Impact of Infectious Diseases on Wild Bovidae Populations in Thailand: Insights from Population Modelling and Disease Dynamics

Wantida Horpiencharoen[1], Jonathan C. Marshall[1], Renata L. Muylaert[1], Reju Sam John[2], David T. S. Hayman[1]

[1]Molecular Epidemiology and Public Health Laboratory, Hopkirk Research Institute, Massey University, Palmerston North 4472 New Zealand

[2]Department of Physics, The University of Auckland, 38 Princes St, Auckland 1010, New Zealand

Abstract

The wildlife and livestock interface is vital for wildlife conservation and habitat management. Infectious diseases maintained by domestic species may impact threatened species such as Asian bovids, as they share natural resources and habitats. To predict the population impact of infectious diseases with different traits, we used stochastic mathematical models to simulate the population dynamics 100 times over 100 years for a model gaur (*Bos gaurus*) population with and without disease. We simulated repeated introductions from a reservoir, such as domestic cattle. We selected six bovine infectious diseases; anthrax, bovine tuberculosis, hemorrhagic septicaemia, lumpy skin disease, foot and mouth disease and brucellosis, all of which have caused outbreaks in wildlife populations. From a starting population of 300, the disease-free population increased by an average of 228% over 100 years. Brucellosis with frequency-dependent transmission showed the highest average population declines (-97%), with population extinction occurring 16% of the time. Foot and mouth disease with frequency-dependent transmission showed the lowest impact, with an average population increase of 200%. Overall, acute infections with very high or low fatality had the lowest impact, whereas chronic infections produced the greatest population decline. These results may help disease management and surveillance strategies support wildlife conservation.

Keywords: Bovine, disease transmission, prediction, population, wildlife conservation

Introduction

Livestock encroachment into wildlife habitats can drive disease transmission between wildlife and domestic livestock, which is a vital issue for both human public health and wildlife conservation. An effect of agricultural expansion and land-use change is to bring wildlife and livestock close to each other and increase the contact frequency and time between domestic and wildlife populations (1-3). This increased contact may increase the risk of disease transmission as they can share the same natural resources (e.g. grassland and water) (4).

Infectious diseases can cause dramatic declines in wildlife populations, as demonstrated by chytridiomycosis, which has been implicated in the likely extinction of over 200 amphibian

species (5). Most infectious bovid pathogens are capable of infecting both domestic and wild
species. For example, bighorn sheep populations declined from ovine respiratory disease
(*Mycoplasma ovipneumoniae*) acquired when sharing the grazing areas with domestic sheep (6).
Similarly, bovine brucellosis has been transmitted from domesticated yak to wild yak in China (7),
and between bison, elk and domestic cattle in the USA (8). Brucellosis affects these bison and elk
populations both indirectly and directly as the seropositive animals may be culled for
management and directly as the pathogen affects animal reproductive systems (9). Critically, the
impact of infectious diseases is determined by disease-specific traits, such as infection fatality
rates (10).

There are five wild bovids species (gaur, banteng, wild water buffalo, mainland serow
and Chinese goral) that remain in Thailand. They are experiencing dramatic population declines
from habitat destruction, illegal hunting (11), and resource competition with domestic livestock
(12). Infectious diseases transmitted from contact with domestic cattle could cause further
declines. Several diseases circulate in Thai cattle, including endemic diseases like bovine
tuberculosis (bTB) from *Mycobacterium bovis* (13), and new infectious diseases, such as the
recent lumpy skin disease (LSD) (14).

Infectious disease modelling provides a tool to understand disease dynamics better and
predict the potential consequences of infection in a population, helping disease prevention and
control programs (15), particularly as collecting field data or conducting experiments on some
pathogens and hosts is extremely challenging. Models have, for example, been used to determine
the potential impact of disease on endangered species, such as canine distemper in the Amur
tiger (16). Although models contain uncertainty and may not cover all factors, predictions can
guide the policies and help decision-making (17).

Here, we use mathematical models to explore the potential consequences of six major
bovine infectious diseases on endangered Thai wild bovid populations. Our aim is to estimate the
potential population changes after the disease is introduced in the population from a reservoir,
such as domestic cattle. The diseases are anthrax, haemorrhagic septicaemia (HS), bTB, LSD, foot
and mouth disease (FMD) and bovine brucellosis, which all infect a range of bovid species, are
distributed worldwide, including Thailand, and have different characteristics. Our study
predominantly focuses on the gaur (*Bos gaurus*) population as their populations are well
described plus, of five species of Thai wild bovids, they have the greatest opportunity to interact
with domestic livestock and humans since they are the most likely to share space and resources
(e.g. agricultural areas, watering holes) (18, 19). We hypothesised that acute infections with very
low and very high infection fatality rates would have less impact on populations than those with
moderate mortality or chronic diseases, the latter high fatality case because they 'burn out' by
removing infectious individuals rapidly (10). The study aims to help infectious disease surveillance
and monitoring prioritisation strategies in wildlife and livestock for wild bovid conservation.

Material and methods

1. Model construction

1.1. Population dynamic models

We selected gaur populations as a model system because they are widespread across
Thailand, overlap with livestock and people, and demographic data are available (18, 20).
Further, their demography is similar to other threatened wild bovids (Figure S1 in supplementary
material). We used the same model structure for all species. The demographic parameters for
the remaining four bovid species used in simulations are provided in Table 2 because they exhibit
variations in population sizes, social behaviours, and distribution, making them interesting for
further infectious disease modelling of population impact.

We assumed demographic parameters were otherwise constant. If N is the total animal
population, N_a is the adult population, N_{sa} is the subadult population, N_c is the calf population
and μ is the annual birth rate. Only adult females were assumed to add new calves to the
population, which enter the susceptible class at a birth rate $\mu_b N_a$. Animals can leave their
compartments at the natural death rate (μ_a, μ_{sa} or μ_c) or age from calf to subadult (δ_c) and from
subadult to adult (δ_{sa}). The natural death rate was estimated based on the mortality rate of wild
ungulates and gaur in captivity (21). The initial population was 300 animals, based on the gaur
population size in the Khao Pheang Ma non-hunting area (8 km^2) in Thailand (22, 23). Thus, the
population dynamic model equations at time t can be as following:

91
$$N_t \quad = \quad N_c + N_{sa} + N_a$$

92
$$\frac{dN_c}{dt} \quad = \quad \mu_b N_a + \delta_c N_c - \mu_c N_c$$

93
$$\frac{dN_{sa}}{dt} = \quad \delta_c N_c - \delta_{sa} N_{sa} - \mu_{sa} N_{sa}$$

94
$$\frac{dN_a}{dt} \quad = \quad \delta_{sa} N_{sa} - \mu_a N_a$$

95   (Equation 1)

    1.2. Infectious disease models

We used the same age-structured population as the baseline model (Equation 1) and
incorporated compartments with different parameter values for building the disease models.

We modelled the diseases based on Susceptible-Infected-Recovered (SIR) models and
modified them based on the disease parameters of domestic animals (e.g. dairy cattle,
domesticated buffalo) and wildlife from previous studies and background knowledge. Table 1
presents the diseases and model structures we used, and a flow diagram is in the supplementary
materials. For the compartments used in the models, S denotes the number of susceptible
animals, E denotes the number of exposed animals, I denotes the number of infected animals,
R denotes the number of recovered animals, and M denotes the number of calves with
maternally derived immunity.

We selected six infectious diseases which have been reported to cause outbreaks in wild
ungulates and livestock populations in several places, including Thailand, which are: anthrax
(*Bacillus anthracis*) with an SI structure, bovine tuberculosis (bTB-*Mycobacterium bovis*) with an
SEI structure, haemorrhagic septicaemia (HS-*Pasteurella multocida*) with an $SIRS$ structure,
lumpy skin disease (LSD) with an $SEIRS$ structure, and both foot and mouth disease (FMD) and
brucellosis (*Brucella abortus*) with an $SIERMS/E$ structure. Table 2 displays the disease
parameters used in the models. These infections have a range of key parameters of interest. They
include infectious diseases with very short (effectively no) incubation periods (e.g. HS) to long
incubation periods (e.g. bTB), and very high mortality (e.g. anthrax) to low mortality (e.g. LSD,
FMD).

