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Neither Williston nor Dollo: mandibular complexity from stem tetrapods to modern amphibians

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Keywords: Mandible; Williston; Dollo; Macroevolution; Complexity; Tetrapod

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Abstract

Directional trends in evolution have long captured the attention of biologists, and are particularly interesting when they reflect fundamental developmental processes that underlie morphological change. Here, we apply deep time data and a phylogenetic comparative framework to assess two fundamental "laws" – Williston's law of phenotypic simplification and Dollo's law of irreversibility – in the tetrapod mandible, a structure that has sustained the same primary function of feeding for nearly 400 million years. In spite of this conserved function, the tetrapod mandible has undergone numerous morphological and compositional changes during and since the initial water-to-land transition around 390Ma. To quantify these shifts, we reconstructed the compositional evolution of the mandible with 31 traits scored in 568 species from early tetrapods through to modern amphibians, thereby capturing immense developmental and ecological diversity as well as an excellent fossil record. Mandibular complexity and jaw disparity are highest at the base of the tetrapod tree and generally decrease through time, with stasis dominating over the last ~160M years. Nonetheless, we find a lack of support for Williston's and Dollo's laws, with loss and gain of jaw components equally likely throughout the course of early tetrapod and amphibian evolution. Combined, our results demonstrate that evolutionary patterns of mandibular complexity are more nuanced than either Williston's or Dollo's laws allow. Thus, laws of simplification are too crude to capture the evolutionary processes underlying the evolution of even a functionally conserved structure through deep time.

Summary

The lower jaw is a key innovation in vertebrate evolution with a unifying primary function: feeding. In spite of this conserved function, the jaw is extremely diverse in shape and composition. In limbed vertebrates (tetrapods), the jaw evolves from a complex structure comprising multiple elements and high numbers of teeth towards a simpler structure comprising few elements and generally fewer teeth. Superficially, this pattern suggests support for both Williston's and Dollo's laws of phenotypic simplification and irreversibility, respectively. However, we find a lack of support for either law in the jaw of the earliest tetrapods and amphibians, adding to growing literature refuting overly simplified "laws" governing organismal evolution.

Main Text

Introduction

A key goal in evolutionary biology is to characterise how phenotypic diversity changes across clades and through time. These patterns of diversity can be modelled by contrasting evolutionary processes, each exemplifying the spread of trait adaptations in different ways. Many studies have emphasised the adaptative radiation-type evolutionary process in which morphological disparity rapidly increases from a single lineage possessing a key innovation or ecological opportunity (1, 2). Alternatively, under the neutral theory of evolution (3, 4), diversity can gradually accumulate via genetic drift. In contrast, several central tenets of evolutionary biology propose the exact opposite pattern, in which early members of a clade exhibit the highest disparity, experimenting with diverse structures and functions, and this variation is subsequently whittled down over time (5, 6).

The processes underlying this latter mode of evolution have been characterised in several ways. Traits that consistently endure in a lineage are hypothesised to become fixed or 'canalised', ceasing to produce variants upon which natural selection acts to generate further diversity (7). Patterns of loss of structures in vertebrate evolution is formalised by Williston's law which states that organisms decrease the number of bony structures through time but contiguously increase functional specialisation, as has been suggested by studies of the tetrapod (limbed vertebrate) skull (8, 9). For Williston, structural reduction was founded in Dollo's law of irreversibility, which states that once a trait is lost, it cannot be regained in the same form due to the high energetic

cost involved in gaining novel structures (10, 11) and the rapid degradation of genetic and
developmental pathways that are released from selection pressure (12). Dollo's law can be
qualitatively observed in many clades – e.g. snakes have not regained limbs, nor birds teeth –
and has been supported in multiple quantitative studies, from vertebrates, to parasites, to plants
(13–17). Conversely, the number of exceptions have been growing in recent years, including
regains of wings in stick insects (18), coiled shells in slipper limpets (19), the tadpole stage in
marsupial frogs (20), larval feeding in marine snails (21), oviparity in sand boas (22), and teeth
across multiple frog families (23, 24) among others. However, these studies are overwhelmingly
focused on extant taxa, thereby excluding the vast history of evolutionary change captured in the
fossil record, which includes direct data on the polarity and timing of morphological shifts. Here,
we pair a dataset containing a large extinct component with a modern phylogenetic framework in
order to test these laws in the lower jaw of early tetrapods and amphibians.
The tetrapod lower jaw is an ideal structure in which to investigate both of these fundamental
biological laws, as it maintains the same primary function (feeding) through tetrapod evolution.
Maintenance of a single primary function could indicate a static evolutionary history or could
indicate constraint due to adherence to the aforementioned evolutionary laws. We focus on this
simple structure in the lineage from earliest tetrapods to modern amphibians, thereby capturing
an exceptionally taxonomically (25), ecologically (26, 27), and developmentally (28) diverse
dataset, with a good early fossil record outside crown amphibians (temnospondyls and
polyphyletic lepospondyls).
Whilst we know that the structure, morphology, and feeding mechanisms have changed across
Tetrapoda since the water-to-land transition 390 million years ago, lower jaw composition has not
been comprehensively studied across the clade. Moreover, although the evolution of lower jaw
composition has previously been an area of focussed research in the earliest tetrapods (29), little
work has focussed on lissamphibians (crown amphibians; 29, 30). Nonetheless, the
lissamphibian lower jaw is highly variable in its composition, ranging from three elements in
Gymnophiona (caecilians and their ancestors; 31, 32), to four elements in Salientia (frogs, toads,
and their ancestors; 33), and between three and five elements in Caudata (salamanders, newts,
and their ancestors; 34). Moreover, the developmental variation of lissamphibians, with
paedomorphic, metamorphic, and direct developing taxa, alongside their ecological range,
taxonomic diversity, and excellent fossil record, makes them an ideal group for testing for
persistent trends in morphological evolution.
Here, we use a sample of 568 species of early tetrapods and amphibians, to reconstruct shifts in
lower jaw composition over nearly 400 million years of evolutionary history. We tested the
predictions that jaw complexity, as captured by number of elements, decreases through time and
that losses occur more frequently than gains for three key traits (number of elements, teeth, and
tooth-bearing elements in a hemimandible). In doing so, we estimate ancestral states and test for
correlated evolution of traits. Finally, we estimate disparity through time to assess whether
amphibian jaw disparity accumulated gradually or via a rapid adaptive radiation. Combined, these
analyses allow us to assess if the water-to-land transition of tetrapods was associated with
greater innovation in jaw composition and to establish whether either or both of Williston's and
Dollo's laws have shaped early tetrapod and amphibian lower jaw evolution.
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95 **Results**
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Ancestral states and transition rates of key traits
Dollo's law and Williston's law both predict a higher occurrence of element losses than gains.
Evolutionary modelling of the numbers of teeth, tooth-bearing elements, and elements in the
hemimandible fails to unambiguously support different rates for gains and losses for any of these
traits. Rather, the marginal likelihood scores for three evolutionary models – equal rates (ER),
Markov (Mk), and unequal rates (freeK) – are equivalent for each character (Table S1). We also
did not find strong support for any model of correlation among these three characters, suggesting,

