Cell autonomous role of leucine-rich repeat kinase in protection of dopaminergic neuron survival

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Abstract

Mutations in leucine-rich repeat kinase 2 (LRRK2) are the most common genetic cause of Parkinson's disease (PD), which is the leading neurodegenerative movement disorder characterized by the progressive loss of dopaminergic (DA) neurons in the substantia nigra pars compacta (SNpc). However, whether LRRK2 mutations cause PD and degeneration of DA neurons *via* a toxic gain-of-function or a loss-of-function mechanism is unresolved and has pivotal implications for LRRK2-based PD therapies. In this study, we investigate whether LRRK2 and its functional homologue LRRK1 play an essential, intrinsic role in DA neuron survival through the development of DA neuron-specific *LRRK* conditional double knockout (cDKO) mice. We first generated and characterized floxed *LRRK1* and *LRRK2* mice and then confirmed that germline deletions of the floxed *LRRK1* and *LRRK2* alleles result in null mutations, as evidenced by the absence of *LRRK1* and *LRRK2* mRNA and protein in the respective homozygous deleted mutant mice. We further examined the specificity of Cre-mediated recombination driven by the *dopamine transporter-Cre* (*DAT-Cre*) knockin (KI) allele using a GFP reporter line and confirmed that *DAT-Cre*-mediated recombination is restricted to DA neurons in the SNpc. Crossing these validated floxed *LRRK1* and *LRRK2* mice with *DAT-Cre* KI mice, we then generated DA neuron-restricted *LRRK* cDKO mice and further showed that levels of LRRK1 and LRRK2 are reduced in dissected ventral midbrains of *LRRK* cDKO mice. While DA neuron-restricted *LRRK* cDKO mice of both sexes exhibit normal mortality and body weight, they develop age-dependent loss of DA neurons in the SNpc, as demonstrated by the progressive reduction of DA neurons in the SNpc of *LRRK* cDKO mice at the ages of 20 and 24 months but the unaffected number of DA neurons at the age of 15 months. Moreover, DA neurodegeneration is accompanied with increases of apoptosis and elevated microgliosis in the SNpc as well as decreases of DA terminals in the striatum, and is preceded by impaired motor coordination. Taken together, these findings provide the unequivocal evidence for the importance of LRRK in DA neurons and raise the possibility that LRRK2 mutations may impair its protection of DA neurons, leading to DA neurodegeneration in PD.

Introduction

Parkinson's disease (PD) is the most common movement disorder and is characterized by the progressive loss of dopaminergic (DA) neurons in the substantia nigra pars compacta (SNpc). Dominantly inherited missense mutations in the *leucine-rich repeat kinase 2* (*LRRK2*) gene are the most common cause of both familial and sporadic PD, highlighting the importance of LRRK2 in PD pathogenesis (1-16). LRRK2 is a large protein of 2,527 amino acid residues containing multiple functional domains including several leucine-rich repeats (LRRs), a GTPase-like domain of Ras-of-complex (Roc), a C-terminal of Roc (COR) domain, and a serine/threonine MAPKKK-like kinase domain. LRRK1, a homologue of LRRK2, belongs to the evolutionarily conserved Roco protein family and contains similar LRRs, Roc, COR, and kinase domains (17-19). LRRK proteins are broadly expressed with LRRK2 being most abundant in the kidney (20). While most PD mutations are found in LRRK2, rare variants in the Roc, COR, and kinase domains of LRRK1 have been reported and may be associated with PD (21). Furthermore, the Roc-COR domain of LRRK2 forms dimers and exhibits conventional Ras-like GTPase properties, and the R1441/C/G/H and I1371V mutations destabilize dimer formation and decrease GTPase activity (22, 23). Recent high-resolution cryoEM structural studies of full-length LRRK2 demonstrated its existence as dimers and pathogenic mutations such as R1441/C/G/H and Y1699I at the Roc-COR interface, whereas the G2910S mutant and wild-type LRRK2 share similar structure (24).

Previous genetic studies demonstrated that LRRK2 plays essential roles in the autophagy-lysosomal pathway (25-28). Consistent with high levels of LRRK2 expression in kidneys, *LRRK2*-/- mice develop age-dependent phenotypes in the kidney, including autophagy-lysosomal impairments and increases of α-synuclein, apoptosis, and inflammatory responses (25, 26, 29). It has also been reported that LRRK2 is important for maintaining lung homeostasis, and *LRRK2* deficiency results in impaired autophagy in alveolar type II epithelial cells (28). It was proposed that the lack of brain phenotypes in *LRRK2*-/- mice might be due to the presence of LRRK1 that compensates functionally in the absence of LRRK2 (25, 26). Indeed, *LRRK* double knockout (DKO) mice develop age-dependent, progressive loss of DA neurons in the SNpc, beginning at 14 months of age (30). However, *LRRK* DKO mice also exhibit

lower body weight and increased mortality, raising the possibility that DA neurodegeneration in *LRRK* DKO mice may be secondary to poor health.

In this study, we investigate whether LRRK2 and its functional homologue LRRK1 play an essential, intrinsic role in DA neuron survival through the development of DA neuron-specific *LRRK* conditional DKO (cDKO) mice. We first generated and confirmed floxed *LRRK1* and *LRRK2* mice, and then crossed them with the *CMV-Cre* deleter to create germline deletions of the floxed *LRRK1* and *LRRK2* alleles, followed by Northern and Western analyses to confirm the absence of *LRRK1* and *LRRK2* mRNA and protein in the respective homozygous deleted mutant mice. We also crossed a GFP reporter mouse with *DAT-Cre* KI mice and confirmed that Cre-mediated recombination occurs in most, if not all, DA neurons in the SNpc and is restricted to DA neurons. We then crossed these thoroughly validated floxed *LRRK1* and *LRRK2* mice with *DAT-Cre* KI mice to generate DA neuron-restricted *LRRK* cDKO mice and further confirmed the reduction of LRRK1 and LRRK2 in dissected ventral midbrains of *LRRK* cDKO mice. While DA neuron-restricted *LRRK* cDKO mice of both sexes exhibit normal mortality and body weight, they develop age-dependent loss of DA neurons in the SNpc, as demonstrated by the progressive reduction of TH+ DA neurons or NeuN+ neurons in the SNpc of cDKO mice at the ages of 20 and 24 months. Moreover, DA neurodegeneration is accompanied with increases of apoptosis and elevated microgliosis in the SNpc as well as decreases of DA terminals in the striatum, and is preceded by impaired motor coordination. Interestingly, quantitative electron microscopy (EM) analysis showed a similar number of electron-dense vacuoles in SNpc neurons of *LRRK* cDKO mice relative to controls, in contrast to age-dependent increases of vacuoles in SNpc neurons of germline *LRRK* DKO mice (30, 31). These findings provide the unequivocal evidence for the importance of LRRK in DA neurons and raise the possibility that LRRK2 mutations may impair this crucial physiological function, leading to the loss of DA neurons in Parkinson's disease.

Results

Generation and molecular characterization of the floxed and deleted *LRRK1* and *LRRK2* alleles

LRRK2 and its homologue LRRK1 share several functional (LRRs, GTPase Roc, COR, and kinase) domains (Figure 1A). To investigate the intrinsic role of LRRK in DA neurons, we generated floxed *LRRK1* (*fLRRK1*) and floxed *LRRK2* (*fLRRK2*) mice, which permit deletions of *LRRK1* and *LRRK2* selectively in DA neurons by *DAT-Cre*, in which Cre recombinase is expressed under the control of the endogenous *DAT* promoter (32). We introduced two *loxP* sites in introns 26 and 29 of *LRRK1* through homologous recombination and site-specific recombination by FLP recombinase to remove the positive selection *PGK-neo* cassette, which is flanked by two *FRT* sites (Figure 1B, C; Figure 1-figure supplement 1, 2). The embryonic stem (ES) cells carrying the targeted allele or the floxed allele were identified and validated by Southern analysis using the 5' and 3' external probes as well as the *neo* probe (Figure 1D; Figure 1-figure supplement 3). The validated ES cells carrying the floxed *LRRK1* allele were injected into mouse blastocysts to generate floxed *LRRK1* mice, which were further confirmed by Southern using the 5' and 3' external probes (Figure 1D). In the presence of Cre recombinase, the floxed *LRRK1* genomic region containing part of intron 26 (1,288 bp), exons 27-29, which encode the kinase domain, and part of intron 29 (1,023 bp) is deleted (Figure 1C), and the removal of exons 27-29 (625 bp) results in a frameshift of downstream codons.

The *LRRK2* targeting vector contains the 5' homologous region, a *loxP* site 1,768 bp upstream of the transcription initiation site, exons 1-2, the *PGK-neo* cassette flanked by two *loxP* sites and two *FRT* sequences, and the 3' homologous region (Figure 1B, E; Figure 1-figure supplement 4, 5). The ES cell clones carrying the targeted allele were identified and validated by Southern analysis using the 5’ external probe (Figure 1F; Figure 1-figure supplement 6) and genomic PCR followed by sequencing. Mice carrying the *LRRK2* targeted allele were further verified by Southern analysis using the 5' and 3' external probes (Figure 1-figure supplement 6) as well as the *neo* probe and were then crossed with the *Actin-FLP* mice (33) to remove the *PGK-neo* cassette flanked by two *FRT* sites to generate floxed *LRRK2* mice (Figure 1E). The resulting floxed *LRRK2* mice were confirmed by Southern analysis using the 5’ and 3’ external probes (Figure 1F). In the presence of Cre recombinase, the floxed *LRRK2* region containing the promoter and exons 1-2 are deleted, likely resulting in a null allele, as we previously targeted a very

similar region (~2.5 kb upstream of the transcription initiation site and exons 1-2) to generate germline deletion of *LRRK2* (25).

Dopaminergic neurons in the SNpc are a small neuronal population embedded in the ventral midbrain, making it difficult to confirm whether DA neuron-specific deletions of the floxed *LRRK1* and *LRRK2* regions result in null alleles. We previously generated three independent *LRRK1* knockout (KO) mice, and only one KO line (Line 2) represents a *LRRK1* null allele (30), whereas deletion of *LRRK2* promoter region and exons 1-2 resulted in a *LRRK2* null allele (25). We therefore generated germline deleted *LRRK1* and *LRRK2* mice from floxed *LRRK1* and *LRRK2* mice, respectively, by crossing them to germline deleter *CMV-Cre* transgenic mice (34).

We then performed Northern analysis of *LRRK1* using both an upstream probe specific for exons 2-3 and a downstream probe specific for exons 27-29 (Figure 1G; Figure 1-figure supplement 7, 8). Because of the low expression level of *LRRK1* mRNA and the relative abundance of *LRRK1* mRNA in the lung (20), we enriched polyA+ RNA from the lung of the mice carrying homozygous germline deleted (Δ/Δ) *LRRK1* alleles derived from the floxed *LRRK1* alleles. Using the exons 2-3 probe, the *LRRK1* transcripts in wild-type mice are of the expected size of ~7.4 kb, whereas the detected *LRRK1* transcripts in *LRRK1* Δ/Δ mice are smaller and expressed at lower levels, consistent with the deletion of exons 27-29 (625 bp), resulting in a frameshift of downstream codons and the likely degradation of the truncated *LRRK1* mRNA (Figure 1G; Figure 1-figure supplement 8). Using a probe specific for exons 27-29, there is no *LRRK1* transcript in *LRRK1* Δ/Δ mice (Figure 1G; Figure 1-figure supplement 8). Extensive RT-PCR analysis of total RNA isolated from the kidney, brain, and lung of *LRRK1* Δ/Δ mice using an exon 32-specific primer for RT and exon-specific primer sets for PCR (*e.g.* exons 4-8, 11-17, 20-25, and 25-31), followed by sequencing confirmation of the PCR products, indicated normal *LRRK1* splicing in *fLRRK1/fLRRK1* mice and the lack of *LRRK1* exons 27-29 in *LRRK1* Δ/Δ mice (Figure 1-figure supplement 9).

