Altered reactivity to threatening stimuli in *Drosophila* models of Parkinson's disease, revealed by a trial-based assay

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Abstract

The fruit fly *Drosophila melanogaster* emerges as an affordable, genetically tractable model of behavior and brain diseases. However, despite the surprising level of evolutionary conservation from flies to humans, significant genetic, circuit-level and behavioral differences hinder the interpretability of fruit fly models for human disease. Therefore, to allow a more direct fly-versus-human comparison, we surveyed the rarely exploited, rich behavioral repertoire of fruit flies with genetic alterations relevant to Parkinson's disease (PD), including overexpression of human mutant Parkin or α-synuclein proteins and mutations in

dopamine receptors. Flies with different genetic backgrounds displayed variable behaviors, including freezing, slowing and running, in response to predator-mimicking passing shadows used as threatening stimuli in a single-animal trial-based assay. We found that the expression of human mutant Parkin in flies resulted in reduced walking speed and decreased reactivity to passing shadows. Flies with dopamine receptor mutations showed similar alterations, consistent with the motor and cognitive deficits typical in humans with PD. We also found age-dependent trends in behavioral choice during the fly lifespan, while dopamine receptor mutant flies maintained their decreased general reactivity throughout all age groups. Our data demonstrate that single-trial behavioral analysis can reveal subtle behavioral changes in mutant flies that can be used to further our understanding of disease pathomechanisms and help gauge the validity of genetic *Drosophila* models of neurodegeneration, taking us one step closer to bridging the gap in fly-to-human translation.

Introduction

Parkinson's disease (PD) affects over 6 million people worldwide, representing the second most common neurodegenerative disease. PD is slowly progressing and typically leads to years of aggravating disability, thereby placing a huge burden on families, health care systems and the society, measured in hundreds of billions of dollars annually (Olesen et al., 2012; Obeso et al., 2017; Przedborski, 2017; Bloem et al., 2021). Therefore, a hitherto elusive disease-modifying therapy is of prominent priority, envisioned to be fueled by research into basic mechanisms of the disease.

Patients with PD display a diverse set of motor and non-motor symptoms with an underlying progressive loss of midbrain dopaminergic neurons (Schapira, 2009; Schapira et al., 2017; Bloem et al., 2021), mechanisms of which are widely studied in animal models of the disease. Primate models offer the advantage of more direct translatability but are restricted to hard-to-handle neurotoxin approaches and do not promise fast progress. Unlike the genetic mouse models of Alzheimer's disease, most of the successful rodent models of PD are also toxin-based, making it difficult to exploit and further our understanding of the genetic bases of the disease (Cannon and Greenamyre, 2010; Bové and Perier, 2012; Breger and Fuzzati Armentero, 2019). This left a niche for the genetically tractable, affordable fruit flies as genetic models of PD (Feany and Bender, 2000; Guo, 2012; Hewitt and Whitworth, 2017), building on the homologies between the vertebrate basal ganglia and the fruit fly central complex (Strausfeld and Hirth, 2013). However, substantial genetic, anatomical, physiological, and behavioral discrepancies between insects and mammals call for better validation methods of fruit fly models for human disease. Therefore, to provide means for a more direct comparison between flies and mammals, we developed a

single-animal trial-based behavioral assay that facilitates fine-grained assessment of phenotypical behavioral changes in fruit flies with genetic alterations relevant to understanding human PD.

Genes linked to familial forms of PD may serve as an ideal basis for genetic disease models. Notably, mutations in the *PARK2* gene, which encodes the Parkin protein involved in maintaining mitochondrial integrity, are associated with autosomal-recessive forms of PD (Guo, 2012). Parkin loss-of-function mutant flies were found to have advanced mitochondrial aging, structural mitochondrial damage and a consequential selective loss of dopaminergic neurons (Cackovic et al., 2018), leading to motor deficits assessed by a climbing assay (Chambers et al., 2013; Cackovic et al., 2018), as well as non-motor PD phenotypes including memory deficits (Julienne et al., 2017). Mutations in the *SNCA* gene of α-Synuclein (α-Syn) are also associated with familiar forms of PD (Polymeropoulos et al., 1997) and α-Syn has been shown to accumulate in Lewy-bodies and -neurites (Spillantini et al., 1997). α-Syn proteins have been found in the pre-synaptic terminals in humans and mice (Kahle et al., 2000) and are thought to be involved in regulating dopamine (DA) synthesis under physiological conditions by reducing tyrosine hydroxylase activity (Perez et al., 2002). Expression of human α-Syn in flies has been proposed as a genetic model of PD, showing age-dependent loss of dopaminergic neurons and locomotor dysfunction in a climbing assay (Feany and Bender, 2000; Haywood and Staveley, 2006). However, other studies found normal locomotion and dopaminergic cell counts in these flies, casting doubts on the validity of this model (Pesah et al., 2005; Nagoshi, 2018).

Despite the observed homology between mammalian and fruit fly DA systems in motor control and the establishment of *Drosophila* PD models based on human genetic information derived from familial PD patients, the role of *Drosophila* DA receptors in locomotor control is not well characterized. Nevertheless, it has been demonstrated that the D1-like DA receptor mediates ethanol-induced locomotion in the ellipsoid body (Kong et al., 2010). Furthermore, *Dop1R1* has been shown to be involved in turning behavior for goal-directed locomotion (Kottler et al., 2019) and startle-induced negative geotaxis (Sun et al., 2018). To shed light on the possible roles of DA receptors in threat-induced motor behaviors, we tested the behavioral responses of three dopamine receptor (*Dop1R1*, *Dop1R2* and *DopEcR*) insertion mutant lines to predator-mimicking passing shadows and compared them to established fruit fly PD models with partially known locomotor deficits.

We found that flies expressing the R275W mutant allele of human Parkin ('Parkin flies') showed slower average locomotion speed, which, in contrast, was not a characteristic of flies expressing the A53T mutant allele of the human *SNCA* gene ('α-Syn flies'). Parkin flies also showed less behavioral reactivity to passing

shadows compared to controls, whereas α-Syn flies showed increased durations of stopping after the stimuli. Dopamine receptor mutant flies showed reduced speed and less behavioral reactivity similar to Parkin flies. *Dop1R1* mutant flies exhibited more pronounced behavioral alterations than the other two receptor mutants in most of the parameters tested. These data demonstrate that mutations in DA receptor genes lead to specific patterns of behavioral deficits in *Drosophila*; hence, these dopamine receptor paralogs may have different functions in behavioral control. The modest phenotype of A53T α-Syn compared to Parkin flies suggests that the latter should be favored as a genetic model of human PD, at least with respect to the motor deficits examined in this study. We further propose that single-trial analyses such as those we present here help us gain a better understanding of the behavioral changes in fruit fly models of PD and are strong tools for validating *Drosophila* models of human diseases.

Results

A single-animal trial-based assay to test behavioral responses to predator-mimicking passing shadows

We designed a behavioral apparatus to examine the responses of individual flies to predator-mimicking passing shadows. To do this, we designed a transparent plexiglass arena (Fig. 1a), featuring 13 tunnels (53 mm x 5 mm) for simultaneous tracking of 13 individual flies. The height of the tunnels (1.5 mm) was designed to allow free walking in two dimensions but prevent jumps and flight. To simulate predatory threat, we created passing 'shadows' (Fig. 1a, right; see Methods) with a sliding red screen presented on a 10.1-inch display placed on the top of the arena. A high frame-rate camera was placed under the arena (Fig. 1a, left), allowing us to simultaneously record the movement of the animals and the shadows (Supplementary Movie 1 and 2). All 13 tunnels housed a single fly in each session, enabling the collection of single-animal data from 13 flies in parallel. The locomotion of individual flies was tracked by custom software developed using the Bonsai visual programming environment (Fig. S1, see Methods). We recorded 40-minute-long sessions, consisting of 40 trials of 2-second-long shadow presentations separated by pseudorandom inter-trial-intervals to prevent the animals from learning temporal expectations of the shadow presentations (Fig. 1b).

Fruit flies exhibit a rich behavioral repertoire upon threatening stimuli, including freezing (or stopping) and various escape behaviors such as jumping, slow or fast take-off and running, modulated by walking speed at the time of the threatening stimulus (Zacarias et al., 2018). To study these behaviors, we calculated the speed and acceleration of fruit flies based on the tracked x-y position of their center of

mass (Fig. 2a) and aligned these signals to the presentations of the shadow stimuli. To identify separate response types to the threating stimuli, we applied hierarchical clustering on these stimulus-aligned speed traces (see Methods). This analysis consistently revealed three main stereotypical behavioral responses across flies in addition to the trials where no significant response was evoked (Fig. 2b). Since jumping and flying were not possible in the arena, fruit flies were restricted to choose among freezing, slowing (also observed in (Zacarias et al., 2018)) and running. While this clustering approach was sufficient to reveal response types qualitatively, cluster boundaries were sensitive to genotype-specific differences across groups of flies (Király and Hangya, 2022). Therefore, for rigorous group comparisons, we determined exact definitions of each response type based on fly speed and acceleration (Fig. 2c). Reactions were considered robust if the absolute value of the acceleration reached 200 mm/s^2; otherwise, the trial was classified as a 'no reaction' trial. We considered the reaction as a 'stop' if the animal was moving before shadow presentation and its speed decreased to zero in the first second relative to the shadow presentation. If fly speed decreased to a non-zero value, the trial was defined as 'slow down', and if the fly accelerated after the shadow presentation, the trial was classified as 'speed up' (see also Methods).

Parkinson's model and dopamine receptor mutant fruit flies showed reduced walking speed and decreased reactivity to threatening stimuli

To generate PD fly models, we expressed the human mutant Parkin (275W) and α-Syn (A53T) coding transgenes (*UAS-Parkin-275W* and *UAS-α-Syn-A53T*, respectively), applying the UAS-Gal4 system (Duffy, 2002). Transgenes were driven by *Ddc-Gal4* inducing the expression of the correspondent human mutant proteins in dopaminergic and serotoninergic fly neurons. Parkin (275W) and α-Syn (A53T) flies were compared to control animals from the same genetic background without mutant transgenes (*Ddc-Gal4* females were crossed with *isogenic* w^{1118} males; the examined F1 generation flies are referred to as *iso* w^{1118}; Fig. 3) or mutants overexpressing GFP (Fig. S2). The same level of eye pigmentation and vision of the compared genotypes were achieved by the prior replacement of the *w** mutant X chromosome of the applied *Ddc-Gal4* stock for that of the wild-type. We also used Mi{MIC} random insertion lines for dopamine receptor mutants, namely y^1 *w*; Mi{MIC}Dop1R1*^{MI04437} (BDSC 43773), y^1 *w*; Mi{MIC}Dop1R2*^{MI08664} (BDSC 51098) (Pimentel et al., 2016; Harbison et al., 2019), and w^{1118}*; PBac{PB}DopEcR*^{c02142}*/TM6B,* Tb^1 (BDSC 10847) (Ishimoto et al., 2013; Petruccelli et al., 2016, 2020), referred to as Dop1R1, Dop1R2 and DopEcR, respectively. The dopamine receptor mutant groups were

compared to their parental strains without the mutations (y^1 w^{67c23} served as control for *Dop1R1* and *Dop1R2* and w^{1118} for *DopEcR*; Fig. 3).

To test whether mutant flies showed differences in overall locomotion independent of the threatening stimuli, we analyzed average fly speed in the 200 ms time windows before stimulus presentation. We found that Parkin flies showed reduced mean speed compared to controls which expressed the Gal4 driver alone (24.96% reduction, $p = 6.55 \times 10^{-6}$, Mann-Whitney U-test; Fig. 3a, top), while α-Syn flies did not show significant reduction (p = 0.799, Mann–Whitney U-test). We observed similar changes in dopamine receptor mutant flies, where the *Dop1R1* and *DopEcR* defective lines showed a robust mean speed decrease compared to the control groups (*Dop1R1*, 19.68% decrease compared to y^1 w^{67c2}, p = 0.0016; *DopEcR*, 32.97% decrease compared to w^{1118}, p = 0.0034; Fig. 3a, bottom), while the *Dop1R2* mutant flies only showed a non-significant speed decrease (21.73% compared to y^1 w^{67c23}, p = 0.1009, Mann–Whitney U-test).