Table 1 Diseases, pathogens, and the structures adopted in the modelling procedures.

Disease	Pathogens	Model structure
Anthrax	*Bacillus anthracis*	$S \rightarrow I$
Bovine TB	*Mycobacterium tuberculosis*	$S \rightarrow E \rightarrow I$
Haemorrhagic septicaemia	*Pasteurella multocida*	$S \rightarrow I \rightarrow R \rightarrow S$
Lumpy Skin Disease	*Capripoxvirus*	$S \rightarrow E \rightarrow I \rightarrow R \rightarrow S$
Foot and Mouth Disease	*Aphthovirus*	$S \rightarrow E \rightarrow I \rightarrow R \rightarrow M \rightarrow S/E$
Bovine brucellosis	*Brucella abortus*	$S \rightarrow E \rightarrow I \rightarrow R \rightarrow M \rightarrow S/E$

1.3. Mode of transmission

Different transmission types can provide different model results (24). Here, we
considered two disease transmission modes; 1) density-dependent (DD) and 2) frequency-
dependent (FD). DD transmission is assumed when the contact rate is proportionate to the
population density, while FD transmission is assumed when the contact rate is independent of
the population density (24, 25). We assumed the transmission modes for each pathogen and
then compared them by introducing both transmission modes because, for some infections,

there is no clear evidence of which type suits the pathogen's transmissions and these represent

extreme situations of population change for both transmission modes (10).

Transmission is often likely a mix of both DD and FD in many cases such as FMD and bTB

(26, 27). For the transmission rate (β), we used parameter values based on the reference studies

with the reported FD or DD transmission (Table 2), which differs among infectious diseases.

However, to test the sensitivity of the results to these assumptions, we also rescaled the β rate

to all models to examine the consistency of the results between FD and DD using Equations 2:

$$\beta_{DD} = \frac{\beta_{FD}}{N}$$

$$\beta_{FD} = \beta_{DD} \; x \; N \qquad\qquad\qquad \text{(Equation 2)}$$

1.4. Infection reintroduction

To model the repeated introduction of an infection from a reservoir such as domestic

cattle (e.g. for FMD) or the environment (e.g. anthrax), we repeatedly reintroduced infection

into our population at rate ϵ independently of any infection in the population. This

reintroduction means the impact of infections are not simply estimated by the basic

reproductive number (R_0), the average number of secondary cases caused by a primary case in

a completely susceptible population.

Anthrax (Bacillus anthracis)

To model anthrax, we initially assumed the transmission is FD. We used an SI model (28,

29) with the transmission rate for FD at 0.01, then rescaled to FD using Equation 2. We assumed

that S animals are exposed to infected animals and then become infectious (I) at rate β. All

infected animals (I) die (100% mortality) (30) at disease-induced death rate (ρ) and the infectious

rate (γ), $\gamma\rho I$.

Bovine tuberculosis (bTB - *Mycobacterium tuberculosis*)

Bovine TB is a chronic and zoonotic infection in livestock and wildlife worldwide (31).

We first assumed DD transmission and used an SEI structure for modelling. The flow of the model

starts from S, which are exposed to I animals and become exposed (E) at transmission rate (β);

then E animals enter the I compartment at the incubation rate (σ). As we assume lifelong

infection without recovery (31); I animals either die with an age-specific disease-induced fatality

rate (ρ) or natural death rate (μ). S and E adults give birth with the normal birth rate $\mu_b(S_a + E_a)$

but I adults are assumed to have a lower fecundity rate (reduced by 27%, (32) at $\mu_{bI}I_a$. Bovine TB

has a long incubation period from several months up to 7 years (33), so here we used 5 months

based on the mean incubation period in the African buffalo (31). We also assumed there's no

vertical or pseudo-vertical (e.g. in utero or calf rearing) transmission as it is uncommon for bTb
(34, 35).

Haemorrhagic septicaemia (HS - *Pasteurella multocida*)

HS is a fatal septicaemic disease in cattle and buffalo. We assume DD transmission based
on a previous HS modelling study (36). We used an $SIRS$ model and excluded an E class as the
disease can show acute clinical signs with a short incubation period of $\sim$ 18-20 hours (37)
animals become I at the transmission rate (β). I animals may die from HS at the disease-induced
fatality rate (ρ) or survive and become recovered (R) at infectious rate ($1/\gamma$). We calculated the
fatality rate in the cattle population to range from 0.53% to 5.84%. This was determined by
dividing the number of deaths from HS (0.21%, assumed from the percentage of deaths from
bovine respiratory disease, (38) by the minimum (3.59%) and maximum (40%) prevalence of
seropositive animals from *P. multocida* infected herds (39, 40). Therefore, we used two infection
fatality rates (0.53 and 5.83%), since case fatality is underestimated, as a large proportion of
animals are infected but do not develop clinical signs of diseases. R animals reenter S when they
lose immunity at the immunity loss rate (ω). We used the proportion of susceptible animals (0.6)
to calculate R_0 and therefore the β rate using the equation: $R_0 = (1/(1-I)) = 1/S$.

Lumpy skin disease (LSD - Lumpy skin disease virus, *Capripoxvirus*)

We used an $SEIRS$ structure for LSD. We inserted E and R compartments as the disease
has an incubation period of between 7 and 14 days and a recovery period of around 4 - 6 months.
We initially assumed the transmission is DD as the cattle density could be one of the risk factors
to increase the transmission rate within-herd. However, we used both FD and DD β values
because the published work has reported differences in incidence rates associated with different
transmission modes (41). We assumed different birth rates for I females (μ_{bI}) from the natural
birth rate, because LSD can reduce the fertility rate by 10% (42). Also, we applied the highest
fatality rate in calves (5%) and lower mortality rates to subadults (3%) and adults (1%) (43).

Foot and mouth disease (FMD - Foot-and-mouth disease virus, *Aphthovirus*) & Bovine Brucellosis (*Brucella abortus*)

We initially assumed the transmission was FD for both FMD and brucellosis. An
$SEIRMS/E$ model was applied for FMD and brucellosis. We considered the $SEIR$ model
appropriate for both diseases. Recovered FMD and brucellosis cows can pass immunity to their
offspring. Therefore, we added a maternally-derived immunity (M) compartment, which refers
to the calves born with maternally-derived immunity from recovered mothers (R_a). We assumed
that if recovered adults (R_a) calve at the birth rate (μ_b), a calf will receive maternal immunity and
stay in M compartment for an average of 6 months (44) before immunity wanes and they become
susceptible again (S_m) at a loss of immunity rate (ω_m). S_m calves can either become an exposed

calf (E_c) if there is contact with I or enter a susceptible subadult (S_{sa}) compartment at a loss of
immunity rate plus calf ageing rate: $1/\delta_c = 1/(\delta_m + \omega_m)$ or $1/\delta_m = 1/(\delta_c - \omega_m)$ if they have no contact
with I to ensure that animals spend the same average time in the calf age class ($_c$).

Vertical transmission from mothers to calves can be a consequence of infection among
infected mothers with different probabilities for FMD (~ 0.5) and brucellosis (~ 0.9). Infectious
adults are assumed to produce an infectious calf (I_c) entering I at the birth rate $\mu_b I$. The
proportion of infected females producing infected calf denotes α. So, an infected female can
produce an infected calf at a rate $\alpha\mu_{bI}I_a$ and produce a susceptible calf at a rate $(1-\alpha)\mu_{bI}I_a$.

