Insect population dynamics under *Wolbachia*-induced cytoplasmic incompatibility: puzzle more than buzz in *Drosophila suzukii*

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Abstract

In theory, the introduction of individuals infected with an incompatible strain of *Wolbachia pipientis* into a recipient host population should result in the symbiont invasion and reproductive failures caused by cytoplasmic incompatibility (CI). Modelling studies combining *Wolbachia* invasion and host population dynamics show that these two processes could interact to cause a transient population decline and, in some conditions, extinction. However, these effects could be sensitive to density dependence, with the Allee effect increasing the probability of extinction, and competition reducing the demographic impact of CI. We tested these predictions with laboratory experiments in the fruit fly *Drosophila suzukii* and the transinfected *Wolbachia* strain *w*Tei. Surprisingly, the introduction of *w*Tei into *D. suzukii* populations at carrying capacity did not result in the expected *w*Tei invasion and transient population decline. In parallel, we found no Allee effect but strong negative density dependence. From these results, we propose that competition interacts in an antagonistic way with *Wolbachia*-induced cytoplasmic incompatibility on insect population dynamics. If future models and data support this hypothesis, pest management strategies using *Wolbachia*-induced CI should target populations with negligible competition but a potential Allee effect, for instance at the beginning of the reproductive season.

Keywords: Density dependence, Allee effect, intraspecific competition, *Drosophila suzukii*, *Wolbachia*, cytoplasmic incompatibility, eco-evolutionary dynamics.

Introduction

Wolbachia pipientis is an endosymbiotic bacterium found in a large number of nematode and arthropod species, including insects [1,2]. The overall record of interactions between *Wolbachia* and their hosts is highly variable, ranging from parasitism to mutualism. The ability of *Wolbachia* to manipulate the reproduction of their hosts at the advantage of its own transmission is the most widespread facet of these interactions [3]. Cytoplasmic incompatibility (CI) is one such manipulation that occurs, in its simplest form, when infected males cross with uninfected females. *Wolbachia*-induced sperm modification in males cannot be restored by females. In diploid species, these crosses result in embryo death while others produce viable offspring [4]. Hence, infected females

have more offspring on average than uninfected females because they can produce offspring with all types of

males, be they infected or not. Since *Wolbachia* is transmitted maternally and vertically, these females are more

likely to transmit the incompatible strain, thus promoting its spread [5].

The high invasion success of *Wolbachia* combined with its ability to reduce the transmission of viruses or other

pathogens [6,7] has aroused the interest of epidemiologists. Disease-vector control strategies have been

developed based on the introduction of individuals artificially infected with a virus-blocking incompatible strain of

Wolbachia into a target population. The reproductive advantage of infected females favors the invasion of the

virus-blocking strain, which increases in frequency until it becomes the majority or fixed strain. In the event of a

resident strain not inducing CI is present in the population, it would be replaced by the introduced incompatible

strain, which results in a host population infected with a *Wolbachia* strain with traits of interest for pest control.

For instance, such a replacement strategy has been shown to be effective in suppressing dengue transmission by

the mosquito *Aedes aegypti* [8].

In order to optimize the success of such *Wolbachia* replacement programs, it is necessary to identify the factors

that influence *Wolbachia* invasion dynamics*.* Historically, most theoretical models of CI dynamics are analogous

to population genetic models and have focused on the evolution of the frequencies of an incompatible strain

within the host population, neglecting the specificities of host population dynamics [9–13]. Only a limited number

of host demographic parameters directly affecting changes in frequencies were considered and assumed to be

constant [8,13,14]. These models predict the existence of a critical infection frequency, referred to as the invasion

threshold, above which an incompatible strain will deterministically invade the host population. The threshold

depends on three parameters: the intensity of infection-induced CI, the success of maternal transmission and the

costs resulting from infection, such as reduced fertility. More recently, Hancock *et al.* [15–17] added density

dependence on host demographic parameters in order to study *Wolbachia* invasion dynamics with more realistic

assumptions. They concluded from different models that the intensity of competition, by varying the reproductive

success of individuals, significantly reduces the ability of *Wolbachia* to invade a population [16,17].

The ability of *Wolbachia* to invade may serve another pest management strategy, alternative to the drive of

interesting characters. In a landmark publication, Dobson *et al.* [18] have proposed a novel CI-based insect pest

suppression strategy based on *Wolbachia* invasion. They extended classical CI models by assuming realistic host

population dynamics with density dependent survival of immature stages and parameter values taken from the

insect literature. They simulated the introduction of an incompatible *Wolbachia* strain into a host population that

had reached carrying capacity. While the incompatible strain spreads through the population, the frequency of

incompatible crosses increased, resulting in a transient reduction in host population size. Population size declined

to a minimum when the proportions of infected and uninfected individuals were similar and then increased again

when the incompatible strain became the majority. One of their predictions is that the minimum population size

reached during invasion is influenced not only by *Wolbachia*'s own invasion parameters, but also by demographic

parameters intrinsic to host populations. For instance, under conditions of high host reproductive rate and

intraspecific competition, the demographic consequences of CI may go unnoticed.

The transient population decline caused by *Wolbachia*-induced cytoplasmic incompatibility, as predicted by

Dobson *et al.* [18], is a form of demographic destabilization that, in conjunction with other processes, could

cascade towards population extinction. As other process, the Allee effect is an excellent candidate. It is

characterized by population growth rate decreasing with the decrease of the population size, which triggers an

extinction vortex if the growth rate becomes negative below a critical number of individuals (the Allee threshold)

or if the growth rate remains positive but combines with stochasticity. The Allee effect has thus been envisioned

as a catalyst for extinction upon which novel tactics could be developed to eradicate pest populations [19–23].

Building on these ideas, Blackwood *et al.* [24] assumed an Allee effect to investigate the conditions under which

CI expression would serve pest eradication. To this end, they simulated *Wolbachia* introductions into populations

affected by a more or less intense Allee effect. When the Allee effect was strong and the population reproduction

rate low, the model predicted that the minimum population size reached during *Wolbachia* invasion would fall

below the Allee threshold, leading to population extinction. On the other hand, at high reproduction rates, the

transient reduction induced by CI was not large enough to bring the population size below the Allee threshold.

The theoretical models of Dobson *et al.* [18] and Blackwood *et al.* [24] are of major interest for insect pest

management but, to our knowledge, they have not been tested yet. We are unaware of any empirical evidence of

either transient population declines caused by the invasion of an incompatible *Wolbachia* strain and subsequent

increase in CI or population extinctions if CI combines with an Allee effect. We therefore initiated a research

program using these models as a general theoretical framework and *Drosophila suzukii* as a model organism.