Next, we tested whether mutant flies showed a difference in their reaction to threatening stimuli. We found that PD model flies showed line-specific alterations in their freezing behavior. Parkin flies froze after stimuli less frequently, stopping in 37.72% of trials compared to the 43.48% observed in the *iso* w^{1118} animals (p = 0.0043, Mann Whitney U-test; Fig. 3b, top). However, in the stop trials, they showed normal duration of pauses in locomotion (p = 0.8018 compared to *iso* w^{1118}, Mann Whitney U-test; Fig. 3c, top). In contrast, α-Syn flies showed an unchanged stopping frequency when compared to controls (p = 0.0768 compared to *iso* w^{1118}, Mann-Whitney U-test), but their stop durations showed a large increase (116.98% increase compared to *iso* w^{1118}, $p = 2.18 \times 10^{-11}$, Mann-Whitney U-test). Interestingly, in contrast to PD model flies, dopamine receptor mutant flies did not show significant differences in their freezing behavior relative to controls (stop proportion: *Dop1R1*, p = 0.0631 compared to y^1 w^{67c23}; *Dop1R2*, p = 0.1838 compared to y^1 w^{67c23}; *DopEcR*, p = 0.1445 compared to w^{1118}; Mann-Whitney U-test; Fig. 3b, bottom; stop duration: *Dop1R1*, p = 0.893 compared to y^1 w^{67c23}; *Dop1R2*, p = 0.154 compared to y^1 w^{67c23}; *DopEcR*, p = 0.3576 compared to w^{1118}; Mann-Whitney U-test; Fig. 3c, bottom).

α-Syn flies showed a reduced aptitude to increase their speed, or 'run', upon encountering threatening stimuli (26.67% decrease compared to *iso* w^{1118}, $p = 8.4 \times 10^{-6}$, Mann-Whitney U-test; Fig. 3d, top). Similar results were found in *Dop1R1* (25% decrease compared to y^1 w^{67c23}, p = 0.0122; Mann-Whitney U-test; Fig. 3d, bottom) and *Dop1R2* (41.67% decrease compared to y^1 w^{67c23}, p = 0.0024, Mann-Whitney U-test), but not in *DopEcR* mutant flies (compared to w^{1118}, p = 0.6598, Mann-Whitney U-test). Frequency of slowing, i.e., reducing their speed without freezing in a full stop, was moderately decreased in Parkin (14.38%

decrease compared to *iso* w^{1118}, p = 0.0723, Mann-Whitney U-test; Fig. 3e, top) and *DopEcR* mutant flies (15.28% decrease compared to *iso* w^{1118}, p = 0.020, Mann-Whitney U-test; Fig. 3e, bottom).

Overall, all mutations tested resulted in decreased reactivity to threatening stimuli, confirmed by a significant increase in the proportion of trials where no reactions were detected (Fig. 3f). This effect was significant for Parkin flies (30% increase compared to *iso* w^{1118}, p = 0.0043, Mann-Whitney U-test), as well as for *Dop1R1* (50% increase compared to y^1 w^{67c23}, p = 0.0173, Mann-Whitney U-test) and *DopEcR* mutant flies (50% increase compared to *iso* w^{1118}, p = 0.0167, Mann-Whitney U-test), but not for α-Syn (α-Syn vs. *iso* w^{1118}, p = 0.151, Mann-Whitney U-test) and *Dop1R2* mutant flies (*Dop1R2* vs. y^1 w^{67c23}, p = 0.1903, Mann-Whitney U-test).

While *Ddc-Gal4* lines are widely used (Wang et al., 2007), they express mutations both in dopaminergic and serotonergic neurons, preventing the assessment of whether the behavioral differences were due to one of those cell types alone. To address this, we tested *NP6510-Gal4-R275W* (Parkin) and *NP6510-Gal4-A53T* (α-Syn) as well as *TH-Gal4-R275W* (Parkin) and *TH-Gal4-A53T* (α-Syn) flies where expression of the mutation was restricted to dopaminergic cells only (Riemensperger et al., 2013). Results from these tests were largely similar to those of the *Ddc-Gal4* lines (Fig. S3), reproducing the decreased speed (*NP6510-Gal4-R275W*, 30.3% decrease compared to *NP6510-Gal4- iso* w^{1118}, p = 1.55 x 10^{-6}; *NP6510-Gal4-A53T*, 20.4% decrease compared to *NP6510-Gal4- iso* w^{1118}, p = 5.82 x 10^{-10}; *TH-Gal4-R275W*, 13.08% decrease compared to *TH-Gal4- iso* w^{1118}, p = 1.49 x 10^{-6}; *TH-Gal4-A53T*, 17.57% decrease compared to *TH-Gl4- iso* w^{1118}, p = 6.75 x 10^{-7}; Mann–Whitney U-test) and decreased overall reactivity (*NP6510-Gal4-R275W*, 57.14% increase in 'No reaction' compared to *NP6510-Gal4- iso* w^{1118}, p = 3.49^{-8}; *NP6510-Gal4-A53T*, 14.28% increase in 'No reaction' compared to *NP6510-Gal4- iso* w^{1118}, p = 0.1658; *TH-Gal4-R275W*, 66.7% increase in 'No reaction' compared to *TH-Gal4- iso* w^{1118}, p = 2.77 x 10^{-10}; *TH-Gal4-A53T*, 66.7% increase in 'No reaction' compared to *TH-Gal4- iso* w^{1118}, p = 1.11 x 10^{-6}; Mann–Whitney U-test) of PD-model flies. Nevertheless, *TH-Gal4* and *NP6510-Gal4* mutants showed an increased propensity to stop (*NP6510-Gal4-R275W*, 16.47% increase compared to *NP6510-Gal4- iso* w^{1118}, p = 4.4 x 10^{-11}; *NP6510-Gal4-A53T,* 20.17% increase compared to *NP6510-Gal4- iso* w^{1118}, p = 1.38 x 10^{-17}; *TH-Gal4-R275W*, 16.32% increase compared to *TH-Gal4- iso* w^{1118}, p = 1.55 x 10^{-4}; *TH-Gal4-A53T*, 22.41% increase compared to *TH-Gal4- iso* w^{1118}, p = 8.82 x 10^{-10}, Mann–Whitney U-test). Stop duration showed a significant increase not only in α-Syn but also in Parkin fruit flies (*NP6510-Gal4-R275W*, 42.6% increase compared to *NP6510-Gal4- iso* w^{1118}, p = 5.99^{-5}; *NP6510-Gal4-A53T*, 69.5% increase compared to *NP6510-Gal4- iso* w^{1118}, p = 4.33 x 10^{-11}; *TH-Gal4-R275W*, 33.62% increase compared to *TH-Gal4- iso* w^{1118}, p = 1.63 x 10^{-6}; *TH-Gal4-A53T*, 37.48% increase compared

211 to *TH-Gal4- iso w*^{1118}, p = 3.89 x 10^{-7}; Mann–Whitney U-test). Overall, this suggests that mutations in the
212 dopaminergic neurons alone were sufficient to produce the above-described phenotypical changes.

213 After these detailed comparisons of central tendencies of behavioral data, we also tested whether
214 behavioral variability within fly response types showed differences across mutant groups. We found that
215 stop duration showed a significantly larger dispersion in α-Syn flies compared to controls (p = 0.0001,
216 permutation test; Fig. S4), while no other significant difference in data variance was detected.

217

218 **Reaction to threatening stimuli depends on fly walking speed**

219 It has been shown that fruit flies may exhibit a different choice of escape behavior based on their
220 momentary speed when encountering the threat (Zacarias et al., 2018). Therefore, we tested whether
221 mutations in genes relevant to PD caused a change in this speed - behavioral response relationship. We
222 calculated the probability of stopping, slowing, speeding up and no reaction as a function of speed for all
223 mutants, as well as the ratio of each response type conditioned on walking speed at the time of shadow
224 presentations (Fig. 4a-b). These analyses confirmed that 'running' and 'no reaction' was most likely at
225 slow (< 5 mm/s) or zero walking speeds, while slowing down was more frequent as the walking speed
226 increased. Freezing was most frequently observed in the 5-13 mm/s speed range. We did not observe
227 significant difference between groups in their speed - behavioral response relationship for any of the
228 response types (stops, p = 0.4226; slow down, p = 0.6025; speed up, p = 0.9074; no reaction p = 0.4692;
229 two-way ANOVA genotype × speed interaction).

230 Since the distribution of the expression of various escape behaviors depended on walking speed, which
231 was also different across the mutant lines tested (Fig. 3), we asked whether the difference in baseline
232 speed could explain the observed differences in behavioral reactions. Therefore, we performed
233 simulations where reaction probabilities were based on the baseline walking speed distribution of each
234 mutant line to test whether behavioral responses could be predicted based on speed alone (Fig. S5). In
235 most of the groups we found a significant difference between the predicted and the measured response
236 type distributions, suggesting a role for other behavioral differences among the mutant lines beyond the
237 baseline speed differences (Parkin, p = 0.0487; α-Syn, p < 0.000001; *Dop1R1*, p = 0.332; *Dop1R2*, p =
238 0.006157; *DopEcR*, p = 0.003901, chi-square test).

239

240 **Changes in escape behavior from the first to the fourth week of life**

241 The age of *Drosophilae* may have a significant influence on their responses to threatening stimuli. To test
242 this, we examined control and dopamine receptor mutant flies (w^{1118}, y^{1} w^{67c23}, *Dop1R1*, *Dop1R2*, *DopEcR*)
243 in 5 age groups: from 1-day-old to 4-week-old (Fig. 5; Fig. S6). The animals were kept under conditions
244 that accelerated their aging (29°C and 70% humidity). Our data showed that the behavior of 1-day-old
245 flies was significantly different from all other age groups. One-day-old flies stopped significantly more
246 often, and the frequency of stopping gradually decreased with age in all groups (two-way ANOVA, age, p
247 $= 1.87 \times 10^{-20}$, $f = 26.11$; genotype, $p = 1.3 \times 10^{-29}$, $f = 38.42$; genotype × age, $p = 3.18 \times 10^{-6}$, $f = 3.54$; 1-
248 day-old vs. 1-week-old, $p = 1.1 \times 10^{-4}$; 1-week-old vs. 2-week-old, $p = 0.0074$, 2-week-old vs. 3-week-old,
249 $p = 0.1015$; 3-week-old vs. 4-week-old, $p = 0.0079$, Mann-Whitney test for post hoc analysis). In contrast,
250 the probability of slowing down showed a substantial increase from 1-day-old to 1-week-old animals, and
251 then remained relatively stable until the 4th week (two-way ANOVA, age, $p = 9.11 \times 10^{-15}$, $f = 18.72$;
252 genotype, $p = 1.4 \times 10^{-21}$, $f = 27.59$; genotype × age, $p = 0.0028$, $f = 2.291$; 1-day-old vs. 1-week-old, $p = 5.9$
253 $\times 10^{-10}$; 1-week-old vs. 2-week-old, $p = 0.992$; 2-week-old vs. 3-week-old, $p = 0.0305$; 3-week-old vs. 4-
254 week-old, $p = 0.033$; Mann-Whitney test for post hoc analysis). The probability of speeding up showed a
255 similar trend, except that the 4-week-old animals showed a significant decrease compared to 3-week-old
256 flies (two-way ANOVA, age, $p = 2.48 \times 10^{-15}$, $f = 19.44$; genotype, $p = 2.28 \times 10^{-44}$, $f = 59.48$; genotype × age,
257 $p = 1.1 \times 10^{-7}$, $f = 4.12$; 1-day-old vs 1-week-old, $p = 2.64 \times 10^{-10}$; 1-week-old vs. 2-week-old, $p = 0.572$; 2-
258 week-old vs. 3-week-old, $p = 0.3445$; 3-week-old vs. 4-week-old, $p = 0.0179$; Mann-Whitney test for post
259 hoc analysis). Thus, different types of escape reactions showed remarkable tendencies across the fly
260 lifespan, while dopamine receptor mutant flies maintained their decreased general reactivity throughout
261 all age groups.

