Mapping the landscape pool of ardeid species to landscape structure associated with Japanese
encephalitis virus in Australia

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Abstract

The geographic extent of Japanese encephalitis virus (JEV), a zoonotic, mosquito-borne virus,
increased dramatically in south-eastern Australia over the summer of 2021-2022 resulting in
outbreaks in piggeries, 45 cases of human disease, and simultaneous detection of JEV by mosquito
and arbovirus surveillance programs. Preliminary investigation of the landscape epidemiology of the
piggery outbreaks showed that they were associated with particular landscape structure as well as
intermediate ardeid species richness. However, no exploration of the ways in which individual
species or species composition may couple with JEV-associated landscape composition and
configuration has been undertaken and, therefore, key questions regarding the landscape
epidemiology and infection ecology of JEV in Australia remain unanswered. The current study sought
to interrogate how individual species presence and their functional traits were distributed across the
landscape, and how these distributions aligned with the landscape structure of JEV occurrence.
Inhomogeneous Poisson point process models and multivariate hierarchical models were used to
investigate the landscape-level ardeid species pools and the distribution of their pool-weighted traits
with respect to JEV outbreaks. The distributions of six species were strongly associated with the
distribution of JEV outbreaks reported in 2022: *Casmerodius albus* (*Ardea alba*), *Ardea pacifica*,
Botaurus poiciloptilus, *Egretta novaehollandiae*, *Ixobrychus minutus* (*Ixobrychus dubius*), and
Nycticorax caledonicus. Some JEV-delimiting landscape features, including transient wetlands and
proximity to waterways, were associated with the presence of nearly all ardeid species, while others,
such as crop cultivation, habitat fragmentation, and water flow accumulation were associated with
only a subset of these species. In addition, hand-wing index was associated with species presence in
landscapes associated with JEV outbreaks, which may suggest the importance of dispersal capacity in
ardeid community assembly in landscapes that share environmental features with JEV occurrence.
This study provides a preliminary selection of target species for the development of wild waterbird
JEV surveillance, while uniquely accounting for sympatry and the potential for functional modulation
of environmental filtering in high-risk landscapes, despite the broad landscape scale that necessarily
constrained the analysis. Nevertheless, while the results provide an actionable One Health
framework with which to develop wildlife surveillance, it must be noted that they do not describe
community assembly rules, nor do they provide a description of community composition at local
scale. The findings advance the current understanding of the landscape epidemiology of JEV in
Australia by connecting landscapes with individual species from a family of established maintenance
hosts and, given the large geographic extent of the potential circulation of JEV in maintenance hosts,
provides valuable evidence-based information for targeted surveillance strategies.

Introduction

In early 2022 Japanese encephalitis virus (JEV) was detected for the first time in south-eastern regions of Australia, resulting in the declaration of a Communicable Disease Incident of National Significance. A total of 45 cases of human disease were reported, including seven fatalities[1]. Outbreaks were also reported in piggeries with unprecedented geographic distribution, high numbers of affected piggeries, and high incidence within piggeries [2–4]. Previous JEV occurrence in Australia had been limited to isolated cases in the Torres Strait Islands[5] and the Cape York Peninsula of northern Queensland[6]. The extensive geographic distribution of these outbreaks, which included eastern and southern Australia, was of particular concern because it deviated markedly from anticipated emergence events for JEV[7] and suggested extensive circulation of the virus in maintenance communities prior to emergence in pigs and humans[8]. Furthermore, these outbreaks followed an extensive period of anomalous weather due to three consecutive La Niña phases of the El Niño Southern Oscillation (ENSO). The anomalies associated with these shifts included high levels of precipitation and major flooding events across eastern Australia, generating considerable disturbance among natural communities[9]. It has been hypothesised that the anomalous La Niña-associated weather patterns may have contributed to the widespread distribution of JEV outbreaks in 2022 due to modulation of the distribution of vector populations or maintenance host populations[8]. Notably, an extensive investigation of landscape composition and configuration and La Niña-associated anomalies showed that landscape structure was substantially more influential to outbreak occurrence than were specific precipitation or temperature anomalies associated with La Niña[10]. However, certain aspects of high-risk landscape structure, such as the composition of transient wetlands and the propensity to flooding, would also have been affected by both proximal and distal precipitation events, and so La Niña-associated weather anomalies may have influenced outbreak occurrence indirectly, particularly if the influence varied by scale.

Japanese encephalitis virus circulates in wild ardeid bird maintenance hosts[11–17] vectored by mosquitoes (primarily *Culex annulirostris* in Australia[18]) and can spillover to pigs[19–24] and

humans, which are competent and non-competent hosts, respectively. Previous work has examined

the distribution of ardeid species richness as a key nidus of the maintenance community, identifying

a strong quadratic relationship between richness and JEV piggery outbreak occurrence, whereby

intermediate richness was associated with greatest risk(10). Nevertheless, neither the association

between outbreaks and individual species, nor the influence of the structure and functional

distribution of species assemblages were assessed with respect to the 2022 piggery outbreaks in

Australia, and the latter has not been assessed with respect to outbreaks anywhere in the Indo-

Pacific region to date. Due to the logistics and resources required to sample birds across such vast

landscapes, a better understanding of how particular species and species community structure align

with the landscape structure associated with JEV outbreaks would be particularly useful to identify

target species to monitor JEV circulation, and other flaviviruses with avian reservoirs, in these

landscapes.

The study of the functional ecology of viral maintenance hosts has increased in recent years

as a means to help identify potential hosts based on specific biological and ecological traits,

frequently focusing on life history due to the association between fast-living and host competence in

many r-selected species(25–33). These species, which are typically generalist and frequently more

resilient to anthropogenic pressure(34–36), tend to invest more in earlier and more fecund

reproductive effort, and less in immune function and adult survival(37). As such, many of these

species also present as optimal hosts for pathogens(38). Nevertheless, the association between fast-

living species and host competence is by no means universal, with some systems demonstrating the

opposite or ambiguous associations, such as ebolaviruses(39), Rift Valley fever virus (RVFV)(40), and

Ross River virus (RRV)(41). In addition to life history, other host traits, including population density,

diet composition, and dispersal ability, can also be influential to the infection ecology of some

pathogens(35,36). Japanese encephalitis virus host functional ecology has hitherto gone unexplored

among ardeid species and thus represents a critical knowledge gap for this system.

The aims of this study were to identify preliminary target host species in specific landscapes

for One Health surveillance monitoring of JEV. Initially, we sought to determine what, if any, ardeid

species were distributed in close association with the distribution of JEV piggery outbreaks after

adjusting each species' presence for sympatric ardeid species. Secondly, the study interrogated how

the structure of species pools at landscape scale were coupled with landscape composition and

configuration known to be associated with JEV outbreaks. The study then explored the extent to

which pool-weighted species' trait means were coupled with these same JEV landscapes as well as

the individual species present therein.

Methods

<u>Data Sources</u>

Animal data

Australian piggery outbreaks (N = 54) reported to the World Organisation for Animal Health

(WOAH) by the Australian government between 19 January, 2022 and 4 April, 2022[42] have been

described in a previous epidemiological investigation[10]. Briefly, outbreak occurrence was defined

as one or more cases reported at the level of the piggery. An additional 11 independently-

documented JEV occurrence locations (eight piggery outbreaks reported by a commercial pork

producer, and 3 positive mosquito pools detected through the NSW Arbovirus Surveillance and

Mosquito Monitoring Program[43]) were used to externally validate model performance.

To model the habitat suitability of each of the 14 extant species of ardeid birds in mainland

Australia[44], 791,416 observations of these species recorded between 1 January, 2010 and 31

December, 2020 were acquired from the Global Biodiversity Information Facility (GBIF)[45]. The

phylogenetic tree of these species was obtained from the VertLife project using the Hackett

backbone and constructed based on the methods described by Jetz et al. (Jetz et al., 2012). Any

taxonomic binomial discrepancies between the VertLife tree and the named species obtained from

GBIF used for the species distribution models described below were resolved, with alternate

binomials appearing in parentheses in Table S1. Data obtained from the national pig herd dataset,

which is used in the Australian Animal Disease Spread Model, were used to quantify domestic pig

density and piggery density for the JEV models described below. This dataset comprises >8000

registered pig herds of all types (including commercial, boar studs, smallholder and pig keepers)(46).