Similarly, Northern analysis of *LRRK2* using a probe specific for exons 1-5 and RT-PCR followed by sequencing confirmed the absence of *LRRK2* mRNA in *LRRK2* Δ/Δ brains and normal *LRRK2* transcripts in *fLRRK2/fLRRK2* brains (Figure 1H; Figure 1-figure supplement 10, 11). Furthermore, Western analysis confirmed the absence of LRRK1 and LRRK2 proteins in the brain of *LRRK1* Δ/Δ and *LRRK2* Δ/Δ mice, respectively (Figure 1I). Taken together, our Northern, RT-PCR followed by sequencing, and Western analyses demonstrated that deletion of the floxed *LRRK1* and *LRRK2* alleles results in null mutations. Thus, floxed *LRRK1* and *LRRK2* alleles can be used to generate DA neuron-specific *LRRK* cDKO mice.

Generation and molecular characterization of DA neuron-specific *LRRK* cDKO mice

To generate DA neuron-specific *LRRK* cDKO mice, we used *DAT-Cre* KI mice, which express Cre recombinase under the control of the endogenous *DAT* promoter (32). To confirm if Cre-mediated recombination occurs broadly and specifically in DA neurons of the SNpc, we crossed *DAT-Cre* mice with the *Rosa26-CAG-LSL-ZsGreen1* reporter mouse (35). Upon Cre expression, Cre recombinase removes the floxed "stop" cassette, resulting in the expression of EGFP. We found that Cre-mediated recombination (GFP+) occurs in TH+ cells in the SNpc (Figure 2A). Quantification of GFP+ and/or TH+ cells in the SNpc showed that 99% of TH+ DA neurons are also GFP+, demonstrating that Cre-mediated recombination takes place in essentially all DA neurons in the SNpc (Figure 2B).

Having confirmed *fLRRK1* and *fLRRK2* mice as well as *DAT-Cre* mice, we then bred them together to generate *LRRK* cDKO mice (*fLRRK1/fLRRK1; fLRRK2/fLRRK2; DAT-Cre/+*), which were further bred with *fLRRK1/fLRRK1; fLRRK2/fLRRK2* mice to generate cDKO and littermate controls (*fLRRK1/fLRRK1; fLRRK2/fLRRK2*). It was previously reported that germline *LRRK* DKO mice failed to gain weight as they aged (30). However, DA neuron-specific *LRRK* cDKO and littermate control mice have similar body and brain weights at the ages of 2-24 months (Figure 2C, D). Western analysis showed a significant reduction of LRRK1 and LRRK2 proteins in the dissected ventral midbrain but not in the

cerebral cortex of DA neuron-specific *LRRK* cDKO mice at 2-3 months of age, relative to littermate controls (Figure 2E, F), further validating these DA neuron-specific *LRRK* cDKO mice.

Age-dependent loss of TH+ DA neurons in the SNpc of *LRRK* cDKO mice

To determine whether the inactivation of LRRK selectively in DA neurons of the SNpc affects their survival, we performed TH immunostaining and quantified TH+ DA neurons in the SNpc of *LRRK* cDKO mice and littermate controls using stereological methods. The morphology of TH+ DA neurons in *LRRK* cDKO mice at the ages of 15, 20, and 24 months appears normal (Figure 3A). Quantification of TH+ neurons in the SNpc using serial sections emcompassing the entire SNpc revealed that the number of DA neurons in the SNpc at the age of 15 months is similar between cDKO mice (10,000 ± 141) and littermate controls (10,077 ± 310; $F_{1,46} = 16.59$, $p = 0.0002$, two-way ANOVA with Bonferroni's *post hoc* multiple comparisons, $p > 0.9999$; Figure 3B). However, at the age of 20 months, the number of TH+ neurons in the SNpc of cDKO mice (8,948 ± 273) is significantly reduced, compared to controls (10,244 ± 220; $p = 0.0041$), and is further decreased at the age of 24 months (Control: 9,675 ± 232, cDKO: 8,188 ± 452; $p = 0.0010$; Figure 3B). Similar genotypic differences were observed in an independent quantification by another investigator, also conducted in a genotype blind manner, using the fractionator and optical dissector to randomly sample 25% area of the SNpc (Figure 3-figure supplement 1).

We further performed TH and NeuN double immunostaining of *LRRK* cDKO and control mice at 24 months of age (Figure 3C). Quantification of NeuN+ and TH+/NeuN+ cells in the SNpc using serial sections encompassing the entire SNpc showed that the number of NeuN+ neurons is also significantly reduced in the SNpc of cDKO mice (17,923 ± 813), compared to controls (21,907 ± 469, $p = 0.0006$, Student's *t*-test; Figure 3D). The number of TH+/NeuN+ cells is also lower in the SNpc of *LRRK* cDKO mice (10,500 ± 644), compared to control mice (14,102 ± 310, $p = 0.0001$; Figure 3D). These data indicate that the reduction in TH+ cells is not due to decreases of TH expression in DA neurons, but rather a result of the loss of DA neuron cell bodies in the SNpc of *LRRK* cDKO mice.

We further evaluated apoptosis in the SNpc of *LRRK* cDKO and littermate controls at the age of 24 months using an antibody specific for active Caspase-3 to label apoptotic cells. We observed increases in apoptotic DA neurons, labeled by active Caspase-3+ and TH+ immunoreactivity, in the SNpc of *LRRK* cDKO mice (Figure 4A). Quantification of active Caspase-3+/TH+ apoptotic DA neurons shows a significant increase in the SNpc of *LRRK* cDKO mice (323 ± 38), compared to controls (157 ± 8, p = 0.0004, Student's *t*-test; Figure 4B). These results further support that LRRK plays an intrinsic role in the survival of DA neurons in the SNpc during aging.

Age-dependent loss of TH+ DA terminals in the striatum of *LRRK* cDKO mice

To determine whether loss of DA neurons in the SNpc is accompanied with loss of DA terminals in the striatum, we performed TH immunostaining and quantified TH immunoreactivity in the striatum of *LRRK* cDKO and littermate control mice at the ages of 15 and 24 months (Figure 5A). Quantitative analysis showed normal levels of TH immunoreactivity in the striatum of cDKO mice at 15 months of age but reduced levels of TH immunoreactivity in the striatum of cDKO mice at 24 months of age (-19%, p = 0.0215, Student's *t*-test), suggesting an age-dependent loss of TH+ dopaminergic terminals in the striatum (Figure 5B).

Unaffected TH+ noradrenergic neurons in the LC of *LRRK* cDKO mice

Previously, we reported the reduction of noradrenergic neurons in the locus coeruleus (LC) of *LRRK* DKO mice (30). We therefore quantified TH+ noradrenergic neurons in the LC of *LRRK* cDKO mice at the age of 24 months. The number of TH+ cells in the LC is similar between cDKO and littermate controls (Control: 3,418 ± 86, cDKO: 3,350 ± 99, p = 0.6110, Student's *t*-test; Figure 6A, B). Further examination of the GFP reporter line crossed with *DAT-Cre* showed the lack of Cre-mediated recombination in the LC (Figure 6C), further support the cell intrinsic nature of DA neuron loss in the SNpc of aged *LRRK* cDKO mice.

Quantitative EM analysis of the SNpc in *LRRK* cDKO mice

We previously reported age-dependent increases of electron-dense autophagic and autolysosomal vacuoles as well as the presence of large lipofuscin granules in the surviving SNpc neurons of *LRRK* DKO mice beginning at 10 months of age (30, 31). To determine whether selective inactivation of LRRK in DA neurons similarly results in the accumulation of electron-dense vacuoles in the SNpc, we performed EM analysis in the SNpc of *LRRK* cDKO mice and littermate controls at the age of 25 months (Figure 7). We observed various electron-dense double membrane autophagosomes and single membrane autolysosomes as well as lipofuscin granules composed of lipid-containing residues of lysosomal digestion in the SNpc of *LRRK* cDKO and littermate control mice (Figure 7C-F). Interestingly, the number of electron-dense vacuoles in the SNpc is similar between *LRRK* cDKO mice and littermate controls at the age of 25 months (Control: 6.72 ± 0.43, cDKO: 6.99 ± 0.52, $p = 0.6839$, Student's *t*-test; Figure 7G). We also found no significant difference in the area of electron-dense vacuoles in the SNpc between *LRRK* cDKO and littermate controls (Control: 4.43 ± 0.44 μm^2; cDKO: 4.60 ± 0.49 μm^2, $p = 0.8048$; Figure 7G). The difference in accumulation of electron-dense vacuoles in the SNpc between germline DKO mice and DA neuron-restricted cDKO suggests that LRRK in non-DA neurons, possibly microglia, may play a more prominent role in the regulation of the autophagy-lysosomal pathway.

Enhanced microgliosis in the SNpc of *LRRK* cDKO mice

To determine whether selective inactivation of LRRK in DA neurons results in elevated microgliosis in the SNpc of *LRRK* cDKO mice. We performed immunohistochemical analysis of Iba1, which labels microglia, and TH, which marks DA neurons and processes, thus showing the boundary of the SNpc (Figure 8A-C). We found that the number of Iba1+ immunoreactive microglia is significantly increased in the SNpc of *LRRK* cDKO mice at 15 months of age (2541 ± 193), compared to controls (1737 ± 83; $F_{1,45} = 102.6$, $p < 0.0001$, two-way ANOVA with Bonferroni's *post hoc* multiple comparisons, $p = 0.0017$; Figure 8A, D). The number of Iba1+ immunoreactive microglia in the SNpc of *LRRK* cDKO is further increased compared to controls at the age of 20 (Control: 2426 ± 68, cDKO: 3639 ± 127, $p < 0.0001$; Figure 8B, D) and 25 months (Control: 2640 ± 187, cDKO: 4089 ± 100, $p < 0.0001$; Figure 8C,

D). These results show that despite the selective inactivation of LRRK in DA neurons of *LRRK* cDKO mice, microgliosis accompanies DA neuronal loss in the SNpc.

Impaired motor coordination of *LRRK* cDKO mice

To determine whether *LRRK* cDKO mice show motor deficits, we performed behavioral analysis of *LRRK* cDKO and littermate controls at the ages of 10 and 22 months using two versions of the elevated beam walk test with varying width of the beam (Figure 9). *LRRK* cDKO mice at 10 months of age displayed significantly more hindlimb slips/errors (4.4 ± 0.5) and longer traversal time (7.3 ± 0.3) in the 10 mm beam walk test, relative to littermate controls, which exhibited fewer slips (2.0 ± 0.3, $p = 0.0005$, Student's *t*-test) and shorter traversal time (5.8 ± 0.4, $p = 0.0075$; Figure 9A). In the 20 mm beam walk, which is less challenging than the narrower beam walk, both *LRRK* cDKO (1.5 ± 0.2) and littermate controls (1.0 ± 0.2, $p = 0.0733$) at 10 months of age performed well with few hindlimb slips and with no difference between *LRRK* cDKO and control mice. In addition, the traversal time is similar between *LRRK* cDKO (5.2 ± 0.3) and control littermates (5.1 ± 0.4, $p = 0.9796$; Figure 9A). These results show that *LRRK* cDKO mice at 10 months of age already exhibit deficits in motor coordination. However, in the pole test *LRRK* cDKO and control mice showed similar turning time (Control: 1.2 ± 0.1; cDKO: 1.5 ± 0.1; $p = 0.1219$) and descending time (Control: 4.7 ± 0.1; cDKO: 4.7 ± 0.2; $p = 0.8620$; Figure 9B). The *LRRK* cDKO and control mice at 10 months of age exhibit similar body weight (Figure 9C).

