Txnip deletions and missense alleles prolong the survival of cones in a retinitis pigmentosa mouse model

Yunlu Xue[1,2], Constance L. Cepko[1,3,*]

1. Departments of Genetics and Ophthalmology, Blavatnik Institute, Harvard Medical School, Boston, MA 02115
2. Present address: Lingang Laboratory, Shanghai, China, 200031
3. Howard Hughes Medical Institute, Boston, MA 02115

*Correspondence: cepko@genetics.med.harvard.edu

Abstract

Retinitis pigmentosa (RP) is a prevalent inherited retinal degenerative disease worldwide, affecting 1 in 4,000 people. The disease is characterized by an initial loss of night vision followed by a loss of daylight and color vision. Many of the RP disease genes are expressed in the rod photoreceptors, the cell type that initiates dim light vision. Following loss of rods, the cone photoreceptors, which initiate daylight vision, also are affected and can die leading to total loss of vision. The reasons for loss of cone vision are not entirely clear, but appear to be due to loss of the rods. Previously we showed that overexpressing Txnip, an α-arrestin protein, in mouse models of RP using AAV gene therapy prolonged the survival of RP cones (Xue et al. 2021). At least part of the mechanism for cone survival was a switch in the fuel source, from glucose to lactate. In addition, the mitochondria of cones were both morphologically and functionally improved by delivery of Txnip. We have gone on to test several alleles of Txnip for the ability to prolong cone survival in *rd1*, a mouse model of RP. In addition, proteins that bind to Txnip and/or have homology to Txnip were tested. Five different deletions of Txnip were expressed in cones or the retinal pigmented epithelium (RPE). Here we show that the C-terminal half of Txnip (149-397aa) is sufficient to remove GLUT1 from the RPE cell surface, and improved *rd1* cone survival when expressed specifically in the RPE. Overexpressing Arrdc4, an α-arrestin that shares 60% similar protein sequence to Txnip, reduced *rd1* cone survival. Reduction of the expression of HSP90AB1, a protein that interacts with Txnip and regulates metabolism, improved the survival of *rd1* cones alone and was additive for cone survival when combined with Txnip. However, full length Txnip with a single amino acid change, C247S, remains the most highly efficacious form of the gene for cone rescue. The above observations suggest that only a subset of the hypothesized and known activities of Txnip play a role in promoting RP cone survival, and that the activities of Txnip in the RPE differ from those in cone photoreceptors.

Introduction

Retinitis pigmentosa (RP) is an inherited retinal degenerative disease that affects one in ~4,000 people worldwide (Hartong et al. 2006). The disease first manifests as poor night vision, likely due to the fact that many RP disease genes are expressed in rod photoreceptors, which initiate night vision. Cone photoreceptors, which are required for daylight, color and high acuity vision, also are affected, so are the retina pigmented epithelial (RPE) cells (Chrenek et al. 2012; Wu et al. 2021; Napoli et al. 2021; Napoli and Strettoi 2022), which support both rod and cone photoreceptors. However, cones and RPE cells typically do not express RP disease genes. Nonetheless, RP cones lose function and die after most of the rods in their immediate neighborhood die. While it is not entirely clear what causes cone death, there are data suggesting problems with metabolism, oxidative stress, lack of trophic factors, oversupply of chromophore, and inflammation (Mohand-Said et al. 1998; Komeima et al. 2006; Punzo et al. 2009; Zhao et al. 2015; Xue et al. 2023). We have been pursuing gene therapy to address some of these problems. Our hope is to create therapies that are disease-gene agnostic by targeting common problems for cones across disease-gene families. One of our strategies is aimed at cone metabolism. Several lines of evidence suggest that RP cones do not have enough glucose, their main fuel source (Reviewed in Xue and Cepko 2023). We found that overexpression of Txnip, an α-arrestin protein with multiple functions, including glucose metabolism, prolonged the survival of cones and cone-mediated vision in three RP mouse strains (Xue et al. 2021). Regarding the mechanism of rescue, we found that it relied upon the use of lactate as a fuel source by cones. In addition, cones treated with Txnip showed improved mitochondrial morphology and function. As Txnip is known to bind directly to thioredoxin, we tested a Txnip allele with a single amino acid (aa) change, C247S, which abolishes the interaction with thioredoxin (Patwari et al. 2006). This allele provided better rescue than the wildtype (wt) Txnip allele, ruling out its interaction with thioredoxin as required for cone rescue. These findings inspired us to further modify Txnip in various ways to look for better rescue, as well as to explore potential mechanisms for Txnip's action in cones. To this end, we also tested a related α-arrestin protein, as well as an interacting partner, for rescue effects.

Results

Arrdc4 reduces *rd1* cone survival

As Txnip is a member of the α-arrestin protein family, we explored whether another family member might prolong RP cone survival. There are six known α-arrestins in mammals (Puca and Brou 2014). Among them, arrestin domain containing protein 4 (Arrdc4) is the closest to Txnip in amino acid sequence, sharing ~60% similar amino acids with Txnip (**Figure 1A**).

Arrdc4 is thought to have functions that are similar to those of Txnip in regulating glucose metabolism *in vitro* (Patwari et al. 2009; Dagdeviren et al. 2020). Like Txnip and other α-arrestins, Arrdc4 is composed of three parts: a N-terminal arrestin (Arrestin N-) domain, a C-terminal arrestin (Arrestin C-) domain, and an intrinsically disordered region (IDR) at the C-terminus. Because an IDR lacks a stable 3D structure under physiological conditions, previous studies using crystallography did not reveal the full structure of the TXNIP protein (Hwang et al. 2014). None of the other α-arrestins have been characterized structurally. To begin to examine potential similarities in structure among some of these family members, we utilized an artificial intelligence (AI) algorithm, AlphaFold-2, to visualize the predicted 3D full structure of ARRDC4 (Jumper et al. 2021). Similar to TXNIP, ARRDC4 is predicted to have a "W" shaped arrestin structure, which is composed of the Arrestin N- and C-domains, plus a long IDR which looks like a tail (**Figure 1B**).

Arrdc4 was tested for its ability to prolong cone survival in *rd1* mice using AAV-mediated gene delivery, as was done for Txnip previously (Xue et al. 2021). Expression of Arrdc4 was driven by a cone-specific promoter, RO1.7, derived from human red opsin (Wang et al. 1992; Busskamp et al. 2010; Ye et al. 2016). The vector was packaged into the AAV8 serotype capsid. AAV-Arrdc4 was injected sub-retinally into P0 *rd1* mouse eyes along with AAV-H2BGFP, which is used to trace the infection and to label the cone nuclei for counting. At P50, the treated retinas were harvested and flat-mounted for further quantification of cones within the central retina, the area that first degenerates. Unlike Txnip, the cone counts were much lower in Arrdc4 treated retina relative to the AAV-H2BGFP control (**Figure 1C&D**).