The mathematical ordinary differential equations for the calf population (X_c) are:

$$\frac{dS_c}{dt} = \underbrace{\mu_b(S_a + E_a) + (1-\alpha)\mu_b I_a}_{\text{birth}} - \underbrace{\beta_c S_c(I_c + I_{sa} + I_a)}_{\text{transmission rate}} - \underbrace{\delta_c S_c}_{\text{ageing rate}} - \underbrace{\mu_c S_c}_{\text{natural death}} + \underbrace{\omega_c R_c}_{\text{recovey rate}} - \underbrace{\epsilon S_c}_{\text{force of infection}}$$

$$\frac{dE_c}{dt} = \beta_c S_c(I_c + I_{sa} + I_a) + \beta_c S_m(I_c + I_{sa} + I_a) - \sigma_c E_c - \delta_c E_c - \mu_c E_c + \epsilon S_c$$

$$\frac{dI_c}{dt} = \sigma_c E_c - (1-\rho_c)\gamma_c I_c - \rho_c\gamma_c I_c - \delta_c I_c - \mu_c I_c + \alpha\mu_{bI}I_a$$

$$\frac{dR_c}{dt} = (1-\rho_c)\gamma_c I_c - \omega_c R_c - \delta_c R_c - \mu_c R_c$$

$$\frac{dM}{dt} = \mu_b R_a - \omega_M M - \mu_c M$$

$$\frac{dS_m}{dt} = \omega_M M - \delta_m S_m - \beta_m S_m(I_c + I_{sa} + I_a)$$

202   (Equation 3)

The system of ordinary differential equations and all other disease model equations and
diagrams can be found in the supplementary materials. Note that all equations are subsets or
variants of Equation 3.

2. Model simulations

Due to the small population size of gaur and other endangered bovids, we were
interested in how infections might lead to their decline. Additionally, as we aimed to allow
infections to go extinct in populations if they could not be sustained. Therefore, we chose
stochastic models for this study because they effectively capture the stochastic nature of wildlife
populations using random values. This randomness introduces variation in population sizes,
which significantly affects small population sizes and long-term simulations (45). First, we built
the population dynamics model without infectious disease classes and parameters as a baseline
model (10). Then, we introduced an infectious adult ($I_a = 1$) to the susceptible (S) population. We
assumed that I would infect S at a transmission rate, β, and enter the next compartment based
on the model structure. Demography (birth rate, natural death rate and ageing rate), the external
force of infection (ϵ), and disease-induced fatality (ρ) were included in all disease models. The
stochastic simulation was performed using the Poisson distribution to calculate the probability

of events by multiplying the rate parameters i with a time step through Gillespie's τ-leap
algorithm ($\tau = 1$) (Equation 4).

$$Prob_i = Poisson(\tau * rate_i * X)$$
(Equation 4)

Where X is a state (e.g. S, I, R). All models were simulated for 100 years, and the
stochastic models were simulated 100 times to generate the mean and to understand the
uncertainty. We modelled the population change for 100 years, as long-term simulations of at
least 10 years or three generations of species is recommended to explore population trends, and
short-term time series may lead to misleading conclusions (46).

The parameter values used for modelling were collected from the literature review and
observational data (Table 2).

3. Measuring impact

We compared the difference in total population (N) between no infection and disease
models by calculating the average percentage of the population change using the total
population at the start ($N_{t=0}$) minus the total population at the end, ($N_{t=100}$) of the simulation
time, divided by $N_{t=0}$ and converted this to a percentage, then divided by 100 times of simulations,
using the following equation:

$$\bar{x} = (\sum_{i=1}^{100} \frac{N_t - N_{t=0}}{N_{t=0}}/100) \times 100$$

(Equation 5)

We used a principal component analysis (PCA) to find which diseases showed similar
traits group by four disease parameters (transmission rate, incubation rate, infectious rate and
fatality rate) which were included in all models and then coloured the values based on the
percentage of the total population change. We performed PCA in R software using the PCATools
package (47). The highest percentage of the first two axes contributed most to the population
percentage changes.

4. Code availability

We used R Core Team (48) to simulate all the models and for further analysis. The R
code for reproducing the analyses is available at a GitHub repository
https://github.com/Wantidah/InfectiousModel.

Results

1. Disease free model

248  We developed stochastic models for a gaur population, including a baseline model
249 without infection and six infectious disease models. The baseline model of the gaur population
250 demonstrated significant population growth, increasing from 300 to an average of 685 (range
251 113 - 1469) additional animals, which is approximately a 228% (38 - 489%) increase over the 100
252 years simulated. The average adult and subadult populations consistently increased, while the
253 calf population slightly decreased from 95 to 82 animals (19 - 279) on average (Figure 1: A&I).
254 This gave us a disease-free population to model the impact of disease introduction into.

255  The population dynamics of the four other wild bovid species in Thailand show similar
256 trends to the gaur population (Figure S1, Figure 1), so we assumed there will be similar trends
257 for the other two large bovids (banteng, wild water buffalo) that have similar herd sizes,
258 population demography (e.g. age-structured, birth rate, death rate) and social behaviours to gaur
259 (49).

260  However, the population dynamics may differ from the medium-sized bovids (Chinese
261 goral and mainland serow) that live in smaller groups or even pairs and can be isolated from each
262 other (50).

2. Disease impacts

264  Brucellosis had the greatest impact on population decline, while FMD had the lowest
265 impact. Our PCA quantitatively shows that pathogens with longer incubation periods, chronic
266 infection and medium to low fatality lead to greater population decline in smaller populations of
267 endangered bovids than a high fatality or high transmission rate alone. Most diseases were
268 grouped by similar traits which can see in the PCA biplot for brucellosis, bTB and LSD, while a few
269 variations were seen for HS, and the FMD FD model was something of an outlier, (rescaling DD)
270 at β = 6552 (Figure S17 and Figure S18). The greatest contribution to the percentage population
271 change in the first axis was the infectious rate (55%) and fatality rate (42%). For the second axis,
272 is the incubation period (74%). The first axis, PC1, has 43.25% and second axis PC2 has 31.61% of
273 the variance explained.

274  Using different parameter values, fatality rates and modes of transmission yielded
275 different effects on the modelled populations for HS, FMD, LSD and brucellosis. FD brucellosis
276 had the largest population impact, yet DD brucellosis suppressed population growth but led to a
277 stable population. In contrast, FD transmission of HS, LSD and FMD showed a continued
278 population increase. Anthrax and bTB showed only a slight difference in the average population
279 change between the two transmission modes (Figure 3; (A)). Simply rescaling the β with
280 modelling FD or DD transmission had limited changes, which demonstrated consistency in the
281 population change within the same infectious disease. Rescaling the β also reduced the
282 probability of local extinction in the gaur population (N = 0) for FD brucellosis. Figure 2 and Figure
283 3 present the results of rescaling.

284   For LSD, FMD and brucellosis highlighting differences in population trends between FD
285   and DD transmissions. We selected some important model results in Figure 1, and all modelling
286   results and diagrams for infectious diseases and population changes can be found in the
287   supplementary material, Figure S2 – Figure S16.

Anthrax

There is no substantial impact on the population after introducing anthrax into the
population, with a similar population change observed between FD and DD models. Both
transmission rates showed an increase in population, with a 57% increase for FD and 51% for
rescaled DD model. No massive die offs were predicted, with only 1-3 infectious animals
predicted for each outbreak for both transmission modes, consistent with the low transmission
rate (β = 0.01) applied and a rare case of animal-to-animal transmission.

Bovine tuberculosis

There was uncertainty regarding the mode of transmission in bTB models, however, (31)
showed limited qualitative differences in model outcomes when they used FD or DD
transmission. Here, we saw similar results in that overall the populations tended to decline
gradually through the simulation period with a 88 - 89% decline from the initial population.
Rescaling the β transmission parameter also led to limited qualitative differences in population
trends, but we did see differences in predicted classes; for example, this increased or decreased
the number of infected individuals over time (i.e., higher or lower prevalence) (see
supplementary materials, Figure S5 – S6).

Haemorrhagic septicaemia

305   For HS, we found that the impact of infection was less dependent on the mode of
306   transmission than case fatality. A ten-fold increase in fatality rate led to a decline in the total
307   population change (Figure S8 – S9).