We then performed the beam walk test in naïve *LRRK* cDKO and control mice at 22 months of age (Figure 9D). The aged *LRRK* cDKO and control mice performed similarly in hindlimb errors ($p = 0.7022$) and traversal time ($p = 0.8139$) in 10 mm beam walk test. The hindlimb slips (Control: 12.0 ± 3.0; cDKO: 13.8 ± 3.3) and traversal time (Control: 15.3 ± 2.0; cDKO: 16.2 ± 2.8) of both genotypic groups were much higher, compared to the respective younger mice ($p < 0.0001$; Figure 9D). Moreover, higher percentage of the aged mice (2-3 out of 8-9 per genotypic group) failed the test, compared to the younger mice (0-2 mice out of 18-20 per genotypic group, Figure 9A, D). These results show that aged mice of both genotypes perform poorly in the challenging narrow beam walk test, suggesting that it is not optimal

to reveal subtle differences in motor coordination between the genotypic groups. The performance of *LRRK* cDKO and control mice at 22 months of age in the 20 mm beam walk test was also not significantly different with similar hindlimb errors (p = 0.7142) and traversal time (p = 0.2223; Figure 9D). In the pole test, *LRRK* cDKO mice and littermate controls also displayed similar turning time (p = 0.1184) and descending time to their home cage (p = 0.1413; Figure 9E) as well as body weight (Figure 9F).

Discussion

LRRK2 mutations represent ~5% familial and 1-2% of sporadic PD cases, and the G2019S mutation is the most common mutation but with higher age of onset and incomplete penetrance (74% at age 79) (36). Despite the importance of LRRK2 in PD pathogenesis, whether LRRK2 mutations cause PD and DA neurodegeneration *via* a loss- or gain-of-function mechanism remains unresolved, even though the distinction between these two pathogenic mechanisms is critical for directing LRRK2-based PD therapeutic development. Neither *LRRK2*-deficient mice nor *LRRK2* R1441C and G2019S KI mice develop dopaminergic neurodegeneration (25, 37, 38). It was proposed that the lack of brain phenotypes in *LRRK2*-/- mice might be due to the presence of LRRK1 (25, 26), which is broadly expressed in the brain including the midbrain (https://www.proteinatlas.org/ENSG00000154237-LRRK1/brain). Indeed, germline deletions of LRRK2 and LRRK1 results in age-dependent loss of DA neurons and increases of apoptosis and microgliosis in the SNpc of *LRRK* DKO mice (30, 31). However, the pleiotropic roles of LRRK1 and LRRK2 in the kidney (25, 26, 29), lung (28), and bone (39, 40), and the observed earlier mortality and lower body weight in *LRRK* DKO mice raised the possibility that DA neurodegeneration in *LRRK* DKO mice may be due to poor health.

To address this question, we generated floxed *LRRK1* and *LRRK2* mice (Figure 1, Figure1-figure supplement 1-10) and DA neuron-specific *LRRK* cDKO mice using the *DAT-Cre* KI allele (32) to delete floxed *LRRK1* and *LRRK2* regions selectively in DA neurons (Figure 2). Using an EGFP reporter mouse, we found that *DAT-Cre* drives Cre-mediated recombination in almost all DA neurons in the SNpc. Unlike

germline DKO mice (30), DA neuron-specific *LRRK* cDKO mice exhibit normal body weight and mortality during mouse lifespan. Importantly, *LRRK* cDKO mice develop age-dependent, progressive DA neurodegeneration, as evidenced by the normal number of DA neurons at the age of 15 months and the progressive reduction of DA neurons in the SNpc at the ages of 20 and 24 months (Figure 3), accompanied with increases of apoptotic DA neurons in the SNpc (Figure 4) and decreases of DA terminals in the striatum (Figure 5), whereas the number of noradrenergic neurons in the LC is unchanged, consistent with the lack of Cre-mediated recombination in the LC by *DAT-Cre* (Figure 6). Moreover, *LRRK* cDKO mice exhibit impaired performance in a narrow beam walk, a behavioral paradigm sensitive to altered motor coordination (Figure 9). Furthermore, microgliosis is elevated in the SNpc of *LRRK* cDKO mice, even though LRRK expression in microglia is unaffected in these mice, suggesting a cell extrinsic microglial response to pathophygiological changes in DA neurons (Figure 8). The molecular mechanism by which LRRK supports cell autonomous DA neuron survival is unknown. Single-cell RNA sequencing of dissected ventral midbrains may permit identification of genes and pathways that are selectively altered in DA neurons of aged *LRRK* cDKO mice before and after the onset of DA neurodegeneration.

Interestingly, quantitative EM analysis showed similar numbers of electron-dense vacuoles in the SNpc of *LRRK* cDKO and control mice at 25 months of age (Figure 7), in contrast to age-dependent increases of vacuoles in the SNpc of *LRRK* DKO mice (30, 31), suggesting non-DA neurons (*e.g.* microglia), in which LRRK expression is unaffected in DA neuron-specific *LRRK* cDKO mice but is absent in germline DKO mice, may contribute to this phenotypic difference. Development of microglia- and/or astrocyte-specific *LRRK* cDKO mice would permit further dissection of cell autonomous and non-cell autonomous roles of LRRK in DA neuron survival and determine whether glial LRRK plays a more important role in the regulation of the autophagy-lysosomal pathway and protein turnover. It would also be interesting to test whether reducing LRRK expression or function affects accumulation and aggregation of human mutant α-synuclein and tau in the aging brain, as both Lewy bodies and tauopathies are associated with PD patients carrying LRRK2 mutations (3).

The toxic gain-of-function pathogenic mechanism was initially proposed based on *in vitro* kinase assays using recombinant mutant LRRK2, which showed that R1441C and G2019S resulted in increases of autophosphorylation and phosphorylation of a generic substrate (41). Subsequent biochemical studies further reported that LRRK2 mutations (*e.g.* R1441C/G, G2019S) enhanced kinase activity, leading to elevated levels of pSer1292-LRRK2 (42) as well as pT73-Rab10 and pS106-Rab12 (43). While these biochemical changes are excellent biomarkers of LRRK2 mutations, there is no experimental evidence showing that increased phosphorylation of LRRK2 substrates drives DA neurodegeneration.

The loss-of-function pathogenic mechanism was prompted by mouse knockout findings showing that germline inactivation of LRRK results in age-dependent loss of DA neurons in the SNpc and DA terminals in the striatum (30, 31). The current study demonstrated a cell intrinsic role of LRRK in support of DA neuron survival in the SNpc of the aging brain, providing an unequovacal genetic evidence of an essential, cell autonomous requirement of LRRK in DA neurons. Furthermore, most transgenic mice overexpressing mutant LRRK2 did not produce neurodegeneration or DA neuron loss (44-52), though two studies reported that overexpression of LRRK2 G2019S but not R1441C resulted in DA neurodegeneration (53, 54). However, these transgenic mouse results are inconsistent with the human genetic findings, as R1441C is a more potent causative mutaion (highly penetrant, in a critical amino acid residue harboring 3 distinct PD mutations), whereas G2019S is known to be a weaker mutation with lower penetrance and older age of disease onset.

A genomic database analysis is sometimes quoted as the conclusive evidence for PD not caused by LRRK2 loss-of-function (55). However, the report was not a linkage study of large pedigrees carrying clear-cut loss-of-function mutations, such as large exonic deletions. Rather, it was a predictive analysis based on several available sequencing databases (141,456 individuals sequenced in the Genome Aggregation Database, 49,960 exome-sequenced individuals from the UK Biobank, and more than 4 million participants in the 23andMe genotyped dataset). As a result, the authors were careful in stating their conclusions. Therefore, there is no clear evidence to conclude whether loss-of-function mutations in LRRK2 cause PD or do not cause PD.

Genetically, it is well-known that missense mutations may gain a toxic function or may cause a partial loss of function *in cis* and a dominant negative inhibition of wild-type protein *in trans*, further reducing function. Missense mutations in the *PSEN* genes linked to familial Alzheimer's disease are good examples of such a loss-of-function coupled with a dominant negative mechanism (56-62). While PD-linked *LRRK2* mutations are mostly dominantly inherited missense mutations, though homozygous R1441H carriers have been reported (12), variants in LRRK1 have also been reported (21, 63). The notion that mutant LRRK2 may act in a dominant negative manner to inhibit the activity of wild-type LRRK2 is supported by structural and functional studies of LRRK2 as homodimers or heterodimers with LRRK1(22, 24, 64-68). Furthermore, LRRK2 G2385R variant, a risk factor of PD (69-72), has been reported as a partial loss-of-function mutation (73, 74). Future studies are needed to reconcile the biochemical findings of elevated kinase activity by LRRK2 mutations and the genetic findings of an essential physiological requirement of LRRK in DA neuron survival, and to determine whether LRRK2 mutations impair this relevant physiological function in protecting DA neurons while increasing kinase activity.

Materials and methods

Key Resource Table

Antibodies				
Reagent Name	**Source**	**Catalog #**	**Identifiers**	**Titer**
Rabbit anti-LRRK1	Alomone Lab	ANR-101	AB_2756700	WB: 1:1000
Rabbit anti-LRRK2	abcam	Ab133474	AB_2713963	WB: 1:1000
Mouse anti-α-Vinculin	Millipore	05-386	AB_309711	WB: 1:2000
Rabbit anti-GFP	abcam	ab290	AB_303395	IF: 1:1000
Mouse anti-TH	SantaCruz	Sc-25269	AB_628422	IF: 1:50
Rabbit anti-TH	abcam	Ab112	AB_297840	IHC: 1:750
Rabbit anti-NeuN	Cell Signaling Technology	12943S	AB_2630395	IF: 1:400
Rabbit anti-cleaved caspase-3	Cell Signaling Technology	9661	AB_2341188	IF: 1:150
Rabbit anti-Iba1	Wako	019-19741	AB_839504	IF: 1:500
Goat anti-rabbit IRdye800	LI-COR Biosciences	926-32211	AB_2651127	WB: 1:20000
Goat anti-mouse IRdye680	LI-COR Biosciences	926-68020	AB_2651128	WB: 1:20000
Alexa Fluor 488 goat anti-mouse IgG	ThermoFisher Scientific	A-11001	AB_2534069	IF: 1:250
Alexa Fluor 555 goat anti-rabbit IgG	ThermoFisher Scientific	A-32732	AB_2633281	IF: 1:250
Alexa Fluor 488 goat anti-rabbit IgG	ThermoFisher Scientific	A-11034	AB_2576217	IF: 1:250
Alexa Fluor 555 goat anti-mouse IgG	ThermoFisher Scientific	A-21424	AB_141780	IF: 1:250
Goat Biotinylated anti-Rabbit IgG (H&L)	Vector Laboratories	BA-1000	AB_2313606	IHC: 1:250
Oligonucleotides				
LRRK1 PCR Forward Primer for Left arm amplification: P1 5'-GACATCCGCGGCACCATGTGAGTGGCAGCTGTGGTGAGAAC-3' *Sac*II sequence is underlined.				
LRRK1 PCR Reverse Primer for Left arm amplification: P2 5'-GACATGCGGCCGCAAGCTTTTTAATAGCCGTTCTTTCTTAGAGAAGGCAG-3' *Not*I and *Hind*III sequences are underlined.				
LRRK1 PCR Forward Primer for Left-Middle arm amplification: P3 5'-CCAGTCACTTCTCCACCTCAGGGAAAATGG-3'				
LRRK1 PCR Reverse Primer for Left-Middle arm amplification: P4 5'-CCTTGTGGTACCCGGACCTTCTATCACCTTTATCC-3' *Kpn*I sequence is underlined.				
LRRK1 PCR Forward Primer for Right-Middle arm amplification: P5 5'-GGTCCGGGTACCACAAGGTGCTGGTTAAGTGCC-3' *Kpn*I sequence is underlined.				

