Finding Drug Repurposing Candidates for Neurodegenerative Diseases using Zebrafish Behavioral Profiles

Thaís Del Rosario Hernández*, Sayali V Gore, Jill A Kreiling, Robbert Creton

Department of Molecular Biology, Cell Biology and Biochemistry, Brown University, Providence, Rhode Island, USA

Corresponding author: Thais_Del_Rosario@brown.edu

Abstract

Drug repurposing can accelerate drug development while reducing the cost and risk of toxicity typically associated with de novo drug design. Several disorders lacking pharmacological solutions and exhibiting poor results in clinical trials - such as Alzheimer's disease (AD) - could benefit from a cost-effective approach to finding new therapeutics. We previously developed a neural network model, Z-LaP Tracker, capable of quantifying behaviors in zebrafish larvae relevant to cognitive function, including activity, reactivity, swimming patterns, and optomotor response in the presence of visual and acoustic stimuli. Using this model, we performed a high-throughput screening of FDA-approved drugs to identify compounds that affect zebrafish larval behavior in a manner consistent with the distinct behavior induced by calcineurin inhibitors. Cyclosporine (CsA) and other calcineurin inhibitors have garnered interest for their potential role in the prevention of AD. We generated behavioral profiles suitable for cluster analysis, through which we identified 64 candidate therapeutics for neurodegenerative disorders.

Introduction

Alzheimer's Disease (AD) is the most common cause of dementia among the aging population("2023 Alzheimer's disease facts and figures," 2023), characterized by senile plaques resulting from accumulation of extracellular amyloid β (Aβ), the formation of intracellular neurofibrillary tangles (NFT) through tau hyperphosphorylation, and overall neuronal degeneration(Anand et al., 2014; Braak & Braak, 1991). Symptoms of AD do not emerge until late in the disease course, obscuring the underlying mechanisms of AD pathology ("2023 Alzheimer's disease facts and figures," 2023). There is an absence of disease-modifying therapeutics, and current FDA-approved drugs for AD only provide symptomatic relief or marginally prevent the progression of the disease (Mullane & Williams, 2013). Although there is a growing need for both preventative and remedial AD treatments, clinical trials are majorly unsuccessful in remedying AD symptomatology (Mullane & Williams, 2013).

Drug discovery is an extensive and expensive process that often results in compounds that do not make it to market due to safety and/or efficacy concerns, adverse side effects, and incompatibility with the comorbidities present in the target population (Arrowsmith & Harrison, 2012). Drug repurposing is a valuable method to take advantage of the additional targets of currently approved drugs that have already been determined safe for human use. It has been used as a powerful tool to find alternative uses for pharmaceutical compounds that are already on the market, for diseases such as Parkinson's disease (amantadine), tuberculosis (cycloserine), and attention deficit hyperactivity disorder (atomoxetine)(Al-Subari et al., 2020; Hauser et al., 2017; Roy & Chaguturu, 2017). Cyclosporine A (CsA) is one such drug repurposing candidate in the context of AD. CsA is a calcineurin inhibitor currently used for chronic immunosuppression to prevent allograft rejection (Taglialatela et al., 2015). A human population study found that organ transplant recipients maintained on calcineurin inhibitors, including CsA, had lower incidences of AD compared to the general population (Taglialatela et al., 2015).

We previously developed a deep neural network model Z-LaP Tracker, based on the markerless position estimation software DeepLabCut, to quantify relevant behaviors in zebrafish larvae (Gore et al., 2023). Zebrafish are an established model for the study of a wide variety of pathologies, including neurodegenerative disorders, in part due to their homology with the human genome (Howe et al., 2013; Kalueff et al., 2014). They have comparable behavioral responses to mammalian systems, exhibiting a wide range of behaviors quantifiable by well-established behavioral test batteries (Kalueff et al., 2013; Kalueff et al., 2014; Norton, 2013). In particular, zebrafish larvae are an attractive model for high throughput screens of sizable compound libraries. Small molecule drugs can be easily administered through the water and the larvae can be placed in multi-well plates for phenotypic screening. In the current study, we used a high throughput, whole-organism approach to screen a small-molecule library of FDA-approved drugs and identify CsA-like compounds (Fig 1). We generated a list of potential drug repurposing candidates for the prevention and treatment of neurological disorders, including AD.

Results

Behavioral Screening of FDA-Approved Drugs

Zebrafish larvae were exposed to the 876 compounds included in the Cayman Chemical FDA-approved Drug Library for a total of 6 hours. Each compound was evaluated using 48 zebrafish larvae, and each experiment included DMSO and egg water controls. In total, we examined 50,496 larvae: 42,048 larvae treated with small-molecule compounds, and 8,448 DMSO-treated larvae. Zebrafish larvae were exposed to all compounds and controls at a concentration of 10μM for 6 hours. At this concentration 23 compounds

resulted in a ≥50% mortality rate, but none of the compounds had a 100% mortality rate. We did not include larvae if they moved less than 1% of the time throughout the experiment. Additionally, low likelihood (< 0.50) data points were filtered out and not included in behavioral profile generation. We generated behavioral profiles by comparing differences in behavior between DMSO-vehicle controls and each compound.

Our behavioral assay features a 3-hour Microsoft PowerPoint presentation with an initial 60 minutes of no stimuli, 80 minutes of moving lines - alternating direction every 10 minutes and color every 20 minutes -, followed by a 10 minute period with no stimuli, and 30 minutes of sound stimuli alternating frequency every 10 minutes (Fig 2A). This assay is analyzed in 10-minute intervals called periods, and behaviors are averaged per period.

The resulting behavioral profiles consisted of 25 behaviors representing overall activity, reactivity, swimming patterns, and optomotor response (Fig 2B). Namely, we measured (1) Activity during the 1st hour, (2) Activity during Period 15, (3) Habituation, (4) Startle response, (5) Excitability, (6) Optomotor response to moving red lines, (7) Optomotor response to moving green lines, (8) Optomotor response to moving blue lines, (9) Optomotor response to faster moving red lines, (10) Combined optomotor response to red, green, and blue moving lines, (11) Scoot movement during the 1st hour, (12) Scoot movement during the presentation of moving lines of any color or speed, (13) Burst movement during the first hour, (14) Burst movement during the presentation of moving lines of any color or speed, (15) Percent edge location during the first hour, (16) Percent edge location during the presentation of moving lines of any color or speed, (17) Percent clockwise orientation during the 1st hour, (18) Percent clockwise orientation during the presentation of moving lines of any color or speed, (19) Upward orientation during moving red lines, (20) Upward orientation during moving green lines, (21) Upward orientation during moving blue lines, (22) Upward orientation during faster moving red lines, (23) Combined upward orientation during red, green, and blue moving lines, (24) Turn angle during the 1st hour, and (25) Absolute turn angle during the 1st hour (Supplementary Table 1).

Fig. 2. Overview of 25 parameters measured during 3-hour behavioral assay. (A) 3-hour timeline of the presentation shown to treated zebrafish larvae. 25 parameters of behavior are measured during 18 periods of visual and acoustic stimuli. **(B)** Behavioral profiles of treatments using 12 compounds from the Cayman Chemical FDA-approved Drug Library. Each treatment is compared to DMSO-vehicle controls and the resulting differences are color-coded based on an increase (red) or decrease (green) in a particular behavioral parameter. The values represent differences as compared to DMSO controls in percentage points. Treatments are identified by the Cayman Chemical library's plate number (M1-M11) and well number (A1-H11).

Clustering Results

K-means Cluster Analysis

We used the K-means clustering algorithm to identify compounds in the Cayman Chemical FDA-approved Drug Library that would cluster together with calcineurin-inhibitor CsA. We identified the optimal number of clusters (k = 4) using the elbow

method. We found that CsA clusters with 58 other compounds (Fig 3). The overall behavioral profile of CsA-like compounds features increased activity during the first hour and period 15, decreased habituation and startle response, increased excitability, decreased optomotor response during all visual stimuli, increased Scoot movement, and a reduced orientation response during all visual stimuli (Fig 4). The Pearson correlation value for this cluster was 0.62.

Fig. 3. K-means cluster analysis. 876 compound treatments and DMSO controls were assigned to k=4 clusters. Cluster 4 contains cyclosporine A and 58 compound treatments with similar behavioral profiles.

Fig. 4. Behavioral profiles of CsA-like compounds. Identified 64 compounds that induce behavioral profiles similar to CsA when administered to 5 dpf zebrafish larvae. Each behavioral profile is composed of 25 parameters measuring activity, reactivity, swimming patterns, and optomotor response.

Hierarchical Cluster Analysis

We used agglomerative hierarchical clustering to group the behavioral profiles generated by exposure to the Cayman Chemical FDA-approved Drug Library. We found that CsA and other compounds with similar behavioral profiles formed a distinct cluster featuring the same behavioral patterns found in our previous screening of FDA-approved drugs (Fig 5A) (Tucker Edmister, Del Rosario Hernández, et al., 2022). We identified 53 CsA-like compounds, of which 47 were also found with the K-means clustering method (Fig 5B). The correlation value for this cluster was 0.60. Multiple CsA-like subclusters with increasing correlation values can be identified in our hierarchical analysis (Supplementary Figure 1).

Fig. 5. Hierarchical cluster analysis. (A) Overview of the behavioral profiles elicited by 876 compound treatments. A high-resolution version of this image is included in the supplementary information (Supplementary Figure 2). **(B)** Cluster of 53 compounds that induce CsA-like behavioral profiles. Red indicates an increase in

a behavioral value relative to DMSO controls, while green indicates a decrease. **(C)** Pearson pairwise correlations of CsA and the 47 CsA-like compounds identified by both K-means and hierarchical clustering.

Using K-means and hierarchical cluster analysis, we found a total of 64 compounds displaying CsA-like behavioral paradigms (Table 1). These 64 compounds affect manifold biological functions and are used to treat a wide range of diseases. We further analyzed the composition of our clusters of interest by calculating the Pearson correlation between the behavioral profiles of the compounds in the Cayman Chemical FDA-approved Drug Library and CsA. We visualized the 47 compounds found by both clustering methods showing their degree of similarity to CsA (Fig 5C).

Table 1. List of compounds inducing CsA-like behavioral profiles, identified by K-means and hierarchical clustering analyses.

Mode of action	Type	Receptor subtype	ID	Compound Name	Clustering method
Dopamine	Antagonist	D1/D2	M8H2	Fluphenazine (hydrochloride)	Both
			M7D4	Perphenazine	Both
		D2	M3H3	Ziprasidone (hydrochloride hydrate)	Both
			M7D9	Loxapine (succinate)	Both
		D2/D3	M4C11	Paliperidone	K-Means
		Non-selective	M1G5	Haloperidol	Both
			M2H8	Thioridazine (hydrochloride)	K-Means
			M6E8	Droperidol	Both
			M11C9	(±) - Asenapine	Both
	Agonist	D2	M3B9	Bromocriptine (mesylate)	Both
			M6H6	Aripiprazole	Both
	Inhibitor	Dopamine-β-hydroxylase	M4A10	Disulfiram	Both
	Dual Action	Non-selective	M11A9	Ergoloid (mesylates)	Hierarchical Clustering
Adrenergic Receptors	Agonist	β2-Adrenergic Receptors	M4D5	Formoterol (hemifumarate hydrate)	Both
			M4D6	Isoproterenol (hydrochloride)	Hierarchical Clustering
			M10D8	Arformoterol (tartrate)	Both
			M10G3	Salbutamol (hydrochloride)	K-Means
			M11A2	Dipivefrin (hydrochloride)	Both
		α-Adrenergic Receptors	M7G9	(±)-Epinephrine (hydrochloride)	Hierarchical Clustering
	Antagonist	α1A-Adrenergic Receptors	M3H7	Trifluoperazine (hydrochloride)	Both
			M6D5	Doxazosin (mesylate)	Both
		α2-Adrenergic Receptors	M6H7	Mirtazapine	K-Means
Calcium Channels and	Inhibitor	Calcineurin	M1G11	Cyclosporin A	Both
			M1H10	Pimecrolimus	Both

(Continued)

