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Do Reticulate Relationships Between Tropical Trees Drive Diversification?

Insights from *Inga* (Fabaceae)

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ABSTRACT

Evolutionary radiations underlie much of the species diversity of life on Earth,

particularly within the world's most species-rich tree flora – that of the Amazon rainforest.

Hybridisation catalyses many radiations by generating genetic and phenotypic novelty that

promote rapid speciation, but the influence of hybridisation on Amazonian tree radiations has

been little studied. We address this using the ubiquitous, species-rich neotropical tree genus

Inga, which typifies rapid radiations of rainforest trees. We assess patterns of gene tree

incongruence to ascertain whether hybridisation catalysed rapid radiation in *Inga*. Given the

importance of insect herbivory in structuring rainforest tree communities (and hence the

potential for hybridisation to promote adaptation through admixture of defence traits), we

also test whether introgression of loci underlying chemical defences against herbivory

facilitated rapid speciation in *Inga*. Our phylogenomic analyses of 189/288 *Inga* species

using >1300 target capture loci showed widespread introgression in *Inga* and closely related

genera. Specifically, we found widespread phylogenetic incongruence explained by

introgression, with phylogenetic networks recovering multiple introgression events across

Inga and related genera. In addition, most defence chemistry loci showed evidence of

positive selection and marginally higher levels of introgression. Interestingly, we recovered

around 20% admixed variation between multiple *Inga* species, a similar proportion to that

found in other radiations catalysed by 'ancient' hybridisation. Overall our results suggest that

introgression has occurred widely over the course of *Inga's* history, likely facilitated by

extensive dispersal across Amazonia, and that in some cases introgression of chemical

defence loci may facilitate adaptation in *Inga*.

Keywords: Hybridisation; Introgression; Radiation; Diversification; Phylogenomics;

Amazon; Rainforest; Trees

Rapid evolutionary radiations that generate exceptionally species-rich groups are a

fundamental component of biodiversity (Hughes et al. 2015). Hybridisation (interbreeding

between species) underlies rapid radiation in many taxa (e.g. Barrier et al. 1999; Meier et al.

2017; Lamichhaney et al. 2018) because it allows admixture of existing genetic variation,

which can generate genomic and phenotypic novelty more rapidly than *de novo* mutation

alone (Rieseberg and Willis 2007). Under this model, described as 'combinatorial' speciation

(Marques et al. 2019), genetic and phenotypic variation generated by hybridisation may

confer adaptation to new environments (e.g. adaptive introgression and/or transgressive

segregation (Rieseberg et al. 1999; Suarez-Gonzalez et al. 2018)) or lead to re-sorting of

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intrinsic incompatibilities (e.g. Bateson-Dobzhansky-Muller incompatibilities) that promote

reproductive isolation and hence rapid speciation (Schumer et al. 2015). A relatively small

number of hybridisation events at the 'base' of a clade can catalyse the rapid diversification

of many species from a few initial lineages (Seehausen 2004), as demonstrated in Lake

Victoria's cichlid fishes (Meier et al. 2017) and Hawaiian silverswords (Barrier et al. 1999).

It is possible to detect genetic admixture and infer past reticulation events in a clade

through examining gene tree conflict (Doyle 1992; Naciri and Linder 2015). Specifically, the

proportions of different conflicting gene tree topologies can provide evidence for whether

introgression (interspecific transfer of genetic material following persistent hybridisation) or

incomplete lineage sorting (ILS) underlie observed incongruence (e.g. Green et al. 2010;

Durand et al. 2011; Pease et al. 2018). This incongruence can also help estimate the relative

genetic contributions of progenitor lineages to introgressant descendants (Patterson et al.

2012). There is a growing body of work that explores incongruence to better understand

introgression across the tree of life, particularly in plants (e.g. in oaks (McVay et al. 2017)

and willows (Wagner et al. 2020)), but only recently have such studies been undertaken in the

most species-rich flora on Earth - that of the tropical Americas (e.g. Morales☐Briones et al.

2018; Schley et al. 2020; Larson et al. 2021; reviewed in Schley et al. 2022).

The flora of neotropical rainforests is remarkable in its species diversity (Antonelli and

Sanmartín 2011; Ulloa Ulloa et al. 2017; Raven et al. 2020), particularly at local scales –

there are more tree species in a single hectare of the Amazon (c. 655 spp. (Valencia et al.

2004)) than in all of Europe (c. 454 spp. (Rivers et al. 2019)). Many species-rich neotropical

plant groups arose through recent, rapid radiations (e.g. Erkens et al. 2007; Koenen et al.

2015), but the influence of hybridisation on tropical plant radiations has been little studied,

and virtually not at all in tropical rainforest trees (reviewed in Abbott (2017)). The prevailing

view, based largely on morphological patterns, has been that hybrids between tropical tree

species are exceptionally rare (e.g. Ashton 1969), but this is challenged by recent genomic

data for a few Amazonian tree genera (e.g. *Brownea* (Schley et al. 2020); *Eschweilera*

(Larson et al. 2021)).

The tree genus *Inga* is widespread and abundant in neotropical rainforests, and was the

first documented example of recent, rapid radiation in the Amazonian tree flora (Richardson

et al. 2001). *Inga* exhibits the highest diversification rate of any Amazonian tree genus (Baker

et al. 2014), with *ca.* 300 species arising in the last *ca.* 10 Ma (Ringelberg et al. 2023).

Similar recent, rapid radiation events in other genera gave rise to a large portion of the

Amazonian tree flora, with over half the tree species in the Amazon belonging to genera with

at least 100 species (Dexter and Chave 2016). *Inga* is an ideal study system with which to test

the combinatorial speciation model as a catalyst of radiations in the Amazon due to the large

volume of previous work examining the diversification, ecology and biogeography of the

group (Richardson et al. 2001; Kursar et al. 2009; Dexter et al. 2017; Forrister et al. 2019)

coupled with its ubiquity, abundance and species diversity in the Amazon (Pennington, T. D.

1997). Previous phylogenetic work on *Inga,* based on Sanger sequencing of relatively few

species, revealed low resolution of species-level relationships (Richardson et al. 2001; Kursar

et al. 2009; Dexter et al. 2010), with resolution improving in later phylogenomic studies

using 22 *Inga* species (Nicholls et al. 2015). Here we generate the most comprehensive

phylogenetic tree of *Inga* to date, comprising >1300 loci and 189 species, to help understand

how hybridisation contributed to diversification of the genus.

One factor that could promote hybridisation more widely amongst *Inga* species is their

remarkable level of co-occurrence in local rainforest communities. Up to 19 species of *Inga*

can coexist in 1ha, and such high local diversity is also typical of many other species-rich

Amazonian tree genera (e.g. *Protium* and *Eschweilera* (Valencia et al. 1994)). These

dispersal-assembled local communities of *Inga* (Dexter et al. 2017) are largely structured by

insect herbivore pressure, such that co-occurring *Inga* species differ more in their chemical

defences against herbivores than would be expected by chance (Kursar et al. 2009; Endara et

al. 2022). This is because growing in proximity to congeners with similar chemical defences

results in higher herbivore pressure and hence lower survival probability of individuals

(Forrister et al. 2019). This suggests that herbivore pressure plays a role in divergence and

coexistence between *Inga* species.

Divergent adaptation in chemical defence traits is well documented in *Inga* over

evolutionary timescales (Forrister et al. 2023), likely being driven by the negative density-

dependent processes mentioned above over ecological timescales (Forrister et al. 2019).

Higher densities of conspecifics with the same defences leads to increased mortality from

herbivores (Janzen 1970; Connell 1971), suggesting that it is adaptive to possess rare defence

chemistry phenotypes, as fewer local herbivores can overcome them. Therefore, the transfer

of defence chemistry loci between *Inga* species via hybridisation, followed by positive

selection on those loci, may be predicted to facilitate colonisation of new ecological spaces

and eventually lead to speciation. To our knowledge this has never been investigated from a

genomic perspective. Therefore, here we aim to test the significance of hybridisation in

Inga's diversification and we hypothesise that:

1)   Reticulation occurred at the base of the *Inga* radiation, as well as throughout
120   it, and hence may have contributed to the rapid diversification of the genus throughout
121   its history
122   2)   Genomic loci with the strongest evidence for introgression and natural
123   selection will be related to chemical defences against herbivores

127   **MATERIALS & METHODS**

128   *Taxon Sampling and DNA Sequencing*

We performed phylogenomic analyses on target capture sequencing data from 189 of

the 282 accepted *Inga* species (67%), 109 of which were sequenced for this study, and 80 of

which were taken from previous work (one from Nicholls et al. 2015; 79 from Ringelberg et

al. 2023). Our sampling was based on a list of all accepted *Inga* species compiled using the

World Checklist of Vascular Plants (WCVP 2020) (as of June 2021) and a monograph of

Inga (Pennington, T. D. 1997). Preliminary analyses identified three nested clades within

Inga, which reflect major transitions in substitution rate (the 'fast clade'), shifts in the trait of

indumentum as a defence for young shoots (the 'hairy clade') and shifts in ecology and

defence chemistry (the 'Red hair' clade) (Coley et al. 2018; Ringelberg et al. 2023), and so

downstream analyses are divided into these three nested, monophyletic groups.

To contextualise our *Inga* analyses at broader phylogenetic scales, we collated

sequence data from many outgroup species generated by previous phylogenomic studies

(Koenen et al. 2020; Ringelberg et al. 2022). We included all seven genera and 32 other

species from the 'Inga clade' (*sensu* Koenen et al. 2020) within which *Inga* is nested. In

addition, we included a further 73 species comprising all 42 genera from the 'Ingoid clade'

(sensu Koenen et al. 2020) within which the Inga clade is nested. Finally, we included 22

species from the broader Mimosoid legume clade, giving a total of 127 outgroup species. A

list of accessions sampled including species, sampling location, data source and voucher

information is detailed in Supplementary Table S1, available on Dryad.

