Title Page

Title

Phylogenomics supports island contribution to metapopulation dynamics in a predominantly continental bird species

Short Running Title

Phylogenomics in continent-island system

Authors

AOKI Daisuke[1,*], SENZAKI Masayuki[2], ANDO Haruko[3], ODAYA Yoshiya[4], HEIM Wieland[5,6], KITAZAWA Munehiro[3,7], WULF Tom[8], TRENSE Daronja[9], BASTARDOT Marc[10], FUKUDA Atsunori[11], TAKAHASHI Masao[12,13], KONDO Natsuko[3]

[1.] Department of Wildlife Biology, Forestry and Forest Products Research Institute, Tsukuba, Japan 305-8687

[2.] Faculty of Environmental Earth Science, Hokkaido University, Sapporo, Japan, 060-0810

[3.] Biodiversity Division, National Institute for Environmental Studies, Tsukuba, Japan, 305-8506

[4.] Abiko City Museum of Birds, Abiko, Japan, 270-1145

[5.] Swiss Ornithological Institute, Sempach, Switzerland, 6204

[6.] Department of Evolutionary Biology and Environmental Studies, University of Zurich, Zurich, Switzerland, 8006

[7.] Graduate School of Agriculture, Hokkaido University, Sapporo, Japan, 060-0809

[8.] Department of Agriculture, Ecotrophology, and Landscape Development, Anhalt University of

Applied Science, Bernburg, Germany, 06406

[9.] Department of Biology, Institute of Integrated Natural Sciences, Koblenz-Landau University,

Koblenz, Germany, 56070

[10.] Department of Ecology and Evolution, University of Lausanne, Lausanne, Switzerland, 1015

[11.] The Wild Bird Society of Japan, Ibaraki Branch, Mito, Japan, 310-0055

[12.] Iwate Prefectural Museum, Morioka, Iwate, Japan, 020-0102

[13.] Oosekka-land, Hachinohe, Aomori, Japan, 031-0823

* Corresponding author: Daisuke AOKI, aokid@ffpri.affrc.go.jp, +81-29-829-8258

Current address

HEIM Wieland: Institute of Biology and Environmental Sciences, University of Oldenburg,

Oldenburg, Germany, 26129

ODAYA Yoshiya: Chiba Natural History Museum and Institute, Chiba, Japan, 260-0852

OCRID

AOKI Daisuke: 0000-0002-5780-8487

SENZAKI Masayuki: 0000-0002-3843-5793

ANDO Haruko: 0000-0001-7827-3883

ODAYA Yoshiya: 0009-0004-1057-4005

HEIM Wieland: 0000-0002-3262-2491

KITAZAWA Munehiro: 0000-0003-0167-4464

TRENSE Daronja: 0000-0003-3598-8699

FUKUDA Atsunori: 0009-0009-2479-2886

TAKAHASHI Masao: 0000-0001-8385-6098

KONDO Natsuko: 0000-0002-0883-0506

Author Contributions

DA and MS conceived the ideas and designed the study with contributions by WH; MS, YO,

WH, MK, and AF led the field work with substantial support by DA and MT; HA, NK, and DA

led the laboratory procedures; DA conducted bioinformatics and analyzed the data with supports

by HA and NK; DA coordinated the study, prepared the draft, led the writing and revised the

manuscript with assistance from MS. All authors gave final approval for publication and agreed

to be held accountable for the work performed therein.

Acknowledgements

Some tissue and blood samples were kindly provided by the Yamashina Institute for

Ornithology. We thank many members of our field teams; Toshio Sadakuni for sampling in

Kushiro, Akio Miya and Jun-ichi Ebina for sampling at Hotokenuma, Aomori; Sergei M.

Smirenski and Yury Anisimov for sampling in Russia; Shigeo Ozawa, Ryosuke Abe, Koichi

Shirakawa, Daisuke Akoshima, Ayumu Hamachi, Hiroko Shirakawa, Mikiya Oikawa, and

Shiomi Hakataya for sampling in the Kanto region. We also thank Nobuyoshi Nakajima for his

support on sequencing runs on MiSeq. Our field surveys were conducted with permission from

the landowners of each site, including Tomatoh. DNA samples were collected with permission

from the Ministry of the Environment Government of Japan (permission # includes 21-31-0172-

74, 19-129, 1810254, 2012217-16). We also acknowledge the provision of rings by the Bird

Ringing Centre of Russia. Transportation of DNA samples was conducted under a Material

Transfer Agreement between the National Institute for Environmental Studies and the Amur Bird

Project based on the Access and Benefit-Sharing (ABS) agreement in the Nagoya Protocol.

Bioinformatics was mostly performed on the supercomputer of AFFRIT, MAFF, Japan.

Funding

This study was partly conducted with the support of a grant-in-aid for Scientific Research (C) to

DA (no. 22K20670) and MS (no. 23H02243) from the Japanese Society for the Promotion of

Science (JSPS).

Conflict of Interest Statement

The authors declare no conflicts of interest.

Abstract

Aim:

Islands have recently been recognized as potential sources of biodiversity, challenging the

traditional view that their small population sizes and low genetic diversity limit such roles. This

raises the question of how insular genetic variation becomes incorporated into continental

populations, contrary to expectations of unidirectional colonization. Here, we investigate

whether and how island-derived genetic variation has influenced a continental population

through population establishment and gene flow in a bird species where frequent trans-ocean

dispersal is expected.

Location:

Continental East Asia (Russian Far East), Japanese Archipelago

Taxon:

Swinhoe's Rail (*Coturnicops exquisitus*)

Materials and Methods:

We apply integrative phylogenomics to reconstruct the spatiotemporal history of the species.

Colonization sequences and gene flow are inferred by comparing four different phylogenetic

reconstruction methods, using mitochondrial sequences obtained by Sanger sequencing and

genome-wide data obtained by genotyping by sequencing (MIG-seq). We assess a history of

colonization and gene flow based on summary statistics, demographic trajectory inference by

Stairway Plot2, demographic modeling by fastsimcoal2, and species distribution modeling.

Results:

Analyses collectively supported asymmetric gene flow from the island to the continental

population, following divergence around the Middle Pleistocene. Post-divergence, the island

maintained a large and stable population, while the continental population underwent a severe

bottleneck, suggesting a significant evolutionary role of the island for the continental population.

Additionally, evidence of recent re-establishment of the island by continental individuals

indicates dynamic exchange and persistence within a continent-island metapopulation.

Main conclusions:

The maintenance of insular genetic variation within a dynamic continent-island metapopulation

may have enabled the island to act as a genetic and demographic reservoir for the continental

population. Thus, continent-island metapopulation dynamics may be a key evolutionary pathway

through which island populations contribute to continental genetic diversity.

Keywords

Coturnicops exquisitus, genetic variation, gene flow, island biogeography, metapopulation,

phylogeography, reverse colonization, Swinhoe's Rail

Main Text

Introduction

Island biological diversity contributes disproportionately to global biodiversity (Whittaker and Fernández-Palacios 2007). Islands, comprising 3.5% of the terrestrial land, contribute nearly 20% of global species richness and more than 60% of terrestrial extinctions (Whittaker et al. 2017). Traditionally, islands were assumed to sustain smaller population sizes than continents due to their small and isolated landmasses (Leroy et al. 2021). Small populations are typically subjected to strong genetic drift, resulting in reduced genetic variation (Leroy et al. 2021; Cerca et al. 2023), which likely increases extinction risks (Frankham 2005) and leads to "evolutionary dead ends" (Vamosi et al. 2014). Consequently, islands have often been viewed as sinks where lineages evolve independently and do not contribute to global diversification (Bellemain and Ricklefs 2008).

Recently, evidence has emerged showing that islands can also serve as sources of continental colonization (Whittaker et al. 2017). Macroevolutionary (i.e., species-level) studies revealed numerous cases of continental species or lineages nested within island radiations–a pattern termed as "reverse colonization" (Bellemain and Ricklefs 2008). A review found that over one-third of the phylogenies across diverse taxa in various continent-island systems exhibited such patterns (Bellemain and Ricklefs 2008). Reverse colonization can also trigger mainland diversification. For example, the passerine radiation–now nearly 60% of global bird species–originated from colonization out of New Guinea (Jønsson and Holt 2015).

