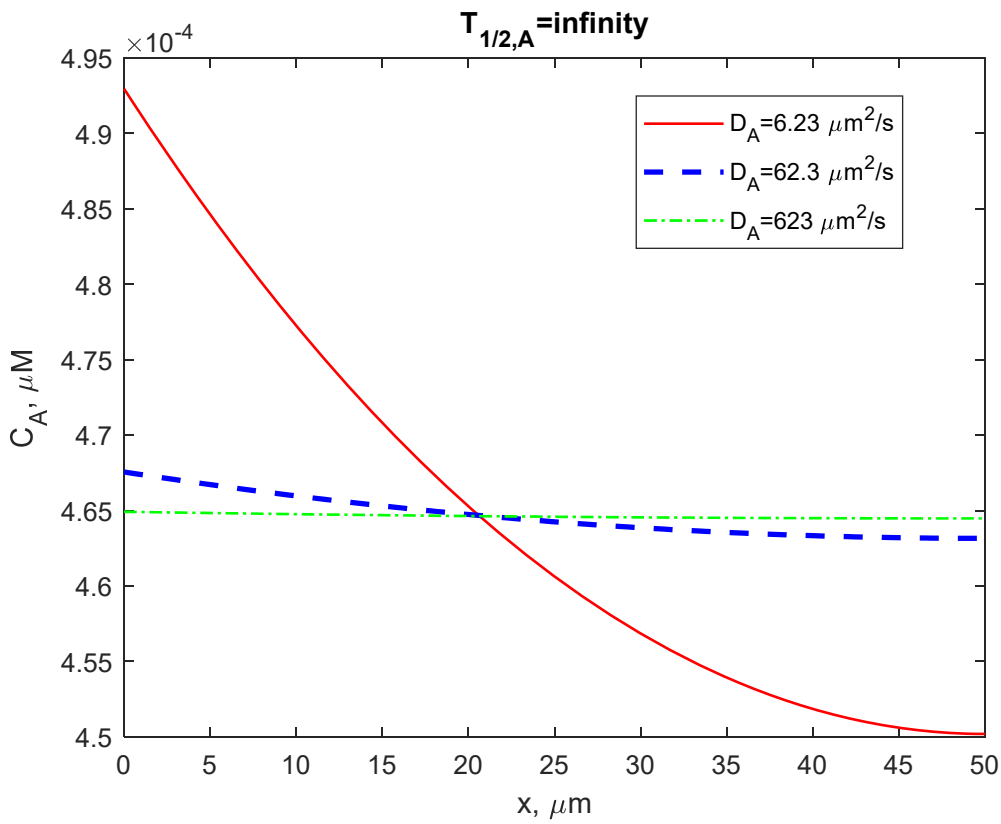

Control volume (CV)
Release of Amyloid-β monomers
$q_A = q_{A,0}$
C_A, concentration of monomers
C_B, concentration of aggregates
Amyloid-β Plaque
$q_A = 0$
$x = 0$
$x = L$

Figure 1

(a)


T1/2,A=infinity
×10-4
4.95
4.9
4.85
4.8
4.75
4.7
4.65
4.6
4.55
4.5
CA, μM
0
5
10
15
20
25
30
35
40
45
50
x, μm
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

(b)

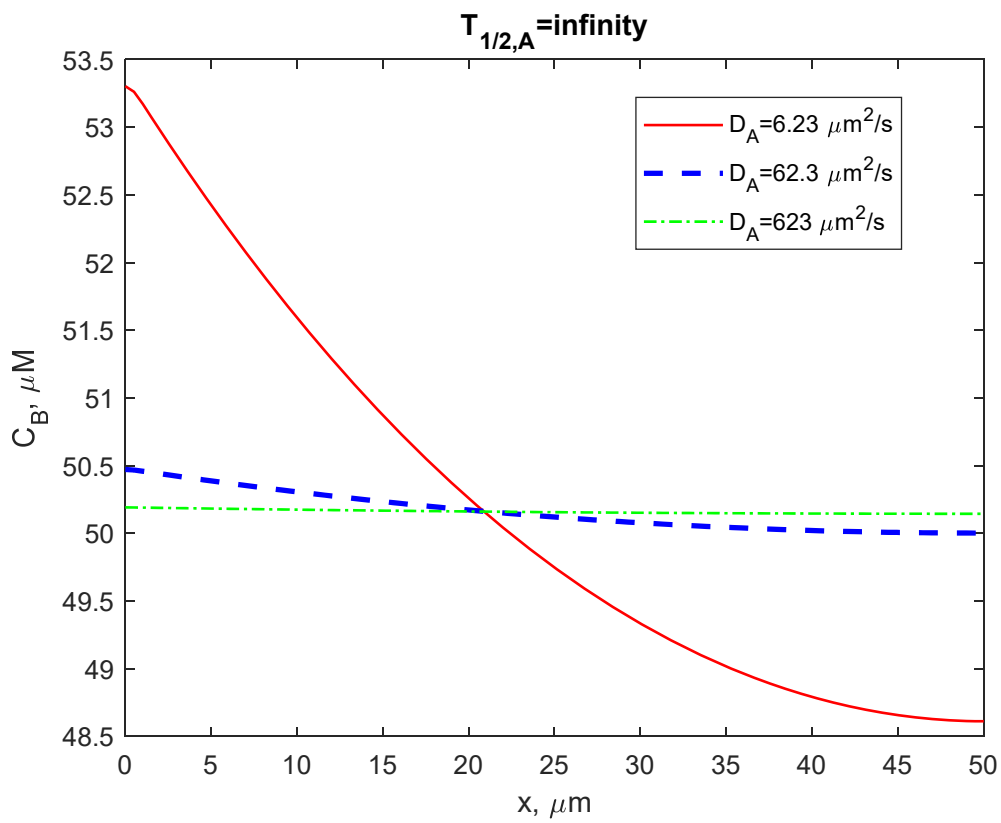
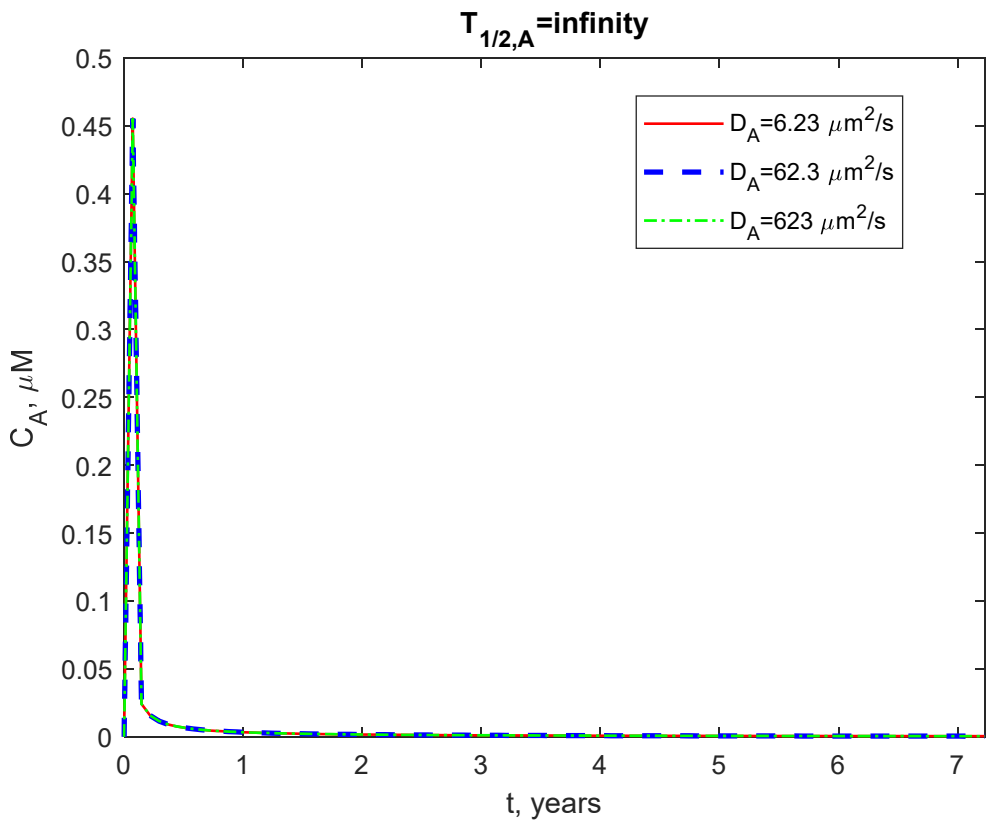

