Allelic variants confer Arabidopsis adaptation to small

regional environmental differences

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Running title: Genetics of local Arabidopsis adaption

Abstract

Natural populations of *Arabidopsis thaliana* provide powerful systems to study adaptation of wild plant species. Previous research has predominantly focused on global populations or accessions collected from regions with diverse climates. However, little is known about the genetics underlying adaptation in regions with mild environmental clines. We have examined a diversity panel consisting of 192 *A. thaliana* accessions collected from the Netherlands, a region with limited climatic variation. Despite the relatively uniform climate, we identified compelling evidence of local adaptation within this population. Notably, semidwarf accessions, due to mutation of the *GIBBERELLIC ACID REQUIRING 5* (*GA5*) gene, occur at a relatively high frequency near the coast and these displayed enhanced tolerance to high wind velocities. Additionally, we evaluated the performance of the population under iron deficiency conditions and found that allelic variation in the *FE SUPEROXIDE DISMUTASE 3* (*FSD3*) gene affects tolerance to low iron levels. Moreover, we explored patterns of local adaptation to environmental clines in temperature and precipitation, observing that allelic variation at *LA RELATED PROTEIN 1C* (*LARP1c*) likely affects drought tolerance. Not only is the genetic variation observed in a diversity panel of *A. thaliana* collected in a region with mild environmental clines comparable to that in collections sampled over larger geographic ranges, it is also sufficiently rich to elucidate the genetic and environmental factors underlying natural plant adaptation.

Introduction

Adaptation is defined as the process through which a population attains higher fitness in its native environment than non-native populations sampled from different sites

(Kawecki and Ebert, 2004). Unraveling the genomic and physiological basis of adaptation

in plants is a central question in modern plant biology. From an evolutionary

perspective, linking plant genotypes to adaptive traits helps to clarify how

environmental gradients drive natural selection of both standing and novel variation,

which ultimately may result in speciation (Sobel et al., 2010). It also enables us to model

whether natural populations are able to adapt to future environmental conditions (Bay

et al., 2017; Exposito-Alonso et al., 2019) and provides insights on processes or even

genes to focus on when breeding crops for enhanced abiotic stress tolerance (Huang

and Han, 2014).

The model plant species *Arabidopsis thaliana* provides an excellent system to

study natural plant adaptation (Weigel and Nordborg, 2015). Recent studies of this

species have shifted from studying the global HapMap population (Li et al., 2010),

traditionally used for genome-wide association studies (GWAS), to populations sampled

from regions containing a range of contrasting climates, including Sweden (Long et al.,

2013), the Iberian Peninsula (Tabas-Madrid et al., 2018), and the south-west of France

(Frachon et al., 2018), as these local populations are expected to minimize genetic

heterogeneity and thus provide more statistical power for detecting quantitative trait

loci (QTLs) (Brachi et al., 2013; Korte and Farlow, 2013). While studies on such regional

populations have successfully identified several adaptive loci (Frachon et al., 2018;

Tabas-Madrid et al., 2018; Arteaga et al., 2022), we know comparatively little about the

loci driving adaptation in regions having a milder diversity in environmental conditions.

The Netherlands, a region which has remained largely under-sampled so far,

provides an excellent opportunity to address this issue. This region covers an area

comparable in size to the south-west of France, but much smaller than Sweden and the

Iberian Peninsula. Compared to the other three regions, the Netherlands has only mild

climatic clines (https://www.knmi.nl/klimaat-viewer), but past work indicated that local

adaptation of *A. thaliana* may still be expected (Barboza et al., 2013). The Netherlands

contains a moderate frequency of accessions with short inflorescences, mediated by the

same loss-of-function allele of the *GA5* gene that managed to spread over more than

100 kilometres in the western part of the country (Barboza et al., 2013). There is reason

to believe that more such signatures of local adaptation can be found, for instance

based on the small differences in average temperature and annual precipitation, or the

soil type on which *A. thaliana* grows in the Netherlands (De Vries et al., 2003). While it

can be found on the clay and peat regions in the west and north of the country, it

mainly grows on sandy soils, which largely occur in the east and south, and in the

coastal dunes. It is often found in roadsides and gardens in which substantial movement

of soils and seeds from other regions has occurred in the past.

To comprehensively address the contribution of genetic variation to plant

adaptation in the Netherlands, we generated the Dutch *Arabidopsis thaliana* Map

(DartMap) panel, a collection of 192 *A. thaliana* accessions sampled from different sites

in the country. We show that this panel contains a surprisingly high level of genetic

diversity and provide evidence of local adaptation to mild climatic clines. Moreover, we

present strong evidence that variation at two loci is involved in mediating adaptation to

wind tolerance and iron deficiency. Our work demonstrates that plant populations

sampled in small geographic regions with a low diversity in environmental conditions

can contain a level of genetic variation that is sufficiently rich to facilitate local

adaptation.

Results

The degree of *A. thaliana* genetic diversity in the Netherlands is typical of that of most European regions

The global *A. thaliana* collection of the 1,001 Genomes (1001G) Project (The 1001 Genomes Consortium, 2016) includes only 11 lines from the Netherlands, which likely represents only a small portion of the region's genetic variation due to limited dispersal of minor alleles (Tabas-Madrid et al., 2018). Therefore, we assembled and analysed the DartMap panel and used it together with the Dutch 1001G accessions to characterize the overall degree of genetic diversity of *A. thaliana* in the Netherlands. In total, we identified 2,712,612 SNPs and 353,974 indels in the DartMap panel and the Dutch 1001G accessions (collectively referred to as DartMap + 1001G panel for brevity) relative to the *A. thaliana* Col-0 nuclear genome reference sequence. Moreover, we detected 487 SNPs and 209 indels in the chloroplast sequence, and 137 SNPs and 15 indels in the mitochondrial sequence.

Furthermore, we examined copy number variation in the DartMap panel, excluding the Dutch 1001G accessions because they were sequenced on a different sequencing platform. We detected 29,155 copy number variants (CNVs) relative to the Col-0 reference genome. Most of these are deletions, less than 500 bp (Figures S1a and S1b), which is in line with the distribution of structural variants found in an earlier study on the full 1001G collection (Göktay et al., 2021). We found that 448 genes overlap with CNVs predicted to disrupt gene function and present at moderate frequencies within the DartMap panel (Table S1). We only considered deletions here, as only those were called with high precision (Figure S2). The 448 genes are significantly enriched for genes involved in disease resistance ($P = 4.39 \times 10^{-6}$).

To contextualize the degree of genetic diversity of the DartMap + 1001G panel, we

compared it to similarly sized collections from Sweden (Long et al., 2013) and the

Iberian peninsula (Tabas-Madrid et al., 2018), using the same SNP and indel calling

approach. Considering only SNPs and indels that are polymorphic in the DartMap +

1001G panel (i.e. at least one accession contains a reference allele), the Swedish

collection contains a similar number of variants, while the Iberian one is more diverse

(Table 1). This disparity can be attributed to admixture of "relict" accessions in the

Iberian collection, which originated from *A. thaliana* populations in Europe before the

last ice age, and the genetically distinct "non-relict" group that repopulated Europe

thereafter (The 1001 Genomes Consortium, 2016).

Table 1: **The number of polymorphic bi-allelic SNPs and indels ("Variants") found in**

regional *A. thaliana* populations.

Population	Samples	Variants	Reference
Dutch (DartMap + 1001G)	203	3,057,293	This study, (The 1001 Genomes Consortium, 2016)
Sweden	243	3,084,964	(Long et al., 2013)
Iberian Peninsula	190	4,482,982	(Tabas-Madrid et al., 2018)

Most SNPs and indels detected in the DartMap + 1001G panel are shared among

multiple accessions and the distribution of their minor allele frequencies is similar to

that of the Swedish collection (Figure 1a). In contrast, the Iberian collection contains a

relatively larger fraction of rare variants (Figure 1a), possibly because the mountain

systems in this region strongly limit the spread of minor alleles (Tabas-Madrid et al.,

2018).

To assess the genetic diversity of the DartMap + 1001G panel, we calculated

pairwise genetic distances between individual accessions. Most pairs of accessions differ

at approximately 320,000-340,000 variant sites, similar to the Swedish collection and

most pairs of the Iberian collection (Figure 1b). However, the Iberian collection shows a

right-skewed distribution of pairwise genetic distances, indicating the presence of

accessions that are genetically more distant from each other compared to the majority

(Figure 1b). This tail in the distribution aligns with previous findings of genetic distance

between Iberian accessions and attributed to the presence of the relict accessions in

this region (The 1001 Genomes Consortium, 2016). Taken together, our analyses

indicate that genomic variation of *A. thaliana* in the Netherlands predominantly

originated from the post-glacial expansion event from which most *A. thaliana*

accessions in Europe descended (Lee et al., 2017).

