Context dependent multisensory integration: Mechanosensation depends on luminance for robust performance

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Abstract   Multisensory, goal-directed behaviors are ubiquitous and essential in animals across taxa. Individual sensory modalities may be significantly tuned by environmental context, and yet they must continue to combine and produce multisensory behaviors. This necessitates either behavioral robustness or goal-relevant adaptation in sensorimotor systems. In the case of hover-feeding hawkmoths of the species *Manduca sexta*, proboscis mechanosensation and vision are known to combine linearly in tracking of moving flowers. This tracking behavior is affected by light-levels, and visual compensation is necessary for robust flower-tracking. Whether the mechanosensory response also adjusts to light-level, in order to produce linear combination across luminance contexts, is not known. We applied control theoretic analyses to study integration of mechanosensory and visual information in flower tracking hawkmoths in the context of high luminance. This was performed using a two-part robotic flower system. Under the conditions of high luminance, we verified the linear combination of mechanosensory and visual systems, and the complementary modulation of these systems. We show that both systems tune their frequency response with changing environmental light level,while keeping their combined error low at relevant frequencies. Thus, the animal adapts to changing conditions, across modalities, even when only one of the modalities (vision) is directly affected. This cross-talk between sensorimotor systems is constrained , and possibly driven, by performance requirements.

Introduction

Animal behavior often requires the integration of multiple sensory cues in a task-specific manner. This necessitates sensory-to-motor transformations which may simultaneously depend on the context and activity of other sensory modalities. Multisensory behaviors become especially challenging when a task must be performed across contexts (such as day and night) because the context can affect sensory processing.If the organism maintains performance across contexts which necessarily change one modality, then its other modalities must either be robust to these changes or also adjust when the context changes. Many organisms modulate one sensory modality when the reliability or salience of another modality changes. But how do organisms that behaviorally integrate multiple cues with different weights across frequencies tune the overall dynamic frequency as contexts change?

Control theory and system identification can help us identify how multiple modalities have changed across their dynamic range and whether their integration is robust or adaptive. Control theoretic experiments have revealed principles of multisensory integration, especially in reference-tracking tasks, such as cockroach wall-following (*Cowan et al.* (*2006*)), knifefish refuge-tracking (*Sutton et al.* (*2016*)), blowfly gaze stabilization (*Hengstenberg* (*1988*),*Hengstenberg et al.* (*1986*)), and hawkmoth flower-tracking (*Sprayberry and Daniel* (*2007*)). The electric knifefish, *eigenmannia virescens*, combines visual and electrosensory cues to track refuges, linearly superimposing the two modalities but adjusting their weights based on salience (*Sutton et al.* (*2016*)). Visual and tactile integration studies in human posture control (*Oie et al.* (*2002*)) and height-estimation (*Ernst and Banks* (*2002*)) have shown evidence for re-weighting of modalities depending on their salience. These are statistically-optimal strategies for the specific cues considered, which therefore have amplitude-dependent gains. However, they do not examine the strategy for combining cues across their dynamic range of frequencies. While the more general case of nonlinear integration is important (*Huston and Krapp* (*2009*),*Cellini and Mongeau* (*2022*)). in some animals the combination of cues is effectively linear, such as for hawkmoth flower-tracking (*Roth et al.* (*2016*)) and in *Drosophila* wing motor output (*Rauscher and Fox* (*2021*)). The respective contributions to behavior of the different sensory modalities in these studies was investigated in a single behavioral and environmental context. However, sensory systems are tuned to their environment and have characteristic timescales *Roth et al.* (*2016*); *Dahake et al.* (*2018*). Two strategies are possible for maintaining this performance across contexts. A robust combination means that one modality does not necessarily have to change in conjuction with the other to maintain behavioral performance across the dynamic range. An adaptive strategy means that there must be concomitant changes in one modality when the other adjusts for context.

Manduca sexta is a crepuscular hawkmoth, capable of hover feeding in mid-air while tracking flower movement. Flower-tracking behavior involves closed-loop flight control, with frequency-response dynamics that depend on sensory modalities. Using dummy flowers and controlled sensory stimuli, we know that these flower-tracking dynamics are linear (*Sprayberry and Daniel* (*2007*),*Sponberg et al.* (*2015*), *Dahake et al.* (*2018*),*Stöckl et al.* (*2017*)), and this tracking is performed using visual and proboscis mechanosensory information. Visual and antennal mechanosensory contributions to flower-tracking are widely separated in frequencies (*Dahake et al.* (*2018*)). Proboscis mechanosensation, however, has significant overlap with visual flower-tracking response. *Roth et al.* (*2016*) used robotic flowers to infer a template for multisensory integration of visual and proboscis mechanosensory inputs, as a linear combination of the two systems. They also found a frequency dependent separation of the role of each sensory system, with the visual system sensitive to relatively high frequencies. These results were tested for a single, low luminance level. Decrease in luminance results in an increase in time-lag of the moth's tracking of a moving flower (*Sponberg et al.* (*2015*)). Therefore, a change in the visual context has an effect on the moth's behavior, while also directly affecting at least one of the two sensory systems used in this behavior. Animal locomotor behavior is often performed in changing environmental conditions, such as light-level, terrain roughness or wind-speed, which are very different contexts. Changing context can result in a change in the gain (magnitude) or the sensitivity of a sensory response. Context dependence of neural systems has been demonstrated multiple times, such as by *Maimon et al.* (*2010*), *Olberg* (*1983*), *Sponberg et al.* (*2015*),*Stöckl et al.* (*2016*) and *Theobald et al.* (*2010*). In *Maimon et al.* (*2010*), changing the context, sometime termed the behavioral state, of a fly from stationary or flapping changed the gain of motion-processing neurons. In addition to internal state, informational cues from the environment, such as different odors or light-levels, help determine the context for a behavior and change the tuning of sensory systems. In moths, this is seen in the gating of flip-flop activity by pheromones to drive cast-and-surge behavior (*Olberg* (*1983*)), and luminance-dependent adaptations for flower-tracking and optomotor correction (*Sponberg et al.* (*2015*),*Stöckl et al.* (*2016*),*Theobald et al.* (*2010*)).

It is not surprising that the gain and phase response of the visual system depends on back-

ground. Here, we test whether overall luminance (i.e. the changing context) also affects the hawk-moths' mechanosensory response to a moving flower separately from its impacts on vision and how the two cues are combined in these different settings. To maintain tracking performance across the range of frequencies at which the moth track natural, wind-blown flowers *Sponberg et al.* (*2015*), the animal may transform mechanosensory cues robustly, in which case we would expect to see the gain response of the mechanosensory pathway remain constant but to not significantly degrade performance. Alternatively, the mechanosensory system itself may also be luminance-dependent in a way that it adapts sensorimotor control to maintain performance in the new context. In either case, both visual and mechanosensory systems depend on luminance, their linear combination across light-levels may persist or may switch to a non-linear integration. Finally, these changes may degrade behavioral performance, or the moths may track flowers equally well or better despite the frequency-dependent modulation of one or both sensory modalities.

