Bacteria face trade-offs in the decomposition of complex biopolymers
Running title: Trade-offs in degradation of polymers

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Although depolymerization of complex carbohydrates is a growth-limiting bottleneck for microbial decomposers, we still lack understanding about how the production of different types of extracellular enzymes affect individual microbes and in turn the performance of whole decomposer communities. In this work we use a theoretical model to evaluate the potential trade-offs faced by microorganisms in biopolymer decomposition which arise due to the varied biochemistry of different depolymerizing enzyme classes. We specifically consider two broad classes of depolymerizing extracellular enzymes, which are widespread across microbial taxa: exo-enzymes that cleave small units from the ends of polymer chains and endo-enzymes that act at random positions generating degradation products of varied sizes. Our results demonstrate a fundamental trade-off in the production of these enzymes which can be explained by intrinsically different temporal depolymerization dynamics. As a consequence, specialists that produce either exo- or only endo-enzymes limit their growth to high or low substrate conditions, respectively. Conversely, generalists that produce both enzymes in an optimal ratio expand their niche and benefit from the synergy between the two enzymes. Finally, our results show that, in spatially-explicit environments, consortia composed of endo- and exo-specialists can only exist under oligotrophic conditions. In summary, our analysis demonstrates that the (evolutionary or ecological) selection of a depolymerization pathway will affect microbial fitness under low- or high substrate conditions, with impacts on the ecological dynamics of microbial communities. It provides a possible explanation why many polysaccharide degraders in nature show the genetic potential to produce both of these enzyme classes.

Keywords: Microbial interactions, polymer degradation, microbial communities, extracellular enzymes, public-goods, polysaccharides

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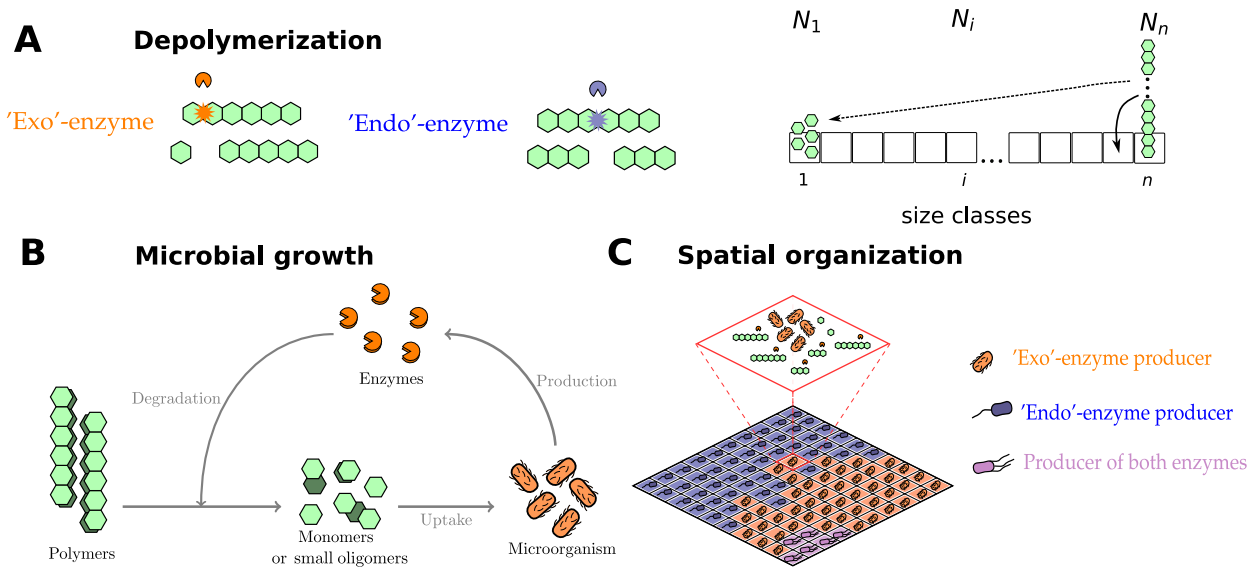


FIG. 1. **Features included in the microbial degradation model.** (A) Depolymerization dynamics due to activity of exo or endo-enzymes. On the right we show the division of polysaccharides into different classes (from 1 to n) according to their length (N_1 representing the concentration of monomers – they all belong to class 1, N_i represents the concentration of a oligomer of size i). Arrows represent possible fluxes between these size classes which result from enzymatic reactions. (B) Microbial growth dynamics, which includes depolymerization of a complex substrate, uptake of small subunits (monomers) and enzyme production. (C) Schematic representation of the spatial grid model, where each microsite represents a microhabitat with a small microbial population. This population can colonize neighboring habitats after its biomass riches a certain threshold.

I. INTRODUCTION

The decomposition of organic matter by microbes is a rate-limiting process in the global carbon cycle [49]. Across all ecosystems, this process is governed by microbially derived extracellular enzymes, sometimes also known as eco-enzymes [49]. This is because organic matter is mostly composed of large structural polysaccharides such as cellulose or chitin, which cannot be taken up directly by cells, and must be broken into smaller pieces by extracellular enzymes before utilization [4]. On the one hand, since these polysaccharides possess a diverse range of crystalline structures and linkages, one type of enzyme is often insufficient to efficiently process them. On the other hand, the production of each additional enzyme adds to the metabolic costs of microorganisms, requiring a careful assessment of the expected return on such investment [26, 40, 45, 48]. For challenging environments with high molecular diversity and spatio-temporal heterogeneity, such as soil, enzyme production can pose a great burden on microbial metabolism [31], which would be exacerbated if microorganisms produce multiple enzyme classes for each nutrient source [53]. Despite these constrains we know numerous examples of microorganisms which display genetic potential to produce a variety of enzymes involved in depolymerization and examples of enzymes with consecutive, redundant, or even synergistic actions [6, 22, 28, 52]. Moreover, it has been shown that microbial populations go through diversification depending on the chain length, concentration and solubility of the polymer found in their surrounding environment [21]. Hahemann et al observed that trough adaptive radiation a population of closely related bacteria subdivides into specialized ecophysiological types adapted to the given environmental opportunities, showing a great flexibility and variability of the depolymerization pathways and hinting to the existence of trade-offs for the production of the enzymes engaged in depolymerization.