Figure 1: **Comparing the genetic diversity of three regional populations.** (a) Minor allele

frequency of bi-allelic SNPs and indels. (b) Genome-wide similarity between pairs of

samples (bars are overlapping).

Genetic diversity of the *A. thaliana* in the Netherlands is shaped by ancient and contemporary forces

We explored the demographic and evolutionary history of the DartMap + 1001G panel using population structure analyses based on the genetic distances between individual accessions, which have proven useful for this purpose in previous studies of *A. thaliana* (The 1001 Genomes Consortium, 2016; Tabas-Madrid et al., 2018). We computed population structure of the DartMap + 1001G panel through hierarchical clustering based on pairwise organellar (Figure 2a, Table S2) and nuclear (Figure 2b, Table S2) genome-wide similarity, including all accessions of the 1001G collection sampled in Belgium, France, Germany, and the United Kingdom to evaluate relatedness between Dutch *A. thaliana* accessions and those of nearby countries. The dendrograms based on chloroplast and mitochondrial variation consist of three nearly equidistant groups (Figure S3) with identical spatial distributions across the Netherlands (Figure 2a), as expected for organellar sequences that are both predominantly maternally inherited.   A previous study showed that European accessions contain several highly diverged chloroplast haplogroups of which the estimated time of divergence considerably predates the post-glacial expansion event (Hsu et al., 2019). A principal component analysis, based on chloroplast variation between individual accessions of the DartMap + 1001G panel and of a global collection of *A. thaliana*, separates Dutch accessions in three different groups (Figure 2c) that correspond to different subsets of chloroplast haplogroups identified in the same study (Figure 2d). This implies that the distinct organellar groups of the DartMap + 1001G panel represent ancient variation, while the nuclear groups reflect more recent genomic differentiation, explaining the limited overlap in spatial distribution between the two (Figure 2a and b).

The spatial distribution of the nuclear groups provides insights into the factors

that shaped genomic variation in the DartMap + 1001G panel following the postglacial

expansion event. Based on nuclear variation, the panel can be divided into two main

groups, predominantly separated by the Rhine and Meuse river branches (group 1 to the

north and group 2 to the south) (Figure 2b). The rivers likely acted as natural dispersion

barriers (Pico et al., 2008), restricting genetic flow between the two groups. It is worth

noting that *A. thaliana* is generally not found on clay soil accompanying the rivers, except

on sandy river dunes and residential areas where it has been introduced through human-

induced transport of soil from other regions. Group 1 can be further divided into three

subgroups (Figure 2b), of which two contain accessions that are closely related to each

other (subgroups 1a and 1b) and less related to the rest of the population (Figure S4).

Subgroup 1a contains eight accessions of the Dartmap panel with short inflorescences,

referred to as "semidwarfs", and one accession (Tha-1) of the 1001G panel, previously

described as a semidwarf (Barboza et al., 2013). The accessions from subgroup 1a are

mainly found in the western part of the Netherlands, close to the North Sea coast (Figure

2b). Subgroup 1b is predominantly found on the Dutch islands in the north, implying that

these islands were colonized by a limited number of founder accessions, mostly from

group 1. Group 2 consists of accessions that are genetically similar to accessions from

Germany (subgroup 2a), respectively France and the United Kingdom (subgroup 2b)

(Figure S5), suggesting that these are part of a large genetic group spread over

Northwestern Europe.

Besides large clusters, we identified 27 pairs of accessions that are near duplicates

in terms of nuclear similarity (Table S3), with differences of fewer than 50,000 variant

sites between them. This level of similarity is significantly lower than the median pairwise

similarity of 354,388 variant sites (Figure 1a). Strikingly, they include 6 out of 8 of the

semidwarf accessions of subgroup 1a, the furthest of which are separated by more than

100 km (Table S3) (Flood et al., 2016a).

Figure 2: **Population structure of *A. thaliana* in the Netherlands.** (a-b) Geographical

location of Dutch accessions clustered based on organellar (a) and nuclear (b) genome-

wide similarity. Colours are used to distinguish accessions of different clusters. The main

rivers of the Netherlands are depicted by blue lines. (c) Principal component analysis of

chloroplast variants in Dutch accessions and the global collection of *A. thaliana* analysed

in a previous study (Hsu et al., 2019). (d) Samples in (c) coloured according to the

chloroplast haplogroups reported in a previous study (Hsu et al., 2019).

Semidwarf *ga5* mutants display tolerance to windy conditions

The widespread geographical distribution of highly identical genotypes that all have the same semidwarf phenotype suggests there may be a selective advantage to this trait, similar to the near-identical and widespread atrazine-resistant genotypes distributed along the United Kingdom railway system, which were identified previously (Flood et al., 2016a).

An earlier study already indicated a higher frequency of semidwarfs in the Netherlands compared to the global estimated frequency of at least 1% (Barboza et al., 2013). Next to the eight semidwarfs found in subgroup 1a, we identified a ninth semidwarf accession, Snh-1, in subgroup 2b. Almost all examined semidwarf accessions found in the field are loss-of-function mutants at the *GA5* locus, which encodes the *GIBBERELLIN 20-OXIDASE 1* (*GA20OX1*) gene (Barboza et al., 2013). Further investigation of the genomic sequences of *GA5* revealed that the eight semidwarf accessions in subgroup 1a, that are genotypically nearly identical, share a previously described *ga5* splice site mutation specific to Dutch semidwarfs (Barboza et al., 2013), while Snh-1 contains a 52-bp deletion in the last exon of *GA5*.

The appearance of a relatively high frequency of natural *ga5* mutants suggests that semidwarfism provides an adaptive advantage. Dwarfism in plants is a common adaptation to high altitude (Billings and Mooney, 1968; Grace, 1988; Luo et al., 2015), although it remains unclear which aspect of high altitude causes this. The majority of the Dutch semidwarfs are collected within 15 km of the coast, often only a few meters above sea level, so there is no association with high altitude. However, the locations where the semidwarfs were collected share an important characteristic providing a potential selective force, namely the presence of high wind speeds (Figure 3a). High

wind speeds are also a characteristic of high-altitude environments. We therefore

investigated whether the presence of high wind speeds could create a selective

environment favouring semidwarf *A. thaliana* over normal-sized plants.

We grew six Dutch semidwarf accessions and five unrelated tall accessions (Table

S4), in a climate-controlled growth chamber equipped to provide controlled wind

speeds typically experienced in coastal areas of the Netherlands in March/April.

Moreover, we included Col-0, L*er* and the *ga5* loss-of-function mutants in Col-0 and L*er*

backgrounds for a direct comparison of near isogenic lines differing only at the *GA5*

locus. Despite our preselection for similar flowering time (Table S4), based on previous

data collected on greenhouse-grown plants, the semidwarfs flowered significantly later

in the controlled growth room conditions than the taller accessions (Figure 3b). This

likely had little effect on our final results though, as all accessions were exposed to

windy conditions for the same time and all plants started to flower before the end of

the experiment. The tall accessions were strongly affected by the wind, as evidenced by

a significant reduction in stem length (approximately 30% on average) and fresh weight

(approximately 36% on average) (Figure 3c and 3d). In contrast, the semidwarfs also

showed a significant reduction in stem length, but the average reduction was only

around 10%. No significant decrease in fresh weight was observed (Figure 3d). Similar

effects were observed for Col-0 and L*er*, where the wind treatment significantly

decreased their stem length and fresh weight, whereas the respective *ga5* loss-of-

function mutants remained unaffected (Figure 3c and d). Collectively, these results

strongly suggest that the semidwarf phenotype caused by the *ga5* mutation provides a

strong advantage with respect to plant biomass production, when growing in conditions

of high wind speeds.

Figure 3: **Phenotypic response to windy conditions.** (a) Average annual wind speed

(between 1981-2010) in the Netherlands as measured at ground level. Green circles

indicate semidwarf accessions that share the same splice site mutation, black triangles

indicate all other DartMap accessions. Figure adapted from

https://www.knmi.nl/klimaat-viewer/kaarten/wind/gemiddelde-

windsnelheid/jaar/Periode_1981-2010. (b-d) Plant responses to absence or presence of

windy conditions (grey and orange respectively) for (b) flowering time, (c) inflorescence

length one week after the first flower opened, (d) fresh weight at flowering *$P < 0.05$;

**$P < 0.01$; ***$P < 0.001$, two-tailed Student's *t*-test. NL – Dutch accessions; *GA5* – wild-

type allele, *ga5* – mutant allele.