LRRK1 PCR Reverse Primer for Right-Middle arm amplification: P6 5'-AGCAGACCTCTTGCCTTCTACTACTGACTG-3'
LRRK1 PCR Forward Primer for Right arm amplification: P7 5'- GACATGTCGACGGATCCGTAGGGAAGACCCACTAGGAGGAAGAAAG-3' *Sal*I sequence is underlined.
LRRK1 PCR Reverse Primer for Right arm amplification: P8 5'- GACATAAGCTTTGGTACCTTTCTAAAGGCAGCATTTTGCTTGC-3' *Hind*III sequence is underlined.
LRRK1 PCR Forward Primer for 5' Southern Probe: P15 5'-CAGAATACCCCATGCTGGGGAATTGC-3'
LRRK1 PCR Reverse Primer for 5' Southern Probe: P16 5'-CCGTTTCTAGGATTCTAATTTTTC-3'
LRRK1 PCR Forward Primer for 3' Southern Probe: P17 5'-AACTCCTTCCTGGTGCTGGCAGGCCTGGCTG-3'
LRRK1 PCR Reverse Primer for 3' Southern Probe: P18 5'-ACAAGTGACGTGACCATGGACGGAGCTGCG-3'
Neo PCR Forward Primer for Southern Probe: P23 5'-ATTCGGCTATGACTGGGCAC-3'
Neo PCR Reverse Primer for Southern Probe: P24 5'-GACCACCAAGCGAAACATCG-3'
LRRK1 PCR Forward Primer for exons 2-3 Northern Probe: P31 5'-CAGGATGAGCGTGTGTCTGCAG-3'
LRRK1 PCR Reverse Primer for exons 2-3 Northern Probe: P32 5'-CCTTCTCCTGTGAGGATTCGCTCT-3'
LRRK1 PCR Forward Primer for exons 27-29 Northern Probe: P33 5'-CTGGCCTACCTGCACAAGAA-3'
LRRK1 PCR Reverse Primer for exons 27-29 Northern Probe: P34 5'-CCTTCCCATCCCAGAACACC-3'
GADPH PCR Forward Primer for Northern Probe: P37 5'-ACCACAGTCCATGCCATCAC-3'
GADPH PCR Reverse Primer for Northern Probe: P38 5'-TCCACCACCCTGTTGCTGTA-3'
LRRK1 RT Primer in exons 32: P47 5'-GGCTCAGGTCATGCTCAGTT-3'
LRRK1 PCR Forward Primer for exons 4-8 RT-PCR: P53 5'-TTTTGGACACGCCGAAGTAGT-3'
LRRK1 PCR Reverse Primer for exons 4-8 RT-PCR: P54 5'-AGCCGCTCCAGGTAGTTTTT-3'
LRRK1 PCR Forward Primer for exons 11-17 RT-PCR: P57 5'-GGACCTCTCCAGAAACCAGC-3'
LRRK1 PCR Reverse Primer for exons 11-17 RT-PCR: P58 5'-GCAGGGTTGCTATCCTCTCC-3'
LRRK1 PCR Forward Primer for exons 20-25 RT-PCR: P63 5'-GCGGTCAGTGGCAAAGAATG-3'
LRRK1 PCR Reverse Primer for exons 20-25 RT-PCR: P64 5'-AATGCTGTTCTCACCCTCCG-3'
LRRK1 PCR Forward Primer for exons 25-31 RT-PCR: P41 5'-GAATTCTGCTAATGCCCCAGC-3'
LRRK1 PCR Reverse Primer for exons 25-31 RT-PCR: P42 5'-AGGCTGTAGATATAGATTTTCTGGT-3'

LRRK2 PCR Forward Primer for Left arm amplification: P9 5'-GAACACACAAGGCTATGGCTATTGTC-3'
LRRK2 PCR Reverse Primer for Left arm amplification: P10 5'-GTAGGACTATCATCCACCTGTAGGACTCC-3'
loxP site introducing oligo for *LRRK2*: P39 5'-GATCCATAACTTCGTATAATGTATGCTATACGAAGTTATGCTAGCA-3' *BamH*I and *Nhe*I sequences are underlined.
loxP site introducing oligo for *LRRK2*: P40 5'-CTAGTGCTAGCATAACTTCGTATAGCATACATTATACGAAGTTATG-3' *Spe*I and *Nhe*I sequences are underlined.
LRRK2 PCR Forward Primer for Middle arm amplification: P11 5'-GTCCTACAGGTGGATTCTAGACCTACAAGG-3' *Xba*I sequence is underlined.
LRRK2 PCR Reverse Primer for Middle arm amplification: P12 5'-GACGCGGCCGCGTTTAGTAGCTAGCATGACTATGAAGGAG-3' *Not*I and *Nhe*I sequences are underlined.
LRRK2 PCR Forward Primer for Right arm amplification: P13 5'-GCACTTGAGTCTTAATCTTGGGCAC-3'
LRRK2 PCR Reverse Primer for Right arm amplification: P14 5'-CATTCGAGCAGCTAAGCCTGTAATC-3'
LRRK2 PCR Forward Primer for 5' Southern Probe: P19 5'-CATGGGAGAGAGGGTTTCTCACTTACT-3'
LRRK2 PCR Reverse Primer for 5' Southern Probe: P20 5'-CTTGGACAGCATTGTCAGCCTAGAC-3'
LRRK2 PCR Forward Primer for 3' Southern Probe: P21 5'-GCCAAGCAGTTATTGATGCTGTAGC-3'
LRRK2 PCR Reverse Primer for 3' Southern Probe: P22 5'-CGAGCTGTAAGATGAGCTGGGTACT-3'
LRRK2 PCR Forward Primer for Northern Probe: P35 5'-AGGAAGGCAAGCAGATCGAG-3'
LRRK2 PCR Reverse Primer for Northern Probe: P36 5'-GGCTGAATATCTGTGCATGGC-3'
LRRK2 RT Primer in exons 51: P93 5'-TCGTGTGGAAGATTGAGGTCC-3'
LRRK1 PCR Forward Primer for Genotyping: P25 5'-ATTGGTCTTTGAAGAGACAGCATCTGG-3'
LRRK1 PCR Reverse Primer for Genotyping: P26 5'-TTTCCCTGAGGTGGAGAAGTGACTGG-3'
LRRK1 PCR Reverse Primer for Genotyping: P27 5'-TCACGTCGTCTAAGCCTCCT-3'
LRRK1 PCR Forward Primer for Genotyping: P28 5'-CTTCCTCAGAAGTTAGGTAAACATTG AGTG-3'
LRRK1 PCR Reverse Primer for Genotyping: P29 5'-CTAAGTGACACCGTGTTTCCAAAGTC-3'
LRRK1 PCR Reverse Primer for Genotyping: P30 5'-GGAAAGTTTCACAATTGGAAAAATAAAAATATTTACTGCAGATA-3'
DAT-IRES-Cre PCR Forward Primer for Genotyping: JKM1823 5'-TGGCTGTTGGTGTAAGTGG-3'
DAT-IRES-Cre PCR Reverse Primer for Genotyping: JKM1824 5'-GGACAGGGACATGGTTGACT-3'

DAT-IRES-Cre PCR Reverse Primer for Genotyping: JKM1825 5'-CCAAAGACGGCAATATGGT-3'

Mice

All animal use was approved by the IACUC committees of Harvard Medical School and Brigham and Women's Hospital, and conformed to the USDA Animal Welfare Act, PHS Policy on Humane Care and Use of Laboratory Animals, the "ILAR Guide for the Care and Use of Laboratory Animals" and other applicable laws and regulations. Mice were housed in constant humidity- and temperature-controlled rooms and maintained on a 12 hr light/dark cycle and were given standard rodent chow and water. Mice of both sexes at multiple ages, from 2 months to 25 months, were used. Floxed *LRRK1* (*fLRRK1*) and the resulting germline deleted *LRRK1*$^{\Delta}$ alleles, and floxed *LRRK2* (*fLRRK2*) and the resulting germline deleted *LRRK2*$^{\Delta}$ alleles were generated and thoroughly validated at the genomic DNA, mRNA, and protein levels. *DAT-IRES-Cre* (The Jackson Laboratory, IMSR_JAX:006660), *ACT-FLPe* (IMSR_JAX:005703), *CMV-Cre* transgenic (IMSR_JAX:006054), and *Rosa26-CAG-LSL-ZsGreen1* mice (IMSR_JAX:007906) used in the current study were previously characterized and reported (32-35). *LRRK1/LRRK2* cDKO mice (*fLRRK1/fLRRK1; fLRRK2/fLRRK2; DAT-IRES-Cre/+*) and littermate controls (*fLRRK1/fLRRK1; fLRRK2/fLRRK2*) were obtained from multiple breeding cages of *fLRRK1/fLRRK1; fLRRK2/fLRRK2* and cDKO mice. All mice were maintained on the C57BL6 and 129 hybrid genetic background (F1: IMSR_JAX:101043). All phenotypic analyses were performed in a genotype blind manner, as previously described (75).

Generation of targeted and floxed *LRRK1* and *LRRK2* alleles

The generation and validation of the *LRRK1* and *LRRK2* targeting vectors, the targeted, floxed, and deleted *LRRK1* and *LRRK2* alleles by Southern, Northern, RT-PCR, and the sequences of the floxed *LRRK1* and *LRRK2* alleles are included in **Figure 1-figure supplement 1-11**.

LRRK1

To generate the *LRRK1* targeting vector, we first PCR amplified the *left middle* homologous region (2,079 bp) containing partial intron 26, exon 27, and partial intron 27 of *LRRK1* from mouse BAC DNA (clone RP23-213J23, BACPAC Resources Center) using primers, P3 and P4. The PCR fragment was subcloned into the *pGEM-T* vector (A1360, Promega) to generate pLM1 (for details, see Figure 1-figure supplement 1). The *right middle* homologous region (3,403) containing *LRRK1* genomic region from partial intron 27 to partial intron 29 was amplified by PCR using primers, P5 and P6, and then subcloned into the *pGEM-T* vector (pLM2). The *left middle* homologous region containing *pGEM-T* plasmid was then digested with *Not*I and *Kpn*I (endogenous *Kpn*I site in the intron 27) and subcloned into the *Not*I and *Kpn*I sites of the *right middle* homologous region containing *pGEM-T* plasmid (pLM3). After that, the *middle* homologous region (5.5 kb), from partial intron 26 to partial intron 29, was released by *Not*I and *Sac*II digestions and was blunted by Klenow, and then subcloned into the *Sma*I site of *PGKneoF2L2DTA* vector (#13445, addgene) to generate the *middle* homologous region-*PGKneoF2L2DTA* plasmid (pLM4).

The *left* homologous region (2,016 bp) containing partial intron 25, exon 26, and partial intron 27 of *LRRK1* was PCR amplified from mouse BAC DNA (clone RP23-213J23, BACPAC Resources Center) using primers, P1 and P2. The PCR fragment was digested with *Sac*II (introduced by P1) and *Not*I (introduced by P2 along with *Hind*III), and was subcloned into the *Sac*II and *Not*I sites of the *PGKneoF2L2DTA* vector (pLM5). The *right* homologous region (3,131 bp) containing partial intron 29, exon 30, and partial intron 30 of *LRRK1* was PCR amplified from mouse BAC DNA (clone RP23-213J23) using primers, P7 and P8. The PCR fragment was digested with *Sal*I (introduced by P7) and *Hind*III (introduced by P8), and was subcloned into the *Sal*I and *Hind*III sites of the *PGKneoF2L2DTA* vector (pLM6).

The *left* homologous region in the *PGKneoF2L2DAT* vector (pLM5) was digested with *Sac*II and *Not*I, and subcloned into the *Sac*II and *Not*I sites of the *right* homologous region containing *pGKneoF2L2DTA* plasmid (pLM6) to generate pLM7. Finally, the *loxP-middle* homologous region-*PGK-Neo-loxP* fragment was released from pLM4 by *Not*I and *Sal*I digestions, and subcloned into the *Not*I and

*Sal*I sites of pLM7 to generate the final target vector (pLM8), which contains two *loxP* sites (intron 26 and intron 29). Upon Cre-mediated recombination, the endogenous *LRRK1* genomic sequence flanked by the 5' *loxP* site (1,288 bp upstream of *LRRK1* exon 27) to the 3' *loxP* site (1,023 bp downstream of exon 29) is excised. The *PGK-neo* cassette flanked by two *FRT* sites is under the control of the mouse *phosphoglycerate kinase 1* (*PGK*) promoter and contains the *bovine growth hormone* polyA signal. To enhance the ratio of ES cells carrying homologous recombination events instead of random insertion of the targeting vector (76), the negative selection *PGK-DTA* cassette, which encodes diphtheria toxin A chain, was also included in the *LRRK1* targeting vector.