Although reverse colonization operates at the species level (i.e., a new continental species arises from an ancestral island species), its microevolutionary (i.e., population-level)

mechanisms remain poorly understood. Speciation requires several population-level processes,

including the establishment and persistence of populations–processes influenced by

spatiotemporal dynamics of migration between the ancestral and derived populations (Harvey et

al. 2019). For instance, depending on divergence levels, gene flow from ancestral populations

may lead to genetic or demographic rescue (Hufbauer et al. 2015), introduce novel genetic

variation (Cerca et al. 2023), or cause incompatibilities (Bank et al. 2012), thereby facilitating or

impeding divergence. Given the distinct population genetic processes and environments

experienced by island and continental populations (Barton 1996; Weigelt et al. 2013), these

population-level processes are likely important at the initial stage of reverse colonization. To our

knowledge, there has been only one study that modeled the population genetic process of

continental establishment by insular organisms (Patiño et al. 2015). This study, however, used

bryophytes, whose genetic variation may not be sensitive to demographic changes due to their

ability for asexual reproduction (Ellegren and Galtier 2016), leaving why many sexually

reproductive animals exhibit reverse colonization uncovered. Furthermore, gene flow in

continent-island systems has mostly been studied in the context of divergence-with-gene-flow–

how island populations diverge despite gene flow (Warren et al. 2015)–with a focus on genomic

regions resisting homogenization (Feder et al. 2012). As a result, studies have been biased

towards the effects of continent-derived gene flow, with limited exploration of migration

dynamics and their demographic consequences in continent-island systems as a whole.

Phylogeography is a powerful framework to examine population establishment and gene

flow directionality in continent-island systems, as it infers historical processes shaping present

genetic patterns (Lomolino et al. 2017). When integrated with demographic reconstruction and

species distribution modeling (SDM), it becomes especially effective in reconstructing

spatiotemporally explicit histories. Phylogenetic trees provide patterns that emerge as a result of

demographic processes, while the processes themselves remain unmodeled (Harvey et al. 2019).

In contrast, demographic reconstruction using high-throughput sequencing explicitly models

these processes (Nadachowska-Brzyska et al. 2022). SDMs estimate the timing and extent of

historical range shifts (Harvey et al. 2019). Therefore, these methods aid inference on the

direction, intensity, and timing of population establishment or gene flow.

Here, we apply integrative phylogeography to examine the direction and demographic

consequences of population establishment and gene flow in a continent-island system. We

selected Swinhoe's Rail (*Coturnicops exquisitus*), the smallest Asian rail, as the ideal model.

Demographic processes shape the present genetic variation of birds more strongly than in many

other taxa (Ellegren and Galtier 2016), and therefore, they are expected to underpin important

demographic processes in continent-island systems. Rails are known for repeated transoceanic

dispersals, as inferred from phylogenetic reconstruction at the species level (Garcia-R and

Matzke 2021). Swinhoe's Rail mainly breeds in continental East Asia (Heim et al. 2019), while a

limited number of island populations have recently been discovered (Senzaki et al. 2020).

Therefore, we expect that Swinhoe's Rail is prone to frequent colonization events across the sea,

even at the population level, and it is ideal to examine how a currently small island population

has historically contributed to the continental population establishment, persistence, and

evolution.

In this study, we reconstruct the phylogeographic history of Swinhoe's Rail to test

whether island genetic variation has contributed to continental populations via population

establishment and/or gene flow. We infer the direction of establishment by examining 1) whether

the insular population forms an early-branching clade with nested continental populations, 2) the

demographic stability of each population, and 3) habitat suitability at the time of divergence. We

assume that derived populations arose through bottlenecks from stable ancestral ones (e.g.,

Yeung et al. 2011). We determine the direction of gene flow by 1) analyzing phylogenetic

signals of gene flow, 2) estimating gene flow by demographic modeling, and 3) predicting

historical range shifts that could enable migration by the SDM. Finally, we discuss how these

events impacted population size and genetic variation on the continent.

Materials and Methods

Field sampling

Swinhoe's Rail is a poorly studied species. Previously known breeding ranges were limited to

the continent (Fig. 1a), while evidence of breeding is now absent from most of the region (Heim

et al. 2019). The currently recognized breeding areas were re-discovered recently, including the

Amur region (Heim et al. 2019) and Lake Baikal (Anisimova et al. 2019) in Russia, as well as

parts of Japan, namely Aomori of Honshu (Miya et al. 2005) and Hokkaido (Senzaki et al. 2020)

(Fig. 1a). Despite the species' cryptic behavior and preference for inaccessible wetlands, we

successfully developed sampling strategies (Heim et al. 2019; Senzaki et al. 2020) and collected

feather, buccal swab, or blood samples covering the entire extent of the currently known

breeding range: Amur (n = 6) and Baikal (n = 1) for continental Russia, Kushiro (n = 12) and

Tomakomai (n = 8) for Hokkaido, and Aomori (n = 4) for Honshu during their breeding season

(Fig. 1a, Table S1). We also collected samples of wintering birds in the Kanto region of Honshu

(n = 37) and used a genetic sample from a migratory bird in Ishikari, Hokkaido (Supplementary

Methods), to assess migratory connectivity and historical range shifts. Additionally, we retrieved

mitochondrial data from GenBank for phylogenetic analyses (Table S1).

Laboratory procedures & Bioinformatics

The whole genomic DNA was extracted from each tissue and used in two experiments: Sanger

sequencing of the partial mitochondrial Cytochrome *b* (*Cytb*) region for 49 samples, and

multiplexed inter-simple sequence repeats genotyping by sequencing (MIG-seq; Suyama and

Matsuki 2015) for 52 samples (Table S1). MIG-seq, a polymerase chain reaction-based

genotyping by sequencing approach, is effective for low-quality DNA at the cost of reduced

genome coverage and is appropriate for swab and dried feather samples. It yielded 4,834,210

read pairs (9,668,420 reads), which were trimmed and aligned to a reference genome of a closely

related species, *Laterallus jamaicensis coturniculus* (WGS Accession ID, JAKCOX01, Table

S1). Three individuals with over 30% missing sites in ANGSD pre-runs (one from Amur and two

from Kanto) were excluded (Fig. S1–S4), yielding a final dataset of 7,244,578 paired and 37,186

unpaired reads for 49 samples. See Supplementary Methods for parameter settings.

We accounted for genotype uncertainty by calculating genotype likelihoods (GLs) with

ANGSD (Korneliussen et al. 2014) (see Supplementary Methods, Fig. S1–S2). GLs were first

calculated separately for five regional populations, using half the sample size of each population

as '-minInd' and 2 as '-setMinDepthInd', retaining sites with depth ≥ 2 in over half the samples.

For downstream analyses, GLs were recalculated on intersecting sites among sample subsets

using different filtering parameters, yielding varying site counts (Table 1, S2, Fig. S4). For

analyses of variant sites, we set 1e-06 for '-SNP-pval' and 0.05 for '-minMaf'. For SNAPP

analyses, genotypes were called using '-doGeno' with a posterior cutoff of 0.9. Populations were

defined either by five sampling regions (Fig. 1a) or by two genetically supported breeding

groups–"island" (Kushiro + Tomakomai) and "continental" (Russia + Aomori) (Table 1). This

grouping was robustly supported by multiple analyses (Fig. 1b–c, 2b, S5–S10). The 'SAMtools'

model was used in the entire GL calculations. Linkage disequilibrium (LD) was assessed and

pruned using ngsLD (Fox et al. 2019) for some analyses (Table 1). Demographic inference used

folded site frequency spectra (SFS). Full workflows and parameters are provided in

Supplementary Methods and Fig. S4.

Genetic structure

We examined global genetic structure via principal component analysis (PCA) using PCAngsd

(Meisner and Albrechtsen 2018), ADMIXTURE analysis with NGSadmix (Skotte et al. 2013)

with K=1–6, NeighborNet (Bryant and Moulton 2004) based on pairwise genetic distance from

ngsDist (Vieira et al. 2015), and the D-statistic using 'doabbababa2' in ANGSD (Soraggi et al.

2018) (see Supplementary Methods for details). Two Kanto samples, closely related to one

Tomakomai and one Kushiro sample, showing kinship coefficients (KING, Waples et al. 2019) >

0.088 based on estimates from ngsRelate (Hanghøj et al. 2019), were excluded to meet the

assumptions of these analyses. We included two American *Laterallus* group species (*Laterallus*

jamaicensis, Accession SRX14572910; *Atlantisia rogersi,* Accession SRX6608087) as

outgroups in ngsDist and supplementary PCA/ADMIXTURE analyses (Fig. S5–S7). After

confirming their monophyly, only *L. jamaicensis* was used as an outgroup for phylogenetic

reconstruction.