There are several reasons for choosing *Drosophila suzukii* as a model organism. First, *D. suzukii* is a major invasive

pest of berry and stone fruit crops [25]. Native from Southeast Asia, *D. suzukii* was first reported in the USA and

Europe in the late 2000s and continues to spread worldwide (*e.g.* Africa; [26–28]). Unlike other *Drosophila* species,

the serrated ovipositor of *D. suzukii* allows females to lay eggs under the skin of ripening or mature fruits, causing

significant damage to cultivated fruits and consequently, huge economic losses [25]. This calls for research to

develop environmentally friendly methods to control this major agricultural pest. Second, *D. suzukii* is naturally

infected with *Wolbachia*. Contrary to the common observation of "most-or-few" infection frequencies, the

prevalence of the resident *w*Suz strain in European populations is highly variable, averaging 46% [29]. This

infection polymorphism could be a consequence to the fact that *w*Suz induced no or weak CI [29,30] as well as the

variable benefits and costs of *w*Suz protection from viruses [31]. Third, we capitalized on a strong body of

experimental work based on the transinfection of *D. suzukii* with various *Wolbachia* strains, including *w*Ha and

*w*Tei, which cause cytoplasmic incompatibility [32–34]. Finally, the expression of negative density dependence on

individual performance has already been observed in *D. suzukii* [35]. Under experimental conditions with high

larval densities, some individuals were observed to leave the breeding site, which was interpreted as a response

to intraspecific competition for resources [36]. While the Allee effect has not yet been demonstrated in *D. suzukii*,

it has been documented in other closely related *Drosophila* species [37].

The aim of this study is to estimate the strength of positive and negative density dependence in *D. suzukii* and to

test the effect of *Wolbachia*-induced CI on the dynamics of experimental populations. In this article, we report the

absence of an Allee effect under our laboratory conditions. In contrast, we found strong negative density

dependence resulting in logistic population growth. Finally, we found no effect of introducing an incompatible

Wolbachia strain on *D. suzukii* population dynamics. We discuss these results in the light of the models developed

by Dobson *et al.* [18] and Blackwood *et al.* [24] and propose that density dependent competition may have much

stronger influences than expected on *Wolbachia* invasion and the demographic consequences of cytoplasmic

incompatibilities.

Materials and methods

Insect rearing

Two lines of *D. suzukii* have been maintained in the laboratory since 2018, *i.e.*, for around 24 generations before experiments. These lines were infected with either *w*Suz, the strain naturally present in *D. suzukii* [29,30,38] or the transinfected *Wolbachia* strain *w*Tei, the natural symbiont of *D. teissieri*. The latter was selected because of the incompatibility induced by transinfected *D. suzukii* males when crossed with *w*Suz-infected females [32,34]. The two lines were reared on cornmeal-based food medium [39] containing, per 1 L of water: 10 g agar, 80 g corn flour, 80 g brewer's yeast and 60 ml moldex (100 g methyl hydroxybenzoate in 1 L 70% ethanol). Rearing, as well as all experiments, occurred in a built-in climatic chamber at 23-24°C, 50-60% humidity and a photoperiod of 12 h-12 h.

Experiment 1. Estimation of population size

In order to estimate accurately the number of flies in population cages, we developed a rapid photo-based census method. Prior to experiments, we assessed the relevance of this method. For this, between 4 and 396 individuals were introduced into 59 cubic experimental cages (21 cm x 21 cm x 21 cm) made of fine-mesh fabric and a transparent plastic front panel. The cages were then photographed using a Nikon N750D camera and a Nikon lens (AF-S Micro Nikkor 60 mm f/2.8G ED). Visible Individuals were counted on a computer using the software ImageJ 1.52 [40] on three predefined areas (S1 Fig): (i) individuals appearing on the front and on the rear panels (area 1), (ii) individuals appearing on the whole surface except the left side and the cage floor (area 2), or (iii) only individuals appearing on the front panel (area 3). Areas 1 and 2 were used for the experiments 2 and 3, respectively. Area 3 was not used in the present study but it will be used in follow-up articles. The reasons for this choice are given in the Results section.

For each area, linear regressions between the numbers of individuals on pictures and the total numbers in the cage were fitted, and the coefficients of determination R^2 estimated. Kolmogorov-Smirnoff tests, from the

139 DHARMa package in R, were used to test normality of the residual distribution. These and the following statistical

140 analyses were carried out using R 4.2.2 software [41].

Experiment 2. Early dynamics of small populations

Founding populations

This experiment aimed at quantifying a demographic Allee effect, *i.e.*, positive density dependence at low

population size. To this end, we founded *D. suzukii* populations with different numbers of male-female pairs and

estimated subsequent growth rates. Founding pairs were formed with 1 male and 1 female, all virgins and less

than 24 hours old. Populations were founded with either 1, 3, 10, 30 or 100 pairs, and each of these initial

population size was replicated between 8 and 13 times (13 populations with 1 pair, 13 with 3 pairs, 10 with 10

pairs, 8 with 30 pairs, and 8 with 100 pairs, for a total of 52 populations). Replicated populations were distributed

across four blocks spaced one week apart. For each population, the pairs were introduced into a cage as described

above. Weekly, flies were provided with a new oviposition patch consisting of 100 ml (75 mm diameter) rearing

medium supplemented with a fresh raspberry. As the generation time was around two weeks in our rearing

conditions, each patch was exposed three weeks to ascertain that eggs laid in the patch yielded new adults

emerging into the cages. Immature drosophila developed by feeding on the rearing medium whereas adult flies

were supplied with water and honey three times a week. To estimate population size, each cage was photographed

once a week. Populations were followed nine weeks post foundation; generations were completely overlapping,

but if they were discrete, nine weeks would represent 4 to 5 generations of *D. suzukii*.

Fitting an exponential growth model

The early dynamics of populations founded with a small number of individuals is expected to depend on initial

population size and the strength of an Allee effect. In the absence of an Allee effect, exponential population growth

is expected. With a weak Allee effect, exponential growth is also expected, but the growth rate should be lower

for initially small populations (*e.g.*, 1 pair, 3 pairs, etc.) than for larger ones (*e.g.*, 100 pairs). With a strong Allee

effect, populations of initial size smaller than the Allee threshold should go extinct whereas populations initially

above the Allee threshold should increase exponentially, with a growth rate again depending on population size.

We tested these predictions by counting populations that went extinct and estimating the growth rate of those

that persisted. The growth rate (*r*) was estimated by fitting a simple exponential growth model such as:

$$N_t = N_0\, e^{r\,t} \tag{1}$$

to each population time series, excluding the last week at which the largest populations had stopped increasing.

In practice, we fitted weekly counts with a generalized linear model based on a Poisson distribution and a log link

function, *i.e.*:

$$\log\,(N_t) = \log\,(N_0) + r\,t \tag{2}$$

The initial number of individuals observed just after the populations had been founded was defined as an offset

(log (N_0)), that is, a parameter included in the model but not estimated. The number of individuals estimated

immediately after release (week 1) was missing for all populations from the first block. It was impossible to add

the initial number as an offset to the model. Therefore, the five populations of the first block were discarded due

to missing values. Growth curves and estimated growth rates *r* were obtained for each population. A linear

regression was then used to analyze the effect of the number of founders on growth rate.