Results

In multisensory behaviors, probing a system's response to conflicting sensory information can separate the sensorimotor transform for each cue and show how they combine. We used sensory-conflict and coherent-motion stimuli, as described in Roth et al (*Roth et al.* (*2016*))[see Methods and Materials], but now across contexts. Briefly, our setup consists of a two-part robotic flower, with a facade (V) and a nectary (M) for visual and proboscis mechanosensory inputs, respectively (Figure 1a). In order to test for context dependence, we performed these experiments at the flower illumination value of 300 lux (early dusk) instead of the previously used value of 0.3 lux (starlit night). Based on the observations at 0.3 lux, we hypothesized that at 300 lux the two modalities combine linearly, as per the control diagram in Figure 1a. For system-identification, we provided sum-of-sines stimuli (see Methods and Materials) in M-moving (facade stationary), V-moving (nectary stationary) and coherent motion (both moving together) cases (Figure 1b). Moth trajectories in response to these motion stimuli were recorded. In the frequency domain, like in previous studies, we found that Fourier amplitude peaks are present selectively at each of the driving-frequencies, up to at least 3 Hz for all 12 individuals and all 3 stimulus conditions (Figure 1b). Using combinations of these three experiments we can isolate the dynamics of the mechanosensory system separate from the visual system across luminance conditions.

Figure 1. (a) Feedback control diagram for multisensory integration. Under conditions of high-luminance (300 lux, lighter shades, dashed boxes) and low-luminance (0.3 lux, darker shades), the visual (yellow-orange) and mechanosensory (blue-cyan) systems are summed linearly. Mechanosensory and visual position stimuli (r_m and r_v) result in moth positional response (y, purple-pink). The sensory (S_m and S_v), control (C) and plant (P) blocks correspond to sensory, neural and biomechanical subsystems. (b) Time-domain and frequency-domain flower-tracking at high (300 lux) luminance. The coherent MV response (pink), as well as M-moving (cyan) and V-moving (yellow) responses, to motion stimuli (black) show a reliance on both sensory systems. Responses have significant peaks at several driving-frequencies

Mechanosensory and Visual Systems Combine Linearly at High Luminance

We first test whether the visual and mechanosensory cues are still combined linearly across con-
texts (luminance levels). Under each stimulus condition (V-moving, M-moving, and MV-moving),
we quantify the moth's response to flower motion at each frequency in stimulus using the gain
and phase delay in the response. Gain and phase define points in the complex plane as polar co-
ordinates. Frequency responses provide a simple way to test the hypothesis because if the two
cues combine linearly then simply adding (in the complex plane) the response to the V-moving
and M-moving experiments should exactly predict the MV-moving experiments. This is not triv-
ial because in the V-moving and M-moving cases the other cue (the nectary or the facade of the
flower, respectively) is stationary and so the moth must balance conflicting cues in each case. In
order to check consistency of linearity, we first measured the M-moving and V-moving closed-loop
transfer-functions (Hm and Hv), defined by the gain and phase response in the M-moving and V-
moving conditions. These tell us how the moths respond to sensory conflict, in closed-loop. The
mechanosensory and visual closed-loop transfer functions are given by:

$$H_{m,h}(\omega) = \frac{Y_{m,h}(\omega)}{X_{m,h}(\omega)} \qquad H_{v,h}(\omega) = \frac{Y_{v,h}(\omega)}{X_{v,h}(\omega)} \qquad (1)$$

Here, $Y_{m,h}(\omega)$ and $X_{m,h}(\omega)$ are the measured lateral positions of the moth and flower respectively, in
the M-moving case, while $Y_{v,h}(\omega)$ and $X_{v,h}(\omega)$ are for the V-moving case, all in the frequency-domain.
The subscript 'h' denotes high luminance (300 lux). We compare these high-luminance transfer
function measurements with their low-luminance counterparts. If one or both of these transfer
functions change with luminance, then the system has luminance-dependent adaptation.
We find that the closed-loop response (H_m and H_v) for both M-moving and V-moving stimuli, changes
with luminance. For mechanical-moving stimuli, the gain is significantly higher (p=0.95, t-test on

gain and phase at each frequency) in the low luminance condition (Figure 2a) than for high luminance. The opposite is true for visual-moving stimuli (Figure 2b), where the gain is higher for high luminance, over the range of ecologically relevant frequencies (see *Sponberg et al.* (*2015*) for natural frequencies of flowers). Despite these differences, the M-moving response has a low-pass characteristic for both luminance conditions, while the V-moving response is more akin to band-pass (i.e. has a distinct peak in gain) for both luminance conditions. The V-moving peak-gain frequency shifts from 2.9Hz (at 0.3 lux) to 1.9Hz (at 300 lux). Therefore, the moth's multisensory flower-tracking system has quantitative luminance-dependent adaptation, while keeping the qualitative shapes of the transforms. Given this adaptation, it is not trivially obvious that H_m and H_v will continue to combine linearly.

Figure 2. Closed-loop Mechanosensory (a) and Visual (b) motion frequency response functions. Gain and phase of the M-moving (blue-cyan) and V-moving (yellow-orange) transfer functions in (a) and (b) respectively, show luminance-dependence. The lighter shades correspond to 300 lux and the darker shades correspond to 0.3 lux. Mean and 95% confidence intervals are shown, calculated as described in Methods and Materials. Illustrations above the plots show top-views of the moth, nectary and facade, with arrows indicating motion

To test linear combination, in addition to sensory-conflict, we also measured the response of moths to coherent motion of stimuli (MV-moving):

$$H_h(\omega) = \frac{Y_h(\omega)}{X_h(\omega)} \qquad (2)$$

Here, $Y_h(\omega)$ and $X_h(\omega)$ are the frequency domain representations of the moth's and coherent-stimulus' positions, respectively, as a function of frequency (ω). At low luminance, Roth et al *Roth et al.* (*2016*) found that mechanosensory and visual contributions sum linearly, as in Figure 1a (control diagram) and Figure 3a (measured and predicted bode plots). To ascertain whether linear summation of modalities holds true at high luminance (300 lux) as well, we calculated the mean linear-sum prediction:

$$< H_h(\omega) >=< H_{m,h}(\omega) > + < H_{v,h}(\omega) > \qquad (3)$$

Figure 3. Linear Sum Validity Across Luminance Contexts: Gain and phase plots for predicted (brown) and experimental (violet-pink) coherent-tracking transfer function at (a) 0.3 lux (H_l) and (b) 300 lux (H_h). Mean and 95% confidence intervals of the mean are shown, as described in Methods and Materials

On comparing this with the mean transfer function for coherent motion ($H_h(\omega)$), we find that
the two are not significantly different in both gain and phase (Figure 3b, 95% confidence intervals
of mean, at each frequency). Next, we evaluated the linear-sum prediction individually for each
moth, which states that H_m and H_v sum to give H, the coherent tracking response. For all indi-
viduals considered, the linear-sum prediction matches the experimental coherent motion result
over the range of ecologically relevant frequencies. Hence, frequency-dependent re-weighting of
mechanical and visual systems maintains the linearity of their integration, at every frequency. The
summation-template for the eyes and proboscis, described by *Roth et al.* (*2016*) (Figure 1a), per-
sists at illumination levels 1000-fold higher than the one used in their study.