The *LRRK1* targeting vector was linearized by *Xho*I digestion, and then electroporated into MKV6.5 embryonic stem (ES) cells (a gift from R. Jaenisch), which were derived from B6/129 F1 mice (The Jackson Laboratory, IMSR_JAX:101043). G418 was applied to the culture at 150 µg/ml 24 h later, and after 6 days of G418 selection, the surviving ES clones (480) were picked and screened by Southern analysis using *Hind*III digestion of genomic DNA followed by hybridization with the 5' external and 3' external probes to confirm proper recombination events in the 5' and 3' homologous regions, respectively. 25 ES cell clones were confirmed to carry the proper homologous recombination events at the 5' homologous region, giving rise to the 17.0 kb and the 4.8 kb bands, which represent the wild-type and the targeted allele, respectively (Figure 1D), and the 3' homologous region, giving rise to the 17.0 kb and the 14.1 kb bands, which represent the wild-type and the targeted allele, respectively. We then expanded the selected 4 ES cell clones (3A11, 3D8, 3H1, and 3H6) and further verified by Southern using the 5' and 3' external, and *neo* probes. Two ES cell clones (3D8 and 3H6) were transfected with *pCAGGS-flpE-puro* (#20733, addgene) to delete the *PGK-neo* cassette by FLP recombinase. Three resulting ES cell clones (3D8C5, 3D8E5, and 3H6G7) were confirmed by Southern analysis using the 5' and 3' external probes and the *neo* probe to confirm the floxed *LRRK1* allele by the deletion of the *PGK-neo* cassette.

LRRK2

To generate the *LRRK2* targeting vector, the *left* homologous region (2,579 bp) containing the *LRRK2* promoter region was PCR amplified from mouse BAC DNA (clone RP23-526A2, BACPAC Resources Center) using primers, P9 and P10, and subcloned into the *pGEM-T* vector to generate pLRRK2#1 (for details see Figure 1-figure supplement 4). The *BamH*I-*loxP*-*Nhe*I-*Spe*I fragment (56 bp), which was generated by annealing two complementary oligos, P39 and P40, was introduced to the *BamH*I and *Spe*I sites of pLRRK2#1 to generate pLRRK2#2. The *middle* homologous region, containing genomic sequences from the promoter region to partial intron 2, was PCR amplified from mouse BAC DNA (clone RP23-526A2) using primers, P11 and P12, digested with *Xba*I and *Not*I, and subcloned into the *Xba*I and *Not*I sites of the *pGEM-T* vector to generate pLRRK2#3, which was digested with *Xba*I and *Not*I, and subcloned into the *Xba*I and *Not*I sites of pLRRK2#2 to generate pLRRK2#4.

The *right* homologous region (3,503 bp), which contains partial intron 1, exon 2, and partial intron 2, was PCR amplified from mouse BAC DNA (clone RP23-526A2) using primers, P13 and P14, and the PCR fragment was subcloned into the *EcoR*V site of *pBluescript II KS* (+) vector (212207, Agilent) to generate pLRRK2#5. The *right* homologous region in pLRRK2#5 was digested with *BamH*I followed by Klenow, and then digested with *Cla*I. The released fragment was subcloned into the *EcoR*V and *Cla*I sites of the *PGKneoF2L2DTA* vector to generate pLRRK2#6. *LFNT-tk/pBS* plasmid (a gift from S. Tonegawa) was digested with *Sac*II (followed by Klenow) and *Not*I (followed by *Ssp*I digestion to make it easier to isolate the *Not*I-*Sac*II/KN fragment) to release the *loxP-FRT-PKG-neo-loxP-FRT* fragment (2,928 bp), and then subcloned into the *Xba*I (followed by Klenow) and *Not*I sites of pLRRK2#6 to generate pLRRK2#7. Finally, pLRRK2#4 containing the *left-loxP-middle* homologous regions was digested with *EcoR*V and *Not*I, and subcloned into *Ale*I and *Not*I sites of pLRRK2#7 containing the *loxP-FRT-PKG-neo- loxP-FRT-right* homologous region to generate the final targeting vector, which contains the *loxP* sites in the promoter region (1,768 bp upstream of the transcription initiation site) and in *LRRK2* intron 2 (878 bp downstream of exon 2). Upon Cre-mediated recombination, the floxed endogenous *LRRK2* genomic sequences from the 5' *loxP* site (1,768 bp upstream of *LRRK2*

transcription) to the 3' *loxP* site in intron 2 (878 bp downstream of exon 2) are excised. The negative selection *PGK-DTA* cassette is also included in the *LRRK2* targeting vector.

The *LRRK2* targeting vector was linearized by *Ahd*I digestion, and then electroporated into MKV6.5 ES cells. G418 was applied to the culture at 150 ug/ml 24 h later, and after 6 days of G418 selection, the surviving ES clones (480) were picked and screened by Southern analysis using *Nhe*I digestion of genomic DNA followed by hybridization with the 5' external probe (753 bp, PCR amplified using P19 and P20). Southern analysis confirmed that two ES cell clones (N24 and N34) carry the proper recombination event in the 5' homologous arm, giving rise to 11.6 kb and 3.6 kb bands, which represent the wild-type and targeted alleles, respectively (Figure 1F), followed by genomic PCR confirmation for the proper recombination in the right arm.

Three (3D8C5, 3A11, 3H6) and one (N24) ES clones for *LRRK1* and *LRRK2*, respectively, were microinjected into C57BL/6 mouse blastocysts to generate chimera mice, which were bred with B6/129 F1 mice to produce heterozygous floxed *LRRK1* and targeted *LRRK2* mice. Floxed *LRRK1* mice were confirmed by Southern analysis using the 5' and 3' external probes (Figure 1D). Targeted *LRRK2* mice were confirmed by Southern (data shown in Figure 1-figure supplement 6; *Nhe*I digestion followed by hybridization with the 5' external probe and *Sph*I digestion followed by hybridization with the 3' external probe). Targeted *LRRK2* mice were then bred with *Actin-FLP* deleter mice (IMSR_JAX:005703) (33) to generate floxed *LRRK2* mice, which were confirmed by Southern analysis using the 5' and 3' external probes following *Nhe*I digestion (Figure 1F). Heterozygous *fLRRK1*/+ and *fLRRK2*/+ mice were crossed with each other to obtain homozygous single floxed mice (*fLRRK1/fLRRK1* and *fLRRK2*/*fLRRK2*) and double floxed mice (*fLRRK1/fLRRK1*; *fLRRK2*/*fLRRK2*).

Generation of deleted *LRRK1/LRRK2* alleles and DA neuron-specific cDKO mice

In order to ensure that Cre-mediated deletion of the floxed *LRRK1* and *LRRK2* alleles results in null alleles, we crossed floxed *LRRK1* and *LRRK2* mice with germline deleter, *CMV-Cre* transgenic mice (*B6.C-Tg(CMV-cre)1Cgn/J*; IMSR_JAX:006054) (34), to generate *LRRK1* deleted (Δ/Δ) mice (by

removing exons 27-29) and *LRRK2* deleted (Δ/Δ) mice (by removing the promoter region and exons 1-2) for further molecular characterization. To generate DA neuron-specific *LRRK1*/*LRRK2* cDKO mice, we used *DAT-Cre* KI mice (*B6.SJL-Slc6a3tm1.1(cre)Bkmn/J;* IMSR_JAX:006660), which express Cre recombinase under the control of the endogenous *DAT* promoter (32). We crossed double floxed mice with *DAT-Cre* mice to generate *LRRK* cDKO mice (*fLRRK1*/*fLRRK1; fLRRK2*/*fLRRK2*; *DAT-Cre/+*). *LRRK* cDKO and littermate control mice used in the phenotypic analysis were obtained by crossing *fLRRK1*/*fLRRK1; fLRRK2*/*fLRRK2; DAT-Cre/+* with *fLRRK1*/*fLRRK1; fLRRK2*/*fLRRK2* mice. We only used cDKO and control mice that carry all floxed *LRRK1* and *LRRK2* alleles for phenotypic analysis.

Southern analysis

For the identification and validation of the targeted and floxed *LRRK1* alleles, we used the 5' external, 3' external, and *neo* probes. Genomic DNA from ES cells or mouse tails was digested with *Hind*III. The 5' external probe (377 bp), which is 839 bp upstream of the 5' homologous region, was PCR amplified from mouse BAC DNA (clone RP23-213J23) using primers P15 and P16. The 3' external probe (305 bp), which is 2,736 bp downstream of the 3' homologous region, was PCR amplified from mouse BAC DNA (clone RP23-213J23) using primers P17 and P18. The *neo* probe (363 bp) was PCR amplified from pSoriano plasmid using primers P23 and P24. Following *Hind*III digestion, the presence of the 17.0 kb product using either the 5' or 3' external probe represents the wild-type allele, whereas the 4.8 kb (the 5' external probe) and the 12.2 kb (the 3' external probe) products represent the floxed *LRRK1* allele. Genomic DNA digested by *Hind*III and hybridized with the *neo* probe further confirmed the wild-type and floxed alleles (no band) and the targeted *LRRK1* allele (14.1 kb).

For the identification and validation of the targeted and floxed *LRRK2* alleles, we used the 5' and 3' external probes as well as the *neo* probe. Genomic DNA from ES cells or mouse tails was digested with *Nhe*I or *Sph*I followed by hybridization with the 5' or 3' external probe or the *neo* probe. The 5' external probe (753 bp), which is 84 bp upstream of the 5' homologous region, was PCR amplified from mouse BAC DNA (clone RP23-526A2) using primers P19 and P20. The 3' external probe (622 bp),

which is 38 bp downstream of the 3' homologous region, was PCR amplified from mouse BAC DNA (clone RP23-526A2) using primers P21 and P22. Following *Nhe*I digestion, the presence of the 11.5 kb product using either the 5' or 3' external probe represents the wild-type allele, whereas the 3.6 kb (the 5' external probe) and the 5.2 kb (the 3' external probe) products represent the floxed *LRRK2* allele.

PCR genotyping

Genomic PCR was performed to determine the presence of the deleted, the floxed, and/or the wild-type alleles. For *LRRK1*, the following primers were used: P25 (5'-ATTGGTCTTTGAAGAGACAGCATCTGG, forward primer, 392 nt downstream of exon 26), P26 (5'-TTTCCCTGAGGTGGAGAAGTGACTGG, reverse primer, 567 nt downstream of exon 26), and P27 (5'-TCACGTCGTCTAAGCCTCCT, reverse primer, 1,218 nt downstream of exon 29). The PCR products from P25 and P26 are 266 bp and 405 bp, which represent the wild-type and the *fLRRK1* alleles, respectively, whereas the PCR product from P25 and P27 is 583 bp, which represents the deleted *LRRK1* allele.

For *LRRK2*, the following primers were used: P28 (5'-CTTCCTCAGAAGTTAGGTAAACATTGAGTG, forward primer, 2,069 nt upstream of exon 1), P29 (5'-CTAAGTGACACCGTGTTTCCAAAGTC, reverse primer, 1,739 nt upstream of exon 1), and P30 (5'-GGAAAGTTTCACAATTGGAAAAATAAAAATATTTACTGCAGATA, reverse primer at 2,848 nt downstream of exon 2). The PCR products from P28 and P29 are 305 bp and 367 bp, representing the wild-type and the *fLRRK2* alleles, respectively, whereas the PCR product from P28 and P30 is 587 bp, which represents the deleted *LRRK2* allele.