DNA was extracted from 20 mg of dried leaf material with the DNeasy Plant Mini Kit

(Qiagen, Hilden, Germany) using modifications described in Nicholls et al. (2015). DNA

library preparation, enrichment and sequencing were carried out either by Arbor BioSciences

(Ann Arbor, MI, USA) or the University of Exeter sequencing service (Exeter, UK)

following the NEBnext Ultra II FS protocol (New England Biolabs, Ipswich, MA, USA).

Targeted bait capture was performed with a subfamily-specific 'Mimobaits' bait set (Nicholls

et al. 2015; Koenen et al. 2020) using the MyBaits protocol v.2 and 3 (Arbor Biosciences,

Ann Arbor, MI, USA). The Mimobaits set targets 1320 loci, including 113 genes associated

with anti-herbivore defence chemistry in *Inga* (hereafter 'defence chemistry' loci). Other loci

targeted by the Mimobaits set are 'single-copy phylogenetically informative' loci (1044),

'differentially expressed' loci (109) and 'miscellaneous' loci (54), that latter of which are

unannotated but contain phylogenetic signal. Enriched libraries were sequenced using the

NovaSeq 6000 platform with a paired-end 150bp run on two S1 flow cell lanes.

162   *Sequence Assembly, Trimming and Alignment*

All analyses were conducted on the UK Crop Diversity Bioinformatics HPC Resource.

DNA sequencing reads were quality-checked with FASTQC 0.11.3 (Andrews 2010) and were

trimmed using TRIMMOMATIC 0.3.6 (Bolger et al. 2014) to remove adapter sequences and to

quality-filter reads. TRIMMOMATIC settings permitted < 2 mismatches, a palindrome clip

threshold of 30 and a simple clip threshold of 10. Bases with a quality score < 28 and reads

shorter than 36 bases long were removed from the dataset. Following quality-filtering, reads

were mapped to target loci using BWA 0.7.17 (Li and Durbin 2009), loci were assembled

using SPADES 3.11.1 (Bankevich et al. 2012) with a coverage cut-off of 5× and exons were

extracted with EXONERATE (Slater and Birney 2005), all of which are implemented in the

HYBPIPER pipeline 1.2 (Johnson et al. 2016). Recovery of loci was visualised using the

'gene_recovery_heatmap.R' script distributed with HYBPIPER. To improve the signal:noise

ratio and remove possible paralogues from our *Inga* dataset we used the Putative Paralogs

Detection pipeline 1.0.1, (PPD; https://github.com/Bean061/putative_paralog; Zhou et al.

(2022)), described in Supplementary Methods on Dryad.

We conducted all subsequent analyses on three data subsets to include a broad range of

phylogenetic scales and to assess the influence of paralogy on our analyses. The first dataset

included one accession for each of 189 *Inga* species with *Zygia* sp. '*mediana*' as the outgroup

(dataset 'Singlesp'). The second dataset included one accession from each of 127 outgroup

species and proportional per-clade sampling of 50 *Inga* species (dataset 'Outgroup'), with

species chosen based on locus recovery. The third and final dataset comprised the same 189

accessions as the 'Singlesp' *Inga* dataset, but loci were assembled and cleaned using the PPD

pipeline, with paralogous loci removed (dataset 'PPD').

Targeted loci for each of the three datasets were aligned by gene region using 1000

iterations in MAFFT 7.453 (Katoh and Standley 2013) with the

'—*adjustdirectionaccurately*' flag to incorporate reversed sequences. These alignments were

then cleaned using the '-*automated1*' flag in trimAl 1.3 (Capella-Gutiérrez et al. 2009), and

realigned with MAFFT using the '—*auto*' flag. This resulted in 1305 refined alignments for

the 'Singlesp' dataset, 1044 for the 'Outgroup' dataset and 1267 loci for the 'PPD' dataset.

Alignment summaries were then generated with AMAS (Borowiec 2016).

193   *Phylogenomic Analyses*

Gene trees were inferred for each locus alignment across the three datasets using IQ-

TREE (Nguyen et al. 2015) by selecting the best-fit substitution model (-*MFP*) while

reducing the impact of severe model violations (-*bnni*) with 1000 ultrafast bootstrap

replicates. Following this, a 'species tree' was generated based on the best-scoring IQtrees

using ASTRALMP 5.15.5 under the default parameters (Zhang et al. 2018) for all three

datasets. Following concatenation of all locus alignments within the 'Singlesp' data subset

using AMAS, we used the same parameters as above to infer a phylogenetic tree from the

concatenated supermatrix using IQ-TREE. Finally, we visualised shared genetic variation

among species by building neighbour net plots with uncorrected P-distances in SPLITSTREE

v4.14.6 (Huson and Bryant 2005) for each of the three datasets.

Analysing Incongruence and Reticulation

To assess incongruence among our gene trees, we estimated three metrics

implemented in the QUARTET SAMPLING method 1.3.1

(https://www.github.com/fephyfofum/quartetsampling; Pease et al. (2018)) based on each

dataset's ASTRAL species tree, using 100 replicate runs. For each node, we estimated

Quartet Concordance (QC) to assess whether there was incongruence, Quartet Differential

(QD) to assess whether one incongruent topology was favoured and Quartet Informativeness

(QI) to assess whether data were sufficiently informative to distinguish well-supported

incongruence from lack of signal (QI not shown as all datasets were informative).

Having assessed incongruence across our three datasets, we then visually investigated

gene tree conflict of certain higher-level relationships highlighted in the Quartet Sampling

analysis with DISCOVISTA (Sayyari et al. 2018) (clades defined in Supplementary Table S2,

available on Dryad). We interpreted gene tree conflict as "low" if the proportion of gene trees

supporting the species tree topology was >30 percent higher than for both alternative

topologies after Kuhnhäuser et al. (2021). We interpreted gene tree conflict as suggesting

introgression if one conflicting topology was above the 33% 'equal frequency' threshold.

We then assessed whether the incongruence we inferred was caused by introgression

or ILS using the *Dtrios* function in DSUITE (Malinsky et al. 2021). We used Patterson's D-

statistic (i.e. the 'ABBA-BABA' test; Green et al. 2010; Durand et al. 2011) and estimated

the proportion of shared variation between species using the F_4 ratio (Patterson et al. 2012),

where for both metrics values closer to 1 indicate more introgression. We generated an input

VCF file for *Dtrios* for each of the three datasets by calling SNPs *de-novo* using BWA 0.7.17,

SAMTOOLS 1.13 (Danecek et al. 2021) and BCFTOOLS 1.13 (Li 2011) as in the 'Speciation

Genomics' github (https://speciationgenomics.github.io), using our target bait set sequences

as the reference. The resulting VCFs were filtered to contain sites with > 8x coverage. We

used the ASTRAL species tree topology from each of the three datasets to inform taxon

relationships for each set of tests and assessed significance of each test using 20 jackknife

resampling runs. Finally, we visualized our D-statistic and F_4 ratio estimates with Ruby

scripts available from https://github.com/mmatschiner. Since F_4 ratios may be correlated

among closely-related species, we additionally estimated the F_{branch} statistic (Malinsky et al.

2018) for each combination of taxa to control for this, and plotted them using the dtools.py

utility distributed with DSUITE.

To investigate species relationships and model historical migration (i.e. introgression)

events, we used TREEMIX v1.13 (Pickrell and Pritchard 2012a) on representative subsets of

our three datasets, down-sampled due to computational limitations. Input VCFs were filtered

to remove all sites with missing data using VCFTOOLS *--max missing* (Danecek et al. 2011) as

suggested for TREEMIX (Pickrell and Pritchard 2012b). We ran TREEMIX with between 0 and

10 migration edges (M), using 5 separate iterations per M value. Since TREEMIX was

designed for population-level sampling, but was the only tractable network method for a

dataset of this size, we used the *-noss* flag to account for having one accession per species.

The best-fit number of migration edges was chosen based on the highest Δm value with the

package *OptM* (Fitak 2021) in R v 4.2.1 (R Development Core Team 2013), following which

the iteration with the highest log likelihood was plotted using the R scripts in the Speciation

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Genomics github (https://speciationgenomics.github.io/Treemix/). Details of SNP calling and

accession selection for TREEMIX are found in Supplementary Methods on Dryad.

Assessing Per-Locus Introgression and Selection

We assessed whether defence chemistry loci experienced elevated introgression and

selection relative to other loci in our target capture dataset. To do this we first estimated the

per-locus proportion of introgression between subclades in our target-capture data using the f

$_{dM}$ statistic (Malinsky et al. 2015) implemented in DSUITE. Given that we observed the most

introgression in the 'hairy clade' of *Inga*, we performed the analysis on five subsets of our

'Singlesp' dataset, containing five different combinations of taxa to represent all subclades

within the Hairy clade (selected subclades shown in Supplementary Fig. S1; subclade

selection described in Supplementary Methods, available on Dryad). For each subset, we

calculated f_{dM} using nonoverlapping windows of 50 informative SNPs with a rolling mean of

one window. We took the absolute values of all f_{dM} scores, as we were only interested in

comparing the magnitude of introgression across loci, and calculated a mean f_{dM} score per-

locus for downstream analyses.

