Title: Cellular characterization of the mouse collecting lymphatic vessels reveals that lymphatic muscle cells are the innate pacemaker cells

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Running title: Lymphatic Muscle as Innate Pacemaker Cells

Impact: Lymphatic muscle cells, but not CD34+ adventitial cells, exhibited pacemaker behaviors such as pressure-dependent depolarization, pressure-dependent calcium mobilization during diastole, and propagated contraction waves induced by focal, optogenetic depolarization via enforced channel-rhodopsin2.

Abbreviations: Ano1, Anoctamin 1; ChR2, channel rhodopsin; cLV, collecting lymphatic vessel; IALV, Inguinal Axillary Lymphatic Collecting Vessel; ICC, Interstitial Cell of Cajal; ICLC, Interstitial Cell of Cajal Like Cell; LEC, Lymphatic Endothelial Cell; LMC, Lymphatic Muscle Cell; STDs, Spontaneous transient depolarizations; STMs, Spatio-Temporal Maps

Keywords: Lymphatic collecting vessel, lymphatic muscle cell, pacemaking, interstitial cells of Cajal like cells, mesenchymal stem cells

Abstract

Collecting lymphatic vessels (cLVs) exhibit spontaneous contractions with a pressure-dependent frequency, but the identity of the lymphatic pacemaker cell is still debated. Here we combined immunofluorescence and scRNAseq analyses with electrophysiological methods to examine the cellular constituents of the mouse cLV wall and assess whether any cell type exhibited morphological and functional processes characteristic of pacemaker cells. We employed inducible Cre (iCre) mouse models to target specific cell populations including c-kitCreER^{T2} to target interstitial cells of Cajal like cells; *PdgfrβCreER*^{T2} to target pericyte-like cells; *PdgfrαCreER*^{TM} to target CD34^{+} adventitial cells; and *Myh11CreER*^{T2} to target lymphatic muscle cells (LMCs) directly. These inducible Cre lines were crossed to the fluorescent reporter ROSA26mT/mG, the genetically encoded Ca^{2+} sensor GCaMP6f, and the light-activated cation channel rhodopsin2 (ChR2). Only LMCs consistently, but heterogeneously, displayed spontaneous Ca^{2+} events during the diastolic period of the contraction cycle, and whose frequency was modulated in a pressure-dependent manner. Further, optogenetic depolarization with ChR2 only induced propagated contractions in LMCs. Membrane potential recordings in LMCs demonstrated that the rate of diastolic depolarization significantly correlated with contraction frequency. These findings support the conclusion that LMCs, or a subset of LMCs, are responsible for mouse cLV pacemaking.

Introduction

The spontaneous contractions of collecting lymphatic vessels (cLV) are an integral component to fluid and macromolecule homeostasis as they provide the force to transport fluid from the interstitial spaces back to the blood circulation (Scallan et al., 2016). In humans, spontaneous contractile activity is estimated to account for 2/3 of lymph transport (Engeset et al., 1977) and this function is significantly compromised in patients suffering from lymphedema, whose cLVs typically display weak and irregular or entirely absent contractile activity (Olszewski, 2002). *Ex vivo* studies, in which the intraluminal pressure can be precisely controlled, have refined our understanding of the pressure-dependent regulation of contraction frequency (Benoit et al., 1989; Gashev et al., 2004), with some mouse cLVs displaying a 10-fold increase in contraction frequency over a 10 cmH_2O pressure gradient (Scallan and Davis, 2013; Zawieja et al., 2018a). The observation that cLVs, often cannulated at various lengths for *ex vivo* preparations, retain a consistently tunable contraction frequency points to the presence of (a) pacemaker cell(s) innate to the structure of the cLV wall and with a seemingly ubiquitous presence along the length of the vessel (Zawieja et al., 1993; Castorena-Gonzalez et al., 2018b). Furthermore, isolated cLVs typically display single pacemaker initiation sites unless damaged or electrically uncoupled by pharmacological inhibition of gap junctions or genetic deletion of *Gjc1* (Connexin 45, Cx45) (Behringer et al., 2017; Castorena-Gonzalez et al., 2018b; Castorena-Gonzalez et al., 2020). In sum, this suggests the pacemaker cell(s) is(are) likely both ubiquitous and continuous, to allow for electrical conduction via gap junctions, along the length of the cLV and prevent colliding contractile waves which would impair lymph transport.

Investigations into the cLV pacemaker identity have focused largely on cells termed interstitial cells of Cajal like cells (ICLC; or telocytes) (McCloskey et al., 2002; Briggs Boedtkjer et al., 2013), as they display some morphological and cell marker expression profiles similar to the interstitial cells of Cajal (ICC), which are bona fide pacemakers in the gastrointestinal (GI) tract. ICC are classically identified by either methylene blue staining and expression of CKIT, and coordinate GI smooth muscle contraction (Maeda et al., 1992; Ward et al., 1994; Ordog et al., 1999). ICC also express the canonical Ca^{2+} activated chloride channel Anoctamin 1 (Ano1) (Gomez-Pinilla et al., 2009), which is required for pacemaker activity (Hwang et al., 2009; Zhu et al., 2009; Singh et al., 2014). Previous reports in sheep mesenteric lymphatic vessels identified a population of cKIT^{+}, VIMENTIN^{+}, ICLC in the vessel wall between the endothelial and LMC layer and running along the axis of the vessel (McCloskey et al., 2002).

Investigations in the human thoracic duct also identified a significant population of ICLC in close proximity to the lymphatic muscle cells (LMCs) evident by methylene blue staining, immunostaining for CD34, VIMENTIN, and cKIT, as well as the gold standard of electron microscopy (Briggs Boedtkjer et al., 2013). However, neither study could determine if these cells had functional electrical communication with the LMCs or demonstrate either a membrane electrical clock or internal Ca^{2+} clock to drive the rhythmic lymphatic vessel contractions observed *ex vivo*. LMCs share a functional similarity to ICC in that they also display the *Ano1* mediated Ca^{2+} activated chloride current (Van Helden, 1993; Toland et al., 2000; Mohanakumar et al., 2018) (Zawieja et al., 2019), that regulates pacemaking. Spontaneous transient depolarizations, presumably Ano1 dependent, were recorded in mesenteric cLVs from guinea pigs (Van Helden, 1993; von der Weid et al., 2008) providing a mechanism for membrane potential instability to drive AP initiation. Furthermore, computational models have proposed LMC sarcoplasmic reticulum (SR) Ca^{2+} release as the oscillator mechanism driving pacemaking (Imtiaz et al., 2007). SR Ca^{2+} release has also been implicated in pericyte regulation of arterioles (Hashitani et al., 2015; van Helden and Imtiaz, 2019), in microvascular vasomotion (Boedtkjer et al., 2008; Aalkjaer et al., 2011; van Helden and Imtiaz, 2019), and in the contraction waves of atypical muscle cells of the lower urinary tract (Grainger et al., 2022).

Presently, no investigations have clearly identified the cellular identities of possible pacemaker cells within the cLVs of the mouse. Mouse cLVs exhibit contractile parameters and conduction speed equivalent to those of human vessels (Castorena-Gonzalez et al., 2018b) and their simplified architecture, compared to larger mammals, in combination with the genetic tools developed for the mouse model, allowed us to test for a fundamental pacemaker cell in the cLV. In this study we utilized multiple genetic mouse models, confocal imaging of fluorescent reporters, cell specific expression of GCaMP6f for Ca^{2+} imaging, and optogenetic light-activated depolarization to both visualize and test the functional aspects of putative pacemaker cells, along with membrane potential recordings in LMCs in pressure-challenged cLVs. We also performed immunostaining and single cell RNA sequencing (scRNAseq) of isolated cLVs to provide greater detail to the heterogenous cellular populations found within the mouse cLVs. Despite identifying a significant population of $CD34^{+}Pdgfr\alpha^{+}$ adventitial cells along the length of mouse cLVs, the results of our functional studies support a myogenic (LMC) origin of pacemaking in cLVs.

Results

Methylene Blue Staining Reveals a Minor Population of Cells in Mouse cLVs

Methylene blue staining was used to identify an ICLC population in the human lymphatic thoracic duct (Briggs Boedtkjer et al., 2013). In our isolated and cleaned lymphatic inguinal axillary collecting vessels (IALVs), methylene blue stained a significant number of cells with variable density along the length of the IALV and heterogenous cell morphologies (Figure 1A-C). A significant portion of the stained cells resembled lymphatic vessel-associated macrophages with an elongated shape, while other cells were smaller and circular (Figure 1D-F). Methylene blue also appeared to stain mast cells as there were large ovoid cells with intracellular granules on the adventitial surface of the vessel. In addition, methylene blue stained a minor population of cells that exhibited long and thin axon-like extensions which appeared to have a slight helical orientation, with a small central body and nucleus (Figure 1C). None of these cell populations were aligned with the longitudinal axis of the vessel that would permit efficient coupling or regulation across the circumferential layer of LMCs required for coordinated propagation along the length of the vessel.

Immunofluorescence Imaging of IALVs Stained for ICLC, LEC, and LMC Markers

We next stained IALVs for the putative telocyte/ICLC markers cKIT, CD34, and the intermediate filament VIMENTIN, which have been previously utilized for ICLC identification in human and sheep lymphatic tissues (McCloskey et al., 2002; Briggs Boedtkjer et al., 2013). Additionally, an antibody to the intermediate filament Desmin was used to label muscle cells (McCloskey et al., 2002). IALVs stained for cKIT (Figure 2B) showed robust signal in large ovoid cells with a non-segmented circular nucleus (Figure 2A), characteristic of mast cells that were in the outer part of the adventitia. Similarly, cKIT stained

populations of elongated cells as well as circular cells with variable densities throughout the IALV wall, similar to methylene blue$^+$ cell populations (Figure 2B, J). Staining for CD34 revealed a large population of cells that were seemingly contiguous along the length of the vessel. The CD34$^+$ cells generally had multiple lobular processes and a "oak leaf" like appearance, typically a characteristic of fibroblasts, though some contained short, thin dendrite-like extensions (Figure 2C, G, K). The CD34$^+$ cells were negative for Desmin [Figure 2H), which primarily stained the circumferential LMCs (Figure 2F; note that the largely non-circumferential cell organization in this region is typical for a lymphatic endothelial valve site (Bridenbaugh et al., 2013a)]. Furthermore, CD34$^+$ cells and cKIT$^+$ cells were separate populations (Figure 2D, L). A VIMENTIN antibody labeled lymphatic endothelial cells (LECs) which exhibited a horizontal cobblestone morphology in parallel with the vessel axis (Figure 2E, I), while also co-labeling the majority of the CD34$^+$ cells (Figure 2H) and cKIT$^+$ cells (Figure 2L). Videos of the half vessel z-stacks are provided (Supplemental Movies 1-3 for Figure 2D, H, and L respectively).

Of the cells stained in Figure 2, the CD34$^+$ population was intriguing due to its high density and distribution throughout the length of the IALV, which potentially would be conducive to effective regulation of LMC excitability. In addition to CD34 and VIMENTIN, PDGFRα staining is also commonly ascribed to both telocytes (Vannucchi et al., 2013; Xiao et al., 2013; Zhou et al., 2015) as well as fibroblasts (Kimura et al., 2021; Clayton et al., 2022). We performed immunofluorescence imaging for PGDFRα counterstained with CD34 and markers for LMCs, LECs, and pericytes. As noted in Figure 2, CD34$^+$ cells (Figure 3A) did not co-label LMCs (Figure 3D) which were smooth muscle actin$^+$ (SMA, Figure 3B) and Calponin$^+$ (Figure 3C). However, nearly all CD34$^+$ (Figure 3E) cells were also PDGFRα$^+$ (Figure 3F, H). Occasionally some overlap of PDGFRα and SMA$^+$ signal was noted (Figure 3G, H). LECs staining with CD31 (PECAM, Figure 3I) revealed the expected rectangular elongated cobblestone morphology that was distinct from the PDGFRα+ cells (Figure 3J, L). Staining for Calponin also specifically labeled LMCs (Figure 3K) but not PDGFRα$^+$ cells (Figure 3L). Lastly, we stained for PDGFRα, CD34, and the commonly used pericyte marker PDGFRβ (Figure 3 M-P). As above, CD34 and PDGFRα were highly colocalized (Figure 3Q, R, T), and many of the CD34$^+$ and PDGFRα$^+$ cells were also PDGFRβ$^+$ (Figure 3P). PDGFRβ also stained some circumferential LMCs (Figure 3Q). During the imaging of mouse IALVs for these markers, we also observed that the lymphatic secondary endothelial valves were populated by elongated cells that stretched the length of the valve leaflet and were positive for CD34, PDGFRα, and PDGFRβ, with varying intensities. These cells could be observed in most, if not all, the valves we assessed and found within both leaflets of the valve (Figure 3R,S). These cells had long, thin extensions that were branched, along with apparent dendrite-like extensions with a morphology that closely resembled those described for pericytes or telocytes (Popescu and Faussone-Pellegrini, 2010). PDGFRα$^+$ or CD34$^+$ cells with this morphology were only observed in the valve leaflets, and thus seemed insufficient to regulate pacemaking as normal contractions are observed in cLVs without secondary valves (Van Helden, 1993; Gashev et al., 2002). Representative z-stacks demonstrating these valve-located "telocyte" shaped cells (Figure 3R,S) are provided as Supplemental Movies 4 and 5.

We next determined the degree of colocalization between the CD34 and PDGFRα signal given the significant overlap in their staining profile. Colocalization analysis of PDGFRα (SuppFigure 1A) and CD34 (SuppFigure 1B) and their colocalization (SuppFigure 1C) was determined with the FIJI BIOP-JACoP tool. The Pearson's coefficient was 0.83 (SuppFigure 1D) and Mander's coefficient of overlap was 0.80 for the PDGFRα$^+$ signal and 0.87 for the CD34 signal (SuppFigure 1E). Colocalization between Myh11 and PDGFRα was significantly lower (SuppFigure 1D-F) with a Pearson's coefficient of 0.30 (SuppFigure 1G), whereas the Mander's coefficient for Myh11 overlap with PDGFRα was 0.077 and 0.043 for PDGFRα signal overlap with Myh11 (SuppFigure 1H). The high degree of colocalization CD34 and PDGFRα signal informed our use of the commercially available transgenic *PdgfrαCreER*^{TM} mouse model to target these cells. The vast majority of the PDGFRα$^+$ cells were located in the adventitial layer (SuppFigure 2A-D), which varied between 1-3 PDGFRα$^+$ cells thick (SuppFigure 2E). Under this layer, we observed only a single layer of largely circumferential LMCs stained by Myh11 (SuppFigure 2B) sitting atop a single layer of CD31$^+$ LECs (SuppFigure 2A). We also observed occasional PDGFRα$^+$ cells

or their extensions located in the sub-endothelial space (SuppFigure 2 E', E") positioned between the LECs and the LMCs.

Use of iCre-Mediated Recombination of Rosa26mT/mG to Delineate and Characterize Specific IALV Cell Types

After confirming the presence of VIMENTIN$^+$, cKIT$^+$, and CD34$^+$ PDGFRα$^+$ positive cells within the mouse IALV, we sought to further investigate these cell populations by using constitutive and inducible Cre recombinase expressing mouse lines. IALVs from the constitutively active *PdgfrαCre*-ROSA26mTmG and *Ng2Cre*-ROSA26mTmG mice had GFP fluorescence in the majority of LMCs as well as in the fibroblast-shaped cells found within the IALV wall (Figure 4 A,B). While informative of expression of the LMC progenitor cells, neither constitutive Cre would be useful in delineating cell types. In contrast to the constitutively active *PdgfrαCre*, the tamoxifen inducible *PdgfrαCreER^{TM}* line drove significant recombination in only the fibroblast-shaped cells previously stained for CD34 and PDGFRα but not in LMCs or LECs (Figure 4C). *PdgfrβCreER^{T2}*, commonly used to label pericytes, drove recombination in both a minor population of the LMCs and the fibroblast-shaped cells. *cKitCreER^{T2}*, which capably drives recombination in the ICCs of the GI tract (Baker et al., 2016), drove recombination only in a small population of irregularly-spaced, large ovoid cells on the surface of the IALV (Figure 4E), although recombination in 1 or 2 LECs could occasionally be detected (not shown). Finally, *Myh11CreER^{T2}* drove recombination in nearly all LMCs which were largely circumferentially oriented with dendrite-like, cell-cell contacts visible between them and without significant GFP fluorescence in either LECs or the fibroblast-shaped CD34$^+$ PDGFRα$^+$ cell population (Figure 4F). Additionally, some LMCs maintained the bipolar shape but had secondary extensions forming a "Y" shape in which an adjacent LMC typically filled the inner void. A very minor population of recombined cells in the *Myh11CreER^{T2}*-ROSA26mTmG IALVs were smaller and irregularly patterned with multiple fine axon-like projections or ruffled edges (Figure 4F).

To complement the morphological and cell density findings obtained with confocal microscopy, we digested IALVs from the iCre-ROSA26mTmG lines, and the *Prox1-eGFP* line as a control, into single cell suspensions and sorted the respective GFP$^+$ populations (Figure 4G-J) for RT-PCR profiling (Figure 4K). We first focused on determining the molecular fidelity of the sorted cells based on the gene promoters used to drive each "iCre" model to discern cellular overlap. In agreement with the confocal images, sorted GFP$^+$ cells from *PdgfrβCreER^{T2}*-ROSA26mT/mG IALVs expressed *Pdgfrβ* but also *Myh11* and *Pdgfrα*. In contrast, GFP-sorted cells from *PdgfrαCreER^{TM}* IALVs expressed *Pdgfrα* and *Pdgfrβ*, but with no detectable expression of *Myh11*. GFP$^+$ cells from sorted *Myh11CreER^{T2}*-ROSA26mTmG IALVs had high expression for *Myh11* as well as *Pdgfrβ*, but did not express *Pdgfrα*. IALVs from *cKitCreER^{T2}*-ROSA26mTmG mice were not pursued for FACS due to the exceptionally sparse recombination observed along the IALV.

Characterization of the cellular components of the mouse IALVs by scRNAseq and FACs-RT-PCR

The results from the immunofluorescence staining, ROSA26mTmG reporter imaging, and FACs-RT-PCR experiments suggested that both LMCs and AdvCs can express *Pdgfrβ*. To provide further clarity and detail to the cellular populations within the mouse cLV wall and potential subsets within those broad cell types we performed scRNAseq on isolated and cleaned inguinal axillary cLVs from male and female mice. The resulting UMAP projection (Figure 5A) revealed a host of cell types which had 3 main clusters corresponding to LECs, LMCs and AdvCs (Figure 5A). We assessed the expression of genes that correspond to the markers from our earlier immunofluorescence staining as well as cell identification markers commonly used within the literature to identify each cell cluster (Figure 5B). Cell identity was confirmed by commonly used markers (Figure 5B) and the top differentially expressed genes (SuppFigure 3A). Dot plots for the LEC markers *Prox1 (*Figure 5*C)* and *Flt4 (*Figure *5D)*, LMC markers *Myh11* (Figure 5E) and *Cnn1* (Figure 5F), and the AdvCs markers *Pdgfrα* (Figure 5G) and *Lumican* (Figure 5H) were quite specific for labelling their respective cell clusters. Very few *Kit* (Figure 5I) expressing cells were observed in accordance with our imaging results. *Pdgfrβ* was observed in both LMC and AdvC

clusters (Figure 5J) while the remaining cell clusters were of immune origin as they expressed the gene encoding the hematopoietic marker *CD45* (Figure 5K). Notably, the previous genes suggested to identify LMCs in a previous scRNASeq study (Kenney et al., 2022), *Dpt*, *Pi16*, and *Ackr3*, were largely absent in LMCs and instead were expressed in a minor population of AdvCs (SuppFigure3B). We provide a further sub-clustering breakdown of the LECs (SuppFigure 4), LMCs (SuppFigure5), AdvCs (SuppFigure 6), and a detailed expression profile of the immune cell clusters (SuppFigure7). Further assessment of the LEC subcluster included a putative lymphatic endothelial "up valve" cell population in sub cluster 8 which expressed high levels of *Prox1*, *Cldn11*, *Itga9*, *Gja4*, and *Neo1* and "down valve" population in cluster 6 which expressed *Clu*, *Adm*, *Gja4* and *Lypd6* (SuppFigure 4C) which mapped well to a previous RNAseq dataset (Gonzalez-Loyola et al., 2021; Petkova et al., 2023; Yoon et al., 2024). The top differentially expressed genes in the putative down valve population in cluster 8 included *Irx3*, *Neo1*, *Tub*, *Ano4*, and *Fxyd2* and we noted *Cacna1e*, *Fgf14*, and *Irf1* in the up-valve cluster 6. Analysis of the LMC subclusters did not reveal any significant differences in the expression of known pacemaking associated genes *Ano1* or *Itpr1* at our initial conditions of Log2FC of 0.5. However, we provide an overview of the typical ion channel families expressed in LMCs in SuppFigure 5B-I. The AdvC cells could be further subclustered into multiple populations (SuppFigure 6A,C) with little evidence of LMC gene contamination as these cells lacked *Myh11*, *Kcnma1*, and *Tagln* despite expression of *Cacna1c*, *Ano1*, and *Cx45*. Over 75% of AdvCs expressed *Pdgfrα* (SuppFigure 6) and 65% of the total AdvCs expressed both *Pdgfrα* and *CD34*. Our immunofluorescent colocalization of *Pdgfrα* and *CD34 was also supported as* 72% of *Pdgfrα expressing* AdvCs also co-expressed *CD34* (SuppFigure 6D). The vast majority of AdvCs expressing *Pdgfrβ* (SuppFigure 6E) or *Cspg4* (SuppFigure 6F) also expressed *Pdgfrα.* Expression of *Ano1*, *Cx45*, and *Cacna1c*, was also observed in some of the AdvCs and most of those cells also co-expressed *Pdgfrα*, supporting further use of the *PdgfrαCreER*^{TM} line (SuppFigure 6G-I).

While scRNASeq highlighted the depth of heterogeneity of the cellular composition of the mouse cLV, we wanted to validate the actual recombined cell populations from the iCre-ROSA26mTmG models. We profiled each iCre driven recombination of ROSAmTmG via FACs-purified cells and RT-PCR for common markers for endothelial cells, muscle cells, and pericytes. *Nos3* (eNOS) expression was observed only in the *Prox1-eGFP* sorted cells, and LECs also expressed *Vim*, *Mcam*, and had weak but detectable signal for *CD34* (Figure 6A). *Myh11CreER*^{T2} sorted cells showed expression of smooth muscle actin (*Acta2*), the alpha subunit of the L-type voltage gated Ca^{2+} channel *Cacna1c* (*Cav1.2*), *Desmin* (*Des*), *M*cam, and *Vimentin* (*Vim*, Figure 6B). In addition to the genes expressed under *Myh11CreER*^{T2} recombination, *Cdh5*, *CD34*, and *Cspg4* (*Ng2*) were detected in cells sorted from *PdgfrβCreER*^{T2} IALVs (Figure 6C). As expected, the GFP^{+} cells sorted from *PdgfrαCreER*^{TM} IALVs expressed mRNA for *CD34*, weak signal for *Cspg4*, and *Vimentin*, but not *Desmin*, *Acta2*, nor the pericyte marker *Mcam* (Figure 6D). *Cacna1c* was expressed in cells FACS purified from both *PdgfrβCreER*^{T2} and *Myh11CreER*^{T2} IALVs and sorted cells from *PdgfrαCreER*^{TM} IALVs without any evidence that *Myh11* expressing muscle cells contaminated the latter. These findings confirmed the separate cell populations achieved with *PdgfrαCreER*^{TM} and *Myh11CreER*^{T2} mediated recombination, at least as it pertains to the ROSA26mTmG reporter. These findings were largely validated by our scRNASeq dataset. *Cdh5* (Figure 6E) and *Nos3* (Figure 6F) were almost exclusively expressed in the LEC clusters while *Acta2* (Figure 6G) was highly expressed in the LMC cluster. We also observed that *Cacna1c* was highly expressed in the LMCs (Figure 6H) and some AdvCs. *Cd34* was widely expressed in AdvCs matching our immunofluorescence data. *Cd34* expression was also seen in LECs (Figure 6I) although we did not observe a signal in LECs in our earlier immunofluorescence staining (Figure 3). *Cspg4* was observed in a minor population of AdvCs (Figure 6J). The intermediate filament *Vim* (Figure 6K) was ubiquitously expressed across all clusters expressed but *Des* was primarily expressed in LMCs and some subsets of AdvCs (Figure 6K, L). The endothelial and pericyte marker *Mcam* (also referred to as CD146) was expressed in LECs and LMCs but was largely absent in AdvCs (Figure 6M). We followed up the identification of Cav1.2 expression in the *PdgfrαCreER*^{TM} sorted cell population by assessing the expression of other genes involved in either electrical conduction (Cx45) (Figure 6N) or pacemaking

(Ano1) (Figure 6O) of IALVs. Expression of *Ano1* and *Cx45* was observed in Pdgfr*αCreER*^{TM} ROSA26mtmG FACS-purified cells (Figure 6P).

Inducible Deletion of Either Cav1.2, Ano1, or Cx45 with *PdgfrαCreER*^{TM} Did Not Affect cLV Pacemaking

The expression of the genes critically involved in cLV function—*Cav1.2*, *Ano1*, and *Cx45*—in the *PdgfrαCreER*^{TM}-ROSA26mTmG purified cells and scRNAseq data prompted us to generate *PdgfrαCreER*^{TM}*-Ano1*$^{fl/fl}$, *PdgfrαCreER*^{TM}*-Cx45*$^{fl/fl}$, and *PdgfrαCreER*^{TM}*-Cav1.2*$^{fl/fl}$ mice for contractile tests. We isolated popliteal cLVs and tested their pacemaker and contractile function in response to a physiological pressure range of 0.5-10 cmH_2O, under normal conditions. However, we did not detect any significant differences in pacemaking or contractile function as assessed by contraction frequency, ejection fraction, and vessel tone in popliteal cLVs studied from *PdgfrαCreER*^{TM}*-Ano1*$^{fl/fl}$ mice (Figure 7A-C) or *PdgfrαCreER*^{TM}*-Cx45*$^{fl/fl}$ mice (Figure7D-F). There was no difference in contraction frequency of cLVs from *PdgfrαCreER*^{TM}*-Cav1.2*$^{fl/fl}$ mice compared to floxed control mice, however, we noted a mild but statistically significant increase in ejection fraction at the lowest pressure, 0.5 cmH_2O (Figure 7H). Additionally, vessels isolated from *PdgfrαCreER*^{TM}*-Cav1.2*$^{fl/fl}$ mice also had a statistically significant increase in vessel tone (Figure 7I) noted at the 2-way level although we did not resolve significance at any specific pressure with this sample. No differences in normalized contraction amplitude, fractional pump flow, or diastolic diameter were observed (SuppFigure 8). In total, despite the presence of transcript for these critical genes in *Pdgfrα*$^{+}$ cells, *PdgfrαCreER*^{TM} mediated deletion of *Cx45*, *Cav1.2* or *Ano1* failed to recapitulate previous reports of the significant contractile defects using the *Myh11CreER*^{T2} line to delete the same genes (Castorena-Gonzalez et al., 2018b; Zawieja et al., 2019; To et al., 2020; Davis et al., 2022).

PDGFRα^{+} Adventitial Fibroblasts Express Markers Associated with Multipotency

Despite the lack of cLV pacemaking deficits in the *PdgfrαCreER*^{TM} genetic knockout lines, we were curious to discern further insight into the role or function of the PDGFRα^{+} CD34^{+} cells since they comprise a significant portion of the lymphatic cLV wall. We performed RT-PCR on FACS purified cells from *Prox1-eGFP*, *Myh11CreER*^{T2}-ROSA26mTmG, and *PdgfrαCreER*^{TM}-ROSA26mTmG IALVs for multipotency markers including Krüppel-like factor 4 (*Klf4*), stem cell antigen 1 (*Sca1*, also referred to as *Ly6a*), and *Gli1,* with *CD34* and *Pdgfrα* used to assess purity. Recombined (GFP^{+}) cells from *Myh11CreER*^{T2}-ROSA26mTmG had weak expression of *Klf4* and *Gli1 but* were negative for *Ly6a* (SuppFigure 9A). *PdgfrαCreER*^{TM} recombined cells strongly expressed *Klf4*, *Ly6a*, and *Gli1 (*SuppFigure 9A). LECs sorted from *Prox1-eGFP* IALVs were positive for *Klf4*, weak for *Ly6a*, and positive for *CD34* but negative for *Gli1* and *PDGFRα* (SuppFigure 9B). The unrecombined population (tdTomato^{+}) cells in the *Myh11CreER*^{T2}- ROSA26mTmG IALVs (SuppFigure 9B) showed expression for all the markers as expected. *PdgfrαCreER*^{TM} *recombined cells* also expressed the mesenchymal stromal cell markers *CD29*, *CD105*, and CD44 (SuppFigure 9C, positive control in 9D). However, expression of these genes was not homogenous across all the AdvCs population based on our scRNAseq analysis (SuppFigure 9E-J). We performed immunofluorescence staining for one of these multipotent markers, Ly6a (SuppFigure 9K) in the adventitial cells with PDGFRα (SuppFigure 9L) and counter staining for LMCs with MYH11 (SuppFigure 9M). The morphology and staining pattern of Sca1 overlapped significantly with PDGFRα staining and not MYH11 staining (SuppFigure 9N, Supplemental Movie 6).

Optogenetic Stimulation of iCre-driven Channel Rhodopsin 2

We next used optogenetic methods to test whether the cell populations recombined by either *cKitCreER*^{T2}, *PdgfrαCreER*^{TM}, or *Myh11CreER*^{T2} could elicit a coordinated contraction. The ChR2-tdTomato construct appeared more sensitive to recombination than ROSA26mTmG, in some cases resulting in LMC expression of ChR2-tdTomato in *PdgfrαCreER*^{TM} and *cKitCreER*^{T2} popliteal cLVs based on cell morphology. Care was taken to image each vessel for tdTomato (Figure 8A,C,E) prior to stimulation at its respective sites under brightfield conditions for diameter tracking (Figure 8B,D,F) to

ensure fidelity of the cell types and morphologies observed in Figure 3 and Figure 4. As with ROSA26mTmG, *cKitCreER*^{T2} drove the ChR2-tdTomato expression primarily in large ovoid cells found on the adventitia of the vessel. Cells were stimulated by positioning an optical laser fiber (tip diameter 2-3 μm) near a ChR2+ cell, with an illumination field of 10-50 μm. Localized photo-stimulation of these cells did not initiate coordinated contractions (Figure 8G-J,S). Similarly, photo-stimulation of ChR2-tdTomato expressing cells driven by *PdgfrαCreER*^{TM} failed to initiate a coordinated contraction (Figure 8K-N, T). In contrast, localized photo-stimulation of LMCs, using *Myh11CreER*T2 to express Chr2-tdTomato, resulted in a propagated contraction in the popliteal vessel (Figure 8O-R, U). In total, only 3.25% of photo-stimulation events for *cKitCreER*^{T2}-ChR2-TdTomato and 3.03% of photo-stimulation events for *PdgfrαCreER*^{TM}-ChR2-tdTomato were associated with a contraction, while 88.5% of photo-stimulation events for *Myh11CreER*^{T2}-ChR2-tdTomato induced contractions (Figure 8V). The optogenetic triggering of contractions observed in *PdgfrαCreER*^{TM}-ChR2-tdTomato and *cKitCreER*^{T2}-ChR2-TdTomato vessels is likely due to the happenstance of spontaneous contractions occurring during the time and proximity of optogenetic stimulation (see Methods). As a control, we also used non-induced (no tamoxifen) *Myh11CreER*^{T2}-ChR2-tdTomato cLVs and contractions were associated with only 7% of photo-stimulation events, in line with the *PdgfrαCreER*^{TM} and *cKitCreER*^{T2} results (Figure 8V). As mast cells are not ascribed any tissue specific pacemaking behavior, these similar low percentages observed between these three groups are suggestive of random coincidence. Brightfield videos of the photo-stimulation and representative traces for *cKitCreER*^{T2}-ChR2-TdTomato, *PdgfrαCreER*^{TM}-ChR2-tdTomato, *Myh11CreER*^{T2}-ChR2-tdTomato are provided in Supplemental Movies 7-9.