Table 1. Continued

Mode of action	Type	Receptor subtype	ID	Compound Name	Clustering method
Related Pathways			M11F7	FK-506 (Tacrolimus)	Both
		L-type Calcium Channel	M3B3	Nimodipine	Both
			M4A6	Amiodarone (hydrochloride)	Both
			M5H3	Nicardipine (hydrochloride)	Both
			M11C10	Dronedarone (hydrochloride)	Both
		Calmodulin	M3E3	Loperamide (hydrochloride)	Both
(Steroid) Hormones	Modulator	Estrogen	M1E2	Tamoxifen (citrate)	Both
			M7E7	Toremifene (citrate)	Both
	Activator	Estrogen	M6D7	Mitotane	Both
		Progesterone	M8C8	Megestryl Acetate	Both
	Agonist	Progesterone	M9A6	Desogestrel	Both
5-HT (Serotonin) Receptors	Agonist	5-HT1B/5-HT1D	M10C4	Dihydroergotamine (mesylate)	Hierarchical Clustering
	Inhibitor	5-HT2A	M1G7	Clozapine	Both
			M5G7	Amoxapine	Hierarchical Clustering
			M11H11	Nefazodone (hydrochloride)	Both
		5-HT1A/5-HT2A	M6F4	Flibanserin	Both
		5-HT1A	M8F5	Brexpiprazole	Both
PPARs	Agonist	PPARγ	M1A3	Trepostinil	K-Means
			M1F6	Rosiglitazone (maleate)	Both
			M11C2	Rosiglitazone	Both
		PPARα	M11E7	Fenofibrate	Both
COX	Inhibitor	COX-1/COX-2	M4C7	Celecoxib	Both
			M11B3	(±)-Flurbiprofen	Both
		COX-2	M11F11	Oxaprozin	Both
Angiotensin Receptors	Antagonist	AT1	M1D2	Eprosartan (mesylate)	K-Means
			M1D11	Telmisartan	Both

Table 1. Continued

Mode of action	Type	Receptor subtype	ID	Compound Name	Clustering method
VMAT	Inhibitor	VMAT1/VMAT2	M5C2	Reserpine	Both
		VMAT2	M7B9	Tetrabenazine	Both
Retinoids	Agonist	Non-selective RAR	M7E6	Acitretin	Both
		RARa, RARb, RARg	M8H3	Tazarotene	Both
ACh	Blocker		M3C2	Meclizine (hydrochloride)	Both
	Agonist	nAChRs	M5C8	(±)-Nicotine	Both
Copper	Chelator	Copper(II)	M10G9	Trientine (hydrochloride)	Both
			M7G2	Bacitracin (zinc)	Both
CMV	Inhibitor	DNA Terminase Complex	M5H5	Letermovir	K-Means
		Viral DNA Polymerase	M9H8	L-Valacyclovir (hydrochloride)	Hierarchical Clustering
Thrombopoietin Receptors	Agonist		M2A8	Eltrombopag	K-Means
PDE	Inhibitor	PDE3A	M3H4	Cilostazol	K-Means
5α-Reductase	Inhibitor		M4G6	Dutasteride	Both
Alkylating Agents	Alkylation	DNA	M9E9	Chlorambucil	K-Means
Vitamin D	Agonist	Vitamin D3 Receptor	M11D3	25-hydroxy Vitamin D3 (Calcifediol)	K-Means

Novel Compounds Displaying CsA-like Behavior

Our K-means clustering and Hierarchical clustering analyses revealed a total of 64 compounds with behavioral paradigms similar to CsA. Of these, 47 were found in both cluster analysis methodologies. We performed statistical analyses on the effects of these compounds in larval behavior. When compared against DMSO-vehicle controls, 89% of the CsA-like compounds induced statistically significant changes in behavior. These changes were observed throughout most of the 25 behavioral measures, excluding clockwise movements and turn angle. Overall, 40% of the compounds screened in this study elicited at least one significantly different behavior compared to DMSO controls. The statistical analyses performed on the behavioral profiles of all the compounds found in the Cayman Chemical FDA-approved Drug Library can be found in Supplementary Table 2.

Identification of Predominant Targets and Pathways in CsA-like Clusters

To further investigate potential similarities between the identified CsA-like compounds, we queried the Disease-Gene Interaction Database (DGIdb), Therapeutic Targets Database (TTD), Guide to Pharmacology (GtoPdb), Kyoto Encyclopedia of Genes (KEGG), Protein ANalysis THrough Evolutionary Relationships (PANTHER), WikiPathways, and Reactome databases and matched each compound in the Cayman Chemicals Library of FDA-approved Drugs to primary and secondary target genes, as well as their respective molecular pathways and associated mechanisms of action. Our multi-database exploration approach enabled us to fill in information gaps between sources and resulted in an unbiased collection of information, which would otherwise not be possible without an extensive literature review.

We compared the most common molecular targets perturbed by the cluster of CsA-like compounds to the overall target composition of all the compounds in the Cayman Chemical FDA-approved Drug Library. We found that the CsA-like cluster compounds acted on an entirely different composition of targets than the predominant targets found in the compilation of all compounds in the small-molecule library. Specifically, the most common targets of the entire library were genes encoding P450 enzymes (Fig 6A). In contrast, the CsA-like drugs predominantly target genes related to neuromodulation (dopamine receptors, serotonin receptors, adrenergic receptors) or associated with neurodegenerative disorders (ATXN2, KCNH2, mTOR) (Fig 6B).

Fig. 6. Predominant targets and pathways perturbed by CsA-like compounds. (A) Top 10 targets affected by the 876 compounds in the Cayman Chemical FDA-approved Drug Library. **(B)** Top 10 targets affected by CsA and the 64 CsA-like compounds found in our clustering analyses. **(C)** Top 25 Wikipathways, Reactome, KEGG, and PANTHER pathways containing targets affected by CsA-like compounds.

We queried all the targets affected by the 64 CsA-like compounds and matched them to biological pathways from Wikipathways, Reactome, KEGG, and PANTHER (Fig 6C). We selected the top 25 pathways by percentage of CsA-like compounds compared to other compounds not found in our clustering analyses. The pathways' categories correspond with the predominant targets from our previous query, continuing to demonstrate an emphasis on neurological function and modulation.

Additionally, we utilized Ingenuity Pathway Analysis (IPA) to map the predicted relationships between CsA-like compounds and AD-related targets. We generated a custom pathway and overlaid connections with the Molecule Activity Prediction (MAP) tool available in the IPA software (Fig 7). The resulting network predicts a heavily inhibited AD node through diverse downstream effects triggered by the activation of CsA-like molecules.

Fig. 7. IPA analysis showing the predicted effect of CsA-like compounds on Alzheimer's disease. Orange lines and nodes indicate predicted activation, blue lines and nodes indicate predicted inhibition, and yellow lines indicate inconsistent findings. Red nodes indicate activation through custom input using the Molecule Activity Predictor (MAP). Purple outlines indicate AD-related nodes.

Discussion

In this study, we evaluated 876 FDA-approved compounds in 5dpf zebrafish larvae. We generated behavioral paradigms for each compound and clustered them to find CsA-like compounds. Using K-means and hierarchical clustering, we found a total of 64 compounds that evoke CsA-like behavior in 5 dpf zebrafish larvae. We chose to use two clustering analyses to classify our behavioral profiles. Due to the large number of measurements obtained in the current study, we sought to reduce the number of volatile factors during our analysis - factors such as overrepresentation of behaviors and outliers within the data could all have an unexpected effect on clustering results. We employed and compared the results of both clustering methods to maximize the generation of recognizably stable patterns from our behavioral data. Since each method employs different criteria to cluster the behavioral profiles of FDA-approved drugs, they serve as mutual confirmation of their results - evidenced by the high amount of overlap between the identified clusters. They also provide us with more conservative measures to evaluate the produced clusters, such as a list of compounds whose patterns persisted through clustering methods.

In a previous study, we screened 190 compounds included in the Tocriscreen FDA-Approved Drugs Library using the same methods described in the current study and found 32 compounds displaying CsA-like behavior (Gore et al., 2023). These compounds act on a variety of molecular targets, pathways, and diseases, yet induce analogous patterns in zebrafish larval behavior during our 25-behavior screening. During our current screening of 876 compounds, we found a total of 64 compounds displaying CsA-like behavior, of which 11 were previously found in our previous screening, and 2 were not present during our current screening. There are 19 compounds that were included in both screenings, but only found to cluster with CsA during our previous screening. There is a majority but not a complete overlap between the compounds found in the Tocriscreen and the Cayman Chemical libraries, given that a total of 172 compounds are shared between the two libraries. This gives rise to some questions about the reproducibility of the "missing" compounds - whether they truly display a CsA-like behavioral profile. A possible explanation may be that some of these compounds display behavioral profiles akin to other compounds within their target group, which is made more evident in a larger screening and results in tighter biological classification-based clusters, and the rest induce behaviors highly deviated from the range typically found in CsA-like compounds (Supplementary Figure 3).

We classified the 64 CsA-like compounds found during our clustering analyses and noticed that they targeted 5 main categories: dopamine receptors, adrenergic receptors, calcium channels and related pathways, steroid hormones, and 5-HT receptors. To further investigate the clinical significance of our results, we manually curated literature pertaining to these 64 compounds in association with AD or Alzheimer's-like pathology (Table 2).

Table 2. Literature search for CsA-like compounds' association with Alzheimer's disease.

Compound	Description	Effect	Link to Alzheimer's Disease
Fluphenazine	D1/D2 Receptor Antagonist	-	
Perphenazine	D1/D2 Receptor Antagonist	↑	Suppressed tau-induced lethargy, tau aggregation, and neuron loss in *C. elegans (McCormick et al., 2013).*
Ziprasidone	D2 Receptor Antagonist	-	
Loxapine	D2 Receptor Antagonist	-	
Paliperidone	D2/D3 Receptor Antagonist	-	
Haloperidol	Non-selective Dopamine Receptor Antagonist	↑	Reduced tau phosphorylation in a tau mouse model(Koppel et al., 2016). Inhibited Aβ formation in cultured mammalian cells (Higaki et al., 1997).
Thioridazine	Non-selective Dopamine Receptor Antagonist	-	
Droperidol	Non-selective Dopamine Receptor Antagonist	↑	Inhibited Aβ formation in cultured mammalian cells (Higaki et al., 1997).
Asenapine	Non-selective Dopamine Receptor Antagonist	-	
Bromocriptine	D2 Receptor Agonist	↑	Improved Aβ1-42 induced neuroinflammation, neuronal apoptosis, and memory deficits in mice(Liu et al., 2023). Reduced Aβ-42 in human iPSC-derived neurons(Kondo et al., 2017). Inhibited the binding of Aβ oligomers to EphB2(Suzuki et al., 2016).
Aripiprazole	D2 Receptor Agonist	↑	Decreased Aβ accumulation and inhibited neuroinflammation in the brains of 5xFAD mice (Jeong et al., 2021). Inhibited Aβ and P-tau in N2a Swe cells(Heo et al., 2020).
Disulfiram	Dopamine-β-hydroxylase inhibitor	↑	Prevented Aβ aggregation SH-SY5Y human neuronal cells; also reduced plaque-burden in the dentate gyrus and ameliorated behavioral deficits in 5xFAD mice (Reinhardt et al., 2018).

(Continued)

Table 2. Continued

Compound	Description	Effect	Link to Alzheimer's Disease
Ergoloid	Non-selective Dopamine Receptor Effector	↑	FDA-approved drug to improve cognitive function in AD, with varying efficacy(Schneider et al., 2000).
Formoterol	β2-adrenergic Receptor Agonist	↑	Improved cognition and decreased oxidative stress, neuro-inflammation, and apoptotic parameters in streptozotocin-induced sporadic AD mouse model(Abdel Rasheed et al., 2018).
Isoproterenol	β2-adrenergic receptor agonist	↑↓	Restored lysosomal proteolysis, calcium homeostasis, and normal autophagy flux in PSEN1 Knock-out cells and fibroblasts from PSEN1 familial AD patients (Lee et al., 2020). Injection into the basolateral amygdala rescues the memory deficit caused by Aβ in rats(Ishikawa et al., 2022). Reduced intracellular Zn2+ level increased by Aβ in mouse brain(Kawano et al., 2022). Bilateral injection into rat hippocampus results in hyper phosphorylation of tau and disturbance of spatial memory retention(Sun et al., 2005).
Arformoterol	β2-adrenergic Receptor Agonist		-
Salbutamol	β2-adrenergic Receptor Agonist	↑	Impeded the aggregation of tau in vitro(Townsend et al., 2020).
Dipivefrin	β2 adrenergic Receptor Agonist		-
Epinephrine	α-adrenergic Receptor Agonist		-
Trifluoperazine	α1A-adrenergic Receptor antagonist	↑	Inhibited H2O2-induced cell viability loss, intracellular reactive oxygen species (ROS) generation, and reduced cell apoptosis in H2O2 in PC12 cells(Liu et al., 2011).
Doxazosin	α1A-adrenergic Receptor Antagonist	↑	Prevented GSK-3β activation and Tau hyper phosphorylation on an in vitro model of organotypic hippocampal cultures exposed to amyloid-β(Coelho et al., 2019).
Mirtazapine	α2-adrenergic Receptor		-