T1/2,A=infinity
53.5
53
52.5
52
51.5
51
50.5
50
49.5
49
48.5
CB, μM
0
5
10
15
20
25
30
35
40
45
50
x, μm
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

Figure 2

(a)


T1/2,A=infinity
0.5
0.45
0.4
0.35
0.3
0.25
0.2
0.15
0.1
0.05
0
CA, μM
0
1
2
3
4
5
6
7
t, years
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

(b)

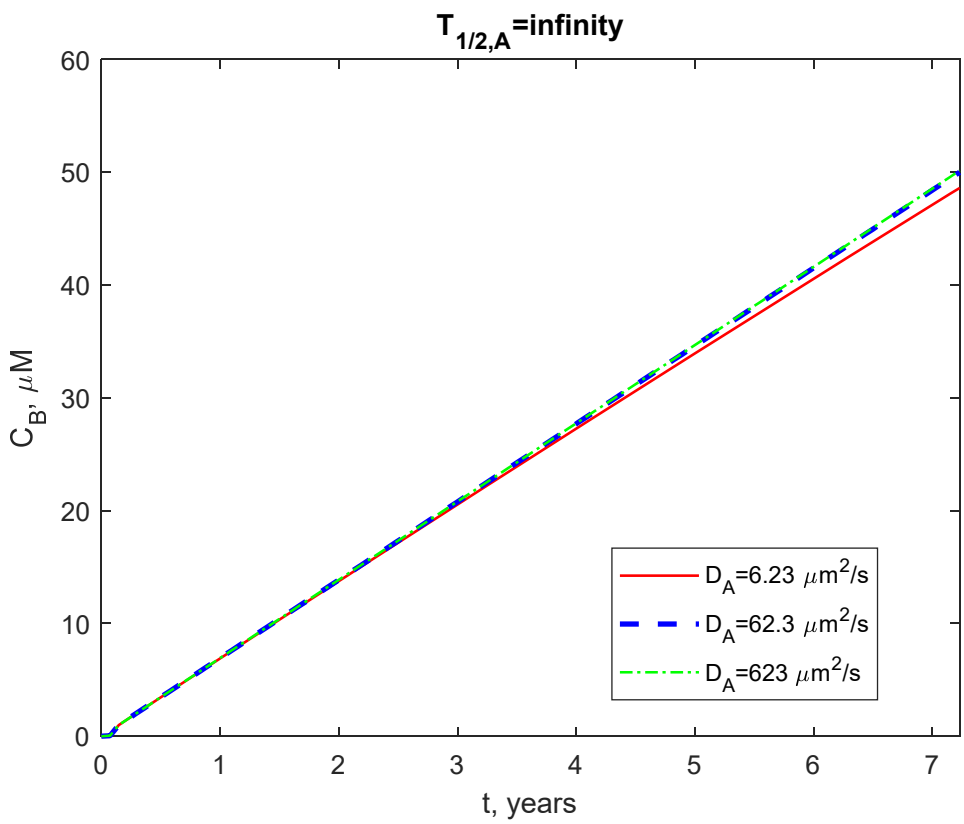
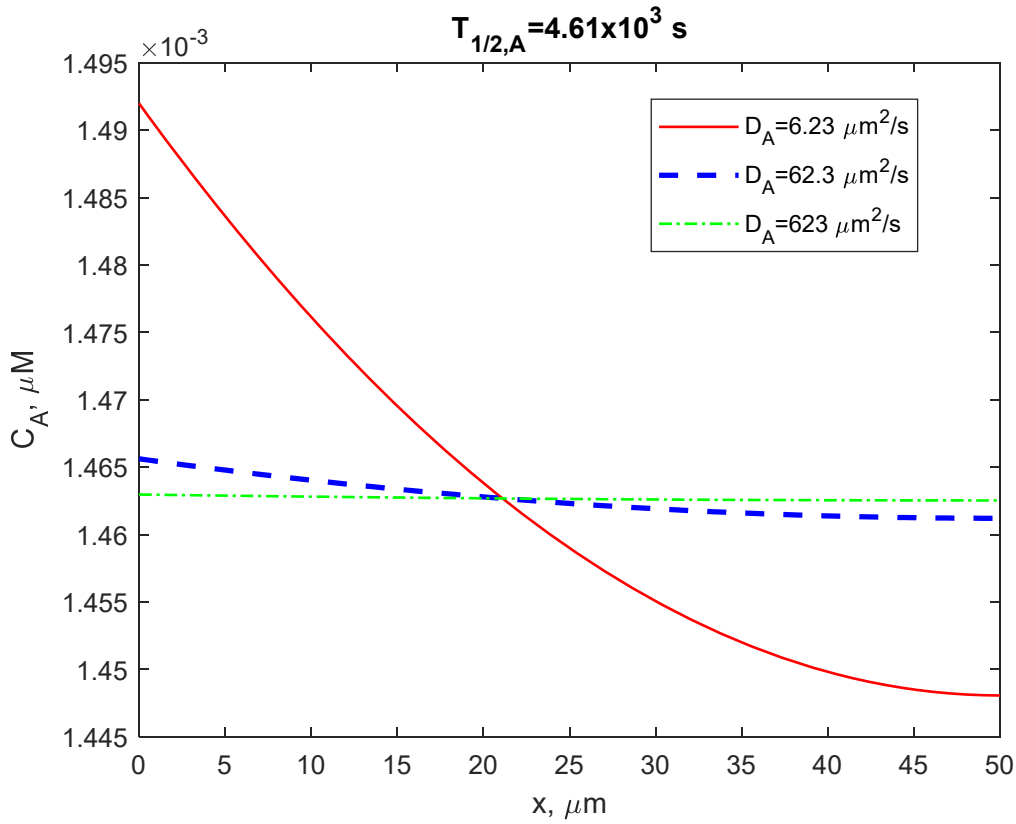