Flowering time variants display a latitudinal cline

The transition from vegetative to reproductive stage is a key life history trait in *A.*

thaliana, influenced by seasonal cues like day length and temperature (Andrés and

Coupland, 2012). Previous GWAS of flowering time yielded few significant associations,

mainly due to allelic heterogeneity (Atwell et al., 2010; Tabas-Madrid et al., 2018; Zhang

and Jiménez-Gómez, 2020). We expect this should be less of a problem in a regional

population with more closely related individuals. Therefore, we performed GWA

analysis of flowering time, with and without vernalization, using the DartMap panel. We

discovered a highly significant association between flowering time and a 16-bp indel

near the end of the first exon of the *FRIGIDA* (*FRI*) gene (position 269960 on

chromosome 4 of Col-0), irrespective of vernalization treatment (Figure 4a). This indel is

known to cause a premature stop codon in the *FRI* coding sequence, leading to a non-

functional *FRI* allele (Koornneef et al., 1994; Johanson et al., 2000; Andrés and Coupland,

2012). *FRI* is a key regulator of flowering time in *A. thaliana* and loss-of-function alleles

are widespread in nature (Zhang and Jiménez-Gómez, 2020). As the variant we picked

up by GWA analysis is unlikely to be the only loss-of-function mutation to occur in the

Netherlands, we examined all polymorphisms in the *FRI* coding sequences (CDS) in the

DartMap and found a total of 41 *FRI* alleles. When compared to the *FRI-H51* allele, that

encodes a fully functional protein and is regarded as the ancestral *FRI* allele based on

sequence comparisons with *A. lyrata* (Le Corre et al., 2002; Zhang and Jiménez-Gómez,

2020), another 11 loss-of-function *fri* alleles are identified across 88 accessions. The

accessions with a loss-of-function allele are on average early flowering (Figure 4b).

Although the Netherlands has a small latitudinal range, we examined if there is a

latitudinal cline in the diversity panel regarding flowering time, consistent with previous

findings in Northern European and Mediterranean accessions (Stinchcombe et al., 2004).

While we did not observe such a cline when considering the entire population, when

excluding the accessions carrying the *fri* loss-of-function alleles, we indeed found such a

latitudinal cline (Figure 4c). This means other loci, next to *FRI*, are involved in controlling

flowering time in the DartMap collection. These analyses collectively demonstrate the

suitability of the DartMap panel for conducting GWA analyses on phenotypic variation

relevant to local adaptation.

Figure 4: **GWA analysis of flowering time in the DartMap panel upon vernalization.** (a)

Manhattan plot for days until flowering after a six-week vernalization treatment,

identifying one highly associated polymorphism located in the *FRIGIDA* gene (arrow).

The red line corresponds to a permutation-based threshold level of significance at $P <$

0.05. (b) Flowering time distribution of three *FRIGIDA* (*FRI*) variants in the DartMap

panel. Accessions with a functional *FRI* allele (*FRI Functional*), accessions with the Col-0

variant (Col-*fri*) that corresponds to a 16-bp deletion causing a premature stop codon in

the *FRIGIDA* coding sequence and accessions with a predicted functional knock-out

allele of *FRI* (*fri* Predicted KO). (c) Average days until flowering for DartMap accessions

relative to their latitudinal site of origin with linear fit for accessions with a predicted

loss-of-function *fri* allele (red, R^2 = 0.016, *P* = 0.128) or a functional *FRI* allele (blue, R^2 =

0.082, *P* = 0.002).

Local adaptation to the Dutch climate

The Dutch climate is characterized as a temperate maritime climate, which is relatively

uniform across the Netherlands. However, there are mild and gradual clines in

environmental variables, with slightly higher temperatures and less precipitation in

inland areas compared to coastal areas. One advantage of having a densely and

uniformly sampled population is the potential to detect whether local adaptation to

these gradual environmental clines has influenced genetic diversity. To test this

hypothesis, we conducted a genome-environment association (GEA) analysis on 19

quantitative climatic variables related to temperature and precipitation (Table S5 and

Figure 5a as example). We identified a few QTLs for these variables, but the most

notable QTL is located on Chr. 4 (Figure 5b). We named it *qDPT* (QTL for the Dutch

Precipitation and Temperature climatic variables). This QTL is associated with seven

climatic variables reflecting aspects of temperature and precipitation (Table S5).

Further examination of *qDPT* revealed that it appears to consist of two separate

QTLs, approximately 230 kilobase pairs apart (Figure S6a). The SNPs that are found most

frequently with the highest association in each QTL are SNP $4^{16770900}$ and SNP $4^{17000635}$

with minor allele frequencies (MAF) of 14.2% and 17.5%, respectively. To determine

whether the -log(P) peaks represent distinct QTLs or a single one, we calculated pairwise

r^2 values between SNP $4^{16770900}$ and all other biallelic SNPs in the selected genomic

region (16,700,000–17,200,000 bp). This revealed only weak evidence of linkage

disequilibrium (LD) (pairwise $r^2 = 0.52$) between the two SNPs most strongly associated

with the trait (Figure S6b). Hence our conclusion these are two separate QTLs. For

convenience, we will refer to them as '*qDPT-A*' and '*qDPT-B*', with their corresponding

SNPs labelled as A/a for SNP $4^{16770900}$ and B/b for SNP $4^{17000635}$.

The GEA analyses provide insights into whether specific alleles are correlated

with particular climatic variables. A strong association at a specific locus can indicate

that allelic variation at that locus is important for local adaptation (Ferrero-Serrano and

Assmann, 2019). However, such a strong correlation could also arise by accident, if the

climatic variable happens to align with the population structure. We confirm that this is

not a case of such a potential statistical artifact. First of all, kinship correction is used to

perform the GEA analyses, and this should drastically reduce the chance of finding loci

that are strongly associated due to the population structure. Secondly, we observe that

the geographical distribution of the accessions contributing to the population structure

appear to be mostly affected by the river systems (Figure 2b), which can be regarded as

a South – North cline. The climatic variables that map at *qDPT* follow a mostly East –

West cline. Moreover, the minor allele of *qDPT-A* is found in five out of six nuclear

subclusters (the common allele is found in all subclusters), and for *qDPT-B* both alleles

are found all subclusters. In conclusion, identification of these QTLs is not an artifact

due to the general population structure and thus likely to represent a locus associated

with plant adaptation to the environment.

It is not immediately apparent to which specific climatic variable(s) this locus

contributes. The climate in the Netherlands is primarily influenced by the North Sea,

which affects various environmental factors such as temperature, wind speed and

precipitation patterns and variability. Therefore, it is not surprising that the same peak

is consistently observed for several climatic variables, given the high correlation among

the variables associated with *qDPT*. Consequently, unveiling the particular climatic

variable or combination of variables to which the adaptation is associated with this

specific locus may be challenging. However, we noted that the climatic variables

associated to *qDPT* are all connected to seasonality, while annual means in precipitation

and temperature are not associated. This suggests that the adaptation is related to

conditions that prevail during certain parts of the year, such as one or two seasons, or

to adaptations that enable the plant to effectively tolerate greater variability. To explore

which climatic factor may be involved in this regard, we tested if allelic variation at *qDPT*

resulted in a differential response to a gradual decrease in water availability, which is

likely to be seasonal.

As *qDPT-A* and *qDPT-B* are in close proximity, we selected the best possible set of

accessions distinguishing best the combinations of alleles for these QTLs aiming to

disentangle the individual contributions of the QTLs. Given the available genotypes of

accessions in the DartMap population, this results in 13 accessions with 'AB', 7 with 'Ab',

1 with 'aB' and 7 with 'ab' (Table S6). Initially, all accessions were grown under well-

watered conditions. Subsequently, after two weeks, half of the replicates for each

accession were no longer watered for eleven days. We refer to this as the drought

treatment. We regularly measured the projected leaf area (PLA) during this period in the

control and the drought treatment. Due to an uneven distribution of accessions across

the genotypic combinations for these QTLs, we compared accessions with 'Ab'

genotypes against those with 'ab' genotypes (7 accessions per group) to assess the

effect of *qDPT-A*. For *qDPT-B*, all 28 accessions (14 accessions per *qDPT-B/b* allele) were

included in the analysis. Our results indicated a significant effect of *qDPT-B* on the PLA

response to the drought treatment (two-way ANOVA with *qDPT-B/b* allele and days

after last watering as variables; P = 0.003) (Figure 5c and 5d). However, we did not

observe a significant effect for *qDPT-A*. Subsequently, we examined the distribution of

qDPT-B alleles across the Netherlands and the 1001G panel. We found that SNP $4^{17000635}$

(*qDPT-B*) has a much higher MAF in the Netherlands (MAF = 17.5%) (Figure 5e)

compared to the 1001G panel (MAF = 3.1%) (Figure 5f).