Testing density dependence

A complementary method to test for positive density dependence is to assess the relation between the proportion

p of populations growing between *t* and *t + 1* and the population size at time *t*. Analyzing *p* rather than individual

counts circumvents the high sampling variance at low numbers and the consequent spurious estimations of growth

rates [42]. With an Allee effect, we expect *p* to increase with population size at low population size, with $p = 0.5$

being the Allee threshold. Independently of the experimental manipulation of initial population size, we associated

all possible pairs of N_t and N_{t+1} for each of the 47 populations at each weekly interval. A population was considered

stable or growing for $N_{t+1} \geq N_t$ (further referred to as a growing population given the low number of pairs, 14 of

410 pairs, with $N_{t+1} = N_t$) or decreasing otherwise. We then calculated the proportion *p* of growing populations

within 18 classes of population sizes (1-49, 50-99, etc.). This proportion was fitted with a logistic regression to test

three alternative hypotheses: (i) a first null model suggests no relation between population size and growth

$$p = \beta_0 \, , \tag{3}$$

(ii) a second standard model suggesting a linear relation

$$p = \beta_0 + \beta_1 \, N_t \, , \tag{4}$$

and (iii) a third polynomial model allowing a non-linear relationship

$$p = \beta_0 + \beta_1 \, N_t + \beta_2 \, N_t^{\,2} \tag{5}$$

The Eq (5) is well suited to describe bell-shaped curves that characterize populations with both an Allee effect and negative density dependence. The Akaike Information Criterion (AIC) was used to select the best model.

Experiment 3. *Wolbachia* invasion and consequent population dynamics

Experimental populations

The aim of this experiment was to assess changes in *D. suzukii* population dynamics caused by the introduction of individuals infected with an incompatible *Wolbachia* strain. To this end, we founded eight populations with 100 *w*Suz-infected individuals (sex ratio 1:1) aged 3 to 5 days. Each population was studied continuously across three distinct periods, for a total of 23 weeks: a first period of population growth (5 to 8 weeks); a seven-week period of dynamic equilibrium around the carrying capacity, starting upon observation of the first decrease in number; a post-introduction period (8 to 11 weeks) starting upon the introduction of infected flies. Populations were maintained as in the previous experiment, with a new reproduction patch provided weekly and exposed during three weeks, and water and honey to feed adults three times a week. Cages were again photographed every week for estimation of population sizes.

Introducing *Wolbachia* and characterizing infection status

Individuals introduced after the seven-week period of dynamic equilibrium were infected by either *w*Suz for the

four "control" populations (without cytoplasmic incompatibility) or *w*Tei for the four "CI" populations (with

cytoplasmic incompatibility). Theoretically, *Wolbachia* invasion depends on the intensity of CI, transmission

efficiency, and infection cost [9–13]. Assuming perfect transmission, an incompatible strain should invade if it is

introduced above a threshold equal to the ratio of infection cost (parameter *sf* in the models referenced above)

to incompatibility (parameter *sh*). In our system of *D. suzukii* and *w*Tei, incompatibility is incomplete, with an egg

hatch rate of about 33% after incompatible crosses (*i.e.*, *sh* = 0.67; [34]). In parallel, the analyses of traits such as

fecundity, longevity, egg hatch rate and developmental time all suggest an absence of cost of *w*Tei infection on *D.*

suzukii [32,34]. This apparent absence of infection costs (*sf* = 0), even with incomplete incompatibility, suggests a

theoretical invasion threshold of 0%. In practice, this would mean just enough individuals to compensate for the

stochastic events that are not considered in the theoretical models. A number of individuals close to 10% of the

mean number of individuals over equilibrium period should be sufficient to trigger invasion. Before introduction,

we verified the fixation of *w*Suz for all populations by genotyping 20 individuals per population (methods below).

After introduction, we monitored the evolution of *w*Tei frequencies over time in the four CI populations. For this,

we randomly sampled 20 individuals per week and stored them in 99% ethanol at - 20°C until analysis. Individual

infection status was assessed *via* HRM (High Resolution Melting), a genotyping method that allows the detection

of changes in the melting profiles of double-stranded DNA (S2 Fig, [43]). For each individual, DNA was extracted

using Macherey Nagel's NucleoSpin kit. *Wolbachia*-specific primers 81f (5'-TGGTCCAATAAGTGATGAAGAAAC-3';

[44]) and 2R (5'-CAGCAATTTCAGGATTAG-3'; [43]) were used to amplify the *wsp* ("*Wolbachia Surface Protein*")

gene. PCRs were performed in a 10 µL volume containing 500 nM primers, 1X Precision Melt Supermix (Biorad),

containing dNTPs, iTaqTM DNA polymerase, MgCl2, EvaGreen buffer, stabilizers, and 2 µL of 10-fold diluted DNA

[43]. Cycle conditions were 95°C for 120 s, then 30 s at 95°C, 30 s at 54°C and 30 s at 72°C for 40 cycles, followed

by 30 s at 95°C and 60 s at 60°C (Biorad CFX96). The temperature ramp rate was 1.6°C/s.

Estimating carrying capacity

The eight populations were studied over enough time to estimate the carrying capacity *K* in our environmental

conditions. For this, we fitted non-linear regressions to the number of adults observed each week, from foundation

to the introduction of infected flies (first and second time periods). Three candidate population growth models

were fitted: Gompertz, logistic, and Weibull. SSlogis, SSgompertz and SSweibull functions in R were used to obtain

the initial parameter values for each candidate model (so-called "selfStart" models [41,45]). All these models share

one parameter, the asymptotic population size defined as the carrying capacity. The AIC and root-mean-square

error (RMSE) were then used to select the best of the three models fitted to each population.

Testing the consequences of CI on population dynamics

Evolution of the frequency of wTei-infected individuals was analyzed using a generalized linear mixed model based

on a negative binomial distribution and a logit link function, with time as a fixed effect and population as a random

effect. We excluded the first period of population growth to focus on the influence of introducing incompatible

Wolbachia on the dynamics of populations that had reached carrying capacity. The main prediction tested here is

that the introduction of individuals infected by the incompatible wTei *Wolbachia* strain should lead to a decline in

population size, co-occurring with the invasion of wTei and subsequent increase in the proportion of incompatible

crosses. This decline should not be observed in control populations with introductions of individuals infected by

wSuz. We thus fitted the following model:

$$N_{ijt} = \beta_0 + \beta_1\, t + \beta_2\, intro + \beta_3\, t\, intro + \beta_4\, t\, intro\, strain + \varepsilon_{ijt} \qquad (6)$$

249   where N_{ijt} is the number of individuals in populations of introduction status i (before *vs* after introduction; variable

250   *intro* in the model) having undergone treatment j (introduction of wTei or wSuz; variable *strain* in the model) at

251   time t relative to the introduction (rescaled so that t = 0 upon introduction). This model allowed to test four

252   predictions (S3 Fig).