262

263 **Discussion**

264 We tested *Drosophila* mutant strains relevant for PD in a single-trial single-animal behavioral assay. Our
265 tests revealed strain-specific behavioral alterations in flies' reactions to predator-mimicking passing
266 shadows, serving as proof of principle demonstration of the viability of single-trial approaches in
267 *Drosophila*, a method widely used in mammalian studies. Specifically, we found reduced walking speed,
268 decreased freezing frequency and decreased overall reactivity in Parkin flies. In contrast, α-Syn flies
269 merely showed an increased freezing duration without a concomitant change in freezing frequency,

suggesting that Parkin flies better recapitulate some behavioral features of human PD progression. *Dop1R* mutant flies resembled Parkin flies in their decreased walking speed and decreased reactivity to the threatening stimuli. The distribution of the behavioral response fruit flies chose to execute depended on their speed at the time of stimuli, and this speed - behavioral response relationship was robust across the tested genotypes. Nevertheless, differences in walking speed alone did not explain the strain-specific behavioral alterations. Age-dependence of behavioral choice was also found to be stereotypic across the tested genotypes.

Recent studies increasingly recognize the importance of investigating subtle changes in fly behavior to better understand the manifestations of locomotor and other disorders (Geissmann et al., 2017; Zacarias et al., 2018; Seidenbecher et al., 2020). Along these lines, Aggarwal and colleagues introduced an automated climbing assay for fruit flies and revealed subtle motor deficits in different mutant strains (Aggarwal et al., 2019). We expanded the scope of these studies by examining a diverse behavioral repertoire of *Drosophila* in response to threatening stimuli. Many earlier studies used looming stimulus, that is, a concentrically expanding shadow, mimicking the approach of a predator from above, to study escape responses in flies (Card and Dickinson, 2008; de Vries and Clandinin, 2012; Zacarias et al., 2018; Ache et al., 2019; Oram and Card, 2022) as well as rodents (Lecca et al., 2017; Braine and Georges, 2023; Heinemans and Moita, 2024). These assays have the advantage of closely resembling naturalistic, ecologically relevant threat-inducing stimuli, and allow a relatively complete characterization of the fly escape behavior repertoire (Zacarias et al., 2018). As a flip side of their large degree of freedom, they do not lend themselves easily to provide a fully standardized, scalable behavioral assay. Therefore, Gibson et al. suggested a novel threat-inducing assay operating with moving overhead translational stimuli, that is, passing shadows, and demonstrated that they induce escape behaviors in flies akin to looming discs (Gibson et al., 2015). This assay, coined ReVSA (repetitive visual stimulus-induced arousal) by the authors, had the advantage of scalability, while constraining flies to a walking arena that somewhat restricted the remarkably rich escape types flies otherwise exhibit. Here we carried this idea one step further by using a screen to present the shadows instead of a physically moving paddle and putting individual flies to linear corridors instead of the common circular fly arena. This ensured that the shadow reached the same coordinates in all linear tracks concurrently and made it easy to accurately determine when individual flies encountered the stimulus, aiding data analysis and scalability. We found the same escape behavioral repertoire as in studies with looming stimuli and ReVSA (Gibson et al., 2015; Zacarias et al., 2018), with a similar dependence on walking speed (Zacarias et al., 2018; Oram and Card, 2022), confirming that looming stimuli and passing shadows can both be considered as threat-inducing visual stimuli.

302

303 Mutations in *PARK2* gene lead to impaired ubiquitination and a consequential mitochondrial dysfunction
304 (Guo, 2010, 2012), and early-onset PD in human patients. This led to the widespread use of Parkin flies as
305 a genetic model of PD (Guo, 2010; Chambers et al., 2013; Aggarwal et al., 2019). Of note, flies that lack
306 Parkin display a significant degeneration of dopaminergic neurons (Whitworth et al., 2005). We found
307 similarities between Parkin and *Dop1R* mutant flies in their altered responses to predator-mimicking
308 passing shadows: reduced walking speed and decreased overall reactivity, suggesting that the lack of
309 dopamine action through *Dop1R* may be one of the common pathways underlying motor deficits. This is
310 in line with a recent study demonstrating impaired startle-induced geotaxis and locomotor reactivity in
311 *Dop1R* mutant flies (Sun et al., 2018), similar to what had been shown for Parkin flies (Aggarwal et al.,
312 2019), and a demonstration of dopaminergic control over walking speed (Marquis and Wilson, 2022). In
313 contrast, flies expressing human α-syn showed moderate changes in motor behavior except for a marked
314 prolongation of freezing duration upon threating stimuli. This is in accordance with studies suggesting that
315 α-syn misexpression is not fully penetrant under some conditions (Pesah et al., 2005; Nagoshi, 2018), and
316 calls for the use of other genetic *Drosophila* models, including Parkin flies, in studying the pathophysics of
317 human PD.

318 The dopamine/ecdysteroid receptor *DopEcR* is a G-protein-coupled dual receptor for dopamine and the
319 steroid hormone ecdysone. It has been proposed that the *DopEcR* may serve as an integrative hub for
320 dopamine-mediated actions and stress responses in fruit flies (Petruccelli et al., 2020). We found that
321 *DopEcR* mutant flies showed decreased mean walking speed, decreased probability of slowing down and
322 decreased overall behavioral reactivity in response to predator-mimicking passing shadows. This pattern
323 of alterations was largely similar to those observed in Parkin and *Dop1R* mutant flies, suggesting that the
324 *DopEcR* may convey similar dopamine-mediated functions as *DopR1* at least in those motor aspects tested
325 in the present study. A recent work demonstrated that serotonin also modulates both walking speed and
326 startle response in flies, suggesting a complex neuromodulatory control over *Drosophila* motor behavior
327 (Howard et al., 2019). We recognize a limitation in using PBac{PB}DopEcRc02142 over the TM6B, Tb1
328 balancer chromosome, as the balancer itself may induce behavioral deficits in flies. We consider this
329 unlikely, as the PBac{PB}DopEcRc02142 mutation demonstrates behavioral effects even in heterozygotes
330 (Ishimoto et al., 2013). Additionally, to our knowledge, no studies have reported behavioral deficits in flies
331 carrying the TM6B, Tb1 balancer chromosome over a wild-type chromosome.

In humans, degeneration of midbrain dopaminergic neurons has been found as a direct cause of PD symptoms and DA supplementation with its precursor levodopa has been one of the most successful therapeutic approaches to date. The fruit fly and mammalian DA systems show a number of homologies which are thought to serve as a good basis for *Drosophila* PD models (Feany and Bender, 2000; Hewitt and Whitworth, 2017). Specifically, a deep-running evolutional conservation has been revealed between the arthropod central complex and the vertebrate basal ganglia, where GABAergic and dopaminergic neurons play key roles in motor control in both phylae (Strausfeld and Hirth, 2013). Clusters of fly dopaminergic neurons that project to the ellipsoid body, the fan-shaped body and the lateral accessory lobes are thought to share homologies with the striatum-projecting dopaminergic neurons of the substantia nigra in mammals. Both fruit fly central complex and vertebrate basal ganglia mediate a range of functions from motor control and sensorimotor integration to action selection and decision making to motivation (Strauss, 2002; Claridge-Chang et al., 2009; Kong et al., 2010) and learning (McCurdy et al., 2021; Zolin et al., 2021; Fisher et al., 2022; Qiao et al., 2022; Taisz and Jefferis, 2022; Yamada et al., 2023). It has been found that fruit flies lacking the DA-synthetizing tyrosine hydroxylase enzyme in the central nervous system show reduced activity and locomotor deficit that worsen with age, and also exhibit marked impairments in associative learning based on both appetitive and aversive reinforcement (Riemensperger et al., 2011). This is in line with mammalian studies emphasizing the role of the dopaminergic nigrostriatal pathway in controlling goal-directed action (Eban-Rothschild et al., 2016), reinforcement learning (Schultz et al., 1997; Lak et al., 2014), and more recently, conveying aversive information (Menegas et al., 2015, 2018). Our findings are consistent with the apparent homologies across fruit fly and mammalian dopaminergic systems and suggest that a more in-depth investigation of specific behavioral changes and underlying dopamine-related dysfunctions may yield translatable results. We also observed age-related changes in motor behavior in response to threatening stimuli, particularly in 4-week-old flies, which may parallel the age-dependent worsening of PD symptoms in humans.

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Data and code availability

All analysis functions are available at https://github.com/hangyabalazs/Drosophila-behavior-analysis. All data are available at https://figshare.com/s/40539ee040a269bf05e5.

Methods

Animals

We used 2-weeks-old *Drosophila melanogaster* (both males and females) raised at 24°C and 60% humidity in a natural light-dark cycle for experiments presented in Figures 1-4. For age-group comparisons (Fig.5.), fruit flies aged 1, 7, 14, 21 and 28 days were used. These flies were raised at 29°C and 70% humidity to accelerate aging.

Three sets of mutant groups were used for behavioral comparison. In the first set of animals we used *w*; UAS-Parkin-R275W* (created in the laboratory of Kah-Leong Lim, Neurodegeneration Research Laboratory, National Neuroscience Institute, Singapore)(Wang et al., 2007; Kovács et al., 2017) and *w*; UAS-alpha-Syn-A53T* (BDSC 8148) from the Eotvos Lorand University (Szinyákovics et al., 2023), with y^1, v^1; *UAS-GFP* (BDSC 35786) and *+; P{Ddc-GAL4.L}4.36* (modified version of BDSC 7009, the original first chromosome w^{1118} has been replaced to w^+ in this study) as controls. For the second set of animals we used *Dop1R1*, *Dop1R2* and *DopEcR* mutant flies kindly donated by the Anne Von Phillipsborn lab, DANDRITE, Aarhus University and y^1 *w*; Mi{MIC}Dop1R1*^{MI04437} (BDSC 43773), y^1 *w*; Mi{MIC}Dop1R2*^{MI08664} (BDSC 51098), and w^{1118}*; PBac{PB}DopEcR*^{c02142}*/TM6B, Tb*^{1} (BDSC 10847) with w^{1118} (BDSC 5905) y^1 and y^1 w^{67c23} (BDSC 6599) as controls from the Bloomington stock center. Table 1. shows the number of recorded flies for the age group comparisons, and Table 2. shows the number of recorded flies for comparing the mutant groups. For the third set of flies we used w[*]; P{w[+mC]=ple-GAL4.F}3 (*TH-Gal4*), Bloomington Drosophila

Stock Center: 8848 (Friggi-Grelin et al., 2003), and y[*] w[*]; P{w[+mW.hs]=GawB}NP6510 (*NP6510-Gal4*), Kyoto DGGR: 113956 (Riemensperger et al., 2013; Majcin Dorcikova et al., 2023). Since it was important to ensure that the animals' light perception was not impaired, we replaced the white loss-of-function mutant allele of the transgenic *Gal4* lines with a wild-type allele. As controls, we used transheterozygous +/ *TH-Gal4* and +/ *NP6510-Gal4* strains in all cases.

	Dop1R1	*Dop1R2*	*DopEcR*	w^{1118}	y^{1} w^{67c23}
1-day-old	35	36	36	35	41
1-week-old	36	54	36	36	36
2-week-old	36	36	36	36	30
3-week-old	35	35	29	36	36
4-week-old	30	36	34	30	30

Table 1. Number of recorded flies for age group comparisons.

2-week-old	No. of flies
Dop1R1	45
Dop1R2	47
DopEcR	35
w^{1118}	46
y^{1}, w^{67c23}	40
Parkin (Ddc)	78
α-Syn (Ddc)	69
iso w^{1118}	92
+GFP	65
Parkin (NP6510)	170
α-Syn (NP6510)_	181
+/ NP6510-Gal4	176
Parkin (TH)	322
α-Syn (TH)	176
+/ TH-Gal4	282

Table 2. Number of recorded flies for comparing the mutant groups.