Inference of phylogenetic relationship

Phylogenetic trees from different datasets and methods can reflect gene flow and incomplete

lineage sorting (ILS) differently (Twyford and Ennos 2012). Thus, we inferred a phylogenetic

relationship through four approaches: a mitochondrial phylogenetic tree (mtDNA tree) using

Cytb data, a maximum likelihood tree (RAxML tree), a graph-based population tree based on

allele frequency (TreeMix), and a coalescent-based population tree based on single nucleotide

polymorphisms (SNPs) (SNAPP tree) using MIG-seq genome-wide nuclear data.

mtDNA tree: The time-calibrated Bayesian inference was performed on 772 bps of partial *Cytb,*

a widely used marker suitable for avian phylogeography (e.g., Weir and Schluter 2004), using an

uncorrelated lognormal relaxed clock model and coalescent exponential population prior in

BEAST v.2.7.3 (Bouckaert et al. 2019). Samples included 12 haplotypes from 54 Swinhoe's rails

and three outgroups (two *C. noveboracensis*, one *L. jamaicensis*; Table S1). The standard

mitochondrial molecular clock rate for birds, 1.05%/lineage/Myr (Weir and Schluter 2008), was

applied to estimate divergence times. Markov chain Monte Carlo (MCMC) was run for 2 million

generations, sampling every 1,000. Convergence was checked by following Aoki et al. (2018),

and a maximum clade credibility tree with mean heights was drawn after discarding the first 10%

as burn-in. We confirmed the inferred tree by a haplotype network. See Supplementary Methods

for settings.

RAxML tree: A maximum likelihood tree was reconstructed from both variant and invariant

sites using RAxML-ng (Kozlov et al. 2019) under the 'GTGTR4+FO+G' model. Genotype

probabilities (GPs) calculated in a VCF file output of ANGSD were used to prepare a CATG

input file. For missing sites, a low, uninformative probability of 0.01 was assigned. With '--prob-

msa' enabled, the tree was reconstructed considering genotype uncertainty. Standard non-

parametric bootstrapping with 100 replicates was simultaneously conducted.

TreeMix: We used TreeMix (Pickrell and Pritchard 2012) to infer a population-level tree

incorporating gene flow and drift. Allele frequencies were estimated based on GLs using

glactools (Renaud 2017), and an ACF file was converted to TreeMix input by the 'acf2treemix'

program with a '--noprivate' option, to exclude population-specific alleles. Migration edges

ranging from zero to five were tested across ten replicates with a block size of 20 (based on the

LD decay; Supplementary Methods) for jackknife sampling. The best number of migration edges

scoring the highest Δm score was determined by an R package 'OptM' v. 0.1.6 (Fitak 2021),

selecting the run with the highest model likelihood as the best-fit model. We repeated the same

analyses without grouping (i.e., individual-level tree) to confirm that inferred gene flow reflects

actual migration rather than population grouping artifacts.

SNAPP tree: SNAPP infers a population tree based on a coalescent model using unlinked SNPs

without missing data (Bryant et al. 2012). We selected three or four samples scoring the lowest

missing proportion from each breeding population (three from Aomori due to the small sample

size), whose missing proportions ranged between 2% and 12%. Unlinked hard-called SNPs

without missing data were obtained from these samples by ANGSD. We estimated tree topology

while simultaneously estimating divergence times (Stange et al. 2018). We calibrated the split

between Swinhoe's Rail and *L. jamaicensis* using two log-normal priors (lognormal(5.11, 0.09)

and lognormal(8.95, 0.0225) in million years ago [Mya]), which corresponded to its divergence

time estimates by our clock-calibrated mtDNA tree (Fig. 2a) and fossil-calibrated mtDNA tree

by Garcia-R et al. (2020), respectively. We ran 500,000 MCMC iterations to get posterior trees

and analyzed the posterior trees using 'TreeSetAnalyser' to derive a 95% credible set.

Demographic inferences

We examined the historical demography of the continental and island populations via summary

statistics, demographic trajectory inferences (Stairway Plot 2, SWP2), and parameter estimations

in demographic models (fastsimcoal2, FSC2). These methods are complementary to each other

because SWP2 is a nonparametric model, flexibly inferring complex effective population size

(N_e) trajectories yet without modeling gene flow, while FSC2 models both gene flow and N_e

changes but in user-specified simpler models (Nadachowska-Brzyska et al. 2022). We also

estimated the contemporary N_e of the island population by converting the estimated census

population size (N_c) in a plain of Hokkaido in the early nineteenth century (Kitazawa et al.

2022), using N_c:N_e = 1:0.44, a mean ratio for birds (Frankham 1995).

Summary statistics: We calculated Watterson's theta θ_W (Watterson 1975) and Tajima's D

(Tajima 1983) per reference scaffold and population. Tajima's D values around zero suggest

demographic stability, and values below zero (especially <-1) indicate a strong ancient

bottleneck (Gattepaille et al. 2013). Furthermore, the large and small variance of Tajima's D

among scaffolds reflects moderate bottleneck and population expansion, respectively (Gattepaille

et al. 2013). Per-scaffold estimates of these indices were compared between populations using

linear mixed models. We also estimated the fixation index (F_{ST}) using two-dimensional SFS

between four breeding populations. Details in Supplementary Methods.

SWP2: We inferred demographic trajectories of the continental and island populations using

one-dimensional folded SFS in Stairway Plot 2 v. 2.1.1 (Liu and Fu 2020). We assumed a

mutation rate of 1.91e-09 bp/year (Nam et al. 2010) and converted it to bp/generation by using a

generation time of 1 year (a value is for its congener, *C. noveboracensis*; Alvo and Robert 1999).

Default settings were used. The timing at which populations started to show diverging

demographic trends was interpreted as a sign of population divergence.

FSC2: We tested five models differing in the presence and timing of migration matrices (Fig.

3c), using two-dimensional SFS. We estimated t_{MIG} at which both the continental and island

populations experienced demographic changes and migration matrices may have switched, by

fixing their divergence time at 0.5 Mya, which was roughly but collectively agreed by multiple

approaches (Fig. 2a, 2d, 3f). Between the two populations, we modeled no migration throughout

("no migration"), migration only after ("recent migration") or before ("ancient migration") t_{MIG},

and migration throughout at the same rates ("constant migration") or different rates ("different

migration") across t_{MIG} (Fig. 3c). We conducted 100 replicates of 1 million coalescent

simulations across 40–100 optimization ECM cycles for parameter estimates, in fastsimcoal2 v.

2.7.0.9 (Excoffier et al. 2021). The same mutation rate as the SWP2 analyses was used. A

simulation with the smallest difference between 'MaxEstLhood' and 'MaxObsLhood' was

selected from each model, and models with the ΔAIC (delta Akaike Information Criterion, the

difference between the lowest AIC and an AIC in focus) <2 were considered best-fit. The ranges

of parameter values for simulation are in Table S5.

Species distribution modeling

SDMs were run using MaxEnt (Phillips et al. 2006) to model the species' breeding extent. We

compiled 48 breeding and 111 non-breeding records from previously published sources (Table

S6), including both previously known and recently discovered sites (Fig. S20). After spatial

thinning (one record per 0.5º grid cell), 33 breeding points were retained as an occurrence input

(Fig. 4b). Background points were sampled to cover one-fourth of the total number of girds in a

1,000-km buffered convex hull around the full dataset (both breeding and non-breeding records),

resulting in 2,104 points (Fig. S20). We extracted time slices of 22 environmental variables

(Krapp et al. 2021) using an R package 'pastclim' v. 1.2.3 (Leonardi et al. 2023) and selected

eight covariates with low collinearity, satisfying that any pair scores variance inflation factor

<10. Twelve models with different parameter settings were tested via 'ENMevaluate' in an R

package ENMeval v. 2.0.4 (Kass et al. 2021), and the best model was selected based on the

lowest ΔAIC for a small sample size (ΔAICc). The best-fit model was projected across time

slices from the present and 0.5 Mya at 1,000-year intervals (Fig. 4c). We compared the historical

changes in the potential habitat of this species on the island and the continent (Fig. S23), by

calculating three indices, habitat suitability (sum of values of the predicted presence grids),

availability (the total number of predicted presence grids), and quality (suitability divided by

availability). We also calculated centroid distances (weighted by suitability) between island and

continental habitats to track historical changes in the level of geographic isolation. Effects of

negative multivariate environmental similarity surface (MESS) were controlled for these

calculations. See Supplementary Methods for details.

Results

Phylogenetic relationships and gene flow

Multiple phylogenetic methods collectively inferred population divergence. A bifurcation

between an island clade (Tomakomai and Kushiro) and a continental clade (Russia and Aomori)

was supported by TreeMix (Fig. 2c), SNAPP tree (Fig. 2d), and the D-statistic (Fig. S10). This

structure was further supported by ADMIXTURE at K = 2 that scored the highest ΔK value (Fig.

1c, S6–7), the first axis of PCA explaining >99% of variation (Fig. 1b, S5), NeighborNet (Fig.

