Differential conformational dynamics in two type-A RNA-binding domains drive the double-stranded RNA recognition and binding

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ABSTRACT

TAR RNA binding protein (TRBP) has emerged as a key player in the RNA interference (RNAi) pathway. While facilitating Dicer-mediated pre-miRNA cleavage and recruitment of the RISC complex, TRBP binds to different pre-miRNAs & siRNAs, each varying in sequence, structure, or both. Our hypothesis posits that TRBP displays dynamic adaptability to accommodate this heterogeneity in target RNA structures. Thus, it is crucial to ascertain the role of intrinsic and RNA-induced protein dynamics in RNA recognition and

binding. Prior investigations from the lab have elucidated the role of intrinsic and RNA-induced conformational exchange in the double-stranded RNA-binding domain 1 (dsRBD1) of TRBP in shape-dependent RNA recognition. Building further on this, the current study delves into the intrinsic and RNA-induced conformational dynamics of the dsRBD2 of TRBP, juxtaposed with dsRBD1. Remarkably, dsRBD2 exhibits a higher binding affinity to a short (12bp) dsRNA when compared with dsRBD1, which can be attributed to the presence of residues critical to RNA binding and structural plasticity in dsRBD2. Utilizing state-of-the-art NMR spin relaxation, relaxation dispersion experiments, and MD simulations, we report that dsRBD2 depicts constrained conformational plasticity when compared to dsRBD1, as evidenced by the fast (ps-ns) and slower (μs-ms) conformational dynamics measurements. Although, in the presence of RNA, dsRBD2 undergoes induced conformational exchange within the designated RNA-binding regions and the other residues, the amplitude of the motions remains modest when compared to those observed in dsRBD1. The culmination of our findings leads us to propose a dynamics-driven model of the two tandem domains of TRBP, substantiating their contributions to the versatility of dsRNA recognition and binding.

INTRODUCTION

RNA-binding proteins (RBPs) play a crucial role in every aspect of RNA biology, including folding, splicing, processing, transport, and localization (1–5). RBPs can be broadly classified into single-stranded RNA-binding proteins (ssRBPs) and double-stranded RNA-binding proteins (dsRBPs). While ssRBPs are generally sequence-specific as the interaction points include exposed nucleobases (6, 7); dsRBPs tend to target a particular structural fold, especially the A-form RNA duplex via phosphate backbone and sugar hydroxyl protons (8).

dsRBPs interact with highly structured dsRNAs via their double-stranded RNA binding motifs/domains (dsRBMs/dsRBDs).

dsRBDs are 65-68 amino acid long domains (9) that contain a highly conserved secondary structure (α_1-β_1-β_2-β_3-α_2), where the two α-helices are packed against an antiparallel β-sheet formed by three β-strands (10, 11). All the dsRBDs have three common RNA-binding regions: (1) the middle of helix α1 (E residue), (2) the N-terminal residues of helix α2 (KKxAK motif), and (3) the loop between β1 and β2 strands (GPxH motif), forming a canonical RNA-binding surface that interacts with the consecutive minor, major, and minor groove of a dsRNA (8, 12). dsRBDs interact with dsRNAs in a non-sequence-specific manner targeting the sugar 2'-OH groups (dsRBDs do not target dsDNAs) and phosphate backbone (13). dsRBDs are often arranged in a modular fashion to create highly versatile RNA-binding surfaces. dsRBDs have been classified into type A and type B (8, 9, 14, 15). Type A dsRBDs are highly homologous (> 59%) to the consensus sequence and are indispensable to RNA-binding, whereas type B has only conserved C-terminal end, binds weakly to the RNA, and in some cases stabilizes the protein-protein complex (16–18).

dsRBDs, both within and across proteins, exhibit significant variations in their binding modes and affinities towards specific RNA targets. For example, *X. laevis* RNA-binding protein A (XlrbpA) dsRBD1 and dsRBD2, classified as type A dsRBDs, demonstrate notably stronger RNA-binding affinity than XlrbpA-dsRBD3, which falls under type B (14). In Protein Kinase R (PKR), these two types of dsRBDs also demonstrate a differential dynamic behavior, i.e., dsRBD1 (type A) shows plasticity on μs and ps-ns timescale, while dsRBD2 (type B) predominantly depicts dynamics on the ps-ns timescale dynamics (15, 19). Furthermore, the average order parameter (S^2) for the individual α-helices and β-sheets is observed to be lower for dsRBD1 than for dsRBD2, suggesting the latter as a more rigid domain (15, 19). Similarly, TRBP dsRBD2 shows a significantly stronger binding affinity

(four times) than dsRBD1 for pre-miR-155 (20). Additionally, studies have reported that dsRBD1 and dsRBD2 of TBBP1 display a similar binding affinity for siRNA, whereas, for TAR RNA, dsRBD1 has a higher binding affinity than dsRBD2 (21, 22). It is intriguing to notice that the two type-A dsRBDs originating from the same protein demonstrate a markedly different binding affinity for target RNAs across various cases, as elucidated above. Recent investigations have suggested that the dissimilar behaviour of dsRBDs in Dicer (23), PKR (19), and double-stranded-RNA-binding protein 4 (DRB4) (24) might be attributed to protein dynamics. Although variations in the binding affinity for the two type-A dsRBDs of TRBP have been documented for several target RNAs, a comprehensive exploration of the differences in the conformational dynamics between these two domains remains to be undertaken.

In this study, we have investigated the role of differential conformational dynamics of the two type A dsRBD domains of TRBP2 (isoform 1 of TRBP) in RNA recognition and binding. First discovered as a TAR RNA-binding protein (39 kDa) involved in HIV-I replication (25), TRBP was later found to be indispensable to the RNAi pathway (26). It is involved in Dicer-mediated pre-miRNA/pre-siRNA cleavage, recruitment of Ago (27), and guide strand selection (28, 29). TRBP and its homologs across different species such as Loquacious (Loqs in *D. melanogaster*), R2D2 (*D. melanogaster*), DRB1-3,5 (*A. thaliana*), RNAi defective 4 (RDE-4 in *C. elegans*), Xlrbpa (*X. laevis*) (30), and the PKR activator (PACT) protein (mammals) (31) have conserved arrangement of three consecutive dsRBDs; of which, the two N-terminal domains (type A) are known to bind dsRNAs, whereas the third domain is known for protein-protein interactions. We have recently established the role of TRBP2-dsRBD1 dynamics in dsRBD-dsRNA interactions and proposed that dsRBD1 adopts a conformationally dynamic structure to recognize a set of topologically different dsRNA structures (32). The μs timescale motions were found to be present all along the dsRBD1

backbone with higher frequency motions (k_{ex} > 50 kHz) in the RNA-binding sites. The data suggested that the presence of conformational exchange in the µs timescale could help the dsRBD1 to dynamically tune itself for targeting conformationally distinct dsRNA substrates.

In this work, we have compared TRBP2-dsRBD1 with TRBP2-dsRBD2 in terms of the structure, dynamics (intrinsic and RNA-induced), and dsRBD-dsRNA interactions in the two type-A domains. We have measured motions in the TRBP2-dsRBD2 at ps-ns and µs-ms timescale dynamics by NMR relaxation dispersion experiments in apo-state and studied its perturbation in the presence of an A-form duplex RNA. We also compared the apo- and RNA-bound conformational dynamics measured in dsRBD2 with that of dsRBD1. Based on our observations, we propose that the differential protein dynamics and its perturbation in the presence of RNA in the two dsRBDs enables them to recognize a variety of RNA substrates and may lead to diffusion along the length of the RNA.

MATERIALS AND METHODS

Protein overexpression and purification

The TRBP2-dsRBD1 (1–105 aa) and TRBP2-dsRBD2 (154–234 aa) cDNA clones were obtained as a kind gift from N. L. Prof. Jennifer Doudna (University of California, Berkeley, CA, USA). The residue numbers in the two constructs (Figure 1A) have been mentioned in reference to the full-length TRBP2 sequence (1–366 aa, Uniprot ID: Q15633-1). The cDNA for TRBP2-dsRBD2 was cloned in pHMGWA vector (Amp^R), and was expressed as a fusion protein having N-terminal His_6-Maltose binding protein (MBP) tag-TEV protease cleavage site followed by the protein of interest. Treatment with TEV protease during purification resulted in non-native Ser-Asn-Ala residues at the N-terminal to TRBP2-dsRBD2 (154–234), which were excluded from all the NMR-based dynamics studies. For the NMR experiments, ^{15}N-labeled and ^{15}N-^{13}C labeled (as required) TRBP2-dsRBD2 protein

was prepared using $^{15}NH_4Cl$ and ^{13}C-glucose (Cambridge Isotope Laboratories) as a sole source of nitrogen and carbon, respectively, in M9 minimal medium.