Table 2. Continued

Compound	Description	Effect	Link to Alzheimer's Disease
Pimecrolimus	Calcineurin Inhibitor	↑	Reduced Aβ secretion in AD model neurons(Nishioka et al., 2016).
Tacrolimus	Calcineurin Inhibitor	↑	Prevented age- and AD-associated microstructural changes in the hippocampus, parahippocampal cortex, and prefrontal cortex of the middle-aged beagle brain(Hamsanandini et al., 2021). Reversed learning and memory impairment caused by Aβ accumulation in Tg2576 APP mouse model(Dineley et al., 2007). Ameliorated plaque-associated synapse loss in plaque bearing mouse model(Rozkalne et al., 2011).
Nimodipine	L-type Calcium Channel Inhibitor	↑↓	Protected microglia from Aβ-dependent cytotoxicity and inhibited Aβ-stimulated IL-1β synthesis in vivo(Sanz et al., 2012). No effect on amyloid pathology of 5xFAD mice(Sadleir et al., 2022). Facilitated the clearance of Aβ across the BBB in an in vitro model(Bachmeier et al., 2011).
Amiodarone	L-type Calcium Channel Inhibitor	↑	Inhibited β-secretase cleavage of APP and Aβ generation in HEK293-APP cells (Mitterreiter et al., 2010).
Nicardipine	L-type Calcium Channel Inhibitor	↑	Facilitated the clearance of Aβ across the BBB in an in vitro model(Bachmeier et al., 2011).
Dronedarone	L-type Calcium Channel Inhibitor		-
Loperamide	Calmodulin Inhibitor		-
Tamoxifen	Estrogen Modulator	↑	Enhanced spatial and contextual memory and increased ACh levels in Aβ1–42 injected-breeding-retired-female mice(Pandey et al., 2016). Aβ induced cell death in a mouse HT-22 cell line(Gursoy et al., 2002). Long-term use of tamoxifen in patients with breast cancer is associated with a lower risk of dementia(Sun et al., 2016).
Toremifene	Estrogen Modulator	↑	Reduced Aβ secretion in AD model neurons(Nishioka et al., 2016).
Mitotane	Estrogen Activator		-

Table 2. Continued

Compound	Description	Effect	Link to Alzheimer's Disease
Megestrol Acetate	Progesterone Activator	-	
Desogestrel	Progesterone Agonist	-	
Dihydroergotamine	5-HT1B/5-HT1D Agonist	↑	Inhibited the binding of Aβ oligomers to EphB2(Suzuki et al., 2016).
Clozapine	5-HT2A Inhibitor	↑	Improved Aβ-induced memory impairment and suppressed Aβ levels and plaque deposition in the brain of a transgenic mouse model of AD (Choi et al., 2017).
Amoxapine	5-HT2A Inhibitor	↑	Suppressed the level of Aβ in HEK293-APPsw cells (Li et al., 2017).
Nefazodone	5-HT2A Inhibitor		-
Flibanserin	5-HT1A/5-HT2A Inhibitor		-
Brexpiprazole	5-HT1A Inhibitor		-
Treprostinil	PPARγ Agonist		-
Rosiglitazone (maleate)	PPARγ Agonist	↑	Attenuated learning and memory deficits, and reduced Aβ 42 levels in Tg2576 mice(Pedersen et al., 2006).
Rosiglitazone	PPARγ Agonist	↑	Elicits neuroprotection on SH-SY5Y cells (K et al., 2021). Facilitated Aβ clearance in mice overexpressing mutant human APP (Escribano et al., 2010). Reduced spatial memory impairment, Aβ oligomers and aggregates, and astrocytic and microglia activation in a double transgenic AD mouse model(Toledo & Inestrosa, 2010).
Fenofibrate	PPARα Agonist	↑↓	Reduced the release of Aβ-42 in APP/PS1 transgenic mice (Zhang et al., 2014). Inhibited the Aβ-induced phenotype in a *C. elegans* AD model (Leiteritz et al., 2020). Raised Aβ-42 in APP transfected H4 cells(Kukar et al., 2005).
Celecoxib	COX-1/COX-2 Inhibitor	↑↓	Cleared Aβ in the neurons of APP/PS1 transgenic mice(Guo et al., 2017). Attenuated AlCl3-induced intellectual impairment and the associated neurodegenerative changes in rats(Abdel-Aal et al., 2021).

Table 2. Continued

Compound	Description	Effect	Link to Alzheimer's Disease
			No beneficial effects found in randomized control trials (Martin et al., 2008; Miguel-Álvarez et al., 2015).
Flurbiprofen	COX-1/COX-2 Inhibitor	↑	Lowered Aβ in H4 cell lines and in APP mice (Eriksen et al., 2003). Reduced Aβ-42 in both Neuro-2a cells and rat primary cortical neurons(Gasparini et al., 2004).
Oxaprozin	COX-2 Inhibitor	↑	Epidemiological studies found reduced AD incidence(Peter et al., 2002).
Eprosartan	AT1 Antagonist	↑	Restored and beneficially affected cerebral blood flow and connectivity (Wiesmann et al., 2017). Did not alter the level of Aβ or APP in the brains of 3xTg-AD mice(Ferrington et al., 2011).
Telmisartan	AT1 Antagonist	↑	Attenuated STZ induced impaired learning and memory as well as biochemical changes in AD mouse model(Singh et al., 2012). Improved cognitive decline and attenuated the Aβ-induced increase in expression of cytokines in Aβ 1-40 ICV injected mice(Tsukuda et al., 2009). Reduced amyloid burden in the cortex and hippocampus of 5XFAD mice (Torika et al., 2017).
Reserpine	VMAT1/VMAT2 Inhibitor	↑	Demonstrated neuroprotective activity against Aβ toxicity and anti-oxidative stress in PC12 cell cultures (Kashyap et al., 2020). Alleviated AB proteotoxicity in AD *C. elegans* model (Arya et al., 2009; Saharia et al., 2012).
Tetrabenazine	VMAT2 Inhibitor		-
Acitretin	Non-selective RAR Agonist	↑	Reduced Aβ40 and Aβ42 in APP/PS1-21 transgenic mice (Tippmann et al., 2009). Increased CSF APPs-α levels compared with the placebo group in a clinical study on AD patients(Endres et al., 2014).
Tazarotene	RARa, RARb, RARg Agonist		-

Table 2. Continued

Compound	Description	Effect	Link to Alzheimer's Disease
Meclizine	Ach Blocker	↑	Restored cognition and biochemical alterations in STZ-treated mice(Singh et al., 2020).
Nicotine	nAChRs Agonist	↑↓	Attenuated icv-STZ-induced impairments in recognition memory and was associated with higher neuronal density in rats(Esteves et al., 2017). Reduced the levels of Aβ and BACE1 peptides in hippocampal area CA1 and prevented Aβ-induced impairment of learning and short-term memory in rat AD model(Srivareerat et al., 2011). Reduced Aβ 1–42 positive plaques in the brains of APPsw mice (Nordberg et al., 2002). Increased the aggregation and phosphorylation state of tau in 3x-TgAD mice(Oddo et al., 2005). Exacerbated cognitive impairment and tau phosphorylation in Aβ25–35 injected rats(Deng et al., 2010).
Trientine	Copper (II) Chelator	↑	Decreased Aβ deposition and synapse loss in the brains of APP/PS1 mice (Wang et al., 2013).
Bacitracin	Copper (II) Chelator	↑	Reduced pathology in a transgenic *C. elegans* model of proteotoxicity associated with AD (Lublin et al., 2011).
Letermovir	DNA Terminase Complex Inhibitor	-	
L-Valacyclovir	Viral DNA Polymerase Inhibitor	-	
Eltrombopag	Thrombopoietin Receptor Agonist	-	
Cilostazol	PDE3A Inhibitor	↑	Attenuated learning and memory impairment induced by Aβ25-35 in mice(Hiramatsu et al., 2010). Suppressed Aβ-induced apoptosis and oxidative stress, and increased cell viability of SH-SY5Y cells(Oguchi et al., 2017). Decreased accumulation of Aβ1–42 in activated N2aSwe cells(Park et al., 2016).
Dutasteride	5α-Reductase Inhibitor	-	

Table 2. Continued

Compound	Description	Effect	Link to Alzheimer's Disease
Chlorambucil	DNA Alkylator	-	
Calcifediol	Vitamin D3 Receptor Agonist	↑	Improved cognitive function in a randomized controlled trial targeting elderly subjects with mild cognitive impairment (Yang et al., 2020). Reduced total AB levels in SH-SY5Y cells transfected with human APP695(Grimm et al., 2017).

↑: Drug demonstrates a potentially beneficial effect on Alzheimer's disease treatment.

↓: Drug demonstrates a potentially exacerbating effect on Alzheimer's disease pathologies.

Dopaminergic system dysfunction has been associated with AD through multiple post-mortem and in vivo studies measuring dopamine levels, cortical plasticity, and dopaminergic neuron degeneration (D'Amelio et al., 2018; Kemppainen et al., 2003; Nobili et al., 2017; Storga et al., 1996). These findings propose a strong link between dopaminergic deficit and AD in both early and late stages of the disease. A study using dopamine agonist rotigotine found an increase in cholinergic activity and normalized levels of LTP-like cortical plasticity (Koch et al., 2014). We found a total of 9 dopamine receptor antagonists and 2 dopamine receptor agonists of interest during our screening. The majority of the dopamine receptor antagonists in our list have not been studied in the context of AD; however, some dopamine receptor antagonists - including Haloperidol - have been chosen for clinical trials (D'Amelio et al., 2018). Furthermore, the 4 remaining dopamine-associated compounds found in our CsA-like cluster have all been linked to AD pathology, mostly through the inhibition of Aβ aggregation and improvement of cognitive function.

Dopamine-related antipsychotics, both typical and atypical, are the primary pharmacological option used to treat the neuropsychiatric and behavioral symptoms often present in AD (Calsolaro et al., 2019). These can include agitation, depression, psychosis, and overall behavioral disturbance. Several of the compounds listed in the present study are used to treat these symptoms in AD patients, namely, fluphenazine (Gottlieb et al., 1988), perphenazine (Pollock et al., 2002), ziprasidone (Kasckow et al., 2004), loxapine (Petrie et al., 1982; Raskind, 1998), and haloperidol (Devanand et al., 1998). It is important to note that many of these compounds have multiple targets and can simultaneously affect a variety of pathways. For instance, there is evidence of interaction between the dopamine and the serotonin pathways, implying the possibility of cross-pathway alteration in AD pathology (Ceyzériat et al., 2021). This is specifically utilized in the prescription of atypical antipsychotics, distinctively known as serotonin-dopamine antagonists. Due to their relevant multi-target capabilities and current use in patients with AD, we believe that this class of drugs would benefit from further studies exploring their possible benefits in the treatment of multiple stages of AD progression.

β-adrenergic receptors (βARs) are G protein-coupled receptors (GPCRs) responsible for regulating synaptic plasticity and memory formation(Kemp & Manahan-Vaughan, 2008). Specifically, β2-adrenergic receptor (β2AR) agonists are emerging therapeutic targets for neurological diseases (Li, 2023). During our screening, we found 5 β2AR agonists of interest with varying levels of supporting literature in the context of AD. Interestingly, there has been evidence of a reduction in AD prevalence in patients prescribed with β-blockers to treat hypertension (Khachaturian et al., 2006; Rosenberg et al., 2008). This protective trend also applies to other antihypertensive drugs (Ding et al., 2020; Wei et al., 2015), which include α1-adrenergic receptor antagonists such as Doxazosin. However, clinical studies often show contradictory results regarding the effects of β-blockers on cognitive impairment (Lebouvier et al., 2020). Studies performed in AD animal models also highlight the conflicting nature of β2AR signaling in the context of AD (Branca et al., 2014; Ni et al., 2006). Due to the opposing effects of β1- and β2-blockers in memory

and cognitive function (Ramos et al., 2005; Ramos et al., 2008), it is difficult to predict their effectiveness in treating AD pathogenesis. Comparably, α-adrenergic receptors (αARs) have been extensively linked to cognition as well as glucose metabolism - both part of AD symptomatology (Gannon et al., 2015). αAR antagonists, in particular, have been investigated both in vitro and in vivo as promising therapeutic targets of AD(Karczewski et al., 2018; Katsouri et al., 2013; Yu et al., 2022), and could prove interesting targets for future experiments.

The potential role of calcium homeostasis dysregulation in AD has been broadly explored through the impairment of mitochondrial function in the presence of excessive calcium levels (Calvo-Rodriguez & Bacskai, 2021), the induction of mitochondrial calcium overload by Aβ oligomers (Calvo-Rodríguez et al., 2016; Sanz-Blasco et al., 2008), and the direct effect of insoluble tau on calcium dyshomeostasis (Esteras et al., 2021; Mahoney et al., 2020). L-type calcium channels are substantially expressed on neurons (Zamponi, 2016), thus making L-type calcium channel blockers such as nimodipine, amiodarone, nicardipine, and dronedarone attractive targets for calcium regulation in the context of AD. Lebouvier et al. outline three potential mechanisms of action of L-type calcium channel blockers: (i) suppression of Aβ-induced calcium release, (ii) inhibition of amyloidogenesis, and (iii) increasing Aβ transcytosis (Lebouvier et al., 2020). Given our initial interest in calcineurin inhibitor CsA, we also emphasize both Pimecrolimus and Tacrolimus as compounds of interest due to both their shared mechanism of action and comprehensive supporting research on the subject of neurodegenerative diseases.