Arena

We designed an arena made of four layers of plexiglass and metal that formed 13 tunnels (Fig. 1a). Each tunnel was 52mm x 5mm × 1.5mm, in which individual flies were able to move freely in two dimensions but could not fly. The bottom layer was a standard transparent plexiglass layer. The second layer contained the tunnel walls made of metal to prevent horizontal spread of light. The third layer was the top of the tunnels, made of transparent plexiglass, which could slide above the tunnel. It also contained a

small hole on the side, where the flies could be inserted into the tunnel through a pipette. The fourth layer was a metal cover designed with cut-outs that matched the shape and position of the tunnels to prevent flies in the neighboring tunnels from detecting the shadow before it reached their position.

Movement tracking

Flies were tracked by using a FLIR Camera (FLIR Blackfly S USB3 FLIR Systems, Wilsonville, OR, US) placed under the arena (Fig. 1a). The frame rate was set to 100 frames/second to allow the detection of position with high temporal resolution. The recorded images were processed by Bonsai (2.5.2) tracking software (Lopes et al., 2015), using customized Bonsai code. The code extracts each tunnel area separately from the image and detects flies by selecting the biggest and darkest pixel object in the tunnel area in real time. Fly position coordinates were calculated from the centroid. The software stored x-y position coordinates along with their timestamps in csv files, which were later processed in Matlab R2016a (Mathworks, Natick, MA, US).

Predator-mimicking shadow stimuli

A 10.1-inch screen (HannStar HSD101PWW1-A00,) was placed on the top of the arena. The screen presented a yellow background. We implemented 'shadow' stimuli in red color, as these stimuli were suggested to be perceived by fruit flies as dark shadows and also enabled continuous tracking of the animals (Sharkey et al., 2020). The red color screen represented a large enough contrast change to evoke escape behaviors of flies but also provided sufficient brightness to enable continuous motion tracking. A passing shadow stimulus was chosen, as it was affecting all tunnels simultaneously. To mimic an approaching shadow, a red screen slid in from the side, stayed on for 2 seconds, then slid out. The shadow stimuli were separated by pseudorandomized inter-trial intervals with an approximately exponential distribution with a mean of 52.2 seconds, preventing flies from anticipating the next shadow stimulus.

Identification of behavioral response types to the threatening stimuli

Animal speed was calculated as a function of time in a -200 ms to 1000 ms window around each shadow stimulus in 10 ms time bins. These speed functions were normalized by subtracting the average speed in the 200 ms window before the shadow presentation to reveal stimulus induced absolute instantaneous speed changes. Principal component analysis (PCA) was used to reduce dimensionality of the normalized speed functions in the 1 s interval following shadow presentation in a way that the variance between trials is maximally preserved in the low-dimensional representation. Agglomerative hierarchical clustering was

performed in the space spanned by the first three principal components to identify trials with distinct characteristic response types.

Behavior detection

After identifying the four most frequently observed animal responses to shadow presentations, we algorithmically defined them based on the speed and acceleration thresholds. First, we examined the speed of the animal in the 200 ms time window before the shadow (v_{pre}); stopping/freezing or slowing down was only possible if the fly had been moving, defined by $v_{pre} > 0$ mm/s. Second, we examined the acceleration (a_{post}) in the 1 s interval following the shadow using a 100 ms moving average window to characterize the flies' response. Reactions were considered robust if the absolute value of the acceleration reached 200 mm/s^2. A trial was characterized as a 'stop' trial, if the average v_{pre} was above 0 mm/s, the fly's first reaction was deceleration, and 0 mm/s speed was reached before accelerating again. If the first reaction was deceleration but 0 mm/s speed was not reached, the trial was classified as a 'slow down' trial. In 'speed up' trials, the first response following the shadow was acceleration. If the absolute value of a_{post} did not reach the 200 mm/s^2 threshold during the 1 s interval following the shadow, the trial was labeled as a 'no reaction' trial.

Data analysis and statistics

Data analysis was performed in Matlab R2016a (Mathworks, Natick, MA, US) using custom-written code. Statistical comparison between groups of flies was performed by two-sided Mann-Whitney U-test. Exact p values are reported in the Results section. We performed multi-group comparison in the 'Changes in escape behavior from the first to the fourth week of life' section using two-way ANOVA, as it is reportedly robust to non-normality of the data (Mooi et al., 2018; Knief and Forstmeier, 2021), while there is a lack of a consensual non-parametric alternative. Nevertheless, Kruskal-Wallis tests for the main effect of age showed significant effects in all genotypes in accordance with the ANOVA, confirming the results (Stop frequency, *DopEcR* p = 0.0007; *Dop1R1*, p = 0.004; *Dop1R2*, p = 9.94×10^{-5}; w^{1118}, p = 9.89×10^{-13}; $y^1\ w^{67}c^{23}$, p = 2.54×10^{-5}; Slowing down frequency, *DopEcR*, p = 0.0421; *Dop1R1*, p = 5.77×10^{-6}; *Dop1R2*, p = 0.011; w^{1118}, p = 2.62×10^{-5}; $y^1\ w^{67c23}$, p = 0.0382; Speeding up frequency, *DopEcR*, p = 0.0003; *Dop1R1*, p = 2.06×10^{-7}; *Dop1R2*, p = 2.19×10^{-6}; w^{1118}, p = 0.0044; $y^1\ w^{67c23}$, p = 1.36×10^{-5}). We used Mann-Whitney U-test for post hoc analyses to maintain consistent use of tests across figures. Since there is no straightforward way of correcting for multiple comparisons in this case, we reported uncorrected p values and suggest considering them at p = 0.01, which minimizes type I errors. Nevertheless, Tukey's tests with the 'honest

significance' approach yielded similar results. The effect of movement speed on the distribution of behavioral response types was tested using a nested Monte Carlo simulation framework (Fig. S5). This simulation aimed to model how different movement speeds impact the probability distribution of response types, comparing these simulated outcomes to empirical data. This approach allowed us to determine whether observed differences in response distributions were solely due to speed variations across genotypes or if additional behavioral factors contributed to the differences. First, we calculated the probability of each response type at different specific speed values (outer model). These probabilities were derived from the grand average of all trials across each genotype, capturing the overall tendency at various speeds. Second, we simulated behavior of virtual flies (n = 3000 per genotypes, which falls within the same order of magnitude as the number of experimentally recorded trials from different genotypes) by drawing random velocity values from the empirical velocity distribution specific to the given genotype and then randomly selecting a reaction based on the reaction probabilities associated with the drawn velocity (inner model). Finally, we calculated reaction probabilities for the virtual flies and compared them with real data from animals of the same genotype. Differences were statistically tested by Chi-squared test.

Figure Legends

Fig. 1. Behavioral apparatus and experimental design.

a, Top left, schematic of the experimental setup. Top right, schematic of the fly arena. Bottom left, photograph of the experimental setup. Bottom right, a photograph of the fly arena. **b,** Timeline of a session showing the shadow presentations in red over the yellow background.

Fig. 2. Behavioral characterization of the escape behavior repertoire of individual fruit flies.

a, Schematic for calculating speed and acceleration based on tracked position coordinates. **b,** Four characteristic escape behaviors categorized by PCA for an example session of a control fly (w1118; from left; stop, slow down, speed up, no reaction). Top, color-coded heatmaps indicating the walking speed of the fly (blue, low speed; yellow, high speed), aligned to shadow presentations (purple line). Bottom, average moving speed triggered on the shadow presentations (purple line). **c,** Threshold-based classification of behavioral responses (from left, stop, slow down, speed up, no reaction). Top, single-trial example for each response type. Time intervals for calculating the average speed before the shadow presentation (purple) as well as the speed and acceleration after the shadow presentation (light green) are marked. Threshold values for each response type are displayed above the graphs. Middle, average

494 walking speed across the trials from all sessions of w^{1118} flies, sorted by the type of behavioral response.
495 Line and errorshade show mean ± SE. Bottom, scatter plots showing minimal (Stop and Slow down trials)
496 or average (Speed up and No reaction trials) post-stimulus speed vs. average pre-stimulus speed,
497 representing the same trials as above.

498 **Fig. 3. PD transgenic and dopamine receptor mutant fruit flies showed reduced walking speed and**
499 **decreased reactivity to threatening stimuli.**

500 **a**, Distribution of the mean speed measured in the time window [-0.2, 0] seconds relative to shadow
501 presentation for the different mutant groups. Top, Parkin flies showed reduced mean speed compared to
502 controls (Parkin vs. *iso* w^{1118}; $p = 6.55 \times 10^{-6}$, Mann–Whitney U-test). Bottom, *Dop1R1* and *DopEcR* mutant
503 flies showed reduced mean speed compared to controls (y^1w^{67c23} and w^{1118} respectively; p = 0.0016, p =
504 0.0034; Mann-Whitney U-test). **b,** Distribution of the proportion of stop trials in different mutant groups.
505 Parkin flies showed a reduced tendency for stopping compared to *iso* w^{1118} (p = 0.0043, Mann Whitney U
506 test). **c,** Distribution of stop duration in different mutant groups. α-Syn flies showed increased stop
507 durations compared to *iso* w^{1118} ($p = 2.18 \times 10^{-11}$, Mann-Whitney U-test). **d,** Distribution of the proportion
508 of speed up trials in different mutant groups. Top, α-Syn flies showed a reduced tendency to speed up
509 compared to their controls ($p = 8.4 \times 10^{-6}$, Mann-Whitney U-test). Bottom, *Dop1R1* and *Dop1R2* mutant
510 flies also showed a significantly reduced tendency to speed up compared to their controls (p = 0.0122 and
511 p = 0.0024, respectively; Mann-Whitney U-test). **e,** Distribution of the proportion of slow down trials in
512 different mutant groups. Bottom, *DopEcR* showed a 15.04% decrease compared to w^{1118} (p = 0.020, Mann-
513 Whitney U-test). **f,** Distribution of the proportion of 'no reaction' trials in different mutant groups. Top,
514 Parkin mutants showed reduced reactivity compared to *iso* w^{1118} controls (p = 0.0043, Mann-Whitney U-
515 test). Bottom, *Dop1R1* and *DopEcR* mutants also showed reduced reactivity compared to y^1w^{67c23} and w^{1118}
516 controls, respectively (p = 0.0173 and p = 0.0167, respectively; Mann-Whitney U-test). Box-whisker plots
517 show median, interquartile range and non-outlier range. *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$. Exact
518 genotypes: *iso* w^{1118}: *+; +; Ddc-Gal4/+.* Parkin: *+; +; Ddc-Gal4/UAS-Parkin-R275W*. α-Syn: *+; +; Ddc-*
519 *Gal4/UAS-α-Syn-A53T*. y^1 w^{67c23}: y^1 w^{67c23}. *Dop1R*: y^1 *w*; Mi{MIC}Dop1R1*^{MI04437}. *Dop1R2*: y^1 *w*;*
520 *Mi{MIC}Dop1R2*^{MI08664}. *DopEcR*: w^{1118}*; PBac{PB}DopEcR*^{c02142}*/TM6B, Tb*^{1}.

521 **Fig. 4. Reaction to threatening stimuli depends on fly walking speed**.

522 **a,** Probability of a given response type as a function of average fly speed before the shadow presentation
523 (200 ms pre-stimulus time window). From left, stop, slow down, speed up and no reaction trials are

quantified. Top, Parkin and α-Syn flies. Bottom, *Dop1R1*, *Dop1R2* and *DopEcR* mutant flies. **b,** Proportion of a given response type as a function of average fly speed before the shadow presentation. Top, Parkin and α-Syn flies. Bottom, *Dop1R1*, *Dop1R2* and *DopEcR* mutant flies. Exact genotypes: *iso* w^{1118}: *+; +; Ddc-Gal4/+.* Parkin: *+; +; Ddc-Gal4/UAS-Parkin-R275W*. α-Syn: *+; +; Ddc-Gal4/UAS-α-Syn-A53T*. y^1 w^{67c23}: y^1 w^{67c23}. *Dop1R*: y^1 *w*; Mi{MIC}Dop1R1*^{MI04437}. *Dop1R2*: y^1 *w*; Mi{MIC}Dop1R2*^{MI08664}. *DopEcR*: w^{1118}*; PBac{PB}DopEcR*^{c02142}*/TM6B,* Tb^1.