TRBP2-dsRBD1 was overexpressed and purified as previously described (33). Briefly, the pHMGWA plasmid containing the cDNA for the TRBP2-dsRBD2 was transformed into *E. coli* BL21 (DE3) cells and plated on an LB agar plate (containing 100 μg/mL ampicillin) and incubated at 37 ºC for 12 h. A single isolated colony was inoculated into 20 mL LB broth (containing 100 μg/mL ampicillin) and incubated for 12 h at 37 ºC, 225 rpm to initiate a starter culture that was eventually used to inoculate 2 L LB broth (containing 100 μg/mL ampicillin) and incubated at 37 ºC, 225 rpm till the OD_{600} reached to 0.8–1.0. For induction, IPTG was added at the final concentration of 1 mM, and the culture was further incubated at 37 ºC, 225 rpm for another 8 h. The cells were harvested by centrifuging the culture at 4500 *g*, 4 ºC for 20 min and resuspended in 25 mL of buffer A (20 mM Tris-Cl, pH 7.5, 500 mM NaCl, 10% glycerol, 5 mM DTT, 10 mM Imidazole). To the resuspended cells, lysozyme (Sigma-Aldrich) was added to a final concentration of 50 μg/mL and incubated in ice for 30 min. Post-incubation, 1% Triton X-100, 100 μl of 1 mM PMSF, and 10 X Protease Inhibitor Cocktail (PIC, Roche) (2 mL per 4 g of cell pellet) were added to the cell suspension. The partially lysed cells were sonicated using an ultrasonic sonicator microprobe at 60% amplitude, with 5 s ON and 10 s OFF pulse, for a period of 60 min in an ice bath for complete lysis. The cell lysate was centrifuged at 15,000 x *g* for 2 h, at 4ºC, to obtain the total soluble protein (TSP). The TSP was then circulated through a pre-equilibrated (buffer A) Ni-NTA column (HisTrap, 5 ml, GE HealthCare) for 4 h at 4 ºC. After equilibrating with the TSP, the column was washed with 40 CVs of buffer A containing 30 mM imidazole to remove the impurities. The fusion protein was eluted with the elution buffer B (buffer A containing 300 mM imidazole). Nucleic acids were removed from the eluted protein using polyethyleneimine (PEI) precipitation. To remove imidazole, the protein solution was

dialyzed against cleavage buffer C (20 mM Tris-Cl, pH 7.5, 25 mM NaCl, 10% glycerol, 5 mM DTT). His_6-MBP-tag was cleaved using TEV protease at a final concentration of 1:100 (protease: protein) at 4 ºC for 18 h with intermittent mixing. The completion of the cleavage reaction was tested and confirmed by SDS-PAGE analysis. The cleavage mix was then passed through a sulphopropyl sepharose column (HiPrep SP FF 16/10 20ml, GE HealthCare) and washed with buffer C. TRBP2-dsRBD2 protein was eluted from the cation-exchange column using a salt gradient ranging from 25 mM to 1 M NaCl in the buffer C. To further purify, the protein was subjected to size-exclusion chromatography using sephacryl S-100 HR 16/60 column (GE HealthCare). The final purified protein was concentrated to 1 mM using Amicon (3 kDa cutoff, Merck) and exchanged with NMR buffer D (10 mM sodium phosphate, pH 6.4, 100 mM NaCl, 1 mM EDTA, 5 mM DTT) before recording any experiment.

Design and preparation of RNA

RNA duplexes miR-16-1-A, miR-16-1-M, miR-16-1-B, and miR-16-1-D were designed and procured as mentioned previously (32). The shorter 12 bp RNA duplex RNA oligo (D12) was designed from miR-16-1-D and was procured from either Integrated DNA Technologies (Coralville, IA, USA) or GenScript Biotech Corporation (Piscataway NJ, USA). All the RNA sequences used in this study have been listed in Supplementary Table S1.

The preparation of RNA duplexes and the confirmation of their formation was performed, as described previously (32). The annealing of D12 RNA was confirmed by ^{1}H NMR by observing the imino protons (Figure S1). The annealed samples were maintained in buffer D for all data measurements.

Size-Exclusion Chromatography – Multiple Angle Light Scattering

Size-exclusion chromatography coupled with multiple angle light scattering (SEC-MALS) experiments were performed using an S75 column (Superdex 75 10/300 GL 24 ml, GE Healthcare), Agilent HPLC system (Wyatt Dawn HELIOs II) and a refractive index detector (Wyatt Optilab T-rEX). The system was first calibrated by injecting 100 μl of 30 μM Bovine Serum Albumin solution (ThermoScientific). Post calibration, 100 μl protein samples were injected (in duplicate) at a concentration of 0.8 mM for TRBP2-dsRBD2. The respective molar mass values of the peaks were calculated using the Zimm model in ASTRA software version 7 (Wyatt Technologies).

Isothermal titration calorimetric binding assays

All isothermal titration calorimetry (ITC) experiments were performed using a MicroCal PEAQ-ITC calorimeter (Malvern Panalytical, Malvern, UK) operating at 25ºC. The final RNAs and protein solutions used for the assays were prepared in buffer D. The D12 dsRNA was used at a concentration of 10 or 20 μM in the sample cell. TRBP2-dsRBD1 concentration was varied from 5–19 folds of RNA, whereas, in the case of TRBP2-dsRBD2, it was varied from 10–18 folds. The first injection was 0.4 μl (discarded for data analysis), which was followed by eighteen 2 μl injections.

Data were fitted with a single-site binding model using the MicroCal PEAQ-ITC analysis software (Malvern Panalytical, Malvern, UK) to extract the binding affinity (K_D), stoichiometry (n), enthalpy (ΔH), and entropy (ΔS). These parameters were used to derive the Gibbs free energy change of binding using the equation:

$\Delta G = -RT \ln (K_D)$,

and the overall change in the entropy of the system, using the experimental temperature and the relationship:

$\Delta G = \Delta H - T\Delta S$.

The final values of the kinetic and thermodynamic parameters are given as the average of triplicate measurements (Supplementary Table S2).

NMR Spectroscopy

All the NMR experiments were recorded at 298 K either on: 1) Ascend^{TM} Bruker AVANCE III HD 14.1 Tesla (600 MHz) NMR spectrometer equipped with a quad-channel ($^{1}H/^{13}C/^{15}N/^{19}F$) Cryoprobe (in-house); or on 2) Ascend^{TM} Bruker Avance AV 18.89 Tesla (800 MHz) NMR spectrometer equipped with a triple-channel ($^{1}H/^{13}C/^{15}N$) Cryoprobe and a Broad Band Inverse probe (located at National Facility for High-Field NMR at TIFR, Mumbai). The ^{1}H-^{15}N HSQC spectrum was collected with 2048 and 128 points and 12 ppm and 28 ppm spectral width in ^{1}H and ^{15}N dimensions, respectively, giving an acquisition time of 100 ms in the direct dimension. An inter-scan delay of 1.0 s and 4 scans were used on a 1 mM ^{15}N-labeled TRBP2-dsRBD2 sample in a 5 mm Shigemi tube (Shigemi Co., LTD., Tokyo, Japan) (34). ^{1}H-^{15}N TOCSY-HSQC (mixing times = 60, 80, and 120 ms) and ^{1}H-^{15}N NOESY-HSQC (mixing times = 150, 300, and 400 ms) were recorded on a 600 MHz NMR spectrometer on ^{15}N-labeled TRBP2-dsRBD2. Further, triple resonance experiments like HNCO, HNCACO, HNCA, HNCOCA, HNN, CBCANH, and CBCA(CO)NH were carried out using 1.2 mM of ^{15}N-^{13}C-labeled TRBP2-dsRBD2 sample in a 5 mm Shigemi tube. All the NMR data were processed via TopSpin/NMRPipe (35) and were analyzed in SPARKY/CARA (36, 37).

For NMR-based titration assays, ^{1}H-^{15}N-HSQCs were measured on TRBP2-dsRBD2 (50 μM) with increasing concentrations of duplex RNAs from 0.05–0.2 (in the case of D12 RNA 0.05–5 equivalents) equivalents of the protein. After every RNA addition, the protein was allowed to equilibrate for 30 mins before acquiring ^{1}H-^{15}N HSQC. Peak intensities were

plotted against the RNA:protein concentration for each residue and were fit to the one-site binding isotherm using the below equation (38):

$$\Delta\delta_{obs} = \Delta\delta_{max}\{([P]_t + [L]_t + K_d) - [([P]_t + [L]_t + K_d)^2 - 4[P]_t[L]_t]^{\frac{1}{2}}\} / 2[P]_t$$

where, $\Delta\delta_{obs}$ is the change in the observed chemical shift from the apo state, $\Delta\delta_{max}$ is the maximum chemical shift change on saturation with ligand, $[P]_t$ and $[L]_t$ are the total concentration of protein and ligand, respectively, and K_d is the dissociation constant.

Apo-TRBP-dsRBD2 nuclear spin relaxation experiments (R_1, R_2, and [^{1}H]-^{15}N-nOe) were recorded at two different field strengths (600 and 800 MHz NMR spectrometers) on a 1 mM ^{15}N-TRBP2-dsRBD2 sample in a 5 mm Shigemi tube. ^{15}N longitudinal relaxation rates (R_1) were recorded with 8 inversion recovery delays of 10, 30*, 50, 100, 200, 300, 450*, and 600 ms. ^{15}N transverse relaxation rates (R_2) were recorded with 8 CPMG (Carr-Purcell-Meiboom-Gill) delays of 17, 34*, 51, 68, 85, 102, 136*, and 170 ms. Steady-state [^{1}H]-^{15}N heteronuclear nOe experiments were recorded with and without ^{1}H saturation with a relaxation delay of 5 s.

All D12-bound TRBP-dsRBD2 nuclear spin relaxation experiments were recorded in a similar fashion as the apo-protein on a 1 mM ^{15}N-TRBP2-dsRBD2 in the presence of 50 μM D12 RNA in a 3 mm NMR tube. The ^{15}N-R_1 rates were measured with 5 inversion recovery delays of 10, 30*, 70, 150, and 600 ms on 600 MHz spectrometer and 10, 70*, 150, 300, and 600* ms on 800 MHz spectrometer. The ^{15}N-R_2 rates were measured with 5 CPMG delays of 17, 51, 85, 136*, and 170 ms on 600 MHz spectrometer and with 4 CPMG delays of 17, 34*, 51, and 68 ms on 800 MHz spectrometer with a CPMG loop length of 17 ms.