T1/2,A=infinity
60
50
40
30
20
10
0
CB, μM
0
1
2
3
4
5
6
7
t, years
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

Figure 3

(a)


T1/2,A=4.61x10^3 s
×10^-3
1.495
1.49
1.485
1.48
1.475
1.47
1.465
1.46
1.455
1.45
1.445
CA, μM
0
5
10
15
20
25
30
35
40
45
50
x, μm
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

(b)

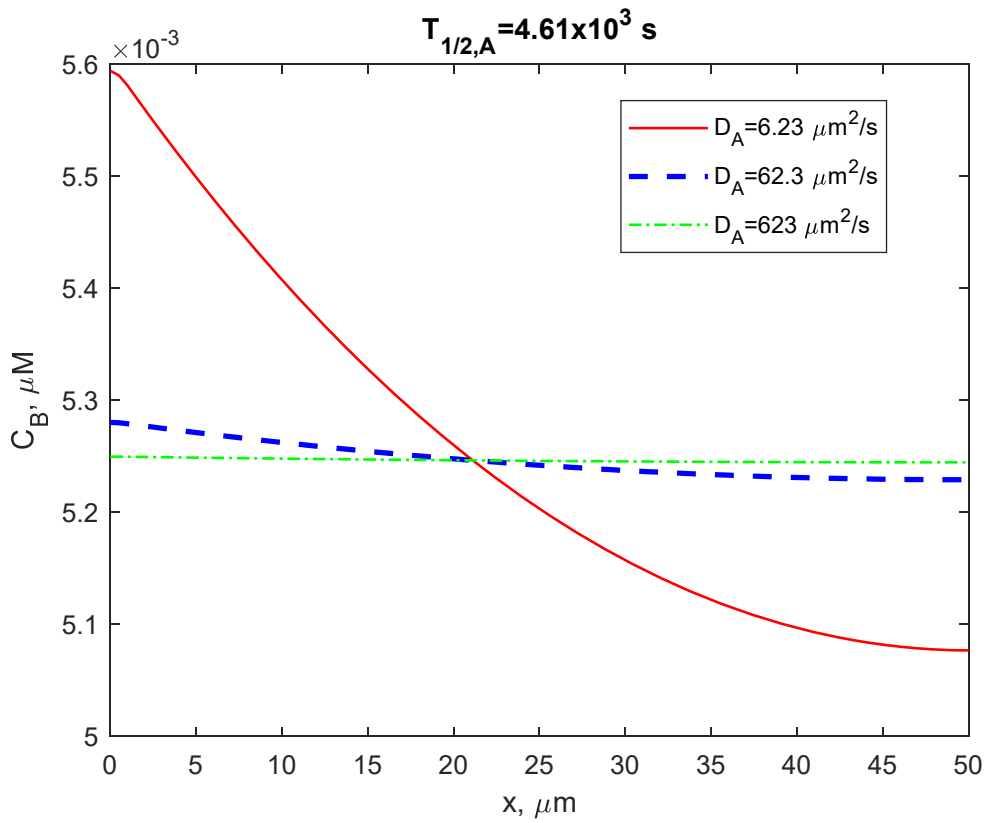
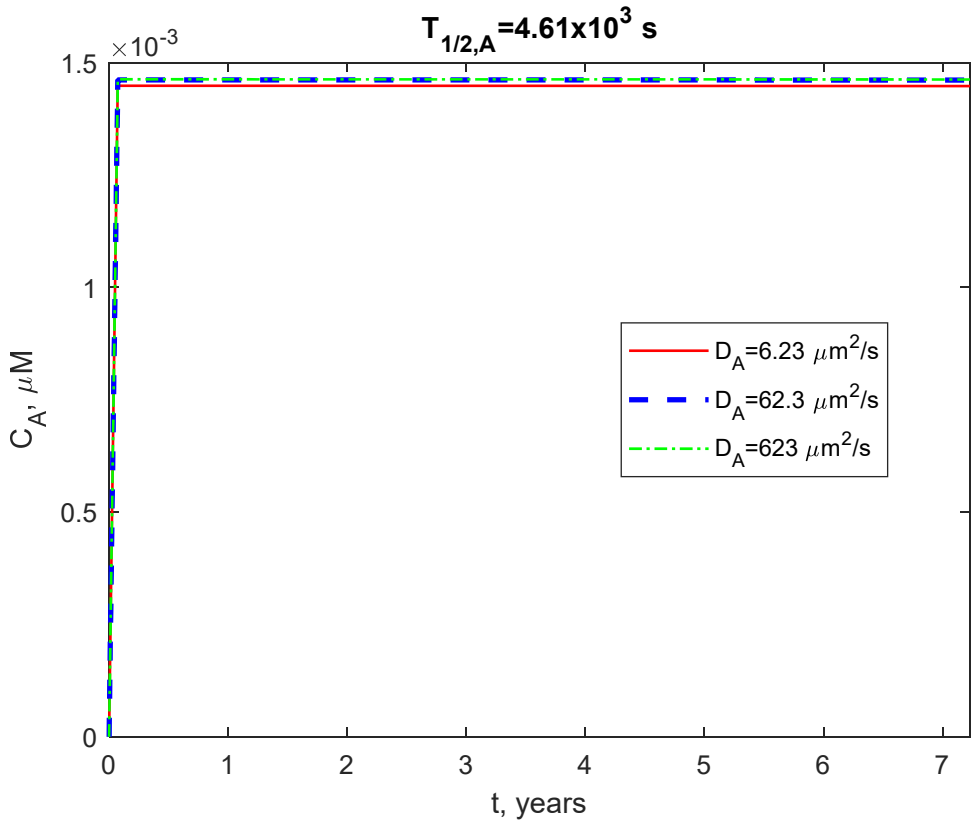

T1/2,A=4.61x10^3 s
×10^-3
5.6
5.5
5.4
5.3
5.2
5.1
5
CB, μM
0
5
10
15
20
25
30
35
40
45
50
x, μm
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

Figure 4

(a)


T1/2,A=4.61x10^3 s
×10^-3
1.5
1
0.5
0
CA, μM
0
1
2
3
4
5
6
7
t, years
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

(b)

