Navigating Pubertal Goldilocks: The Optimal Pace for Hierarchical Brain Organization

Hanna Szakács[a,b], Murat Can Mutlu[c,d], Giulio Balestrieri[e], Ferenc Gombos[a,f], Jochen Braun[c,d], Morten L. Kringelbach[g,h,i], Gustavo Deco[e,j,k], Ilona Kovács[f,l,*]

[a]Laboratory for Psychological Research, Pázmány Péter Catholic University, 1 Mikszáth square, Budapest 1088, Hungary
[b]Semmelweis University Doctoral School, Division of Mental Health Sciences, 26 Üllői road Budapest 1085, Hungary
[c]Institute of Biology, Otto-von-Guericke University, 44 Leipziger Straße
[d]Center for Behavioral Brain Sciences, Otto-von-Guericke University, 44 Leipziger Straße 39120 Magdeburg, Germany
[e]Center for Brain and Cognition, Computational Neuroscience Group, Universitat Pompeu Fabra, 25-27 Ramon Trias Fargas Barcelona 08005, Spain
[f]HUN-REN-ELTE-PPKE Adolescent Development Research Group, 1 Mikszáth Kálmán square Budapest 1088, Hungary
[g]Centre for Eudaimonia and Human Flourishing, Linacre College, University of Oxford, Oxford, UK
[h]Department of Psychiatry, University of Oxford, Oxford, UK
[i]Center for Music in the Brain, Department of Clinical Medicine, Aarhus University, Aarhus, Denmark
[j]Department of Information and Communication Technologies, Universitat Pompeu Fabra, 122-140 Carrer de Tànger, Barcelona 08018, Spain
[k]Institució Catalana de la Recerca i Estudis Avançats (ICREA), 23 Passeig de Lluís Companys Barcelona 08010, Spain
[l]Institute of Psychology, Faculty of Education and Psychology, Eötvös Loránd University, 25-27 Kazinczy street, Budapest 1075, Hungary

*corresponding author: Ilona Kovács, Institute of Psychology, Faculty of Education and Psychology, Eötvös Loránd University, 25-27 Kazinczy street, Budapest 1075, Hungary, e-mail: kovacs.ilona@ppk.elte.hu

Abstract

Adolescence is a timed process with an onset, tempo, and duration. Nevertheless, the temporal
dimension, especially the pace of maturation, remains an insufficiently studied aspect of
developmental progression. This study focuses on the modifications due to the different timings
of developmental shifts during adolescence and addresses the impact of adolescent maturation
on brain development. To reveal potential relationships between pubertal pace and the
advancement of brain organisation, we analyse the connection between skeletal age-based
maturation stages and hierarchical organisation in the temporal dynamics of resting-state EEG
recordings (alpha frequency range). By adopting skeletal maturity as a proxy for pubertal
progress and employing entropy production to measure hierarchical brain organisation, our
findings indicate that an average maturational trajectory optimally aligns with cerebral
hierarchical order. Adaptive developmental plasticity may not fully compensate for accelerated
or decelerated timelines, potentially increasing the risk of behavioural problems and psychiatric
disorders consequent to such alterations.

Main

Similar to early childhood development, adolescence is a time-bound event with an onset,
tempo, and duration. While the interaction between genetic and environmental factors is
established as a fundamental contributor to diversities in adult behavioural consequences, the
temporal aspect, specifically, the speed of maturation, remains an insufficiently explored
feature of developmental progression[1,2]. In the current study, we focus on the alterations of
hierarchical brain organisation, related to different timings of pubertal maturation.

Adolescent cerebral maturation involves intricate sequences of neurodevelopment[3,4],
interwoven with alterations by age and stress[5–7]. Normative trajectories delineating volumetric[8]
and synaptic density[9] shifts underscore the ongoing developmental dynamics of the adolescent
brain. It is important to acknowledge, however, that the comprehensive volumetric
transformations and mean synaptic density reflect the cumulative outcome of anatomical
reconfiguration within the human brain, where the different components undergo
heterochronous maturation[10,11]. Increasingly earlier onset of puberty referred to as worldwide
secular trends[12–14], or delayed onset due to, e.g., malnutrition or eating disorders[15,16] might have
significant consequences on developmental trajectories, impacting a wide range of
physiological and psychological processes. For instance, puberty-associated reductions in
cortical thickness[17] or synaptic density[18] may manifest prematurely or belatedly, deterring the
adaptability of regions maturing at later stages to the atypical developmental milieu. Premature
or postponed puberty can also generate challenges in behavioural adjustment, potentially
leading to social isolation and heightened anxiety, particularly in the absence of adequate
coping mechanisms[19], thereby emphasising the critical role of adolescence in mental health[8,20].
Consequently, it is essential to investigate the impact of pubertal timing on the developing
brain.

Uncertainty around the impact of atypical pubertal timelines arises from ineffective maturity
measures and difficulty dissociating chronological age from pubertal maturity[21,17,22–24]. While
puberty onset and its potential impact on developmental trajectories have long been a topic of
interest, with significant individual variability suspected[23,25,26], a precise assessment of the
relationship between maturity and brain development remains noticeably absent from the
literature, which is due to the lack of a reliable method to assess maturity levels. The Tanner
Scale[27–29], considered as the benchmark, relies on subjective evaluations of physical attributes
such as breast and testicle size, rendering it susceptible to evaluator subjectivity. This scale
originates from a post-war longitudinal study carried out within an orphanage during 1949-
1971, thus failing to account for contemporary nutritional conditions and secular trends in
growth[13,28,30–32]. Self- and parent-report versions of this method have also been deemed
unreliable[29,28,33].

Recently, we have introduced ultrasonic bone age as a promising alternative for maturity
assessments in human developmental research[34], with evidence for selective maturity-
dependent effects in cognitive[34,35], motor[36], and emotional development[37]. Operating without

harmful radiation, this technology gauges acoustic conductivity at hand and wrist growth zones
(*Fig. 1a*) to estimate bone age, demonstrating robust correlation with pubertal hormone
levels[28,33,38,39], X-ray estimations[40], and displaying high reliability[40]. This method overcomes
traditional pubertal staging accuracy issues by providing an objective, continuous measure with
unprecedented accuracy. In the current study, skeletal maturity serves as a proxy for individual
pubertal maturity status. Addressing the impact of pubertal timing on brain development, the
degree of maturity of each participant is calculated based on the difference between their bone
age (BA) and chronological age (CA). It is widely acknowledged that the timing of puberty
can differ significantly among individuals and follows a normal distribution[41–44], therefore, to
allow for statistical comparison across groups with different maturity levels, in our cross-
sectional study, we selected an equal number of decelerated (BA < CA), average (BA = CA)
and accelerated (BA > CA) maturity participants after screening them for bone age (*Fig. 1*,
please also consult *Fig. 2a* under the *Results* section*,* and the *Methods/Participants* section for
details). This approach not only disentangles chronological age from pubertal maturity (*Fig.*
2b) but also provides insights into how adolescents with varying maturity levels navigate
pubertal goldilocks.

As illustrated in *Fig. 1*, beyond bone development, the refinement of brain structure is also part
of the anatomical changes associated with puberty and adolescence. One of the significant
maturational events during this phase is the targeted elimination of synapses, known as synaptic
pruning[45–48] (*Fig. 1b*), which occurs in various species[49,50] and across the nervous system[45]
during development to adapt to environmental conditions[48,50]. This process is vital for optimal
brain function and shaping a healthy adult brain[47,51]. The timing of synaptic pruning has been
linked to puberty onset in rodents[52,53] and humans[18], suggesting that early or late puberty may
lead to a correspondingly early or delayed initiation of synaptic pruning. However, it is not
clear whether synaptic pruning follows a normal course or becomes dysregulated when it
begins earlier or later (*Fig. 1b*). As insufficient synaptic pruning or „underpruning" has been
tied to neurodevelopmental disorders[4,22] and excessive pruning or „overpruning" to psychiatric
conditions such as schizophrenia[54,55], it is important to investigate whether untimely maturation
may lead to the emergence of alterations in brain organisation, thereby contributing to the
current psychological challenges faced by adolescents[56–60].

The temporal aspect of synaptic pruning holds significance within hierarchical
neurodevelopment as well. Typically, transmodal association regions undergo peak synaptic
density later than sensory zones, experiencing pruning throughout adolescence[11,45,61]. This
design permits a broad experience-dependent refinement window but also introduces the
prospect of prolonged vulnerability[11,62]. It is also becoming increasingly clear that
developmental alterations in the functional[63,64] and evolutionary[65] hierarchies of the brain
appear to mirror anatomical maturation patterns. Consequently, our primary objective is to
investigate the precise influence of pubertal maturation on the configuration of associative
brain regions, which manifest the most substantial developmental shifts during adolescence.
For this purpose, we assess resting-state brain activity and extract the degree of hierarchical
arrangement among participant groups exhibiting diverse levels of physical maturation.

Fig. 1: Anatomical aspects of skeletal and brain maturation.

a, Bone development exhibits strong correlation with pubertal hormone levels. As bones form, acoustic conductivity within the growth zones undergoes alteration. This change in conductivity serves as the foundation for contemporary ultrasonic bone age assessments - a method offering objective, non-invasive estimation of biological age (BA) in children and adolescents. While a large proportion of the population aligns their BA with chronological age (CA), there are also accelerated individuals whose BA exceeds their CA, and decelerated ones, whose BA is less than their CA (the illustration does not show these variations, only examples of average bone development). **b**, Brain development is also profoundly influenced by the dynamic shifts in pubertal hormone levels. For example, a targeted process of synaptic pruning (reduction in dendritic spine density) is initiated by puberty onset, and pruning then continues until adulthood throughout the nervous system. However, as our conceptual figure suggests, synaptic pruning may not adhere to its expected course, but it may undergo dysregulation in cases of accelerated or decelerated pubertal maturation. Untimely maturation with dysregulated synaptic pruning may lead to the emergence of alterations in hierarchical brain organisation, which is the main issue addressed in the current study.