For *DAT-IRES-Cre*, the following primers were used: JKM1823 (5'-TGGCTGTTGGTGTAAAGTGG, forward primer at exon 16 and 3'-UTR), JKM1824 (5'-GGACAGGGACATGGTTGACT, reverse primer at 3'-UTR), and JKM1825 (5'-CCAAAAGACGGCAATATGGT, reverse primer at *IRES* sequence). The PCR product from JKM1823

and JKM1824 is 264 bp, which represents the wild-type allele, whereas the PCR product from JKM1823 and JKM1825 is 152 bp, which represents the *DAT-IRES-Cre* KI allele.

Northern analysis

Total RNA was isolated from brains, kidneys, or lungs using TRI reagent (T9424, Millipore-Sigma) according to the manufacturer's instruction. For the *LRRK1* Northern analysis, polyA+ RNA was enriched from ~500 μg total RNA using the Poly(A)Purist™ MAG Kit (AM1922, ThermoFisher) according to the manufacturer's instruction. For the *LRRK2*, ~10 μg of total RNA was used for Northern analysis. RNA was separated in formaldehyde agarose gel, and transferred into Amersham Hybond-nylon membrane (RPN303N, GE Healthcare). Probes were synthesized using Prime-It II random labeling kit (#300385, Agilent) and then used for membrane hybridization at 55ºC overnight.

The cDNA probe specific for *LRRK1* exons 2-3 (383 bp) was PCR amplified using primers P31 (5'-CAGGATGAGCGTGTGTCTGCAG) and P32 (5'-CCTTCTCCTGTGAGGATTCGCTCT). The cDNA probe specific for *LRRK1* exons 27-29 (550 bp) was PCR amplified using primers P33 (5'-CTGGCCTACCTGCACAAGAA) and P34 (5'-CCTTCCCATCCCAGAACACC).

The cDNA probe specific for *LRRK2* exons 1-5 (437 bp) was PCR amplified using primers P35 (5'-AGGAAGGCAAGCAGATCGAG) and P36 (5'-GGCTGAATATCTGTGCATGGC). The probe specific for *GAPDH* exons 5-7 (452 bp) was PCR amplified using primers P37 (5'-ACCACAGTCCATGCCATCAC) and P38 (5'-TCCACCACCCTGTTGCTGTA).

Hybridization was performed using α-^{32}P-dCTP-labeled probes specific to each gene. Specific signals were detected by autoradiography with Hyperfilm (E3018, Amersham).

RT-PCR

Total RNA was isolated from brains, kidneys, or lungs using TRI reagent (T9424, Millipore-Sigma,) according to the manufacturer's instruction. Approximately 1 μg of RNA was reverse-transcribed using Superscript III (18080093, Thermo Fisher Scientific) according to the manufacturer's instructions. For RT-PCR analysis of *LRRK1* transcripts in mice carrying the homozygous floxed or deleted alleles, we

used primers P53 and P54 for exons 4-8 (714 bp), P57 and P58 for exons 11-17 (818 bp), P63 and P64 for exons 20-25 (922 bp), or P41 and P42 for exons 25-31 to confirm normal splicing of *LRRK1* mRNA in *fLRRK1/fLRRK1* mice and truncated *LRRK1* transcripts lacking exons 27-29 in Δ/Δ mice (for details see Figure 1-figure supplement 9). For RT-PCR analysis of *LRRK2* transcripts, we used primers P35 and P36 in exons 1-5 to confirm normal splicing of *LRRK2* mRNA (437 bp) in *fLRRK2/fLRRK2* mice and the absence of RT-PCR products in *LRRK2* Δ/Δ mice (Figure 1-figure supplement 11). The identity of the PCR products was confirmed by sequencing.

Western analysis

Fresh tissues were collected and homogenized in an ice-cold stringent RIPA buffer (50 mM Tris-Cl (pH 7.6), 150 mM NaCl, 0.5 mM EDTA, 1% NP40, 0.5% sodium deoxycholate, 0.1% SDS, 1mM PMSF supplement with protease inhibitor cocktail (P8340, Sigma) and phosphatase inhibitor cocktail (P0044, Sigma)), followed by sonication. Homogenates were centrifuged at 14,000xg for 20 min at 4ºC to separate supernatants (RIPA buffer-soluble fraction). An equal amount (10-40 μg per lane) of total proteins from each preparation were loaded and separated on NuPAGE gels (Invitrogen), and transferred to nitrocellulose membranes. The membranes were blocked in Intercept (TBS) Blocking Buffer (927-60001, Li-Cor) for 1 hr at room temperature, and incubated at 4ºC overnight with specific primary antibodies. Primary antibodies used were rabbit anti-LRRK1 (ANR-101, Alomone Lab, RRID: AB_2756700), rabbit anti-LRRK2 (ab133474, abcam, RRID: AB_2713963), mouse anti-α-Vinculin (05-386, Millipore, RRID: AB_309711). Membranes were then incubated with dye-coupled secondary antibodies, goat anti-mouse IRdye680 (#925-68070, LI-COR, RRID: AB_2651128) or goat anti-rabbit IRdye800 (#925-32211, LI-COR, RRID: AB_2651127). Signals were quantified using the Odyssey Infrared Imaging System (LI-COR).

Histological analysis

Mice were anesthetized with ketamine (100 mg/kg) + xylazine (10 mg/kg) + acepromazine (3 mg/kg), and transcardially perfused with phosphate-buffered saline solution (PBS, pH 7.4) containing

0.25 g/L heparin (H3149, Sigma) and 5 g/L procaine (P9879, Sigma). Brains were post-fixed in 4% formaldehyde in PBS (pH 7.4) (15710, Electron Microscopy Sciences) at 4˚C overnight and then processed for paraffin embedding following standard procedures. For frozen sections, post-fixed brains were immersed in a sucrose series solution (15% and 30% sucrose in PBS) at 4ºC overnight for cryoprotection, and then brains were embedded in an OCT compound (4583, Sakura). Serial coronal sections (16 μm) of paraffinized brains or frozen brains were obtained using Leica RM2235 or Leica CM1860, respectively. Coronal sections containing the SNpc or LC were selected for immunohistochemical analysis.

Histological analyses were performed as described previously (30, 31, 75). Briefly, for DAB-derived TH-immunohistochemistry, coronal sections were deparaffinized, alcohol-dehydrated, and then subjected to permeabilization with a solution containing 0.1% Triton X-100 in TBS followed by antigen retrieval for 5 min in 10 mM sodium citrate buffer, pH 6.0. Endogenous peroxidase activity was quenched by incubating in 0.3% H_2O_2 in methanol. Sections were then blocked with a solution containing 5% normal goat serum (S-1000, Vector Laboratories) and 0.1% Triton X-100 in TBS for 1 hr at room temperature. After blocking, sections were incubated with the primary antibody, rabbit anti-TH (1:750, ab112, abcam, RRID: AB_297840) overnight at 4ºC. Sections were washed three times in 0.1% Triton X-100 in TBS followed by 1 hr incubation with goat biotinylated anti-rabbit IgG secondary antibody (1:250, BA-1000, Vector Laboratories, RRID: AB_2313606) at room temperature and 30 min incubation with Vectastain Elite ABC reagent (PK-6100, Vector Laboratories) and then developed using chromogenic DAB substrate (SK-4100, Vector Laboratories).

For immunofluorescence staining of paraffin sections, coronal sections were deparaffinized, alcohol-dehydrated, and then subjected to permeabilization with a solution containing 0.1% Triton X-100 in PBS followed by antigen retrieval for 5 min in 10 mM sodium citrate buffer, pH 6.0 except those for cleaved-caspase3 immunostaining which were performed antigen retrieval for 10 min. Sections were then blocked with a solution containing 10% normal goat serum and 0.1% Triton X-100 in PBS for 1 hr at room temperature. After blocking, sections were incubated with primary antibodies overnight at 4ºC. The

primary antibodies used were mouse anti-TH (1:50, sc-25269, Santa Cruz, RRID: AB_628422), rabbit anti-NeuN (1:400, 12943S, Cell Signaling Technology, RRID: AB_2630395), rabbit anti-cleaved caspases-3 (1:150, 9661, Cell Signaling Technology, RRID: AB_2341188), or rabbit anti-Iba1 (1:500, 019-19,751, Wako, RRID: AB_839504). Sections were washed three times in 0.1% Triton X-100 in PBS followed by 1 hr incubation with fluorophore-conjugated secondary antibodies, goat anti-mouse IgG Alexa Fluor 488 (1:500, A11001, ThermoFisher, RRID: AB_2534069) and goat anti-rabbit IgG Alexa Fluor 555 (1:500, A32732, ThermoFisher, RRID: AB_2633281) at room temperature. Fluorescence images were taken (a stack of 2-3 confocal images spaced at 4 μm), projected with maximal intensity projection mode, and analyzed by FV1000 confocal microscope system (Olympus).

For immunofluorescence staining of cryopreserved sections, coronal brain sections were washed with PBS to rinse out OCT, and then blocked with a solution containing 5% normal goat serum and 0.1% Triton X-100 in TBS for 1 hr at room temperature. After blocking with 10% NGS, sections were incubated with primary antibodies, rabbit anti-GFP (1: 1000, ab290, abcam, RRID: AB_303395), and mouse anti-TH (1:50, sc-25269, Santa Cruz, RRID: AB_628422), overnight at 4ºC. Sections were incubated with fluorophore-conjugated secondary antibodies, goat anti-rabbit IgG Alexa Fluor 488 (1:500, A11034, ThermoFisher, RRID: AB_2576217) and goat anti-mouse IgG Alexa Fluor 555 (1:250, A21424, ThermoFisher, RRID: AB_141780) for 1 hr at room temperature. Fluorescence images were taken and analyzed by FV1000 confocal microscope system (Olympus).

The number of GFP+ and GFP+/TH+ cells in the SNpc of *DAT-IRES-Cre/+; Rosa26-CAG-LSL-ZsGreen1/+* reporter mice were quantified using 3 comparable coronal sections (16 μm in thickness, spaced 320 μm apart) per brain (n = 3 brains, one hemisphere). The percentage of GFP+ DA neurons in the SNpc was obtained by dividing the sum of GPF+/TH+ neurons by the sum of total TH+ neurons quantified.

Quantification of DA neurons in the SNpc

Quantification of TH+ DA neurons in the SNpc or LC was performed as previously described (30, 31, 75, 77). Briefly, TH+ neurons in the SNpc, which were marked based on morphological features as previously described (78), were quantified in every 10th serial coronal section (16 μm in thickness) throughout the SNpc (a total of 6-9 sections, spaced 160 μm apart). Total number of TH+ cells in the SNpc was calculated as follows: [total number of TH+ DA neurons quantified in all 6-9 sections] x 10 (every 10th section sampled) x 2 (both hemispheres). Total number of TH+ noradrenergic neurons in the LC (a total of 6-9 sections, spaced 80 μm apart) was calculated as follows: [total number of TH+ noradrenergic neurons quantified in all 6-9 sections] x 5 (every 5th section sampled) x 2 (both hemispheres).

DA neuron quantification was also performed independently by another investigator, also in a genotype blind manner, using stereological quantification of 25% of the SNpc area. The fractionator with 100 μm x 100 μm was set up in the SNpc, and TH+ DA neurons were counted using the optical dissector method with 50 μm x 50 μm sample box (25% of the total area). Total number of TH+ cells in the SNpc was calculated as follows: [total number of TH+ DA neurons quantified in sample boxes of all 6-9 sections] x 10 (every 10th section sampled) x 4 (1/4 area sampled: 50 x 50/100 x 100) x 2 (both hemispheres).

Quantification of NeuN+ neurons, apoptotic cells, Iba1+ microglia in the SNpc

NeuN+, active Caspase-3+, or Iba1+ cells in the SNpc, which was marked by TH immunoreactivity, were quantified using serial coronal sections (16 μm in thickness, every 10th section, a total of 6-9 sections per brain). Compared to DAB immunostaining followed by counting under the stereomicroscope, which captures TH-positive DA neurons in a single cell layer of the brain section, immunofluorescent staining picks up more cells (*e.g.* TH+ or NeuN+) in multiple layers of the brain section under the confocal microscope by the maximal intensity projection mode. Thus, more TH-positive DA neurons in the SNpc were captured and quantified under confocal microscopy compared to DAB staining. The total number of NeuN+ cells in the SNpc was calculated by multiplying the total number of

NeuN+ cells in all sections counted] x 10 (every 10th section sampled) x 2 (both hemispheres). The total number of active Caspase-3+ or Iba1+ cells in the SNpc, which was marked by TH immunoreactivity, was calculated as follows: [total number of active Caspase-3+ or Iba1+ cells in all sections counted from one hemisphere] x 10 (every 10th section sampled).

