Diverse prey capture strategies in teleost larvae

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Summary

Animal behavior is adapted to the sensory environment in which it evolved, while also being constrained by physical limits, evolutionary history, and developmental trajectories. The hunting behavior of larval zebrafish (*Danio rerio*), a cyprinid native to streams in Eastern India, has been well characterized. However, it is unknown if the complement and sequence of movements employed during prey capture by zebrafish is universal across freshwater teleosts. Here, we explore the syntax of prey capture behavior in larval fish belonging to the clade *Percomorpha*, whose last common ancestor with cyprinids lived ~240 million years ago. We compared the behavior of four cichlid species from Lake Tanganyika endemic to deep benthic parts of the lake (*Lepidiolamprologus attenuatus*, *Lamprologus ocellatus*, and *Neolamprologus multifasciatus*) or inhabiting rivers (*Astatotilapia burtoni*) with that of medaka (*Oryzias latipes*), a fish found in rice paddies in East Asia. Using high speed videography and neural networks, we tracked eye movements and extracted swim kinematics during hunting from these five species. Notably, we found that the repertoire of hunting movements of cichlids is broader than that of zebrafish, but shares basic features, such as eye convergence, positioning of prey centrally in the binocular visual field, and discrete prey capture bouts, including two kinds of capture strikes. In contrast, medaka swim continuously, track the prey monocularly without eye convergence, and position prey laterally before capturing them with a side swing. This configuration of kinematic motifs suggests that medaka may judge distance to prey by motion parallax, while cichlids and zebrafish may use binocular visual cues. Together, our study documents the diversification of locomotor and oculomotor adaptations among hunting teleost larvae.

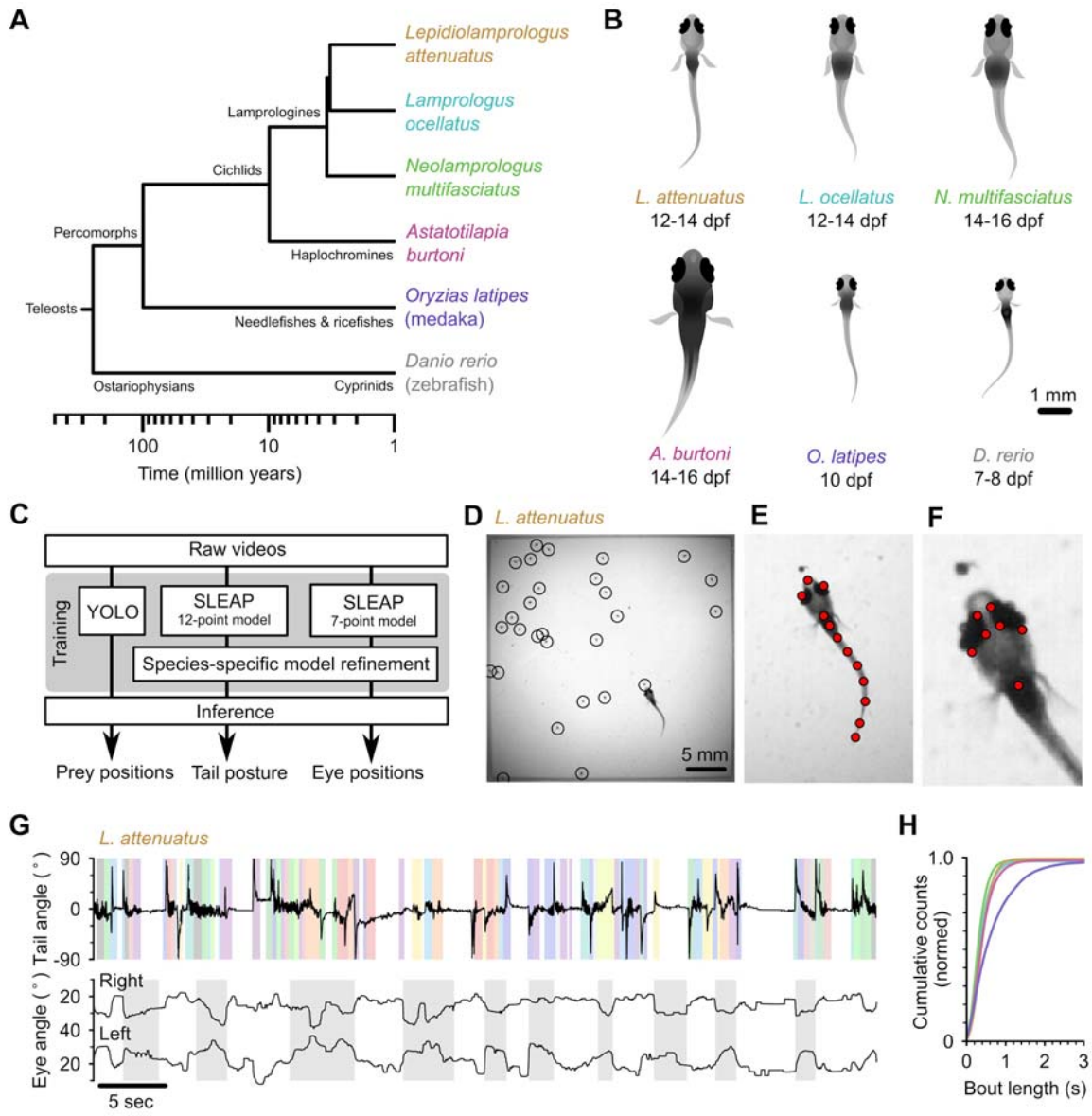

Figure 1. Tracking prey capture behavior in five species of fish larvae. (**A**) Phylogenetic relationship between percomorph species used in this study, and their relationship to zebrafish (a cyprinid). Based on Betancur-R et al. (2017). (**B**) Schematics of larvae of each species studied here (and zebrafish, *Danio rerio*, for reference). Scale bar: 1 mm. (**C**) Outline of tracking procedure. Raw video frames were analyzed with three different neural networks to extract tail pose, eye pose and prey position in each frame. Kinematic features were extracted from raw pose data and used for subsequent analyses. (**D-F**) Tracking output for each neural network for a single frame from a recording of an *L. attenuatus* larva, showing identified artemia (D), tail points (E) and eye points (F). (**G**) One minute of tail and eye tracking from *L. attenuatus* showing tail tip angles (top trace) and right and left eye angles (bottom traces) relative to midline. Bottom trace, gray shaded boxes: automatically identified prey capture periods, when the eyes are converged (see **Figure 2**). Top trace, shaded boxes: automatically identified and classified bouts. Color corresponds to cluster identity in **Figure 4**. (**H**) Cumulative distribution of bout lengths across species for all fish combined. Colors as in (A) and (B). Medaka bouts (purple line) are longer than cichlid bouts.

Introduction

In recent decades, biologists have increasingly relied on a handful of genetically tractable species to study questions related to behavioral mechanisms and their underlying neural circuitry (Devineni & Scaplen, 2022; Piggott et al., 2011; Zhu & Goodhill, 2023). However, it is often not clear to what extent these findings in model organisms can be generalized to other taxa or even closely related species. For example, recent comparative behavioral studies in drosophilids that diverged less than 40 mya found differences in both locomotion (York et al., 2022) and sequencing of spontaneous behaviors (Hernández et al., 2021). For vertebrates, the zebrafish larva has become a dominant model for understanding the neuronal circuits and pathways controlling innate behavior (Baier & Scott, 2009; Friedrich et al., 2010; Gahtan & Baier, 2004; Orger & de Polavieja, 2017; Portugues & Engert, 2011). While many aspects of the visuomotor transformations and underlying neural circuitry for prey capture have been revealed in this species (reviewed by Zhu & Goodhill, 2023), it is not known if this is the general solution for larval teleosts or a derived adaptation to the zebrafish's specific ecological niche.

