The evolution of transposable elements in *Brachypodium distachyon* is governed by purifying selection, while neutral and adaptive processes play a minor role

Robert Horvath[1*], Nikolaos Minadakis[1], Yann Bourgeois[2,3], Anne C. Roulin[1*]

[1] Department of Plant and Microbial Biology, University of Zurich, Zollikerstrasse 107, 8008 Zurich, Switzerland

[2] DIADE, University of Montpellier, CIRAD, IRD, Montpellier, France

[3] University House, Winston Churchill Ave, Portsmouth PO1 2UP, United Kingdom

[*] **Corresponding author**: robert.horvath@bluewin.ch; anne.roulin@botinst.uzh.ch

Abstract

Background: Understanding how plants adapt to changing environments and the potential contribution of transposable elements (TEs) to this process is a key question in evolutionary genomics. While TEs have recently been put forward as active players in the context of adaptation, few studies have thoroughly investigated their precise role in plant evolution. Here we used the wild Mediterranean grass *Brachypodium distachyon* as a model species to identify and quantify the forces acting on TEs during the adaptation of this species to various conditions, across its entire geographic range.

Results: Using sequencing data from more than 320 natural *B. distachyon* accessions and a suite of population genomics approaches, we reveal that putatively adaptive TE polymorphisms are rare in wild *B. distachyon* populations. After accounting for changes in past TE activity, we show that only a small proportion of TE polymorphisms evolved neutrally (< 10%), while the vast majority of them are under moderate purifying selection regardless of their distance to genes.

Conclusions: TE polymorphisms should not be ignored when conducting evolutionary studies, as they can be linked to adaptation. However, our study clearly shows that while they have a large potential to cause phenotypic variation in *B. distachyon*, they are not favored during evolution and adaptation over other types of mutations (such as point mutations) in this species.

Key words: Transposable elements, adaptation, *Brachypodium distachyon*, natural selection, age-adjusted SFS

Background

Transposable elements (TEs) are an intrinsic part of eukaryotic genomes and their evolution [1-12]. In addition to modulating genome size, the ability of TEs to create genetic diversity through insertion and excision events can lead to new phenotypes on which selection can act. TEs can alter phenotypes through various mechanisms, including the functional disruption of genes [1, 2], large-scale changes in the regulatory apparatus [3, 4], alteration of epigenetic landscapes [5, 6], ectopic recombination and structural rearrangements [7, 8]. In plants, the dynamics of TE loss and proliferation play a major role in genome evolution [e.g., 9-12]. TEs therefore constitute potentially important drivers of plant evolution, both in nature and during domestication [13].

Beyond their influence on genome structure, and given that their transpositional activity can be stress-inducible [for review 14], TEs are often regarded as more likely than classical point mutations to produce the diversity needed for individuals to respond quickly to challenging environments [15-17]. For instance, punctual TE polymorphisms can lead to gains of fitness and evolve under positive selection [2, 20-25]. TE polymorphisms can even induce more extreme changes in gene expression than single nucleotide polymorphisms (SNPs) in plants [18, 19].

Despite such evidence, whether TE polymorphisms are major contributors to adaptation to changing environments is still debated. Indeed, TE transposition can be disruptive, and purifying selection has been shown to play an important role in TE evolution [e.g., 30, 32]. Based on simulations, it has been suggested that the persistence of TE polymorphisms within a genome without an uncontrolled accumulation, can only be achieved if weak purifying selection is the main force governing TE evolution [33-36]. The uncertainty surrounding this important question in evolutionary genomics results from the limited number of studies that comprehensively tested

the extent to which selection shapes TE allele frequencies, both in plants [25, 26] and animals

[27-31] and characterized the distribution of fitness effects of new TE insertions. To clarify this

question, we used the plant model system *Brachypodium distachyon* [37] to disentangle the

effects of purifying and positive selection on TE polymorphisms in natural populations.

B. distachyon is a wild annual grass endemic to the Mediterranean basin and Middle East.

Recent genetic studies based on more than 320 natural accessions spanning from Spain to Iraq

(hereafter referred to as the *B. distachyon* diversity panel) revealed that *B. distachyon* accessions

cluster into three main genetic lineages (the A, B and C genetic lineages), which further divide

into five main genetic clades that display little evidence for historical gene flow (Fig. 1A; [38, 39]).

Niche modeling analyses suggest that the species moved southward during the last glacial period

and recolonized Europe and the Middle East within the last five thousand years [39].

Consequently, while some *B. distachyon* genetic clades currently occur in the same broad

geographical areas (Fig. 1A), natural accessions are adapted to a mosaic of habitats [38, 39].

These past and more recent shifts in the species distribution led to clear footprints of positive

selection in the genome [39, 40] and make *B. distachyon* an ideal study system to investigate the

contribution of TEs to the adaptation of plants in the context of environmental changes.

In *B. distachyon*, TEs are exhaustively annotated and account for approximately 30% of

the genome [37]. Recent TE activity has been reported for many families, but despite past

independent bottlenecks and expansions experienced by the different genetic clades, no lineage-

specific TE family activity has been observed [32]. Rather, TE activity tends to be homogeneous

throughout the species range and across genetic clades, indicating a high level of conservation of

the TE regulatory apparatus [32]. While purifying selection shapes the accumulation patterns of

TEs in this species [32], some TE polymorphisms have been observed in the vicinity of genes [32],

potentially affecting gene expression [41]. These early studies, based on a relatively small number

of accessions originating exclusively from Spain and Turkey, suggested that TE polymorphisms

could contribute to functional divergence and local adaptation in *B. distachyon* [32].

To test this hypothesis, we used the *B. distachyon* diversity panel to identify TE

polymorphisms in a large set of 326 natural accessions spanning the whole species distribution.

We combined a set of population genomic analyses to assess the proportion of TE polymorphisms

associated with positive or purifying selection as well as neutral evolution. We also quantified

the strength of purifying selection through forward simulations. Altogether, our work provides

the first quantitative estimate of the adaptive, neutral, and disruptive potential of TEs, while

accounting for changes in TE activity, in a plant harboring a relatively small genome. Altogether,

our result advocate against an extended role of TEs in recent adaptation.

Results

Genetic variation in *Brachypodium distachyon*

Using the *B. distachyon* diversity panel (Fig. 1A), we identified 97,660 TE polymorphisms in our

B. distachyon dataset, of which 9,172 were retrotransposons, 52,249 were DNA-transposons and

36,239 were unclassified. We also identified 9 million SNPs across the 326 samples, including

182,801 synonymous SNPs. A Principal Component Analysis (PCA) performed either with SNPs or

TE polymorphisms reflects the previously described population structure of *B. distachyon* [38,

39], with the first two components of the PCA splitting the data according to the demographic

structure (Additional file 1: Fig. S1). Investigating the genetic variation caused by

retrotransposons and DNA-transposons revealed that the observed diversity in retrotransposons

strongly correlated with the demographic structure (Mantel test; r = 0.79, *p* value = 0.001), while

the observed diversity in DNA-transposons only had a weaker correlation (Mantel test; r = 0.36,

p value = 0.001) with the demographic structure (Additional file 1: Fig. S2).

Fig 1. Distribution of the studied accessions and TE polymorphism frequencies. Panel A: Map showing the geographical distribution of the accessions used in the current study. The phylogenetic tree illustrates the phylogeny between the five genetic clades. This panel was made based on the data and results published by Stritt et al. [38] and Minadakis et al. [39]. Panel B: Observed (blue) and simulated (gray) X^tX values of TE polymorphisms in *B. distachyon*. Dotted lines show the 2.5% and 97.5% quantiles of the simulated X^tX values. Panel C-G: Folded site frequency spectrum of TE polymorphisms and synonymous SNPs in all clades. C: A_East; D: A_Italia; E: B_West; F: B_East; G: C.

From the initial TE and SNP dataset, we could estimate the time of origin in generations

(age) of 50,891 TE polymorphisms and 108,855 synonymous SNPs based on pairwise differences

in identity by descent (IBD) regions around the focal mutation (see Materials and methods). The

results of the age estimate analysis were checked by contrasting the observed correlation

between allele age and frequency of synonymous SNPs to the theoretical predictions of Kimura

and Ohta [42] for neutrally evolving mutations. We found that, the observed correlation matched

expectations (Additional file 1: Fig. S3), with older alleles found on average at higher frequencies

than younger ones. Furthermore, most TE polymorphisms in our dataset were young and only a

few were very old (Additional file 1: Fig. S4).

