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Reconfiguration of brain-wide neural activity after early life adversity.

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Abstract

Early life adversity (ELA) predisposes individuals to physical and mental disorders lifelong. How ELA affects brain functions, leading to these vulnerabilities, is a mystery. To understand ELA's impacts, investigations into neural activity affected by ELA must go beyond localized areas toward simultaneous recordings from multiple widely distributed regions. Such studies will expose relative activity between regions and discover shifts in regional activity in response to different experiences. Here, we performed longitudinal manganese-enhanced magnetic resonance imaging (MEMRI) to measure brain-wide neural activity in ELA-exposed mice across a series of experiences in adulthood. To ascertain whether ELA resulted in atypical brain activity, results were compared to those of the standard mouse (Std). MEMRI captured activity in the freely moving home cage condition, and short- and long-term after acute exposure to TMT, an innate predator threat. Images were normalized and aligned then analyzed with statistical mapping and automated segmentation. We found that neural activity in the home cage was greater in ELA compared to Std in multiple striatal-pallidal and hypothalamic regions. Upon acute threat, neural activity in Std increased in these regions to become more similar to that in ELA, while new hyperactive responses in ELA emerged in the midbrain and hindbrain. Nine days after acute threat, heightened neural activity in ELA remained within locus coeruleus and increased within posterior amygdala, ventral hippocampus, and dorsomedial and ventromedial hypothalamus. These results reveal functional imbalances that arise between multiple brain-systems after ELA, which are dependent upon context and cumulative experiences into adulthood.

Significance Statement

Early life adversity (ELA) is a crucial determinant of adult health. Yet, the neurobiological basis for this association remains elusive. Neural activity differs within localized brain regions in rodents who experienced ELA, but how this localized activity contributes to overall brain states has hitherto been unknown. Here we used longitudinal manganese-enhanced MRI to reveal brain-wide activities altered by ELA. Computational analyses revealed widespread reconfiguration of segment-wise neural activity throughout the brain across a series of conditions: freely moving, experiencing threat or its aftermath. These brain state dynamics were unique to ELA. Reconfiguration of brain states after ELA, shown here for the first time, may explain vulnerability to mental and physical disorders after childhood adversity.

Main Text

Introduction

Early life adversity (ELA) poses risks for neuropsychiatric disorders, especially after a second

traumatic event, such as a life-threatening experience (1-5). ELA's long-term consequences on

global neural activity into adulthood could serve as a biological basis for poor outcomes (6-8). We

are studying the brain states of adult mice who experienced early life adversity and how their

dynamics progress from being in the home cage to short- and long-term after acute threat. To

ascertain how a potentially normal mouse is affected by ELA into adulthood, we compared ELA-

exposed mice to the standard mouse reared under generalized conditions (9, 10).

We invoked ELA in mice using a well-established paradigm: fragmented maternal care induced by

limited bedding (11, 12). Most studies using preclinical paradigms focus on localized brain regions

with invasive methods (13-27). Only a few have examined ELA's brain-wide consequences (28-

31). Lacking are any studies that examine the brain-wide effects of threat in the adult previously

exposed to ELA. Many questions remain that cannot be answered by localized techniques, such

as whether brain-wide activity is changed in an adult who has experienced ELA, and whether

activity in response to threat is also affected. To investigate these questions and overcome inherent

limitations of other techniques, we employed longitudinal manganese-enhanced MRI (MEMRI)

together with computational data-driven analysis.

Longitudinal MEMRI detects brain-wide activity before, during and after acute threat in awake freely

moving mice (32, 33). Predator odor is a widely accepted paradigm for naturalistic innate threat in

rodents (56-58). MEMRI does not require invasive or destructive procedures (33). Mn(II) is safe,

even continuous infusion of low doses has minimal effects on behavior (33, 34). We chose to deliver

Mn(II) via two single bolus injections in order to avoid surgical procedures required for continuous

infusion and to allow for Mn(II) clearance between imaging sessions (32). Systemic Mn(II)

distributes throughout the brain parenchyma (35), enters active neurons through voltage-gated

calcium channels (36, 37) and accumulates in neurons in an activity-dependent manner (33, 38,

39). Regional accumulations of Mn(II) throughout the brain produce robust hyperintense MR signals

with large effect sizes at 100 μm^3 resolution (32, 33, 35, 40-42). These signals represent a

composite picture of regional neural activity occurring between the time of injection and imaging.

Alignment of multiple MR images, collected repeatedly from large cohorts of individuals as they

progress in parallel through a series of experiences, produces a dataset that enables extraction of

statistically significant voxel-wise information about brain-wide activity over time, not possible with

most other techniques (32, 33, 43-54). Such processing is a well-accepted practice in human
imaging that yields much useful information and is routinely repurposed for rodent imaging (51, 55-
58). To ensure reliability and reproducibility, computational processing is verified and validated at
each step. Thus, MEMRI can uniquely image enduring consequences of ELA on brain-wide activity
in the adult and its dynamics in response to threat.

Here, we examine the consequences of ELA on adult brain-wide activity during normal home cage
exploration in the adult and investigate how this activity changes in response to acute threat, short
and long term, with data-driven and hypothesis-based analyses of MEMRI images captured
longitudinally. Our alignments and computational processing pipeline were rigorously verified at
every step. We validated Mn(II) dose by showing similar intensity measurements across all
individuals. We verified that Mn(II) signal replicated neural activity detected by other methods by
measuring intensities in circuits known to be involved in predator odor responses and by correlation
of Mn(II)-dependent enhancements with c-fos expression. We used within- and between-group
statistical parametric mapping and automated segmentation with our new *InVivo* mouse brain atlas
to quantify relative activity within and between 101 segments brain-wide under each condition. We
found that volumes of activities in multiple segments throughout the brain differed significantly
between standard mice and those exposed to ELA undergoing the same experiential timeline.

Results

To witness neural activity in the home cage condition (HC) of adult mice who experienced ELA,
and their responses to predator stress short- and long-term, we adapted our longitudinal imaging
procedure (**Fig. 1A**) (32). We compared ELA mice (ELA, n = 12) to pool-weaned JAX-reared mice
of the same strain (Standard, Std, n = 12) to determine whether brain-wide activity of adult mice
who had experienced ELA differed from a phenotypically "normal" adult mouse.

Successful Replication of Behavioral Paradigms. To test whether our protocols for these two
paradigms, fragmented care (ELA) and predator stress (TMT), faithfully reproduced previously
reported behaviors (59-61), we compared videos of adult mice as they explored a custom arena
(**Fig. 1B-D**). For the effect of ELA experience, we measured the time ELA mice spent in the light at
baseline (BL) and compared that to Std mice. As expected, adult ELA mice spent 10% more time
in the light at BL than Std, demonstrating a persisting effect of ELA experience in an unprovoked
situation ($p < 0.05$ FDR, Cohen's d = 1.58) (**Fig. 1B**). For the effect of TMT, we measured time
spent in the light and compared between BL and TMT. As expected, TMT decreased time spent in
the light in both ELA and Std mice by more than 20%, demonstrating an acute effect of TMT ($p <$

0.05 FDR, Cohen's d_{Std} = 2.29, d_{ELA} = 3.42). To assess mouse recovery from MR anesthesia before

being presented with TMT, we measured distance traveled in 1 min epochs and tracked

movements from the beginning of a 10 min exposure to TMT (**Fig. 1C-D**). During the first minute of

TMT-exposure, distance traveled was equal to or greater than that at BL. Motility then decreased

by 50% or more over the 10 min TMT-exposure period ($p < 0.05$, FDR). These data support the

contention that our procedures faithfully reproduced ELA and TMT paradigms, giving us confidence

in assigning behavioral characteristics to unknown brain-wide activity.

Longitudinal Imaging. Our MR imaging protocol captures 8 sequential images across 12

individuals for each group (ELA and Std), yielding a total of 192 raw images with >2 million voxels

each (**Fig. 1E**). To ensure no differences between groups were introduced by processing, all

images were processed as a batch through our semi-automated processing pipeline (32, 33). This

pipeline yielded a combined dataset of skull-stripped, intensity normalized and spatially aligned

images ready for statistical evaluation. Quality assurance considerations included reviewing

imaging parameters, performing SNR measurements to confirm similar Mn(II) doses, visualizing

modal scaling output in voxel-by-intensity histograms to monitor equivalent intensity normalization,

calculating Jaccard Similarity and Mutual Information Indices for alignment accuracy, and

determining that smoothing does not significantly change regional signal intensities. All images

reported here met our quality assurance standards (*SI Appendix* **Fig. S1-2** and **Table S1**).