Confocal Ca^{2+} Imaging of GCaMP6f Expression Driven by *cKitCreER*^{T2}, *PdgfrαCreER*^{TM}, and *Myh11CreER*^{T2} Over the Lymphatic Contraction Cycle

Subcellular calcium transients are observed in many pacemaker cells. We imaged IALVs from *cKitCreER*^{T2}-GCaMP6f mice, which primarily resulted in expression of GCaMP6f in the large ovoid cells in the adventitia (Figure 9A), although we occasionally observed GCaMP6f expression in both LEC and LMCs (Figure 9A) as depicted in the maximum projection of the acquisition period (Supplemental Movie 10) and the spatio-temporal maps (STMS). The aberrant expressions of GCaMP6f in cells that demonstrated the typical cobblestone morphology of LECs or the circumferential LMCs that exhibited Ca^{2+} flashes and diastolic Ca^{2+} transients (Figure 9D,E green arrows) prior to contraction were not included in the *cKitCreER*^{T2}-GCaMP6f analysis. Of 39 *cKitCreER*^{T2}-GCaMP6f cells analyzed, only 1 *cKitCreER*^{T2}-GCaMP6f cell exhibited a spontaneous Ca^{2+} transient during the recording period (Figure 9B,C Cell 7). However, the Ca^{2+} transient in that cell did not align temporally with the "Ca^{2+} flash" of the LMC with incidental GCaMP6f expression (Figure 9C,D). Despite the lack of Ca^{2+} transients under the baseline conditions throughout the IALV contraction cycle, many *cKitCreER*^{T2}-GCaMP6f cells exhibited a robust and prolonged Ca^{2+} event in response to stimulation with the mast cell activator compound 48-80 (Figure 9F, G, H). Notably, the Ca^{2+} events in the ovoid cells elicited by administration of compound 48-80 did not acutely alter the LMC Ca^{2+} activity (Figure 9I,J). Similarly, the majority of *PdgfrαCreER*^{TM}-GCaMP6f expressing cells also largely lacked Ca^{2+} transients and resulted in incidental LMC GCaMP6f expression (Figure 10B, Supplemental Movie 11). Some cells exhibited high basal Ca^{2+} levels (Figure 10A,D) sustained throughout the recording, but without oscillations (Figure 10B,C). In contrast, spurious GCaMP6f expression in a circumferentially oriented LMC displayed Ca^{2+} flashes associated with contraction (Figure 10B,C). Of the 21 *PdgfrαCreER*^{TM} -GCaMP6f cells assessed, only 3 exhibited Ca^{2+} transients and those were singular events contained within a single cell within the 20 sec imaging period (Figure 10E,F). The lack of either global or consistent Ca^{2+} transients within either *cKitCreER*^{T2}-GCaMP6f or *PdgfrαCreER*^{TM}-GCaMP6f IALVs was in stark contrast to Ca^{2+} imaging of *Myh11CreER*^{T2}-GCaMP6f IALVs. *Myh11CreER*^{T2} drove GCaMP6f expression in nearly all circumferential LMCs (Figure 11A), which exhibited global and nearly synchronous Ca^{2+} flashes in 100% of the analyzed cells (Figure 11B, C). Additionally, non-synchronous stochastic and localized Ca^{2+} transients were commonly observed in the LMCs during diastole (Figure 11D, E, Supplemental Movie 12). Many LMCs exhibited Ca^{2+} transients during each diastolic period while other LMCs displayed few Ca^{2+} transients or lacked

diastolic Ca^{2+} transients during the recording period (Figure 11B). In aggregate, only 1 of 39 *cKitCreER^{T2}*-GCaMP6f cells and 3 of 21 *PdgfrαCreER^{TM}*-GCaMP6f cells displayed a Ca^{2+} transient during recording, while 20 of 43 LMCs displayed at least one diastolic transient apart from 43 of 43 LMCs with global Ca^{2+} flashes.

Pressure Dependency of Subcellular Ca^{2+} Transients in LMCs

We next sought to test whether diastolic Ca^{2+} transients were pressure-dependent, given that cLVs exhibit pressure-dependent chronotropy (Zawieja et al., 2019). GCaMP6f expressing LMCs were studied at intraluminal pressures of 0.5 -5 cmH_2O in the presence of nifedipine, which blocked the Ca^{2+} flashes but not local Ca^{2+} transients (Figure 12A). As intra-luminal pressure was increased, there was a marked increase in the occurrence of Ca^{2+} transients (Figure 12B, Supplemental Movies 13-15). These calcium transients were converted into particles (PTCLs) for further analysis as previously described (Drumm et al., 2019a). Activity maps of Ca^{2+} PTCL activity were generated (Figure 12C) and PTCL area (Figure 12D) and frequency were determined at each pressure (Figure 12E). The maps show that as pressure increased, the area of the LMC layer displaying a Ca^{2+} transient increased (as evident by the increase in PTCL area) as did the distribution of Ca^{2+} PTCLs across the LMC layer (Figure 12C). Across 11 experiments, the area of the field of view activated by PTCLs/frame increased from 73.2 ± 17.7 μm^2/frame at 0.5 cmH_20 to 108.6 ± 20.5 μm^2/frame at 2 cm H_20 and was further enhanced to 139.2 ± 26.9 μm^2/frame at 5 cm H_2O (Figure 12F). The number of PTCLs per frame also increased with pressure, from 2.9 ± 0.4 at 0.5 cmH_20 to 4.1 ± 0.5 and 5.2 ± 0.6 PTCL/frame at 2 and 5 cmH_20 respectively (Figure 12G).

Contraction Frequency is Set by the Diastolic Depolarization Rate

To assess how pressure regulates LMC membrane potential, we first recorded membrane potential in cells exhibiting action potentials (APs) using a microelectrode filled with biocytin-AF488 to label the impaled cell. In each case (n=3 IALVs) the labeled cell was a LMC wrapping circumferentially around the vessel (Figure 13A, B) and as these recordings were made over the course of many minutes the direct neighboring circumferential LMCs also exhibited fluorescence, albeit weaker in intensity, as expected for cells couple by gap junctions (Figure 13A). In all the recorded cells exhibiting APs, we noted a diastolic depolarization preceding the sharp upstroke achieved once threshold was met at each pressure (Figure 13C). The AP frequency and rate of the diastolic depolarization increased with pressure (Figure 13D, E). Linear regression of a plot of each AP frequency and diastolic depolarization rate at each pressure demonstrated a tight associated between the two parameters. However, we did not observe a significant effect of pressure on minimum membrane potential (Figure 13G), threshold potential (Figure 13H), AP upstroke (Figure 13I), AP peak potential (Figure 13J), plateau potential (Figure 13K), or the time spent over threshold (Figure 13L).

Discussion

The identification of the cellular origin and signaling mechanisms underlying cLV pacemaking will reveal novel targets for pharmacological intervention in treating lymphedema and the associated lymphatic contractile dysfunction. In this study we tested proposed pacemaker cell types based on 3 parameters: 1) that the pacemaker cells are located along the entire length of the cLV, to accommodate spontaneous contractions and coordinated electrical conduction; 2) that depolarization of the pacemaker cell can drive a coordinated and propagated contraction of the vessel; and 3) that the presence of Ca^{2+} transients precedes or coincides with contraction, as commonly observed in other pacemaker cells. We used confocal microscopy and a combination of immunofluorescence and fluorescent reporters under the control of various inducible Cres to identify and target both muscle and non-muscle cells, previously labeled as ICLCs, which co-express the markers CD34 and PDGFRα. Our cell characterizations were supplemented by scRNAseq analysis of isolated and cleaned mouse IALVs which supported our finding of three major cell types including LECs, LMCs, and AdvCs each of which could be further sub-clustered into transcriptionally unique populations. From our initial fluorescence imaging studies, a role for

intrinsic pacemaking by LMCs (Van Helden, 1993; von der Weid et al., 2008), or by a novel population of $CD34^+$ lymphatic ICLC (McCloskey et al., 2002; Briggs Boedtkjer et al., 2013), also referred to as telocytes, were further examined and found to co-express *Pdgfrα*. We utilized *PdgfrαCreER*^{TM} to further test whether these cells exhibited pacemaker capabilities. However, these PDGFRα^+ cells had minimal Ca^{2+} activity despite ongoing contractions and optogenetic stimulation of ChR2 in these cells failed to drive a spontaneous contraction. In contrast, photo-stimulation of LMCs expressing ChR2 elicited robust, propagated contractions with similar characteristics and propagation to spontaneous contractions in the same vessels. Furthermore, Ca^{2+} imaging in LMCs revealed diastolic Ca^{2+} transients in diastole that increased in frequency and spatial spread as pressure was elevated. We also demonstrated that the primary component of the AP driving the frequency change with pressure is diastolic depolarization, which we have previously reported to be dependent on ANO1 (Zawieja et al., 2019) and IP3R1 (Zawieja et al., 2023). Notably, we recently reported that diastolic Ca^{2+} transients are abrogated in IALVs from *Myh11CreER*^{T2}*-Itpr1* inducible knockout mice, supporting a IP_3R1-ANO1 axis as the pressure-dependent pacemaker mechanism in LMCs. These results, in addition to the recent findings using targeted deletion of Cx45 (Castorena-Gonzalez et al., 2018b) or Cav1.2 (To et al., 2020; Davis et al., 2022) in lymphatic muscle support the model of LMCs as the intrinsic pacemaker as has been previously proposed (Van Helden, 1993; Van Helden et al., 1996; Van Helden and Zhao, 2000).

Pacemaking in Smooth Muscle

In many smooth muscle organs, regulation of a coordinated contraction is a complex and multicellular phenomenon. Multiple cell types integrate physical and biological information into electrical activity to be transmitted to the force-producing smooth muscle cells, sometimes across great distances relative to cell size, to regulate Ca^{2+} influx by voltage-dependent Ca^{2+} channels required for contraction. The intestine is one such documented tissue in which $cKIT^+$ ICCs and PDGFRα^+ interstitial cells form an electrical syncytium to regulate intestinal motility (Sanders et al., 1999; Sanders et al., 2014). The pacemaking function of intestinal ICCs relies heavily on ANO1, a Ca^{2+} activated Cl^- channel, which is required for slow wave activity in the ICCs. Both cKIT and ANO1 can be used as a marker for ICCs in the intestine (Hwang et al., 2009; Cobine et al., 2017; Malysz et al., 2017), $cKIT^+$ and $VIMENTIN^+$ ICLCs have been observed in sheep lymphatic vessels (McCloskey et al., 2002), yet these cell populations did not form gap junctions with the smooth muscle to form electrical connections (Briggs Boedtkjer et al., 2013) as occurs in the intestinal ICCs. Our cKIT staining and *cKitCreER*^{T2}-ROSA26mTmG reporter studies on mouse IALVs revealed a sparse population of large ovoid cells previously classified as mast cells (Chatterjee and Gashev, 2012; Zawieja et al., 2019). Their identity as mast cells was further supported by a sustained global Ca^{2+} event after stimulation with the mast cell degranulating agent compound 48-80. However, both VIMENTIN and CD34 showed robust staining throughout the mouse lymphatic vessel wall. LECs stained for VIMENTIN, as did non-muscle stellate shaped cells, with many co-expressing CD34. Other smaller circular cells some of which were also $cKIT^+$ as well and some whose morphology was similar to that of the macrophage staining profile of the GFP^+ cells in IALVs from MacGreen mice, were also $VIMENTIN^+$, consistent with previous reports of macrophage staining in cLVs (Bridenbaugh et al., 2013b; Chakraborty et al., 2015; Zawieja et al., 2016). While $VIMENTIN^+$ cells had a perinuclear staining profile, CD34 demarcated the cell membrane and was useful for assessing the morphology of these cells. Of particular interest, the $VIMENTIN^+CD34^+$ cells densely populated the length of the mouse IALV, with a majority displaying a flattened stellate morphology characterized by a classic rounded oak-leaf appearance, although some displayed fine dendrite-like extensions. Contrasting with the previous findings in the human thoracic duct (Briggs Boedtkjer et al., 2013), we did not observe a significant population of $CD34^+$ cells with a bipolar morphology oriented axially along the vessel. However, z-stack reconstructions of sections of the mouse IALV that included the secondary valves revealed interstitial $CD34^+$PDGFRα^+ cells that resembled those bipolar cells with multiple axon-like extensions throughout the endothelial leaflets; these were similar to interstitial cells that were previously reported in in collecting vessel valves (Leak and Burke, 1968) and lymphovenous valves (Geng et al., 2016). While these cells have not been frequently described in the valves of peripheral cLVs, we observed

them in each of the valve regions imaged and, in addition, they were labeled with other Cre drivers, including *NG2Cre-ROSA26mTmG* and *PdgfrβCreER*^{T2}*-ROSA26mTmG* (data not shown). Whether these cells regulate leaflet extracellular matrix deposition or lymphatic valve integrity is unknown, but a possible role as a critical pacemaker can be excluded as vessel segments without valves display normal contractile behavior (Van Helden, 1993; Gashev et al., 2002). Instead, the majority of the CD34$^+$PDGFRα$^+$ cells were found in the adventitia, in 2-3 layers, overtop the LMCs, and they were consistently observed in high density along the IALV. Some CD34$^+$PDGFRα$^+$ cells or their extensions were present between the lymphatic endothelial and muscle layers as had been previously reported with electron microscopy of human lymphatic vessels (Briggs Boedtkjer et al., 2013). Thus, while some of these AdvCs may be contained within the extracellular matrix that retracts onto the vessel during microdissection, many others are intimately dispersed within the vessel wall.

PDGFRα$^+$CD34$^+$ Cells are Not Involved in cLV Pacemaking Under Physiological Conditions

Co-expression of CD34 and PDGFRα has recently been ascribed as a delineating feature of telocytes, although PDGFRα routinely labels fibroblasts and specific interstitial cells in the GI tract involved in purinergic neurotransmission in the GI tract (Kurahashi et al., 2011; Kurahashi et al., 2013; Clayton et al., 2022). CD34 expression is also ascribed to some multipotent cell populations of various origins (Sidney et al., 2014). For example, PDGFRα$^+$ fibroblasts appear to be progenitors of the smooth muscle fibers associated with the lacteal, the lymphatic capillary in the villus (Sanketi et al., 2024). It remains controversial to what extent telocytes are distinct from or are components/subtypes of either cell type and morphological discrimination between the populations typically requires electron microscopy imaging (Clayton et al., 2022). Mesenchymal stromal cells (Andrzejewska et al., 2019) and fibroblasts (Muhl et al., 2020; Buechler et al., 2021; Forte et al., 2022) are not monolithic in their expression patterns displaying both organ directed transcriptional patterns as well as intra-organ heterogeneity (Lendahl et al., 2022) as readily demonstrated by recent single cell RNA sequencing studies that provided immense detail about the subtypes and activation spectrum within these cells and their plasticity (Luo et al., 2022a). We were able to distinguish up to 10 subclusters of AdvCs, the majority of which expressed or co-expressed CD34 and PDGFRα. These cells were consistently negative for smooth muscle markers such as *Des*, *Cnn1*, *Acta2*, *Myh11* or the pericyte marker *Mcam*. However, PDGFRβ expression was noted in our scRNAseq analysis and in our RT-PCR of sorted *PdgfrαCreER*^{TM}-ROSA26mTmG cells. PDGFRβ protein expression was confirmed with variable immunofluorescence staining amongst the PDGFRα stained cells as well as LMCs. The *PdgfrβCreER*^{T2}ROSA26mTmG mice had only modest recombination in both the LMC and PDGFRα$^+$ cell populations, but potentially highlighted a myofibroblast-like cell subpopulation, cells that might lie on the spectrum of differentiation from lymphatic muscle and PDGFRα$^+$ cells, or perhaps a cell with pacemaker activity as PDGFRβ is widely used as a pericyte marker and some pericytes display pacemaker activity (Hashitani et al., 2015). Adding to this intrigue, the *PdgfrαCreER*^{TM} sorted cells expressed transcripts for *Cacna1c*, the voltage-gated L-type Ca^{2+} channel critical for lymphatic contractions (Zawieja et al., 2018a; To et al., 2020); *Ano1*, the ion channel underlying pressure-dependent chronotropy (Mohanakumar et al., 2018; Zawieja et al., 2019); and *Cx45*, the primary connexin mediating electrical conduction in mouse lymphatic collecting vessels (Castorena-Gonzalez et al., 2018b; Hald et al., 2018). Expression of these genes in certain sub-populations of the AdvCs was also apparent in our scRNAseq analysis. Thus, the presence of those gene transcripts does not appear to be due to muscle cell contamination or incidental recombination in LMCs as we did not detect LMC markers in the RT-PCR profiling of the sorted PDGFRα$^+$ cells nor were GFP-expressing cells with an LMC morphology observed in imaging of *PdgfrαCreER*^{TM}-ROSA26mTmG vessels. Critically, however, deletion of *Cav1.2*, *Cx45*, or *Ano1* through *PdgfrαCreER*^{TM}-mediated recombination neither recapitulated the previous phenotypes achieved with *Myh11CreER*^{T2} (Castorena-Gonzalez et al., 2018b; Zawieja et al., 2019; To et al., 2020; Davis et al., 2022) nor significantly affected pacemaking in mouse popliteal cLVs. This finding is in stark contrast to the complete lack of contractions observed in *Myh11CreER*^{T2}*-Cav1.2*$^{fl/fl}$ vessels (To et al., 2020; Davis et al., 2023b) or the vessels from vascular muscle specific *Itga8CreER*^{T2}*-Cav1.2*$^{fl/fl}$ mice (Davis et al., 2022; Warthi et al., 2022), and the significant loss in pressure-induced chronotropic

modulation of pacemaker function in IALVs with *Myh11CreER*^{T2}-mediated deletion of Ano1 that we have previously reported (Zawieja et al., 2019) . While a sub-population of CD34^{+}PDGFRα^{+} cells may share expression of critical pacemaker genes identified in the LMCs, they do not appear to be involved in cLV pacemaking or contractile function under physiological states. Instead, CD34^{+}PDGFRα^{+} cells co-stained significantly with Sca1^{+}, suggesting they may be primed to act as resident multipotent cells (Song et al., 2020; Kimura et al., 2021). To this point, the *PdgfrαCreER*^{TM} FACS purified cells also expressed markers associated with "stemness" such as *CD34*, *Klf4*, *Gli1*, *CD29*, *CD105*, *CD44*, and *Vimentin*, in addition to *Sca1*, and it is likely that the *PdgfrαCreER*^{TM} population includes various distinct subpopulations (Jolly et al., 2022) expressing these markers. These cells may play a role in rebuilding the lymphatic collecting vessel vasculature following collecting vessel damage or lymph node resection and further studies are required to assess their functional multipotency.

SR Ca^{2+} Cycling in Pacemaking

The use of the mouse IALV model, in addition to the simplicity of the vessel architecture, provided the use of genetic tools that previously had been instrumental in identifying the cKIT^{+} ICC as the pacemaker cells of the GI tract (Ward et al., 1994; Huizinga et al., 1995; Torihashi et al., 1995). Through the use of the respective *PdgfrαCreER*^{TM} and *Myh11CreER*^{T2} drivers, we were able to specifically image Ca^{2+} in each respective cell type in pressurized, contracting vessels. Pacemaking initiating cells have an inherently unstable membrane potential, oftentimes utilizing the oscillatory nature of Ca^{2+} release from the sarcoendoplasmic reticulum coupled to Ca^{2+} sensitive electrogenic exchangers or ion channels to drive depolarization (Van Helden, 1993; Hashitani et al., 2015; Baker et al., 2021b; Sanders et al., 2022). One such example is the pacemaker ICC in the gastric corpus which exhibits abundant Ca^{2+} transients that couple to ANO1-mediated chloride currents in both the intervening period between slow waves as well as the plateau phase of the slow wave (Baker et al., 2021a), however such activity is not characteristic of all pacemaker ICC types. The identification of a Ca^{2+}-activated chloride current in LMCs (Van Helden, 1993; Toland et al., 2000) and its correspondence with subcellular Ca^{2+} transients (Van Helden, 1993; Ferrusi et al., 2004; von der Weid et al., 2008) led Van Helden to postulate that LMCs had intrinsic pacemaking capability (Van Helden, 1993; Van Helden et al., 1996). We have previously reported that mouse LMCs in pressurized vessels routinely display subcellular Ca^{2+} release events that reflect the kinetics and characteristics of Ca^{2+} puffs and waves in addition to the coordinated global Ca^{2+} flash associated with Ca^{2+} influx during an AP (Castorena-Gonzalez et al., 2018b; Zawieja et al., 2018a; Zawieja et al., 2019). Here we confirmed the consistent presence of subcellular Ca^{2+} transients only in LMCs with GCaMP6f driven by *Myh11CreER*^{T2} but not in the cells with GCaMP6f driven by *PdgfrαCreER*^{TM}. Critically, we also demonstrated that the Ca^{2+} transients increased in both frequency and spatial spread as pressure was elevated in the vessel, as would be expected to account for the pressure-dependent chronotropy observed in lymphatic collecting vessels. This underscores the finding that genetic deletion of *Ano1* in the LMCs dramatically reduced contraction frequency and abolished pressure-dependent chronotropy in those vessels (Zawieja et al., 2019). This phenotype was largely replicated with a similar reduction in frequency and loss of pressure-dependent chronotropy in our recent study utilizing *Myh11CreER*^{T2} to drive deletion of IP3R1 from LMCs (Zawieja et al., 2023) in which these diastolic Ca^{2+} transients were absent. This fits with the central role of IP3R and subcellular Ca^{2+} release as critical components of intrinsic LMC pacemaking (Van Helden et al., 1996; von der Weid et al., 2008). In addition to the transcriptional heterogeneity identified by scRNASeq, we also noted heterogeneity in the propensity of LMCs to display diastolic Ca^{2+} transients under control conditions or show the sustained Ca^{2+} oscillations that occur in the presence of nifedipine. We did not detect any significant difference in the expression of *Itpr1*, the gene encoding the IP3R1, across our LMCs subclusters. However, when using less stringent conditions we identified that the LMC cluster "0" had significantly increased expression of *Itprid2* (Log2FC of 0.26) which encodes the KRas-induced actin-interacting protein (KRAP). KRAP has recently been implicated in IP3R1 immobilization and licensing and was required for IP3R1-mediated Ca^{2+} puffs (Thillaiappan et al., 2021; Atakpa-Adaji et al., 2024). Whether the higher expression of KRAP results in a greater probability of these LMCs to display IP3R1-dependent Ca^{2+} oscillations in LMCs

requires further investigation. Of note, LMCs also express the components for store operated calcium entry including *Stim1*, *Stim2*, *Orai1*, *Orai3*, *Saraf*, and *Stimate*, which may be involved in maintaining IP3R1-dependent SR Ca2+ release oscillations.

The membrane potential recordings we made in this study suggest that the regulation of pressure-dependent chronotropy is through modulation of the diastolic depolarization rate in LMCs, as previously demonstrated in rat mesenteric lymphatic vessels (Zawieja et al., 2018b). The appearance of the diastolic depolarization may depend on the method of vessel stretch employed as it is not always observed in wire myograph preparations (von der Weid et al., 2014). Notably, in this study *PdgfrαCreER*^{TM} mediated deletion of Ano1 had no effect on contractile parameters. The lack of Ca^{2+} transients in PDGFRα^{+} cells across any stage of the lymphatic contraction cycle also diminishes any expected role for this cell type to perform as the pacemaker for the mouse IALV. We recently showed that pressure-dependent Ca^{2+} mobilization from the SR, through IP3R1 (Zawieja et al., 2023), sets the basis for LMC pacemaking as previously proposed (Van Helden, 1991; von der Weid et al., 2008). However, the mechanisms driving IP3R1 activation and Ca^{2+} oscillations remain to be fully addressed.

A pacemaker cell would be expected to be electrically coupled to the LMC layer to permit the nearly synchronous conduction velocity of the contraction wave (Zawieja et al., 1993; Castorena-Gonzalez et al., 2018b; Hald et al., 2018) and to transmit depolarization into coupled LMCs to activate the voltage dependent Ca^{2+} channels that are responsible for lymphatic muscle APs. Connexins are the molecular constituents of gap junctions and, as stated above, we detected Cx45 expression in *PdgfrαCreER*^{TM} sorted cells. However, we did not detect any impairment in pacemaking, nor were contraction conduction speed deficits or multiple pacemakers noted in the *PdgfrαCreER*^{TM} -Cx45fl/fl popliteal cLVs, in contrast to the development of multiple pacemaker sites and the lack of entrainment that characterize cLVs from *Myh11CreER*^{T2}-Cx45$^{fl/fl}$ mice (Castorena-Gonzalez et al., 2018b). Admittedly, we did not perform an exhaustive assessment of the connexin expression profile of the CD34^{+}PDGFRα^{+} cells, and Cx45 may not be the dominant connexin expressed in the CD34^{+}PDGFRα^{+} cells, or heterotypic connexons could exist (Koval et al., 2014). However, electron microscopy studies of the putative ICLC in the human thoracic duct did not detect any gap junctions, although peg-and-socket connections were observed (Briggs Boedtkjer et al., 2013). We utilized optogenetics to directly depolarize the specific cell populations in both the *PdgfrαCreER*^{TM} and *Myh11CreER*^{T2} mouse models in an attempt to drive contractions. Local photo-stimulation of the PDGFRα cells failed to initiate contraction while the stimulation of *Myh11CreER*^{T2} recombined cells resulted in contractions that were indistinguishable from the spontaneously occurring contractions. These results give functional credence to the lack of hetero-cellular coupling via gap junctions that was previously reported (Briggs Boedtkjer et al., 2013). Just as critically, our results also highlight the regenerative nature of the lymphatic muscle AP. Local, optogenetic-initiated depolarization of either a single or a few LMCs to threshold was sufficient to drive a coordinated contraction along the vessel conducted activity at the tissue level.

Conclusions

Our present findings lend further support to the hypothesis that the LMCs are intrinsic pacemakers (van Helden et al., 2006; Mitsui and Hashitani, 2020) and that mouse cLVs do not require an ICC-like cell network to drive propagated contractions. These findings also underscore the significance of lymphatic muscle Ca^{2+} handling as the driver of lymphatic pacemaking, which can be compromised in disease states leading to impaired lymphatic contractile activity (Stolarz et al., 2019; Lee et al., 2020; Van et al., 2021). Further studies delineating the specific SR Ca^{2+} release and influx pathways, and the contributions of Ca^{2+} sensitive ion channels are required to develop sophisticated in silico models and identify potential therapeutic targets to rescue lymphatic pacemaking in lymphedema patients (Olszewski, 2002, 2008).

Limitations

One fundamental assumption underlying our conclusions is that there is a conserved pacemaking pathway and cell type regulating lymphatic collecting vessel contractions across species, specifically pertaining to the capability of lymphatic muscle to maintain pacemaking and coordination despite changes in tissue complexity and cLV wall thickness. It is worth noting that lymphatic collecting vessels in mice have similar pressure-dependent chronotropy and contraction conduction velocity as recorded in rats and human vessels (Castorena-Gonzalez et al., 2018b). These similarities exist despite the fact that mouse lymphatic collecting vessels are typically encircled by a single layer of lymphatic muscle while larger species may have multiple layers of LMCs in the wall. It is possible that vessels with multiple layers of LMCs need a network of ICLC to coordinate their activity. The simplicity in the makeup of the mouse cLV and the use of cell targeting Cre models provides great control over experimental variables, but other cell types may be required for coordination of LMC pacemaking in other species where the lymphatic cLV walls are larger and thicker and contain multiple muscle cell layers. Our scRNAseq analysis also is likely biased using ROSAmTmG mice with FACS purification to remove debris and concentrate specific cell types from these pooled small vessels. Larger and more complex cells, with attributes that can be ascribed to ICCs, are more likely to be lost in this methodology (e.g.., depending on the FACS gating parameters) and this procedure can also elicit a stress response in the transcriptome of the analyzed cells. However, we also did not observe long and complex cells, aside from the circumferential LMCs, in our immunofluorescence and recombination reporter imaging experiments. Immediate and early gene expression motifs driven by a stress response may be a component of the differences in sub-clusters that were identified. Future scRNAseq or snRNAseq studies with deeper sequencing will be required to ensure that the full transcriptomic heterogeneity is accounted under different cellular stress conditions.

Our data demonstrate that limited staining of a few cell markers alone is insufficient to identify discrete cell populations in mouse cLVs. Additionally, mRNA expression does not equal protein translation nor guarantee specific function as we did not readily detect endothelial CD34 with immunofluorescence despite detecting transcript; additionally, *PdgfrαCreER*^{TM}-mediated deletion of Ano1, Cx45, or Cav1.2 had no effect on cLV pacemaking. Further experimentation is also required to fully characterize expression of multipotent cell markers and function of CD34^{+}PDGFRα^{+}Sca1^{+} cells invested within the mouse cLVs, although doing so was beyond the scope of this study assessing pacemaker identity. Tangentially, another limitation of our approach pertains to the specificity and recombination efficiency of inducible Cre recombinase models, which can be a notable confounding variable (Chakraborty et al., 2019). We observed that our inducible Cre models led to a degree of nonspecific recombination within the mouse cLV, with GCaMP6f and ChR2 particularly susceptible to recombination compared to the ROSA26mT/mG reporter. Recombination in multiple cell types was expected with the constitutive Cre models we employed (*Ng2Cre* and *PdgfrαCre*), as vascular and lymphatic muscle precursor cells can transiently express *Nestin*, *Pdgfrα*, and *Ng2* (Hill et al., 2015; Castorena-Gonzalez et al., 2018b; Kenney et al., 2020). We also observed that *PdgfrβCreER*^{T2} drove recombination in a sub population of LMCs and PDGFRα^{+} cells. These appeared to be two distinct populations that only share expression for PDGFRβ based on our scRNAseq dataset, but which may exist along a continuum of differentiation. PDGFB-PDGFRβ signaling is critical for normal mural cell recruitment to both the blood and lymphatic vasculature (Gaengel et al., 2009; Wang et al., 2017) and proliferating vascular smooth muscle cells and pericytes have both been documented to express PDGFRβ (Andrae et al., 2008; Pitulescu and Adams, 2014). Ideally, novel Cre or combinatorial Cre models that specifically target LMCs or sub populations of LMCs may be developed to further tease out the functional roles of these cells.

Materials and Methods

Mice

Wild-type (WT) male mice (25-35 g) on the C57BL/6J background , ROSA26mT/mG reporter (Muzumdar et al., 2007) (Strain#007676), transgenic *PdgfrαCre* (Strain#013148), CSFR1-EGFP

(MacGreen) (Sasmono et al., 2003) (Strain#018549), genetically encoded Ca^{2+} sensor GCaMP6f (Chen et al., 2013) (Strain#028865), transgenic *PdgfrαCreER*^{TM} (Kang et al., 2010) (Strain#018280), NG2-Cre (Strain #:008533) (Zhu et al., 2008), and ChR2/tdTomato fusion mice (Madisen et al., 2012) (Strain#012567) were purchased from The Jackson Laboratory (Bar Harbor, MA, USA). *PdgfrβCreER*^{T2} (Gerl et al., 2015) mice were a gift from Ralf Adams (Mac Planck Institute) and kindly provided by Lorin Olson (Oklahoma Medical Research Foundation) and are currently available at Jax (Strain#029684). The *Myh11CreER*^{T2} mice (Wirth et al., 2008) were a gift from Stefan Offermanns, Max-Planck-Intstitut fur Herz-und Lungendforschung, Bad Nauheim, Germany, and are currently available at Jax (Strain #019079, Y-Linked). *c-KitCreER*^{T2} mice (Heger et al., 2014) were a gift from Dieter Saur (Technical University of Munich). *Prox1-eGFP* mice (Choi et al., 2011) were a gift from Young-Kwon Hong (University of Southern California). For genotyping, we isolated genomic DNA from mouse tail clips using the HotSHOT method (Truett et al., 2000). Specific mouse genotypes were confirmed via PCR using 2x PCR Super Master Polymerase Mix (Catalog # B46019, Bimake, Houston, TX) performed as specified by the provider. Mice used for this study were 3-12 months of age. All animal protocols were approved by the University of Missouri Animal Care and Use Committee and conformed to the US Public Health Service policy for the humane care and use of laboratory animals (PHS Policy, 1996).

iCre Tamoxifen Induction

Mice harboring *PdgfrαCreER*^{TM}, *PdgfrβCreER*^{T2}, *Myh11CreER*^{T2}, and *cKitCreER*^{T2} were crossed with ROSA26mT/mG mice to generate *PdgfrαCreER*^{TM}-ROSA26mT/mG, *PdgfrβCreER*^{T2}-ROSA26mT/mG, *Myh11CreER*^{T2}-ROSA26mT/mG, and *cKitCreER*^{T2}-ROSA26mT/mG mice, respectively. The resulting iCre-ROSA26mT/mG mice were induced with tamoxifen 2-4 weeks after weaning for confocal imaging. Mice aged 2-6 months were injected with tamoxifen for contraction studies, FACs analysis, GCaMP6f imaging, and Chr2 induction. Tamoxifen induction was performed via consecutive 100 μL i.p. injections of tamoxifen ranging from 1 to 5 days at concentrations ranging from 0.2 -10 mg/mL in safflower oil, using a titrated induction protocol to determine the extent of recombination in specific cell populations. We used our maximal induction protocol, 100 μL of tamoxifen at 10 mg/mL over 5 consecutive days, for *cKitCreER*^{T2}-GCaMP6f, *Myh11CreER*^{T2}-GCaMP6f, and *PdgfrαCreER*^{TM} -GCaMP6f mice. Due to the paucity of recombined cells in the *cKitCreER*^{T2}-ROSA26mT/mG reporter mice, we used our maximal tamoxifen induction protocol for *cKitCreER*^{T2}-ChR2/tdTomato mice as this still resulted in the ability to excite single recombined cells. *Myh11CreER*^{T2}-ChR2/tdTomato mice were induced with one 100 μL i.p. injection of tamoxifen at 0.2 mg/mL while *PdgfrαCreER*^{TM}-ChR2/tdTomato were induced with 1 injection at 0.4 mg/mL tamoxifen to get mosaic induction sufficient for single cell stimulation. All mice, regardless of induction duration, were given at least 2 weeks to recover following tamoxifen injection.