Fig. 5. Changes in escape behavior from the first to the fourth week of life

Proportion of responses as a function of age for different groups of mutants and controls. From left to right, stop, slow down, speed up and no reaction trials are quantified. Lines and errorshades show mean and standard error. Exact genotypes: *iso* w^{1118}: *+; +; Ddc-Gal4/+.* Parkin: *+; +; Ddc-Gal4/UAS-Parkin-R275W*. α-Syn: *+; +; Ddc-Gal4/UAS-α-Syn-A53T*. y^1 w^{67c23}: y^1 w^{67c23}. *Dop1R*: y^1 *w*; Mi{MIC}Dop1R1*^{MI04437}. *Dop1R2*: y^1 *w*; Mi{MIC}Dop1R2*^{MI08664}. *DopEcR*: w^{1118}*; PBac{PB}DopEcR*^{c02142}*/TM6B,* Tb^1.

Supplementary Figure Legends

Fig. S1. Picture of the arena from the camera view

The 13 tunnels of the arena from the camera view with the tracking lines showing that the animals can move in 2 dimensions. The tracking line color indicates time, with dark blue and red marking the beginning and the end of the tracking, respectively.

Fig. S2. Parkin and α-Syn fly group escape reaction compared to GFP mutant flies

a, Distribution of the mean speed measured in the 0.2 second before shadow presentation for Parkin, α-Syn and +GFP mutant groups. Parkin and α-Syn flies showed reduced mean speed compared to +GFP controls (Parkin vs. +GFP, $p = 1.58 \times 10^{-11}$; α-Syn vs. +GFP, $p = 2.85 \times 10^{-4}$; Mann-Whitney U-test). **b,** Distribution of the proportion of stop trials in different mutant groups. Parkin flies showed a reduced tendency for stopping compared to the +GFP group (Parkin vs. +GFP, $p = 7.1 \times 10^{-5}$; α-Syn vs. +GFP, p = 0.67; Mann-Whitney U-test). **c,** Distribution of stop duration in different mutant groups. α-Syn flies showed increased stop durations compared to GFP (Parkin vs. +GFP, p = 0.17; α-Syn vs. +GFP, $p = 3.98 \times 10^{-10}$; Mann-Whitney U-test). **d,** Distribution of the proportion of speed up trials in different mutant groups. Both Parkin and α-Syn flies showed a reduced tendency to speed up compared to +GFP controls (Parkin vs. +GFP, $p = 8.3 \times 10^{-5}$; α-Syn vs. +GFP, $p = 1.96 \times 10^{-12}$; Mann-Whitney U-test). **e**, Distribution of the proportion of slow down trials in different mutant groups. Parkin flies showed a reduced tendency to slow down compared to +GFP controls (Parkin vs. +GFP, p = 0.044; α-Syn vs. +GFP, p = 0.465; Mann-

554 Whitney U-test). **f**, Distribution of the proportion of 'no reaction' trials in different mutant groups. Both
555 Parkin and α-Syn flies showed reduced reactivity compared to +GFP controls (Parkin vs. +GFP, $p = 9.4 \times$
556 10^{-13}; α-Syn vs. +GFP, $p = 1{,}3 \times 10^{-8}$; Mann-Whitney U-test). *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$. Exact
557 genotypes: *iso* w^{1118}: *+; +; Ddc-Gal4/+.* Parkin: *+; +; Ddc-Gal4/UAS-Parkin-R275W*. α-Syn: *+; +; Ddc-*
558 *Gal4/UAS-α-Syn-A53T*.

559 **Fig. S3. Restricting Parkin and α-Syn mutations to dopaminergic neurons**

560 **a,** Distribution of the mean speed measured in the 0.2 second before shadow presentation for Parkin
561 (R275W), α-Syn (A53T) and control (*iso* w^{1118}) groups. Top, *NP6510-Gal4* lines; bottom, *TH-Gal4* lines in all
562 panels. *NP6510-Gal4-R275W*, 30.3% decrease compared to *NP6510-Gal4-iso* w^{1118}, $p = 1.55 \times 10^{-6}$;
563 *NP6510-Gal4-A53T*, 20.4% decrease compared to *NP6510-Gal4-iso* w^{1118}, $p = 5.82 \times 10^{-10}$; *TH-Gal4-R275W*,
564 13.08% decrease compared to *TH-Gal4-iso* w^{1118}, $p = 1.49 \times 10^{-6}$; *TH-Gal4-A53T*, 17.57% decrease
565 compared to *TH-Gl4-iso* w^{1118}, $p = 6.75 \times 10^{-7}$; Mann–Whitney U-test. **b,** Distribution of the proportion of
566 stop trials in different mutant groups. *NP6510-Gal4-R275W*, 16.47% increase compared to *NP6510-Gal4-*
567 *iso* w^{1118}, $p = 4.4 \times 10^{-11}$; *NP6510-Gal4-A53T,* 20.17% increase compared to *NP6510-Gal4-iso* w^{1118}, $p = 1.38$
568 $\times 10^{-17}$; *TH-Gal4-R275W*, 16.32% increase compared to *TH-Gal4-iso* w^{1118}, $p = 1.55 \times 10^{-4}$; *TH-Gal4-A53T*,
569 22.41% increase compared to *TH-Gal4-iso* w^{1118}, $p = 8.82 \times 10^{-10}$, Mann–Whitney U-test. **c,** Distribution of
570 stop duration in different mutant groups. *NP6510-Gal4-R275W*, 42.6% increase compared to *NP6510-*
571 *Gal4-iso* w^{1118}, $p = 5.99^{-5}$; *NP6510-Gal4-A53T*, 69.5% increase compared to *NP6510-Gal4-iso* w^{1118}, $p = 4.33$
572 $\times 10^{-11}$; *TH-Gal4-R275W*, 33.62% increase compared to *TH-Gal4-iso* w^{1118}, $p = 1.63 \times 10^{-6}$; *TH-Gal4-A53T*,
573 37.48% increase compared to *TH-Gal4-iso* w^{1118}, $p = 3.89 \times 10^{-7}$; Mann–Whitney U-test. **d,** Distribution of
574 the proportion of speed up trials in different mutant groups. *NP6510-Gal4-R275W*, 9.1% decrease
575 compared to *NP6510-Gal4-iso* w^{1118}, $p = 0.0052$; *NP6510-Gal4-A53T*, 9.1% decrease compared to *NP6510-*
576 *Gal4-iso* w^{1118}, $p = 3.14 \times 10^{-4}$; *TH-Gal4-R275W*, 13.3% decrease compared to *TH-Gal4-iso* w^{1118}, $p = 1.25 \times$
577 10^{-9}; *TH-Gal4-A53T*, 20% decrease compared to *TH-Gal4-iso* w^{1118}, $p = 1.33 \times 10^{-15}$; Mann–Whitney U-test.
578 **e,** Distribution of the proportion of slow down trials in different mutant groups. *NP6510-Gal4-R275W*,
579 22.2% decrease compared to *NP6510-Gal4-iso* w^{1118}, $p = 8.23^{-5}$; *NP6510-Gal4-A53T*, 39.1% decrease
580 *NP6510-Gal4-iso* w^{1118}, $p = 2.42 \times 10^{-10}$; *TH-Gal4-R275W*, 22.8% decrease compared to *TH-Gal4-iso* w^{1118},
581 $p = 2.85 \times 10^{-8}$; *TH-Gal4-A53T*, 31.49% decrease compared to *TH-Gal4-iso* w^{1118}, $p = 1.95 \times 10^{-11}$; Mann–
582 Whitney U-test. **f,** Distribution of the proportion of 'no reaction' trials in different mutant groups. *NP6510-*
583 *Gal4-R275W*, 57.14% increase compared to *NP6510-Gal4-iso* w^{1118}, $p = 3.49^{-8}$; *NP6510-Gal4-A53T*, 14.28%
584 increase compared to *NP6510-Gal4-iso* w^{1118}, $p = 0.1658$; *TH-Gal4-R275W*, 66.7% increase compared to

585 *TH-Gal4-iso* w^{1118}, p = 2.77 x 10^{-10}; *TH-Gal4-A53T*, 66.7% increase compared to *TH-Gal4-iso* w^{1118}, p = 1.11
586 x 10^{-6}; Mann–Whitney U-test.

587 **Fig. S4. Stop duration showed large variance in α-Syn flies**

588 **a,** Distributions of stop duration in *iso* w^{1118} controls, Parkin and α-Syn flies. **b,** Permutation tests revealed
589 a significantly increased variance in α-Syn compared to control flies (p = 0.0001). Histograms show
590 shuffled variance difference between Parkin (left) or α-Syn (right) and control groups. Observed difference
591 in variance is indicated by a blue vertical line. The critical value corresponding to p = 0.05 is indicated by
592 a green dashed vertical line.

593 **Fig. S5. Differences in walking speed do not explain different escape behaviors**

594 Real and simulated proportion of response types are shown for the mutant groups tested. For the
595 simulations, random velocity values were drawn from the velocity distribution of each genotype and then
596 a response type was randomly selected based on the response type distributions associated with the
597 drawn velocity. Bar plots show the proportion of response types color-coded. For each mutant group, the
598 right bar shows the measured proportion of escape responses, while the left bar shows the simulated
599 distribution. We found significant differences between the real and simulated distributions for the
600 following groups: Parkin (p = 0.0487), α-Syn (p < 0.000001), *Dop1R2* (p = 0.006157) and *DopEcR* (p =
601 0.003901). We found no significant differences for the *Dop1R1* group (p = 0.332; Chi-square test for all
602 the statistics). *, p < 0.05; **, p < 0.01; ***, p < 0.001. Exact genotypes: *iso* w^{1118}: *+; +; Ddc-Gal4/+.* Parkin:
603 *+; +; Ddc-Gal4/UAS-Parkin-R275W*. α-Syn: *+; +; Ddc-Gal4/UAS-α-Syn-A53T*. y^1 w^{67c23}: y^1 w^{67c23}. *Dop1R*: y^1
604 *w*; Mi{MIC}Dop1R1*^{MI04437}. *Dop1R2*: y^1 *w*; Mi{MIC}Dop1R2*^{MI08664}. *DopEcR*: w^{1118};
605 *PBac{PB}DopEcR*^{c02142}*/TM6B,* Tb^1.

606 **Fig. S6. Changes in escape behavior from the first to the fourth week of life**

607 **a,** Average walking speed as a function of age for different groups of controls (left) and mutants (right).
608 Lines and errorshades show mean and standard error. Mean speed showed significant differences among
609 age groups (two-way ANOVA, age, f = 41.38, p = 9.39 x 10^{-32}; genotype, f = 42.19, p = 2.46 x 10^{-32}; genotype
610 × age, f = 4.36, p = 2.667 x 10^{-8};Tukey's post hoc test: 1 day old vs. 1 week old, p = 9.92 x 10^{-9}; 1 week old
611 vs. 2 weeks old, p = 7.34 x 10^{-7}; 2 weeks old vs. 3 weeks old, p = 0.085; 3 weeks old vs. 4 weeks old, p =
612 0.0043). **b,** Stop duration as a function of age for different groups of controls (left) and mutants (right).
613 We found that age did not affect the duration of stops (two-way ANOVA, age, f = 0.85, p = 0.49; genotype,
614 f = 0.91, p = 0.46; genotype × age, f = 2.53, p = 0.0008; Tukey's post hoc test: 1 day old vs. 1 week old, p =

0.98; 1 week old vs. 2 weeks old, p = 0.992; 2 weeks old vs. 3 weeks old, p = 0.578; 3 weeks old vs. 4 weeks old, p = 0.9759). Lines and errorshades show mean and standard error.

Supplementary Movie 1

Example recording of *iso* w^{1118} files in the arena. Flies could walk freely inside the tunnels, speed up, slow down, stop, turn, climb the side walls, flip, or groom their wings.