For ^{15}N relaxation dispersion measurement, a constant time CPMG (Carr-Purcell-Meiboom-Gill) experiment (39) was recorded on a 600 MHz NMR spectrometer. CPMG relaxation dispersion experiments for apo and D12-bound ^{15}N-TRBP2-dsRBD2 were

acquired separately, consisting of 15 data points with ν_{cpmg} values of 25, 50, 75, 125*, 175, 275, 375*, 525, 675, 825*, and 1000 Hz at a constant relaxation time – T_{relax} (40 ms).

Heteronuclear Adiabatic Relaxation Dispersion (HARD) experiments (40, 41) were recorded on apo and D12-bound ^{15}N-TRBP2-dsRBD2 on the 600 MHz NMR spectrometer, as described previously (32). The relaxation delays used for $R_{1\rho}$ were 0, 16, 32, 64, 96, and 128 ms for apo-; and 0, 16, 32, 64, and 112 ms for D12-bound ^{15}N TRBP2-dsRBD2. For $R_{2\rho}$ experiments, the relaxation delays used were 0, 16, 32, and 64 ms for apo and 0, 16, and 32 ms for D12-bound ^{15}N-TRBP2-dsRBD2. R_1 experiments were acquired similarly to $R_{1\rho}$ and $R_{2\rho}$ experiments without using the adiabatic pulse during evolution. The delays used for the R_1 experiment were 16, 48, 96, 160, 224, 320, 480, and 640 ms for both apo- and RNA-bound protein samples.

All relaxation experiments were measured using a single scan interleaving method, and the order of delays was randomized. The time periods marked with an asterisk have been recorded in duplicates for error estimation. An inter-scan delay of 2.5 s was used for all the above-mentioned relaxation experiments (unless mentioned otherwise).

NMR Relaxation Data Analysis

NMR spectra were processed with either TopSpin 3.5pl6 or NMRPipe and visualized through SPARKY (version 3.115). For the backbone resonance assignment, CARA was used. All the input NMR spectra were fed in the form of either ucsf or 3rrr file formats. Manual peak picking was done for double and triple-resonance NMR experiments using a synchroscope in CARA. Further identification, confirmation, and assignment of residues were done in a stripscope mode. The chemical shifts of the assigned residues were used as inputs in the CS-Rosetta program in the BMRB server

(https://csrosetta.bmrb.wisc.edu/csrosetta/submit) to get an ensemble of the 10 lowest energy structures the structure.

Different relaxation rates like $R_1/R_2/R_{1\rho}/R_{2\rho}$ were calculated using mono-exponential decay fitting of peak height against the corresponding set of relaxation delays in a Mathematica script (42). Steady-state [^{1}H]-^{15}N nOes for individual residues were calculated as a ratio of the corresponding residue peak height in the spectra recorded in the presence and absence of ^{1}H saturation.

Additional analysis of ^{15}N-relaxation data (R_1, R_2, [^{1}H]-^{15}N nOe) was conducted using the extended model-free formalism via Relax v4.0.3 software (43) in a similar fashion carried out previously (32). The structure of TRBP-dsRBD2 available with PDB ID 2CPN was employed for this analysis.

The effective transverse relaxation rates (R_{2eff}) from CPMG relaxation dispersion experiments, the R_1, $R_{1\rho}$, and $R_{2\rho}$ relaxation rates obtained from the HARD experiments, the rotational correlation time, and the subsequent k_{ex}, and p_A were calculated by fitting the R_1, $R_{1\rho}$, and $R_{2\rho}$ data to a two-state model using numerical fittings as described previously (32, 42). The errors in different fitted parameters were obtained using 500 Monte-Carlo simulations in addition to the duplicate relaxation data points.

Molecular dynamics simulations

All the ensemble cluster analysis was done using UCSF Chimera (44) on the CS-Rosetta (45, 46) structure of TRBP2-dsRBD2 to select the lowest energy structures. Those were used as the seed structure for apo TRBP2-dsRBD2 molecular dynamics (MD) simulation. Briefly, GROMACS V.2019.6 (https://www.gromacs.org) was used for all the atomistic MD simulations. All-atom additive CHARMM36 protein force field was used to build the topologies (47), and initial structure solvation was done TIP3P water molecules in a

cubic simulation box with a minimum distance of 1.2 nm from protein in all directions. Microsecond timescale simulations were set up in triplicates in a similar way as described previously (32). All analysis was performed using GROMACS and VMD.

RESULTS AND DISCUSSION

Structural and dynamical comparison of apo TRBP2-dsRBD1 and TRBP2-dsRBD2

Primary sequence comparison of TRBP2-dsRBD1 and TRBP2-dsRBD2 domain constructs (Figure 1A) revealed 30% identity and 38% similarity. The consensus for dsRNA-binding has been marked in red (Figure 1B). The two reported RNA-binding regions 1 (E) and 3 (KKxAK) of both the domains were matched to the consensus. While region 2 (GPxH) of TRBP2-dsRBD2 was an exact match to the consensus, dsRBD1 harbored a mutation in region 2 (P56Q). Proline is a rigid amino acid with one less dihedral angle; it imparts flexibility to the backbone by causing secondary structure breaks (48, 49). Thus, the presence of Pro186 in the β_1-β_2 loop of dsRBD2 may perturb the β_1-β_2 loop region plasticity, thereby making it more accessible to the incoming RNA partner. Additionally, dsRBD2 also contains a KR-helix motif in α_2-helix which is also known to increase its binding affinity. Owing to the tightly conserved RNA-binding regions and the presence of the KR-helix motif, dsRBD2 could make stronger contact with the RNA.

The SEC-MALS study of TRBP2-dsRBD2 at experimental concentrations showed a single monomeric species in solution (Figure S2). The ^{1}H-^{15}N HSQC spectrum of TRBP2-dsRBD2 indicated a well-folded protein. It (154–234 aa) was compared to previously assigned spectra of the TRBP2-D1D2 construct (19–228 aa) (20) and TRBP2-dsRBD2 construct (157–228 aa) (21) to transfer the backbone amide resonance assignments. Resonance assignments were further confirmed using a set of double and triple resonance experiments, and a total of 73 non-proline residues were assigned as shown in the

representative ^{1}H-^{15}N HSQC (Figure S3). Overall, 89% ^{1}H (151/169), 74% ^{13}C (124/168), and 87% ^{15}N (73/84) resonances from the backbone, and 43% ^{1}H (174/402) and 25% ^{13}C (59/238) from the side chains were assigned. Similar to the CS-Rosetta structure calculated previously for dsRBD1 (Figure 1C) by our group (33), the CS-Rosetta structure calculated for the dsRBD2 (Figure 1D) also matched with the previously reported structures of the core dsRBD2 (159–227 aa) and was found to adapt the characteristic αβββα fold of a dsRBD. An alignment of core structures between dsRDB1 and dsRBD2 (Figure 1E) yielded an RMSD of 0.894 □, indicating a close match between the two domains. Despite their core length being identical (69 aa), the individual secondary structure spans were found to be different (Supplementary Table 3). The flexible regions, like loops 1 and 2, are longer in dsRBD2, whereas the structured regions β_2, β_3, and α_2 are longer in dsRBD1. Most importantly, loop 2 (β_1-β_2 loop) — critical for RNA-binding (8)— is equal to the canonical length in dsRBD2, while in dsRBD1, it is shorter by 1 residue.

The MD simulation data of RMSD against the simulation time of TRBP2-dsRBD2 showed that the protein remained stable during the course of triplicate simulations, as seen previously in the case of dsRBD1 (Figure 2B). Comparing the core residue-specific RMSF plots of these two type A dsRBD domains (Figure 2A), we observe that the average RMSF of the core domain of dsRBD2 is 0.22 nm, which is almost half as compared to dsRBD1 RMSF average of 0.43. This, in turn, indicates that dsRBD2 is overall more rigid amongst the two. The most interesting fact is that loop 2 of dsRBD2 is very flexible compared to the rest of the core, with an average RMSF of 0.49 nm (more than double the core average). Hence, we propose that this long, conserved, and dynamic loop 2 might help dsRBD2 to establish strong contacts in the minor grooves of dsRNA partners. An effect of conserved insertions in the β_1-β_2 loop on RNA binding has been previously discussed in the crystal structure of full-length *Arabidopsis* HEN1 in complex with a small RNA duplex (50).

To gain insights into the picosecond to nanosecond (ps-ns) timescale motions of apo-dsRBD2, NMR spin relaxation data (R_1, R_2, and [^{1}H]-^{15}N NOE) was recorded on 600 and 800 MHz NMR spectrometers. The set of plots for R_1, R_2, and [^{1}H]-^{15}N NOE at both 600 and 800 MHz magnetic fields showed the expected trend (Figure S4). R_1 in the structured residues showed lower values at a higher magnetic field and similar values in the terminal regions at the two magnetic fields. The increase of core R_1 rates (decreasing T_1) with increasing magnetic field indicates that most of the core residues lie on the right-hand side of the T_1 minima in the standard plot of T_1 vs. $\omega\tau_c$, where ω is the frequency of motions, and τ_c is rotational correlation time (51). The core residues had average R_1 rates of 1.44 ± 0.05 Hz (Figure 6). The R_2 rates of a few N-terminal (S156, N159) and core residues (V180, M194, R197, V198, G207, and A217) marginally increased (0.7–3 Hz) when measured at 800 MHz than when measured at 600 MHz, suggesting a very insignificant presence of the R_{ex} component in the linewidth of these residues in the experimental conditions (Figure S4). The terminal (N159, T227) and the loop residues (G185, E191, F200, K210) exhibited relative flexibility as indicated by lower R_2 and nOe values (Figure 6). A few residues lying in different secondary structures, e.g., Q165, L167, and V168 in α_1; T193 and T195 in β_2; L212, A213, N216, and L223 in α_2 exhibited only slightly higher (> 1 Hz) than the average R_2 rate (11.02 ± 0.39 Hz), indicating a thin conformational exchange in the secondary structured regions (Figure 6).