Brain States in Mice Exposed to ELA. To detect Mn(II)-enhancements in ELA mice across

conditions from the home cage (HC) to post-threat (TMT) to nine days after threat (D9), we

compared post-Mn(II) images for each condition to each mouse's respective pre-Mn(II) image using

voxel-wise statistical parametric mapping (SPM) (**Fig. 2**). SPM of images at HC revealed large

volumes of statistically significant signals in multiple brain regions identified by anatomy,

demonstrating that the unprovoked ELA brain was already highly active (T = 2.72, $p < 0.01$, cluster

corr.) (**Fig. 2A**). Larger volumes of SPM signal were seen within regions of the olfactory bulb,

striatum/pallidum, hypothalamus, hindbrain, and cerebellum, and smaller volumes within

subregions of hippocampus, amygdala, and hindbrain. Immediately after TMT, signals in these

regions were maintained or expanded in volume, with sizable increases in the mid- and hindbrain

(e.g., LC) (**Fig. 2B**). By D9, activity decreased overall in olfactory, hindbrain, and cerebellum, or

increased overall in hippocampus, amygdala, and midbrain (e.g., PAG) (**Fig. 2C**). Other subregions

also changed, such as a decrease of the volume of statistical signal in the lateral septum (LS) and

appearance of new signal in the paraventricular nucleus of the thalamus (PVT). Direct SPM paired

t-test comparisons between ELA post-Mn(II) images at each condition added to these findings from

activation volumes, to also identify statistical differences in signal intensities within many regions

across conditions (*SI Appendix* **Fig. S3**). These findings indicate that ELA-exposed brains are already highly active before a second provocation in adulthood, and that short- and long-term brain responses to TMT have unique patterns of activity, with regions emerging or disappearing between conditions.

Magnitude of Signal Enhancements. To determine whether differences in statistical maps accurately represent intensity changes across conditions, and to test whether signal intensities increase in response to TMT within expected regions, we measured and compared signal intensities within 3 x 3 x 3 voxel regions of interest (ROI) between conditions (**Fig. 3A**, and *SI Appendix* **Tables S2** and **S3**). We selected these ROIs due to their involvement in predator odor responses (62, 63), to determine the magnitude of intensity difference suggested by SPM. Our previous, voxel-wise SPM of ELA images at HC and TMT revealed significantly enhanced signal intensities after TMT in all four expected regions (p<0.05 cluster corr.). At HC, dlOB-DII and vlAON, were already enhanced and only minimally changed across conditions. Region of interest (ROI) measurements within these locations revealed that these SPM signals at HC represented large signal differences over pre-Mn(II) (6.8%, dlOB-DII; 8.0%, vlAON) and large Cohen's D effect sizes (d = 1.6, dlOB-DII; d = 1.5, vlAON; p < 0.05 FDR). In aOC and aCEA, SPM signals were minimal at HC and increased from HC to TMT. These SPM signal changes were recapitulated in ROI comparisons, with aOC and aCEA showing small magnitudes of signal enhancement over pre-Mn(II) at HC (1.4 and 3.0%), and larger magnitudes at TMT (3.5 and 6.2%). Thus, in this case SPMs accurately represented signal intensity differences. Furthermore, signal intensity increased in all four regions expected to respond to TMT (+2.1-3.9%, d = 0.85-1.20, p < 0.05 FDR), demonstrating robust TMT-induced enhancement.

Statistical mapping of ELA mice at HC (Fig. 2) indicated signal enhancements within regions we previously identified to respond to TMT in Std mice (32). To determine the degree of signal intensity differences between ELA and Std at HC and test whether ELA signal truly exceeded Std in these regions, we measured and compared signal intensities within 3 x 3 x 3 voxel ROIs (**Fig. 3B**, *SI Appendix* **Tables S2** and **S4**). Magnitudes of signal enhancement at HC over pre-Mn(II) corresponded to the presence or absence of significant SPM signal for both Std and ELA (non-enhanced: SI = 2.0-3.6%, d = 0.5-0.7; enhanced: SI = 6.1-17.9%, d = 0.96-5.17). Direct comparisons between groups revealed that ELA intensities were heightened at HC in all four measured regions (SI = 4.1-6.6%, d > 1.00, p < 0.05 FDR). Thus, ELA signal intensities within threat-responsive regions exceeded that of Std even before threat. ELA also displayed a lower variance than Std.

Together these observations support the conclusion that differences in SPM-identified activation
volumes, at $p < 0.01$ and cluster size ≥ 8 voxels, reliably represent intensity differences between
post-Mn(II) images. However, when SPM analysis reveals overlapping anatomy of activation
volumes between groups or conditions, direct measurements of post-Mn(II) intensities may be
necessary to uncover potential differences in activity levels that are not reflected solely by activation
volumes.
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Mn(II)-Enhanced Signal Corresponds to c-fos Expression. We next sought to test whether
different degrees of Mn(II)-enhancement corresponded to differences in neural excitation, as
detected by c-fos expression (33, 42, 64, 65) (**Fig. 4**). To test this hypothesis, from a subset of mice
from both groups ($n_{Std} = 5$ and $n_{ELA} = 6$), we selected pairs of regions with varying numbers of c-
fos+ nuclei, where anatomy was also identifiable in MR slices: hypothalamus (HYP) and thalamus
(THA); superior central raphe (CS) and midbrain (MB) (**Fig. 4A-B**). In both groups, regions with
statistically greater numbers of c-fos+ nuclei (HYP vs THA; CS vs MB) also displayed statistically
greater Mn(II)-enhanced signal intensities (**Fig. 4C-D**, $p < 0.001$). Robust effect sizes were
observed for both modalities (c-fos: d = 3.75-5.03; MEMRI: d = 1.88-6.37). Statistical differences
in MR signal intensities from these subsets of mice were replicated in a second subset from each
group, demonstrating test-retest reliability (d = 2.94-8.43, $p < 0.001$, FDR) (***SI Appendix* Fig. S4**).
To further test whether intensities correlated linearly with c-fos counts, we correlated MEMRI and
c-fos data across the four regions with bootstrap resampling and permutation testing. In both
groups, we found strong correlations (Std: r = 0.636, $p < 0.0001$; ELA: r = 0.582, $p < 0.0013$) (**Fig.**
4E and ***SI Appendix* Fig. S5**). Thus, our data suggest that the degree of MEMRI signal intensities
reflects differences in neural excitation, and further that MEMRI data reflect cumulative activity
within a given volume, i.e., density of active neurons as well as the level of activity of those neurons.
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Deeper Dives into Brain States of Adult Mice Exposed to ELA. To assign neural activity in ELA
mice to specific anatomical regions, we aligned statistical maps of enhanced voxels with our *InVivo*
Atlas (***SI Appendix* Table S5**). Then we quantified activity in those anatomically defined segments
at each condition with a new approach, fractional activation volumes (FAV) (32) (**Fig. 5**). The FAV
is the fraction of activated to total voxels in each segment. Activated voxels were those found to be
statistically enhanced in SPM comparisons of post-Mn(II) to pre-Mn(II) images at each condition.
FAVs were visualized as column graphs. Comparing column graphs between conditions
demonstrated successive changes in FAVs as mice progressed through successive conditions.
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Overlays of statistical maps show that much of the activity at HC coincides with that of TMT in ELA
mice ($p < 0.01$ cluster corr., d = 0.79-8.19) (**Fig. 5A** and ***SI Appendix* Table. S6A)**. Some regions

expand while a few others contract from HC to TMT. By FAV analysis of ELA, we see that at HC
large FAVs were prominent in various domains (OLF, STR/PAL, HYP, and CB) with segments
(MOB, MS, BST, and AHN) almost totally active. Immediately after TMT, FAV values were retained
or increased, with the largest increases in CEA and LC. Several STR/PAL and THA segments
decreased in FAV after TMT, including LSc, LGd and MH. No relationships between segment
volume and FAV were observed. Differences in signal intensities between conditions in these
segments were also statistically significant by SPM comparisons between successive post-Mn(II)
images, with the greatest differences in the LC (*SI Appendix* **Fig. S6** and **Table S6B**).