Lumpy skin disease

309   For LSD, we found that the two published transmission parameter values (0.008 and
310   0.032) led to differing outcomes that also depended on the mode of transmission (41). Whilst
311   rescaling the parameters did not lead to qualitative differences, the use of the parameter values
312   estimated from direct density-dependent transmission within herds from

313 Table 2 Parameters and variables.

Symbol	Description	Species						Units	References
		Gaur	Banteng	Buffalo	Serow	Goral			
N	Starting total population	300	470	69	120 (assume)	292 (assume)		animal	
μ_b	Birth rate	0.34	0.35	0.40	0.70	0.50		year^{-1}	(22, 51, 52)
μ_c	Calf death rate	0.27	0.26	0.27	0.50	0.45		year^{-1}	(21, 53, 54)
μ_{sa}	Subadult death rate	0.15	0.26	0.15	0.15	0.28		year^{-1}	(21, 55-57)
μ_a	Adult death rate	0.17	0.15	0.20	0.28	0.18		year^{-1}	(21, 55-57)
δ_c	Calf ageing	0.0027	0.0027	0.0027	0.0027	0.0027		day^{-1}	(58)
δ_{sa}	Subadult ageing	0.0009	0.0009	0.0009	0.0009	0.0009		day^{-1}	(58)

Symbol	Description	Disease						Units	References
		Antrax	bTB	HS	LSD	FMD	Brucellosis		
β	Disease transmission rate	0.01- 3x10^{-5}	1.4x10^{-3}	0.330	0.008-0.032	0.15-0.026	5.5x10^{-3} - 5.5x10^{-6}	day^{-1}	(29, 31, 36, 41, 59-61)
σ	1/Incubation period	0.14	6.7x10^{-3}	-	0.14	0.13	0.07	day^{-1}	(31, 41, 62-65)
γ	1/Infectious period	1	-	0.33	0.03	0.20	0.0014	day^{-1}	(66-69)
ρ_c	Disease-induced fatality in calf	1	0	0.53 - 5.84	0.05	0.10	0.10	day^{-1}	(30, 32, 38, 43, 70, 71)
ρ_{sa}	Disease-induced fatality in subadult	1	0	0.53 - 5.84	0.03	0.05	0.05	day^{-1}	(30, 32, 38, 43, 70, 71)
ρ_a	Disease-induced fatality in adult	1	0.11	0.53 - 5.84	0.01	0.03	0.03	day^{-1}	(30, 32, 38, 43, 70, 71)
α	Infected female will produce infected calf	0	-	-	-	0.50	0.9	day^{-1}	(61)
μ_{bI}	Birth rate for infectious individuals	-	6.8x10^{-4}	-	8x10^{-4}	8x10^{-4}	5x10^{-4}	day^{-1}	(32, 42, 61)
ω_c	Losing of immunity for calf	-	-	5.6x10^{-3}	5.6x10^{-3}	8.3x10^{-3}	5.6x10^{-3}	day^{-1}	(66, 72-74)
ω_{sa}	Losing of immunity for subadult	-	-	5.6x10^{-3}	5.6x10^{-3}	8.3x10^{-3}	5.6x10^{-3}	day^{-1}	(66, 72-74)
ω_a	Losing of immunity for adult	-	-	5.6x10^{-3}	5.6x10^{-3}	1.8x10^{-3}	5.6x10^{-3}	day^{-1}	(66, 72-74)
ω_m	Waning of maternal immunity	-	-	-	-	6.9x10^{-3}	5.6x10^{-3}	day^{-1}	(66, 73)
ϵ	External force of infection rate	2x10^{-5}	2x10^{-5}	2x10^{-5}	2x10^{-5}	2x10^{-5}	2x10^{-5}	day^{-1}	(10)

314
315
[To interpret the parameters, any rate r can be converted to probability $P(t)$ using $1 - exp^{rt}$, where t is the time period, e.g. for ϵ, $P(t) = 1 - exp^{\epsilon t} = 1 - exp^{2x10^{-5}*365} = 0.007$, or if the total S in the population is 300, ≈ 2 events per year. The dashed line (-) means no parameters were used in the models.]

(41) led to a population decline. However, the estimate from the indirect transmission

(presumably via mechanical transmission from flies) did not and the modelled population still

grew by 155% with FD LSD (Figure S11).

Foot and mouth disease

The least impact on the modelled population was seen in the FMD model with FD

transmission, which predicted the total population growing by 200%, around 28% less than the

disease-free population. Frequency-dependent FMD transmission with a β transmission rate of

0.115 and the rescaled DD parameter 3e-4 similarly had limited impact on the population growth

with an increasing population over time (Figure 2). Increasing β in the DD model, however,

decreased the total population by - 80% at β = 21, which had a greater impact on the population

change from 130% at β = 3e-4. FMD also showed a periodic pattern with outbreaks around every

3 - 5 years (Figure 1). Increasing the β rate from 0.11 to 21 in FD FMD models led to similar

dynamics close to DD transmission (Figure S13 – Figure S14).

Brucellosis

Brucellosis with FD transmission led to a 97% decrease in the average population change

(Figure 2 (H)) and was most likely to drive the population to local extinction with 16% of the total

simulations leading to extinction, mostly occurring from year 80-100 (Figure 1 (H, P), and Figure

S15).

Figure 1 Modelled gaur population dynamics with and without disease: A-H are single example stochastic simulations for 100 years; I-P are 100 stochastic simulations for 100 years. Mean values are in solid lines. A and I are no infection models and the others are the infectious disease models where bTB is bovine tuberculosis; HS haemorrhagic septicaemia; LSD lumpy skin disease; and FMD foot and mouth disease. The entire model results, including all disease parameters used in the simulations, can be found in the supplementary materials.

Figure 2 Population dynamics for LSD, FMD and brucellosis with different transmission modes and rescaled β transmission coefficient values to isolate the effect of the mode of transmission. A, D, G (left) are DD models; B, E, H (centre) are FD models, and C, F, I (right) are DD models with rescaled β transmission of FD parameters. Rescaling LSD (A-B) and FMD (D-E) parameters have limited impact over the period modelled, but rescaling the brucellosis β shows a reduction in FD transmission.

Figure 3 Overall modelled gaur population changes for each infection. Shown are the 100 results after 100 years of 100 stochastic simulations. The x-axis is the type of disease transmission and y axis is the population change in percentage. A) (top) compares no infection, FD and DD for anthrax, HS and bTB; B) (bottom) compares FD and DD transmission and DD* that uses the rescaled β transmission from the FD model with DD transmission for LSD, FMD, and brucellosis.

Discussion

Interactions between wildlife and livestock can facilitate the transmission of emerging infectious diseases (75), making this interface an essential area of concern to public health, animal production and wildlife conservation. We identified the potential consequences and severity of six bovine infectious diseases present in Thailand (anthrax, HS, bTB, LSD, FMD and brucellosis) in a model wild bovid population, using different infectious disease model compartments based on the current literature (Table 2). Brucellosis had the greatest population impacts and FMD the lowest, despite the same model structures being used for these two pathogens. Overall, our base model predicted population growth with varying impacts of diseases, and our analyses matched our expectation that those acute infections with very high fatality rates (anthrax and HS) have less impact than chronic infections with lower infectious rates (bTB, brucellosis), as infected individuals are rapidly removed from populations (10). Therefore, our analyses suggest that pathogens with longer incubation periods, chronic infection and low to medium fatality rates have a greater negative impact on population growth in small populations of endangered bovids (Figure 1 – Figure 3, Figure S5 – S6 and S15). This is most likely

because these traits allow infections to persist, allowing long-term infection effects on
demographic structures (e.g. reduced birth rate, increased death rate).

We used 100% fatality rates in all infected animals as the worst-case scenario for the
anthrax model, which led to limited impact over the 100 years, likely because of this rapid
removal of infected individuals (I), despite repeated reintroduction of infection (10). We first
considered anthrax transmission between infected and susceptible animals as FD transmission,
assuming contact rate is more influential than host density (29). However, the transmission mode
could also be DD, based on the density of spores in the contaminated environment (e.g. infected
carcass, soil) (28) and the cattle density that could contribute to between-species (76). Thus, we
modelled repeated introductions through ϵ to cover the external force of infections, including
the risk of disease transmission from cattle, other than within-herd transmission.