<u>Evaluation of cone survival using Txnip deletion alleles expressed in the RPE</u>

We previously showed that overexpressing the Txnip wt allele in the RPE using an RPE-specific promoter, derived from the Best1 gene (Esumi et al. 2009), did not improve RP cone survival. The wt allele removes the glucose transporter from the plasma membrane, thus preventing the RPE from taking up glucose for its own metabolism, and preventing it from serving as a conduit for glucose to flow from the blood to the cones. However, a double mutant, Txnip.C247S.LL351 and 352AA, improved cone survival when expressed only in the RPE. The C247S mutation eliminates the interaction with thioredoxin, and enhances the Txnip rescue when expressed in cones (Xue et al. 2021). The LL351 and 352AA mutations eliminate a clathrin-binding site, which reduces the ability of Txnip to remove the glucose transporter member 1 (GLUT1, encoded by *Slc2a1* gene) from the cell surface (Wu et al. 2013). Accordingly, we proposed a model in which Txnip.C247S.LL351 and 352AA promotes the use of lactate by the RPE, as we found was the case when Txnip was expressed in cones. Although the RPE normally uses lactate in wt animals, in RP, it is hypothesized that it retains the glucose that it normally would deliver to cones (Reviewed in Hurley 2021). The retention of

glucose by the RPE is thought to be due to a reduction in lactate supply, as rods normally provide lactate for the RPE, and with rod loss that source would be greatly diminished. If the RPE can utilize lactate in RP, perhaps using lactate supplied by the blood, and the LL351 and 352AA mutation impairs the ability of Txnip to remove the glucose transporter from the plasma membrane, this allele of Txnip may then allow glucose to flow from the blood to the cones via the GLUT1 transporter. The expression of Txnip.C247S.LL351 and 352AA allele thus has the potential to address the proposed glucose shortage of RP cones. However, we noted two caveats. One is that survival of cones was not as robust as when Txnip was expressed directly in cones. In addition, the *rd1* retina in the FVB strain used here, even without any treatment, shows holes in the cone layer, which appear as “craters”. A RP rat model presents a similar pattern (Ji et al. 2012, 2014; Zhu et al. 2013). When Txnip.C247S.LL351 and 352AA is expressed in the RPE, there are more craters in the photoreceptor layer. We note that these craters are common only in the *rd1* allele on the FVB background, i.e. not as common on other inbred mouse strains that also harbor the *rd1* allele, so the meaning of this observation is unclear.

Arrestins are well-known for their protein-protein interactions via different domains. Different regions of Txnip are known to physically bind to different protein partners to affect several different functions. For example, the N-terminus is sufficient to interact with KPNA2 for Txnip’s localization to the nucleus (Nishinaka et al. 2004), while the C-terminus of Txnip is critical for interactions with COPS5, to inhibit cancer cell proliferation (Jeon et al. 2005). The C-terminus of Txnip is also necessary for inhibition of glycolysis, at least *in vitro,* through an unclear mechanism (Patwari et al. 2009). Based on these studies, we made several deletion alleles of Txnip, and expressed them in the RPE. We assayed their ability to clear GLUT1 from the RPE surface (**Figure 2A**), as well as promote cone survival (**Figure 2B-G**). The 149-397aa portion of Txnip.C247S (C.Txnip.C247S) had the highest activity for GLUT1 removal from the RPE surface *in vivo*. As predicted by ColabFold, an AI algorithm based on AlphaFold-2 (Mirdita et al. 2022), the Arrestin C-domain of Txnip, which is part of C.Txnip.C247S, interacts with the intracellular C-terminal IDR of GLUT1 (**Figure 2-figure supplement 1**). These results suggest that the C-terminal portion of Txnip is sufficient to bind and clear GLUT1 from cell surface.

Cone survival was assayed *in vivo* following infection of *rd1* with these missense and deletion alleles at P0 and sacrifice at P50 (**Figure 2B-G**). Best1-Txnip.C247S, Best1-N.Txnip (1-228aa), and Best1-sC.Txnip (255-397aa, sC: short C-) did not show significant improvement in cone survival. However, Best1-C.Txnip.C247S (149-397aa), Best1-C.Txnip.C247S.LL351 and 352AA (149-397aa), and Best1-nt.Txnp.C247S^{320} (1-320aa, nt: no-tail) promoted significant cone survival compared to the corresponding control retinas. Best1-N.Txnip and Best1-sC.Txnip treated *rd1* retina did not have increased numbers of craters, while all other vectors increased the number of craters. These results

suggest that the C-terminal portion of Txnip expressed in the RPE is required for RP cone survival, for a function(s) that is unrelated to the removal of GLUT1 or to the mechanism that leads to an increase in craters.

Evaluation of Txnip deletions for autonomous cone survival

Our previous study used the human red opsin promoter, "RedO", in AAV to drive the expression of Txnip in *rd1* cones, with some low level of expression in some rods. This same strategy was used to evaluate whether the aforementioned deletion alleles of Txnip could prolong cone survival. Neither N.Txnip (1-228aa) nor C.Txnip.C247S (149-397aa) promoted significant improvement in *rd1* cone survival. However, nt.Tnxip.C247S^{301} (1-301aa) and nt.Txnip.C247S^{320} (1-320aa) promoted survival of *rd1* cones: 47% and 63% more cones than the control GFP virus, respectively (**Figure 3A&B**). In comparison, the full-length Txnip.C247S promoted an increase of 97% in cones in our previous study (Xue et al. 2021). These results show that the full-length Txnip provides the most benefit in terms of RP cone survival. To determine if expression of this allele might give increased survival when expressed in both the RPE and in cones, we used a CMV promoter to drive expression, as CMV expresses highly in both cell types (Xiong et al. 2015). CMV-Txnip.C247S provided a 38% rescue (**Figure 3A&C**), which is lower than RedO-Txnip.C247S (97%) alone. These and previous results are summarized in **Figure 4**.

Inhibiting Hsp90ab1 prolongs *rd1* cone survival

To further investigate the potential mechanism(s) of cone survival induced by Txnip, we considered the list of protein interactors that were identified in HEK293 cells using biotinylated protein interaction pull-down assay plus mass spectrometry (Forred et al. 2016). Forred et al. identified a subset of proteins that interact with Txnip.C247S, the mutant that provides better cone rescue than the wt Txnip allele (Xue et al. 2021). As we found that Txnip promotes the use of lactate in cones, and improves mitochondrial morphology and function, we looked for Txnip interactors that are relevant to mitochondria. We identified two candidates, PARP1 and HSP90AB1. PARP1 mutants have been shown to protect mitochondria under stress (Szczesny et al. 2014; Hocsak et al. 2017). Accordingly, in our previous study, we crossed the null PARP1 mice with *rd1* mice, to ask if mitochondrial improvements alone were sufficient to induce cone rescue. We found that it was not. In our current study, we thus prioritized HSP90AB1 inhibition, which had been shown to improve skeletal muscle mitochondrial metabolism in a diabetes mouse model (Jing et al. 2018).

Three shRNAs targeting different regions of the mRNA of *Hsp90ab1* (shHsp90ab1) were delivered by AAV into the retinas of wt mice. Knock-down was evaluated using an AAV encoding a FLAG-tagged Hsp90ab1 that was co-injected with the AAV-shRNA. All three shRNAs reduced the

HSP90AB1-FLAG signal compared to the shNC, the non-targeting control shRNA (**Figure 5A&B**), suggesting that they are able to inhibit the expression of HSP90AB1 protein *in vivo*. The promotion of cone survival was then tested in *rd1* mice using these shRNA constructs. The two shRNAs with the most activity in reducing the FLAG-tagged HSP90AB1 signal, shHsp90ab1$^{(\#a)}$ and shHsp90ab1$^{(\#c)}$, were found to increase the survival of *rd1* cones at P50 (**Figure 5C&D**). To determine if this effect was capable of increasing the Txnip effect, the shRNAs were co-injected with Txnip.C247S. A slight additive effect of shHsp90ab1 and Txnip.C247S was observed (**Figure 5E&F**). We also asked if there might be an effect of the knock-down of *Hsp90ab1* on a *Parp1* loss of function background. We did not observe any rescue effect of the shRNAs on this background (**Figure 5G&H**).

Discussion

In RP, the RPE cells and cones degenerate due to non-autonomous causes after the death of rods. Although the causes of cone death are not entirely clear, one model proposes that they do not have enough glucose, their main fuel source (Punzo et al. 2009; Hurley 2021; Xue and Cepko 2023). In a previous study, we found that Txnip promoted the use of lactate within cones and led to healthier mitochondria. The mechanisms for these effects are unclear, and we sought to determine what domains of Txnip might contribute to these effects, as well as explore alleles of Txnip that might be more potent for cone survival. We further tested the rescue effects of several alleles when expressed in the RPE, a support layer for cones, through which nutrients, such as glucose, flow to the cones from the choriocapillaris. The results suggest that Txnip has different mechanisms for Txnip-mediated cone survival when expressed in the RPE versus in cones.