The overall contribution of TEs to clade differentiation and adaptation is limited

To examine the overall contribution of TEs to evolution and adaptation in *B. distachyon*, we first

identified regions of the genomes that were likely affected by recent selective sweeps. The fast

increase in the frequency of a beneficial allele is expected to lead to a longer than average

haplotype around the mutation under positive selection. Such events (known as selective

sweeps) can be identified by computing the integrated haplotype score (iHS) around focal

mutations [43]. We therefore computed iHS along the genome for the four derived genetic

clades. Regions of the genomes with significantly higher iHS than average are expected to harbor

mutations that were under positive selection during evolution and adaptation. We hypothesized

that if TEs constitute an important part of the genetic makeup that led to adaptation in a given

genetic clade, then they should be more frequently fixed or at higher frequencies in regions with

high iHS than in the corresponding regions that did not experience recent selective sweeps in

other clades.

First, we tested if more TE polymorphisms were fixed in a specific region of the genome

if a genetic clade had a high iHS, and presumably experienced a selective sweep, than in other

genetic clades. An analysis of covariance (ANCOVA) revealed that the number of fixed TE

polymorphisms per clade did not significantly differ between high iHS regions and the same

regions in other clades (Table 1). These results indicate that there is no correlation between the

overall number of fixed TE polymorphisms per clade in a region and recent selective sweeps.

However, the number of fixed TEs in genomic regions along the genome was significantly affected

by the total number of TEs in the region, the TE superfamily, the TE age, the genetic clade and

the overall genetic features of the region (e.g., recombination rate, see Materials and methods)

but not by the iHS itself (Table 1). Similarly, we tested if the allele frequency of TE polymorphisms

was significantly higher in a specific region of the genome if a genetic clade had a high iHS than

in other genetic clades. A second ANCOVA revealed that the allele frequency of TE

polymorphisms was significantly influenced by the TE superfamily, TE age, clade and overall

genetic features of the region but not by the iHS (Table 2). These results indicate that TEs in high

iHS regions did not experience a significant increase in their frequency and that TEs in high iHS

regions are experiencing the same selective constraints as other TEs.

Table 1 ANCOVA predicting the number of fixed TE polymorphisms per clade in candidate regions
under positive selection.

Variable	Sum of squares	degrees of freedom	F value	P value
Total number of TEs in the region	28969.6	1	35405.64	< 0.001
TE superfamily	887.5	14	77.48	< 0.001
Clade	587	3	239.13	< 0.001
Genomic region	136.7	80	2.09	< 0.001
TE age	45.5	2	27.81	< 0.001
High iHS	0	1	0.03	0.869

149 **Table 2** ANCOVA predicting the allele frequency of TE polymorphisms per clade in candidate
150 regions under positive selection.

Variable	Sum of squares	degrees of freedom	F value	P value
TE superfamily	453.2	14	247.3	< 0.001
Clade	17.7	3	45.18	< 0.001
Genomic region	147	80	14	< 0.001
TE age	2	2	7.7	< 0.001
High iHS	0.1	1	0.79	0.374

A complementary approach to explore the impact of positive selection on TEs consists in

investigating their genetic differentiation among populations. Using the five genetic clades as

focal populations, we computed X^tX values, a standardized measure of genetic differentiation

corrected for the neutral covariance structure across populations [44, 45], for each TE

polymorphism. Mutations affected by positive selection are expected to be over-differentiated

between clades and display significantly higher X^tX values than other mutations [45]. In contrast,

a low X^tX value implies that the mutation is less differentiated than other mutations and

potentially evolves under balancing selection, whereas purifying selection and a neutral

evolution are not expected to impact the differentiation of a mutation among populations [44].

We contrasted the observed X^tX values computed for each TE polymorphism to a simulated

pseudo-observed dataset (simulated observations under the demographic model inferred from

the covariance matrix of the SNP dataset, for more details see [45]) and found that only a small

fraction of the TE polymorphisms (0.06%) displayed X^tX values higher than the 97.5% quantile of

the simulated values (Fig. 1B). This indicates that only a few TE polymorphisms are over-

differentiated among genetic clades and might have been affected by positive selection.

However, a relatively larger portion of the TE polymorphisms (4.3%) displayed X^tX values smaller

than the 2.5% quantile of the simulated values (Fig. 1B), indicating that balancing selection might

also shape TE frequency in *B. distachyon.*

To further examine the contribution of TEs to adaptation, we tested whether and how

many TE polymorphisms were significantly associated with environmental factors. If the presence

of a TE provides an advantage in a certain environment and contributes to adaptation, we

expected a correlation between the environment and the presence/absence of this TE. In this

context, we performed genome-environment association analyses (GEA) using all TEs and SNPs

identified across the 326 samples and 32 environmental factors associated with precipitation,

solar radiation, temperature, elevation and aridity (see in Materials and methods for the full list).

The GEA revealed that only nine of the 97,660 TE polymorphisms were significantly associated

with some environmental factors (Additional file 2: Table S1), confirming that TEs only had a

limited contribution to adaptation in *B. distachyon.* Importantly, two of these nine TEs were

found in a gene, and three were in the vicinity of genes (less than 2 kilobase (kb) away, Additional

file 2: Table S1).

Purifying selection dominates the evolution of TE polymorphisms in *B. distachyon*

To further characterize the forces governing the evolution of TE polymorphisms in *B. distachyon*,

we examined the genome-wide frequency distribution of TEs. We first computed the folded site

frequency spectrum (SFS) and found that the folded SFS of TE polymorphisms was shifted toward

a higher proportion of rare minor alleles compared to neutral sites in all genetic clades (Fig. 1C-

G). Splitting the TE data into DNA-transposons and retrotransposons resulted in similar folded

SFS and shifts in both TE classes (Additional file 1: Fig. S5 and S6).

These shifts could be the result of purifying selection as the analyses presented above

indicate that positive selection has a negligible effect on TE polymorphism frequencies in

B. distachyon. However, in contrast to SNPs, TEs do not evolve in a clock-like manner, as their

transposition rate is known to vary between generations [46, 47]. Changes in transposition rate

and purifying selection can lead to similar shifts in the SFS but can be disentangled using age-

adjusted SFS [48]. In brief, if TE polymorphisms are evolving neutrally, they are expected to

accumulate on average at the same rate in a population as neutral SNPs of the same age. Hence,

Δ frequency, the difference between the average frequency of TE polymorphisms and neutral

sites in a specific age bin, will remain close to 0 regardless of the polymorphisms' age. In contrast,

if TE polymorphisms evolve under purifying selection, they will tend to occur at lower frequencies

than neutral SNPs of the same age, as selection will prevent them from accumulating in the

population. Consequently, the Δ frequency will reach negative values for older TE polymorphisms

[48].

Because this model does not allow for back mutations, as typically observed for DNA-

transposons that can excise from the genome, we primarily investigated the age-adjusted SFS of

retrotransposons in the four derived clades. This analysis revealed that retrotransposons are

indeed prevented by natural selection from randomly accumulating, as older retrotransposons

are significantly less frequent than neutral SNPs of the same age (Fig. 2; one-sided Wilcoxon test,

Bonferroni corrected p value < 0.01).

As previous studies showed that the distance between TE polymorphisms and the next

gene can impact the strength of selection affecting TEs [6, 49, 50], we further split our

retrotransposon polymorphisms into three categories based on their distance to the next gene:

retrotransposons (i) in and up to 1 kb away from genes, (ii) between 1 kb and 5 kb away and (iii)

more than 5 kb away. The age-adjusted SFS of all three categories displayed the same pattern as

that observed for the whole retrotransposon polymorphism dataset: older retrotransposon

polymorphisms were significantly less frequent than neutral sites of the same age regardless of

their distance to genes (one-sided Wilcoxon test, Bonferroni corrected p value < 0.01), indicating

 that retrotransposons more than 5 kb away from genes are also affected by purifying selection

 (Fig. 2).