Bovine tuberculosis causes chronic, fatal infection and reduces pregnancy rates and,
therefore, the population growth of wild bovids (32, 77). There is no current evidence of bTB
infection driven population declines in Asian wild bovids, however our study found that,
regardless of both transmission modes, the long-term effect of bTB would be to reduce the
expected total population by around 88-89%. This is similar to findings by Jolles et al. (2005) (32),
who showed that bTB persisted in African buffalo populations and reduced adult buffalo
numbers primarily through mortality of animals more than 4.5 years old. The transmission
coefficient (β) was noted as one of the most important parameters for bTB in African buffalo (31).
In our work, we found consistent population dynamics between FD and DD transmission, defined
by similar trends and percentages of population change, when converting β between the original
value (from several studies) and the rescaled values (Figure 2 – Figure 3). This is likely due to the
duration of the infection, which might increase the probability of contact with infectious animals
in the population and the number of transmissions.

For HS, many animals are infected but do not develop clinical signs, making it difficult to
detect an infected animals. Further, variable clinical signs make positive cases difficult to detect,
therefore the case fatality rate is normally substantially higher than the actual infection fatality
rate. In our study, we calculated the fatality rate using the prevalence of seropositive animals
(max = 40%) from reported studies of cattle populations, and this substantially decreased the
case fatality from 90% to around 6% of animals (40, 78). Our model shows that changing the
mortality from 0.53% to 5.8% affects the total population numbers more than changing the
transmission modes, by more strongly reducing the population sizes (Figure S8 – Figure S9). HS
antibodies were found in free-ranging buffalo in Asia so this population might be a reservoir, but
this needs further investigation (37). HS is endemic among cattle in Thailand (78), and mortality
in wild ungulates has been reported historically (79), so the mortality and infection status of HS
should be considered in the mitigation plans for endangered species (e.g. wild water buffalo and
banteng).

Both FMD and LSD with FD transmission had the least impact on populations, with acute, short infections with lower overall mortality (74, 80). FMDV in particular, is highly contagious among cattle with a very high β coefficient compared to the other diseases (27). Yet, although Beck-Johnson, Gorsich (27) found little effect of FMD with either transmission modes within-herds, our results showed that DD transmission led to greater population declines, as did rescaling the parameter used for FD models. The reason for the latter observation is not clear but might be because the dynamics with reintroduction allow more infection to persist and so suppress the population (Figure 2). Note that with reported wild bovid herd sizes, acute transmission is unlikely to allow FMDV to persist, but reintroductions from cattle reservoirs (modelled through ϵ) are likely (73). Our result for FMD DD transmission also showed cyclic patterns in outbreaks consistent with seasonal patterns of outbreaks observed in Thailand (81).

We found that brucellosis with FD transmission and its reported β rate might cause extinction 16% of the time, whereas DD transmission (β = 5e-6) may suppress population growth, but not enough to cause population declines and even with the published FD rate rescaled (β = 1e-5) and used in a DD model, this caused declines but not extinctions (Figure 3; (B)). Brucellosis has caused population declines among African buffalo, especially when there is co-infection with tuberculosis (82). However, brucellosis only caused limited population growth impacts in American bison, even though the disease persisted in the population over time (61, 83). In Dobson and Meagher's study (61), their FD brucellosis models showed bison populations would increase in numbers, whereas our models predicted a decrease, perhaps because our model species' population size and structure differed from their study. Notably, *Brucella* can infect multiple species, and the transmission source may not be obvious when multiple species interact. For example, brucellosis outbreaks in Yellowstone National Park, USA, were not from wild bison as first thought, with elk the likely primary host (84). Understanding the potential transmission among and from other wild Asian ungulates may be necessary to fully understand potential brucellosis impacts.

We assumed a single, closed (no migration) population with constant natural birth and death rates. Therefore, our models explore the intrinsic population dynamics without considering the influence of other positive (e.g. conservation) or negative factors (e.g. habitat destruction, competition). Furthermore, it is unclear what population changes occur during migration (85), so a closed population model can only simulate within-herd dynamics and reflect the population impacts in a small population, such as in small protected areas (20, 22).

Selecting the appropriate transmission mode for modelling is challenging (86). The infectious disease parameter values themselves are mostly estimated from livestock outbreak data, which can vary among the regions. Rarely is infection 'natural' without intervention through disease control (27). Although the transmission type for some pathogens have been recorded as FD or DD in previous studies, these were mainly conducted under farm husbandry

or experimental conditions in captive or closed systems. These conditions significantly allow
animal density to affect contact rates. However, our study focused on wildlife populations that
are distributed in areas in which the frequency of contact could have more influence. Moreover,
some infectious diseases can display aspects of FD and DD depending on the conditions, such as
within- or between herd transmission, herd size, density, contact with other reservoirs and
contact mode (indirect, direct). For example, the bTB transmission rate can be increased
correlated to herd size if the area is stable because the density of animals is increased (87). Also,
the transmission mode for anthrax spores from animal to animal is FD, but from the environment
to an animal is based on the density of the spores in the areas. We, therefore, took the strategy
of assuming the most extreme scenarios, fully FD and DD, and used both for modelling.

453   Our models also added the external force of infection (ϵ), which represents the re-
introduction of pathogens. ϵ is assumed to include transmission from other sources of infection
other than just infectious animals, such as transmission due to environmental factors (e.g. soil,
carcasses) or vectors (e.g. blood-sucking fly) to susceptible animals (10). This transmission can
theoretically cause population extinctions if agents have high case fatality rates. Here, we chose
a relatively high reintroduction rate ($\sim$ 2 per year into the initial population), which likely
represents a worst-case scenario. However, to improve this study, we encourage adding the
specific environmental factors for each disease and incorporating spatial analyses (88, 89).

461   Further studies might also consider adding the potential reservoir hosts and their
dynamics into the models by building two or more host models to examine the transmission
route among the potential hosts (90-94). Modelling coinfection is another important point as
there are interactions between infections such as FMDV and HS, which seen as a secondary
infection in FMD outbreaks (95), or between brucellosis and bTB (82). However, our analyses
provide an approach to understanding the *relative* likely impact of common endemic and
emerging diseases with different traits and is a tool for understanding gaps in disease surveillance
and control systems by using the prediction modelling before implementing actions. Future
analyses could also determine the impact of using an Erlang-distributed waiting time, rather than
an exponential distribution, on those parameters with large amounts of variation, particularly
the incubation period (96). Another further analysis is a sensitivity analysis that can be applied
to identify the degree of influence of the disease parameters on the model output, in this case,
population change. It also suggests which state of disease transmission should prompt action and
aids in selecting optimal control measures (97).

475   Strengthening disease surveillance and mitigation programs may be further achieved by
targeting virulent diseases through passive and active surveillance data, such as collecting the
frequency of infections, number and species of wild ungulates, behaviour and time spent
together between wild and domestic livestock (particularly in the high-risk areas) (98, 99). It may
be useful for disease mitigation to largely focus on domestic animal disease control and

preventing transmission to wildlife as an amenable approach (98). Moreover, conserving wildlife
habitat can reduce the probability of contact and the risk of disease transmission between
wildlife and domestic livestock (3, 100). Limiting the contact between wildlife and livestock could
reduce species extinction (101).

With applications in wildlife conservation, a reproducible modelling framework is
advantageous for targeting pathogens that threaten other wildlife populations with similar
assumptions. Although our infectious disease modelling focused on the traits of pathogens in
one species population, our method and framework may be applicable to other wildlife
populations by incorporating their population demographics and disease parameters. This
framework is also beneficial for endangered species, enabling the simulation of various scenarios
and the identification of potential disease threats, along with estimating the recovery period
after introducing the infection.

Conclusion

Our study has provided a prediction of the potential consequence of disease in wild
bovid populations considering six important bovine infectious diseases; anthrax, HS, bTB, LSD,
FMD and brucellosis. The baseline population model shows a natural population growth of ~
228%, suggesting maintaining healthy vulnerable populations could allow them to reestablish
and overcome current levels of extinction threats while diseases and other factors may regulate
population growth. The inclusion of different disease traits has consequences on the population
numbers depending on the transmission, incubation, fatality and infectious rates. Brucellosis and
bTB models show the greatest, long-term impact among all the models, whereas FMD and LSD
showed the least impact, suggesting common but more chronic or 'slow' infections with
relatively high mortality may pose the greatest threat to smaller, threatened bovid populations.