The average R_1 (dsRBD1 = 1.52 ± 0.03 Hz; dsRBD2 = 1.44 ± 0.05 Hz) and R_2 rates (dsRBD1 = 9.83 ± 0.17 Hz; dsRBD2 = 11.02 ± 0.39 Hz) for the two domains were very similar (Figure S15 of (32)). In contrast, dsRBD1 had significantly lower average heteronuclear steady-state nOe values (0.63 ± 0.02) than dsRBD2 (0.75 ± 0.03), indicating a more rigid core at the picosecond timescale motions in dsRBD2, thereby supporting the observations made in the MD simulation study.

The extended model-free analysis of the relaxation data for the two core domains allowed us to compare the order parameters S^2, R_{ex} components, and global tumbling time (τ_c). The overall order parameters for dsRBD1 followed the expected trend of higher values in the secondary structured regions and lower values in the terminals and loops (Figure S5). Interestingly, the secondary structural motifs α_1, loop 2, and α_2, containing RNA-binding regions of dsRBD2, showed a higher rigidity than dsRBD1. The presence of conserved Pro in loop 2 (L2) of dsRBD2 and a longer length of L2 in dsRBD2 might be responsible for the additional observed flexibility in dsRBD2, as also observed in the MD simulations (Figures 2 and S3). Loop 1 (W173, L175) of dsRBD2 containing the stabilizing tryptophan (absent in dsRBD1) also showed higher rigidity than dsRDB1, corroborated by the low RMSF values for residues in this region from the MD simulation study. The global tumbling time (τ_c) of dsRBD2 was 6.90 ns, and that of dsRBD1 was 7.64 ns suggesting a more compact structure of dsRBD2 than dsRBD1. This result corroborates the CD-based determination of the melting point of the two constructs. While TRBP2-dsRDB2 exhibited a higher T_m (55°C), TRBP2-dsRBD1 melted at 45°C (data not shown), thus suggesting a stronger network of interactions in TRBP2-dsRDB2 (21). Indeed, it has been shown previously that the tryptophan (W173) present in the α_1-β_1 of dsRBD2 (W is absent in dsRBD1 at this position) induced local hydrophobic and cation-π interactions with K171, E199, R200, F201, and V225, thereby enhancing the overall thermal stability of dsRBD2 (21).

Only 4 residues of dsRBD2 could fit into a model-free model containing the exchange term (R_{ex}) as opposed to 11 residues in dsRBD1, suggesting that there is significantly lower conformational flexibility at the μs-ms timescale in dsRBD2. While dsRBD2 depicted a low frequency (< 2 Hz) contribution to the line width of resonances in the RNA-binding regions (E166, N216, A219, and L223), dsRBD1 had a larger contribution (2–8 Hz) to the line width of resonances present all along the backbone (Figure S5). In summation, the plasticity at the

ps-ns timescale is present in both the dsRBDs of TRBP, while the amplitude of motions is found to be largely restricted for the dsRBD2.

Similar to dsRBD1, dsRBD2 does not exhibit motions at a slower ms timescale as probed by the CPMG relaxation dispersion experiments. No dispersion was observed in the $R_{2\mathrm{eff}}$ rates with increasing ν_{CPMG}, suggesting the absence of motions sensitive to this experiment (Figure S6) (32).

The heteronuclear adiabatic relaxation dispersion (HARD) NMR experiment was used to study NMR spin relaxation in a rotating frame. The dispersion in relaxation rates is created by changing the shape of a hyperbolic secant (HSn, where n = stretching factor) adiabatic pulse that is used to create the spin-lock. HARD experiments are sensitive to the conformational exchange processes occurring on the 10 μs–10 ms timescales. The $R_{1\rho}$ and $R_{2\rho}$ relaxation rates showed that with the increase in applied spin-lock field strength from HS1 to HS8, the $R_{1\rho}$ rates increased and the $R_{2\rho}$ rates decreased for both dsRBD1 and dsRBD2 (Figure 3). The extent of dispersion was much less in the core residues of dsRBD2, indicating the absence of higher conformational dynamics in the dsRBD2. The $R_{1\rho}$ and $R_{2\rho}$ rates were then fit to Block-McConnel equations by using the geometric approximation approach (52) to extract the rate of exchange (k_{ex}) and the excited state population (p_{B}) (Figures 4A and 4C). The number of residues with $k_{\mathrm{ex}} > 5{,}000$ Hz and > 10% p_{B} (red and green, medium and big spheres) differed vastly between the two domains. Only 3 residues with such conformational exchange in dsRBD2 were identified: R200 (loop 3), L212 and R224 (α_2); whereas in dsRBD1, the number is 15, distributed along the entire backbone of the protein (Figure 6 of (32)), similar to PKR-dsRBD1, where 75% of the residues from dsRBM1 showed $R_{\mathrm{ex}} > 1$ Hz (19). Interestingly, all three conserved RNA-binding regions in dsRBD1: α_1 (L35, Q36), β_1-β_2 loop (E54), and α_2 (N61) were brimming with a $k_{\mathrm{ex}} > 50{,}000$ Hz. To sum it up, most of the dsRBD2 core showed low conformational exchange ($k_{\mathrm{ex}} < 5000$ Hz), depicted in blue small

spheres, while the dsRBD1 domain was undergoing significantly higher conformational exchange (k_{ex} > 5000 Hz). Taken together, the intrinsic k_{ex} profile of TRBP2-dsRBD1 and TRBP-dsRBD2 revealed the presence of 10 μs–10 ms timescale conformational dynamics distributed throughout the core dsRBD rather than being localized in the RNA binding regions; however, the amplitude of motions was found much smaller in dsRBD2.

A similar extensive fast and slower timescale conformational dynamics has also been studied in the case of DRB4 protein (24), where the first domain was found to have a large conformational exchange when compared with that of the second domain. The conformational plasticity of dsRBD1 enables it to initiate primary interaction with various structurally and sequentially different dsRNAs in the TRBP2 protein (32).

dsRBD2 has a higher affinity for a short double-stranded A-form RNA duplex

The NMR investigation of the interaction between the TRBP-dsRBD2 and the four miR-16-1 mutants (22–23 bp) dsRNAs revealed subtle structural perturbations in TRBP2-dsRBD2 (Figure S7). These changes were indicated by minute chemical shift perturbations (< 0.1 ppm) in the presence of RNA, consistent with previous findings by Yamashita et al., where authors compared the solution structure of TRBP-dsRBD2 structure with the crystal structure of GC10RNA-bound TRBP-dsRBD2 (21, 53). There were two intriguing observations in the ^{1}H-^{15}N HSQC-based titrations. First, the amide signals were getting broadened at as low as 0.1 RNA equivalents suggesting the slow-to-intermediate timescale of binding as has also been observed previously by our group and other research groups in TRBP2 dsRBD1, Staufen dsRBD3, hDus2 dsRBD, MLE dsRBD2, DBR4 dsRBD1, PKR dsRBD, and Dicer dsRBD (23, 24, 32, 54–58). Second, not only the reported RNA-binding residues but the entire backbone was undergoing line broadening, suggesting the presence of RNA-induced motions in the entire backbone. Since the protein is not saturated with the

RNAs at 0.1 RNA equivalents (considering a reported K_d = 1.7 μM for a 22 bp dsRNA (59), [protein] = 50 μM, [RNA] = 5 μM, fraction bound of protein < 10%) we can rule out an increase in the size due to the formation of a stable protein-RNA complex causing line-broadening. Upon excess addition of RNA (to 2 equivalents of miR-16-1-M), the backbone amide signals were not recovered, indicating the RNA-protein interaction does not seem to come out of the local minima of slow-to-intermediate exchange regime (Figure S7).

The amide NMR signals were, however, partially recovered by shortening the length of the RNA duplex to 12 bp D12 RNA (60, 61). The line broadening was delayed till 0.35 equivalents of D12 RNA as compared to 0.1 equivalents for the longer RNAs (Figure S8). The excessive line broadening all along the backbone and recovery of the same by reducing the length of the RNA hints towards the presence of the phenomenon of protein sliding over the length of the RNA, as has been reported for Loqs-PD dsRBD and MLE dsRBD (56, 62–64). To extract the binding constant (K_d) for the D12-TRBP-dsRBD2 interaction, residue-wise peak intensities were plotted against the RNA concentration and tried to fit to the binding isotherm for one-site binding (Figure S9). Due to extensive line broadening, there was a lack of data at the inflection point, affecting the data fitting. Hence the fitted parameters had large errors and remained inconclusive.

The ITC-based study was performed with D12 duplex RNA to get the apparent RNA-binding affinity (K_d) of the two N-terminal dsRBDs, thermodynamic parameters (ΔH, ΔG, TΔS), and stoichiometry (n). Our studies showed that TRBP2-dsRBD2 binds to D12 RNA relatively strongly with a K_d of 1.1 ± 0.37 μM as compared to TRBP2-dsRBD1 (Figure 5), which did not show significant heat exchange during the titration, and data could not be fitted to any binding model. The differential potential against the time plot indicates that RNA got saturated with TRBP2-dsRBD2 as early as the eighth injection. The integrated change in injection enthalpy (ΔH) versus the molar ratio of the reactants yielded an ΔH of –10.15 ± 0.5

kcal/mol, suggesting an enthalpy-driven binding event. ITC study was repeated for TRBP2-dsRBD1 with as high as 22 folds excess of protein, but RNA remained unsaturated. These results strongly suggest that TRBP2-dsRBD2 binds more efficiently to a small perfect A-form duplex, D12 RNA, than TRBP2-dsRBD1.