The C-terminal portion of Txnip.C247S (149-397aa) expressed within the RPE, but not within cones, delayed the degeneration of cones (**Figure 2**). The full length Txnip.C247S expressed within cones, but not within the RPE, was the most effective configuration for cone survival (**Figure 3**). The expression of full length Txnip.C247S in both the RPE and cones did not provide better rescue than in cones alone. As Txnip has several domains, that presumably interact with different partners, it is possible that these different effects on cone survival are due to the interaction of different Txnip domains with different partners in the RPE versus the cones, or different results from the interactions of the same domains and partners in the two cell types. The N-terminal half of Txnip (1-228aa) might exert harmful effects in the RPE, that negate the beneficial effects from the C-terminal half. This hypothesis is supported by the observation that N.Txnip led to an obvious thinning of the outer nuclear layer of the wt retina, reflecting a loss of photoreceptors (**Figure 2A**). A negative effect of the N-terminal half of Txnip in the RPE is further supported by the observation that its removal, in the C-

terminal 149-397 allele, led to better cone survival when expressed in the RPE. In cones, the C-terminal half, including the C-terminal IDR tail, may cooperate with the N-terminal half, or negate its negative effects, to benefit RP cone survival. However, the C-terminal half is not sufficient for cone rescue when expressed in cones, as the 149-397 allele did not rescue.

The C-terminal half of Txnip apparently affects cone survival differently when expressed within the two cell types. This notion is informed by the different rescue effects of expression of the 149-397 allele, which rescues cones when expressed in the RPE, but not when expressed in cones. This domain loses the cone rescue activity if it loses aa 149-254, when expressed in the RPE, as shown by the 255-397 allele. In cones, the rescue activity is present in the 1-301 and the 1-320 allele, but is lost in the 149-397 allele. It is possible that effects on protein structure cause this loss, or that an interaction between N-terminal and C-terminal domains are required for cone rescue within cones.

One Txnip function that likely is important to these effects in the two cell types is Txnip's removal of the glucose transporter from the plasma membrane. The LLAA Txnip mutant is unable to effectively remove the transporter, due to its loss of interaction with clathrin. When this mutant allele is expressed in the RPE, it leads to improved cone survival, in contrast to the wt allele. This might be due to better health in the RPE, when it is able to take up glucose to fuel its own metabolism, and/or to provide glucose to cones. When the LLAA allele is expressed in cones, it also promotes cone survival, though not as well as the wt allele (Xue et al. 2021). The wt allele might be more beneficial in cones if it is part of the mechanism that forces cones to rely more heavily on lactate vs. glucose. All of these observations of cone rescue from expression within cones suggest that cone rescue relies on activities that reside in both the N and C-terminal portions, including the ability of Txnip to interact with clathrin. However, it will be important to probe structural alterations and stability of Txnip in cones and RPE when these various alleles are expressed to further support these hypotheses.

Regarding potential protein interactions beyond the glucose transporter, the interaction of Txnip with thioredoxin is apparently negative, as we found in our previous study with the C247S allele. This is most easily understood by the release of thioredoxin from Txnip, whereupon it can play its anti-oxidation role, which would be important in the RP retina which exhibits oxidative damage. It also would free Txnip to interact with other partners. Another partner interaction suggested by previous studies and explored here is the interaction with HSP90AB1 (**Figure 5**). Knocking down Hsp90ab1 improved mitochondrial metabolism of skeletal muscle in a diabetic mouse model (Jing et al. 2018). Here we found that it enhanced cone survival in *rd1* mice. This rescue seems to be dependent on PARP1, another binding partner of Txnip. As shown by PARP1 knock out mice, PARP1 is deleterious to mitochondrial heath under stressful conditions (Szczesny et al. 2014; Hocsak et al. 2017; Xue et al. 2021). When we examined a possible rescue effect of PARP1 loss on *rd1* cone survival, we did not see a benefit, indicating that the Txnip-mediated rescue is not due solely to its beneficial effects on

mitochondria, nor does Txnip-mediated rescue rely upon PARP1 (Xue et al. 2021). These results indicate that the Txnip rescue is more complex than inhibition of HSP90AB1, and a PARP1-independent mechanism is involved. It is possible that HSP90AB1 directly interacts with PARP1, and this interaction is critical for shHsp90ab1 to benefit RP cones. We looked into the predicted 3D structures of HSP90AB1 and PARP1 using AlphaFold2 (**Figure 5-figure supplement 1**), but did not gain additional insight into such interactions. Despite the unclear structural interactions, combining Hsp90ab1 inhibition with Txnip.C247S could be a potential combination therapy to maximize the protection of RP cones.

Material and Methods

Key Resources Table				
Reagent type (species) or resource	**Designation**	**Source or reference**	**Identifiers**	**Additional information**
Antibody	Rabbit anti-GLUT1	Abcam	ab116730	IHC (1:500)
Genetic reagent (M. musculus)	Arrdc4 cDNA	GeneCopoeia	Cat. #: Mm26972 NCBI: NM_001042592.2	
Genetic reagent (M. musculus)	Hsp90ab1 cDNA	GeneCopoeia	Cat. #: Mm03161 NCBI: NM_008302.3	
software, algorithm	Protein 3D structure prediction	AlphaFold-2	TXNIP (M. musculus); ARRDC4 (M. musculus); HSP90AB1 (M. musculus); PARP1 (M. musculus)	(Jumper et al. 2021) https://alphafold.ebi.ac.uk
software, algorithm	Protein 3D interaction prediction	ColabFold	AlphaFold2_mmseqs2	(Mirdita et al. 2022) https://github.com/sokrypton/colabfold
software, algorithm	Protein 3D structure viewer	RCSB PDB	Mol* 3D Viewer	To visualize the 3D structure of proteins in .pdb files https://www.rcsb.org/3d-view

The material and methods in this study are similar to those used in our previous study (Xue et al. 2021). The cone number of the central retina is defined as the counts of H2BGFP-positive cells within the central portion of the retina. New reagents and algorithms used in this study are listed in the Key resources table above. Txnip deletions were cloned from the Txnip plasmid using Gibson assembly (**Figure 4**). The following sense strand sequences were used to knock down the Hsp90ab1: siHsp90ab1(#a) 5′ - GCATCTACCGCATGATTAAAC-3′ ; siHsp90ab1(#b) 5′ - CCAGAAGTCCATCTACTATAT-3′ ; siHsp90ab1(#c) 5′ - CCTGAGTACCTCAACTTTATC-3′ .

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Figure 1 Effect of Arrdc4 on cone survival in retinitis pigmentosa (RP) mice.

A. Amino acid sequences of mouse TXNIP and mouse ARRDC4. In the full length alignment (421 amino acid), Identity: 172 / 421, 40.86%; Similarity: 246 / 421, 58.43%; Gaps: 28 / 421, 6.65%. Color code: identical, black; similar, blue; not similar, red.

B. Predicted 3D protein structures of mouse TXNIP and mouse ARRDC4 by artificial intelligence (AI) algorithm AlphaFold-2. Abbreviations: Arr N-, N-terminal arrestin domain; Arr C-, C-terminal arrestin domain.

C. Representative P50 *rd1* flat-mounted retinas after P0 subretinal infection with AAV8-RO1.7-Arrdc4 (1×10^9 vg/eye), plus AAV8-RedO-H2BGFP (2.5×10^8 vg/eye), or control eyes infected with AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye alone.