We then compared segmental FAVs between TMT and D9 of ELA mice to learn how TMT affected
brain-wide activity long term (**Fig. 5B**). Segmentation analysis revealed that FAV decreased in
three domains (OLF, HB, and CB) with the most substantial change in CB. Most other segments
either retained or further increased FAV, particularly within AMY, THA, HYP and MB domains. In
many segments where FAV had increased following TMT, FAV at D9 either persisted (LC) or
increased (CEA), suggesting a longer lasting effect of the threat experience. Segments where FAV
decreased acutely after TMT typically remained low (LSc). Others increased at D9 (MH). These
results suggest that brain activity in some segments does not return to pre-threat levels.

To test whether activities do not return to pre-threat levels, we compared HC with D9 using a similar
strategy in both ELA and Std mice (**Fig. 5C and 5D**, also *SI Appendix* **Table S6A**). Both overlays
of SPMs and columns graphs of FAV show that at D9 after threat, activity in both ELA and Std was
increased substantially compared to HC in many domains (AMY, STR/PAL, HYP, and MB). The
magnitude of FAV increase in these domains was twice as large on average in Std than in ELA
(+0.28 vs +0.11). This pattern also appeared in pairwise comparisons of post-Mn(II) intensities
between conditions (*SI Appendix* **Fig. S7** and **Table S6B**). Neither the brain-wide activity of ELA
nor Std returned to pre-threat levels at D9. These data suggest regions in the brain remain
hyperactive one week or more after acute threat.

Visualization of brain states at HC and D9 revealed qualitative differences between groups. At HC,
before threat, FAVs of ELA were substantially greater than those of Std across many brain
segments. These larger FAVs in ELA could not be attributed to procedural differences, such as
MnCl$_2$ dosing, MR parameters, imaging timing, image processing, segmentation analysis (*SI*
Appendix **Fig. S2** and **Table S1**), differences in pre-Mn(II) signal intensities between groups (*SI*
Appendix **Fig. S8**) or differences in anatomy between groups (*SI Appendix* **Fig. S9**). While Std
exhibited larger FAV increases from HC to D9 than ELA in most segments, ELA exhibited larger
increases than Std in a subset of segments, including RE and LC. In other segments, ELA displayed

the opposite directional change from HC to D9 compared to Std, including PA, LSr, IPN, and PCG.

Except for IPN, these segments also responded acutely to TMT in ELA (**Fig. 5B**). These findings

suggest that brain-wide activity and its evolution from threat-naïve (HC) to threat-experienced (D9)

conditions differs between groups and that ELA mice resemble a post-threat-like state already at

HC. Furthermore, those segments that respond to TMT in ELA appear to be major contributors to

differences between groups at D9. These qualitative observations prompted a direct between-

group statistical analysis.

ELA Affected Many Segments Within Each Anatomical Domain. To determine whether signal

intensities between ELA and Std differed statistically between groups, and to map the anatomical

distribution of those differences, we performed a direct SPM comparison. First, we performed an

unpaired t-test comparing baseline pre-Mn(II) images between groups, which revealed small

intensity differences (*SI Appendix* **Fig. S7**). For subsequent comparisons, post-Mn(II) images were

adjusted to correct for these baseline differences. We then compared ELA and Std post-Mn(II)

images at each condition by an unpaired SPM t-test ($p < 0.05$, cluster corr., d = 0.70-5.30) (**Fig. 6**

and **Movie S1**). Segmentations of SPM difference maps were used to calculate the proportion of

statistically enhanced voxels (either ELA > Std or Std > ELA) per total voxels in each segmented

region at each condition, the fractional difference volume (FDV) (**Fig. 6A-D**, and *SI Appendix*

Table S7). At all three conditions, ELA exhibited greater signal intensities than Std (i.e., heightened

activity) in most segments but to a lesser extent at D9, supporting observations from the within-

group SPMs shown in **Fig. 5**. From HC to D9, the total volume of enhanced voxels in ELA relative

to Std decreased by 27%.

Closer examination of FDVs within different anatomical domains identified heightened activity at

HC in ELA compared to Std within OLF, STR/PAL, and HYP domains (**Fig. 6A**, yellow columns).

At TMT, differences in these domains decreased, although heightened HYP activity in ELA was

somewhat preserved and new differences became noticeable in the CB (**Fig. 6B**). At D9,

heightened activity in the HYP domain of ELA returned or even increased (**Fig. 6C**). While

heightened activity was present in most HYP segments for each condition, many other domains in

ELA exhibited smaller volumes of heightened activity over Std at TMT and D9, particularly

segments within STR/PAL. In contrast, ELA exhibited diminished activity compared to Std within

only a few segments of CTX, THA, HYP, MB and CB domains (**Fig. 6A-C**, blue columns). Thus,

ELA in general displayed heightened brain-wide activity over Std at HC. The number and degree

of differences lessened upon experiencing threat in most segments but not in all. Next, we will take

a deeper dive into segmental activity dynamics.

Examining segment-wise FDV across conditions revealed at least five patterns of segmental
dynamics (**Fig. 6D**). To facilitate interpretation of activity differences across conditions, we plotted
FDVs indicated by arrows in **Fig. 6A-C** as line graphs (**Fig. 6D**). These graphs represent the
dynamic patterns of heightened activity in either ELA (yellow) or Std (blue). The *first* pattern across
many segments was heightened activity in ELA at HC, which became more similar between groups
at TMT (**Fig. 6D**, blue). Some of these segments continued to be similar between groups at D9
(e.g., MS) while others returned to heightened activity in ELA (e.g., VMH). A *second* pattern
followed an opposite dynamic in response to TMT. In these segments, volumes of heightened ELA
activity expanded acutely with TMT and returned to HC levels at D9 (CEA and CS, red). A *third*
pattern appeared in a few segments, where FDVs of ELA rose only at D9 (PA and DMH, green). A
fourth pattern emerged in two segments (DP and ARH, black). Similar to pattern one, ELA activities
in these two segments were greater than those of Std at HC and/or D9. Dissimilar to pattern one,
ELA activities decreased in these segments below those of Std at TMT. A comparison between
male and female found that ELA females might contribute to between-group differences at D9 in
VMH, ARH and PA more than ELA males, with sex-differences in PA also present in pre-Mn(II)
images (***SI Appendix* Fig. S10**). A single segment followed a *fifth* pattern (LC, purple), where
activity was greater in Std at HC and then reversed after TMT through D9 to become heightened
in ELA. Together these data demonstrate that brain-wide activity of ELA mice is reconfigured
across conditions and that dynamics in response to threat, both brain-wide and segment-wise,
differ dramatically from Std.

Discussion

In this study we mapped the spatial-temporal transformations that ELA imposes on brain-wide
activity, as detected initially when in a home cage (HC) environment and then short and long-term
after challenge by an acute threat as compared to a Standard, phenotypically "normal," mouse.
Three different brain states (66), as defined here by widely distributed patterns of fractional
activation volumes among segments, emerged within each group. In all three conditions, brain
states of adult mice reared with the fragmented maternal care model for ELA differed from brain
states of adult Std mice, with greater overall neural activity after ELA (**Fig. 6E** and ***SI Appendix***
Fig. S11). Even without provocation, ELA mice at HC had heightened fractional activation volumes
in regions known to be activated by predator odor or altered in reward processing: olfactory and
hypothalamic segments (65, 67-72), as well as striatum-pallidum (32, 73, 74). Distribution of
heightened activity expanded immediately after threat to engage subcortical structures as well as
monoaminergic nuclei and deeper brainstem segments in ELA, as also occurred in Std (32). Neither
group regained HC activity patterns nine days after threat. Across all conditions, activity in

hypothalamus of ELA was higher than Std. At D9, new differences emerged in the ventral hippocampus and posterior amygdala, where activity was greater in ELA. Our longitudinal brain-wide imaging strategy coupled with our computational processing and analytics empowered us to follow these brain states in the same individuals as they progress through this series of experiences in sample sizes sufficient for statistical significance. Our processing pipeline, aligning 192 whole-brain images captured with $100\mu m^3$ resolution, allows precise localization of statistically significant activity common across multiple individuals. This unique capability is not possible with most other physiological approaches.