Supplementary Movie 2

Example recording of Parkin flies in the arena. Similar behaviors can be observed as in *iso* w^{1118} flies (see Supplementary Movie 1).

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Figures

Figure 1

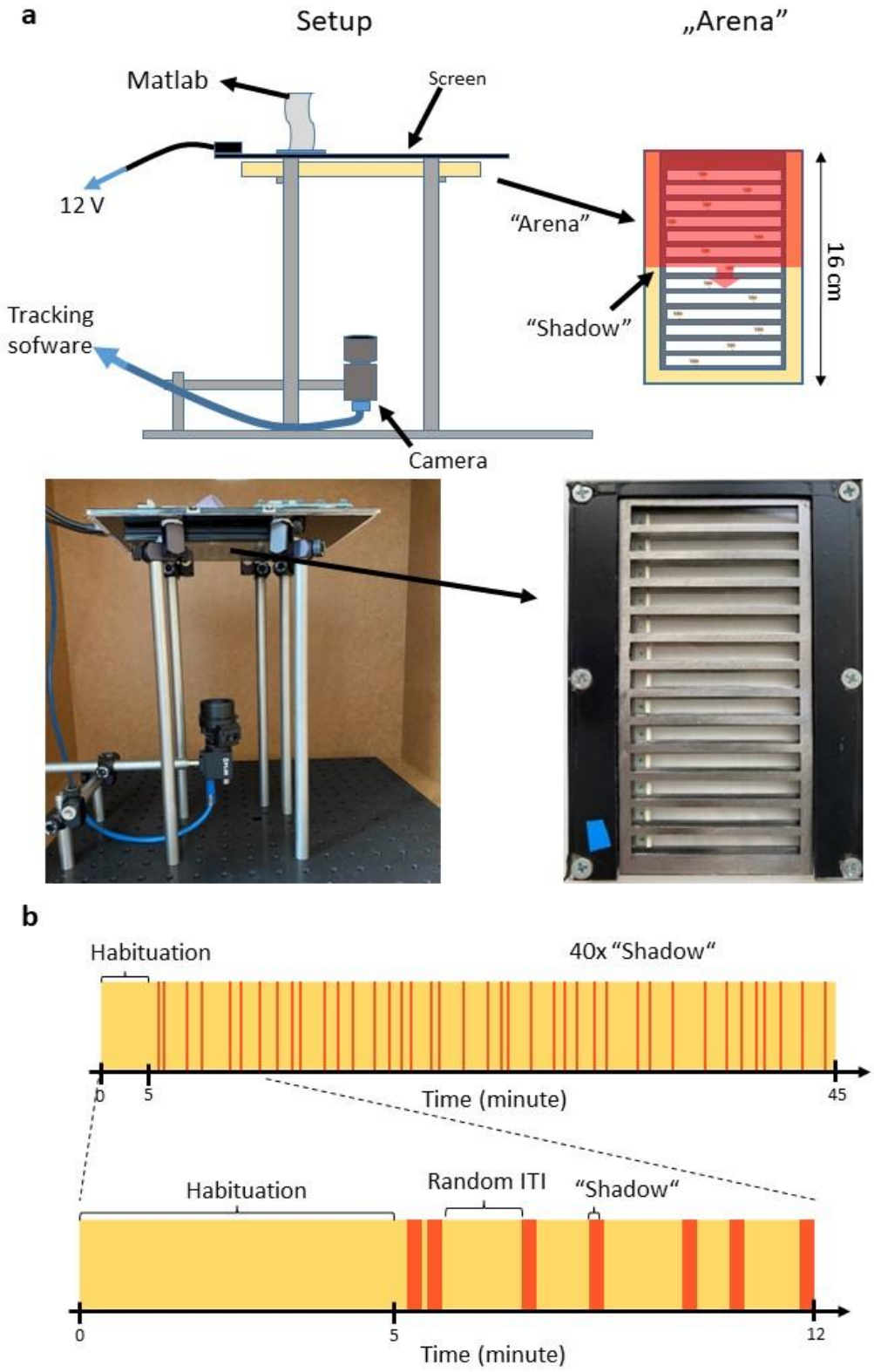


Figure 2

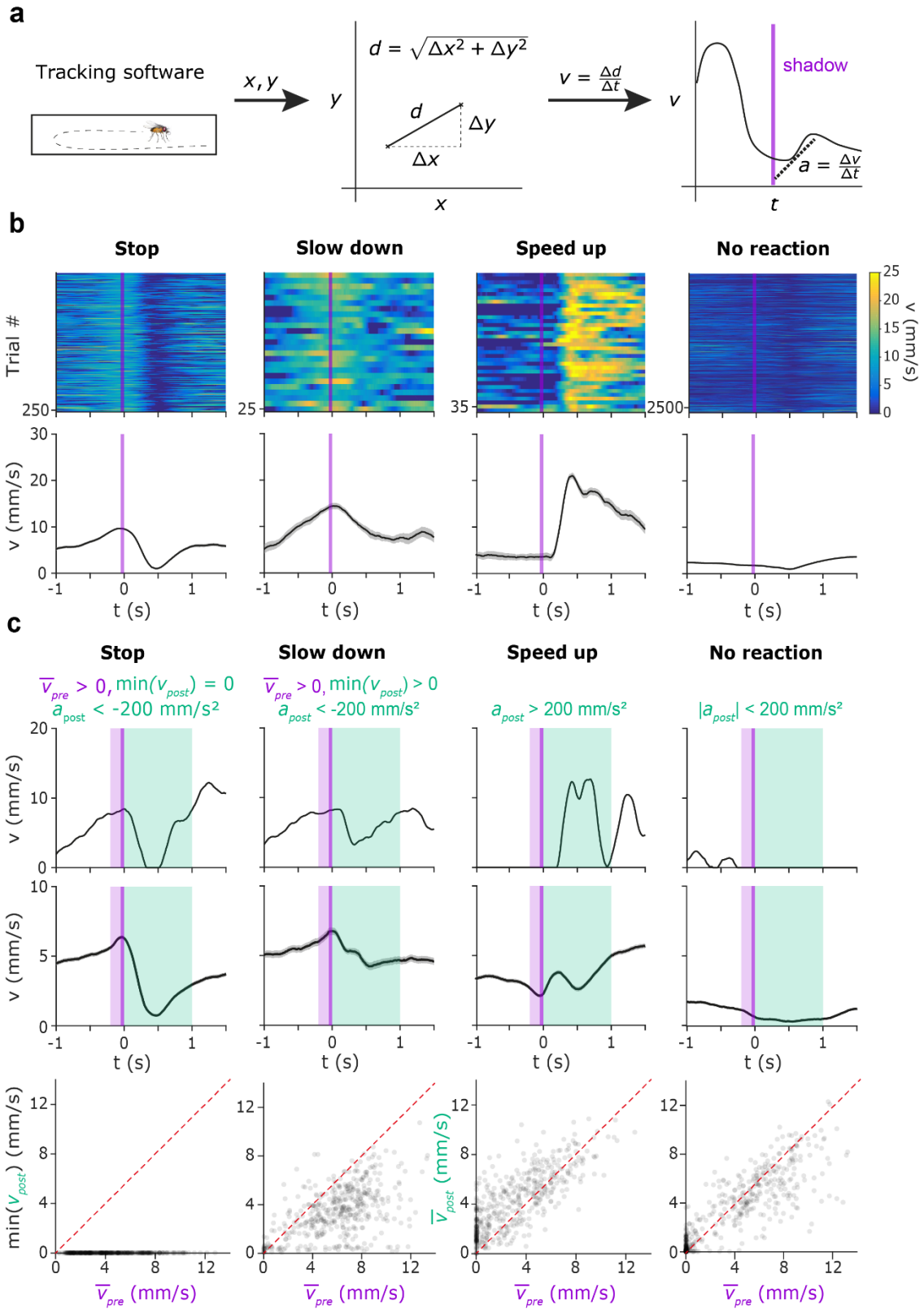

a
Tracking software
x, y
y
x
d = √(Δx² + Δy²)
d
Δy
Δx
v = Δd/Δt
v
t
shadow
a = Δv/Δt
b
Stop
Slow down
Speed up
No reaction
Trial #
250
25
35
2500
v (mm/s)
t (s)
c
Stop
Slow down
Speed up
No reaction
v̄pre > 0, min(vpost) = 0
apost < -200 mm/s²
v̄pre > 0, min(vpost) > 0
apost < -200 mm/s²
apost > 200 mm/s²
|apost| < 200 mm/s²
min(vpost) (mm/s)
v̄post (mm/s)
v̄pre (mm/s)