D. Quantification of H2BGFP-positive cones within the center of P50 *rd1* retinas transduced with Arrdc4, and control (same as in C). The number in the round brackets '()' indicates the number of retinas within each group. Error bar: standard deviation. **** p< or <<0.0001.

RedO: red opsin promoter; RO1.7: a 1.7kb version of red opsin promoter. AAV: adeno-associated virus.

Figure 2 Txnip deletions expressed only within RPE cells: effects on GLUT1 removal and cone survival.

A. Glucose transporter 1 (GLUT1) expression in P20 wildtype eyes infected with control (AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye), or a Txnip allele (2.5×10^8 vg/eye) plus RedO-H2BGFP (2.5×10^8 vg/eye), as indicated in each panel. Txnip deletions are detailed in **Figure 4**. Magenta: GLUT1; green: RedO-H2BGFP for infection tracing; gray: DAPI.

B, D, F. Representative P50 *rd1* flat-mounted retinas after P0 infection with one of seven different Txnip alleles expressed only within the RPE, as indicated in the figure, or control eyes infected with AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye alone.

C, E, G. Quantification of H2BGFP-positive cones within the center of P50 *rd1* retinas transduced with indicated vectors, as shown in B, D, F. The number in the round brackets '()' indicates the number of retinas within each group. Error bar: standard deviation.

C.Txnip.CS: C-terminal portion of Txnip.C247S; C.Txnip.CS.LLAA: C-terminal portion of Txnip.C247S.LL351 and 352AA; nt.Txnip.CS^{320}: no tail Txnip (1-320aa). NS: not significant, p>0.05, *p<0.05, **p<0.01, **** p< or <<0.0001. Best1: Best1 promoter.

Figure 2 Supplement 1 Predicted protein-protein interaction of TXNIP and GLUT1 by an algorithm based on AlphaFold-2.

Figure 3 Tests of Txnip alleles on cone survival.

A. Representative P50 *rd1* flat-mounted retinas after P0 infection with one of 5 different Txnip alleles (AAV8-RedO- N.Txnip /C.Txnip.C247S/ nt.Txnip.C247S$^{1-301}$/nt.Txnip.C247S$^{1-320}$ or AAV8-CMV-Txnip.C247S, ≈1 × 10^9 vg/eye, plus AAV8-RedO-H2BGFP, 2.5 × 10^8 vg/eye), or control eyes infected with AAV8-RedO-H2BGFP, 2.5 × 10^8 vg/eye alone.

B, C. Quantification of H2BGFP-positive cones within the center of P50 *rd1* retinas transduced with AAV8-RedO- N.Txnip /C.Txnip.C247S/ nt.Txnip.C247S$^{1-301}$/nt.Txnip.C247S$^{1-320}$ or AAV8-CMV-Txnip.C247S, and control (same as in A). The number in the round brackets '()' indicates the number of retinas within each group. Error bar: standard deviation. NS: not significant, **** p< or <<0.0001.

Figure 4 Summary of various alleles of TXNIP in this and previous study (Xue et al. 2021).
"RPE GLUT1 Removal" refers to the amount of GLUT1 immunohistochemical signal following expression in the RPE using the Best1 promoter. "Cone Rescue: Expression in RPE" refers to cone rescue following expression only in the RPE using the Best1 promoter. "Cone Rescue: Expression in Cones" is due to expression only in cone photoreceptors using the RedO promoter.
Abbreviations: Y (x%): Yes with x% increase compared to AAV-H2BGFP control; N: No; I: Impaired ability to remove GLUT1; NT: Not tested. N.Txnip, N-terminal portion of TXNIP; C.TXNIP.C247S, C-terminal portion of TXNIP.C247S mutant allele; sC.TXNIP: a shorter version of C-terminal portion of TXNIP; nt.Txnip.C247S, no tail version TXNIP.C247S mutant allele; Arrestin N-, N-terminal arrestin domain; Arrestin C-, C-terminal arrestin domain; PPxY, a motif where P is proline, x is any amino acid and Y is tyrosine.

Figure 5 Effect of knockdown of Hsp90ab1 in RP cones *in vivo*.

A. AAV8-RO1.7-Hsp90ab1-FLAG (1 × 10^9 vg/eye) co-injected with shNC (non-targeting shRNA control, AAV8-RedO-shRNA, 1 × 10^9 vg/eye) or co-injected with Hsp90ab1 shRNAs #a, #b, #c (AAV8-RedO-shRNA, 1 × 10^9 vg/eye) in P20 wt retina, all also injected with AAV8-RedO-H2BGFP (2.5 × 10^8 vg/eye) to track the infection. Magenta: anti-FLAG; green: anti-GFP; gray: DAPI. Right panel:

B. The quantification of FLAG intensity from multiple fields of inner segment regions in A. The number in the square brackets '[]' indicates the number of images taken from regions of interest of one retina, in each condition.

C. Representative P50 *rd1* flat-mounted retinas injected with shNC, shHsp90ab1(#a), shHsp90ab1(#b), or shHsp90ab1(#c) (AAV8-RedO-shRNAs, 1×10^9 vg/eye, plus AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye).

D. Quantification of H2BGFP-positive cones within the center of P50 *rd1* retinas transduced with shNC, shHsp90ab1(#a, #b, #c) (same as in C).

E. Representative P50 *rd1* flat-mounted retinas with H2BGFP (gray)-labeled cones transduced with Txnip.C247S or Txnip.C247S + shHsp90ab1 (AAV8-RedO-Txnip.C247S, 1×10^9 vg/eye; AAV8-RO1.7-shHsp90ab1(#a or #c), 1×10^9 vg/eye; plus AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye).

F. Quantification of H2BGFP-positive cones within the center of P50 *rd1* retinas transduced with Txnip.C247S or Txnip.C247S + shHsp90ab1 (same as in E).

G. Representative P50 *Parp1*$^{-/-}$ *rd1* flat-mounted retinas with H2BGFP (gray)-labeled cones transduced with shNC (non-targeting shRNA control, AAV8-RedO-shRNA, 1×10^9 vg/eye; plus AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye) or shHsp90ab1 (AAV8-RedO-shRNA #a or #c, 1×10^9 vg/eye; plus AAV8-RedO-H2BGFP, 2.5×10^8 vg/eye).

H. Quantification of H2BGFP-positive cones within the center of P50 *Parp1*$^{-/-}$ *rd1* retinas transduced with shNC or shHsp90ab1 (same as in G).

Error bar: standard deviation. NS: not significant, $p>0.05$, *$p<0.05$, **$p<0.01$, ***$p<0.001$ **** $p<$ or $<<0.0001$.

Figure 5 Supplement 1 Predicted 3D protein structures of HSP90AB1 and PARP1.