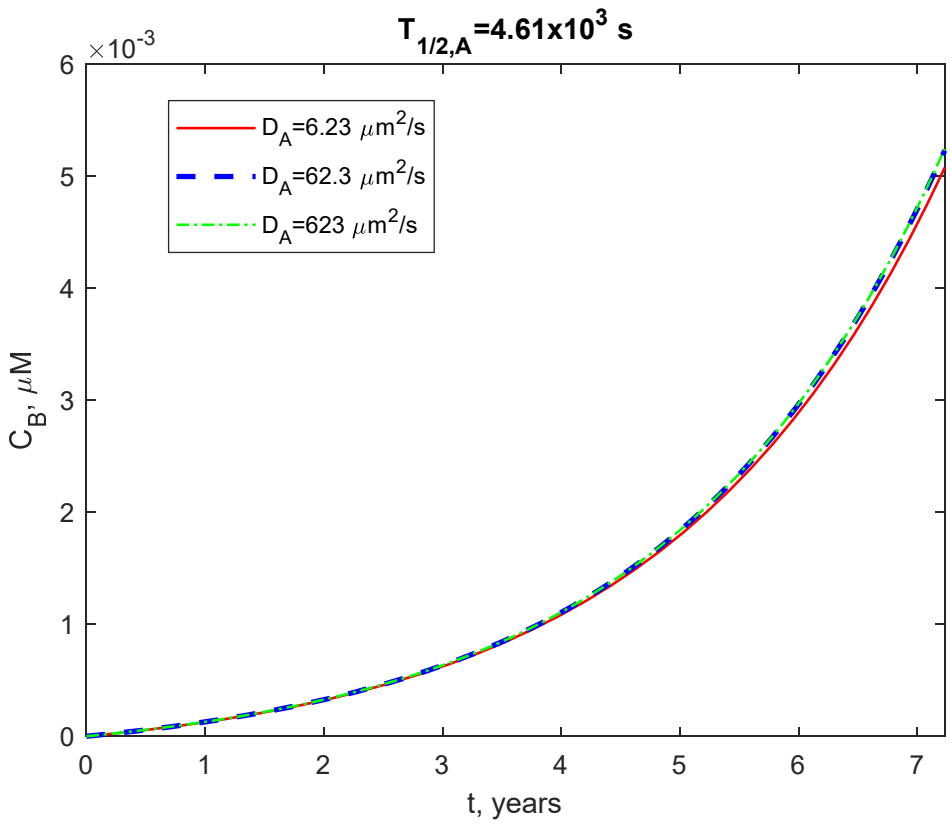
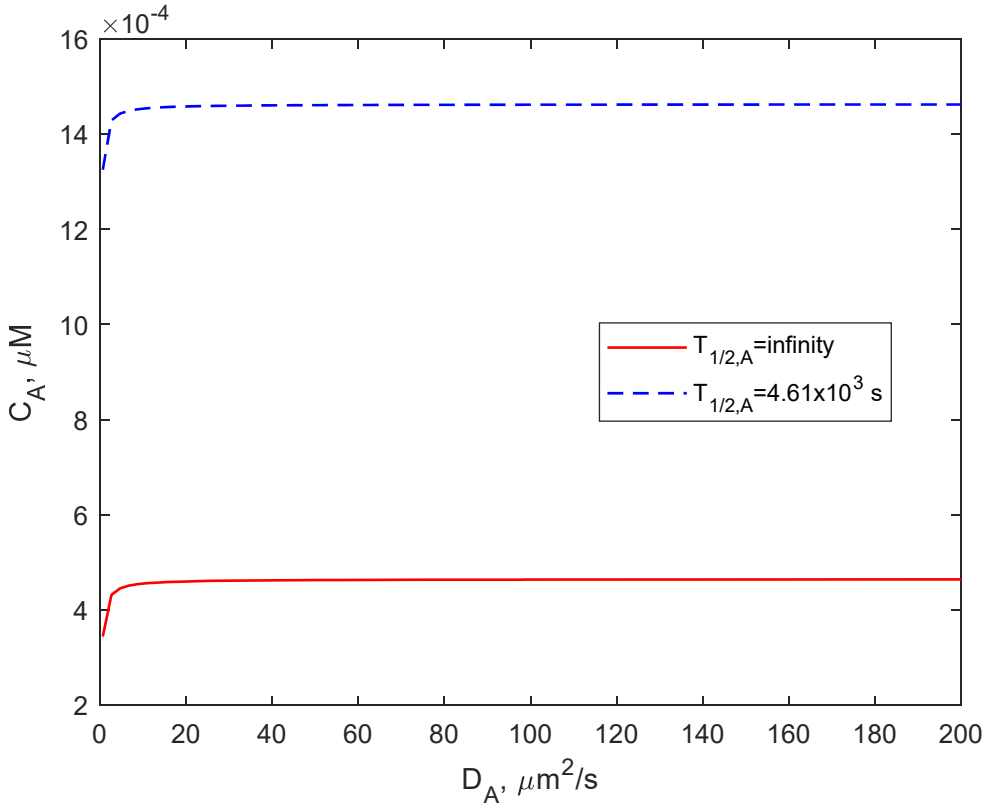

T1/2,A=4.61x10^3 s
×10^-3
6
5
4
3
2
1
0
CB, μM
0
1
2
3
4
5
6
7
t, years
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

Figure 5

(a)


×10^-4
16
14
12
10
8
6
4
2
C_A, μM
0
20
40
60
80
100
120
140
160
180
200
D_A, μm^2/s
T_1/2,A=infinity
T_1/2,A=4.61x10^3 s

(b)

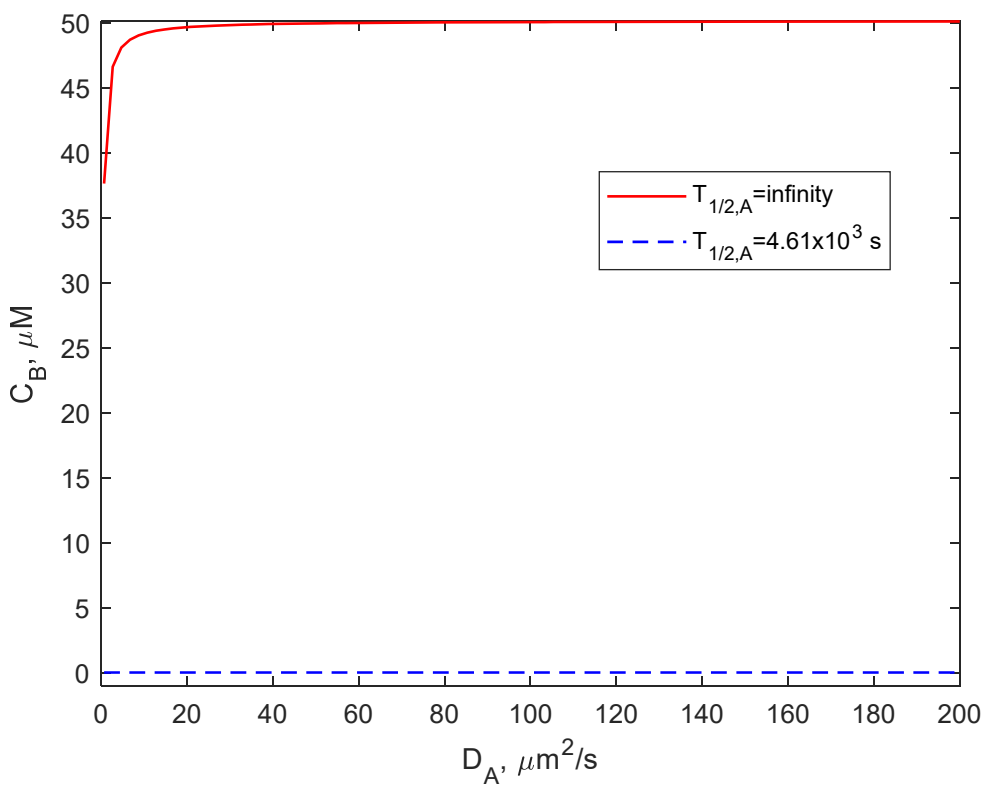
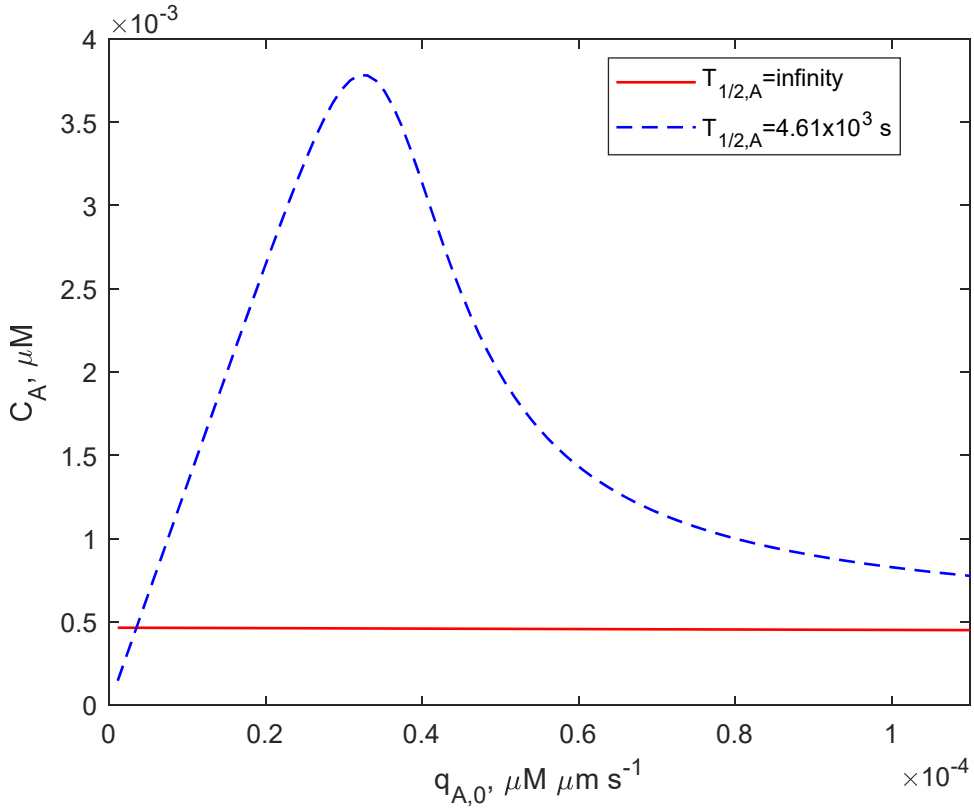