Recent progress in neuroimaging has enabled the creation of normative brain-growth charts[8,66],
similar to those applied for anthropometric attributes like height and weight. Despite their
reliance solely on chronological age and their predominant cross-sectional nature, these
investigations reveal the spatiotemporal dynamics of developmental plasticity and have even
identified a sensorimotor-to-associative sequence of refinement[10,11]. The latter findings align
with the progress of phylogenetic brain development from less variable unimodal areas towards
heteromodal association cortices[65,67,68]. It is important to recognise that the most recently
evolved associative networks within the brain might be particularly vulnerable to diverse

pubertal timelines, given their ontogenetic maturation coincides more closely with the
adolescent phase. Irregular timelines could also relate more to alterations in the finely tuned
intrinsic activity of these networks, as opposed to extensive anatomical reconfigurations. While
fMRI has proven useful in assessing intrinsic resting-state activity, it is sensitive to neural,
vascular, and respiratory influences. Therefore, direct measures of neural activity might offer
better options for investigating this intrinsic behaviour. Resting-state EEG provides direct
measures of intrinsic cortical activity, allowing for the detection of changes in oscillatory
activity and functional connectivity that are not captured by fMRI. In this study, we rely on
resting-state EEG in the alpha frequency range (see *Methods*) to compare the progression of
hierarchical brain organisation across groups with early, on-time and late maturing groups as
defined by the difference between their bone age and chronological age.

To reveal any relationships between pubertal pace and the advancement of brain organisation,
we analyse how skeletal age-based maturity levels link to high-level features of resting-state
EEG-dynamics. Specifically, we introduce "entropy production" to capture the degree of
functional hierarchical organisation of the brain. In thermodynamics and systems biology, the
asymmetry and directionality of flow in the state space of the components of a living system is
known as "breaking of the detailed balance" and give rise to non-equilibrium states which can
be captured by the level of non-reversibility in time[69,70]. In other words, different levels of
hierarchical organisation allow the orchestration of the whole-brain dynamics accordingly in
different ways. Hierarchy is built up by the underlying asymmetry in the directionality of
information flow between different regions. Thermodynamics establishes a direct link between
hierarchy, non-equilibrium, and time irreversibility. This comes from the fundamental idea of
the second law of thermodynamics, explicitly stating that a system will go from order to
disorder over time. We capture these different levels of organisation and non-equilibrium
through the level of irreversibility, i.e., through the "arrow of time."[69,71]

Asymmetric flow can be observed in state spaces at various scales, from the molecular[72] to the
macroscopic brain dynamics[70]. In general, asymmetric flow reflects a degree of non-
reversibility in the dynamics of the system. In the case of the brain, such non-reversibility is
thought to reflect the hierarchical organisation of brain activity[69,73]. Entropy production is a
direct measure of non-reversibility and indirectly captures the functional hierarchy expressed
by brain activity[69,71,74] (see *Methods*). The purpose of our study is to investigate the relationship
between entropy production, as a proxy of hierarchical brain activity, and the level of physical
maturation.

Results

As it is explained in *Fig. 2b*, we dissociated biological and chronological age through the
assignment of participants into one-year-wide hexagonal bins to ensure the absence of overlap
between maturity groups with approximately equal numbers of participants. We then analysed
the data through various grouping approaches illustrated in *Fig. 2c-d*, allowing for the analysis
of chronological age and maturity levels independently. Results are discussed according to
those groupings. We have obtained and analysed closed-eye resting state high-density EEG

recordings in the alpha frequency range (8-12 Hz) in 61 adolescent, and 26 emerging adult
female participants (see *Fig. 2* and *Methods/Participants*). To measure the level of hierarchical
brain activity in resting-state EEG recordings, we parsed the recorded neural data into 12
topographically distinct, phase-based activity patterns, referred to as "brain states" (see *Fig. 6*),
utilising a modified *k*-means clustering algorithm[75]. Each brain state captures patterns where
two major electrode sets record congruent signals, but these sets are in opposition to one
another. After extracting the brain state patterns, we studied the sequence in which these brain
states occurred, specifically focusing on the asymmetrical nature of their transitions. A
sequence of consecutive brain states is illustrated in *Fig. 7a* as a representative example in the
Methods/Data Analysis section, while *Fig. 7b* presents a table that counts the observed
occurrences of each transition within this example, highlighting their asymmetric quality.

Fig. 2 Participant grouping by adolescent age and maturity.

a, The black curve illustrates the normal distribution of diverse maturity levels in a standard population, where maturity is represented along the horizontal axis as the discrepancy between biological age (BA) and chronological age (CA). Positive differences indicate accelerated, while negative differences indicate decelerated maturity. In the present study, to explore the influence of varying maturity levels, we initially assessed bone age using an ultrasonic device to estimate the biological age of participants. We then selected an approximately equal number of participants with decelerated, average, and accelerated maturity levels. **b,** Dissociation of biological and chronological age is achieved through the assignment of participants into one-year-wide hexagonal bins which ensures the absence of overlap between maturity groups. The data were then analysed through various grouping approaches: **c,** The effect of chronological age is analysed by averaging across different maturity groups within specific age groups; **d,** The effect of maturity is analysed by averaging across age groups within distinct maturity groups; **e,** By comparing data at the bin-level, the interplay between age and maturity effects is revealed.

Chronological age related entropy production

To evaluate the effect of chronological age on entropy production, we averaged across maturity
groups within a 12-year-old (N=31, mean age=12.48y, SD=0.27y) and a 13-year-old (N=30,
mean age=13.58y, SD=0.29y) group. In order to have normative data from young adults, we
also included 22-year-old participants (N=26, mean age=21.69y, SD=0.64y). For further
details see *Fig. 2c*, *Fig. 3c*, and *Methods*/*Participants*. Importantly, adult data should only be
compared to age-averaged results of adolescents as the adult cohort has not been selected with
respect to maturity. Given that bone formation is completed by adulthood, bone age assessment
is irrelevant in adults. Therefore, adult data were included in the statistical analysis only where
the basis of the analysis is chronological age, irrespective of maturity.

Fig. 3a and *3b* provide a detailed look at brain state dynamics within each age-group.
Logarithmic irreversibility transition matrices in *Fig. 3a* illustrate a contribution to entropy
production of brain state transitions in age groups, derived from the log ratio of the forward
and reverse transition probabilities, estimated from transition counts. Matrices were generated
through averaging individual matrices within each age group, followed by normalisation of
values to a -1 to +1 range (using the largest absolute value of the global extremum), indicated
by the colour scale. Each matrix holds 12x12 values, with the starting brain states on the
horizontal and the subsequent states in brain state switching on the vertical axis. Log ratios of
the forward and reverse transition probabilities are non-symmetric. Positive and negative
values indicate a preference for forward and reverse transitions, respectively. The relative
similarity in the variability in hue and intensity indicates comparable entropy production
between age groups, with a slight indication of age-related improvement. Graphs in *Fig. 3b*
show total entropy production contributions of brain state pairs, computed by summing the log
ratio values of forward and reverse transition probabilities within each pair, creating symmetric
matrices. Graph values are generated by averaging the individual symmetric matrices within
age groups, then normalising them between 0 and 1. The nodes represent the 12 brain states,
with the most dominant state (highest percentage of total activity) positioned at the 12 o'clock
position, and subsequent states are arranged clockwise in order of descending dominance. The
edges indicate pairwise contributions to entropy production, where increasing thickness and
darkness represent greater contribution. The slight increase in line thickness and darkness
across age-groups may indicate a subtle rise in contributions to entropy production with age.

Group averages and variances of entropy production were computed via inverse-variance
weighting to minimise the biassing effect on the weighted arithmetic mean introduced by
variance. 95% confidence intervals of group-level entropy production were also calculated
using the weighted group average. Weighted group averages revealed a tendency of increase
in entropy production with age, as it is demonstrated in *Fig. 3d*. Specifically, the 12-year-old
group exhibited an average mean entropy production of 0.02300 bit (95% CI: 0.02064-
0.02537), while the average mean entropy production of the 13-year-old group was 0.02415
bit (95% CI: 0.02166-0.02663), and the average mean entropy production of the 22-year-old
group was 0.02538 bit (95% CI: 0.02271-0.02805). To determine whether the group
differences are statistically significant, we utilised two-sample z-tests to compare the group

averages (see *Methods* for further details). While a slight increase in groupwise average entropy production with chronological age is observed, pairwise comparison of group averages were not statistically significant ($p>0.05$ in each comparison). Hedges' g effect size estimates revealed small effect sizes in all cases: between the 12-year-old and 13-year-old (g=0.17), and between 13-year-old and 22-year-old (g=0.18) groups, and in the comparison of the 12-year-old and 22-year-old group (g=0.35).

Our statistical results indicate that, while there is a slight increase in entropy production with increasing chronological age, this variable independently does not have a substantial impact on entropy production, further warranting the inspection of maturity in adolescence.

Fig. 3 Chronological age related entropy production.

a, Logarithmic irreversibility transition matrices illustrate two distinct, non-symmetric values in each brain state pair, derived from the log ratio of the forward and reverse transition probabilities. The matrices inform about the preference for directionality in brain state switching. The extent of deviation from 0 reflects the strength of directional preference (forward transitions in red, backward transitions in blue), indicating the level of irreversibility and hierarchical organisation in the brain. The relative similarity in the variability in hue and intensity indicates comparable entropy production between age groups. **b**, Graphs represent the cumulative contribution to entropy production of both forward and reverse transitions and reveal the level of irreversibility in the dynamics of each brain state pair within age groups. Nodes represent the 12 brain states, edges indicate pairwise contributions to entropy production, where increasing thickness and darkness represent greater contribution, therefore larger irreversibility. There seems to be a subtle rise in contributions to entropy production with age. **c**, The effect of chronological age is analysed by averaging across different maturity groups within specific age groups. **d**, Bar plots illustrate the average entropy production and respective 95% confidence interval for each age group calculated via inverse-variance weighting. Differences among groups were statistically assessed using two-sample z-tests, accompanied by Hedges' g effect size calculations, yielding no statistically significant difference among the age groups in pairwise comparisons ($p > 0.05$) and small effect sizes in all comparisons.