We found dynamic changes in segment-wise fractional activation volumes between conditions throughout the brain in both cohorts. When a segment transitions from lower to higher FAV/FDV, we predict that segment to have a more dominant functional role within its connectome. For example in ELA, the increased volume of activity In CEA (from 43% to 53%) as the mouse progresses from home cage to immediately after threat may contribute to heightened activity throughout threat circuity (75). That ELA mice sustained heightened CEA activity at all three conditions, suggests that ELA leads to persistent engagement of threat circuitry. This sustained engagement of threat circuitry after ELA may sensitize CS and LC to threat, promoting monoaminergic modulation of segmental activities toward the patterns observed at D9 (76-79). Differing levels of activity among segments of a single network across different conditions could produce different internal states (80, 81) and/or outcomes from that same network, be they actions, decisions, emotions, thoughts, memories, or creative works, such as music, poetry, or art.

Results presented here should be considered in light of several limitations. 1) Theoretically, small variations in rearing environment could contribute to differences between groups not attributable to ELA. However, multi-site studies in mice find that different breeding/rearing facilities have no effect on stress reactivity (82) and minimal explanatory value for at least 14 other physiological measures (83). JAX-reared mice are the phenotypical standard and the most commonly used mouse by researchers globally (9), making them an invaluable comparison group to generalize findings across research facilities (84). 2) Although we found minimal sex differences between male and female ELA mice, we cannot fully attribute heightened activity in HYP and vHIP/AMY segments to ELA because we only had a small sample size and because our Std group was all male. Other studies suggest a combinatorial effect of ELA and sex is likely within these threat responsive regions (71, 85, 86). 3) To what degree does Mn(II) signal faithfully represent neural activity? Staining by c-fos shown here and previous reports (32, 36-39, 42, 87, 88) contend that Mn(II) tags active neurons, though minor non-calcium-dependent mechanisms may exist (88, 89). The amount that accumulates and the rate of Mn(II) transport within axons cannot explain changes in segmental

signal intensities between HC and TMT, nor the new pattern of segmental activity at D9. 4)

Spatiotemporal resolution of MEMRI is less granular than some other methods. At 100 µm^3 voxels,

our MEMRI is better than current BOLD imaging but still not at the level of single neurons studied

with electrophysiology. MEMRI temporal resolution is less than either BOLD imaging or

electrophysiology. However, MEMRI's capacity for repeated longitudinal brain-wide imaging across

a long time span of neural activity in awake, freely moving mice makes these tradeoffs worthwhile.

MEMRI pinpoints regions worthy of study at greater resolutions by other techniques. 5) Small

anatomical variations could have influenced our results (90-93). However, our Jacobian

deformation analysis reported minimal alterations in anatomical volumes, and our SPM analysis

accounted for any signal differences by preserving intensity concentrations within voxels (94, 95).

6) We recognize that statistical methods are limited due to the choice of thresholding and pre-

preprocessing, which is particularly difficult with MRI (33, 96-98). We minimized these potential

issues by rigorously examining each processing step for image quality, by ensuring sample size

was sufficient to detect moderate to large signal enhancements from Mn(II), and by selecting

statistical thresholds based on effect size and false positive rate corrections. 7) Atlas segmentation

yields volumes that vary widely, 10^1-10^4 voxels. This may lead us to miss some large segments

where small- to medium-sized activations only produce small fractional volumes.

In conclusion, ELA has a brain-wide impact on balance of neural activity among widely distributed

sub-regions and their dynamics across a series of experiences, from freely moving exploration in

the home cage to immediate- and post-threat conditions. We identify specific subcomponents of

this brain-wide network that are differentially activated depending upon condition in adult mice who

experienced ELA and compare these with the standard, commonly studied, adult mouse. Future

studies dissecting the functional connections suggested by these MEMRI results will provide

valuable information on coordination of coactivated regions. A next step will be to determine the

functional significance of these disparate segmental activities, how they interact in a serially

dependent manner to reconfigure future brain states, and whether these interactions contribute to

vulnerabilities for mental health disorders and/or compulsive drug use. Ultimately this longitudinal

MEMRI technique will provide insights into neurophysiological/biochemical mechanisms

contributing to the reconfiguration of brain states by ELA and inspire reparative/restorative

interventions to protect from the harm of ELA and secondary traumatic events later in adulthood.

Materials and Methods

Mice. Animal protocols were approved by Institutional Animal Care and Use Committees at both

California Institute of Technology (Caltech) and University of New Mexico. Numbers were

determined by statistical power analyses. Both Std and ELA reared groups of C57BL/6J mice

(Stock No: 000664) were bred at Jackson Labs (JAX). Adults from each group went through parallel

acute threat timelines, were handled by the same experimentalists, and imaged at Caltech (n = 24,

12 in each group), in a series of experiments performed over a period of 3.8 years. For ELA, we

followed a fragmented care protocol (11, 12). Longitudinal MEMRI commenced at ages 11-13 wk.

This age was chosen because brain development is considered largely complete by 3 months (99).

Equal numbers of male (n = 6) and female (n = 6) ELA pups were used from 5 weaning groups.

Twelve male Std mice selected from 5 different weaning cages were subjected to the longitudinal

experimental protocol. No litter/cage effects were detected for either group.

Video Recordings and Acute Threat Exposure. We followed our previously published

longitudinal imaging protocol (32). Briefly, prior to imaging and during TMT exposure mice were

tracked in a custom arena using Noldus Ethovision (XT15) (32, 100, 101). Prior to TMT exposure,

mice were bright, alert and responsive, and displayed normal motility. Percentage of time spent in

light side of the arena and distance travelled in the arena were calculated and statistically evaluated

with FDR correction in R v4.2.1 (*nlme* and *emmeans* packages) (102-107). Graphs were generated

in R (*ggplot2*) (108).

Longitudinal MEMRI. After capture of a non-enhanced image, mice were injected i.p. with 0.3

mmol/kg of $MnCl_2$-$4H_2O$ in sterile buffered saline and then imaged with contrast according to the

timeline previously reported (32). Between scanning sessions, video recordings, or $MnCl_2$

injections, mice were returned to their home cage. Mn(II) kinetics were followed by ROI analysis

across the timeline to test for expected intensity patterns. For optimization of MEMRI imaging

parameters, refer to our review (33).

MR Image Pre-Processing. MR images from both groups were pre-processed together in batch

according to standardized protocols (33, 49, 51). Briefly, raw MR image quality was visually

assessed and SNR quantified. Image artifacts were corrected when needed and outlier images

excluded from statistical analyses. Std and ELA pre- and post-Mn(II) images were then skull-

stripped, intensity normalized and anatomically aligned together in batch. Each processing step

was verified and validated (***SI Appendix* Fig. S1-2**).

MR Image Analysis. *Comparison of Anatomy Between Groups.* For comparison of anatomy

between groups, we calculated Jacobian Determinants of the Deformation fields generated by the

affine-nonlinear alignment step. Average Jacobian determinant values were calculated using our

atlas-based segmentation process (see below). Two sample t-tests between Std and ELA values
were performed for each segment, FDR correction.

Measurement and Analysis of MR Intensities in ROIs. Measurements were made from pre- and
post-Mn(II)-enhanced MR images that had been skull-stripped, scaled, and aligned. Coordinates
(x/y/z) for regions of interest (3 x 3 x 3 voxels) were selected by projecting the annotated atlas
labels onto the aligned grayscale template in FSLeyes and placing the crosshairs within segments
that appeared different in SPM comparisons to pre-Mn(II) and that corresponded to previously
reported TMT-responsive regions (32, 75). Mean intensities from ROI measurements were
compared by paired and two-sample t-tests in R (*t.test)*, and effect sizes and FDR adjustments
were calculated (*effectsize* and *p.adjust)*.

Statistical Parametric Mapping (SPM). For SPM, all images were smoothed to meet SPM
assumptions with a 0.15 mm^3 Gaussian kernel, the minimum size allowable (56, 95). We adjusted
for small baseline differences between groups in their pre-Mn(II) images by subtracting the average
voxel-wise differences of intensities from each of the ELA images. After this adjustment, an
unpaired t-test of pre-Mn(II) images yielded no voxels with significant differences between groups.
Voxel-wise T-maps of statistically significant differences between images of different conditions
within group (paired t-test) or between groups (unpaired t-test) were calculated using SPM12 (95).
Statistical maps were cluster corrected (*cluster corr.*) in SPM12 with a minimum cluster size of 8
voxels (95). Fiber tracts and ventricles segments were masked from statistical maps using our
InVivo Atlas. Setting statistical thresholds included consideration of minimal detectable effects and
smallest cluster size needed to remove false positives while minimizing loss of signal.