50
45
40
35
30
25
20
15
10
5
0
C_B, μM
0
20
40
60
80
100
120
140
160
180
200
D_A, μm^2/s
T_1/2,A=infinity
T_1/2,A=4.61x10^3 s

Figure 6

(a)


×10^-3
4
3.5
3
2.5
2
1.5
1
0.5
0
C_A, μM
0
0.2
0.4
0.6
0.8
1
×10^-4
q_A,0, μM μm s^-1
T_1/2,A=infinity
T_1/2,A=4.61x10^3 s

(b)

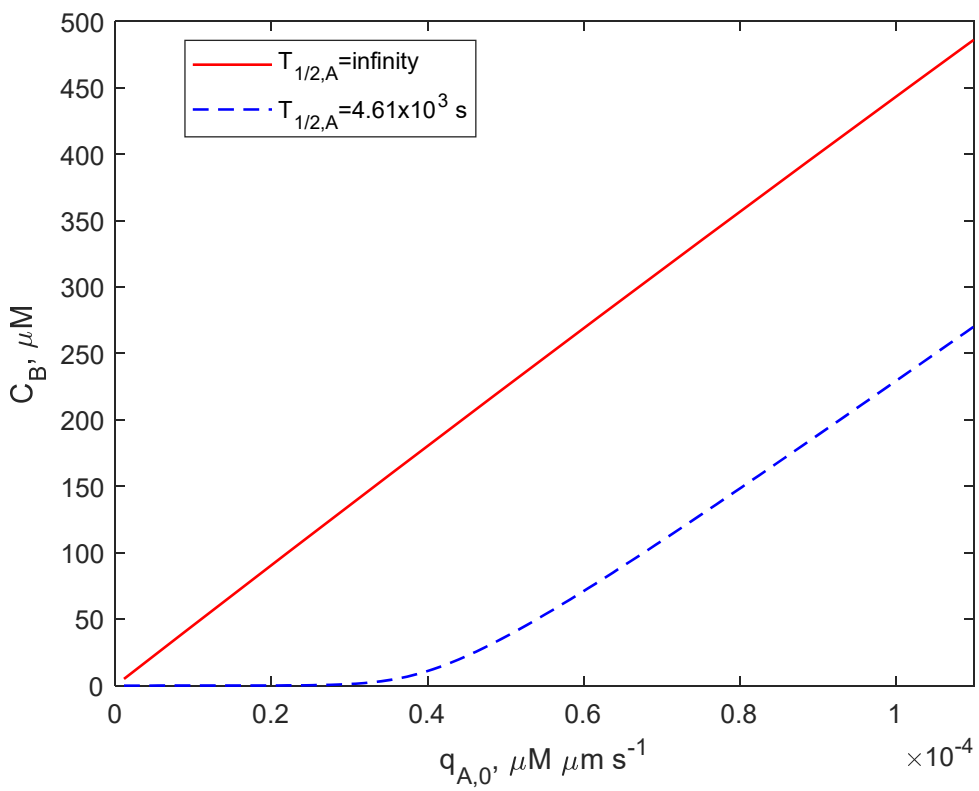
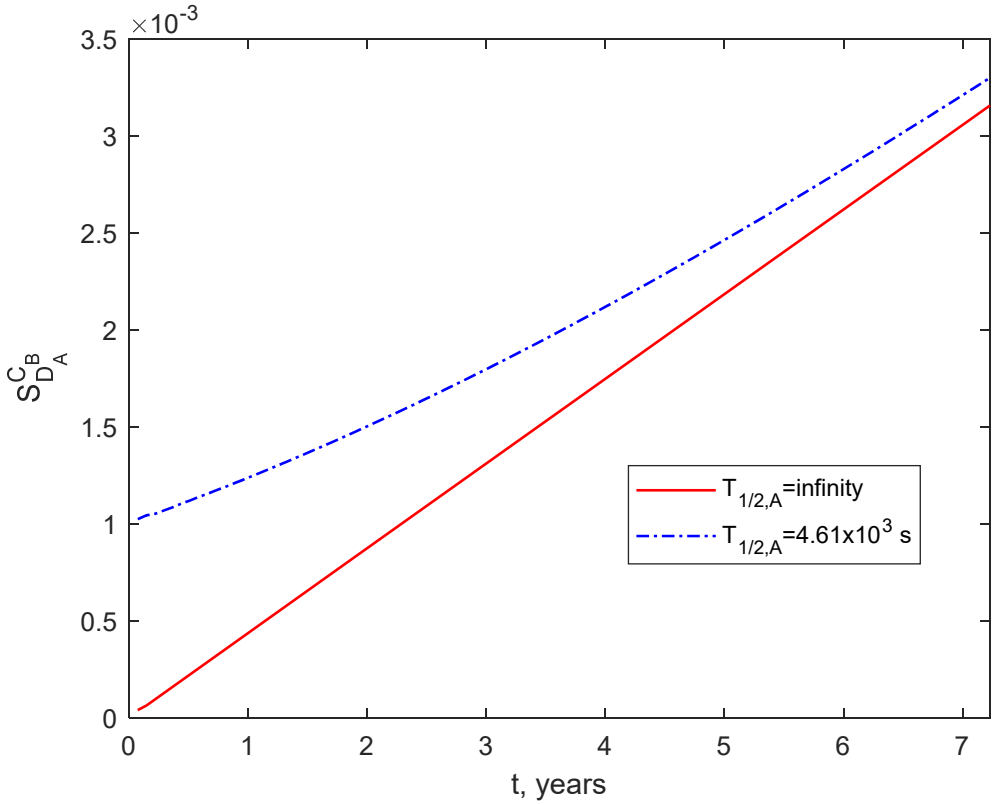