Acknowledgements

This work was supported by funds from the New Zealand Ministry of Foreign Affairs and
Trade Manaaki New Zealand Scholarship (WH), Royal Society Te Apārangi Rutherford Discovery
Fellowship (RDF-MAU1701) and the Percival Carmine Chair in Epidemiology and Public Health
(both DTSH). We thank Prof Richard Laven from Massey University for helpful comments on HS
and Dr Rosemary Barraclough from ^{m}EpiLab for revising the manuscript draft. The authors thank
Massey University's subscription to New Zealand eScience Infrastructure (NeSI) which enabled
us to use high-performance computing facilities https://www.nesi.org.nz.

References
1.   Byrne AW, Barrett D, Breslin P, O'Keeffe J, Murphy KJ, Conteddu K, et al. Disturbance
Ecology Meets Bovine Tuberculosis (bTB) Epidemiology: A Before-and-After Study on the

Association between Forest Clearfelling and bTB Herd Risk in Cattle Herds. Pathogens.
2022;11(7).

2.   Hassell JM, Begon M, Ward MJ, Fèvre EM. Urbanization and disease emergence: dynamics
at the wildlife–livestock–human interface. Trends in ecology & evolution. 2017;32(1):55-
67.

3.   Hayman DTS. Conservation as vaccination: Integrated approaches to public health and
environmental protection could prevent future disease outbreaks. EMBO Rep.
2016;17(3):286-91.

4.   Jori F, Hernandez-Jover M, Magouras I, Dürr S, Brookes VJ. Wildlife–livestock interactions
in animal production systems: what are the biosecurity and health implications? Animal
Frontiers. 2021;11(5):8-19.

5.   Skerratt LF, Berger L, Speare R, Cashins S, McDonald KR, Phillott AD, et al. Spread of
Chytridiomycosis Has Caused the Rapid Global Decline and Extinction of Frogs. EcoHealth.
2007;4(2):125.

6.   Carpenter TE, Coggins VL, McCarthy C, O'Brien CS, O'Brien JM, Schommer TJ. A spatial risk
assessment of bighorn sheep extirpation by grazing domestic sheep on public lands.
Preventive Veterinary Medicine. 2014;114(1):3-10.

7.   Shi Q, Guo Y, Engelhardt SC, Weladji RB, Zhou Y, Long M, et al. Endangered wild yak (Bos
grunniens) in the Tibetan plateau and adjacent regions: Population size, distribution,
conservation perspectives and its relation to the domestic subspecies. Journal for Nature
Conservation. 2016;32:35-43.

8.   Cotterill GG, Cross PC, Merkle JA, Rogerson JD, Scurlock BM, du Toit JT. Parsing the effects
of demography, climate and management on recurrent brucellosis outbreaks in elk.
Journal of Applied Ecology. 2020;57(2):379-89.

9.   Schumaker BA, Peck DE, Kauffman ME. Brucellosis in the Greater Yellowstone area:
disease management at the wildlife–livestock interface. Human-Wildlife Interactions.
2012;6(1):48-63.

10.   Hayman DTS, Sam John R, Rohani P. Transmission models indicate Ebola virus persistence
in non-human primate populations is unlikely. Journal of The Royal Society Interface.
2022;19(187):20210638.

11.   Jenks KE, Kitamura S, Lynam AJ, Ngoprasert D, Chutipong W, Steinmetz R, et al. Mapping
the distribution of dholes, *Cuon alpinus* (Canidae, Carnivora), in Thailand. Mammalia.
2012;76(2):175-84.

12.   Chaiyarat R, Srikosamatara S. Populations of domesticated cattle and buffalo in the
Western Forest Complex of Thailand and their possible impacts on the wildlife
community. Journal of environmental management. 2009;90(3):1448-53.

13.   Singhla T, Boonyayatra S. Prevalence, Risk Factors, and Diagnostic Efficacy of Bovine
Tuberculosis in Slaughtered Animals at the Chiang Mai Municipal Abattoir, Thailand. Front
Vet Sci. 2022;9:846423.

14.   Punyapornwithaya V, Seesupa S, Phuykhamsingha S, Arjkumpa O, Sansamur C, Jarassaeng
C. Spatio-temporal patterns of lumpy skin disease outbreaks in dairy farms in
northeastern Thailand. Front Vet Sci. 2022;9:957306.

15.   Kirkeby C, Brookes VJ, Ward MP, Dürr S, Halasa T. A Practical Introduction to Mechanistic
Modeling of Disease Transmission in Veterinary Science. Front Vet Sci. 2020;7:546651.

16.   Gilbert M, Sulikhan N, Uphyrkina O, Goncharuk M, Kerley L, Castro EH, et al. Distemper,
559   extinction, and vaccination of the Amur tiger. Proceedings of the National Academy of
560   Sciences. 2020;117(50):31954-62.
561   17.   Singer A, Salman M, Thulke HH. Reviewing model application to support animal health
562   decision making. Prev Vet Med. 2011;99(1):60-7.
563   18.   Chaiyarat R, Prasopsin S, Bhumpakphan N. Food and nutrition of Gaur (*Bos gaurus* C.H.
564   Smith, 1827) at the edge of Khao Yai National Park, Thailand. Scientific Reports.
565   2021;11(1):3281.
566   19.   Chockcharoen R, Pharejaem T, Saisamorn A, Pattanavibool A. Gaur recovery and
567   management in Khao Phang Ma Non-Hunting Area, northeastern Thailand. BULLetin.
568   2020;3:15-22.
569   20.   Prayoon U, Suksavate W, Chaiyes A, Winitpornsawan S, Tunhikorn S, Faengbubpha K, et
570   al. Past, present and future habitat suitable for gaur (*Bos gaurus*) in Thailand. Agriculture
571   and Natural Resources. 2021;55(5):743–56-–56.
572   21.   Ahrestani FS, Iyer S, HeitkÖNig IMA, Prins HHT. Life-history traits of gaur *Bos gaurus*: a
573   first analysis. Mammal Review. 2011;41(1):75-84.
574   22.   Duengkae P, Ariyaraphong N, Tipkantha W, Jairak W, Baicharoen S, Nguyen DHM, et al.
575   Coincidence of low genetic diversity and increasing population size in wild gaur
576   populations in the Khao Phaeng Ma Non-Hunting Area, Thailand: A challenge for
577   conservation management under human-wildlife conflict. PLOS ONE.
578   2022;17(8):e0273731.
579   23.   Laichanthuek P, Sukmasuang R, Duengkae P. Population and habitat use of gaur (*Bos
580   gaurus*) around Khao Phaeng Ma area, Nakhon Ratchasima province. Journal of Wildlife in
581   Thailand. 2017;24:83 - 95.
582   24.   McCallum H, Barlow N, Hone J. How should pathogen transmission be modelled? Trends
583   in Ecology & Evolution. 2001;16(6):295-300.
584   25.   Smith KF, Acevedo-Whitehouse K, Pedersen AB. The role of infectious diseases in
585   biological conservation. Animal Conservation. 2009;12(1):1-12.
586   26.   Ciaravino G, García-Saenz A, Cabras S, Allepuz A, Casal J, García-Bocanegra I, et al.
587   Assessing the variability in transmission of bovine tuberculosis within Spanish cattle
588   herds. Epidemics. 2018;23:110-20.
589   27.   Beck-Johnson LM, Gorsich EE, Hallman C, Tildesley MJ, Miller RS, Webb CT. An exploration
590   of within-herd dynamics of a transboundary livestock disease: A foot and mouth disease
591   case study. Epidemics. 2023;42:100668.
592   28.   Friedman A, Yakubu AA. Anthrax epizootic and migration: persistence or extinction. Math
593   Biosci. 2013;241(1):137-44.
594   29.   Saad-Roy CM, van den Driessche P, Yakubu AA. A Mathematical Model of Anthrax
595   Transmission in Animal Populations. Bull Math Biol. 2017;79(2):303-24.
596   30.   Sinkie ZM. Modeling And Simulation Study Of Anthrax Attack On Environment.
597   2016;3(4):5.
598   31.   Cross PC, Getz WM. Assessing vaccination as a control strategy in an ongoing epidemic:
599   Bovine tuberculosis in African buffalo. Ecological Modelling. 2006;196(3):494-504.
600   32.   Jolles AE, Cooper DV, Levin SA. Hidden Effects of Chronic Tuberculosis in African Buffalo.
601   Ecology. 2005;86(9):2358-64.