- H1: Control and CI populations were at equilibrium before introduction so that there should be no effect of
  time during the seven weeks prior to the introduction ($\beta_1 = 0$).
- H2: Introductions should result in an immediate but small increase in population size ($\beta_2 \neq 0$).
- H3: Introductions may destabilize populations from equilibrium and hence, induce a change in the effect of
  time on population size ($\beta_3 \neq 0$).
- H4: The latter change is expected to be affected by the strain, with wTei, contrary to wSuz, inducing a
  decrease in population size ($\beta_4 \neq 0$).

260   These four hypotheses were tested with likelihood ratio tests between models with and without the variable or
261   interaction concerned.

Results

Estimation of population size (experiment 1)

In order to estimate population size, we counted adult flies on pictures of three different areas of the experimental
cages. Whatever the area considered, we found a clear relationship between the number counted on pictures and
the total number of flies in the cages (Fig 1). Linear regressions were characterized by high coefficients of
determination: R^2 = 0.99 for areas 1 and 2 and R^2 = 0.86 for the last area. Kolmogorov-Smirnov tests confirmed the
normal distribution of errors, and hence, an absence of sampling bias (Fig 1; KS test: p = 0.83, 0.85 and 0.34 for
areas 1, 2 and 3 respectively). Given the high coefficients of determination obtained for areas 1 and 2 and the
small number of populations involved, these two areas were chosen to estimate the population size in experiments
2 and 3 respectively. However, should the number of experimental populations increase, we would recommend
the use of area 3, because counting higher numbers of insects in the other two areas would be too time-consuming
to characterize many populations on a weekly basis. Therefore, area 3 was not used in the present study, but it
will be used in the future.

Early dynamics of small populations (experiment 2)

None of the newly founded populations went extinct, even introducing a single pair. Population growth during the
first eight weeks was well described by an exponential model, regardless of the number of founding pairs (Fig 2).
The estimated growth rate r was influenced by initial population size (Anova, $F_{4,42}$ = 14.6, p < 0.001). Contrary to
expectation with an Allee effect, the growth rate r was highest at smallest population sizes, from 1.73 ± 0.03 for
single pairs to 1.4 ± 0.04 for 100 pairs.

The detailed analysis of week-to-week variations in population sizes confirmed the absence of Allee effect. Among

the three alternative models, the second order polynomial logistic regression best fitted the proportion of

increasing populations (AIC = 61.3, *versus* 82.9 for the null model and 63.5 for linear regression). The resulting

model (logit p = 1.814 + 2.014 10^{-3} N_t - 1.056 10^{-5} N_t^2) suggests that the proportion of populations growing between

consecutive weeks (p) peaks at a population size of about N_t = 95 individuals. This is the population size at which

the derivative of the above equation equals zero. However, the predicted value of p for this "optimal" population

size (p = 0.87) is not different from that predicted for N_t = 0, *i.e.* the intercept: p = 0.86 [0.77-0.92]. Thus, despite

a second-order logistic regression best fitting the data, there is no evidence of an Allee effect. This also appears

clearly in Fig 3.

Altogether, these results suggest an absence of Allee effect in our conditions. Rather, the decrease in population

growth with increasing population size reflects negative density dependence.

292 Reaching carrying capacity (Experiment 3)

293   The eight experimental populations studied over a longer time period were well described by Gompertz, Logistic

294   or Weibull population growth models (Table 1). Notwithstanding slightly different shapes, these three models are

295   characterized by a plateau, the carrying capacity K (Fig 4). Although the estimated values of K varied between

296   populations, they were of similar order of magnitude, between 455 and 593 (Table 1). This carrying capacity was

297   reached after five to eight weeks**.**

299 Consequences of cytoplasmic incompatibilities at carrying

300 capacity (Experiment 3)

To test the effect of introducing CI into a recipient population of *D. suzukii*, we fitted a generalized linear model to

population size, with a negative binomial distribution of errors and a log link function (Fig 5). The variance in

numbers within replicate was low (5.3 10^{-12}) so we added no random effect. Hypotheses H1 to H4 were tested *via*

likelihood ratio tests for comparisons of models with and without parameters under scrutiny.

• H1: as expected from the design of the experiment, prior to the introductions of infected flies, the

numbers of individuals varied at random around the carrying capacity (β_0 = 6.21; test for parameter β_1

= - 0.02; $LR_{1,135}$ = 0.98, p = 0.32).

• H2: as expected, introducing infected flies had an immediate, positive influence on population size (test

for parameter β_2 = 0.24; $LR_{1,135}$ = 6.13, p = 0.01).

• H3: following the immediate increase in numbers, the introduction of infected individuals had no

general influence on the variation over time (test for parameter β_3 = - 0.009; $LR_{1,135}$ = 0.18, p = 0.67).

• H4: most importantly, this absence of effect of introduction on population dynamics was consistent for

the two strains of *Wolbachia*, *i.e.*, the interaction *time* × *strain* after introduction was not significant

(test for parameter β_4 = - 0.02; $LR_{1,135}$ = 3.11, p = 0.08). A trend in the expected direction can be observed

and could be responsible for this marginally significant test, but this trend seems to be partly driven by

only few data points (populations 6 and 8; Fig 5).

Given the importance of this latter test and the low p-value, we complemented the parametric analysis with a chi-

square test to compare the proportion of weekly observations where population sizes were below and above the

carrying capacity. Before introduction, 53% (15/28) and 64% (18/28) observations were below K for control and CI

populations respectively, and these proportions did not differ (χ^2 = 0.30, p = 0.59). After introduction 38% (14/37)

observations were below K for CI populations, a proportion that, contrary to theoretical predictions, was not

higher than that observed for control populations (47%, 18/38; χ^2 = 0.36, p = 0.55).

Altogether, these results suggest that introducing the incompatible *Wolbachia* strain wTei into a population of *D.*

suzukii at carrying capacity produced an immediate and limited burst in population size but no consistent decrease

in population size over time.

Invasion dynamics of the incompatible strain (Experiment 3)

After introducing flies infected by wTei at an initial frequency of 10% of the estimated population size, the

prevalence of wTei did not change consistently over time (χ^2 = 3.55, df = 1, p = 0.06). The prevalence of wTei

reached up to 25% in two populations, which probably produced the marginally significant test, but it was below

the initial frequency introduced in the other two (Fig 6). For populations with weak evidence for invasion of wTei,

weekly variations in the frequency of infected individuals were high, which could reflect random effects such as

sampling variance, drift, or the combination of both processes.