Lymphatic Vessel Isolation

We utilized both popliteal and inguinal-axillary lymphatic collecting vessels (IALVs) in this study, which were isolated as described previously (Zawieja et al., 2018a). In brief, mice were anaesthetized with a cocktail of 100/10 mg/mL ketamine/xylazine and shaved along the flank or the legs for IALVs and popliteal cLVs respectively. The IALV (also referred to as the flank cLV) is located adjacent to the thoracoepigastric vein and connects the inguinal and axillary lymph nodes. A cut was made along the dorsal midline and the skin retracted and pinned out to reveal the thoracoepigastric vascular bed. The thoracoepigastric vascular bed and connected perivascular adipose containing the IALV vessels was dissected out and pinned onto a Sylgard coated dish in Krebs buffer. Popliteal lymphatic vessels were exposed through a superficial incision in the leg, removed and transferred to the Krebs-albumin filled dissection chamber. After removal, the vessel was carefully cleaned of adipocytes and excess matrix using fine forceps and scissors through micro-dissection. For immunofluorescence, sections containing 2-3 valves were isolated, while shorter IALV sections consisting of 1-2 valves were isolated for GCaMP6f Ca^{2+} imaging. Similarly, popliteal cLVs were isolated (Castorena-Gonzalez et al., 2018a) following an incision

along the skin overlying the saphenous, removed and transferred to the Krebs-albumin filled dissection chamber; these vessels were used for ChR2 optogenetic depolarization experiments.

Lymphatic Vessel Isobaric Function

PdgfrαCreER^{TM} mice were crossed with *Ano1*$^{fl/fl}$, *Cx45*$^{fl/fl}$, and *Cav1.2*$^{fl/fl}$ mice to generate *PdgfrαCreER*^{TM}*-Ano1*$^{fl/fl}$, *PdgfrαCreER*^{TM}*-Cx45*$^{fl/fl}$, and *PdgfrαCreER*^{TM}*-Cav1.2*$^{fl/fl}$ mice. These mice and their respective fl/fl controls were injected with tamoxifen as described above for 5 days and given two weeks to recover. The popliteal vessels were isolated, cleaned, and prepared for isobaric contractile tests as previously reported (Davis et al., 2023a). Once equilibrated, inner diameter was tracked over a physiological pressure range (stepped from 3 to 2, 1, 0.5, 3, 5, 8, and 10 cmH_2O) with 2min of recording at each pressure. Following the pressure step protocol the vessels were equilibrated in with Ca^{2+}-free Krebs buffer (3mM EGTA) and diameter at each pressure recorded under passive conditions (DMAX). The contractile parameters end diastolic diameter (EDD), end systolic diameter (ESD), and contraction frequency (FREQ) were recorded with a custom LabVIEW program and the following contractile parameters assessed:

1) Contraction Amplitude (AMP) = EDD−ESD

2) Normalized Contraction Amplitude = ((EDD−ESD)/DMAX) × 100

3) Ejection Fraction (EF) = $(EDD^2-ESD^2)/EDD^2$

4) Fractional Pump Flow (FPF) = EF × FREQ

5) Tone = ((DMAX−EDD)/DMAX) × 100

Methylene Blue Staining

Isolated IALVs sections were transferred into a Krebs-BSA buffer filled 3-mL observation chamber, with a cover slip bottom, and cannulated onto two glass micropipettes (30-80 μm, outer diameter) held in place by pipette holders on a Burg-style V-track mounting system. The pipette holders were attached to a 3-way valve stop cock with polyethylene tubing filled with Krebs-BSA buffer. Vessels were pressurized to approximately 5 cmH_2O by raising the 3-way valve and the vessels were stretched to remove any slack. For methylene blue (Sigma, M9140) staining, IALVs from wild type C57Bl6 mice were stained with 50 μM methylene blue in Krebs-BSA buffer for two hours at room temperature and covered in foil to limit light-induced phototoxicity. After the staining period, the vessel chambers were washed three times with Ca^{2+} free PSS to remove methylene blue. Brightfield images and manual Z-stack videos were collected on an inverted Leica DMi1 4X or 20X air objective, or a Leica DMi8 with a 25X water objective or an inverted DMi8 using a Leica Flexacam C1 color camera for image acquisition. Some methylene blue images were also collected using a color Nikon DS-Fi3 camera. The collected z-stacks were analyzed using Image J and the "Stack Focuser" plugin (https://imagej.nih.gov/ij/plugins/stack-focuser.html). To accentuate the methylene blue stained cells, the color image stack was split into red, green, and blue channel stacks. The blue channel stack was then divided by the green channel stack using the "Image Calculator" function. The resulting 32-bit image was then converted into 16-bit image to permit the use of the Stack Focuser plugin with the 'n kernel value' set to 11.

Fluorescence Confocal Imaging

IALVs vessels from each respective iCre-ROSA26mT/mG mouse were prepared in a similar manner (excluding the addition of methylene blue). We performed confocal imaging to acquire z-stacks of 7-10 overlapping regions of interests to allow for manual stitching, with 1 μm z-steps at (20X) or 0.5 μm

steps at 40X. We imaged through to the midpoint of the vessel except when imaging the valve interstitial cells, in which case the entire vessel was imaged. Max projections were made using FIJI. Following live imaging, the vessels were pressurized to 5 cmH_2O and fixed with 4% paraformaldehyde for 30 min at room temperature. IALVs were then washed with PBS containing 0.1% Triton X-100 (PBST) 3 times and blocked for a minimum of 2 hr with Blockaid® (B-10710, ThermoFisher Scientific). IALVs were then stained with the corresponding primary antibodies in BlockAid® Solution: anti-smooth muscle actin (SMA) 1:500 (Sigma, A2547), anti-GFP 1:200 (ThermoFisher, A11122), anti-cKIT 1:100 (Cell Signaling, 3074), anti-VIMENTIN 1:100 (Thermofisher , OMA1-06001), anti-desmin 1:200 (Invitrogen, PA5-16705), anti-GFP 1:200 (Abcam, ab13970, anti-CD34 1:200 (Invitrogen, 14-0341-82), anti-PDGFRA 1:200 (R&DSystems, AF1062), anti-PDGFRβ 1:200 (eBiosciences, 14-1402-82), anti-calponin 1:500 (Abcam, AB46794), anti-MYH11 1:500 (Abcam, AB124679), anti-Sca1 1:200 (Biolegend, 108101). IALVs were then washed in PBS and incubated overnight with the corresponding donkey secondary antibodies (ThermoFisher®) at 1:200. After a final wash, IALVs were re-cannulated and pressurized for imaging using the spinning disk confocal microscope and Hamamatsu Orca Flash4 camera using a 20X air objective (Olympus UplanApo, 0.75) or 40X (Olympus UApo A340, 1.15) water objective. Images were taken as described above, and the resulting stacks were turned into a max projection using FIJI. Colocalization analysis of the max projections of CD34 and PDGFRα was performed using the BIOP JACoP colocalization plugin (Bolte and Cordelieres, 2006) with both Pearson's and Mander's coefficients reported.

LMC Dissociation and FACS Collection-

IALVs vessels *PdgfrαCreER^{TM}*-ROSA26mT/mG, *PdgfrβCreER^{T2}*-ROSA26mT/mG, *Myh11CreER^{T2}*-ROSA26mT/mG, Macgreen, and *Prox1-eGFP* mice were dissected and cleaned of excess adventitia and adipose tissue in Krebs buffer. Isolated vessels were then transferred into a low Ca^{2+} PSS solution supplemented with 0.1 mg/mL bovine serum albumin (BSA, Amersham Life Science, Arlington Heights, IL). Primary LMCs were collected by enzymatic dissociation of IALVs. The dissected vessels were cleaned in room temperature Krebs-BSA buffer and then transferred into a 1-mL tube of low-Ca^{2+} PSS on ice, washed, and equilibrated for 10 min. Vessels were then digested in low-Ca^{2+} PSS with 26 U/mL papain (Sigma, P4762) and 1 mg/mL dithioerythritol for 30 min at 37°C with gentle agitation every few minutes. This solution was then decanted and replaced with low-Ca^{2+} PSS with containing 1.95 collagenase H (U/mL, Sigma), 1.8 mg/mL collagenase F (Sigma), and 1mg/mL elastase (Worthington LS00635) and incubated for 3 - 5 min at 37°C. The mixture was then spun down at 1000 rpm for 4 min, the digestion buffer removed, and replaced with low-Ca^{2+} PS. This process was repeated twice to remove residual digestion buffer. The vessel was then triturated with a fire-polished Pasteur pipette to dissociate the cells into a single cell suspension, passed through a Falcon cap strainer (35 μm), and resuspended in ice-cold low-Ca^{2+} PSS for sorting. For iCre-ROSA26mT/mG mice, GFP^+RFP^- cells or GFP^+ cells from Macgreen and *Prox1-eGFP* mice were then FACS purified straight into RNA isolation buffer for RT-PCR analysis. FACs was performed with a Beckman-Coulter MoFlo XDP instrument using an excitation laser (488 m) and emission filter (530/40 m). Sorting was performed using 70-µm nozzle at a sheath pressure of 45 p.s.i. and sort rate of 100 events/s and with an efficiency of >90%. To maximize cell yield, we isolated both the left and right full-length IALVs vessels from 2 mice for digestions and subsequent FACS collection. For *Myh11CreER^{T2}*-ROSA26mT/mG and *PdgfrαCreER^{TM}*-ROSA26mT/mG, the yield averaged 1,000-2,000 cells per mouse. For *Prox1-eGFP* mice, LEC yield was typically 1,500-2,000 cells per mouse.

RT-PCR Profiling of FACS Purified Cells- Total RNA was extracted from FACS purified GFP^+ cells from the isolated IALVs vessels using the Arcturus PicoPure RNA isolation kit (ThermoFisher Scientific, Waltham, MA) per the listed instructions. Prior to elution in 20 µl of water, on-column DNAse digestion (Qiagen, Valencia, CA) was performed to ensure removal of genomic DNA contaminants. RNA was converted into cDNA using SuperScript III First-Strand Synthesis System (Thermo Fisher Scientific, Waltham, MA) using oligo (dT) and random hexamer priming following the manufacturer's protocol. Each RT reaction used approximately 50-100 cells worth of RNA based on the sorted cells count number. Our

PCR reaction mixture contained first-strand cDNA as the template, 2 mM $MgCl_2$, 0.25 μM primers, 0.2 mM deoxynucleotide triphosphates; and GoTaq® Flexi DNA polymerase (Promega, Madison, WI). The PCR program comprised an initial denaturation step at 95°C for four min; followed by 35 repetitions of the following cycle: denaturation (94° C, 30 s), annealing (58° C, 30 s), and extension (72° C, 30 s). This was followed by a final elongation step for 5 min at 72° C. PCR amplification products were separated on a 2% agarose gel by electrophoresis, stained with SYBR-Safe (Thermo Fisher Scientific, Waltham, MA), and visualized by UV trans-illumination. All primers were designed to amplify an intron-spanning region. Endpoint RT-PCR Primer sequences, amplicon size, accession numbers, and source are listed in Table 1.

scRNASeq Analysis of Mouse IALVs

For scRNASeq analyses of isolated IALVs we used a total of 10 *Rosa26mTmG* mice, without Cre expression and without tamoxifen treatment, with equivalent representation of sex (5 males and 5 females with ages between 10-12 months). Full length IALVs from both the left and right side of each *ROSA26mTmG* mouse were isolated and cleaned of excessive matrix and adipose tissue. Isolated vessels were digested into single cell suspensions as described above and the cells were kept on ice following single cell suspension until all the tissues had been processed. Cells from all vessels were combined and sorted for tdTomato expression to remove debris and concentrate the cells for downstream single cell 3' RNA-Seq libraries creation with 10x Genomics Chromium Chip and Chromium Next GEM Single Cell 3' RNA-Seq reagents. Samples were sequenced with the NovaSeq 6000 S4-PE100 flow cell.

Mus musculus genome GRCm39 and annotation GTF (v106) from Ensembl (https://useast.ensembl.org/Mus_musculus/Info/Index) were used to build the reference index and the reads were processed using Cell Ranger (v7.0.1; Zheng et al., 2017) with default parameters. The quality control and filtering steps were performed using R (v4.2.1; https://www.r-project.org/). Ambient RNA was removed from the Cell Ranger output with SoupX (Young and Behjati, 2020). Doublet score for each cell was estimated using scDBlFinder (v1.12.0; Germain et al., 2021). Non-expressed genes (sum zero across all samples) and low-quality cells (>10% mitochondrial genes, < 500 genes, < 1,000 UMIs per cell and doublet score <0.5) were removed with custom R scripts. Of 10,188 cells, 7,435 passed our inclusion criteria and included three dominant clusters including LECs (2,962 cells), LMCs (978 cells), and fibroblast (2,261 cells) with the remaining cells comprising immune cells (1,147 cells) and some mammary epithelial cell contamination (87 cells). Cells passing filtering were normalized/scaled (SCTransformation), dimensionally reduced (t-distributed stochastic neighbor embedding (t-SNE) and uniform manifold approximation and projection (UMAP)) clustered, and hierarchically analyzed with Seurat (Hao et al., 2021; Hao et al., 2024) with default parameters. To select the optimal cluster resolution, we used Clustree with various resolutions. We examined the resulting tree to identify a resolution where the clusters were well-separated and biologically meaningful, ensuring minimal merging or splitting at higher resolutions. Our goal was to find a resolution that captured relevant cell subpopulations while maintaining distinct clusters without excessive fragmentation. Initial clustering of the entire population of cells was done at a resolution of 0.8 and 18 PCs to achieve the UMAP of 19 cell clusters as shown in Figure 5a. We used a resolution of 0.5 for sub-clustering LMCs (original groups 5 and 6), 0.87 for LECs (original groups 0,1,2,11), and 1.0 for fibroblasts (3,7,8,9,10,13). Marker gene expression profile on cell clusters and gene co-expression was visualized using Seurat and ShinyCell R application (Ouyang et al., 2021). The full scRNAseq raw dataset has been uploaded to the NIH GEO under the accession number GSE277843. Differential gene expression within subclusters of LECs, LMCs, and AdvCs was performed using Seurat's "Find Markers" function and with a minimum of either 40% or 50% cell expression and average log fold change (Log2FC) minimum of 0.5 or 1. When assessing the LMC IP3 receptor genes *Itpr1-3* and *Itprid2* a percent cell expression of 40% and Log2FC of 0.25 was used. In the volcano plot for LEC subcluster 8 differential gene expression, listed genes had a cutoff of a Log2FC of 2 or -2 to be displayed on the plot.

***Ex vivo* Ca^{2+} imaging with the genetically encoded GCaMP6f Indicator**

cKitCreER^{T2}, *Myh11CreER^{T2}*, and *PdgfrαCreER^{TM}* mice were crossed with GCaMP6f mice in a similar manner as described for ROSA26mT/mG. *cKitCreER^{T2}*-GCaMP6f, *PdgfrαCreER^{TM}*-GCaMP6f, and *Myh11CreER^{T2}*-GCaMP6f were induced with tamoxifen (10 mg/mL) for 5 consecutive days by i.p. injection. IALVs isolated from *cKitCreER^{T2}*-GCaMP6f, *PdgfrαCreER^{TM}*-GCaMP6f, and *Myh11CreER^{T2}*-GCaMP6f were cannulated as described above. The cannulated vessel, with micropipette holders, observation chamber and V-track mounting system, was transferred to the stage of the spinning disk confocal with a Prime95B scMOS camera (Photometrics), a Cascade II EMCCD (Photometrics), or an Ixon888 EMCCD camera (Andor) for Ca^{2+} imaging (Castorena-Gonzalez et al., 2018b). Pressures for the input and output cannula were connected to a T-junction which was set briefly to 8 cmH_2O and the vessel lengthened to remove axial slack. A peristaltic pump maintained constant perfusion of the observation chamber with Krebs buffer at a rate of 0.5 mL/min while the vessel equilibrated at 37°C for 30-60 min with pressures set to 3 cmH_2O. Spontaneous contractions were allowed to stabilize over a period of 30 min to verify normal function and then were blunted with 2 μM wortmannin to limit movement associated with contractions during Ca^{2+} imaging. A Windows-based computer was used to digitize the pressure transducer signals and video image of the vessel from a firewire camera at 30-40 Hz (Davis et al., 2012). A custom-written LabVIEW program (National Instruments; Austin, TX) detected the inner diameter of the vessel from the video (Davis, 2005). Once contractions were <5 μm in amplitude, Ca^{2+} recordings were made at 20FPS for 20-40 s.

Ca^{2+} Imaging and Analysis in IALVs Over the Contraction Cycle

Background noise was determined by using the histogram feature of FIJI in a rectangle in a region of the field of view without sample. This value was subtracted from the entire field of view. In some cases, the vessel movement due to contraction was offset with video stabilization with the FIJI plugin Image Stabilizer. A max projection was used to create non-overlapping ROIs of GCaMP6f^{+} cells for each iCre-GCaMp6f IALV. From these cell ROIs, the "reslice z" function was used to create pseudo-linescan STMs, which were divided by their baseline values to obtain F/F_0 values for each individual cell. At least 3 cells, except in the case of 1 *cKitCreER^{T2}*-GCaMp6f IALV, in which only two cells were observed, were analyzed in this manner for each vessel segment. Max projections of the image stack were then used to create non-overlapping cell masks of 3-5 muscle cells per field of view of one vessel. Ca^{2+} traces for those cells contained 5-10 contraction cycles and Ca^{2+} transients and were characterized for peak intensity (expressed as a baseline-referenced ratio, F/F_0), frequency, and duration in seconds.

Analysis of Subcellular Ca^{2+} Transients in *Myh11CreER^{T2}*-GCaMP6f IALVs

For *Myh11CreER^{T2}*-We performed Ca^{2+} imaging as above in the presence of 1 μM nifedipine to stop the "Ca^{2+} flashes" associated with APs (Zawieja et al., 2018a) and focus on the subcellular activity at 3 different experimental pressures of 0.5, 2, and 5 cmH_2O. For this protocol, we used a particle analysis approach to analyze all Ca^{2+} transients in the field of view. Ca^{2+} transients in intact vessels were quantified by particle analysis as previously described (Drumm et al., 2017; Drumm et al., 2019b). Movies of Ca^{2+} transients in intact vessels were imported into custom built Volumetry software (version G8d) and background subtracted. Movies were smoothed using a Gaussian filter: 1.5 x 1.5 mM, StdDev 1.0). Raw Ca^{2+} transients were converted to Ca^{2+} particles (PTCLs) using a flood-fill algorithm as previously described (Drumm et al., 2017; Drumm et al., 2019b). PTCLs <10 μM^2 were rejected to facilitate the removal of noise and then the total PTCL area and PTCL count could be tabulated for each recording.

Membrane Potential Recordings in IALVs

Mouse IALVs were isolated and cleaned as described above. IALVs were pressurized in our isobaric myography apparatus and allowed to equilibrate to ensure typical contractile activity was evident. A bolus of wortmannin at 2 μM was then applied to the bath to blunt contraction amplitude below 5 μm. Intracellular recordings of lymphatic muscle were made with microelectrodes (250–300 MΩ) filled with 1 M KCl and an SEC-05x amplifier (NPI) connected to a Grass S48 stimulator, viewed with a Tektronix TDS3052 digital oscilloscope. Membrane potential and diameter were simultaneously recorded using a

custom Labview program. Membrane potential and APS were allowed to stabilize and then pressure was slowly raised from 0.5cmH_2O to 2cmH_2O and then 5cmH_2O. In some cases, the electrode dislodged due to the intrinsic contractions of the vessel or wall displacement as pressure was modulated. In these situations, we attempted to re-impale the cell or one of the neighboring cells. Only vessels in which a recording with a minimum of 3 stable APs was successfully recorded at 2 of the 3 experimental pressures were used for subsequent analysis.

We also confirmed LMC impalement using microelectrode filled with 1 M KCl and (100 µg/ml) Biocytin-AF488 (A12924, ThermoFisher) to label impaled cells that displayed APs, over a 10-minute recording period. Following the impalement and loading with Biocytin-AF488 the vessel was transferred to our imaging apparatus for confocal imaging and 3D reconstruction using the Andor Dragonfly 200 and IMARIS. Image stacks were taken with a 25x water objective at 0.5-micron intervals throughout the diameter of the vessel.

Light Activation of ChR2 to stimulate Popliteal Collecting Lymphatic Vessel Contractions

As the IALV has a nearly continuous contractile cycle, we utilized the popliteal vessel for its much slower contraction frequency in the experiments testing our ability to trigger a propagated contraction upon stimulation of the enforced expression of ChR2. Popliteal vessels were isolated from *cKitCreER*^{T2}-ChR2/tdTomato, *PdgfrαCreER*^{TM} -ChR2/tdTomato, or *Myh11CreER*^{T2}-ChR2/tdTomato mice (3-9 months of age) as previously described (Scallan and Davis, 2013), although we intentionally retained some connective tissue and adipose tissue to ensure we had a sufficient population of recombined cells to test in the adventitia layer of the vessel. Contractions were allowed to stabilize over a 30-min equilibration period with pressure set to 3 cmH_2O. If basal contraction frequency was too high, we applied pinacidil to the bath in 100 nM increments, without exceeding 600 nM, to further slow contraction frequency to around 6 contractions per minute. Pinacidil at sub 1 µM doses can slow contraction frequency without causing overt hyperpolarization of membrane potential (Davis et al., 2020). Supplemental 100 nM doses of pinacidil were applied throughout the experiment to maintain a spontaneous contraction frequency below 6 per minute to allow ample diastolic time for ChR2 stimulation. Throughout this protocol the popliteal was allowed to contract spontaneously to ensure we had not overly inhibited APs by the pacemaking cells with pinacidil. Occasionally, spontaneous contractions occurred just prior to light-evoked contractions, resulting in a potential false positive, so we performed multiple stimulations over a period of 5 - 10 min, typically waiting at least 3 s after any spontaneous contraction before stimulating. Care was made to align the light fiber in such a way that only part of the vessel would be directly illuminated and so target cells of interest would be directly activated by 473 nm light using a Laser diode (Doric LD Fiber Light Source, Quebec, Canada), through an optical probe with a 10-µm tip (Doric, OPT_200_0.22.010). To further limit the excitation field, the optical probe was coated with black acrylic paint using an eyelash brush so that the uncoated opening was ~2-3 µm. With the probe positioned within 5 µm of one side of the vessel wall, the spread of light covered an area ~10-100 µm wide on the back side of the vessel (depending on the diode amplitude setting). Light pulses, 200 ms in length, were triggered by a Grass S9 stimulator (Harvard Apparatus, Holliston, MA) connected to the external TTL input of the laser diode. Pulse amplitude was adjusted between 40-90 mA using the Laser Diode Module Driver (Doric). A contraction was considered to be triggered if it occurred within 50ms of stimulation. We performed photo-stimulation from 2-4 sites within each vessel, with 6-14 stimulations per site. If a photo-stimulation was triggered incidentally after the initiation of a "spontaneous contraction" it was discarded from the analysis. For *Myh11CreER*^{T2}-ChR2-tdTomato 6 vessels from 3 separate mice were tested. For *PdgfrαCreER*^{TM}-ChR2-tdTomato 6 vessels from 4 separate mice were tested with a max of two vessels per mouse. For *cKitCreER*^{T2}- ChR2-tdTomato 7 vessels from 3 separate mice were assessed. Diameter was recorded to align photo-activation with the contraction cycle in a custom Labview program.

Solutions and Chemicals.

Krebs buffer was composed of (in mM): 146.9 NaCl, 4.7 KCl, 2 $CaCl_2$, 1.2 $MgSO_4$, 1.2 $NaH_2PO_4 \cdot H_2O$, 3 $NaHCO_3$, 1.5 NaHEPES, and 5 d-glucose (pH = 7.4 at 37°C). Krebs-BSA buffer was prepared with the addition of 0.5% (w/v) bovine serum albumin (BSA) while Krebs Ca^{2+}-free replaced $CaCl_2$ with 3mM EGTA. Tamoxifen was dissolved to 10mg/ml in a Safflower Oil-Ethanol (95%-5% v/v) solution with rocking agitation, separated into aliquots, and stored at -20 °C. Wortmannin was dissolved in DMSO to a stock solution of 1mM. Pinacidil was dissolved in DMSO to a stock concentration of 1 µM. Nifedipine was dissolved in DMSO to a stock concentration of 1 mM. All chemicals were obtained from Sigma (St. Louis, MO), except for BSA (US Biochemicals; Cleveland, OH), $MgSO_4$ and NaHEPES (Fisher Scientific; Pittsburgh, PA).

Statistical Tests

Statistical differences in the isobaric contractile tests for popliteal cLVs isolated from *PdgfrαCreER^{TM}-Ano1$^{fl/fl}$*, *PdgfrαCreER^{TM}-Cx45$^{fl/fl}$*, and *PdgfrαCreER^{TM}-Cav1.2$^{fl/fl}$* mice over the various contractile parameters were assessed via 1) repeated measures two-way ANOVAs with Sidak's multiple comparison tests performed using Prism9 (Graphpad). Data are plotted as mean ± SEM and significance determined at $p < 0.05$ and $0.10 > p > 0.05$ were reported. Cre negative mice were used for controls and experiment order was not randomized aside from random mouse selection from cages housing both Cre^+ or Cre^- mice. Experimental sample size was determined by the results from our previous experiments assessing *Cx45*, *Ano1*, *Cacna1c* with *Myh11CreER^{T2}* mice. Data from cLVs in which a negative tone value was recorded at any pressure, which typically indicated incomplete relaxation or occluding bubbles in the cannula, were not included in the tone analysis. Vessels that failed to contract at a given pressure had no value recorded for ejection fraction or normalized amplitude and REML mixed effects model was used in place of repeated measures 2-way ANOVA. We used a categorical Chi-squared statistical test for the experiments assessing our ability to trigger a contraction with activation of ChR2+ cells. Ca^{2+} particle area and frequency were compared using 1-way ANOVA with Tukey's post-hoc test. Significance was determined at $p < 0.05$. A mixed effects analysis with Tukey's multiple comparison post-hoc test was used to compare AP parameters across pressure using Prism9 (Graphpad).

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Data availability
Single-cell mRNA sequencing data generated to support this study have been deposited in NCBI GEO under accession number GEO: GSE277843. The authors declare that all other data supporting the findings

of this study are available within the paper, its supplementary information files, and the uploaded scRNAseq dataset.

References

Aalkjaer, C., D. Boedtkjer, and V. Matchkov. 2011. Vasomotion - what is currently thought? *Acta Physiol (Oxf)*. 202:253-269.

Andrae, J., R. Gallini, and C. Betsholtz. 2008. Role of platelet-derived growth factors in physiology and medicine. *Genes Dev*. 22:1276-1312.

Andrzejewska, A., B. Lukomska, and M. Janowski. 2019. Concise Review: Mesenchymal Stem Cells: From Roots to Boost. *Stem Cells*. 37:855-864.

Atakpa-Adaji, P., A. Ivanova, K. Kujawa, and C.W. Taylor. 2024. KRAP regulates mitochondrial Ca2+ uptake by licensing IP3 receptor activity and stabilizing ER-mitochondrial junctions. *J Cell Sci*. 137.

Baker, S.A., B.T. Drumm, D. Saur, G.W. Hennig, S.M. Ward, and K.M. Sanders. 2016. Spontaneous Ca(2+) transients in interstitial cells of Cajal located within the deep muscular plexus of the murine small intestine. *J Physiol*. 594:3317-3338.

Baker, S.A., S.J. Hwang, P.J. Blair, C. Sireika, L. Wei, S. Ro, S.M. Ward, and K.M. Sanders. 2021a. Ca(2+) transients in ICC-MY define the basis for the dominance of the corpus in gastric pacemaking. *Cell Calcium*. 99:102472.

Baker, S.A., W.A. Leigh, G. Del Valle, I.F. De Yturriaga, S.M. Ward, C.A. Cobine, B.T. Drumm, and K.M. Sanders. 2021b. Ca(2+) signaling driving pacemaker activity in submucosal interstitial cells of Cajal in the murine colon. *Elife*. 10.

Basciani, S., S. Mariani, M. Arizzi, M. Brama, A. Ricci, C. Betsholtz, C. Bondjers, G. Ricci, A. Catizone, M. Galdieri, G. Spera, and L. Gnessi. 2004. Expression of platelet-derived growth factor (PDGF) in the epididymis and analysis of the epididymal development in PDGF-A, PDGF-B, and PDGF receptor beta deficient mice. *Biol Reprod*. 70:168-177.

Behringer, E.J., J.P. Scallan, M. Jafarnejad, J.A. Castorena-Gonzalez, S.D. Zawieja, J.E. Moore, Jr., M.J. Davis, and S.S. Segal. 2017. Calcium and electrical dynamics in lymphatic endothelium. *J Physiol*. 595:7347-7368.

Benoit, J.N., D.C. Zawieja, A.H. Goodman, and H.J. Granger. 1989. Characterization of intact mesenteric lymphatic pump and its responsiveness to acute edemagenic stress. *Am J Physiol*. 257:H2059-2069.

Boedtkjer, D.M., V.V. Matchkov, E. Boedtkjer, H. Nilsson, and C. Aalkjaer. 2008. Vasomotion has chloride-dependency in rat mesenteric small arteries. *Pflugers Arch*. 457:389-404.

Bolte, S., and F.P. Cordelieres. 2006. A guided tour into subcellular colocalization analysis in light microscopy. *J Microsc*. 224:213-232.

Bridenbaugh, E.A., I.T. Nizamutdinova, D. Jupiter, T. Nagai, S. Thangaswamy, V. Chatterjee, and A.A. Gashev. 2013a. Lymphatic muscle cells in rat mesenteric lymphatic vessels of various ages. *Lymphat Res Biol*. 11:35-42.

Bridenbaugh, E.A., W. Wang, M. Srimushnam, W.E. Cromer, S.D. Zawieja, S.E. Schmidt, D.C. Jupiter, H.C. Huang, V. Van Buren, and D.C. Zawieja. 2013b. An immunological fingerprint differentiates muscular lymphatics from arteries and veins. *Lymphat Res Biol*. 11:155-171.

Briggs Boedtkjer, D., J. Rumessen, U. Baandrup, M. Skov Mikkelsen, N. Telinius, H. Pilegaard, C. Aalkjaer, and V. Hjortdal. 2013. Identification of interstitial Cajal-like cells in the human thoracic duct. *Cells Tissues Organs*. 197:145-158.