Dynamics perturbations of TRBP2-dsRBD2 at various timescales in the presence of RNA ligand

Minute structural changes in TRBP2-dsRBD2 in the presence of RNA necessitated a deeper insight into the perturbations of conformational dynamics at multiple timescales. In the presence of D12 RNA, the average R_1 rates (apo = 1.44 ± 0.05 Hz; bound = 1.37 ± 0.07 Hz) and nOe values (apo = 0.75 ± 0.029; bound = 0.72 ± 0.076) remained unperturbed, while there was a significant increase in the average R_2 rates (apo = 11.02 ± 0.39 Hz; bound = 20.04 ± 1.11 Hz) (Figure 6). The apparent increase in the R_2 rates (= R_2* + R_{ex}) hinted towards a perturbation in the μs-ms timescale dynamics, to which only R_2 rates are sensitive. This phenomenon could be attributed to either an increase in intrinsic R_2* resulting from an increase in the residence time of the RNA on the protein, or an increase in the R_{ex} component caused by a chemical exchange between apo- and D12-bound state, or RNA-induced conformational exchange in the protein. Interestingly, such a perturbation in R_2 rates was not found for the dsRBD1 upon RNA-binding (32). The terminals (N159, Q154, Q155, S156, E157, V225, V228, L230, A232), a few residues in the vicinity of loop regions (E177, E183, G185, E191, S209) harbored a lower than average bound-state R_2 rates than the rest of the core indicating a faster motion induced in these residues. On adding RNA, the R_2 rates and nOe values of the N- and C-terminal residues (S156, E157, T227, V228, L230, A232), β1 (E177), the loop 2 region (E183, G185, 189, E191), and the loop 4 region (S209) decreased, indicating further enhanced flexibility.

The extended model-free analysis of apo and RNA-bound TRBP2-dsRBD2 suggested that the anisotropic (ellipsoid) diffusion model was the best fit for the global motion of them both. The global tumbling time (τ_c) of core TRBP2-dsRBD2 in the presence of RNA was 10.9 ns as against 6.90 for the apo-dsRBD2 core, indicating an apparent increase in molecular weight. The overall higher S^2 values for TRBP2-dsRBD2 indicated a rigidification of the backbone amide vectors in the presence of D12 RNA (Figure S10). A few residues in the α1 (E166) and α2 (N216, A219, L223) regions exhibited a R_{ex} component > 1 Hz, lying in the vicinity of the reported RNA-binding regions 1 and 3. The number of residues with a significant (> 2 Hz) R_{ex} — implying the presence of μs-ms timescale motions — increased in the case of bound-state. R_{ex} was found to be induced in the α1 (V168), β1 (Y178), β2 (T195), β3 (I204), and α2 (N216, L223) region, thereby indicating that not only the RNA-binding residues but the rest of the core might play a role while interacting with dsRNA substrates. The presence of R_{ex} at multiple sites (in addition to RNA-binding regions) rules out the chemical exchange between apo- and D12-bound state contributing to R_{ex}, thereby implying a presence of RNA-induced conformational exchange is the predominant contributor to R_{ex} and to increased line broadening (or apparent R_2 rates, discussed in the NMR-based titration paragraph).

The effective transverse relaxation rates, R_{2eff}, for TRBP2-dsRBD2, were plotted against the CPMG frequencies (ν_{CPMG}) in the presence of D12 RNA (Figure S11). None of the residues in either condition showed significant dispersion (> 5 Hz) in the R_{2eff} rates with the increase in ν_{CPMG}, suggesting motions in the timescale sensitive to this experiment remain absent in D12RNA-bound TRBP2-dsRBD2.

Most importantly, HARD NMR experiments were recorded for TRBP2-dsRBD2 in the presence of D12-RNA and showed fascinating results (Figure 4 and S12). In the presence of RNA, there was massive induction of 10 μs–10 ms timescale conformational dynamics as

reflected by the increase in the number of residues having significant k_{ex}. The extent of enhancement in k_{ex} varied along the backbone of the core protein. For incidence, a k_{ex} of 5000–50,000 kHz with 10–20% p_B (green spheres) was observed in α_1 (G162, L167), loop3 (R200), and α_2 (A213, A219) and 20–40% p_B in α_1 (V168), and β_3 (E203), depicted by the medium and big green spheres respectively (Figures 4B and 4D). The residues V198 in loop 3 and M221 in the α_2 region exhibited the presence of the highest frequency of motion with a k_{ex} > 50,000 Hz. Intriguingly, among these residues, L167, V168 (α_1), and A213 (α_2) lie in the reported RNA-binding regions 1 and 3, respectively. L167 and V168 are adjacent to the key RNA binding residue E165 (region 1), which interacts directly with the RNA minor groove. A213 precedes the important KR-helix motif in the α_2 region of dsRBD2. The K214 and R215 residues make ionic interactions with the negatively charged phosphate backbone of the RNA major groove. The interaction between the RNA and the RNA-binding residues of the protein might induce conformational exchange within the nearby residues like in the case of G162 (region 1), A219 and M221 (region 2). Additionally, residues like R197 (β_2); V198 and R200 (L3); E203 and G207 (β_3) are further from RNA binding regions with k_{ex} > 5000 Hz. Thus, the ligand affected not only the RNA-binding residues but the entire protein.

Notable perturbations in terms of Δk_{ex} ($k_{bound} - k_{apo}$) > 10,000 Hz were mapped on the structure of the protein (Figure 7). There was an enhancement of k_{ex} in the residues lying in α1 (G162, L167), loop 3 (V198), end of β3 (G203), and α2 (A213, A219) region. Concomitantly, suppression of exchange was observed in β_1 (V180), loop 3 (R200), and α_2 (A212, A224) regions. A tantalizing relay of exchange was seen between two residue pairs in close spatial proximity. For instance, exchange was induced in V198 and quenched in R200 in the loop 3 region. Similarly, in the case of A213 and L212 falling in the N-terminal RNA-binding region 3, and A219 and R224 lying in the C-terminal of α_2, the former of the pairs underwent an increase in k_{ex} while the latter observed a decrease. Thus, the enhancement of

RNA-induced conformational exchange was accommodated by allosteric quenching of the same. Altogether, there was a significant induction of 10 μs–10 ms timescale motions over the entire backbone of TRBP2-dsRBD2 in the presence of RNA. This induction of motions could be ascribed to either an exchange between the apo- and RNA-bound state of the protein, conformational dynamics in the RNA-bound state of the protein, or the previously reported sliding motion of the protein on the dsRNA (56, 62–64). Interestingly, in TRBP2-dsRBD2, the extent of dynamics perturbation, in terms of Δk_{ex}, was significantly lower (10–50 kHz) than TRBP-dsRBD1 where at least 10 residues underwent a $\Delta k_{ex} > 50$ kHz in the presence of small dsRNA. Moreover, a relay of exchange was evident only in loop 3 and α_2 regions of dsRBD2 and not in α_1 region, which was observed earlier in dsRBD1. Summing up, in the presence of RNA, dsRBD2 undergoes enhanced conformational exchange in the 10 μs–10 ms timescale. When compared to dsRBD1, dsRBD2 samples limited conformational space both in the absence and presence of D12-RNA.

Thus, our study has led us to propose a model involving two tandem type A dsRBDs in TRBP2. According to our model, these dsRBDs work synergistically to recognize, bind, and assist associated proteins (like Dicer) in processing the incoming dsRNA ligand (Figure 8). The first dsRBD, dsRBD1, has remarkable flexibility, which allows it to recognize a wide range of structurally and sequentially diverse dsRNAs. On the other hand, dsRBD2 firmly adheres to the RNA ligand due of its conserved RNA-binding stretches and relatively high rigidity. During their interaction with the RNA, both dsRBDs experience enhanced conformational changes on a fast timescale (picoseconds to nanoseconds) and a moderate timescale (microseconds to milliseconds), as observed through nuclear spin relaxation and rotating frame relaxation dispersion measurements. These dynamic changes likely enable the dsRBDs to slide along the length of the RNA, which explains the broadening of the signals observed during titration with longer RNAs. This sliding movement is likely crucial for other

associated proteins, such as Dicer (65–67), to carry out precise microRNA biogenesis. By understanding these processes, we can gain insights into the intricate mechanisms involved in RNA processing, which has implications in different biological processes.

CONCLUSION

It has been long established that despite engaging with a wide array of dsRNA molecules exhibiting diverse structures and sequences, dsRBDs remain unaffected in their own structural conformation. However, the more significant question that has persisted is whether protein dynamics itself plays a crucial role in these intriguing interactions. Though variability in the binding affinity for the two type-A dsRBDs of TRBP has been reported, the specific contribution of conformational dynamics has remained unexplored until now. In the present investigation, we have exclusively delved into the role of conformational dynamics of the two type-A dsRBDs of TRBP2 in RNA recognition and binding. Our findings have unveiled that TRBP2-dsRBD2 samples a limiting conformational space in solution, and it is dsRBD1 that recognizes sequentially and structurally diverse RNA substrates through its high conformational plasticity. While dsRBD1 explores and engages dsRNA via its dynamically interactive RNA-binding surface, dsRBD2 holds the RNA in position via stronger canonical contacts. Once bound to the RNA, the ensuing conformational exchange in both dsRBD1 and dsRBD2 might facilitate the domains to diffuse over the length of the RNA freely, thus playing a pivotal role in assisting Dicer-mediated differential cleavage of RNA. Thus, this study not only adds valuable insights into the mechanics of RNA-protein interactions but also underscores the significance of conformational dynamics in dictating the functional outcome in such intricate biological processes.