Discussion

The endosymbiotic bacterium *Wolbachia pipientis,* widespread in insects, often causes cytoplasmic

incompatibility, a reproductive failure that occurs when males and females do not have the same infection status

[3]. This has led to novel and interesting ideas for insect pest management based on the introduction of an

incompatible *Wolbachia* strain into a target population, with the assumption that the resulting incompatibilities

will affect insect population dynamics. The reasoning is that if the incompatible strain is introduced at an initial

frequency above its invasion threshold, it will invade the population, which should then lead to an increase in the

proportion of incompatible crosses and, in turn, a decrease in the population growth rate. This basic idea has been

formalized in the theoretical models of Dobson *et al.* [18] and Blackwood *et al.* [24] which predict that the

introduction of one or more incompatible *Wolbachia* strains into a target population could lead to a transient

decrease in population size [18]. Furthermore, assuming that a pre-existing Allee effect is enhanced by

complementary tactics such as mating disruption, the unwanted population could be driven to extinction [24].

Although these models are of great interest for their efforts to link *Wolbachia* invasion to insect population

dynamics, their hypotheses and predictions have not been tested with appropriate experiments. Using *Drosophila*

suzukii as a model organism, we present here a first step in this direction.

Our main result is that introducing the incompatible *Wolbachia* strain wTei into a recipient *Drosophila suzukii*

population at carrying capacity does not result in an observable decrease in population size. This finding, with this

specific *Wolbachia* strain and insect host, contradicts theoretical predictions [18,24], and moderates the

perspective on using cytoplasmic incompatibility as a lever for insect pest management. In the remainder of the

discussion, we explore the various hypotheses that could explain this result.

The most trivial hypothesis to explain the lack of influence of *Wolbachia*-induced CI on population dynamics is that

contrary to expectations, the introduced incompatible *Wolbachia* did not invade the recipient populations and

therefore, did not produce enough incompatible crosses to produce an effect. This is a hypothesis that we cannot

firmly reject. Although the prevalence of individuals infected with *w*Tei reached up to 25% in some populations,

weekly estimates revealed important variations, with frequencies that were sometimes lower than the 10% fixed

at introduction. This result contrasts with the published data, showing that incompatible *Wolbachia* strains

introduced above their invasion threshold readily invade recipient populations. In the mosquito *Aedes aegypti*,

the introduction of 20% of *Wolbachia*-infected females (with strain *w*AlbB) into an experimental population of

uninfected individuals resulted in the fixation of the introduced strain in seven generations [46]. In another study

on the same species, 65% of *w*Mel-infected mosquitoes were introduced into two populations of uninfected

individuals, and again, the introduced strain became fixed 3 to 8 generations after introduction (30 to 80 days

respectively [47]). In the fruit fly *Drosophila simulans*, caged populations were established with a mixture of

uninfected individuals and either 35% of *w*Au-infected individuals or 4% of *w*Ri-infected individuals. A significant

increase in *w*Au and *w*RI frequencies was observed in all populations from the 10th generation onwards, except in

two populations where *w*Ri was either lost or at low frequency [48]. With a similar invasion dynamic, our 4-5

generations experiment (seven to ten weeks) should have revealed the invasion of *w*Tei more clearly than it did.

In this respect, our results are surprising.

Invasion may have failed if the number of infected drosophila introduced was too low to trigger invasion. We

showed in the methods section that the absence of infection costs (*sf* = 0) suggests a theoretical invasion threshold

close to 0% [9-13]. This means that in practice the introduction of very few individuals infected with *w*Tei should

result in the invasion of this strain. Under these conditions, the number of flies we introduced experimentally -

between 76 and 100 individuals in populations that were an order of magnitude larger - should have been high

enough to induce invasion.

The observed frequencies of *w*Tei in the experimental populations with CI can alternatively be interpreted as a

successful but slow invasion. In this context, the generational overlap inherent in our biological model could have

allowed older males, inducing lower CI, to participate in reproduction. Indeed, CI intensity decreases rapidly with

male age in several *Drosophila* species [49–51]. In *D. suzukii*, CI intensity is reduced to 27% in mass crosses between

*w*Suz-infected females and *w*Tei-infected males aged 9-10 days, compared to 53% when males are 2-3 days old

[32,34]. If older males are as competitive for access to females as younger ones, the negative relationship between

CI intensity and male age could reduce the demographic consequences of CI, particularly in species such as *D.*

suzukii, where individuals can survive for over 58 days at 23-24°C [34].

A final hypothesis, not exclusive to the previous arguments, is that the influence of negative density dependence

on host population dynamics overwhelms that of *Wolbachia*-induced CI. In our experiment, after an initial phase

of exponential growth, the populations were well described by logistic-type models characterized by a carrying

capacity estimated between 450 and 600 adults. This upper limit to population growth is probably a consequence

of competition among larvae for limited food resources. Indeed, intraspecific competition certainly occurs in

natural populations of *D. suzukii*. Our data suggest that the number of larvae developing in small fruits can be high

(for instance, we could find up to 21 individuals at emergence from raspberry or blackberry exposed 48 h in the

field; S1 Table). This suggests that larval intraspecific competition may be severe *in natura*, in particular when fruit

abundance is weak, *i.e.*, at the beginning or at the end of the season. Laboratory experiments are consistent.

According to Bezerra Da Silva *et al*. [36], the recurrent observation of *D. suzukii* pupae outside oviposition patches

is a sign of high sensitivity to larval competition. The number of eggs and larvae developing in damaged fruits

affects development time and further reduces adult size and fecundity [52,53]. In adults, increasing density results

in females laying in fewer eggs laid per fruit, a behavior interpreted as an adaptation to alleviate competition

among immature stages [54].

If larval survival depends on the intensity of competition within the fruit, a reduction in egg hatch rate resulting

from *Wolbachia*-induced CI should, at high densities, alleviate competition and therefore, cause an immediate

increase in larval survival. Reduced competition would offset incompatibility, and if this were true, CI would not

cause the predicted decline in population size. Such a verbal argument echoes a specific section of the theoretical

analysis of Dobson *et al.* ([18], Fig 2B), where the effect of density on immature survival is scaled by the parameter

γ [55]. Increasing the influence of density by increasing γ made the effects of introducing incompatible strains

disappear [18]. Other modelling studies have highlighted similar influence of competition among hosts on

Wolbachia invasion [15–17]. More generally, the strength of competition may alter the benefits of gene drive [56]

and of the sterile insect technique [57] with undesirable consequences such as increasing (rather than decreasing)

the size of pest populations.

The extent to which negative density dependent survival may jeopardize attempts to reduce pest reproduction

appears to be a key issue. However, empirical evidence for the critical role of density dependence in *Wolbachia*

invasion and consequences for host population dynamics is completely lacking. In *D. suzukii*, we suggest that

negative density dependence may explain the discrepancy between prior expectations and observed population

dynamics. However, further species-specific models and experiments are needed to better understand the

interplay between density dependence, the invasion of incompatible *Wolbachia* strains and the resulting

population dynamics. Follow-up experiments could manipulate intraspecific competition to understand how

competition interacts with CI. If there is an antagonistic interaction between the two processes, the population

consequences of CI should only appear when flies are released before the populations reach their carrying

capacity.