The traditional view of degradation of complex substrates assumes that at the core of this process we find a series of depolymerization steps catalyzed by glycoside hydrolases (EC 3.2.1.*, e.g. cellulose and chitin) or other lyases (e.g. alginate). The hydrolysis itself can proceed in two different ways: where small products are cleaved from the chain ends (effectuated by exo-enzymes); or where a polymer molecule is randomly broken at any position (as done by endo-enzymes), see Fig. 1A. These enzymes appear to be redundant to a certain degree, since both have the same objective – production of small units ready for uptake.

Before we engage with the main research question proposed in this study, we made a survey to illustrate the prevalence of different strategies in polymer degrading microorganisms, differentiating specialists – producers exclusively of

endo or exo enzymes, from generalists – producers of both enzyme types. Although a large diversity of carbohydrate-active enzymes (CAZymes) are known, the exact functions of most of them have never been tested biochemically. Chitinases are an informative test-case and the action of different chitinase types are depicted in Fig. 2 [38]. In this view endo-chitinases are assumed to cleave any bond, while the exo-chitinases act by cutting off dimers from the ends of the polymeric chain. Monomers are then generated through the activity of N-acetylglucosaminidases (also known as chitobiases) that specifically cleave dimers into monomers. According to their sequence diversity and evolutionary origin, microbial chitinases belong to three major glycoside hydrolases (GH) families: 18, 19 and 20 [1, 8, Note2]. Proteins belonging to the GH18 and GH19 families are mostly extracellular and display both endo- and exo-chitinase activity [5, 8], while GH20 family proteins are mainly periplasmic N-acetylglucosaminidases (chitobiases).

A potential source of information regarding the prevalence of endo and exo-chitinases in microorganisms can be environmental genomic data. However, predicting such strategies from genomes turned out to be extremely challenging. Functional classifications, e.g., Enzyme Commission (EC) numbers or KEGG identifiers, are still evolving and have ambiguous information about exo/endo activity. The KEGG database defines 3 main coarse-grained chitinase groups: K01183 (mainly GH18), K01207 (chitobiase, GH20), and K03791 (mainly GH19). We checked for these identifiers in the genome annotations of bacteria from all environments (using Anotree) and of soils only (JGI), see SM for details. According to these databases, chitobiase (N-acetylglucosaminidase) is widespread in bacteria (Fig. 6B) and predominantly can be the only type of chitinase that a bacteria has. Conversely, the other two types of sequences were found in distinct but overlapping groups (Fig. 2C). Notably K03791 (GH19) was mostly found in Proteobacteria and was absent in Firmicutes. Although such analysis illustrates that microorganisms exhibit a plethora of diverse chitinase combinations, it still fails to provide a proper comparison of prevalence of exo and endo-activity.

To complement our sequence-based overview we performed a literature review of experimentally characterized microorganisms where endo/exo-chitinase activity has been distinguished (see details in the Supplement). We find again that large part of chitin degraders seem not to specialize in a single enzyme class of chitinases, showing both exo- and endo-activity types (Fig. 2D). Together, our sequence- and literature based analyses raise a series of questions, such as:(i) What are the advantages of producting different enzyme classes at the same time? (ii) Which trade-offs in the production of different enzyme classes are faced by microorganisms individually or within communities? And (iii) how do these trade-offs affect microbial growth in different conditions, especially in complex environments like soils?

Despite its importance, depolymerization has so far not been explicitly included in current microbial models [19, 24] or into the models of organic matter decomposition in general [42, 43, 46, 47]. In this work we intend to bridge this gap, by including the different forms of hydrolysis into a microbial growth model [24, 25], see Fig. 1B. This allows us to theoretically analyse the possible trade-offs faced by microorganisms growing on biopolymers and strategies found in nature. We assume that potential trade-offs may be different depending whether microbes live in well mixed or spatially explicit environments, so we include both in our analysis (Fig. 1C). In spatially structured systems, which host a significant portion of microbes on earth (e.g. soil) [10], the diffusive exchange of products is known to play a central role in self-organization within consortia [11, 13, 29, 30, 37]. To model such systems, we incorporate polymer degradation by means of population balance equations into an individual-based spatially explicit microscale microbial community model. This approach allows us to track the spatial distribution of poly and oligosaccharide molecules of different sizes in space and time, as well as the microbes feeding on them. Using a theoretical approach, we analyse the advantages and the disadvantages of these two depolymerization types and discuss possible synergies among them.