We propose that the region encompassing the genomic locus with a LOD > 6,

which is consistently observed across different mappings, is the most likely location for

the gene(s), allelic variation of which causes the QTL. This region comprises 13 genes

spanning from At4g35820 to At4g35940 (Table S7). Within this region, there are a few

genetic variants that are in linkage disequilibrium (r^2 > 0.4) with SNP $4^{17000635}$ and that

potentially impact the encoded protein sequence of the gene they reside in.

Noteworthy are two non-synonymous variants in *OSCA4.1* (At4g35870), also known as

EARLY-RESPONSIVE TO DEHYDRATION STRESS 4 (*ERD4*). The first variant constitutes a

change from methionine (neutral-polar) to isoleucine (hydrophobic) at the 10th position,

while the second variant changes an isoleucine to leucine (both hydrophobic) at the

641st position. These substitutions constitute relatively minor sequence differences that

are unlikely to affect the functionality of the protein encoded by the *ERD4* gene. We

tested T-DNA insertion lines for 11 of the 13 candidate genes in response to control and

drought treatments and measured F_v/F_m as a proxy of plant stress (Maxwell and

Johnson, 2000) (Figure 5g). For the remaining two genes, At4g35930 and At4g35940, no

T-DNA insertion mutant lines were available. The mutant of *LARP1c* (At4g35890) has

significantly lower F_v/F_m values in the drought treatment compared to Col-0, while it

does not differ from Col-0 in the control treatment (Figure 5g). Furthermore, the mutant

PLA is slightly higher, though not significantly different from Col-0 PLA under the control

treatment, but it is significantly impaired under the drought treatment (Figure 5h).

These results pinpoint *LARP1c* as the most prominent candidate gene to underlie the

difference in response to drought.

Figure 5: **Genome-environment association (GEA) analysis on climatic variables related**

to precipitation and temperature. (a) Geographical distribution of precipitation

seasonality (coefficient of variation) expressed in Seasonality Index (SI) units for the

DartMap. SI values <0.19 are defined as 'precipitation throughout the year', whereas

values between 0.20 and 0.39 are defined as 'precipitation throughout the year, but

with a definite wetter season'. (b) Manhattan plot for the GEA related to precipitation

seasonality (coefficient of variation). (c) Response to the drought treatment for

accessions differing at their *qDPT-B* allele. Boxplots represent the normalised projected

leaf area (PLA) calculated per accession by dividing the PLA in the withholding watering

treatment over the PLA in the control treatment for a set of 14 randomly chosen

accessions per allele (depicted as an adenine (A) or guanine (G) nucleotide at SNP

$4^{17000635}$). (d) Pictures of six representative accessions per *qDPT-B* allele in the control

and drought treatment, taken 11 days after the start of the treatment. (e) Geographical

distribution of the two alleles ('A' and 'G') across the Netherlands and (f) Europe. (g))

F_v/F_m as a proxy of plant stress of Col-0 and candidate gene T-DNA insertion lines. (f)

The response of PLA of Col-0 and the *larp1c* mutant in the control and drought

treatment.

Natural allelic variation for *FSD3* affects iron deficiency tolerance

Although the Netherlands is relatively uniform in climate, the soil type composition

differs substantially across the country, with clay, peat and sand forming the major

types (Hartemink and Sonneveld, 2013). This could be an important driver for local

adaptation in the Netherlands, as nutrient composition and especially nutrient

availability (Veer, 2006), differs between soil types. Iron (Fe) in particular is known to

vary in availability and shortage of Fe will severely limit plant growth (Hell and Stephan,

2003). We determined the light-use efficiency of photosystem II (PSII) electron transport

(Φ_{PSII}) of the DartMap diversity panel grown under control and Fe-deficient conditions,

through chlorophyll fluorescence measurements. Low Φ_{PSII} can serve as an early

indicator of Fe deficiency (Terry, 1980), as Fe is an essential component of iron-sulphur,

and heme proteins that play pivotal roles in photosynthesis and respiration (Hänsch and

Mendel, 2009).

The low Fe supply leads to only mild visible symptoms, such as reduced plant

growth and mild leaf chlorosis, but causes extensive variation for Φ_{PSII}, much more than

in the control condition (Figure 6a). GWA mapping identified a single associated region,

on chromosome 5, with the most significantly associated SNP located at position

7,856,132 (Figure 6b). The same locus, and the same SNP, is also found when the

average Φ_{PSII} values in the Fe-deficient condition, and the ratio between Φ_{PSII} in Fe

deficiency versus control, are used as traits for GWA mapping (Figure S7). To identify

candidate genes for the causal allelic variant, we limited the associated region to 22 kb,

based on the genome positions of SNPs that are in linkage disequilibrium (LD) (at $r^2 >$

0.55) with the most significantly associated SNP. This region is predicted to contain 10

genes (Figure 6c).

We identified three common haplotypes for this genomic region in the DartMap

panel. Accessions that share the haplotype with the Col-0 variant associated with

SNP5^{7856132} are referred to as haplotype 1, whereas accessions containing the non-

reference variant are further distinguished into two haplotypes that are referred to as

haplotypes 2 and 3 (Table S8). Haplotype 1 associates with higher Φ_{PSII} in Fe-deficient

conditions relative to the non-reference haplotypes, which do not significantly differ in

Φ_{PSII} (Figure 6d). The higher Φ_{PSII} of accessions with haplotype 1 correlates with a

significantly higher projected leaf area in Fe-deficient conditions than haplotypes 2 and

3 (Figure 6e), while these do not differ significantly for projected leaf area.

In total, 40 sequence variants are in LD with SNP5^{7856132} at a cut-off of $r^2 > 0.55$,

of which 27 are intergenic, 7 intronic and 6 exonic (Table S9). Only two of the exonic

variants encode a non-synonymous amino acid substitution, but in both cases the

substitutions are from a leucine to an isoleucine or vice versa. These are very similar

amino acids, and neither of these substitutions, one in *RIBF1* (At5g23330), the other in

DUF789 (At5g23380), appear to substantially change the function of these proteins, as

predicted by InterPro (Mitchell et al., 2019).

We thus hypothesized that non-protein-coding variants that affect either levels

or timing of gene expression should be causing the observed QTL. We measured

expression of each candidate gene by means of RT-qPCR in both control and Fe-

deficient conditions in shoots of six accessions of each haplotype (Table S9). Two known

marker genes for Fe deficiency in leaves, *FE SUPEROXIDE DISMUTASE 1* (*FSD1)* and

FERRIC REDUCTION OXIDASE 3 (*FRO3)* (Waters et al., 2012), indeed showed the

expected lower (*FSD1*) or higher (*FRO3*) expression under the Fe deficiency treatment

compared to control (Figure S8a). Two of the 10 candidate genes, *FE SUPEROXIDE*

DISMUTASE 3 (*FSD3)* and *PRENYLCYSTEINE ALPHA-CARBOXYL METHYLTRANSFERASE*

(STE14A), are about 2-fold higher expressed in accessions with haplotype 1 compared to

accessions with either of the other two haplotypes (Figure 6f and Figure S8b). The Φ_{PSII}

in Fe deficiency correlates well with expression of these genes (Figure 6g). None of the

eight other candidate genes showed such response of expression between haplotypes

and correlation with the Φ_{PSII} in Fe deficiency (Figure S8b).

Since the absolute expression levels of *STE14A* are very low, we focused on *FSD3*

as the most likely candidate to underlie the QTL. Two distinct splice variants are

described for this gene that influence functionality regarding chloroplast development

(Lee et al., 2019). We measured the relative gene expression of each splice variant

independently, but found highly similar splice variant ratios between the different

haplotypes (Figure S8c), indicating that natural variation affects gene expression of *FSD3*

only quantitatively and not through altering differential splicing. Analysis of the non-

protein-coding sequences surrounding *FSD3* identified a variable $(TTC)_n/(GAA)_n$

microsatellite repeat directly downstream of the *FSD3* 3' UTR, which lacks either two or

four of such repeats in haplotypes 2 and 3 relative to haplotype 1. As there are no

differences in either Φ_{PSII} or *FSD3* expression between haplotype-2 and-3 accessions we

distinguished two alleles for the *FSD3* locus based on their functionality in Fe deficiency:

an Fe deficiency tolerant allele, which has the Col-0 reference sequence, and a sensitive

allele lacking two or four $(TTC)_n/(GAA)_n$ repeats. We subsequently confirmed the causal

nature of the allelic variation at *FSD3* to explain the QTL using quantitative

complementation (Weigel, 2012), in which we test if the difference in Φ_{PSII} between F_1

progeny derived from crosses between accessions with a tolerant allele crossed with

both Col-0 and the *fsd3* mutant, and F_1 progeny derived from similar crosses with

accessions carrying the sensitive allele. In Fe-deficient conditions, the F_1 progeny

derived from the *fsd3* mutant and accessions with a tolerant *FSD3* allele significantly

outperform the F_1 progeny derived from crosses with accessions with a sensitive *FSD3*

allele ($P = 0.046$) (Figure 6h). This is not found under control conditions (Figure 6i).