Figure 3

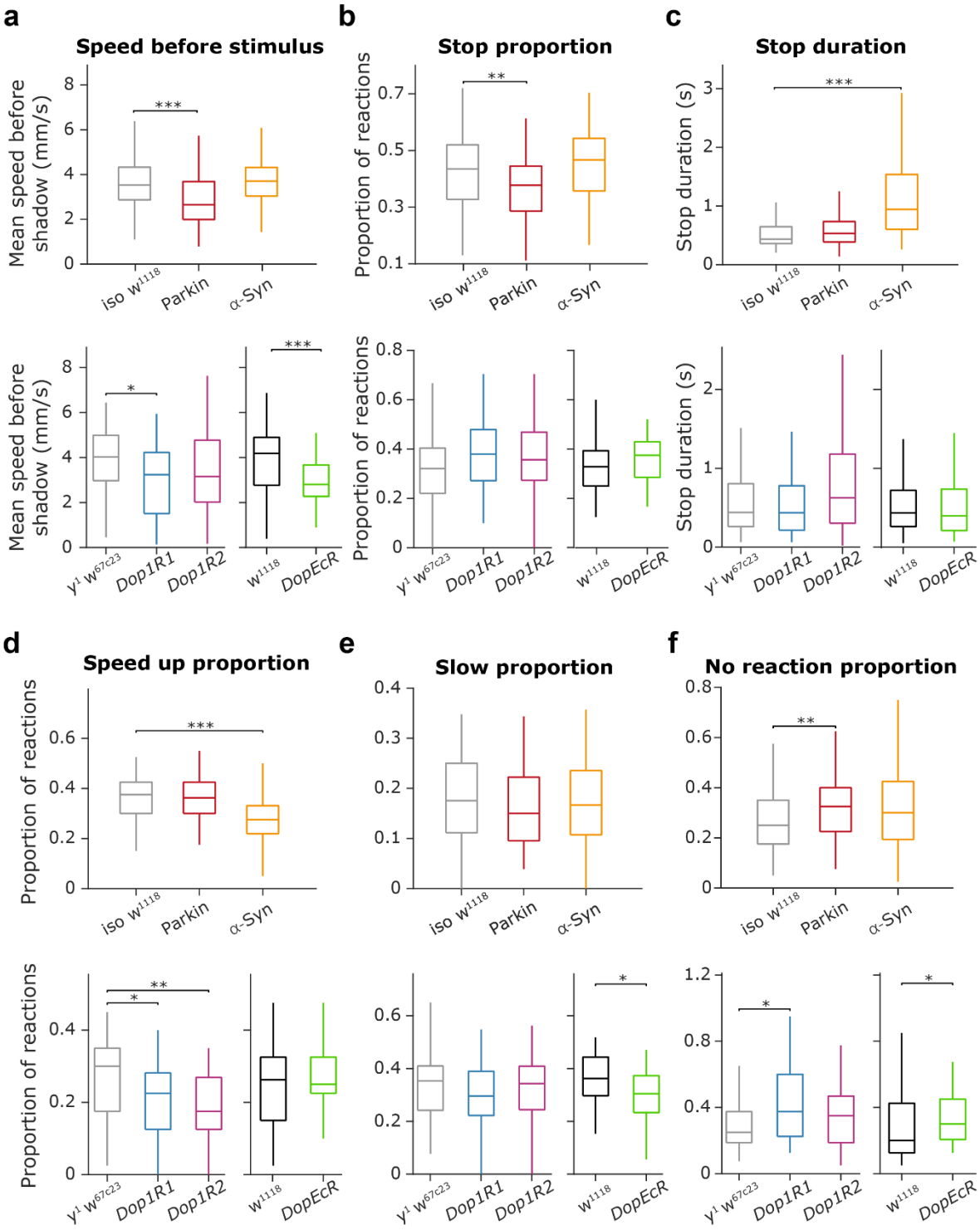
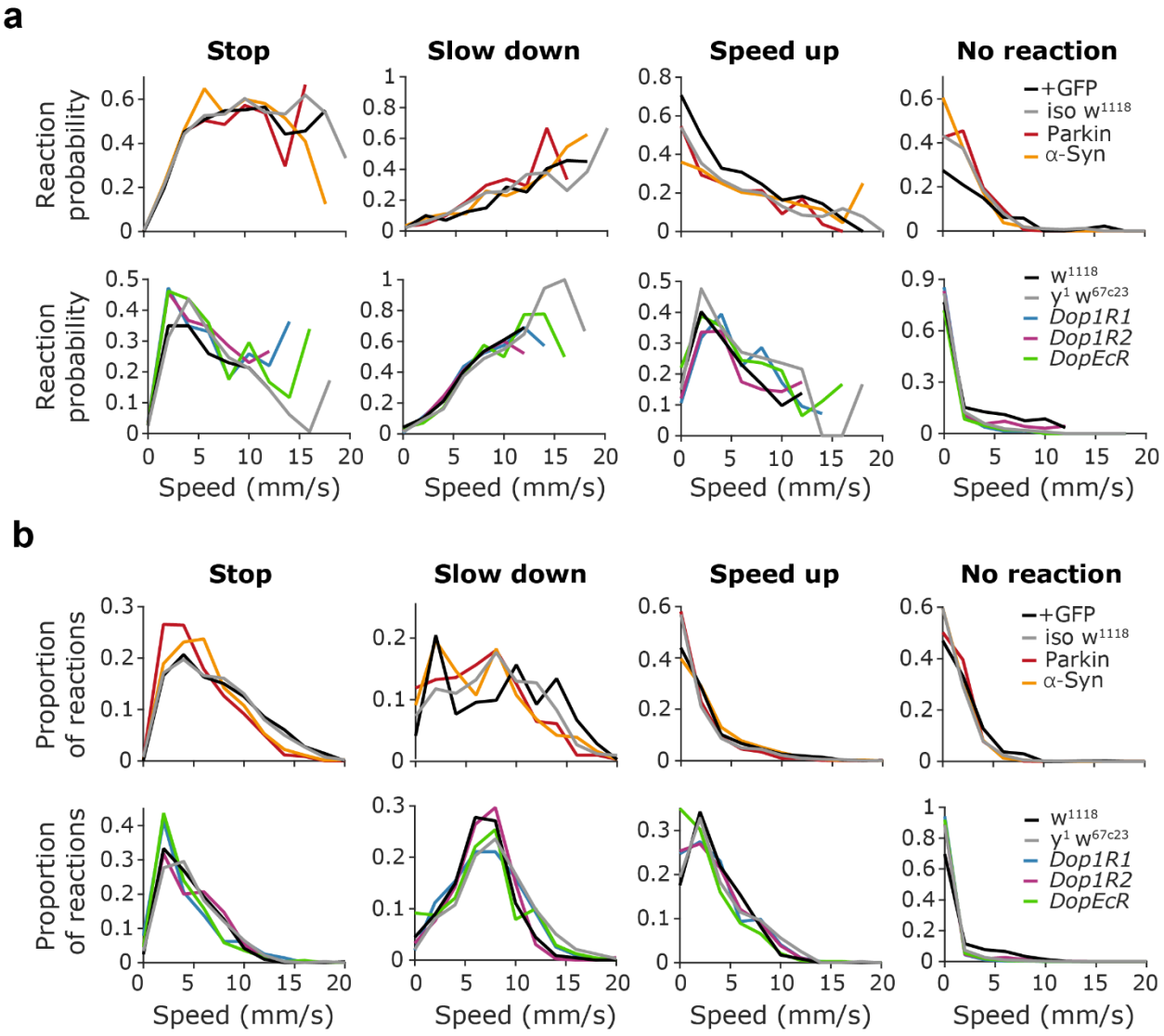
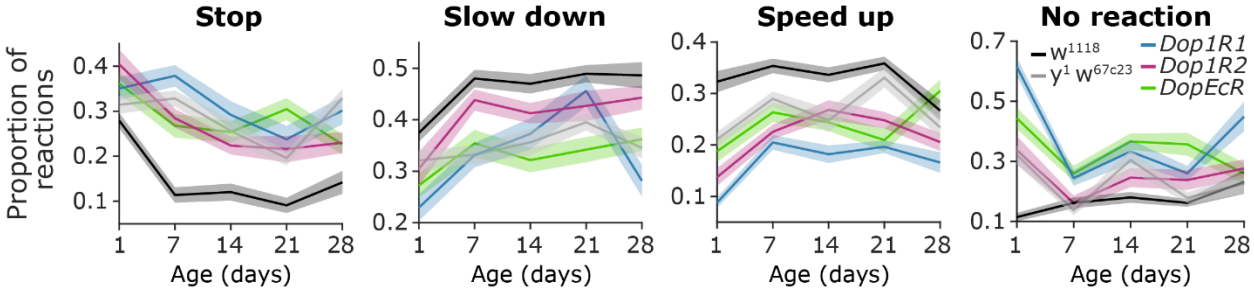

a
Speed before stimulus
Mean speed before shadow (mm/s)
b
Stop proportion
Proportion of reactions
c
Stop duration
Stop duration (s)
iso w1118
Parkin
α-Syn
y1 w67c23
Dop1R1
Dop1R2
w1118
DopEcR
d
Speed up proportion
e
Slow proportion
f
No reaction proportion

Figure 4


a
Stop
Slow down
Speed up
No reaction
Reaction probability
+GFP
iso w1118
Parkin
α-Syn
w1118
y1 w67c23
Dop1R1
Dop1R2
DopEcR
Speed (mm/s)
b
Proportion of reactions

802 **Figure 5**



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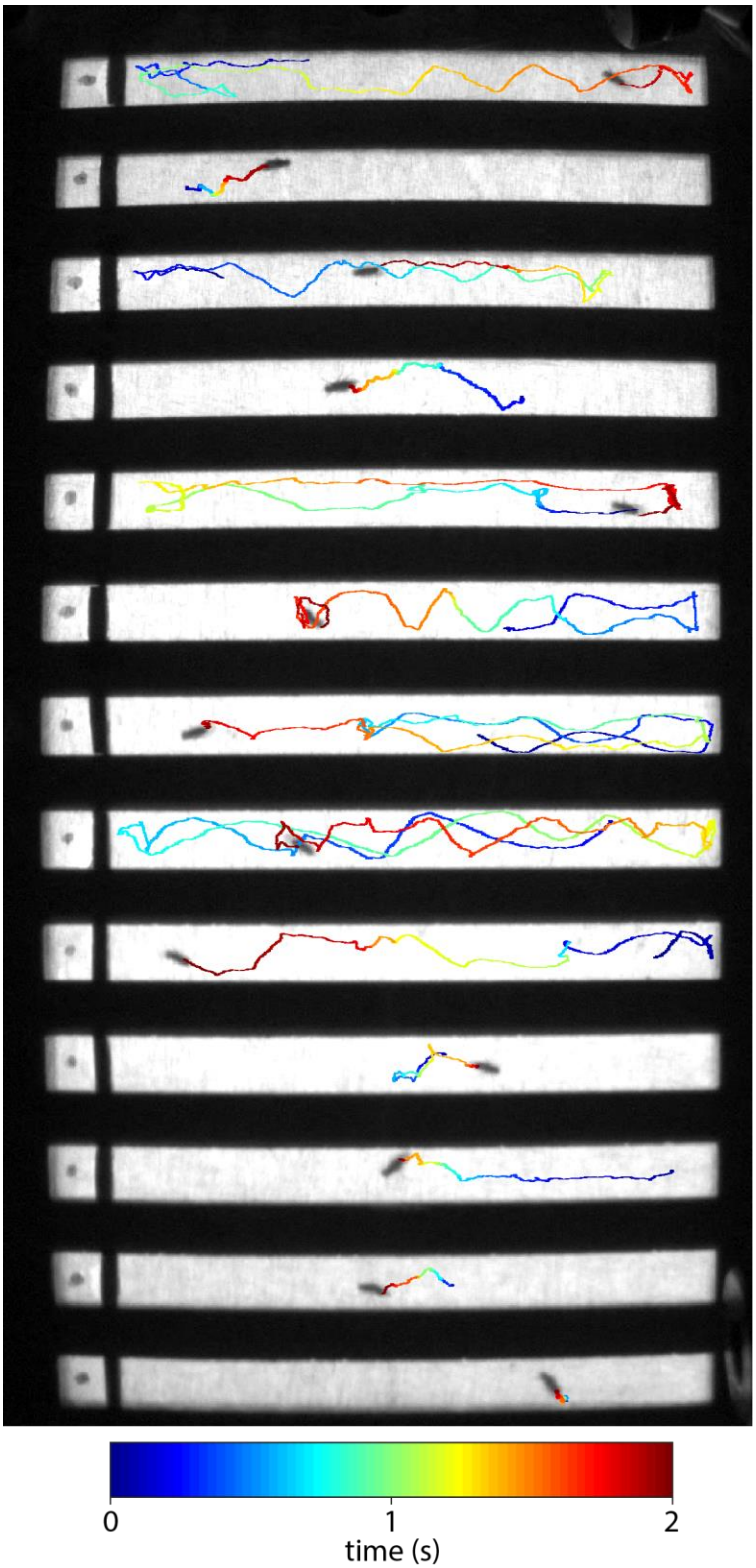
Supplementary Figures

Figure S1



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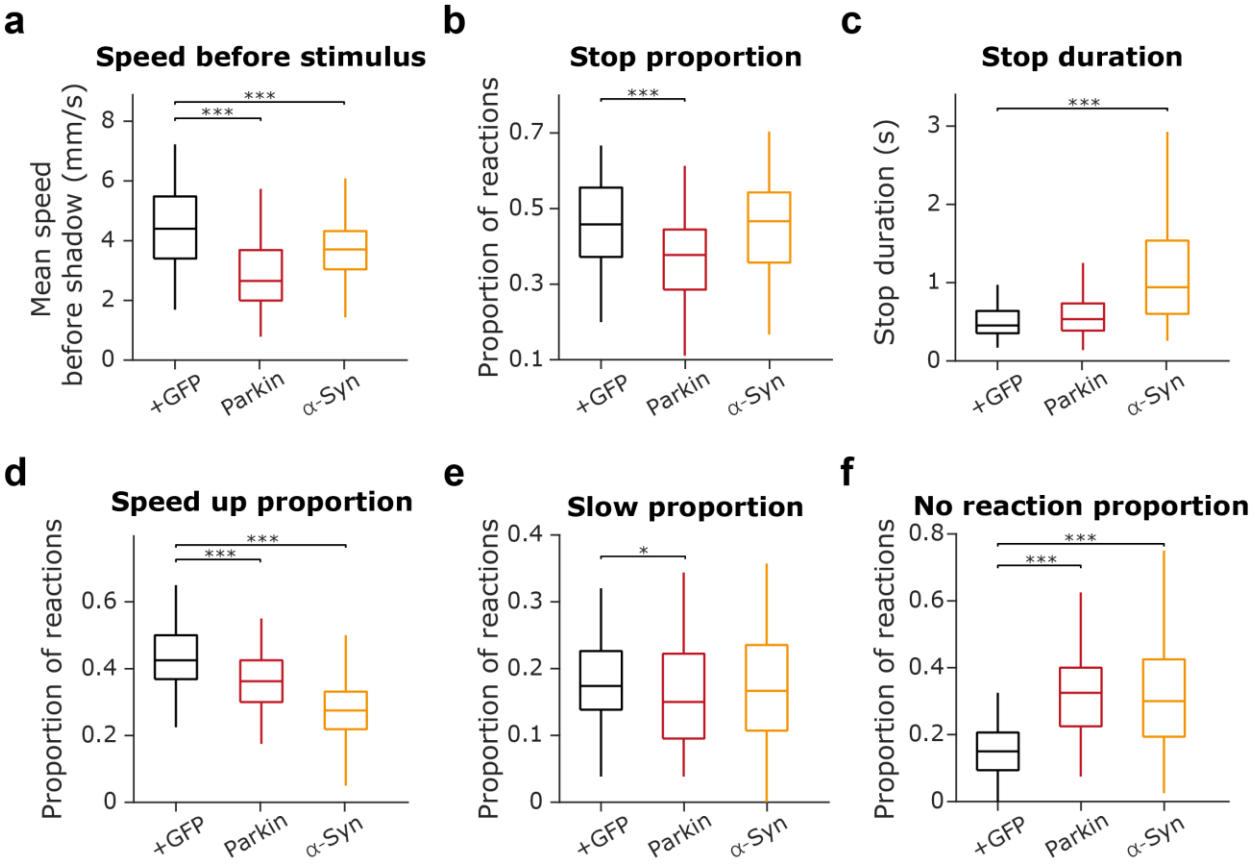