II. METHODS

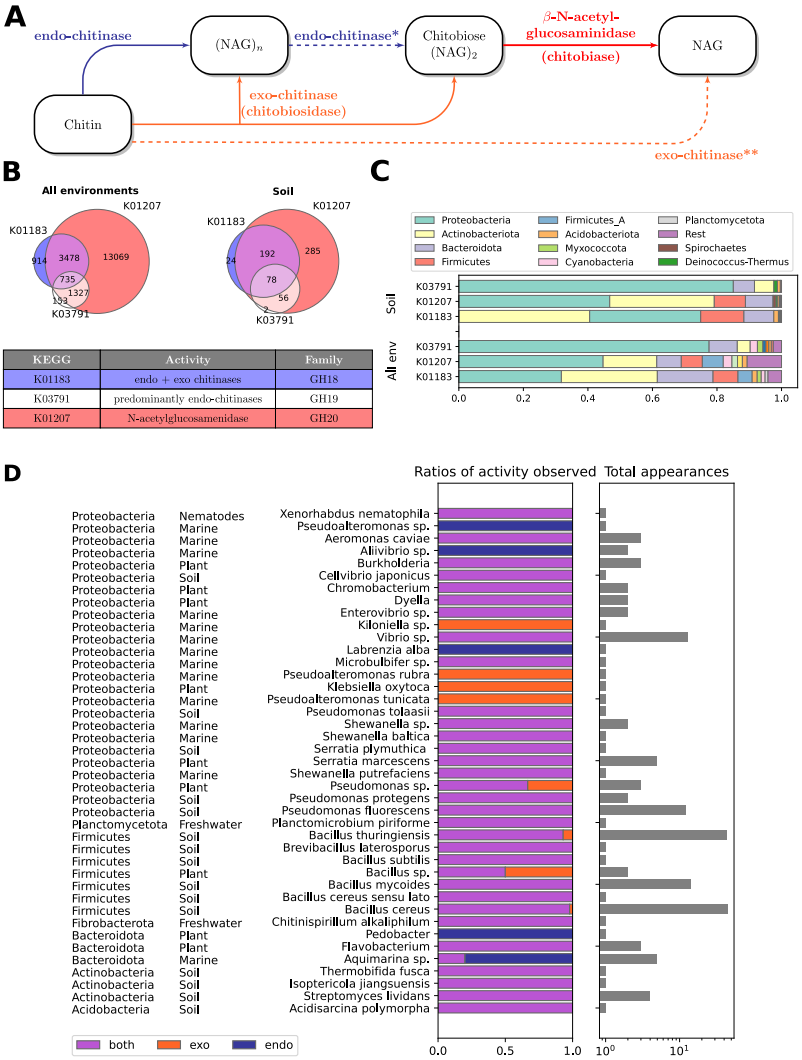
Enzyme kinetics and population balance equations (PBE). As explained previously the exo-enzymes E_{exo} cleaves long polymeric chain N_i into small organic molecules (monomers, N_1) which can be taken up by the cell, following:

$$\mathrm{E_{exo}} + \mathrm{N}_i \underset{\mathrm{k_2}}{\overset{\mathrm{k_1}}{\rightleftharpoons}} (\mathrm{E_{exo}N}_i) \xrightarrow{\mathrm{k_3}} \mathrm{E_{exo}} + \mathrm{N}_{i-1} + \mathrm{N}_1 \tag{1}$$

On the other hand endo-enzymes E_{endo} are able to cleave any bond:

$$\mathrm{E_{endo}} + \mathrm{N}_{i+j} \underset{\mathrm{k_2}}{\overset{\mathrm{k_1}}{\rightleftharpoons}} (\mathrm{E_{endo}N}_{i+j}) \xrightarrow{\mathrm{k_3}} \mathrm{E_{endo}} + \mathrm{N}_i + \mathrm{N}_j \tag{2}$$

We model the degradation of chitin by means of population balance equations (PBE), following the framework proposed by Suga et al [50]. This approach divides the available polymers in groups depending on their size (monomrs, dimers, trimers, n-mers, etc), we denote the number of polymers of size i as N_i. The number of polymers of each size class evolves, $\frac{dN_i}{dt}$ following the fragmentation dynamic established by the enzyme which catalyzes the reaction, see



KEGG	Activity	Family
K01183	endo + exo chitinases	GH18
K03791	predominantly endo-chitinases	GH19
K01207	N-acetylglucosamenidase	GH20

FIG. 2. **Mode of action and distribution of different classes of chitinases.** (A) Schematic representation of the hydrolysis of chitinase into NAG (N-acetylglucosamine), which involves: endo- and exo-chitinases, as well as a N-acetylglucosaminidase. *Endo-chitinases are also able to cut dimers from the chain however this reaction may have a different turnover rate. **There is a potential for certain types of exo-chitinases to cleave directly monomers from the long chains of polymers. (B, C) Distribution of chitinases within different microbiomes, the chitinases were classified according to genetic sequence. (D) Recordings of exo or endo-chitinase activity in chitin degrading microorganisms, the data is based on a literature survey (details in the SM). We indicate the number of strains used for the classification (total appearances), and the origin of the bacteria (e.g. marine, soil, carnivorous plant etc).

SM. This dynamics decreases the numbers of polymers of one class and correspondingly adds to other size classes. One important property of the system is that the total amount of monomers in the system is conserved $\sum_{i=1}^{n} iN_i = M$. We use Michaelis-Menten kinetics for both enzyme types; the reaction rate v_{exo_i} of exo-enzyme is given by

$$v_{exo_i} = k_{exo} \frac{E_{exo} N_i}{k_m + \sum_{j=1}^{n} N_j}; \tag{3}$$

and the reaction rate for the endo enzyme is:

$$v_{endo_i} = k_{endo} \frac{(i-1) E_{endo} N_i}{k_m + \sum_{j=1}^{n} (j-1) N_j}, \tag{4}$$

in both equations the k_m is the half saturation rate of both enzymes, which is set to 0.5 nmol/mm^3. The turnover

number (catalytic constant) of the two enzymes are given by k_{exo} and k_{endo}. For exo-enzymes k_{exo} is fixed to 0.82 nmol nmol C^{-1} h^{-1}. Conversely the turnover number of endo-enzymes (k_{endo}) can be similar or lower than of exo-enzymes depending on the settings.The full set of equations used can be found in the SM.

Microbial growth dynamics Microorganisms are modeled following the dynamics established in [24, 25]. In this model microorganisms grow by taking up label substrate, released trough the degradation of large polysacharides by extracellular enzymes, produced by these microorganisms (the general framework is presented in Fig. 1B). This approach has the advantage of explicitly including the stoichiometric C and N fluxes. The microbial metabolism is divided into uptake, maintenance, enzyme production and growth. Microorganisms are able to uptake only the smallest substrate units (with a fixed C:N ratio characteristic of the substrate CN_s), in our case those are monomers, although these settings can be extended to include other sizes as well. Additional nitrogen is taken up from dissolved inorganic nitrogen pool (DIN). All uptake is modeled accounting for receptors on the cell membrane, for details see Supplementary Material. A fraction of carbon from the uptake is used for maintenance. Another fraction of carbon together with nitrogen is used for enzyme production (CN_e, fixed C:N ratio of enzymes). The rest is used for growth (CN_m fixed C:N ratio characteristic to microbes). Each metabolism step, such as growth and enzyme production, includes an additional fraction of respired carbon. If there are leftovers of carbon or nitrogen those are respired or eliminated, respectively. In the absence of nutrients in the environment the microbial biomass decreases due to a requirement of maintenance. We set up a lower limit, after which the microorganism is considered dead. Random death events are also included in the model. The enzymes, on the other hand, have a fixed lifetime, established by their decay rate r_e. Additional details and parameters used can be found in the SM.