Reporting bias can occur with respect to wild bird observations, especially differential

reporting between regional and remote locations compared with more accessible locations.

Therefore, this study selected background points proportional to the human footprint (HFP), a robust

indicator of landscape accessibility(47), to correct for such bias in the ardeid suitability models. The

HFP data were retrieved from the Socioeconomic Data and Applications Center (SEDAC) registry

maintained by the Center for International Earth Science Information Network (CIESIN)(48).

Reporting bias may also affect piggery outbreak data, whereby large commercial herds may be more

likely to capture and record cases due to more systematic collection of production data and

increased probability of disease detection in larger herds. Therefore, background points for JEV

models were selected proportional to herd size to correct for potential piggery reporting bias.

Environmental data for habitat suitability models

Land cover classification based on the European Space Agency and Climate Change Initiative

was used to classify wetlands(49,50) and land cover(51) at 3 arc seconds in 2010, as this time point

corresponded to the beginning of the period of recorded ardeid observations described above.

Waterways were represented by a separate high-resolution (3 arc seconds) image produced in

collaboration between CIESIN and the WorldPop project (52,53). Mean annual precipitation, mean

annual temperature, and isothermality were obtained from WorldClim (54). The Priestley-Taylor α

coefficient (P-Tα) is the ratio of actual evapotranspiration to potential evapotranspiration and was

used to describe water stress in the landscape(55,56). The P-Tα data were acquired from the

Consultative Group for International Agricultural Research (CGIAR) Consortium for Spatial

Information at a resolution of 30 arc seconds(57).

Environmental data for landscape-level metrics

All JEV outbreaks were reported to a scale of at least 5 arc minutes (~ 10 km), which thus

represents the landscape-level scale of the current investigation. Land cover classes were

represented using the previously validated Copernicus land cover data at 100 m resolution[58,59].

The Joint Research Centre's Global Surface Water product (based on Landsat 5, 7, and 8 imagery) was

used to represent high resolution (30 m) surface water seasonality in the year prior to piggery

outbreaks[60,61]. Each pixel in the raster tiles represents the total number of months (0 – 12) with

surface water present during 2021. The heterogeneity of water transience across the landscape was

based on five derived rasters: 1) permanent surface water presence over the 12 months of 2021, 2)

surface water absence over the 12 months of 2021, 3) surface water presence for 1 to 3 months of

the year, 4) surface water presence for 4 to 6 months of the year, and 5) surface water presence for 7

to 9 months of the year. Water movement through and accumulation in the landscape was

quantified using hydrological flow accumulation, which describes the total upland area draining into

each 500m by 500m parcel and was acquired from the Hydrological Data and Maps based on SHuttle

Elevation Derivatives at multiple Scales (HydroSHEDS) information system[62].

Ardeid trait data

Bird species' diet composition and body mass were obtained from the Elton Traits

database[63]. Diet composition may be an important characteristic of host species because this can

delineate generalist versus specialist species and potential resilience to human pressure[64]. Only

the dietary proportions of fish and invertebrates demonstrated heterogeneity among the ardeid

species, so only these dietary elements were investigated. Body mass is an important life history

characteristic as well as being a potential modulator of immune function[32,33]. Body mass was log

transformed prior to analysis. The TetraDENSITY database was used to obtain species mean

population density[65], the AVONET database was used to obtain the hand-wing index[66], and the

Lislevand bird traits database was used to obtain the reproductive characteristic, egg mass[67].

These species traits were included since population density may influence the dynamics of JEV

circulation in populations and communities(68–71), hand-wing index has been shown to be the best

metric to represent bird dispersal(72,73) particularly with respect to dispersal in response to climate

and habitat loss(74,75), and egg mass is an important indicator of reproductive life history. The

International Union for the Conservation of Nature (IUCN) Red List was used as a source of an

additional life history trait for each bird species: species' generation length (76). The data for

population density, hand-wing index, and egg mass were missing for 7, 3, and 3 species, respectively,

so a random forest machine learning algorithm, which was previously shown to be a robust approach

to this application(31,77), was used to impute the missing data. The rfImpute function in the

randomForest package was used to implement the algorithm(78).

<u>Statistical Analysis</u>

Species distribution modelling

All 14 species of ardeid birds extant in mainland Australia were modelled using the GBIF data

(Table S1). The modelling of habitat suitability for these species has been previously described(10).

Briefly, an ensemble of boosted regression trees (BRT), random forests (RF), and generalised additive

models (GAM), each with five-fold cross-validation, was used to model landscape suitability at fine-

scale (30 arc seconds). Observation data were thinned so that only one observation per pixel was

included in the analysis to avoid overfitting (Table S1). Environmental features in each suitability

model comprised mean annual temperature, mean annual precipitation, isothermality, P-Tα, and

proximity to surface water. Each model was evaluated according to performance and fit, using the

area under the receiver operating characteristic curve (AUC) and deviance, respectively. Suitability

estimates for each species were then derived from an ensemble of the three models (BRT, RF, and

GAM) using their AUC-weighted mean(79). Potential spatial sampling bias associated with the GBIF

observations was corrected by sampling background points proportional to HFP as an indicator of

accessibility, and thus occurrence reporting. All species' landscape suitability estimates were then

summed as an estimate of local species richness across the geographic extent of Australia.

Adjusted individual species presence at landscape-scale was computed by applying

interspecific biotic constraints at a scale of 5 arc minutes (~ 10 km), which was the scale of piggery

outbreak reporting as well as the scale at which key landscape structure influencing outbreak

occurrence was previously described(10). These biotic constraints were based on spatially-explicit

species assemblage modelling (SESAM) to adjust each species' habitat suitability estimate by

sympatric species (80–82). The SESAM framework is a frequently used approach to estimate

community composition from habitat suitability but is not intended to represent community

structure in the current study in which local communities were not observed. Here, the approach is

applied to construct a landscape-level species pool, given environmental filtering, which can then be

mapped to specific landscapes that are associated with JEV outbreaks.

The SESAM framework was applied as follows. First, each species' habitat suitability was

derived from the ensemble species distribution models described above. Second, a crude estimate of

species richness was calculated by summing all of the individual species' habitat suitabilities. Third,

each species is contrasted with all other species present within each landscape parcel to determine

whether a given species should be retained or excluded from the parcel. This final step uses the

probability ranking rule(81,83) to rank each species present in the parcel based on their suitability

estimate, which was computed in step one. Species are then selected for inclusion in the parcel

beginning with the species with the highest suitability and then continuing down the list of lower

ranked species until the sum of selected species equals the expected species richness value for each

parcel, which is based on the crude richness estimate obtained in step two. After attaining this

threshold sum, the SESAM adjusted-species richness has been achieved and no further species will

be included in that particular parcel. Subsequently, the species pool available for community

assembly was estimated with each species retained in a given landscape parcel under the SESAM

model as follows. Each 1 km^2 pixel of each species' habitat suitability estimate (step one, above) that

was greater than or equal to the true skill statistic (TSS)(84) was classified as present and summed

across all pixels within the 5 arc minute landscape parcel. The TSS was used to designate each parcel

as present or absent based on the habitat suitability because it has been shown to be independent of

prevalence(84). This summed metric thus provided an estimate for landscape-level species

"abundance" available to community assembly within each particular landscape parcel. Similarly,

species relative abundance was computed by dividing each individual species abundance by the total

abundance of all species for each parcel. The "relative abundance" of each species available thus

provides an estimate of the potential influence of a species for community assembly within

landscape parcels, rather than an actual estimate of relative abundance of a species in a local

community. This proxy of relative influence within the landscape-level species pool is the metric used

for identifying ardeid species targets for JEV surveillance in this study. The sdm package(79) was used

for fitting all species distribution models and deriving the habitat suitability ensembles. The SESAM

analysis was conducted using the ecospat package(82).

Because of the scaling up of species presence to compute proxies of abundance and relative

abundance at the landscape scale of 5 arc minutes, the analyses were limited to a single bird family,

the Ardeidae, to limit the degree of assumptions regarding environmental filtering, dispersal, and

biotic interactions of species between bird families. A narrower, single family-focus was considered

to be particularly necessary because of the direct examination of the associations between the

distribution of species pool-weighted mean traits (see description below), species composition, and

JEV risk, all at the broad landscape scale of JEV outbreak reporting. Although this was deemed an

acceptable compromise given that many of these species have been demonstrated to be competent

hosts, it is also recognised that additional non-ardeid waterbirds, such as the Australian white ibis,

have been recommended for JEV monitoring(8) despite the current lack of evidence regarding their

host status. As such, the current study's narrower focus limits the breadth of the conclusions to the

Ardeidae.