S8), and F_{ST} (Fig. S9). On the other hand, some phylogenies indicated ILS and/or post-

divergence gene flow. In the mtDNA tree, divergence between the two clades was supported,

while Russian samples appeared in both clades, and most Tomakomai and Kushiro samples

clustered in the island clade (except one from Kushiro) (Fig. 2a, S11). This pattern suggests

either ILS due to the island population being derived from a large continental population or post-

divergence gene flow from the island to the continental clade. Low bootstrap supports in the

RAxML tree (Fig. 2b, S12–13) and low posterior probabilities for a bifurcation tree (41.5%) in

the SNAPP tree (Fig. 2d) can also reflect this uncertainty. The divergence time of the two clades

was estimated to be relatively old, around 0.5 Mya (ranging between 0.2–1.0 depending on

analyses), corresponding to the Middle Pleistocene (the Chibanian epoch), collectively supported

by different calibration methods, data, and algorithms, including the mtDNA tree (Fig. 2a), the

SNAPP tree (Fig. 2d), and SWP2 (Fig. 3f).

Gene flow was also inferred by the best-fit population-level TreeMix tree with one

migration edge scoring the highest Δm value (Fig. 2c, S14, S16), which was also supported by

our individual-level tree (Fig. 2c, S15), indicating that gene flow inferred in our population-level

analyses is not the artifact of population grouping (see Supplementary Discussion, Fig. S16–

S19). Meanwhile, gene flow was directed from the common ancestor of this species to Russia.

On our individual-level tree, the Tomakomai samples were located at the basal position. This

suggests that ancient variation may have been introduced to the Russian population via gene flow

from a population preserving ancient genetic variation, such as the Tomakomai population

(Supplementary Discussion, Fig. S17–S19). This gene flow possibly occurred relatively recently

(e.g., post-expansion of the continental population), as migration was not towards the ancestor of

the continental cluster (Fig. 2c), but to Russia (at the population-level) or to an ancestor of one

Aomori and one Russian sample (at the individual-level). One Russian individual clustered with

island samples (Fig. 2c, inset), which was inferred to have the island cluster in our

ADMIXTURE analysis at K = 3 (Fig. 1c), also supporting recent gene flow.

Aomori samples consistently clustered with and showed low genetic differentiation from

Russian samples in all methods–phylogenetic trees (Fig. 2), PCA, ADMIXTURE (Fig. 1b–c),

NeighborNet (Fig. S8), F_{ST} (Fig. S9), and the D-statistic (Fig. S10)–suggesting recolonization of

Aomori by the continental population. This was unlikely to be a sample size artifact, as

supported patterns held across genetic distance (NeighborNet) and F_{ST} that consider sample size

and were robust to excluding Kanto wintering samples (i.e., smaller effect of sample size) (Fig.

1c, S5–S7). Additionally, the Baikal sample, collected ~1,500 km from Amur, showed no major

differentiation, despite the limited sample size. Together with the genetic position of Aomori, the

present range of the continental population is a result of a recent expansion.

Historical demography and contemporary genetic variation

The continental population had marginally higher genetic diversity than the island population (*p*

= 0.089; Fig. 3a, Table S3). FSC2 results suggested that gene flow from the island contributed

significantly to contemporary continental diversity. The best-fit FSC2 model (ΔAIC <2.0) was

"different migration" model (ΔAIC = 0.0, Akaike model weight = 0.747), with "constant

migration" as the second-best yet with a marginal ΔAIC value (ΔAIC = 2.17, Akaike model

weight = 0.253). Both supported post-last-glacial-maximum (LGM, ~0.02 Mya) gene flow (t_{MIG}

= 0.014–0.019 Mya) from the island to the continent, at rates 1.8–2.7 times higher than in the

opposite direction (Fig. 3c–e, Table S4). In contrast, gene flow before the LGM was inconclusive

due to significant differences between the two models.

Our demographic analyses suggested that the island population remained relatively

stable, or, if any, experienced a modest level of expansion since the population split until the

LGM, supported by Tajima's *D* values close to zero (Fig. 3b, Table S3), and FSC2 (0.5–2.3-fold

change), and SWP2 (~1.9-fold change) (Fig. 3d–e, Table S4). Meanwhile, different methods

showed discordant results for the demographic history of the continental population. Expansion

after a strong ancient population bottleneck was suggested by the significantly low value of

Tajima's D close to -1 (median value = -0.845) with a smaller inter-scaffold variance than the

island population (Fig. 3b) and the best-fit FSC2 models (nearly 98% population decline, Fig.

3d–e), while expansion soon after the population split around 0.5 Mya without a population

bottleneck was suggested by SWP2.

A sharp post-LGM bottleneck was suggested for the island population (97–99%

reduction; Fig. 3d–e) by the best-fit FSC2 models. Our SWP2 results did not contradict it despite

a large uncertainty towards the present (Fig. 3f). This was also supported by the small

contemporary effective population size of Ishikari Plain in Hokkaido (including Tomakomai) (N_e

= 4,840) calculated from the census population size estimates around the early nineteenth

century, which well corroborated the results of the FSC2.

SDMs

The best-fit model (response curve 'LQH' + regularization multiplier 2.0, AICc = 515.86,

Akaike model weight = 0.77, the Area Under the Curve (AUC) = 95.2%) accurately predicted

the present breeding ranges of Swinhoe's Rail (Fig. 4a–b, S21). During historical periods, some

areas did not resemble any climate in the modeling domain of this species, indicated by the

negative values in the MESS analyses (dark gray-shaded grids in Fig. 4a, S22, see

Supplementary Methods). We restricted our interpretations to results robust to different

treatments (inclusion or removal) of negative MESS grids.

The island served as a relatively stable glacial refugium, albeit smaller in extent than the

continent. Continental habitats underwent drastic and repeated changes in habitat suitability and

availability since 0.5 Mya (Fig. 4d). Habitat quality remained consistently higher on the island

than on the continent throughout this period (Fig. S24). After the LGM, the island habitat

contracted, corresponding to both SWP2 and FSC2, whereas the continental habitat expanded,

only supporting the post-LGM demography of FSC2 (Fig. 4d, S22, S24). The temporal change in

the level of habitat isolation between the island and the continent well supported the times of

divergence and gene flow estimated by our genetic analyses. The 0.38–0.43 Mya interval showed

stable isolation without periods of extreme proximity between the two regions, corresponding to

the divergence period (Fig. 4e, S22, S25). The LGM was the most recent period of reduced

isolation, consistent with post-LGM gene flow. Overall trends in these measures were similar

between different treatments of negative MESS grids, although the amplitude of their changes

differed between each other (Supplementary Results and Discussion, Fig. S24–S25).

Discussion

Using an integrative phylogeography in Swinhoe's Rail, we found that the contemporary

continental population exhibited larger effective population size than the island population,

consistent with general expectations (Leroy et al. 2021). However, multiple lines of evidence

suggested that, historically, the island population was more stable and possibly larger than the

continent, and that gene flow occurred from the island to the continent. These findings suggest

that the island served as an important refugium for this species. In contrast, the direction of

population establishment at the time of divergence remains unclear. Some phylogenetic trees

supported bifurcating topologies at the population level, but individual-level analyses did not

support reciprocal monophyly, leaving colonization direction unresolved.

Gene flow and incomplete lineage sorting in the continent-island system

Incongruent phylogenetic patterns across analyses suggest a complex history of this

species in the continent-island system, involving two non-mutually exclusive mechanisms, ILS

and post-divergence gene flow. In recently diverged populations like Swinhoe's Rail, gene

genealogies may remain shared between two geographic populations (Twyford and Ennos 2012).

For example, higher uncertainties were involved in the inference of the ancestral divergence and

gene flow, including low posterior supports for bifurcation in the SNAPP tree and lack of

reciprocal monophyly in the individual-level trees. These may reflect ILS, low marker

resolution, or a combination of the two (Maddison and Knowles 2006), where ILS, derived from

ancestral genetic variation, can obscure ancient evolutionary events (Twyford and Ennos 2012).

Therefore, ancient gene flow inferred by FSC2 may be confounded with ILS, and thus its

presence remains inconclusive. Nevertheless, the absence of monophyly due to ILS does not

negate the existence of the two genetic clusters (i.e., continental and island populations),

especially given multiple lines of support, and ILS is commonly retained to various degrees

across populations and species (Maddison and Knowles 2006).

By contrast, recent gene flow is more readily distinguished from ILS (Twyford and

Ennos 2012), and multiple results are consistent with this notion, supporting the presence of

post-LGM gene flow from the island to the continent. First, the continental population showed

signatures of strong historical bottlenecks–e.g., low Tajima's D, FSC2 results, and SDM-inferred

habitat instability–reducing the likelihood of retained ancestral variation. This is because a strong

reduction in population size likely causes a fixation of genetic variants. Given the historically

stable and relatively large island population, the island population may have preserved ancient

genetic variations, causing ILS (see Supplementary Results & Discussion). Second, two-

dimensional-SFS-based approaches, such as FSC2, and TreeMix– both of which explicitly

account for ILS by estimating the level of genetic drift (Pickrell and Pritchard 2012; Murray et

al. 2025)–consistently inferred gene flow to the continent. Therefore, while the relative

contribution of gene flow versus ILS to the continental genetic variation remains unclear, our

findings at least support the presence of recent gene flow from the island to the continent.