We then assessed whether each of our target-capture loci experienced positive selection

on at least one branch and at least one site using BUSTED (Murrell et al. 2015). We prepared

our target capture alignments for BUSTED analysis by trimming non-homologous sequence

fragments (i.e. intronic regions captured either side of exons), masking misaligned amino

acid residues and producing codon-aware alignments using OMM_MACSE (Ranwez et al.

2011; Ranwez et al. 2021).

Using these codon-aware alignments, we first tested for the presence of selection in

each locus across all taxa in our *Inga* 'Singlesp' dataset. We accounted for false positives by

adjusting the selection test P-values output by BUSTED with a 5% FDR (false discovery

rate) in R (Benjamini and Hochberg 1995). We assessed whether there were associations

between locus selection result ('under selection' if the BUSTED FDR P-value < 0.05) and

locus annotation ('defence chemistry', 'differentially expressed', 'single-copy

phylogenetically informative' and 'miscellaneous') using χ^2 tests in R. We also visualised

associations between selection result and locus annotation using the R package *corrplot* (Wei

et al. 2017). We repeated the BUSTED analysis using identical parameters for five down-

sampled alignments comprising the same 'hairy clade' taxon subsets for which we estimated f

$_{dM}$ (subclades defined in Supplementary Fig. S1, available on Dryad).

Finally, we used analysis of covariance (ANCOVA) to assess whether variation in our

per-locus f_{dM} estimates was explained by interactions between three variables. The variables

were locus annotation, selection result and the length of the locus alignment (to control for

differences in the number of sites between loci). Response variables were square-root

transformed to improve normalcy, and the heteroscedasticity of residuals was examined using

the *plot()* function in R. Box plots of per-locus f_{dM} estimates were generated using *ggplot2*

(Wickham 2016) in R, with locus annotation and selection result as grouping variables. In

total, we estimated both the proportion of introgression and selection for 1167 loci.

290   **RESULTS**

291   *Phylogenomic Analyses*

We achieved a mean of 72.05% reference length recovery onto the *Mimobaits* bait

sequences (78.99%, excluding outgroups, across 1305 loci). Overall, the 'Singlesp' dataset

had 1.53×10^6 sites, with a mean of 0.31% variable sites and 2.30% missing data across all

loci. The PPD dataset had 1.89×10^6 sites, with a mean of 0.30% variable sites and 3.02%

missing data. Finally, the outgroup dataset had 1.44×10^6 sites, with a mean of 0.66% variable

sites and 12.45% missing data. The 'Outgroup' dataset comprised only of the 1044 'single-

copy phylogenetically informative' loci sequenced by previous studies (Koenen et al. 2020;

Ringelberg et al. 2022). Heatmaps showing % recovery per locus are available in

Supplementary Fig. S2, along with summaries of sites, variability and missing data per locus

in Table S3, both available on Dryad.

Our ASTRAL analyses indicated that most bipartitions were well supported across the

three datasets, with local posterior probabilities (LPP) >0.8 (Supplementary Fig. S3ai; Fig.

S3b; Fig. S3c, available on Dryad) and a quartet score of 0.47, 0.69 and 1.36 for the single-

accession-per-species, PPD and outgroup datasets, respectively. Within *Inga,* relationships

between 'Red hair' clade species were less well supported than the rest of the tree, and the

concatenated IQTREE analysis of the Singlesp dataset recovered a nearly identical topology to

the ASTRAL tree with high support (BS >90, Supplementary Fig. S3ai; Fig. S3aii, available

on Dryad). The removal of paralogs with PPD did not materially influence the resolution or

topology of the ASTRAL *Inga* tree (Supplementary Fig. 3c, available on Dryad). Within the

outgroup tree, most generic splits are well supported (LPP >0.9), and *Inga* was monophyletic.

However, three *Zygia* species clustered with other genera (*Z. inundata* and *Z. sabatieri* with

Inga; Z. ocumarensis with *Macrosamanea* (Supplementary Fig. S3b, available on Dryad)).

Genetic variation visualized with SPLITSTREE showed many shared splits within

Inga, Zygia and *Macrosamanea,* as well as between genera (Supplementary Fig. S4a, Fig.

S4b available on Dryad). Shared splits are particularly evident within the 'Hairy' clade of

Inga, along with the 'Red hair' clade nested within it (Supplementary Fig. S4a, available on

Dryad). SPLITSTREE analysis of the Outgroup dataset showed that *Zygia inundata* and *Z.*

sabatieri clustered with *Inga,* while *Z. ocumarensis* clustered with *Macrosamanea*

(Supplementary Fig. S4b, available on Dryad). Removal of 38 paralogous loci with PPD

resulted in a different clustering of *Inga* species, with the long branch leading to *Inga*

gereauana bisecting the Red hair and Hairy clades (Supplementary Fig. S4c, available on

Dryad).

325   *Incongruence is Common Within and Among Genera*

Our analyses recovered evidence of phylogenetic incongruence both within and

between Ingoid clade genera (Fig.1a; Fig. 1b). Negative Quartet Concordance (QC) scores

and DiscoVista analyses suggested that a single conflicting topology was disproportionately

represented at several incongruent nodes, particularly within the 'Hairy' and 'Red hair' clades

of *Inga* (Fig. 1a nodes ii-v; see also QD trees in Supplementary Fig. S6a, available on Dryad).

However, most nodes in the *Inga* ASTRAL tree showed multiple conflicting topologies (i.e.,

QC scores between 0 and 0.5; Fig. 1a). Removal of paralogs with PPD did not materially

influence the results (Supplementary Fig. S5; Fig. S6c). All nodes were shown to be

informative with QI scores.

Most nodes in the outgroup tree recovered much higher QC scores, indicative of more

phylogenetic concordance (Fig. 1b). This was with the exception of a few nodes involving

Zygia, Macrosamanea, *Abarema* and the clade containing *Jupunba,* that had negative QC

scores (Fig. 2a nodes ii-iv; QD trees in Supplementary Fig. S6b, available on Dryad) or

showed multiple conflicting topologies (QC <= 0 => 0.5). For both trees, the deepest

divergences revealed higher proportions of concordant topologies (Fig. 1a node i; Fig. 1b

node i).

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a)

b)

Figure 1

a: *Inga* single accession per species ASTRAL tree with QC values plotted on each node. Nodes of interest are additionally annotated with DiscoVista plots, showing the relative proportions of different discordant topologies. Quartet frequencies are represented as bar graphs, with red bars (left) representing the main topology from the ASTRAL analysis, and with blue and turquoise bars (middle and right) representing alternative topologies. Dashed horizontal lines mark the expectation for equal frequencies of the three possible topologies (Y = 0.333), i.e. maximal gene tree conflict. Node i indicates low proportions of both conflicting alternative topologies. Nodes ii-v indicate one major conflicting topology.

b: *Inga* outgroup ASTRAL tree with QC values plotted on each node. Nodes of interest are additionally annotated with DiscoVista plots, showing the relative proportions of different discordant topologies. Quartet frequencies are represented as bar graphs, with red bars (left) representing the main topology from the ASTRAL analysis, and with blue and turquoise bars (middle and right) representing alternative topologies. Dashed

horizontal lines mark the expectation for equal frequencies of the three possible topologies (Y = 0.333), i.e.
maximal gene tree conflict. Node i indicates low proportions of both conflicting alternative topologies. Nodes ii
- iv indicate one major conflicting topology.

Reticulation Occurs at Multiple Phylogenetic Scales

The overrepresentation of one incongruent topology that we inferred for several nodes

(Fig. 1aii-v; Fig. 1bii-iv) was reinforced by the high *D*-statistics, F_4 ratios and F_{branch} scores

that we calculated for all three datasets. This suggests reticulation contributed to

incongruence at these nodes. Across all methods (D statistics, F_4 ratios and F_{branch})

introgression is evident across entire clades for all three datasets.

Within *Inga*, *D*-statistics up to 0.2 were observed most frequently in the 'Red hair' clade

and the sister group to the 'Hairy' clade (Fig. 2a) although significant *D*-statistics were

evident across the *Inga* tree, particularly within some early-diverging *Inga* lineages.

Similarly, the highest F_4 ratios we recovered suggested that species in the 'Red hair' clade

shared up to 20% of their sequence variation with the earliest diverging lineages in the 'Hairy'

clade. Only the F_4 ratio varied significantly between 'Singlesp' and 'PPD' runs, and revealed

higher proportions of introgression in the 'PPD' run between the 'Red hair' and early-

diverging 'Hairy' subclades (Supplementary Fig. S7, available on Dryad). Estimates of

shared ancestry made using F_{branch}, which controls for correlated introgression in closely-

related lineages, showed again that species in the 'Red hair' clade experienced the most

introgression with other lineages, both within and outside the 'Red hair' clade – particularly

Inga cardozana and *Inga pauciflora* (Supplementary Fig. S8a, available on Dryad).

In the broader Ingoid clade, both *D*-statistics and F_4 ratios suggested that most

significant introgression occurred within *Zygia*, as well as between *Zygia, Inga* and other

Ingoid clade genera (*D* = 0.4, F_4 ratio = ca. 20%; Fig. 2b). Both *D*-statistics and F_4 ratios

showed that significant introgression also occurred between *Jupunba* and *Hydrochorea*, as

well as within *Jupunba* and *Osodendron* (Fig. 2b). F_{branch} scores recovered more limited

evidence of introgression, with the highest scores occurring within closely related species

pairs in *Macrosamanea, Robrichia, Zygia* and *Jupunba* (Supplementary Fig. S8b, available

on Dryad).