Spatially explicit dynamics. The spatial model is also an extension of [24, 25]. The spatial model consists of a 100×100 spatial grid representing a 1 mm^2 soil area. Each microsite represented in this grid is considered to be 10 μm^3. Each grid cell may contain: a single type of microorganisms, enzymes, substrate (in the form of polymers of all sizes) and dissolved inorganic nitrogen. It is important to emphasize that we consider that everything is well mixed within a single microsite. Microorganisms are not allowed to move but can colonize the neighboring sites by division/reproduction. When microbial biomass within a microsite exceeds a certain limit, the biomass is divided in half, half of this biomass is transferred to an unoccupied neighboring site. If all the surrounding sites are occupied the division does not occur, and the biomass continues growing. Enzymes are also transferred together with microorganisms; we assume that they are attached to the cell wall and do not diffuse away. Finally, polymers of size larger than 10, are insoluble and do not diffuse. Each one of the smaller sizes diffuses. We consider that the diffusion rate of each n-mer depends on its size, taking into account Rouse-Zimm relation for polymer diffusion [12] we can write the diffusion of an oligomer of size i as $D_i = D_0(i^{-1/x})$ with $x = 1$, where D_0 is the diffusion rate of the monomer. All parameters can be found in the SM.

III. RESULTS

Degradation by endo-enzymes is characterized by a nonlinear monomer release rate. In our first simulation scenario we aimed to characterized the activity of enzymes without the presence of microorganisms. For that we introduced a pool of substrate and a small, but fixed, concentration of endo or exo-enzymes and recorded the degradation dynamics. Our results show that the main difference between the two enzyme types during degradation is the rate of monomer release. In the absence of microbes, exo-enzymes show a constant rate of monomer release (Fig. 3A, B). This rate depends on the number of polymer chains, with larger number of chains leading to faster degradation (Fig. 3B). Note that for exo-enzymes the number of polymer chains never changes and as degradation proceeds, with each monomer cleaved out, each one of these chains becomes shorter and shorter. For the endo-enzymes, on the other hand, the release rate is not constant: the process starts with only a few monomers produced, but this number increases nonlinearly with time. Moreover, the duration of this initial monomer production lag depends on the concentrations of polymers, with high concentrations leading to fewer monomers released in the starting period (Fig. 3C). Despite having this initial delay, the endo-enzymes are much more efficient in the presence of long chains, as we show in Fig. 3A, where we show that for a chain size of $n = 200$ the total time necessary to degrade the total polymer pool T_{deg} under endo-activity is significantly shorter. This difference in T_{deg} between endo and exo-enzymes decreases for small chains, see Fig. S3 in Supplementary Material. In summary, the model shows that specialists producing exo or endo-enzymes would experience different monomer release rates. Next we discuss how these differences are reflected on the microbial growth of such specialists.

Specializing in exo or endo-enzyme production decreases the ecological niche of microorganisms. Although having both endo and exo-enzyme types may appear redundant, since they are simply different ways to degrade the same substrate, our results show that specializing in one or the other enzyme production can lead to significant disadvantages. To understand them, we added microbial growth dynamics to the enzyme model. First we analyze the microbial growth in well mixed conditions. We initialize the population with a small amount of enzymes

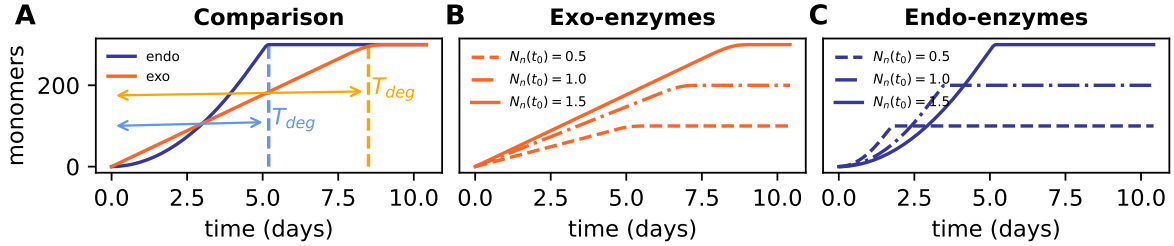


FIG. 3. **Differences in the dynamics of release of monomers by exo and endo-enzymes.** (A) Comparison between depolymerization dynamics of exo and endo enzymes, the system starts with M = 300 nmol/mm^3 of monomers all connected into chains of size n = 200 (i.e. the concentration of polymers in the initial pool is $N_n(t_0) = 1.5$ nmol/mm^3). T$_{deg}$ indicates the time until the available pool of substrate is fully degraded. Note T$_{deg}$ is shorter for endo-enzymes. (B) Monomer release by exo- (in concentration $C_e = 3$ nmol C/mm^3) and (C) endo-enzymes ($C_e = 3$ nmol C/mm^3), with same turnover rates $k_{exo} = k_{endo} = 0.82$ nmol nmol C^{-1} h^{-1}, for different initial concentrations of polymers (in nmol /mm^3) of size n= 200, note the differences in the final amount of monomers obtained. The solid lines in (A, B, C) represent the monomer release dynamics for identical initial pools.

(in an analogous way the population can be initialized with a small amount of monomers). This is necessary to trigger enzyme production within the population which shortly after becomes either self-sustained or dies out [39].