A. Predicted 3D protein structures of HSP90AB1 by AI algorithm AlphaFold-2 from two angles of view.

B. Predicted 3D protein structures of PARP1 by AI algorithm AlphaFold-2 from two angles of view.

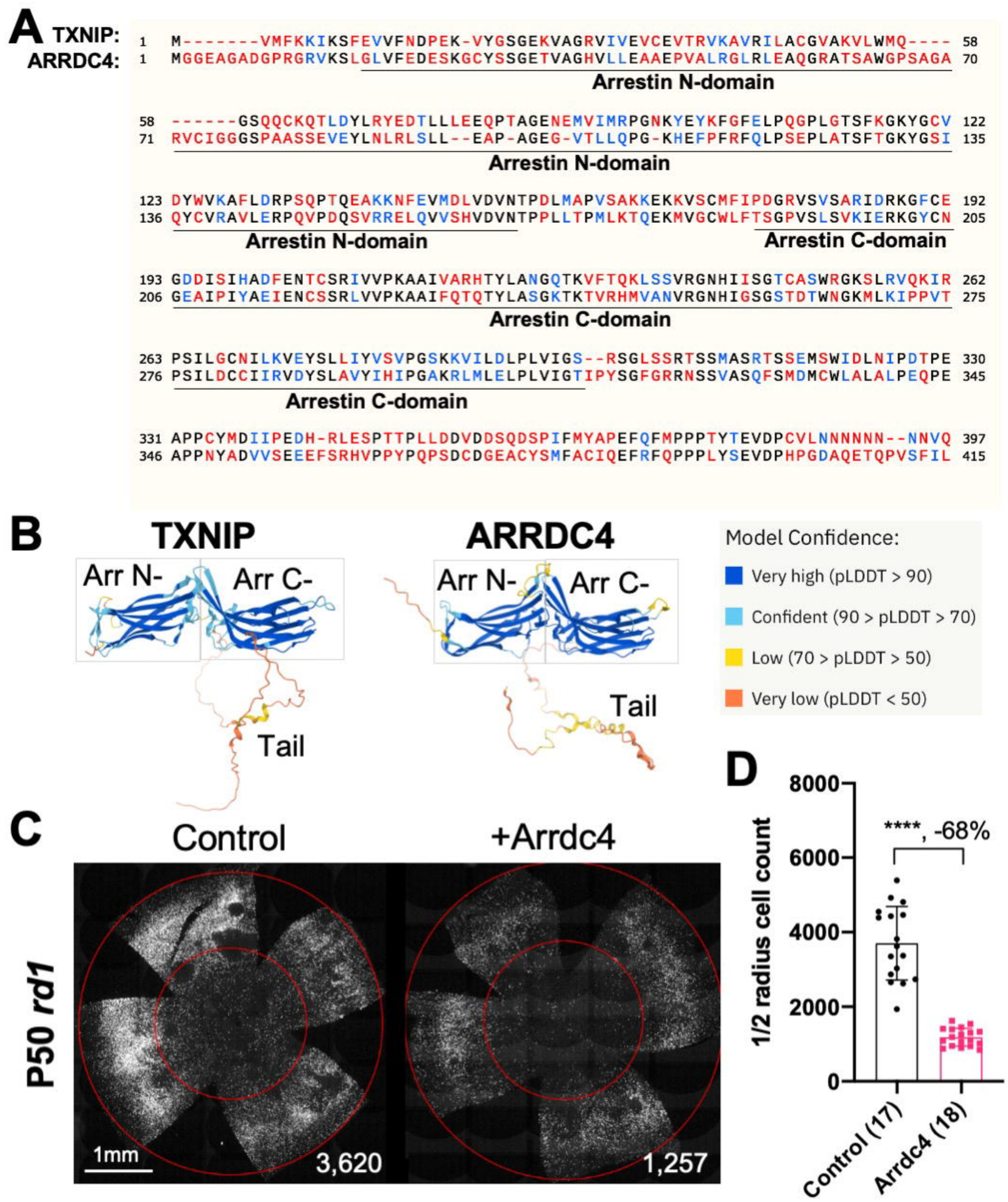
A
TXNIP:
ARRDC4:
Arrestin N-domain
Arrestin C-domain
B
TXNIP
ARRDC4
Arr N-
Arr C-
Tail
Model Confidence:
Very high (pLDDT > 90)
Confident (90 > pLDDT > 70)
Low (70 > pLDDT > 50)
Very low (pLDDT < 50)
C
Control
+Arrdc4
P50 rd1
1mm
3,620
1,257
D
1/2 radius cell count
8000
6000
4000
2000
0
****, -68%
Control (17)
Arrdc4 (18)