Discrepancies in population size histories between FSC2 and SWP2 may also reflect this

gene flow. In demographic trajectory inference methods, such as SWP2, migration can inflate

effective population size estimates, even without true expansion (Nadachowska-Brzyska et al.

2022). Therefore, the genetic bottleneck inferred in FSC2 might have been concealed in SWP2

by gene flow to the continental population. Alternatively, our simplified FSC2 models may have

failed to capture complex fluctuating histories and rather showed averaged trends (long-term N_e,

Nadachowska-Brzyska et al. 2022), which may have exaggerated the impact of a bottleneck.

However, because a long-term N_e reflects a harmonic mean of a demographic trajectory

(Nadachowska-Brzyska et al. 2022), the effect of past bottlenecks on the continental genetic

variation should not be undervalued.

Population persistence through a continent-island metapopulation

Historically large island populations and gene flow from islands have rarely been expected in

continent-island systems. This is because small island size is typically assumed to limit

population size (Leroy et al. 2021), leading to low dispersal potential and resultant gene flow

(Poethke and Hovestadt 2001; Matthysen 2005). However, based on our FSC2 estimates,

ancestral island population retained a large N_e that was comparable to that of avian continental

populations (mean N_e = 362,456) (Leroy et al. 2021) and large tropical island populations, such

as New Guinean birds (mean N_e = 325,000) (Pujolar et al. 2022), whereas the N_e of the ancestral

continental population was as small as that of oceanic island populations (mean N_e = 94,944)

(Leroy et al. 2021). These differences may reflect contrasting habitat stability; our SDMs

suggested more stable conditions on the island throughout glacial cycles than on the continent

(Fig. 4d, S24), which could have allowed the persistence of a stable population and promoted

dispersal (Ronce et al. 2000), leading to gene flow to the continent. In a large population,

adaptive or ancient variation can be more likely to be preserved than in a small population where

weakly deleterious mutations accumulate (Leroy et al. 2021; Lai et al. 2019). Asymmetric gene

flow from such a large population may restore genetic variation and result in demographic and

evolutionary consequences for a small population (Fitzpatrick et al. 2016). Therefore, the island

population could have potentially contributed to the evolution and persistence of the continental

population in this species. However, it remains unknown to what extent gene flow has

contributed to the continental expansion, as the signature of gene flow was not found originating

from the entire continental population (Fig. 2c).

Aomori of Honshu Island was inferred to have been recolonized recently in the wave of

continental expansion, suggesting the possibility that the island population has also been

sustained by migrants from the continent. Honshu might have been occupied previously by the

present Hokkaido ("island") population during a glacial period, supported by 1) the potential

presence of suitable habitats on Honshu but not on Hokkaido during some glacial maxima

inferred by the SDM (Fig. S22), and 2) a high affinity of Kanto samples towards Hokkaido

breeding birds (Fig. 1b–c), given that wintering regions of short-distance seasonal migrants may

reflect the past breeding range (Zink and Gardner 2017). Indeed, the population contraction of

Swinhoe's Rail in Japan reflects not only the post-LGM environmental changes but also

anthropogenic factors, such as habitat loss and fragmentation due to land use changes (Kitazawa

et al. 2022). This was also supported by our SDMs that predicted present suitable areas larger

than currently recognized (Fig. 4a–b) and that the current Aomori habitat originated from an

abandoned rice paddy (Ministry of the Environment, 2023). Hence, continental immigration

resulting in re-establishment rather than gene flow may reflect the extinction of local island

populations under anthropogenic impacts.

Altogether, our findings suggest that Swinhoe's Rail has persisted as a dynamic

continent-island metapopulation, where the Japanese Archipelago was not merely a one-way sink

but played a role as pivotal as the continent, functioning as potentially a demographic and

genetic source. Following divergence during a period of prolonged geographic isolation (~0.5

Mya; Fig. 5a), spatiotemporal climatic heterogeneity likely led the island and the continent to

shift roles within the metapopulation: the island supported relatively a large, stable population

that retained ancient genetic variation, while the continent underwent drastic and repeated

demographic changes that resulted in smaller long-term population size (Fig. 5b–c). Source-sink

roles may have shifted over time via gene flow and population re-establishment, potentially

promoting persistence of the metapopulation in the evolutionary timescale (Fig. 5b–c). Our

findings were different from many previous studies that found islands to harbor small genetic

variation and N_e and act only as recipients of continental migrants. This is possibly because these

patterns were often observed in systems with strong phenotypic or genomic divergence on

islands (e.g., Wang et al. 2016; Delmore et al. 2020; Recuerda et al. 2021), where post-

divergence gene flow from islands may be prevented or overlooked. In contrast, studies have

reported that multiple island populations are maintained with gene flow on archipelagic or land-

bridge islands (Seki et al. 2007; Mapel et al. 2021), and our results may be an expansion of such

systems to the continent-island setting. Some Japanese endemic lineages have extended their

ranges to the continental Russian Far East (e.g., *Gallinago hardwickii*, Nechaev and Fujimaki

1998) or Korean Peninsula (e.g., *Cettia diphone*, Clement and Kirwan 2020). Gene flow from

Japanese lineages to their continental counterparts has also been suspected, yet without explicit

inferences of the underlying historical processes (*Locustella amnicola* and *L. fasicolata*,

Drovetski et al. 2015; *Lanius cristatus*, Aoki et al. 2021). Similar continent-island systems are

recognized in other regions (Bellemain et al. 2008; Garcia-R et al. 2017), suggesting the

universality of such a system across the globe.

A recent review emphasizes the importance of long-term population persistence in

dynamic metapopulations for promoting speciation and adaptive radiation (Harvey et al. 2019).

Our view also emphasizes the long-term consequences of metapopulation between continental

and island populations, where source and sink relationships dynamically change, differently from

the traditional perspective that focuses on the persistence of a metapopulation in the ecological

timescale (Hanski 1998). A metapopulation structure across the continent-island system can be

perceived as a transitional state for insular populations to undergo reverse colonization, thereby

promoting long-term persistence and adaptation to the continental environment. Therefore, we

highlight that our insights obtained from the population-level study may shed light on the origin

of reverse colonization at the species level. Future research needs to incorporate more

polymorphic sites to resolve ILS and gene flow and estimate the number, direction, and intensity

of past migration, to understand the relative impact of these elements on the present genetic

variation. This would disentangle the evolutionary importance of how a continent-island

metapopulation may facilitate expansion, adaptation, and diversification on the continent,

extending our microevolutionary idea to a macroevolutionary framework.

Conclusion

Historical demography is fundamental to understanding a determinant of population genetic structure that is significantly linked with subsequent speciation and diversification (Harvey et al. 2019). In this study, we reconstructed the demographic history of a rail species in a continent-island system. We found strong evidence of asymmetrical gene flow from the Japanese Archipelago to the continental population, which may have influenced the continental genetic variation and demography. Furthermore, our results suggested that this species has persisted as a metapopulation spanning both the islands and the continent. These findings challenge conventional views of continent-island dynamics, particularly given that breeding populations of this species in Japan were only recently documented (Senzaki et al. 2020). To fully understand the processes that have given rise to the present biodiversity, evolutionary studies on the current and apparent species distribution are insufficient. Rather, reconstructing the long-term, spatially explicit population histories of organisms can shed light on the previously unfilled gap between micro- and macroevolutionary concepts and facilitate our understanding of global biodiversity patterns.

References

Alvo R., and M. Robert. 1999. COSEWIC status report on the yellow rail *Coturnicops noveboracensis* in Canada. Committee on the Status of Endangered Wildlife in Canada, Ottawa.

Anisimova, V., Y. Anisimov, M. Bastardot, and W. Heim. 2019. First observation of Swinhoe's Rail *Coturnicops exquisitus* in the delta of Selenga river. Baikal Zoological Journal 2:112–113.

Aoki, D., G. Kinoshita, A. P. Kryukov, I. Nishiumi, S.-I. Lee, and H. Suzuki. 2018. Quaternary-related genetic differentiation and parallel population dynamics of the Eurasian Jay (*Garrulus glandarius*) in the circum-Japan Sea region. J. Ornithol. 159:1087–1097.

Aoki, D., H. Sakamoto, M. Kitazawa, A. P. Kryukov, and M. Takagi. 2021. Migration-tracking integrated phylogeography supports long-distance dispersal-driven divergence for a migratory bird species in the Japanese archipelago. Ecol. Evol. 11:6066–6079.

Bank C., R. Bűrger, and J. Hermisson. (2012) The limits to parapatric speciation: Dobzhansky-Muller incompatibilities in a continent-island model. Genetics, 191:845–863.