Quantification of TH+ DA terminals in the striatum

For quantification of TH immunoreactivity in the striatum, we performed immunostaining using every 10th serial coronal sections (16 μm in thickness) throughout the entire striatum (a total of 12-15 sections, spaced 160 μm apart). The images of TH immunoreactivity in the striatum were captured under 2X objective lens (Olympus BX40, 8-bit RGB camera) using identical exposure time, sensitivity, brightness, contrast, and gamma (CellSens Entry Software) and then analyzed using the Fiji version of ImageJ, and the optical density was determined as previously described (31, 79, 80). The mean value of TH immunoreactivity in the striatum of control mice of each age group was set as 100%.

Quantitative EM analysis

The collection and quantification of the EM images were performed as described previously (30, 31). Mice were perfused with PBS containing 0.25 g/L heparin and 5 g/L procaine followed by a fixative solution containing 2.5% paraformaldehyde and 2.5% glutaraldehyde in 0.1 M sodium cacodylate buffer (pH 7.4) (#1549, Electron Microscopy Science). Brains were dissected and post-fixed overnight in a fixative solution at 4°C. The dissected tissues were trimmed to 1-2 mm^3 cubes followed by osmication and uranyl acetate staining, dehydration in graded alcohol, and embedded in TABB 812 Resin (Marivac Ltd.) at the Harvard Medical School EM facility. 0.5 μm sections were stained with toluidine blue and viewed under the light stereomicroscope (Nikon Eclips E600) to find the SNpc area for EM viewing. Then adjacent sections were cut with 80 nm in thickness with the Leica Ultracut S microtome, picked up on formvar-carbon coated slot Copper grids, stained with 0.2% Lead Citrate, and viewed and imaged under the JEOL 1200x Electron Microscope. A minimum of 10 micrographs containing the entire cell body in the SNpc area were analyzed for each brain. The image was analyzed by using the Fiji version of

ImageJ. The number of electron-dense autophagic and lysosomal vacuoles (>0.5 μm in diameter) in individual neuronal profiles was quantified. We previously calculated the diameter of electron-dense autophagic/lysosomal vacuoles by measuring the longest side manually (30, 31). In the current study, we used Feret's Diameter (81) to accurately measure the longest distance between any of two points in the electron-dense autophagic/lysosomal vacuoles, resulting in a higher number of electron-dense vacuoles quantified. The number and the area of electron-dense vacuoles (>0.5 μm in diameter) in individual neuronal profiles was quantified, and the average number or area of vacuoles per mouse was calculated. Experiments were done in a genotype blind manner (after scarifying mice, the brain samples were coded and sent to the Harvard Medical School EM Core, where the images were captured, and the vacuoles were quantified by another independent experimenter).

Behavioral analysis

Naive *LRRK* cDKO mice and littermate controls at 10 and 22 months of age were used. Mice were acclimated in the behavior facility for a minimum of 7 days and were then individually handled daily for 5 consecutive days before testing. Mice were coded so the experimenter was unaware of their genotypes until the data analysis was complete. In the beam walk test, a Plexiglas beam of 100 cm in length (Plastic Zone), 20 mm or 10 mm in width, was raised 60 cm above a table, and safety bedding was placed under the beam to avoid any harm in case of falls. Mice were placed onto the starting point of the beam in bright light, and the time (in seconds) to reach their home cage on the other darker side of the beam (~80 cm in distance) as well as the hindpaw slips (number of hindlimb errors) were recorded. Mice were trained three trials per day for 2 consecutive days to traverse the 20 mm beam (without the wire mesh) to their home cage. On the test day, mice were trained further with two additional trials on the 20 mm beam (without the wire mesh). Mice were then tested in two successive trials on the 20 mm beam (with the wire mesh) followed by two consecutive test trials on the 10 mm beam (with the wire mesh). All test trials were videotaped, and the travel time and the number of hindlimb errors were recorded. Between

trials mice were placed in the home cage for 2 min to rest. Mice that fell off the beam during both trials or stalled on the beam for more than 120 sec during the test were excluded.

In the pole test, mice were placed on top of the pole (60 cm in height, 10 mm in diameter) with their head facing upward, and the base of the pole was placed in the home cage. Mice were trained three trials per day for 2 days to traverse the pole to the cage floor and were further trained two trials before testing on the test day. Mice were then tested for 2 trials, and the time to turn around (turning time) and the time to descend the pole (descending time) were recorded. Between trials, mice were placed in the home cage for 2 min to rest. Mice that stalled on the top of the pole for more than 120 sec were excluded.

Experimental design and statistical analysis

All experiments were performed in a genotype blind manner with the exception of molecular analysis (Southern, Northern, RT-PCR, and Western). All statistical analyses were performed using Prism 9 (GraphPad software) or Excel (Microsoft). All data are presented as the means ± SEM. The exact sample size of each experiment is indicated in the figure or the legend. The slight difference in sample size among various histological analysis (*e.g.* at 24M, 11 control and 8 cDKO brains analyzed for TH+ cells but 9 control and 8 cDKO analyzed for NeuN+, Caspase-3+, Iba1+ cells in the SNpc) is due to the paraffin blocks used for quantification (each paraffin block contains 7 brains of mixed genotypes; for details see Source Data files) or specific brain sections being damaged, thus, excluded in the analysis.

Statistical analyses were conducted using an unpaired two-tailed Student's *t*-test for the comparison of a given variable in two genotypes or two-way ANOVA followed by Bonferroni's *post hoc* comparisons for the comparison of more than two conditions. Statistical outliers were identified and excluded using the ROUT method with 1% the maximum desired false discovery rate developed by Prism, and the only statistical outliers of the current study were identified in the behavioral analysis and were marked in Figure 9 - Source Data. All statistical analysis was performed on data from ≥ 4 mouse brains per genotype per age group, and experiments were performed and repeated on different days, often by

independent investigators. Statistical significance is indicated as *$p < 0.05$, **$p < 0.01$, ***$p < 0.001$, ****$p < 0.0001$, and values not significantly different are often not noted.

Data availability

All data generated and analyzed in this study are included in the manuscript and the supporting information. Source Data files are provided for key findings shown in Figures 1, 3, 4, 8, and 9.

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Author Contributions

J.S. conceived and directed the project. L.M. (LRRK1) and Y.T. (LRRK2) generated the targeting vectors and performed ES cell experiments and Southern blotting. G.H. performed Northern analysis and RT-PCR followed by sequencing of floxed and deleted *LRRK1* and *LRRK2* mice, and J.K. performed Western analysis of deleted *LRRK1* and *LRRK2* mice as well as cDKO mice. J.K. and G.H. performed quantitative histological and EM analyses of the SNpc, including independent quantification of TH+ neurons in the SNpc and quantification of TH+ terminals in the striatum. P.C. performed the GFP reporter analysis of *DAT-Cre* mice in the SNpc and the LC. G.H. and A.S. performed independent behavioral analysis. J.K., G.H., A.S., P.C. analyzed the data and made the figures, and J.K., G.H., A.S., and J.S. wrote the paper.

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Figure Legends

Figure 1. Generation and characterization of floxed and deleted *LRRK1* and *LRRK2* alleles

(**A**) Schematic illustrations of human LRRK1 and LRRK2 proteins showing similar functional domains. $LRRK1_{2015}$ protein is derived from exons 2-34 (Ensembl Genome Database: ENSG00000154237). $LRRK2_{2527}$ protein is derived from exons 1-51 (ENSG00000188906). LRR: Leucine rich repeats; Roc: Ras-of-complex; COR: C-terminal of Roc; KIN: Kinase domain.

(**B**) Schematic illustrations of the gene structures of mouse *LRRK1* and *LRRK2*. The boxes in blue are exons that encode the LRRK1 and LRRK2 proteins, and the gray boxes represent the 5' and 3' UTRs. The exons are not drawn in scale. The start codon ATG is in exon 2 of *LRRK1* and exon 1 of *LRRK2*. The exons 27-29 of *LRRK1* and the promoter/exons 1-2 of *LRRK2* are flanked with *loxP* sites (green arrowheads).

(**C**) Targeting strategy for the generation of the targeted, floxed, and deleted *LRRK1* alleles. The red boxes represent the targeted exons 27-29, and the blue boxes represent the untargeted exons. The locations and sizes of the 5' and 3' external probes are shown. The targeting vector contains the 5' and 3' homologous regions (marked by dashed lines) and the middle region (from intron 26 to intron 29), which includes a *loxP* site (green arrowhead) in intron 26 (1,288 bp upstream of exon 27) and the *PGK-neo* selection cassette flanked by two *FRT* (FLP recognition target) sequences (gray circles) followed by another *loxP* site in intron 29 (1,023 bp downstream of exon 29). A negative selection cassette encoding diphtheria toxin fragment A (DTA) is also included in the targeting vector to reduce ES cells bearing randomly inserted targeting vectors. ES cells carrying the correctly targeted *LRRK1* allele were transfected with *pCAG-FLP* to remove the *PGK-neo* cassette and generate the floxed *LRRK1* allele. Floxed *LRRK1* mice were bred with *CMV-Cre* transgenic mice to generate germline deleted *LRRK1* mice. Detailed strategy for generating targeting vector and DNA sequence of floxed *LRRK1* allele can be found in Figure 1-figure supplement 1 and 2, respectively.

(**D**) Southern analysis of the targeted and floxed *LRRK1* alleles. Genomic DNA from ES cells or mouse tails was digested with *Hind*III and hybridized with the 5' or 3' external probe. For the 5' probe, the resulting 17.0 kb and 4.8 kb bands represent the wild-type (WT) and the targeted (T) or floxed (F) alleles, respectively. For the 3' probe, the resulting 17.0 kb and 12.2 kb bands represent the WT and the floxed alleles, respectively. Detailed Southern strategy can be found in Figure 1-figure supplement 3.

(**E**) Targeting strategy for the generation of the targeted, floxed, and deleted *LRRK2* alleles. The red boxes represent *LRRK2* exons 1 and 2, and the start codon ATG resides in exon 1. The locations and sizes of the 5' and 3' external probes are shown. The targeting vector contains the 5' and 3' homologous regions (marked by dashed lines) and the middle region (from the promoter to intron 2), which includes a *loxP* site (green arrowhead) upstream (1,768 bp) of the transcription initiation site and the *PGK-neo* selection cassette flanked by two *FRT* sequences (gray circles) and two *loxP* sites (green arrowheads) in intron 2 (878 bp downstream of exon 2). A negative selection cassette encoding DTA is also included in the targeting vector. Mice carrying the correctly targeted *LRRK2* allele were crossed with *Actin-FLP* deleter mice to generate floxed *LRRK2* mice, which were bred with *CMV-Cre* transgenic mice to generate germline deleted *LRRK2* mice. Detailed strategy for generating targeting vector and DNA sequence of floxed *LRRK2* allele can be found in Figure 1-figure supplement 4 and 5, respectively.

(**F**) Southern analysis of the targeted and floxed *LRRK2* alleles. Genomic DNA from ES cells or mouse tails was digested with *Nhe*I and hybridized with the 5' or 3' external probe. For the 5' probe, the resulting 11.5 kb and 3.6 kb bands represent the wild-type (WT) and the targeted (T) or floxed (F) alleles, respectively. For the 3' probe, the resulting 11.5 kb and 5.2 kb bands represent the WT and the floxed alleles, respectively. Detailed Southern strategy can be found in Figure 1-figure supplement 6.