surprisingly, that numbers of jaw elements and teeth are largely independent through early tetrapod
and amphibian evolution (Table S2).
Ancestral state reconstructions estimate that the highest number of tooth-bearing elements occur
at the base of the tree, with the earliest tetrapods having up to five tooth-bearing elements (Figure
1). The number of tooth-bearing elements rapidly decreases, with most temnospondyls and
lepospondyls having three or fewer tooth-bearing elements. The number of tooth-bearing elements
in Lissamphibia is relatively static at either zero in frogs or one in caecilians and adult salamanders.
A similar pattern is seen in the ancestral state estimates of the number of teeth in the jaw (Figure
S1).
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115 *Figure 1: Stochastic character map of the number of tooth-bearing elements in the mandible. Tip*
116 *colours reflect observed character states, internal node colours and branch colours on the inset*
117 *reflect mapped ancestral state estimates.*
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The number of total elements in the jaw is the highest at the base of the tree, with the earliest
tetrapod jaws consisting of eleven elements (or twelve - *Eusthenopteron foordi*; 35) (Figure 2). The
albanerpetonid temnospondyls, which persist to the Pliocene and are possibly nested within
Lissamphibia (37), are a stark contrast to the extant clades, and retain ten elements in their jaws

as in most other temnospondyls (except *Cacops aspidephorus* and *Plagiosuchus pustuliferus*
which have eleven elements; 37, 38).
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127 *Figure 2: Stochastic character map of the number of overall elements in the mandible. Tip colours*
128 *reflect observed character states, internal node colours and branch colours on the inset reflect*
129 *mapped ancestral state estimates.*
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131 ***Variation and disparity in jaw composition through time***
132 Compositional variation in the jaw is highly phylogenetically structured (Figure 3). In particular,
133 lissamphibian orders are tightly clustered in distinct regions, while the early tetrapods,
134 temnospondyls, and lepospondyls show greater disparity and broadly overlap with each other. The
135 volume of morphospace occupied per group is highest in the early tetrapods, followed by roughly

comparable morphospace occupation of temnospondyls, lepospondyls, and salamanders, and finally, is lowest in caecilians and frogs (Figure 3 [inset]).

Figure 3: PCoA of all tetrapod jaw characters, with colours indicating the individual clades and (inset) estimated disparity of each group using PCOA output with bootstrapping (n=100). Vectors used here and in subsequent figures are from Phylopic under CC1.0 Universal Public Domain Dedication and Attribution 3.0 Unported licenses. Oophaga pubilio *(Salientia) by Chuanxin Yu,* Triturus marmoratus *(Caudata) by Beth Reinke,* Dermophis mexicanus *(Gymnophiona) by Jose Carlos Arenas-Monroy,* Balanerpeton woodi *(Temnospondyli) and* Acanthostega gunnari *(Devonian tetrapod) by Scott Harman.*

Reconstructing subclade disparity through time demonstrates that disparity of jaw form was higher during the initial diversification of the Devonian tetrapods than at any other point in tetrapod evolution (Figure 4). Disparity initially declines around the Late Devonian extinction event (~ 371.87 Ma: 40) and declines sharply at the Devonian-Carboniferous boundary. Disparity rapidly increases to a value near one during Romer's Gap (41), followed by another sharp decline. Jaw disparity continues to fluctuate between 345-220 Ma, with the lepospondyls, temnospondyls, and lissamphibians diverging and the early tetrapods, lepospondyls, and the majority of the temnospondyls disappearing during that window. Despite the fluctuations observed during the first ~150 million years of tetrapod evolution, jaw disparity seems to be relatively unaffected by post-Devonian mass extinction events, with only small increases or decreases around the Permian-Triassic extinction (~ 252.28 Ma: 42), the Triassic-Jurrasic extinction (~ 201.58 Ma: 43), and the

159 Cretaceous-Palaeogene extinction (~ 66.043 Ma: 44). From approximately 180 to 160 Ma, jaw
160 disparity declines gradually and then is relatively static to the present.
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Figure 4: (left axis) Disparity of jaw composition through time plotted against (right axis) number of
elements in hemimandible. Disparity near 1 implies disparity is concentrated within groups whereas
disparity near 0 indicates disparity is concentrated between groups. Vertical dotted lines indicate
a) Late Devonian extinction, b) Permian-Triassic extinction, Triassic-Jurassic extinction, d)
Cretaceous-Paleogene extinction. Shaded line indicates 'Romer's Gap', 360-345Ma. Icons indicate
oldest known fossil per clade; key and colours same as Figure 3.