500
450
400
350
300
250
200
150
100
50
0
C_B, μM
0
0.2
0.4
0.6
0.8
1
×10^-4
q_A,0, μM μm s^-1
T_1/2,A=infinity
T_1/2,A=4.61x10^3 s

Figure 7

(a)


×10^-3
3.5
3
2.5
2
1.5
1
0.5
0
0
1
2
3
4
5
6
7
t, years
$S^{C_B}_{D_A}$
T 1/2,A=infinity
T 1/2,A=4.61x10^3 s

(b)

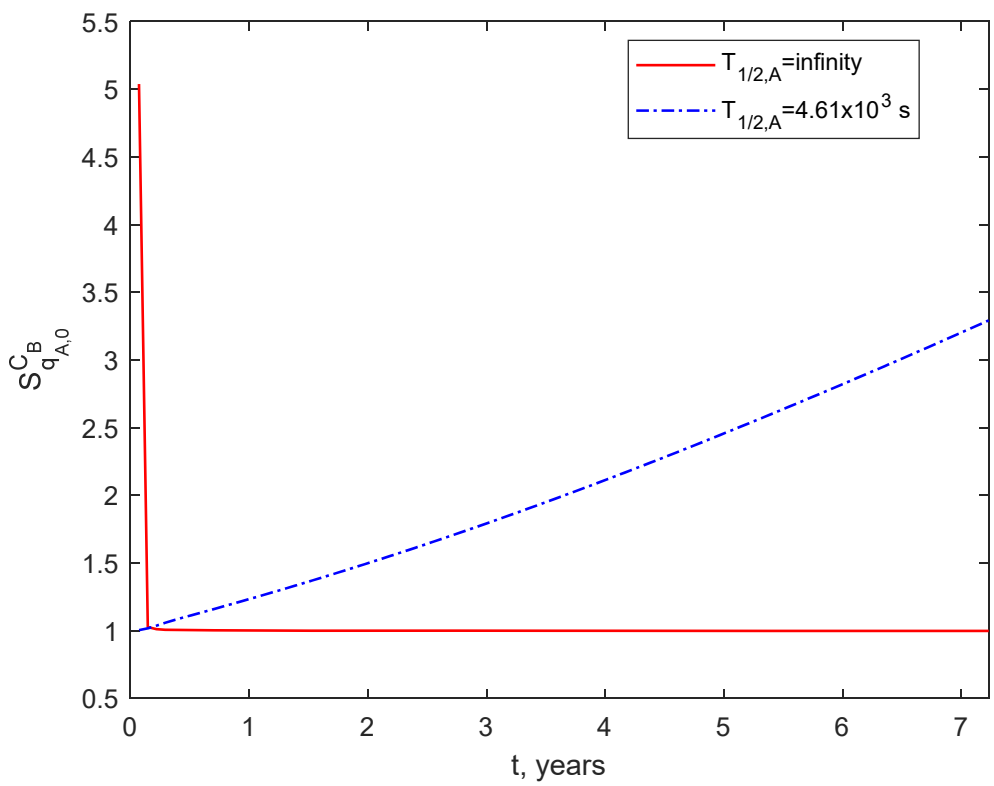

5.5
5
4.5
4
3.5
3
2.5
2
1.5
1
0.5
0
1
2
3
4
5
6
7
t, years
$S^{C_B}_{q_{A,0}}$
T 1/2,A=infinity
T 1/2,A=4.61x10^3 s

Figure 8

Effect of diffusivity of amyloid beta monomers on the formation of senile plaques

Andrey V Kuznetsov

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Raleigh, NC 27695-7910, USA; e-mail: avkuznet@ncsu.edu

Supplemental Materials

S1. Dimensionless equations

S1.1. Dimensionless form of Eqs. (3)-(7)

Eqs. (1)-(7) are converted into their dimensionless form as follows:

$$\frac{\partial C_A^*}{\partial t^*} = D_A^* \frac{\partial^2 C_A^*}{\partial x^{*2}} - C_A^* - C_A^* C_B^* - C_A^* \frac{\ln(2)}{T_{1/2,A}^*}, \tag{S1}$$

$$\frac{\partial C_B^*}{\partial t^*} = C_A^* + C_A^* C_B^*, \tag{S2}$$

$$\text{At } t^* = 0: \; C_A^* = 0, \; C_B^* = 0, \tag{S3}$$

$$\text{At } x^* = 0: \; -D_A^* \frac{\partial C_A^*}{\partial x^*} = q_{A,0}^*, \tag{S4}$$

$$\text{At } x^* = 1: \; \frac{\partial C_A^*}{\partial x^*} = 0. \tag{S5}$$

S1.2. Dimensionless form of Eqs. (8) and (9)

The dimensionless form of Eqs. (8) and (9) is as follows:

$$\frac{dC_A^*}{dt^*} = -C_A^* - C_A^* C_B^* - C_A^* \frac{\ln(2)}{T_{1/2,A}^*} + q_{A,0}^*, \tag{S6}$$

$$\frac{dC_B^*}{dt^*} = C_A^* + C_A^* C_B^*. \tag{S7}$$

Eqs. (S6) and (S7) should be solved with the initial conditions provided in Eq. (S3).

S1.3. Dimensionless form of Eqs. (16) and (17)

The dimensionless form of Eqs. (16) and (17) is as follows:

$$C_A^* = \frac{q_{A,0}^*}{1 + q_{A,0}^* t^*}, \tag{S8}$$

$$C_B^* = q_{A,0}^* t^* . \tag{S9}$$

S2. Tables

Table S1. Dimensionless independent variables used in the model.

Symbol	Definition
t^*	$t k_1$
x^*	x / L

Table S2. Dimensionless dependent variables used in the model.