Here, we applied advances in computational ethology, such as automated tracking with deep neural networks (Mathis et al., 2018; Pereira et al., 2019) and unsupervised analyses (Berman et al., 2014; Marques et al., 2018; Mearns et al., 2020; Wiltschko et al., 2015) to compare hunting behavior of zebrafish to the Japanese rice fish, medaka (*O. latipes*) (Mano & Tanaka, 2012) and four cichlids from Lake Tanganyika (El Taher et al., 2021; Higham et al., 2007). Medaka and cichlids belong to a diverse clade of teleosts known as percomorphs, whose last common ancestor with ostariophysi, such as zebrafish and Mexican cavefish, lived ~240 mya (Betancur-R et al., 2017) (**Fig. 1A**). While medaka and cichlids are only distantly related (~100 mya; Betancur-R et al., 2017), the haplochromine species and the three lamprologine species studied here diverged recently, within the past 3 million years (Ronco et al., 2021). We discovered that swim kinematics and the use of eye movements differ qualitatively and quantitatively between these species. This divergence may be driven, in part, by differences in the sensorineural mechanisms underlying prey detection.

Results

<u>Artificial neural networks track swimming behavior and eye movements in percomorph larvae</u>

At five to seven days post fertilization (dpf), zebrafish larvae robustly feed on small prey items such as paramecia. To stage-match larvae of different species to zebrafish, we first identified when they started feeding (**Fig. 1B**). Cichlid larvae started feeding later than zebrafish, from 12-14 dpf, and medaka started feeding at approximately 10 dpf. We next adapted an experimental paradigm used to study prey capture in zebrafish for these other species (Mearns et al., 2020). Individual larvae were placed in chambers with prey items (either artemia or paramecia). We recorded each animal for 15 minutes using a high-speed camera (**Supp. Videos 1-5**, **Methods**). We then used neural networks to extract tail pose, eye movements, and prey locations from

videos (**Fig. 1C**). We trained a 12-point SLEAP model (Pereira et al., 2022) to track the tail, and a 7-point model to track the eyes of larvae (**Fig. 1E,F**). We tracked prey position using YOLO (Redmon et al., 2016; **Fig. 1D**). We subsequently used these estimated pose dynamics and prey location information to compare hunting and swimming across species.

Larvae of different species have diverse swim patterns

At the first-feeding stage, zebrafish larvae swim in discrete, discontinuous bouts (Budick & O'Malley, 2000), which merge into continuous swimming at the juvenile stage (Westphal & O'Malley, 2013). In contrast, *Danionella cerebrum*, a close relative of zebrafish, exhibits slow continuous swimming as larvae (Rajan et al., 2022). It is not known how larval swimming patterns differ between more distantly related species.

We found marked variation in the continuity of swimming in early percomorph larvae (**Fig. 1G; Supp. Fig. 1**). On one extreme, medaka (OL) and *A. burtoni* (AB) swam with a "motorboating" style characterized by sustained, uninterrupted tail undulations over many seconds with only short breaks between swimming episodes, similar to *Danionella*. On the other extreme, *L. ocellatus* (LO) and *N. multifasciatus* (NM) had an intermittent swimming style, often resting at the bottom of the chamber for minutes at a time with quiescent periods interrupted by short, rapid bursts of activity. *L. attenuatus* (LA) showed a behavior intermediate to these extremes, alternating between rapid tail beating and quiescence, each lasting on the order of a few seconds, a discontinuous style similar to zebrafish.

Analyses across the animal kingdom suggest that behavior is organized into sub-second kinematic motifs (Berman et al., 2014; Stephens et al., 2008; Tinbergen, 1951; Wiltschko et al., 2015). Inspecting periods of swimming across percomorph species revealed significant substructure within swimming episodes (**Fig. 1G; Supp. Fig. 1**). We segmented continuous tail traces into discrete bouts using a change detection algorithm adapted from Mearns et al., 2020 (see **Methods**), which revealed changes in swim kinematics occuring on the order of tens to hundreds of milliseconds (median seconds: 0.33, LO; 0.29, NM; 0.38, LA; 0.36, AB; 0.49, OL). Furthermore, medaka exhibited significantly longer swims than cichlids ($p < 0.001$, KS test) (**Fig. 1H**). These results highlight differences in how fish larvae pattern their swim episodes, which may relate to different sensorimotor strategies for prey detection and tracking.

Larvae of different species use diverse strategies to hunt prey

Zebrafish larvae converge their eyes at the onset of hunting episodes, and keep their eyes converged over the course of prey capture (Bianco et al., 2011; Patterson et al., 2013). Eye convergence aids prey capture by bringing prey into a binocular zone in the central visual field (Gahtan et al., 2005; Gebhardt et al., 2019). In contrast, some fish species such as the blind cave fish, *Astyanax mexicanus*, capture prey that are positioned laterally to their mouth (Lloyd et al., 2018; Espinasa et al., 2023). This shift in capture strategy appears to have happened over a

relatively short timescale when cavefish diverged from their surface ancestors, and consequently cavefish still converge their vestigial eyes even though they do not rely on vision for prey capture (Espinasa & Lewis, 2023).

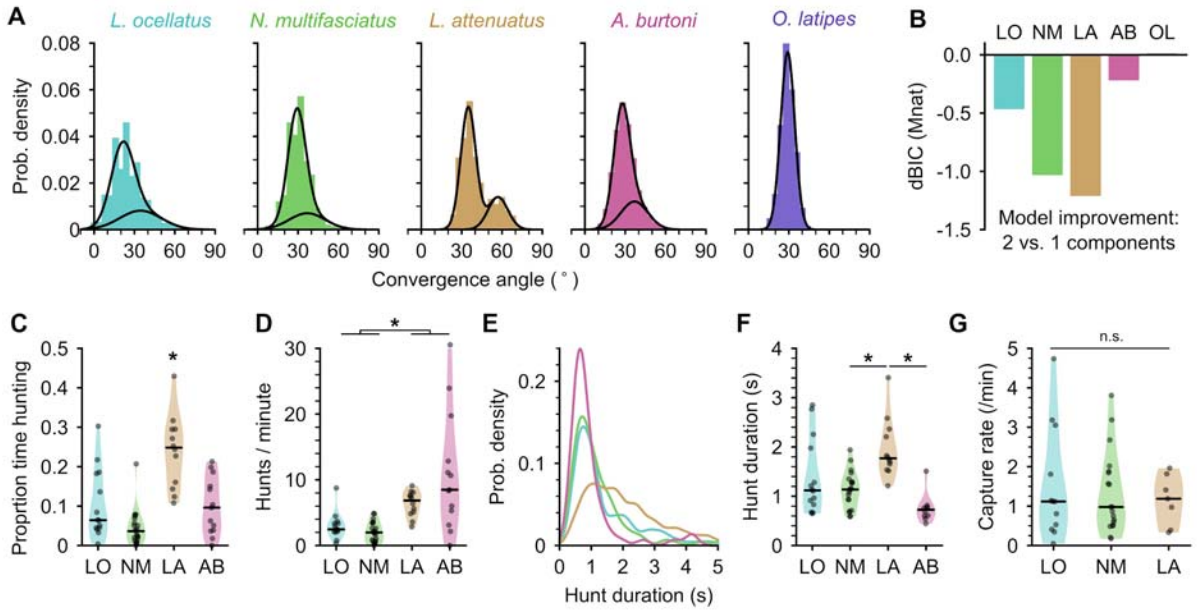

Figure 2. Eye movements during prey capture and statistics of hunting sequences. (**A**) Histograms of eye convergence angles for each species. The convergence angle is the angle between the long axes of the eyes. Shaded bars: normalized binned counts of convergence angles from all fish. Black lines: a best fit gaussian mixture model for each species (with one or two components). For all cichlid species, the data are better modeled as being drawn from two underlying distributions. For medaka (*O. latipes*), the data are better modeled with a single underlying distribution. (**B**) Improvement in fit for a two component over single component Gaussian mixture model, assessed using Bayesian inference criterion (BIC). The BIC is a measure of model fit, while punishing over-fitting. Lower values are better. Two mixtures provide a better fit over a single mixture for all species except medaka (OL). (**C**) Proportion of time spent by cichlids engaged in prey capture within the first five minutes of being introduced to the behavior arena. Points are single animals; black bar is the median. (**D**) Hunting rate, measured as the number of times eye convergence is initiated per minute, within the first five minutes. Points are single animals; black bar is the median. (**E**) Kernel density estimation of hunt durations for each species (all animals pooled). *A. burtoni* (pink) hunts skew shorter, *L. attenuatus* (brown) hunts tend to be longer. (**F**) Median hunt duration for each animal compared across species. Black bar is the median across animals. (**G**) Capture rate (number of artemia consumed) per minute over the first five minutes for three cichlid species. Black bar is the median across animals.