Buechler, M.B., R.N. Pradhan, A.T. Krishnamurty, C. Cox, A.K. Calviello, A.W. Wang, Y.A. Yang, L. Tam, R. Caothien, M. Roose-Girma, Z. Modrusan, J.R. Arron, R. Bourgon, S. Muller, and S.J. Turley. 2021. Cross-tissue organization of the fibroblast lineage. *Nature*. 593:575-579.

Castorena-Gonzalez, J.A., M. Li, and M.J. Davis. 2020. Effects of Elevated Downstream Pressure and the Role of Smooth Muscle Cell Coupling through Connexin45 on Lymphatic Pacemaking. *Biomolecules*. 10.

Castorena-Gonzalez, J.A., J.P. Scallan, and M.J. Davis. 2018a. Methods for Assessing the Contractile Function of Mouse Lymphatic Vessels Ex Vivo. *Methods in molecular biology*. 1846:229-248.

Castorena-Gonzalez, J.A., S.D. Zawieja, M. Li, R.S. Srinivasan, A.M. Simon, C. de Wit, R. de la Torre, L.A. Martinez-Lemus, G.W. Hennig, and M.J. Davis. 2018b. Mechanisms of Connexin-Related Lymphedema. *Circ Res*. 123:964-985.

Chakraborty, R., F.Z. Saddouk, A.C. Carrao, D.S. Krause, D.M. Greif, and K.A. Martin. 2019. Promoters to Study Vascular Smooth Muscle. *Arterioscler Thromb Vasc Biol*. 39:603-612.

Chakraborty, S., S.D. Zawieja, W. Wang, Y. Lee, Y.J. Wang, P.Y. von der Weid, D.C. Zawieja, and M. Muthuchamy. 2015. Lipopolysaccharide modulates neutrophil recruitment and macrophage polarization on lymphatic vessels and impairs lymphatic function in rat mesentery. *Am J Physiol Heart Circ Physiol*.ajpheart 00467 02015.

Chatterjee, V., and A.A. Gashev. 2012. Aging-associated shifts in functional status of mast cells located by adult and aged mesenteric lymphatic vessels. *Am J Physiol Heart Circ Physiol*. 303:H693-702.

Chen, T.W., T.J. Wardill, Y. Sun, S.R. Pulver, S.L. Renninger, A. Baohan, E.R. Schreiter, R.A. Kerr, M.B. Orger, V. Jayaraman, L.L. Looger, K. Svoboda, and D.S. Kim. 2013. Ultrasensitive fluorescent proteins for imaging neuronal activity. *Nature*. 499:295-300.

Cheng, X., J. Liu, M. Asuncion-Chin, E. Blaskova, J.P. Bannister, A.M. Dopico, and J.H. Jaggar. 2007. A novel Ca(V)1.2 N terminus expressed in smooth muscle cells of resistance size arteries modifies channel regulation by auxiliary subunits. *J Biol Chem*. 282:29211-29221.

Choi, I., H.K. Chung, S. Ramu, H.N. Lee, K.E. Kim, S. Lee, J. Yoo, D. Choi, Y.S. Lee, B. Aguilar, and Y.K. Hong. 2011. Visualization of lymphatic vessels by Prox1-promoter directed GFP reporter in a bacterial artificial chromosome-based transgenic mouse. *Blood*. 117:362-365.

Clayton, D.R., W.G. Ruiz, M.G. Dalghi, N. Montalbetti, M.D. Carattino, and G. Apodaca. 2022. Studies of ultrastructure, gene expression, and marker analysis reveal that mouse bladder PDGFRA(+) interstitial cells are fibroblasts. *Am J Physiol Renal Physiol*. 323:F299-F321.

Cobine, C.A., E.E. Hannah, M.H. Zhu, H.E. Lyle, J.R. Rock, K.M. Sanders, S.M. Ward, and K.D. Keef. 2017. ANO1 in intramuscular interstitial cells of Cajal plays a key role in the generation of slow waves and tone in the internal anal sphincter. *J Physiol*. 595:2021-2041.

Davis, M.J. 2005. An improved, computer-based method to automatically track internal and external diameter of isolated microvessels. *Microcirculation*. 12:361-372.

Davis, M.J., J.A. Castorena-Gonzalez, H.J. Kim, M. Li, M. Remedi, and C.G. Nichols. 2023a. Lymphatic contractile dysfunction in mouse models of Cantu Syndrome with K(ATP) channel gain-of-function. *Function (Oxf)*. 4:zqad017.

Davis, M.J., J.A. Castorena-Gonzalez, and S.D. Zawieja. 2023b. Electric field stimulation unmasks a subtle role for T-type calcium channels in regulating lymphatic contraction. *Sci Rep*. 13:15862.

Davis, M.J., H.J. Kim, M. Li, and S.D. Zawieja. 2022. A vascular smooth muscle-specific integrin-alpha8 Cre mouse for lymphatic contraction studies that allows male-female comparisons and avoids visceral myopathy. *Front Physiol*. 13:1060146.

Davis, M.J., H.J. Kim, S.D. Zawieja, J.A. Castorena-Gonzalez, P. Gui, M. Li, B.T. Saunders, B.H. Zinselmeyer, G.J. Randolph, M.S. Remedi, and C.G. Nichols. 2020. Kir6.1-dependent KATP channels in lymphatic smooth muscle and vessel dysfunction in mice with Kir6.1 gain-of-function. *J Physiol*.

Davis, M.J., J.P. Scallan, J.H. Wolpers, M. Muthuchamy, A.A. Gashev, and D.C. Zawieja. 2012. Intrinsic increase in lymphangion muscle contractility in response to elevated afterload. *Am J Physiol Heart Circ Physiol*. 303:H795-808.

Drumm, B.T., G.W. Hennig, S.A. Baker, and K.M. Sanders. 2019a. Applications of Spatio-temporal Mapping and Particle Analysis Techniques to Quantify Intracellular Ca2+ Signaling In Situ. *J Vis Exp*.

Drumm, B.T., G.W. Hennig, S.A. Baker, and K.M. Sanders. 2019b. Applications of spatio-temporal mapping and particle analysis techniques to quantify intracellular Ca 2+ signaling in situ. *Journal of Visualized Experiments*. 2019:1-13.

Drumm, B.T., G.W. Hennig, M.J. Battersby, E.K. Cunningham, T.S. Sung, S.M. Ward, K.M. Sanders, and S.A. Baker. 2017. Clustering of Ca2+ transients in interstitial cells of Cajal defines slow wave duration. *The Journal of General Physiology*. 149:703-725.

Drumm, B.T., B.E. Rembetski, C.A. Cobine, S.A. Baker, G.P. Sergeant, M.A. Hollywood, K.D. Thornbury, and K.M. Sanders. 2018. Ca(2+) signalling in mouse urethral smooth muscle in situ: role of Ca(2+) stores and Ca(2+) influx mechanisms. *J Physiol*. 596:1433-1466.

Engeset, A., W. Olszewski, P.M. Jaeger, J. Sokolowski, and L. Theodorsen. 1977. Twenty-four hour variation in flow and composition of leg lymph in normal men. *Acta physiologica Scandinavica*. 99:140-148.

Ferrusi, I., J. Zhao, D. van Helden, and P.Y. von der Weid. 2004. Cyclopiazonic acid decreases spontaneous transient depolarizations in guinea pig mesenteric lymphatic vessels in endothelium-dependent and -independent manners. *Am J Physiol Heart Circ Physiol*. 286:H2287-2295.

Forte, E., M. Ramialison, H.T. Nim, M. Mara, J.Y. Li, R. Cohn, S.L. Daigle, S. Boyd, E.G. Stanley, A.G. Elefanty, J.T. Hinson, M.W. Costa, N.A. Rosenthal, and M.B. Furtado. 2022. Adult mouse fibroblasts retain organ-specific transcriptomic identity. *Elife*. 11.

Gaengel, K., G. Genove, A. Armulik, and C. Betsholtz. 2009. Endothelial-mural cell signaling in vascular development and angiogenesis. *Arterioscler Thromb Vasc Biol*. 29:630-638.

Gashev, A.A., M.J. Davis, M.D. Delp, and D.C. Zawieja. 2004. Regional variations of contractile activity in isolated rat lymphatics. *Microcirculation*. 11:477-492.

Gashev, A.A., M.J. Davis, and D.C. Zawieja. 2002. Inhibition of the active lymph pump by flow in rat mesenteric lymphatics and thoracic duct. *J Physiol*. 540:1023-1037.

Geng, X., B. Cha, M.R. Mahamud, K.C. Lim, R. Silasi-Mansat, M.K.M. Uddin, N. Miura, L. Xia, A.M. Simon, J.D. Engel, H. Chen, F. Lupu, and R.S. Srinivasan. 2016. Multiple mouse models of primary lymphedema exhibit distinct defects in lymphovenous valve development. *Dev Biol*. 409:218-233.

Gerl, K., L. Miquerol, V.T. Todorov, C.P. Hugo, R.H. Adams, A. Kurtz, and B. Kurt. 2015. Inducible glomerular erythropoietin production in the adult kidney. *Kidney Int*. 88:1345-1355.

Gomez-Pinilla, P.J., S.J. Gibbons, M.R. Bardsley, A. Lorincz, M.J. Pozo, P.J. Pasricha, M. Van de Rijn, R.B. West, M.G. Sarr, M.L. Kendrick, R.R. Cima, E.J. Dozois, D.W. Larson, T. Ordog, and G. Farrugia. 2009. Ano1 is a selective marker of interstitial cells of Cajal in the human and mouse gastrointestinal tract. *Am J Physiol Gastrointest Liver Physiol*. 296:G1370-1381.

Gonzalez-Loyola, A., E. Bovay, J. Kim, T.W. Lozano, A. Sabine, F. Renevey, S. Arroz-Madeira, A. Rapin, T.P. Wypych, G. Rota, S. Durot, D. Velin, B. Marsland, G. Guarda, M. Delorenzi, N. Zamboni, S.A. Luther, and T.V. Petrova. 2021. FOXC2 controls adult lymphatic endothelial specialization, function, and gut lymphatic barrier preventing multiorgan failure. *Sci Adv*. 7.

Grainger, N., C.C. Shonnard, S.K. Quiggle, E.B. Fox, H. Presley, R. Daugherty, M.C. Shonnard, B.T. Drumm, and K.M. Sanders. 2022. Propagation of Pacemaker Activity and Peristaltic Contractions in the Mouse Renal Pelvis Rely on Ca(2+)-activated Cl(-) Channels and T-Type Ca(2+) Channels. *Function (Oxf)*. 3:zqac041.

Hald, B.O., J.A. Castorena-Gonzalez, S.D. Zawieja, P. Gui, and M.J. Davis. 2018. Electrical Communication in Lymphangions. *Biophys J*. 115:936-949.

Hao, Y., S. Hao, E. Andersen-Nissen, W.M. Mauck, 3rd, S. Zheng, A. Butler, M.J. Lee, A.J. Wilk, C. Darby, M. Zager, P. Hoffman, M. Stoeckius, E. Papalexi, E.P. Mimitou, J. Jain, A. Srivastava, T. Stuart, L.M. Fleming, B. Yeung, A.J. Rogers, J.M. McElrath, C.A. Blish, R. Gottardo, P. Smibert, and R. Satija. 2021. Integrated analysis of multimodal single-cell data. *Cell*. 184:3573-3587 e3529.

Hao, Y., T. Stuart, M.H. Kowalski, S. Choudhary, P. Hoffman, A. Hartman, A. Srivastava, G. Molla, S. Madad, C. Fernandez-Granda, and R. Satija. 2024. Dictionary learning for integrative, multimodal and scalable single-cell analysis. *Nat Biotechnol*. 42:293-304.

Hashitani, H., R. Mitsui, S. Masaki, and D.F. Van Helden. 2015. Pacemaker role of pericytes in generating synchronized spontaneous Ca2+ transients in the myenteric microvasculature of the guinea-pig gastric antrum. *Cell Calcium*. 58:442-456.

Heger, K., B. Seidler, J.C. Vahl, C. Schwartz, M. Kober, S. Klein, D. Voehringer, D. Saur, and M. Schmidt-Supprian. 2014. CreER(T2) expression from within the c-Kit gene locus allows efficient inducible gene targeting in and ablation of mast cells. *Eur J Immunol*. 44:296-306.

Hill, R.A., L. Tong, P. Yuan, S. Murikinati, S. Gupta, and J. Grutzendler. 2015. Regional Blood Flow in the Normal and Ischemic Brain Is Controlled by Arteriolar Smooth Muscle Cell Contractility and Not by Capillary Pericytes. *Neuron*. 87:95-110.

Huizinga, J.D., L. Thuneberg, M. Kluppel, J. Malysz, H.B. Mikkelsen, and A. Bernstein. 1995. W/kit gene required for interstitial cells of Cajal and for intestinal pacemaker activity. *Nature*. 373:347-349.

Hwang, S.J., P.J. Blair, F.C. Britton, K.E. O'Driscoll, G. Hennig, Y.R. Bayguinov, J.R. Rock, B.D. Harfe, K.M. Sanders, and S.M. Ward. 2009. Expression of anoctamin 1/TMEM16A by interstitial cells of Cajal is fundamental for slow wave activity in gastrointestinal muscles. *J Physiol*. 587:4887-4904.

Imtiaz, M.S., J. Zhao, K. Hosaka, P.Y. von der Weid, M. Crowe, and D.F. van Helden. 2007. Pacemaking through Ca2+ stores interacting as coupled oscillators via membrane depolarization. *Biophys J*. 92:3843-3861.

Jolly, A.J., S. Lu, K.A. Strand, A.M. Dubner, M.F. Mutryn, R.A. Nemenoff, M.W. Majesky, K.S. Moulton, and M.C.M. Weiser-Evans. 2022. Heterogeneous subpopulations of adventitial progenitor cells regulate vascular homeostasis and pathological vascular remodelling. *Cardiovasc Res*. 118:1452-1465.

Kang, S.H., M. Fukaya, J.K. Yang, J.D. Rothstein, and D.E. Bergles. 2010. NG2+ CNS glial progenitors remain committed to the oligodendrocyte lineage in postnatal life and following neurodegeneration. *Neuron*. 68:668-681.

Kenney, H.M., R.D. Bell, E.A. Masters, L. Xing, C.T. Ritchlin, and E.M. Schwarz. 2020. Lineage tracing reveals evidence of a popliteal lymphatic muscle progenitor cell that is distinct from skeletal and vascular muscle progenitors. *Sci Rep*. 10:18088.

Kenney, H.M., C.L. Wu, A.E. Loiselle, L. Xing, C.T. Ritchlin, and E.M. Schwarz. 2022. Single-cell transcriptomics of popliteal lymphatic vessels and peripheral veins reveals altered lymphatic muscle and immune cell populations in the TNF-Tg arthritis model. *Arthritis Res Ther*. 24:64.

Kimura, K., K. Ramirez, T.A.V. Nguyen, Y. Yamashiro, A. Sada, and H. Yanagisawa. 2021. Contribution of PDGFRalpha-positive cells in maintenance and injury responses in mouse large vessels. *Sci Rep*. 11:8683.

Koval, M., S.A. Molina, and J.M. Burt. 2014. Mix and match: investigating heteromeric and heterotypic gap junction channels in model systems and native tissues. *FEBS Lett*. 588:1193-1204.

Kramann, R., R.K. Schneider, D.P. DiRocco, F. Machado, S. Fleig, P.A. Bondzie, J.M. Henderson, B.L. Ebert, and B.D. Humphreys. 2015. Perivascular Gli1+ progenitors are key contributors to injury-induced organ fibrosis. *Cell Stem Cell*. 16:51-66.

Kurahashi, M., Y. Nakano, L.E. Peri, J.B. Townsend, S.M. Ward, and K.M. Sanders. 2013. A novel population of subepithelial platelet-derived growth factor receptor alpha-positive cells in the mouse and human colon. *Am J Physiol Gastrointest Liver Physiol*. 304:G823-834.

Kurahashi, M., H. Zheng, L. Dwyer, S.M. Ward, S.D. Koh, and K.M. Sanders. 2011. A functional role for the 'fibroblast-like cells' in gastrointestinal smooth muscles. *J Physiol*. 589:697-710.

Leak, L.V., and J.F. Burke. 1968. Ultrastructural studies on the lymphatic anchoring filaments. *The Journal of cell biology*. 36:129-149.

Lee, Y., S. Chakraborty, and M. Muthuchamy. 2020. Roles of sarcoplasmic reticulum Ca(2+) ATPase pump in the impairments of lymphatic contractile activity in a metabolic syndrome rat model. *Sci Rep*. 10:12320.

Lendahl, U., L. Muhl, and C. Betsholtz. 2022. Identification, discrimination and heterogeneity of fibroblasts. *Nat Commun*. 13:3409.

Li, S.W., C.Y. Wang, Y.J. Jou, T.C. Yang, S.H. Huang, L. Wan, Y.J. Lin, and C.W. Lin. 2016. SARS coronavirus papain-like protease induces Egr-1-dependent up-regulation of TGF-beta1 via ROS/p38 MAPK/STAT3 pathway. *Sci Rep*. 6:25754.

Luo, H., X. Xia, L.B. Huang, H. An, M. Cao, G.D. Kim, H.N. Chen, W.H. Zhang, Y. Shu, X. Kong, Z. Ren, P.H. Li, Y. Liu, H. Tang, R. Sun, C. Li, B. Bai, W. Jia, Y. Liu, W. Zhang, L. Yang, Y. Peng, L. Dai, H. Hu, Y. Jiang, Y. Hu, J. Zhu, H. Jiang, Z. Li, C. Caulin, J. Park, and H. Xu. 2022a. Pan-cancer single-cell analysis reveals the heterogeneity and plasticity of cancer-associated fibroblasts in the tumor microenvironment. *Nat Commun*. 13:6619.

Luo, Y., J. Wang, K. Li, M. Li, S. Xu, X. Liu, Z. Zhang, X. Xu, Y. Zhang, J. Pan, P. Liu, S. Gao, Z. Miao, and Y. Yu. 2022b. Single-cell genomics identifies distinct B1 cell developmental pathways and reveals aging-related changes in the B-cell receptor repertoire. *Cell Biosci*. 12:57.

Madisen, L., T. Mao, H. Koch, J.M. Zhuo, A. Berenyi, S. Fujisawa, Y.W. Hsu, A.J. Garcia, 3rd, X. Gu, S. Zanella, J. Kidney, H. Gu, Y. Mao, B.M. Hooks, E.S. Boyden, G. Buzsaki, J.M. Ramirez, A.R. Jones, K. Svoboda, ..., and H. Zeng. 2012. A toolbox of Cre-dependent optogenetic transgenic mice for light-induced activation and silencing. *Nat Neurosci*. 15:793-802.

Maeda, H., A. Yamagata, S. Nishikawa, K. Yoshinaga, S. Kobayashi, K. Nishi, and S. Nishikawa. 1992. Requirement of c-kit for development of intestinal pacemaker system. *Development*. 116:369-375.

Majesky, M.W., H. Horita, A. Ostriker, S. Lu, J.N. Regan, A. Bagchi, X.R. Dong, J. Poczobutt, R.A. Nemenoff, and M.C. Weiser-Evans. 2017. Differentiated Smooth Muscle Cells Generate a Subpopulation of Resident Vascular Progenitor Cells in the Adventitia Regulated by Klf4. *Circ Res*. 120:296-311.

Malysz, J., S.J. Gibbons, S.A. Saravanaperumal, P. Du, S.T. Eisenman, C. Cao, U. Oh, D. Saur, S. Klein, T. Ordog, and G. Farrugia. 2017. Conditional genetic deletion of Ano1 in interstitial cells of Cajal impairs Ca(2+) transients and slow waves in adult mouse small intestine. *Am J Physiol Gastrointest Liver Physiol*. 312:G228-G245.

McCloskey, K.D., M.A. Hollywood, K.D. Thornbury, S.M. Ward, and N.G. McHale. 2002. Kit-like immunopositive cells in sheep mesenteric lymphatic vessels. *Cell Tissue Res*. 310:77-84.

Mitsui, R., and H. Hashitani. 2020. Synchrony of spontaneous Ca(2+) activity in microvascular mural cells. *Journal of smooth muscle research = Nihon Heikatsukin Gakkai kikanshi*. 56:1-18.

Mohanakumar, S., J. Majgaard, N. Telinius, N. Katballe, E. Pahle, V.E. Hjortdal, and D.M.B. Boedtkjer. 2018. Spontaneous and alpha-adrenoceptor-induced contractility in human collecting lymphatic vessels require chloride. *Am J Physiol Heart Circ Physiol*.

Muhl, L., G. Genove, S. Leptidis, J. Liu, L. He, G. Mocci, Y. Sun, S. Gustafsson, B. Buyandelger, I.V. Chivukula, A. Segerstolpe, E. Raschperger, E.M. Hansson, J.L.M. Bjorkegren, X.R. Peng, M. Vanlandewijck, U. Lendahl, and C. Betsholtz. 2020. Single-cell analysis uncovers fibroblast heterogeneity and criteria for fibroblast and mural cell identification and discrimination. *Nat Commun*. 11:3953.

Muzumdar, M.D., B. Tasic, K. Miyamichi, L. Li, and L. Luo. 2007. A global double-fluorescent Cre reporter mouse. *Genesis*. 45:593-605.

Olszewski, W.L. 2002. Contractility patterns of normal and pathologically changed human lymphatics. *Ann N Y Acad Sci*. 979:52-63; discussion 76-59.

Olszewski, W.L. 2008. Contractility patterns of human leg lymphatics in various stages of obstructive lymphedema. *Ann N Y Acad Sci*. 1131:110-118.

Ordog, T., S.M. Ward, and K.M. Sanders. 1999. Interstitial cells of cajal generate electrical slow waves in the murine stomach. *J Physiol*. 518:257-269.

Petkova, M., M. Kraft, S. Stritt, I. Martinez-Corral, H. Ortsater, M. Vanlandewijck, B. Jakic, E. Baselga, S.D. Castillo, M. Graupera, C. Betsholtz, and T. Makinen. 2023. Immune-interacting lymphatic endothelial subtype at capillary terminals drives lymphatic malformation. *J Exp Med*. 220.

Pitulescu, M.E., and R.H. Adams. 2014. Regulation of signaling interactions and receptor endocytosis in growing blood vessels. *Cell Adh Migr*. 8:366-377.

Popescu, L.M., and M.S. Faussone-Pellegrini. 2010. TELOCYTES - a case of serendipity: the winding way from Interstitial Cells of Cajal (ICC), via Interstitial Cajal-Like Cells (ICLC) to TELOCYTES. *J Cell Mol Med*. 14:729-740.

Sanders, K.M., S.A. Baker, B.T. Drumm, and M. Kurahashi. 2022. Ca(2+) Signaling Is the Basis for Pacemaker Activity and Neurotransduction in Interstitial Cells of the GI Tract. *Adv Exp Med Biol*. 1383:229-241.

Sanders, K.M., T. Ordog, S.D. Koh, S. Torihashi, and S.M. Ward. 1999. Development and plasticity of interstitial cells of Cajal. *Neurogastroenterol Motil*. 11:311-338.

Sanders, K.M., S.M. Ward, and S.D. Koh. 2014. Interstitial cells: regulators of smooth muscle function. *Physiol Rev*. 94:859-907.

Sanketi, B.D., M. Mantri, L. Huang, M.A. Tavallaei, S. Hu, M.F.Z. Wang, I. De Vlaminck, and N.A. Kurpios. 2024. Villus myofibroblasts are developmental and adult progenitors of mammalian gut lymphatic musculature. *Dev Cell*. 59:1159-1174 e1155.

Sasmono, R.T., D. Oceandy, J.W. Pollard, W. Tong, P. Pavli, B.J. Wainwright, M.C. Ostrowski, S.R. Himes, and D.A. Hume. 2003. A macrophage colony-stimulating factor receptor-green fluorescent protein transgene is expressed throughout the mononuclear phagocyte system of the mouse. *Blood*. 101:1155-1163.

Scallan, J.P., and M.J. Davis. 2013. Genetic removal of basal nitric oxide enhances contractile activity in isolated murine collecting lymphatic vessels. *J Physiol*. 591:2139-2156.

Scallan, J.P., S.D. Zawieja, J.A. Castorena-Gonzalez, and M.J. Davis. 2016. Lymphatic pumping: Mechanics, mechanisms and malfunction. *J Physiol*.

Sidney, L.E., M.J. Branch, S.E. Dunphy, H.S. Dua, and A. Hopkinson. 2014. Concise review: evidence for CD34 as a common marker for diverse progenitors. *Stem Cells*. 32:1380-1389.

Singh, R.D., S.J. Gibbons, S.A. Saravanaperumal, P. Du, G.W. Hennig, S.T. Eisenman, A. Mazzone, Y. Hayashi, C. Cao, G.J. Stoltz, T. Ordog, J.R. Rock, B.D. Harfe, J.H. Szurszewski, and G. Farrugia. 2014. Ano1, a Ca2+-activated Cl- channel, coordinates contractility in mouse intestine by Ca2+ transient coordination between interstitial cells of Cajal. *J Physiol*. 592:4051-4068.

Song, K., Y. Qing, Q. Guo, E.K. Peden, C. Chen, W.E. Mitch, L. Truong, and J. Cheng. 2020. PDGFRA in vascular adventitial MSCs promotes neointima formation in arteriovenous fistula in chronic kidney disease. *JCI Insight*. 5.

Spandidos, A., X. Wang, H. Wang, S. Dragnev, T. Thurber, and B. Seed. 2008. A comprehensive collection of experimentally validated primers for Polymerase Chain Reaction quantitation of murine transcript abundance. *BMC Genomics*. 9:633.

Spandidos, A., X. Wang, H. Wang, and B. Seed. 2010. PrimerBank: a resource of human and mouse PCR primer pairs for gene expression detection and quantification. *Nucleic Acids Res*. 38:D792-799.

Stolarz, A.J., M. Sarimollaoglu, J.C. Marecki, T.W. Fletcher, E.I. Galanzha, S.W. Rhee, V.P. Zharov, V.S. Klimberg, and N.J. Rusch. 2019. Doxorubicin Activates Ryanodine Receptors in Rat Lymphatic Muscle Cells to Attenuate Rhythmic Contractions and Lymph Flow. *J Pharmacol Exp Ther*. 371:278-289.

Thillaiappan, N.B., H.A. Smith, P. Atakpa-Adaji, and C.W. Taylor. 2021. KRAP tethers IP(3) receptors to actin and licenses them to evoke cytosolic Ca(2+) signals. *Nat Commun*. 12:4514.

To, K.H.T., P. Gui, M. Li, S.D. Zawieja, J.A. Castorena-Gonzalez, and M.J. Davis. 2020. T-type, but not L-type, voltage-gated calcium channels are dispensable for lymphatic pacemaking and spontaneous contractions. *Sci Rep*. 10:70.

Toland, H.M., K.D. McCloskey, K.D. Thornbury, N.G. McHale, and M.A. Hollywood. 2000. Ca(2+)-activated Cl(-) current in sheep lymphatic smooth muscle. *Am J Physiol Cell Physiol*. 279:C1327-1335.

Torihashi, S., S.M. Ward, S. Nishikawa, K. Nishi, S. Kobayashi, and K.M. Sanders. 1995. c-kit-dependent development of interstitial cells and electrical activity in the murine gastrointestinal tract. *Cell Tissue Res*. 280:97-111.

Truett, G.E., P. Heeger, R.L. Mynatt, A.A. Truett, J.A. Walker, and M.L. Warman. 2000. Preparation of PCR-quality mouse genomic DNA with hot sodium hydroxide and tris (HotSHOT). *Biotechniques*. 29:52, 54.

Van Helden, D.A., P.Y. Von Der Weid, and M. Crowe. 1996. Intracellular Ca Release: A basis for electrical pacemaking in lymphatic smooth muscle. *In* Smooth muscle excitation. T.B. Ed. Bolton and T. Tomita, editors. London: Academic Press. 355–373.

Van Helden, D.F. 1991. Spontaneous and noradrenaline-induced transient depolarizations in the smooth muscle of guinea-pig mesenteric vein. *J Physiol*. 437:511-541.

Van Helden, D.F. 1993. Pacemaker potentials in lymphatic smooth muscle of the guinea-pig mesentery. *J Physiol*. 471:465-479.

van Helden, D.F., K. Hosaka, and M.S. Imtiaz. 2006. Rhythmicity in the microcirculation. *Clin Hemorheol Microcirc*. 34:59-66.

van Helden, D.F., and M.S. Imtiaz. 2019. Venous Vasomotion. *Adv Exp Med Biol*. 1124:313-328.

Van Helden, D.F., and J. Zhao. 2000. Lymphatic vasomotion. *Clin Exp Pharmacol Physiol*. 27:1014-1018.

Van, S., S. Pal, B.R. Garner, K. Steed, V. Sridharan, S. Mu, N.J. Rusch, and A.J. Stolarz. 2021. Dantrolene Prevents the Lymphostasis Caused by Doxorubicin in the Rat Mesenteric Circulation. *Front Pharmacol*. 12:727526.

Vannucchi, M.G., C. Traini, M. Manetti, L. Ibba-Manneschi, and M.S. Faussone-Pellegrini. 2013. Telocytes express PDGFRalpha in the human gastrointestinal tract. *J Cell Mol Med*. 17:1099-1108.

von der Weid, P.Y., S. Lee, M.S. Imtiaz, D.C. Zawieja, and M.J. Davis. 2014. Electrophysiological properties of rat mesenteric lymphatic vessels and their regulation by stretch. *Lymphat Res Biol*. 12:66-75.

von der Weid, P.Y., M. Rahman, M.S. Imtiaz, and D.F. van Helden. 2008. Spontaneous transient depolarizations in lymphatic vessels of the guinea pig mesentery: pharmacology and implication for spontaneous contractility. *Am J Physiol Heart Circ Physiol*. 295:H1989-2000.

Wang, X., and B. Seed. 2003. A PCR primer bank for quantitative gene expression analysis. *Nucleic Acids Res*. 31:e154.

Wang, Y., Y. Jin, M.A. Mae, Y. Zhang, H. Ortsater, C. Betsholtz, T. Makinen, and L. Jakobsson. 2017. Smooth muscle cell recruitment to lymphatic vessels requires PDGFB and impacts vessel size but not identity. *Development*. 144:3590-3601.

Ward, S.M., A.J. Burns, S. Torihashi, and K.M. Sanders. 1994. Mutation of the proto-oncogene c-kit blocks development of interstitial cells and electrical rhythmicity in murine intestine. *J Physiol*. 480 (Pt 1):91-97.

Warthi, G., J.L. Faulkner, J. Doja, A.R. Ghanam, P. Gao, A.C. Yang, O.J. Slivano, C.T. Barris, T.C. Kress, S.D. Zawieja, S.H. Griffin, X. Xie, A. Ashworth, C.K. Christie, W.B. Bryant, A. Kumar, M.J. Davis, X. Long, L. Gan, E.J. Belin de Chantemele, Q. Lyu, and J.M. Miano. 2022. Generation and Comparative Analysis of an Itga8-CreER (T2) Mouse with Preferential Activity in Vascular Smooth Muscle Cells. *Nat Cardiovasc Res*. 1:1084-1100.

Wirth, A., Z. Benyo, M. Lukasova, B. Leutgeb, N. Wettschureck, S. Gorbey, P. Orsy, B. Horvath, C. Maser-Gluth, E. Greiner, B. Lemmer, G. Schutz, J.S. Gutkind, and S. Offermanns. 2008. G12-G13-LARG-mediated signaling in vascular smooth muscle is required for salt-induced hypertension. *Nat Med*. 14:64-68.

Xiao, J., F. Wang, Z. Liu, and C. Yang. 2013. Telocytes in liver: electron microscopic and immunofluorescent evidence. *J Cell Mol Med*. 17:1537-1542.

Yang, L., Y. Zhu, D. Tian, S. Wang, J. Guo, G. Sun, H. Jin, C. Zhang, W. Shi, M.E. Gershwin, Z. Zhang, Y. Zhao, and D. Zhang. 2021. Transcriptome landscape of double negative T cells by single-cell RNA sequencing. *J Autoimmun*. 121:102653.