Atlas Alignment, Segmentation, and Calculation of Fractional Volumes. Our updated *InVivo* Atlas
grayscale image was inversely aligned to the dataset, and the warp field applied to the label image.
This permitted automated segmentation of the images into 117 regions (32). Segment masks were
created and applied using *fslmaths*. The number of statistically enhanced voxels were identified as
those from SPM paired or unpaired t-tests exceeding thresholds set according to our criteria
described above. The number of total voxels within each segment were counted using *fslstats* (24).
The fraction of enhanced voxels from SPM paired (FAV) or unpaired (FDV) t-tests to total voxels
within each segment were calculated in R and visualized as column graphs and line plots using
ggplot2 (107-109).

Immunohistochemistry, c-fos Quantification and Correlation to MEMRI. Brains were prepared
for histology after D9 according to our protocol (5 Std and 6 ELA) (32, 110). We selected 1-3

sections also identifiable in MR slices that had regions with both very high and very low numbers

of c-fos stained nuclei. Nuclei were counted in a 3 x 3 grid of 100 μm^2 squares from 1-3 x 35 μm

thick sections of each brain, and MR intensities (SI) measured from 100 μm^3 voxels in

corresponding slices. Counts and SI were visualized in quartile boxplots with R (*ggplot2*). A square

root transformation linearized c-fos+ count data for determination of effect size and statistical

significance. Regions counted in the same section or measured in the same slice were compared

by linear mixed models and by post-hoc paired t-tests with FDR correction. Counts and SI of each

group were averaged by region and individual and then correlated, with p-value determined by

permutation testing.

Please see *SI Appendix* **Extended Methods** for additional methodological details.

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Figures and Tables

Figure 1.

Fig. 1. Longitudinal imaging procedures. *(A)* Schematic of the overall timeline (open arrows, top).

Boxes diagram events. *Left box*: ELA paradigm; *Middle box*: Home Cage and Acute Threat, timeline

of MR and video capture: Baseline, BL; Home Cage, HC; Acute Threat, TMT. *Right box*: Day 8

Baseline, D8-BL; Day 9 Post-Threat, D9-H. *(B-D)* Video tracking results: *(B)* Percent time spent in

the light (quartile boxplots, individuals indicated by dot plot overlays) (cyan, Std; yellow, ELA; BL

and TMT as per schematic). *(C)* Distance traveled after imaging during TMT exposure. Line graphs

of distance travelled per 1-min epochs with error bars representing standard error of the mean.

Repeated measures ANOVA with fixed effects of group and condition. For (B-C), paired and

unpaired t-tests with FDR correction for multiple comparisons; Cohen's D effect sizes (d); * $p <$

0.05, n = 22 (10 Std, 12 ELA). *(D)* Representative tracking of one mouse from each group, Std and

ELA, at BL (gray) and TMT (red). Shaded areas represent dark side of the arena. *(E)* Overview of

MR data processing and analysis used in this study (also see *SI Appendix* **Fig. S1-2**) (33, 49, 51).

Figure 2.

Mn(II) Accumulations across Conditions after ELA

Fig. 2. Statistical parametric maps of MEMRI in ELA mice change as they progress from home
cage to acute threat to post threat. *(A-C)* Shown are statistical maps of voxel-wise paired t-tests of
post-Mn(II) images of ELA mice at each successive condition compared to their pre-Mn(II) images.
Maps are overlaid on the grayscale dataset average of all pre-Mn(II) images (coronal slices, top
rows; sagittal, bottom rows). Slice position is indicated on the grayscale image (top left, white lines).
White arrows indicate notable differences between conditions. MOB, main olfactory bulb; LS, lateral
septum; BST, bed nuclei of the stria terminalis; PVT, paraventricular thalamus; HIP, hippocampus;
LC, locus coeruleus; CEA, central amygdala; and CB, cerebellum. For t-tests, n = 12, p < 0.01, T
= 2.72, cluster corr. For comparisons between conditions of ELA mice see *SI Appendix,* **Fig. S3**.
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Figure 3.

Fig. 3. Measurements of MEMRI signal intensities in candidate regions of interest in systems
known to be involved in predator odor-type threat responses. *(A) Top*: Representative slices from
two voxel-wise statistical maps (ELA HC & ELA TMT) of post-Mn(II) > pre-Mn(II) superimposed
onto the grayscale *InVivo* atlas. SPM paired t-tests: $p < 0.01$ cluster corr. ROI locations selected
where TMT signal did not overlap with HC (yellow crosshairs). *Bottom*: Quartile boxplots of signal
intensity as a percent over pre-Mn(II) measured in each ROI. Effect sizes of the differences in signal
between conditions at each ROI (Cohen's D, d) and average signal intensity differences (ΔSI)
between the two conditions are indicated. Paired t-test, * $p < 0.05$, ** $p < 0.01$, FDR. dlOB-DII,
dorsal lateral DII olfactory subdomain; vlAON, ventrolateral anterior olfactory nucleus; aOC,
anterior olfactory cortex; and aCEA, anterior central amygdala. *(B) Top*: Similar comparison but
between groups at one condition, HC. Slices of paired t-tests of HC greater than pre-Mn(II) from
either Std or ELA are superimposed on the grayscale atlas. Crosshairs indicate ROI locations,
selected where ELA signal did not overlap with Std. *Bottom*: Quartile boxplots of unpaired t-tests
of ROI measurements: * $p < 0.05$, FDR. mOB-DII, medial DII olfactory subdomain; dmNDB,
dorsomedial diagonal band nucleus; aLS, anterior lateral septum; dmACB, dorsomedial nucleus
accumbens. See ROI coordinates and summary statistics in *SI Appendix,* **Tables S2-4**.

Figure 4.

Fig. 4. MEMRI and c-fos staining correlate. (*A*) Coronal slices of the average intensities of day-9

post-Mn(II) images (color code, bottom) (n = 6). Minimum (300 a.u.) based average noise and

maximum (11,000 a.u.) from intensity histograms. Small white boxes indicate regions (3 x 3 x 1

voxels) where signal intensities (SI) were measured. HYP, hypothalamus; THA, thalamus; CS,

superior central raphe; MB, midbrain. Mag bar = 500 μm. (*B*) Two representative histological

sections from one of the mice contributing to *(A)* stained for c-fos corresponding to anatomy of

slices in *(A)* (large white boxes). Purple boxes indicate locations of 3 x 3 grid of 100 μm² squares

where numbers of c-fos+ nuclei were counted. Mag bar = 500 μm. See *SI Appendix* **Fig. S5A-B**

for corresponding images from Std. *(C)* Statistical comparisons of MEMRI SI in four locations for

both groups. Shown are quartile boxplots with dot plot overlays of MEMRI SI from ELA (yellow, left)

and Std (cyan, right). Paired t-tests were performed between regions within the same slice. Note

large Cohen's D effect sizes (d) for paired t-tests between active and inactive regions. For evidence

of test-retest reliability of MEMRI SI, see *SI Appendix* **Fig. S4**. *(D)* c-fos counts paralleled MEMRI

SI with high or low numbers of c-fos+ nuclei. Shown are quartile boxplots with dot plot overlay of

raw c-fos+ counts from ELA (purple, left) and Std (pink, right). A square root transformation

linearized c-fos+ counts for statistics. Note similar large Cohen's D effect sizes (d) between active

and inactive regions. *(E)* Strong Pearson correlations between MEMRI SI and numbers of c-fos+

nuclei (linearized) when paired by individual. Scatter plots with linear regressions (black lines) of

MEMRI SI (vertical axes) by c-fos+ counts (horizontal axes). Points represent the average values

for each of the 4 regions across 5 Std (*top*, *cyan*) and 6 ELA (*bottom*, *yellow*) brains. Note strong

correlations in both groups (r_{Std} = 0.636, r_{ELA} = 0.582). For paired t-tests in *(C-D)*, *** p < 0.001,

FDR corrected. For *(E)*, p < 0.001 by permutation testing and bootstrapping. Also see *SI Appendix*

Fig. S5C-D.

Figure 5.