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172 **Discussion**
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In this study, we find a pattern of declining disparity, with near stasis occurring over the last
~160M years, indicating that jaw composition has simplified over time from the earliest tetrapods
to modern amphibians, and that canalisation occurred early in the evolution of each crown
amphibian order. We estimate the ancestral jaw at the root of the tetrapod clade as bearing
eleven elements, including five tooth-bearing elements, and a moderate (between one and 50)
number of teeth. Each subsequent clade has reduced the number of elements, tooth-bearing
elements, and teeth in the jaw. Surprisingly, however, gain or loss of elements or teeth do not
occur at different rates throughout early tetrapod and amphibian evolution, and largely evolve
independently. This contravenes both Dollo's law – assuming the gains are homologous to former
structures that were lost – and Williston's law – wherein only losses occur, resulting in a trend
towards structural simplification.
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Considering Williston: decreasing lower jaw complexity through time
Faster taxonomic and phenotypic diversification have long been recognised near the origin of
clades such as Tetrapoda, with the first forays out of the water associated with key adaptations to

life on land (45–48). We identified high levels of within-group disparity characterising the water-to-
land transition and numerous other late Devonian and Early Carboniferous peaks indicative of the
early innovation of jaw expression of the earliest tetrapods. These modifications to the jaw played
a crucial part in the water-to-land radiation, allowing early tetrapods to transition from feeding in
the water to feeding on land (49–51). High levels of within-group disparity in the early tetrapod
jaw correspond to high disparity previously identified in early tetrapod humerus (52) and cranium
(53). Additionally, our disparity analysis shows a rapid peak and trough coincident with the period
referred to as Romer's Gap, a well-known period with few fossils from 360 to 345 Ma (41).
However, this volatility in disparity could also reflect relatively lower rates of diversification and
extinction in the early Carboniferous compared to other times in early tetrapod and amphibian
evolution.
Late Permian and Early Triassic peaks of within-group disparity likely reflect adaptive radiations
of earlier fauna into ecological gaps created by the Permo-Triassic mass extinction (e.g.
stereospondyl temnospondyls, 54, 55, and more broadly across Temnospondyli, 56). The
subsequent decline of within-group disparity corresponds with the extinction of the lepospondyls
and most temnospondyls, with the remaining albanerpetonid temnospondyls retaining the general
temnospondyl jaw bauplan (high element count etc), as well as establishment of the specific jaw
structures for the crown lissamphibian clades. Canalisation of the lissamphibians is supported
through the strong phylogenetic clustering in the PCoA, indicating that each lissamphibian clade
rapidly reaches a stable within-group expression of jaw composition (Figure 3). Overall, the
compositional structure of the jaw is most diverse in the earliest tetrapods, becomes more
canalised within the temnospondyls and lepospondyls, and shows highly phylogenetically
restricted, static compositions in the lissamphibians.
In spite of the canalisation observed overall across the lissamphibian orders, the extent of
compositional variation of the salamander jaw is more comparable to that of the temnospondyls
and lepospondyls than to the frogs and caecilians (Figure 3 [inset]). Salamanders have
particularly variable jaw compositions, with the number of elements and the number of tooth-
bearing elements in the adult jaw varying between the main salamander families (35). The
variation in the salamander jaw may be a reflection of the varied functional requirements on the
jaw at different developmental stages (57) and the variation in life history modes observed in
salamanders, from paedomorphic to biphasic to direct developing (58–60). Indeed, cranial
elements are highly variable in salamanders, with the pterygoid, maxilla, prefrontal,
orbitosphenoid, and nasal being absent in adult forms of many species, reflecting differences in
life history strategies (61). Similarly, the tooth-bearing elements of the skull, including the dentary
and coronoid ossify at the first stage of ossification during development (Boisvert, 2004 in 57),
and indeed the coronoid of some salamanders bears teeth in early larval stages before
reabsorption of the teeth and coronoid element in later developmental stages (35). The degree of
flexibility in jaw composition across adult salamanders could thus reflect the variety of life history
pathways in the clade.
Similar variation in jaw expression is seen in the polyphyletic lepospondyls (Table 1). The jaw of
aïstopod *Andersonerpeton* remains similar to that of earlier tetrapods and temnospondyls,
retaining eleven elements including an adsymphysial covered with denticles, and three coronoids,
also covered with denticles, and fang pairs on the anterior two coronoids (62). However, the
aïstopod *Phlegethontia* possesses only two elements in the jaw: a dentate dentary and an
edentulous posteriomedial element (63). This variation could be simply a reflection of the
uncertainties around lepospondyl affinities resulting in pooling taxa that do not form a
monophyletic clade (e.g. 64, 65). Equally, as suggested for salamanders, the variation of the
lepospondyls could be a reflection of the varied ecologies and developmental modes proposed
for Lepospondyli (66, 67). Greater clarity on lepospondyl ontogeny and development would likely
strengthen these comparisons further.
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Table 1: Presence of dentition across different jaw elements in lepospondyls (L) and salamanders (S). Presence of denticles has been noted for lepospondyls ('Denticles'). Huskerpeton *has an edentulous posterior coronoid ('No teeth'). Absence of elements is noted with '-' for both lepospondyls and salamanders.*

Taxon	Dentary	Anterior coronoid	Intercoronoid	Posterior coronoid	Adsymphyseal	References
Andersonerpeton longidentatum (L)	x	x	X	*Denticles*	*Denticles*	(62)
Diplocaulus minimus (L)	x	-	-	x	-	(68)
Euryodus dalyae (L)	x	x	X	x	-	(69)
Huskerpeton englehorni (L)	x	x	-	*No teeth*	-	(70)
Pantylus cordatus (L)	x	?	?	x	-	(71, 72)
Ambystoma mexicanum (S)	x	x	-	-	-	(73)
Jeholotriton paradoxicus (S)	x	x	-	-	-	(74)
Kokartus honorarius (S)	x	-	-	x	-	(75)
Marmorerpeton wakei (S)	x	x	-	?x	-	(73)
Necturus sp. (S)	x	-	-	x	-	(73)
Qinglongtriton gangouensis (S)	x	-	-	x	-	(76)
Proteus anguinus (S)	x	-	-	x	-	(73)