Yoon, J.H., H. Jin, H.J. Kim, S.P. Hong, M.J. Yang, J.H. Ahn, Y.C. Kim, J. Seo, Y. Lee, D.M. McDonald, M.J. Davis, and G.Y. Koh. 2024. Nasopharyngeal lymphatic plexus is a hub for cerebrospinal fluid drainage. *Nature*. 625:768-777.

Young, M.D., and S. Behjati. 2020. SoupX removes ambient RNA contamination from droplet-based single-cell RNA sequencing data. *Gigascience*. 9.

Zawieja, D.C., K.L. Davis, R. Schuster, W.M. Hinds, and H.J. Granger. 1993. Distribution, propagation, and coordination of contractile activity in lymphatics. *Am J Physiol*. 264:H1283-1291.

Zawieja, S.D., J.A. Castorena, P. Gui, M. Li, S.A. Bulley, J.H. Jaggar, J.R. Rock, and M.J. Davis. 2019. Ano1 mediates pressure-sensitive contraction frequency changes in mouse lymphatic collecting vessels. *J Gen Physiol*. 151:532-554.

Zawieja, S.D., J.A. Castorena-Gonzalez, J. Scallan, and M.J. Davis. 2018a. Differences in L-type calcium channel activity partially underlie the regional dichotomy in pumping behavior by murine peripheral and visceral lymphatic vessels. *Am J Physiol Heart Circ Physiol*.

Zawieja, S.D., J.A. Castorena-Gonzalez, K.H.T. To, P. Gui, T.L. Domeier, and M.J. Davis. 2018b. Electrical Pacemaking in Lymphatic Vessels. *In* Signal Transduction in Smooth Muscle. M. Trebak and S. Earley, editors. CRC Press- Taylor & Francis Group. 324-359.

Zawieja, S.D., G.A. Pea, S.E. Broyhill, A. Patro, K.H. Bromert, M. Li, C.E. Norton, J.A. Castorena-Gonzalez, E.J. Hancock, C.D. Bertram, and M.J. Davis. 2023. IP3R1 underlies diastolic ANO1 activation and pressure-dependent chronotropy in lymphatic collecting vessels. *J Gen Physiol*. 155.

Zawieja, S.D., W. Wang, S. Chakraborty, D.C. Zawieja, and M. Muthuchamy. 2016. Macrophage alterations within the mesenteric lymphatic tissue are associated with impairment of lymphatic pump in metabolic syndrome. *Microcirculation*. 23:558-570.

Zhou, Q., L. Wei, C. Zhong, S. Fu, Y. Bei, R.I. Huica, F. Wang, and J. Xiao. 2015. Cardiac telocytes are double positive for CD34/PDGFR-alpha. *J Cell Mol Med*. 19:2036-2042.

Zhu, M.H., T.W. Kim, S. Ro, W. Yan, S.M. Ward, S.D. Koh, and K.M. Sanders. 2009. A Ca(2+)-activated Cl(-) conductance in interstitial cells of Cajal linked to slow wave currents and pacemaker activity. *J Physiol*. 587:4905-4918.

Zhu, X., D.E. Bergles, and A. Nishiyama. 2008. NG2 cells generate both oligodendrocytes and gray matter astrocytes. *Development*. 135:145-157.

Table 1 Primer list for RT-PCR						
Gene	**Strand**	**Accession #**	**Sequence (5'-3')**	**Size**	**Exon**	**Source**
Prox1	s	NM_008937	GTA AGA CAT CAC CGC GTG C	218	1	NIH Primer Tool
	as		TCA TGG TCA GGC ATC ACT GG		2	
CD11b (Itgam)	s	NM_008401	ATG GAC GCT GAT GGC AAT ACC	203	13	MGH Primer Bank ID 668048a1
	as		TCC CCA TTC ACG TCT CCC A		14	
Pdgfrα	s	NM_011058	AGA GTT ACA CGT TTG AGC TGT C	252	8	MGH Primer Bank 26349287a1
	as		GTC CCT CCA CGG TAC TCC T		10	
						-
Myh11	s	NM_013607	AAG CTG CGG CTA GAG GTC A	238	33	MGH Primer Bank ID 7305295a1
	as		CCC TCC CTT TGA TGG CTG AG		34	
CD117 (cKIT)	s	NM_021099	CGC CTG CCG AAA TGT ATG ACG	162	21	(Drumm et al., 2018)
	as		GGT TCT CTG GGT TGG GGT TGC		23	
Pdgfrβ	s	NM_008809	AGC TAC ATG GCC CCT TAT GA	367	16	(Basciani et al., 2004)
	as		GGA TCC CAA AAG ACC AGA CA		19	
CD144 (VE-cadherin)	s	NM_009868	CTT CCT TAC TGC CCT CAT TGT	313	3	IDT Primer Quest
	as		CTG TTT CTC TCG GTC CAA GTT		5	
Nos3 (eNOS)	s	NM_008713	CTG CCA CCT GAT CCT AAC TTG	143	22	IDT Real time primer tool
	as		CAG CCA AAC ACC AAA GTC ATG		23	
Acta2 (Smooth Muscle Actin)	s	NM_007392	GAG CTA CGA ACT GCC TGA C	129	7	IDT TaqMan Mm.PT.58.16320644
	as		CTG TTA TAG GTG GTT TCG TGG A		8	

CaV 1.2 exon1b	s	NM_001159533	ATG GTC AAT GAA AAC ACG AGG ATG		1	(Cheng et al., 2007)
	as		GGA ACT GAC GGT AGA GAT GGT TGC	234	2	
CD34	as	NM_133654	GGT ACA GGA GAA TGC AGG TC	119	1	IDT Mm.PT.58.8626728
	s		CGT GGT AGC AGA AGT CAA GT		2	
Cspg4 (Ng2)	as	NM_139001	CTT CAC GAT CAC CAT CCT TCC	132	5	IDT Mm.PT.58.29461721
	s		CCC GAA TCA TTG TCT GTT CCC		6	
-	-	-	-	-	-	
Vimentin	s	NM_011701	CTG TAC GAG GAG GAG ATG CG	249	1	(Li et al., 2016)
	as		AAT TTC TTC CTG CAA GGA TT		3	
Desmin	s	NM_010043	GTG GAT GCA GCC ACT CTA GC	218	3	MGH Primer Bank ID 33563250a1
	as		TTA GCC GCG ATG GTC TCA TAC		4	
CD146 (Mcam)	s	NM_023061	CCC AAA CTG GTG TGC GTC TT	220	1	MGH Primer Bank 10566955a1
	as		GGA AAA TCA GTA TCT GCC TCT CC		3	
KLF4	s	NM_010637	ATT AAT GAG GCA GCC ACC TG	400	1	(Majesky et al., 2017)
	as		GGA AGA CGA GGA TGA AGC TG		3	
Ly6a (Sca1)	s	NM_001271416	CTC TGA GGA TGG ACA CTT CT	400	2	(Majesky et al., 2017)
	as		GGT CTG CAG GAG GAC TGA GC		4	
Gli1	s	NM_01029	ATC ACC TGT TGG GGA TGC TGG AT	316	8	(Kramann et al., 2015)
	as		CGT GAA TAG GAC TTC CGA CAG		10	
CD29 (Itgb1)	s	NM_010578	TCG ATC CTG TGA CCC ATT GC	170	14	NIH Primer Tool

	as		AAC AAT TCC AGC AAC CAC GC		15	
CD105 (Endoglin)	s	NM_007932	TGA GCG TGT CTC CAT TGA CC	416	11	NIH Primer Tool
	as		GGG GCC ACG TGT GTG AGA A		15	
CD44	s	NM_009851	CAC CAT TTC CTG AGA CTT GCT	148	18	IDT Mm.PT.58.12084136
	as		TCT GAT TCT TGC CGT CTG C		19	
CD31 (Pecam1)	s	NM_008816	CTG CCA GTC CGA AAA TGG AAC	218	7	MGH Primer Bank ID 6679273a1
	as		CTT CAT CCA CTG GGG CTA TC		8	
GJC1 (Connexin 45)	s	NM_008122	GGT AAC AGG AGT TCT GGT GAA	140	2	IDT Mm.PT.58.8383900
	as		TCG AAA GAC AAT CAG CAC AGT		3	
Anoctamin 1 (TMEM16A)	s	NM_178642	GGC ATT TGT CAT TGT CTT CCA G	141	25	IDT Real time primer tool
	as		TCC TCA CGC ATA AAC AGC TC		26	
CD45 (Ptprc)	s	NM_001111316	ATG CAT CCA TCC TCG TCC AC	225	29	NIH Primer Tool
	as		TGA CTT GTC CAT TCT GGG CG		31	
MGH Harvard Primer Bank (Wang and Seed, 2003; Spandidos et al., 2008; Spandidos et al., 2010)						

Figure 1 Methylene blue staining of isolated mouse IALVs
Representative image of an isolated and cleaned IALV after methylene blue staining which revealed cells of various morphology. (B) is the zoomed in image of the yellow dotted box in A which contained large ovoid cells with granular staining (B, yellow asterisks). Fine cellular extensions (red asterisks) stained by methylene blue in some cells were visualized with color channel separation and division (C). (D, E) Similar as B and C, but in a separate vessel which stained with a higher density of methylene blue stained cells some of which had limited cellular processes. F) Focal reconstruction from imaging a methylene blue stained IALV using an upright microscope and immersion objective.

Figure 2 Staining Mouse IALVs for ICLC Markers
Representative immunofluorescent max projections of half vessel confocal image stacks imaged from mouse IALVs stained for ICLC markers. DAPI (A), cKIT (B), and CD34 (C) and their merged image (D). Representative max projections of the intermediate filament VIMENTIN (E), the intermediate filament desmin (F), CD34 (G) and their merged image (H). Representative max projection of VIMENTIN (I), cKIT (J), CD34 (K) and their merged image (L). Scale bar = 100 µm for all images.

Figure 3 Immunofluorescence Labeling of Mouse IALVs with Markers for ICLC, LMC, LEC, and Immune Cell Populations

We stained isolated mouse IALVs with cellular markers used to differentiate various cell types observed in cLVs. Half vessel image stacks were taken with confocal microscopy and the resulting representative max projections are shown. (A) CD34 stained cells and LMC staining with SMA (B) and calponin (C) and the corresponding merged (D) image. There was significant overlap in (E) CD34 staining along with the fibroblast marker PDGFRα compared to LMC staining with SMA (G) and the merged (H) image. The endothelial marker CD31 (I) to delineate LECs with PDGFRα staining (J), and the LMC marker calponin (K) with the merged image (L) revealed 3 separate populations of cells. PDGFRβ (O) stained many cells that were CD34 (M) and PDGFRα (N) positive, as seen in the merge imaged (P), in addition to PDGFRβ signal detected in the LMC layer (Q). Max projections of only the luminal frames of a z-stack at lymphatic valve locations revealed PDGFRβ, CD34, and PDGFRα labeling in bipolar shaped cells with long extensions that traveled throughout the valve leaflets (V, W). d Control IALV (Y) stained only with secondary antibody. Scale bar = 100 µm for all images.

SuppFigure 1 Colocalization of CD34 and PDGFRα

Representative max projections and their corresponding threshold adjusted image for colocalization analysis for PDGFRα (A), CD34 (B), and their colocalized signal (C) and for comparison we tested Myh11 (D) and PDGFRα (E) colocalization (F) using the FIJI BIOP-JACoP colocalization plugin on the z-stacks acquired by confocal microscopy. Pearson's coefficient (G) and Mander's coefficients (H) were calculated from n=3 separate stained IALVS, each from a separate mouse for CD34 and PDGFRα and n=4 for Myh11 and PDGFRα. Magnification for A-C 40X and 25x for D-F. Significant differences in colocalization below 0.05 are signified by the overhead lines.

SuppFigure 2 PDGFRα^{+} Cells Reside Primarily in the Mouse Lymphatic Collecting Vessel Adventitia and Some in the Subendothelial Space

Max projection of confocal imaging of an IALV stained for LECs with CD31 (A), LMCs with MYH11(B), and for PDGFRα (C) with the corresponding merge file (D). Orthogonal views of the z-stack with (E) showing a single slice in the z stack and E' and E" the orthogonal views. White dotted boxes outline locations where PDGFRα signal is observed between LMC and LEC layers. Scale bar is 100 µm in (D) and 50 µm in (E).

Figure 4 iCre-ROSA26mTmG Labelling and Fidelity to Target Putative Pacemaker Cell Populations
Stitched montages of serial max projections of GFP and tdTomato signal from live IALVs isolated from *PdgfrαCre*-ROSA26mTmG (A), *Ng2Cre*-ROSA26mTmG (B), *PdgfrαCreER^{TM}*-ROSA26mTmG (C), *PdgfrβCreER^{T2}*-ROSA26mTmG (D), *cKitCreER^{T2}*-ROSA26mTmG (E), and *Myh11CreER^{T2}*-ROSA26mTmG (F). IALVs were digested into single cells and GFP^{+} cells were purified via FACS from *Prox1-eGFP* (G), *Myh11CreER^{T2}*-ROSA26mTmG (H), *PdgfrαCreER^{TM}*-ROSA26mTmG (I), and *PdgfrβCreER^{T2}*-ROSA26mTmG (J) mice. Representative gels demonstrating RT-PCR products corresponding to the respective genes used in the promoter of each specific transgene employed to drive either eGFP or Cre mediated recombination of ROSA26mTmG from each GFP^{+} sorted population (K-N) to assess fidelity. Images are representative of IALVs from at least 3 separate mice. FACs and RT-PCR was repeated at least 3 times for each mouse.

Figure 5 scRNAseq analysis of mouse IALVs from ROSA26mTmG mice.
IALVs were cleaned and isolated from 8 ROSA26mTmG mice and digested into a single cell suspension for scRNAseq analysis with the 10X platform. A) UMAP of the various cell populations that compromise the mouse IALV though some mammary epithelia contamination was present (populations 18,19). B) Heat map of commonly used genes for cell identification for each of the cell clusters. Dot plots to assess cell cluster expression of the genes shown in Figure 4 using a dot plot for the LEC markers *Prox1* (C) and *Flt4* (D, VEGFR3), LMC markers *Myh11* (E) and caponin1 (F, *Cnn1*), fibroblast markers *Pdgfrα* (G) and *Lum* (H, *Lumican*), ICC marker *Kit* (I), the pericyte and smooth muscle precursor marker (*Pdgfrβ*), and the hematopoietic marker *Ptprc* (K, CD45).

SuppFigure 3. scRNASeq Analysis of the mouse IALV cell populations.
Heatmap of top 4-5 differentially expressed genes, based on p value, for each major cell cluster identified. LECs (Clusters 0,1,2, 11), LMCs (Cluster 5,6), and IALV adventitial cells (AdvC, 3,7,8,9,10,13) were comprised of multiple clusters. B) Bubble plot of common identification genes reveal that the previous reported LMC transcriptome markers Dpt, Pi16, and Ackr3 are specific for a sub population of the Adv and not LMCs.

SuppFigure 4. Subclusters of IALV LECs revealed by scRNAseq.
The LECs were further sub-clustered to reveal 10 putative LEC subclusters (0-9) as shown in the UMAP (A) and the top differentially expressed genes amongst those sub-clusters are provided in the adjacent heatmap (B). (C) Bubble plot showing sub-cluster 8 was significantly enriched for previously documented lymphatic endothelial cell up valve genes including *Itga9*, *Cldn11*, and *Neo1* and Cluster 6 had down valve gene signature including *Clu* and *Adm*. The top 30 differentially expressed genes in cluster 8, both positive and negative fold change regulated, are labeled in the volcano plot(D).

SuppFigure 5. Subclusters of IALV LMCs revealed by scRNAseq.
The LMCs could be subclustered into 4 putative subclusters (0-3) as shown in the UMAP (A). We profiled these subclusters based on their expression of the typical smooth muscle markers (B), SR associated genes (C), voltage gated Ca^{2+} channels, (D) Voltage gated Na^{+} channels and Na^{+} transporters implicated in lymphatic pacemaking (E), voltage gated K^{+} channels (F), Ca^{2+} activated K^{+} channels (G), inward rectifying K^{+} channels and two-pore domain K^{+} channels (H), and Cl^{-} channels (I).

SuppFigure 6. Subclusters of IALV AdvCs revealed by scRNAseq.
AdvCs also could be further subclustered into multiple populations as shown in the UMAP (A). Bubble plot of genes used as Cre drivers and genes associated with pacemaking revealed subcluster 10 had expression of *Ano1*, *Cx45*, and *Cacna1c (CaV1.2)* but with only minimal evidence of LMC contamination as indicated by muscle signature genes *Myh11*, *Kcnma1*, and *Tagln*. C) Heatmap of the top differentially expressed genes among each of the subclusters. We assessed co-expression of *Pdgfrα* with CD34 (D) to confirm our immunofluorescence imaging (Sugg Figure 1), and assessed the co-expression of *Pdgfrα* with the pericyte markers *Pdgfrβ* (E) and *Cspg4* (F). We further assessed co-expression of *Pdgfrα* the genes linked with contractile dysfunction *Ano1* (G), *Gcj1* (H), and *Cacna1c* (I) to ensure *PdgfrαCreER*^{TM} would target the AdvCs expressing these genes. The cyan colored slice of the pie chart indicates the minor population of cells expressing these genes that did not express *Pdgfrα.*

SuppFigure 7. Immune cell populations associated with the mouse IALV.
Lymphatic vessels are host to numerous immune cell populations, including monocyte, macrophage, and dendritic cell populations are revealed by immunofluorescent staining for eGFP in the "Macgreen" (Csf1r-eGFP) reporter mice (A). Staining for Pdgfrα (B) demonstrates that AdvCs are distinct from the GFP^+ cells nor do they stain for the hematopoietic marker *Ptprc* (CD45) (C, D). Bubble plot of our scRNASeq analysis of IALVs revealed macrophages (cluster 4), moDCs (cluster14) and cDC1 cells (17) based off identifying gene markers (B). C) Bubble plot of T-cell markers revealed multiple populations of T cells including naive double negative T-cells (Yang et al., 2021) and naive $CD4^+$ and $CD8^+$ T-cells. A bubble plot for B-cell markers showed that cluster 15 had an expression profile for immature and mature B2 B-cells (D)(Luo et al., 2022b).

Figure 6 RT-PCR Profiling of FACs Purified Cells from iCre-ROSA26mTmG
Expanded RT-PCR profiling of genes to discriminate LECs, LMCs, and other cell types in our GFP^+ sorted cells from *Prox1-eGFP* (A), *Myh11CreER*^{T2}-ROSA26mTmG (B), *PdgfrβCreER*^{T2}-ROSA26mTmG (C), and *PdgfrαCreER*^{TM}-ROSA26mTmG (D). Dot plots for the genes assessed in A-D in our IALV scRNAseq analysis confirmed those results. In addition to a population of AdvCs expressing Cacna1c, we also noted expression of Cx45 (N) which was also observed in LECs) and Ano1 (O) in the AdvC clusters. We confirmed this expression using GFP^+ cells sorted from *PdgfrαCreER*^{TM}-ROSA26mTmG IALVs for RT-PCR (P) and ruled out hematopoietic or LEC contamination. All RT-PCRs were performed 2-4 times for each gene over each sorted cell population collected from different mice.

Figure 7 Isobaric contractile Assessment of popliteal cLV from *PdgfrαCreER^{TM}* driven deletion of *Ano1, CX45,* and *CaV1.2*

Summary of the contractile parameters recorded from popliteal cLVs in *PdgfrαCreER^{TM}-Ano1*$^{fl/fl}$, *PdgfrαCreER^{TM}-Cx45*$^{fl/fl}$ mice, *PdgfrαCreER^{TM}-Cav1.2*$^{fl/fl}$ mice. Contraction frequency (A, D, G), ejection fraction (B, E, H), and vessel tone (C, F, I) were assessed. No statically significant differences observed in cLVs isolated from *PdgfrαCreER^{TM}-Ano1*$^{fl/fl}$ and *PdgfrαCreER^{TM}-Cx45*$^{fl/fl}$ mice across these three parameters. Mean and SEM shown, n=6 popliteal vessels from 3 mice *PdgfrαCreER^{TM}-Ano1*$^{fl/fl}$ mice and n=10 popliteal vessels from 6 mice *Ano1*$^{fl/fl}$ mice. Mean and SEM shown, n=5 popliteal vessels from 3 mice *PdgfrαCreER^{TM}-CX45*$^{fl/fl}$ mice and n=8 popliteal vessels from 11 mice *CX45*$^{fl/fl}$ mice. Mean and SEM shown, n=6 popliteal vessels from 3 mice *PdgfrαCreER^{TM}-Cav1.2*$^{fl/fl}$ mice and n=9 popliteal vessels from 20 mice *Cav1.2*$^{fl/fl}$ mice. The contractile data from control *Cav1.2*$^{fl/fl}$ vessels was previously published but was separated by sex (Davis et al., 2022) while they are combined here. * Denotes significance at $p < 0.05$ which $0.10 > p > 0.05$ are reported as text. Normalized contraction amplitude, fractional pump flow, end diastolic diameter can be found in SuppFigure 8.

SuppFigure 8 Contractile indices from isobaric myography on cLVs from *PdgfrαCreER^{TM}* driven deletion of *Ano1*, *CX45*, and *CaV1.2*

Summary of the contractile parameters recorded from popliteal cLVs in *PdgfrαCreER^{TM}-Ano1*$^{fl/fl}$, *PdgfrαCreER^{TM}-Cx45*$^{fl/fl}$ mice, *PdgfrαCreER^{TM}-Cav1.2*$^{fl/fl}$ mice. No differences in normalized contraction amplitude (A, D, G), fractional pump flow (B, E, H), or end diastolic diameter (C, F, I) were observed. The contractile data from control *Cav1.2*$^{fl/fl}$ vessels was previously published but was separated by sex (Davis et al., 2022) while they are combined here.

SuppFigure 9 PDGFRα AdvCs Include Multipotent Cell
Representative RT-PCR results profiling purified GFP^+ cells purified from IALVs isolated from *PdgfrαCreER*TM-ROSA26mTmG via FACS. PDGFRα cells expressed the multipotent markers *Klf4*, *Ly6a*, *Gli1*, *CD29*, *CD105*, and *CD44* (A) with total brain cDNA serving as a positive control (B). Representative RT-PCR results showing lack of expression of some of these markers in the GFP^+ cells purified from *Myh11CreER*^{T2}--ROSA26mTmG (C) or *Prox1-eGFP* mice, in contrast to the RFP^+ population from *Myh11CreER*^{T2}--ROSA26mTmG mice (D). RT-PCRs were repeated at least 2 times from separate purified cells populations from different mice. Dot plots of only the AdvCs cluster highlights populations of cells that express genes associated with multipotency such as *Ly6a* (E), *Klf4* (F), *Gli1* (G), *Itgb1* (H, *CD29*), *Eng* (I, *CD105*), *CD44* (J). Expression of protein for Ly6a was confirmed with immunofluorescence. Representative max projections of IALVs stained for Ly6a (K), PDGFRα (L), MYH11 (M) and the corresponding merged file (N). Scale bar is 100 µm.

Figure 8 ChR2-Mediated Depolarization Only in LMCs Triggers Contraction
Representative max projections of tdTomato-ChR2 signal in popliteal cLVs isolated from *cKitCreER*^{T2}-ChR2-tdTomato (A), *PdgfrαCreER*^{TM}-ChR2-tdTomato (C), and *Myh11CreER*^{T2}- ChR2-tdTomato (E) with their corresponding brightfield image (B, D, F) respectively. Time-lapse brightfield images every 0.5 s starting at stimulation t=0 for *cKitCreER*^{T2}-ChR2-tdTomato (G-J), *PdgfrαCreER*^{TM}-ChR2-tdTomato (K-N), and *Myh11CreER*^{T2}- ChR2-tdTomato (O-R). The I bar denotes the inner diameter at t=0 over time and white asterisks denote the contraction. Representative diameter trace for the popliteal cLV demonstrate spontaneous contractions with the dotted boxes indicating the optical stimulation event in the respective brightfield images of the time lapse images. Isolated cLVs from *cKitCreER*^{T2}-ChR2-tdTomato (S), *PdgfrαCreER*^{TM}-ChR2-tdTomato (T), and *Myh11CreER*^{T2}- ChR2-tdTomato (U) were stimulated with light pulses (red dashed lines) and the summation of contraction triggering for each genotype (V). Mean and SEM are shown, **** denotes $p<0.0001$. Contraction recorded from at least 6 popliteal cLVs from 3 mice per genotype.

Figure 9 *cKitCreER*T2 Drives GCaMP6f Expression Primarily in Mast Cells in Mouse IALVs
Representative max projection of GCaMP6f signal over time in an IALV isolated from a *cKitCreER*^{T2}-GCaMP6f mouse with ROI indicated around individual cells, primarily large ovoid cells, but also including a circumferential LMC (Cell10) and a horizontal LEC (Cell 11). Of cells 1-9, only cell 7 had any Ca^{2+} activity (red arrows) during the recording time as indicated by the STMs from each ROI (B) and their normalized F/F_0 plots in (C). In contrast, the LMC in ROI 10 had both rhythmic global Ca^{2+} events (D) that spanned the cell axis (vertical axis) in the STM (E) in addition to localized Ca^{2+} events intervening the time between global events (green arrows). Representative max projection of GCaMP6f signal over time after stimulation with C48-80 (F) with many large ovoid cells displaying long lasting global Ca^{2+} events (G, H) while not immediately affecting the LMC Ca^{2+} dynamics (I, J).

Figure 10 Lack of coordinated Ca^{2+} Activity Across Contraction Cycle in PDGFRα Cells
Representative max projections of GCaMP6f signal over time in an IALVs isolated from *PdgfrαCreER*^{TM}-GCaMP6f mice (A, D). ROIs were made around cells and GCaMP6f recorded over time to generate the corresponding STMs (B, E) for each cell and plots (C, F) respectively. Once again, incidental recombination occurred in a LMC which displayed rhythmic Ca^{2+} flashes (C) while the slight undulation in the other cells is due to movement artifact (B). Red arrows indicate the limited local Ca^{2+} activity observed in two cells from a *PdgfrαCreER*^{TM}-GCaMP6f IALV.

Figure 11 Heterogeneous Diastolic Ca^{2+} Transient Activity in LMCs
Representative max projections of GCaMP6f signal over time in an IALVs isolated from *Myh11CreER^{T2}*-GCaMP6f mice (A). LMCs were outlined with ROIs to assess GCaMp6F signal over time. Rhythmic global flashes (B) were entrained across all the LMCs in the FOV (C) with many cells exhibiting diastolic Ca^{2+} release events. Cells exhibiting at least one diastolic Ca^{2+} event, within the context of our focal plane constraints, over the recorded time were denoted by the red asterisks. The plot in (D) magnifies the first diastolic period, seconds 1-3 of C to assist in visualizing the lack of coordination of the diastolic events. (E) Max projection of the pseudo-linescan analysis across the axis of the vessel to highlight diastolic Ca^{2+} transients in all cells in the field of view and their lack of coordination across the cells (x-axis). The white dotted box shows the first diastolic period plotted in (D).

Figure 12 Pressure Dependency of Mouse LMC Diastolic Ca^{2+} Transients
Representative max projection of GCaMP6f signal over 20 s in an IALVs isolated from *Myh11CreER*^{T2}-GCaMP6f mice in the presence of the L-type blocker nifedipine (1μM) (A) pressurized to 0.5 cmH_2O, 2 cmH_2O, 5 cmH_2O. The local diastolic Ca^{2+} transients persist in the presence of nifedipine and increase with increasing pressure as demonstrated in the whole vessel STMs (B). Particle occurrence maps highlight the Ca^{2+} activity in each LMC as pressure is raised (C). Representative particle analysis plots for particle area (D) and particle counts/frame at each pressure (E). Summary files for particle area (F) and count /frame (G0. * Denotes $p<0.05$, Mean and SEM shown with n=12 separate IALVs from 8 MYH11-CreER^{T2}-GCaMP6f

Figure 13 Pressure-Dependent Diastolic Depolarization in LMCs
Intracellular recordings of LMC action potentials (AP) were confirmed by loading (greater than 10minutes) the impaling electrode with 1M KCl 100ug/ml AF488-Biocytin while recording APs followed by imaging on a spinning disk confocal microscope. 3D reconstruction of the z-stack confirmed the circumferential pattern of the impaled LMC that was strongly labeled by AF488-Biocytin (A, B), which also labeled neighboring LMCs, likely through gap junctions as AF488-Biocytin is <1kDa. In a separate set of experiments APs were recorded at 3 different pressures, 0.5 cmH_2O, 2 cmH_2O, and 5cmH_2O. We plotted the representative recordings from 1 cell at each pressure (C). AP frequency was significantly increased with pressure (D) as was the diastolic depolarization rate. Plotting the AP frequency and diastolic depolarization rate from all recordings at each pressure (F) highlights the significant effect diastolic depolarization rate has on the AP frequency. Minimum membrane potential (G), threshold membrane potential of AP initiation (H), upstroke constant (I), peak membrane potential (J), plateau membrane potential (K), and time over threshold (L) are also reported, although not significant.