Maturity related entropy production

To analyse the relationship between maturity and group-level average entropy production, we
sorted adolescent participants according to their level of maturity, irrespective of chronological
age, forming three distinct groups: decelerated maturity (CA-1.5≤BA<CA-0.5; N=21, mean
age=12.98y, SD=0.60y, mean BA-CA=-0.92, SD=0.24), average maturity (CA-
0.5≤BA≤CA+0.5; N=21, mean age=12.97y, SD=0.60y, mean BA-CA=0.03, SD=0.29), and
accelerated maturity (CA+1.5≥BA>CA+0.5; N=19, mean age=13.12y, SD=0.68y, mean BA-
CA=0.88, SD=0.31). Participant groups are illustrated both in *Fig. 2d* and *Fig. 4c*.

Fig. 4a and *4b* offer detailed insight into brain state dynamics within each maturity group. As
it is described above, with respect to *Fig. 3a*, logarithmic irreversibility transition matrices in
Fig. 4a illustrate a contribution to entropy production of brain state transitions, in this case
within decelerated, average, and accelerated maturity groups among adolescent participants.
The larger hue and intensity variation in the matrix of the average maturity group indicates
higher entropy production relative to the decelerated and accelerated groups. Graphs in *Fig. 4b*
represent the total contribution of both forward and reverse transitions to entropy production
(see the description related to *Fig. 3b* above), indicating higher entropy production in the
average maturity group, apparent in the thicker and darker edges of the graph compared to that
of the decelerated and accelerated groups.

Using inverse-variance weighting, we calculated the group-level average and variance of
entropy production to minimise the distorting effect of variance, and determined the 95%
confidence intervals based on the weighted group average. The inverse-variance weighted
group average and confidence interval of the decelerated maturity (mean=0.02146 bit, 95% CI:
0.01866-0.02426), average maturity (mean=0.02794 bit, 95% CI: 0.02475-0.03113), and
accelerated maturity (mean=0.02208 bit, 95% CI: 0.01913-0.02503) groups are illustrated in
Fig. 4d. It is clear that the average maturity group expresses higher average entropy production
than both accelerated and decelerated maturity groups. We utilised two-sample z-tests to
determine the statistical significance of the apparent differences, reported with Hedges' g effect
size tests. Results showed that the average maturity group expresses significantly higher
groupwise average entropy production than both accelerated (p=0.0082, g=0.83) and
decelerated (p=0.0028, g=0.92) maturity groups. Effect sizes were large in both pairwise tests.
229
The average maturity group consistently expressed significantly higher (p<0.05) average
entropy production than both decelerated and accelerated groups with additional clustering
approaches, specifically, working with 6, 8 and 10 brain states, and 12 brain states that are
specific to each unique group; see the Open Science Framework link under *Data availability*
for further details.

Our findings show that maturity significantly influences entropy production, irrespective of
chronological age, with the group-level entropy production being significantly higher in the
average maturity group.

Fig. 4 Maturity related entropy production.

a, Logarithmic irreversibility transition matrices illustrate two distinct, non-symmetric values in each brain state pair, derived from the log ratio of the forward and reverse transition probabilities. The matrices inform about the preference for directionality in brain state switching. The extent of deviation from 0 reflects the strength of directional preference (forward transitions in red, backward transitions in blue), indicating the level of irreversibility and hierarchical organisation in the brain. The larger hue and intensity variation in the matrix of the average maturity group indicates higher entropy production relative to the decelerated and accelerated groups. **b**, Graphs represent the cumulative contribution to entropy production of both forward and reverse transitions and reveal the level of irreversibility in the dynamics of each brain state pair within maturity groups. Nodes represent the 12 brain states, edges indicate pairwise contributions to entropy production, where increasing thickness and darkness represent greater contribution, therefore larger irreversibility. Graphs indicate higher entropy production in the average maturity group, apparent in the thicker and darker lines. **c**, The effect of maturity is analysed by averaging across age groups within distinct maturity groups. **d**, Bar plots illustrate the average entropy production and respective 95% confidence interval for each maturity group, calculated via inverse-variance weighting. Using two-sample z-tests, we compared the average group to both decelerated (p=0.0028) and accelerated (p=0.0082) groups, observing significant differences in each comparison with large effect sizes (Hedges' g).

Chronological age and maturity related entropy production

To study the interplaying effect of chronological age and maturity, we divided adolescents into
two age brackets, namely 12-year-olds (N=31, mean age=12.48y, SD=0.27y) and 13-year-olds
(N=30, mean age=13.58y, SD=0.29y), and within these brackets, we further assorted
participants into three distinct maturity groups, forming six groups, referred to as bins, in total.

In the 12-year-old group, we identified three maturity subgroups: decelerated maturity (CA-
$1.5 \leq BA < CA-0.5$; N=10, mean age=12.41y, SD=0.26y, mean BA-CA=-0.86, SD=0.21);
average maturity ($CA-0.5 \leq BA \leq CA+0.5$; N=11, mean age=12.48y, SD=0.28y, mean BA-
CA=0.04, SD=0.31); and accelerated maturity ($CA+1.5 \geq BA > CA+0.5$; N=10, mean
age=12.54y, SD=0.27y, mean BA-CA=0.91, SD=0.34). Similarly, in the 13-year-old group,
we established three maturity subgroups: decelerated maturity ($CA-1.5 \leq BA < CA-0.5$; N=11,
mean age=13.49y, SD=0.24y, mean BA-CA=-0.97, SD=0.27); average maturity (CA-
$0.5 \leq BA \leq CA+0.5$; N=10, mean age=13.52y, SD=0.31y, mean BA-CA=0.03, SD=0.28); and
accelerated maturity ($CA+1.5 \geq BA > CA+0.5$; N=9, mean age=13.76y, SD=0.26y, mean BA-
CA=0.85, SD=0.28). Participant bins are illustrated both in *Fig. 2e* and in *Fig. 5c*.

Fig. 5a and *5b* offer detailed insight into brain state dynamics within each bin. As it is described
above, with respect to *Fig. 3a*, logarithmic irreversibility transition matrices in *Fig. 5a* illustrate
a contribution to entropy production of brain state transitions, in this case within decelerated,
average, and accelerated maturity groups within age groups. In both age brackets, the larger
hue and intensity variation in the matrix of the average maturity group indicates higher entropy
production relative to the decelerated and accelerated groups. Graphs in *Fig. 5b* represent the
total contribution of both forward and reverse transitions to entropy production (see the
description related to *Fig. 3b* above), indicating higher entropy production in the average
maturity group in both age groups, apparent in the thicker and darker edges of the graph
compared to that of the decelerated and accelerated groups.

We utilised inverse-variance weighting to calculate group averages and variances of entropy
production, minimising the biassing effect that variance introduces to the weighted arithmetic
mean. Additionally, we determined the 95% confidence intervals for group-level entropy
production based on the weighted group average. As it is suggested in *Fig. 5d*, the average
maturity group expresses higher average entropy production at the group-level than decelerated
and accelerated groups in the same chronological age category. We performed two-sample z-
tests to determine the statistical significance of differences. Two-sample z-tests, accompanied
by Hedges' g effect size estimations, confirmed statistically significant differences among the
12-year-old maturity bins (see *Fig. 5d*); we compared the average entropy production of the
average maturity (mean=0.02823 bit, 95% CI: 0.02374-0.03271) bin to both decelerated
(mean=0.02032 bit, 95% CI: 0.01633-0.02431; *p*=0.0098, *g*=1.120) and accelerated
(mean=0.02145 bit, 95% CI: 0.01750-0.02540; *p*=0.0261, *g*=0.963) bins, revealing significant
differences and large effect sizes in these comparisons. Performing two-sample z-tests on the
maturity bins of the 13-year-olds did not yield a significant result (*p*>0.05 in all comparisons).

Effect size measurements revealed a medium effect size (g=0.61) between the average maturity
(mean entropy production=0.02728 bit, 95% CI: 0.02277-0.03179) and decelerated (mean
entropy production=0.02299 bit, 95% CI: 0.01899-0.02700) bins, and a large effect size in the
comparison of average versus accelerated (mean entropy production=0.02257 bit, 95% CI:
0.01813-0.02701; g=0.67) bins.

Our results show that maturity strongly influences entropy production among 12-year-old
participants. While we did not find significant differences between the maturity groups in the
13-year-old age bracket, the higher group-level entropy production in the average maturity
group is still present in the logarithmic irreversibility transition matrices (*Fig. 5a*), in the graphs
of *Fig. 5b*, and in the effect size calculations, indicating a persistent pattern of increased entropy
production associated with average maturity.

Fig. 5 Chronological age and maturity related entropy production.

a, Logarithmic irreversibility transition matrices illustrate two distinct, non-symmetric values in each brain state pair, derived from the log ratio of the forward and reverse transition probabilities. The matrices inform about the preference for directionality in brain state switching. The extent of deviation from 0 reflects the strength of directional preference (forward transitions in red, backward transitions in blue), indicating a higher level of irreversibility and hierarchical organisation in the brain. The hue and intensity diversity in the matrices in the average maturity groups across both age groups indicates higher entropy production values compared to decelerated and accelerated maturity groups. **b**, Graphs represent the cumulative contribution to entropy production of both forward and reverse transitions and reveal the level of irreversibility in the dynamics of each brain state pair within age groups. Nodes represent the 12 brain states, edges indicate pairwise contributions to

entropy production, where increasing thickness and darkness represent greater contribution, therefore larger irreversibility. Graphs indicate higher entropy production in both average maturity groups, apparent in the thicker and darker lines. **c**, The effect of maturity is analysed by averaging across age groups within distinct maturity groups. **d**, Bar plots illustrate the average entropy production and respective 95% confidence interval for each maturity group, calculated via inverse-variance weighting. Using two-sample z-tests, we compared the average group to both decelerated (p=0.0098) and accelerated (p=0.0261) groups, observing significant differences in each comparison with large effect sizes (Hedges' g) in 12-year-olds, however, the analysis of 13-year-olds did not yield significant results.

Discussion

A novel question was addressed regarding the role of adolescent maturation in brain
development. While individual differences in maturity among teenagers are well-established,
it has been unclear whether there is a difference in brain development between groups of
adolescents with diverse maturity levels. The introduction of skeletal age into research now
allows for conclusive answers to this question in a field obscured by methodology issues
around maturity assessments. Our results suggest that an average maturational trajectory might
be optimal in terms of hierarchical brain organisation. The significance of this study stems from
the possibility that developmental plasticity may not fully compensate for accelerated or
decelerated timelines, and a heightened risk of behavioural problems and psychiatric disorders
may arise from such alterations.