Fig. 2 Age-adjusted SFS of retrotransposons. The top row shows the age-adjusted SFS of all retrotransposons (colored), non-synonymous SNPs (light gray) and high effect SNPs (dark gray) in the four derived clades. The bottom row shows the age-adjusted SFS of retrotransposons based on their distance to the next gene in the four derived clades. The X axes show the age range of the mutations in each bin, and the age range of each bin was chosen so that each bin represents the same number of retrotransposon observations in the top row. The different columns show the four derived clades: A: A_East; B: A_Italia; C: B_West; D: B_East. Boxplots are based on 100 estimations of Δ frequency. Significant deviations of Δ frequency estimates from 0 in the age-adjusted SFS of retrotransposons are shown with asterisks (one-side Wilcoxon tests, Bonferroni corrected *p* value < 0.01: ***).

Retrotransposon polymorphisms tended to be more deleterious than SNPs predicted to

have a high impact on fitness. Indeed, the age-adjusted SFS of retrotransposons resulted in a

larger deviation of Δ frequency from 0 than for non-synonymous SNPs and high effect SNPs (Fig.

2). In addition, Δ frequency in the oldest (last) age bin was significantly smaller than in all other

age bins in the A_East, B_East and B_west clades (one-sided Wilcoxon test, Bonferroni corrected

p value < 0.01). In the A_Italia clades the oldest age bin was not significantly different form the

second oldest age bin (two-sided Wilcoxon test, Bonferroni corrected p value N.S.). While older

non-synonymous SNPs and high effect SNPs were generally less frequent than neutrally evolving

SNPs at the same age, the negative Δ frequency trend was reversed for the oldest non-

synonymous SNPs and high effect SNPs (Fig. 2). In all clades, Δ frequency in the oldest age bin

was significantly higher than at least the lowest Δ frequency observed in the other age bins for

non-synonymous SNPs, as well as high effect SNPs (one-sided Wilcoxon test, Bonferroni

corrected p value < 0.01). This might be because not all predicted non-synonymous SNPs and

high effect SNPs might result in fitness effects. Those SNPs can therefore evolve neutrally or

nearly neutrally and persist as polymorphic SNPs much longer in a population than those

affecting fitness negatively. Hence, the last age bin of the non-synonymous and high effect SNP

age-adjusted SFS likely harbors mainly neutrally and nearly neutrally evolving mutations, and

consequently, Δ frequency in not the smallest in the last age bin in these age-adjusted SFS.

To assess whether similar forces may drive retrotransposon and DNA-transposon

evolution, we repeated the analysis for DNA-transposons. The age-adjusted SFS of DNA-

transposons revealed very similar patterns, with Δ frequency showing significant deviations from

0 in older age bins (one-sided Wilcoxon test, Bonferroni corrected p value < 0.01) and DNA-

transposon polymorphisms being more deleterious than non-synonymous SNPs and high effect

SNPs (Additional file 1: Fig. S9).

Forward simulations allow us to quantify the strength of purifying selection

To evaluate to what extent the proportion of neutrally evolving mutations in the focal group of

mutations affects the shape of the age-adjusted SFS, we ran forward simulation with mutations

under multiple selective constraints, and we tested what ratio of neutral to selected mutations

can lead to an age-adjusted SFS similar to that observed for retrotransposons in *B. distachyon*.

Specifically, we investigated the conditions under which we observed a Δ frequency in the oldest

age bin significantly smaller than Δ frequency in all other age bins. Our simulations revealed that

the shape of the age-adjusted SFS of retrotransposons could only be reproduced if less than 10%

of the mutations were neutrally evolving for most of the selective constraint investigated

(Additional file 1: Fig. S7 and Additional file 2: Table S2).

Finally, we used the results from our simulations to narrow down the selection strength

affecting retrotransposons in *B. distachyon* by investigating the age of the oldest

retrotransposons in our dataset. The main difference between the age-adjusted SFS of mutations

evolving under weak and strong purifying selection is that the oldest mutations are much older

in the simulation with weak purifying selection than in the simulation with strong purifying

selection. This age difference arises because mutations under strong purifying selection are

removed from the population more effectively and, therefore, cannot persist as long in the

population. Examining the age of the last retrotransposon bins in the age-adjusted SFS revealed

that the ages of the oldest retrotransposons were the most similar to the expected ages of the

oldest mutations in our simulations, with a scaled selection coefficient (S) of -5 and -8 (Fig. 3),

indicating that retrotransposons in *B. distachyon* are under moderate purifying selection. In

simulations with a nearly neutral selection coefficient (S = -1), the simulated mutations were

much older than the oldest observed retrotransposons (Fig. 3). Conversely, in simulations with a

strong purifying selection coefficient (S < -10), they were much younger than the oldest observed

retrotransposons (Fig. 3).

Fig. 3 Relative age difference ((mutation age in simulations - observed mutation age)/maximum absolute age difference) between simulated and observed data in the last bin of the age-adjusted SFS. A: 25% quantile; B: 50% quantile; C: 75% quantile.

Discussion

B. distachyon is a widely used model species in evolutionary genomics, molecular ecology,

developmental biology and crop functional genomics [for review 51, 52] with past and ongoing

TE movements in its genome [32]. In this study, we used a diversity panel containing next-

generation sequencing data from over 320 individuals sampled across the whole geographical

range of *B. distachyon* to examine the role of TEs during evolution and adaptation. We

investigated the frequency with which positive selection led to an increase in the frequency and

fixation of TEs and quantified the strength of purifying selection on TE polymorphisms.

Accounting for population structure and fluctuant transposition rates, we demonstrate that TEs

are rarely part of the genetic makeup that was positively selected during environmental

adaptation in *B. distachyon*. Furthermore, we show that the majority of TE polymorphisms found

in the natural population of this model species are under weak to moderate purifying selection,

with only a small minority of TE polymorphisms evolving neutrally.

Rare instances of positive selection on TEs

By combining complementary approaches, we were able to demonstrate that TEs are rarely the

target of positive selection in *B. distachyon*. We first probed for footprints of positive selection

on TE polymorphisms using the five genetic clades as focal populations. In conducting this

analysis, we did not find TE polymorphisms to be at high frequencies or fixed at higher rates than

expected, in regions of the genome presumably harboring selective sweeps in at least one of the

genetic clades (high iHS regions). This suggested that TEs were rarely the target of positive

selection, which we confirmed with a genome-wide scan for overly differentiated TE

polymorphisms using X^tX analysis. Indeed, this approach revealed that only a very small

proportion of TE polymorphisms are more differentiated than expected under a neutral scenario.

296   Importantly, the X^tX analysis also revealed that a non-negligible fraction of the TE

polymorphisms is less differentiated than expected and are shared among genetic clusters. This

could be the result of selection favoring the same TE polymorphisms in different accessions to

adapt to similar environmental constraints across genetic clades. To test this scenario, we

performed GEA with 32 environmental factors, and found only nine TE polymorphisms

significantly associated with any of these, and representing a very small proportion (< 0.01%) of

all the TE polymorphisms we identified. Interestingly though, these nine TE polymorphisms were

associated with environmental variables pertaining to precipitation, temperature and altitude,

which are known to drive adaptation in *B. distachyon* [39]. Some insertions were found within or

in close proximity of genes, making these polymorphisms very good candidates for future

functional validation.

307   Single TE insertions can have a drastic impact on phenotypic variation and be affected by

positive selection [for review, see 16, 53, 54]. For instance, TEs have increased in frequency

through positive selection in humans [23] or during range expansion in *Arabidopsis* [25] and

D. melanogaster [20, 22]. Evidently, *B. distachyon* exhibits a different pattern, as causal

mutations for adaptation in this grass species are rarely TEs. Only a few studies have thoroughly

quantified the extent to which positive selection influences the evolution of TEs [25, 26, 28, 30,

31]. But two of these drew similar conclusions to us, in the green anole *Anolis carolinensis* [30]

and in the invasive species *Drosophila Suzukii* [31]. In addition, a large number of candidate genes

for adaptation were identified with a similar approach focusing on SNPs [39], indicating that

population structure or demographic events are not limiting factors for the methods we used.

Altogether, these observations call for a closer investigation of which forces, e.g., purifying

selection or neutral evolution, are important in shaping TE allele frequency in natural

populations.