In well mixed conditions exo-producers grow well only in high nutrient concentrations (Fig. 4A). For them the quantity of polymer chains, rather than their size, becomes the limiting factor for nutrient uptake. If the uptake rate is insufficient for the maintenance and further enzyme production of microorganisms, they will not survive, which is evident in the low-nutrient conditions (Fig. 4A). Endo-producers face opposite challenges and cannot grow in high nutrient concentrations. While environments rich with polymers potentially offer enough nutrients, these chains must first be broken down to monomers for the uptake, a process which usually takes multiple steps for endo-enzymes. As we have shown in the previous subsection larger nutrient amounts result in longer delays before sufficient monomers appear to sustain endo-enzyme producers (Fig. 3C). If this delay is too long, the microorganisms cannot grow under those conditions, leading to population collapse (Fig. 4A). In other words there is an intrinsic time period during which the microbes are able to wait for the return on their investment into enzymes. If they do not get this return on time the population is unable to survive.

Most microbial communities do not experience well-mixed conditions, and must deal with the challenges imposed by spatial structure. In this context, an important constrain is the loss of breakdown products due to diffusion. We measured the extent of this loss and its effect on growth by using a spatially explicit model, in which we tracked the biomass of a single microsite (i.e. a grid cell), with microorganisms that are unable to divide and spread to neighboring sites but oligomers which can diffuse away (Fig. 4B, C). We observed that while exo-producers are only mildly affected, endo-producers lose nearly half of their potential biomass even when diffusion is weak (Fig. 4B, C). To be able to understand if this loss may affect the spatial self-organization of the microbial community we followed with further spatial analysis, now allowing microbial reproduction and spatial spread of the population. We have chosen a diffusion rate (3 μm^2/h) that allows endo-producers to also accumulate sufficient biomass for reproduction to occur. In the spatial context, endo-producers are still limited to nutrient-poor environments, while exo-producers thrive in conditions with high polymer availability (Fig. 4D, E). Additionally, the varying nutrient levels have an impact on the spatial organization pattern. Since endo-producers have limited growth, they expand in a narrow ring as the population consumes all the nutrients during habitat expansion (Fig. 4E). On the other hand, exo-producers take longer to deplete all the available resources and move forward before the microsite is fully depleted (Fig. 4D). Our findings also indicate a significant delay before endo-producers develop into a sizable population, and their spread through the grid is considerably slower.

Advantages of being a generalist: niche expansion and synergy. Our results suggest that microorganisms might benefit from producing both endo- and exo-enzymes simultaneously. To assess whether this is the case, we examine the effects of varying the proportions of produced endo ((1 - fr_{exo}) and exo-enzyme (fr_{exo}) fractions in microbes able to produce both enzymes. As before, first we address the catalytic efficiency of a constant pool with these two enzymes, which we uncouple from microbial growth. Fig. 5 compares the degradation efficiency, evaluated trough the degradation time T_{deg} necessary to degrade the whole substrate pool into monomers, considering different enzyme fractions. In a process where the two types of enzymes have the same turnover rates, the endo-enzymes finish to degrade the pool of polymers faster, with smaller T_{deg} (Fig. 5A), especially when longer chains are present. When considering the more characteristic scenario where endo-enzymes have longer intrinsic turnover rates than exo-enzymes a different picture emerges. In such cases, combining endo and exo-enzymes can result in a synergistic effect that speeds up degradation, particularly when the two enzymes exist in a specific proportion ($fr_{exo} \sim 0.8$, Fig. 5B). Note

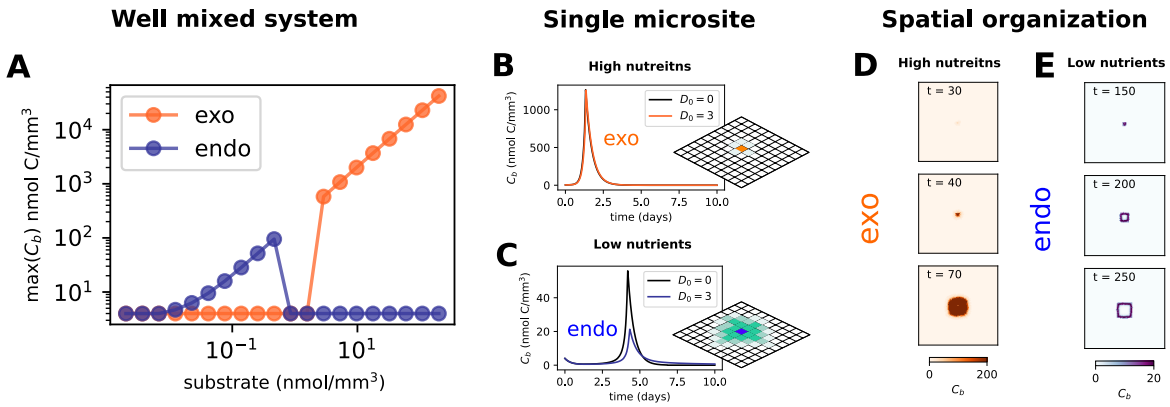


FIG. 4. **Differences between endo and exo-enzyme producers: their growth conditions; how much products they loose to diffusion and how they organize in space.** We show results for degradation of polysacharide chains of size $n = 500$ by microorganisms in different set-ups. For comparison, the turnover rates of the two enzymes are the same, $k_{exo} = k_{endo} = 0.82$ h^{-1} (A) Maximum biomass (max(($C_b(t)$) achieved during degradation of different pools of polymers in well mixed conditions. The results show that high nutrient conditions hinder the growth of endo-producers and promote the growth of exo-producers. (B, C) Time evolution of biomass $C_b(t)$ in a single microsite of a spatially explicit model. We assume conditions where microorganisms are unable to divide, but oligomers up to size 10 (see Methods) are able to diffuse. We compare the settings with no diffusion ($D_0 = 0$) and with diffusion ($D_0 = 3\ \mu m^2/h$). (D, E) Snapshots of the spatial distribution of microbes (biomass concentration C_b(x, y, t)) at indicated times. High nutrient concentration: M= 1500 nmol/mm^3; Low nutrient concentration: M = 150 nmol/mm^3.

that this effect is prominent only for the case when monomer pool is trapped into longer chains and the exo-enzyme activity is limited by the availability of polymer ends (see $n = 500$ in Fig. 5B).