Key Resources Table				
Reagent type (species) or resource	Designation	Source or reference	Identifiers	Additional information
Mouse Strain C57BL/6J	C57BL/6J	The Jackson Laboratory	Jax Strain #:000664	
Mouse B6.129(Cg)-Gt(ROSA)26Sortm4(ACTB-tdTomato,-EGFP)Luo/J Strain C57BL/6J	ROSA26mT/mG	The Jackson Laboratory	Jax Strain #007676	
Mouse C57BL/6-Tg(Pdgfra-cre)1Clc/J Strain C57BL/6NJ	PdgfrαCre	The Jackson Laboratory	Jax Strain #013148	
Mouse B6.Cg-Tg(Csf1r-EGFP)1Hume/J Strain C57BL/6NJ	CSFR1-EGFP	The Jackson Laboratory	Jax Strain #018549	
Mouse B6N.Cg-Tg(Pdgfra-cre/ERT)467Dbe/J Strain C57BL/6NJ	PdgfrαCreER™	The Jackson Laboratory	Jax Strain #018280	
Mouse B6;FVB-Ifi208Tg(Cspg4-cre)1Akik/J Mixed Strain C57BL/6 and FVB	NG2-Cre	The Jackson Laboratory	Jax Strain #:008533	

Mouse B6.Cg-Gt(ROSA)26Sortm27.1(CAG-COP4*H134R/tdTomato)Hze/J Strain C57BL/6J	ChR2/tdTomato	The Jackson Laboratory	Jax Strain #012567	
Mouse B6.Cg-Tg(Pdgfrb-cre/ERT2)6096Rha/J Strain C57BL/6J	PdgfrβCreER^{T2}	The Jackson Laboratory	Jax Strain #029684	
Mouse B6.FVB-Tg(Myh11-icre/ERT2)1Soff/J *Strain C57BL/6N*	Myh11CreER^{T2}	The Jackson Laboratory	Jax Strain #019079	
Mouse Kittm1(cre/ERT2)Dsa Strain 129S/SvEv 129S6/SvEvTac C57BL/6	c-KitCreER^{T2}	Dieter Saur (Technical University of Munich)		
Mouse Tg(Prox1-EGFP)KY221Gsat/Mmucd Strain FVB/N-Crl:CD1(ICR))	Prox1-eGFP	Young-Kwon Hong (University of Southern California	MMRRC ID #31006	
Mouse GCaMP6f Strain C57BL/6J	GCaMP6f	Jax	Jax Strain #028865	
antibody	anti-smooth muscle actin (SMA)	Sigma	A2547	1:500

antibody	anti-GFP	ThermoFisher ,	A11122	1:200
antibody	anti-cKIT	Cell Signaling	3074	1:100
antibody	anti-VIMENTIN	Thermofisher	OMA1-06001	1:100
antibody	anti-desmin	Invitrogen	PA5-16705	1:200
antibody	anti-GFP	Abcam	ab13970	1:200
antibody	anti-CD34	Invitrogen	14-0341-82	1:200
antibody	anti-PDGFRA	R&DSystems	AF1062	1:200
antibody	anti-PDGFRβ	eBiosciences	14-1402-82	1:200
antibody	anti-calponin	Abcam	AB46794	1:500
antibody	anti-Sca1	Biolegend	108101	1:200
sequenced-based reagent	*Prox1*-Forward NM_008937	NIH Primer Tool, this paper	GTA AGA CAT CAC CGC GTG C	
sequenced-based reagent	*Prox1*-Reverse NM_008937	NIH Primer Tool, this paper	TCA TGG TCA GGC ATC ACT GG	
sequenced-based reagent	*CD11b (Itgam)* Reverse	MGH Primer Bank ID 668048a1	ATG GAC GCT GAT	

	NM_008401		GGC AAT ACC	
sequenced-based reagent	*CD11b (Itgam)* Forward NM_008401	MGH Primer Bank ID 668048a1	TCC CCA TTC ACG TCT CCC A	
sequenced-based reagent	*Pdgfrα* Forward NM_011058	MGH Primer Bank ID 26349287a1	AGA GTT ACA CGT TTG AGC TGT C	
sequenced-based reagent	*Pdgfrα* Reverse NM_011058	MGH Primer Bank ID 26349287a1	GTC CCT CCA CGG TAC TCC T	
sequenced-based reagent	*Myh11* Forward NM_013607	MGH Primer Bank ID 7305295a1	AAG CTG CGG CTA GAG GTC A	
sequenced-based reagent	*Myh11* Reverse NM_013607	MGH Primer Bank ID 7305295a1	CCC TCC CTT TGA TGG CTG AG	
sequenced-based reagent	*CD117 (cKIT)* Forward NM_021099	(Drumm et al., 2018)	CGC CTG CCG AAA TGT ATG ACG	
sequenced-based reagent	*CD117 (cKIT)* Reverse NM_021099	(Drumm et al., 2018)	GGT TCT CTG GGT TGG GGT TGC	
sequenced-based reagent	*Pdgfrβ* Forward NM_008809	(Basciani et al., 2004)	AGC TAC ATG GCC CCT TAT GA	
sequenced-based reagent	*Pdgfrβ* Reverse NM_008809	(Basciani et al., 2004)	GGA TCC CAA AAG ACC AGA CA	
sequenced-based reagent	*CD144 (VE-cadherin)* Forward NM_009868	IDT Primer Quest Tool, this paper	CTT CCT TAC TGC CCT CAT TGT	
sequenced-based reagent	*CD144 (VE-cadherin)* Reverse	IDT Real time primer tool, this paper	CTG TTT CTC TCG	

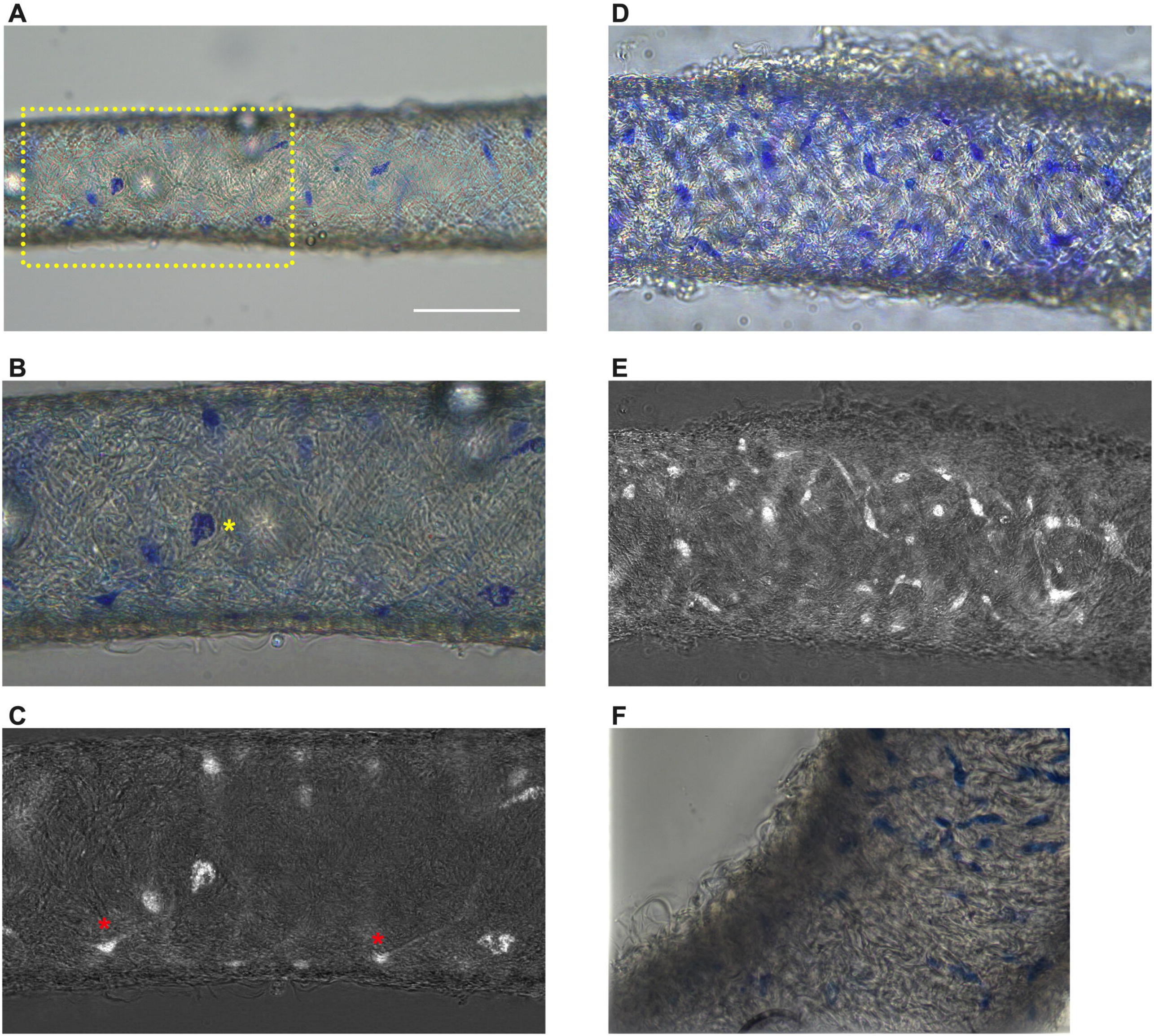
	NM_009868		GTC CAA GTT	
sequenced-based reagent	*Nos3 (eNOS)* Forward NM_008713	IDT Real time primer tool, this paper	CTG CCA CCT GAT CCT AAC TTG	sequenced-based reagent
sequenced-based reagent	*Nos3 (eNOS)* Reverse NM_008713	IDT Real time primer tool, this paper	CAG CCA AAC ACC AAA GTC ATG	sequenced-based reagent
sequenced-based reagent	*Acta2 (Smooth Muscle Actin)* Forward NM_007392	IDT TaqMan Mm.PT.58.16 320644	GAG CTA CGA ACT GCC TGA C	sequenced-based reagent
sequenced-based reagent	*Acta2 (Smooth Muscle Actin)* Reverse NM_007392	IDT TaqMan Mm.PT.58.16 320644	CTG TTA TAG GTG GTT TCG TGG A	sequenced-based reagent
sequenced-based reagent	*Cacna1c (CaV 1.2)* Forward NM_0011595 33	Cheng et al., 2007)	ATG GTC AAT GAA AAC ACG AGG ATG	sequenced-based reagent
sequenced-based reagent	*Cacna1c (CaV 1.2)* Reverse NM_0011595 33	Cheng et al., 2007)	GGA ACT GAC GGT AGA GAT GGT TGC	sequenced-based reagent
sequenced-based reagent	*CD34* Forward NM_133654	IDT Mm.PT.58.86 26728	GGT ACA GGA GAA TGC AGG TC	sequenced-based reagent
sequenced-based reagent	*CD34* Reverse NM_133654	IDT Mm.PT.58.86 26728	CGT GGT AGC AGA AGT CAA GT	sequenced-based reagent
sequenced-based reagent	*Cspg4 (Ng2)* Forward NM_139001	IDT Mm.PT.58.29 461721	CTT CAC GAT CAC CAT CCT TCC	sequenced-based reagent

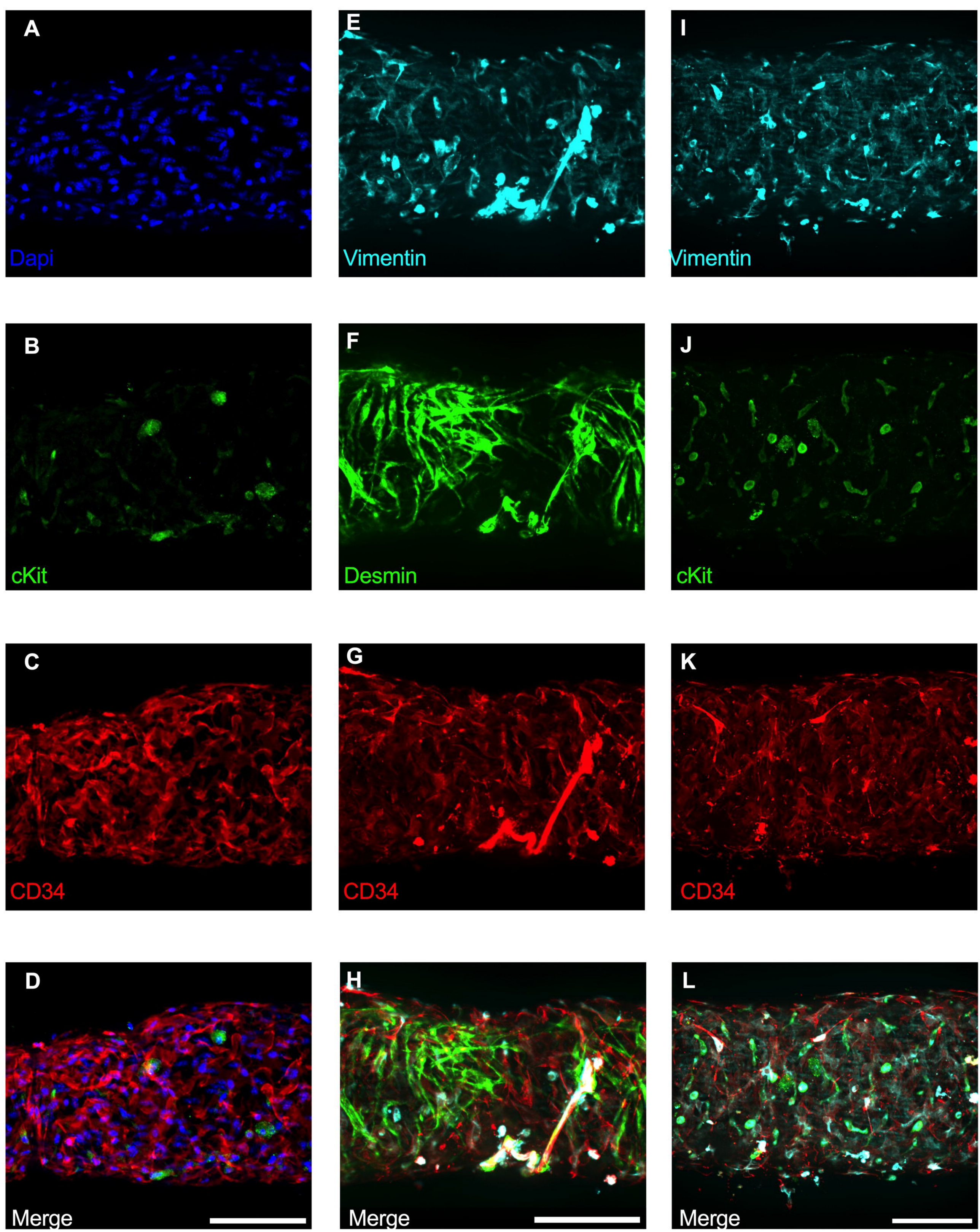
sequenced-based reagent	*Cspg4 (Ng2)* Reverse NM_139001	IDT Mm.PT.58.29 461721	CCC GAA TCA TTG TCT GTT CCC	sequenced-based reagent
sequenced-based reagent	*Vimentin* Forward NM_011701	(Li et al., 2016)	CTG TAC GAG GAG GAG ATG CG	sequenced-based reagent
sequenced-based reagent	*Vimentin* Reverse NM_011701	(Li et al., 2016)	AAT TTC TTC CTG CAA GGA TT	sequenced-based reagent
sequenced-based reagent	*Desmin* Forward NM_010043	MGH Primer Bank ID 33563250a1	GTG GAT GCA GCC ACT CTA GC	sequenced-based reagent
sequenced-based reagent	*Desmin* Reverse NM_010043	MGH Primer Bank ID 33563250a1	TTA GCC GCG ATG GTC TCA TAC	sequenced-based reagent
sequenced-based reagent	*CD146 (Mcam)* Forward NM_023061	MGH Primer Bank ID 10566955a1	CCC AAA CTG GTG TGC GTC TT	sequenced-based reagent
sequenced-based reagent	*CD146 (Mcam)* Reverse NM_023061	MGH Primer Bank ID 10566955a1	GGA AAA TCA GTA TCT GCC TCT CC	sequenced-based reagent
sequenced-based reagent	*KLF4* Forward NM_010637	(Majesky et al., 2017)	ATT AAT GAG GCA GCC ACC TG	sequenced-based reagent
sequenced-based reagent	*KLF4* Reverse NM_010637	(Majesky et al., 2017)	GGA AGA CGA GGA TGA AGC TG	sequenced-based reagent
sequenced-based reagent	*Ly6a (Sca1)* Forward NM_001271416	(Majesky et al., 2017)	CTC TGA GGA TGG ACA CTT CT	sequenced-based reagent
sequenced-based reagent	*Ly6a (Sca1)* Reverse NM_001271416	(Majesky et al., 2017)	GGT CTG CAG GAG GAC TGA GC	sequenced-based reagent

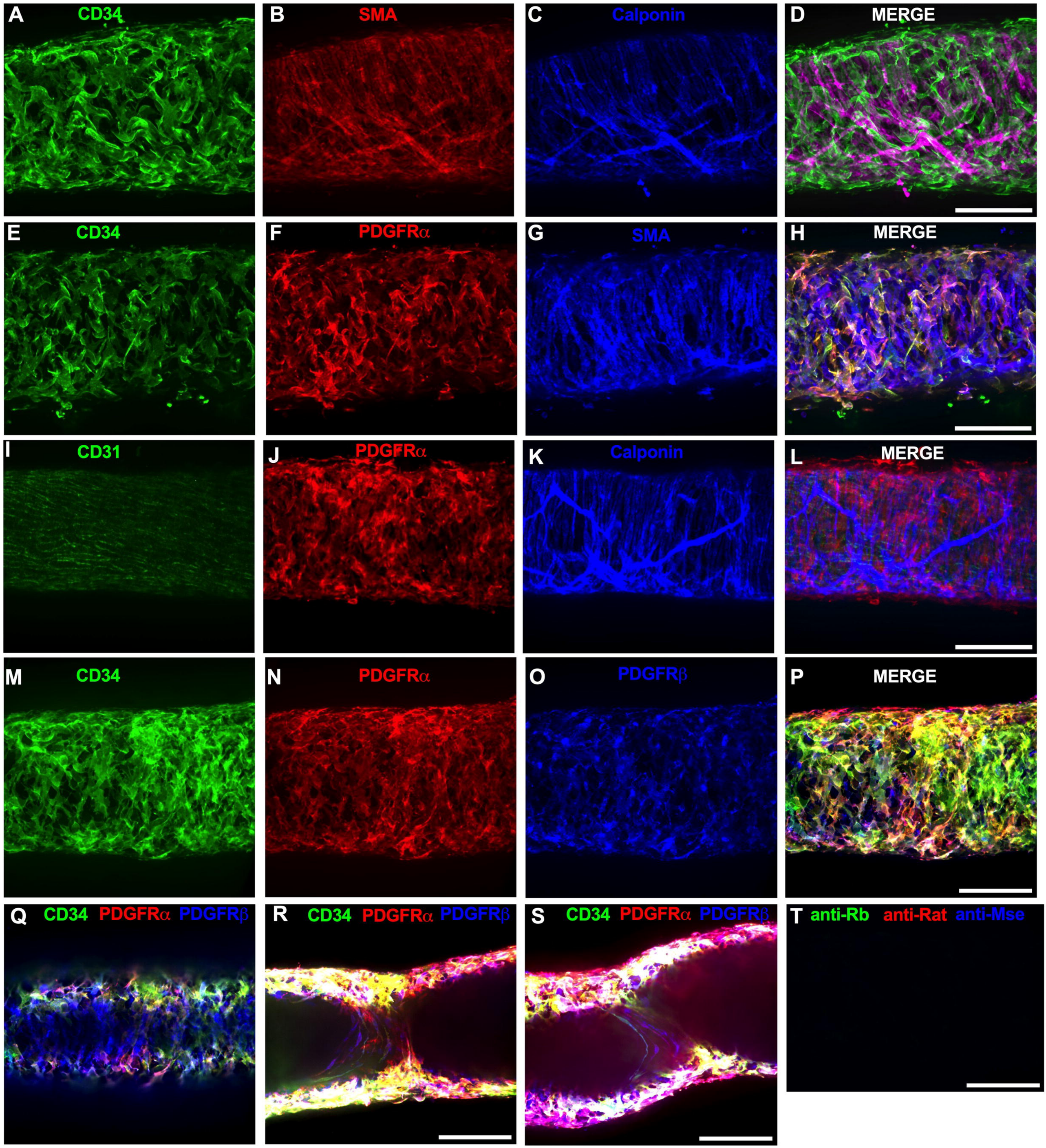
sequenced-based reagent	*Gli1* Forward NM_01029	(Kramann et al., 2015)	ATC ACC TGT TGG GGA TGC TGG AT	sequenced-based reagent
sequenced-based reagent	*Gli1* Reverse NM_01029	(Kramann et al., 2015)	CGT GAA TAG GAC TTC CGA CAG	sequenced-based reagent
sequenced-based reagent	*CD29 (Itgb1)* Forward NM_010578	NIH Primer Tool, this paper	TCG ATC CTG TGA CCC ATT GC	sequenced-based reagent
sequenced-based reagent	*CD29 (Itgb1)* Reverse NM_010578	NIH Primer Tool, this paper	AAC AAT TCC AGC AAC CAC GC	sequenced-based reagent
sequenced-based reagent	*CD105 (Endoglin)* Forward NM_007932	NIH Primer Tool, this paper	TGA GCG TGT CTC CAT TGA CC	sequenced-based reagent
sequenced-based reagent	*CD105 (Endoglin)* Reverse NM_007932	NIH Primer Tool, this paper	GGG GCC ACG TGT GTG AGA A	sequenced-based reagent
sequenced-based reagent	*CD44* Forward NM_009851	IDT Mm.PT.58.12 084136	CAC CAT TTC CTG AGA CTT GCT	sequenced-based reagent
sequenced-based reagent	*CD44* Reverse NM_009851	IDT Mm.PT.58.120 84136	TCT GAT TCT TGC CGT CTG C	sequenced-based reagent
sequenced-based reagent	*CD31 (Pecam1)* Forward NM_008816	MGH Primer Bank ID 6679273a1	CTG CCA GTC CGA AAA TGG AAC	sequenced-based reagent
sequenced-based reagent	*CD31 (Pecam1)* Reverse NM_008816	MGH Primer Bank ID 6679273a1	CTT CAT CCA CTG GGG CTA TC	sequenced-based reagent

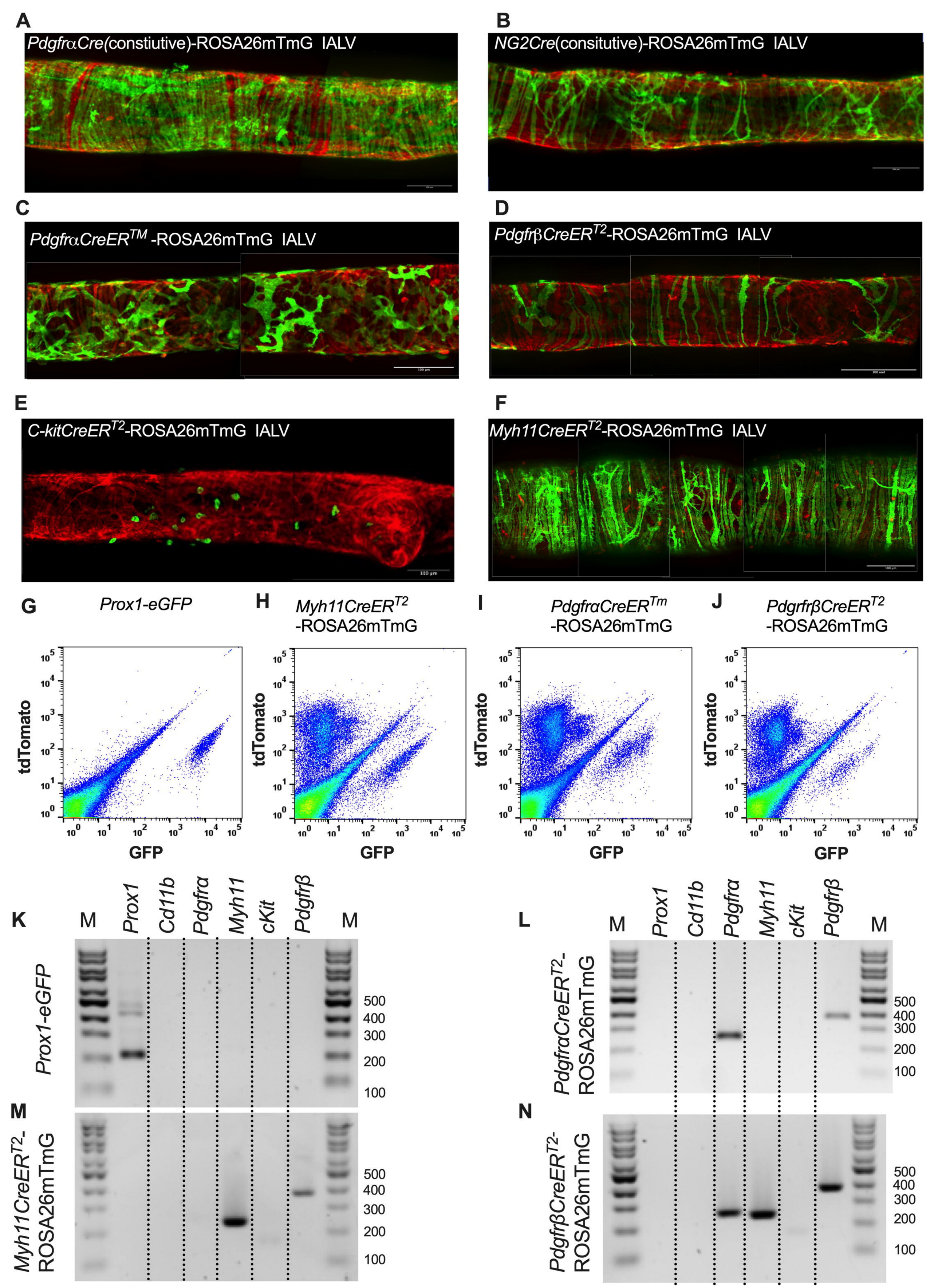
sequenced-based reagent	*GJC1 (Connexin 45)* Forward NM_008122	IDT Mm.PT.58.8383900	GGT AAC AGG AGT TCT GGT GAA	sequenced-based reagent
sequenced-based reagent	*GJC1 (Connexin 45)* Reverse NM_008122	IDT Mm.PT.58.8383900	TCG AAA GAC AAT CAG CAC AGT	sequenced-based reagent
sequenced-based reagent	*Anoctamin 1 (TMEM16A) Forward* NM_178642	IDT Real time primer tool, this paper	GGC ATT TGT CAT TGT CTT CCA G	sequenced-based reagent
sequenced-based reagent	*Anoctamin 1 (TMEM16A) Reverse* NM_178642	IDT Real time primer tool, this paper	TCC TCA CGC ATA AAC AGC TC	sequenced-based reagent
sequenced-based reagent	*CD45 (Ptprc)* Forward NM_001111316	NIH Primer Tool, this paper	ATG CAT CCA TCC TCG TCC AC	sequenced-based reagent
sequenced-based reagent	*CD45 (Ptprc)* Reverse NM_001111316	NIH Primer Tool, this paper	TGA CTT GTC CAT TCT GGG CG	sequenced-based reagent
commercial dye	Methylene Blue	Sigma	M9140	
commercial reagent	Blockaid	ThermoFisher	A11122	
Commercial enzyme	Collagenase H	Sigma	C8051	
Commercial enzyme	Collagenase F	Sigma	C7926	
Commercial reagent	Dithioerythritol (DTT)	Sigma	D8161	

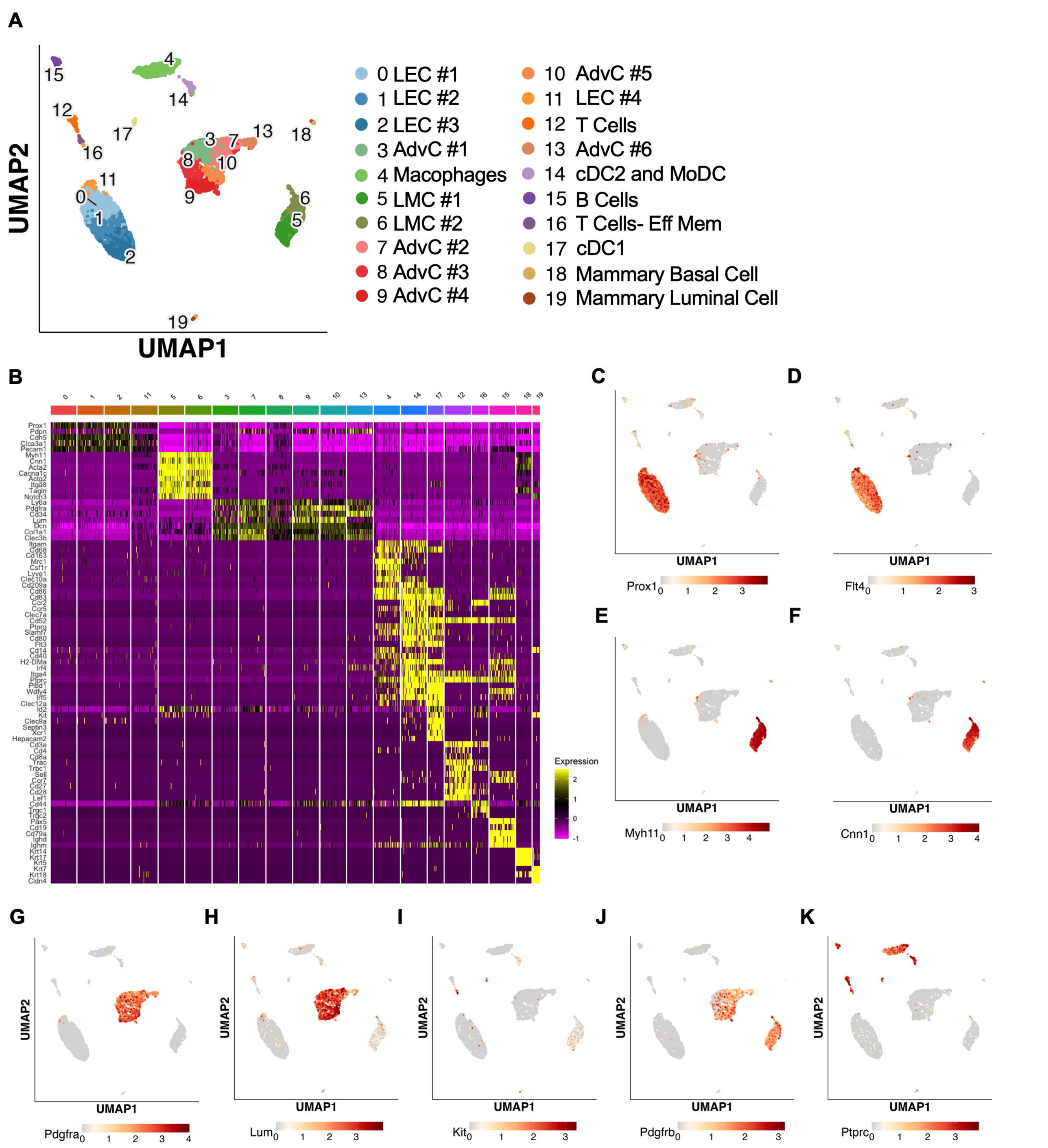
Commercial enzyme	elastase	Worthington	LS00635	
Commercial enzyme	papain	Sigma	P4762	
antibody	Donkey anti-mouse AF647	ThermoFisher	A32787	1:200
antibody	Donkey anti-Rat AF555	ThermoFisher	A48270	1:200
antibody	Donkey anti-Rabbit AF488	ThermoFisher	A21206	1:200
antibody	Donkey anti-Goat AF647	ThermoFisher	A21447	1:200
antibody	Donkey anti-Goat AF555	ThermoFisher	A21432	1:200
Commercial RNA isolation kit	Arcturus PicoPure RNA isolation kit	ThermoFisher	KIT0204	
Software	Prism 10	GraphPad		
Software	Volumetry software (version G8d)	Grant Hennig		(Drumm et al., 2017; Drumm et al., 2019b)