Considering Dollo: gains and losses in the lower jaw

Against this backdrop of overall simplification of the composition of the jaw, we tested for differences in transition rates for three key structures (elements, tooth-bearing elements, and teeth) to assess support for a bias towards loss of elements. We found no strong support for gains and losses of these structures occurring at different rates, meaning that early tetrapods and amphibians were as likely to gain components of the jaw as they were to lose them.
Unexpectedly, we also did not find support for correlations among the numbers of elements, tooth-bearing elements, or teeth, indicating that these characters evolved independently through early tetrapod and amphibian evolution.
Whilst loss of elements, the reduction or complete loss of dentition and associated dentigerous elements are immediately obvious, there have been notable, and in some cases, pervasive gains of jaw components. The regain of 35 dentary teeth in the frog *Gastrotheca guentheri* is the most well-known, being the only known frog to ever have had teeth in its lower jaw (23). A second row of mandibular teeth are present in caecilians (inner mandibular [= splenial] teeth), situated on the pseudodentary, a fusion of the dentary, splenial, and (?posterior) coronoid (77). This is reconstructed as a gain on the dentary rather than regain on the coronoid due to the placement of the teeth, although this requires validation with developmental data. Similarly, as noted above, the presence and distinction of the anterior and posterior coronoids are uncertain in salamanders and deserve further study (e.g. 73).
The most pervasive gain is the mentomeckelian(s), bony ossifications found at the jaw symphysis of all three lissamphibian orders (although the salamander families Plethodontidae, Sirenidae, and Amphiumidae lack mentomeckelians; 56). The mentomeckelian is formed from the rod-shaped Meckel's cartilage which runs the length of the jaw, which also ossifies across tetrapods at the posterior jaw to form the articular. The mentomeckelian has been described in Late Jurassic stem-caudatan *Lingolongtriton daxishanensis* (78) but has not yet been identified in the

albanerpetontid temnospondyls, which are typically reconstructed as the most closely related
clade to the lissamphibians (37).
The mentomeckelian has been identified in other jawed vertebrates, most notably in some jawed
fishes (mentomandibular: 79), and comparable structures are present in some extant lizards and
birds (80) and the captorhinid *Moradisaurus grandis* (81). These structures have been suggested
to be homologous to the predentary of ornithischian dinosaurs (80). Given this homology
argument, it is possible that the mentomeckelian may also be homologous to the adsymphysial,
which occurs at the anterior jaw symphysis in earlier tetrapods. Alternatively, the mentomeckelian
could be an intermediary state between a dermal bone at the anterior jaw and the loss of extra
ossified elements at the symphysis, as derived salamanders and the majority of amniotes have
neither a mentomeckelian nor another dermal anterior element. Concentrated study on the
potential homology of these structures across vertebrates will further illuminate whether this
structure has a common origination – therefore supporting Dollo's law – or whether they are
examples of structural and/or functional convergence.
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Declines through time, but neither Williston nor Dollo
Overall, we identify decreasing complexity and disparity of jaw composition from the earliest
tetrapods to modern lissamphibians, with the largest shifts occurring in the late Palaeozoic. Jaw
composition is strongly phylogenetically structured, and within-group diversity is particularly low in
frogs and caecilians, suggesting canalization of the jaw bauplan in these extant clades.
Salamanders and lepospondyls exhibit the greatest extent of compositional variation, particularly
in the number of elements and tooth-bearing elements, which could be related to feeding diversity
through ontogeny, in addition to the variety of developmental and life history pathways observed
and hypothesised for these groups, respectively.
On the surface, this pattern of compositional simplification appears to support Williston's law;
however, this would require unidirectional loss or at least a bias in transition rates favouring
losses. We noted a number of state changes across the tree, typically indicating a character loss,
yet there were also a number of notable gains: the mentomeckelian element, the inner
mandibular (=splenial) dentary tooth row in caecilians, and dentary teeth of *G. guentheri*. Given
these gains, Williston's law is not fully supported, and, as we found no strong support for a model
of different rates between gain and loss of teeth, elements, and tooth-bearing elements, neither is
Dollo's law supported in the evolution of the amphibian jaw. The lack of support for Williston's and
Dollo's laws suggests that the macroevolutionary story of the jaw is more complex than these
laws inherently imply. Given the growing number of exceptions to these "laws", including the work
presented here, perhaps it is time to consider the validity or utility of such evolutionary
fundamentals. Diversity of amphibian jaw composition is neither unidirectional nor as static as first
appears, but is rather a complex interplay between the ancestral tetrapod structure and the
development and life histories of these complex vertebrates.
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311   **Materials and Methods**
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Sampling and character collation
We scored 31 discrete and meristic lower jaw characters across 200 fossil and 368 extant
tetrapods (n=568), including 41 early tetrapods, 90 temnospondyls, 41 lepospondyls, 263 frogs (3
fossils), 48 caecilians (2 fossils), and 85 salamanders (23 fossils). We sampled to a minimum of
family level for extant species, and attempted to record every described fossil early tetrapod,
temnospondyl, lepospondyl, and amphibian jaw regardless of level of completeness.
Characters were scored based mainly on published literature, with some data on extant
amphibians scored from high-resolution micro-CT scans (SI). For each species, the characters
were scored for only one side (hemimandible). The characters scored include: the presence or
absence of elements within the jaw and tooth counts for each dentigerous element and for the
hemimandible overall. Where elements were partially or fully fused, they were recorded as their
individual components. Where characters were not preserved they were marked as missing, with