We found that all four cichlid species examined here converged their eyes during prey capture, but medaka did not (**Fig. 2A**). Two peaks in the distribution of eye convergence angles were visible in LA (**Fig. 2A**, center), but other cichlids also spent significant time with their eyes converged more than 50 degrees. To test whether two eye convergence states also existed in these species, we fitted Gaussian mixture models to the eye convergence data of each species

with one or two components. For all species except medaka, modeling the data using two underlying distributions provided a better fit than a single distribution (**Fig. 2B**). The unimodal distribution of eye vergence angles in medaka was also present when we plotted the joint distribution of left and right eye angles (**Supp. Fig. 2**), indicating that this species does not perform eye convergence.

Using eye convergence to identify hunting periods, we found that different cichlid species spent different amounts of time engaged in prey capture (**Fig. 2C**). LO, NM, and AB spent less than 10% of the time hunting (median proportions: 0.064, LO; 0.036, NM; 0.096, AB). In contrast, LA, a piscivorous species as adults, spent about a quarter of the time actively engaged in prey capture, with all individuals tested spending at least 10% of their time and one individual spending over 40% of the time hunting (median: 0.25, p-values < 0.05 comparing LA to all other species, Mann-Whitney U test with Bonferroni correction).

Surprisingly, although LO, NM and AB all spent comparable total time engaged in prey capture, AB initiated hunts at a much higher rate, similar to LA (**Fig. 2D**) (median hunts/min: 2.45, LO; 1.98, NM; 6.85, LA; 8.49, AB; p-values < 0.05 comparing either LO or NM to LA or AB, Mann-Whitney U test with Bonferroni correction). AB hunts were also shorter than other species, while LA hunts tended to be longer (**Fig. 2E-F**). LO and NM individual hunt durations were similar to each other and intermediate to the other species (medians across fish (seconds): 1.12, LO; 1.14, NM; 1.77, LA; 0.73, AB). Despite these differences in time engaged in prey capture, species consumed prey at similar rates of approximately one artemia per minute (**Fig. 2G**).

Taken together, our results highlight differences among closely related cichlids in hunting behavior. LA are the most persistent hunters, initiating prey capture often and spending a longer time engaged in the behavior once initiated (**Supp. Vid. 1**). In contrast, AB prey capture dynamics are characterized by a high rate of short-duration hunting episodes (**Supp. Vid. 4**). Both LA and AB prey capture behaviors are examples of an active hunting strategy. On the other hand, LO and NM initiate prey capture rarely, spending much time resting at the bottom of the chamber, and moving only occasionally when they dart towards prey (**Supp. Vid. 2&3**), characteristic of a sit-and-wait predation strategy.

<u>Cichlids center prey within a strike zone</u>

Zebrafish larvae strike at prey once it is localized in the center of their visual field and ~0.5 mm away (the “strike zone”) (Mearns et al., 2020; Patterson et al., 2013). This centering behavior is impaired when animals are blinded in one eye (Mearns et al., 2020). To test whether cichlids similarly center prey within a strike zone, we identified the most likely targeted artemia during each hunting episode and studied how the prey moved through the visual field as hunting sequences progressed (**Fig. 3A**, see **Methods**) (n events: 176, LO; 361, NM; 756, LA). This revealed that, within each prey capture sequence, prey became increasingly localized to the near central visual field over time. Computing kernel density estimates of prey distributions at the

beginning, middle, and end of each hunting episode revealed that cichlids initiate prey capture at a wide range of distances, and that they center prey in the visual field in the early stages of a hunting sequence (**Fig. 3B**). For instance, the azimuthal angle of prey only decreased over the start of the hunting episode (**Fig. 3D**) (p-value > 0.05, bootstrap test difference between medians at middle and end of prey capture). They then subsequently close the distance between themselves and their prey over the latter half of a hunt sequence (p-value < 0.001, bootstrap test difference between medians). On average, the cichlids' eyes converged when prey were ~5 mm away (median distance, mm: 5.43, LO; 3.98, NM; 4.17, LA). However, all species could detect prey up to 15 mm away (**Fig. 3C**). LA often initiated prey capture when artemia were already in the center of the visual field (**Fig. 3A,B&D**).

In all cichlid species analyzed, prey were highly localized to a central strike zone ~1-2 mm away at the end of prey capture (**Fig. 3B-C**). The tails of the distributions in **Fig. 3C** are likely due to a significant number of aborted hunting events. Together, these results demonstrate that cichlids are able to detect prey at a large distance and possess the fine sensorimotor control required to localize prey to a strike zone.

Fish larvae of different species share a common pose space

Zebrafish larvae have a unique swim repertoire during prey capture, which is distinct from exploratory swim bouts (Borla et al., 2002; McElligott & O'Malley, 2005). During prey capture, these swims each mediate distinct transformations of the visual scene: J-turns center prey in the visual field, approach swims bring prey to the strike zone, and strikes are deployed to capture prey. The identification and classification of these distinct swims has been aided by finding low-dimensional representations of behavior (Johnson et al., 2020; Marques et al., 2018; Mearns et al., 2020).

Due to neural and mechanical constraints, it is often possible to capture variation in tail shape over time with a greatly reduced number of dimensions, which serves to eliminate tracking noise and aid analysis (Berman et al., 2014; Mearns et al., 2020; Stephens et al., 2008; York et al., 2022). Performing principal components analysis (PCA) on all species combined revealed that tail pose dynamics are similarly low-dimensional (**Fig. 4A**, black line; 3 PCs explain > 90% variance). These results also hold when each species is analyzed individually (**Fig. 4A**, colored lines). As with zebrafish larvae (Girdhar et al., 2015; Mearns et al., 2020), the principal components correspond to a harmonic series (**Fig. 4B**), with the first PC capturing "turniness", and PCs 2 and 3 capturing tail oscillations during locomotion. The first three PCs were similar across species (**Fig. 4C**), suggesting that animals that share a common body plan exhibit similar low-dimensional pose-spaces.