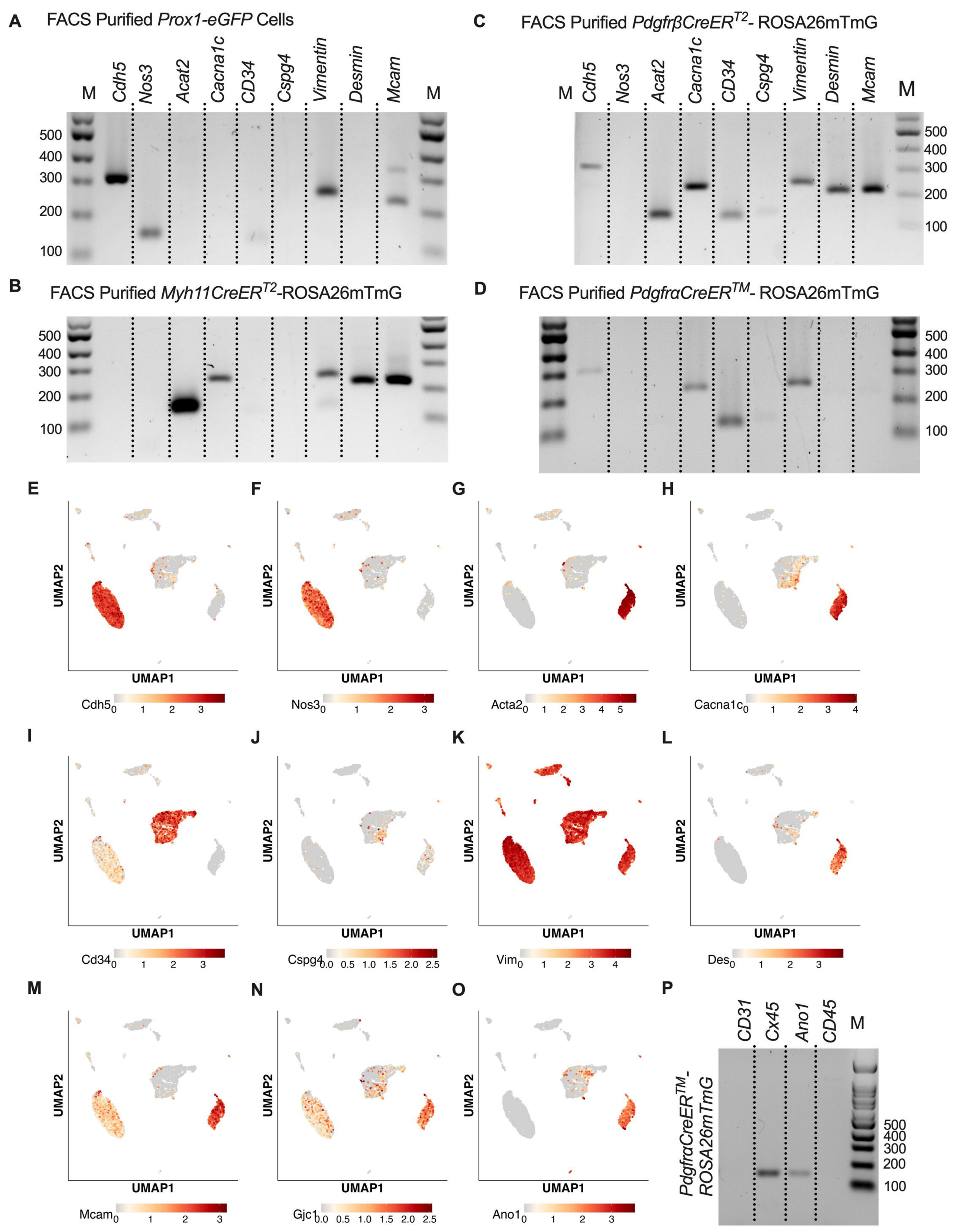

A
D
B
E
C
F

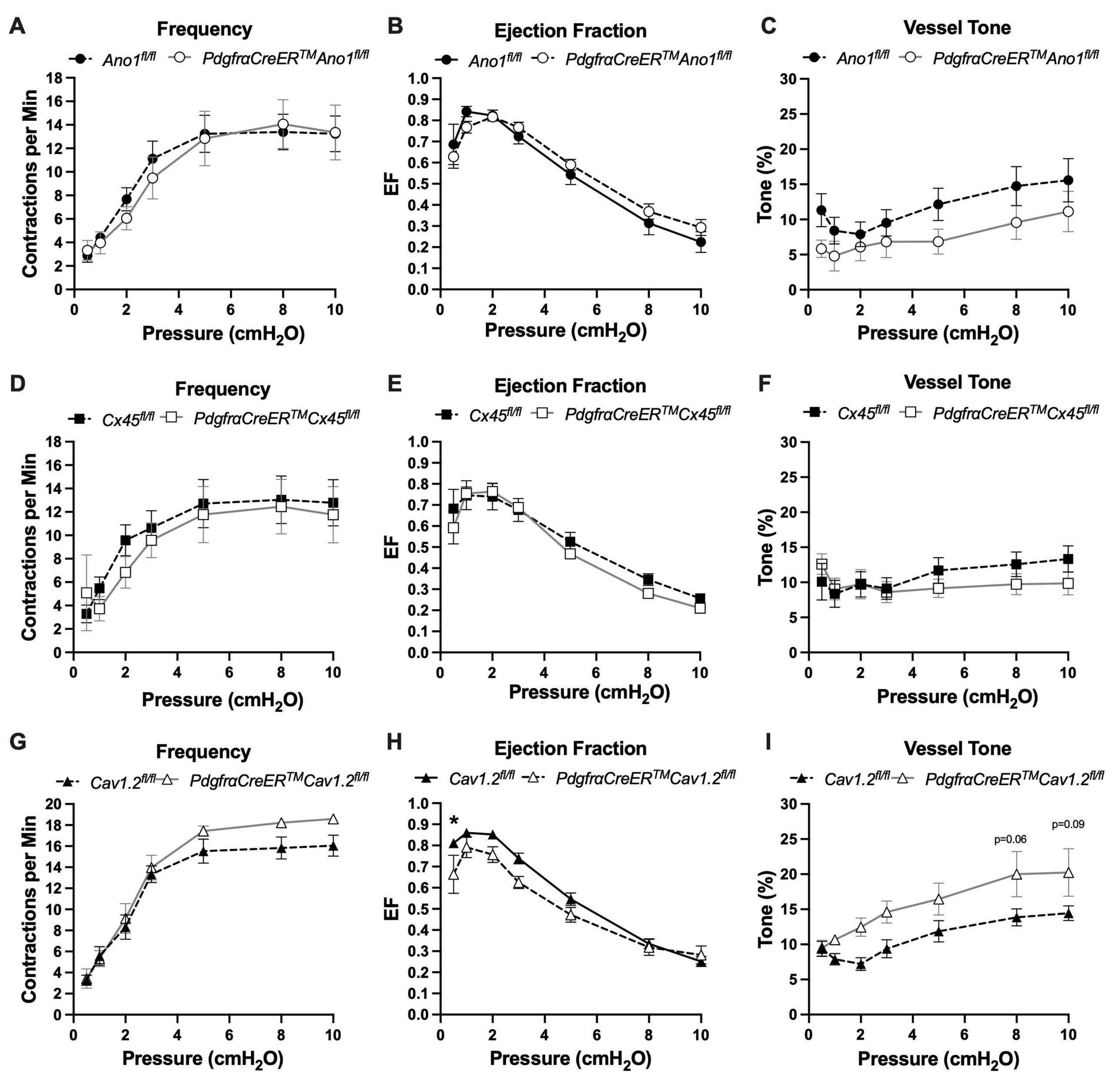

A
Dapi
E
Vimentin
I
Vimentin
B
cKit
F
Desmin
J
cKit
C
CD34
G
CD34
K
CD34
D
Merge
H
Merge
L
Merge

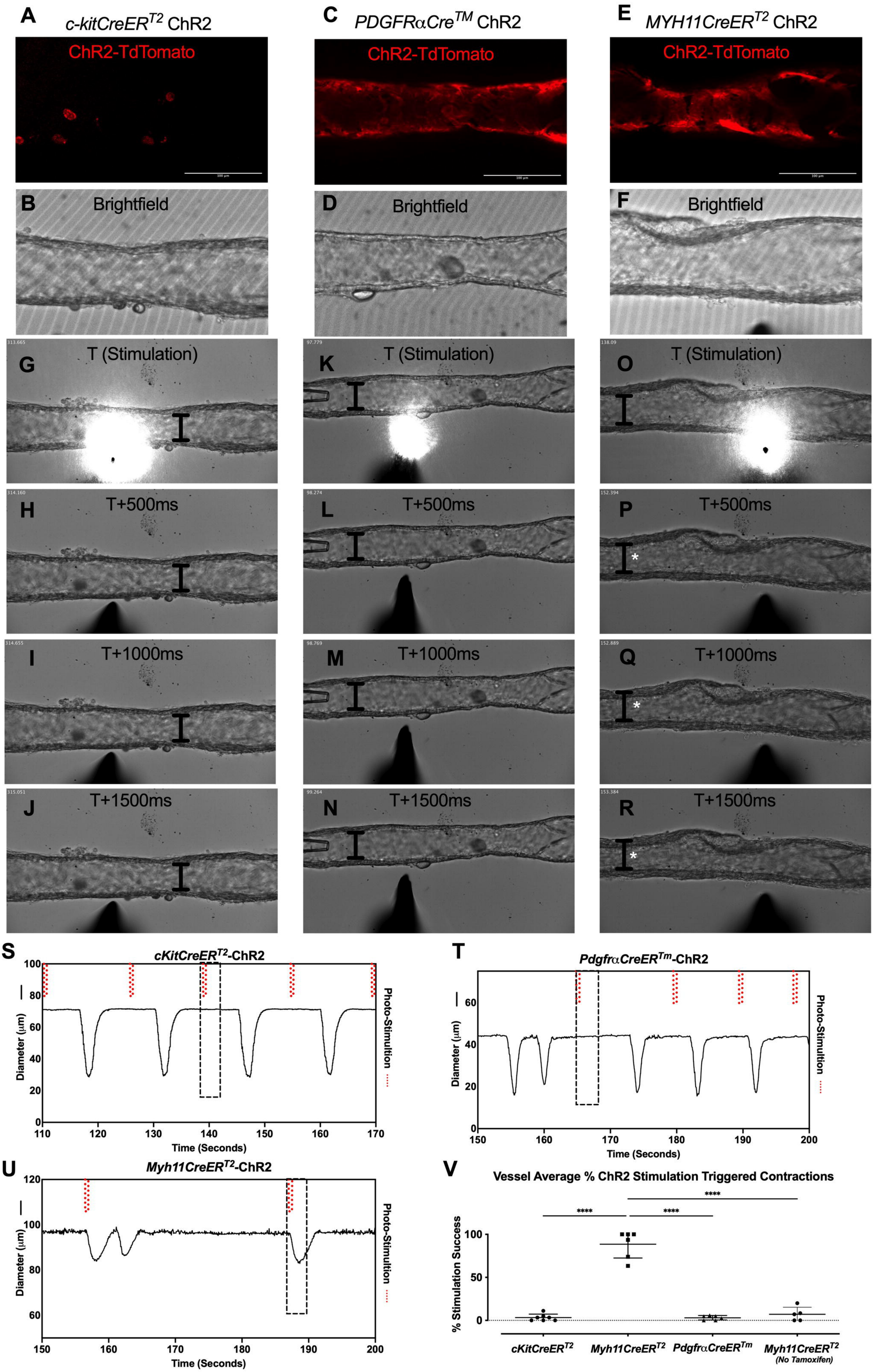

A
CD34
B
SMA
C
Calponin
D
MERGE
E
CD34
F
PDGFRα
G
SMA
H
MERGE
I
CD31
J
PDGFRα
K
Calponin
L
MERGE
M
CD34
N
PDGFRα
O
PDGFRβ
P
MERGE
Q
CD34 PDGFRα PDGFRβ
R
CD34 PDGFRα PDGFRβ
S
CD34 PDGFRα PDGFRβ
T
anti-Rb anti-Rat anti-Mse


A
PdgfrαCre(constiutive)-ROSA26mTmG IALV
B
NG2Cre(consitutive)-ROSA26mTmG IALV
C
PdgfrαCreER^{TM} -ROSA26mTmG IALV
D
PdgfrβCreER^{T2}-ROSA26mTmG IALV
E
C-kitCreER^{T2}-ROSA26mTmG IALV
F
Myh11CreER^{T2}-ROSA26mTmG IALV
G
Prox1-eGFP
H
Myh11CreER^{T2} -ROSA26mTmG
I
PdgfraCreER^{Tm} -ROSA26mTmG
J
PdgrfrβCreER^{T2} -ROSA26mTmG
tdTomato
GFP
K
M
Prox1
Cd11b
Pdgfrα
Myh11
cKit
Pdgfrβ
M
Prox1-eGFP
500
400
300
200
100
L
PdgfraCreER^{T2}-ROSA26mTmG
M
N
Myh11CreER^{T2}-ROSA26mTmG
PdgfrβCreER^{T2}-ROSA26mTmG

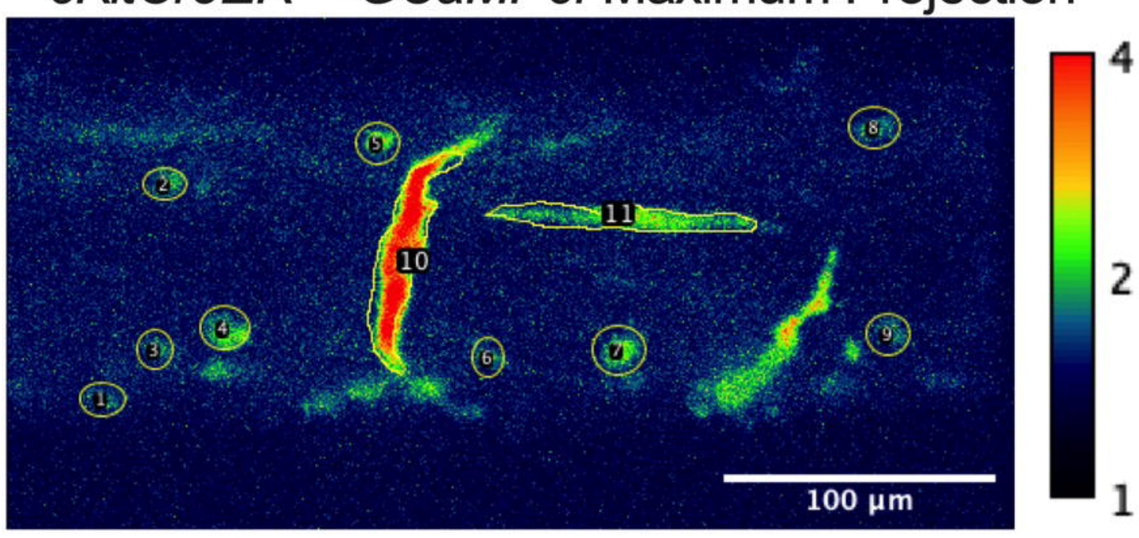

A
UMAP2
UMAP1
0 LEC #1
1 LEC #2
2 LEC #3
3 AdvC #1
4 Macophages
5 LMC #1
6 LMC #2
7 AdvC #2
8 AdvC #3
9 AdvC #4
10 AdvC #5
11 LEC #4
12 T Cells
13 AdvC #6
14 cDC2 and MoDC
15 B Cells
16 T Cells- Eff Mem
17 cDC1
18 Mammary Basal Cell
19 Mammary Luminal Cell
B
Expression
C
Prox1
D
Flt4
E
Myh11
F
Cnn1
G
Pdgfra
H
Lum
I
Kit
J
Pdgfrb
K
Ptprc


A
FACS Purified Prox1-eGFP Cells
M
Cdh5
Nos3
Acat2
Cacna1c
CD34
Cspg4
Vimentin
Desmin
Mcam
M
500
400
300
200
100
B
FACS Purified Myh11CreER^{T2}-ROSA26mTmG
C
FACS Purified PdgfrβCreER^{T2}- ROSA26mTmG
D
FACS Purified PdgfraCreER^{TM}- ROSA26mTmG
E
F
G
H
I
J
K
L
M
N
O
P
UMAP2
UMAP1
Cdh5$_0$ 1 2 3
Nos3$_0$ 1 2 3
Acta2$_0$ 1 2 3 4 5
Cacna1c$_0$ 1 2 3 4
Cd34$_0$ 1 2 3
Cspg4$_{0.0}$ 0.5 1.0 1.5 2.0 2.5
Vim$_0$ 1 2 3 4
Des$_0$ 1 2 3
Mcam$_0$ 1 2 3
Gjc1$_{0.0}$ 0.5 1.0 1.5 2.0 2.5
Ano1$_0$ 1 2 3
CD31
Cx45
Ano1
CD45
M
PdgfraCreER^{TM}-
ROSA26mTmG
500
400
300
200
100


A
Frequency
Ano1fl/fl
PdgfrαCreERTMAno1fl/fl
Contractions per Min
Pressure (cmH2O)
B
Ejection Fraction
EF
C
Vessel Tone
Tone (%)
D
Cx45fl/fl
PdgfrαCreERTMCx45fl/fl
E
F
G
Cav1.2fl/fl
PdgfrαCreERTMCav1.2fl/fl
H
*
I
p=0.06
p=0.09

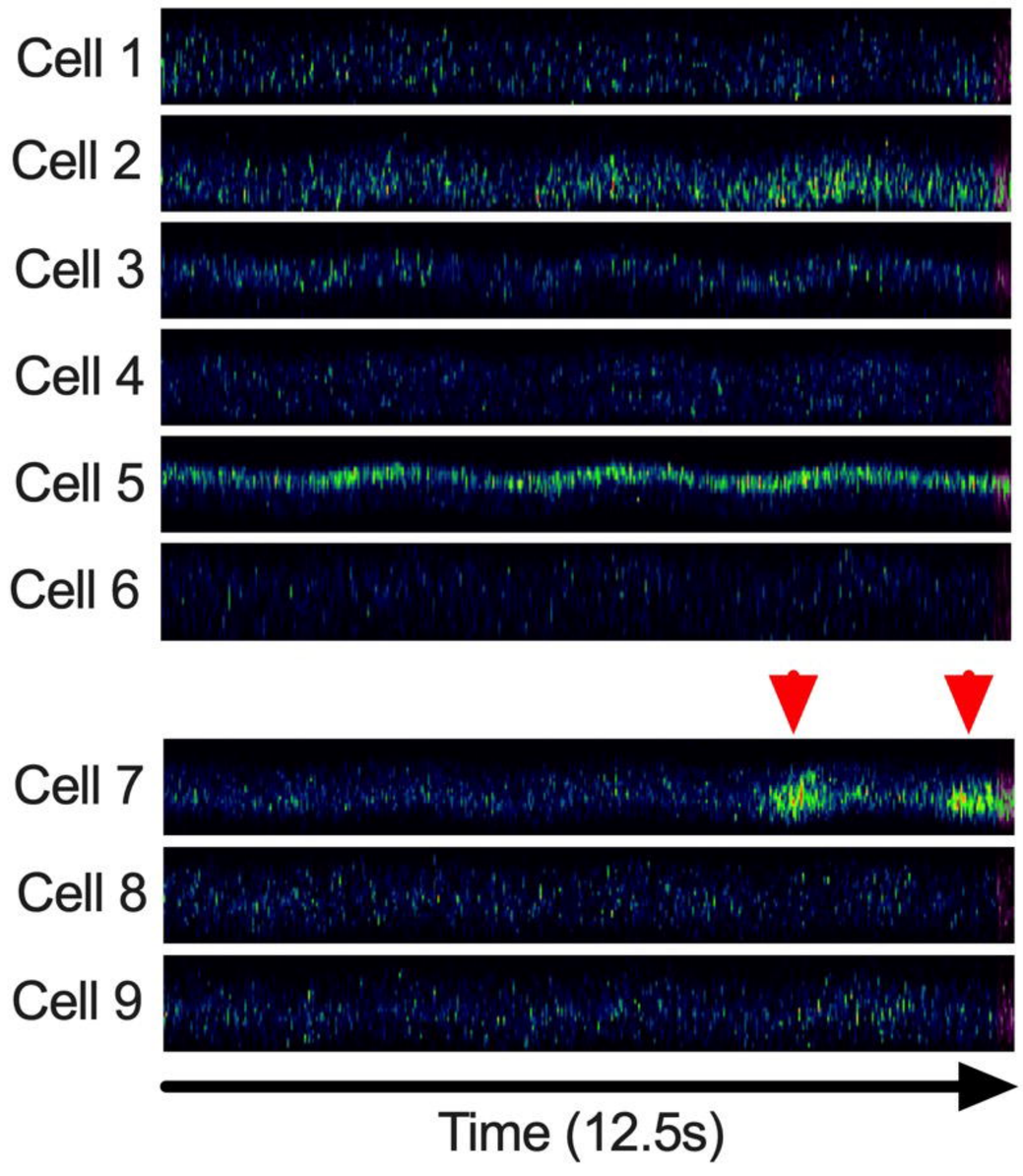

A
c-kitCreER^{T2} ChR2
ChR2-TdTomato
B
Brightfield
C
PDGFRαCre^{TM} ChR2
ChR2-TdTomato
D
Brightfield
E
MYH11CreER^{T2} ChR2
ChR2-TdTomato
F
Brightfield
G
T (Stimulation)
H
T+500ms
I
T+1000ms
J
T+1500ms
K
T (Stimulation)
L
T+500ms
M
T+1000ms
N
T+1500ms
O
T (Stimulation)
P
T+500ms
Q
T+1000ms
R
T+1500ms
S
cKitCreER^{T2}-ChR2
Diameter (μm)
Time (Seconds)
Photo-Stimultion
T
PdgfrαCreER^{Tm}-ChR2
Diameter (μm)
Time (Seconds)
Photo-Stimultion
U
Myh11CreER^{T2}-ChR2
Diameter (μm)
Time (Seconds)
Photo-Stimultion
V
Vessel Average % ChR2 Stimulation Triggered Contractions
% Stimulation Success
****
****
****
cKitCreER^{T2}
Myh11CreER^{T2}
PdgfrαCreER^{Tm}
Myh11CreER^{T2}
(No Tamoxifen)