the exception of total number of elements. For incomplete fossils, the total number of elements in
the jaws were inferred to have no more elements than their closest relatives based on
phylogenetic bracketing, as indicated in SI. Where data was provided in the literature, we also
recorded heterodonty/homodonty, presence or absence of mandibular fenestrae, and presence of
Meckel's cartilage separate to elements formed from Meckelian ossification (articular and
mentomeckelian). We also collected data on tooth implantation type, state of symphyseal fusion,
and length (elongate vs short) and width (gracile vs robust) of the jaw, but were unable to capture
this data for the majority of the species, and so discounted these characters from our analysis of
the 31 characters presented here.
In the tooth counts, we included anything described as a tooth from the main tooth row, marginal
teeth, accessory teeth, fangs, and fang-like teeth. We excluded denticles, as these are often
undescribed, or the total number is unknown due to the preservation of the specimen. In the
earliest tetrapods, fields of denticles are sometimes located on the prearticular element in
addition to other elements (e.g. 29, 82, 83). As we excluded denticles in the overall tooth count,
we also excluded the prearticular from counts of the number of tooth-bearing elements.
We created a composite phylogeny using Mesquite 3.61 (84) and the R packages 'ape' (85),
'geiger' (86), and 'phytools' (87) in the R computing environment version 4.1.2 (88). We started
with a fossil tree for a backbone phylogeny (52) and grafted clades based on the published
literature (SI). Placeholder tips for the extant clades were inserted using Mesquite. We estimated
divergence times and branch lengths using the function 'bin_cal3TimePaleoPhy' from the R
package 'paleotree' (89). The extant clades were then taken from a recent molecular analysis
(90) and pasted onto the placeholder branches.
Variation and disparity in jaw composition through time
To identify the major axes of variation in jaw composition, we conducted a Principal Coordinates
analysis (PCoA) on all 31 characters for the full dataset of 568 species. We used the 'vegdist'
function with the method 'gower' from the R package 'vegan' (91) and used the 'pcoa' function
from the R package 'ape'. We applied Lingoes correction for negative eigenvalues, which
returned the relative weightings of each axis of the PCoA. We assessed the disparity in PCoA
scores between clades using the package 'dispRity' (92). We used the 'dispRity.per.group'
function to estimate disparity for each group with bootstrapping (n=100).
To reconstruct overall disparity in jaw morphology through time, we used the 'dtt' function from
the R package 'geiger' on the full dataset (31 characters, 568 species), applying 1,000 iterations
to generate a null distribution.
Given the high proportion of zero values in our data (e.g. zero teeth and tooth-bearing elements
in most frogs), we were unable to explicitly assess evolutionary trends through traditional
evolutionary models.
Ancestral states and transition rates of key traits
To reconstruct changes in key jaw traits across the phylogeny, we selected the three traits that
were best represented in the data: number of elements, number of tooth-bearing elements, and
number of teeth in the hemimandible. The number of overall teeth had three states (0 teeth, 1-50
teeth, 51+ teeth), the number of elements had ten states (between 3 and 12 elements), and the
number of dentigerous elements had six states (between 0 and 5 elements). We only included
taxa for which the data on all three characters were available (491 species, SI). We quantified
transition rates among the states by fitting three different evolutionary models for each trait in
RevBayes (93): an equal rates model (ER), a Markov (Mk) model, and an unequal rates model
(freeK). The models ran for at least 10,000 generations, and the first 2,000 were discarded as
burn-in. We used the R packages 'convenience' (94) to test for convergence of the model runs
and 'RevGadgets' (95) and 'plotrix' (96) to visualise the results.
We then used the 'make.simmap' function from the R package 'phytools' to generate a
distribution of 100 stochastic character maps, using an equal rates model, based on the outcome
of the evolutionary modelling analyses. We estimated ancestral states for the same three key
characters as above (number of elements, tooth-bearing elements, and teeth), calculating the
averaged probability of the state of the internal nodes across the 100 iterations, and mapping the
averaged nodes onto one random stochastic character map.

We also tested the correlation between the number of elements and the number of tooth-bearing
elements (491 species) using the R package 'corHMM' (97). corHMM uses a Markov model to
describe the discrete transitions between the observed states. We ran four models, across two
different model structures: correlated ER (equal rates), independent ER, correlated ARD (all rates
differ), and independent ARD. We compared model fit using AICc scores.

Acknowledgments

For thought-provoking and valuable discussion on modelling discrete data we thank Gustavo Burin, Natalie Cooper, Julien Clavel, and Thomas Guillerme and for suggestions on coding we are grateful to Liam Revell and Katherine Corn. For discussions on amphibian jaws we thank the members of the NHM Herpetology Group, Susan Evans, Marc Jones, and Anne Claire-Fabre.

Author Contributions: Conceptualisation, EW, RF, and AG; Methodology, EW, RF, and AG ; Investigation, EW; Formal Analysis, EW, RF, and AG; Writing – Original Draft, EW; Writing – Review & Editing, EW, RF, and AG; Supervision, RF and AG.

Competing Interest Statement: We declare no competing interests.

Supplementary Information:
Composite tree references

Table S1: Marginal likelihood scores for RevBayes evolutionary models

Table S2: AICc scores for evolutionary (corHMM) models

Figure S1: Ancestral states reconstruction for number of teeth in hemimandible

Data, composite tree file, R scripts, and RevBayes scripts are available at https://github.com/emilycwatt

References

1. J. B. Yoder, *et al.*, Ecological opportunity and the origin of adaptive radiations. *J. Evol. Biol.* **23**, 1581–1596 (2010).

2. J. T. Stroud, J. B. Losos, Ecological Opportunity and Adaptive Radiation. *Annu. Rev. Ecol. Evol. Syst.* **47**, 507–532 (2016).

3. M. Kimura, *The Neutral Theory of Molecular Evolution* (Cambridge University Press, 1983).

4. W.-C. Ho, Y. Ohya, J. Zhang, Testing the neutral hypothesis of phenotypic evolution. *Proc. Natl. Acad. Sci.* **114**, 12219–12224 (2017).

5. G. G. Simpson, *The Major Features of Evolution* (Columbia University Press, 1953) https:/doi.org/10.7312/simp93764 (June 5, 2023).

6. L. J. Harmon, *et al.*, Early bursts of body size and shape evolution are rare in comparative data. *Evolution* **64**, 2385–2396 (2010).

7. G. Gibson, G. Wagner, Canalization in evolutionary genetics: a stabilizing theory? *BioEssays* **22**, 372–380 (2000).

8. S. W. Williston, *Water reptiles of the past and present* (The University of Chicago Press, 1914) (March 6, 2023).

9. W. K. Gregory, 'Williston's law' relating to the evolution of skull bones in the vertebrates. *Am. J. Phys. Anthropol.* **20**, 123–152 (1935).