Barton N. H. 1996. Natural selection and random genetic drift as causes of evolution on islands. Phil. Trans. R. Soc. Lond. B, 351:785–795.

Bellemain, E., E. Bermingham, and R. E. Ricklefs. 2008. The dynamic evolutionary history of the bananaquit (*Coereba flaveola*) in the Caribbean revealed by a multigene analysis. BMC Evol. Biol. 8:240.

Bellemain, E., and R. E. Ricklefs. 2008. Are islands the end of the colonization road? Trends Ecol. Evol. 23:461–468.

BirdLife International, and Handbook of the Birds of the World. 2024. Bird species distribution

maps of the world. Version 2024.2. Available at

http://datazone.birdlife.org/species/requestdis.

Bouckaert, R., T. G. Vaughan, J. Barido-Sottani, S. Duchêne, M. Fourment, A. Gavryushkina, J.

Heled, G. Jones, D. Kühnert, N. De Maio, M. Matschiner, F. K. Mendes, N. F. Müller, H. A.

Ogilvie, L. du Plessis, A. Popinga, A. Rambaut, D. Rasmussen, I. Siveroni, M. A. Suchard,

C.-H. Wu, D. Xie, C. Zhang, T. Stadler, and A. J. Drummond. 2019. BEAST 2.5: An

advanced software platform for Bayesian evolutionary analysis. PLoS Comput. Biol.

15:e1006650.

Bryant, D., R. Bouckaert, J. Felsenstein, N. A. Rosenberg, and A. RoyChoudhury. 2012.

Inferring species trees directly from biallelic genetic markers: bypassing gene trees in a full

coalescent analysis. Mol. Biol. Evol. 29:1917–1932.

Bryant, D., and V. Moulton. 2004. Neighbor-net: an agglomerative method for the construction

of phylogenetic networks. Mol. Biol. Evol. 21:255–265.

Cerca, J., D. D. Cotoras, V. C. Bieker, R. De-Kayne, P. Vargas, M. Fernández-Mazuecos, J.

López-Delgado, O. White, M. Stervander, A. J. Geneva, J. E. Guevara Andino, J. I. Meier,

L. Roeble, B. Brée, J. Patiño, J. M. Guayasamin, M. de L. Torres, H. Valdebenito, M. D. R.

Castañeda, J. A. Chaves, P. J. Díaz, L. Valente, M. L. Knope, J. P. Price, L. H. Rieseberg, B.

G. Baldwin, B. C. Emerson, G. Rivas-Torres, R. Gillespie, and M. D. Martin. 2023.

Evolutionary genomics of oceanic island radiations. Trends Ecol. Evol. 38:631–642.

Clement, P. and G. M. Kirwan. 2020. Japanese Bush Warbler (*Horornis diphone*), version 1.0. In

del Hoyo, J., A. Elliott, J. Sargatal, D. A. Christie, and E. de Juana eds. Birds of the World.

Cornell Lab of Ornithology, Ithaca, NY, USA.

Delmore, K., J. C. Illera, J. Pérez-Tris, G. Segelbacher, J. S. Lugo Ramos, G. Durieux, J.

Ishigohoka, and M. Liedvogel. 2020. The evolutionary history and genomics of European

blackcap migration. Elife 9:e54462.

Drovetski, S. V., G. Semenov, Y. A. Red'kin, V. N. Sotnikov, I. V. Fadeev, and E. A. Koblik.

2015. Effects of asymmetric nuclear introgression, introgressive mitochondrial sweep, and

purifying selection on phylogenetic reconstruction and divergence estimates in the pacific

clade of *Locustella* warblers. PLoS One. 10:e0122590.

Ellegren, H., and N. Galtier. 2016. Determinants of genetic diversity. Nat. Rev. Genet. 17:422–

433.

Excoffier, L., N. Marchi, D. A. Marques, R. Matthey-Doret, A. Gouy, and V. C. Sousa. 2021.

fastsimcoal2: demographic inference under complex evolutionary scenarios. Bioinformatics

37:4882–4885.

Feder, J. L., S. P. Egan, and P. Nosil. 2012. The genomics of speciation-with-gene-flow. Trends

Genet. 28:342–350.

Fitak, R. R. 2021. OptM: estimating the optimal number of migration edges on population trees

using Treemix. Biol Methods Protoc 6:bpab017.

Fitzpatrick, S. W., J. C. Gerberich, L. M. Angeloni, L. L. Bailey, E. D. Broder, J. Torres-

Dowdall, C. A. Handelsman, A. López-Sepulcre, D. N. Reznick, C. K. Ghalambor, and W.

Chris Funk. 2016. Gene flow from an adaptively divergent source causes rescue through

genetic and demographic factors in two wild populations of Trinidadian guppies. Evol.

Appl. 9:879–891.

Fox, E. A., A. E. Wright, M. Fumagalli, and F. G. Vieira. 2019. ngsLD: evaluating linkage

disequilibrium using genotype likelihoods. Bioinformatics 35:3855–3856.

Frankham, R. 1995. Effective population size/adult population size ratios in wildlife: a review.

Genet. Res. 66:95–107.

Frankham, R. 2005. Genetics and extinction. Biol. Conserv. 126:131–140.

Garcia-R, J. C., L. Joseph, G. Adcock, J. Reid, and S. A. Trewick. 2017. Interisland gene flow

among populations of the buff-banded rail (Aves: Rallidae) and its implications for insular

endemism in Oceania. J. Avian Biol. 48:679–690.

Garcia-R, J. C., E. M. Lemmon, A. R. Lemmon, and N. French. 2020. Phylogenomic

reconstruction sheds light on new relationships and timescale of rails (Aves: Rallidae)

evolution. Diversity 12:70.

Garcia-R, J. C., and N. J. Matzke. 2021. Trait-dependent dispersal in rails (Aves: Rallidae):

Historical biogeography of a cosmopolitan bird clade. Mol. Phylogenet. Evol. 159:107106.

Gattepaille, L. M., M. Jakobsson, and M. G. B. Blum. 2013. Inferring population size changes

with sequence and SNP data: lessons from human bottlenecks. Heredity 110:409–419.

Hanghøj K., I. Moltke, P. A. Andersen, A. Manica, T. S. Korneliussen. 2019. Fast and accurate

relatedness estimation from high-throughput sequencing data in the presence of inbreeding.

GigaScience 8:giz034

Hanski, I. 1998. Metapopulation dynamics. Nature 396:41–49.

Harvey, M. G., S. Singhal, and D. L. Rabosky. 2019. Beyond reproductive isolation:

demographic controls on the speciation process. Annu. Rev. Ecol. Evol. Syst. 50:75–95.

Heim, W., D. Trense, A. Heim, and J. Kamp. 2019. Discovery of a new breeding population of

the vulnerable Swinhoe's Rail *Coturnicops exquisitus* confirmed by genetic analysis. Bird

Conserv. Int. 29:454–462.

Hufbauer, R. A., M. Szűcs, E. Kasyon, C. Youngberg, M. J. Koontz, C. Richards, T. Tuff and B.

A. Melbourne. 2015. Three types of rescue can avert extinction in a changing environment. Proc. Natl. Acad. Sci. U.S.A. 112:10557–10562.

Jønsson, K. A., and B. G. Holt. 2015. Islands contribute disproportionately high amounts of evolutionary diversity in passerine birds. Nat. Commun. 6:8538.

Kass, J. M., R. Muscarella, P. J. Galante, C. L. Bohl, G. E. Pinilla-Buitrago, R. A. Boria, M. Soley-Guardia, and R. P. Anderson. 2021. ENMeval 2.0: Redesigned for customizable and reproducible modeling of species' niches and distributions. Methods Ecol. Evol. 12:1602–1608.

Kitazawa, M., Y. Yamaura, M. Senzaki, M. Hanioka, H. Ohashi, M. Oguro, T. Matsui, and F. Nakamura. 2022. Quantifying the impacts of 166 years of land cover change on lowland bird communities. Proc. Biol. Sci. 289:20220338.

Korneliussen, T. S., A. Albrechtsen, and R. Nielsen. 2014. ANGSD: Analysis of Next Generation Sequencing Data. BMC Bioinformatics 15:356.

Kozlov, A. M., D. Darriba, T. Flouri, B. Morel, and A. Stamatakis. 2019. RAxML-NG: a fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. Bioinformatics 35:4453–4455.

Krapp, M., R. M. Beyer, S. L. Edmundson, P. J. Valdes, and A. Manica. 2021. A statistics-based reconstruction of high-resolution global terrestrial climate for the last 800,000 years. Sci Data 8:228.

Lai, Y., C. K. L. Yeung, K. E. Omland, E. Pang, Y. Hao, B. Liao, H. Cao, B. Zhang, C. Yeh, C. Hung, H. Hung, M. Yang, W. Liang, Y. Hsu, C. Yao, L. Dong, K. Lin, & S. Li. 2019. Standing genetic variation as the predominant source for adaptation of a songbird, Proc. Natl. Acad. Sci. U.S.A. 116:2152–2157.