A *cKitCreER*^{T2}-*GCaMP6f* Maximum Projection

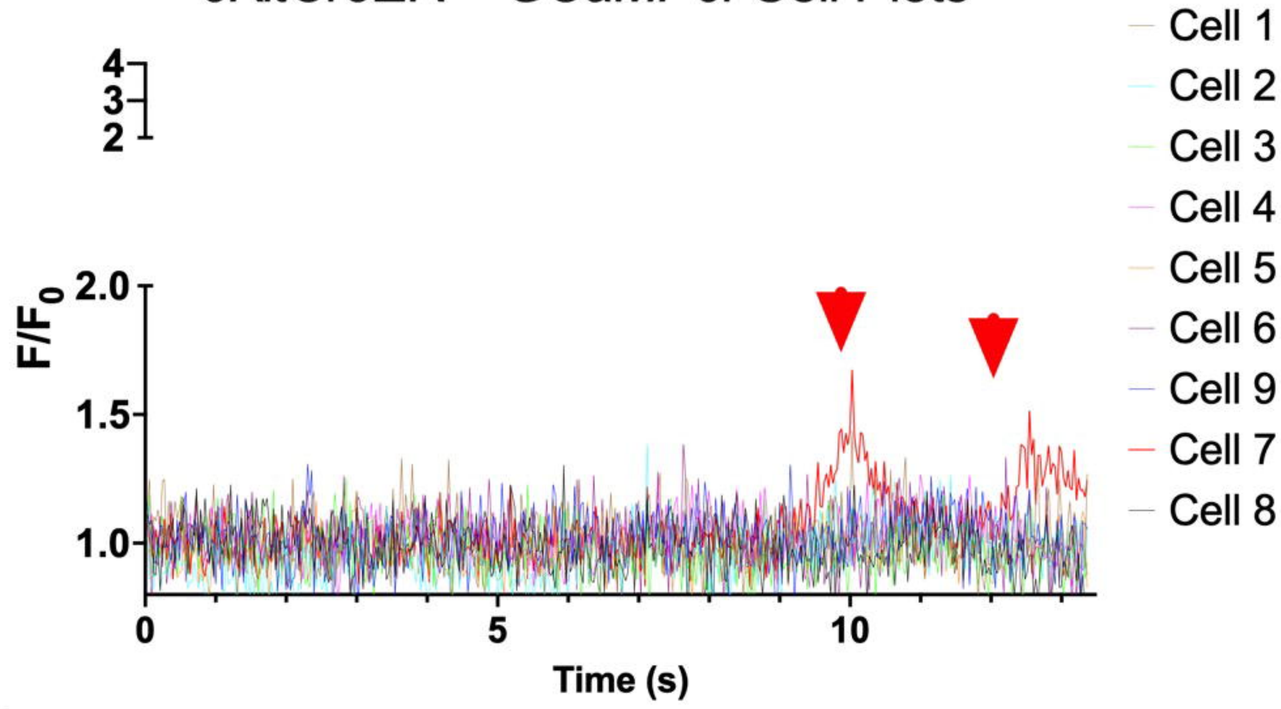


cKitCreER^{T2}-*GCaMP6f* STMs

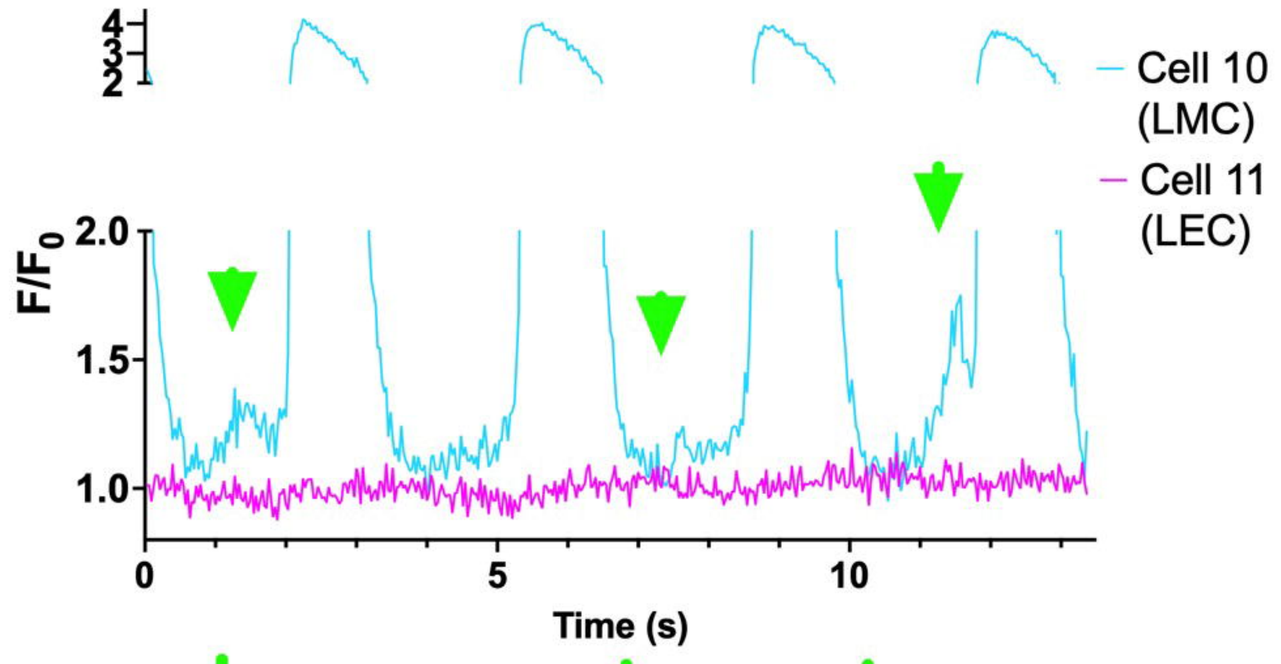
B



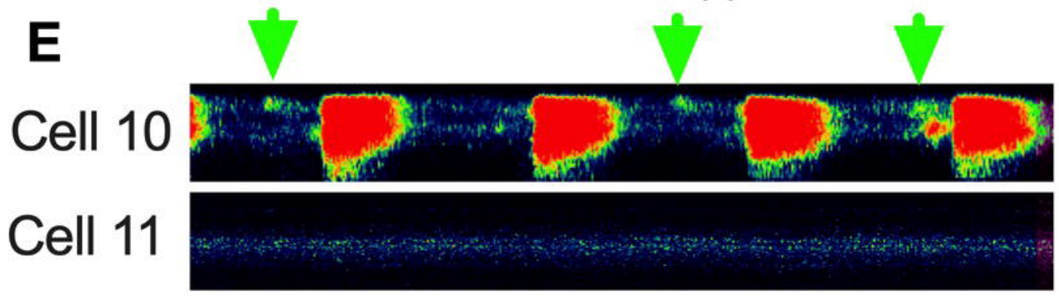
C *cKitCreER*^{T2}-*GCaMP6f* Cell Plots



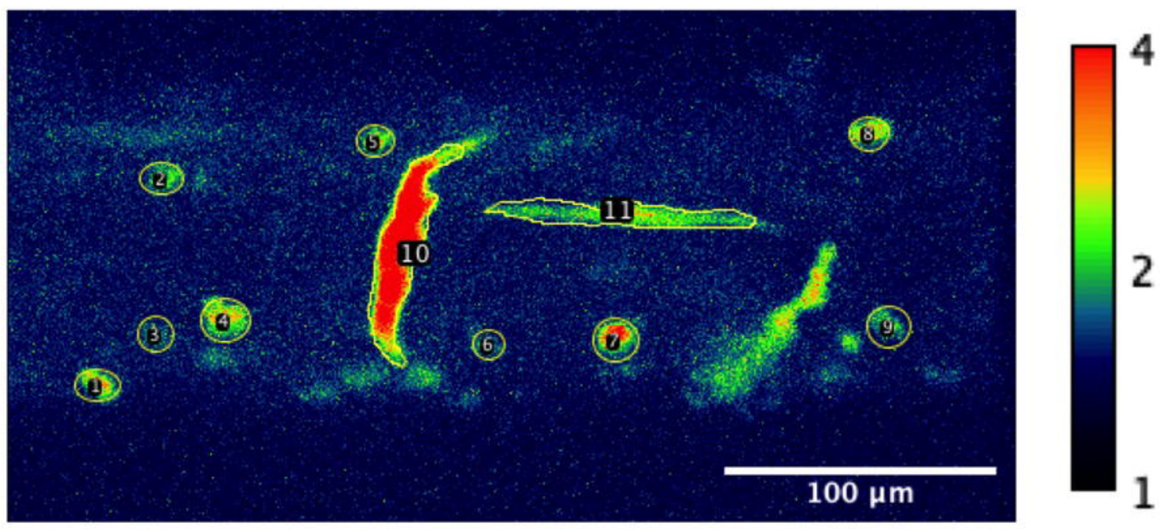
D Incidental LEC and LMC



E

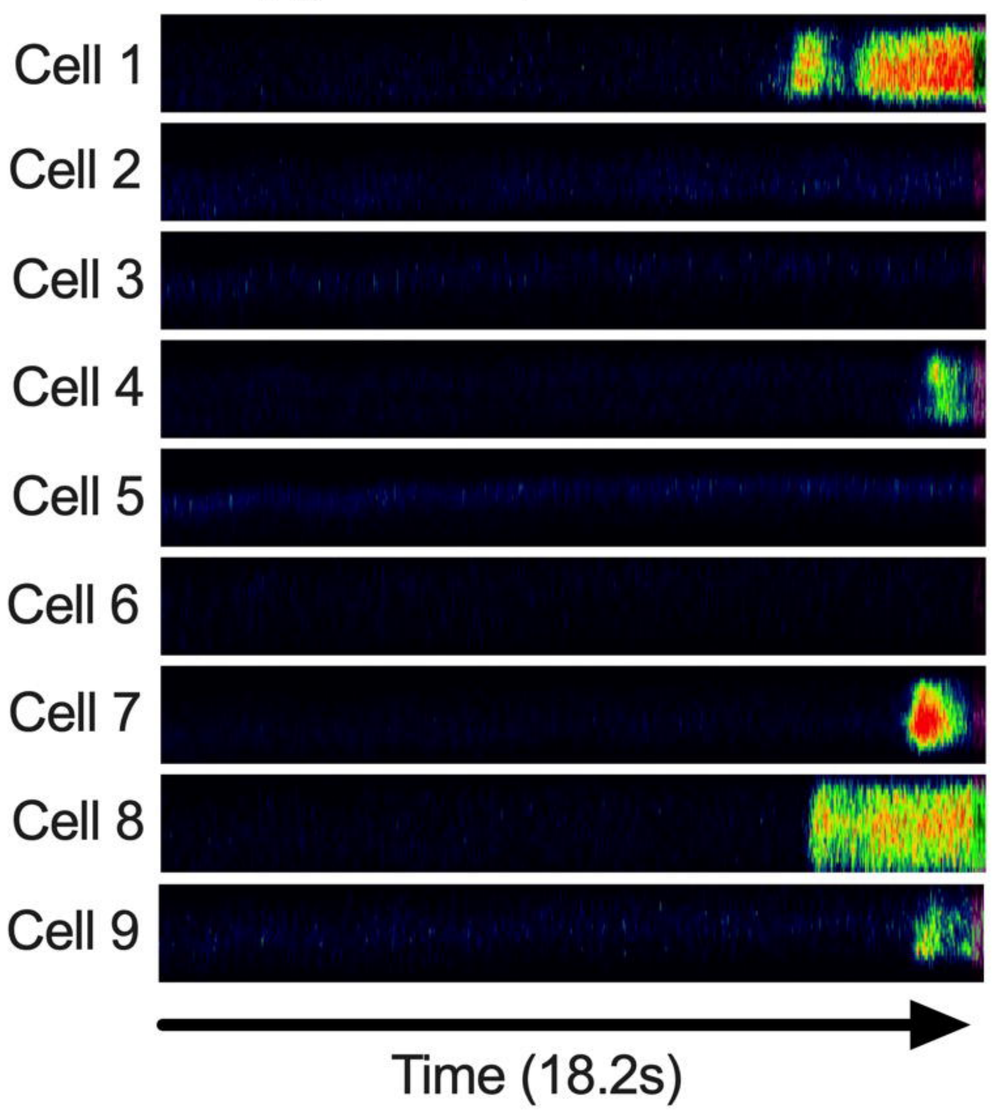


E Maximum Projection: 100ug/ml C48-80

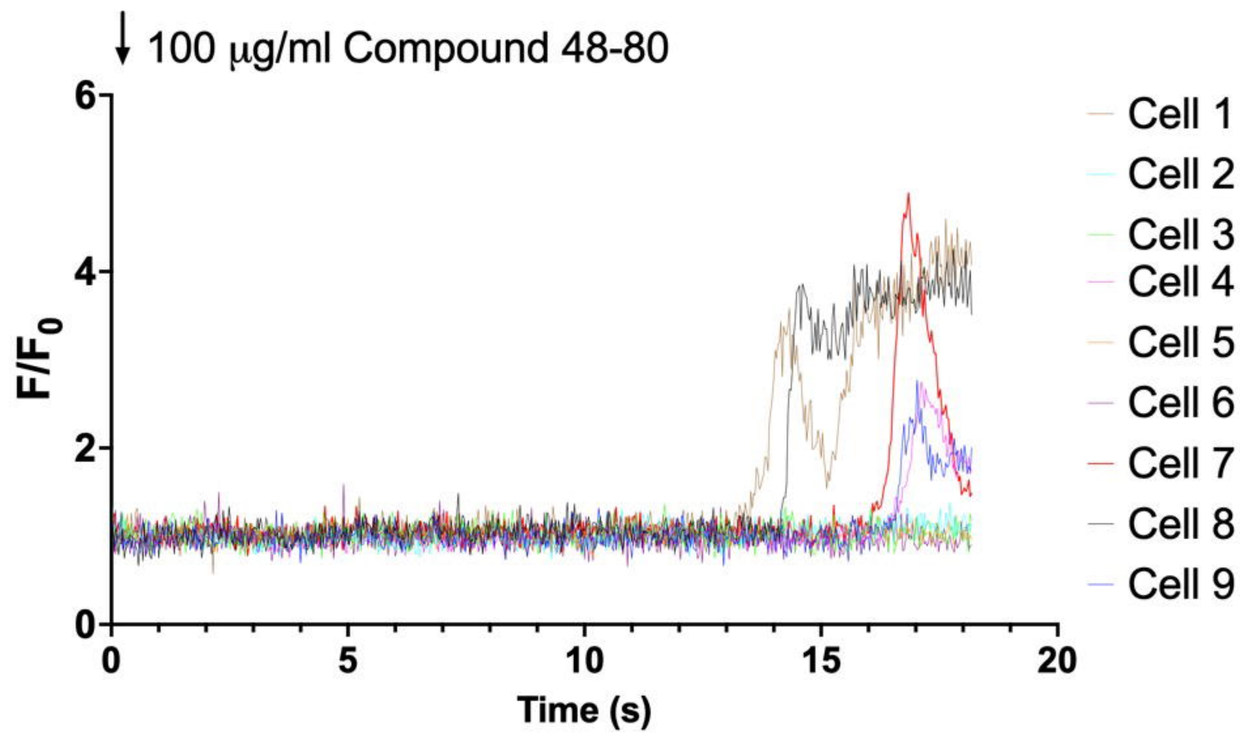


cKitCreER^{T2}-*GCaMP6f* STMs
100µg/ml Compound 48-80

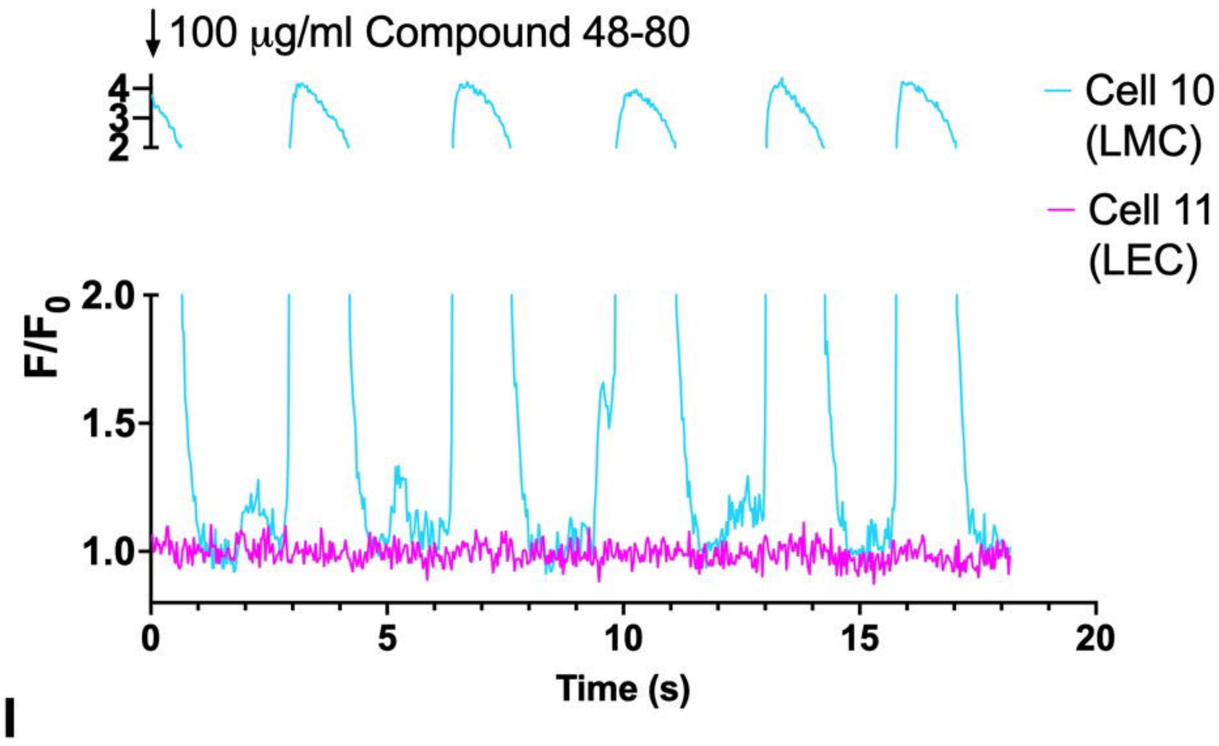
F



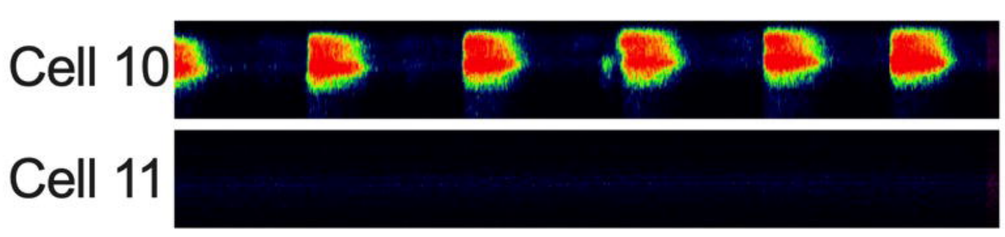
G *cKitCreER*^{T2}-*GCaMP6f* Cell Plots



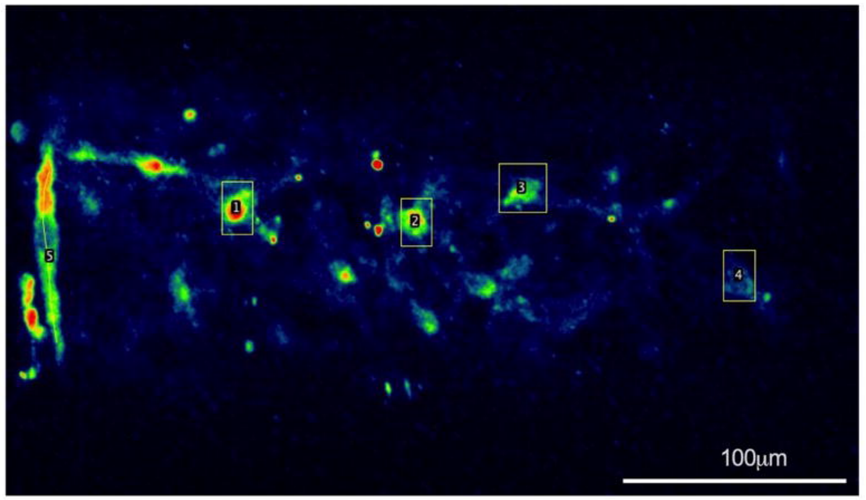
H Incidental LEC and LMC



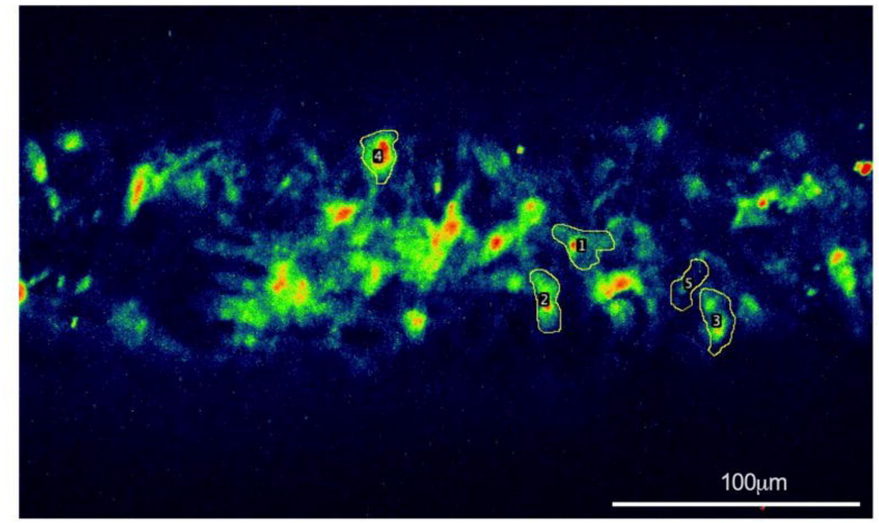
I



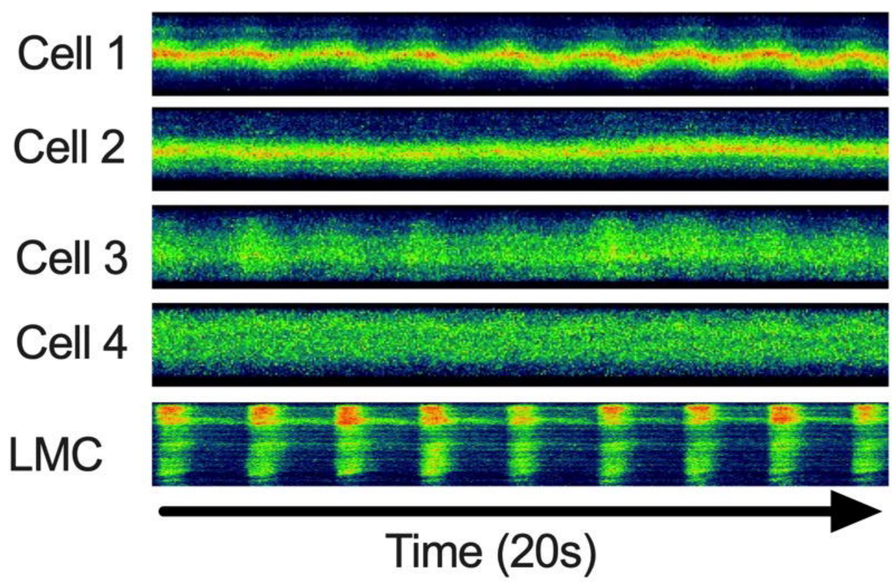
A *PdgfrαCreER*^{TM}-GCaMP6f Maximum Projection



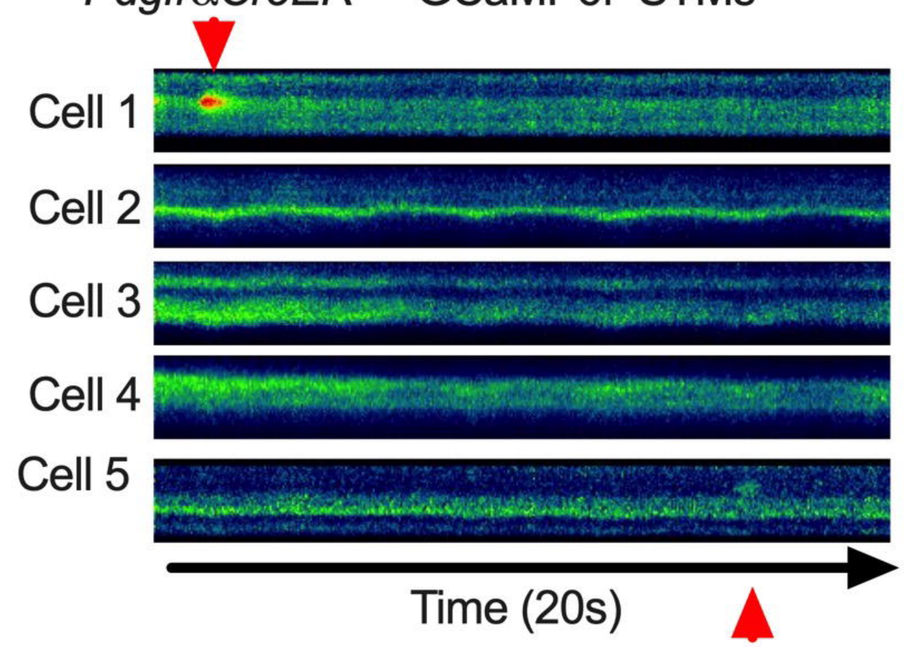
D *PdgfrαCreER*^{TM}-GCaMP6f Maximum Projection



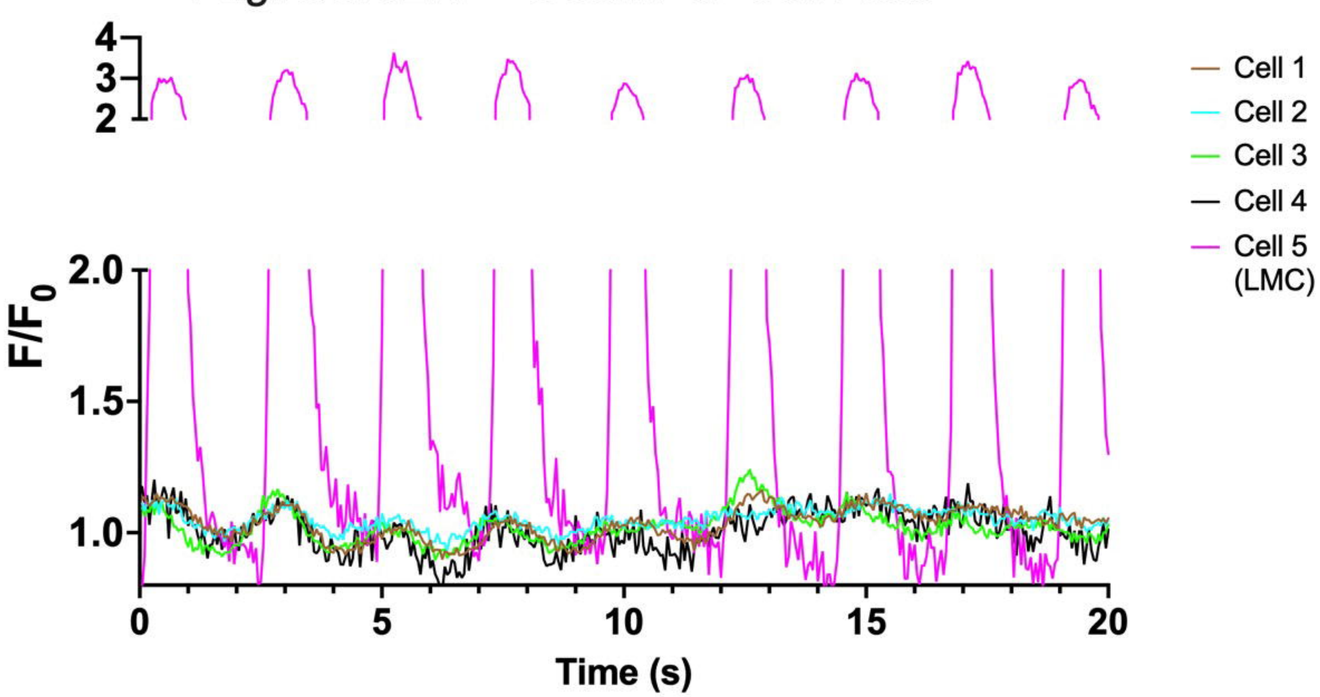
B *PdgfrαCreER*^{TM}-GCaMP6f STMs



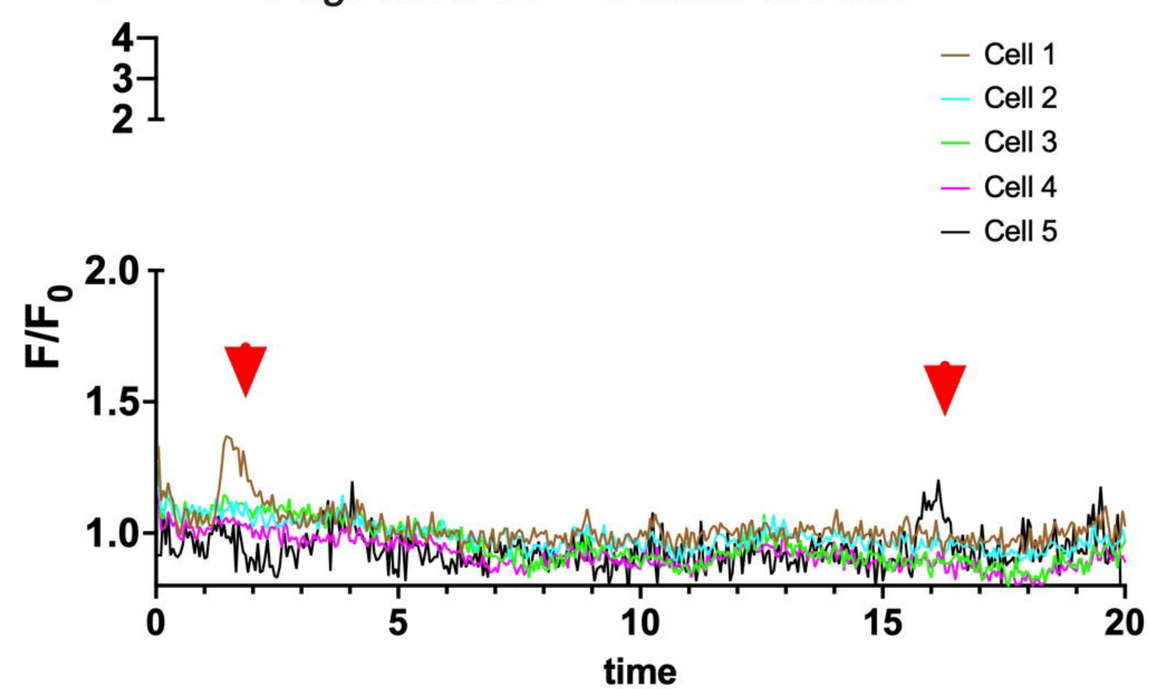
E *PdgfrαCreER*^{TM}-GCaMP6f STMs

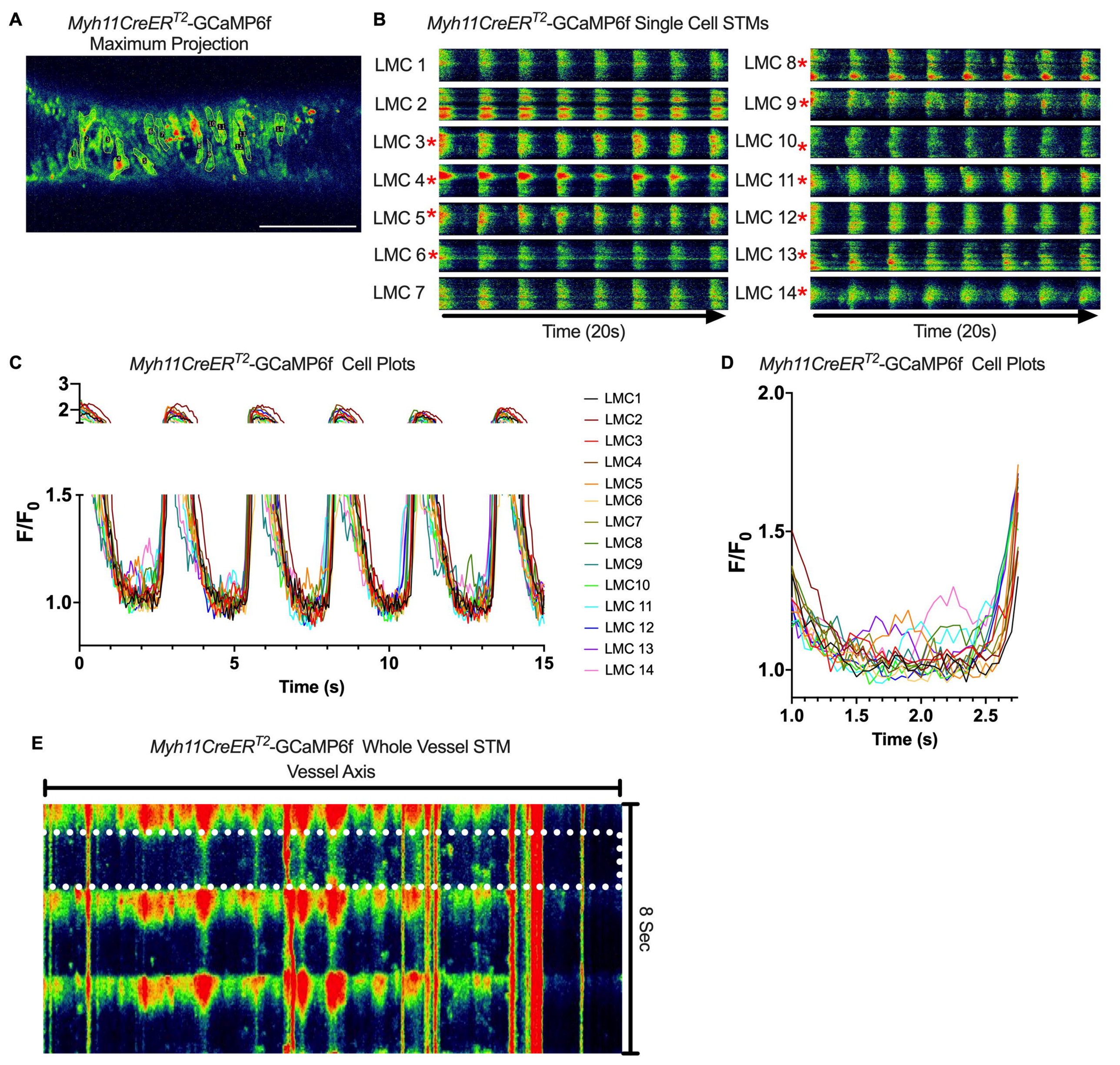


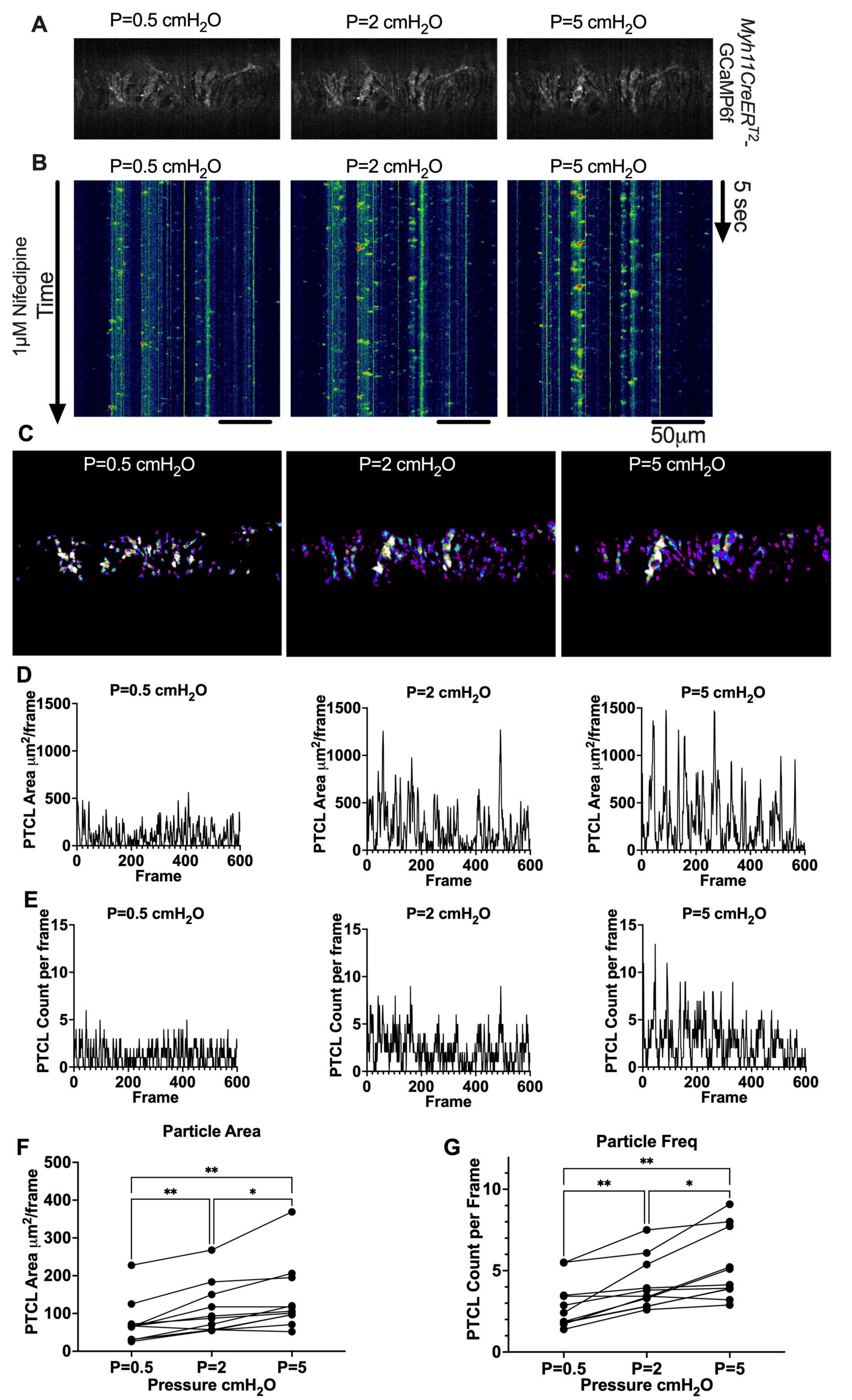
C *PdgfrαCreER*^{TM}-GCaMP6f Cell Plots

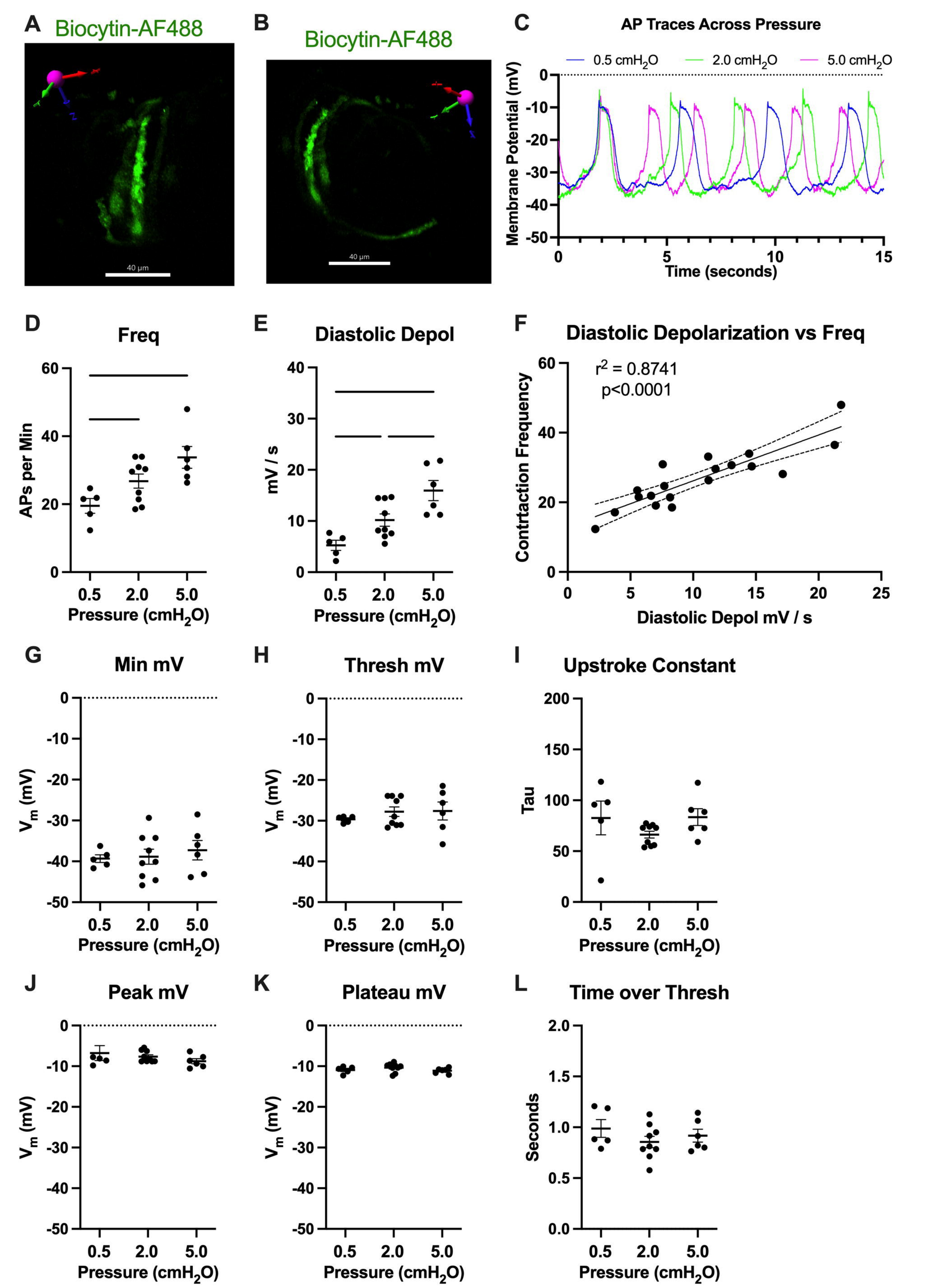


F *PdgfrαCreER*^{TM}-GCaMP6f Plots




A
Myh11CreER^{T2}-GCaMP6f
Maximum Projection
B
Myh11CreER^{T2}-GCaMP6f Single Cell STMs
LMC 1
LMC 2
LMC 3*
LMC 4*
LMC 5*
LMC 6*
LMC 7
LMC 8*
LMC 9*
LMC 10*
LMC 11*
LMC 12*
LMC 13*
LMC 14*
Time (20s)
Time (20s)
C
Myh11CreER^{T2}-GCaMP6f Cell Plots
F/F$_0$
Time (s)
LMC1
LMC2
LMC3
LMC4
LMC5
LMC6
LMC7
LMC8
LMC9
LMC10
LMC 11
LMC 12
LMC 13
LMC 14
D
Myh11CreER^{T2}-GCaMP6f Cell Plots
F/F$_0$
Time (s)
E
Myh11CreER^{T2}-GCaMP6f Whole Vessel STM
Vessel Axis
8 Sec


A
P=0.5 cmH2O
P=2 cmH2O
P=5 cmH2O
Myh11CreERT2-GCaMP6f
B
1μM Nifedipine
Time
5 sec
50μm
C
D
PTCL Area μm2/frame
Frame
E
PTCL Count per frame
F
Particle Area
PTCL Area μm2/frame
Pressure cmH2O
G
Particle Freq
PTCL Count per Frame


A
Biocytin-AF488
40 µm
B
Biocytin-AF488
40 µm
C
AP Traces Across Pressure
0.5 cmH2O
2.0 cmH2O
5.0 cmH2O
Membrane Potential (mV)
0
-10
-20
-30
-40
-50
0
5
10
15
Time (seconds)
D
Freq
APs per Min
60
40
20
0
0.5
2.0
5.0
Pressure (cmH2O)
E
Diastolic Depol
mV / s
40
30
20
10
0
0.5
2.0
5.0
Pressure (cmH2O)
F
Diastolic Depolarization vs Freq
r2 = 0.8741
p<0.0001
Contrtaction Frequency
60
40
20
0
0
5
10
15
20
25
Diastolic Depol mV / s
G
Min mV
Vm (mV)
0
-10
-20
-30
-40
-50
0.5
2.0
5.0
Pressure (cmH2O)
H
Thresh mV
Vm (mV)
0
-10
-20
-30
-40
-50
0.5
2.0
5.0
Pressure (cmH2O)
I
Upstroke Constant
Tau
200
150
100
50
0
0.5
2.0
5.0
Pressure (cmH2O)
J
Peak mV
Vm (mV)
0
-10
-20
-30
-40
-50
0.5
2.0
5.0
Pressure (cmH2O)
K
Plateau mV
Vm (mV)
0
-10
-20
-30
-40
-50
0.5
2.0
5.0
Pressure (cmH2O)
L
Time over Thresh
Seconds
2.0
1.5
1.0
0.5
0.0
0.5
2.0
5.0
Pressure (cmH2O)