To address how well adolescents with different maturational pace can navigate pubertal
goldilocks, we relied on skeletal age as a proxy for pubertal progress and resting-state EEG-
derived entropy production as a gauge of hierarchical brain organisation. As illustrated in *Fig.*
2, we employed the strategy of selecting an equal number of participants with accelerated,
average, and decelerated maturity levels. We have also completed our sample by a group of
emerging adult participants who were not selected by maturity levels since bone age
assessments are not informative at this age. The dissociation between biological age and
chronological age illustrated in *Fig. 2b* allowed us to independently analyse the impact of age
and maturity on the extent of resting-state EEG-derived entropy production.

First, we analysed the relationship between chronological age and entropy production by
averaging across the three maturity groups within the 12 and 13-year-old participant groups
(*Fig. 2c* and *Fig. 3c*) and comparing them to the emerging adult group. Despite a noticeable
increasing trend in entropy production with age (*Fig. 3d*), chronological age in itself did not
have a statistically significant effect on entropy production. The similar logarithmic
irreversibility transition matrices of the age-groups (*Fig. 3a*), and the relatively similar
dispersion of entropy production among brain state pairs (*Fig. 3b*) support the statistical results.
These findings suggest that the level of hierarchy in the adolescent brain may not be
meaningfully influenced by chronological age alone. It is important to note that we lack
information about the adolescent maturational trajectories of emerging adult participants, while
adolescents have been screened and selected for the maturity groups in an approximately equal
number. Therefore, the distributions of individuals with different maturity levels might be

different between the emerging adults and adolescents (*Fig. 2a*), and their direct comparison
should be approached with caution.

To investigate the influence of maturity on entropy production, we categorised adolescent
participants into three distinct groups based on their level of maturity (decelerated, average,
and accelerated maturity groups in *Fig. 2d* and *Fig. 4c.*). Statistical analysis of group averages
(*Fig. 4d*) revealed significantly higher entropy production in the average maturity group with
large effect sizes, pointing at a considerably higher level of hierarchical organisation when
chronological age and bone age align. A closer look at the logarithmic irreversibility transition
matrices (*Fig. 4a*) illustrating the strength of directional preference in switching among the 12
states provides further insight into the role of maturity in hierarchical organisation. The average
maturity group has a relatively higher number of brain state pairs among which a strong
directionality is established. The contribution of brain state pairs also reflects an elevated level
of hierarchy in the average maturity group (*Fig. 4b*). The dynamics between brain states
underlying the entropy production levels in each group indicate well-established causal
interactions, and hierarchical brain organisation in the average maturity group, while these
relationships seem relatively less dense in the decelerated and accelerated groups. These
findings indicate that physical maturity is inherently linked to the shaping of irreversible, causal
interactions within the cortex[69] during adolescence. Furthermore, it suggests that the optimal
maturational pace for fostering these relationships is one in which chronological and biological
age are closely matched.

To see whether the maturity related effect is stable across different chronological ages, we
compared the three maturity groups within the 12- and 13-year-old age groups (*Fig. 2d* and
Fig. 5c). The analysis revealed that the average maturity group displays significantly higher
entropy production compared to both the decelerated and accelerated maturity groups among
the 12-year-olds (*Fig. 5d*). While this tendency is still apparent in the 13-year-old group,
statistically significant differences between maturity groups were not detected. However, the
logarithmic irreversibility transition matrices of each bin (*Fig. 5a*) make a case for the optimal
pace of maturation even in the absence of statistical significance, as the average maturity
groups in both age brackets convey a larger number of brain state pairs where a strong
preference of switching direction is established. The contribution of each brain state pair to
entropy production (*Fig. 5b*) also highlights a larger number of well-established irreversible
interactions in the average maturity groups. Taken together, the statistical results (Fig. 5d) may
suggest that the clear advantage of an on-time maturational pace is limited to a specific
chronological age window with a declining tendency in older age groups, however, this
possibility may need further investigation as the graphical analyses presented in Fig. 5a and 5b
suggest a persistent pattern of increased entropy production associated with average maturity.

To conclude, we would like to re-emphasise that maturation plays a pivotal role in shaping
both brain structure and behaviour. This process ensures that the brain efficiently coordinates
distributed computations across its entirety, allowing not only survival but also optimal
functioning. This efficiency hinges on a hierarchical orchestration of state-dependent, self-
organised brain activity over time. This orchestration facilitates near-optimal information

processing and transfer while minimising energy consumption. Therefore, a paramount
concern of a comprehensive theory of brain function is the exploration of hierarchy. This entails
unravelling the dynamic and evolving self-organisation of brain states, spanning from stability
to transitions, as they navigate through probabilistic and sometimes chaotic shifts. Here, we
assumed that the inherent hierarchy of spontaneous brain activity might be reflective of the
level of maturation. Harnessing the principles of thermodynamics, we quantified hierarchy by
computing the level of irreversibility in brain signals during resting state conditions.
Specifically, we captured the level of irreversibility through entropy production[69,71,74,76,77] as
computed through a sophisticated clustering strategy. Note that resting states cover a rich range
of the dynamical repertoire of the brain and thus those are ideal to characterise the most general
and wide variety of interaction across the whole brain, i.e., the underlying hierarchical
orchestration.

In terms of limitations, the current project is confined to the examination of adolescent females,
a limitation that might be apparent. However, a selection was essential when aiming to
distinctly disentangle the impacts of maturation and chronological age on brain development.
Involving both sexes would have made it difficult to carry out the detailed analysis within the
narrow temporal windows of biological and chronological age depicted in *Fig. 2b*. As male
pubertal onset occurs approximately 1-1.5 years later than that of females[32,78], this analysis
would necessitate a considerably larger sample size. We opted for inclusion of females since
menarche age provides valuable supplementary data affirming the accuracy of bone age
assessments (see *Methods*). Given the conclusive findings with females, underscoring a
substantial role of biological age in brain development, it is plausible to assume a comparable
effect might manifest in males. Indeed, we would expect that maturity is a relevant factor in
the general brain development of males as well; however, owing to the distinct gonadal
hormone levels governing female and male developmental trajectories[21,79], disparities are also
anticipated, including the timing of the impact of maturity levels.

An additional limitation of our study pertains to its cross-sectional design within a relatively
narrow age spectrum, which prevents an exploration of potential long-term consequences due
to variations in pubertal pace. The evolutionarily expanded heteromodal association cortices,
mentioned in the introduction, hosting a range of higher order functions[11,65,67,68,80] may have a
potential to compensate when confronted with divergent maturational paces. Nevertheless, the
consequences of accelerated or delayed development on relevant and precisely timed
adolescent brain alterations such as cortical volume reduction[81,82], grey matter thinning[17,83],
and synaptic pruning[18,45] remain unknown. The human brain could conceivably adapt to
distinct maturational tempos, and with only transient disparities, developmental trajectories
might still converge within the normative range. While the distinctions in hierarchical brain
organisation uncovered in our study may be transient, even these potentially short-term
developmental discrepancies are essential to discuss since they could temporarily place the
child outside the typical range and cause heightened stress levels. Subsequent investigations
must undoubtedly address the longitudinal trajectories of brain development within cohorts
characterised by diverse rates of maturation. It needs to be considered, however, that any such
study faces a serious trade-off: observing long-term alterations due to accelerated puberty onset

is more likely with age, but direct assessment of pubertal timing becomes less reliable with age. Although a long-term follow-up between puberty onset and adulthood seems necessary to complement our work, in the current study we have chosen to work with a cross-sectional sample to get a first glimpse on the impact of different maturational tempos on brain development.

With respect to the broader relevance of our findings, although the participants enrolled in the present study were deliberately chosen to fall within a normative range of maturational speed and were further screened for favourable socioeconomic circumstances, as well as the absence of developmental or neurological disorders, the findings bear more general implications, especially for clinical domains. We found an advantage of on-time maturation with respect to hierarchical brain organisation within our non-clinical cohort, indicating that significant deviations towards either accelerated or delayed maturational speeds might amplify disparities, leading developmental trajectories into the clinical spectrum. The acceleration of pubertal development is a growing concern today, particularly given the surge in medical referrals for young girls experiencing precocious puberty globally, with some instances manifesting as early as 6 or 7 years old[84–95]. Premature puberty carries the risk of compromised adult stature, psychosocial challenges, and potential health complications later in life[44,96,97]. The sudden peak in the lower left segment of the puberty onset age distribution curve (akin to that depicted in *Fig. 2a*), corresponding to instances of early puberty onset, suggests that there may be significant implications for the broader population involving a rapid secular trend. The possibility that developmental plasticity may not fully compensate for these changes is highly relevant. Secular trends in pubertal onset age persist across both developing and modern societies. While advancements in living conditions primarily account for trends in developing countries today[13–15,98,99], negative anthropogenic influences, such as endocrine disrupting chemicals, have shaped the trend in developed nations in the last decades[98]. The recent pandemic has emerged as a novel potential factor influencing the age of pubertal onset, and it is essential to further investigate its impact on the general population.

In present times, within the most affluent nations, important developmental events, such as birth and puberty can be artificially timed, inducing intraspecies heterochrony which denotes variations in the timing of developmental events. Genetic-induced heterochrony introduces natural variability throughout evolution[100–102]. It remains uncertain whether anthropogenic-induced heterochrony (e.g., accelerated trends in pubertal timing, or artificially delayed puberty) will yield compromised mental functioning and health, or foster adaptive diversity in successive generations. Therefore, our study seems to be very timely, and can be considered as a first step towards investigating the potential outcomes of "anthropogenic heterochrony". Mental health problems have surged following successive lockdowns, demonstrating the psychological toll on adolescents worldwide[57–60]. The findings of our study will aid in devising interventions more attuned to developmental contexts, thus lowering the risk of developing disabling psychopathologies in future generations.