10. L. Dollo, Les lois de l'évolution. *Bull. Société Belge Géologie* **VII**, 164–166 (1893).

11. S. C. Stearns, The evolutionary links between fixed and variable traits. *Acta Palaeontol. Pol.* **3–4**, 215–232 (1993).

12. C. R. Marshall, E. C. Raff, R. A. Raff, Dollo's law and the death and resurrection of genes. *Proc. Natl. Acad. Sci. U. S. A.* **91**, 12283–12287 (1994).

13. M. S. Y. Lee, R. Shine, Reptilian viviparity and Dollo's law. *Evolution* **52**, 1441–1450 (1998).

14. R. H. Cruickshank, A. M. Paterson, The great escape: do parasites break Dollo's law? *Trends Parasitol.* **22**, 509–515 (2006).

15. B. Igic, L. Bohs, J. R. Kohn, Ancient polymorphism reveals unidirectional breeding system shifts. *Proc. Natl. Acad. Sci.* **103**, 1359–1363 (2006).

16. F. Galis, J. W. Arntzen, R. Lande, Dollo's law and the irreversibility of digit loss in *Bachia*. *Evolution* **64**, 2466–2476 (2010).

17. W.-E. Lönnig, K. Stüber, H. Saedler, J. H. Kim, Biodiversity and Dollo's Law: To What Extent can the Phenotypic Differences between Misopates orontium and Antirrhinum majus be Bridged by Mutagenesis? *Biomediation Biodivers. Bioavailab.* **1**, 1–30 (2007).

18. M. F. Whiting, S. Bradler, T. Maxwell, Loss and recovery of wings in stick insects. *Nature* **421**, 264–267 (2003).

19. M. Pagel, Limpets break Dollo's Law. *Trends Ecol. Evol.* **19**, 278–280 (2004).

20. J. J. Wiens, C. A. Kuczynski, W. E. Duellman, T. W. Reeder, Loss and re-evolution of complex life cycles in marsupial frogs: does ancestral trait reconstruction mislead? *Evolution* **61**, 1886–1899 (2007).

21. R. Collin, O. R. Chaparro, F. Winkler, D. Véliz, Molecular Phylogenetic and Embryological Evidence That Feeding Larvae Have Been Reacquired in a Marine Gastropod. *Biol. Bull.* **212**, 83–92 (2007).

22. V. J. Lynch, G. P. Wagner, Did egg-laying boas break Dollo's law? Phylogenetic evidence for reversal to oviparity in sand boas (*Eryx*: Boidae). *Evolution* **64**, 207–216 (2010).

23. J. J. Wiens, Re-Evolution of Lost Mandibular Teeth in Frogs After More Than 200 Million Years, and Re-Evaluating Dollo's Law. *Evolution* **65**, 1283–1296 (2011).

24. D. J. Paluh, W. A. Dillard, E. L. Stanley, G. J. Fraser, D. C. Blackburn, Re-evaluating the morphological evidence for the re-evolution of lost mandibular teeth in frogs. *Evolution* (2021) https:/doi.org/10.1111/evo.14379 (November 10, 2021).

25. AmphibiaWeb, <https://amphibiaweb.org> University of California, Berkeley, CA, USA (2023).

26. K. D. Wells, "The Ecology and Behavior of Amphibians" in *The Ecology and Behavior of Amphibians*, (University of Chicago Press, 2007) https:/doi.org/10.7208/9780226893334 (July 17, 2023).

27. B. F. Oliveira, V. A. São-Pedro, G. Santos-Barrera, C. Penone, G. C. Costa, AmphiBIO, a global database for amphibian ecological traits. *Sci. Data* **4**, 170123 (2017).

28. R. P. Elinson, E. M. del Pino, Developmental diversity of amphibians. *WIREs Dev. Biol.* **1**, 345–369 (2012).

29. P. E. Ahlberg, J. A. Clack, Lower jaws, lower tetrapods - a review based on the Devonian genus *Acanthostega*. *Earth Environ. Sci. Trans. R. Soc. Edinb.* **89**, 11–46 (1998).

30. R. Carroll, R. Holmes, The skull and jaw musculature as guides to the ancestry of salamanders. *Zool. J. Linn. Soc.* **68**, 1–40 (1980).

31. R. A. Nussbaum, The evolution of a unique dual jaw-closing mechanism in caecilians: (Amphibia: Gymnophiona) and its bearing on caecilian ancestry. *J. Zool.* **199**, 545–554 (1983).

32. M. H. Wake, J. Hanken, Development of the skull of Dermophis mexicanus (amphibia: Gymnophiona), with comments on skull kinesis and amphibian relationships. *J. Morphol.* **173**, 203–223 (1982).

33. W. E. Bemis, K. Schwenk, M. H. Wake, Morphology and function of the feeding apparatus in *Dermophis mexicanus* (Amphibia: Gymnophiona). *Zool. J. Linn. Soc.* **77**, 75–96 (1983).

34. M. Fabrezi, S. B. Emerson, Parallelism and convergence in anuran fangs. *J. Zool.* **260**, 41–51 (2003).

35. C. Rose, "The Developmental Morphology of Salamander Skulls" in *Amphibian Biology*, H. Heatwole, Ed. (Surrey Beatty and Sons, 1994), pp. 1684–1781.

36. L. B. Porro, E. J. Rayfield, J. A. Clack, Computed tomography, anatomical description and three-dimensional reconstruction of the lower jaw of *Eusthenopteron foordi* Whiteaves, 1881 from the Upper Devonian of Canada. *Palaeontology* **58**, 1031–1047 (2015).

37. R. Matsumoto, S. E. Evans, The first record of albanerpetontid amphibians (Amphibia: Albanerpetontidae) from East Asia. *PLOS ONE* **13**, e0189767 (2018).

38. R. Damiani, R. R. Schoch, H. Hellrung, R. Werneburg, S. Gastou, The plagiosaurid temnospondyl *Plagiosuchus pustuliferus* (Amphibia: Temnospondyli) from the Middle Triassic of Germany: anatomy and functional morphology of the skull. *Zool. J. Linn. Soc.* **155**, 348–373 (2009).