Figure 6: **Genome Wide Association (GWA) analysis of the response of photosynthesis**

efficiency to iron deficiency. (a) Distribution of Φ_{PSII}, the light-use efficiency of

photosystem II (PSII) electron transport, when grown in control (21 µM Fe^{2+}, grey) and

low iron supply (1 µM Fe^{2+}), orange). (b) Manhattan plot of the SNP variant associations

[-log(P) > 2] when using the residual values of the linear regression of Φ_{PSII} in control

conditions versus iron deficiency conditions as trait for GWA analysis. The red line

indicates a permutation-based threshold level of significance of $P < 0.05$. (c) The

genomic region surrounding the significantly associated region on chromosome 5 (green

box in (b)). Red dotted lines demarcate the area in linkage disequilibrium with the most

significant SNP, containing candidate genes depicted below (in grey). (d-e) Phenotypic

distributions for Φ_{PSII} (d) and projected leaf area (e) per haplotype. (f) Relative gene

expression levels per haplotype for *FSD3*. (g) Photosynthesis efficiency relative to the

average relative *FSD3* expression level per accession with a linear fit (*FSD3*: R^2 = 0.74, *P*

= 2.94 × 10^{-6}). (h-i) Φ_{PSII} of F_1 progeny of crosses between Col-0 (tolerant *FSD3* allele) or

the *fsd3* knockout mutant with accessions homozygous for the tolerant allele (N = 5)

(blue) and similarly with accessions homozygous for the sensitive allele (N = 9) (red) in

iron deficiency (h) and control conditions (i). Error bars represent SE. **P* < 0.05; **P <

0.01; ***P < 0.001, two-tailed Student's *t*-test.

Discussion

Here we present the DartMap panel, a new biodiversity panel consisting of 192

Dutch *A. thaliana* accessions, and we demonstrate its potential for detecting genetic

signs of local adaptation. The genetic variation observed in this panel largely falls within

the reported diversity of the 1001G collection, which is expected for accessions

collected in Northwestern Europe, outside the European relict regions (The 1001

Genomes Consortium, 2016). The genetic diversity is unexpectedly high considering the

relatively small geographical area without remarkable environmental clines. It illustrates

the high level of standing genetic variation that exists among local populations of this

predominantly self-pollinating species (Bomblies et al., 2010; The 1001 Genomes

Consortium, 2016). Despite, or perhaps due to, the absence of strong environmental

clines, the DartMap panel is very suitable for GWAS. The genetic architecture underlying

natural adaptation is strongly dependent on the local environment (Brachi et al., 2013;

Fournier-Level et al., 2013). Consequently, adaptive loci may exhibit genetic or allelic

heterogeneity in regions with contrasting climates, which hampers their detection

through GWAS. A milder diversity in environmental conditions can help to alleviate this

limitation, as exemplified by the successful identification of the well-known *FRI* gene,

which controls flowering time variation. This gene was not previously detected by GWAS

in other collections such as the Iberian collection (Tabas-Madrid et al., 2018) or the large

1001G panel (The 1001 Genomes Consortium, 2016), primarily due to allelic

heterogeneity. As such, the DartMap should be regarded as a valuable complement to

collections sampled from regions with contrasting climates, and not a replacement.

With the availability of several regional diversity panels, it will be easier to identify such

adaptive loci when taking into account that each region may favour its own adaptive

variation.

Our finding that genes involved in disease resistance are overrepresented among

those impacted by CNV is consistent with previous work: a similar phenomenon was

found for defence-response genes in a global set of 1,301 natural *A. thaliana* accessions

(Göktay et al., 2021) and this same category of genes was found to be enriched in

regions highly rearranged between seven diverse *A. thaliana* genomes (Jiao and

Schneeberger, 2020). These observations suggest that CNVs may act as a genomic

response to biotic stress. The next step would be to investigate whether such structural

variants correlate to variation in the prevalence and abundance of known pathogens of

A. thaliana (Bartoli et al., 2018).

The prevalence of plants with short inflorescences and the predominance of a

near-isogenic genotype at various dispersed locations, strongly indicate an adaptive

advantage for *ga5* mutants in the coastal regions of the Netherlands. Allelic

heterogeneity and strong selection of one successful, near-isogenic genotype

represented multiple times in the DartMap panel was the reason we could not identify

the *GA5* locus through GWAS, but only based on prior knowledge on possible causal loci

determining plant semidwarfism (Barboza et al., 2013). Signatures of selection were

previously identified in *GA5* among *A. thaliana* accessions collected globally (Barboza et

al., 2013), but the environmental cue driving such selection remained elusive. Later (Luo

et al., 2015) identified several *ga5* loss-of-function plants in two Alpine populations of *A.*

thaliana and noted a decrease in plant height with increasing altitude although the

adaptive advantage to high altitude environments specifically was not conclusively

demonstrated. Our findings demonstrate that high-wind treatment significantly impacts

the growth vigour of tall Dutch *A. thaliana* accessions, whereas it has no significant

effect on the vigour of predominantly coastally located semidwarf plants. Arguably, we

determined this for plants that are growing under optimal water and nutrient supply.

Under natural circumstances, with additional growth-limiting factors and probably

periods with much stronger winds, this reduction in vigour in the tall accessions is likely

to be more detrimental, and also likely to affect their fitness (Younginger et al., 2017) or

fecundity, which we could not establish under our artificial windy conditions. The

observed phenotypic responses in terms of biomass and stem length to windy

conditions are similar between the natural Dutch accessions and the Col-0 and Ler

reference strains, as well as their corresponding *ga5* loss-of-function lines. Considering

that wind speed generally increases with altitude in mountainous areas

(https://map.neweuropeanwindatlas.eu), the selective advantage of the Alpine

semidwarfs is also likely to be due to tolerance to wind speed. The selective pressure

seems high, given the two independent *ga5* alleles we found in the Netherlands, and

others found elsewhere in the world (Barboza et al., 2013; Luo et al., 2015). It is

remarkable that the near-isogenic genotype collected at several Dutch sites spread over

such large distances, without any obvious means of transport. This is in contrast to a

previous case of whole-genome hitchhiking, due to selection on herbicide tolerance,

where the spread was clearly linked to transportation along railway tracks (Flood et al.,

2016a). However, given the high population density of the Netherlands, we assume that

human-mediated movement of soil and seeds would also be sufficient to achieve

dispersal of the near-isogenenic *ga5* semidwarfs.

The presence of a North-South cline in flowering time and the association

between windy conditions and inflorescence height highlight the importance of

relatively mild environmental clines for local adaptation. These patterns can be

detected in a uniformly and densely sampled population like the DartMap. This is

further exemplified by the strong association on chromosome 4 related to the mild cline

in seasonality, where the alternative allele of *qDPT-B* is only found at a high local

abundance in the West of the Netherlands. However, it should be noted that such

associations do not necessarily provide evidence of causality. Demographic processes or

genetic drift can produce similar patterns to those created by selection (Rellstab et al.,

2015). To further investigate whether this association represents a case of local

adaptation, common garden experiments or reciprocal transplanting experiments can

be conducted (Hancock et al., 2011; Rellstab et al., 2015; de Villemereuil et al., 2016;

Terés et al., 2019). Nonetheless, the high local relative abundance observed in the West

of the Netherlands compared to rest of the world, along with the differential sensitivity

to drought, suggests the presence of an adaptive locus. However, it seems unlikely that

this adaptation is solely attributed to the drought response measured in our

experiments, as similar climate characteristics in other regions would also be expected

to exhibit a higher relative abundance. Therefore, it is more plausible that a

combination of different climatic factors and/or other unconsidered factors such as

urbanization contribute to this adaptation.

Identifying the causal gene in genotype-environment association analysis can be

challenging because allelic variation may affect tolerance to an experimentally

measured trait that may not necessarily reflect the trait under natural selection.