Leonardi, M., E. Y. Hallett, R. Beyer, M. Krapp, and A. Manica. 2023. pastclim 1.2: an R

package to easily access and use paleoclimatic reconstructions. Ecography 2023:e06481.

Leroy, T., M. Rousselle, M.-K. Tilak, A. E. Caizergues, C. Scornavacca, M. Recuerda, J. Fuchs,

J. C. Illera, D. H. De Swardt, G. Blanco, C. Thébaud, B. Milá, and B. Nabholz. 2021. Island

songbirds as windows into evolution in small populations. Curr. Biol. 31:1303–1310.e4.

Lisiecki, L. E., and M. E. Raymo. 2005. A Pliocene-Pleistocene stack of 57 globally distributed

benthic δ^{18}O records. Paleoceanography 20:PA1003. American Geophysical Union (AGU).

Liu, X., and Y.-X. Fu. 2020. Stairway Plot 2: demographic history inference with folded SNP

frequency spectra. Genome Biol. 21:280.

Lomolino, M. V., B. R. Riddle, and R. J. Whittaker. 2017. Biogeography: Biological Diversity

across Space and Time. 5th Edition. Sinauer Associates, Maryland.

Maddison W. P., and L. L. Knowles. 2006. Inferring phylogeny despite incomplete lineage

sorting. System. Biol. 55:21–30.

Mapel, X. M., E. F. Gyllenhaal, T. H. Modak, L. H. DeCicco, A. Naikatini, R. B. Utzurrum, J. O.

Seamon, A. Cibois, J.-C. Thibault, M. D. Sorenson, R. G. Moyle, L. N. Barrow, and M. J.

Andersen. 2021. Inter- and intra-archipelago dynamics of population structure and gene

flow in a Polynesian bird. Mol. Phylogenet. Evol. 156:107034.

Matthysen, E. 2005. Density-dependent dispersal in birds and mammals. Ecography 28:403–416.

Meisner, J., and A. Albrechtsen. 2018. Inferring population structure and admixture proportions

in low-depth NGS data. Genetics 210:719–731.

Ministry of the Environment, Government of Japan. 2023. Hotokenuma. Available at

https://www.env.go.jp/nature/ramsar/conv/ramsarleaflet/14_Hotokenuma.pdf. Accessed on

June 27th, 2025.

Miya, A., S. Sannohe, J. Ebina, and H. Sekishita. 2005. A survey report on Yellow Rail

(*Coturnicops noveboracensis exquisitus*) observed during the breeding season at

Hotokenuma Reclaimed Land in Misawa, Aomori Prefecture. Jap. J. Ornithol. 54:104–107.

Murray, C. S., M. Karram, D. J. Bass, M. Doceti, D. Becker, J. C. B. Nunez, A. Ratan, A. O.

Bergland. 2025. Trans-specific polymorphisms between cryptic *Daphnia* species affect

fitness and behavior. 34:e17632

Nadachowska-Brzyska, K., M. Konczal, and W. Babik. 2022. Navigating the temporal

continuum of effective population size. Methods Ecol. Evol. 13:22–41.

Nam, K., C. Mugal, B. Nabholz, H. Schielzeth, J. B. W. Wolf, N. Backström, A. Künstner, C. N.

Balakrishnan, A. Heger, C. P. Ponting, D. F. Clayton, and H. Ellegren. 2010. Molecular

evolution of genes in avian genomes. Genome Biol. 11:R68.

Nechaev, V. A., and Y. Fujimaki. 1998. Present status of the Latham's snipe *Gallinago*

hardwickii on Sakhalin. Res. Bull. Obihiro. Univ. 21:61–65.

Patiño, J., M. Carine, P. Mardulyn, N. Devos, R. G. Mateo, J. M. González-Mancebo, A. J.

Shaw, and A. Vanderpoorten. 2015. Approximate Bayesian computation reveals the crucial

role of oceanic islands for the assembly of continental biodiversity. Syst. Biol. 64:579–589.

Poethke H. J., and T. Hovestadt. 2001. Evolution of density- and patch-size-dependent dispersal

rates. Proc. R. Soc. Lond. B. 269:637–645.

Phillips, S. J., R. P. Anderson, and R. E. Schapire. 2006. Maximum entropy modeling of species

geographic distributions. Ecol. Modell. 190:231–259.

Pickrell, J. K., and J. K. Pritchard. 2012. Inference of population splits and mixtures from

genome-wide allele frequency data. PLoS Genet. 8:e1002967.

Pujolar, J. M., M. P. K. Blom, A. H. Reeve, J. D. Kennedy, P. Z. Marki, T. S. Korneliussen, B.

G. Freeman, K. Sam, E. Linck, T. Haryoko, B. Iova, B. Koane, G. Maiah, L. Paul, M. Irestedt, and K. A. Jønsson. 2022. The formation of avian montane diversity across barriers and along elevational gradients. Nat. Commun. 13:268.

Recuerda, M., J. C. Illera, G. Blanco, R. Zardoya, and B. Milá. 2021. Sequential colonization of oceanic archipelagos led to a species-level radiation in the common chaffinch complex (Aves: *Fringilla coelebs*). Mol. Phylogenet. Evol. 164:107291.

Renaud, G. 2017. glactools: a command-line toolset for the management of genotype likelihoods and allele counts. Bioinformatics 34:1398–1400.

Ronce, O., F. Perret, and I. Olivieri. 2000. Evolutionarily stable dispersal rates do not always increase with local extinction rates. Am. Nat. 155:485‑96.

Seki, S.-I., H. Takano, K. Kawakami, N. Kotaka, A. Endo, and K. Takehara. 2007. Distribution and genetic structure of the Japanese wood pigeon (*Columba janthina*) endemic to the islands of East Asia. Conserv. Genet. 8:1109–1121.

Senzaki, M., M. Kitazawa, T. Sadakuni, and M. Takahashi. 2020. Breeding evidence of the vulnerable Swinhoe's Rail (*Coturnicops exquisitus*) in Japan. Wilson J. Ornithol. 132:711–717.

Skotte, L., T. S. Korneliussen, and A. Albrechtsen. 2013. Estimating individual admixture proportions from next generation sequencing data. Genetics 195:693–702.

Soraggi, S., C. Wiuf, and A. Albrechtsen. 2018. Powerful inference with the D-statistic on low-coverage whole-genome data. G3 8:551–566.

Spratt, R. M., and L. E. Lisiecki. 2016. A Late Pleistocene sea level stack. Clim. Past 12:1079–1092.

Stange, M., M. R. Sánchez-Villagra, W. Salzburger, and M. Matschiner. 2018. Bayesian

divergence-time estimation with genome-wide single-nucleotide polymorphism data of sea

catfishes (Ariidae) supports Miocene closure of the Panamanian Isthmus. Syst. Biol.

67:681–699.

Suyama, Y., and Y. Matsuki. 2015. MIG-seq: an effective PCR-based method for genome-wide

single-nucleotide polymorphism genotyping using the next-generation sequencing platform.

Sci. Rep. 5:16963.

Tajima, F. 1983. Evolutionary relationship of DNA sequences in finite populations. Genetics

105:437–460.

Taylor, B., and C. J. Sharpe. 2020. Swinhoe's Rail (*Coturnicops exquisitus*), version 1.0. *in* J. del

Hoyo, A. Elliott, J. Sargatal, D. A. Christie, and E. de Juana, eds. Birds of the World.

Cornell Lab of Ornithology, Ithaca, NY.

Twyford, A. D., and R. A. Ennos. 2012. Next-generation hybridization and introgression.

Heredity 108:179–189.

Vamosi, J. C., W. S. Armbruster, and S. S. Renner. 2014. Evolutionary ecology of specialization:

insights from phylogenetic analysis. Proc. R. Soc. B 281:20142004.

Vieira, F. G., F. Lassalle, T. S. Korneliussen, and M. Fumagalli. 2015. Improving the estimation

of genetic distances from Next-Generation Sequencing data. Biol. J. Linnean Soc. 117:139–

149.

Wang, N., B. Liang, J. Wang, C.-F. Yeh, Y. Liu, Y. Liu, W. Liang, C.-T. Yao, and S.-H. Li.

2016. Incipient speciation with gene flow on a continental island: Species delimitation of the

Hainan Hwamei (*Leucodioptron canorum owstoni*, Passeriformes, Aves). Mol. Phylogenet.

Evol. 102:62–73.

Waples R. K., A. Albrechtsen, and I. Motke. 2019. Allele frequency-free inference of close

familial relationships from genotypes or low-depth sequencing data. Mol. Ecol. 28:35–48

Warren, B. H., D. Simberloff, R. E. Ricklefs, R. Aguilée, F. L. Condamine, D. Gravel, H.