Mechanosensory and visual systems are separately modulated by luminance

The observation that H_v and H_m change with luminance does not tell us whether both visual and
mechanosensory systems are individually modified. This is due to the nature of closed-loop mea-
surements, since H_m is affected by the visual sense as well due to sensory conflict. In order to test
if the observed luminance-dependent adaptation was due to shifts in only the visual system, or in-
volved changes in both vision and mechanosensation, we must mathematically "open" the closed
loop behavior. We do this by combining the behavior results from the M-moving, V-moving and
coherent-motion conditions and factoring out the changes due only to the mechanosensory and
only to the visual systems, which we define as the open-loop mechanosensory transform, G_m, and
the open-loop visual transform, G_v, respectively (Figure 4). Opening the loop to evaluate G_m and
G_v requires comparing across stimulus conditions. These transforms tell us how moths react to
mechanosensory and visual errors, individually (Figure 4).

The (experimentally measured, Figure 2) closed-loop transfer functions H_m and H_v are related to
the open-loop transfer functions (G_m and G_v) by:

$$H_m(\omega) = \frac{G_m(\omega)}{1 + G_m(\omega) + G_v(\omega)} = \frac{Y_m(\omega)}{X_m(\omega)} \qquad H_v(\omega) = \frac{G_v(\omega)}{1 + G_m(\omega) + G_v(\omega)} = \frac{Y_v(\omega)}{X_v(\omega)} \qquad (4)$$

184   Thus, we can infer the open-loop transfer functions using the closed-loop transfer functions mea-
185   sured from conflict (Eq 4) and coherent (Eq 2 and 3) trials:

$$G_m(\omega) = S_m(\omega)C(\omega)P(\omega) = \frac{H_m(\omega)}{1 - H(\omega)} \qquad G_v(\omega) = S_v(\omega)C(\omega)P(\omega) = \frac{H_v(\omega)}{1 - H(\omega)} \qquad (5)$$

186   In the following equations, the subscript 'h' indicates that they pertain to the high luminance ex-
187   periments (300 lux).

$$E_h(\omega) = \begin{bmatrix} 1 - H_h(\omega) & 1 - H_h(\omega) \\ 1 - H_{m,h}(\omega) & -H_{m,h}(\omega) \\ -H_{v,h}(\omega) & 1 - H_{v,h}(\omega) \end{bmatrix} \qquad G_h(\omega) = \begin{bmatrix} G_{m,h}(\omega) \\ G_{v,h}(\omega) \end{bmatrix} \qquad \Psi_h(\omega) = \begin{bmatrix} H_h(\omega) \\ H_{m,h}(\omega) \\ H_{v,h}(\omega) \end{bmatrix} \qquad (6)$$

$$E_h(\omega)G_h(\omega) = \Psi_h(\omega) \qquad G_h(\omega) = E_h(\omega)\backslash\Psi_h(\omega) \qquad (7)$$

For each frequency (ω), and for every individual moth, we thus have an estimate of the open-loop mechanosensory (G_m) and visual (G_v) transfer functions (Eq. 7). Here, we have used MATLAB's *ml-divide* function ("\" operator). We performed a similar calculation on low luminance data (Methods and Materials), but with unpaired data across individuals

On comparing the estimates for open-loop mechanosensory transfer functions, $G_{m,h}$ and $G_{m,l}$ (Fig-

Figure 4. Both visual and mechanosensory open-loop systems change with luminance: Frequency response functions for (a) mechanosensory ($G_{m,l}$: blue, and $G_{m,h}$: cyan) and (b) visual ($G_{v,l}$: orange and $G_{v,h}$: yellow) open-loop transfer functions. The high luminance (300 lux, lighter shades) open-loop transfer functions (mean and 95% confidence intervals) are calculated for each individual, as described in the text. The low-luminance (0.3 lux, darker shades) open-loop transfer functions (mean and 95% confidence intervals) are calculated through permutation, as described in the text. The open-loop transfer functions transform the stimulus-error (E_m and E_v) into moth positional response (Y_m and Y_v) by action of sensory, neural and biomechanical blocks (S, C and P). High to low luminance transformation is composed of frequency dependent gain changes (K_m and K_v)

ure 4b), we find that mechanosensory gain is decreased for high luminance, and this decrease is most apparent between 0.5Hz and 2.9Hz. At 1.3Hz, the mechanosensory gain falls from 1 to around 0.4. The opposite trend is seen for the open-loop visual transfer functions $G_{v,h}$ and $G_{v,l}$. Visual gain is increased in the high luminance condition, while the phase is not significantly different between high and low luminance. The increase in visual gain is most apparent between 0.7Hz and

2.9Hz. The difference in mean gain for several frequencies up to 4.3Hz in both mechanosensory and visual cases is significant (t-test, p=0.95, evaluated at each frequency individually).

The mechanosensory open-loop gain changes with luminance. Therefore, we conclude that both mechanosensory and visual systems are individually dependent on light-level. The change in the two systems varies in a frequency-dependent manner. This is not an intuitively obvious result, since there is currently no known direct mechanism for modulation of proboscis mechanosensory output by luminance. Our result does not necessitate a change in the sensory response of the proboscis afferent neurons, but does predict a luminance-dependent change in the quantitative, motor response to proboscis stimulation. The shift, opposite to visual change, in these behavioral mechanosensory responses, may manifest itself in a number of ways, possibly through changes in neuromodulator profiles at multisensory-integration sites in the brain, and in the flight-control networks of the thorax.

Visual and mechanosensory errors are changed, and coherent error is maintained constant across luminance levels

Since the mechanosensory and visual transfer functions depend on luminance, we next test whether flower-tracking performance is affected by these modifications. We defined complex tracking-errors for coherent and conflict stimuli, as was done by *Roth et al.* (*2016*). The coherent and conflict errors are defined in the complex plane, where perfect tracking of the moving stimulus is gain 1 and phase 0 (at point 1+0j). Deviation from this perfect tracking point is the complex tracking error.