Landscape metric analysis

The metrics used to delineate high-risk landscape structure have been described in detail

previously in an analysis of the landscape composition and configuration associated with JEV

outbreaks in piggeries(10). Briefly, increasing cultivated land and fragmented grassland were both

associated with JEV outbreaks and these were computed as follows. Total cultivated land

composition was computed by summing the area of all crop patches in each landscape parcel

(landscape-level scale of 5 arc minutes). Grassland fragmentation was assessed by measuring mean

patch edge relative to patch area for all grassland patches in each parcel using the related

circumscribing circle (RCC). The RCC metric has been shown to be robust to patch size(85) in contrast

to the perimeter-to-area ratio (PAR), wherein similarly shaped patches can give different values for

different sizes and can translate to significant error when summarising PAR at landscape scale(85).

These composition and configuration metrics were previously computed for all land cover classes as

referenced above, but only those shown to be associated with JEV outbreak occurrence are

investigated with respect to ardeid species here as this was the stated aim of the current study. The

landscapemetrics package(86) was used to calculate RCC in R v. 4.3.1.

Functional metrics for species traits

As a crude measure of the species pool mean traits at landscape scale, the commonly used

community-weighted mean was calculated for each trait among all species identified as present

within each landscape parcel(87,88). As with the community composition metrics used to represent

abundance and relative abundance in the species pool at landscape scale described above, these

"community-weighted means" are not intended to represent local community trait means, but rather

to identify whether any species traits shared among the pool stood out as potentially influential for

community assembly at landscape scale, and particularly in high-risk JEV landscapes. Because of the

limited scope for evaluation of local community functional structure in this context, measures of

functional divergence and convergence were not assessed. The FD package(89) was used to compute

weighted means for species traits at landscape scale in R v. 4.3.1.

Point process modelling

Each of the 14 individual ardeid species' landscape-level relative abundances within the pool

were interrogated with respect to JEV outbreaks by fitting inhomogeneous Poisson point process

models (PPMs)(90,91). This allowed the spatial dependence of the outbreaks' distribution to be

evaluated against each species' distribution relative to other species across the landscape. To control

for potential reporting bias in outbreak surveillance, the background points used in these models

were sampled proportional to mean pig density, as described above. For the ardeid species PPMs, a

simple univariate PPM of JEV outbreaks was created for each of the 14 extant ardeid species' relative

abundance. Similarly, for the species trait PPMs, a simple univariate PPM of JEV outbreaks was

created for each of the species' traits; however, this univariate trait analysis was followed by a

multiple PPM to determine which traits among the species pool were independently associated with

JEV outbreaks. Correlation between the spatial distribution of traits was low (Pearson's r ≤ 0.24) and

all variance inflation factors were ≤ 1.46, indicating collinearity was not a concern for the trait

multiple PPM. All PPMs were fit at a scale of 5 arc minutes, which was the scale of outbreak

reporting(10). The Akaike information criterion (AIC) was used to evaluate model fit. The spatstat R

package was used to fit the PPMs(92).

Multivariate analysis for species assemblage composition

A hierarchical modelling of species communities (HMSC) framework(93,94) was used to

assess species pool composition, and individual species presence, in relation to the specific features

of landscape composition and configuration that have been shown to delineate JEV outbreak risk

(Figure S1)(10). In addition, the HMSC framework accounted for the potential modulation of species

presence in high-risk landscape structure by their phylogeny and functional traits, associations with

the latter representing species "responses" to the landscape features, however these do not

represent estimates of actual responses within communities and so this interpretation must be

tempered (see below). Models were fit under two domains: the first comprised each of the specific

landscape features shown to be associated with JEV outbreaks (Figure S1) and the second comprised

JEV outbreak occurrence itself as the single landscape feature of interest. The two separate domains

were evaluated since the landscape features associated with JEV outbreaks did not explain all of the

variability in JEV outbreak occurrence, indicating that there are additional unmeasured landscape

features that cannot be captured by the first domain, but which may still be relevant to species

presence in the landscape. The models were fit with the probit link function to represent species

presence, rather than species abundance, as the former is more appropriate given the focus of

investigation at landscape scale as already described above. The probit function is preferred over the

logit function in HMSC models because it provides a more pragmatic fit to the hierarchical

multivariate data(93). These models are fitted using Markov chain Monte Carlo methods and were

run here with four chains and 1000 samples each and a burn-in set to 20% of the initial samples. The

adjusted r^2 was used to assess model fit for each of the two model domains. Model convergence was

demonstrated for each species (Gelman diagnostic ≈ 1 for each species). Although interspecific

interaction within communities was not observed in this study, the HMSC framework does allow for

further accounting of important phylogenetic and functional influence on the sharing of space

between sympatric species given environmental filtering, which cannot be evaluated with the PPMs

described above. Nevertheless, it must be noted that since biotic filtering has not been directly

observed, which necessarily occurs at the level of the local community, true community-level

functional responses to environmental conditions are not captured at this landscape scale. Instead,

these multivariate analyses allow correlative associations to be identified that are anticipated to be

useful in characterising how community assembly could potentially operate in landscapes associated

with JEV outbreaks, and thereby help to inform sampling targets within such landscapes so that

community monitoring of JEV hosts can be systematically structured for One Health surveillance

across the large geographic extent of eastern Australia. The Hmsc package(94) was used for the

HMSC models.

Results

The PPMs identified landscape-level distributions of six species within the pool that were

strongly associated with JEV outbreaks: *Casmerodius albus*, *Ardea pacifica*, *Botaurus poiciloptilus*,

Egretta novaehollandiae, *Ixobrychus minutus*, and *Nycticorax caledonicus* (Table S2, Table 1, and

Figure 1). Of all the species traits interrogated under the PPMs, only the distributions of mean

population density and mean hand-wing index demonstrated independent associations with the

distribution of JEV outbreaks (Table S2, Table 2 and Figure 2), whereby increasing species pool-

weighted mean population density and increasing species pool-weighted mean hand-wing index

were both associated with increasing JEV outbreaks at landscape scale.

The HMSC model was in close agreement with respect to the structure of the species pool,

identifying the same species presences associated with JEV occurrence as those identified by the

PPMs, while also identifying one additional species, *Mesophoyx intermedia* (Figure 3). The median

adjusted r^2 for the landscape feature-based model and JEV-based model was 15.8% and 5%,

respectively. Furthermore, the HMSC model showed that whilst some high-risk landscape features

were common to all ardeids (the presence of transient wetlands and proximity to waterways, as

expected), other high-risk features were specific to individual species, or small clusters of species

(Figure 3). For example, the presence of *Ardea pacifica*, *Botaurus poiciloptilus*, *Egretta*

novaehollandiae, and *Ixobrychus minutus* was each associated with increasing presence of croplands,

whereas the presence of *Botaurus poiciloptilus*, *Ixobrychus minutus*, and *Nycticorax caledonicus* was

each associated in increasing grassland fragmentation. Additionally, the presence of five

(*Casmerodius albus*, *Ardea pacifica*, *Egretta novaehollandiae*, *Mesophoyx intermedia*, and *Nycticorax*

caledonicus) of the seven species associated with high-risk JEV landscapes was specifically associated

with greater water flow accumulation (Figure 3). All species whose presence was not associated with

JEV outbreaks were not associated, or were negatively associated, with these landscape features

(except *Butorides striata*, which was associated with grassland fragmentation).

Species presence within these high-risk landscape features did not appear to be modulated

by their functional traits after having accounted for phylogenetic structure (left panel, Figure 4).

However, interrogating landscapes defined solely by their JEV outbreak occurrence, rather than the

individual landscape features associated with outbreaks, showed that species presence in landscapes

characterised by outbreak occurrence was associated with greater species hand-wing index and body

mass (right panel, Figure 4).