Fig. 5. Statistical maps show progressive modifications of activity across sequential conditions. *(A-D)* Voxel-wise paired t-tests of post-Mn(II) compared to pre-Mn(II) images at two conditions: in ELA mice, *(A)* HC (blue) and TMT (red), *(B)* TMT (red) and D9 (green), and *(C)* HC (blue) and D9 (green); and in Std mice, *(D)* HC (blue) and D9 (green). *Top*: Pairs of statistical maps were projected onto the grayscale *InVivo* Atlas with segments outlined to show the anatomical overlaps between conditions. *Bottom*: Corresponding column graphs show segment-wise fractional activation volumes (FAV, vertical axes) quantified from segmentations of statistical maps. Anatomical domains (color-coded on horizontal axes) are divided into 101 segments. Arrows point to examples of segments with FAV alterations between conditions and color-coded to indicate direction of change. For paired t-tests, $p < 0.01$, cluster *corr.* for multiple comparisons. ELA, $T_{(11)} = 2.72$, $d \geq 0.79$, n = 12; Std, $T_{(10)} = 2.76$, $d \geq 0.83$, n = 11. See *SI Appendix* **Table S5** for list of domains and segments, and **Table S6A-B** for FAV values.

Figure 6.

Fig. 6. ELA brain states differ from Std at every condition. *(A-C)* Fractional Difference Volumes (FDVs) from segmentation of unpaired SPM t-tests between signal intensities of ELA and Std at *(A)* HC, *(B)* TMT and *(C)* D9 conditions. All ELA images were adjusted for the voxel-wise average differences between groups' pre-Mn(II) images. FDV color-code represents where ELA signal intensities were heightened (yellow) or diminished (blue) relative to Std. Arrows in column graphs point to example segments with notable FDV dynamics as in *(D)*: blue, MS and VMH; red, CEA and CS; green, PA and DMH; black, DP and ARH; and purple, LC. *(D)* Shown are examples of various dynamics in FDV in 9 segments, which were highlighted by arrows in column graphs *(A-C)*. Line graphs show FDVs (vertical axes) of each segment across three successive conditions (horizontal axes). Graphs are grouped and color-coded according to the dynamical pattern of ELA and Std FDV values. *(A-D)* Unpaired t-tests, $p < 0.05$, cluster corr.; HC: $T_{(21)} = 1.72$, $n_{Std} = 11$, $n_{ELA} = 12$; TMT and D9: $T_{(22)} = 1.72$, $n = 12$ per group. *(E)* Three schematics summarizing differences in activity between ELA and Std across the 10 segmented anatomical domains at each condition: HC (*top*), TMT (*middle*) and D9 (*bottom*). Each color gradient corresponds to magnitudes and direction of segmental fractional difference volumes (FDV), and proportion of coloring indicating proportion of segments following that pattern within each domain; See **SI Appendix Movie 1** for a slice-by-slice walkthrough of between group SPMs at each condition projected on brain anatomy, **Table S8** for FDV values and **Table S6** for list of domains and segments.

References

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1. Felitti VJ*, et al.* (1998) Relationship of Childhood Abuse and Household Dysfunction to Many of the Leading Causes of Death in Adults. *American Journal of Preventive Medicine* 14(4):245-258.

2. Merrick M*, et al.* (2019) Vital Signs: Estimated Proportion of Adult Health Problems Attributable to Adverse Childhood Experiences and Implications for Prevention — 25 States, 2015–2017. *MMWR. Morbidity and Mortality Weekly Report* 68.

3. Zarse EM*, et al.* (2019) The adverse childhood experiences questionnaire: Two decades of research on childhood trauma as a primary cause of adult mental illness, addiction, and medical diseases. *Cogent Medicine* 6(1):1581447.

4. Spadoni AD*, et al.* (2022) Contribution of early-life unpredictability to neuropsychiatric symptom patterns in adulthood. *Depression and Anxiety* 39(10-11):706-717.

5. Yehuda R*, et al.* (2010) Putative biological mechanisms for the association between early life adversity and the subsequent development of PTSD. *Psychopharmacology* 212(3):405-417.

6. Chen Y & Baram TZ (2016) Toward Understanding How Early-Life Stress Reprograms Cognitive and Emotional Brain Networks. *Neuropsychopharmacology* 41(1):197-206.

7. Kraaijenvanger EJ*, et al.* (2020) Impact of early life adversities on human brain functioning: A coordinate-based meta-analysis. *Neuroscience & Biobehavioral Reviews* 113:62-76.

8. Malave L, Van Dijk MT, & Anacker C (2022) Early life adversity shapes neural circuit function during sensitive postnatal developmental periods. *Translational Psychiatry* 12(1):306.

9. Garcia-Arocena D (2015) More researchers are using B6J mice than ever before. (The Jackson Laboratory).

10. Laboratory TJ (2024) Mouse Room Conditions.

11. Molet J, Maras PM, Avishai-Eliner S, & Baram TZ (2014) Naturalistic rodent models of chronic early-life stress. *Developmental Psychobiology* 56(8):1675-1688.

12. Rice CJ, Sandman CA, Lenjavi MR, & Baram TZ (2008) A Novel Mouse Model for Acute and Long-Lasting Consequences of Early Life Stress. *Endocrinology* 149(10):4892-4900.

13. Bercum FM, Navarro Gomez MJ, & Saddoris MP (2021) Elevated fear responses to threatening cues in rats with early life stress is associated with greater excitability and loss of gamma oscillations in ventral-medial prefrontal cortex. *Neurobiol Learn Mem* 185:107541.

14. Bolton JL, *et al.* (2018) Anhedonia Following Early-Life Adversity Involves Aberrant
Interaction of Reward and Anxiety Circuits and Is Reversed by Partial Silencing of
Amygdala Corticotropin-Releasing Hormone Gene. *Biological Psychiatry* 83(2):137-147.

15. Bolton JL, *et al.* (2022) Early stress-induced impaired microglial pruning of excitatory
synapses on immature CRH-expressing neurons provokes aberrant adult stress
responses. *Cell Rep* 38(13):110600.

16. Brosens N, Simon C, Kessels HW, Lucassen PJ, & Krugers HJ (2023) Early life stress
lastingly alters the function and AMPA-receptor composition of glutamatergic synapses in
the hippocampus of male mice. *J Neuroendocrinol* 35(12):e13346.

17. de Carvalho G, Khoja S, Haile MT, & Chen LY (2023) Early life adversity impaired dorsal
striatal synaptic transmission and behavioral adaptability to appropriate action selection
in a sex-dependent manner. *Front Synaptic Neurosci* 15:1128640.

18. Guadagno A, *et al.* (2018) Reduced resting-state functional connectivity of the basolateral
amygdala to the medial prefrontal cortex in preweaning rats exposed to chronic early-life
stress. *Brain Struct Funct* 223(8):3711-3729.

19. Guadagno A, Verlezza S, Long H, Wong TP, & Walker CD (2020) It Is All in the Right
Amygdala: Increased Synaptic Plasticity and Perineuronal Nets in Male, But Not Female,
Juvenile Rat Pups after Exposure to Early-Life Stress. *The Journal of neuroscience : the*
official journal of the Society for Neuroscience 40(43):8276-8291.

20. Guadagno A, Wong TP, & Walker CD (2018) Morphological and functional changes in
the preweaning basolateral amygdala induced by early chronic stress associate with
anxiety and fear behavior in adult male, but not female rats. *Prog Neuropsychopharmacol*
Biol Psychiatry 81:25-37.

21. Gunn BG, *et al.* (2013) Dysfunctional astrocytic and synaptic regulation of hypothalamic
glutamatergic transmission in a mouse model of early-life adversity: relevance to
neurosteroids and programming of the stress response. *The Journal of neuroscience :*
the official journal of the Society for Neuroscience 33(50):19534-19554.

22. Johnson FK, *et al.* (2018) Amygdala hyper-connectivity in a mouse model of
unpredictable early life stress. *Transl Psychiatry* 8(1):49.

23. Kooiker CL, Chen Y, Birnie MT, & Baram TZ (2023) Genetic Tagging Uncovers a Robust,
Selective Activation of the Thalamic Paraventricular Nucleus by Adverse Experiences
Early in Life. *Biol Psychiatry Glob Open Sci* 3(4):746-755.

24. Levis SC, *et al.* (2022) Enduring disruption of reward and stress circuit activities by early-
life adversity in male rats. *Transl Psychiatry* 12(1):251.

25. Malter Cohen M*, et al.* (2013) Early-life stress has persistent effects on amygdala
function and development in mice and humans. *Proc Natl Acad Sci U S A* 110(45):18274-
18278.