Positive density dependence, *i.e.*, the Allee effect, is also being proposed as an important process in pest

management. The basic reasoning is that eradication can be achieved *via* a combination of treatments that reduce

population size (pesticides, predators, etc.) and tactics that enhance a pre-existing Allee effect (for instance,

mating disruption). Theoretical models, verbal arguments and reviews are numerous [19,20,58,59] and, for insect

pests, well supported by widespread evidence of Allee effects [21,60]. A keystone example is that of the gypsy

moth *Lymantria dispar*, whose spread is slowed by a combination of insecticides, sterile insect technique and

mating disruption [61]. From this perspective, an important finding of our study is the absence of a demographic

Allee effect in *D. suzukii*. Despite founding populations with the smallest possible number of individuals (one male

and one female) we observed no positive relationship between population size and growth rate, and no extinction.

This finding contrasts with observations of Allee effects in closely related species such as *D. melanogaster* [37,62].

In our experimental conditions with population cages, relatively high densities may have prevented an Allee effect.

Alternatively, a component Allee effect could have occurred at the individual level, but was offset by negative

density dependence and thus did not translate to the population level.

Conclusion

In conclusion, many of our results are puzzling with respect to prior expectations. The absence of a transient decrease in *D. suzukii* population size after the introduction of the incompatible *Wolbachia* strain *w*Tei is most easily explained by the slow invasion of this introduced strain, which, in itself, lacks clear explanations. Nevertheless, there were some incompatible crosses in populations where the proportion of *w*Tei was sometimes higher than 20%, but these did not influence the host population dynamics. We therefore conclude that the strong density dependent competition could be a key driver of insect population dynamics, counterbalancing the predicted effect of cytoplasmic incompatibility, and possibly hampering pest management programs based on reproductive failure. *Wolbachia*-based methods, just like any other methods that reduce pest realized fecundity, should be implemented when populations are not under an overwhelming competition, for instance, at the beginning of the season.

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References

1.   Hilgenboecker K, Hammerstein P, Schlattmann P, Telschow A, Werren JH. How many species are infected with *Wolbachia*? – a statistical analysis of current data: *Wolbachia* infection rates. FEMS Microbiol Lett 2008; 281(2): 215-20.

2.   Weinert LA, Araujo-Jnr EV, Ahmed MZ, Welch JJ. The incidence of bacterial endosymbionts in terrestrial arthropods. Proc R Soc B. 2015; 282(1807): 20150249.

3.   Werren JH, Baldo L, Clark ME. *Wolbachia*: master manipulators of invertebrate biology. Nat Rev Microbiol. 2008; 6(10): 741-51.

453   4.   Werren JH. Biology of *Wolbachia*. Annu Rev Entomol. 1997; 42(1): 587-609.

454   5.   Turelli M, Hoffmann AA. Rapid spread of an inherited incompatibility factor in California *Drosophila*.

455   Nature. 1991; 353(6343): 440-2.

456   6.   Flórez LV, Biedermann PHW, Engl T, Kaltenpoth M. Defensive symbioses of animals with prokaryotic and

457   eukaryotic microorganisms. Nat Prod Rep. 2015; 32(7): 904-36.

458   7.   Ross PA, Turelli M, Hoffmann AA. Evolutionary ecology of *Wolbachia* releases for disease control. Annu

459   Rev Genet. 2019; 53(1): 93-116.

460   8.   Hoffmann AA, Montgomery BL, Popovici J, Iturbe-Ormaetxe I, Johnson PH, Muzzi F, *et al.* Successful

461   establishment of *Wolbachia* in *Aedes* populations to suppress dengue transmission. Nature. 2011;

462   476(7361): 454-7.

463   9.   Caspari E, Watson GS. On the evolutionary importance of cytoplasmic sterility in mosquitoes. Evolution.

464   1959; 13(4): 568-70.

465   10.   On the dynamics of symbiote-dependent cytoplasmic incompatibility in culicine mosquitoes.

466   J Invertebr Pathol. 1978; 31(1): 10-8.

467   11.   Hoffmann AA, Turelli M, Harshman LG. Factors affecting the distribution of cytoplasmic incompatibility

468   in *Drosophila simulans*. Genetics. 1990; 126(4): 933-48.

469   12.   Hurst L D. The incidences and evolution of cytoplasmic male killers. Proc R Soc Lond B. 1991; 244(1310):

470   91-9.

471   13.   Turelli M. Evolution of incompatibility-inducing microbes and their hosts. Evolution. 1994; 48(5):

472   1500-13.

473   14.   Walker T, Johnson PH, Moreira LA, Iturbe-Ormaetxe I, Frentiu FD, McMeniman CJ, *et al.* The *w*Mel

474   *Wolbachia* strain blocks dengue and invades caged *Aedes aegypti* populations. Nature. 2011; 476(7361):

475   450-3.

476   15.   Hancock PA, Sinkins SP, Godfray HCJ. Population dynamic models of the spread of *Wolbachia*. Am Nat.

477   2011; 177(3): 323-33.

478   16.   Hancock PA, White VL, Callahan AG, Godfray CHJ, Hoffmann AA, Ritchie SA. Density-dependent

479   population dynamics in *Aedes aegypti* slow the spread of *w*Mel *Wolbachia*. J Appl Ecol. 2016; 53(3):

480   785-93.

481   17.   Hancock PA, White VL, Ritchie SA, Hoffmann AA, Godfray HCJ. Predicting *Wolbachia* invasion dynamics

482   in *Aedes aegypti* populations using models of density-dependent demographic traits. BMC Biol. 2016;

483   14(1): 96.

484   18.   Dobson SL, Fox CW, Jiggins FM. The effect of *Wolbachia*-induced cytoplasmic incompatibility on host

485   population size in natural and manipulated systems. Proc R Soc Lond B. 2002; 269(1490): 437-45.

486   19.   Liebhold AM, Tobin PC. Population ecology of insect invasions and their management. Annu Rev

487   Entomol. 2008; 53(1): 387-408.

488   20.   Tobin PC, Berec L, Liebhold AM. Exploiting Allee effects for managing biological invasions. Ecol Lett. 2011;

489   14(6): 615-24.

490   21.   Fauvergue X. A review of mate-finding Allee effects in insects: from individual behavior to population

491   management. Entomol Exp Appl. 2013; 146(1): 79-92.

492   22.   Liebhold AM, Berec L, Brockerhoff EG, Epanchin-Niell RS, Hastings A, Herms DA, *et al.* Eradication of

493   invading insect populations: from concepts to applications. Annu Rev Entomol. 2016; 61(1): 335-52.