Coherent motion error:

$$E_{coherent}(\omega) = (1 + 0j) - H(\omega) \tag{8}$$

Error with respect to the moving stimulus:

$$E_{m,nectary}(\omega) = (1 + 0j) - H_m(\omega) \qquad E_{v,facade}(\omega) = (1 + 0j) - H_v(\omega) \tag{9}$$

Error with respect to stationary stimulus:

$$E_{m,facade}(\omega) = (0) - H_m(\omega) \qquad E_{v,nectary}(\omega) = (0) - H_v(\omega) \tag{10}$$

$E_{coherent}$, E_m and E_v correspond to coherent (MV) motion, M-moving and V-moving conditions, respectively. The magnitude of coherent motion errors is interpreted as a performance metric. We found that coherent tracking error is not significantly different at 0.3 lux and at 300 lux (Figure 5a). The error magnitudes, when plotted for coherent motion (Figure 5a) show that error is small for low frequencies, greater than 1 at intermediate frequencies, and close to 1 for high frequencies (matching the trend seen in *Sponberg et al.* (*2015*) and *Roth et al.* (*2016*)). This means that the moth tracks slow-frequencies of flowers (less than 1Hz, similar to natural flower motion *Sponberg et al.* (*2015*)) very well, at the expense of poor tracking at higher frequencies. Both high and low-luminance coherent-motion errors are very similar, with a characteristic overshoot around 2.3Hz. Hence, despite modulation of individual sensory responses, the combined response error is indistinguishable for luminance changes of 3 orders of magnitude.

Figure 5. (a) Coherent tracking error is not significantly different for 0.3 lux and 300 lux. This is seen from the error magnitude for high and low luminance, violet-pink (b) Mechanical motion case: error magnitudes (with respect to facade and nectary) show speed-dependent differences in tracking error across contexts (c) Visual motion case: error magnitudes (with respect to facade and nectary) show speed-dependent differences in tracking error across contexts. Darker shades (i) correspond to 0.3 lux and lighter shades (ii) correspond to 300 lux. Means and 95% confidence intervals are plotted, as described in Materials and Methods

The individual error magnitudes of E_m and E_v can tell us which modality the hawkmoths weigh more strongly at different luminances. In the M-moving case, at 0.3 lux the mechanosensory error ($E_{m,nectary}$) is lower than the visual error ($E_{m,facade}$) up to 2.3Hz (Figure 5b). Meanwhile, the visual error is smaller at 300 lux, at all frequencies tested. In the V-moving case, a similar trend holds true (Figure 5c). Here, $E_{v,nectary}$ is smaller than $E_{v,facade}$ for most frequencies at 0.3 lux, while $E_{v,facade}$ is smaller up to 1.9Hz at 300 lux. This relates to the greater reliance on the visual system at 300 lux, and the slower-speed tuning of the proboscis mechanosensory system. These observations are consistent with visual response having a higher weight at high luminance, and mechanosensory response at low luminance. Thus, despite a switch in the higher weighted modality, we find that coherent tracking performance is very similar across luminance conditions (Figure 5a), for the entire range of ecologically relevant speeds (less than 2Hz *Sponberg et al.* (*2015*)). As the luminance decreases, the moth's eyes and early visual system compensate using a spatial and temporal summation strategy (*Sponberg et al.* (*2015*), *Stöckl et al.* (*2016*)), which may contribute to the increased error in response to facade motion. By increasing the relative weight of proboscis cues in this condition, coherent tracking performance is maintained.

Luminance Dependent Delay is the Result of Multisensory Reweighting

Hawkmoths are known to effectively slow their brains to perform flower-tracking in low-luminance conditions (*Sponberg et al.* (*2015*),*Stöckl et al.* (*2016*), *Stöckl et al.* (*2017*)). While a number of possible feedback-control models may fit this description, the result is consistent with a sensorimotor delay of 10ms for a luminance change from 300 lux to 0.3 lux. However this effective or emergent delay may results from multiple changes in the underlying sensory processing such as spatial and temporal summation at low light levels (*Warrant* (*1999*), *Stöckl et al.* (*2016*), *O'Carroll and Warrant* (*2011*)). Adding a delay term (τ=10ms) to the coherent open-loop response at high luminance, we

257 compare it with the coherent closed-loop response at low luminance (Equation 13, where s=$j\omega$).

$$H_{delayed}(\omega) = \frac{\frac{H_h(\omega)e^{-\tau s}}{1-H_h(\omega)}}{1 + \frac{H_h(\omega)e^{-\tau s}}{1-H_h(\omega)}} \tag{11}$$

258 As in (*Sponberg et al.* (*2015*)), the delay has the effect of increasing the gain peak over the range of
259 frequencies from (1.3 to 2.9 Hz), and the phase response rotates to drop by 3.6 degrees per unit
260 frequency (deg/Hz), especially evident after 6.1Hz (Figure 6b). This prediction is not significantly
261 different from the experimental data from low luminance trials at these frequencies. Unlike in
262 (*Sponberg et al.* (*2015*)), the secondary peak present at 0.3 lux between 2.9Hz and 8.9Hz, disappears
263 for 300 lux conditions. This may be attributed to the larger size of the flower in our studies, but
264 the mechanism behind and consequence of the secondary peak is unknown. This implies that the
265 delay model is a simplification of a mechanism with additional features.

Figure 6. (a) Control diagram models for pure delays. Pure delay of 10ms is incompatible with multisensory data (Figure 3, Figure 4), while frequency-dependent gain-modulation (K_m and K_v in Figure 4) is consistent (b) Bode plots for high-luminance (300 lux, pink), low-lumiance (0.3 lux, purple) and delay-prediction (green dashed) transfer functions. Inset has linear y-axis, showing gain between 1.1Hz and 3.2Hz

266   While a delay in the coherent response is consistent with these features of coherent motion
267 responses across the luminance levels, this does not mean that the visual and mechanosensory
268 systems are purely delayed an equal amount. That would be incompatible with the observed
269 shift in gains of the open-loop responses (Figure 6a). Instead, a frequency-dependent gain ad-
270 justment in both vision and mechanosensation can account for the change (in Figure 4, K_m and
271 K_v are frequency-dependent gain knob). A more general model consisting of both variable-gain
272 and delay tuning of both systems, may give comparable results. The pure-delay motif for coher-
273 ent flower motion found in *Sponberg et al.* (*2015*) results from a linear combination of gain and
274 phase tuning of both open-loop, mechanosensory and visual sensorimotor branches. Specifically,
275 the delay resulting from the visual system's spatio-temporal summation at low luminance com-
276 plemented by frequency-dependent gain and phase adjustments in the mechanosensory systems.
277 This is reminiscent of *Stöckl et al.* (*2017*), where gain and delay tuning was found to account for
278 inter-species differences. Therefore, both within-species differences (this study) across contexts,
279 and between-species differences (*Stöckl et al.* (*2017*)), are consistent with similar types of control
280 adjustments.