Morlon, N. Mouquet, J. Rosindell, J. Casquet, E. Conti, J. Cornuault, J. M. Fernández-

Palacios, T. Hengl, S. J. Norder, K. F. Rijsdijk, I. Sanmartín, D. Strasberg, K. A. Triantis, L.

M. Valente, R. J. Whittaker, R. G. Gillespie, B. C. Emerson, and C. Thébaud. 2015. Islands

as model systems in ecology and evolution: prospects fifty years after MacArthur-Wilson.

Ecol. Lett. 18:200–217.

Watterson, G. A. 1975. On the number of segregating sites in genetical models without

recombination. Theor. Popul. Biol. 7:256–276.

Weigelt P., W. Jetz, and H. Kreft. 2013. Bioclimatic and physical characterization of the world's

islands. Proc. Natl. Acad. Sci. U.S.A. 110:15307–15312.

Weir J. T., and D. Schluter. 2004. Ice sheets promote speciation in boreal birds. Proc. R. Soc.

Lond. B. 271:1881–1887.

Weir, J. T., and D. Schluter. 2008. Calibrating the avian molecular clock. Mol. Ecol. 17:2321–

2328.

Whittaker, R. J., and J. M. Fernández-Palacios. 2007. Island Biogeography: Ecology, Evolution,

and Conservation. Oxford University Press, Oxford.

Whittaker, R. J., J. M. Fernández-Palacios, T. J. Matthews, M. K. Borregaard, and K. A. Triantis.

2017. Island biogeography: Taking the long view of nature's laboratories. Science

357:eaam8326.

Yeung, C. K. L., P.-W. Tsai, R. T. Chesser, R.-C. Lin, C.-T. Yao, X.-H. Tian, and S.-H. Li.

2011. Testing founder effect speciation: divergence population genetics of the spoonbills

Platalea regia and *Pl. minor* (Threskiornithidae, Aves). Mol. Biol. Evol. 28:473–482.

Zink, R. M., and A. S. Gardner. 2017. Glaciation as a migratory switch. Sci Adv 3:e1603133.

Data Availability Statement

Scripts and data for analysis are available on Dryad: 10.5061/dryad.37pvmcvs1 (private link:

https://datadryad.org/stash/share/2PRy64eM1UIjsjCJlNLx4B9ownPLtYVob4aFp_1MXys). Raw

sequence reads are deposited to Sequence Read Archives (see Table S1). Settings and procedures

for these processes are fully described in the main and supplementary texts and the original

scripts.

Tables

Table 1 Dataset and the number of sites used in each analysis

Analyses	Linkage	Site type	Outgroup	Populations	# Sites	# Samples	Notes
PCA, ADMIXTURE	unlinked	V	Non	4 breeding[1], 1 wintering[2]	1,147	47	-
NeighborNet (ngsDist)	linked	V	Laja/Atro	4 breeding	1,958	32	
RAxML, D-statistic	linked	V/I	Laja	4 breeding	108,095	31	-
TreeMix	linked	V	Laja	4 breeding	1,669 (683)	31	Population-specific private alleles were removed by glactools. The number of sites inside the bracket is for the individual-level tree.
SNAPP	unlinked	V	Laja	4 breeding	206	16	The top three/four samples with the lowest missing allele proportions in each population were selected. Excess heterozygous sites were removed by BCFtools
Diversity indices, FSC2, SWP2	linked	V/I	Non	2 genetic[3]	130,368	30	-

808 Abbreviation: PCA, principal component analysis; FSC2, fastsimcoal2; SWP2, Stairway Plot 2;

809 V, variants; V/I, both variants and invariants; Laja, *Laterallus jamaicensis*; Atro, *Atlantisia*

810 *rogersi*; [1] Tomakomai, Kushiro, Russia, and Aomori; [2] Kanto; [3] Island, and Continental.

Figures

Figure legends

Figure 1 A map of sampling localities of Swinhoe's rails (a), and their population genetic

structure inferred by (b) a principal component analysis (PCA) and (c) an ADMIXTURE

analysis conducted using genotype likelihoods of 1,147 single nucleotide polymorphisms

(SNPs). (a, b) The colors of dots and inertia ellipses represent different regional populations.

Dark-grayed regions are the expert range map of this species in BirdLife International and

Handbook of the Birds of the World (2024), which also includes previously known ranges. (c)

Genetic assignment of individuals to each number of genetic clusters (K = 2 to 5) is shown.

Colors do not correspond to the sampling localities. Lambert Azimuthal Equal Area projection

was used for the map. The inset photo by Marc Bastardot.

Figure 2 Phylogenetic trees of Swinhoe's Rail reconstructed using different methods and genetic

markers. (a) A mitochondrial phylogenetic tree (mtDNA tree) using 772 bp of the *Cytb* gene.

Posterior probabilities >0.9 were indicated by black dots on nodes. Numbers at the nodes

indicate median divergence time estimates with 95% highest posterior density within brackets

(bold) and posterior probabilities (italic). (b) A maximum likelihood (ML) tree (RAxML tree)

was reconstructed using 108,095 sites including both variant and invariant sites. Only nodes with

bootstrap values >70 are annotated with black circles. (c) A population-level tree reconstructed

by TreeMix using the allele frequency of 1,669 polymorphic loci, excluding population-specific

private alleles. Branch lengths indicate drift parameters, the strength of drift specific to each

branch. The inset indicates an individual-level tree reconstructed using non-private 683

polymorphic loci. The colored arrows indicate the estimated gene flow. (d) Posterior population-

level trees reconstructed by SNAPP analyses using 206 single nucleotide polymorphisms (SNPs)

with no missing data. Colors of trees indicate percentages of each posterior tree in the credible

set obtained by the 'TreeSetAnalyser' program. Values at each node indicate their divergence

time estimates with credible intervals within brackets. Two different divergence time estimates

were obtained for different calibration constraints. Values beneath each node indicate posterior

probabilities.

Figure 3 Demographic inference of the two genetic populations of Swinhoe's Rail based on site

frequency spectra, using (a, b) the summary statistics, (c–e) fastsimcoal2 (FSC2), and (f)

Stairway Plot 2. Fixed values in each model are italicized. Effective population size and time are

scaled by mutation rate (1.91e-09 bp/generation) and generation time (1 year). Effective

population size shows the number of diploid individuals. (a, b) Comparison of scaffold-level

statistics between the island and continental populations for (a) Watterson's theta and (b)

Tajima's *D*. Statistically significant differences in the values of each index were annotated with

asterisks (* $p < 0.05$). (c) Generalized schematic representation of the five models tested.

Parameters with uppercase represent the effective population size we modeled. Hatched and

dotted arrows represent different migration matrices within the model. (d, e) The two best-fit

FSC2 models with estimated parameters. Numbers by or on branches and arrows represent

effective population size and the number of migrants per year, respectively. The size of each box

is not scaled to the corresponding population size. Divergence time is shown in million years. (f)

Changes in effective population size (in millions) over time (in million years) are shown for the

island and continental populations. Bold lines and thin ribbons around them show median and

95% confidence intervals, respectively. The gray range indicates the approximate period of

population divergence.

Figure 4 A predicted present distribution of Swinhoe's Rail (a) by the species distribution model

using its present occurrence records (b). (a) Color gradients from indigo to yellow represent

environmental suitability for rails. Regions with dark gray shades are where model extrapolation

was restricted due to the absence of the climate in the modeling domain (grids with negative

MESS values). (b) The red grids indicate the used occurrence records at the resolution of the

environmental layers, and gray areas indicate the expert range map of this species. (c) A

temporal plot for a proxy for paleoclimatic temperature (δ^{18}O; lower values indicate higher

temperatures) (Lisiecki and Raymo 2005; Spratt and Lisiecki 2016). (d–e) Historical changes in

the total suitability of the habitat in the continent (blue) and the island (yellow) (d) and the

suitability-weighted centroid distance between the two (e). Black solid and dotted lines indicate

the overall median and standard deviation of the distance, and a red bar indicates the period of

the smallest amplitude in the temporal change. Grids covered by negative MESS values were

also included in the calculation. Gray bars correspond to the estimated period of the population

split (left) and gene flow (LGM, right). Lambert Azimuthal Equal Area projection was used for

maps.

Figure 5 Schematic representation of our continent-island metapopulation scenario of Swinhoe's

Rail; geographic isolation occurred around 0.5Mya (a), and source-sink switching dynamics

occurred in the continent-island system between the glacial (b) and interglacial period (c). Gene

flow (a dotted arrow) and re-establishment or expansion of population (solid arrows) may have

facilitated persistence and evolution of the metapopulation. Filled circles represent stable (solid)

and unstable (hatched) populations.

Figures

Figure 1

Figure 2

883   **Figure 3**

Figure 4

888   **Figure 5**