Figure 1

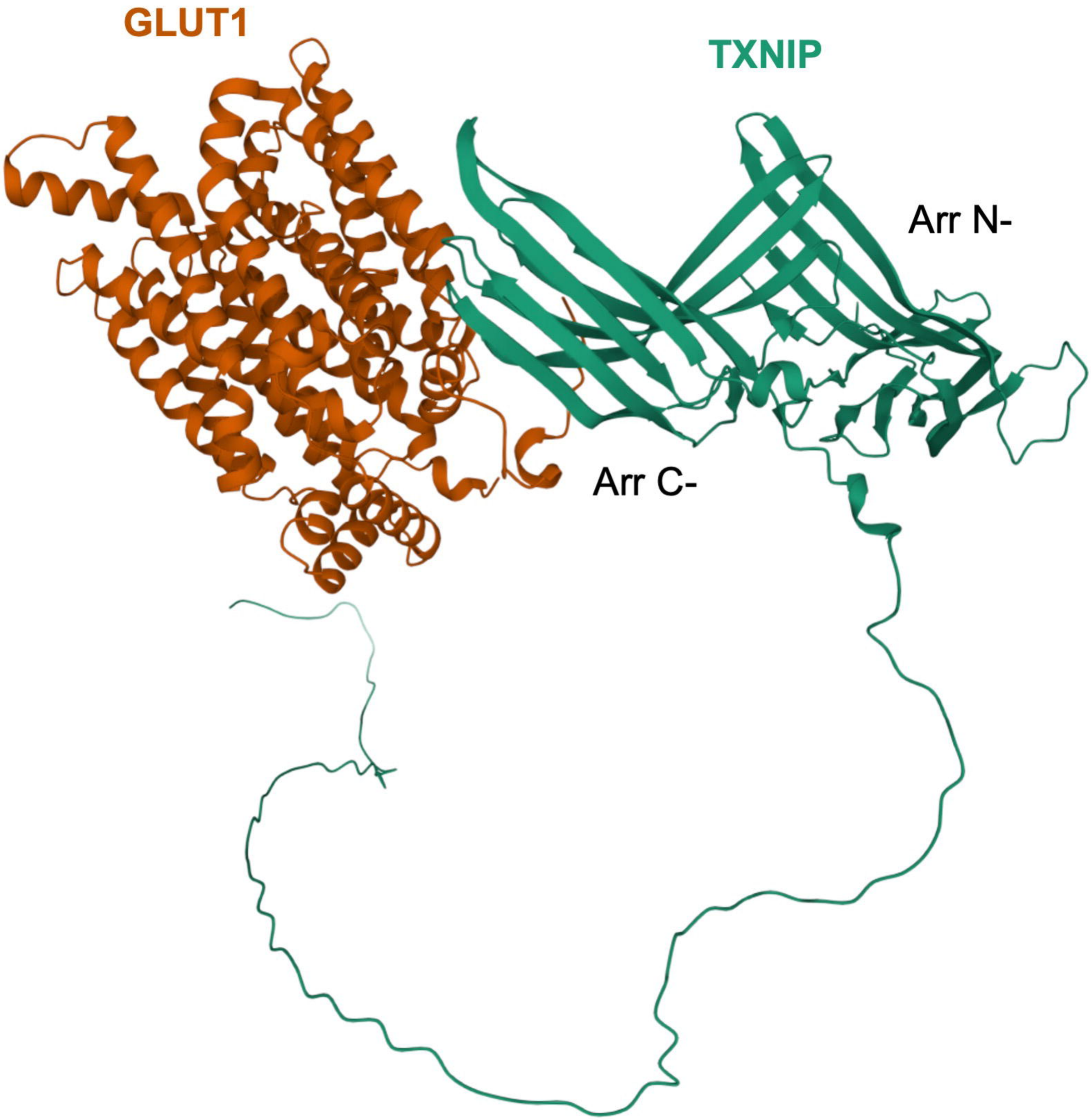

Figure 2 Supplement 1

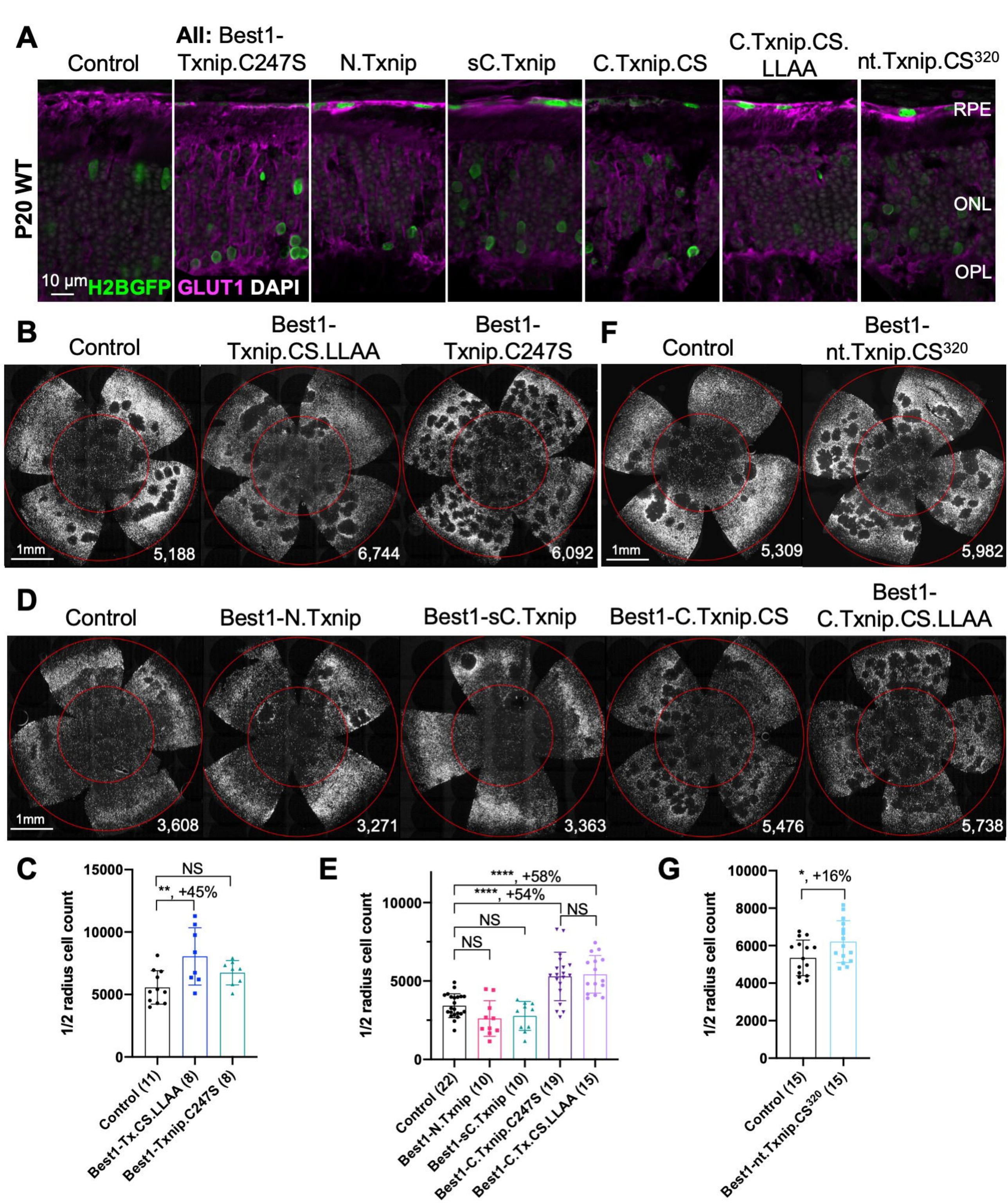
A
All: Best1-Txnip.C247S
Control
N.Txnip
sC.Txnip
C.Txnip.CS
C.Txnip.CS. LLAA
nt.Txnip.CS320
P20 WT
RPE
ONL
OPL
10 µm
H2BGFP
GLUT1
DAPI
B
Control
Best1-Txnip.CS.LLAA
Best1-Txnip.C247S
1mm
5,188
6,744
6,092
F
Control
Best1-nt.Txnip.CS320
1mm
5,309
5,982
D
Control
Best1-N.Txnip
Best1-sC.Txnip
Best1-C.Txnip.CS
Best1-C.Txnip.CS.LLAA
1mm
3,608
3,271
3,363
5,476
5,738
C
1/2 radius cell count
15000
10000
5000
0
NS
**, +45%
Control (11)
Best1-Tx.CS.LLAA (8)
Best1-Txnip.C247S (8)
E
1/2 radius cell count
10000
5000
0
****, +58%
****, +54%
NS
NS
NS
Control (22)
Best1-N.Txnip (10)
Best1-sC.Txnip (10)
Best1-C.Txnip.C247S (19)
Best1-C.Tx.CS.LLAA (15)
G
1/2 radius cell count
10000
8000
6000
4000
2000
0
*, +16%
Control (15)
Best1-nt.Txnip.CS320 (15)

Figure 2

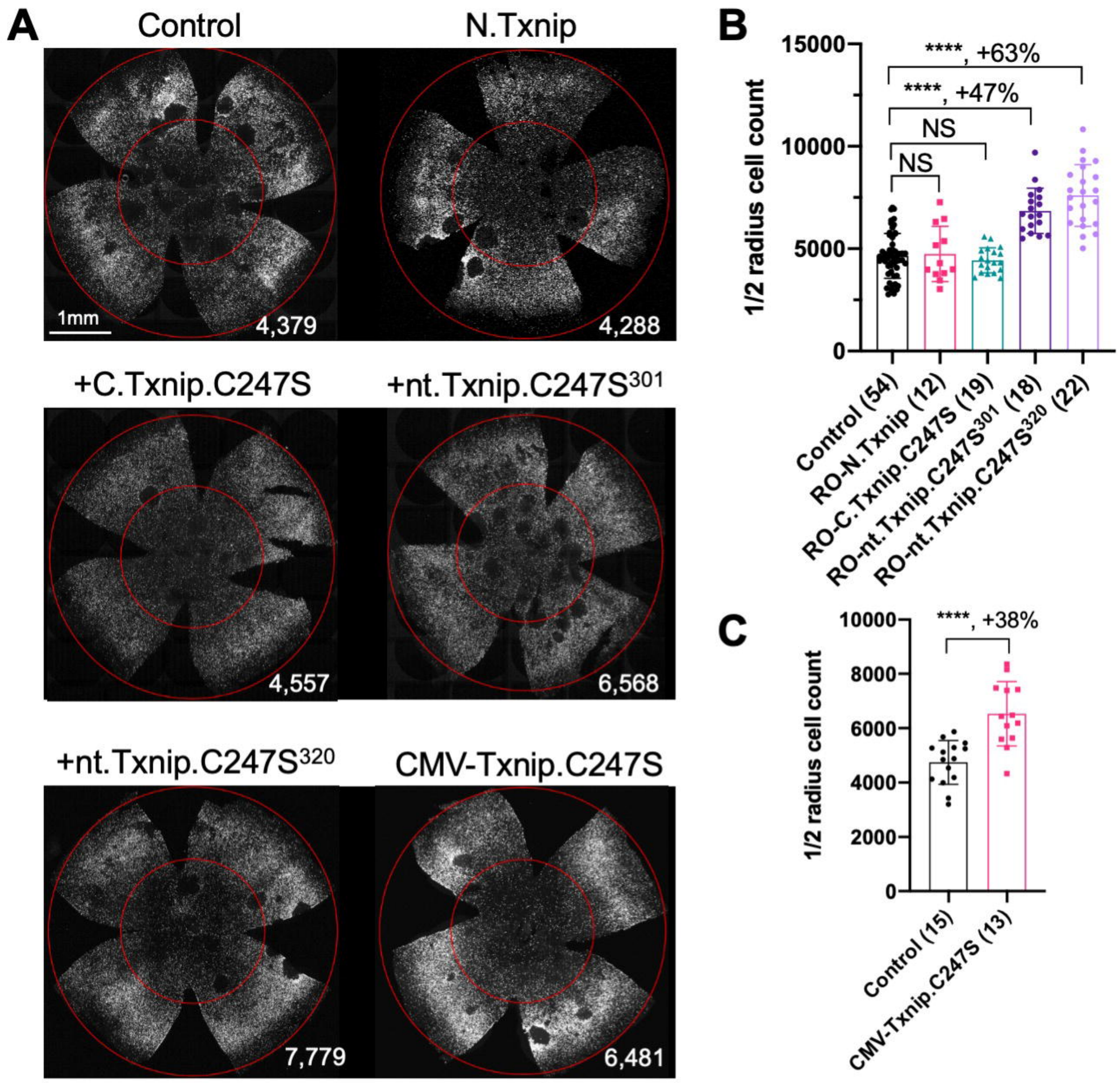
A
Control
N.Txnip
1mm
4,379
4,288
+C.Txnip.C247S
+nt.Txnip.C247S301
4,557
6,568
+nt.Txnip.C247S320
CMV-Txnip.C247S
7,779
6,481
B
1/2 radius cell count
15000
10000
5000
0
****, +63%
****, +47%
NS
NS
Control (54)
RO-N.Txnip (12)
RO-C.Txnip.C247S (19)
RO-nt.Txnip.C247S301 (18)
RO-nt.Txnip.C247S320 (22)
C
1/2 radius cell count
10000
8000
6000
4000
2000
0
****, +38%
Control (15)
CMV-Txnip.C247S (13)