 Discussion

 Multisensory Integration is Context Dependent for *Manduca sexta* Flower Tracking

We found that *Manduca sexta's* proboscis-mechanosensory flower-tracking response is luminance-
dependent, adapting in a manner separate from changes to vision. The mechanosensory and
visual systems continue to combine linearly across light levels, and their coherently-combined ef-
fect is equivalent to a delay in the moth's response. These luminance-dependent adaptations result
in comparable flower-tracking performance across contexts. Thus, a linear combination of cues,
individually re-weighted depending on luminance context, enables coherent tracking of flower mo-
tion in *Manduca sexta*. This multisensory integration and re-weighting of cues may be important
because *Manduca sexta* experiences luminance changes across this range during foraging. The
difference in peak-response frequencies of the mechanosensory and visual systems, combined
with their frequency overlap, allows for the adaptive nature of the context-dependent response.
Degradation of one system, such as increases in visual delay and loss of resolution with luminance
decrease, can be partially accommodated by tuning the complementary (mechanosensory) system
gain, at the appropriate speeds.
Multisensory integration may not be essential for flower-tracking in all species. Indeed, the diurnal
hawkmoth, *Macroglossum stellatarum* relies mostly on its visual sense, and very little on proboscis
cues, in response to looming flowers (***Farina et al.*** (***1994***)), a fact underlined by its region-specific
optical specializations (***Farina et al.*** (***1995***), ***Warrant et al.*** (***1999***)). In *Manduca sexta*, its longer pro-
boscis may make for a more reliable organ for sensing (***Stöckl et al.*** (***2017***)), which is especially desir-
able in low or variable light conditions. However, even *M. sexta* shows redundancy between the two
systems (***Roth et al.*** (***2016***)). This means that either system can, on its own, successfully produce
flower-tracking. Having two redundant systems allows for compensation when visual salience is
affected, such as when flying into shade or between evenings and late nights. The conditions which
may lead to decreased mechanosensory salience are less well understood.
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Manduca sexta feed from flowers moving in the wind, performing an energetically expensive
and challenging hovering behavior. The energetic gains from successful feeding, however, have
been estimated to more than make up for this loss (***Sprayberry and Daniel*** (***2007***),***Stöckl and Kel-***
ber (***2019***)), and are known to have fitness benefits in terms of the number of eggs laid (***Levin et al.***
(***2016***), ***von Arx et al.*** (***2013***)). In order to feed uninterrupted, they would benefit from tracking the
frequencies of natural flower motion ***Sponberg et al.*** (***2015***). The two sensorimotor systems used
for flower tracking have appreciable luminance-dependent response-gain modulation at these fre-
quencies (Figure 4). Hence the integration of visual and mechanosensory cues has an essential
impact on the successful tracking of moving flowers.
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The multisensory aspect of flower-tracking may be related to the use of the mechanosensory
and visual systems in associated hawkmoth behaviors, such as proboscis placement (***Goyret and***
Kelber (***2012***),***Goyret*** (***2010***)), flower handling and learning (***Goyret and Raguso*** (***2006***),***Deora et al.***
(***2021***)). Switching between these behaviors seems to be gated by olfaction and other multimodal
cues (***Raguso and Willis*** (***2002***)). We find that luminance acts as an additional environmental cue,
controlling the method of execution of a behavior, by re-weighting of sensorimotor gains. The
re-weighting of sensory systems has been observed previously, in refuge-keeping of electric knife-
fish. Depending on the salience of electrosensory (conductance) and visual (light-level) information,
Sutton et al. (***2016***) found that the gain ratio of their two systems was modulated, with the infor-
mation being combined in a roughly linear manner. While this study was done only at distinct
probe frequencies, and not across the dynamic range, the re-weighting shows us that the same
effective dynamics may emerge across distantly related taxa, when engaging in a similar behavior.
This raises the question, is there a performance benefit to the linear combination of multisensory
information?

331

332 Sensorimotor Timescales of Visual and Mechanosensory Modalities Shift Inversely

333 Taken together our results show not only the visual tracking of object, but also the mechanosens-
334 ing of objects changes with the background luminance. Each of the visual and mechanosensory
335 responses show frequency tuning across overlapping frequency ranges (Figure 4). The width of
336 the frequency bands over which each system's gain is larger than the other's, tells us the relative
337 speed-tuning of the mechanosensory and visual systems. We see that the proboscis mechanosen-
338 sory system operates over a slower timescale than the compound-eye visual system, with a gain
339 crossover point defining each system's bandwidth. As luminance increases, this crossover point
340 moves to lower speeds. Thus we say that the visual system's bandwidth increases, that of the
341 mechanosensory system decreases. In other words, the visual system slows down (as evidenced
342 by its peak shifting) enough to compensate for the loss of mechanosensory gain, at high luminance.
343 The two systems continue to fulfill their roles of high- and low-speed response, respectively, but
344 with their frequency bandwidths rescaled accordingly.

345

346 Bandwidth separation and complementary action are usually seen when two sensory systems
347 have very different frequency tuning. Difference in timescales of visual (slow) and gyroscopic hal-
348 tere (fast) responses in dipterans, enable optomotor reflexes and successful object-fixation behav-
349 ior (*Mureli and Fox* (*2015*)). The optomotor response range of *drosophila* is expanded by the use
350 of these two, differently tuned systems (*Sherman and Dickinson* (*2003*)). The use of high-speed
351 mechanosensors also allows *drosophila* to stabilize their visually-controlled flight speed (*Fuller*
352 *et al.* (*2014*)). In *Manduca sexta*, the function of high-frequency gyroscopic mechanosensation is
353 performed by the antennae (*Sane et al.* (*2007*)). Similarly, in the closely-related species, Macroglos-
354 sum stellatarum, *Dahake et al.* (*2018*) noted the separation of antennae-guided stabilization, and
355 visually-guided flower-tracking roles. In their study, the visual system could not compensate for
356 the loss of antennae, even in bright lighting conditions. This led to the hypotheses of either spatial
357 separation of the two systems, via parallel and independent descending pathways, or temporal
358 separation (multiplexing) due to a difference in response speeds. Thus, they found that antennal
359 mechanosensation is separate in its frequency response from visual and proboscis mediated cues.
360 We find that the proboscis and eyes compensate for each other, and work in combination across
361 contexts. This is possible, and even necessary due to the overlap in frequency bands between
362 the eyes and proboscis. The shift in frequency bands in our system may be investigated further
363 through electrophysiology in the proboscis and descending neurons

364

365 The proboscis and eyes, unlike the halteres-eyes and antennae-eyes systems, have consider-
366 able frequency-overlap, and fulfill similar roles. A closer analogy would be the widefield motion (op-
367 tomotor) and flower-tracking systems, in the related hover-feeding species, *M. stellatarum*. When
368 *M. stellatarum* feeds from a looming flower (*Farina et al.* (*1995*)), distance-stabilization (flower dis-
369 tance) and drift-compensation (widefield motion) responses have very different bandpass shapes,
370 while sharing a common range of frequencies. Similarly, the response to rotating widefield motion
371 (*Kern* (*1998*),*Kern and Varju* (*1998*)) is strongly peaked. These behaviors may be in conflict or co-
372 herent, depending on the context of motion (intent) of the moth and changes in its environment.
373 Thus, even more extreme rescaling of sensory information has been observed here, dependent
374 on internal and external context (albeit for the same sensory system, i.e. vision).