However, in our study we found that the mutant of *La-RELATED PROTEIN 1c* (*LARP1c*) is

more sensitive to drought compared to other tested mutants and Col-0. *LARP1c*

encodes a protein with a conserved La-motif domain, which is associated with RNA

recognition and is widely found among eukaryotes (Bayfield et al., 2010). Moreover, the

sequence variant with the highest statistical association in *qDPT-B* (SNP $4^{17000635}$) is

located in the 3' UTR of *LARP1c*. Although neither of these observations provide

conclusive evidence on whether this gene is underlying the *qDPT-B*, we consider *LARP1c*

as the most plausible candidate gene. In *A. thaliana, LARP1c* is known to be involved in

seed germination and the regulation of leaf senescence, which can be induced by

various environmental stresses (Zhang et al., 2012; Yan et al., 2023). Additionally, each

of the three LARP1 (1a, 1b, 1c) proteins in *A. thaliana* localise at the processing bodies

(Zhang et al., 2012; Yan et al., 2023) and are proposed to play a role in RNA processes in

response to environmental perturbations. For instance, *LARP1a* is required for

thermotolerance of *A. thaliana* to long exposure of moderately high temperatures,

where it is involved in regulating the RNA decay machinery together with the

cytoplasmic exoribonuclease XRN4 (Merret et al., 2013). However, the specific functions

of *LARP1c* in plants are still relatively understudied, and there may be other roles yet to

be explored (Yan et al., 2023). *LARP1c* may thus also be involved in the response to

drought and possibly also other environmental factors.

In addition to the mild and gradual environmental clines, we also examined the

panel for sensitivity to low iron availability, as we expected to find variation for this trait

given the distinct differences in soil type across the Netherlands. This identified non-

coding variation at *FSD3* to cause a major QTL affecting sensitivity to low iron supply.

FSD3 is long known to encode an essential chloroplast-localized iron superoxide

dismutase. Its function is to protect developing chloroplasts against excessive damage

by superoxide (Myouga et al., 2008). However, natural genetic variation in *FSD3* has not

been previously observed. The QTL affecting sensitivity to low iron supply is influenced

by subtle allelic variation in *FSD3*, specifically related to the number of repeats in a

microsatellite region directly downstream of the transcribed region. This variation

results in a two-fold difference in gene expression between contrasting alleles, which

strongly correlates with photosynthesis efficiency under iron-deficient conditions. When

plants are supplied with sufficient iron, there is no phenotypic difference between

contrasting haplotypes. FSD3 is one of two chloroplast-localized iron superoxide

dismutases, the other being FSD2 (Gallie and Chen, 2019). While both can form

heterocomplexes and *FSD3* overexpression can partly complement the *fsd2* mutant

phenotype (Gallie and Chen, 2019), they are not genetically redundant. The *fsd2* loss-of-

function mutant displays a compact rosette phenotype, with pale green leaves, and the

fsd3 loss-of-function mutant is barely viable, with a very small and pale rosette (Myouga

et al., 2008). Since iron is an essential co-factor for iron superoxide dismutases, a

decrease in iron supply is likely to affect their function. Under the mild iron deficiency

conditions we tested, this decrease in function was apparently just enough to affect

photosynthesis efficiency significantly more in the genotypes with lower *FSD3*

expression.

We found no reason to suggest strong selection for either of the two alleles.

Since the FSD3 function is essential for proper chloroplast development and tolerance

to reactive oxygen species generated by photosynthesis, lower *FSD3* expression is likely

disadvantageous. The impact of lower *FSD3* expression may even extend beyond

chloroplast development and photosynthesis efficiency. For instance, hydrogen

peroxide, which is produced when superoxides are converted by superoxide dismutases,

can function as a signaling molecule in the systemic response to iron deficiency (Le et

al., 2016). Therefore, plants with lower *FSD3* expression may have a delayed response in

detecting iron deficiency. Nevertheless, a large ionome survey of 1135 global *A. thaliana*

accessions did not reveal any clear indications of local adaptation to iron deficiency

(Campos et al., 2021). This suggests that establishing a direct link between allelic

variation and local adaptation to iron deficiency may be challenging. Factors such

seasonal variation in iron availability depending on precipitation or limited distribution

and heterogeneity of the allelic variation could contribute to the difficulty in establishing

such a link.

In conclusion, our work demonstrates that regions with relatively mild

environmental clines can still harbor a wide range of adaptive genetic variants. This

highlights the value of establishing regional collections as excellent tools for studying

local adaptation of *A. thaliana*. Such a collection does not need to be limited to regions

with strong environmental clines as were put forward in previous studies (Brachi et al.,

2013; Frachon et al., 2018). Even a diversity panel from a relatively small geographic

region with mild environmental clines, such as the Netherlands, provides enough

variation to pick up small differences in flowering time and sensitivity to iron deficiency.

Given the decreasing costs of whole genome sequencing, it becomes increasingly easier

to characterize the genetic variation in such populations in great detail. Our study may

serve as a blueprint of this approach, supporting the view that collections of wild plant

species can be powerful systems to determine the genomic basis of natural adaptation

(Weigel and Nordborg, 2015; Sollars et al., 2017; Monnahan et al., 2019).

Methods

Plant material

A set of 192 lines was selected from a large collection of around 2,000 natural accessions sampled throughout the Netherlands, with the aim to obtain a uniform geographical spread of accessions and generate a local diversity panel, the Dutch *Arabidopsis thaliana* Map (DartMap) panel, suitable for GWAS (Table S2). While accessions were selected from a larger set of about 2,000 collected individuals, the selected panel has a relatively lower density of accessions in the north of the Netherlands and in the very south, due to limited availability or collector capacity. Seeds collected from individual plants in the field were used to propagate each accession as a line for two generations through single-seed-descent upon self-fertilization in a greenhouse, before DNA isolation.

DNA isolation

Genomic DNA was extracted from one or more inflorescences (open flowers and above) from a single plant per accession. The material was frozen in liquid nitrogen, ground to a fine dust and incubated in 300μL 2x CTAB buffer (2% CTAB, 1.4m NaCl, 100mm Tris, 20mm EDTA, pH 8) for 30 minutes at 65°C. An equal volume of chloroform was added, mixed, centrifuged (3250 rpm for 15 min) and the supernatant was collected. The DNA was precipitated by adding an equal volume of ice-cold isopropanol, incubated overnight at -20°C and centrifuged (3250 rpm for 15 min). The precipitate was washed twice with 70% ethanol and air dried. DNA was dissolved in milliQ water and treated with RNase (Promega) for 30 minutes at 37°C.

Genomic DNA sequence data

Genomic DNA of the DartMap panel was sequenced at GenomeScan B.V., Leiden.

Libraries were prepared using the NEBNext Ultra DNA Library Prep kit for Illumina,

according to described procedures. 500-700 bp insert size fragments were sequenced

using a Illumina Hiseq 4000 device, aiming for at least 30x coverage of paired-end reads

(2x151 bp) of each accession, and used to generate the alignment files necessary for

calling genomic variants (see Note S1 for details). We additionally generated alignments

for the DartMap panel combined with 11 Dutch accessions included in the 1,001

Genomes (1001G) Project (The 1001 Genomes Consortium, 2016) (Sequence Read

Archive ID: SRP056687). This combined set of samples is referred to as the "DartMap +

1001G panel", to clearly distinguish it from the DartMap panel containing accessions

collected in this study only.

Single nucleotide polymorphism (SNP) and indel calling

Genetic variants were called using a workflow based on the Genome Analysis Toolkit

(GATK) Best Practices (Van der Auwera et al., 2013). Base quality scores of aligned reads

were recalibrated using GATK (version 4.0.2.1) BaseRecalibrator with default

parameters, using a set of variants called in a world-wide panel of 1135 *A. thaliana*

accessions as known sites (The 1001 Genomes Consortium, 2016) (obtained from

ftp.ensemblgenomes.org/pub/plants/release-37/vcf/arabidopsis_thaliana). SNPs and

short insertions/deletions (indels) were called in each sample using GATK

HaplotypeCaller, allowing a maximum of three alternate alleles at each site. Samples

were jointly genotyped using GATK GenomicsDBImport and GATK GenotypeGVCFs with

default parameters. This last step generated three different VCF files containing calls for

the nuclear genome, the mitochondrial genome, and the chloroplast genome

respectively. We filtered these sets to remove putative false positive calls (see Note S1

for details).

Copy number variation (CNV) calling

We called CNVs in the DartMap panel, defined here as deletions, duplications, and

insertions of at least 50 bp, by applying the Hecaton workflow (Wijfjes et al., 2019) to

the alignment files of each accession. Accessions of the 1001G were excluded from this

analysis, as they were sequenced using a different sequencing platform, which can

strongly affect CNV calling: a linear model of genetic distance between accessions based

on CNVs as a function of differences in coverage, sequencing technology, insert size, and

read length (see Note S1 for details) explained 30% of the variation in the number of

CNVs found in each sample (R^2 = 0.3). CNVs called in each accession were genotyped

and merged to generate a single VCF file as output (see Note S1 for details).