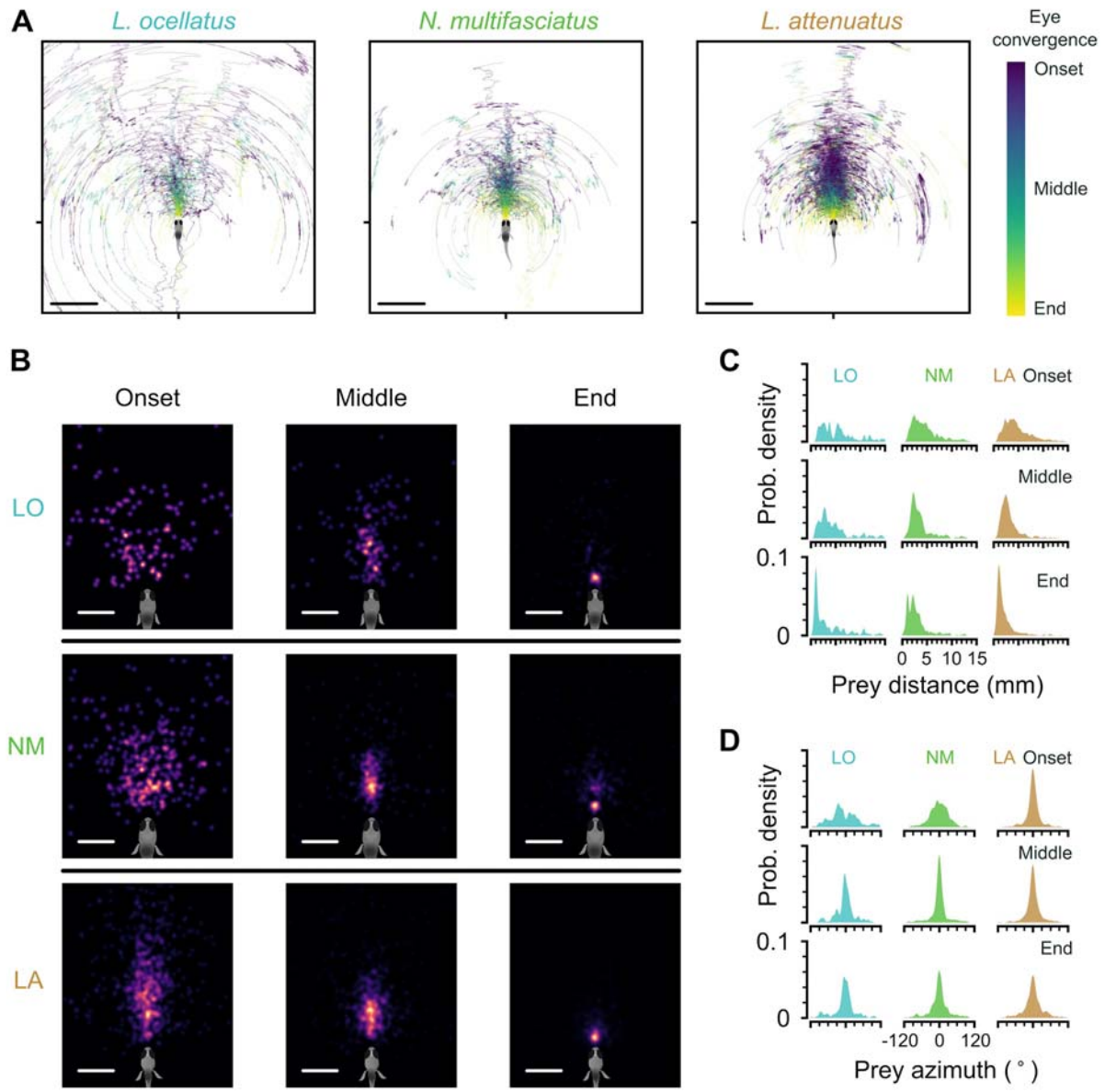

Figure 3. Location of prey in the visual field during prey capture in cichlids. (**A**) Trajectories of prey in the visual field for all automatically identified and tracked hunting events. Each prey is represented by a single line that changes in color from blue to yellow from the onset of eye convergence to when the eyes de-converge. Scale bar: 5 mm. (**B**) Kernel density estimation of the distribution of prey in the visual field across all hunting events. Rows: individual species. Columns: snapshots showing the distribution of hunted prey items at the beginning, middle and end of hunting sequences. Scale bar: 2 mm. (**C-D**) Kernel density estimation of prey distance (C) and azimuthal angle from the midline (D) at the onset (top), in the middle (center) and at the end (bottom) of hunting episodes. Each column shows the distribution from all events for a single species.

Hunting sequences in cichlids share kinematic motifs with zebrafish but are more complex

A common low-dimensional pose space in drosophilids has aided with the identification of evolutionary trajectories in behavior (York et al., 2022). We therefore wanted to investigate the trajectories through pose space of cichlids and medaka to understand if they share common behaviors during hunting , or if their behaviors have diverged over evolution.

For each species, we projected the time series of rostrocaudal tail angles for each distinct bout onto the first three principal components (Mearns et al., 2020). We then performed affinity propagation to identify clusters corresponding to different swim types and performed t-SNE to visualize these clusters in a low-dimensional embedding (**Fig. 4D,E, Supp. Fig. 3**). We found between 11 (LO) and 32 (AB) behavioral clusters per species. AB and LA held the richest behavioral repertoires, reflecting the more elaborate prey pursuit behaviors in these species (**Fig. 4E, Supp. Fig. 3**). The number of clusters we find is on par with or greater than the bout types that have been found in zebrafish larvae (Marques et al., 2018; Mearns et al., 2020).

Similar to zebrafish, we found that cichlids have specific kinematic motifs for prey capture (**Fig. 4F**). Some of these swims share characteristics with hunting bouts previously described in zebrafish, such as J-turns and approach swims, but we also identify new types that do not have clear analogs in zebrafish (**Fig. 4G**). For example, cichlid larvae often "hover" in place, oscillating their tail without any forward movement (**Fig. 4G**). Such behaviors tend to be longer lasting than approach swims. During hunting, cichlids will alternate between approaches and these hover swims.

Another feature of cichlid hunting behavior not present in zebrafish is tail coiling. Here, the tail coils into an S-shape over many hundreds of milliseconds leading up to a strike. The coil is released in a spring-loaded attack to capture prey (**Fig. 4I**). In addition to such capture springs, we have also observed suction captures and ram-like attack swims in cichlids.

Finally, we found that cichlids exhibit a range of post-strike maneuvers not present in zebrafish. These include swimming backwards to re-center prey in the visual field in cases where capture strikes miss (**Supp. Vid. 7**), and expelling prey ("spitting") that has already been captured (**Supp. Vid. 6**). Spitting behavior often triggered another attempt to hunt and capture the same prey, and was particularly common in LA. This behavior likely explains the high incidence of hunt initiations when prey are already in the central visual field in this species (**Fig. 3A,B&D**). The function of spitting is not clear, and it might indicate that the prey are slightly too large for LA to swallow. Alternatively, such a behavior may possibly be adaptive, providing already sated animals the opportunity to practice hunting without overeating. We also find that cichlid captures often occur in two phases, with the prey first being caught by the pharyngeal jaws (a second, internal set of jaws present in cichlids; Liem 1973) before being ingested. Together, these results reveal a richer behavioral repertoire for pursuing, capturing, and reorienting towards missed prey in cichlids than in zebrafish.

Cichlids share two common capture strikes that are distinct from medaka side-swing strikes

Studies in invertebrates have demonstrated that behavior can evolve through changing the kinematics of a behavior performed in a given sensory context (Ding et al., 2016), changing which sensory cues drive behavior (Seeholzer et al., 2018), or changing the transition frequencies between a common set of kinematic motifs (Hernández et al., 2021). It is not known to what extent these factors play a role in the evolution of vertebrate behaviors, nor the evolutionary timescales over which behaviors diverge to the point of becoming distinct. To address this question, we focused on the capture strike kinematics of cichlids and medaka. The capture strike of zebrafish larvae has been extensively characterized, consisting of two distinct types: the attack swim and S-strike. These strikes are deployed variably, dependent on experience and different prey distances (Lagogiannis et al., 2020; McClenahan et al., 2012; Mearns et al., 2020; Patterson et al., 2013).

Medaka swimming is nearly continuous, and their eyes do not converge during prey capture. Nevertheless, medaka may still perform different behaviors during spontaneous swimming and prey capture. Investigating clusters of behavior in medaka revealed types of swim that are similar to prey capture bouts of cichlids and zebrafish. These included J-turns, approach swims, and slow swims (**Fig. 4H**). The presence of these swims suggests that medaka, like other species studied, may also have a separate suite of behaviors reserved for prey capture. Remarkably, we found that medaka did not strike at prey in the central visual but rather captured prey in the lateral visual field with a unique side-swing behavior (**Fig. 4H-J**). Although the eyes do not converge, there is sometimes a slight nasalward rotation of the ipsilateral eye leading up to and during the side swing (**Fig. 4I**).

To confirm that side-swing behavior was unique to medaka, we clustered capture strikes of cichlids and medaka after projecting their tail kinematics into the common pose space, setting the number of clusters to the number of species (**Fig. 4A**). We reasoned that if bouts were similar within each species but different between species, cluster identity would correlate with species identity. In contrast, if species shared similar kinematic motifs, clusters would contain bouts from all species. We found that strikes from all cichlid species were similar to each other, while the medaka side-swing strike was unique (**Fig. 4K**).

This analysis uncovered at least two kinematically distinct capture strikes in cichlids. The slower of these behaviors is similar to the zebrafish attack swim, characterized by a symmetric undulation of the tail about the midline which propels the fish towards the prey, while the faster corresponds to the capture spring. The S-shape of the tail leading up to the capture spring shares structural similarities with the S-strike of zebrafish larvae (Mearns et al., 2020), differing primarily in the time course over which they occur. The S-strike of zebrafish may represent an accelerated form of the capture spring, forming and releasing over tens of milliseconds rather

than hundreds of milliseconds seen in cichlids or, vice versa, the capture spring may represent an augmentation of the S-strike.