Discussion

This study presents a description of the structure of the ardeid species pool at landscape

scale to inform how the landscape and functional structure of ardeid presence may be leveraged for

JEV surveillance in this natural nidus of virus circulation. The results showed that six of the 14 extant

ardeid species were strongly associated with the distribution of JEV outbreaks in piggeries reported

in 2022. While some JEV-associated landscape features, such as transient wetlands and proximity to

waterways, were commonly associated with species presence among most ardeids, other features of

high-risk landscape structure were specific to smaller groups of species, such as crop cultivation,

habitat fragmentation, and water flow accumulation. Interestingly, species traits did not provide

additional insight beyond that provided by phylogeny when considering the individual landscape

features associated with JEV outbreaks. However, hand-wing index was associated with species

presence in landscapes demarcated solely by outbreak occurrence and simultaneously demonstrated

an association between the distribution of species-pool weighted mean hand-wing index and JEV

outbreaks. These findings suggest that whilst high-risk JEV landscapes can vary with respect to the

ardeid species present, JEV surveillance across broad regions of eastern and southern Australia can

be informed using a combination of landscape structure and targeting of individual species in these

landscapes.

Previous work in Australia(10) and across the Indo-Pacific region(95,96) has demonstrated an

association between ardeid species richness and JEV outbreaks, albeit one that is quadratic in

nature. As such, low risk was associated with low and high ardeid richness, while high risk was

associated with intermediate richness. Nevertheless, the ways in which individual species, or species

community structure, may respond to specific landscape composition and configuration associated

with JEV occurrence has hitherto remained unknown. This marks a critical gap in our current

understanding of the infection ecology of JEV and highlights the need for wild bird monitoring to fill

that gap. However, given the extraordinarily broad geographic distribution of JEV outbreaks in

Australia, developing a systematic sampling strategy to answer key maintenance host ecology

questions that is representative of this breadth of occurrence is exceedingly challenging. To help

inform the development of such a sampling strategy, the current work sought to describe ardeid

composition as a representation of the available species pool for community assembly at the more

practical (and translatable) scale of the landscape. This also allowed for species presence to be

directly evaluated with respect to the landscape structure previously associated with JEV outbreaks,

and therefore that landscape structure can inform species selection for surveillance. The current

study identified six species that were individually associated with JEV outbreaks and found that

species presence is likely to be heterogeneous across the different landscape features associated

with those outbreaks, and that these associations also reflected differences in functional traits.

Moreover, the distinct approaches applied using both the PPMs and HMSC models were in strong

agreement with respect to the species identified in association with JEV outbreaks.

Associations between the dominant traits of species present in the landscape pool may

provide insight into potential drivers of species structure in landscapes delineating JEV risk. For

example, landscapes associated with JEV occurrence were also characterised by species

demonstrating a high level of dispersal capacity, as indicated by greater hand-wing index. This may

indicate the importance of dispersal among species in the context of seasonal or transient wetlands,

which were strongly associated with species presence in the current study, and JEV outbreaks

previously(10). Moreover, both modelling frameworks led to similar conclusions with respect to

hand-wing index. Such a scenario is supported by analyses of population structure among birds in

Australia following La Nina precipitation anomalies(8,97), and reflects more general evidence for the

importance of hand-wing index as an indicator of dispersal capacity in response to climate gradients

and habitat fragmentation(73–75). Interestingly, the two modelling approaches did not agree with

respect to population density and body mass; the PPM identified a positive association between

species pool-weighted mean population density and JEV outbreaks, and the HMSC model showed

that the association between species' presence and JEV outbreaks was modulated by species pool-

weighted mean body mass rather than population density. The association between JEV outbreaks

and higher population density is not surprising as this may reflect pathogen transmission

dynamics(68–70), but it is not clear why this association did not persist across modelling frameworks.

The discrepancy may result from the ability of the HMSC model to account for phylogenetic structure

in species presence, for which the PPM could not account, and the subsequent modulation by traits.

It should also be noted that the association between hand-wing index and species presence in high-

risk landscapes was not an artifact of body mass because these traits were not correlated (Pearson's

r = -0.21), which is as expected since hand-wing index adjusts for body size(72). While these findings

are intriguing, it is critical to emphasise that they are only correlative of possible drivers of

community assembly at landscape scale and do not describe realised community structure. It is not

possible to infer local community composition or functional responses to environmental conditions

from the data presented here. Rather, the true utility of this work is to describe preliminary,

evidence-based, configurations of ardeid species pools for the coupling of high-risk JEV landscape

structure with potential maintenance hosts across extraordinarily broad geographic extent, and

thereby inform the development of broad-scale sampling strategies for One Health JEV surveillance

in Australia.

Beyond the implications of these findings for potential wild bird surveillance programs, there

may be further potential utility for the design of other arbovirus surveillance programs by state and

territory health authorities in Australia. Arbovirus surveillance is currently conducted in many regions

throughout Australia, typically comprising mosquito and sentinel chicken monitoring. The selection

of monitoring sites is often informed by historic arbovirus activity or generically according to

potential mosquito and waterbird habitats. The results of the current investigation suggest that

consideration of the presence of wildlife hosts in specific landscapes may provide additional

information in identifying appropriate arbovirus surveillance locations more generally. Nevertheless,

whether wild host monitoring is employed specifically in the service of JEV surveillance, or more

generally for broad arbovirus surveillance, concomitant consideration of mosquito communities, and

specifically the nature of the alignment between their community composition and high-risk

landscapes and key maintenance hosts, will also be essential for maximally beneficial surveillance

output. The current study was not able to interrogate vector communities specifically, and as such

this remains a key knowledge gap in the infection ecology of JEV.

471   In addition to the specific constraints to interpretation of the results described above,

further description of additional limitations is included below. First, both the recording of ardeid

observations and the reporting of piggery outbreaks may be affected by reporting biases. We have

corrected for potential biases by sampling background points for ardeid SDMs proportional to the

HFP, and sampling background points for JEV piggery outbreaks proportional to mean pig density, as

described above. Nevertheless, it is possible that residual bias may persist. Second, notwithstanding

the inclusion of each of the ardeid species found in Australia in the habitat suitability modelling, it is

important to note that suitability represents the fundamental niche only, because it does not include

direct observational evaluation of biotic interactions to more formally define the realised niche or

subsequent community structure. However, this study did adjust the suitability estimates of

individual species by the estimates of all other species present at local scale using the SESAM

framework to represent species presence more appropriately in the landscape-level species pool.

Nevertheless, we must acknowledge that neither local biotic interactions nor dispersal history are

represented in this framework and should be considered in future work. Third, as noted above,

species other than ardeids may provide additional maintenance hosts to JEV-associated landscapes.

However, due to the current study's aim of coupling high-risk landscape structure with individual

target species and species pools at landscape scale, the analysis was constrained to the single,

established family of maintenance hosts to limit the extent of the assumptions regarding interspecific

interactions since direct community observation at local scale was not possible. Therefore, this study

is a preliminary, albeit actionable, representation of possible target species for specific landscapes

and should be expanded to include additional non-ardeid species as work progresses on demarcating

local community structure in high risk-landscapes through the implementation of monitoring

programmes. Finally, previous work showed that the piggery outbreaks were not associated with

feral pigs(10). Nevertheless, JEV has been identified in feral pigs, albeit only very sporadically(98). As

such, it is possible that there may be multiple landscapes of JEV risk, those pertaining to domestic

piggeries and those pertaining to shrub and forested bushlands. If the latter are in fact present, then

different surveillance programs targeting different maintenance hosts may be required.

In conclusion, this study has identified an initial selection of target ardeid species for

sampling in association with particular configurations of landscape risk. Although preliminary, it is

anticipated that this evidence-based approach to early JEV surveillance development will provide a

more efficient and productive yield than a simple random or convenience-based approach to

maintenance host sampling, particularly given the wide geographic extent in which JEV outbreaks

occurred across Australia. As wild bird monitoring progresses and local community structure

becomes better articulated with respect to JEV circulation, we expect that this selection will be

updated to reflect currently unrecognised local heterogeneities in landscape and community

composition, as well as novel maintenance hosts.

Declarations of interest: none.

Funding: This research did not receive any specific grant from funding agencies in the public,
commercial, or not-for-profit sectors

Acknowledgements: We thank Australian Pork Limited for supporting (non-financially) this research.
We thank SunPork Group for information about case farms. We thank Richard Bradhurst from the
University of Melbourne who facilitated the sharing of the data products.

Data Availability Statement: All data described in this study are publicly available for download at the
cited sources in the text except the pig density data, which contains sensitive information to the
Australian agricultural industry (e.g. locations of piggeries). Provision of this data for third parties
requires their application to Australian Pork Limited.