The primary steroid hormones estrogen, progesterone, and testosterone can be found in various regions of the central nervous system (Garcia-Ovejero et al., 2005). Steroidogenesis has been proven to be a natural mechanism to combat neurodegenerative conditions, and in vivo studies have shown an increase in AD-like neuropathology following gonadectomy (Carroll & Rosario, 2012). While hormone therapy seems promising as a preventive therapy in neurodegeneration, epidemiological studies and clinical trials reveal controversial results (Henderson, 2014). Treatment timing and dosage are important factors to consider in future research involving these agents.

The serotonergic system has been associated with cognitive function and performance in neurological diseases including schizophrenia, epilepsy, and AD (Švob Štrac et al., 2016). Neurotransmitter serotonin (5-HT) is involved in the regulation of various physiological processes including cognition and emotional behavior (Ciranna, 2006). Cerebrospinal fluid 5-HT levels in AD patients were significantly decreased compared to healthy controls (Tohgi et al., 1992), and post mortem studies show a decrease in brain 5-HT levels (Palmer et al., 1987). Given the overwhelming evidence of serotonergic influence in cognitive function, there has been interest in various 5-HT receptor (5-HTR) agonists and antagonists for the treatment of AD, including 5-HT2A (Garcia-Romeu et al., 2022; Lu et al., 2021) and 5-HT6 (Geldenhuys & Van der Schyf, 2009; Kucwaj-Brysz et al., 2021). We have identified 6 5-HTR antagonists with various 5-HT receptor targets: dihydroergotamine, clozapine, amoxapine, nefazodone, flibanserin, and brexpiprazole.

Considering that multiple 5-HTRs have a demonstrated beneficial effect on cognitive processes, a multi-receptor approach through one or more compounds might be valuable in future studies.

In addition to the aforementioned 5 categories, we also found compounds belonging to 13 other classes, namely, peroxisome proliferator-activated receptor (PPAR) inhibitors, cyclooxygenase (COX) inhibitors, angiotensin receptor inhibitors, vesicular monoamine transporter type (VMAT) inhibitors, retinoid acid receptor agonists, acetylcholine (ACh) effectors, copper chelators, cytomegalovirus (CMV) inhibitors, thrombopoietin receptor agonists, phosphodiesterase (PDE) inhibitors, 5α-reductase inhibitors, alkylating agents, and vitamin D agonists. Collectively, these 64 compounds comprise a heterogeneous mix of thoroughly studied and seldom explored potential therapeutic targets for neurodegenerative diseases. One of the main advantages of high-throughput behavioral screening approaches for drug repurposing is the lack of assumption regarding mechanism. In this study we present a diverse collection of compounds with seemingly diverging mechanisms of action, yet all evoking similar behavioral profiles in zebrafish larvae. Because AD is a multi-faceted neurodegenerative disease, it can greatly benefit from multi-targeted approaches to potentially ameliorate its pathology and impede further progression.

Methods

Animal Handling and Husbandry

All of the research in this study has been conducted in accordance with federal regulations and guidelines for the ethical and humane use of animals and has been reviewed and approved by Brown University's Institutional Animal Care and Use Committee (IACUC). All behavioral experiments were performed on 5 days post-fertilization (dpf) zebrafish larvae (Danio rerio). Wild-type adult zebrafish used for breeding were housed at Brown University's Animal Care facilities in 15- and 30-gallon tanks containing a mixed male and female population and kept on a 14 hr light and 10 hr dark cycle. During the light cycle, adult zebrafish were fed with Gemma Micro 300 and frozen brine shrimp. Adult zebrafish were bred in a group setting of approximately 40 zebrafish per tank, and embryos were collected and grown to 5 dpf as previously described(Pelkowski et al., 2011; Thorn et al., 2019; Tucker Edmister, Del Rosario Hernández, et al., 2022; Tucker Edmister, Ibrahim, et al., 2022). Embryos and larvae (0-5 dpf) were housed in 2L tanks with egg water containing 60 mg/L sea salt (Instant Ocean) and 0.25 mg/L methylene blue in deionized water. Zebrafish larvae used in this study were not fed, as they can obtain proper nutrition from their yolk sacs (Clift et al., 2014). Additionally, because sexual dimorphism is not apparent at this stage, larvae were not differentiated by sex (Liew & Orbán, 2014).

Pharmacological Treatment

Zebrafish larvae were treated with 876 FDA-approved compounds from the Cayman Chemical FDA-Approved Drugs Screening Library (Cayman Chemical, Ann Arbor, Michigan, Item No. 23538). Each compound was originally provided in 10mM stocks dissolved in dimethyl sulfoxide (DMSO) and we diluted these compounds in egg water to a 10μM final concentration. Zebrafish larvae at 5 dpf were exposed to 100μL of the treatment or control solutions for a total of 6 hours. Egg water and 1μl/ml DMSO were used as control treatments.

Behavioral Imaging

Zebrafish larvae were imaged using our previously described imaging setup and protocols (Pelkowski et al., 2011; Thorn et al., 2019; Tucker Edmister, Del Rosario Hernández, et al., 2022; Tucker Edmister, Ibrahim, et al., 2022). Briefly, larvae were imaged after 3 hours of exposure while placed in 96-well opaque plates (PerkinElmer, 6006290). The imaging system contains a glass stage capable of holding 4 plates at once. The plates were placed in a temperature-controlled imaging cabinet kept at 28.5°C. A high-resolution camera (18-megapixel Canon EOS Rebel T6 with an EF-S 55–250 mm f/4.0–5.6 IS zoom lens) captures a picture of the plates every 6 seconds. An M5 LED pico projector (Aaxa Technologies) with a 900 lumens LED light source was used to display a 3-hour Microsoft PowerPoint presentation featuring visual stimuli in the form of moving lines, as well as audio stimuli (100 ms, 400Hz) repeating at 1- and 20-second intervals(Gore et al., 2023).

Image Analysis

Larvae pose estimation and subsequent behavioral quantification were performed using our automated image processing framework, Z-LaP Tracker, which contains a model trained with open-source software DeepLabCut(10, 66). Briefly, we trained a deep neural network to recognize three main features of zebrafish larvae: the right eye, left eye, and yolk sac. This model allows us to identify larvae even in changing background conditions. The quantified larval behaviors generated by Z-LaP Tracker were further evaluated and summarized using Excel templates (10). We evaluated a total of 25 behaviors encapsulating activity, reactivity, swimming patterns, and optomotor response (10). The differences of these values compared to DMSO-vehicle controls formed behavioral paradigms for each compound.

Cluster Analyses

We evaluated a total of 876 compounds from the Cayman Chemical FDA-Approved Drugs Screening Library. We exposed 5 dpf zebrafish larvae to each compound (n=48

larvae) and averaged the results of each compound. Additionally, we normalized the data by calculating differences in behavior in comparison to the DMSO-vehicle control, creating behavioral profiles suitable for cluster analysis.

We performed K-means, a distance-based algorithm that seeks to partition each data point into one of k number of clusters by iteratively minimizing the distance between a point and its corresponding cluster mean. We used principal component analysis (PCA) as a dimensionality reduction method, and we used the elbow method to determine an optimal number of clusters k (k=4). PCA transforms the data into a two-dimensional form where K-means can be applied, and the clustering results plotted in a geometrical space.

We also performed hierarchical clustering, an unsupervised clustering method that allows us to visualize hierarchical relationships between individual compounds and their assigned clusters. Specifically, we used agglomerative hierarchical clustering with Euclidean distance as a distance metric, and complete linkage to measure dissimilarities between clusters.

All clustering methods were performed using R (R Studio 2022.12.0).

Library Annotation and IPA Analysis

Biological targets and pathways were assigned to each of the compounds in the Cayman Chemicals FDA-approved drug library, based on hits from the Disease-Gene Interaction Database (DGIdb)(Freshour et al., 2021), Therapeutic Targets Database (TTD)(Chen et al., 2002) , Guide to Pharmacology (GtoPdb)(Harding et al., 2022), Kyoto Encyclopedia of Genes (KEGG)(Kanehisa & Goto, 2000), Protein ANalysis THrough Evolutionary Relationships (PANTHER)(Mi & Thomas, 2009), WikiPathways(Slenter et al., 2018), and Reactome(Jassal et al., 2020) databases. Each database was queried for primary and secondary target genes associated with the compounds, as well as their respective molecular pathways and associated mechanisms of action. Cross-database datasets were generated with available matching information (i.e., UniProt ID, Gene Symbol, Ligand ID) to annotate the library. QIAGEN Ingenuity Pathway Analysis (IPA) was used to further analyze compounds of interest. Specifically, we used the "Build a Pathway" tool to input previously identified targets from our database query associated with compounds of interest. We then added "Alzheimer's disease" and its related targets as a node in our pathway. Connections between elements were automatically generated using IPA's "Connect" tool. We generated an overlay using the "Molecule Activity Predictor (MAP)" to indicate activation or inhibition of pathway components and connections.

Statistical analyses

Statistical tests and graphs were generated using Microsoft Excel 2016, R, and BioRender. Due to the nature of our data, we used non-parametric Welch's unequal variance t-test along with a Bonferroni correction for multiple comparisons. In the current screen, we compared 876 drugs to the DMSO-vehicle controls and differences were considered significant when $p < 5.7 \times 10\text{-}5$ $p < 1.1 \times 10\text{-}5$ (0.01/876), or $p < 1.1 \times 10\text{-}6$ (0.001/876). Pearson correlation coefficients were calculated using R.

Acknowledgments

We are grateful for valuable intellectual input from members of the Creton lab. This work was supported by the National Institutes of Health, Grant R01GM136906 (R.C.) and Grant R01GM136906-03S1 (R.C., J.A.K.).

Author contributions

Conceptualization: RC, JAK
Methodology: RC, SVG
Investigation: TDRH, SVG
Visualization: TDRH
Supervision: RC, SVG, JAK
Writing—original draft: TDRH
Writing—review & editing: RC, SVG, TDRH

Competing interests

Brown University submitted a patent application for the treatment of neurodegenerative disease using CsA-type compounds (application 63/193,935, Robbert Creton - inventor, Sara Tucker Edmister, Rahma Ibrahim, Rohit Kakodkar and Jill A. Kreiling - contributors). The authors declare that they do not have other competing interests.

Data Sharing and Availability

The DeepLabCut model used for pose estimation can be found on our GitHub repository (https://github.com/brown-ccv/Automated-Analysis-of-Zebrafish), along with installation and usage instructions. All data are available in the main text or the supplementary materials.

Supplementary Materials

Fig. S1.
(A) Subclusters of CsA-like compounds identified with hierarchical clustering. **(B)** Box-whisker plots of Pearson correlation values of compounds with the same primary target; mechanisms of action are marked (+) for agonists and (-) for antagonists. Pearson correlation values for the subclusters of CsA-like compounds are also plotted. The mean correlation of biologically unrelated compounds is plotted as a red line (0.186).

Fig. S2.
Hierarchical cluster analysis of the behavioral profiles for all screened compounds.

Fig. S3.
Behavioral profiles of 19 compounds previously identified as CsA-like during the screening of the Tocriscreen FDA-Approved Drugs Library. **(A)** 12 compounds displaying dissimilar behavioral effects from CsA. Behavioral profiles induced by biologically similar compounds, namely **(B)** β2-adrenergic receptor agonists, **(C)** α1-adrenergic receptor blockers, **(D)** androgen blockers, **(E)** PDE5 inhibitors, and **(F)** VEGFR inhibitors.

Table S1.
Difference values of drug-treated larvae vs DMSO-treated larvae for all 876 compounds.

Table S2.
Statistical analyses performed on the behavioral profiles of all 876 compounds.

References

2023 Alzheimer's disease facts and figures. (2023). Alzheimer's & Dementia, 19(4), 1598-1695. https://doi.org/https://doi.org/10.1002/alz.13016

Abdel-Aal, R. A., Hussein, O. A., Elsaady, R. G., & Abdelzaher, L. A. (2021). Celecoxib effect on rivastigmine anti-Alzheimer activity against aluminum chloride-induced neurobehavioral deficits as a rat model of Alzheimer's disease; novel perspectives for an old drug. Journal of Medical and Life Science, 3(4), 44-82. https://doi.org/10.21608/jmals.2021.210630

Abdel Rasheed, N. O., El Sayed, N. S., & El-Khatib, A. S. (2018). Targeting central β2 receptors ameliorates streptozotocin-induced neuroinflammation via inhibition of glycogen synthase kinase3 pathway in mice. Progress in Neuro-Psychopharmacology and Biological Psychiatry, 86, 65-75. https://doi.org/https://doi.org/10.1016/j.pnpbp.2018.05.010

Al-Subari, S., Butt, I., Budhathoki, P., Khadka, S., Shrestha, D., Alamgeer, & Alhouzani, T. (2020). Repurposing Drugs for COVID-19: An Approach for Treatment in the Pandemic. Alternative therapies in health and medicine, 26.