Symbol	Definition
C_A^*	$C_A k_2 / k_1$
C_B^*	$C_B k_2 / k_1$

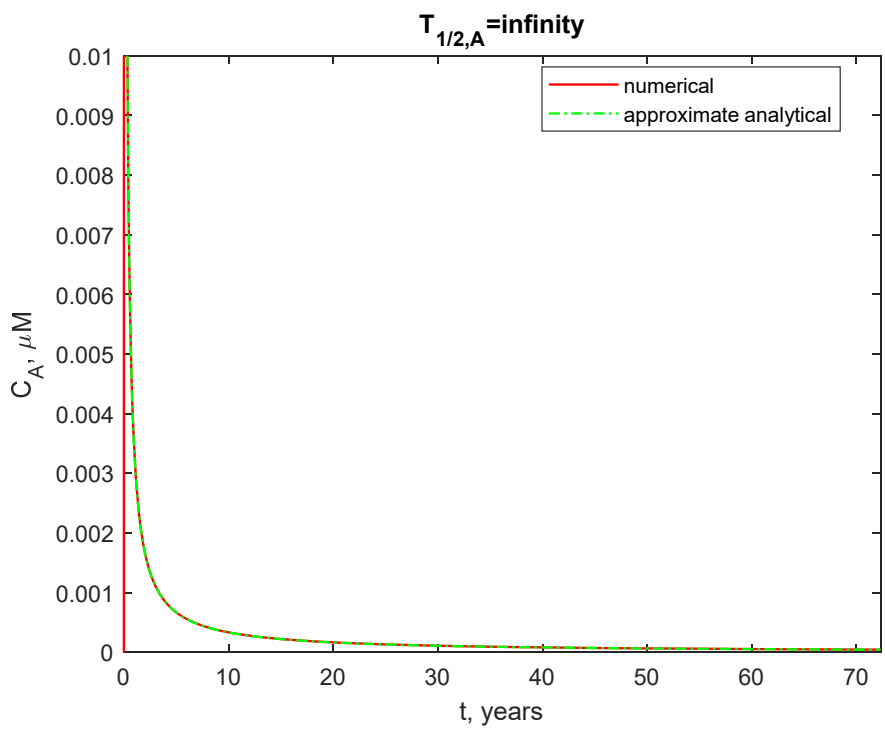
Table S3. Dimensionless parameters used in the model.

Symbol	Definition	Value(s) obtained using the dimensional parameter values provided in the last column of Table 3
D_A^*	$D_A / \left(L^2 k_1\right)$	1.12×10^7

$q^*_{A,0}$	$q_{A,0}k_2 / \left(k_1^2 L\right)$	4.21×10^5
$T^*_{1/2,A}$	$T_{1/2,A}k_1$	1.02×10^{-5} , 1.02×10^{1} (to represent $T^*_{1/2,A}\to\infty$)

S3. Supplementary figures

(a)



(b)

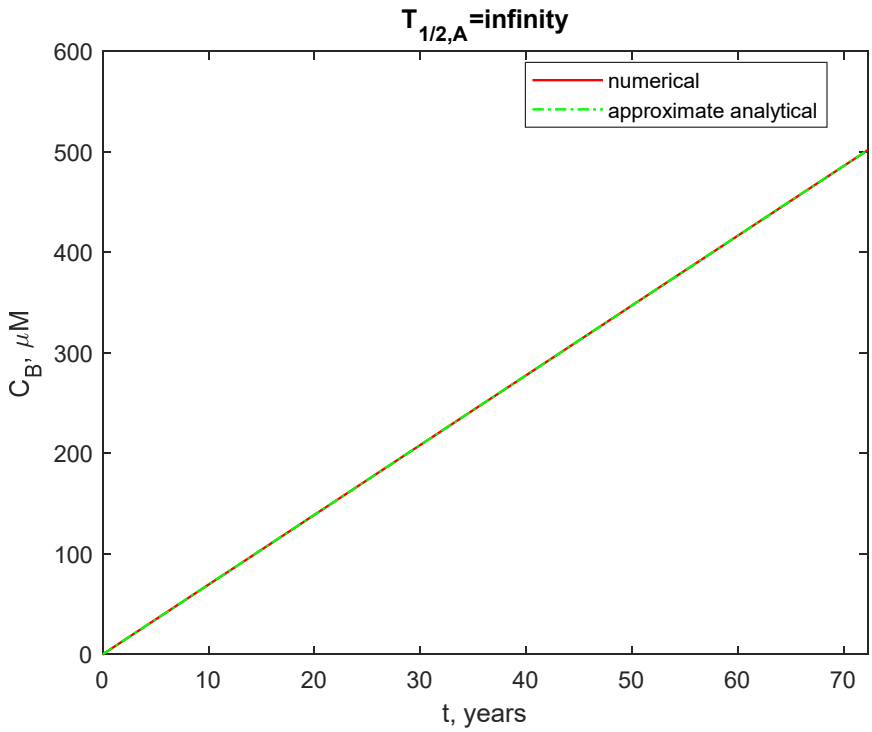
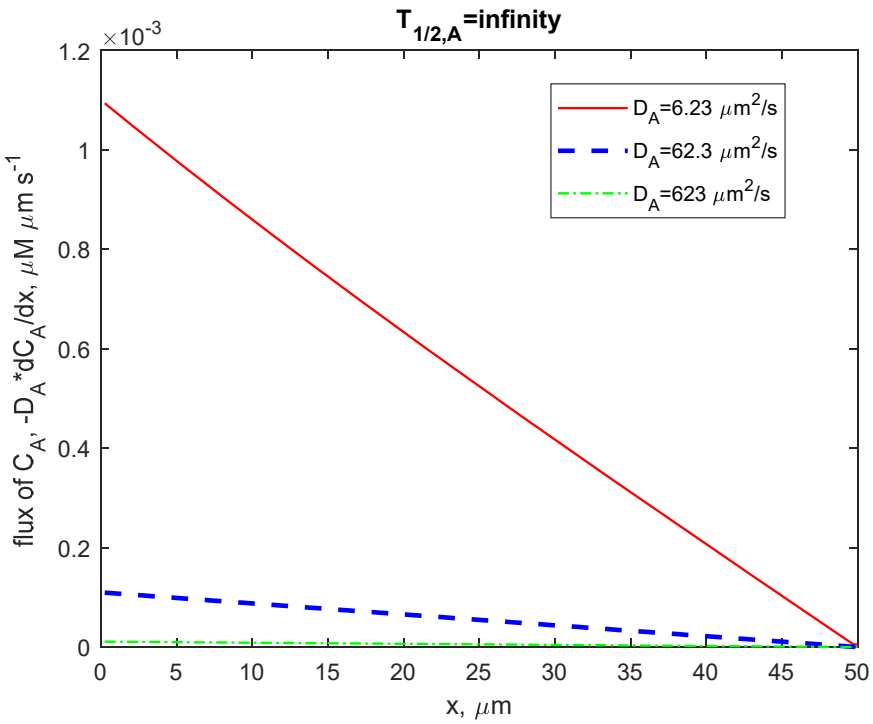


Fig. S1. A comparison between the numerical solutions of Eqs. (8) and (9) and the approximate solutions given by Eqs. (16) and (17). (a) Molar concentration of Aβ monomers, C_A, as a function of time (in years). (b) Molar concentration of Aβ aggregates, C_B, as a function of time (in years). A hypothetical scenario spanning 70 years.