375 Luminance dependent delays in visual processing can arise form multisensory reweight-
376 ing

377 Luminance-dependent changes in the visual system have been studied extensively, and at various
378 stages, in insects. Matched filtering (*Kohn et al.* (*2018*)) has been found to result in speed tuning of
379 sensory systems depending on their natural ecology, which may in turn span different luminance

contexts. Light-level affects the temporal frequency tuning of lobula plate tangential cells (LPTCs) (*O'Carroll and Warrant* (*2011*)) of *Deilephila elpenor*, and summation of spatial information by lamina monopolar cells (LMCs) (*Stöckl et al.* (*2020*)), in addition to physical dark-adaptation in the eyes. The result of such adaptation, along with possible differences in wing kinematics and control, can result as an effective delay in flower tracking for *M. sexta* (*O'Carroll and Warrant* (*2011*); *Sponberg et al.* (*2015*)).

Our observation, that coherent motion tracking is consistent with previously characterized delay (*Sponberg et al.* (*2015*)), gives us a model for how the visual and mechanical systems must combine: the result of their respective luminance-tuning adds to give an overall system delay. Flower-tracking behavior has been previously studied in the linear regime for three species of hawkmoths (*Stöckl et al.* (*2017*)), and inter-species differences are matched to natural ecology, with species-dependent gain scaling and luminance-tuned delays. We found that gain and delay tuning of sub-systems, can account for luminance dependent tracking differences within a species as well. Electrophysiological studies of sensory integration, combined with analyses of flight EMGs and dynamics, may shed light on how this template is achieved.

We found that the delay is the result of multisensory integration and control, and the proboscis mechanosensory system has appreciable luminance-dependent gain-modulation. The corresponding changes in the proboscis, if any, have yet to be studied. Alternatively, an inverse set of changes closer to flight control centers, could also account for this gain tuning, without direct modulation of proboscis sensors. If true, this could be a general locomotion strategy used by animals, whereby multiple sensory systems can robustly combine to control different speeds of action, through changes in a subset of the sensors, and corresponding changes in output control sites.

Methods and Materials

Robotic flower for sensory conflict experiments

Following the methods described by *Roth et al.* (*2016*), we used a two-part robotic flower to generate coherent motion as well as sensory conflict stimuli. The motion of the flower facade provided visual stimuli, and the nectary motion provided mechanosensory cues. The robotic flower's dimensions matched those used by *Roth et al.* (*2016*). The facade and nectary were 3d printed out of ABS P430 using a Stratasys® uPrint® 3D printer, and were mounted on 16 cm carbon fiber rods. The flower facade had a slit wide enough to not interfere with proboscis motion (2mm). The rubbing of the facade against the proboscis would suggest a phase-leading derivative (velocity) response, where as we observe, low phase lags consistent with a proportional (positional) response. Thus, movement of the facade did not deflect the stationary proboscis in the absence of nectary movement. Two stepper motors (57sth56-2804mb, Dongzheng. Motor Co. LTD; China) were used to provide independent movement stimuli. The nectary was painted black to minimize visual distraction. Our robotic flower differed from the ones used by *Sponberg et al.* (*2015*), in its diameter (9.5cm vs 4.5cm). The larger diameter of our robotic flower was necessary to generate the sensory conflict stimuli. Hence, the applicability of the control-system delay theory to our paradigm would require further evaluation.

Sum-of-sines Stimuli

Sum of sines stimuli were provided, with 20 frequency components (being prime multiples of 0.1Hz), logarithmically spaced, from 0.2Hz to 19.9Hz. This stimulus was presumed to be pseudo-random, so that it could not be predicted by the moth. Stimulus amplitudes were chosen such that velocity amplitudes were equal at each frequency. This velocity was similar to the one used by *Sponberg et al.* (*2015*). Phase difference for each component sine waveform was randomized, and kept constant across trials for a given luminance level. We tested 3 stimulus conditions: Coherent motion, M-moving and V-moving. In coherent motion, both the facade and nectary moved

428 together. In the M-moving case, the facade was stationary, while the nectary was moved as a sum
429 of sines. Conversely, in the V-moving case, the facade was moved while the nectary remained sta-
430 tionary.
431

Animal Handling and Video Recording

432
433 We obtained *Manduca sexta* pupae from the University of Washington. Within 2 to 3 days post-
434 eclosion, 8 moths were placed successively in a flight chamber, with the robotic flower facade being
435 illuminated at 300lux. The light source used was the LED light, Neewer CN-126, with 2 attached
436 neutral density filter layers. For ease of tracking, we painted a white spot (Prang White Paint, Dixon
437 Ticonderoga Co.) on each moth's thorax before the start of every trial. The MATLAB plug-in, DLT-
438 DV was used to track the position of the thorax of each individual. We painted a black spot on
439 top of the flower facade, and a white spot on top of the nectary (out of view of the feeding moth)
440 for automated tracking. Tracks were validated manually. Movement stimuli were set to ramp
441 up linearly in amplitude, over 2 seconds during the beginning and end of recordings, to prevent
442 sudden impulsive stimuli and motor saturation. Sum of sines stimuli were provided for 20 seconds,
443 in order to get four repeats of the lowest frequency. A Fastec camera was used to record the
444 trials (top view), at 2000fps. A seven-component scent (*Campos et al.* (*2015*) 0.9-1.7uL) was placed
445 on a Q-tip™ to increase the likelihood of feeding behavior. Stimulus order was randomized, and
446 each individual was made to fly once every day, over 3-4 days. Hence, each feeding individual was
447 presented with all 3 stimuli. Individuals that did not feed on at least 3 of these days were excluded
448 from analysis. Individuals were weighted before and after each trial, and the decrease in nectary
449 weight was compared with increase in moth weight, while the nectary was refilled for each trial.
450 The procedure was repeated for 10 more feeding individuals. Out of 18 trials, 12 were identified
451 to coherently track and feed from flowers, and did not suffer excessive visible wing-damage, and
452 these were included in our analysis.
453 Low-luminance (0.3 lux) tracking results were taken directly from Roth et al's dataset.