33. Barlow ND, Kean JM, Hickling G, Livingstone PG, Robson AB. A simulation model for the spread of bovine tuberculosis within New Zealand cattle herds. Prev Vet Med. 1997;32(1-2):57-75.

34. Menzies FD, Neill SD. Cattle-to-cattle transmission of bovine tuberculosis. Vet J. 2000;160(2):92-106.

35. Phillips CJ, Foster CR, Morris PA, Teverson R. The transmission of *Mycobacterium bovis* infection to cattle. Res Vet Sci. 2003;74(1):1-15.

36. Deepak DGaSSA. Modeling Vaccination Programs in Outbreaks of Hemorrhagic Septicemia in India. Journal of Animal Research. 2020;10.

37. De Alwis MC. Haemorrhagic septicaemia--a general review. Br Vet J. 1992;148(2):99-112.

38. Baptista AL, Rezende AL, Fonseca PA, Massi RP, Nogueira GM, Magalhães LQ, et al. Bovine respiratory disease complex associated mortality and morbidity rates in feedlot cattle from southeastern Brazil. J Infect Dev Ctries. 2017;11(10):791-9.

39. Khan A, Sadique U, Ahmad R, Khan H, Mohammad Y, Zubair M. Sero-surveillance of hemorrhagic septicemia in cattle and buffaloes in district Malakand, NWFP. J Agric Biol Sci. 2006;1.

40. Nobrega D, Andres-Lasheras S, Zaheer R, McAllister T, Homerosky E, Anholt RM, et al. Prevalence, Risk Factors, and Antimicrobial Resistance Profile of Respiratory Pathogens Isolated From Suckling Beef Calves to Reprocessing at the Feedlot: A Longitudinal Study. Front Vet Sci. 2021;8:764701.

41. Magori-Cohen R, Louzoun Y, Herziger Y, Oron E, Arazi A, Tuppurainen E, et al. Mathematical modelling and evaluation of the different routes of transmission of lumpy skin disease virus. Vet Res. 2012;43(1):1.

42. Abdulqa HY, Rahman HS, Dyary HO, Othman HH. Lumpy Skin Disease. Reproductive Immunology: Open Access. 2016;1(4).

43. Das M, Chowdhury MSR, Akter S, Mondal AK, Uddin MJ, Rahman MM, et al. An updated review on lumpy skin disease: perspective of Southeast Asian countries. J adv biotechnol exp ther. 2021;4(3):322-33.

44. Laubscher L, Hoffman L. An Overview of Disease-Free Buffalo Breeding Projects with Reference to the Different Systems Used in South Africa. Sustainability. 2012;4(11):3124-40.

45. Wei F, Geritz SA, Cai J. A stochastic single-species population model with partial pollution tolerance in a polluted environment. Applied Mathematics Letters. 2017;63:130-6.

46. White ER. Minimum Time Required to Detect Population Trends: The Need for Long-Term Monitoring Programs. BioScience. 2018;69(1):40-6.

47. Blighe K, Lun A. PCAtools: PCAtools: Everything Principal Components Analysis. R package version 2.14.0 ed2019.

48. R Core Team. R: A language and environment for statistical computing. 4.1 ed. Vienna, Austria: R Foundation for Statistical Computing; 2022.

49. Srikosamatara S. Density and biomass of large herbivores and other mammals in a dry tropical forest, western Thailand. Journal of Tropical Ecology. 1993;9(1):33-43.

50. Chen Y, Xiao Z, Zhang L, Wang X, Li M, Xiang Z. Activity Rhythms of Coexisting Red Serow and Chinese Serow at Mt. Gaoligong as Identified by Camera Traps. Animals. 2019;9(12):1071.

51.   Khiowsree N, Sukmasuang R, Kutintara U. Population monitoring and distribution range of
wild buffalo (*Bubalus bubalis* LINNAEUS, 1758) in Huai Kha Khaeng wildlife sanctuary,
Uthai Thani province. Journal of Wildlife in Thailand. 2015;22(1):112 - 8.
52.   Srikosamatara S, Suteethorn V. Populations of gaur and banteng and their management in
Thailand. Natural History Bulletin of the Siam Society. 1995;43(1):55-83.
53.   Choquent D. Growth, body condition and demography of wild banteng (*Bos javanicus*) on
Cobourg Peninsula, northern Australia. Journal of Zoology. 1993;231(4):533-42.
54.   McCullough DR. The african buffalo: a study of resource limitation of population. Univ.
Chicago Press, Chicago, xii + 355 pp., 1977. Price $20.00. Journal of Mammalogy.
1979;60(2):438-9.
55.   Gardner P, Pudyatmoko S, Bhumpakphan N, Yindee M, Ambu D, Goossens B. Banteng Bos
javanicus d'Alton, 1823., Ecology, Evolution and Behaviour of Wild Cattle: Implications for
Conservation, ed. M. Melletti and J. Burton. Published by Cambridge University Press;
2014.
56.   Heinen JT. Population viability and management recommendations for wild water buffalo
Bubalus bubalis in Kosi Tappu Wildlife Reserve, Nepal. Biological Conservation.
1993;65(1):29-34.
57.   Tang S, Li C, Liu W, Bao W. Viability analysis of Chinese goral population in Saihanwula
National Nature Reserve, Inner Mongolia. Scientia Silvae Sinicae. 2019;55(3):118-24.
58.   Jones KE, Bielby J, Cardillo M, Fritz SA, O'Dell J, Orme CDL, et al. PanTHERIA: a species-
level database of life history, ecology, and geography of extant and recently extinct
mammals. Ecology. 2009;90(9):2648-.
59.   Cabezas AH, Sanderson MW, Volkova VV. A Meta-Population Model of Potential Foot-
and-Mouth Disease Transmission, Clinical Manifestation, and Detection Within U.S. Beef
Feedlots. Frontiers in Veterinary Science. 2020;7.
60.   Wongsathapornchai K, Salman M, Edwards JR, Morley PS, Keefe TJ, Campen HV, et al. Use
of epidemiologic risk modeling to evaluate control of foot-and-mouth disease in southern
Thailand. American journal of veterinary research. 2008;69(2):240-51.
61.   Dobson A, Meagher M. The Population Dynamics of Brucellosis in the Yellowstone
National Park. Ecology. 1996;77(4):1026-36.
62.   WHO. Anthrax in humans and animals: World Health Organization International Office of
Epizootics; 2008.
63.   WOAH. Haemorrhagic Septicaemia: WOAH - World Organisation for Animal Health; 2021
[Available from: https://www.woah.org/app/uploads/2021/09/haemorrhagic-
septicemia.pdf.
64.   Sun GQ, Li MT, Zhang J, Zhang W, Pei X, Jin Z. Transmission dynamics of brucellosis:
Mathematical modelling and applications in China. Comput Struct Biotechnol J.
2020;18:3843-60.
65.   Hugh-Jones ME, Tinline RR. Studies on the 1967--68 foot and mouth disease epidemic:
incubation period and herd serial interval. J Hyg (Lond). 1976;77(2):141-53.
66.   Hobbs NT, Geremia C, Treanor J, Wallen R, White PJ, Hooten MB, et al. State-space
modeling to support management of brucellosis in the Yellowstone bison population.
Ecological Monographs. 2015;85(4):525-56.