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Figure 2

a: Heatmaps of minimum per-triplet D statistic (D_{min}, above) and F_4 ratios (below) plotted for the single-
accession-per-species *Inga* dataset. Taxa P2 and P3 are displayed on the *x*- and *y*-axes in the same order as in
Figure 1a. The colour of each square signifies the *D*-statistic or F_4 ratio estimate (blue = low estimate;
red = high estimate). The saturation of these colours represents the significance for that test (see log(P) in inset
box, bottom right). Clades are marked in the same colours as Fig. 1a, representing the 'Fast', 'Hairy' and 'Red
hair' clades.

b: Heatmaps of minimum per-triplet D statistic (D_{min}, above) and F_4 ratios (below) plotted for the Outgroup
dataset. Taxa P2 and P3 are displayed on the *x*- and *y*-axes in the same order as in Figure 1b. The colour of each
square signifies the *D*-statistic or F_4 ratio estimate (blue = low estimate; red = high estimate). The saturation of
these colours represents the significance for that test (see log(P) in inset box, bottom right). Clades are marked
in the same colours as Fig. 1b, representing the 'Ingoid clade', 'Inga clade' and the genus *Inga*.

Our TREEMIX analyses largely recovered the same clades as our ASTRAL trees (Fig.

1a; Fig. 1b) for both the 'Singlesp' and 'Outgroup' datasets (Fig. 3a; Fig 3b). Within *Inga,*

around 96% of variance was explained when no migration (i.e. introgression) was allowed

(Supplementary Fig. S9a; Table S4, available on Dryad). Adding migration events to the tree

produced better fit, with four edges providing the largest improvement in likelihood ($\Delta m;$

Fig. 3a). We inferred moderate migration from early-diverging *Inga* lineages into the 'Fast'

clade (from the first branch after *I. huberi/glomeriflora* into the base of *I. ruiziana*). We also

inferred more significant migration within the 'Fast' clade (the group subtended by *I.*

tomentosa into *I. nobilis*), from the 'Fast' clade into the 'Hairy' clade (*I. nobilis* into *I.*

tenuicalyx) and from the 'Hairy' clade into the base of the 'Red hair' clade (*I. chrysantha* into

the group subtended by *I. cayennensis*).

Removal of paralogs from the *Inga* dataset with PPD resulted in a similar amount of

variance being explained with no migration edges (ca. 96%), however 6 migration edges

were best supported by TREEMIX for the paralog-free runs (Supplementary Fig. S9c;

Supplementary Table S4, available on Dryad). This run inferred introgression events

involving some of the same branches as the Singlesp dataset (*Inga nobilis*; the base of the

subclade containing *I. leiocalycina;* the base of the Hairy clade; the stem lineage of *I.*

tenuistipula/huberi/glomeriflora) as well as two others (the base of the *Inga umbellifera*

subclade and the split between *I. tessmanii/I. jinicuil*) shown in Supplementary Fig. S10

(available on Dryad).

Among Ingoid clade 'Outgroup' species a high proportion of variance was explained

(99.6%) when no migration was allowed (Supplementary Table S4; Fig. S9b, available on

Dryad). Adding migration events to the tree produced slightly better fit, with two edges

providing the largest improvement in likelihood (Δm) (Fig. 3b). The inferred migration

events occurred from *Jupunba oxyphyllidia* into *Jupunba abbottii* and from *Zygia*

obolingoides into *Zygia morongii*.

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Figure 3

a: *Treemix* plot for the single-accession-per-species *Inga* dataset with M=4 migration edges. Text colours of taxa reflect the clade that they belong to (Brown = 'Fast' clade, light green = 'Hairy' clade, dark green = 'Red hair' clade). Arrows indicate 'migration' (i.e. introgression) events between different branches, and the colour of the arrow depicts the 'migration weight' (i.e. the proportion of shared variation). The drift parameter corresponds to the amount of genetic drift along each population. The scale bar shows ten times the average standard error of the entries in the sample covariance matrix.

b: *Treemix* plot for the outgroup dataset with M=2 migration edges. Text colours of taxa reflect the clade that they belong to (Red = Ingoid clade, orange = Inga clade, green = *Inga*). Arrows indicate 'migration' (i.e. introgression) events between different branches, and the colour of the arrow depicts the 'migration weight' (i.e. the proportion of shared variation). The drift parameter corresponds to the amount of genetic drift along each population. The scale bar shows ten times the average standard error of the entries in the sample covariance matrix.

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Our estimates of introgression and selection varied widely across the 1167 target

capture loci we analysed. Per-locus introgression (f_{dM}) varied between 0.0003 – 0.77 across

all five data subsets, approaching the maximum f_{dM} score of 1 in some datasets (Table S5;

Supplementary Fig. S11, available on Dryad). The 'poeppigiana + hairy clade small +

leiocalycina' subset produced the highest f_{dM} scores overall (Table S5; all clade subsets

illustrated in Supplementary Fig. S1, available on Dryad). Across the analyses, the highest f

$_{dM}$ scores were observed in the single-copy phylogenetically informative loci, which were the

most numerous (Supplementary Fig. S11, available on Dryad).

In total, BUSTED inferred that 768 out of 1167 loci (65.7%) showed evidence of positive

selection in at least one taxon after accounting for false discovery rate. Across all taxa, the χ^2

test revealed that there was a significant association between locus annotation and selection

result ($\chi^2 = 9.575$, df = 3, P-value = 0.022). Furthermore, across all taxa a greater number of

'defence chemistry' loci showed evidence of selection than the null expectation, whereas

fewer 'differentially expressed' and 'miscellaneous' loci than the null expectation showed

evidence of selection (Supplementary Fig. S12, available on Dryad).

Similarly, in the data subset containing the 'leiocalycina + vulpina + red hair'

subclades, more 'defence chemistry' loci showed evidence of selection than the null

expectation, whereas fewer 'miscellaneous' loci showed evidence of selection than the null

expectation (Fig. 4a). However, none of the five data subsets for which we estimated

selection returned significant χ^2 results when this association was tested statistically

(Supplementary Table S6, available on Dryad).

Our ANCOVA analyses showed significant differences in f_{dM} score means between different

locus annotations in the 'leiocalycina + vulpina + red hair' data subset (P=<0.01, d.f. = 3, F =

6.76) (Fig. 4b; 4c showing clade subset tested). Moreover, ANCOVA revealed a marginal

difference in f_{dM} means between locus annotations when loci experienced selection ($P= 0.05$,

df $= 3$, $F = 2.60$) (Fig. 4b). For both 'defence chemistry' and 'differentially expressed loci',

the mean f_{dM} score was markedly higher in loci inferred to be under selection – i.e., loci with

more evidence of introgression tended to show more evidence of selection (Fig. 4b). By

contrast, 'miscellaneous' loci inferred to be under selection had markedly lower f_{dM} scores

(Fig. 4b). ANCOVA did not recover significant differences in f_{dM} scores for any of the other

four data subsets (Supplementary Table S7, available on Dryad).

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Figure 4: Per-locus introgression (f_{dM}) and selection (BUSTED) results summarised for the data subset containing the 'leiocalycina + vulpina + red hair' subclades.

a) Correlation plot showing associations between selection ('under selection' if FDR-corrected *P*-value of BUSTED analysis <0.05) and locus annotation (Defence chemistry; Differentially expressed; Miscellaneous; Single-copy phylogenetically informative). The size of each circle represents the Pearson's residual from χ^2 analysis (P_R in the figure legend). The colour of each circle indicates whether a proportion is above (positive = blue) or below (negative = red) the null expectation of χ^2.

b) Box plots of per-locus proportion of introgression (f_{dM}), grouped by selection result ('under selection' if the *P*-value of BUSTED analysis <0.05) and locus annotation (*x*-axis). Sample sizes are shown vertically above each corresponding box. Medians are shown as horizontal lines in each box.

c) The ASTRAL phylogenetic tree of *Inga* indicating which subclades were used for these analyses – the leiocalycina, vulpina and red hair subclades and highlighted and labelled.

DISCUSSION

Diversification of Inga *and the Ingoid Clade*

Our analyses recovered well-supported phylogenetic trees for *Inga* as well as the

broader Ingoid clade within which it is nested (LPP >0.8, BS >90; Supplementary Fig. S3ai-

ii; Fig. S3b; Fig. S3c available on Dryad). The phylogenetic tree of *Inga* inferred by this

study marks a great advance in the resolution of inter-species relationships when compared to

previous phylogenetic work using fewer loci and species (e.g. Richardson et al. 2001; Kursar

et al. 2009; Dexter et al. 2010; Nicholls et al. 2015). Thus, our phylogenetic tree provides the

best available framework to investigate the role of hybridisation in this diverse group.

Our analyses revealed three nested clades within *Inga* (Fig. 1a). The deepest-level

group is the 'fast' clade, at the base of which there was a substitution rate shift inferred by

other studies (Ringelberg et al. 2023). Nested within the 'Fast' clade is the 'Hairy' clade, in

which many species possess indumentum on young leaves as a defence against herbivores

(Agrawal 1999; Coley et al. 2018). Finally, nested within the 'Hairy' clade is the 'Red hair'

clade, containing species that possess dense indumentum on young leaves and highly diverse

defence chemistry (Coley et al. 2018). SPLITSTREE inferred a high degree of shared genetic

variation within the rapidly diversifying 'Hairy' and 'Red hair' clades of *Inga*, as well as

within and between other Ingoid clade genera (*Macrosamanea, Zygia* and *Jupunba*)

(Supplementary Fig. S4a; S4b, available on Dryad). Of particular interest was the grouping of

multiple *Zygia* species with other South American genera (e.g. *Zygia ocumarensis* with

Macrosamanea; Zygia sabatieri and *Z. inundata* with *Inga*). The non-monophyly of *Zygia*

was first described by Ferm et al. (2019), and further cases of generic non-monophyly within

the Ingoid clade have been highlighted by Ringelberg et al. (2022).