Anand, R., Gill, K. D., & Mahdi, A. A. (2014). Therapeutics of Alzheimer's disease: Past, present and future. Neuropharmacology, 76, 27-50. https://doi.org/https://doi.org/10.1016/j.neuropharm.2013.07.004

Arrowsmith, J., & Harrison, R. (2012). Drug Repositioning: Bringing New Life to Shelved Assets and Existing Drugs. In (pp. 7-32). https://doi.org/10.1002/9781118274408.ch1

Arya, U., Dwivedi, H., & Subramaniam, J. R. (2009). Reserpine ameliorates Abeta toxicity in the Alzheimer's disease model in Caenorhabditis elegans. Exp Gerontol, 44(6-7), 462-466. https://doi.org/10.1016/j.exger.2009.02.010

Bachmeier, C., Beaulieu-Abdelahad, D., Mullan, M., & Paris, D. (2011). Selective dihydropyiridine compounds facilitate the clearance of β-amyloid across the blood-brain barrier. Eur J Pharmacol, 659(2-3), 124-129. https://doi.org/10.1016/j.ejphar.2011.03.048

Braak, H., & Braak, E. (1991). Neuropathological stageing of Alzheimer-related changes. Acta Neuropathologica, 82(4), 239-259. https://doi.org/10.1007/BF00308809

Branca, C., Wisely, E. V., Hartman, L. K., Caccamo, A., & Oddo, S. (2014). Administration of a selective β2 adrenergic receptor antagonist exacerbates neuropathology and cognitive deficits in a mouse model of Alzheimer's disease. Neurobiology of Aging, 35(12), 2726-2735. https://doi.org/https://doi.org/10.1016/j.neurobiolaging.2014.06.011

Calsolaro, V., Antognoli, R., Okoye, C., & Monzani, F. (2019). The Use of Antipsychotic Drugs for Treating Behavioral Symptoms in Alzheimer's Disease. Front Pharmacol, 10, 1465. https://doi.org/10.3389/fphar.2019.01465

Calvo-Rodriguez, M., & Bacskai, B. J. (2021). Mitochondria and Calcium in Alzheimer's Disease: From Cell Signaling to Neuronal Cell Death. Trends in Neurosciences, 44(2), 136-151. https://doi.org/https://doi.org/10.1016/j.tins.2020.10.004

Calvo-Rodríguez, M., García-Durillo, M., Villalobos, C., & Núñez, L. (2016). Aging Enables Ca 2+ Overload and Apoptosis Induced by Amyloid-β Oligomers in Rat Hippocampal Neurons: Neuroprotection by Non-Steroidal Anti-Inflammatory Drugs and R-Flurbiprofen in Aging Neurons. Journal of Alzheimer's Disease, 54, 207-221. https://doi.org/10.3233/JAD-151189

Carroll, J. C., & Rosario, E. R. (2012). The potential use of hormone-based therapeutics for the treatment of Alzheimer's disease. Curr Alzheimer Res, 9(1), 18-34. https://doi.org/10.2174/156720512799015109

Ceyzériat, K., Gloria, Y., Tsartsalis, S., Fossey, C., Cailly, T., Fabis, F., Millet, P., & Tournier, B. B. (2021). Alterations in dopamine system and in its connectivity with serotonin in a rat model of Alzheimer's disease. Brain Commun, 3(2), fcab029. https://doi.org/10.1093/braincomms/fcab029

Chen, X., Ji, Z. L., & Chen, Y. Z. (2002). TTD: Therapeutic Target Database. Nucleic Acids Res, 30(1), 412-415. https://doi.org/10.1093/nar/30.1.412

Choi, Y., Jeong, H. J., Liu, Q. F., Oh, S. T., Koo, B. S., Kim, Y., Chung, I. W., Kim, Y. S., & Jeon, S. (2017). Clozapine Improves Memory Impairment and Reduces Aβ Level in the Tg-APPswe/PS1dE9 Mouse Model of Alzheimer's Disease. Mol Neurobiol, 54(1), 450-460. https://doi.org/10.1007/s12035-015-9636-x

Ciranna, L. (2006). Serotonin as a modulator of glutamate- and GABA-mediated neurotransmission: implications in physiological functions and in pathology. Curr Neuropharmacol, 4(2), 101-114. https://doi.org/10.2174/157015906776359540

Clift, D., Richendrfer, H., Thorn, R. J., Colwill, R. M., & Creton, R. (2014). High-Throughput Analysis of Behavior in Zebrafish Larvae: Effects of Feeding. Zebrafish, 11(5), 455-461. https://doi.org/10.1089/zeb.2014.0989

Coelho, B. P., Gaelzer, M. M., dos Santos Petry, F., Hoppe, J. B., Trindade, V. M. T., Salbego, C. G., & Guma, F. T. C. R. (2019). Dual Effect of Doxazosin: Anticancer Activity on SH-SY5Y Neuroblastoma Cells and Neuroprotection on an In Vitro Model of Alzheimer's Disease. Neuroscience, 404, 314-325. https://doi.org/https://doi.org/10.1016/j.neuroscience.2019.02.005

D'Amelio, M., Puglisi-Allegra, S., & Mercuri, N. (2018). The role of dopaminergic midbrain in Alzheimer's disease: Translating basic science into clinical practice. Pharmacological Research, 130, 414-419. https://doi.org/https://doi.org/10.1016/j.phrs.2018.01.016

Deng, J., Shen, C., Wang, Y.-J., Zhang, M., Li, J., Xu, Z.-Q., Gao, C.-Y., Fang, C.-Q., & Zhou, H.-D. (2010). Nicotine exacerbates tau phosphorylation and cognitive impairment induced by amyloid-beta 25–35 in rats. European Journal of Pharmacology, 637(1), 83-88. https://doi.org/https://doi.org/10.1016/j.ejphar.2010.03.029

Devanand, D. P., Marder, K., Michaels, K. S., Sackeim, H. A., Bell, K., Sullivan, M. A., Cooper, T. B., Pelton, G. H., & Mayeux, R. (1998). A Randomized, Placebo-Controlled Dose-Comparison Trial of Haloperidol for Psychosis and Disruptive Behaviors in Alzheimer's Disease. American Journal of Psychiatry, 155(11), 1512-1520. https://doi.org/10.1176/ajp.155.11.1512

Dineley, K. T., Hogan, D., Zhang, W. R., & Taglialatela, G. (2007). Acute inhibition of calcineurin restores associative learning and memory in Tg2576 APP transgenic mice. Neurobiol Learn Mem, 88(2), 217-224. https://doi.org/10.1016/j.nlm.2007.03.010

Ding, J., Davis-Plourde, K. L., Sedaghat, S., Tully, P. J., Wang, W., Phillips, C., Pase, M. P., Himali, J. J., Gwen Windham, B., Griswold, M., Gottesman, R., Mosley, T. H., White, L., Guðnason, V., Debette, S., Beiser, A. S., Seshadri, S., Ikram, M. A., Meirelles, O., . . . Launer, L. J. (2020). Antihypertensive medications and risk for incident dementia and Alzheimer's disease: a meta-analysis of individual participant data from prospective cohort studies. Lancet Neurol, 19(1), 61-70. https://doi.org/10.1016/s1474-4422(19)30393-x

Endres, K., Fahrenholz, F., Lotz, J., Hiemke, C., Teipel, S., Lieb, K., Tüscher, O., & Fellgiebel, A. (2014). Increased CSF APPs-α levels in patients with Alzheimer disease treated with acitretin. Neurology, 83(21), 1930-1935. https://doi.org/10.1212/wnl.0000000000001017

Eriksen, J. L., Sagi, S. A., Smith, T. E., Weggen, S., Das, P., McLendon, D. C., Ozols, V. V., Jessing, K. W., Zavitz, K. H., Koo, E. H., & Golde, T. E. (2003). NSAIDs and enantiomers of flurbiprofen target gamma-secretase and lower Abeta 42 in vivo. J Clin Invest, 112(3), 440-449. https://doi.org/10.1172/jci18162

Escribano, L., Simón, A. M., Gimeno, E., Cuadrado-Tejedor, M., López de Maturana, R., García-Osta, A., Ricobaraza, A., Pérez-Mediavilla, A., Del Río, J., & Frechilla, D. (2010). Rosiglitazone rescues memory impairment in Alzheimer's transgenic mice: mechanisms involving a reduced amyloid and tau pathology. Neuropsychopharmacology, 35(7), 1593-1604. https://doi.org/10.1038/npp.2010.32

Esteras, N., Kundel, F., Amodeo, G. F., Pavlov, E. V., Klenerman, D., & Abramov, A. Y. (2021). Insoluble tau aggregates induce neuronal death through modification of membrane ion conductance, activation of voltage-gated calcium channels and NADPH oxidase [https://doi.org/10.1111/febs.15340]. The FEBS Journal, 288(1), 127-141. https://doi.org/https://doi.org/10.1111/febs.15340

Esteves, I. M., Lopes-Aguiar, C., Rossignoli, M. T., Ruggiero, R. N., Broggini, A. C. S., Bueno-Junior, L. S., Kandratavicius, L., Monteiro, M. R., Romcy-Pereira, R. N., & Leite, J. P. (2017). Chronic nicotine attenuates behavioral and synaptic plasticity impairments in a streptozotocin model of Alzheimer's disease. Neuroscience, 353, 87-97. https://doi.org/https://doi.org/10.1016/j.neuroscience.2017.04.011

Ferrington, L., Miners, J. S., Palmer, L. E., Bond, S. M., Povey, J. E., Kelly, P. A., Love, S., Horsburgh, K. J., & Kehoe, P. G. (2011). Angiotensin II-inhibiting drugs have no effect on

intraneuronal Aβ or oligomeric Aβ levels in a triple transgenic mouse model of Alzheimer's disease. Am J Transl Res, 3(2), 197-208.

Freshour, S. L., Kiwala, S., Cotto, K. C., Coffman, A. C., McMichael, J. F., Song, J. J., Griffith, M., Griffith, Obi L., & Wagner, A. H. (2021). Integration of the Drug–Gene Interaction Database (DGIdb 4.0) with open crowdsource efforts. Nucleic Acids Research, 49(D1), D1144-D1151. https://doi.org/10.1093/nar/gkaa1084

Gannon, M., Che, P., Chen, Y., Jiao, K., Roberson, E. D., & Wang, Q. (2015). Noradrenergic dysfunction in Alzheimer's disease. Front Neurosci, 9, 220. https://doi.org/10.3389/fnins.2015.00220

Garcia-Ovejero, D., Azcoitia, I., DonCarlos, L. L., Melcangi, R. C., & Garcia-Segura, L. M. (2005). Glia-neuron crosstalk in the neuroprotective mechanisms of sex steroid hormones. Brain Research Reviews, 48(2), 273-286. https://doi.org/https://doi.org/10.1016/j.brainresrev.2004.12.018

Garcia-Romeu, A., Darcy, S., Jackson, H., White, T., & Rosenberg, P. (2022). Psychedelics as Novel Therapeutics in Alzheimer's Disease: Rationale and Potential Mechanisms. Curr Top Behav Neurosci, 56, 287-317. https://doi.org/10.1007/7854_2021_267

Gasparini, L., Rusconi, L., Xu, H., del Soldato, P., & Ongini, E. (2004). Modulation of beta-amyloid metabolism by non-steroidal anti-inflammatory drugs in neuronal cell cultures. J Neurochem, 88(2), 337-348. https://doi.org/10.1111/j.1471-4159.2004.02154.x

Geldenhuys, W. J., & Van der Schyf, C. J. (2009). The serotonin 5-HT6 receptor: a viable drug target for treating cognitive deficits in Alzheimer's disease. Expert Review of Neurotherapeutics, 9(7), 1073-1085. https://doi.org/10.1586/ern.09.51

Gore, S. V., Kakodkar, R., Del Rosario Hernández, T., Edmister, S. T., & Creton, R. (2023). Zebrafish Larvae Position Tracker (Z-LaP Tracker): a high-throughput deep-learning behavioral approach for the identification of calcineurin pathway-modulating drugs using zebrafish larvae. Sci Rep, 13(1), 3174. https://doi.org/10.1038/s41598-023-30303-w

Gottlieb, G. L., McAllister, T. W., & Gur, R. C. (1988). Depot Neuroleptics in the Treatment of Behavioral Disorders in Patients with Alzheimer's Disease [https://doi.org/10.1111/j.1532-5415.1988.tb06157.x]. Journal of the American Geriatrics Society, 36(7), 619-621. https://doi.org/https://doi.org/10.1111/j.1532-5415.1988.tb06157.x

Grimm, M. O. W., Thiel, A., Lauer, A. A., Winkler, J., Lehmann, J., Regner, L., Nelke, C., Janitschke, D., Benoist, C., Streidenberger, O., Stötzel, H., Endres, K., Herr, C., Beisswenger, C., Grimm, H. S., Bals, R., Lammert, F., & Hartmann, T. (2017). Vitamin D and Its Analogues Decrease Amyloid-β (Aβ) Formation and Increase Aβ-Degradation. International Journal of Molecular Sciences, 18(12).