67.   Mardones F, Perez A, Sanchez J, Alkhamis M, Carpenter T. Parameterization of the
690   duration of infection stages of serotype O foot-and-mouth disease virus: an analytical
691   review and meta-analysis with application to simulation models. Veterinary research.
692   2010;41(4).
693   68.   Mat B, Arikan MS, Akin AC, Çevrimli MB, Yonar H, Tekindal MA. Determination of
694   production losses related to lumpy skin disease among cattle in Turkey and analysis using
695   SEIR epidemic model. BMC Veterinary Research. 2021;17:1-10.
696   69.   Sinkie ZM, Narasimha MS. Modeling and simulation study of anthrax attack on
697   environment. Journal of multidisciplinary Engineering and Technology (JMEST). 2016;3:4.
698   70.   Govindaraj G, Krishnamohan A, Hegde R, Kumar N, Prabhakaran K, Wadhwan VM, et al.
699   Foot and Mouth Disease (FMD) incidence in cattle and buffaloes and its associated farm-
700   level economic costs in endemic India. Preventive Veterinary Medicine. 2021;190:105318.
701   71.   Njeru J, Nthiwa D, Akoko J, Oyas H, Bett B. Incidence of Brucella infection in various
702   livestock species raised under the pastoral production system in Isiolo County, Kenya.
703   BMC Vet Res. 2021;17(1):342.
704   72.   Verma R, Jaiswal T. Haemorrhagic septicaemia vaccines. Vaccine. 1998;16(11-12):1184-
705   92.
706   73.   Jolles A, Gorsich E, Gubbins S, Beechler B, Buss P, Juleff N, et al. Endemic persistence of a
707   highly contagious pathogen: Foot-and-mouth disease in its wildlife host. Science.
708   2021;374(6563):104-9.
709   74.   Tuppurainen ES, Oura CA. Review: lumpy skin disease: an emerging threat to Europe, the
710   Middle East and Asia. Transbound Emerg Dis. 2012;59(1):40-8.
711   75.   Wiethoelter AK, Beltrán-Alcrudo D, Kock R, Mor SM. Global trends in infectious diseases
712   at the wildlife–livestock interface. Proceedings of the National Academy of Sciences.
713   2015;112(31):9662-7.
714   76.   Otieno FT, Gachohi J, Gikuma-Njuru P, Kariuki P, Oyas H, Canfield SA, et al. Modeling the
715   spatial distribution of anthrax in southern Kenya. PLoS Negl Trop Dis.
716   2021;15(3):e0009301.
717   77.   Joly DO, Messier F. The effect of bovine tuberculosis and brucellosis on reproduction and
718   survival of wood bison in Wood Buffalo National Park. Journal of Animal Ecology.
719   2005;74(3):543-51.
720   78.   Singhla T, Tankaew P, Sthitmatee N. Validation of a Novel ELISA for the Diagnosis of
721   Hemorrhagic Septicemia in Dairy Cattle from Thailand Using a Bayesian Approach. Vet Sci.
722   2020;7(4).
723   79.   Robinson S, Milner-Gulland EJ, Grachev Y, Salemgareyev A, Orynbayev M, Lushchekina A,
724   et al. Opportunistic bacteria and mass mortality in ungulates: lessons from an extreme
725   event. Ecosphere. 2019;10(6):e02671.
726   80.   Jori F, Etter E. Transmission of foot and mouth disease at the wildlife/livestock interface
727   of the Kruger National Park, South Africa: Can the risk be mitigated? Prev Vet Med.
728   2016;126:19-29.
729   81.   Chanchaidechachai T, Saatkamp H, de Jong M, Inchaisri C, Hogeveen H, Premashthira S, et
730   al. Epidemiology of foot-and-mouth disease outbreaks in Thailand from 2011 to 2018.
731   Transbound Emerg Dis. 2022;69(6):3823-36.

82. Gorsich EE, Etienne RS, Medlock J, Beechler BR, Spaan JM, Spaan RS, et al. Opposite
outcomes of coinfection at individual and population scales. Proc Natl Acad Sci U S A.
2018;115(29):7545-50.
83. National Academies of Sciences E, Medicine, Division on E, Life S, Board on A, Natural R,
et al. Revisiting Brucellosis in the Greater Yellowstone Area. Washington (DC): National
Academies Press (US); 2017.
84. Godfroid J. *Brucella* spp. at the Wildlife-Livestock Interface: An Evolutionary Trajectory
through a Livestock-to-Wildlife "Host Jump"? Vet Sci. 2018;5(3).
85. Ahrestani FS, Karanth KU. Gaur *Bos gaurus* C.H. Smith, 1827. In: Melletti M, Burton J,
editors. Ecology, Evolution and Behaviour of Wild Cattle. Cambridge: Cambridge
University Press; 2014. p. 174-93.
86. Luis AD, Kuenzi AJ, Mills JN. Species diversity concurrently dilutes and amplifies
transmission in a zoonotic host-pathogen system through competing mechanisms. Proc
Natl Acad Sci U S A. 2018;115(31):7979-84.
87. Alvarez J, Bezos J, de la Cruz ML, Casal C, Romero B, Domínguez L, et al. Bovine
tuberculosis: within-herd transmission models to support and direct the decision-making
process. Res Vet Sci. 2014;97 Suppl:S61-8.
88. Pepin KM, Brown VR, Yang A, Beasley JC, Boughton R, VerCauteren KC, et al. Optimising
response to an introduction of African swine fever in wild pigs. Transbound Emerg Dis.
2022;69(5):e3111-e27.
89. Rohani P, Breban R, Stallknecht DE, Drake JM. Environmental transmission of low
pathogenicity avian influenza viruses and its implications for pathogen invasion. Proc Natl
Acad Sci U S A. 2009;106(25):10365-9.
90. Aranaz A, De Juan L, Montero N, Sánchez C, Galka M, Delso C, et al. Bovine tuberculosis
(*Mycobacterium bovis*) in wildlife in Spain. J Clin Microbiol. 2004;42(6):2602-8.
91. Godfroid J, Garin-Bastuji B, Saegerman C, Blasco JM. Brucellosis in terrestrial wildlife. Rev
Sci Tech. 2013;32(1):27-42.
92. Mohamed A. Bovine tuberculosis at the human-livestock-wildlife interface and its control
through one health approach in the Ethiopian Somali Pastoralists: A review. One Health.
2020;9:100113.
93. Royce K, Fu F. Mathematically modeling spillovers of an emerging infectious zoonosis with
an intermediate host. PLoS One. 2020;15(8):e0237780.
94. Roche X, Rozstalnyy A, TagoPacheco D, Pittiglio C, Kamata A, Beltran Alcrudo D, et al.
Introduction and spread of lumpy skin disease in South, East and Southeast Asia:
Qualitative risk assessment and management: Food & Agriculture Org.; 2021.
95. Chandranaik BM, Hegde R, Shivashankar BP, Giridhar P, Muniyellappa HK, Kalge R, et al.
Serotyping of foot and mouth disease virus and Pasteurella multocida from Indian gaurs
(*Bos gaurus*), concurrently infected with foot and mouth disease and haemorrhagic
septicaemia. Tropical Animal Health and Production. 2015;47(5):933-7.
96. Champredon D, Dushoff J, Earn DJD. Equivalence of the Erlang-Distributed SEIR Epidemic
Model and the Renewal Equation. SIAM Journal on Applied Mathematics.
2018;78(6):3258-78.

97. Nadeem Anjam Y, Shahid I, Emadifar H, Arif Cheema S, ur Rahman M. Dynamics of the
optimality control of transmission of infectious disease: a sensitivity analysis. Scientific
Reports. 2024;14(1):1041.
98. Bengis R, Kock R, Fischer J. Infectious animal diseases: the wildlife/livestock interface.
Revue scientifique et technique (International Office of Epizootics). 2002;21(1):53-65.
99. Silk MJ, Hodgson DJ, Rozins C, Croft DP, Delahay RJ, Boots M, et al. Integrating social
behaviour, demography and disease dynamics in network models: applications to disease
management in declining wildlife populations. Philosophical Transactions of the Royal
Society B. 2019;374(1781):20180211.
100. Caron A, Miguel E, Gomo C, Makaya P, Pfukenyi DM, Foggin C, et al. Relationship between
burden of infection in ungulate populations and wildlife/livestock interfaces.
Epidemiology & Infection. 2013;141(7):1522-35.
101. Pedersen AB, Jones KE, Nunn CL, Altizer S. Infectious diseases and extinction risk in wild
mammals. Conservation Biology. 2007;21(5):1269-79.