Methods

Participants

The study is limited to investigating adolescent and emerging adult females, a deliberate choice
made to disentangle the impacts of maturation and chronological age on brain development.
Inclusion of both genders would have hindered in-depth analysis within the narrow temporal
windows of biological and chronological age depicted in *Fig. 2b*. Given the delayed male
pubertal onset[32,78], a significantly larger sample would be required. We opted for inclusion of
females since menarche age provides valuable supplementary data affirming the accuracy of
bone age assessments in adolescents.

Participants were recruited via school contacts and online advertisements from Budapest,
Hungary. Parents and /or participants provided demographic, educational, and medical history
details. One participant with attention deficit disorder was excluded.

In order to determine the optimal sample size, we performed power calculations for two-sample
z-test for means. A sample size of $N \geq 25$ was found to be required for a power of at least 0.8
to detect a 0.8 effect at $p<0.05$. We aimed for a sample size N=30 in the adolescent and
emerging adult chronological age groups. This yielded 20 subjects in the adolescent maturity
groups with a power of at least 0.7. Ultrasonic bone age screening was conducted, and
adolescent participants were categorised into BA/CA-defined bins, ensuring non-overlapping
maturity groups: average (CA-0.5$\leq$BA$\leq$CA+0.5), accelerated (CA+1.5$\geq$BA>CA+0.5), and
decelerated (CA-1.5$\leq$BA<CA-0.5). A pilot study revealed that around 20% of adolescents
exhibited accelerated or decelerated maturation. Thus, 200 participants were invited (100 per
CA group) for BA screening, aiming for $\geq$10 participants per bin. We excluded extreme cases
(BA>CA+1.5, BA<CA-1.5) to avoid endocrinological complexities. This initial process
resulted in a total of 65 participants between 12 and 14 years of age. However, during EEG
preprocessing (see *Procedure*) 9 recordings were excluded from the study due to excessive
noise. To compensate for this, we conducted further, targeted bone age screenings, and
recorded EEG data of participants who fit into the aforementioned bin criteria, aiming to
maintain a nearly equal distribution of participants across bins. From this additional
recruitment, five new EEG recordings were successfully added. Thus, 61 girls aged 12 to 14
were included in the final cohort as depicted in *Fig. 2b*. The cohort of emerging adult
participants consisted of 30 subjects. EEG data of 26 participants was included after the
preprocessing procedure, as 4 recordings were removed due to noise.

The percentage of postmenarche participants in both age groups was lowest in the decelerated
groups (see the Open Science Framework link under *Data availability*), suggesting that bone
age is indeed a reliable gauge of maturity in adolescence. Mean menarche age of adolescent
participants was 12.06y (SD=0.72). 27 of 61 participants did not provide data on menarche as
they have not reached that stage yet. 88.52% of mothers and 83.61% of fathers of adolescent
participants had a university degree demonstrating high socioeconomic status among the

participating families. The EEG data collection was part of a large-scale study, within which each adolescent participant was also administered the Wechsler Intelligence Scale for Children, 4th edition (WISC-IV)[34], carried out a fine motor task[36], and a Stroop test[35].

The Hungarian United Ethical Review Committee for Research in Psychology (EPKEB) approved the study (reference number 2017/84). Written informed consent was obtained from all subjects and their parents. Participants were given gift cards (approx. EUR 15 value each) for their attendance. All research described in this paper was performed in accordance with the Declaration of Helsinki.

Procedure

Bone age assessment

Body measurements were taken according to the protocol laid down in the International Biological Programme[103] using standard instruments. Skeletal maturity assessment began with an anthropometric procedure that includes the measurement of full body and sitting height (GPM Anthropometer, DKSH, Switzerland Ltd, Zurich, Switzerland), as well as body weight (Seca digital scale). Ultrasonic bone age estimation was conducted using the Sunlight BonAge System (Sunlight Medical Ltd, Tel Aviv, Israel), scanning the speed of sound (SOS) between the distal radius and ulna epiphysis[104]. This bone structure reflects the level of maturation as well as it changes significantly during stages of physical growth. The BonAge device calculates bone age in years and months using software that accounts for ethnicity and gender. Using ultrasound for skeletal maturity assessment is safe for the subjects as opposed to ionising radiation-based techniques, while results obtained from these two types of procedure are highly correlated[40].

Bone age assessment was performed on the left hand and wrist of participants in each case. Participants were instructed to position their hands between the transducers on the unit's armrest. The device then attached to the wrist, applying pressure of approximately 500g. Transducers transmitted 750 kHz frequency ultrasound through the wrist to measure SOS. The transducer on the ulnar side emits ultrasound while the other transducer acts as a receiver. Measurement protocol was repeated five times for accuracy, each measurement lasting about 20 seconds. Measurements were completed either at the high schools of participants, or at the Research Centre for Sport Physiology at the University of Physical Education, Budapest. The entire procedure took 5-10 minutes per person. Trained assistants conducted the measurements, and a biological anthropologist has overseen the measurement procedure and data analysis.

Bone age assessment results are considered valid for 3 months. Subjects were invited for the resting state electroencephalography (EEG) recording session within this timeframe. If a subject participated in laboratory testing 3 months after bone age assessment, the procedure was repeated.

Resting state EEG data recording

Continuous EEG data was recorded using the HydroCel GSN 130, a 128-channel high density
EEG (HD-EEG) system (Electrical Geodesics, Inc., Canada), with the Net Station Acquisition
software (version 5.4) in a windowless, air-conditioned room insulated from external noise and
electromagnetic interference. Before recording, participants were asked to remove all
electronic devices, e.g. phones and smartwatches from their pockets and bodies. Following a
brief guided relaxation, we collected two five-minute (300 second) long resting state recordings
at a 1,000 Hz frequency sampling rate. We aimed to keep impedances below 5 KΩ on all
channels. EEG data recording took place at the Laboratory for Psychological Research,
Pázmány Péter Catholic University, Budapest.

In the first 5-minute long recording segment, participants were instructed to keep their eyes
open, blink minimally and to maintain posture to minimise artefacts generated by muscle
movement[105]. During the second 5 minutes of recording, participants followed the same
protocol, but were instructed to keep their eyes closed. The two conditions are separated since
they exhibit different characteristics, namely a mismatch in topography and activity levels[106].
In the present study, we performed all statistical analyses on closed-eye recordings.

EEG data preprocessing

EEG preprocessing was performed in MATLAB (The MathWorks, Inc.; version: R2021a
Update 4), using the EEGLAB toolbox[107] (version 2021.1). First, we applied a high-pass filter
at 1 Hz (transition bandwidth 1 Hz; passband edge 1 Hz; cutoff frequency -6 dB 0.5 Hz; zero-
phase non-causal filter) to eliminate low frequency noise and direct-current offset, and a 40 Hz
low-pass filter (transition bandwidth 10 Hz; passband edge 40 Hz; cutoff frequency -6 dB 45
Hz; zero-phase non-causal filter) to remove 50 Hz line noise and high frequency components,
using EEGLAB's basic finite impulse response (FIR; version 2.4) filter. Channel locations
were then imported into EEGLAB, the channel layout map was supplied by Electrical
Geodesics, Inc. Following filtering steps, noisy channels, determined by two independent raters
via visual inspection, were excluded manually. As last steps, we removed noisy epochs from
the recordings using the Clean Rawdata (version 2.4) plugin, then re-referenced data to average
reference. The same preprocessing procedure was implemented for all recordings.

Data Analysis

Leading Eigenvector Dynamics Analysis

537   After preprocessing, Leading Eigenvector Dynamics Analysis (LEiDA)[75], adapted to EEG data
538   by our group, was employed to analyse dynamic functional connectivity[69,73] in the developing
539   brain. The steps described below were performed on individual EEG recordings and were
540   uniform in all cases.

First, preprocessed EEG recordings were filtered with a 6th order Butterworth bandpass filter for alpha band (8-12 Hz) without introducing a phase shift, then downsampled to 100 Hz (intervals of 10 ms). Subsequently, signals were detrended and demeaned, followed by a Hilbert-transformation to obtain the phase of all channels at all timepoints t. The nominal recording duration of 300 s yielded a temporal sequence of up to 30,000 phase vectors per subject. However, the EEGLAB toolbox typically excised small parts of the sequence, so that average sequence length was 28,500 vectors (SD=1,600). During preprocessing, we observed a minority of electrodes with interrupted EEG signal during some or all of the recording ("bad channels", range 0 to 12; see *EEG data preprocessing*). To obtain full-size (128x128) coherence matrices at all timepoints and for every subject, we developed a two-step approach termed "patching". In the first step, we formed group-level lists of "bad channels" by combining noisy channels from all subjects in a group, specifically, the 6 bins of adolescent participants (depicted in *Fig. 2e*) and the group of adult subjects. For n remaining "good channels" shared by all subjects of the group, instantaneous phase coherence $PC_{xy,t}$ was computed at every time point for each electrode pair as

$$PC_{xy,t} = cos\left(\phi(E_{x,t}) - \phi(E_{y,t})\right) \tag{1}$$

where $\Phi(E_{x,t})$ and $\Phi(E_{y,t})$ denote the instantaneous phase of electrode x and y, respectively, at each time point t. This resulted in a temporal sequence of n x n phase coherence matrices, sampled at 10 millisecond intervals, for each subject. The leading eigenvector (*1xn*) of each matrix was used to summarise the instantaneous pattern of phase differences between electrodes at every time point. Then, these vectors were clustered with Matlab's built-in *k*-means clustering algorithm, assigning a cluster label to all timepoints in all subjects. In the second step, we patched the missing phase values of "bad channels" in all subjects. To this end, we replaced the missing phase values of a "bad channel" x at a time point t assigned to a particular cluster by randomly sampling a phase value from the ensemble of values recorded in other subjects of the same bin at the same electrode x and at time points assigned to the same cluster. Performing this procedure for all groups, subjects and "bad channels" permitted us to reconstruct full-size phase vectors (1x128) and full-size phase difference matrices (128x128). Full-size leading eigenvectors (1x128) were obtained from full-size phase difference matrices. Note that eigenvectors had the same dimension as the original phase vectors, but represented relative rather than absolute phase.