Moderate purifying selection is the dominant force during TE evolution

Our results suggest that purifying selection is an important factor limiting the ability of TE

polymorphisms to fix and increase their frequency in *B. distachyon*. Indeed, one of the significant

explanatory variables in our ANCOVA models was the genetic clade, a proxy for the effective

population size (N_e), which affects the efficiency with which selection can fix beneficial mutations

and purge deleterious ones. In *B. distachyon*, the number of fixed TE polymorphisms per clade

and the frequency of TE polymorphisms were negatively correlated with N_e, indicating that the

accumulation of TEs is significantly lower in genetic clades with a larger N_e, potentially because

of a greater efficacy of purifying selection.

It is widely accepted that most new TE insertions have a deleterious or no effect on the

fitness of the host [22, 26, 28, 30, 33-36, 55]. To properly quantify the effect of purifying selection

on TE evolution in *B. distachyon*, we used age-adjusted SFS analyses to evaluate the selective

constraint experienced by TE polymorphisms while accounting for previously reported changes

in their activity [32]. While this method can only be applied to retrotransposons (because the

model does not allow back mutations), it provided a first clue on the importance of purifying

selection on TE evolution and revealed that overall, retrotransposons evolved under purifying

selection in all four derived genetic clades. Indeed, the Δ frequency was significantly smaller than

0, especially for older retrotransposons, meaning that old retrotransposons are less common

than neutrally evolving SNPs at the same age. This further demonstrates that even after

accounting for the different genetic clades and using a large sample size, retrotransposons evolve

under purifying selection in *B. distachyon*.

We also revealed that only a minority of retrotransposons evolved neutrally, as the

observed shape of the Δ frequency curve could only be reproduced in our simulation if the

proportion of neutrally evolving mutations in our focal mutations was below 10%. This estimate

gives a first glimpse into the distribution of fitness effects of new TE insertions, a fundamental

parameter in genetics that describes the way in which new TE insertions can contribute to

evolution and adaptation [56]. Here, we show for the first time that new TE insertions have a less

than 10% chance to insert into the genome of *B. distachyon* in a way that will allow them to

evolve neutrally, advocating for a large potential of TEs to create, through their movement, new

phenotypic variation on which selection can act on. PCAs based on TE polymorphisms allowed us

to recover the population structure of *B. distachyon*, implying that demographic history and

hence neutral processes may indeed partially explain the differences in the TE distribution we

observed between genetic clades, as shown in *Arabidopsis thaliana* and *Arabidopsis lyrata* [26,

57], *Drosophila melanogaster* [58], humans [59] and the green anole (*Anolis carolinensis*; [30]).

However, and in line with our simulations, the first two axes of the PCA explain less than 7% of

the variance, indicating that neutrally evolving TEs contribute only mildly to overall TE diversity

in our system.

Because TEs can cause phenotypic variation through new insertions [1-8], it is not

surprising that most new insertions interfere with the function of the genome, especially in a

species with a small genome, such as *B. distachyon* (272 Mb) [37]. The proportion of neutrally

evolving TE polymorphisms is expected to be very small in genes, as insertions in genic regions

are likely to result in loss-of-function [1, 2]. Similarly, TE insertions in close proximity to genes are

expected to be highly disruptive, as regulatory elements such as *cis*-regulatory elements are

predominantly located in the proximity of genes. In *A. thaliana*, for instance, TEs located in the

vicinity of genes (less than 2 kb) globally result in downregulation [60]. Although only specific

families alter gene expression in *B. distachyon* [41], the observed Δ frequency for

retrotransposon polymorphisms in genes and in their 1 kb surroundings matched our

expectations. The fact that TE polymorphisms located more than 5 kb away from genes are also

evolving under purifying selection was more surprising. That said, little is known about the

distance between *cis*-regulatory sequences and genes in *B. distachyon.* In plants, TEs are believed

to affect gene expression in *trans* through the production of small-interfering RNA [61-65].

Hence, the fact that only a small proportion of TEs can accumulate neutrally indicates that, in a

gene-dense genome such as that of *B. distachyon* (42.5% of the genome are genes) [65], TE

insertions in any genomic compartment may result in some *cis-* or *trans*-regulatory effects visible

to selection.

To further ascertain the strength of purifying selection, we used forward simulation and

showed that simulations assuming a moderately weak selection pressure (S = -5 or S = -8) against

TE polymorphisms best fitted our observed data. In theory, no TE polymorphisms under strong

purifying selection should be present in a natural population, as such mutations are expected to

be quickly lost, especially in a predominantly selfing species where most loci are expected to be

homozygous. Therefore, it is not surprising that TE polymorphisms in *B. distachyon* are under

weak to moderate selection, as also shown, for example, for the L1 retrotransposons in humans

[27] or the BS retrotransposon family in *Drosophila melanogaster* [58].

While some of the parameters we chose for our simulations, such as the dominance or

selfing rate, can affect the efficiency of TE purging, it is unlikely that discrepancies in the true and

assumed values for these parameters would have led to drastically different results. For example,

we assumed codominance for all mutations, which might not hold true for each TE

polymorphism. However, because of the high selfing rate observed in *B. distachyon* [38],

heterozygous loci are expected to be rare, and dominance is unlikely to have a strong impact on

our observations. Similarly, with a higher selfing rate, deleterious TE polymorphisms should be

removed more efficiently by purifying selection. To check whether a lower selfing rate could

allow a higher proportion of TE polymorphisms to evolve neutrally, we reran the simulations

assuming fully outcrossing individuals. This also resulted in simulation with weak to moderate

selection strength on TE polymorphisms best fitting the observed data, further strengthening our

results.

While the analyses of positive selection and GEA were based on both DNA-transposons

and retrotransposons, we only used retrotransposons to assess the strength of selection on TE

polymorphisms, as the age-adjusted SFS was developed with the assumption of no back

mutations [48]. Yet, DNA-transposons do not solely transpose through cut and paste mechanisms

as they would otherwise not be so abundant in Eukaryotic genomes. DNA-transposons can also

create extra copies of themselves by transposing during chromosome replication or repair from

a position that has already been replicated, or repaired [66]. We therefore repeated the age-

adjusted SFS analyses using DNA-transposons to evaluate whether DNA-transposons were

affected by similar selective constraints. The folded SFS of DNA-transposons and

retrotransposons display similar shifts toward high proportions of rare alleles and Δ frequency

deviations from 0 in the age-adjusted SFS of DNA-transposons and retrotransposons are

comparable. Hence, we argue that the conclusion drawn for retrotransposons also holds for DNA-

transposons, and that purifying selection affect TEs broadly.

Conclusion

Adaptation to different environmental conditions is a complex process that involves various

mutation types. Here, we show that the vast majority of TE polymorphisms are under purifying

selection in the small genome of *B. distachyon*. Conversely, only a very small proportion of TEs

seem to have contributed to adaptation. The observed lack of neutrally evolving TE

polymorphisms in *B. distachyon* advocates for a large potential of TE polymorphisms to

contribute to the genetic diversity and phenotypic variation on which selection can act and

highlights the need to consider TE polymorphisms during evolutionary studies. Finally, our work

shows that the ability of TEs to cause phenotypic variation does not necessarily translate into

being favored during evolution and adaptation over other mutations with more subtle effects,

such as SNPs.

Materials and methods

Whole-genome resequencing data

In this study, we analyzed a total of 326 publicly available whole-genome sequencing data from *Brachypodium distachyon* accessions sampled around the Mediterranean Basin (Fig. 1A; Additional file 2: Table S3). Our *B. distachyon* dataset consisted of 47 samples published by Gordon et al. [8], 57 samples published by Skalska et al. [67], 65 samples published by Gordon et al. [68], 86 samples published by Stritt et al. [38] and 71 samples published by Minadakis et al. [39], covering all five genetic clades previously described in this species [38, 39]. Each sample was assigned to a genetic clade based on previously published results [39].

Data processing

Raw reads were trimmed using Trimmomatic 0.36 [69] and mapped to the *B. distachyon* reference genome version 3.0 [37] using bowtie2 [70] and yaha [71], and TE polymorphisms were identified using the TEPID pipeline [72] and the recently updated TE annotation by Stritt et al. [73] and Wyler et al. [65]. TE polymorphisms include both TE insertion polymorphisms (TIPs; insertions absent from the reference genome but present in at least one natural accession) and TE absence polymorphisms (TAPs; insertions present in the reference genome but absent from at least one natural accession). The class, superfamily and family of each TE call were assigned based on the TEPID results and the TE annotation from the reference genome. TIPs that were less than 100 base pairs (bp) apart in different samples and assigned to the same TE family were merged.