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Phylogenetic Incongruence is Widespread and Influenced by Introgression

We found phylogenetic incongruence both within and between Ingoid clade genera

using Quartet Concordance (QC) scores and DISCOVISTA (Fig.1a; Fig. 1b). Several nodes of

the phylogenetic trees inferred for the Singlesp and Outgroup datasets showed that one

conflicting topology was overrepresented at several incongruent nodes (Fig. 1a nodes ii-v;

Fig 1b nodes ii-iv; see also QD trees in Supplementary Fig. S6a; Fig. S6b, available on

Dryad), which is indicative of introgression at these nodes (Wendel and Doyle 1998).

However, many deeper nodes showed that several conflicting topologies were equally

frequent (QC scores $0 < 0.5$), which is indicative of incomplete lineage sorting (ILS) as is

common in rapidly-diversifying clades such as *Inga* (Degnan and Rosenberg 2009).

Our explicit tests for introgression using *D*-statistic, F_4 ratios and F_{branch} (Fig. 2a; Fig.

2b; Supplementary Fig. S8a, S8b, available on Dryad) inferred widespread introgression

across *Inga* as well as other rapidly-diversifying Amazonian genera (namely *Zygia* and

Jupunba). Our F_4 ratio estimates suggested that up to 20% of genetic variation was shared

between some *Inga* species, with up to 30% of shared variation being evident in the more

conservative F_{branch} analyses (Supplementary Fig. S8a, available on Dryad). Interestingly, this

is a similar proportion of shared variation as reported in other radiations that were catalysed

by 'ancient' hybridisation (e.g., Lake Victoria cichlid fish (Meier et al. 2017)). TREEMIX

inferred at least four migration events in *Inga* (Fig. 3a) and at least two migration events

between smaller Amazonian tree genera (*Zygia* and *Macrosamanea*) (Fig. 3b; Supplementary

Table S4, available on Dryad). These events represent minimum estimates since not all

species were included in the TREEMIX analyses, and many more such events are likely to

have occurred over *Inga's* evolutionary history given the extensive evidence of introgression

we recovered.

Inga may be representative of other large genera in neotropical rainforests, which

account for half of all Amazonian tree diversity and show high sympatry in local

communities (e.g., *Eschweilera, Protium,* (Bermingham and Dick 2001)). Assuming *Inga* is

representative of these other groups, our analyses suggest that introgression is more

widespread than previously thought in species-rich Amazonian tree genera (Ashton 1969),

particularly in the most widespread, abundant and rapidly-diversifying genera. While

introgression is particularly prominent in certain taxa within these radiations (e.g. *Inga*

cardozana and *Inga tenuicalyx*) we also inferred introgression across *Inga*, between species

that are distantly related (e.g. Fig. 3a; Fig. 3b). Our findings are best described by the

syngameon hypothesis of adaptive radiation (Seehausen 2004), which posits that admixture

between species in geographical proximity occurs continually in an ongoing radiation, often

generating functional diversity and promoting speciation. Rapid radiations may also arise

when admixture between recently divergent lineages at the base of a clade triggers rapid

radiation (Seehausen 2004), but we found comparatively little evidence for this hypothesis

since introgression was not concentrated in early-diverging *Inga* lineages.

Periodical hybridisation within a syngameon may be adaptive for Amazonian tree

species, which are typified by large, disparate populations (ter Steege et al. 2013). This is

because periodical hybridisation can elevate genetic diversity and prevent Allee effects, such

as inbreeding, at low population densities (Cannon and Lerdau 2015; Cannon and Lerdau

2019). *Inga* species are known to be highly dispersible, with the entirety of Amazonia acting

as a species pool for the assembly of local *Inga* communities (Dexter et al. 2017). As such,

the introgression we inferred in *Inga* (as well as other in tree genera such as *Zygia* and

Jupunba) likely results from hybridisation between co-occurring, interfertile species (i.e.

syngameons). Indeed, recent work on Amazonian trees has documented putative syngameons

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in other genera, such as between several *Brownea* species (Schley et al. 2020) and amongst

three species of *Eschweilera* around Manaus in the Brazilian Amazon (Larson et al. 2021).

The multiple introgression events that we inferred could have influenced 'sub-

radiations' within *Inga*, *Zygia* and *Jupunba*, which would also explain why we found

introgression across whole clades (Fig. 2a; Fig. 2b; Supplementary Fig. S8a; Fig. S8b

available on Dryad). Shared variation across entire clades would be expected following

introgression events within ancestral syngameons, as introgressant variants are inherited by

descendent species (e.g. Meyer et al. 2017; Meier et al. 2017; Schley et al. 2020). Such an

effect may also result from methodological artefacts which we could not fully account for in

the D-statistic, F_4 ratio and F_{branch} analyses, such as an incorrectly placed lineage within the

trees or changes in substitution rates (Patterson et al. 2012). However, we were highly

stringent with our accession filtering and selection, and substitution rates only change

materially at the base of the 'fast' clade (Ringelberg et al. 2023). In addition, we inferred

introgression into the base of several clades in the TREEMIX analyses, again suggesting

introgressed loci were inherited by descendent species.

Incomplete lineage sorting, i.e. the retention of ancestral polymorphisms in

descendent lineages (Doyle 1992), is also pervasive in rapid Amazonian tree radiations. This

was shown both by our results (Fig. 1a-b; Fig. 2a-2b; Supplementary Fig. 6a; Fig. S6b

available on Dryad) and previously demonstrated extensively in the Mimosoid legumes, to

which the Ingoid clade belongs (Koenen et al. 2020). ILS arises in these groups because the

probability of coalescence (i.e. sorting of derived alleles into descendent lineages reflecting

speciation history) in *t* generations decreases with increasing effective population size (Fisher

1930; Wright 1931; Kingman 2000). Most rainforest trees have large, widespread populations

(ter Steege et al. 2013), so coalescence and sorting of alleles is unlikely to have yet occurred

across all loci in rapidly-diversifying rainforest tree genera such as *Inga* (discussed in

Pennington, R. T. and Lavin 2016).

Introgression and Selection Influence the Evolution of Defence Chemistry Loci in Inga

We detected multiple introgression events across *Inga,* suggesting that some loci

transferred by introgression are retained over time. It is likely that these loci were not

immediately deleterious and were not subject to purifying selection, perhaps residing in areas

of the genome that are distant from incompatibility loci, allowing them to recombine freely

(Edelman et al. 2019). It is also possible that these regions are adaptive and so are maintained

by positive selection, as shown in temperate tree species (e.g. Rendón-Anaya et al. 2021).

This is particularly interesting in the context of chemical defences against insect herbivores,

since these defences are known to be important for survival, co-existence and ecological

divergence in *Inga* (Kursar et al. 2009; Coley et al. 2018; Forrister et al. 2023).

We found differences in the proportion of loci under selection between locus

annotation classes, with more defence chemistry loci under selection than would be expected

by chance. This was evident across all subsampled species ($\chi^2 = 9.575$, df = 3, p-value =

0.022, Fig. 4a; Supplementary Table S6; Fig. S12, available on Dryad). Previous work using

phylogenetic comparative methods demonstrated divergent evolution in defence chemical

profiles among sister species of *Inga* (Forrister et al. 2023). Our results suggest a potential

mechanism underlying this divergent evolution, given the molecular evidence of positive

selection in defence chemistry loci we observed. This provides an important exemplar for

more broadly understanding the assembly of diverse rainforest tree communities, since

herbivore pressure structures tree communities and divergent defence chemistry facilitates

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coexistence at the community scale for rainforest trees like *Inga* (Kursar et al. 2009; Forrister

et al. 2019).

Our estimates of per-locus introgression (f_{dM}) varied widely across the 1167 target

capture loci we analysed and across data subsets (Table S5; Supplementary Fig. S11,

available on Dryad). Locus annotation best explained variance in introgression across loci in

the 'leiocalcycina + vulpina + red hair' data subset (P=<0.01, d.f. = 3, F = 6.76; Fig. 4b; Fig.

4c; Supplementary Table S7, available on Dryad). Moreover, loci inferred to be under

selection had marginally significant differences in the proportion of introgressed sites (P=

0.05, df = 3, F = 2.60; Supplementary Table S7, available on Dryad).

Altogether, these analyses suggest a plausible role of introgression in generating

adaptive defence chemistry phenotypes in *Inga*, given that f_{dM} scores were marginally higher

in defence chemistry loci that were under selection (Fig. 4b). Moreover, novel defence

chemicals in *Inga* are hypothesised to arise from combinations of different chemical

precursors (Coley et al. 2018), suggesting a potential role for admixture. Thus, assuming

introgression generates a novel defence chemical combination that is effective against

herbivores, this phenotype may rise in frequency in a population and facilitate adaptation

where the abundance of individuals with similar defences is low. In other words,

introgression of rare defence loci that confer resistance to different herbivore communities

may facilitate colonisation of new areas with different suites of herbivores. This is similar to

how introgression of wing pattern mimicry genes facilitates adaptation to local mimicry rings

in *Heliconius* butterflies (The Heliconius Genome Consortium 2012).

In light of the Janzen-Connell hypothesis, where higher densities of conspecifics with

the same defences leads to increased mortality from herbivores (Janzen 1970; Connell 1971),

possession of a rare, introgressant defence chemistry phenotype is likely to be adaptive, as

fewer herbivores in the new area can overcome it. Adaptive introgression facilitating

colonisation of new habitats is well known in plants (Suarez☐Gonzalez et al. 2016),

particularly in the context of defence against herbivores (Whitney et al. 2006), and may have

influenced the rapid radiation of *Inga*.