Linear combination of cues in Individuals

454
455 We collected triads of data at 300 lux, with one trial for each stimulus condition per individual.
456 Therefore, we were able to evaluate the linear combination of cues for each individual moth. The
457 predicted coherent response is the sum of M-moving and V-moving responses:

$$H_h(\omega) = H_{m,h}(\omega) + H_{v,h}(\omega) \tag{12}$$

458 We find this to be the case for every individual tested
459

Frequency-Domain Analysis and Statistics

460
461 Transfer Functions
462 We used the same statistical analysis technique as *Roth et al.* (*2016*) for complex-valued transfer-
463 functions. Briefly, the transfer function H(ω) is given by:

$$H(\omega) = g(\omega)exp(j \cdot \theta(\omega)) \tag{13}$$

464 The log-space representation of this transfer-function is:

$$\ln H = \ln g + j \cdot \theta \tag{14}$$

465 In log-space, we calculate the mean and standard deviation of the gain, and then return to the
466 untransformed complex plane:

$$|H(\omega)|_{\log \text{mean}} = \frac{1}{N} \sum_{k=1}^{N} \ln|H_k| \tag{15}$$

$$|H(\omega)|_{\text{log std}} = \sqrt{\frac{1}{N-1} \sum_{k=1}^{N} (\ln \frac{|H_k(\omega)|}{|H(\omega)|})^2} \tag{16}$$

$$|H(\omega)|_{\text{mean}} = exp|H(\omega)|_{\text{log mean}} \tag{17}$$

$$|H(\omega)|_{\pm std} = exp(|H(\omega)|_{\text{log mean}} \pm |H(\omega)|_{\text{log std}}) \tag{18}$$

470 The phase of the transfer functions is given by:

$$\angle H(\omega)_{mean} = \angle \frac{1}{N} \sum_{k=1}^{N} exp(\angle H_k(\omega) \cdot j) \tag{19}$$

$$\angle H(\omega)_{std} = \sqrt{-2 \ln R} \tag{20}$$

$$R = |\frac{1}{N} \sum_{k=1}^{N} exp(\angle H_k(\omega) \cdot j)| \tag{21}$$

Statistics on predicted values of transfer functions are done by propagation of uncertainty analysis,
similar to *Roth et al.* (*2016*). These include predictions for the linear sum, the open-loop transfer
functions, and the delay predictions.
For low luminance, the data was unpaired. Therefore, to calculate the open-loop frequency re-
sponse functions, $G_{m,l}$ and $G_{v,l}$, we made every possible combination of the 8x8x8 individuals (8
experimental trials each for M, V and MV stimuli). This gave us 512 "paired" triads of data. We then
extended equations (6) and (7) with these 512 rows of triads, and estimated the open-loop func-
tions by solving this version of equation (7). These synthetic transfer functions provide an upper
bound on the variance.

Time Domain and Fourier Amplitude
The time-series of individuals are aligned to the start of stimuli, and a timepoint-specific standard
deviation is found for each stimulus condition (Figure 1b).
The Fourier amplitudes for these timeseries have peaks at driving frequencies. Peak significance
is estimated similar to *Sponberg et al.* (*2015*), assuming white-noise as null signal. The frequency
range for significant peaks depends on the individual and the stimulus condition (coherent or con-
flict). Therefore, only frequencies up to 6.1 Hz are used in analyses, since these frequencies have
significant Fourier peaks.

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References

von Arx M, Sullivan KA, Raguso RA. Dual fitness benefits of post-mating sugar meals for female hawkmoths
(Hyles lineata). Journal of insect physiology. 2013; 59(4):458–465. doi: 10.1016/j.jinsphys.2013.01.006.

Campos EO, Bradshaw HD, Daniel TL. Shape matters: Corolla curvature improves nectar discovery in the
hawkmoth Manduca sexta. Functional Ecology. 2015 4; 29:462–468. doi: 10.1111/1365-2435.12378.

Cellini B, Mongeau JM. Nested mechanosensory feedback actively damps visually guided head movements in
Drosophila. Elife. 2022; 11:e80880. doi: 10.7554/eLife.80880.

Cowan NJ, Lee J, Full RJ. Task-level control of rapid wall following in the American cockroach. Journal of Experi-
mental Biology. 2006 5; 209:1617–1629. doi: 10.1242/jeb.02166.

Dahake A, Stöckl AL, Foster JJ, Sane SP, Kelber A. The roles of vision and antennal mechanoreception in hawk-
moth flight control. eLife. 2018 12; 7. doi: 10.7554/eLife.37606.

Deora T, Ahmed MA, Daniel TL, Brunton BW. Tactile active sensing in an insect plant pollinator. The Journal of experimental biology. 2021 2; 224. doi: 10.1242/jeb.239442.

Ernst MO, Banks MS. Humans integrate visual and haptic information in a statistically optimal fashion. Nature. 2002 1; 415:429–433. doi: 10.1038/415429a.

Farina WM, Kramer D, Varju D. The response of the hovering hawk moth Macroglossum stellatarum to translatory pattern motion. J Comp Physiol A. 1995; 176:551. doi: 10.1007/BF00196420.

Farina WM, Varju D, Zhou Y. The regulation of distance to dummy flowers during hovering flight in the hawk moth Macroglossum stellatarum. J Comp Physiol A. 1994; 174:239–247. doi: 10.1007/BF00193790.

Fuller SB, Straw AD, Peek MY, Murray RM, Dickinson MH. Flying Drosophila stabilize their vision-based velocity controller by sensing wind with their antennae. Proceedings of the National Academy of Sciences of the United States of America. 2014 4; 111:E1182–E1191. doi: 10.1073/pnas.1323529111.

Goyret J. Look and touch: Multimodal sensory control of flower inspection movements in the nocturnal hawkmoth Manduca sexta. Journal of Experimental Biology. 2010 11; 213:3676–3682. doi: 10.1242/jeb.045831.

Goyret J, Kelber A. Chromatic signals control proboscis movements during hovering flight in the hummingbird hawkmoth macroglossum stellatarum. PLoS ONE. 2012 4; 7:34629. doi: 10.1371/journal.pone.0034629.

Goyret J, Raguso RA. The role of mechanosensory input in flower handling efficiency and learning by Manduca sexta. Journal of Experimental Biology. 2006 5; 209:1585–1593. doi: 10.1242/jeb.02169.

Hengstenberg R, Sandeman DC, Hengstenberg B, Horridge GA. Compensatory head roll in the blowfly Calliphora during flight. Proceedings of the Royal Society of London Series B Biological Sciences. 1986; 227(1249):455–482. doi: 10.1098/rspb.1986.0034.