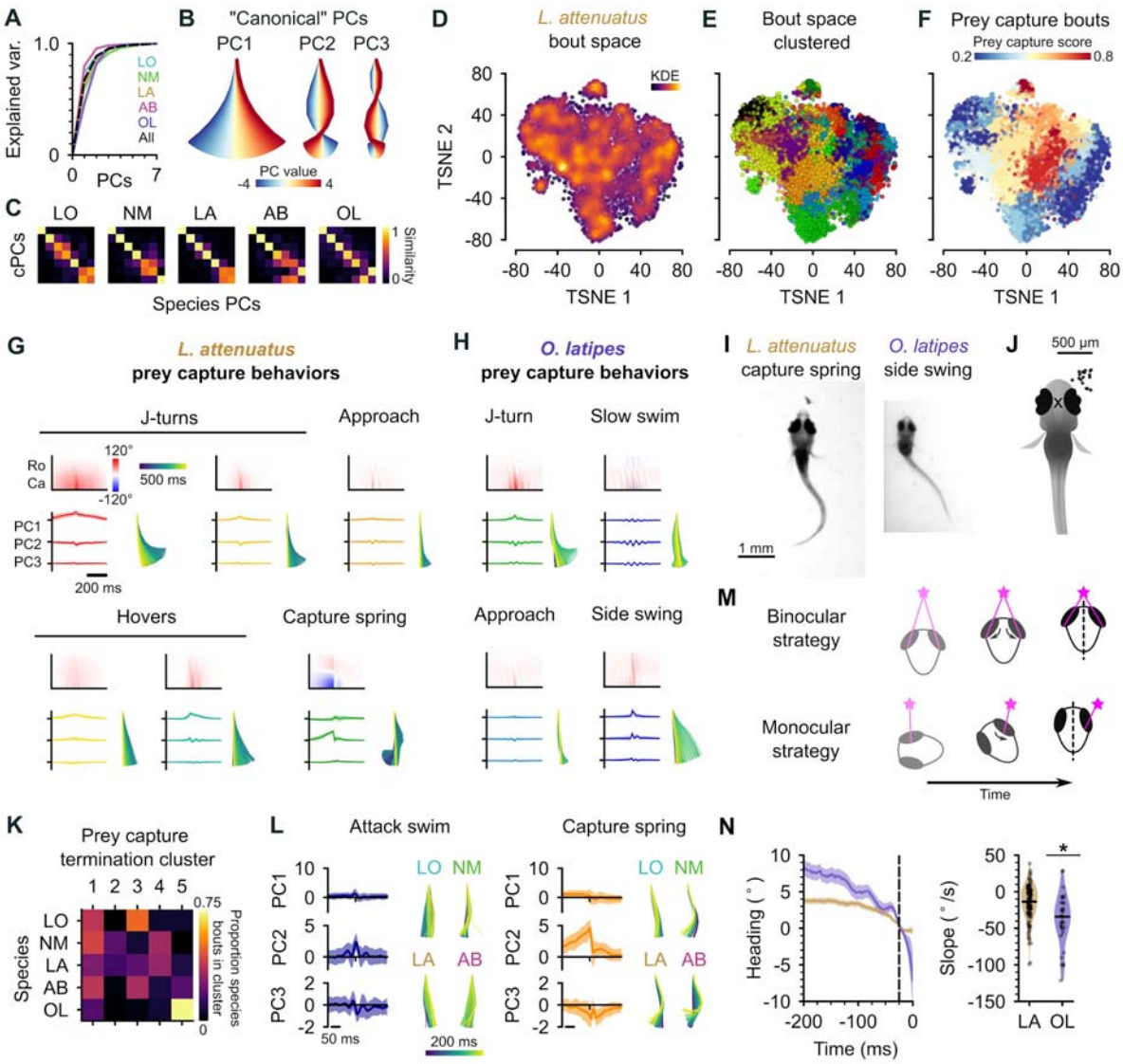

Figure 4. Interspecies comparison of prey capture strategies. (**A**) Cumulative explained variance for the "canonical" principal components (PCs) obtained from all species (black dotted line), and PCs for each species individually (colored lines). In all cases, three PCs explain >90% of the variance in tail shape. (**B**) "Eigenfish" of the first three canonical PCs. Each principal component represents a vector of angles from the base to the tip of the tail (oriented with the fish facing up). At a given moment, the shape of the tail can be described as a linear combination of these vectors. Colors correspond to the tail shape obtained by scaling each PC from -4 to 4 standard deviations from the mean. (**C**) Each species' eigendecomposition compared against the canonical PCs computed for all species together. Color intensity represents the cosine similarity between pairs of vectors. The strong diagonal structure (particularly in the first three PCs) shows that similar PCs are obtained by analyzing species separately or together. (**D**) Behavioral space

of *L. attenuatus*. Each point represents a single bout. Bouts are projected onto the first three PCs, aligned to the peak distance from the origin in PC space and then projected into a two-dimensional space using TSNE. Color intensity represents density of surrounding points in the embedding. (**E**) Clustered behavioral space of *L. attenuatus*. Clusters (colors) are computed via affinity propagation independently of the embedding. (**F**) Prey capture and spontaneous bouts in *L. attenuatus*. Prey capture score is the probability that the eyes are converged at the peak each bout. Bouts are colored according to the mean prey capture score for their cluster. Blue: clusters of bouts that only occur during spontaneous swimming; red: clusters of bouts that only occur during prey capture. (**G-H**) Example prey capture clusters from *L. attenuatus* (G) and medaka, *O. latipes* (H). For each cluster, top left: mean rostrocaudal bending of the tail over time; bottom left: time series of tail pose projected onto first three PCs (mean +/- standard deviation); bottom right: reconstructed tail shape over time for mean bout. (**I**) Representative frames of an *L. attenuatus* capture spring (left) and medaka side swing (right), highlighting differences in tail curvature between these behaviors. (**J**) Location of prey in the visual field (black dots) immediately prior to the onset of a side swing. All events mirrored to be on the right. X marks the midpoint of the eyes, aligned across trials. (**K**) Confusion matrix of clustered hunt termination bouts from all species. Termination bouts from all species were sorted into five clusters based on their similarity. Rows show the proportion of bouts from each species that were assigned to each cluster (columns). Cichlid bouts are mixed among multiple clusters, while medaka bouts (OL) mostly sorted into a single cluster. (**L**) Capture strikes in cichlids. Representative examples of tail kinematics during attack swims (left) and capture springs (right) from each species, including time series of tail pose projected onto the first three PCs (mean +/- standard deviation, all species combined) shown for each type of strike. (**M**) Two hypotheses for distance estimation make different predictions of how heading (black dotted line) changes over time as fish approach prey (pink star). Top: fish maintain prey in the central visual field and use binocular cues to judge distance. Bottom: fish "spiral" in towards prey, using motion parallax to determine distance. Black arrows indicate motion of prey stimulus across the retina. (**N**) Change in heading over time leading up to a capture strike for *L. attenuatus* (brown) and medaka (purple). Left: time series of heading. Zero degrees represents the heading 25 ms prior to the peak of the strike (black dotted line). Mean ± s.e.m. across hunting events. Right: comparison of the rate of heading change, computed as the slope of a line fit to each hunting event 200-25 ms prior to the peak of the strike. The heading decreases more rapidly leading up to a strike in medaka than in *L. attenuatus*.

Different prey capture strategies may reflect different sensorineural solutions to prey detection

Centralized prey and converged eyes in cichlids suggest that the mechanism they use to determine the distance to prey could be similar to that of zebrafish. One possibility is that these species use binocular disparity to judge depth (Qian, 1997). In contrast, we hypothesized that medaka might use a different, monocular strategy to judge the distance to prey, such as motion parallax (Yoonessi & Baker, 2011). These two mechanisms of depth perception make different predictions about how larvae should approach prey to make best use of visual cues (**Fig. 4M**). To investigate these possibilities, we computed the change in heading leading up to a strike as a proxy for the change in visual angle of the prey for cichlids and medaka (**Fig. 4N**). We found that the rate of change in heading was greater in medaka than in cichlids (median degrees/second: -13.4, LA; -34.1, OL; $p < 0.001$, Mann-Whitney U-test). These results are consistent with cichlids using binocular cues, such as binocular disparity, to determine distance to prey, while medaka may employ a monocular strategy.