494   23.   Lampert A, Liebhold AM. Combining multiple tactics over time for cost-effective eradication of invading

495   insect populations. Ecol Lett. 2021; 24(2): 279-87.

496   24.   Blackwood JC, Vargas R, Fauvergue X. A cascade of destabilizations: combining *Wolbachia* and Allee

497   effects to eradicate insect pests. J Anim Ecol. 2018; 87(1): 59-72.

25.   Asplen MK, Anfora G, Biondi A, Choi DS, Chu D, Daane KM, *et al.* Invasion biology of spotted wing
Drosophila (*Drosophila suzukii*): a global perspective and future priorities. J Pest Sci. 2015; 88(3): 469-94.

26.   Hassani IM, Behrman EL, Prigent SR, Gidaszewski N, Ravaomanarivo LHR, Suwalski A, *et al*. First
occurrence of the pest *Drosophila suzukii* (Diptera: Drosophilidae) in the Comoros Archipelago (Western
Indian Ocean). Afr Entomol. 2020; 28(1): 78.

27.   Boughdad A, Haddi K, El Bouazzati A, Nassiri A, Tahiri A, El Anbri C, *et al.* First record of the invasive
spotted wing Drosophila infesting berry crops in Africa. J Pest Sci. 2021; 94(2): 261-71.

28.   Kwadha CA, Okwaro LA, Kleman I, Rehermann G, Revadi S, Ndlela S, *et al.* Detection of the spotted wing
drosophila, *Drosophila suzukii*, in continental sub-Saharan Africa. J Pest Sci. 2021; 94(2): 251-9.

29.   Cattel J, Kaur R, Gibert P, Martinez J, Fraimout A, Jiggins F, *et al. Wolbachia* in european populations of
the invasive pest *Drosophila suzukii*: regional variation in infection frequencies. PLoS ONE. 2016; 11(1):
e0147766.

30.   Hamm CA, Begun DJ, Vo A, Smith CCR, Saelao P, Shaver AO, *et al. Wolbachia* do not live by reproductive
manipulation alone: infection polymorphism in *Drosophila suzukii* and *D. subpulchrella*. Mol Ecol. 2014;
23(19): 4871-85.

31.   Cattel J, Martinez J, Jiggins F, Mouton L, Gibert P. *Wolbachia*-mediated protection against viruses in the
invasive pest *Drosophila suzukii*. Insect Mol Biol. 2016; 25(5): 595-603.

32.   Cattel J, Nikolouli K, Andrieux T, Martinez J, Jiggins F, Charlat S, *et al.* Back and forth *Wolbachia* transfers
reveal efficient strains to control spotted wing Drosophila populations. J Appl Ecol. 2018; 55(5): 2408-18.

33.   Nikolouli K, Colinet H, Renault D, Enriquez T, Mouton L, Gibert P, et al. Sterile insect technique and
Wolbachia symbiosis as potential tools for the control of the invasive species *Drosophila suzukii*. J Pest
Sci. 2018; 91(2): 489-503.

34.   Nikolouli K, Sassù F, Mouton L, Stauffer C, Bourtzis K. Combining sterile and incompatible insect
techniques for the population suppression of *Drosophila suzukii*. J Pest Sci. 2020; 93(2): 647-61.

35.   Hardin JA, Kraus DA, Burrack HJ. Diet quality mitigates intraspecific larval competition in *Drosophila*

523   *suzukii*. Entomol Exp Appl. 2015; 156(1): 59-65.

524   36.   Bezerra Da Silva CS, Park KR, Blood RA, Walton VM. Intraspecific competition affects the pupation

525   behavior of spotted-wing Drosophila (*Drosophila suzukii*). Sci Rep. 2019; 9(1): 7775.

526   37.   Wertheim B, Marchais J, Vet LEM, Dicke M. Allee effect in larval resource exploitation in *Drosophila* : an

527   interaction among density of adults, larvae, and micro-organisms. Ecol Entomol. 2002; 27(5): 608-17.

528   38.   Siozios S, Cestaro A, Kaur R, Pertot I, Rota-Stabelli O, Anfora G. Draft genome sequence of the *Wolbachia*

529   endosymbiont of *Drosophila suzukii*. Genome Announc. 2013; 1(1): e00032-13.

530   39.   Iacovone A, Ris N, Poirié M, Gatti JL. Time-course analysis of *Drosophila suzukii* interaction with

531   endoparasitoid wasps evidences a delayed encapsulation response compared to *D. melanogaster*. PLoS

532   ONE. 2018; 13(8): e0201573.

533   40.   Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. Nat Methods.

534   2012; 9(7): 671-5.

535   41.   R Core Team. R: A Language and Environment for Statistical Computing [Internet]. Vienna, Austria: R

536   Foundation for Statistical Computing; 2022. Available from: https://www.R-project.org/

537   42.   Tobin PC, Whitmire SL, Johnson DM, Bjørnstad ON, Liebhold AM. Invasion speed is affected by

538   geographical variation in the strength of Allee effects. Ecol Lett. 2007; 10(1): 36-43.

539   43.   Henri H, Mouton L. High-Resolution Melting technology: a new tool for studying the *Wolbachia*

540   endosymbiont diversity in the field. Mol Ecol Resour. 2012; 12(1): 75-81.

541   44.   Braig HR, Zhou W, Dobson SL, O'Neill SL. Cloning and characterization of a gene encoding the major

542   surface protein of the bacterial endosymbiont *Wolbachia pipientis*. J Bacteriol. 1998; 180(9): 2373-8.

543   45.   Archontoulis SV, Miguez FE. Nonlinear regression models and applications in agricultural research. Agron

544   J. 2015; 107(2): 786-98.

46.   Xi Z, Khoo CCH, Dobson SL. *Wolbachia* establishment and invasion in an *Aedes aegypti* laboratory
population. Science. 2005; 310(5746): 326-8.

47.   Walker T, Johnson PH, Moreira LA, Iturbe-Ormaetxe I, Frentiu FD, McMeniman CJ, *et al.* The *w*Mel strain
blocks dengue and invades caged populations. Nature. 2011; 476(7361): 450-U101.

48.   Kriesner P, Hoffmann AA. Rapid spread of a *Wolbachia* infection that does not affect host reproduction
in *Drosophila simulans* cage populations. Evolution. 2018; 72(7): 1475-87.

49.   Turelli M, Hoffmann AA. Cytoplasmic incompatibility in *Drosophila simulans*: dynamics and parameter
estimates from natural populations. Genetics. 1995; 140(4): 1319-38.

50.   Reynolds KT, Hoffmann AA. Male age, host effects and the weak expression or non-expression of
cytoplasmic incompatibility in *Drosophila* strains infected by maternally transmitted *Wolbachia*. Genet
Res. 2002; 80(2): 79-87.

51.   Shropshire JD, Hamant E, Cooper BS. Male age and *Wolbachia* dynamics: investigating how fast and why
bacterial densities and cytoplasmic incompatibility strengths vary. mBio. 2021; 12(6): e02998-21.