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AUTHOR CONTRIBUTIONS

J.C. and F.P. designed the study, the approach to characterize the TRBP2-dsRBD2 in apo and RNA-bound state. F.P. performed all the biophysical experiments related to the TRBP2-dsRBD2 protein. F.P. performed all the analysis in active discussion with J.C. D.S. carried out the MD simulation studies. Z.A. and F.P. purified the TRBP-dsRBD1 protein and carried out ITC experiments. H.P. recorded and analyzed dynamics data on dsRBD1. F.P. contributed to the literature survey. F.P. and J.C. have contributed to manuscript writing. All the authors have read and approved the manuscript.

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FIGURES

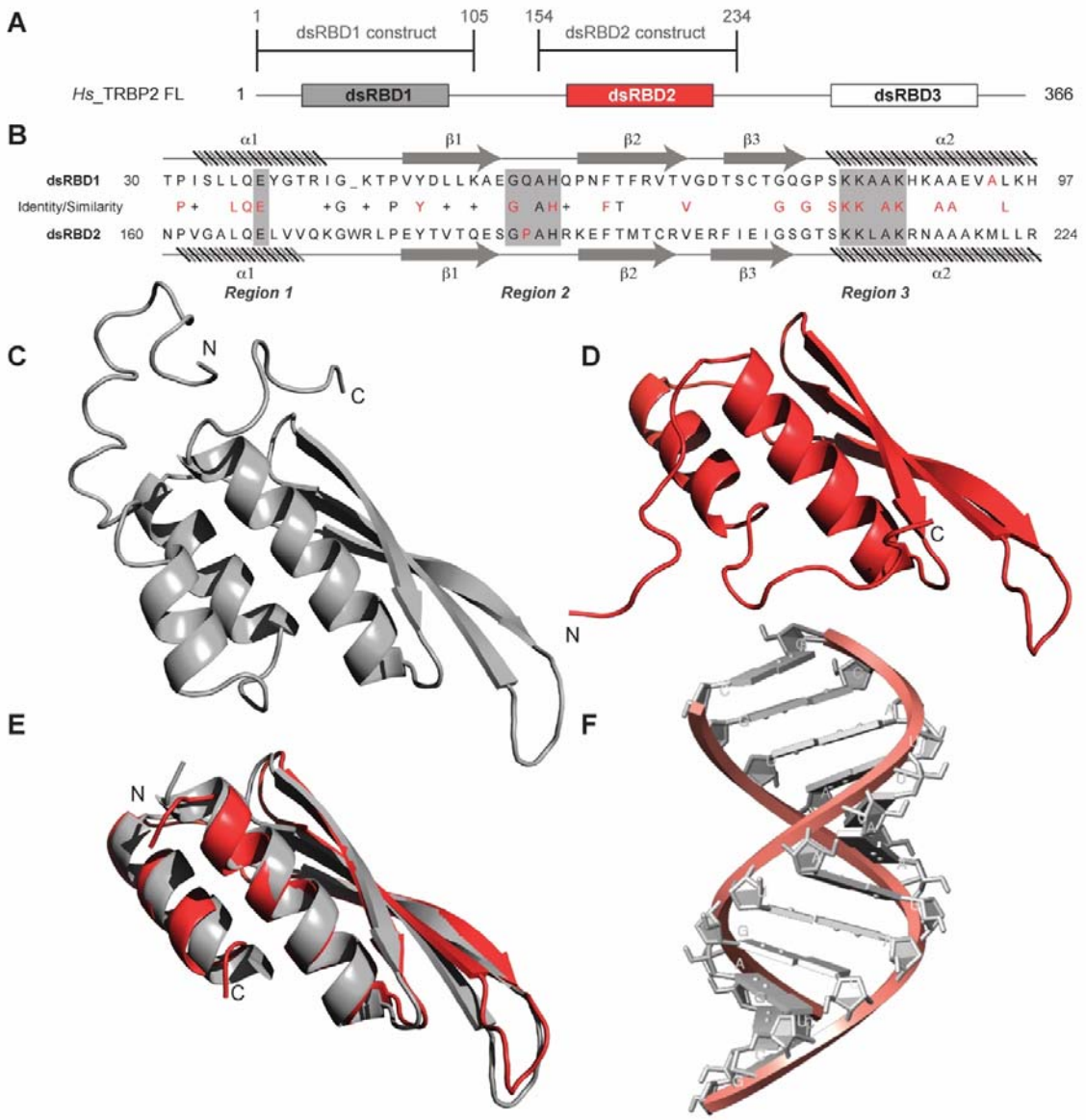

Figure 1: (A) dsRBD constructs [TRBP2-dsRBD1 (1–105 aa) and TRBP2-dsRBD2 (154–234 aa] of Human TRBP2 full-length protein used in this study. (B) Sequence alignment of the two constructs mentioned in (A). CS-Rosetta structures of (C) TRBP2-dsRBD1 (33), (D) TRBP2-dsRBD2, (E) an alignment of core residues of dsRBD1 and dsRBD2, (F) Model structure of 12 bp duplex D12 RNA.

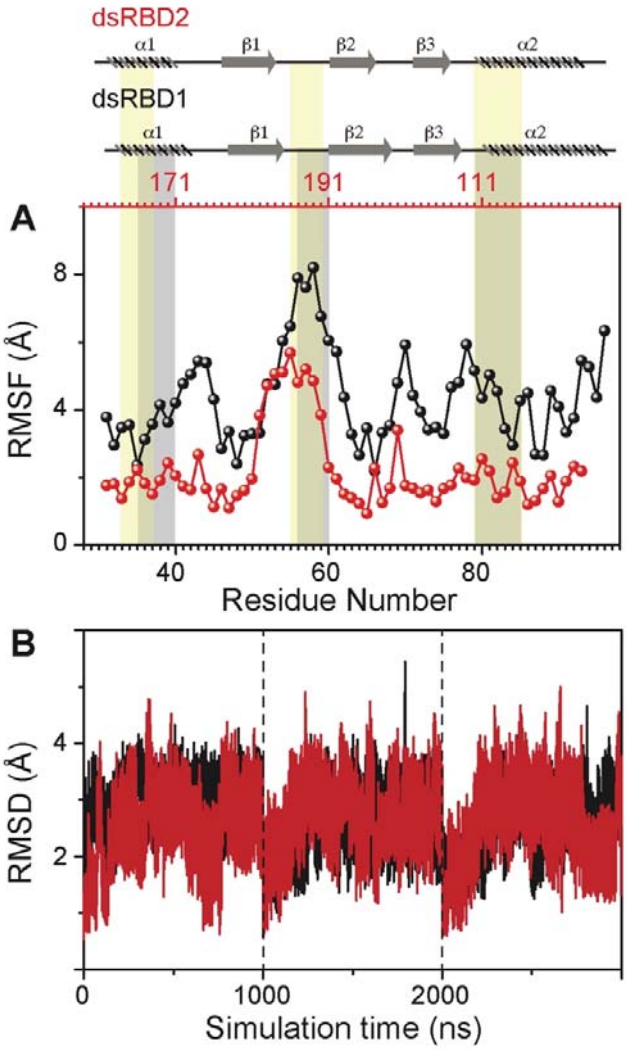

Figure 2: (A) RMSF profile (by C_α) of dsRBD1 (black) and dsRBD2 (red). The secondary structure for the two domains has been shown on the top, and three RNA-binding regions in dsRBD1 and dsRBD2 have been highlighted using vertical grey and yellow bars, respectively. (B) RMSD of profiles of dsRBD1 (black) and dsRBD2 (red) over 1 μs simulation time. RMSD values for data measured in triplicate have been separated by vertical dashed lines.