CONCLUSIONS

Our analyses indicate that rapid Amazonian tree radiations (e.g. *Inga, Zygia* and

Jupunba) display evidence of widespread introgression, in addition to incomplete lineage

sorting. The introgression we inferred may be evidence of 'syngameons' of co-occurring

interfertile species, which are created by dispersal-assembled local tree communities in

neotropical rainforests (Dexter et al. 2017). This introgression has likely influenced

concurrent 'sub-radiations', rather than an 'early burst' resulting from a single hybrid swarm

underlying the entire *Inga* radiation. Functional loci relating to defence chemistry

additionally show evidence of selection, and those loci under selection have slightly higher

proportions of introgression, indicating that introgression likely facilitates adaptation, local

coexistence and diversification in Amazonian trees.

SUPPLEMENTARY MATERIAL

Supplementary material is available from the Dryad Digital Repository:

https://doi.org/xxxxxx

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DATA AVAILABILITY

Data available from the Dryad Digital Repository: https://doi.org/10.5061/dryad.69p8cz92v.

The accession numbers for all data collated from previous studies are found in

Supplementary Table S1 on Dryad. All nucleotide sequence data produced by this study are

available on NCBI GenBank under the accession numbers **XXXX**. In addition, all

phylogenetic trees we produced are available on TreeBASE under the accession numbers

XXXX.

REFERENCES

Abbott R.J. 2017. Plant speciation across environmental gradients and the occurrence and
nature of hybrid zones. *J. Syst. Evol*. 55:238-258.

Agrawal A.A. 1999. Induced responses to herbivory in wild radish: effects on several
herbivores and plant fitness. *Ecology*. 80:1713-1723.

Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data.
Available online at http://www.bioinformatics.babraham.ac.uk/projects/fastqc. 0.11.9.

Antonelli A., Sanmartín I. 2011. Why are there so many plant species in the Neotropics?
Taxon. 60: 403-414.

Ashton P.S. 1969. Speciation among tropical forest trees: some deductions in the light of
recent evidence. *Biol. J. Linn. Soc*. 1:155-196.

Baker T.R., Pennington R.T., Magallon S., Gloor E., Laurance W.F., Alexiades M., Alvarez
E., Araujo A., Arets E.J., Aymard G. 2014. Fast demographic traits promote high
diversification rates of Amazonian trees. *Ecol. Lett*. 17:527-536.

Bankevich A., Nurk S., Antipov D., Gurevich A.A., Dvorkin M., Kulikov A.S., Lesin V.M.,
Nikolenko S.I., Pham S., Prjibelski A.D. 2012. SPAdes: a new genome assembly algorithm
and its applications to single-cell sequencing. *J. Comput. Biol*. 19: 455-477.

Barrier M., Baldwin B.G., Robichaux R.H., Purugganan M.D. 1999. Interspecific hybrid
ancestry of a plant adaptive radiation: allopolyploidy of the Hawaiian silversword alliance
(Asteraceae) inferred from floral homeotic gene duplications. *Mol. Biol. Evol*. 16:1105-1113.

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

Benjamini Y., Hochberg Y. 1995. Controlling the false discovery rate: a practical and

powerful approach to multiple testing. *J. R. Stat. Soc. B*: 289-300.

Bermingham E., Dick C. 2001. The *Inga* - newcomer or museum antiquity? *Science*.

293:2214-2216.

Bolger A.M., Lohse M., Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina

sequence data. *Bioinformatics*. 30:2114-2120.

Borowiec M.L. 2016. AMAS: a fast tool for alignment manipulation and computing of

summary statistics. *PeerJ*. 4: e1660.

Cannon C.H., Lerdau M.T. 2019. Demography and destiny: The syngameon in hyperdiverse

systems. *Proc. Natl. Acad. Sci. U.S.A.* 116: 8105.

Cannon C.H., Lerdau M.T. 2015. Variable mating behaviors and the maintenance of tropical

biodiversity. *Front. Genet*. 6:183.

Capella-Gutiérrez S., Silla-Martínez J.M., Gabaldón T. 2009. trimAl: a tool for automated

alignment trimming in large-scale phylogenetic analyses. *Bioinformatics*. 25: 1972-1973.

Coley P.D., Endara M., Kursar T.A. 2018. Consequences of interspecific variation in

defenses and herbivore host choice for the ecology and evolution of *Inga*, a speciose

rainforest tree. *Oecologia*. 187:361-376.

Connell J.H. 1971. On the role of natural enemies in preventing competitive exclusion in

some marine animals and in rain forest trees. In: Den Boer PJ, Gradwell GR, editors.

Dynamics of Populations. Wageningen, The Netherlands: Centre for Agricultural Publishing

and Documentation

Danecek P., Auton A., Abecasis G., Albers C.A., Banks E., DePristo M.A., Handsaker R.E.,

Lunter G., Marth G.T., Sherry S.T. 2011. The variant call format and VCFtools.

Bioinformatics. 27:2156-2158.

Danecek P., Bonfield J.K., Liddle J., Marshall J., Ohan V., Pollard M.O., Whitwham A.,

Keane T., McCarthy S.A., Davies R.M. 2021. Twelve years of SAMtools and BCFtools.

Gigascience. 10:giab008.

Degnan J.H., Rosenberg N.A. 2009. Gene tree discordance, phylogenetic inference and the

multispecies coalescent. *Trends Ecol. Evol*. 24:332-340.

Dexter K.G., Pennington T.D., Cunningham C.W. 2010. Using DNA to assess errors in

tropical tree identifications: How often are ecologists wrong and when does it matter? *Ecol.*

Monogr. 80: 267-286.

Dexter K.G., Chave J. 2016. Evolutionary patterns of range size, abundance and species

richness in Amazonian angiosperm trees. *PeerJ*. 4:e2402.

Dexter K.G., Lavin M., Torke B.M., Twyford A.D., Kursar T.A., Coley P.D., Drake C.,

Hollands R., Pennington R.T. 2017. Dispersal assembly of rain forest tree communities

across the Amazon basin. *Proc. Natl. Acad. Sci. U.S.A*. 114: 2645-2650.

Doyle J.J. 1992. Gene trees and species trees: molecular systematics as one-character

taxonomy. *Syst. Bot.*: 144-163.

Durand E.Y., Patterson N., Reich D., Slatkin M. 2011. Testing for ancient admixture between

closely related populations. *Mol. Biol. Evol*. 28: 2239-2252.

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

Edelman N.B., Frandsen P.B., Miyagi M., Clavijo B., Davey J., Dikow R.B., García-

Accinelli G., Van Belleghem S.M., Patterson N., Neafsey D.E. 2019. Genomic architecture

and introgression shape a butterfly radiation. *Science*. 366:594-599.

Endara M., Soule A.J., Forrister D.L., Dexter K.G., Pennington R.T., Nicholls J.A., Loiseau

O., Kursar T.A., Coley P.D. 2022. The role of plant secondary metabolites in shaping

regional and local plant community assembly. *J. Ecol*. 110:34-45.

Erkens R.H., Chatrou L.W., Maas J.W., van der Niet T., Savolainen V. 2007. A rapid

diversification of rainforest trees (*Guatteria*; Annonaceae) following dispersal from Central

into South America. *Mol. Phylogenet. Evol*. 44: 399-411.

Ferm J., Korall P., Lewis G.P., Ståhl B. 2019. Phylogeny of the Neotropical legume genera

Zygia and *Marmaroxylon* and close relatives. *Taxon*. 68:661-672.

Fisher R.A. 1930. The genetical theory of natural selection. Oxford, UK.: The Clarendon

Press.

Fitak R.R. 2021. OptM: estimating the optimal number of migration edges on population

trees using Treemix. *Biol. Methods Protoc*. 6:bpab017.

Forrister D.L., Endara M., Soule A.J., Younkin G.C., Mills A.G., Lokvam J., Dexter K.G.,

Pennington R.T., Kidner C.A., Nicholls J.A. 2023. Diversity and divergence: evolution of

secondary metabolism in the tropical tree genus *Inga. New Phytol*. 237:631-642.

Forrister D.L., Endara M., Younkin G.C., Coley P.D., Kursar T.A. 2019. Herbivores as

drivers of negative density dependence in tropical forest saplings. *Science*. 363:1213-1216.

Green R.E., Krause J., Briggs A.W., Maricic T., Stenzel U., Kircher M., Patterson N., Li H.,

Zhai W., Fritz M.H., Hansen N.F., Durand E.Y., Malaspinas A.S., Jensen J.D., Marques-

Bonet T., Alkan C., Prufer K., Meyer M., Burbano H.A., Good J.M., Schultz R., Aximu-Petri

A., Butthof A., Hober B., Hoffner B., Siegemund M., Weihmann A., Nusbaum C., Lander

E.S., Russ C., Novod N., Affourtit J., Egholm M., Verna C., Rudan P., Brajkovic D., Kucan

Z., Gusic I., Doronichev V.B., Golovanova L.V., Lalueza-Fox C., de la Rasilla M., Fortea J.,

Rosas A., Schmitz R.W., Johnson P.L.F., Eichler E.E., Falush D., Birney E., Mullikin J.C.,

Slatkin M., Nielsen R., Kelso J., Lachmann M., Reich D., Paabo S. 2010. A draft sequence of

the Neandertal genome. *Science*. 328: 710-722.