Single nucleotide polymorphisms (SNPs) were called using GATK v.4.0.2.1 [74] using

HaplotypeCaller [75] following Minadakis et al. [39]. The SNP calls were hard filtered using the

following conditions: QD < 5.0; FS > 20.0; SOR > 3.0; MQ < 50.0; MQRankSum < 2.5; MQRankSum

> -2.5; ReadPosRankSum < 2.0; ReadPosRankSum > -2.0. Because *B. distachyon* displays a high

selfing rate [33], most genetic variants are expected to be homozygous within an individual.

Hence, all TE calls were treated as homozygous, and heterozygous SNP calls were removed from

our dataset to reduce false variant calls. Additionally, all sites with multiallelic TE and SNP calls

were removed. SNPs were classified as synonymous, non-synonymous and of high fitness effect

using SnpEff [76]. SNPs and TE polymorphisms were merged into a single vcf file using custom

scripts provided in github (see section Availability of data and materials).

To estimate the age of each SNP and TE polymorphism, the SNPs and TEs found in the

A_East, A_Italia, B_East and B_West clades were polarized using the C clade, which was identified

as the most ancestral *B. distachyon* clade [38] and used as the outgroup throughout this study.

An estimate for the time of origin of all SNPs and TE polymorphisms was calculated with GEVA, a

nonparametric approach that relies on pairwise differences in identity by descent (IBD) regions

around the focal mutation to estimate the time of origin [77]. GEVA was run separately for each

clade using the genetic map produced by Huo et al. [78] and a mutation rate of 7×10^{-9}

substitutions/generation. The theoretical prediction of the correlation between allele age and

allele frequency of neutrally evolving mutations based on N_e [42] was compared to the observed

correlation between allele age and frequency of synonymous SNPs to check the sanity of the age

estimates.

The observed SNP and TE diversity was first examined using a principal component

analysis (PCA), and correlations between TE diversity and genetic clades were tested with a

mantel test using the ade4 package version 1.7-22 [79] in R version 4.1.2 [80]. The folded site

frequency spectrum (SFS) was computed for TE polymorphisms and SNPs using the minor allele

frequency in R version 4.1.2 [80]. Finally, the map of the geographical distribution of the used

accessions was done in R using the rnaturalearth package 0.3.3 [81].

Analyses of positive selection

Regions of the genome affected by positive selection were identified using the integrated

haplotype score (iHS), a measure of the amount of extended haplotype homozygosity along the

ancestral allele relative to the derived allele for a given polymorphic site [82]. iHS was calculated

using the SNP dataset, and regions displaying longer haplotypes and hence high iHS were

identified in R using the rehh package [83, 84]. The threshold to distinguish between regions of

high iHS and other regions was selected such that less than 5% of the *B. distachyon* genome was

classified as high iHS regions in each clade (Additional file 2: Table S4). Candidate regions under

positive selection were defined as all regions that were found to have high iHS in each clade

separately.

A first ANCOVA was used to model the number of fixed TE polymorphisms in each clade

found in the candidate region under positive selection based on the following genetic features:

total number of TEs, TE superfamily, TE age (split into three categories: young: age < 10,000

generations; intermediate: age between 10,000 generations and 60,000 generations; old: age >

60,000 generations), clade, genomic region (a unique ID for each candidate region under positive

selection) and iHS classification of the regions in each clade (high or average). A second ANCOVA

was used to model the allele frequency of TE polymorphisms found in the candidate region under

positive selection based on the following genetic features: TE superfamily, TE age, clade, genomic

region and iHS classification of the regions in each clade. The TE superfamily was included to

account for different evolutionary behaviors of TEs from different superfamilies. Age accounted

for differences in the fixation rate and frequency distribution between young and old TEs. The

clade was included to account for clade-specific differences such as differences in N_e. Finally, a

unique ID for each candidate region under positive selection was included to account for region-

specific differences such as differences in the recombination rate and GC content. In the end,

regions that were found to have a high iHS in some clades were compared to the same regions

in the other clades. All ANCOVAs were run in R using the car package [85].

The standardized allele frequency of a mutation across populations (X^tX) values [44] were

computed for the combined TE and SNP dataset using Baypass version 2.3 [45, 86]. The X^tX values

were used to identify over- and under differentiated TE polymorphisms between clades. A

pseudo-observed dataset (POD) of 100,000 SNPs was simulated under the demographic model

inferred from the covariance matrix of the SNP dataset. The POD was then used to determine the

97.5% (over-differentiated polymorphisms) and 2.5% (under differentiated polymorphisms)

quantiles.

Genome-environment association analyses

We identified TE polymorphisms significantly associated with environmental factors using

genome-environment association analyses (GEA) following Minadakis et al. [39]. GEAs were run

with GEMMA 0.98.5 [87] using the combined TE and SNP vcf file against the following 32

environmental factors extracted by Minadakis et al. [39]: altitude, aridity from March to June,

aridity from November to February, annual mean temperature, mean temperature of warmest

quarter, mean temperature of coldest quarter, annual precipitation, precipitation of wettest

month, precipitation of driest month, precipitation seasonality, precipitation of wettest quarter,

precipitation of driest quarter, precipitation of warmest quarter, precipitation of coldest quarter,

mean diurnal Range, isothermality, temperature seasonality, maximum temperature of warmest

month, minimum temperature of coldest month, temperature annual range, mean temperature

of wettest quarter, mean temperature of driest quarter, precipitation from March to June,

precipitation from November to February, solar radiation from March to June, solar radiation

from November to February, mean temperature between March and June, mean temperature

between November and February, maximum temperature between March and June, maximum

temperature between November and February, minimum temperature between March and June

and minimum temperature between November and February. We applied a False Discovery Rate

(FDR, [88]) threshold of 5% to control for false positive rates.

Age-adjusted frequency spectra and analyses of purifying selection

Footprints of purifying selection on TE polymorphisms were first evaluated using folded SFS. An

age-adjusted site frequency spectrum (age-adjusted SFS) approach was used to further

investigate the impact of purifying selection on retrotransposons while accounting for

nonconstant transposition rates. Briefly, the age-adjusted SFS is a summary statistic that

describes the difference between the average frequency of TEs at a specific age and the average

frequency of neutral sites of the same age [48]. Therefore, the TE dataset was sorted by age and

split into equally large bins with respect to the number of observations in each age bin. Neutral

sites were then randomly down-sampled to match the number of observations in the TE dataset

and its age distribution [48].

The difference between the average TE and neutral site frequency, or Δ frequency, was

computed for each age bin [48]. This method allows for an unbiased comparison between the

allele frequencies of TEs and neutral sites, and is robust to transposition rate changes and

demographic changes [48]. However, the theory behind this method was developed assuming

no back mutations and is therefore best suited for retrotransposons, as DNA-transposons can

exit an insertion site [48]. We used the synonymous SNPs identified with SnpEff as the neutrally

evolving sites. However, because estimating the population wide frequency of TEs is more

challenging than estimating SNP frequencies, putative biases in frequency estimates need to be

assessed before performing age-adjusted SFS analyses. To do so, the SNP dataset was resampled

so that the SNP dataset used in the age-adjusted SFS had a frequency distribution that matched

the observed TE frequency distribution. The age-adjusted SFS of retrotransposons was

contrasted against the age-adjusted SFS of non-synonymous, as well as against high fitness effect

SNPs. Therefore, 10,000 non-synonymous and high fitness effect SNPs were randomly selected

for each clade to reach approximately the same number of retrotransposon polymorphisms, non-

synonymous and high fitness effect SNPs for final comparisons. To estimate the variation in Δ

frequency estimates, all age-adjusted SFS were computed 100 times. All Wilcoxon tests and

Bonferroni *p* value corrections were done in R version 4.1.2 [80].