References

1.   Chief Medical Officer. Australian Department of Health and Aged Care. 2023 [cited 2023 Aug
14]. Statement on the end of Japanese encephalitis virus emergency response | Australian
Government Department of Health and Aged Care. Available from:
https://www.health.gov.au/news/statement-on-the-end-of-japanese-encephalitis-virus-
emergency-response

2.   Australian Department of Health and Aged Care. Japanese encephalitis virus situation
declared a Communicable Disease Incident of National Significance | Australian Government
Department of Health and Aged Care [Internet]. 2022 [cited 2022 Nov 4]. Available from:
https://www.health.gov.au/news/japanese-encephalitis-virus-situation-declared-a-
communicable-disease-incident-of-national-significance

3.   Mackenzie JS, Williams DT, van den Hurk AF, Smith DW, Currie BJ. Japanese Encephalitis Virus:
The Emergence of Genotype IV in Australia and Its Potential Endemicity. Viruses [Internet].
2022 Nov 9 [cited 2022 Dec 18];14(11):2480. Available from:
https://pubmed.ncbi.nlm.nih.gov/36366578/

4.   Williams CR, Webb CE, Higgs S, van den Hurk AF. Japanese Encephalitis Virus Emergence in
Australia: Public Health Importance and Implications for Future Surveillance. Vector Borne
Zoonotic Dis [Internet]. 2022 Nov 1 [cited 2022 Dec 18];22(11):529–34. Available from:
https://pubmed.ncbi.nlm.nih.gov/36354964/

5.   Hanna JN, Ritchie SA, Phillips DA, Shield J, Bailey MC, Mackenzie JS, et al. An outbreak of
Japanese encephalitis in the Torres Strait, Australia, 1995. Med J Aust [Internet]. 1996 Sep 2
[cited 2022 Nov 4];165(5):256–60. Available from:
https://pubmed.ncbi.nlm.nih.gov/8816682/

6.   Hanna JN, Ritchie SA, Phillips DA, Lee JM, Hills SL, Van Den Hurk AF, et al. Japanese
encephalitis in north Queensland, Australia, 1998. Med J Aust [Internet]. 1999 Jun 7 [cited
2022 Nov 4];170(11):533–6. Available from: https://pubmed.ncbi.nlm.nih.gov/10397044/

7.   van den Hurk AF, Pyke AT, Mackenzie JS, Hall-Mendelin S, Ritchie SA. Japanese Encephalitis
Virus in Australia: From Known Known to Known Unknown. Trop Med Infect Dis [Internet].
2019 Feb 20 [cited 2022 Nov 4];4(1). Available from:
https://pubmed.ncbi.nlm.nih.gov/30791674/

8.   Purnell C. The role of waterbirds in Australia's 2022 Japanese Encephalitis outbreak – a rapid
synthesis [Internet]. Carlton; 2022 [cited 2023 Aug 14]. Available from:
https://wildlifehealthaustralia.com.au/Portals/0/Documents/Ongoing%20Incidents/Role_of_
waterbirds_Aus_2022-JEV-
outbreak_RapidSsynthesis_BirdLifeAustralia.pdf?ver=LoZHg8uJJ_EHoY6FVkg9Ww%3D%3D

9.   Bureau of Meteorology A. Recent and historical rainfall maps. 2021 [cited 2022 Nov 9]. Recent
and historical rainfall maps, Australian Bureau of Meteorology. Available from:
http://www.bom.gov.au/climate/maps/rainfall/?variable=rainfall&map=totals&period=12mo
nth®ion=nat&year=2021&month=12&day=31

10.   Walsh MG, Webb C, Brookes V. An evaluation of the landscape structure and La Niña climatic
anomalies associated with Japanese encephalitis virus outbreaks reported in Australian
piggeries in 2022. One Health. 2023 Jun 1;16:100566.

11.   Rodrigues FM, Guttikar SN, Pinto BD. Prevalence of antibodies to Japanese encephalitis and
568   West Nile viruses among wild birds in the Krishna-Godavari Delta, Andhra Pradesh, India.
569   Trans R Soc Trop Med Hyg. 1981;75(2):258–62.

570   12.   Jamgaonkar A V., Yergolkar PN, Geevarghese G, Joshi GD, Joshi M V., Mishra AC. Serological
571   evidence for Japanese encephalitis virus and West Nile virus infections in water frequenting
572   and terrestrial wild birds in Kolar District, Karnataka State, India. A retrospective study. Acta
573   Virol. 2003;47(3):185–8.

574   13.   Buescher EL, Scherer WF, Rosenberg MZ, Kutner LJ. Immunologic studies of Japanese
575   encephalitis virus in Japan. IV. Maternal antibody in birds. Journal of Immunology.
576   1959;83:614–619.

577   14.   Bhattacharya S, Basu P. Japanese Encephalitis Virus (JEV) infection in different vertebrates and
578   its epidemiological significance: a Review. International Journal of Fauna and Biological
579   Studies [Internet]. 2014;1(6):32–7. Available from:
580   https://pdfs.semanticscholar.org/51c4/860d8c66c5fa9d730143e836b8db0b21c1a4.pdf

581   15.   Scherer W. Ecological studies of Japanese encephalitis in Japan. Parts I-IX. American Journal of
582   Tropical Medicine and Hygiene. 1959;8:644–722.

583   16.   Ogata M, Nagao Y, Jitsunari F, Kitamura N, Okazaki T. Infection of herons and domestic fowls
584   with Japanese encephalitis virus with specific reference to maternal antibody of hen
585   (epidemiological study on Japanese encephalitis 26). Acta Med Okayama. 1970;24:175–84.

586   17.   Nemeth N, Bosco-Lauth A, Oesterle P, Kohler D, Bowen R. North American birds as potential
587   amplifying hosts of Japanese encephalitis virus. American Journal of Tropical Medicine and
588   Hygiene. 2012;87(4):760–7.

589   18.   Van Den Hurk AF, Nisbet DJ, Hall RA, Kay BH, Mackenzie JS, Ritchie SA. Vector competence of
590   Australian mosquitoes (Diptera: Culicidae) for Japanese encephalitis virus. J Med Entomol
591   [Internet]. 2003 [cited 2022 Nov 4];40(1):82–90. Available from:
592   https://pubmed.ncbi.nlm.nih.gov/12597658/

593   19.   Baruah A, Hazarika R, Barman N, Islam S, Gulati B. Mosquito abundance and pig seropositivity
594   as a correlate of Japanese encephalitis in human population in Assam, India. J Vector Borne
595   Dis. 2018;55(4):291–6.

596   20.   Borah J, Dutta P, Khan SA, Mahanta J. Epidemiological concordance of Japanese encephalitis
597   virus infection among mosquito vectors, amplifying hosts and humans in India. Epidemiol
598   Infect. 2013;141(1):74–80.

599   21.   Kakkar M, Chaturvedi S, Saxena VK, Dhole TN, Kumar A, Rogawski ET, et al. Identifying
600   sources, pathways and risk drivers in ecosystems of Japanese Encephalitis in an epidemic-
601   prone north Indian district. PLoS One. 2017;12(5):1–17.

602   22.   Chen B, Beaty B. Japanese encephalitis vaccine (2-8 strain) and parent (SA 14 strain) viruses in
603   Culex tritaeniorhynchus mosquitoes. American Journal of Tropical Medicine and Hygiene.
604   1982;31:403–7.

605   23.   Komada K, Sasaki N, Inoue Y. Studies of live attenuated Japanese encephalitis vaccine in
606   swine. Journal of Immunology. 1968;100:194–200.

607   24.   Ghimire S, Dhakal S, Ghimire NP, Joshi DD. Pig Sero-Survey and Farm Level Risk Factor
608   Assessment for Japanese Encephalitis in Nepal. Int J Appl Sci Biotechnol. 2014;2(3):311–4.