Our results show that having the ability to produce the two enzyme classes also allows microorganisms to grow at an expanded range of nutrient concentrations compared to enzyme specialists (Fig. 5C). The proportion in which exo and endo-enzymes are produced is also important in this context, as we show in Fig. 5C.

Finally, diffusion of breakdown products away from microorganisms can also affect the performance of generalists. We quantify this loss by comparing the total biomass of microbes in a single microsite after growth with no diffusion (max(C_b^*)) to the biomass in the presence of diffusion (max(C_b), Fig. 5D). Our simulations reveal that carbon loss increases dramatically with the fraction of endo-chitinase produced, in both high and low nutrient conditions. Overall our simulations reveal that producing both enzymes classes can result in synergistic effects and a broader niche for generalists. However this faster degradation dynamics compared to pure exo-enzyme producers also comes at the cost of increased diffusion-mediated carbon loss.

Conditions for the formation of a consortium of endo and exo producers. In the previous section we considered a single generalist organism producing both enzyme types. A population composed of both exo- and endo-enzyme producing specialists will have both enzyme types, analogous to a generalist organism. We used simulations to study whether the two specialist types can co-exist, and the influence of mixing types on population dynamics. In the case of unequal turnover rates of the two enzymes, as in the previous section the coexistence of the two taxa (of exo and endo producers) is not possible. The faster turnover rate for exo producers gives them an uneven advantage, and they rapidly dominate in all conditions. When the turnover rates are the same, however, coexistence is possible in low nutrient conditions (Fig. 6B, C). We find that the two types spatially segregate but still influence each other. The growth of the consortium is significantly faster compared to pure endo-specialists. In high nutrient conditions, the consortium is quickly dominated by exo-producers, which share little of produced monomers resulting in the extinction of endo-producers (Fig. 6A).

IV. DISCUSSION

We have presented a novel framework to model growth of heterotrophic microbes in the presence of large polymers, which explicitly accounts for the depolymerization dynamics. Our results demonstrate that microbial growth depends not only on the amounts of carbon present in the system but also on the form in which this carbon is found, specifically the polymer chain size. We describe the key features of the degradation by exo and endo-enzyme types and the trade-offs faced by microorganisms specializing in the production of a specific enzyme class. Our model shows