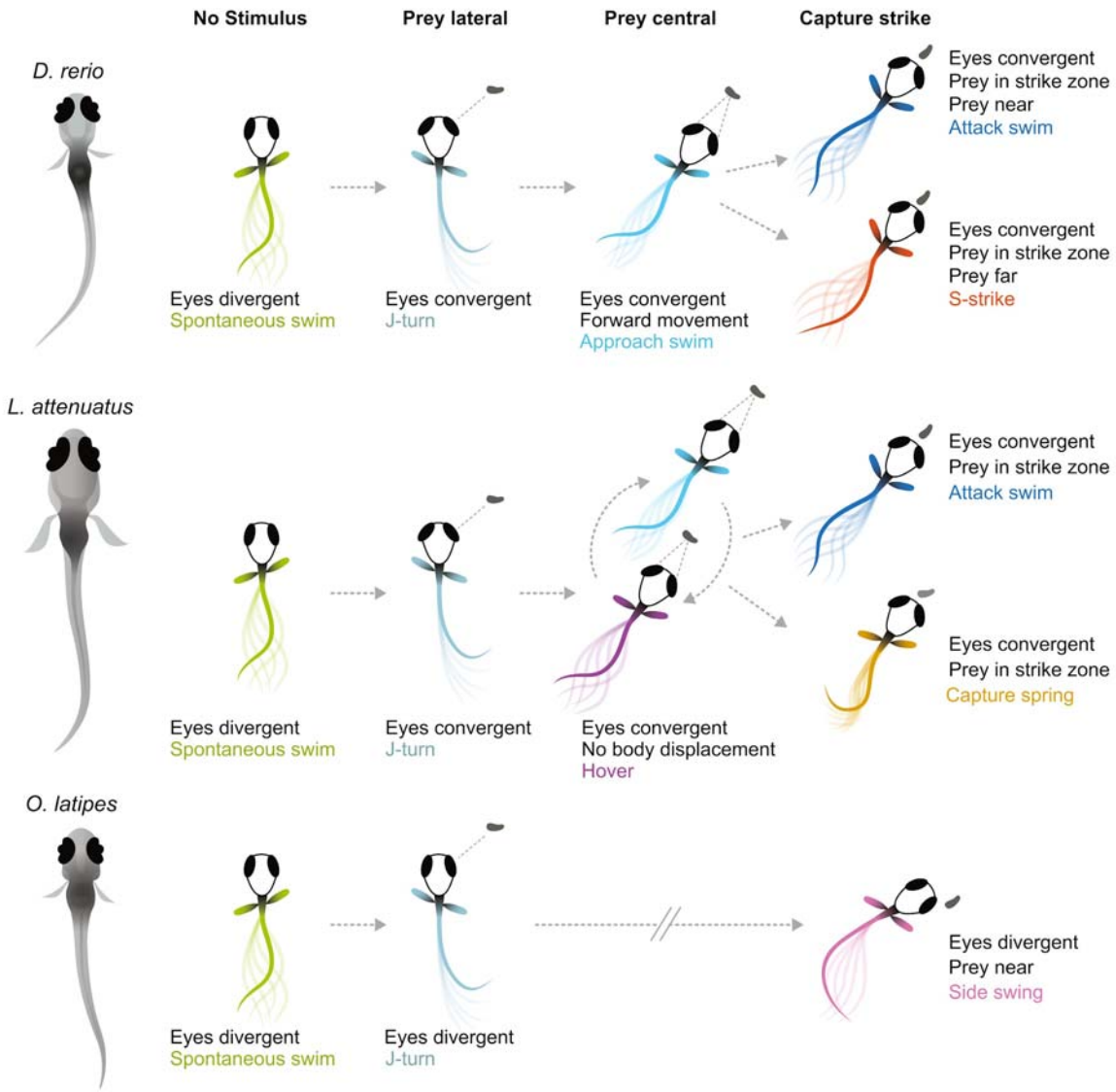

Figure 5. Schematic of hunting strategies in teleost larvae.
Top: prey capture in zebrafish larvae (*Danio rerio*) begins with eye convergence and a J-turn to orient towards the prey. The prey is then approached with a series of low amplitude approach swims. Once in range in the central visual field, the prey is captured with an attack swim or an S-strike. Middle: prey capture in cichlids (represented by *L. attenuatus*) also begins with eye convergence and a J-turn. Prey is approached with a wide variety of tail movements. The prey is captured when it is in the central visual field with either an attack swim, or a capture spring, during which the tail coils over several hundreds of milliseconds. Bottom: prey capture in medaka begins with reorienting J-like turns, but these do not centralize prey in the visual field. Instead, the prey is kept lateral in the visual field and is captured with a side swing.

Discussion

We have found that zebrafish and cichlids share the same general hunting strategy: striking at prey localized to a binocular strike zone while the eyes are converged, with two distinct capture bout types whose kinematics are remarkably similar given these species diverged 240 million years ago (see **Fig. 1A**). In contrast, medaka, which are phylogenetically intermediate to zebrafish and cichlids, deploy a different hunting strategy: striking at prey laterally in the monocular visual field with a side-swing behavior. **Fig. 5** provides a schematic summary of similarities and differences among the teleost larvae investigated. Characterization of prey capture behavior in a wider range of species could reveal whether the similarities between zebrafish and cichlids represent conservation of an ancestral hunting strategy or convergent evolution.

What sensorineural mechanisms might underlie these two different hunting strategies? The side swing of medaka is reminiscent of hunting in (adult) blind cavefish (Lloyd et al., 2018), which position prey laterally prior to strikes using their mechanosensory lateral line. Adult cichlids are known to use their lateral line for prey capture (Schwalbe et al., 2016), and some percomorph species exhibit both an S-shaped and a sideways capture strategy as adults. Therefore, it is conceivable that medaka larvae do not rely on vision to the same extent for hunting, but use their lateral line instead. However, lateral-line use is not uncoupled from eye convergence across the teleost clade. Zebrafish larvae can still hunt in the dark, as do blind zebrafish *lakritz* mutants, albeit with greatly reduced efficacy, and with their eyes converged (Gahtan et al., 2005; Mearns et al., 2020; Patterson et al., 2013). Early-stage cave fish, which evolved from sighted surface ancestors and are completely blind, exhibit vestigial eye convergence movements during prey capture before their eyes fully degenerate (Espinasa et al., 2023; Espinasa & Lewis, 2023). This comparative evidence suggests that the teleost brain employs cues from both sensory modalities, if available, to locate prey, but that eye movements are a poor predictor of the dominance of vision.

On the other hand, species that attack centrally located prey could be using different visual cues to judge prey distance than those that use a side swing to ingest food: binocular disparity (Qian, 1997) vs. motion parallax (Yoonessi & Baker, 2011). Zebrafish use binocular information (Gahtan et al., 2005; Gebhardt et al., 2019; Henriques et al., 2019) and stationary differences such as brightness and contrast (Khan et al., 2023) to estimate depth and distance. Here we have shown that cichlid larvae also likely use binocular cues, while medaka's approach to prey conforms to a monocular strategy (see **Fig. 4M,N**). Strikingly, cichlid and zebrafish larvae swim in bouts with intermittent pauses. We speculate that the pauses may serve to sample the distance to the prey by comparing its position across the two eyes. In contrast, to sample motion cues, it is advantageous to perform continuous glides ("motorboating") and swim lateral to the prey up until it is close enough for a sideways strike. Thus, natural selection may have favored specific locomotor (bouts vs. glides) and oculomotor (convergent vs. divergent) adaptations of behavioral

control depending on the dominant distance measurement mechanism (binocular vs. monocular) (**Fig. 5**). While the behavioral evidence for this conjunction is still circumstantial and calls for further comparative work, this scenario makes testable predictions about its neurobiological and genetic implementation.

We have demonstrated that eye convergence is not always a hallmark of prey capture in fish larvae, and that multiple, kinematically distinct capture swims exist at these early stages in different species. We speculate that ocular vergence angles and swim kinematics for prey capture might co-evolve, with S-shaped captures and attack swims being associated with convergent eyes and side-swing captures occuring in the absence of eye convergence. These two strategies may have been present in a common ancestor of teleosts, with one or the other becoming dominant in certain lineages. Comparing neural circuitry across these (now) experimentally tractable animals could help pinpoint evolvable circuit nodes ("hotspots" of evolutionary change; Roberts et al., 2022; Seeholzer et al., 2018) and genetic loci that underlie prey localization strategies and swim kinematics in fishes.