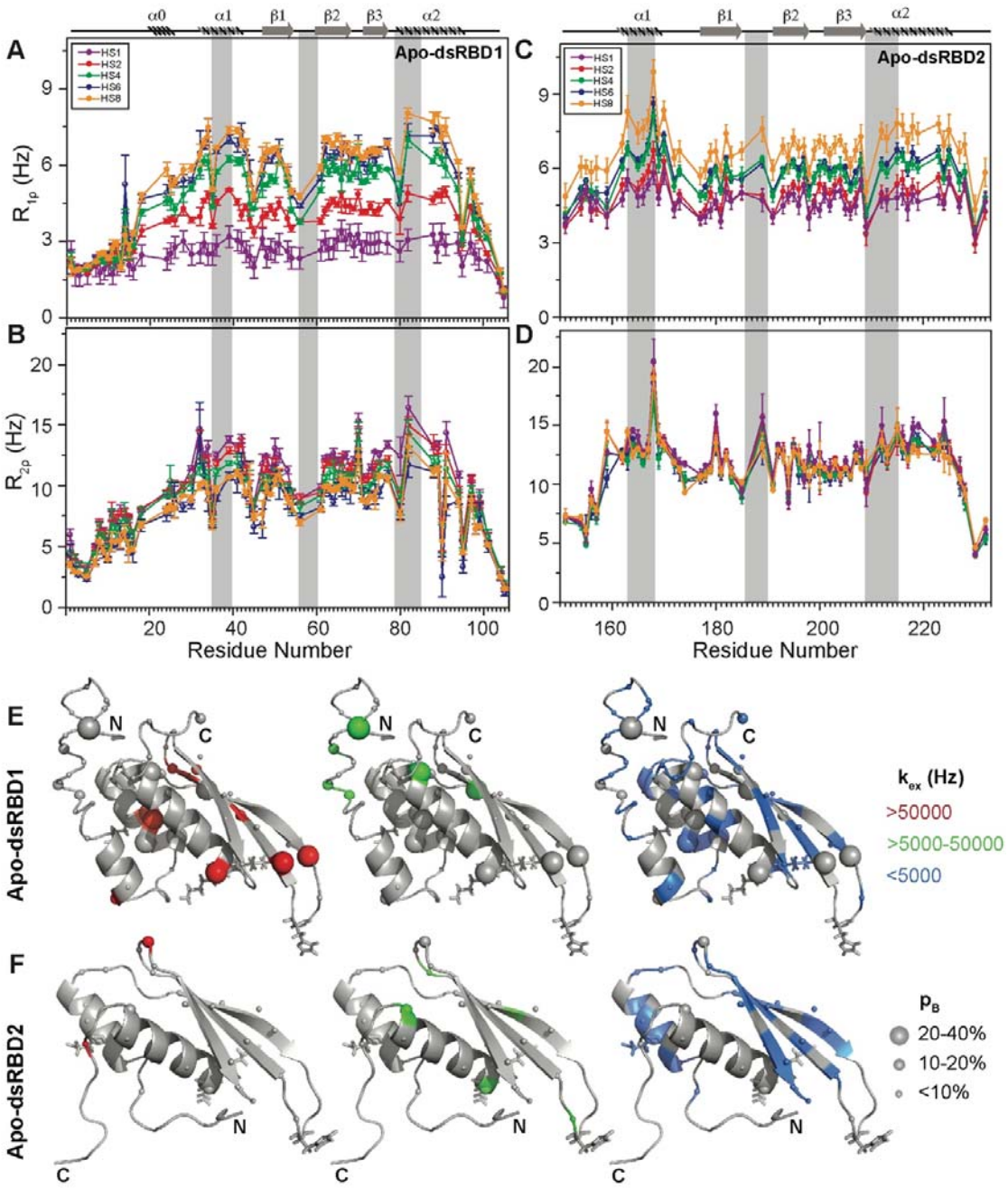

Figure 3: (A) $R_{1\rho}$ data, (B) $R_{2\rho}$ data, recorded on apo ^{15}N-TRBP2-dsRBD1, (C) $R_{1\rho}$ data, (B) $R_{2\rho}$ data, recorded on apo ^{15}N-TRBP2-dsRBD2 using Heteronuclear Adiabatic Relaxation Dispersion (HARD) experiments on a 600 MHz NMR spectrometer, plotted against residue numbers. An increase in the spin-lock field strength is achieved by an increase in the stretching factor of the adiabatic pulse used to create the spin lock, n (in HSn). The secondary structure has been depicted on the top, and three RNA-binding regions have been highlighted using vertical grey bars. Mapping of conformational exchange parameters (rate of exchange

between the ground state and excited state (k_{ex}), and excited state population (p_B)) obtained by fitting the above-described data to a two-state model on the CS-Rosetta structures of (E) apo-dsRBD1, and (F) apo-dsRBD2. Residues have been marked in different colors to highlight the distribution of k_{ex} values, and the diameters of the sphere indicate the extent of p_B along the protein backbone. The RNA-binding residues have been depicted in stick mode.

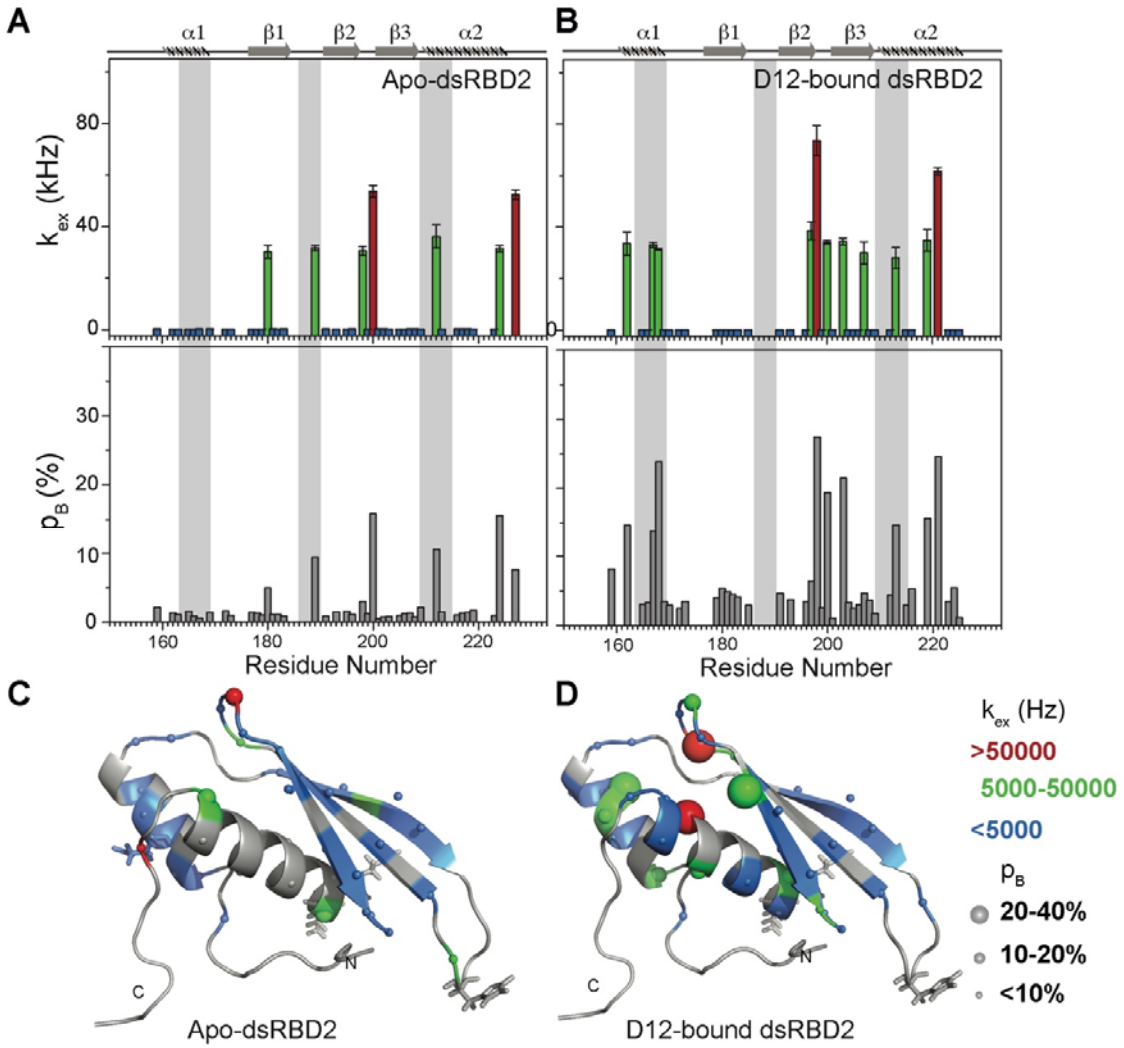

Figure 4: Conformational exchange in (A) apo- and (B) D12-bound- TRBP2-dsRBD2. Top panel: Rate of exchange between the ground state and excited state (k_{ex}); Bottom panel: excited state population (p_B) as obtained by the geometric approximation method, using the HARD experiment, plotted against residue numbers. Mapping of k_{ex}, and p_B, on the CS-Rosetta structure of apo TRBP2-dsRBD2, as extracted for (C) apo TRBP2-dsRBD2 and (D) D12-bound TRBP2-dsRBD2. Different colors highlight the distribution of k_{ex} values, and the sphere's diameter indicates the extent of p_B along the protein backbone. The RNA-binding residues have been depicted in stick mode.

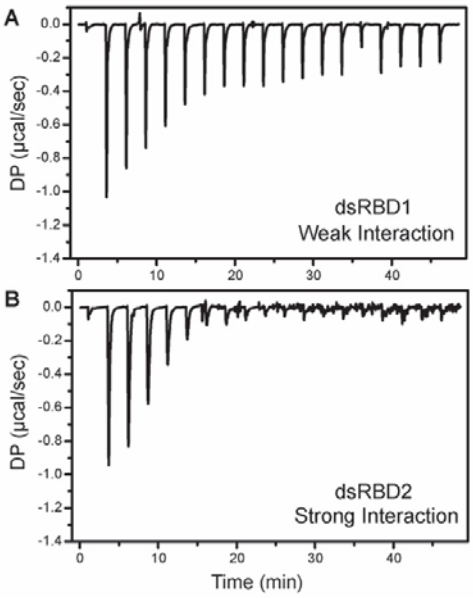

Figure 5: ITC-based binding study of D12 duplex RNA with (A) TBRP2-dsRBD1 and (B) TRBP2dsRBD2. The raw differential potential for each injection has been plotted against the titration time.