Guo, J.-W., Guan, P.-P., Ding, W.-Y., Wang, S.-L., Huang, X.-S., Wang, Z.-Y., & Wang, P. (2017). Erythrocyte membrane-encapsulated celecoxib improves the cognitive decline of Alzheimer's disease by concurrently inducing neurogenesis and reducing apoptosis in APP/PS1

transgenic mice. Biomaterials, 145, 106-127.
https://doi.org/https://doi.org/10.1016/j.biomaterials.2017.07.023

Gursoy, E., Cardounel, A., Al-khlaiwi, T., Al-drees, A., & Kalimi, M. (2002). Tamoxifen protects clonal mouse hippocampal (HT-22) cells against neurotoxins-induced cell death. Neurochemistry International, 40(5), 405-412. https://doi.org/https://doi.org/10.1016/S0197-0186(01)00105-X

Hamsanandini, R., Margo, F. U., Stephanie, M. K., Kathy, B., Jennifer, L. M., Erin Denhart, J., Beverly, M., Jeffrey, S., László, G. P., David, K. P., Christopher, M. N., Craig, E. L. S., & Elizabeth, H. (2021). Tacrolimus Protects against Age-Associated Microstructural Changes in the Beagle Brain. The Journal of Neuroscience, 41(23), 5124. https://doi.org/10.1523/JNEUROSCI.0361-21.2021

Harding, S. D., Armstrong, J. F., Faccenda, E., Southan, C., Alexander, S. P. H., Davenport, A. P., Pawson, A. J., Spedding, M., & Davies, J. A. (2022). The IUPHAR/BPS guide to PHARMACOLOGY in 2022: curating pharmacology for COVID-19, malaria and antibacterials. Nucleic Acids Res, 50(D1), D1282-d1294. https://doi.org/10.1093/nar/gkab1010

Hauser, A. S., Attwood, M. M., Rask-Andersen, M., Schiöth, H. B., & Gloriam, D. E. (2017). Trends in GPCR drug discovery: new agents, targets and indications. Nature Reviews Drug Discovery, 16(12), 829-842. https://doi.org/10.1038/nrd.2017.178

Henderson, V. W. (2014). Alzheimer's disease: review of hormone therapy trials and implications for treatment and prevention after menopause. J Steroid Biochem Mol Biol, 142, 99-106. https://doi.org/10.1016/j.jsbmb.2013.05.010

Heo, H. J., Park, S. Y., Lee, Y. S., Shin, H. K., Hong, K. W., & Kim, C. D. (2020). Combination therapy with cilostazol, aripiprazole, and donepezil protects neuronal cells from β-amyloid neurotoxicity through synergistically enhanced SIRT1 expression. Korean J Physiol Pharmacol, 24(4), 299-310. https://doi.org/10.4196/kjpp.2020.24.4.299

Higaki, J., Murphy Jr, G. M., & Cordell, B. (1997). Inhibition of β-Amyloid Formation by Haloperidol: A Possible Mechanism for Reduced Frequency of Alzheimer's Disease Pathology in Schizophrenia [https://doi.org/10.1046/j.1471-4159.1997.68010333.x]. Journal of Neurochemistry, 68(1), 333-336. https://doi.org/https://doi.org/10.1046/j.1471-4159.1997.68010333.x

Hiramatsu, M., Takiguchi, O., Nishiyama, A., & Mori, H. (2010). Cilostazol prevents amyloid β peptide25-35-induced memory impairment and oxidative stress in mice [https://doi.org/10.1111/j.1476-5381.2010.01014.x]. British Journal of Pharmacology, 161(8), 1899-1912. https://doi.org/https://doi.org/10.1111/j.1476-5381.2010.01014.x

Howe, K., Clark, M. D., Torroja, C. F., Torrance, J., Berthelot, C., Muffato, M., Collins, J. E., Humphray, S., McLaren, K., Matthews, L., McLaren, S., Sealy, I., Caccamo, M., Churcher, C., Scott, C., Barrett, J. C., Koch, R., Rauch, G.-J., White, S., . . . Stemple, D. L. (2013). The

zebrafish reference genome sequence and its relationship to the human genome. Nature, 496(7446), 498-503. https://doi.org/10.1038/nature12111

Ishikawa, Y., Itoh, R., Tsujimoto, R., Tamano, H., & Takeda, A. (2022). Isoproterenol injected into the basolateral amygdala rescues amyloid β1-42-induced conditioned fear memory deficit via reducing intracellular Zn2+ toxicity. Neuroscience Letters, 766, 136353. https://doi.org/https://doi.org/10.1016/j.neulet.2021.136353

Jassal, B., Matthews, L., Viteri, G., Gong, C., Lorente, P., Fabregat, A., Sidiropoulos, K., Cook, J., Gillespie, M., Haw, R., Loney, F., May, B., Milacic, M., Rothfels, K., Sevilla, C., Shamovsky, V., Shorser, S., Varusai, T., Weiser, J., . . . D'Eustachio, P. (2020). The reactome pathway knowledgebase. Nucleic Acids Research, 48(D1), D498-D503. https://doi.org/10.1093/nar/gkz1031

Jeong, Y. J., Son, Y., Park, H. J., Oh, S. J., Choi, J. Y., Ko, Y. G., & Lee, H. J. (2021). Therapeutic Effects of Aripiprazole in the 5xFAD Alzheimer's Disease Mouse Model. Int J Mol Sci, 22(17). https://doi.org/10.3390/ijms22179374

K, C. S., Kakoty, V., Marathe, S., Chitkara, D., & Taliyan, R. (2021). Exploring the Neuroprotective Potential of Rosiglitazone Embedded Nanocarrier System on Streptozotocin Induced Mice Model of Alzheimer's Disease. Neurotox Res, 39(2), 240-255. https://doi.org/10.1007/s12640-020-00258-1

Kalueff, A. V., Gebhardt, M., Stewart, A. M., Cachat, J. M., Brimmer, M., Chawla, J. S., Craddock, C., Kyzar, E. J., Roth, A., Landsman, S., Gaikwad, S., Robinson, K., Baatrup, E., Tierney, K., Shamchuk, A., Norton, W., Miller, N., Nicolson, T., Braubach, O., . . . the Zebrafish Neuroscience Research Consortium, H. (2013). Towards a Comprehensive Catalog of Zebrafish Behavior 1.0 and Beyond. Zebrafish, 10(1), 70-86. https://doi.org/10.1089/zeb.2012.0861

Kalueff, A. V., Stewart, A. M., & Gerlai, R. (2014). Zebrafish as an emerging model for studying complex brain disorders. Trends in Pharmacological Sciences, 35(2), 63-75. https://doi.org/https://doi.org/10.1016/j.tips.2013.12.002

Kanehisa, M., & Goto, S. (2000). KEGG: kyoto encyclopedia of genes and genomes. Nucleic Acids Res, 28(1), 27-30. https://doi.org/10.1093/nar/28.1.27

Karczewski, P., Hempel, P., & Bimmler, M. (2018). Role of alpha1-adrenergic receptor antibodies in Alzheimer's disease. Front Biosci (Landmark Ed), 23(11), 2082-2089. https://doi.org/10.2741/4691

Kasckow, J. W., Mulchahey, J. J., & Mohamed, S. (2004). The Use of Novel Antipsychotics in the Older Patient With Neurodegenerative Disorders in the Long-Term Care Setting. Journal of the American Medical Directors Association, 5(4), 242-248. https://doi.org/https://doi.org/10.1016/S1525-8610(04)70130-9

Kashyap, P., Kalaiselvan, V., Kumar, R., & Kumar, S. (2020). Ajmalicine and Reserpine: Indole Alkaloids as Multi-Target Directed Ligands Towards Factors Implicated in Alzheimer's Disease. Molecules, 25(7). https://doi.org/10.3390/molecules25071609

Katsouri, L., Vizcaychipi, M. P., McArthur, S., Harrison, I., Suárez-Calvet, M., Lleo, A., Lloyd, D. G., Ma, D., & Sastre, M. (2013). Prazosin, an α1-adrenoceptor antagonist, prevents memory deterioration in the APP23 transgenic mouse model of Alzheimer's disease. Neurobiology of Aging, 34(4), 1105-1115. https://doi.org/https://doi.org/10.1016/j.neurobiolaging.2012.09.010

Kawano, Y., Tamura, K., Egawa, M., Tamano, H., & Takeda, A. (2022). Isoproterenol, an adrenergic β receptor agonist, induces metallothionein synthesis followed by canceling amyloid β1-42-induced neurodegeneration. BioMetals, 35(2), 303-312. https://doi.org/10.1007/s10534-022-00365-w

Kemp, A., & Manahan-Vaughan, D. (2008). β-Adrenoreceptors Comprise a Critical Element in Learning-Facilitated Long-Term Plasticity. Cerebral Cortex, 18(6), 1326-1334. https://doi.org/10.1093/cercor/bhm164

Kemppainen, N., Laine, M., Laakso, M. P., Kaasinen, V., Någren, K., Vahlberg, T., Kurki, T., & Rinne, J. O. (2003). Hippocampal dopamine D2 receptors correlate with memory functions in Alzheimer's disease [https://doi.org/10.1046/j.1460-9568.2003.02716.x]. European Journal of Neuroscience, 18(1), 149-154. https://doi.org/https://doi.org/10.1046/j.1460-9568.2003.02716.x

Khachaturian, A. S., Zandi, P. P., Lyketsos, C. G., Hayden, K. M., Skoog, I., Norton, M. C., Tschanz, J. T., Mayer, L. S., Welsh-Bohmer, K. A., Breitner, J. C. S., & for the Cache County Study, G. (2006). Antihypertensive Medication Use and Incident Alzheimer Disease: The Cache County Study. Archives of Neurology, 63(5), 686-692. https://doi.org/10.1001/archneur.63.5.noc60013

Koch, G., Di Lorenzo, F., Bonnì, S., Giacobbe, V., Bozzali, M., Caltagirone, C., & Martorana, A. (2014). Dopaminergic Modulation of Cortical Plasticity in Alzheimer's Disease Patients. Neuropsychopharmacology, 39(11), 2654-2661. https://doi.org/10.1038/npp.2014.119

Kondo, T., Imamura, K., Funayama, M., Tsukita, K., Miyake, M., Ohta, A., Woltjen, K., Nakagawa, M., Asada, T., Arai, T., Kawakatsu, S., Izumi, Y., Kaji, R., Iwata, N., & Inoue, H. (2017). iPSC-Based Compound Screening and In Vitro Trials Identify a Synergistic Anti-amyloid β Combination for Alzheimer's Disease. Cell Reports, 21(8), 2304-2312. https://doi.org/https://doi.org/10.1016/j.celrep.2017.10.109

Koppel, J., Jimenez, H., Adrien, L., Greenwald, B. S., Marambaud, P., Cinamon, E., & Davies, P. (2016). Haloperidol inactivates AMPK and reduces tau phosphorylation in a tau mouse model of Alzheimer's disease. Alzheimers Dement (N Y), 2(2), 121-130. https://doi.org/10.1016/j.trci.2016.05.003

Kucwaj-Brysz, K., Baltrukevich, H., Czarnota, K., & Handzlik, J. (2021). Chemical update on the potential for serotonin 5-HT(6) and 5-HT(7) receptor agents in the treatment of Alzheimer's disease. Bioorg Med Chem Lett, 49, 128275. https://doi.org/10.1016/j.bmcl.2021.128275

Kukar, T., Murphy, M. P., Eriksen, J. L., Sagi, S. A., Weggen, S., Smith, T. E., Ladd, T., Khan, M. A., Kache, R., Beard, J., Dodson, M., Merit, S., Ozols, V. V., Anastasiadis, P. Z., Das, P., Fauq, A., Koo, E. H., & Golde, T. E. (2005). Diverse compounds mimic Alzheimer disease–

causing mutations by augmenting Aβ42 production. Nature Medicine, 11(5), 545-550. https://doi.org/10.1038/nm1235