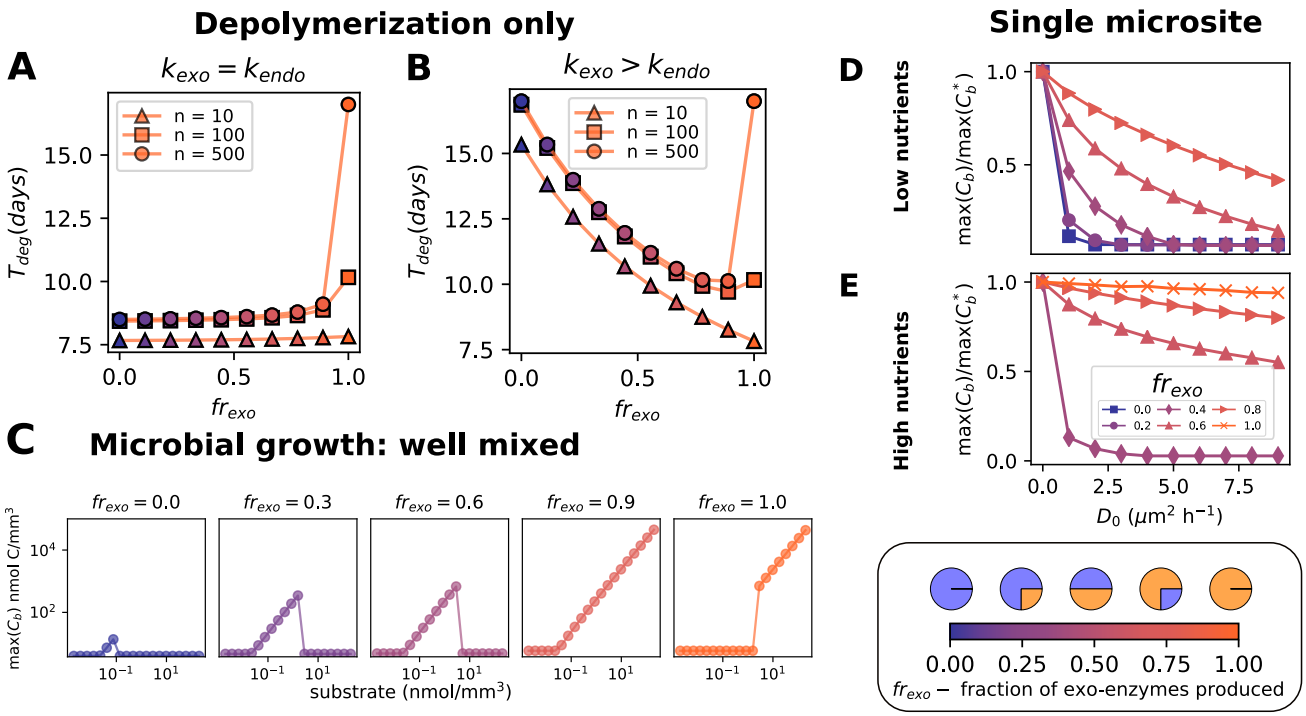


FIG. 5. **Advantages of generalists (producers of both endo and exo enyzmes).** (A, B) Comparison of the total degradation time (T_{deg}) of different proportions of exo and endo-enzymes acting simultaneously. The simulation represents only the depolymerization dynamics in the absence of microbes in homogeneous conditions. The complex substrate pool was initiated with a total of 500 nmol/mm^3 monomers distributed between chains of n-mers (n indicated in the figure labels). A pool with 3 nmol C/mm^2 of two enzymes in different proportions act on the substrate (x-axis and color gradient with purple indicating endo-enzymes and orange indicating exo-enzymes), while the turnover rate or exo-enzyme is $k_{exo} = 0.82$ h^{-1}, the turnover of endo-enzyme is (A) $k_{endo} = k_{exo}$ and (B) $k_{endo} = k_{exo}/2$. (C) Maximum biomass reached from different initial concentrations of polymers for microorganisms producing different fractions of endo and exo enzymes. The parameters are the same as the ones used in simulation in Fig. 4, except for the turnover of endo-enzymes, which is $k_{endo} = k_{exo}/2$. (D, E) Comparison of the maximum growth in the absence of diffusion max(C_b^*) to the maximum growth in the presence of diffusion max(C_b), for different diffusion coefficients D_0 for (D) low, M= 150 nmol/mm^3, and (E) high nutrients, M= 1500 nmol/mm^3. Each curve represents a different proportion of exo to endo-enzymes (fr_{exo}). Note that for low diffusion the ratios of maximum possible biomass would be the same, and with an increase of diffusion the growth decreases due to loss of soluble oligomers.

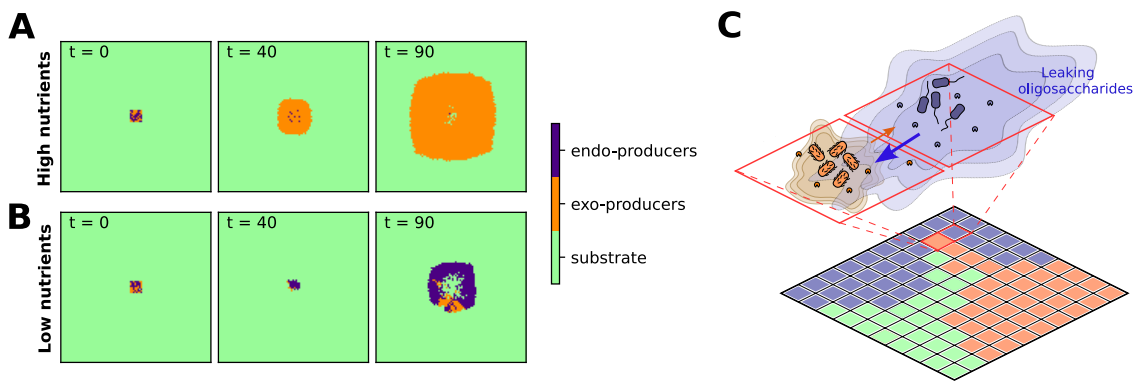


FIG. 6. **Time evolution of consortia of endo and exo producers:** (A) in high nutrient conditions initiated with nutrients concentration of 1500 nmol/mm^3 and (B) in low nutrients condition initiated with 150 nmol/mm^3. The initial carbon is distributed in chains of size $n = 500$. Cooperation is only possible when the turnover rates of the two enzymetypes are comparable, $k_{exo} = k_{endo} = 0.82$ h^{-1}. (C) schematic representation of two interactions within colonies trough the diffusion of degradation products (note that each microsite can be occupied by a single type of microorganism). While endo-producers share more oligomers with their neighbors, exo-producers consume most of the degradation products within a single microsite.

that specialization constrains growth potential. Moreover, our results demonstrate that these disadvantages can be overcome by generalists, that produce both enzymes in a specific ratio. All trade-offs are summarized in Tab. I. Our findings are in agreement with the overwhelming presence of generalists among chitin degraders identified by enzyme activity assays, see Fig. 2 or examples in [8, 9, 44], and thus offer a possible explanation for this observations.

Our work has reveled intrinsic trade-offs with deep consequences for microbial community assembly and organic matter decomposition neglected by previous model-based studies. Current models that target microbial growth [1, 2, 19, 24] or organic matter turnover [46, 47], oversimplify degradation of complex substrates by assuming that the outcome of each reaction between enzymes and polysaccharides necessarily produces units ready for uptake. This indirectly implies kinetics which can only result from exo-enzyme activity. We show that the choice of depolymerization pathways lead to fundamental differences for the microbial activity at the small scale, with impact on organization of microbial communities. Notably, the implication of these trade-offs for large scales are still to be uncovered.

At the center of our findins are the contrasing requirements for growth of exo and endo-enzyme producers. While exo-enzyme producers need a large number of polymer molecules, endo-producers find rich conditions challenging due to a concentration-dependent delay in their ability to produce monomers. Besed on this findings, exo and endo-enzyme producers could thus be tentatively classified as copiotrophs and oligotrophs, respectively. Such classification, however, would not exactly be in agreement with the traditional oligotrophs-copiotrophs categories, which are based on the metabolic trade-off between the microbial resource acquisition and growth [18, 27, 33]. In this conventional view copiotrophs are organisms that highly invest in growth, reproduce quickly and for this reason do well in nutrient rich environments. Conversely, oligotrophs specialize to survive in nutrient low conditions by highly investing in resource acquisition. Note that exo and endo-producers analyzed in our work, apart their investment into different enzymes, share identical metabolic strategies (i.e. they invest in the same way in resource acquisition and growth). Moreover, the two types would perform similarly in the presence of simple substrates (monomers, dimers or other small oligomers) and the difference is only observable in the presence of complex substrates.