Forward simulation

We used SLiM 4.0.1 [89, 90] to run forward simulations and assess the proportion of neutrally

evolving retrotransposons and the average selection strength affecting them. The simulations

were designed to reflect the population size and demographic history of *B. distachyon*. The

simulated genomic fragment was 1 megabase (Mb) long and included neutral (synonymous)

mutations as well as focal mutations that evolved under different selective constraints. The focal

mutations were a mix of neutrally evolving mutations and mutations evolving under a constant

selection pressure. Therefore, the ratio (r) of focal mutations that evolved neutrally was either

0%, 5%, 10%, 25% or 50%. The scaled selection coefficient (S, defined as $N_e s$, with *s* the strength

of selection and N_e the effective population size) affecting the remaining focal mutations was set

at the beginning of the simulation to be either -1, -5, -8, -10, -12, -15, -20 or -50 to cover

effectively neutral (0 > S ≥ -1), intermediate (-1 > S ≥ -10) and strongly deleterious (-10 > S)

selective constraints. The selfing rate was set to 70%, as *B. distachyon* is a highly selfing species

with occasional outcrossing [38, 39]. In addition, a high recombination rate was chosen to

minimize the effects of linked selection in the small genomic fragment simulated. Simulations for

each combination of these two parameters were run 20 times to assess the variation in the

resulting age-adjusted SFS. The shape of the resulting age-adjusted SFS was used to narrow down

the ratio of neutrally evolving TE polymorphisms. Similarly, the age distribution of the mutations

in the oldest bin of the age-adjusted SFS was used to narrow down the strength of selection

affecting TE polymorphisms.

Supentary Information

Supplementary Information

Additional file 1: Supplemental Figures. Figure S1. Principal Component Analyses using TE (left

panel) and SNP (right panel) polymorphisms. **Figure S2.** Principal Component Analyses using

retrotransposon (left panel) and DNA-transposon (right panel) polymorphisms. **Figure S3.**

Observed correlation between age in generations and frequency of synonymous SNPs in the four

derived genetic clades. The red points show the expected age of a neutrally evolving mutation at

a specific frequency based on the predictions of Kimura and Ohta (1973). Panel A: clade A_East;

panel B: clade A_Italia; panel C: clade B_West and panel D: clade B_East. **Figure S4.** Distribution

of the observed TE age scaled by the effective population size (N_e) in the four derived genetic

clades of *B. distachyon*. **Figure S5.** Folded site frequency spectrum of DNA-transposons and

synonymous SNPs in all genetic clades. Panel A: A_East; panel B: A_Italia; panel C: B_West; panel

D: B_East; panel E: C. **Figure S6.** Folded site frequency spectrum of retrotransposons and

synonymous SNPs in all genetic clades. Panel A: A_East; panel B: A_Italia; panel C: B_West; panel

D: B_East; panel E: C. **Figure S7.** Age-adjusted SFS of simulated mutations under negative

selection in the four derived clades. The four columns show the results for the A_East, A_Italia,

B_West and B_East genetic clades, respectively. Each line shows the results for the different

scaled selection coefficients (S). The five colored curves in each plot show the shape of the age-

adjusted SFS with varying ratios of neutrally evolving mutations, and the gray curves show

variation within one standard deviation based on the 20 runs for each simulation. The X axes

show the age bin from the youngest to the oldest, with each age bin including the same number

of observations for each simulation. **Figure S8.** Relative age difference ((mutation age in

simulations - observed mutation age)/maximum absolute age difference) between simulated

data assuming fully outcrossing individuals and observed data in the last bin of the age-adjusted

SFS. A: 25% quantile; B: 50% quantile; C: 75% quantile. **Figure S9.** Age-adjusted SFS of DNA-

transposons (colored), non-synonymous SNPs (light gray) and high effect SNPs (dark gray) in the

four derived clades. The X axes show the age range of the mutations in each bin, and the age

range of each bin was chosen so that each bin represents the same number of DNA-transposons

observations. A: A_East genetic clades; B: A_Italia genetic clades; C: B_West genetic clades; D:

B_East genetic clades. Boxplots are based on 100 estimations of Δ frequency.

Additional file 2: Supplemental Tables. Table S1. List of TEs significantly associated with at least

one environmental factor in the GWAS. **Table S2.** Difference in Δ frequency between the oldest

and second oldest age bin in the different simulations **Table S3.** List of published samples used in

this study. **Table S4.** List of thresholds used and percentage of the genome classified as high iHS

regions in the four derived clades.

Acknowledgments

613   We would like to thank Jeffrey Ross-Ibarra and Mitra Menon as well as Fabrizio Menardo,

614   Michael Thieme, Wenbo Xu, Jigisha, Lars Kaderli and Serafin Schefer for all the discussions and

615   their comments on this project. We thank Emmanuelle Botté for professional editing.

Declarations

618   **Ethics approval and consent to participate**

619   Not applicable.

Consent for publication

622   All authors have read and approved the submission of this manuscript.

Availability of data and materials

625   The datasets supporting the conclusions of this article are publicly available on the European

626   Nucleotide Archive (https://www.ebi.ac.uk/ena/browser/home) and National Center for

627   Biotechnology Information (https://www.ncbi.nlm.nih.gov/sra/), and the archive numbers of

628   the accessions used are listed in the Additional file 1: Table S2. The scripts generated are

629   available on GitHub https://github.com/Roberthorv/TE_in_Brachypodium/tree/main.

Competing interests

632   The authors declare no competing interests.

Funding

This project was funded by the Swiss National Science Foundation (project project

31003A_182785) and the Research Priority Program Evolution in Action from the University of

Zürich. Data analyzed in this paper were generated in collaboration with the Genetic Diversity

Center (GDC), ETH Zürich.

Authors' contributions

A.R. and R.H. conceived the study. R.H. carried out the study, did the TE and SNP calling,

performed the age-adjusted SFS analyses, conducted the $X^t X$ analyses and ran the forward

simulations. N.M. performed the GEA. Y.B. ran the iHS analyses. A.R. acquired the fundings. R.H.

wrote the manuscript and A.R. revised it. All authors discussed the results and commented on

the manuscript.

References

1.   Bhattacharyya MK, Smith AM, Ellis TH, Hedley C, Martin C. The wrinkled-seed character

of pea described by Mendel is caused by a transposon-like insertion in a gene encoding

starch-branching enzyme. Cell. 1990;60(1):115-22.

2.   Hof AE, Campagne P, Rigden DJ, Yung CJ, Lingley J, Quail MA, et al. The industrial melanism

mutation in British peppered moths is a transposable element. Nature. 2016;534:102-

105.

3.   Feschotte C. Transposable elements and the evolution of regulatory networks. Nat. Rev.

Genet. 2008;9:397-405.

4.   Qiu Y, Köhler C. Mobility connects: transposable elements wire new transcriptional

networks by transferring transcription factor binding motifs. Biochem. Soc. Trans.

2020;48(3):1005-1017.

5.   Slotkin RK, Martienssen R. Transposable elements and the epigenetic regulation of the

genome. Nat. Rev. Genet. 2007;8(4):272-85.

6.   Hollister JD, Gaut BS. Epigenetic silencing of transposable elements: a trade-off between

reduced transposition and deleterious effects on neighboring gene expression. Genome

Res. 2009;19(8):1419-1428.

7.   Xiao H, Jiang N, Schaffner E, Stockinger EJ, van der Knaap E. A retrotransposon-mediated

gene duplication underlies morphological variation of tomato fruit. Science.

2008;319(5869):1527-1530.

8.   Gordon SP, Contreras-Moreira B, Woods DP, Des Marais DL, Burgess D, Shu S, et al.

Extensive gene content variation in the Brachypodium distachyon pan-genome correlates

with population structure. Nat. Commun. 2017;8:2184.

9.   Bennetzen JL, Kellogg EA. Do plants have a one-way ticket to genomic obesity? Plant Cell.

1997;9(9):1509-1514.

10.   Vitte C, Panaud O. Formation of solo-LTRs through unequal homologous recombination

counterbalances amplifications of LTR retrotransposons in rice Oryza sativa L. Mol. Biol.

Evol. 2003;20(4):528–540.

11.   Piégu B, Guyot R, Picault N, Roulin A, Sanyal A, Kim H, et al. Doubling genome size without

polyploidization: dynamics of retrotransposition-driven genomic expansions in Oryza

australiensis, a wild relative of rice. Genome Res. 2006;16(10):1262-1269.