Characterizing the impact of deletions

We detected deletions overlapping protein-coding genes (by at least 1 bp) using

bedtools intersect. Genes were obtained from the TAIR10 genomic annotation of

Ensembl Plants (release 40) and from the annotation of the improved *A. thaliana*

mitochondrial assembly (BK010421.1). Gene function was considered disrupted if the

associated gene overlapped with a deletion and at least 76 bp (more than half the

length of a read) of the coding sequence lacked read coverage. Deletions were defined

as having a moderate frequency if they were present in at least 48 samples (25%) of the

DartMap panel. We defined disease resistance genes by first collecting all Uniprot

entries linked to all *A. thaliana* reference genes using the Retrieve/ID mapping tool

(https://www.uniprot.org/uploadlists/), excluding a small number of genes (53) for

which no corresponding entries could be found. Genes were labeled as involved in

disease resistance if the name of their protein product contained the terms "disease

resistance" or "Disease resistance". Enrichment tests were performed using Fisher's

exact test as implemented in the Python SciPy library (Virtanen et al., 2020) (version

1.3.1).

Population structure analysis

We clustered samples based on bi-allelic sites missing calls in fewer than 10% of the

samples and having a non-reference allele in at least 10 of them, to ensure that

accessions are grouped on variation at the population level, rather than rare variants

found in few individuals. For clusters based on mitochondrial and chloroplast variants,

this criterium was relaxed to a minimum of two samples, as a relatively large number of

sites would have been discarded otherwise. To generate a distance metric, we

represented samples by binary vectors with 1s for homozygous variants and 0 for all

other cases and computed the Hamming distance between them, i.e. the number of

variant sites differing between samples, using the dist.gene function of the R package

ape (Paradis and Schliep, 2019) (v5.3). Samples were clustered with the hclust function

in R (R Core Team, 2013), using complete linkage as the agglomeration method, similar

to analyses on the Swedish and Iberian collection (Long et al., 2013; Tabas-Madrid et al.,

2018) (Figure S9). We cut trees into different numbers of groups using the R function

cutree, to explore the geographical distribution of clusters formed at various

hierarchical levels. Geographical distance between accessions was computed using the

distance function of the geopy library (https://github.com/geopy/geopy) (v2.0.0) with

795   default parameters and their geographical locations were plotted using the basemap

796   library (https://matplotlib.org/basemap/) (v1.2.2).

Principal component analysis

Principal component analysis of chloroplast variants called in the Dutch and global

collection was performed using the SeqArray (Zheng et al., 2017) (v1.28.0) and

SNPrelate (Zheng et al., 2012) (v1.22.0) libraries in R. To remove putative false positive

calls, variants were excluded if they had a quality-by-depth (QD) score lower than 25,

leaving a total of 4095 sites. Furthermore, we removed 10 accessions in which at least

50% of the variants were called as heterozygous, suggesting DNA contamination of the

samples used for sequencing. Results were visualized using ggplot2 (Wickham, 2016)

(v3.3.2). Three outlier accessions were excluded from the final figures to improve the

visualization of Dutch accessions with respect to the global collection. Accessions were

coloured based on their chloroplast haplogroup assigned in a previous study (Hsu et al.,

2019). Haplogroups of accessions that were not included in that paper (i.e. the DartMap

panel) were predicted with a 1-nearest neighbour classifier on the accessions that were,

using the first two principal components as features.

Phenotyping flowering time and stem length

For all experiments, seeds were sown on wet filter paper and stratified for 3 days at 4

°C before they were germinated. Seedlings were grown hydroponically on rockwool

blocks (Grodan Rockwool Group, 40 x 40 mm in size) pre-soaked in a nutrient solution

designed for *A. thaliana* (1.7mm NH_4^+, 4.5mm K^+, 0.4mm Na^+, 2.3mm Ca^{2+}, 1.5mm

Mg^{2+}, 4.4mm NO_3^-, 0.2mm Cl^-, 3.5mm SO_4^{2-}, 0.6mm HCO_3^-, 1.12mm PO_4^{3-}, 0.23mm

SiO_3^{2-}, 21 μM Fe^{2+} (chelated with 3% diethylene triaminopentaacetic acid), 3.4 μM

Mn^{2+}, 4.7 μM Zn^{2+}, 14 μM BO_3^{3-} and 6.9 μM Cu^{2+} at pH 7, EC 1.4 mS cm^{-1}). Plants were

grown in a greenhouse until 97 days after sowing. For each treatment, plants were

equally divided over 20 plastic trays, with 76-78 plants per tray. Eight replicates per

genotype per treatment were used, randomly assigned over four separate trays with

two replicates per tray per genotype. Each tray consisted of a unique, randomly

generated combination of genotypes. Trays were placed in a random order in the

greenhouse, and were moved once every two weeks to a new position to minimize

possible spatial effects in the greenhouse. A vernalisation treatment of six weeks was

performed in a climate-controlled growth chamber set at 4 °C, with 12h/12h light/dark

period, 70 μM $m^{-2} s^{-1}$ irradiance and 70% humidity starting at day 20 after sowing. After

vernalisation, plants were returned to the greenhouse. Flowering time and height up to

the first silique were scored in the greenhouse, either with or without a vernalisation

period of six weeks. Flowering time was determined as the day when the first flower

had opened. Stem length up to the first silique was measured one week after the first

flower had opened, as this trait remains stable thereafter (Barboza et al., 2013). To

obtain the set of phenotypes for further analysis, we calculated the average phenotypic

values per accession per treatment.

Wind simulation treatment

In a separate experiment, plants of six semidwarf accessions and five tall accessions of

the DartMap panel, with similar flowering time, as well as the accessions Col-0 and Ler,

and their respective loss-of-function *ga5* mutants (Table S4) were grown for the first 14

days after sowing in a climate-controlled growth chamber, set at 20/18 °C (day/night)

with 10 h light (from 08:00 - 18:00) and 70% humidity at an irradiance of 200 μM $m^{-2} s^{-1}$

at ambient CO_2 levels. The growth chamber contained two treatment basins. The

treatment basins have the same design as described in a previous study (Flood et al.,

2016b), with the distinction that the two treatment basins are physically separated and

are on either side of the climate chamber. Each treatment basin was divided into five

equally sized blocks to hold plants. Accessions were grown in a randomized complete

block design with six replicates per block. Wind was simulated with two fans that were

positioned at either end of one of the treatment basins. The fans were set to provide a

wind speed that varied around 5.6 m/s for the majority of the treatment basin. The

wind speed varied depending on the distance of the plant to the fan, with plants closest

to the fan experiencing wind speeds up to 7 m/s and plants furthest away of the fan and

at the edge of the treatment basin experiencing 3.5 m/s wind. The wind speed for the

control treatment basin varied between 0.5 and 1.0 m/s. Wind speeds were measured

with a TSI Velocalc (model: 8347-M-GB) air velocity monitor. Wind treatment was

simulated by alternating patterns of six hours with wind by six hours without wind and

was terminated when the first plants in the wind treatment basin had fully ripened

siliques.

Extraction of climatic variables

Climatic variables were obtained from google earth engine using R and RGEE (Aybar et

al., 2020). All bands from Worldclim BIO data (Hijmans et al., 2005) selected from 250m-

by-250m squares using the coordinates from the collection site for each accession as

input.

Drought treatment

To test which of the two nearby QTLs at *qDPT* affected drought tolerance, plants were grown in a greenhouse on ceramic-based granular soil. For the first two weeks, all plants were watered three times a week with the nutrient solution described previously. This watering regime was continued for the control treatment while for the drought treatment excess nutrient solution was removed and watering was ceased. Accessions (Table S6) were grown in a randomised complete block design with five replicates per treatment. Photographs were taken with a Nikon D3000 with the same settings, zoom and distance to the plants and were subsequently analysed to obtain the projected leaf area (PLA) with ImageJ (settings Hue 26-114, Saturation 58-218 and Brightness 121-255). The PLA was then normalised per accession by dividing the PLA in the drought treatment by the PLA in the control treatment to correct for innate differences in size of different accessions. For candidate gene validation T-DNA insertion lines were used (Table S7). Plants were grown in our phenotyping growth chamber, the Phenovator (Flood et al., 2016b), with the same conditions in the growth chamber as described for the wind simulation treatment. The drought treatment was induced in a similar way, by using ceramic-based granular soil. In contrast to the experiment in the greenhouse, the growth substrate was covered with black plastic cover to prevent algal growth that might interfere with high throughput phenotyping. Moreover, we watered plants in the drought treatment only once at the start, as there was less evaporation in this setup compared to the greenhouse. Starting at 10 days after sowing, we measured projected leaf area based on near infrared reflectance (NIR) imaging as described by (Flood et al., 2016b) four times per day, and F_v/F_m using chlorophyll fluorescence once per day.