To identify patterns of phase differences that were common to all subjects – also referred to here as "brain states" and depicted in *Fig. 6* –, the leading eigenvectors at all timepoints and from all subjects were pooled and clustered via a modified *k*-means algorithm[108] such as to obtain polarity invariant topographical maps. Through the modified *k*-means clustering algorithm, each vector is assigned to one of k clusters (or brain states) and is labelled accordingly (see *Fig. 6*), allowing us to represent the continuous EEG activity as a trajectory of, and transition between, brain states. LEiDA was carried out in Matlab (version: 2021a). While the results presented here are based on 12 clusters, analyses were also conducted using 6, 8, and 10 clusters and 12 clusters specific to each unique group (see the Open Science Framework link under *Data availability*). The results of these additional analyses were

consistent with those presented in the *Results* section. We chose to present the results based on
12 brain states since it provides the most detailed description of the data.

A representative example for a sequence of successive patterns of phase differences (brain
states) is shown in *Fig. 7a*. The average duration of brain states was 125 ms, with a standard
deviation of ±0.4 ms over groups (range 12 ms to 13 ms) and ±1.5 ms over subjects (range 100
ms to 170 ms). However, the durations of individual brain states were quite variable (range 50
ms to 500 ms), with the average standard deviation over subjects being ±9 ms (range: 7 ms to
14 ms).

Entropy production calculation

To quantify asymmetry in the temporal dynamic of state space trajectories, we transformed the
temporal sequence of brain states (*Fig. 7a*) of each subject into a table of transition counts N_{ij}
by removing diagonal entries (remaining in the same brain state) and by counting the observed
number of forward and reverse transitions between pairs of different brain states (i,j). A
representative example table is shown in *Fig. 7b*. To avoid singularities in the subsequent
computation of entropy production, all counts were incremented by one, $N'_{ij}=N_{ij}+1$. Any effect
of this change was nullified by the noise floor correction (see below). The corresponding
transition probabilities were then obtained as $P_{ij}=N'_{ij}/\Sigma_{ij} N'_{ij}$. To quantify the asymmetry in the
pattern of state transitions, deemed indicative of the level of hierarchical organisation, we
computed entropy production (also termed relative entropy[70]) for each subject as the Kullback-
Leibler (KL) divergence:

602
$$H = D_{KL} = \sum_{i,j} P_{ij} \, log \frac{P_{ij}}{P_{ji}} \qquad (2)$$

where D_{KL} and H denote the total entropy production (i.e., the asymmetry between forward and
reverse transition probabilities in brain state switching), P_{ij} denotes the transition probability
from brain state i to brain state j, while P_{ji} expresses the probability of switching from brain
state j to brain state i.

To estimate the variance of the observed entropy values, we generated 1,000 synthetic
transition trajectories as Markov chains from the transition probability table observed for each
subject via MATLAB's two built-in functions, one of which creates discrete-time Markov
chains and the other simulates Markov chain state walks. Synthetic trajectories had the same
length as observed trajectories. Converting synthetic chains into entropy production values
with *Eq. 2*, produced a distribution of a 1,000 entropy production values for each subject.

As the finite length of trajectories inflates entropy production values, we additionally
performed a noise floor correction[70]. To this end, we synthesised a further 1,000 synthetic
trajectories of the same length from a symmetric transition table (obtained by averaging the
observed table with its transpose). Although perfectly symmetric transition probabilities should
not produce any entropy, the resulting value distribution was positive, revealing inflation due
to finite trajectory length. To obtain corrected entropy production values, we subtracted the
mean symmetric-table entropy from the all asymmetric-table entropy values. From the

distribution of corrected values, we obtained a mean and estimated variance of entropy
production for every subject. Note that our procedure kept intact the temporal order of
trajectories, taking subject-specific brain dynamics into account, improving on the simpler
approach of Lynn and colleagues[70]. All calculations were performed in Matlab (version:
2021a).

Calculating group averages and confidence intervals

625 Group averages and variances of entropy production were computed by inverse-variance
626 weighting. This was done to address the subject heterogeneity and to attenuate the influence of
627 subjects with high variance on the weighted arithmetic mean. The following formulas were
628 used:
629

$$\bar{H} = \frac{\sum_i H_i / \sigma_i^2}{\sum_i 1/ \sigma_i^2} \qquad (3)$$

$$var\,(\bar{H}) = \frac{1}{\sum_i 1/ \sigma_i^2} \qquad (4)$$

633
634 where $\bar{H}$ is the inverse variance weighted group average entropy production, H_i denotes the
635 mean observed entropy production for subject i, $\sigma^i{}_2$ denotes the observed variance for subject
636 i, and $var(\bar{H})$ denotes the inverse variance weighted group variance. 95% confidence intervals
637 for group-level mean entropy production was also computed based on $\pm 1.96*\sqrt{(var(\bar{H}))}$. Group
638 averages and confidence intervals were calculated in Matlab (version: 2021a).

Statistics

639 Statistical assessment of group differences was carried out using two-sample z-tests[109], using
640 the following formula:

$$Z = \frac{\bar{H}_1 + \bar{H}_2}{\sqrt{(var(\bar{H}_1) + var(\bar{H}_2))}} \qquad (5)$$

642 The significance level (p) in the two-sample z-test is determined as in the following:
643

$$p = 2\Phi(-|Z|) \qquad (6)$$

645
646 where Φ is the standard normal cumulative distribution function.
647
648 Results obtained from two-sample z-tests were reported with Hedges' g effect size estimations.
649 All statistical tests were carried out in Microsoft Excel Professional (version 2021).

Fig. 6 Brain state topographies.

After initial preprocessing, EEG signals were analysed via Leading Eigenvector Dynamics Analysis (LEiDA). First, Hilbert transform was applied to each recording to acquire the phase of each electrode. Then, the instantaneous phase coherence between all electrode pairs was computed, yielding a 128x128 phase distance matrix for each time point t. Leading eigenvectors were extracted from the matrices via eigenvector decomposition. Following LEiDA, leading eigenvectors of all subjects were pooled, then clustered via modified k-means clustering to obtain topographically distinct connectivity patterns, which we refer to here as "brain states". As leading eigenvectors from all subjects are combined, the twelve activity patterns are rendered uniform in each group. Brain states are ranked according to their dominance duration, expressed in percentages accompanying rank labels, with brain state number 01 accounting for the largest percentage of total activity, and brain state number 12 the smallest. The colour scale denotes cosine similarity, with similarly coloured dots indicating electrodes with congruent signal phases. Accordingly, values of 1 (dark red) and -1 (deep blue) represent similar and dissimilar (opposite) phases, respectively. Note that electrodes with similar and dissimilar phases form two spatially contiguous subsets.

Fig. 7 Transitions between brain states.

a, Brain states numbered 1 to 12 are represented here by coloured disks. Only the initial 10s of a recording lasting 284 s are shown, specifically, the initial 1,000 leading eigenvectors (observed at 10 ms intervals) in a sequence of 28,400 eigenvectors. In this particular example, the average duration of brain states was 98 ms (i.e., 9.8 consecutive eigenvectors were assigned to the same state, on average). **b,** Number of observed transitions between different states in the entire sequence of 28,400 states. Previous states are represented by rows, next states by columns. Table entries indicate how often each particular transition was observed. Entropy production quantifies the degree of asymmetry in this matrix.

Data availability

650 Demographic and anthropological datasets, and EEG analyses are available at the Open
651 Science Framework (OSF) platform at this address:
652 https://osf.io/eqmwd/?view_only=d82d42bcb7a1481ca3ad33e721179523

Acknowledgements

The authors thank T. Jáger for generating the figures. K. Utczás and Z. Tróznai for bone age
data collection and analysis, G. Oláh for organising the participant cohort, L. J. Fehér for
additional EEG data collection. We also thank the generous and amazing parents, adolescents
and schools who participated in this project.

The project was supported by the National Research, Development and Innovation Office of
Hungary (Grant K-134370 to I.K.) and by the Hungarian Research Network (HUN-REN-
ELTE-PPKE Adolescent Development Research Group). The funders had no role in study
design, data collection and analysis, decision to publish or preparation of the manuscript.

Author Contributions

661 I.K. and F.G. conceptualised and designed the experiments; S.H. carried out data collection;
662 M.L.K. and G.D. contributed theoretical analysis tools; M.C.M., J.B. and F.G. contributed
663 analytical software; S.H., M.C.M., G.B., and J.B. carried out data analysis; F.G. carried out the
664 statistical analysis; S.H. and I.K. drafted the manuscript. All authors discussed the results,
665 contributed to the text, and approved the final version of the manuscript for submission.

Competing Interests statement

666   The authors declare no competing interests.

References

1. Boyce, W. T., Sokolowski, M. B. & Robinson, G. E. Genes and environments, development and time. *Proceedings of the National Academy of Sciences* **117**, 23235–23241 (2020).
2. Roubinov, D., Meaney, M. J. & Boyce, W. T. Change of pace: How developmental tempo varies to accommodate failed provision of early needs. *Neuroscience & Biobehavioral Reviews* **131**, 120–134 (2021).
3. Spear, L. Adolescent Neurodevelopment. *The Journal ofAadolescent Health: official publication of the Society for Adolescent Medicine* **52**, S7–S13 (2013).
4. Mewton, L. & Squeglia, L. Neurodevelopmental Profiles in Adolescence: Leveraging Data From the Landmark Adolescent Brain Cognitive Development Study. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging* **7**, 343–345 (2022).
5. Tyborowska, A. *et al.* Early-life and pubertal stress differentially modulate grey matter development in human adolescents. *Sci Rep* **8**, 9201 (2018).
6. Gotlib, I. H. *et al.* Effects of the COVID-19 Pandemic on Mental Health and Brain Maturation in Adolescents: Implications for Analyzing Longitudinal Data. *Biological Psychiatry: Global Open Science* **0**, (2022).
7. Tooley, U. A., Bassett, D. S. & Mackey, A. P. Environmental influences on the pace of brain development. *Nat Rev Neurosci* **22**, 372–384 (2021).
8. Bethlehem, R. a. I. *et al.* Brain charts for the human lifespan. *Nature* **604**, 525–533 (2022).
9. Johnson, M. B. & Stevens, B. Pruning hypothesis comes of age. *Nature* **554**, 438–439 (2018).
10. Sydnor, V. J. *et al.* Intrinsic activity development unfolds along a sensorimotor–association cortical axis in youth. *Nat Neurosci* **26**, 638–649 (2023).
11. Sydnor, V. J. *et al.* Neurodevelopment of the association cortices: Patterns, mechanisms, and implications for psychopathology. *Neuron* **109**, 2820–2846 (2021).
12. Herman-Giddens, M. E. *et al.* Secondary Sexual Characteristics and Menses in Young Girls Seen in Office Practice: A Study from the Pediatric Research in Office Settings Network. *Pediatrics* **99**, 505–512 (1997).
13. Parent, A.-S. *et al.* The timing of normal puberty and the age limits of sexual precocity: variations around the world, secular trends, and changes after migration. *Endocr Rev* **24**, 668–693 (2003).
14. Eckert-Lind, C. *et al.* Worldwide Secular Trends in Age at Pubertal Onset Assessed by Breast Development Among Girls. *JAMA Pediatr* **174**, e195881 (2020).
15. Lam, B. Y. H. *et al.* MC3R links nutritional state to childhood growth and the timing of puberty. *Nature* **599**, 436–441 (2021).
16. Neale, J., Pais, S. M. A., Nicholls, D., Chapman, S. & Hudson, L. D. What Are the Effects of Restrictive Eating Disorders on Growth and Puberty and Are Effects Permanent? A Systematic Review and Meta-Analysis. *Journal of Adolescent Health* **66**, 144–156 (2020).
17. Vijayakumar, N. *et al.* A longitudinal analysis of puberty-related cortical development. *NeuroImage* **228**, 117684 (2021).
18. Campbell, I. G., Grimm, K. J., de Bie, E. & Feinberg, I. Sex, puberty, and the timing of