12.   Wendel JF, Jackson SA, Meyers BC, Wing RA. Evolution of plant genome architecture.

Genome Biol. 2016;17(37):1-14.

13.   Lisch D. How important are transposons for plant evolution? Nat. Rev. Genet. 2013;14:49-

61.

14.   Negi P, Rai AN, Suprasanna P. Moving through the stressed genome: emerging regulatory

684   roles for transposons in plant stress response. Front. Plant Sci. 2016;7(1448):1-20.

685   15.   Rey O, Danchin E, Mirouze M, Loot C, Blanchet S. Adaptation to Global Change: A

686   Transposable Element-Epigenetics Perspective. Trends Ecol. Evol. 2016;31(7):514-526.

687   16.   Dubin MJ, Mittelsten Scheid O, Becker C. Transposons: a blessing curse. Curr. Opin. Plant

688   Biol. 2018;42:23-29.

689   17.   Quadrana L, Etcheverry M, Gilly A, Caillieux E, Madoui MA, Guy J, et al. Transposition

690   favors the generation of large effect mutations that may facilitate rapid adaption. Nat.

691   Commun. 2019;10(3421):1-10.

692   18.   Uzunović J, Josephs EB, Stinchcombe JR, Wright SI. Transposable elements are important

693   contributors to standing variation in gene expression in Capsella grandiflora. Mol. Biol.

694   Evol. 2019;36(8):1734-1745.

695   19.   Castanera R, Morales-Díaz N, Gupta S, Purugganan M, Casacuberta JM. Transposons are

696   a major contributor to gene expression variability under selection in rice populationse.

697   Life. 2023;12:RP86324.

698   20.   González J, Karasov TL, Messer PW, Petrov DA. Genome-wide patterns of adaptation to

699   temperate environments associated with transposable elements in Drosophila. PLoS

700   Genet. 2010;6(4):e1000905.

701   21.   Studer A, Zhao Q, Ross-Ibarra J, Doebley J. Identification of a functional transposon

702   insertion in the maize domestication gene tb1. Nat. Genet. 2011;43(11):1160-1163.

703   22.   Barrón MG, Fiston-Lavier AS, Petrov DA, González J. Population genomics of transposable

704   elements in Drosophila. Annu. Rev. Genet. 2014;48:561-581.

23.   Rishishwar L, Wang L, Wang J, Yi SV, Lachance J, Jordan IK. Evidence for positive selection

on recent human transposable element insertions. Gene. 2018;675:69-79.

24.   Niu XM, Xu YG, Li ZW, Bian YT, Hou XH, Chen JF, et al. Transposable elements drive rapid

phenotypic variation in Capsella rubella. Proc. Nati. Acad. Sci. USA. 2019;116(14):6908-

6913.

25.   Jiang J, Xu YC, Zhang ZQ, Chen JF, Niu XM, Hou XH, et al. Forces driving transposable

element load variation during Arabidopsis range expansion. bioRxiv. 2022;1-12.

https://www.biorxiv.org/content/10.1101/2022.12.28.522087v1

26.   Lockton S, Ross-Ibarra J, Gaut BS. Demography and weak selection drive patterns of

transposable element diversity in natural populations of Arabidopsis lyrata. Proc. Natl.

Acad. Sci. U S A. 2008;105(37):13965–13970.

27.   Boissinot S, Davis J, Entezam A, Petrov D, Furano AV. Fitness cost of LINE-1 (L1) activity in

humans. Proc. Natl. Acad. Sci. U S A. 2006;103(25):9590-9594.

28.   Blumenstiel JP, Chen X, He M, Bergman CM. An age-of-allele test of neutrality for

transposable element insertions. Genetics. 2014;196(2):523-38.

29.   Rech GE, Bogaerts-Márquez M, Barrón MG, Merenciano M, Villanueva-Cañas JL, Horváth

V, et al. Stress response, behavior, and development are shaped by transposable element-

induced mutations in Drosophila. PLoS Genet. 2019;15(2):e1007900.

30.   Bourgeois Y, Ruggiero RP, Hariyani I, Boissinot S. Disentangling the determinants of

transposable elements dynamics in vertebrate genomes using empirical evidences and

simulations. PLoS Genet. 2020;16(10):e1009082.

31.   Mérel V, Gibert P, Buch I, Rodriguez Rada V, Estoup A, Gautier M, et al. The Worldwide

Invasion of Drosophila suzukii Is Accompanied by a Large Increase of Transposable

Element Load and a Small Number of Putatively Adaptive Insertions. Mol. Biol. Evol.

2021;38(10):4252-4267.

32.   Stritt C, Gordon SP, Wicker T, Vogel JP, Roulin AC. Recent activity in expanding populations

and purifying selection have shaped transposable element landscapes across natural

accessions of the mediterranean grass Brachypodium distachyon. Genome Biol. Evol.

2018;10(1):304-318

33.   Charlesworth B. Transposable elements in natural populations with a mixture of selected

and neutral insertion sites. Genet. Res. 1991;57(2):127-134.

34.   Charlesworth B, Langley CH, Sniegowski PD. Transposable element distributions in

Drosophila. Genetics. 1997;147(4):1993-1995.

35.   Charlesworth B, Charlesworth D. The population dynamics of transposable elements.

Genet. Res. 1983;42(1):1-27.

36.   Charlesworth B. Background selection and patterns of genetic diversity in Drosophila

melanogaster. Genet. Res. 1996;68(2):131-149.

37.   International Brachypodium Initiative. Genome sequencing and analysis of the model

grass Brachypodium distachyon. Nature. 2010;463:763-768.

38.   Stritt C, Gimmi EL, Wyler M, Bakali AH, Skalska A, Hasterok R, et al. Migration without

interbreeding: Evolutionary history of a highly selfing Mediterranean grass inferred from

whole genomes. Mol. Ecol. 2022;31(1):70-85.

39.   Minadakis N, Williams H, Horvath R, Cakovic D, Stritt C, Thieme M, et al. The demographic

history of the wild crop relative Brachypodium distachyon is shaped by distinct past and

present ecological niches. bioRxiv. 2023. https://doi.org/10.1101/2023.06.01.543285

40.   Bourgeois Y, Stritt C, Walser JC, Gordon SP, Vogel JP, Roulin AC. Genome-wide scans of

selection highlight the impact of biotic and abiotic constraints in natural populations of

the model grass Brachypodium distachyon. Plant J. 2018;96(2):438-451.

41.   Wyler M, Stritt C, Walser JC, Baroux C, Roulin AC. Impact of transposable elements on

methylation and gene expression across natural accessions of Brachypodium distachyon.

Genome Biol. Evol. 2020;12(11):1994-2001.

42.   Kimura M, Ohta T. The age of a neutral mutant persisting in a finite population. Genetics.

1973;75(1):199–212.

43.   Voight BF, Kudaravalli S, Wen X, Pritchard JK. A map of recent positive selection in the

human genome. PLoS Biol. 2006;4:e72.

44.   Günther T, Coop G. Robust identification of local adaptation from allele frequencies.

Genetics. 2013;195(1):205-20.

45.   Olazcuaga L, Loiseau A, Parrinello H, Paris M, Fraimout A, Guedot C, et al. A whole-genome

scan for association with invasion success in the fruit fly Drosophila suzukii using contrasts

of allele frequencies corrected for population structure. Mol. Biol. Evol. 2020;37(8):2369-

2385.

46.   Garcia Guerreiro MP. What makes transposable elements move in the Drosophila

genome. Heredity. 2012;108(5):461–468.

47.   Belyayev A. Bursts of transposable elements as an evolutionary driving force. J. Evol. Biol.

769   2014;27(12):2573–2584.

770   48.   Horvath R, Menon M, Stitzer M, Ross-Ibarra J. Controlling for variable transposition rate

771   with an age-adjusted site frequency spectrum. Genome Biol. Evol. 2022;14(2):evac016.

772   49.   Wright SI, Agrawal N, Bureau TE. Effects of recombination rate and gene density on

773   transposable element distributions in Arabidopsis thaliana. Genome Res.

774   2003;13(8):1897-1903.

775   50.   Horvath R, Slotte T. The Role of small RNA-based epigenetic silencing for purifying

776   selection on transposable elements in Capsella grandiflora. Genome Biol. and Evol.