Lebouvier, T., Chen, Y., Duriez, P., Pasquier, F., & Bordet, R. (2020). Antihypertensive agents in Alzheimer's disease: beyond vascular protection. Expert Review of Neurotherapeutics, 20(2), 175-187. https://doi.org/10.1080/14737175.2020.1708195

Lee, J. H., Wolfe, D. M., Darji, S., McBrayer, M. K., Colacurcio, D. J., Kumar, A., Stavrides, P., Mohan, P. S., & Nixon, R. A. (2020). β2-adrenergic Agonists Rescue Lysosome Acidification and Function in PSEN1 Deficiency by Reversing Defective ER-to-lysosome Delivery of ClC-7. J Mol Biol, 432(8), 2633-2650. https://doi.org/10.1016/j.jmb.2020.02.021

Leiteritz, A., Baumanns, S., & Wenzel, U. (2020). Amyloid-beta (Aβ(1-42))-induced paralysis in Caenorhabditis elegans is reduced through NHR-49/PPARalpha. Neurosci Lett, 730, 135042. https://doi.org/10.1016/j.neulet.2020.135042

Li, S. (2023). The β-adrenergic hypothesis of synaptic and microglial impairment in Alzheimer's disease [https://doi.org/10.1111/jnc.15782]. Journal of Neurochemistry, 165(3), 289-302. https://doi.org/https://doi.org/10.1111/jnc.15782

Li, X., Wang, Q., Hu, T., Wang, Y., Zhao, J., Lu, J., & Pei, G. (2017). A tricyclic antidepressant, amoxapine, reduces amyloid-β generation through multiple serotonin receptor 6-mediated targets. Sci Rep, 7(1), 4983. https://doi.org/10.1038/s41598-017-04144-3

Liew, W. C., & Orbán, L. (2014). Zebrafish sex: a complicated affair. Briefings in Functional Genomics, 13(2), 172-187. https://doi.org/10.1093/bfgp/elt041

Liu, S., Han, Y., Zhang, T., & Yang, Z. (2011). Protective effect of trifluoperazine on hydrogen peroxide-induced apoptosis in PC12 cells. Brain Research Bulletin, 84(2), 183-188. https://doi.org/https://doi.org/10.1016/j.brainresbull.2010.12.008

Liu, X., Cheng, Z.-Y., Li, Y.-F., Liu, C., Wang, C., Gong, X.-J., & He, L. (2023). Dopamine D2 receptor agonist Bromocriptine ameliorates Aβ1-42-induced memory deficits and neuroinflammation in mice. European Journal of Pharmacology, 938, 175443. https://doi.org/https://doi.org/10.1016/j.ejphar.2022.175443

Lu, J., Zhang, C., Lv, J., Zhu, X., Jiang, X., Lu, W., Lu, Y., Tang, Z., Wang, J., & Shen, X. (2021). Antiallergic drug desloratadine as a selective antagonist of 5HT(2A) receptor ameliorates pathology of Alzheimer's disease model mice by improving microglial dysfunction. Aging Cell, 20(1), e13286. https://doi.org/10.1111/acel.13286

Lublin, A., Isoda, F., Patel, H., Yen, K., Nguyen, L., Hajje, D., Schwartz, M., & Mobbs, C. (2011). FDA-approved drugs that protect mammalian neurons from glucose toxicity slow aging dependent on cbp and protect against proteotoxicity. PLoS One, 6(11), e27762. https://doi.org/10.1371/journal.pone.0027762

Mahoney, R., Ochoa Thomas, E., Ramirez, P., Miller, H. E., Beckmann, A., Zuniga, G., Dobrowolski, R., & Frost, B. (2020). Pathogenic Tau Causes a Toxic Depletion of Nuclear

Calcium. Cell Reports, 32(2), 107900. https://doi.org/https://doi.org/10.1016/j.celrep.2020.107900

Martin, B. K., Szekely, C., Brandt, J., Piantadosi, S., Breitner, J. C., Craft, S., Evans, D., Green, R., & Mullan, M. (2008). Cognitive function over time in the Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT): results of a randomized, controlled trial of naproxen and celecoxib. Arch Neurol, 65(7), 896-905. https://doi.org/10.1001/archneur.2008.65.7.nct70006

Mathis, A., Mamidanna, P., Cury, K. M., Abe, T., Murthy, V. N., Mathis, M. W., & Bethge, M. (2018). DeepLabCut: markerless pose estimation of user-defined body parts with deep learning. Nature Neuroscience, 21(9), 1281-1289. https://doi.org/10.1038/s41593-018-0209-y

McCormick, A. V., Wheeler, J. M., Guthrie, C. R., Liachko, N. F., & Kraemer, B. C. (2013). Dopamine D2 receptor antagonism suppresses tau aggregation and neurotoxicity. Biol Psychiatry, 73(5), 464-471. https://doi.org/10.1016/j.biopsych.2012.08.027

Mi, H., & Thomas, P. (2009). PANTHER pathway: an ontology-based pathway database coupled with data analysis tools. Methods Mol Biol, 563, 123-140. https://doi.org/10.1007/978-1-60761-175-2_7

Miguel-Álvarez, M., Santos-Lozano, A., Sanchis-Gomar, F., Fiuza-Luces, C., Pareja-Galeano, H., Garatachea, N., & Lucia, A. (2015). Non-Steroidal Anti-Inflammatory Drugs as a Treatment for Alzheimer's Disease: A Systematic Review and Meta-Analysis of Treatment Effect. Drugs & Aging, 32(2), 139-147. https://doi.org/10.1007/s40266-015-0239-z

Mitterreiter, S., Page, R. M., Kamp, F., Hopson, J., Winkler, E., Ha, H. R., Hamid, R., Herms, J., Mayer, T. U., Nelson, D. J., Steiner, H., Stahl, T., Zeitschel, U., Rossner, S., Haass, C., & Lichtenthaler, S. F. (2010). Bepridil and amiodarone simultaneously target the Alzheimer's disease beta- and gamma-secretase via distinct mechanisms. J Neurosci, 30(26), 8974-8983. https://doi.org/10.1523/jneurosci.1199-10.2010

Mullane, K., & Williams, M. (2013). Alzheimer's therapeutics: Continued clinical failures question the validity of the amyloid hypothesis—but what lies beyond? Biochemical Pharmacology, 85(3), 289-305. https://doi.org/https://doi.org/10.1016/j.bcp.2012.11.014

Ni, Y., Zhao, X., Bao, G., Zou, L., Teng, L., Wang, Z., Song, M., Xiong, J., Bai, Y., & Pei, G. (2006). Activation of β2-adrenergic receptor stimulates γ-secretase activity and accelerates amyloid plaque formation. Nature Medicine, 12(12), 1390-1396. https://doi.org/10.1038/nm1485

Nishioka, H., Tooi, N., Isobe, T., Nakatsuji, N., & Aiba, K. (2016). BMS-708163 and Nilotinib restore synaptic dysfunction in human embryonic stem cell-derived Alzheimer's disease models. Scientific Reports, 6(1), 33427. https://doi.org/10.1038/srep33427

Nobili, A., Latagliata, E. C., Viscomi, M. T., Cavallucci, V., Cutuli, D., Giacovazzo, G., Krashia, P., Rizzo, F. R., Marino, R., Federici, M., De Bartolo, P., Aversa, D., Dell'Acqua, M. C., Cordella, A., Sancandi, M., Keller, F., Petrosini, L., Puglisi-Allegra, S., Mercuri, N. B., . . . D'Amelio, M. (2017). Dopamine neuronal loss contributes to memory and reward dysfunction in

a model of Alzheimer's disease. Nature Communications, 8(1), 14727. https://doi.org/10.1038/ncomms14727

Nordberg, A., Hellström-Lindahl, E., Lee, M., Johnson, M., Mousavi, M., Hall, R., Perry, E., Bednar, I., & Court, J. (2002). Chronic nicotine treatment reduces beta-amyloidosis in the brain of a mouse model of Alzheimer's disease (APPsw). J Neurochem, 81(3), 655-658. https://doi.org/10.1046/j.1471-4159.2002.00874.x

Norton, W. (2013). Toward developmental models of psychiatric disorders in zebrafish [Review]. Frontiers in Neural Circuits, 7. https://doi.org/10.3389/fncir.2013.00079

Oddo, S., Caccamo, A., Green, K. N., Liang, K., Tran, L., Chen, Y., Leslie, F. M., & LaFerla, F. M. (2005). Chronic nicotine administration exacerbates tau pathology in a transgenic model of Alzheimer's disease. Proceedings of the National Academy of Sciences, 102(8), 3046-3051. https://doi.org/10.1073/pnas.0408500102

Oguchi, T., Ono, R., Tsuji, M., Shozawa, H., Somei, M., Inagaki, M., Mori, Y., Yasumoto, T., Ono, K., & Kiuchi, Y. (2017). Cilostazol Suppresses Aβ-induced Neurotoxicity in SH-SY5Y Cells through Inhibition of Oxidative Stress and MAPK Signaling Pathway. Front Aging Neurosci, 9, 337. https://doi.org/10.3389/fnagi.2017.00337

Palmer, A. M., Wilcock, G. K., Esiri, M. M., Francis, P. T., & Bowen, D. M. (1987). Monoaminergic innervation of the frontal and temporal lobes in Alzheimer's disease. Brain Research, 401(2), 231-238. https://doi.org/https://doi.org/10.1016/0006-8993(87)91408-9

Pandey, D., Banerjee, S., Basu, M., & Mishra, N. (2016). Memory enhancement by Tamoxifen on amyloidosis mouse model. Hormones and Behavior, 79, 70-73. https://doi.org/https://doi.org/10.1016/j.yhbeh.2015.09.004

Park, S. Y., Lee, H. R., Lee, W. S., Shin, H. K., Kim, H. Y., Hong, K. W., & Kim, C. D. (2016). Cilostazol Modulates Autophagic Degradation of β-Amyloid Peptide via SIRT1-Coupled LKB1/AMPKα Signaling in Neuronal Cells. PLoS One, 11(8), e0160620. https://doi.org/10.1371/journal.pone.0160620

Pedersen, W. A., McMillan, P. J., Kulstad, J. J., Leverenz, J. B., Craft, S., & Haynatzki, G. R. (2006). Rosiglitazone attenuates learning and memory deficits in Tg2576 Alzheimer mice. Experimental Neurology, 199(2), 265-273. https://doi.org/https://doi.org/10.1016/j.expneurol.2006.01.018

Pelkowski, S. D., Kapoor, M., Richendrfer, H. A., Wang, X., Colwill, R. M., & Creton, R. (2011). A novel high-throughput imaging system for automated analyses of avoidance behavior in zebrafish larvae. Behavioural Brain Research, 223(1), 135-144. https://doi.org/https://doi.org/10.1016/j.bbr.2011.04.033

Peter, P. Z., James, C. A., Kathleen, M. H., Kala, M., Lawrence, M., & John, C. S. B. (2002). Reduced incidence of AD with NSAID but not H₂ receptor antagonists. Neurology, 59(6), 880. https://doi.org/10.1212/WNL.59.6.880

Petrie, W. M., Ban, T. A., Berney, S., Fujimori, M., Guy, W., Ragheb, M., Wilson, W. H., & Schaffer, J. D. (1982). Loxapine in psychogeriatrics: a placebo- and standard-controlled clinical investigation. J Clin Psychopharmacol, 2(2), 122-126.

Pollock, B. G., Mulsant, B. H., Rosen, J., Sweet, R. A., Mazumdar, S., Bharucha, A., Marin, R., Jacob, N. J., Huber, K. A., Kastango, K. B., & Chew, M. L. (2002). Comparison of Citalopram, Perphenazine, and Placebo for the Acute Treatment of Psychosis and Behavioral Disturbances in Hospitalized, Demented Patients. American Journal of Psychiatry, 159(3), 460-465. https://doi.org/10.1176/appi.ajp.159.3.460

Ramos, B. P., Colgan, L., Nou, E., Ovadia, S., Wilson, S. R., & Arnsten, A. F. T. (2005). The Beta-1 Adrenergic Antagonist, Betaxolol, Improves Working Memory Performance in Rats and Monkeys. Biological Psychiatry, 58(11), 894-900. https://doi.org/https://doi.org/10.1016/j.biopsych.2005.05.022

Ramos, B. P., Colgan, L. A., Nou, E., & Arnsten, A. F. (2008). Beta2 adrenergic agonist, clenbuterol, enhances working memory performance in aging animals. Neurobiol Aging, 29(7), 1060-1069. https://doi.org/10.1016/j.neurobiolaging.2007.02.003

Raskind, M. A. (1998). Psychopharmacology of noncognitive abnormal behaviors in Alzheimer's disease. J Clin Psychiatry, 59 Suppl 9, 28-32.