609   25.   Brunner JL, LoGiudice K, Ostfeld RS. Estimating reservoir competence of Borrelia burgdorferi
610   hosts: prevalence and infectivity, sensitivity, and specificity. J Med Entomol [Internet]. 2008
611   Jan [cited 2018 Mar 22];45(1):139–47. Available from:
612   http://www.ncbi.nlm.nih.gov/pubmed/18283955

613   26.   Ostfeld RS, Levi T, Jolles AE, Martin LB, Hosseini PR, Keesing F. Life History and Demographic
614   Drivers of Reservoir Competence for Three Tick-Borne Zoonotic Pathogens. Ganta R, editor.
615   PLoS One [Internet]. 2014 Sep 18 [cited 2019 Jul 26];9(9):e107387. Available from:
616   https://dx.plos.org/10.1371/journal.pone.0107387

617   27.   Han BA, Park AW, Jolles AE, Altizer S. Infectious disease transmission and behavioural
618   allometry in wild mammals. Fenton A, editor. Journal of Animal Ecology [Internet]. 2015 May
619   1 [cited 2020 Sep 4];84(3):637–46. Available from: http://doi.wiley.com/10.1111/1365-
620   2656.12336

621   28.   Han BA, Schmidt JP, Bowden SE, Drake JM. Rodent reservoirs of future zoonotic diseases. Proc
622   Natl Acad Sci U S A. 2015;112(22):7039–44.

623   29.   Han BA, Schmidt JP, Alexander LW, Bowden SE, Hayman DTS, Drake JM. Undiscovered Bat
624   Hosts of Filoviruses. PLoS Negl Trop Dis. 2016;10(7):1–10.

625   30.   Stephens PR, Altizer S, Smith KF, Alonso Aguirre A, Brown JH, Budischak SA, et al. The
626   macroecology of infectious diseases: a new perspective on global-scale drivers of pathogen
627   distributions and impacts. Young H, editor. Ecol Lett [Internet]. 2016 Sep [cited 2018 Feb
628   5];19(9):1159–71. Available from: http://www.ncbi.nlm.nih.gov/pubmed/27353433

629   31.   Plourde BT, Burgess TL, Eskew EA, Roth TM, Stephenson N, Foley JE. Are disease reservoirs
630   special? Taxonomic and life history characteristics. Baker ML, editor. PLoS One [Internet].
631   2017 Jul 13 [cited 2017 Nov 30];12(7):e0180716. Available from:
632   http://dx.plos.org/10.1371/journal.pone.0180716

633   32.   Downs CJ, Schoenle LA, Han BA, Harrison JF, Martin LB. Scaling of Host Competence. Trends
634   Parasitol [Internet]. 2019 Mar [cited 2019 Jul 23];35(3):182–92. Available from:
635   http://www.ncbi.nlm.nih.gov/pubmed/30709569

636   33.   Downs CJ, Dochtermann NA, Ball R, Klasing KC, Martin LB. The effects of body mass on
637   immune cell concentrations of mammals. Am Nat [Internet]. 2019 Sep 10 [cited 2019 Sep
638   26];706235. Available from: https://www.journals.uchicago.edu/doi/10.1086/706235

639   34.   Faust CL, McCallum HI, Bloomfield LSP, Gottdenker NL, Gillespie TR, Torney CJ, et al. Pathogen
640   spillover during land conversion. Ostfeld R, editor. Ecol Lett [Internet]. 2018 Apr 1 [cited 2019
641   Jun 28];21(4):471–83. Available from: http://doi.wiley.com/10.1111/ele.12904

642   35.   Johnson CK, Hitchens PL, Pandit PS, Rushmore J, Evans TS, Young CCW, et al. Global shifts in
643   mammalian population trends reveal key predictors of virus spillover risk. Proc Biol Sci. 2020
644   Apr 8;287(1924):20192736.

645   36.   Gibb R, Redding DW, Chin KQ, Donnelly CA, Blackburn TM, Newbold T, et al. Zoonotic host
646   diversity increases in human-dominated ecosystems. Nature [Internet]. 2020 Aug 20 [cited

2021 Feb 15];584(7821):398–402. Available from: https://doi.org/10.1038/s41586-020-2562-
8

37.   Molles Manuel. Ecology: concepts and applications. 7th ed. New York: Mcgraw-Hill
Education; 2015. 258–281 p.

38.   Johnson PTJ, Rohr JR, Hoverman JT, Kellermanns E, Bowerman J, Lunde KB. Living fast and
dying of infection: host life history drives interspecific variation in infection and disease risk.
Ecol Lett [Internet]. 2012 Mar 1 [cited 2018 Mar 22];15(3):235–42. Available from:
http://doi.wiley.com/10.1111/j.1461-0248.2011.01730.x

39.   Schmidt JP, Maher S, Drake JM, Huang T, Farrell MJ, Han BA. Ecological indicators of mammal
exposure to Ebolavirus. Philosophical Transactions of the Royal Society B: Biological Sciences.
2019;374(1782).

40.   Walsh MG, Mor SM. Interspecific network centrality, host range and early-life development
are associated with wildlife hosts of Rift Valley fever virus. Transbound Emerg Dis [Internet].
2018 May 13 [cited 2018 Oct 28]; Available from:
http://www.ncbi.nlm.nih.gov/pubmed/29756406

41.   Walsh MG. Ecological and life history traits are associated with Ross River virus infection
among sylvatic mammals in Australia. BMC Ecol. 2019;19(1).

42.   World Organisation For Animal Health. WAHIS Inicident Reports. 2022 [cited 2022 Nov 4].
World Animal Health Information System (WAHIS). Available from:
https://wahis.woah.org/#/home

43.   New South Wales Health. Vector-borne Diseases Surveillance and Monitoring Weekly
Reports. 2022 [cited 2022 Dec 18]. Surveillance and monitoring weekly reports season 2021-
22 - Vector-borne diseases. Available from:
https://www.health.nsw.gov.au/environment/pests/vector/Pages/nswasp-weekly-report-
2021-22.aspx

44.   McKilligan N. Herons, Egrets and Bitterns: Their Biology and Conservation in Australia
[Internet]. CSIRO Publishing; 2005 [cited 2022 Nov 4]. Available from:
https://www.publish.csiro.au/book/4841

45.   Global Biodiversity Information Facility. Global Biodiversity Information Facility. 2022 [cited
2022 Nov 4]. GBIF occurrence download—Ardeidae Australia. Available from:
https://doi.org/10.15468/dl.6fsjpk

46.   Bradhurst RA, Roche SE, East IJ, Kwan P, Graeme Garner M. A hybrid modeling approach to
simulating foot-and-mouth disease outbreaks in Australian livestock. Front Environ Sci. 2015
Mar 19;3(MAR):17.

47.   Sanderson EW, Jaiteh M, Levy MA, Redford KH, Wannebo A V., Woolmer G. The Human
Footprint and the Last of the Wild. Bioscience. 2002 Oct 1;52(10):891.

48.   Socioeconomic Data and Applications Center | SEDAC. Methods» Last of the Wild, v2 |
SEDAC [Internet]. [cited 2014 Dec 23]. Available from:
http://sedac.ciesin.columbia.edu/data/collection/wildareas-v2/methods

49.   Lamarche C, Santoro M, Bontemps S, d'Andrimont R, Radoux J, Giustarini L, et al. Compilation
and Validation of SAR and Optical Data Products for a Complete and Global Map of

Inland/Ocean Water Tailored to the Climate Modeling Community. Remote Sensing 2017, Vol
9, Page 36 [Internet]. 2017 Jan 11 [cited 2022 Nov 4];9(1):36. Available from:
https://www.mdpi.com/2072-4292/9/1/36/htm

50.   European Space Agency, Climate Change Initiative. ESA (European Space Agency) CCI (Climate
Change Initiative) Land Cover project led by UCLouvain, 2017: Water Bodies v4.0. 2017 [cited
2022 Nov 4]. ESA (European Space Agency) CCI (Climate Change Initiative) Land Cover project
led by UCLouvain, 2017: Water Bodies v4.0. Available from:
http://maps.elie.ucl.ac.be/CCI/viewer/

51.   European Space Agency, Climate Change Initiative. ESA (European Space Agency) CCI (Climate
Change Initiative) Land Cover project 2017. 2017 [cited 2022 Nov 4]. Land Cover CCI Product -
Annual LC maps from 2000 to 2015 (v2.0.7). Available from:
http://maps.elie.ucl.ac.be/CCI/viewer/

52.   WorldPop. WorldPop⬚:: Geospatial covariate data layers [Internet]. [cited 2022 Dec 2].
Available from: https://hub.worldpop.org/project/categories?id=14

53.   University of Southampton S of G and ES, University of Louisville D of G and G, Universite de
Namur D de G. WorldPop hub [Internet]. [cited 2022 Nov 4]. Available from:
https://hub.worldpop.org/doi/10.5258/SOTON/WP00644

54.   Fick SE, Hijmans RJ. WorldClim 2: new 1-km spatial resolution climate surfaces for global land
areas. International Journal of Climatology [Internet]. 2017 May 15 [cited 2017 Jul 10];
Available from: http://doi.wiley.com/10.1002/joc.5086

55.   Priestley CHB, Taylor RJ. On the assessment of surface heat flux and evaporation using large-
scale parameters. Mon Weather Rev. 1972;100(2):81–92.