777   2017;9(10):2911-2920.

778   51.   Raissig MT, Woods DP. The wild grass Brachypodium distachyon as a developmental

779   model system. Curr. Top. Dev. Biol. 2022;147:33-71.

780   52.   Hasterok R, Catalan P, Hazen SP, Roulin AC, Vogel JP, Wang K, et al. Brachypodium: 20

781   years as a grass biology model system; the way forward? Trends Plant Sci.

782   2022;27(10):1002-1016.

783   53.   Casacuberta E, González J. The impact of transposable elements in environmental

784   adaptation. Mol. Ecol. 2013;22(6):1503-1517.

785   54.   Bourgeois Y, Boissinot S. On the population dynamics of junk: a review on the population

786   genomics of transposable elements. Genes. 2019;10(419):1-23.

787   55.   Langmüller AM, Nolte V, Dolezal M, Schlötterer C. The genomic distribution of

788   transposable elements is driven by spatially variable purifying selection. Nucleic Acids Res.

789   2023;1-11.

56.   Eyre-Walker A, Woolfit M, Phelps T. The distribution of fitness effects of new deleterious

amino acid mutations in humans. Genetics. 2006;173(2):891-900.

57.   Lockton S, Gaut BS. The evolution of transposable elements in natural populations of self-

fertilizing Arabidopsis thaliana and its outcrossing relative Arabidopsis lyrata. BMC Evol.

Biol. 2010;10(10):1-11.

58.   González J, Macpherson JM, Messer PW, Petrov DA. Inferring the strength of selection in

drosophila under complex demographic models. Mol. Biol. Evol. 2009;26(3):513-526.

59.   Xue AT, Ruggiero RP, Hickerson MJ, Boissinot S. Differential effect of selection against

LINE retrotransposons among vertebrates inferred from whole-genome data and

demographic modeling. Genome Biol. Evol. 2018;10(5):1265-1281.

60.   Wang X, Weigel D, Smith LM. Transposon variants and their effects on gene expression in

Arabidopsis. PLoS Genet. 2013;9(2):e1003255.

61.   McCue AD, Nuthikattu S, Slotkin RK. Genome-wide identification of genes regulated in

trans by transposable element small interfering RNAs. RNA Biol. 2013;10(8):1379-1395.

62.   McCue AD, Nuthikattu S, Reeder SH, Slotkin RK. Gene expression and stress response

mediated by the epigenetic regulation of a transposable element small RNA. PLoS Genet.

2012.8(2):e1002474.

63.   McCue AD, Slotkin RK. Transposable element small RNAs as regulators of gene expression.

Trends Genet. 2012;28(12):616-623.

64.   Cho J. Transposon-derived non-coding RNAs and their function in plants. Front. Plant Sci.

2018;9(600):1-6.

65.   Wyler M, Keller B, Roulin AC. Potential impact of TE-derived sRNA on gene regulation in

812   the   grass   Brachypodium   distachyon.   bioRxiv.   2022.

813   https://www.biorxiv.org/content/10.1101/2022.04.05.487121v1

814   66.   Wicker T, Sabot F, Hua-Van A., Bennetzen JL, Capy P, Chalhoub B et al. A unified

815   classification system for eukaryotic transposable elements. Nat. Rev. Genet. 2007;8 973-

816   982.

817   67.   Skalska A, Stritt C, Wyler M, Williams HW, Vickers M, Han J, et al. Genetic and methylome

818   variation in turkish Brachypodium distachyon accessions differentiate two geographically

819   distinct subpopulations. Int. J. Mol. Sci. 2020;21(18):6700.

820   68.   Gordon SP, Contreras-Moreira B, Levy JJ, Djamei A, Czedik-Eysenberg A, Tartaglio VS, et

821   al. Gradual polyploid genome evolution revealed by pan-genomic analysis of

822   Brachypodium hybridum and its diploid progenitors. Nat. Commun. 2020;11:3670.

823   69.   Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence

824   data. Bioinformatics. 2014;30(15):2114-2120.

825   70.   Langmead B, Salzberg S. Fast gapped-read alignment with Bowtie2. Nat. Methods.

826   2012;9:357-359.

827   71.   Faust GG, Hall IM. YAHA: fast and flexible long-read alignment with optimal breakpoint

828   detection. Bioinformatics. 2012;28(19):2417-2424.

829   72.   Stuart T, Eichten SR, Cahn J, Karpievitch Y, Borevitz JO, Lister R. Population scale mapping

830   of transposable element diversity reveals links to gene regulation and epigenomic

831   variation. eLife. 2016;5:e20777.

832 73. Stritt C, Wyler M, Gimmi EL, Pippel M, Roulin AC. Diversity, dynamics and effects of long

833   terminal repeat retrotransposons in the model grass Brachypodium distachyon. New

834   Phytol. 2020;227(6):1736-1748.

835 74. McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, et al. The genome

836   analysis toolkit: a MapReduce framework for analyzing next- generation DNA sequencing

837   data. Genome Res. 2010;20(9):1297-1303.

838 75. Poplin R, Ruano-Rubio V, DePristo MA, Fennell TJ, Carneiro MO, Van der Auwera GA, et

839   al. Scaling accurate genetic variant discovery to tens of thousands of samples. bioRxiv.

840   2017. https://www.biorxiv.org/content/10.1101/201178v3

841 76. Cingolani P, Platts A, Wang le L, Coon M, Nguyen T, Wang L, et al. A program for

842   annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in

843   the genome of Drosophila melanogaster strain w1118; iso-2; iso-3. Fly. 2012;6(2):80-92.

844 77. Albers PK, McVean G. Dating genomic variants and shared ancestry in population-scale

845   sequencing data. PLoS Biol. 2020;18(1):e3000586.

846 78. Huo N, Garvin DF, You FM, McMahon S, Luo MC, Gu YQ, et al. Comparison of a high-

847   density genetic linkage map to genome features in the model grass Brachypodium

848   distachyon. Theor. Appl. Genet. 2011;123:455-464.

849 79. Dray S, Dufour AB. The ade4 Package: Implementing the duality diagram for ecologists. J.

850   Stat. Softw. 2007;22(4):1-20.

851 80. R Core Team. R: A language and environment for statistical computing. R Foundation for

852   Statistical Computing, Vienna, Austria. 2021. https://www.R-project.org/ Accessed 9

853   August 2023.

854   81.   Massicotte P, South A. rnaturalearth: World map data from natural earth. R package

855   version 0.3.3.9000. 2023. https://docs.ropensci.org/rnaturalearth/. Accessed 9 August

856   2023.

857   82.   Sabeti PC, Reich DE, Higgins JM, Levine HZP, Richter DJ, Schaffner SF, et al. Detecting

858   recent positive selection in the human genome from haplotype structure. Nature.

859   2002;419:832-837.

860   83.   Gautier M, Klassmann A, Vitalis R. rehh 2.0: a reimplementation of the R package rehh to

861   detect positive selection from haplotype structure. Mol. Ecol. Resour. 2017;17(1):78-90.

862   84.   Gautier M, Vitalis R. rehh: An R package to detect footprints of selection in genome-wide

863   SNP data from haplotype structure. Bioinformatics. 2012;28(8):1176-1177.

864   85.   Fox J, Weisberg S. An R companion to applied regression. Third Edition. Sage. Thousand

865   Oaks CA. 2019. https://socialsciences.mcmaster.ca/jfox/Books/Companion/. Accessed 9

866   August 2023.

867   86.   Gautier M. Genome-wide scan for adaptive divergence and association with population-

868   specific covariates. Genetics. 2015;201(4):1555-1579.

869   87.   Zhou X, Stephens M. Genome-wide efficient mixed-model analysis for association studies.

870   Nat. Genet. 2012;44(7):821-824.

871   88.   Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful

872   approach to multiple testing. J. R. Stat. Soc. 1995;57(1):289-300.

873   89.   Haller BC, Messer PW. Evolutionary modeling in SLiM 3 for beginners. Mol. Biol. Evol.

874   2019;36(5):1101–1109.

875   90.   Haller BC, Messer PW. SLiM 3: forward genetic simulations beyond the Wright-Fisher

876   smodel. Mol. Biol. Evol. 2019;36(3):632–637.