26. Raineki C, Cortes MR, Belnoue L, & Sullivan RM (2012) Effects of early-life abuse differ
across development: infant social behavior deficits are followed by adolescent
depressive-like behaviors mediated by the amygdala. *The Journal of neuroscience : the*
official journal of the Society for Neuroscience 32(22):7758-7765.

27. Yan CG*, et al.* (2017) Aberrant development of intrinsic brain activity in a rat model of
caregiver maltreatment of offspring. *Transl Psychiatry* 7(1):e1005.

28. Holschneider DP, Guo Y, Mayer EA, & Wang Z (2016) Early life stress elicits visceral
hyperalgesia and functional reorganization of pain circuits in adult rats. *Neurobiol Stress*
3:8-22.

29. Islam R*, et al.* (2024) Early adversity causes sex-specific deficits in perforant pathway
connectivity and contextual memory in adolescent mice. *Biol Sex Differ* 15(1):39.

30. White JD*, et al.* (2020) Early life stress causes sex-specific changes in adult fronto-limbic
connectivity that differentially drive learning. *Elife* 9.

31. Kooiker CL*, et al.* (2024) Paraventricular Thalamus Neuronal Ensembles Encode Early-
life Adversity and Mediate the Consequent Sex-dependent Disruptions of Adult Reward
Behaviors. (Cold Spring Harbor Laboratory).

32. Uselman TW, Barto DR, Jacobs RE, & Bearer EL (2020) Evolution of brain-wide activity
in the awake behaving mouse after acute fear by longitudinal manganese-enhanced MRI.
Neuroimage 222:116975.

33. Uselman TW, Medina CS, Gray HB, Jacobs RE, & Bearer EL (2022) Longitudinal
manganese-enhanced magnetic resonance imaging of neural projections and
activity.e4675.

34. Vousden DA*, et al.* (2018) Continuous manganese delivery via osmotic pumps for
manganese-enhanced mouse MRI does not impair spatial learning but leads to skin
ulceration. *NeuroImage* 173:411-420.

35. Aoki I, Wu YJ, Silva AC, Lynch RM, & Koretsky AP (2004) In vivo detection of
neuroarchitecture in the rodent brain using manganese-enhanced MRI. *Neuroimage*
22(3):1046-1059.

36. Bedenk BT*, et al.* (2018) Mn(2+) dynamics in manganese-enhanced MRI (MEMRI):
Cav1.2 channel-mediated uptake and preferential accumulation in projection terminals.
Neuroimage 169:374-382.

37. Narita K, Kawasaki F, & Kita H (1990) Mn and Mg influxes through Ca channels of motor
nerve terminals are prevented by verapamil in frogs. *Brain research* 510(2):289-295.

38. Lin YJ & Koretsky AP (1997) Manganese ion enhances T1-weighted MRI during brain
activation: an approach to direct imaging of brain function. *Magnetic resonance in*
medicine 38(3):378-388.

39. Svehla P, Bedecarrats A, Jahn C, Nargeot R, & Ciobanu L (2018) Intracellular
manganese enhanced MRI signals reflect the frequency of action potentials in Aplysia
neurons. *J Neurosci Methods* 295:121-128.

40. Yu X, Sanes DH, Aristizabal O, Wadghiri YZ, & Turnbull DH (2007) Large-scale
reorganization of the tonotopic map in mouse auditory midbrain revealed by MRI.
Proceedings of the National Academy of Sciences 104(29):12193-12198.

41. Yu X, Wadghiri YZ, Sanes DH, & Turnbull DH (2005) In vivo auditory brain mapping in
mice with Mn-enhanced MRI. *Nature Neuroscience* 8(7):961-968.

42. Malkova NV, Gallagher JJ, Yu CZ, Jacobs RE, & Patterson PH (2014) Manganese-
enhanced magnetic resonance imaging reveals increased DOI-induced brain activity in a
mouse model of schizophrenia. *Proc Natl Acad Sci U S A* 111(24):E2492-2500.

43. Bearer EL*, et al.* (2007) Role of neuronal activity and kinesin on tract tracing by
manganese-enhanced MRI (MEMRI). *Neuroimage* 37 Suppl 1:S37-46.

44. Bearer EL, Zhang X, & Jacobs RE (2007) Live imaging of neuronal connections by
magnetic resonance: Robust transport in the hippocampal–septal memory circuit in a
mouse model of Down syndrome. *NeuroImage* 37(1):230-242.

45. Bearer EL, Zhang X, Janvelyan D, Boulat B, & Jacobs RE (2009) Reward circuitry is
perturbed in the absence of the serotonin transporter. *Neuroimage* 46(4):1091-1104.

46. Zhang X*, et al.* (2010) Altered neurocircuitry in the dopamine transporter knockout mouse
brain. *PLoS One* 5(7):e11506.

47. Gallagher JJ, Zhang X, Ziomek GJ, Jacobs RE, & Bearer EL (2012) Deficits in axonal
transport in hippocampal-based circuitry and the visual pathway in APP knock-out
animals witnessed by manganese enhanced MRI. *Neuroimage* 60(3):1856-1866.

48. Gallagher JJ*, et al.* (2013) Altered reward circuitry in the norepinephrine transporter
knockout mouse. *PLoS One* 8(3):e57597.

49. Delora A*, et al.* (2016) A simple rapid process for semi-automated brain extraction from
magnetic resonance images of the whole mouse head. *J Neurosci Methods* 257:185-193.

50. Medina CS*, et al.* (2017) Hippocampal to basal forebrain transport of Mn(2+) is impaired
by deletion of KLC1, a subunit of the conventional kinesin microtubule-based motor.
Neuroimage 145(Pt A):44-57.

51. Medina CS, Manifold-Wheeler B, Gonzales A, & Bearer EL (2017) Automated
Computational Processing of 3-D MR Images of Mouse Brain for Phenotyping of Living
Animals. *Curr Protoc Mol Biol* 119:29A 25 21-29A 25 38.

52. Bearer EL, *et al.* (2018) Alterations of functional circuitry in aging brain and the impact of
mutated APP expression. *Neurobiol Aging* 70:276-290.
53. Bearer EL, Barto D, & Jacobs RE (2019) Imaging the evolution acute fear: Longitudinal
whole brain imaging in living mice of neural activity with MEMRI. *Proceedings of the*
International Society for Magnetic Resonance in Medicine... Scientific Meeting and
Exhibition. International Society for Magnetic Resonance in Medicine. Scientific Meeting
and Exhibition, (NIH Public Access).
54. Medina CS, *et al.* (2019) Decoupling the Effects of the Amyloid Precursor Protein From
Amyloid-beta Plaques on Axonal Transport Dynamics in the Living Brain. *Front Cell*
Neurosci 13:501.
55. Phinyomark A, Ibanez-Marcelo E, & Petri G (2017) Resting-State fMRI Functional
Connectivity: Big Data Preprocessing Pipelines and Topological Data Analysis. *IEEE*
Transactions on Big Data 3(4):415-428.
56. Flandin G & Friston K (2008) Statistical parametric mapping (SPM). *Scholarpedia*
3(4):6232.
57. Penny WD, Friston KJ, Ashburner JT, Kiebel SJ, & Nichols TE (2011) *Statistical*
Parametric Mapping: The Analysis of Functional Brain Images (Elsevier Science).
58. Pallast N, *et al.* (2019) Processing Pipeline for Atlas-Based Imaging Data Analysis of
Structural and Functional Mouse Brain MRI (AIDAmri). *Frontiers in Neuroinformatics* 13.
59. Fendt M, Endres T, Lowry CA, Apfelbach R, & McGregor IS (2005) TMT-induced
autonomic and behavioral changes and the neural basis of its processing. *Neuroscience*
and biobehavioral reviews 29(8):1145-1156.
60. Rosen J, Asok A, & Chakraborty T (2015) The smell of fear: innate threat of 2,5-dihydro-
2,4,5-trimethylthiazoline, a single molecule component of a predator odor. 9.
61. Gallo M, *et al.* (2019) Limited Bedding and Nesting Induces Maternal Behavior
Resembling Both Hypervigilance and Abuse. 13.
62. Matsukawa M, Yoshikawa M, Katsuyama N, Aizawa S, & Sato T (2022) The Anterior
Piriform Cortex and Predator Odor Responses: Modulation by Inhibitory Circuits. *Front*
Behav Neurosci 16:896525.
63. Matsumoto H, *et al.* (2010) Spatial Arrangement of Glomerular Molecular-Feature
Clusters in the Odorant-Receptor Class Domains of the Mouse Olfactory Bulb.
103(6):3490-3500.
64. Kimbrough A, *et al.* (2020) Brain-wide functional architecture remodeling by alcohol
dependence and abstinence. 117(4):2149-2159.