39. J. S. Anderson, D. Scott, R. R. Reisz, The anatomy of the dermatocranium and mandible of *Cacops aspidephorus* Williston, 1910 (Temnospondyli: Dissorophidae), from the Lower Permian of Texas. *J. Vertebr. Paleontol.* **40**, e1776720 (2020).

40. A.-C. Da Silva, *et al.*, Anchoring the Late Devonian mass extinction in absolute time by integrating climatic controls and radio-isotopic dating. *Sci. Rep.* **10**, 12940 (2020).

41. M. I. Coates, J. A. Clack, Romer's gap: tetrapod origins and terrestriality. *Bull. Muséum Natl. Hist. Nat.* **17**, 373–388 (1995).

42. S. Shen, *et al.*, Calibrating the End-Permian Mass Extinction. *Science* **334**, 1367–1372 (2011).

43. U. Schaltegger, J. Guex, A. Bartolini, B. Schoene, M. Ovtcharova, Precise U–Pb age constraints for end-Triassic mass extinction, its correlation to volcanism and Hettangian post-extinction recovery. *Earth Planet. Sci. Lett.* **267**, 266–275 (2008).

44. P. R. Renne, *et al.*, Time Scales of Critical Events Around the Cretaceous-Paleogene Boundary. *Science* **339**, 684–687 (2013).

45. E. B. Daeschler, N. H. Shubin, F. A. Jenkins, A Devonian tetrapod-like fish and the evolution of the tetrapod body plan. *Nature* **440**, 757–763 (2006).

46. M. Ruta, P. J. Wagner, M. I. Coates, Evolutionary patterns in early tetrapods. I. Rapid initial diversification followed by decrease in rates of character change. *Proc. R. Soc. B Biol. Sci.* **273**, 2107–2111 (2006).

47. C. A. Boisvert, E. Mark-Kurik, P. E. Ahlberg, The pectoral fin of *Panderichthys* and the origin of digits. *Nature* **456**, 636–638 (2008).

48. J. B. Lemberg, E. B. Daeschler, N. H. Shubin, The feeding system of *Tiktaalik roseae*: an intermediate between suction feeding and biting. *Proc. Natl. Acad. Sci.* **118**, e2016421118 (2021).

49. S. M. Reilly, G. V. Lauder, The Evolution of Tetrapod Feeding Behavior: Kinematic Homologies in Prey Transport. *Evolution* **44**, 1542–1557 (1990).

50. M. J. Markey, C. R. Marshall, Terrestrial-style feeding in a very early aquatic tetrapod is supported by evidence from experimental analysis of suture morphology. *Proc. Natl. Acad. Sci.* **104**, 7134–7138 (2007).

51. J. M. Neenan, M. Ruta, J. A. Clack, E. J. Rayfield, Feeding biomechanics in *Acanthostega* and across the fish–tetrapod transition. *Proc. R. Soc. B Biol. Sci.* **281**, 20132689 (2014).

52. M. Ruta, J. Krieger, K. D. Angielczyk, M. A. Wills, The evolution of the tetrapod humerus: morphometrics, disparity, and evolutionary rates. *Earth Environ. Sci. Trans. R. Soc. Edinb.* **109**, 351–369 (2018).

53. J. R. G. Rawson, B. Esteve-Altava, L. B. Porro, H. Dutel, E. J. Rayfield, Early tetrapod cranial evolution is characterized by increased complexity, constraint, and an offset from fin-limb evolution. *Sci. Adv.* **8**, eadc8875 (2022).

54. A. A. Warren, R. J. Damiani, A. M. Yates, Palaeobiogeography of Australian fossil amphibians. *Hist. Biol.* **15**, 171–179 (2001).

55. S. Dias-Da-Silva, C. Marsicano, C. L. Schultz, Rhytidosteid Temnospondyls in Gondwana: A New Taxon from the Lower Triassic of Brazil. *Palaeontology* **49**, 381–390 (2006).

56. M. Ruta, D. Pisani, G. T. Lloyd, M. J. Benton, A supertree of Temnospondyli: cladogenetic patterns in the most species-rich group of early tetrapods. *Proc. R. Soc. B Biol. Sci.* **274**, 3087–3095 (2007).

57. R. L. Carroll, The Palaeozoic Ancestry of Salamanders, Frogs and Caecilians. *Zool. J. Linn. Soc.* **150**, 1–140 (2007).

58. P. T. Chippindale, R. M. Bonett, A. S. Baldwin, J. J. Wiens, Phylogenetic evidence for a major reversal of life-history evolution in plethodontid salamanders. *Evolution* **58**, 2809–2822 (2004).

59. R. M. Bonett, A. L. Blair, Evidence for complex life cycle constraints on salamander body form diversification. *Proc. Natl. Acad. Sci.* **114**, 9936–9941 (2017).

60. T. Vučić, *et al.*, Testing the evolutionary constraints of metamorphosis: The ontogeny of head shape in *Triturus* newts. *Evolution* **73**, 1253–1264 (2019).

61. A.-C. Fabre, *et al.*, Metamorphosis shapes cranial diversity and rate of evolution in salamanders. *Nat. Ecol. Evol.* **4**, 1129–1140 (2020).

62. J. D. Pardo, A. Mann, A basal aïstopod from the earliest Pennsylvanian of Canada, and the antiquity of the first limbless tetrapod lineage. *R. Soc. Open Sci.* **5**, 181056 (2018).

63. J. S. Anderson, Revision of the Aïstopod Genus *Phlegethontia* (Tetrapoda: Lepospondyli). *J. Paleontol.* **76**, 1029–1046 (2002).

64. P. P. Vaughn, The Paleozoic Microsaurs as Close Relatives of Reptiles, Again. *Am. Midl. Nat.* **67**, 79–84 (1962).

65. J. S. Anderson, The Phylogenetic Trunk: Maximal Inclusion of Taxa with Missing Data in an Analysis of the Lepospondyli (Vertebrata, Tetrapoda). *Syst. Biol.* **50**, 170–193 (2001).