sleep EEG measured adolescent brain maturation. *Proceedings of the National Academy of Sciences* **109**, 5740–5743 (2012).

19. Graber, J. A. Pubertal timing and the development of psychopathology in adolescence and beyond. *Horm Behav* **64**, 262–269 (2013).

20. Marín, O. Developmental timing and critical windows for the treatment of psychiatric disorders. *Nat Med* **22**, 1229–1238 (2016).

21. Blakemore, S.-J., Burnett, S. & Dahl, R. E. The role of puberty in the developing adolescent brain. *Hum Brain Mapp* **31**, 926–933 (2010).

22. Juraska, J. M. & Willing, J. Pubertal onset as a critical transition for neural development and cognition. *Brain Research* **1654**, 87–94 (2017).

23. Foulkes, L. & Blakemore, S.-J. Studying individual differences in human adolescent brain development. *Nature Neuroscience* **21**, 315–323 (2018).

24. Mills, K. L. *et al.* Inter-individual variability in structural brain development from late childhood to young adulthood. *NeuroImage* **242**, 118450 (2021).

25. Fuhrmann, D., Knoll, L. J. & Blakemore, S.-J. Adolescence as a Sensitive Period of Brain Development. *Trends Cogn Sci* **19**, 558–566 (2015).

26. Rosenberg, M. D., Casey, B. J. & Holmes, A. J. Prediction complements explanation in understanding the developing brain. *Nature Communications* **9**, 589 (2018).

27. Tanner, J. M. Growth and maturation during adolescence. *Nutr Rev* **39**, 43–55 (1981).

28. Dorn, L. D. & Biro, F. M. Puberty and Its Measurement: A Decade in Review. *Journal of Research on Adolescence* **21**, 180–195 (2011).

29. Desmangles, J.-C., Lappe, J., Lipaczewski, G. & Haynatzki, G. Accuracy of Pubertal Tanner Staging Self-Reporting. *Journal of pediatric endocrinology & metabolism : JPEM* **19**, 213–21 (2006).

30. Ledford, H. Who exactly counts as an adolescent? *Nature* **554**, 429–431 (2018).

31. Roberts, C. Tanner's Puberty Scale: Exploring the historical entanglements of children, scientific photography and sex. *Sexualities* **19**, 328–346 (2016).

32. Worthman, C. M., Dockray, S. & Marceau, K. Puberty and the Evolution of Developmental Science. *J Res Adolesc* **29**, 9–31 (2019).

33. Walker, I. V., Smith, C. R., Davies, J. H., Inskip, H. M. & Baird, J. Methods for determining pubertal status in research studies: literature review and opinions of experts and adolescents. *Journal of Developmental Origins of Health and Disease* **11**, 168–187 (2020).

34. Kovács, I. *et al.* Ultrasonic bone age fractionates cognitive abilities in adolescence. *Sci Rep* **12**, 5311 (2022).

35. Gervan, P. *et al.* Natural variance in executive function components by pubertal timing in neurotypical population of females. Preprint at https://doi.org/10.31234/osf.io/hauxc (2023).

36. Berencsi, A. *et al.* Musical training improves fine motor function in adolescents. *Trends in Neuroscience and Education* **27**, 100176 (2022).

37. Gerván, P. *et al.* Maturation-dependent vulnerability of emotion regulation as a response to COVID-19 related stress in adolescents. *Journal of Pediatric Nursing: Nursing Care of Children and Families* **67**, 132–138 (2022).

38. Vandewalle, S. *et al.* Associations of sex steroids with bone maturation, bone mineral density, bone geometry, and body composition: a cross-sectional study in healthy male adolescents. *J Clin Endocrinol Metab* **99**, E1272-1282 (2014).

39. Proos, L. A., Lönnerholm, T., Jonsson, B. & Tuvemo, T. Can bone age determination provide criteria for growth hormone treatment in adopted girls with early puberty? *Ups J Med Sci* **111**, 117–129 (2006).

40. Utczas, K., Muzsnai, A., Cameron, N., Zsakai, A. & Bodzsar, E. B. A comparison of

skeletal maturity assessed by radiological and ultrasonic methods. *Am J Hum Biol* **29**, (2017).

41. Marceau, K., Ram, N., Houts, R. M., Grimm, K. J. & Susman, E. J. Individual Differences in Boys' and Girls' Timing and Tempo of Puberty: Modeling Development With Nonlinear Growth Models. *Dev Psychol* **47**, 1389–1409 (2011).

42. Palmert, M. R. & Boepple, P. A. Variation in the Timing of Puberty: Clinical Spectrum and Genetic Investigation. *The Journal of Clinical Endocrinology & Metabolism* **86**, 2364–2368 (2001).

43. Brix, N. *et al.* Timing of puberty in boys and girls: A population-based study. *Paediatric and Perinatal Epidemiology* **33**, 70–78 (2019).

44. Hoyt, L. T., Niu, L., Pachucki, M. C. & Chaku, N. Timing of puberty in boys and girls: Implications for population health. *SSM - Population Health* **10**, 100549 (2020).

45. Huttenlocher, P. R. Synaptic density in human frontal cortex - developmental changes and effects of aging. *Brain Res* **163**, 195–205 (1979).

46. Feinberg, I., Thode, H. C., Chugani, H. T. & March, J. D. Gamma distribution model describes maturational curves for delta wave amplitude, cortical metabolic rate and synaptic density. *J Theor Biol* **142**, 149–161 (1990).

47. Sakai, J. How synaptic pruning shapes neural wiring during development and, possibly, in disease. *Proceedings of the National Academy of Sciences* **117**, 16096–16099 (2020).

48. Averbeck, B. B. Pruning recurrent neural networks replicates adolescent changes in working memory and reinforcement learning. *Proceedings of the National Academy of Sciences* **119**, e2121331119 (2022).

49. Rakic, P., Bourgeois, J.-P., Eckenhoff, M. F., Zecevic, N. & Goldman-Rakic, P. S. Concurrent Overproduction of Synapses in Diverse Regions of the Primate Cerebral Cortex. *Science* **232**, 232–235 (1986).

50. Diniz, C. R. A. F. & Crestani, A. P. The times they are a-changin': a proposal on how brain flexibility goes beyond the obvious to include the concepts of "upward" and "downward" to neuroplasticity. *Mol Psychiatry* **28**, 977–992 (2023).

51. Penzes, P., Cahill, M. E., Jones, K. A., VanLeeuwen, J.-E. & Woolfrey, K. M. Dendritic spine pathology in neuropsychiatric disorders. *Nat Neurosci* **14**, 285–293 (2011).

52. Juraska, J. M. & Drzewiecki, C. M. Cortical reorganization during adolescence: What the rat can tell us about the cellular basis. *Developmental Cognitive Neuroscience* **45**, 100857 (2020).

53. Delevich, K., Klinger, M., Okada, N. J. & Wilbrecht, L. Coming of age in the frontal cortex: The role of puberty in cortical maturation. *Seminars in Cell & Developmental Biology* **118**, 64–72 (2021).

54. Feinberg, I. Schizophrenia: caused by a fault in programmed synaptic elimination during adolescence? *J Psychiatr Res* **17**, 319–334 (1982).

55. Keshavan, M., Lizano, P. & Prasad, K. The synaptic pruning hypothesis of schizophrenia: promises and challenges. *World Psychiatry* **19**, 110–111 (2020).

56. Carey, B. For Some Teens, It's Been a Year of Anxiety and Trips to the E.R. *The New York Times* (2021).

57. Peltier, E. & Kwai, I. Suicide and Self-Harm: Bereaved Families Count the Costs of Lockdowns. *The New York Times* (2021).

58. Kids' mental health is in crisis. Here's what psychologists are doing to help. *https://www.apa.org* https://www.apa.org/monitor/2023/01/trends-improving-youth-mental-health.

59. Thorisdottir, I. E. *et al.* Effect of the COVID-19 pandemic on adolescent mental health and substance use up to March, 2022, in Iceland: a repeated, cross-sectional, population-based study. *The Lancet Child & Adolescent Health* **7**, 347–357 (2023).

60. Cochran, G., Cohen, Z. P., Paulus, M. P., Tsuchiyagaito, A. & Kirlic, N. Sustained increase in depression and anxiety among psychiatrically healthy adolescents during late stage COVID-19 pandemic. *Frontiers in Psychiatry* **14**, (2023).

61. Petanjek, Z. *et al.* Extraordinary neoteny of synaptic spines in the human prefrontal cortex. *Proc Natl Acad Sci U S A* **108**, 13281–13286 (2011).