56.   Khaldi a., Khaldi A, Hamimed A. Using the Priestley-Taylor expression for estimating actual
evapotranspiration from satellite Landsat ETM + data. Proceedings of the International
Association of Hydrological Sciences. 2014;364(June):398–403.

57.   Trabucco A, Zomer RJ. CGIAR Consortium for Spatial Information. 2010. Global soil water
balance geospatial database. Available from: http://www.cgiar-csi.org

58.   Jung M, Dahal PR, Butchart SHM, Donald PF, De Lamo X, Lesiv M, et al. A global map of
terrestrial habitat types. Sci Data [Internet]. 2020 Aug 5 [cited 2023 May 8];7(1):1–8. Available
from: https://www.nature.com/articles/s41597-020-00599-8

59.   Jung M, Dahal PR, Butchart SHM, Donald PF, Lamo X De, Lesiv M, et al. Zenodo. 2020 [cited
2023 May 8]. A global map of terrestrial habitat types. Available from:
https://zenodo.org/record/3925749

60.   Pekel JF, Cottam A, Gorelick N, Belward AS. High-resolution mapping of global surface water
and its long-term changes. Nature [Internet]. 2016 Dec 15 [cited 2023 Apr
25];540(7633):418–22. Available from: https://pubmed.ncbi.nlm.nih.gov/27926733/

61.   Joint Research Centre EC. Global Surface Water Explorer [Internet]. [cited 2023 Apr 25].
Available from: https://global-surface-water.appspot.com/

62.   Lehner B, Verdin K, Jarvis A. HydroSHEDS Technical Documentation [Internet]. 2006. Available
from: http://hydrosheds.cr.usgs.gov

63.　Wilman H, Belmaker J, Simpson J, de la Rosa C, Rivadeneira MM, Jetz W. EltonTraits 1.0:
Species-level foraging attributes of the world's birds and mammals. Ecology [Internet]. 2014
Jul 1 [cited 2019 Jul 23];95(7):2027–2027. Available from: http://doi.wiley.com/10.1890/13-
1917.1

64.　Swihart RK, Gehring TM, Kolozsvary MB, Nupp TE. Responses of 'resistant' vertebrates to
habitat loss and fragmentation: the importance of niche breadth and range boundaries.
Divers Distrib [Internet]. 2003 Jan 1 [cited 2018 Mar 21];9(1):1–18. Available from:
http://doi.wiley.com/10.1046/j.1472-4642.2003.00158.x

65.　Santini L, Isaac NJB, Ficetola GF. TetraDENSITY: A database of population density estimates in
terrestrial vertebrates. Global Ecology and Biogeography. 2018;27(7):787–91.

66.　Tobias JA, Sheard C, Pigot AL, Devenish AJM, Yang J, Sayol F, et al. AVONET: morphological,
ecological and geographical data for all birds. Ecol Lett [Internet]. 2022 Mar 1 [cited 2023 Jul
28];25(3):581–97. Available from: https://onlinelibrary.wiley.com/doi/full/10.1111/ele.13898

67.　Lislevand T, Figuerola J, Székely T. AVIAN BODY SIZES IN RELATION TO FECUNDITY, MATING
SYSTEM, DISPLAY BEHAVIOR, AND RESOURCE SHARING. Ecology [Internet]. 2007 Jun 1 [cited
2021 Nov 10];88(6):1605–1605. Available from: http://doi.wiley.com/10.1890/06-2054

68.　Thrall PH, Biere A, Uyenoyama MK. Frequency-Dependent Disease Transmission and the
Dynamics of the Silene-Ustilago Host-Pathogen System. American Naturalist [Internet].
1995;145(1):43–62. Available from:
http://www.jstor.org/stable/2463146?seq=1#page_scan_tab_contents

69.　Begon M, Bennett M, Bowers RG, French NP, Hazel SM, Turner J. A clarification of
transmission terms in host-microparasite models: numbers, densities and areas. Epidemiol
Infect. 2002;129(1):147–53.

70.　Adler FR, Pearce-Duvet JMC, Dearing MD. How host population dynamics translate into time-
lagged prevalence: An Investigation of Sin Nombre virus in deer mice. Bull Math Biol.
2008;70(1):236–52.

71.　Rodríguez-Pastor R, Escudero R, Lambin X, Vidal MD, Gil H, Jado I, et al. Zoonotic pathogens in
fluctuating common vole (Microtus arvalis) populations: Occurrence and dynamics.
Parasitology. 2019;146(3):389–98.

72.　Lockwood R, Swaddle JP, Rayner JM V. Avian Wingtip Shape Reconsidered: Wingtip Shape
Indices and Morphological Adaptations to Migration. J Avian Biol. 1998 Sep;29(3):273.

73.　Kennedy JD, Borregaard MK, Jønsson KA, Marki PZ, Fjeldså J, Rahbek C. The influence of wing
morphology upon the dispersal, geographical distributions and diversification of the Corvides
(Aves; Passeriformes). Proceedings of the Royal Society B: Biological Sciences [Internet]. 2016
Dec 14 [cited 2023 Aug 2];283(1844). Available from:
https://royalsocietypublishing.org/doi/10.1098/rspb.2016.1922

74.　Sheard C, Neate-Clegg MHC, Alioravainen N, Jones SEI, Vincent C, MacGregor HEA, et al.
Ecological drivers of global gradients in avian dispersal inferred from wing morphology. Nature
Communications 2020 11:1 [Internet]. 2020 May 18 [cited 2023 Aug 2];11(1):1–9. Available
from: https://www.nature.com/articles/s41467-020-16313-6

75.   Weeks TL, Betts MG, Pfeifer M, Wolf C, Banks-Leite C, Barbaro L, et al. Climate-driven
769   variation in dispersal ability predicts responses to forest fragmentation in birds. Nature
770   Ecology & Evolution 2023 7:7 [Internet]. 2023 May 29 [cited 2023 Aug 2];7(7):1079–91.
771   Available from: https://www.nature.com/articles/s41559-023-02077-x

772   76.   International Union for the Conservation of Nature. IUCN Red List of Threatened Species
773   [Internet]. [cited 2020 Sep 4]. Available from: https://www.iucnredlist.org/

774   77.   Stekhoven DJ, Buhlmann P. MissForest--non-parametric missing value imputation for mixed-
775   type data. Bioinformatics [Internet]. 2012 Jan 1 [cited 2017 Nov 30];28(1):112–8. Available
776   from: https://academic.oup.com/bioinformatics/article-
777   lookup/doi/10.1093/bioinformatics/btr597

778   78.   Liaw A, Wiener M. Classification and Regression by randomForest. R News. 2002;2(3):18–22.

779   79.   Naimi B, Araújo MB. sdm: a reproducible and extensible R platform for species distribution
780   modelling. Ecography [Internet]. 2016 Apr 1 [cited 2021 Feb 12];39(4):368–75. Available from:
781   http://doi.wiley.com/10.1111/ecog.01881

782   80.   Guisan A, Rahbek C. SESAM - a new framework integrating macroecological and species
783   distribution models for predicting spatio-temporal patterns of species assemblages. J
784   Biogeogr [Internet]. 2011 Aug 1 [cited 2021 Feb 12];38(8):1433–44. Available from:
785   http://doi.wiley.com/10.1111/j.1365-2699.2011.02550.x

786   81.   D'Amen M, Pradervand JN, Guisan A. Predicting richness and composition in mountain insect
787   communities at high resolution: a new test of the SESAM framework. Global Ecology and
788   Biogeography [Internet]. 2015 Dec 1 [cited 2021 Feb 12];24(12):1443–53. Available from:
789   http://doi.wiley.com/10.1111/geb.12357

790   82.   Di Cola V, Broennimann O, Petitpierre B, Breiner FT, D'Amen M, Randin C, et al. ecospat: an R
791   package to support spatial analyses and modeling of species niches and distributions.
792   Ecography [Internet]. 2017 Jun 1 [cited 2021 Feb 12];40(6):774–87. Available from:
793   http://doi.wiley.com/10.1111/ecog.02671

794   83.   D'Amen M, Dubuis A, Fernandes RF, Pottier J, Pellissier L, Guisan A. Using species richness and
795   functional traits predictions to constrain assemblage predictions from stacked species
796   distribution models. J Biogeogr [Internet]. 2015 Jul 1 [cited 2021 Feb 12];42(7):1255–66.
797   Available from: http://doi.wiley.com/10.1111/jbi.12485

798   84.   Allouche O, Tsoar A, Kadmon R. Assessing the accuracy of species distribution models:
799   prevalence, kappa and the true skill statistic (TSS). Journal of Applied Ecology [Internet]. 2006
800   Sep 12 [cited 2021 Feb 16];43(6):1223–32. Available from:
801   http://doi.wiley.com/10.1111/j.1365-2664.2006.01214.x

802   85.   Csorba P, Szabo S. The Application of Landscape Indices in Landscape Ecology. In:
803   Tiefenbacher JP, editor. Perspectives on Nature Conservation - Patterns, Pressures and
804   Prospects. Rijeka: InTech; 2012.