Iron deficiency treatment

In a separate experiment, plants were grown as described for the wind simulation experiment, but in this case, seven rather than five blocks per treatment basin were used. Plants in the control treatment were watered with the nutrient solution described above, while those under iron deficiency treatment received an adjusted solution with only 1 µM Fe^{2+} instead of 21 µM Fe^{2+}. Either three or four replicates were used per accession per treatment, with half of the accessions having four replicates in control condition and three replicates in the iron deficiency treatment and vice versa for the other half of the accessions. On day 15, plants were transferred to our phenotyping growth chamber, the Phenovator (Flood et al., 2016b), with the same conditions. We measured the steady state quantum yield of photosystem II electron transport (Φ_{PSII}), as a proxy for photosynthesis efficiency, and the projected leaf area, based on near infrared reflectance (NIR) imaging as described by (Flood et al., 2016b), at five resp. seven time points per day. We discarded outlier samples with NIR values smaller than the mean minus two standard deviations per treatment. We corrected the raw phenotypic data (Φ_{PSII} and projected leaf area) for block effects by fitting the following linear mixed model in which random terms are underlined:

$$\underline{Y} = Genotype + Treatment + (Genotype \times Treatment) + \underline{Block} + \underline{\epsilon}$$

The model was analysed using the restricted maximum likelihood procedure with the lme4 package in R (Bates et al., 2015). For the GWAS, we calculated the average Φ_{PSII} per accession per treatment. To obtain a single phenotypic value representing a measure of tolerance per accession, we then plotted Φ_{PSII} for each accession under the iron deficient condition versus control condition and fitted a linear regression through

the entire population. We calculated the residuals of each accession relative to this

linear regression to be used as the phenotypic value for GWA analysis. Additionally, we

used the average Φ_{PSII} values in the iron deficient condition and the calculated ratio

between Φ_{PSII} in iron deficiency versus control as alternative traits for GWAS.

Plant material for expression analysis

Plants that were used for gene expression analysis were grown in the greenhouse on

rockwool blocks that were watered with either the control nutrient solution (21 µM

Fe^{2+}) or the adjusted low iron solution (1 µM Fe^{2+}). For both treatments we used five

plastic trays to contain the nutrient solution. Two replicate plants per accession were

distributed over trays in a complete randomised block design. Whole shoots of two

replicate plants per accession originating from the same tray were pooled and

harvested as one sample for RNA isolation 21 days after germination, while five such

samples per accession per treatment were harvested in total. The material was

snapfrozen in liquid nitrogen immediately after harvesting and stored at -80 °C until

RNA isolation. Accessions were selected for gene expression analysis (Table S8) based

on their haplotype for the region with the highest association in the GWAS. Haplotypes

were determined by looking at pairwise correlations between all variants with a minor

allele frequency > 10% in a window of 20 kb surrounding the SNP with the highest

association.

Quantitative complementation

Quantitative complementation was used to test if the difference in Φ_{PSII} between the

F_1's of Columbia-0 (Col-0) and the knockout mutant *fsd3* in Col-0 background crossed

with accessions homozygous for the tolerant allele was significantly different from the

response in Φ_{PSII} for F_1's of Col-0 and *fsd3* crossed with accessions homozygous for the

sensitive allele. Five accessions with a tolerant, and nine accessions with a sensitive

allele for *FSD3* were crossed to both Col-0 and *fsd3* (Table S8). Col-0 and *fsd3* were used

as pollen donor in all crosses. The *fsd3* T-DNA insertion knockout mutant, SALK_103228,

was ordered from the Arabidopsis Biological Resource Center and insertion of the T-

DNA was validated by PCR as earlier described (Myouga et al., 2008). This T-DNA

insertion line has no *FSD3* transcript (Myouga et al., 2008), and we could only grow it

beyond the seedling stage on a 0.3% (w/v) gel-rite medium supplemented with 3% (w/v)

sucrose.

Plants (N = 10 per cross per treatment) were grown as described in the section

"Plant material for expression analysis". Plants were phenotyped for Φ_{PSII} from 21 DAS

onwards as described by (Baker and Oxborough, 2004) with the PlantScreenTM SC

System (Photosystems Instruments). Phenotyping was done for twenty plants per run

and lasted for three consecutive days overall. We phenotyped crosses with the tolerant

allele and the sensitive allele equally spread out over the three days to prevent possible

phenotypic differences caused by the delay between measurements. Moreover, in each

measurement of twenty plants we included F_1's from both the cross between a given

accession with Col-0 and with *fsd3* in a single measurement. We included one reference

tray that was measured each day in the morning, at the start of the afternoon and at

the end of the afternoon to test for possible differences in Φ_{PSII} during the day, but no

significant differences were found. We corrected the phenotypic data for possible block

effects by fitting a linear model, with random terms underlined:

Y = (FSD3 allele group x Pollen donor) + Block + ε

The model was analysed using the restricted maximum likelihood procedure with the

lme4 package in R (Bates et al., 2015). We tested interaction between the *FSD3* allele

group (natural tolerant or sensitive allele) and the pollen donor (Col-0 or the *fsd3*

mutant) with a two-way ANOVA, for which a Kenward-Roger approximation for the

degrees of freedom was used. Tukey's post-hoc multiple comparison test (α = 0.05) was

used to assess significance.

RNA isolation and gene expression analysis

Total RNA was extracted with the Direct-zol RNA mini-prep kit (Zymo Research)

according to the company's instructions and then subjected to a DNAse (Promega)

treatment at 37 °C for one hour. Total removal of DNA was validated by means of a no-

reverse-transcriptase PCR reaction on a few mixtures of samples. The RNA quality was

assessed for purity (A260/A280) with a spectrophotometer (Thermo scientific Nanodrop

2000) and for possible RNA degradation by means of a visual inspection of the RNA on a

1% (w/v) agarose gel. cDNA was then synthesized from 1 μg total RNA (measured by

spectrophotometer) with the iScript™ cDNA synthesis kit (Bio-RAD) according to the

company's instructions. Reverse transcriptase quantitative PCR (RT-qPCR) reactions

were performed with the SensiFast SYBR Green Mastermix (Bioline) on a Bio-RAD cfx96

machine. RT-qPCR primer sets were designed in parts of the coding region that did not

contain SNPs between the used set of accessions. For each RT-qPCR primer set used, we

first validated its efficiency by means of a standard curve and only used primer sets with

efficiencies ranging between 90% and 110%. Gene expression was normalized, using the

delta cycling threshold (ΔCt) method (Livak and Schmittgen, 2001), to three reference

genes, *SAND¸ YLS8*, and *TIP41-like* as earlier described (Han et al., 2013). Differential

expression was assessed by two-sample t-tests on the ΔCt values. All primer sequences

can be found in Table S10. We included *FSD1* and *FRO3* in the expression analysis as

marker genes for iron deficiency in shoots (Waters et al., 2012).

Genome-wide association (GWA) and genome-environment association

(GEA) analyses

GWA and GEA mapping were performed in GEMMA (Zhou and Stephens, 2012) (version

0.98) with a univariate linear mixed model and the minor allele frequency cut-off set at

0.05. A kinship matrix was constructed in GEMMA to correct for population structure.

CNVs were included in the analysis in a similar fashion as SNPs and indels by converting

them into a reference variant, an alternative variant or heterozygous. To estimate a

suitable genome-wide p-value threshold, we performed 1,000 permutations of the

phenotype values. For each permutation we performed association analysis and

extracted the highest association score. The distribution of these 1,000 highest

association scores was then used to determine the empirical threshold value of $p <$

0.05.

1006

Data availability

Raw sequencing data of the DartMap panel can be found on the repository of the

National Center for Biotechnology Information (NCBI) (BioProject ID: PRJNA727738).

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bodies had no role in the design of the study; the collection, analysis, and interpretation

of data; and in writing the manuscript.

1017   Authors' contributions

SS, DDR and MGMA designed the project with RB and RYW. RB composed the panel.

RYW performed computational analysis of sequencing data and overall patterns of

genetic diversity. Wet-lab experiments were conducted by RB, FFMB, MM, JJV, NG and

LMD. Subsequent data analysis of wet-lab experiments was performed by RB, MM, JJV,

NG and LMD. TPJMT contributed especially to data collection and analysis of sequencing

data of non-Dutch *A. thaliana* accessions. BLS extracted environmental data. RYW, RB,

BLS, JJV, MM, NG, LMD, MK, FAvE, SS, DDR, and MGMA contributed to the experimental

design, interpretation of results, and preparation of the manuscript. All authors read and

approved the final manuscript.

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Competing interests

1037   The authors declare that they have no competing interests.

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