62. Lupien, S. J., McEwen, B. S., Gunnar, M. R. & Heim, C. Effects of stress throughout the lifespan on the brain, behaviour and cognition. *Nat Rev Neurosci* **10**, 434–445 (2009).

63. Dong, H.-M., Margulies, D. S., Zuo, X.-N. & Holmes, A. J. Shifting gradients of macroscale cortical organization mark the transition from childhood to adolescence. *Proc Natl Acad Sci U S A* **118**, e2024448118 (2021).

64. Baum, G. L. *et al.* Development of structure–function coupling in human brain networks during youth. *Proceedings of the National Academy of Sciences* **117**, 771–778 (2020).

65. Buckner, R. L. & Krienen, F. M. The evolution of distributed association networks in the human brain. *Trends Cogn Sci* **17**, 648–665 (2013).

66. Ahmad, S. *et al.* Multifaceted atlases of the human brain in its infancy. *Nat Methods* **20**, 55–64 (2023).

67. Bruner, E., Preuss, T. M., Chen, X. & Rilling, J. K. Evidence for expansion of the precuneus in human evolution. *Brain Struct Funct* **222**, 1053–1060 (2017).

68. Neubauer, S., Hublin, J.-J. & Gunz, P. The evolution of modern human brain shape. *Sci Adv* **4**, eaao5961 (2018).

69. Deco, G. *et al.* The arrow of time of brain signals in cognition: Potential intriguing role of parts of the default mode network. *Network Neuroscience* 1–50 (2022) doi:10.1162/netn_a_00300.

70. Lynn, C. W., Cornblath, E. J., Papadopoulos, L., Bertolero, M. A. & Bassett, D. S. Broken detailed balance and entropy production in the human brain. *Proc Natl Acad Sci U S A* **118**, e2109889118 (2021).

71. Deco, G., Sanz Perl, Y., Bocaccio, H., Tagliazucchi, E. & Kringelbach, M. L. The INSIDEOUT framework provides precise signatures of the balance of intrinsic and extrinsic dynamics in brain states. *Commun Biol* **5**, 1–13 (2022).

72. Yin, H., Artsimovitch, I., Landick, R. & Gelles, J. Nonequilibrium mechanism of transcription termination from observations of single RNA polymerase molecules. *Proceedings of the National Academy of Sciences* **96**, 13124–13129 (1999).

73. Kringelbach, M. L., Perl, Y. S., Tagliazucchi, E. & Deco, G. Toward naturalistic neuroscience: Mechanisms underlying the flattening of brain hierarchy in movie-watching compared to rest and task. *Science Advances* **9**, eade6049 (2023).

74. Sanz Perl, Y. *et al.* Nonequilibrium brain dynamics as a signature of consciousness. *Phys. Rev. E* **104**, 014411 (2021).

75. Cabral, J. *et al.* Cognitive performance in healthy older adults relates to spontaneous switching between states of functional connectivity during rest. *Sci Rep* **7**, 5135 (2017).

76. G-Guzmán, E. *et al.* The lack of temporal brain dynamics asymmetry as a signature of impaired consciousness states. *Interface Focus* **13**, 20220086 (2023).

77. Cruzat, J. *et al.* Temporal Irreversibility of Large-Scale Brain Dynamics in Alzheimer's Disease. *J. Neurosci.* **43**, 1643–1656 (2023).

78. Martos-Moreno, G. Á., Chowen, J. A. & Argente, J. Metabolic signals in human puberty: Effects of over and undernutrion. *Molecular and Cellular Endocrinology* **324**, 70 (2010).

79. Hamson, D. K., Roes, M. M. & Galea, L. A. M. Sex Hormones and Cognition: Neuroendocrine Influences on Memory and Learning. *Compr Physiol* **6**, 1295–1337 (2016).

80. Margulies, D. S. *et al.* Situating the default-mode network along a principal gradient of macroscale cortical organization. *Proceedings of the National Academy of Sciences* **113**,

12574–12579 (2016).

81. Giedd, J. N. Structural magnetic resonance imaging of the adolescent brain. *Ann N Y Acad Sci* **1021**, 77–85 (2004).

82. Gogtay, N. *et al.* Dynamic mapping of human cortical development during childhood through early adulthood. *Proc Natl Acad Sci U S A* **101**, 8174–8179 (2004).

83. Tamnes, C. K. *et al.* Development of the Cerebral Cortex across Adolescence: A Multisample Study of Inter-Related Longitudinal Changes in Cortical Volume, Surface Area, and Thickness. *J. Neurosci.* **37**, 3402–3412 (2017).

84. Stagi, S. *et al.* Increased incidence of precocious and accelerated puberty in females during and after the Italian lockdown for the coronavirus 2019 (COVID-19) pandemic. *Ital J Pediatr* **46**, 165 (2020).

85. Street, M. E., Sartori, C., Catellani, C. & Righi, B. Precocious Puberty and Covid-19 Into Perspective: Potential Increased Frequency, Possible Causes, and a Potential Emergency to Be Addressed. *Frontiers in Pediatrics* **9**, (2021).

86. Chioma, L. *et al.* Sedentary lifestyle and precocious puberty in girls during the COVID-19 pandemic: an Italian experience. *Endocr Connect* **11**, e210650 (2022).

87. Trujillo, M. V., Rungvivatjarus, T. & Klein, K. O. Incidence of central precocious puberty more than doubled during COVID-19 pandemic: Single-center retrospective review in the United States. *Front Pediatr* **10**, 1007730 (2022).

88. Yesiltepe Mutlu, G. *et al.* The effects of the covid-19 pandemic on puberty: a cross-sectional, multicenter study from Turkey. *Italian Journal of Pediatrics* **48**, 144 (2022).

89. Mondkar, S. A. *et al.* Impact of COVID-19 lockdown on idiopathic central precocious puberty – experience from an Indian centre. *Journal of Pediatric Endocrinology and Metabolism* **35**, 895–900 (2022).

90. Gnocchi, M. *et al.* Current evidence on the impact of the COVID-19 pandemic on paediatric endocrine conditions. *Frontiers in Endocrinology* **13**, (2022).

91. Choi, K. H. & Park, S. C. An increasing tendency of precocious puberty among Korean children from the perspective of COVID-19 pandemic effect. *Frontiers in Pediatrics* **10**, (2022).

92. Ariza Jimenez, A. B., Aguilar Gomez-Cardenas, F. J. & de la Camara Moraño, C. Likely impact of COVID-19 on referrals to pediatric endocrinology: Increased incidence of precocious puberty in a third-level hospital. *Endocrinología, Diabetes y Nutrición (English ed.)* **69**, 542–544 (2022).

93. Sonja, B., Dirk, S., Joachim, W., Felix, S. & Bettina, G. COVID-19 and increased prevalence of female precocious puberty in Germany. in *ESPE Abstracts* vol. 95 (Bioscientifica, 2022).

94. Toutoudaki, K. *et al.* Increase of prevalence of Idiopathic Precocious Puberty in Girls, during the Covid-19 Pandemic: What are the possible causes? A systematic review.: https://doi.org/10.54088/jui8. *Developmental and Adolescent Health* **2**, 33–41 (2022).

95. Arcari, A. J. *et al.* Precocious puberty in relation to the COVID-19 pandemic. A survey among Argentine pediatric endocrinologists. *Arch Argent Pediatr* e202202767 (2023) doi:10.5546/aap.2022-02767.eng.

96. Golub, M. S. *et al.* Public health implications of altered puberty timing. *Pediatrics* **121 Suppl 3**, S218-230 (2008).

97. Rudolph, K. D., Troop-Gordon, W., Lambert, S. F. & Natsuaki, M. N. Long-term consequences of pubertal timing for youth depression: Identifying personal and contextual pathways of risk. *Dev Psychopathol* **26**, 1423–1444 (2014).

98. Parent, A.-S., Franssen, D., Fudvoye, J., Pinson, A. & Bourguignon, J.-P. Current Changes in Pubertal Timing: Revised Vision in Relation with Environmental Factors Including Endocrine Disruptors. *Puberty from Bench to Clinic* **29**, 174–184 (2016).

99. Shi, L., Jiang, Z. & Zhang, L. Childhood obesity and central precocious puberty. *Frontiers in Endocrinology* **13**, (2022).

100. Smith, K. K. Heterochrony revisited: the evolution of developmental sequences. *Biological Journal of the Linnean Society* **73**, 169–186 (2001).

101. McNamara, K. J. Heterochrony: the Evolution of Development. *Evo Edu Outreach* **5**, 203–218 (2012).

102. Dobreva, M., Camacho, J. & Abzhanov, A. Time to synchronize our clocks: Connecting developmental mechanisms and evolutionary consequences of heterochrony. *Journal of Experimental Zoology Part B: Molecular and Developmental Evolution* **338**, (2021).

103. Seshachar, B. R. Perspectives in Biology: The International Biological Programme. *Current Science* **36**, 64–66 (1967).

104. Mentzel, H.-J. *et al.* Assessment of skeletal age at the wrist in children with a new ultrasound device. *Pediatr Radiol* **35**, 429–433 (2005).

105. Cao, K., Guo, Y. & Su, S. W. A review of motion related EEG artifact removal techniques. in *2015 9th International Conference on Sensing Technology (ICST)* 600–604 (2015). doi:10.1109/ICSensT.2015.7438469.

106. Barry, R. J., Clarke, A. R., Johnstone, S. J., Magee, C. A. & Rushby, J. A. EEG differences between eyes-closed and eyes-open resting conditions. *Clinical Neurophysiology* **118**, 2765–2773 (2007).

107. Delorme, A. & Makeig, S. EEGLAB: an open source toolbox for analysis of single-trial EEG dynamics including independent component analysis. *Journal of Neuroscience Methods* **134**, 9–21 (2004).

108. Pascual-Marqui, R. D., Michel, C. M. & Lehmann, D. Segmentation of brain electrical activity into microstates: model estimation and validation. *IEEE Trans Biomed Eng* **42**, 658–665 (1995).

109. Lee, C. H., Cook, S., Lee, J. S. & Han, B. Comparison of Two Meta-Analysis Methods: Inverse-Variance-Weighted Average and Weighted Sum of Z-Scores. *Genomics Inform* **14**, 173–180 (2016).