66. N. B. Fröbisch, J. C. Olori, R. R. Schoch, F. Witzmann, Amphibian development in the fossil record. *Semin. Cell Dev. Biol.* **21**, 424–431 (2010).

67. J. C. Olori, Ontogenetic sequence reconstruction and sequence polymorphism in extinct taxa: an example using early tetrapods (Tetrapoda: Lepospondyli). *Paleobiology* **39**, 400–428 (2013).

68. D. Germain, The Moroccan diplocaulid: the last lepospondyl, the single one on Gondwana. *Hist. Biol.* **22**, 4–39 (2010).

69. B. M. Gee, J. J. Bevitt, R. R. Reisz, Computed tomographic analysis of the cranium of the early Permian recumbirostran 'microsaur' *Euryodus dalyae* reveals new details of the braincase and mandible. *Pap. Palaeontol.* **7**, 721–749 (2021).

70. A. K. Huttenlocker, J. D. Pardo, B. J. Small, J. S. Anderson, Cranial morphology of recumbirostrans (Lepospondyli) from the Permian of Kansas and Nebraska, and early morphological evolution inferred by micro-computed tomography. *J. Vertebr. Paleontol.* **33**, 540–552 (2013).

71. R. Broom, On the Cotylosaurian genus *Pantylus* Cope. *Bull. Am. Mus. Nat. Hist.* **32**, 527–532 (1913).

72. A. S. Romer, The cranial anatomy of the Permian amphibian *Pantylus*. *Breviora* **314**, 1–37 (1969).

73. M. E. H. Jones, *et al.*, Middle Jurassic fossils document an early stage in salamander evolution. *Proc. Natl. Acad. Sci.* **119**, e2114100119 (2022).

74. Y. Wang, C. S. Rose, *Jeholotriton paradoxus* (Amphibia: Caudata) from the Lower Cretaceous of southeastern Inner Mongolia, China. *J. Vertebr. Paleontol.* **25**, 523–532 (2005).

75. P. P. Skutschas, T. Martin, Cranial anatomy of the stem salamander Kokartus honorarius (Amphibia: Caudata) from the Middle Jurassic of Kyrgyzstan. *Zool. J. Linn. Soc.* **161**, 816–838 (2011).

76. J. Jia, K.-Q. Gao, A New Basal Salamandroid (Amphibia, Urodela) from the Late Jurassic of Qinglong, Hebei Province, China. *PLOS ONE* **11**, e0153834 (2016).

77. M. H. Wake, The development and replacement of teeth in viviparous caecilians. *J. Morphol.* **148**, 33–63 (1976).

78. J. Jia, K.-Q. Gao, A new stem hynobiid salamander (Urodela, Cryptobranchoidea) from the Upper Jurassic (Oxfordian) of Liaoning Province, China. *J. Vertebr. Paleontol.* **39**, e1588285 (2019).

79. L. Grande, W. E. Bemis, A Comprehensive Phylogenetic Study of Amiid Fishes (Amiidae) Based on Comparative Skeletal Anatomy. an Empirical Search for Interconnected Patterns of Natural History. *J. Vertebr. Paleontol.* **18**, 1–696 (1998).

80. J. Ferigolo, M. C. Langer, A Late Triassic dinosauriform from south Brazil and the origin of the ornithischian predentary bone. *Hist. Biol.* **19**, 23–33 (2007).

81. S. P. Modesto, C. D. Richards, O. Ide, C. A. Sidor, The vertebrate fauna of the Upper Permian of Niger—X. The mandible of the captorhinid reptile *Moradisaurus grandis*. *J. Vertebr. Paleontol.* **38**, e1531877 (2018).

82. E. B. Daeschler, Early tetrapod jaws from the Late Devonian of Pennsylvania, USA. *J. Paleontol.* **74**, 301–308 (2000).

83. P. A. Beznosov, J. A. Clack, E. Lukševičs, M. Ruta, P. E. Ahlberg, Morphology of the earliest reconstructable tetrapod *Parmastega aelidae*. *Nature* **574**, 527–531 (2019).

84. W. P. Maddison, D. R. Maddison, Mesquite: a modular system for evolutionary analysis (2021).

85. E. Paradis, K. Schliep, ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. Version 5.6. *Bioinformatics* **35**, 526–528 (2019).

86. M. Pennell, *et al.*, geiger v2.0: an expanded suite of methods for fitting macroevolutionary models to phylogenetic trees. Version 2.0.7. *Bioinformatics* **30**, 2216–2218 (2014).

87. L. J. Revell, phytools: An R package for phylogenetic comparative biology (and other things). Version 1.0.1. *Methods Ecol. Evol.* **3**, 217–223 (2012).

88. R Core Team, R: A language and environment for statistical computing (2021).

89. D. W. Bapst, paleotree: Paleontological and Phylogenetic Analyses of Evolution Version 3.3.25. *Methods Ecol. Evol.* **3**, 803–807 (2012).

90. W. Jetz, R. A. Pyron, The interplay of past diversification and evolutionary isolation with present imperilment across the amphibian tree of life. *Nat. Ecol. Evol.* **2**, 850–858 (2018).

91. J. Oksanen, *et al.*, vegan: Community Ecology Package (2020).

92. T. Guillerme, dispRity: a modular R package for measuring disparity. *Methods Ecol. Evol.* **9**, 1755–1763 (2018).

93. S. Höhna, *et al.*, RevBayes: Bayesian phylogenetic inference using graphical models and an interactive model-specification language. *Syst. Biol.* **65**, 726–736 (2016).

94. L. Fabreti, S. Hoehna, A. Magee, convenience: Convergence assessment for phylogenetic inference. R package version 1.0.0 (2022).

95. C. Tribble, *et al.*, RevGadgets: Visualization and Post-Processing of "RevBayes" Analyses. Version 1.0.0 (2022).

96. J. Lemon, Plotrix: a package in the red light district of R. Version 3.8.1. *R-News* **6**, 8–12 (2006).

97.   B. O'Meara, J. Oliver, J. Boyko, corHMM: Hidden Markov Models of Character Evolution (2022).