(a)


T1/2,A=infinity
×10-3
1.2
1
0.8
0.6
0.4
0.2
0
0
5
10
15
20
25
30
35
40
45
50
x, μm
flux of CA, -DA*dCA/dx, μM μm s-1
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

(b)

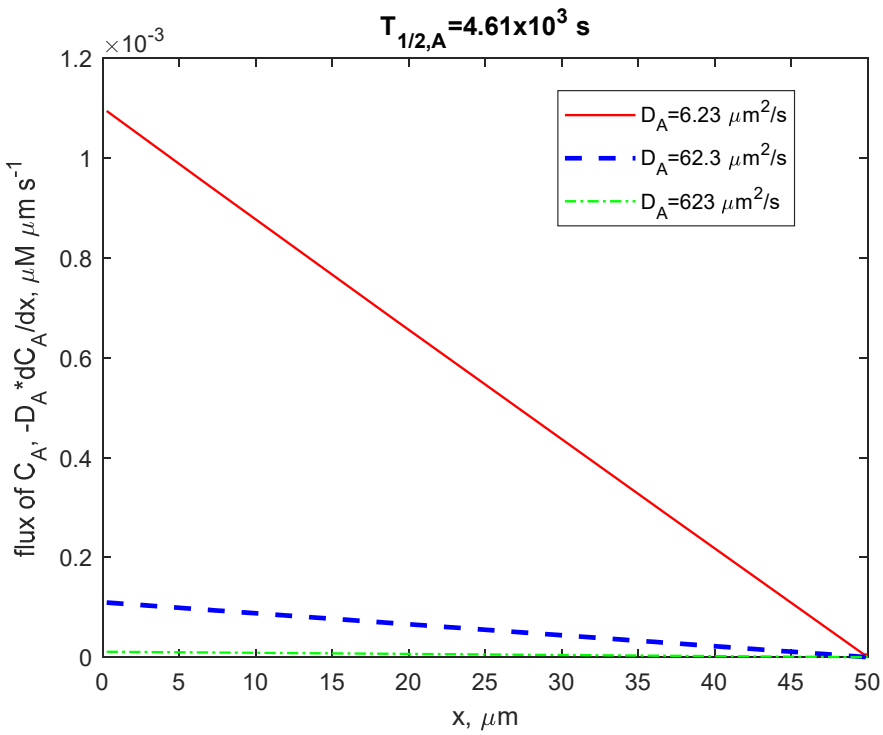
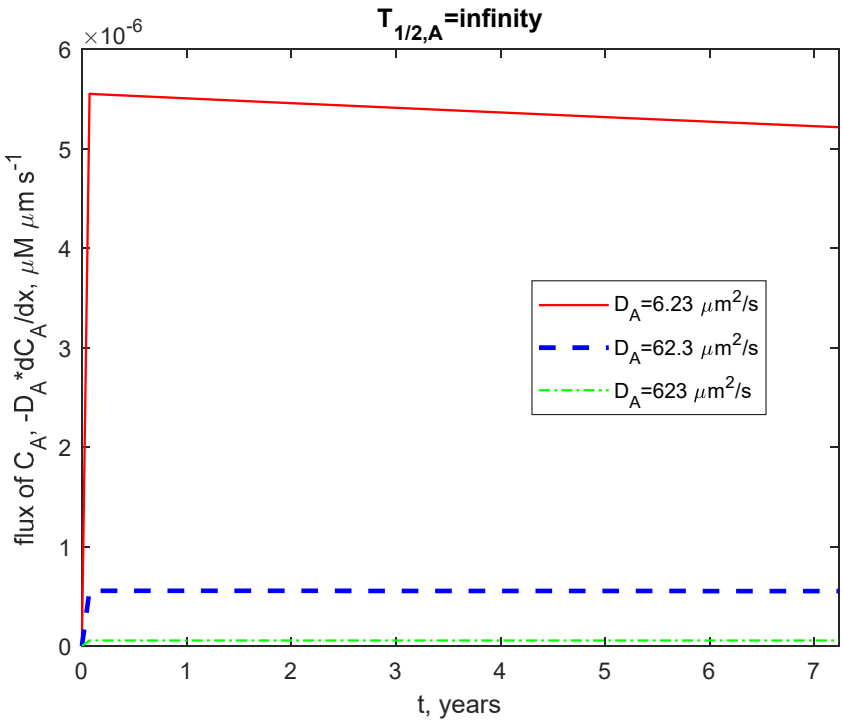

T1/2,A=4.61x103 s
×10-3
1.2
1
0.8
0.6
0.4
0.2
0
0
5
10
15
20
25
30
35
40
45
50
x, μm
flux of CA, -DA*dCA/dx, μM μm s-1
DA=6.23 μm2/s
DA=62.3 μm2/s
DA=623 μm2/s

Fig. S2. Flux of Aβ monomers as a function of the distance from the surface that releases Aβ monomers (such as the cell membrane). The two panels display the results for two different scenarios: (a) $T_{1/2,A} \to \infty$, (b) $T_{1/2,A} = 4.61 \times 10^3$ s. The computational results are presented at $t = 2.28 \times 10^8$ s $= 7.23$ years.

(a)



(b)

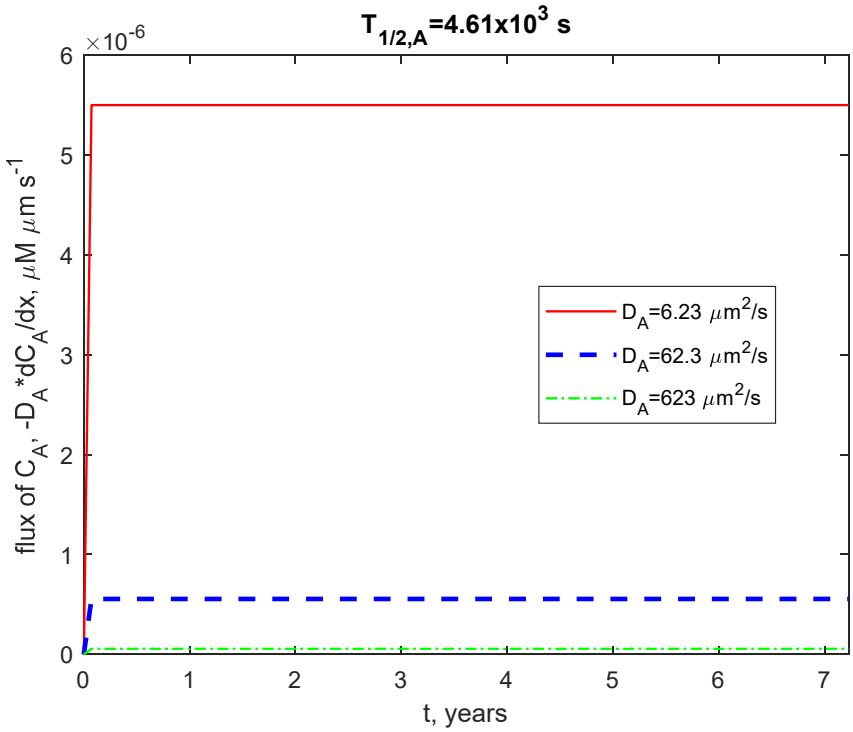


Fig. S3. Flux of Aβ monomers as a function of time (in years). The two panels display the results for two different scenarios: (a) $T_{1/2,A} \to \infty$, (b) $T_{1/2,A} = 4.61 \times 10^3$ s. The computational results are presented at the right-hand side boundary of the CV (at x=L).