Hughes C.E., Nyffeler R., Linder H.P.. 2015. Evolutionary plant radiations: where, when,

why and how? *New Phytol*. 207:249-253.

Huson D.H., Bryant D. 2005. Application of phylogenetic networks in evolutionary studies.

Mol. Biol. Evol. 23: 254-267.

Janzen D.H. 1970. Herbivores and the number of tree species in tropical forests. *Am. Nat*.

104:501-528.

Johnson M.G., Gardner E.M., Liu Y., Medina R., Goffinet B., Shaw A.J., Zerega N.J.,

Wickett N.J.. 2016. HybPiper: Extracting coding sequence and introns for phylogenetics from

high□throughput sequencing reads using target enrichment. *App. Plant. Sci*. 4: 1600016.

Katoh K., Standley D.M.. 2013. MAFFT: Multiple Sequence Alignment Software Version 7:

improvements in performance and usability. *Mol. Biol. Evol*. 30:772-780.

Kingman J.F. 2000. Origins of the coalescent. 1974-1982. *Genetics*. 156: 1461-1463.

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

Koenen E.J., Clarkson J.J., Pennington T.D., Chatrou L.W. 2015. Recently evolved diversity

and convergent radiations of rainforest mahoganies (Meliaceae) shed new light on the origins

of rainforest hyperdiversity. *New Phytol*. 207: 327-339.

Koenen E.J., Kidner C., de Souza É.R., Simon M.F., Iganci J.R., Nicholls J.A., Brown G.K.,

De Queiroz L.P., Luckow M., Lewis G.P. 2020. Hybrid capture of 964 nuclear genes resolves

evolutionary relationships in the mimosoid legumes and reveals the polytomous origins of a

large pantropical radiation. *Am. J. Bot*. 107:1710-1735.

Kuhnhäuser B.G., Bellot S., Couvreur T.L., Dransfield J., Henderson A., Schley R.,

Chomicki G., Eiserhardt W.L., Hiscock S.J., Baker W.J. 2021. A robust phylogenomic

framework for the calamoid palms. *Mol. Phylogenet. Evol*. 157:107067.

Kursar T.A., Dexter K.G., Lokvam J., Pennington R.T., Richardson J.E., Weber M.G.,

Murakami E.T., Drake C., McGregor R., Coley P.D. 2009. The evolution of antiherbivore

defenses and their contribution to species coexistence in the tropical tree genus *Inga. Proc.*

Natl. Acad. Sci. U.S.A. 106: 18073-18078.

Lamichhaney S., Han F., Webster M.T., Andersson L., Grant B.R., Grant P.R. 2018. Rapid

hybrid speciation in Darwin's finches. *Science*. 359: 224-228.

Larson D.A., Vargas O.M., Vicentini A., Dick C.W. 2021. Admixture may be extensive

among hyperdominant Amazon rainforest tree species. *New Phytol*. 232:2520-2534.

Li H. 2011. A statistical framework for SNP calling, mutation discovery, association mapping

and population genetical parameter estimation from sequencing data. *Bioinformatics*.

27:2987-2993.

Li H., Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler

transform. *Bioinformatics*. 25: 1754-1760.

Malinsky M., Challis R.J., Tyers A.M., Schiffels S., Terai Y., Ngatunga B.P., Miska E.A.,

Durbin R., Genner M.J., Turner G.F. 2015. Genomic islands of speciation separate cichlid

ecomorphs in an East African crater lake. *Science*. 350:1493-1498.

Malinsky M., Matschiner M., Svardal H. 2021. Dsuite☐fast D☐statistics and related

admixture evidence from VCF files. *Mol. Ecol. Resour.* 21:584-595.

Malinsky M., Svardal H., Tyers A.M., Miska E.A., Genner M.J., Turner G.F., Durbin R.

2018. Whole-genome sequences of Malawi cichlids reveal multiple radiations interconnected

by gene flow. *Nat. Ecol. Evol*. 2:1940-1955.

Marques D.A., Meier J.I., Seehausen O. 2019. A combinatorial view on speciation and

adaptive radiation. *Trend. Ecol. Evol*. 34: 531-544.

McVay J.D., Hauser D., Hipp A.L., Manos P.S. 2017. Phylogenomics reveals a complex

evolutionary history of lobed-leaf white oaks in western North America. *Genome*. 60:733-

742.

Meier J.I., Marques D.A., Mwaiko S., Wagner C.E., Excoffier L., Seehausen O. 2017.

Ancient hybridization fuels rapid cichlid fish adaptive radiations. *Nat. Comms*. 8: 14363.

Meyer B.S., Matschiner M., Salzburger W. 2017. Disentangling incomplete lineage sorting

and introgression to refine species-tree estimates for Lake Tanganyika cichlid fishes. *Syst.*

Biol. 66:531-550.

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

Morales☐Briones D.F., Liston A., Tank D.C. 2018. Phylogenomic analyses reveal a deep

history of hybridization and polyploidy in the Neotropical genus *Lachemilla* (Rosaceae). *New*

Phytol. 218: 1668-1684.

Murrell B., Weaver S., Smith M.D., Wertheim J.O., Murrell S., Aylward A., Eren K., Pollner

T., Martin D.P., Smith D.M. 2015. Gene-wide identification of episodic selection. *Mol. Biol.*

Evol. 32:1365-1371.

Naciri Y., Linder H.P. 2015. Species delimitation and relationships: the dance of the seven

veils. *Taxon*. 64: 3-16.

Nguyen L., Schmidt H.A., Von Haeseler A., Minh B.Q. 2015. IQ-TREE: a fast and effective

stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol. Biol. Evol*.

32:268-274.

Nicholls J.A., Pennington R.T., Koenen E.J., Hughes C.E., Hearn J., Bunnefeld L., Dexter

K.G., Stone G.N., Kidner C.A. 2015. Using targeted enrichment of nuclear genes to increase

phylogenetic resolution in the neotropical rain forest genus *Inga* (Leguminosae:

Mimosoideae). *Front. Plant Sci*. 6:710.

Patterson N., Moorjani P., Luo Y., Mallick S., Rohland N., Zhan Y., Genschoreck T.,

Webster T., Reich D. 2012. Ancient admixture in human history. *Genetics*. 192:1065-1093.

Pease J.B., Brown J.W., Walker J.F., Hinchliff C.E., Smith S.A. 2018. Quartet Sampling

distinguishes lack of support from conflicting support in the green plant tree of life. *Am. J.*

Bot. 105:385-403.

Pennington R.T., Lavin M. 2016. The contrasting nature of woody plant species in different

neotropical forest biomes reflects differences in ecological stability. *New Phytol*. 210: 25-37.

Pennington T.D. 1997. The Genus *Inga*: Botany. Royal Botanic Gardens, Kew, London, UK:

Kew Publishing

Pickrell J.K., Pritchard J.K. 2012a. Inference of population splits and mixtures from genome-

wide allele frequency data. *PLoS Genet*. 8: e1002967.

Pickrell J.K., Pritchard J.K. 2012b. User Manual for TreeMix v1. Available at:

https://docplayer.net/31797577-User-manual-for-treemix-v1-0-joseph-k-pickrell-jonathan-k-

pritchard.html

R Development Core Team. 2013. R: A language and environment for statistical computing.

Available at: http://www.R-project.org/. 3.6.

Ranwez V., Chantret N., Delsuc F. 2021. Aligning Protein-Coding nucleotide sequences with

MACSE. In: Katoh K, editor. Multiple Sequence Alignment: Methods and Protocols. New

York, NY, USA: Springer p. 51-70.

Ranwez V., Harispe S., Delsuc F., Douzery E.J. 2011. MACSE: Multiple Alignment of

Coding SEquences accounting for frameshifts and stop codons. *PloS one*. 6:e22594.

Raven P.H., Gereau R.E., Phillipson P.B., Chatelain C., Jenkins C.N., Ulloa Ulloa C. 2020.

The distribution of biodiversity richness in the tropics. *Sci. Adv*. 6:eabc6228.

Rendón-Anaya M., Wilson J., Sveinsson S., Fedorkov A., Cottrell J., Bailey M.E., Ruņ⬜is

D., Lexer C., Jansson S., Robinson K.M. 2021. Adaptive introgression facilitates adaptation

to high latitudes in European aspen (*Populus tremula* L.). *Mol. Biol. Evol*. 38:5034-5050.

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

Richardson J.E., Pennington R.T., Pennington T.D., Hollingsworth P.M. 2001. Rapid

diversification of a species-rich genus of neotropical rain forest trees. *Science*. 293: 2242-

2245.

Rieseberg L.H., Archer M.A., Wayne R.K. 1999. Transgressive segregation, adaptation and

speciation. *Heredity*. 83:363-372.

Rieseberg L.H., Willis J.H. 2007. Plant speciation. *Science*. 317:910-914.

Ringelberg J.J., Koenen E.J., Iganci J.R., de Queiroz L.P., Murphy D.J., Gaudeul M.,

Bruneau A., Luckow M., Lewis G.P., Hughes C.E. 2022. Phylogenomic analysis of 997

nuclear genes reveals the need for extensive generic re-delimitation in Caesalpinioideae

(Leguminosae). *PhytoKeys*. 205:3-58.

Ringelberg J.J., Koenen E.J., Sauter B., Aebli A., Rando J.G., Iganci J.R., de Queiroz L.P.,

Murphy D.J., Gaudeul M., Bruneau A. 2023. Precipitation is the main axis of tropical plant

phylogenetic turnover across space and time. *Sci. Adv*. 9:eade4954.