The spatial simulations show additional restrictions for endo-producers, which are subjected to higher losses of intermediate products (diffusing oligomers). Consequently, individual endo-producers experience a reduced return on their enzyme investment, "leaking" large part of the processed resources to the surrounding environment. For this reason in spatial settings such microorganisms should be much more affected by costs of endo-enzyme production. On one hand we can speculate that while this loss penalizes individuals, it may favor larger colonies, as leaking of enzymatic breakdown products to the surrounding environment would go to the benefit of neighboring microbes in the same colony. On the other hand, it also may favor proliferation of "cheaters" who do not degrade the polymer themselves, but are known to "steal" degradation products away from enzyme producers for their own growth [1, 5, 25, 41]. On the other hand, exo-producers are able to uptake the monomers almost at the rate of monomer production, sharing less with their surroundings and consequently cooperate less within colonies. For this reason they would be far less exploited by cheaters. Although we have not explored the cheater-producer feedbacks in our study, such interactions between individuals exploiting the system and those contributing to "public goods" have been extensively analyzed in theoretical models [1] and documented experiments [3, 41], revealing major consequences for organic matter build-up in soils [25]. In summary our results indicate significant differences for spatial-dynamics and organization of exo and endo-producers. These differences are observable in the amount and sizes of diffusing breakdown products and therefore are reflected on potential cellular uptake within consortia. Such effects will have a profound impact on the ecological dynamics, since the diffusion and distribution of soluble byproducts and their complexity were empirically linked to social interactions and spatial self-organization of microorganisms [16, 17], as well as densities of formed colonies and levels of cooperation and competition within them [14, 15]. Finally, we emphasize that our model results for specialists rely on the hypothesis that the enzymes and polymers are relatively well mixed within a single microsite, however small scale (within microsite) spatial organization mixing of extracellular enzymes and polymers may also affect the speed of the degradation process.

For generalists, i.e., microbes that produce both enzymes, we observe a broader niche, suggesting a strong advantage of having both enzyme classes present in the system. We numerically show the synergy between the two enzymes in agreement with previously elaborated conceptual models [28, 52]. Synergies between endo and exo enzymes, where well documented for example for endo and exo-chitinases from *Serratia marcescen* [23]. Moreover, we see that this advantage only appears when most of the resources are used to produce exo-enzymes, and only a small fraction goes into endo-production. Despite these two significant advantages (synergy and niche expansion), in spatial settings the generalists also "leak" larger fractions of products to their surroundings when compared to exo-producers. This offers certain disadvantages, and we see in nature examples to overcome this issue. Microorganisms such as *Vibrio spp.*, for example, have transport systems that can uptake chitin oligomers. The molecular and physiological details of chitin degradation is well documented for these microorganisms and they have endo and exo chitinases within the periplasm and outside the cell. They are known to have large chitoporins that transport molecules with sizes up to 6 NAGs into the periplasm, where additional enzymes proceed with degradation [32, 51]. Since the diffusion rate decreases with oligomer size, it is clearly advantageous to minimize the losses by taken up not only the final but also intermediate

products, preventing them to diffuse away.

Since each one of these enzymes are costly and perform "leaky" functions, they constitute potentital targets for gene loss [20, 34]. According to the black queen hypothesis (BQH) [35, 36], within a community the evolution would favor individuals that lose complex metabolic pathways that produce public goods, as long as these public goods continue to be generated by other community members. This could be the reason for the great variability in the endo/exo-activity pattern for some marine microorganisms such as *Proteobacteria* (e.g. see species from the genera *Pseudoalteromonas*), which we observe in our literature overview (Fig. 2D). Our simulation results, however, suggest that the enzyme losses of endo or exo-enzymes in microbial communities would have different effects depending on the environmental conditions. We observed that consortia combining endo and exo-producers are only stable in low nutrient conditions since in high nutrients exo-producers have high advantages due to fast growth rate and low product loss (lower diffusive losses). These findings hint that if microorganisms are subjected to variable environmental conditions which is common in nature [7], with changing nutrient concentrations, the loss of endo-enzymes would be more likely than of exo-enzymes. As stated previously, however, our overview on enzyme activity (Fig. 2D) suggests that both losses can occur and we can observe microorganisms with endo-activity only. This is the case for the strains of marine bacteria such *Labrenzia alba*, which apparently demonstrates only endo-enzyme activity. If we eliminate the possibility that the absence of exo-activity is linked to cultivation conditions or the employed assay, this would imply that the specific strain must genuinely exhibit oligotrophic characteristics, thriving exclusively in environments with limited nutrients. Other such examples of specialization we see in bacteria found associated to carnivorous plant. This could be because the plant or the established consortia offer alternative nutrient sources that enable endo-producers to endure while awaiting for return on their investment. Moreover, the aquatic settings could facilitate mixing, which attenuates the disadvantages of such specialists within consortia.

Type	Pros	Contras
exo-producers	1. Fast initial growth in nutrient rich conditions 2. Smaller fraction of monomers loss to diffusion. 3. Can expand the niche by growing with endo-producers.	1. Cannot grow in low concentrations of long polysacharides. 2. Degradation time and uptake rate directly depend on the number of polymeric chains.
endo-producers	1. Can grow in nutrient poor conditions. 2. Shorter total degradation times for comparable turnover rates.	1. Cannot start growth in rich conditions. 2. A lag in the initial growth with increase of polymer abundance. 3. Very leaky function. 4. Enzymes may have slower turnover rates. 5. It has difficulty to expand the niche by growing with exo producers.
generalists	1. Synergy between endo and exo enzymes speeds up degradation. 2. Grows in low and high nutrient conditions.	1. Subjected to diffusive loss in spatial settings.

TABLE I. Trade-offs of endo and exo producer specialists and generalists capable to produce both enzymes.

In general, from our review of enzyme activity we see that we have very limited knowledge on the endo and exo-classes of microbial chitinases. Moreover there is still a large number of "unclassified" or uncharacterized sequences when performing genome analysis. Some of these could be associated with chitinases or be alternative chitinases, but have not been functionally characterized or confirmed. We have shown that the different pathways in the degradation dynamics can have fundamentally distinct outcomes for microbes. In this context numerical models can shed light on the constrains and trade-offs to which microorganisms are subjected, offering theories about evolutionary strategies of microorganisms and also possible interactions among them.

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CODE AVAILABILITY

Python and C++ code to reproduce simulations is available at: `https://github.com/kseniaguseva/Microbial_growth_polymer_degradation` and `https://github.com/kseniaguseva/Enzyme_degradation`.

COMPETING INTERESTS

The authors declare no competing interests.

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