Figure 3

	RPE GLUT1 Removal	Cone Rescue: Expression in RPE	Cone Rescue: Expression in Cones
wt Txnip (1-397aa)	Y	N	Y (60%)
Txnip.S308A (1-397aa)	NT	NT	N
Txnip.C247S (1-397aa)	I	N	Y (97%)
Txnip.C247S.LL351&352AA (1-397aa)	N	Y (45%)	Y (43%)
N.Txnip (1-228aa)	N	N	N
C.Txnip.C247S (149-397aa)	Y	Y (54%)	N
C.Txnip.C247S.LL351&352AA (149-397aa)	N	Y (58%)	NT
sC.Txnip (255-397aa)	I	N	NT
nt.Txnip.C247S (1-301aa)	NT	NT	Y (47%)
nt.Txnip.C247S (1-320aa)	N	Y (16%)	Y (63%)
	Cone Rescue		
CMV-Txnip.C247S	Y (38%)		
RedO-Txnip.C247S + Best1-Nrf2	Y (14% increase to RedO-Txnip.C247S)		

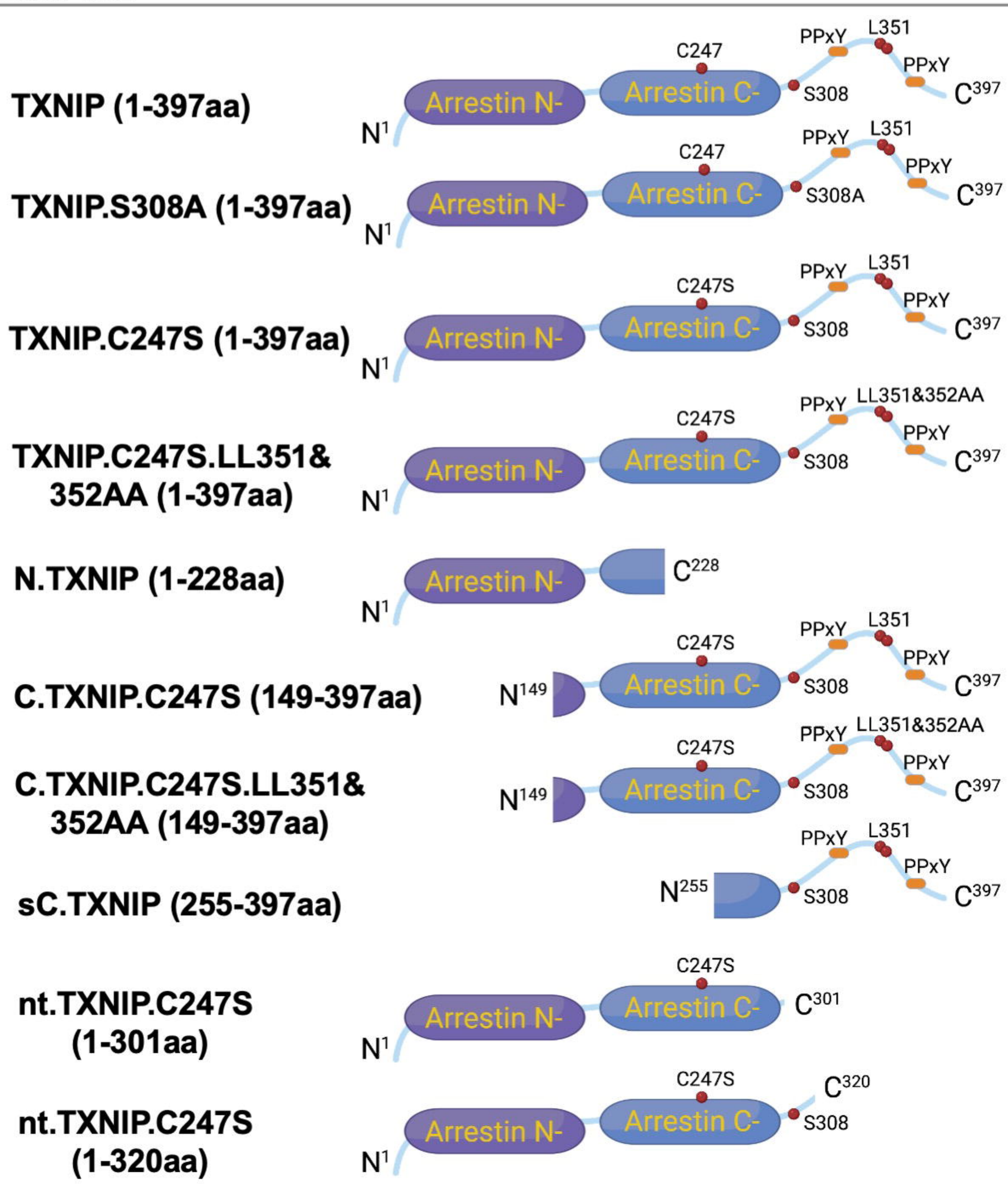

Figure 4

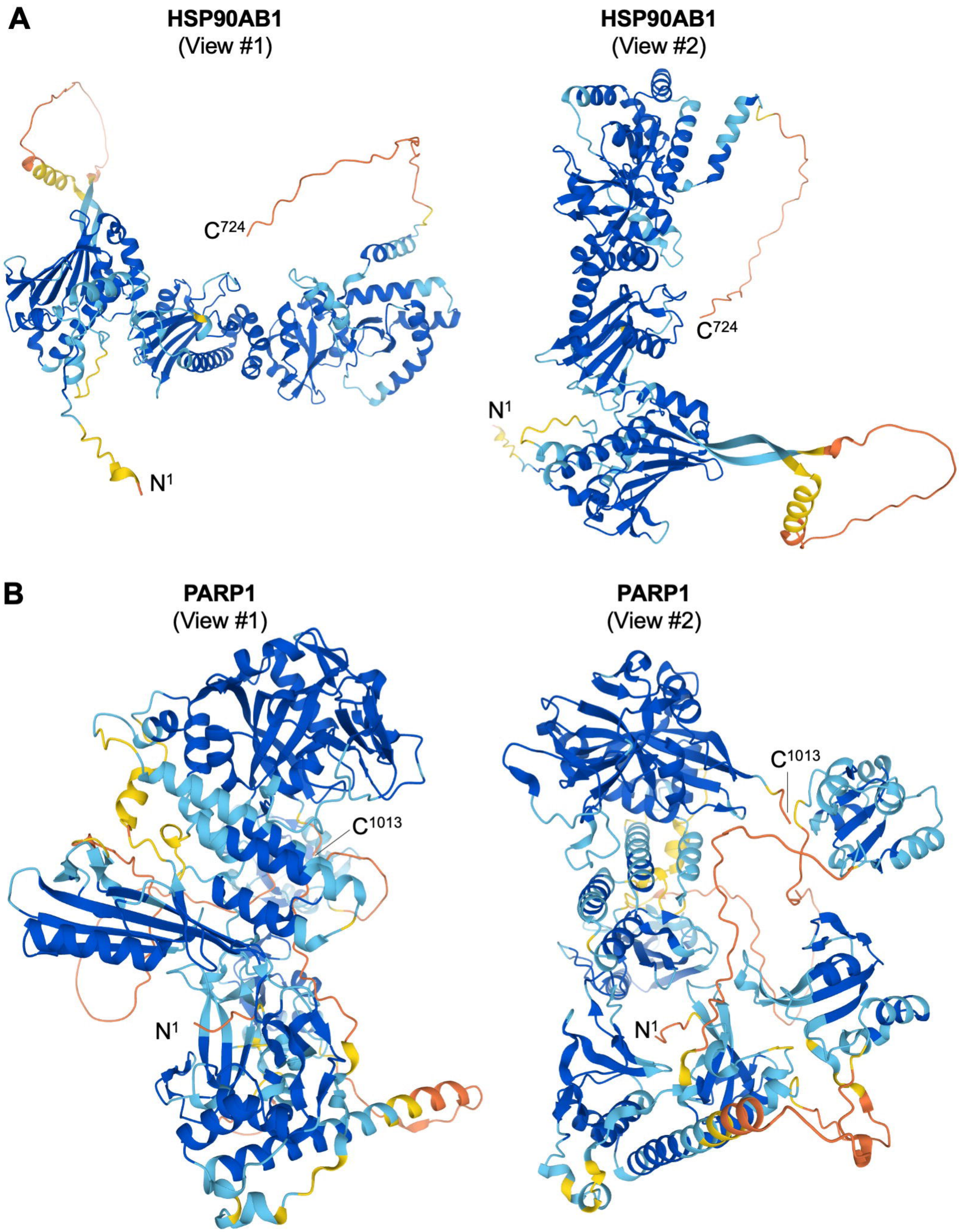

Figure 5 Supplement

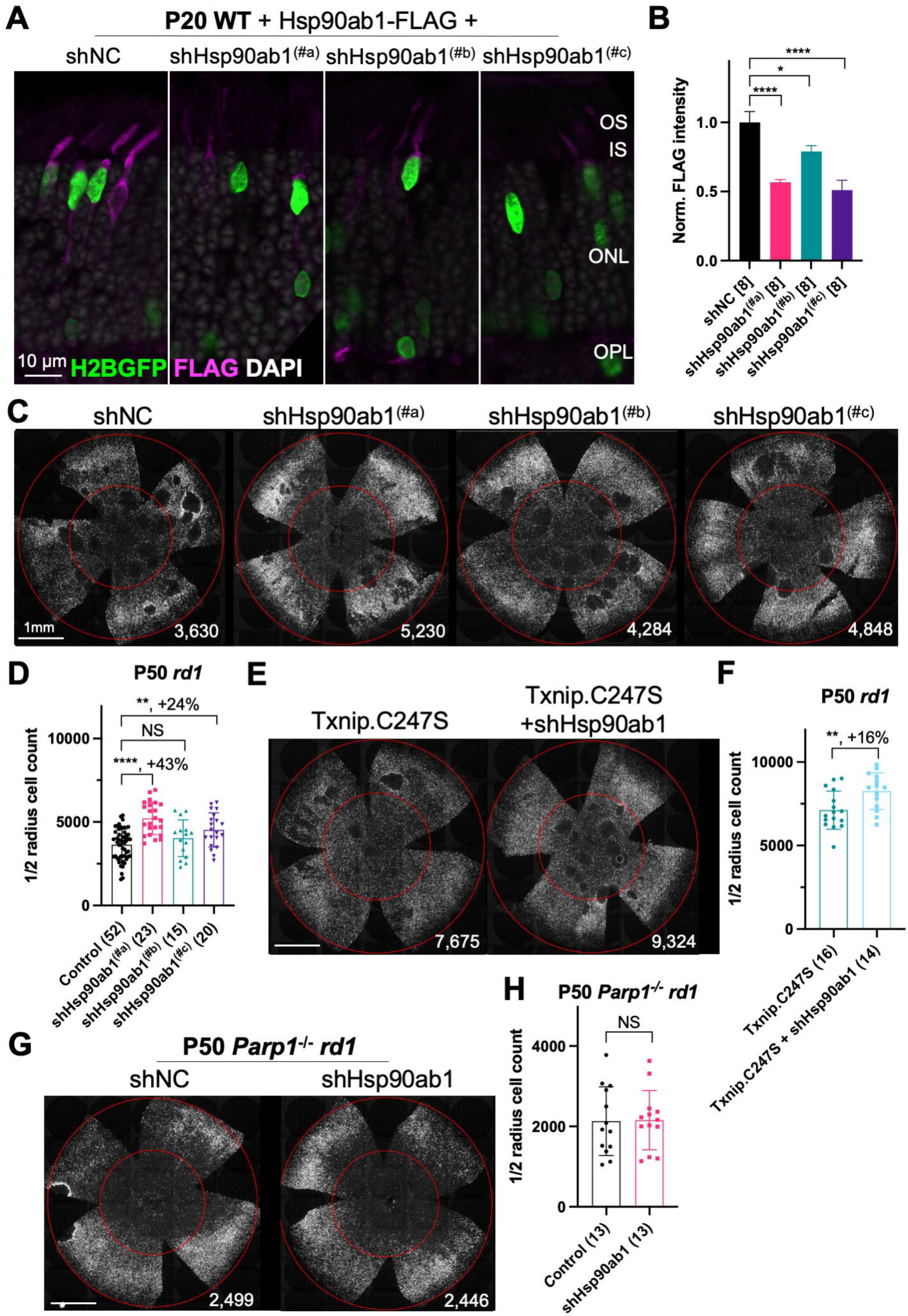
A