52.   Kienzle R, Groß LB, Caughman S, Rohlfs M. Resource use by individual *Drosophila suzukii* reveals a flexible
preference for oviposition into healthy fruits. Sci Rep. 2020; 10(1): 3132.

53.   Reyes-Ramírez A, Belgaidi Z, Gibert P, Pommier T, Siberchicot A, Mouton L, *et al*. Larval density in the
invasive *Drosophila suzukii* : Immediate and delayed effects on life-history traits. Ecol Evol. 2023; 13(8):
e10433.

54.   Elsensohn JE, Schal C, Burrack HJ. Plasticity in oviposition site selection behavior in *Drosophila suzukii*
(Diptera: Drosophilidae) in relation to adult density and host distribution and quality. J Econ Entomol.
2021; 114(4): 1517-22.

55.   Slatkin M, Smith JM. Models of coevolution. Q Rev Biol. 1979; 54(3): 233-63.

56. Dhole S, Lloyd AL, Gould F. Gene drive dynamics in natural populations: the importance of density
dependence, space, and sex. Annu Rev Ecol Evol Syst. 2020; 51(1): 505-31.

57. Yakob L, Alphey L, Bonsall MB. *Aedes aegypti* control: the concomitant role of competition, space and
transgenic technologies. J App Ecol. 2008; 45(4): 1258-65.

58. Blackwood JC, Berec L, Yamanaka T, Epanchin-Niell RS, Hastings A, Liebhold AM. Bioeconomic synergy
between tactics for insect eradication in the presence of Allee effects. Proc R Soc B. 2012; 279(1739):
2807-15.

59. Suckling DM, Tobin PC, McCullough DG, Herms DA. Combining tactics to exploit Allee effects for
eradication of alien insect populations. J Econ Entom. 2012; 105(1): 1-13.

60. Kramer AM, Dennis B, Liebhold AM, Drake JM. The evidence for Allee effects. Pop Ecol. 2009; 51(3):
341-54.

61. Lampert A, Liebhold AM. Combining multiple tactics over time for cost-effective eradication of invading
insect populations. Ecol Lett. 2021; 24(2): 279-87.

62. Elliott KH, Betini GS, Norris DR. Fear creates an Allee effect: experimental evidence from seasonal
populations. Proc R Soc B. 2017; 284(1857): 20170878.

Fig 1. Estimation of population size.

Linear regressions between the number of adult *D. suzukii* censused on pictures of three different areas of the

experimental cages and the total number of individuals in the cage.

588
589
590 **Fig 2. Early dynamics of small *D. suzukii* populations initiated with 1, 3, 10, 30 or 100 male-female pairs.**

591 For each replicated population, the eight-week time series of observed number of individuals (circles) was fitted

592 with an exponential growth model $N_t = N_0 \, e^{\,r\,t}$ (lines). Estimated marginal means of population growth rate *r* and

593 their 95% confidence intervals are shown.

595
596
597 **Fig 3. Proportion of the populations increasing (t to $t + 1$).**

598 For each range of population size at time t (N_t: 0-49, 50-99, etc.) circles represent the proportion p of pairs for

599 which the number of individuals increased ($N_{t+1} \geq N_t$). Circle size represent the number of pairs available to

600 estimate each proportion (largest circle: x pairs; smallest circle: y pairs). The curve represents the second-order

601 polynomial regression that best fitted the data (logit p = 1.814 + 2.014 10^{-3} N_t - 1.056 10^{-5} N_t^2).

603
604
605 **Fig 4. Reaching the carrying capacity.**

606 Observed and predicted time variations of *D. suzukii* population size for eight replicated populations before the

607 introduction of *Wolbachia*-infected individuals. Line types represent the logistic population growth model that

608 best fitted the data (dashed = Weibull, solid = Gompertz, dotted = logistic). Red arrows indicate the time at which

609 infected flies were introduced, *i.e.*, seven weeks after the first observed decrease in population size.

611
612
613 **Fig 5. Impacts of introduced CI-inducing flies.**

614 Observed and predicted number of adult *D. suzukii* in populations with CI (triangles; introduction of individuals

615 infected by *w*Tei) and without CI (circle; introduction of individuals infected by *w*Suz). Time is scaled so that

616 introductions occur at $t = 0$.

Fig 6. Dynamics of *Wolbachia* infection.

Evolution of the proportion of *D. suzukii* infected by the introduced *Wolbachia* strain *w*Tei. Each proportion is presented with its theoretical 95% confidence interval, based on the binomial distribution. The first bar at t = 0 corresponds to the proportion of *w*Tei-infected flies actually introduced (10%). Subsequent prevalence was estimated from the third week post-introduction, after emergence of offspring from the introduced individuals.

628 **Table 1. Gompertz, logistic and Weibull population growth models fitted to each of the eight experimental**

629 **population and values of Akaike Information Criterion (AIC), Root-Mean-Square Error (RMSE) and carrying**

630 **capacity *K*.**

Population	Model	AIC	RMSE	K estimate
	Gompertz	197.9	135.7	597.2 (452.6,741.8)
1	**Logistic**	**197.3**	**133.3**	**593.5 (466.3,720.6)**
	Weibull	_	_	_
	Gompertz	**164.2**	**162.1**	**455.7 (317.1,594.3)**
2	Logistic	_	_	_
	Weibull	_	_	_
	Gompertz	_	_	_
3	Logistic	154.7	109.4	572.7 (478.5,666.9)
	Weibull	**155.7**	**104.9**	**570.7 (475.5,665.9)**
	Gompertz	165.3	66.6	489.0 (433.7,544.3)
4	Logistic	_	_	_
	Weibull	**158.1**	**47.9**	**488.2 (449.6,526.8)**
	Gompertz	_	_	_
5	Logistic	_	_	_
	Weibull	**169.0**	**109.6**	**527.4 (436.2,618.7)**
	Gompertz	182.6	81.5	543.3 (464.9,621.7)
6	**Logistic**	**181.7**	**79.2**	**533.4 (467.4,599.4)**
	Weibull	183.4	78..2	523.8 (462.0,585.5)
	Gompertz	179.5	110.6	520.0 (400.2,639.9)
7	Logistic	178.6	106.9	510.4 (415.5,605.3)
	Weibull	**177.0**	**94.2**	**504.5(427.3,581.7)**
	Gompertz	185.2	135.7	579.3 (459.6,699.0)
8	**Logistic**	**184.7**	**133.4**	**577.6 (464.9,690.3)**
	Weibull	_	_	_

632 *K* (referred as *Asym* in the models) is the parameter common to all three models and that corresponds to the

633 asymptotic value of the population size when time tends to ∞, or the carrying capacity. Values of *K* and 95%

634 confidence intervals were estimated. Models in bold are those that best fit the observed data (lowest AIC and

635 RMSE).