805   86.   Hesselbarth MHK, Sciaini M, With KA, Wiegand K, Nowosad J. landscapemetrics: an open-
806   source R tool to calculate landscape metrics. Ecography. 2019 Oct 1;42(10):1648–57.

807   87.   Lavorel S, Grigulis K, McIntyre S, Williams NSG, Garden D, Dorrough J, et al. Assessing
808   functional diversity in the field – methodology matters! Funct Ecol [Internet]. 2008 Feb 1

[cited 2023 Aug 1];22(1):134–47. Available from:
https://onlinelibrary.wiley.com/doi/full/10.1111/j.1365-2435.2007.01339.x

88. Laliberte E, Legendre P. A distance-based framework for measuring functional diversity from
multiple traits. Ecology [Internet]. 2010 Jan 1 [cited 2023 Aug 1];91(1):299–305. Available
from: https://onlinelibrary.wiley.com/doi/full/10.1890/08-2244.1

89. Laliberté E, Legendre P, Shipley B. FD: measuring functional diversity from multiple traits, and
other tools for functional ecology. R package version 1.0-12.1. CRAN; 2014.

90. Baddeley A, Turner R. Practical Maximum Pseudolikelihood for Spatial Point Patterns (with
Discussion). Aust N Z J Stat [Internet]. 2000 Sep [cited 2014 Oct 23];42(3):283–322. Available
from: http://doi.wiley.com/10.1111/1467-842X.00128

91. Baddeley A, Rubak E, Turner R. Spatial Point Patterns: Methodology and Applications with R
[Internet]. Vol. 11. CRC Press; 2015 [cited 2016 Feb 5]. 810 p. Available from:
https://books.google.com/books?id=rGbmCgAAQBAJ&pgis=1

92. Baddeley A, Turner R. spatstat: An R Package for Analyzing Spatial Point Patterns. Journal of
Statistical Software 12(6) [Internet]. 2005 [cited 2014 Oct 23]; Available from:
http://www.jstatsoft.org/v12/i06/

93. Ovaskainen O, Abrego N. Joint Species Distribution Modelling: With Applications in R
[Internet]. Joint Species Distribution Modelling. Cambridge: Cambridge University Press; 2020
[cited 2023 Aug 1]. Available from: https://www.cambridge.org/core/books/joint-species-
distribution-modelling/0D9FA93EA1DD408332A17266449668B3

94. Tikhonov G, Opedal ØH, Abrego N, Lehikoinen A, de Jonge MMJ, Oksanen J, et al. Joint species
distribution modelling with the r-package Hmsc. Methods Ecol Evol [Internet]. 2020 Mar 1
[cited 2023 Aug 1];11(3):442–7. Available from:
https://onlinelibrary.wiley.com/doi/full/10.1111/2041-210X.13345

95. Walsh MG, Pattanaik A, Vyas N, Saxena D, Webb C, Sawleshwarkar S, et al. A biogeographical
description of the wild waterbird species associated with high-risk landscapes of Japanese
encephalitis virus in India. Transbound Emerg Dis [Internet]. 2022 Sep 1 [cited 2023 Mar
23];69(5):e3015–23. Available from: https://pubmed.ncbi.nlm.nih.gov/35809085/

96. Walsh MG, Pattanaik A, Vyas N, Saxena D, Webb C, Sawleshwarkar S, et al. High-risk
landscapes of Japanese encephalitis virus outbreaks in India converge on wetlands, rain-fed
agriculture, wild Ardeidae, and domestic pigs and chickens. Int J Epidemiol [Internet]. 2022
Mar 31 [cited 2022 Apr 14]; Available from: https://academic.oup.com/ije/advance-
article/doi/10.1093/ije/dyac050/6556297

97. Englert Duursma D, Gallagher R V., Griffith SC. Effects of El Niño Southern Oscillation on avian
breeding phenology. Divers Distrib [Internet]. 2018 Aug 1 [cited 2022 Nov 9];24(8):1061–71.
Available from: https://onlinelibrary.wiley.com/doi/full/10.1111/ddi.12750

98. Australian Veterinary Association. Emergency Animal Diseases. 2023 [cited 2023 Aug 14].
Japanese encephalitis. Available from: https://www.ava.com.au/policy-advocacy/disaster-
response/emergency-animal-diseases/japanese-encephalitis/

Table 1. Univariable regression coefficients and 95% confidence intervals for the associations
between Japanese encephalitis virus outbreaks and each ardeid species as derived from
inhomogeneous Poisson models.

Landscape species pool relative abundance	AIC	Estimate	95% confidence interval
Null model	204.70		
Casmerodius albus (*Ardea alba*) – Great egret	202.26	4.55	0.70 – 8.39
Ardea pacifica – White-necked (Pacific) heron	169.57	2.25	1.53 – 2.96
Botaurus poiciloptilus – Australasian bittern	201.13	3.26	1.17 – 5.34
Egretta novaehollandiae – White-faced heron	193.62	2.85	1.49 – 4.21
Ixobrychus minutus (*Ixobrychus dubius*) – Australian little bittern	195.89	10.43	5.69 – 15.17
Nycticorax caledonicus – Nankeen (rufous) night heron	202.12	2.08	0.41 – 3.76

854   Table 2. Regression coefficients and 95% confidence intervals for the associations between Japanese
855   encephalitis virus outbreaks and species traits as derived from univariable (A) and multiple (B)
856   inhomogeneous Poisson models.

Landscape species pool weighted mean traits	AIC	Estimate	95% confidence interval
Null model	238.05		
A. Univariate PPMs			
Percentage of diet – invertebrates	202.26	4.55	0.70 – 8.39
Hand-wing index	169.57	2.25	1.53 – 2.96
Population density (birds/km^2)	201.13	3.26	1.17 – 5.34
B. Multiple PPM			
Hand-wing index	213.58	0.22	0.104 – 0.335
Population density (birds/km^2)		1.35	0.718 – 1.977

Figure legends

Figure 1. The distribution of the relative abundance in the landscape-scale species pool of each
species identified as a potential target for Japanese encephalitis virus (JEV) surveillance based on the
strong association with the distribution of JEV outbreaks. The overlaid points represent the locations
of the JEV piggery outbreaks.

Figure 2. The distribution of the species pool-weighted mean (PWM) traits for those traits that were
associated with Japanese encephalitis outbreaks in the univariate point process models (proportion
of the diet comprising invertebrates, hand-wing index, and population density) or in the hierarchical
modelling of species communities model (hand-wing index and log body mass). The overlaid points
represent the locations of the JEV piggery outbreaks.

Figure 3. The associations between each species and the landscape features under two model
structures using hierarchical modelling of species communities (HMSC). The first HMSC model (top
panel) shows associations as positive, negative, or neutral "responses" of each species on the y-axis
to each landscape feature associated with JEV outbreaks on the x-axis. The second HMSC model
(bottom panel) shows these associations for JEV outbreak occurrence only and no other landscape
features.

Figure 4. The association between species traits and the landscape features, representing
modulation of species "responses" to the landscape features, under two model structures using
hierarchical modelling of species communities (HMSC). The first HMSC model (left panel) shows
associations as positive, negative, or neutral "modulators" for each trait on the y-axis of each
landscape feature associated with JEV outbreaks on the x-axis. The second HMSC model (right panel)
shows these associations for JEV outbreak occurrence only and no other landscape features.