P20 WT + Hsp90ab1-FLAG +
shNC
shHsp90ab1(#a)
shHsp90ab1(#b)
shHsp90ab1(#c)
OS
IS
ONL
OPL
10 µm
H2BGFP FLAG DAPI
B
Norm. FLAG intensity
1.0
0.5
0.0
****
*
****
shNC [8]
shHsp90ab1(#a) [8]
shHsp90ab1(#b) [8]
shHsp90ab1(#c) [8]
C
shNC
shHsp90ab1(#a)
shHsp90ab1(#b)
shHsp90ab1(#c)
1mm
3,630
5,230
4,284
4,848
D
P50 rd1
1/2 radius cell count
10000
5000
0
**, +24%
NS
****, +43%
Control (52)
shHsp90ab1(#a) (23)
shHsp90ab1(#b) (15)
shHsp90ab1(#c) (20)
E
Txnip.C247S
Txnip.C247S +shHsp90ab1
7,675
9,324
F
P50 rd1
1/2 radius cell count
10000
5000
0
**, +16%
Txnip.C247S (16)
Txnip.C247S + shHsp90ab1 (14)
G
P50 Parp1-/- rd1
shNC
shHsp90ab1
2,499
2,446
H
P50 Parp1-/- rd1
1/2 radius cell count
4000
2000
0
NS
Control (13)
shHsp90ab1 (13)

Figure 5