Reinhardt, S., Stoye, N., Luderer, M., Kiefer, F., Schmitt, U., Lieb, K., & Endres, K. (2018). Identification of disulfiram as a secretase-modulating compound with beneficial effects on Alzheimer's disease hallmarks. Sci Rep, 8(1), 1329. https://doi.org/10.1038/s41598-018-19577-7

Rosenberg, P. B., Mielke, M. M., Tschanz, J., Cook, L., Corcoran, C., Hayden, K. M., Norton, M., Rabins, P. V., Green, R. C., Welsh-Bohmer, K. A., Breitner, J. C. S., Munger, R., & Lyketsos, C. G. (2008). Effects of Cardiovascular Medications on Rate of Functional Decline in Alzheimer Disease. The American Journal of Geriatric Psychiatry, 16(11), 883-892. https://doi.org/https://doi.org/10.1097/JGP.0b013e318181276a

Roy, A., & Chaguturu, R. (2017). Chapter 3 - Holistic Drug Targeting. In B. Patwardhan & R. Chaguturu (Eds.), Innovative Approaches in Drug Discovery (pp. 65-88). Academic Press. https://doi.org/https://doi.org/10.1016/B978-0-12-801814-9.00003-9

Rozkalne, A., Hyman, B. T., & Spires-Jones, T. L. (2011). Calcineurin inhibition with FK506 ameliorates dendritic spine density deficits in plaque-bearing Alzheimer model mice. Neurobiol Dis, 41(3), 650-654. https://doi.org/10.1016/j.nbd.2010.11.014

Sadleir, K. R., Popovic, J., Khatri, A., & Vassar, R. (2022). Oral nimodipine treatment has no effect on amyloid pathology or neuritic dystrophy in the 5XFAD mouse model of amyloidosis. PLOS ONE, 17(2), e0263332. https://doi.org/10.1371/journal.pone.0263332

Saharia, K., Arya, U., Kumar, R., Sahu, R., Das, C. K., Gupta, K., Dwivedi, H., & Subramaniam, J. R. (2012). Reserpine modulates neurotransmitter release to extend lifespan and alleviate age-dependent Aβ proteotoxicity in Caenorhabditis elegans. Exp Gerontol, 47(2), 188-197. https://doi.org/10.1016/j.exger.2011.12.006

Sanz-Blasco, S., Valero, R. A., Rodríguez-Crespo, I., Villalobos, C., & Núñez, L. (2008). Mitochondrial Ca2+ Overload Underlies Aβ Oligomers Neurotoxicity Providing an Unexpected Mechanism of Neuroprotection by NSAIDs. PLoS One, 3(7), e2718. https://doi.org/10.1371/journal.pone.0002718

Sanz, J. M., Chiozzi, P., Colaianna, M., Zotti, M., Ferrari, D., Trabace, L., Zuliani, G., & Di Virgilio, F. (2012). Nimodipine inhibits IL-1β release stimulated by amyloid β from microglia [https://doi.org/10.1111/j.1476-5381.2012.02112.x]. British Journal of Pharmacology, 167(8), 1702-1711. https://doi.org/https://doi.org/10.1111/j.1476-5381.2012.02112.x

Schneider, L., Olin, J. T., Novit, A., & Luczak, S. (2000). Hydergine for dementia. Cochrane Database Syst Rev, 2000(3). https://doi.org/10.1002/14651858.Cd000359

Singh, B., Sharma, B., Jaggi, A. S., & Singh, N. (2012). Attenuating effect of lisinopril and telmisartan in intracerebroventricular streptozotocin induced experimental dementia of Alzheimer's disease type: possible involvement of PPAR-γ agonistic property. Journal of the Renin-Angiotensin-Aldosterone System, 14(2), 124-136. https://doi.org/10.1177/1470320312459977

Singh, H., Sodhi, R. K., Chahal, S. K., & Madan, J. (2020). Meclizine ameliorates memory deficits in streptozotocin-induced experimental dementia in mice: role of nuclear pregnane X receptors. Canadian Journal of Physiology and Pharmacology, 98(6), 383-390. https://doi.org/10.1139/cjpp-2019-0421

Slenter, D. N., Kutmon, M., Hanspers, K., Riutta, A., Windsor, J., Nunes, N., Mélius, J., Cirillo, E., Coort, S. L., Digles, D., Ehrhart, F., Giesbertz, P., Kalafati, M., Martens, M., Miller, R., Nishida, K., Rieswijk, L., Waagmeester, A., Eijssen, L. M. T., . . . Willighagen, E. L. (2018). WikiPathways: a multifaceted pathway database bridging metabolomics to other omics research. Nucleic Acids Research, 46(D1), D661-D667. https://doi.org/10.1093/nar/gkx1064

Srivareerat, M., Tran, T. T., Salim, S., Aleisa, A. M., & Alkadhi, K. A. (2011). Chronic nicotine restores normal Aβ levels and prevents short-term memory and E-LTP impairment in Aβ rat model of Alzheimer's disease. Neurobiology of Aging, 32(5), 834-844. https://doi.org/https://doi.org/10.1016/j.neurobiolaging.2009.04.015

Storga, D., Vrecko, K., Birkmayer, J. G. D., & Reibnegger, G. (1996). Monoaminergic neurotransmitters, their precursors and metabolites in brains of Alzheimer patients. Neuroscience Letters, 203(1), 29-32. https://doi.org/https://doi.org/10.1016/0304-3940(95)12256-7

Sun, L., Wang, X., Liu, S., Wang, Q., Wang, J., Bennecib, M., Gong, C.-X., Sengupta, A., Grundke-Iqbal, I., & Iqbal, K. (2005). Bilateral injection of isoproterenol into hippocampus induces Alzheimer-like hyperphosphorylation of tau and spatial memory deficit in rat. FEBS Letters, 579(1), 251-258. https://doi.org/https://doi.org/10.1016/j.febslet.2004.11.083

Sun, L. M., Chen, H. J., Liang, J. A., & Kao, C. H. (2016). Long-term use of tamoxifen reduces the risk of dementia: a nationwide population-based cohort study. QJM: An International Journal of Medicine, 109(2), 103-109. https://doi.org/10.1093/qjmed/hcv072

Suzuki, K., Aimi, T., Ishihara, T., & Mizushima, T. (2016). Identification of approved drugs that inhibit the binding of amyloid β oligomers to ephrin type-B receptor 2. FEBS Open Bio, 6(5), 461-468. https://doi.org/10.1002/2211-5463.12056

Švob Štrac, D., Pivac, N., & Mück-Šeler, D. (2016). The serotonergic system and cognitive function. 7(1), 35-49. https://doi.org/doi:10.1515/tnsci-2016-0007 (Translational Neuroscience)

Taglialatela, G., Rastellini, C., & Cicalese, L. (2015). Reduced Incidence of Dementia in Solid Organ Transplant Patients Treated with Calcineurin Inhibitors. Journal of Alzheimer's Disease, 47, 329-333. https://doi.org/10.3233/JAD-150065

Thorn, R. J., Dombroski, A., Eller, K., Dominguez-Gonzalez, T. M., Clift, D. E., Baek, P., Seto, R. J., Kahn, E. S., Tucker, S. K., Colwill, R. M., Sello, J. K., & Creton, R. (2019). Analysis of vertebrate vision in a 384-well imaging system. Scientific Reports, 9(1), 13989. https://doi.org/10.1038/s41598-019-50372-0

Tippmann, F., Hundt, J., Schneider, A., Endres, K., & Fahrenholz, F. (2009). Up-regulation of the alpha-secretase ADAM10 by retinoic acid receptors and acitretin. Faseb j, 23(6), 1643-1654. https://doi.org/10.1096/fj.08-121392

Tohgi, H., Abe, T., Takahashi, S., Kimura, M., Takahashi, J., & Kikuchi, T. (1992). Concentrations of serotonin and its related substances in the cerebrospinal fluid in patients with Alzheimer type dementia. Neuroscience Letters, 141(1), 9-12. https://doi.org/https://doi.org/10.1016/0304-3940(92)90322-X

Toledo, E. M., & Inestrosa, N. C. (2010). Activation of Wnt signaling by lithium and rosiglitazone reduced spatial memory impairment and neurodegeneration in brains of an APPswe/PSEN1ΔE9 mouse model of Alzheimer's disease. Molecular Psychiatry, 15(3), 272-285. https://doi.org/10.1038/mp.2009.72

Torika, N., Asraf, K., Cohen, H., & Fleisher-Berkovich, S. (2017). Intranasal telmisartan ameliorates brain pathology in five familial Alzheimer's disease mice. Brain Behav Immun, 64, 80-90. https://doi.org/10.1016/j.bbi.2017.04.001

Townsend, D. J., Mala, B., Hughes, E., Hussain, R., Siligardi, G., Fullwood, N. J., & Middleton, D. A. (2020). Circular Dichroism Spectroscopy Identifies the β-Adrenoceptor Agonist Salbutamol As a Direct Inhibitor of Tau Filament Formation in Vitro. ACS Chemical Neuroscience, 11(14), 2104-2116. https://doi.org/10.1021/acschemneuro.0c00154

Tsukuda, K., Mogi, M., Iwanami, J., Min, L.-J., Sakata, A., Jing, F., Iwai, M., & Horiuchi, M. (2009). Cognitive Deficit in Amyloid-β–Injected Mice Was Improved by Pretreatment With a Low Dose of Telmisartan Partly Because of Peroxisome Proliferator-Activated Receptor-γ Activation. Hypertension, 54(4), 782-787. https://doi.org/10.1161/HYPERTENSIONAHA.109.136879

Tucker Edmister, S., Del Rosario Hernández, T., Ibrahim, R., Brown, C. A., Gore, S. V., Kakodkar, R., Kreiling, J. A., & Creton, R. (2022). Novel use of FDA-approved drugs identified

by cluster analysis of behavioral profiles. Scientific Reports, 12(1), 6120. https://doi.org/10.1038/s41598-022-10133-y

Tucker Edmister, S., Ibrahim, R., Kakodkar, R., Kreiling, J. A., & Creton, R. (2022). A zebrafish model for calcineurin-dependent brain function. Behavioural Brain Research, 416, 113544. https://doi.org/https://doi.org/10.1016/j.bbr.2021.113544

Wang, C. Y., Xie, J. W., Xu, Y., Wang, T., Cai, J. H., Wang, X., Zhao, B. L., An, L., & Wang, Z. Y. (2013). Trientine reduces BACE1 activity and mitigates amyloidosis via the AGE/RAGE/NF-κB pathway in a transgenic mouse model of Alzheimer's disease. Antioxid Redox Signal, 19(17), 2024-2039. https://doi.org/10.1089/ars.2012.5158

Wei, X., Lan, T., Hui-Fu, W., Teng, J., Meng-Shan, T., Lin, T., Qing-Fei, Z., Jie-Qiong, L., Jun, W., & Jin-Tai, Y. (2015). Meta-analysis of modifiable risk factors for Alzheimer's disease. Journal of Neurology, Neurosurgery & Psychiatry, 86(12), 1299. https://doi.org/10.1136/jnnp-2015-310548

Wiesmann, M., Roelofs, M., van der Lugt, R., Heerschap, A., Kiliaan, A. J., & Claassen, J. A. (2017). Angiotensin II, hypertension and angiotensin II receptor antagonism: Roles in the behavioural and brain pathology of a mouse model of Alzheimer's disease. J Cereb Blood Flow Metab, 37(7), 2396-2413. https://doi.org/10.1177/0271678x16667364

Yang, T., Wang, H., Xiong, Y., Chen, C., Duan, K., Jia, J., & Ma, F. (2020). Vitamin D Supplementation Improves Cognitive Function Through Reducing Oxidative Stress Regulated by Telomere Length in Older Adults with Mild Cognitive Impairment: A 12-Month Randomized Controlled Trial. J Alzheimers Dis, 78(4), 1509-1518. https://doi.org/10.3233/jad-200926

Yu, Z. Y., Yi, X., Wang, Y. R., Zeng, G. H., Tan, C. R., Cheng, Y., Sun, P. Y., Liu, Z. H., Wang, Y. J., & Liu, Y. H. (2022). Inhibiting α1-adrenergic receptor signaling pathway ameliorates AD-type pathologies and behavioral deficits in APPswe/PS1 mouse model. J Neurochem, 161(3), 293-307. https://doi.org/10.1111/jnc.15603

Zamponi, G. W. (2016). Targeting voltage-gated calcium channels in neurological and psychiatric diseases. Nature Reviews Drug Discovery, 15(1), 19-34. https://doi.org/10.1038/nrd.2015.5

Zhang, H., Gao, Y., Qiao, P. F., Zhao, F. L., & Yan, Y. (2014). Fenofibrate reduces amyloidogenic processing of APP in APP/PS1 transgenic mice via PPAR-α/PI3-K pathway. Int J Dev Neurosci, 38, 223-231. https://doi.org/10.1016/j.ijdevneu.2014.10.004