a
Speed before stimulus
Mean speed before shadow (mm/s)
0
2
4
6
8
***
***
+GFP
Parkin
α-Syn
b
Stop proportion
Proportion of reactions
0.1
0.3
0.5
0.7
***
+GFP
Parkin
α-Syn
c
Stop duration
Stop duration (s)
0
1
2
3
***
+GFP
Parkin
α-Syn
d
Speed up proportion
Proportion of reactions
0
0.2
0.4
0.6
***
***
+GFP
Parkin
α-Syn
e
Slow proportion
Proportion of reactions
0
0.1
0.2
0.3
0.4
*
+GFP
Parkin
α-Syn
f
No reaction proportion
Proportion of reactions
0
0.2
0.4
0.6
0.8
***
***
+GFP
Parkin
α-Syn

Figure S3

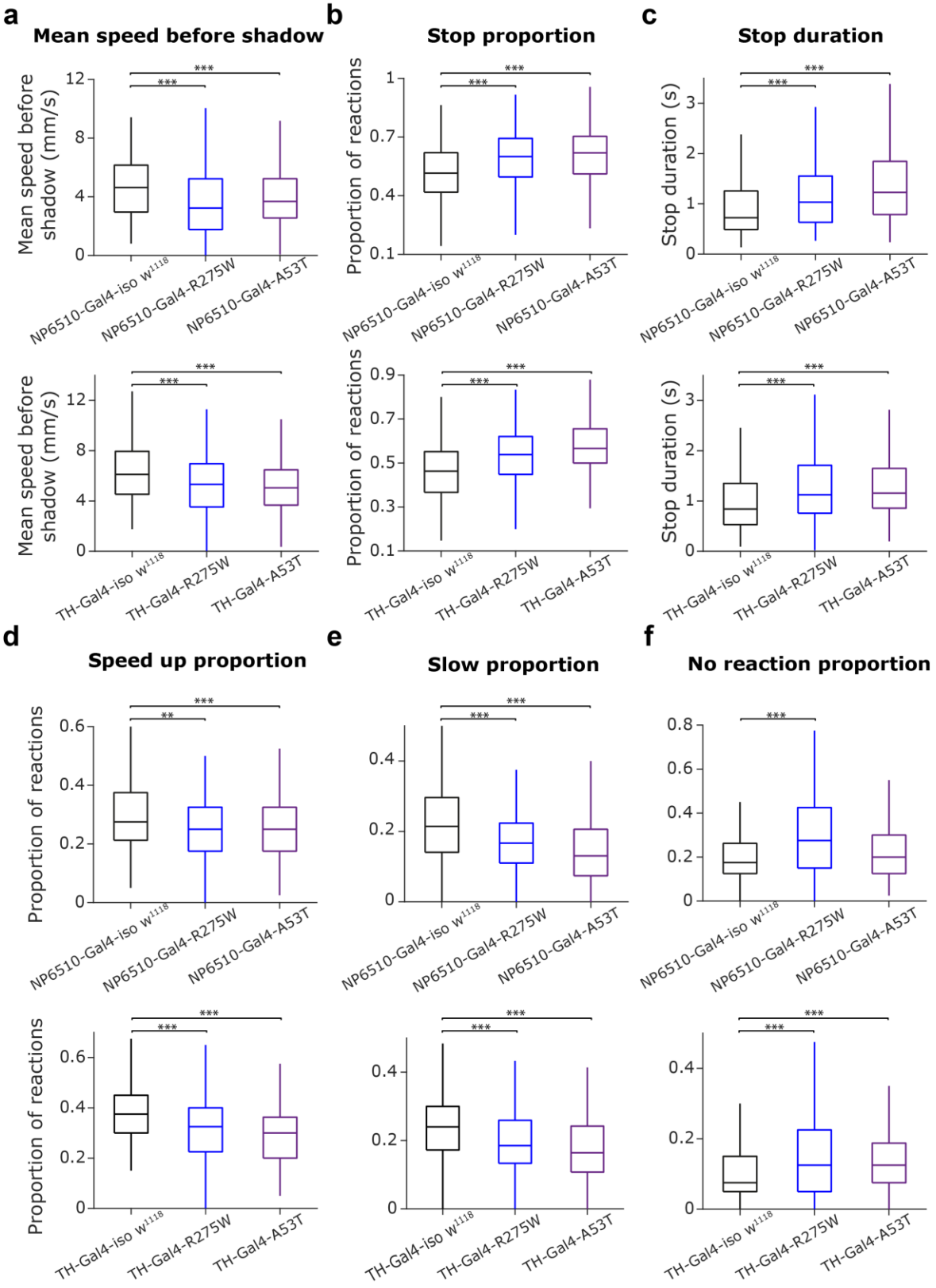
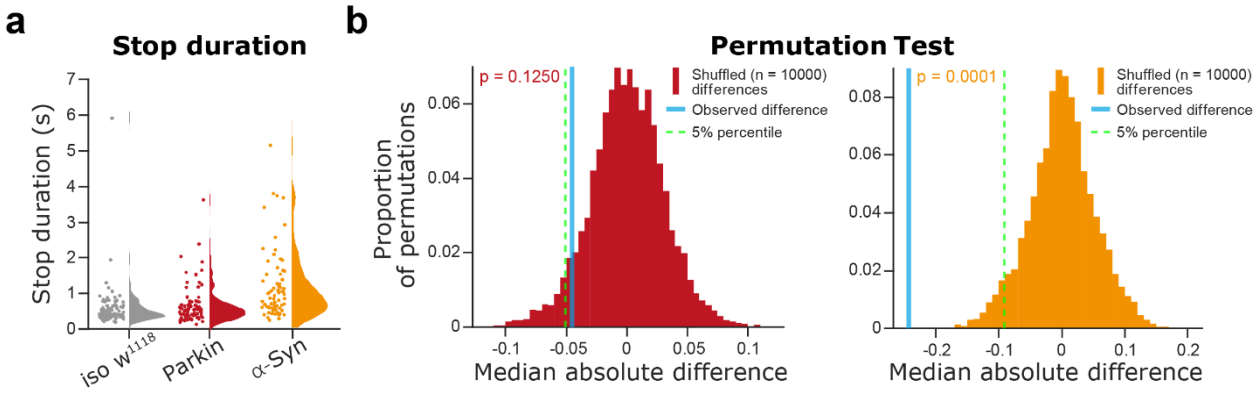
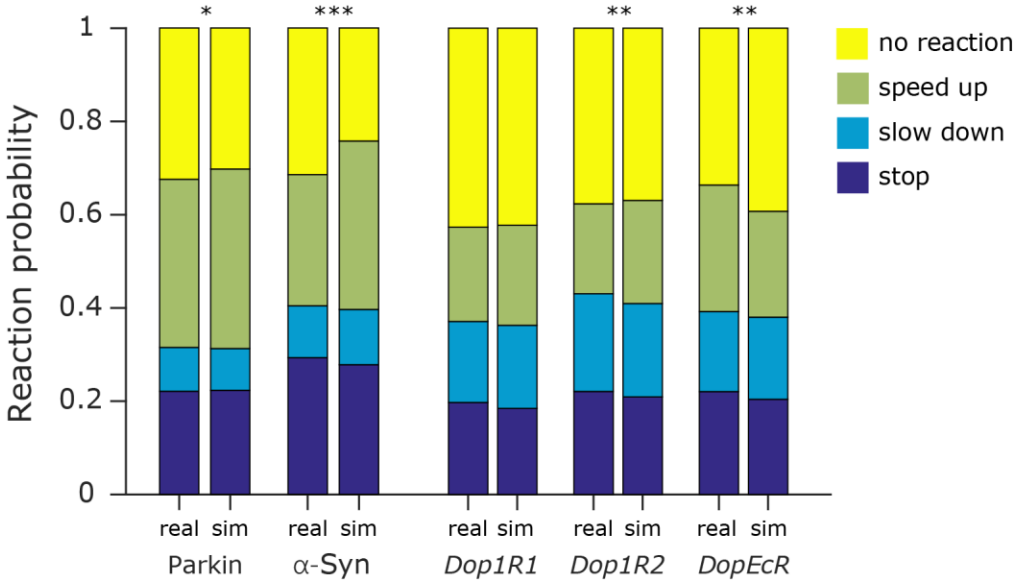

a
Mean speed before shadow
Mean speed before shadow (mm/s)
b
Stop proportion
Proportion of reactions
c
Stop duration
Stop duration (s)
NP6510-Gal4-iso w1118
NP6510-Gal4-R275W
NP6510-Gal4-A53T
TH-Gal4-iso w1118
TH-Gal4-R275W
TH-Gal4-A53T
d
Speed up proportion
e
Slow proportion
f
No reaction proportion
***
**

Figure S4


a
Stop duration
Stop duration (s)
iso w1118
Parkin
α-Syn
b
Permutation Test
p = 0.1250
Shuffled (n = 10000) differences
Observed difference
5% percentile
Proportion of permutations
Median absolute difference
p = 0.0001
Shuffled (n = 10000) differences
Observed difference
5% percentile
Median absolute difference

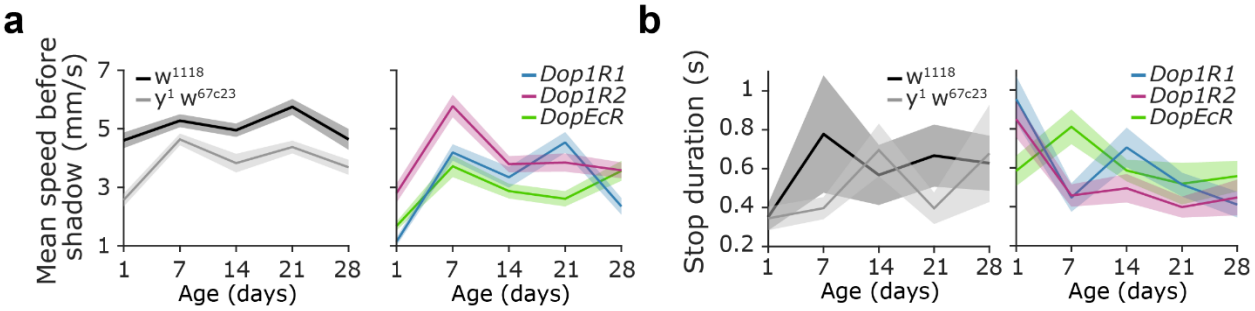
815 **Figure S5**



816

817

Figure S6


a
Mean speed before shadow (mm/s)
7
5
3
1
w1118
y1 w67c23
1 7 14 21 28
Age (days)
Dop1R1
Dop1R2
DopEcR
1 7 14 21 28
Age (days)
b
Stop duration (s)
1
0.8
0.6
0.4
0.2
w1118
y1 w67c23
1 7 14 21 28
Age (days)
Dop1R1
Dop1R2
DopEcR
1 7 14 21 28
Age (days)