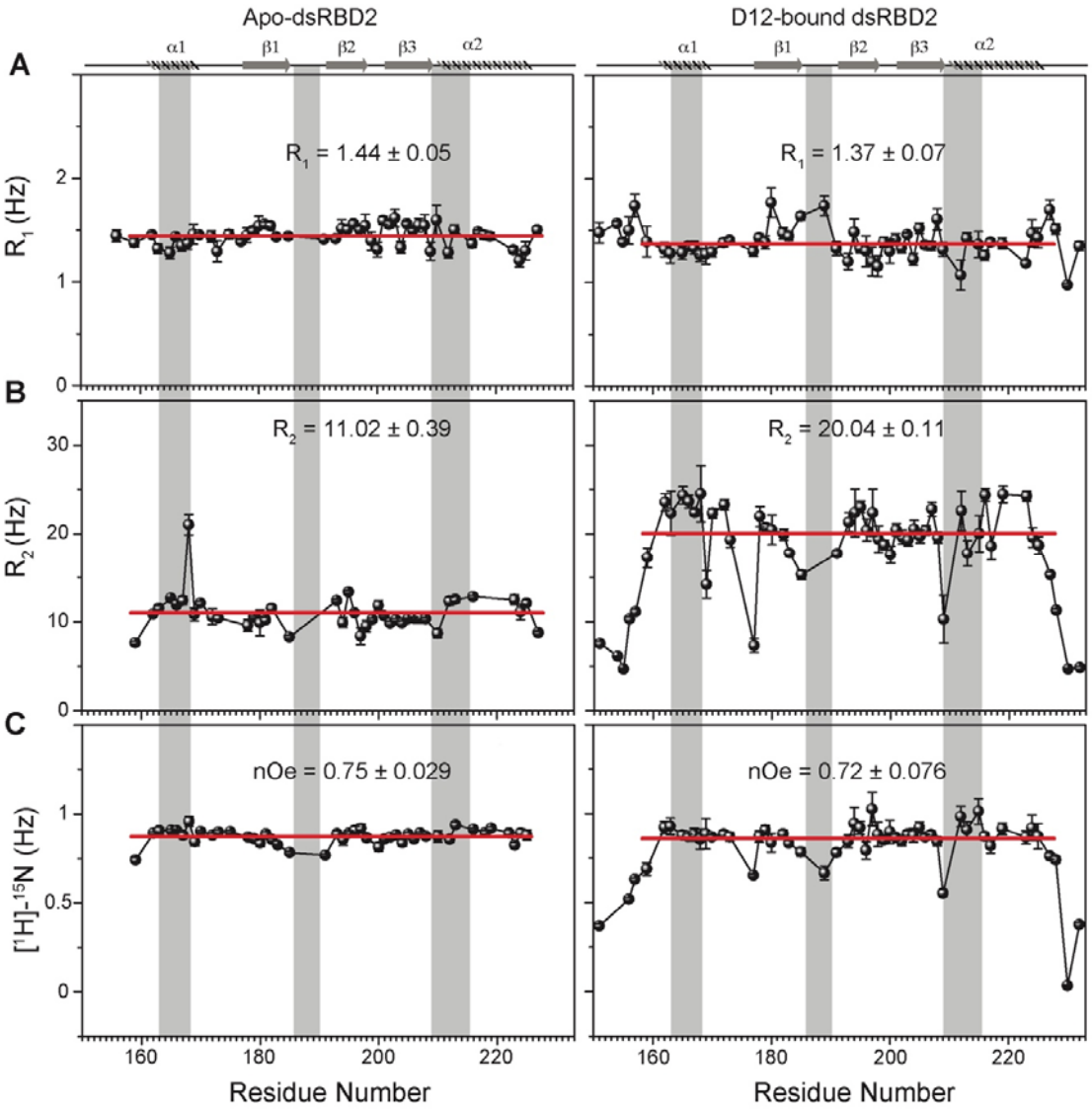

Figure 6: Spin relaxation parameters (A) R_1, (B) R_2, and (C) $[^1H]$-^{15}N nOe plotted against residue numbers for (left panel) apo TRBP2-dsRBD2 and (right panel) D12-bound TRBP2-dsRBD2. The secondary structure of TRBP2-dsRBD2 has been mentioned on the top, and the RNA-binding region of the protein has been marked in grey vertical columns.

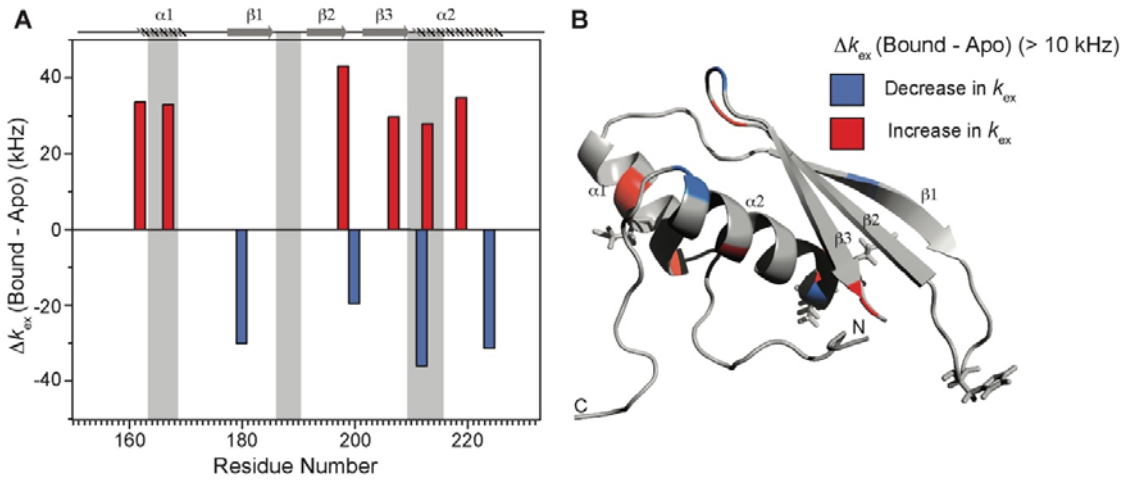

Figure 7: Conformational exchange perturbations in TRBP2-dsRBD2 in the presence of D12 RNA. (A) Δk_{ex} (D12-bound – apo) TRBP2-dsRBD2 plotted against residue numbers. The secondary structure has been shown on the top, and three RNA-binding regions have been highlighted using vertical grey bars. Only residues having significant perturbation (Δk_{ex} > 10 kHz) have been plotted, where an increase is shown in red, and a decrease is shown in blue, (B) An increase in k_{ex} (red) and a decrease (blue) in the presence of D12 RNA indicated on the backbone of the CS-Rosetta structure of apo TRBP2-dsRBD2.

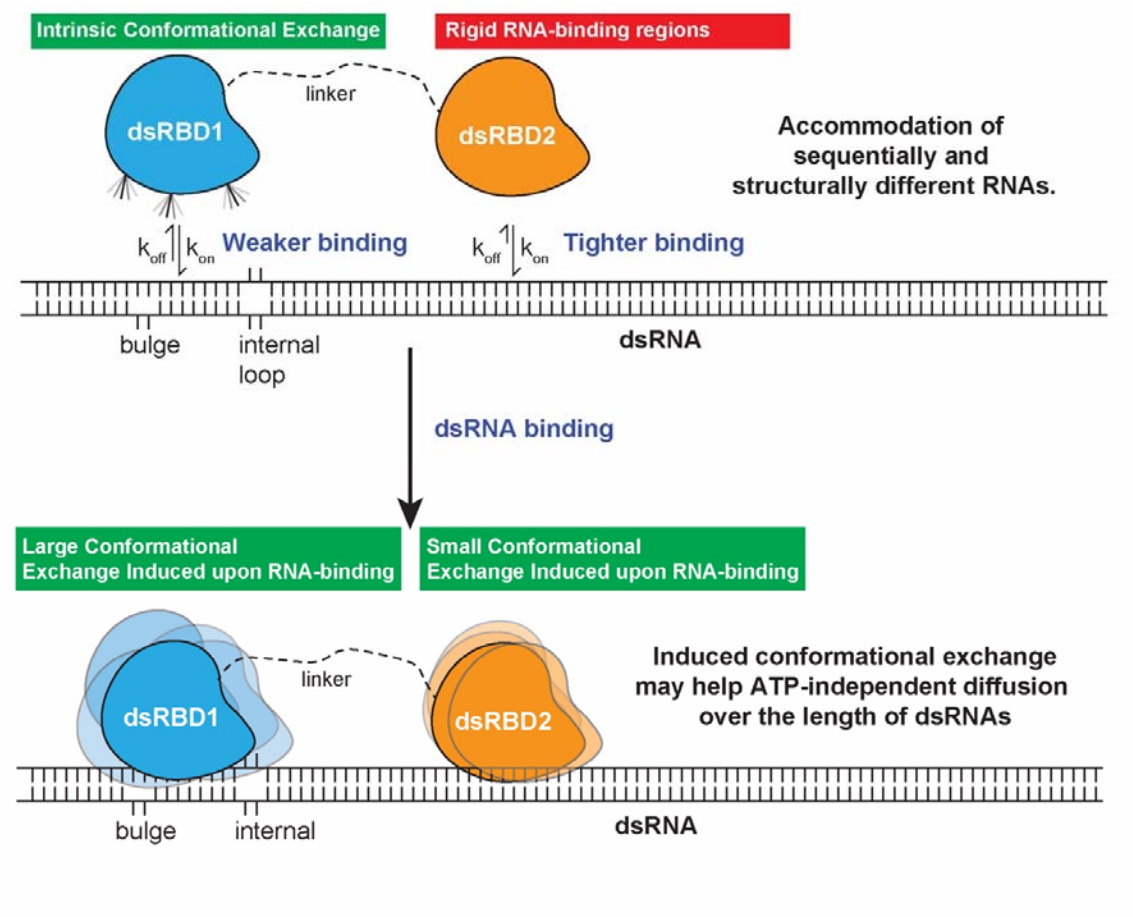

Figure 8: The model proposed for the two type-A dsRBDs in TRBP2 protein. dsRBD2 with rigid and conserved RNA binding regions is able to bind the RNA tightly, whereas dsRBD1 with high conformational exchange is able to recognize different RNA structures (often with bulges and internal loops). Following this, the two dsRBDs upon contacting the RNA undergoes enhanced conformational exchange at different extent. This enhanced conformational exchange coupled with differential binding affinity towards dsRNA might enable the tandem dsRBDs to move along the backbone of the RNA molecule, leading to ATP-independent diffusion.