65. Motta SC*, et al.* (2009) Dissecting the brain's fear system reveals the hypothalamus is
critical for responding in subordinate conspecific intruders. *Proceedings of the National*
Academy of Sciences 106(12):4870-4875.

66. Greene AS, Horien C, Barson D, Scheinost D, & Constable RT (2023) Why is everyone
talking about brain state? *Trends Neurosci* 46(7):508-524.

67. Ben-Shaul Y, Katz LC, Mooney R, & Dulac C (2010) In vivo vomeronasal stimulation
reveals sensory encoding of conspecific and allospecific cues by the mouse accessory
olfactory bulb. 107(11):5172-5177.

68. Canteras NS (2002) The medial hypothalamic defensive system: Hodological
organization and functional implications. *Pharmacology Biochemistry and Behavior*
71(3):481-491.

69. Canteras NS, Chiavegatto S, Ribeiro do Valle LE, & Swanson LW (1997) Severe
Reduction of Rat Defensive Behavior to a Predator by Discrete Hypothalamic Chemical
Lesions. *Brain Research Bulletin* 44(3):297-305.

70. Janitzky K, D'Hanis W, Kröber A, & Schwegler H (2015) TMT predator odor activated
neural circuit in C57BL/6J mice indicates TMT-stress as a suitable model for
uncontrollable intense stress. *Brain research* 1599:1-8.

71. Kunwar PS*, et al.* (2015) Ventromedial hypothalamic neurons control a defensive
emotion state. *eLife* 4:e06633.

72. Pérez-Gómez A*, et al.* (2015) Innate Predator Odor Aversion Driven by Parallel Olfactory
Subsystems that Converge in the Ventromedial Hypothalamus. *Current Biology*
25(10):1340-1346.

73. Koob GF & Volkow ND (2016) Neurobiology of addiction: a neurocircuitry analysis. *The*
lancet. Psychiatry 3(8):760-773.

74. Russo SJ & Nestler EJ (2013) The brain reward circuitry in mood disorders. *Nature*
Reviews Neuroscience 14(9):609-625.

75. Silva BA, Gross CT, & Graff J (2016) The neural circuits of innate fear: detection,
integration, action, and memorization. *Learn Mem* 23(10):544-555.

76. Coletta L*, et al.* (2020) Network structure of the mouse brain connectome with voxel
resolution. 6(51):eabb7187.

77. Salvan P*, et al.* (2023) Serotonin regulation of behavior via large-scale neuromodulation
of serotonin receptor networks. *Nature Neuroscience* 26(1):53-63.

78. Taylor NL*, et al.* (2022) Structural connections between the noradrenergic and cholinergic
system shape the dynamics of functional brain networks. *NeuroImage* 260:119455.

79. Zerbi V*, et al.* (2019) Rapid Reconfiguration of the Functional Connectome after
Chemogenetic Locus Coeruleus Activation. *Neuron* 103(4):702-718.e705.

80.   David & Adolphs R (2014) A Framework for Studying Emotions across Species. *Cell*
803   157(1):187-200.
804   81.   Flavell SW, Gogolla N, Lovett-Barron M, & Zelikowsky M (2022) The emergence and
805   influence of internal states. *Neuron* 110(16):2545-2570.
806   82.   Jaric I*, et al.* (2022) The rearing environment persistently modulates mouse phenotypes
807   from the molecular to the behavioural level. *PLoS Biol* 20(10):e3001837.
808   83.   Jaric I*, et al.* (2024) Using mice from different breeding sites fails to improve replicability
809   of results from single-laboratory studies. *Lab Anim (NY)* 53(1):18-22.
810   84.   Usui T, Macleod MR, McCann SK, Senior AM, & Nakagawa S (2021) Meta-analysis of
811   variation suggests that embracing variability improves both replicability and
812   generalizability in preclinical research. *PLoS Biol* 19(5):e3001009.
813   85.   Chang C-H & Gean P-W (2019) The Ventral Hippocampus Controls Stress-Provoked
814   Impulsive Aggression through the Ventromedial Hypothalamus in Post-Weaning Social
815   Isolation Mice. *Cell Reports* 28(5):1195-1205.e1193.
816   86.   Manzano Nieves G, Bravo M, & Bath KG (2023) Early life adversity ablates sex
817   differences in active versus passive threat responding in mice. *Stress*:1-32.
818   87.   Lehallier B*, et al.* (2012) Brain Processing of Biologically Relevant Odors in the Awake
819   Rat, as Revealed by Manganese-Enhanced MRI. *PLoS ONE* 7(10):e48491.
820   88.   Petrus E, Saar G, Daoust A, Dodd S, & Koretsky AP (2021) A hierarchy of manganese
821   competition and entry in organotypic hippocampal slice cultures. *NMR in Biomedicine*
822   34(4).
823   89.   Bartelle BB, Szulc KU, Suero-Abreu GA, Rodriguez JJ, & Turnbull DH (2013) Divalent
824   metal transporter, DMT1: A novel MRI reporter protein. *Magnetic resonance in medicine*
825   70(3):842-850.
826   90.   Spring S, Lerch JP, & Henkelman RM (2007) Sexual dimorphism revealed in the
827   structure of the mouse brain using three-dimensional magnetic resonance imaging.
828   *NeuroImage* 35(4):1424-1433.
829   91.   Bruce MR*, et al.* (2021) Sexually dimorphic neuroanatomical differences relate to ASD-
830   relevant behavioral outcomes in a maternal autoantibody mouse model. *Molecular*
831   *Psychiatry* 26(12):7530-7537.
832   92.   Holz NE*, et al.* (2023) A stable and replicable neural signature of lifespan adversity in the
833   adult brain. *Nature Neuroscience*.
834   93.   Lerch JP*, et al.* (2011) Maze training in mice induces MRI-detectable brain shape
835   changes specific to the type of learning. *NeuroImage* 54(3):2086-2095.
94.   Ashburner J*, et al.* (2012) SPM8 manual.
95.   Ashburner J*, et al.* (2014) SPM12 manual.

96.   Carter CS, Lesh TA, & Barch DM (2016) Thresholds, Power, and Sample Sizes in
Clinical Neuroimaging. *Biological Psychiatry: Cognitive Neuroscience and Neuroimaging*
1(2):99-100.

97.   Mumford JA (2012) A power calculation guide for fMRI studies. *Social Cognitive and*
Affective Neuroscience 7(6):738-742.

98.   Vandekar SN & Stephens J (2021) Improving the replicability of neuroimaging findings by
thresholding effect sizes instead of p-values. 42(8):2393-2398.

99.   Hammelrath L*, et al.* (2016) Morphological maturation of the mouse brain: An in vivo MRI
and histology investigation. *NeuroImage* 125:144-152.

100.   Brigman JL, Mathur P, Lu L, Williams RW, & Holmes A (2009) Genetic relationship
between anxiety-related and fear-related behaviors in BXD recombinant inbred mice.
20(2):204-209.

101.   Noldus LPJJ, Spink AJ, & Tegelenbosch RAJ (2001) EthoVision: A versatile video
tracking system for automation of behavioral experiments. *Behavior Research Methods,*
Instruments, & Computers 33(3):398-414.

102.   Fox J & Weisberg S (2019) An R Companion to Applied Regression.

103.   Lenth RV (2022) emmeans: Estimated Marginal Means, aka Least-Squares Means.

104.   Pinheiro J, Bates D, & Team RC (2022) nlme: Linear and Nonlinear Mixed Effects
Models.

105.   Pinheiro JC & Bates DM (2000) Mixed-Effects Models in S and S-PLUS.

106.   Posit t (2022) RStudio: Integrated Development Environment for R.

107.   Team RC (2022) R: A Language and Environment for Statistical Computing.

108.   Wickham H (2016) ggplot2: Elegant Graphics for Data Analysis.

109.   Wickham H*, et al.* (2019) Welcome to the Tidyverse. 4(43):1686.

110.   Tyszka JM, Readhead C, Bearer EL, Pautler RG, & Jacobs RE (2006) Statistical diffusion
tensor histology reveals regional dysmyelination effects in the shiverer mouse mutant.
NeuroImage 29(4):1058-1065.