Author contributions

D.S.M. and H.B. conceived the project. D.S.M. and M.W.S. collected the data. S.A.H. trained models and performed tracking. A.V.P. and M.S. maintained cichlid colonies and provided larvae for experiments. D.S.M. and S.A.H. analyzed the data. D.S.M. and H.B. wrote the manuscript with input from all authors.

Competing interests

No competing interests declared.

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References

Baier, H., & Scott, E. K. (2009). Genetic and optical targeting of neural circuits and behavior—Zebrafish in the spotlight. *Current Opinion in Neurobiology*, *19*(5), 553–560. https://doi.org/10.1016/j.conb.2009.08.001

Berman, G. J., Choi, D. M., Bialek, W., & Shaevitz, J. W. (2014). Mapping the stereotyped behaviour of freely moving fruit flies. *Journal of The Royal Society Interface*, *11*(99), 20140672. https://doi.org/10.1098/rsif.2014.0672

Betancur-R, R., Wiley, E. O., Arratia, G., Acero, A., Bailly, N., Miya, M., Lecointre, G., & Ortí, G. (2017). Phylogenetic classification of bony fishes. *BMC Evolutionary Biology*, *17*(1), 162. https://doi.org/10.1186/s12862-017-0958-3

Bianco, I. H., Kampff, A. R., & Engert, F. (2011). Prey Capture Behavior Evoked by Simple Visual Stimuli in Larval Zebrafish. *Frontiers in Systems Neuroscience*, *5*, 101. https://doi.org/10.3389/fnsys.2011.00101

Borla, M. A., Palecek, B., Budick, S., & O'Malley, D. M. (2002). Prey Capture by Larval Zebrafish: Evidence for Fine Axial Motor Control. *Brain Behavior and Evolution*, *60*(4), 207–229. https://doi.org/10.1159/000066699

Budick, S. A., & O'Malley, D. M. (2000). Locomotor repertoire of the larval zebrafish: Swimming, turning and prey capture. *The Journal of Experimental Biology*, *203*(Pt 17), 2565–2579. https://doi.org/10.1242/jeb.203.17.2565

Devineni, A. V., & Scaplen, K. M. (2022). Neural Circuits Underlying Behavioral Flexibility: Insights From Drosophila. *Frontiers in Behavioral Neuroscience*, *15*. https://www.frontiersin.org/articles/10.3389/fnbeh.2021.821680

El Taher, A., Böhne, A., Boileau, N., Ronco, F., Indermaur, A., Widmer, L., & Salzburger, W. (2021). Gene expression dynamics during rapid organismal diversification in African cichlid fishes. *Nature Ecology & Evolution*, *5*(2), 243–250. https://doi.org/10.1038/s41559-020-01354-3

Espinasa, L., Diamant, R., Vinepinsky, E., & Espinasa, M. (2023). Evolutionary modifications of Astyanax larval prey capture (LPC) in a dark environment. *Zoological Research*, *44*(4), 750–760. https://doi.org/10.24272/j.issn.2095-8137.2022.466

Espinasa, L., & Lewis, K.-A. (2023). Eye convergence is evoked during larval prey capture (LPC) without visual stimulus and in blind cavefish. *Subterranean Biology*, *46*, 47–60. https://doi.org/10.3897/subtbiol.46.105707

Frey, B. J., & Dueck, D. (2007). Clustering by Passing Messages Between Data Points. *Science*, *315*(5814), 972–976. https://doi.org/10.1126/science.1136800

Friedrich, R. W., Jacobson, G. A., & Zhu, P. (2010). Circuit Neuroscience in Zebrafish. *Current Biology*, *20*(8), R371–R381. https://doi.org/10.1016/j.cub.2010.02.039

Gahtan, E., & Baier, H. (2004). Of lasers, mutants, and see-through brains: Functional neuroanatomy in zebrafish. *Journal of Neurobiology*, *59*(1), 147–161. https://doi.org/10.1002/neu.20000

Gahtan, E., Tanger, P., & Baier, H. (2005). Visual prey capture in larval zebrafish is controlled by identified reticulospinal neurons downstream of the tectum. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, *25*(40), 9294–9303. https://doi.org/10.1523/JNEUROSCI.2678-05.2005

Gebhardt, C., Auer, T. O., Henriques, P. M., Rajan, G., Duroure, K., Bianco, I. H., & Del Bene, F. (2019). An interhemispheric neural circuit allowing binocular integration in the optic tectum. *Nature Communications*, *10*(1), Article 1. https://doi.org/10.1038/s41467-019-

13484-9
Girdhar, K., Gruebele, M., & Chemla, Y. R. (2015). The Behavioral Space of Zebrafish Locomotion and Its Neural Network Analog. *PLOS ONE*, *10*(7), e0128668. https://doi.org/10.1371/journal.pone.0128668
Henriques, P. M., et al. (2019). Nucleus isthmi is required to sustain target pursuit during visually guided prey-catching. *Current Biology 29*(11), 1771-1786.
Hernández, D. G., Rivera, C., Cande, J., Zhou, B., Stern, D. L., & Berman, G. J. (2021). A framework for studying behavioral evolution by reconstructing ancestral repertoires. *eLife*, *10*, e61806. https://doi.org/10.7554/eLife.61806
Higham, T. E., Hulsey, C. D., Říčan, O., & Carroll, A. M. (2007). Feeding with speed: Prey capture evolution in cichilds. *Journal of Evolutionary Biology*, *20*(1), 70–78. https://doi.org/10.1111/j.1420-9101.2006.01227.x
Johnson, R. E., Linderman, S., Panier, T., Wee, C. L., Song, E., Herrera, K. J., Miller, A., & Engert, F. (2020). Probabilistic Models of Larval Zebrafish Behavior Reveal Structure on Many Scales. *Current Biology*, *30*(1), 70-82.e4. https://doi.org/10.1016/j.cub.2019.11.026
Khan, B., et al. (2023). Zebrafish larvae use stimulus intensity and contrast to estimate distance to prey. *Current Biology 33* (15), 3179-3191.
Liem, K. F. (1973). Evolutionary strategies and morphological innovations: cichlid pharyngeal jaws. *Systematic Zoology 22*(4), 425-441.
Lloyd, E., Olive, C., Stahl, B. A., Jaggard, J. B., Amaral, P., Duboué, E. R., & Keene, A. C. (2018). Evolutionary shift towards lateral line dependent prey capture behavior in the blind Mexican cavefish. *Developmental Biology*, *441*(2), 328–337. https://doi.org/10.1016/j.ydbio.2018.04.027
Maaten, L. van der, & Hinton, G. (2008). Visualizing Data using t-SNE. *Journal of Machine Learning Research*, *9*(86), 2579–2605.
Mano, H., & Tanaka, Y. (2012). Size specificity of predation by Japanese medaka Oryzias latipes on Daphnia pulex. *Journal of Freshwater Ecology*, *27*(2), 309–313. https://doi.org/10.1080/02705060.2012.659222
Marques, J. C., Lackner, S., Félix, R., & Orger, M. B. (2018). Structure of the Zebrafish Locomotor Repertoire Revealed with Unsupervised Behavioral Clustering. *Current Biology: CB*, *28*(2), 181-195.e5. https://doi.org/10.1016/j.cub.2017.12.002
Mathis, A., Mamidanna, P., Cury, K. M., Abe, T., Murthy, V. N., Mathis, M. W., & Bethge, M. (2018). DeepLabCut: Markerless pose estimation of user-defined body parts with deep learning. *Nature Neuroscience*, *21*(9), Article 9. https://doi.org/10.1038/s41593-018-0209-y
McElligott, M. B., & O'Malley, D. M. (2005). Prey Tracking by Larval Zebrafish: Axial Kinematics and Visual Control. *Brain Behavior and Evolution*, *66*(3), 177–196. https://doi.org/10.1159/000087158
Mearns, D. S., Donovan, J. C., Fernandes, A. M., Semmelhack, J. L., & Baier, H. (2020). Deconstructing Hunting Behavior Reveals a Tightly Coupled Stimulus-Response Loop. *Current Biology*, *30*(1), 54-69.e9. https://doi.org/10.1016/j.cub.2019.11.022
New, J. G. (2002). Multimodal Integration in the Feeding Behaviors of Predatory Teleost Fishes. *Brain Behavior and Evolution*, *59*(4), 177–189. https://doi.org/10.1159/000064905
Orger, M. B., & de Polavieja, G. G. (2017). Zebrafish Behavior: Opportunities and Challenges. *Annual Review of Neuroscience*, *40*(1), 125–147. https://doi.org/10.1146/annurev-neuro-