Rivers M., Beech E., Bazos I., Bogunić F., Buira A., Caković D., Carapeto A., Carta A.,

Cornier B., Fenu G. 2019. European Red List of Trees. International Union for Conservation

of Nature and Natural Resources (IUCN).

Sayyari E., Whitfield J.B., Mirarab S. 2018. DiscoVista: interpretable visualizations of gene

tree discordance. *Mol. Phylogenet. Evol*. 122:110-115.

Schley R.J., Pennington R.T., Pérez☐Escobar O.A., Helmstetter A.J., de la Estrella M.,

Larridon I., Sabino Kikuchi I. A. B., Barraclough T.G., Forest F., Klitgård B. 2020.

Introgression across evolutionary scales suggests reticulation contributes to Amazonian tree

diversity. *Mol. Ecol*. 29:4170-4185.

Schley R.J., Twyford A.D., Pennington R.T. 2022. Hybridization: a 'double-edged sword' for

Neotropical plant diversity. *Bot. J. Linn. Soc*. 199:331-356.

Schumer M., Cui R., Rosenthal G.G., Andolfatto P. 2015. Reproductive isolation of hybrid

populations driven by genetic incompatibilities. *PLoS Genet*. 11:e1005041.

Seehausen O. 2004. Hybridization and adaptive radiation. *Trend. Ecol. Evol*. 19:198-207.

Slater G.S.C., Birney E. 2005. Automated generation of heuristics for biological sequence

comparison. *BMC Bioinform*. 6: 31.

Suarez☐Gonzalez A., Hefer C.A., Christe C., Corea O., Lexer C., Cronk Q.C., Douglas C.J.

2016. Genomic and functional approaches reveal a case of adaptive introgression from

Populus balsamifera (balsam poplar) in *P. atrichocarpa* (black cottonwood). *Mol. Ecol*. 25:

2427-2442.

Suarez-Gonzalez A., Lexer C., Cronk Q.C.B. 2018. Adaptive introgression: a plant

perspective. *Biol. Lett*. 14: 10.1098/rsbl.2017.0688.

ter Steege H., Pitman N.C., Sabatier D., Baraloto C., Salomao R.P., Guevara J.E., Phillips

O.L., Castilho C.V., Magnusson W.E., Molino J.F., Monteagudo A., Nunez Vargas P.,

Montero J.C., Feldpausch T.R., Coronado E.N., Killeen T.J., Mostacedo B., Vasquez R.,

Assis R.L., Terborgh J., Wittmann F., Andrade A., Laurance W.F., Laurance S.G., Marimon

B.S., Marimon B.H.Jr., Guimaraes Vieira I.C., Amaral I.L., Brienen R., Castellanos H.,

Cardenas Lopez D., Duivenvoorden J.F., Mogollon H.F., Matos F.D., Davila N., Garcia-

Villacorta R., Stevenson Diaz P.R., Costa F., Emilio T., Levis C., Schietti J., Souza P.,

Alonso A., Dallmeier F., Montoya A.J., Fernandez Piedade M.T., Araujo-Murakami A.,

Arroyo L., Gribel R., Fine P.V., Peres C.A., Toledo M., Aymard C.G.A., Baker T.R., Ceron

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

C., Engel J., Henkel T.W., Maas P., Petronelli P., Stropp J., Zartman C.E., Daly D., Neill D.,

Silveira M., Paredes M.R., Chave J., Lima Filho D. de A., Jorgensen P.M., Fuentes A.,

Schongart J., Cornejo Valverde F., Di Fiore A., Jimenez E.M., Penuela Mora M.C., Phillips

J.F., Rivas G., van Andel T.R., von Hildebrand P., Hoffman B., Zent E.L., Malhi Y., Prieto

A., Rudas A., Ruschell A.R., Silva N., Vos V., Zent S., Oliveira A.A., Schutz A.C., Gonzales

T., Trindade Nascimento M., Ramirez-Angulo H., Sierra R., Tirado M., Umana Medina

M.N., van der Heijden G., Vela C.I., Vilanova Torre E., Vriesendorp C., Wang O., Young

K.R., Baider C., Balslev H., Ferreira C., Mesones I., Torres-Lezama A., Urrego Giraldo L.E.,

Zagt R., Alexiades M.N., Hernandez L., Huamantupa-Chuquimaco I., Milliken W., Palacios

Cuenca W., Pauletto D., Valderrama Sandoval E., Valenzuela Gamarra L., Dexter K.G.,

Feeley K., Lopez-Gonzalez G., Silman M.R. 2013. Hyperdominance in the Amazonian tree

flora. *Science*. 342: 1243092.

The Heliconius Genome Consortium. 2012. Butterfly genome reveals promiscuous exchange

of mimicry adaptations among species. *Nature*. 487:94-98.

Ulloa Ulloa C., Acevedo-Rodriguez P., Beck S., Belgrano M.J., Bernal R., Berry P.E., Brako

L., Celis M., Davidse G., Forzza R.C., Gradstein S.R., Hokche O., Leon B., Leon-Yanez S.,

Magill R.E., Neill D.A., Nee M., Raven P.H., Stimmel H., Strong M.T., Villasenor J.L.,

Zarucchi J.L., Zuloaga F.O., Jorgensen P.M. 2017. An integrated assessment of the vascular

plant species of the Americas. *Science*. 358: 1614-1617.

Valencia R., Balslev H., Miño G.P.Y. 1994. High tree alpha-diversity in Amazonian Ecuador.

Biodivers. Conserv. 3: 21-28.

Valencia R., Foster R.B., Villa G., Condit R., Svenning J., Hernandez C., Romoleroux K.,

Losos E., Magård E., Balslev H. 2004. Tree species distributions and local habitat variation in

the Amazon: large forest plot in eastern Ecuador. *J. Ecol*. 92: 214-229.

Wagner N.D., He L., Hörandl E. 2020. Phylogenomic relationships and evolution of

polyploid *Salix* species revealed by RAD sequencing data. *Front. Plant Sci*. 11:1077.

WCVP. World Checklist of Vascular Plants, version 2.0. Facilitated by the Royal Botanic

Gardens, Kew. Published on the Internet; http://wcvp.science.kew.org/ Retrieved 31st

January 2020. Available from http://wcvp.science.kew.org/.

Wei T., Simko V., Levy M., Xie Y., Jin Y., Zemla J. 2017. Package 'corrplot'. *Statistician*.

56:e24.

Wendel J.F., Doyle J.J. 1998. Phylogenetic incongruence: window into genome history and

molecular evolution. In: Soltis DE, Soltis PS, Doyle JJ, editors. Molecular Systematics of

plants II: DNA sequencing. New York, NY, USA: Springer; p. 265-296.

Whitney K.D., Randell R.A., Rieseberg L.H. 2006. Adaptive introgression of herbivore

resistance traits in the weedy sunflower *Helianthus annuus*. *Am. Nat*. 167:794-807.

Wickham H. 2016. ggplot2: elegant graphics for data analysis. 3.3.6. New York (NY), USA:

Springer-Verlag

Wright S. 1931. Evolution in Mendelian Populations. *Genetics*. 16: 97-159.

Zhang C., Rabiee M, Sayyari E, Mirarab S. 2018. ASTRAL-III: polynomial time species tree

reconstruction from partially resolved gene trees. *BMC Bioinform*. 19:153.

Schley, R.J.; Piñeiro, R.; Nicholls, J.A.; Pezzini, F.F.; Kidner, C; Farbos, A.; Moore, K.; Ringelberg, J.J.; Twyford, A.D.; Dexter, K.G.; Pennington, R.T.

954   Zhou W., Soghigian J, Xiang Q. 2022. A new pipeline for removing paralogs in target

955   enrichment data. *Syst. Biol*. 71:410-425.

a)

D_{min}

F4 Ratio

'Red hair' clade

'Hairy' clade

'Fast' clade

D-statistic

0.0   0.1   0.2

log(p)

-1
-2
-3
-4
-5
-6
-7
-8

0.0   0.1   0.2

f4-ratio

b)

D_{min}

F4 Ratio

Inga

Inga clade

Ingoid clade

D-statistic

0.0   0.1   0.2   0.3   0.4

log(p)

-1
-2
-3
-4
-5
-6
-7
-8

0.0   0.1   0.2

f4-ratio

a)

Migration weight

0.5

Inga_gracilifolia
Inga_kursarii
Inga_glomeriflora
Inga_huberi
Inga_tenuistipula
Inga_brachystachys
Inga_alba
Inga_paraensis
Inga_virgultosa
Inga_umbellifera
Inga_umbratica
Inga_microcalyx
Inga_poeppigiana
Inga_venusta
Inga_chrysantha
Inga_barbourii
Inga_tenuicalyx
Inga_cayennensis
Inga_velutina
Inga_pauciflora
Inga_tonduzii
Inga_chocoensis
Inga_litoralis
Inga_thibaudiana
Inga_ornata
Inga_saltensis
Inga_striata
Inga_ingoides
Inga_cardozana
Inga_lomatophylla
Inga_lineata
Inga_pilosula
Inga_suaveolens
Inga_punctata
Inga_splendens
Inga_spectabilis
Inga_leiocalycina
Inga_ilta
Inga_marginata
Inga_bourgonii
Inga_ruiziana
Inga_alata
Inga_pezizifera
Inga_tomentosa
Inga_multinervis
Inga_acreana
Inga_acrocephala
Inga_melinonis
Inga_nouragensis
Inga_cylindrica
Inga_tessmannii
Inga_jinicuil
Inga_nobilis
Zygia_sp_mediana

10 s.e.

0.00   0.02   0.04   0.06   0.08   0.10   0.12

Drift parameter

Annotation