Hengstenberg R. Mechanosensory control of compensatory head roll during flight in the blowfly Calliphora erythrocephala Meig. Journal of Comparative Physiology A. 1988 3; 163:151–165. doi: 10.1007/BF00612425.

Huston SJ, Krapp HG. Nonlinear integration of visual and haltere inputs in fly neck motor neurons. Journal of Neuroscience. 2009; 29(42):13097–13105. doi: 10.1523/JNEUROSCI.2915-09.2009.

Kern R. Visual position stabilization in the hummingbird hawk moth, Macroglossum stellatarum L. II. Electrophysiological analysis of neurons sensitive to wide-field image motion. Journal of Comparative Physiology - A Sensory, Neural, and Behavioral Physiology. 1998 2; 182:239–249. doi: 10.1007/s003590050174.

Kern R, Varju D. Visual position stabilization in the hummingbird hawk moth, Macroglossum stellatarum L. I. Behavioural analysis. Journal of Comparative Physiology - A Sensory, Neural, and Behavioral Physiology. 1998 2; 182:225–237. doi: 10.1007/s003590050173.

Kohn JR, Heath SL, Behnia R, Eyes matched to the prize: The state of matched filters in insect visual circuits. Frontiers Media S.A.; 2018. doi: 10.3389/fncir.2018.00026.

Levin E, Mitra C, Davidowitz G. Fed males increase oviposition in female hawkmoths via non-nutritive direct benefits. Animal Behaviour. 2016; 112:111–118. doi: 10.1016/j.anbehav.2015.11.019.

Maimon G, Straw AD, Dickinson MH. Active flight increases the gain of visual motion processing in Drosophila. Nature Neuroscience. 2010 3; 13:393–399. doi: 10.1038/nn.2492.

Mureli S, Fox JL. Haltere mechanosensory influence on tethered flight behavior in Drosophila. Journal of Experimental Biology. 2015 8; 218:2528–2537. doi: 10.1242/jeb.121863.

O'Carroll DC, Warrant EJ. Computational models for spatiotemporal filtering strategies in insect motion vision at low light levels. In: *2011 Seventh International Conference on Intelligent Sensors, Sensor Networks and Information Processing*; 2011. p. 119–124. doi: 10.1109/ISSNIP.2011.6146593.

Oie KS, Kiemel T, Jeka JJ. Multisensory fusion: Simultaneous re-weighting of vision and touch for the control of human posture. In: , vol. 14 Brain Res Cogn Brain Res; 2002. p. 164–176. doi: 10.1016/S0926-6410(02)00071-X.

Olberg RM. Pheromone-triggered flip-flopping interneurons in the ventral nerve cord of the silkworm moth, Bombyx mori. Journal of Comparative Physiology A. 1983; 152:297–307. doi: 10.1007/BF00606236.

Raguso RA, Willis MA. Synergy between visual and olfactory cues in nectar feeding by naïve hawkmoths, Manduca sexta. Animal Behaviour. 2002 11; 64:685–695. doi: 10.1006/anbe.2002.4010.

554 **Rauscher MJ**, Fox JL. Haltere and visual inputs sum linearly to predict wing (but not gaze) motor output
555 in tethered flying Drosophila. Proceedings of the Royal Society B: Biological Sciences. 2021 1; 288. doi:
556 10.1098/rspb.2020.2374.

557 **Roth E**, Hall RW, Daniel TL, Sponberg S. Integration of parallel mechanosensory and visual pathways resolved
558 through sensory conflict. Proceedings of the National Academy of Sciences of the United States of America.
559 2016; 113:12832–12837. doi: 10.1073/pnas.1522419113.

560 **Sane SP**, Dieudonné A, Willis MA, Daniel TL. Antennal mechanosensors mediate flight control in moths. Science.
561 2007 2; 315:863–866. doi: 10.1126/science.1133598.

562 **Sherman A**, Dickinson MH. A comparison of visual and haltere-mediated equilibrium reflexes in the fruit fly
563 Drosophila melanogaster. Journal of Experimental Biology. 2003 1; 206:295–302. doi: 10.1242/jeb.00075.

564 **Sponberg S**, Dyhr JP, Hall RW, Daniel TL. Luminance-dependent visual processing enables moth flight in low
565 light. Science. 2015 6; 348:1245–1248. doi: 10.1126/science.aaa3042.

566 **Sprayberry JDH**, Daniel TL. Flower tracking in hawkmoths: Behavior and energetics. Journal of Experimental
567 Biology. 2007 1; 210:37–45. doi: 10.1242/jeb.02616.

568 **Stöckl AL**, Kihlström K, Chandler S, Sponberg S. Comparative system identification of flower tracking perfor-
569 mance in three hawkmoth species reveals adaptations for dim light vision. Philosophical Transactions of the
570 Royal Society B: Biological Sciences. 2017 4; 372. doi: 10.1098/rstb.2016.0078.

571 **Stöckl AL**, Kelber A. Fuelling on the wing: sensory ecology of hawkmoth foraging. Journal of Comparative
572 Physiology A. 2019; 205:399–413. doi: 10.1007/s00359-019-01328-2.

573 **Stöckl AL**, O'Carroll DC, Warrant EJ. Neural summation in the hawkmoth visual system extends the limits of
574 vision in dim light. Current Biology. 2016 3; 26:821–826. doi: 10.1016/j.cub.2016.01.030.

575 **Stöckl AL**, O'Carroll DC, Warrant EJ. Hawkmoth lamina monopolar cells act as dynamic spatial filters to optimize
576 vision at different light levels. Science Advances. 2020 4; 6:eaaz8645. doi: 10.1126/sciadv.aaz8645.

577 **Sutton EE**, Demir A, Stamper SA, Fortune ES, Cowan NJ. Dynamic modulation of visual and electrosensory gains
578 for locomotor control. Journal of the Royal Society Interface. 2016 5; 13. doi: 10.1098/rsif.2016.0057.

579 **Theobald JC**, Warrant EJ, O'Carroll DC. Wide-field motion tuning in nocturnal hawkmoths. Proceedings of the
580 Royal Society B: Biological Sciences. 2010 3; 277:853–860. doi: 10.1098/rspb.2009.1677.

581 **Warrant E**, Bartsch K, Günther C. Physiological optics in the hummingbird hawkmoth: a compound eye without
582 ommatidia. Journal of Experimental Biology. 1999; 202(5):497–511. doi: 10.1242/jeb.202.5.497.

583 **Warrant EJ**. Seeing better at night: life style, eye design and the optimum strategy of spatial and temporal
584 summation. Vision research. 1999; 39(9):1611–1630. doi: 10.1016/S0042-6989(98)00262-4.