071714-033857
Patterson, B. W., Abraham, A. O., MacIver, M. A., & McLean, D. L. (2013). Visually guided gradation of prey capture movements in larval zebrafish. *The Journal of Experimental Biology*, *216*(Pt 16), 3071–3083. https://doi.org/10.1242/jeb.087742
Pereira, T. D., Aldarondo, D. E., Willmore, L., Kislin, M., Wang, S. S.-H., Murthy, M., & Shaevitz, J. W. (2019). Fast animal pose estimation using deep neural networks. *Nature Methods*, *16*(1), Article 1. https://doi.org/10.1038/s41592-018-0234-5
Pereira, T. D., Tabris, N., Matsliah, A., Turner, D. M., Li, J., Ravindranath, S., Papadoyannis, E. S., Normand, E., Deutsch, D. S., Wang, Z. Y., McKenzie-Smith, G. C., Mitelut, C. C., Castro, M. D., D'Uva, J., Kislin, M., Sanes, D. H., Kocher, S. D., Wang, S. S.-H., Falkner, A. L., … Murthy, M. (2022). SLEAP: A deep learning system for multi-animal pose tracking. *Nature Methods*, *19*(4), Article 4. https://doi.org/10.1038/s41592-022-01426-1
Piggott, B. J., Liu, J., Feng, Z., Wescott, S. A., & Xu, X. Z. S. (2011). The Neural Circuits and Synaptic Mechanisms Underlying Motor Initiation in C. elegans. *Cell*, *147*(4), 922–933. https://doi.org/10.1016/j.cell.2011.08.053
Portugues, R., & Engert, F. (2011). Adaptive Locomotor Behavior in Larval Zebrafish. *Frontiers in Systems Neuroscience*, *5*. https://www.frontiersin.org/articles/10.3389/fnsys.2011.00072
Qian, N. (1997). Binocular disparity and the perception of depth. *Neuron*, *18*(3), 359–368. https://doi.org/10.1016/s0896-6273(00)81238-6
Rajan, G., Lafaye, J., Faini, G., Carbo-Tano, M., Duroure, K., Tanese, D., Panier, T., Candelier, R., Henninger, J., Britz, R., Judkewitz, B., Gebhardt, C., Emiliani, V., Debregeas, G., Wyart, C., & Del Bene, F. (2022). Evolutionary divergence of locomotion in two related vertebrate species. *Cell Reports*, *38*(13), 110585. https://doi.org/10.1016/j.celrep.2022.110585
Redmon, J., Divvala, S., Girshick, R., & Farhadi, A. (2016). *You Only Look Once: Unified, Real-Time Object Detection* (arXiv:1506.02640). arXiv. https://doi.org/10.48550/arXiv.1506.02640
Roberts, R. J. V., Pop, S., & Prieto-Godino, L. L. (2022). Evolution of central neural circuits: State of the art and perspectives. *Nature Reviews. Neuroscience*, *23*(12), 725–743. https://doi.org/10.1038/s41583-022-00644-y
Ronco, F., Matschiner, M., Böhne, A., Boila, A., Büscher, H. H., El Taher, A., Indermaur, A., Malinsky, M., Ricci, V., Kahmen, A., Jentoft, S., & Salzburger, W. (2021). Drivers and dynamics of a massive adaptive radiation in cichlid fishes. *Nature*, *589*(7840), Article 7840. https://doi.org/10.1038/s41586-020-2930-4
Schwalbe, M. A. B., Sevey, B. J., & Webb, J. F. (2016). Detection of artificial water flows by the lateral line system of a benthic feeding cichlid fish. *Journal of Experimental Biology*, *219*(7), 1050–1059. https://doi.org/10.1242/jeb.136150
Seeholzer, L. F., Seppo, M., Stern, D. L., & Ruta, V. (2018). Evolution of a central neural circuit underlies Drosophila mate preferences. *Nature*, *559*(7715), 564–569. https://doi.org/10.1038/s41586-018-0322-9
Stephens, G. J., Johnson-Kerner, B., Bialek, W., & Ryu, W. S. (2008). Dimensionality and Dynamics in the Behavior of C. elegans. *PLOS Computational Biology*, *4*(4), e1000028. https://doi.org/10.1371/journal.pcbi.1000028
Tinbergen, N. (1951). *The study of instinct* (pp. xii, 237). Clarendon Press/Oxford University

Press.
Westphal, R. E., & O'Malley, D. M. (2013). Fusion of locomotor maneuvers, and improving sensory capabilities, give rise to the flexible homing strikes of juvenile zebrafish. *Frontiers in Neural Circuits*, *7*, 108. https://doi.org/10.3389/fncir.2013.00108
Wiltschko, A. B., Johnson, M. J., Iurilli, G., Peterson, R. E., Katon, J. M., Pashkovski, S. L., Abraira, V. E., Adams, R. P., & Datta, S. R. (2015). Mapping Sub-Second Structure in Mouse Behavior. *Neuron*, *88*(6), 1121–1135. https://doi.org/10.1016/j.neuron.2015.11.031
Yoonessi, A., & Baker, C. L. (2011). Contribution of motion parallax to segmentation and depth perception. *Journal of Vision*, *11*(9), 13. https://doi.org/10.1167/11.9.13
York, R. A., Brezovec, L. E., Coughlan, J., Herbst, S., Krieger, A., Lee, S.-Y., Pratt, B., Smart, A. D., Song, E., Suvorov, A., Matute, D. R., Tuthill, J. C., & Clandinin, T. R. (2022). The evolutionary trajectory of drosophilid walking. *Current Biology*. https://doi.org/10.1016/j.cub.2022.05.039
Zhu, S. I., & Goodhill, G. J. (2023). From perception to behavior: The neural circuits underlying prey hunting in larval zebrafish. *Frontiers in Neural Circuits*, *17*, 1087993. https://doi.org/10.3389/fncir.2023.1087993

Supplementary Materials

Supplementary Figure 1: Diversity of swimming behavior in teleost larvae. Left: representative five minutes of swimming from each species. Black bar indicates a ten second window expanded on the right. Colors indicated automatically segmented and classified bouts within each swimming episode.

Supplementary Figure 2: 2D histograms of eye angles. Each point represents the angle of the left and right eyes in a given frame. Color corresponds to the probability a point belongs to a convergent state (red: converged; blue: not converged; yellow: equal likelihood).

Supplementary Figure 3: Behavioral spaces for each species. Left: t-SNE embedding and cluster identity of automatically segmented bouts. Right: example clusters from each species, showing average time series in first three PCs (top), rostrocaudal tail bending (middle), and tail kinematics (bottom) for each cluster. For cichlids, behaviors on the right are more likely to occur during prey capture and behaviors on the left are more likely to occur during spontaneous swimming.

Supplementary video 1: Example of *L. attenuatus* freely swimming and hunting artemia, played at half speed.

Supplementary video 2: Example of *L. ocellatus* freely swimming and hunting artemia, played at half speed.

Supplementary video 3: Example of *N. multifasciatus* freely swimming and hunting artemia, played at half speed.

Supplementary video 4: Example of *A. burtoni* freely swimming and hunting artemia, played at half speed.

Supplementary video 5: Example of medaka (*O. latipes*) freely swimming and hunting paramecia, played at half speed. Notice the side-swings of the head during prey capture and the lack of eye convergence.

Supplementary video 6: *L. attenuatus* spitting behavior, played at half speed.

Supplementary video 7: *A. burtoni* swimming backwards during prey capture, played at half speed.