(**G**) Northern analysis of poly(A)+ RNA prepared from the lung of mice carrying homozygous germline deleted (Δ/Δ) *LRRK1* alleles derived from the floxed *LRRK1* alleles using the cDNA probe of exons 2-3 (*left*) and exons 27-29 (*right*). Using the upstream exons 2-3 probe, the *LRRK1* transcripts in wild-type mice are the expected size of ~7.4 kb, whereas the detected *LRRK1* transcripts in *LRRK1* Δ/Δ mice are

truncated, consistent with the deletion of exons 27-29 (625 bp), which results in a frameshift, and are expressed at lower levels, likely due to nonsense mediated degradation of the truncated *LRRK1* mRNA. Using a probe specific for exons 27-29, there is no *LRRK1* transcript in *LRRK1* Δ/Δ mice, as expected. Both blots were hybridized with a *GAPDH* probe as loading controls. Detailed Northern strategy and full-size blots are included in Figure 1-figure supplement 7 and 8, respectively. Extensive RT-PCR analysis of *LRRK1* transcripts in Δ/Δ mice are shown in Figure 1-figure supplement 9.

(**H**) Northern analysis of total RNA prepared from the neocortex of mice carrying homozygous germline *LRRK2* deleted (Δ/Δ) alleles using the cDNA probe of exons 1-5 shows the absence of *LRRK2* transcripts. The blot was hybridized with a *GAPDH* probe as a loading control. The full-size blot is included in Figure 1-figure supplement 10. RT-PCR analysis of *LRRK2* transcripts in Δ/Δ mice are shown in Figure 1-figure supplement 11

(**I**) *Left*: Western analysis of wild-type (+/+) and homozygous *LRRK1* Δ/Δ brains show the absence of LRRK1 protein. *Right*: Western analysis of the neocortex of wild-type (+/+) and homozygous *LRRK2* Δ/Δ mice shows the absence of LRRK2 protein. Vinculin was used as a loading control. Full-size blots can be found in Figure 1-Source Data 1.

Figure 2. Generation and characterization of DA neuron-specific *LRRK* cDKO mice

(**A**) Immunostaining of GFP and/or TH in the SNpc of *DAT-Cre/+; Rosa26-CAG-LSL-ZsGreen1/+* mice at 2 months of age. Cre recombinase is expressed under the control of the *DAT* endogenous promoter and removes the floxed "stop" cassette, resulting in the expression of EGFP under the control of the ubiquitous *CAG* promoter.

(**B**) Quantification of GFP+/TH+ and TH+ cells shows that 99% of TH+ DA neurons (722 ± 46 TH+ cells) in the SNpc are also GFP+ (713 ± 46 cells), indicating that *DAT-Cre* mediated recombination occurs in essentially all TH+ DA neurons. N = 3 mice, 3 comparable sections per hemisphere, 320 μm apart.

(**C**) Similar body weight between *LRRK* cDKO mice and littermate controls at all ages examined ($F_{1,68}$ = 0.001310, p = 0.9712; 2M, 20M : p > 0.9999; 15M: p = 0.7857, 25M: p = 0.8084, two-way ANOVA with Bonferroni's *post hoc* multiple comparisons).

(**D**) Similar brain weight between *LRRK* cDKO and control mice ($F_{1,68}$ = 3.603, p = 0.0619; 2M: p = 0.3893; 15M: p > 0.9999; 20M: p > 0.9999; 25M: p = 0.3223, two-way ANOVA with Bonferroni's *post hoc* multiple comparisons).

(**E**) Western analysis of LRRK1 and LRRK2 proteins in the dissected cerebral cortex (CTX) and ventral midbrain (vMB) of *LRRK* cDKO and littermate controls at 2 months of age.

(**F**) Quantification shows significant decreases of LRRK1 and LRRK2 in the dissected ventral midbrain of *LRRK* cDKO mice (LRRK1, p = 0.0432; LRRK2, p = 0.0162, Student's *t*-test), compared to controls, but not in the dissected cortex of cDKO mice (LRRK1: p = 0.7648; LRRK2: p = 0.2325).

The number in the column indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. *p < 0.05. Scale bar: 100 μm.

Figure 3. Age-dependent loss of DA neurons in the SNpc of *LRRK* cDKO mice

(**A**) TH immunostaining shows TH+ DA neurons in the SNpc of *LRRK* cDKO and littermate controls at the age of 15, 20, and 24 months. Higher power views of the boxed areas show grossly normal DA neuron morphology in *LRRK* cDKO mice.

(**B**) Quantification of TH+ DA neurons in the SNpc reveals similar numbers of DA neurons in *LRRK* cDKO mice (10,000 ± 141) and littermate controls (10,077 ± 310, p > 0.9999) at 15 months of age. At 20 months of age, the number of DA neurons in the SNpc of *LRRK* cDKO mice (8,948 ± 273) is significantly reduced, compared to control mice (10,244 ± 220, $F_{1,46}$ = 16.59, p = 0.0002; p = 0.0041, two-way ANOVA with Bonferroni's *post hoc* multiple comparisons). By 24 months of age, the reduction of

DA neurons in the SNpc of *LRRK* cDKO mice (8,188 ± 452) relative to controls (9,675 ± 232, p = 0.0010) is greater, compared to 20 months of age. Raw quantification data are included in Figure 3-Source Data 1.

(**C**) Immunohistological analysis of TH and NeuN shows TH+ DA neurons (green) and NeuN+ neurons (red) in the SNpc of *LRRK* cDKO mice and controls at 24 months of age.

(**D**) Quantification of NeuN+ cells in the SNpc shows that the number of NeuN+ neurons in *LRRK* cDKO mice (17,923 ± 813) is significantly lower than that in control mice (21,907 ± 469, p = 0.0006, Student's *t*-test), indicating loss of neurons in the SNpc of *LRRK* cDKO mice. All TH+ cells are NeuN+. The number of TH+/NeuN+ cells in the SNpc of *LRRK* cDKO mice (10,500 ± 644) is also lower, compared to control mice (14,102 ± 310, p = 0.0001). There is no significant difference in the number of NeuN+/TH- neurons between littermate controls (7,804 ± 249) and cDKO mice (7,423 ± 344, p = 0.3747).

The number in the column indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. **p < 0.01, ***p < 0.001. Scale bar: 100 μm.

Figure 4. Increases in apoptotic DA neurons in the SNpc of *LRRK* cDKO mice

(**A**) Representative images of TH and active Caspase-3 immunostaining show TH+ DA neurons (green) and active Caspase-3+ apoptotic cells (red) in the SNpc of *LRRK* cDKO and control mice at the age of 24 months.

(**B**) Quantification of active Caspase-3+/TH+ cells shows significant increases of apoptotic DA neurons in the SNpc of *LRRK* cDKO mice (323 ± 38) at 24 months of age, relative to controls (157 ± 8, p = 0.0004, Student's *t*-test). Raw quantification data are included in Figure 4-Source Data 1.

The number in the column indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. ***p < 0.001. Scale bar: 100 μm.

Figure 5. Age-dependent loss of TH+ DA terminals in the striatum of *LRRK* cDKO mice

(**A**) Representative TH immunostaining images in the striatum of *LRRK* cDKO mice and littermate controls at the ages of 15 and 24 months.

(**B**) Quantification of TH immunoreactivity in the striatum of *LRRK* cDKO and control mice shows similar TH immunoreactivity at the age of 15 months ($p = 0.8766$, Student's *t*-test), but there is a significant decrease in TH immunoreactivity in the striatum of *LRRK* cDKO mice at 24 months of age (-19%, $p = 0.0215$), compared to controls.

The number in the column indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. $^*p < 0.05$. Scale bar: 1 mm.

Figure 6. Normal number of TH+ noradrenergic neurons in the LC of *LRRK* cDKO mice

(**A**) Representative images of TH+ noradrenergic neurons in the locus coeruleus (LC) of *LRRK* cDKO mice and littermate controls at 24 months of age.

(**B**) Quantification of TH+ cells shows similar number of TH+ noradrenergic neurons in the LC of *LRRK* cDKO mice (3,350 ± 99) and controls (3,418 ± 86, $p = 0.6110$, Student's *t*-test).

(**C**) *Top*: Immunostaining of TH and GFP in the LC of *DAT-Cre/+; Rosa26-CAG-LSL-ZsGreen1/+* mice at 2 months of age. There is no GFP+ (green) cell in the LC, indicating that *DAT-Cre* is not expressed in the LC. *Bottom*: Higher power views of the boxed areas.

The number in the column indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. Scale bar: 100 μm.

Figure 7. Unchanged number of electron-dense vacuoles in the SNpc of *LRRK* cDKO mice

(**A**, **B**) Representative EM images showing electron-dense vacuoles (arrowheads) in SNpc neurons of cDKO mice and littermate controls at the age of 25 months.

(**C-F**) Higher power views showing various electron-dense vacuoles, autolysosomes (single arrows), autophagosomes (double arrows), and lipid-containing vacuoles (asterisks) in SNpc neurons of littermate control (**C**, **D**) and cDKO (**E**, **F**) mice.

(**G**) *Left*: The average number of electron-dense vacuoles (>0.5 μm in diameter) in the SNpc neuronal profiles per mouse is not significantly different between *LRRK* cDKO mice and littermate controls at the age of 25 months (Control: 6.72 ± 0.43; cDKO: 6.99 ± 0.52, p = 0.6839, Student's *t*-test). *Right*: The total area of electron-dense vacuoles (>0.5 μm in diameter) in the SNpc neuronal profiles per mouse is similar between *LRRK* cDKO and littermate controls (Control: 4.43 ± 0.44 μm^2; cDKO: 4.60 ± 0.49 μm^2, p = 0.8048).

The value in parentheses indicates the number of mice (left) and neuron profiles (right) used in the quantification. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. Scale bar: 1 μm.

Figure 8. Elevated microgliosis in the SNpc of *LRRK* cDKO mice

(**A-C**) Representative images of Iba1+ microglia (red, marked by yellow arrowheads) and TH+ dopaminergic neurons (green) in the SNpc of *LRRK* DKO mice and controls at 15 (**A**), 20 (**B**), and 24 (**C**) months of age.

(**D**) Quantification of Iba1+ microglia shows significant increases in the number of Iba1+ microglia in the SNpc of *LRRK* cDKO mice, compared to control mice at the ages of 15 months (Control: 1737 ± 83, cDKO: 2541 ± 193; $F_{1,45} = 102.6$, $p < 0.0001$, p = 0.0017, two-way ANOVA with Bonferroni's *post hoc* multiple comparisons), 20 months (Control: 2426 ± 68, cDKO: 3639 ± 127, $p < 0.0001$), and 24 months (Control: 2640 ± 187, cDKO: 4089 ± 100, $p < 0.0001$). Raw quantification data are included in Figure 8-Source Data 1.

The number in the column indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. **p < 0.01, ****p < 0.0001. Scale bar: 100 μm.

Figure 9. Impairment of motor coordination in *LRRK* cDKO mice

(**A**) In the 10 mm beam walk test, compared to control mice, *LRRK* cDKO mice at 10 months of age exhibit markedly more hindlimb slips (Control: 2.0 ± 0.3; cDKO: 4.4 ± 0.5; p = 0.0005, Student's *t*-test) and longer traversal time (Control: 5.8 ± 0.4; cDKO: 7.3 ± 0.3; p = 0.0075). In the less challenging 20 mm beam walk test, there is no significant difference in the number of hindlimb slips (Control: 1.0 ± 0.2; cDKO: 1.5 ± 0.2; p = 0.0733) and traversal time (Control: 5.1 ± 0.4; cDKO: 5.2 ± 0.3; p = 0.9796) between *LRRK* cDKO and control mice.

(**B**) In the pole test, *LRRK* cDKO and control mice at 10 months of age display similar turning time (Control: 1.2 ± 0.1; cDKO: 1.5 ± 0.1; p = 0.1219) and descending time (Control: 4.7 ± 0.1; cDKO: 4.7 ± 0.2; p = 0.8620).

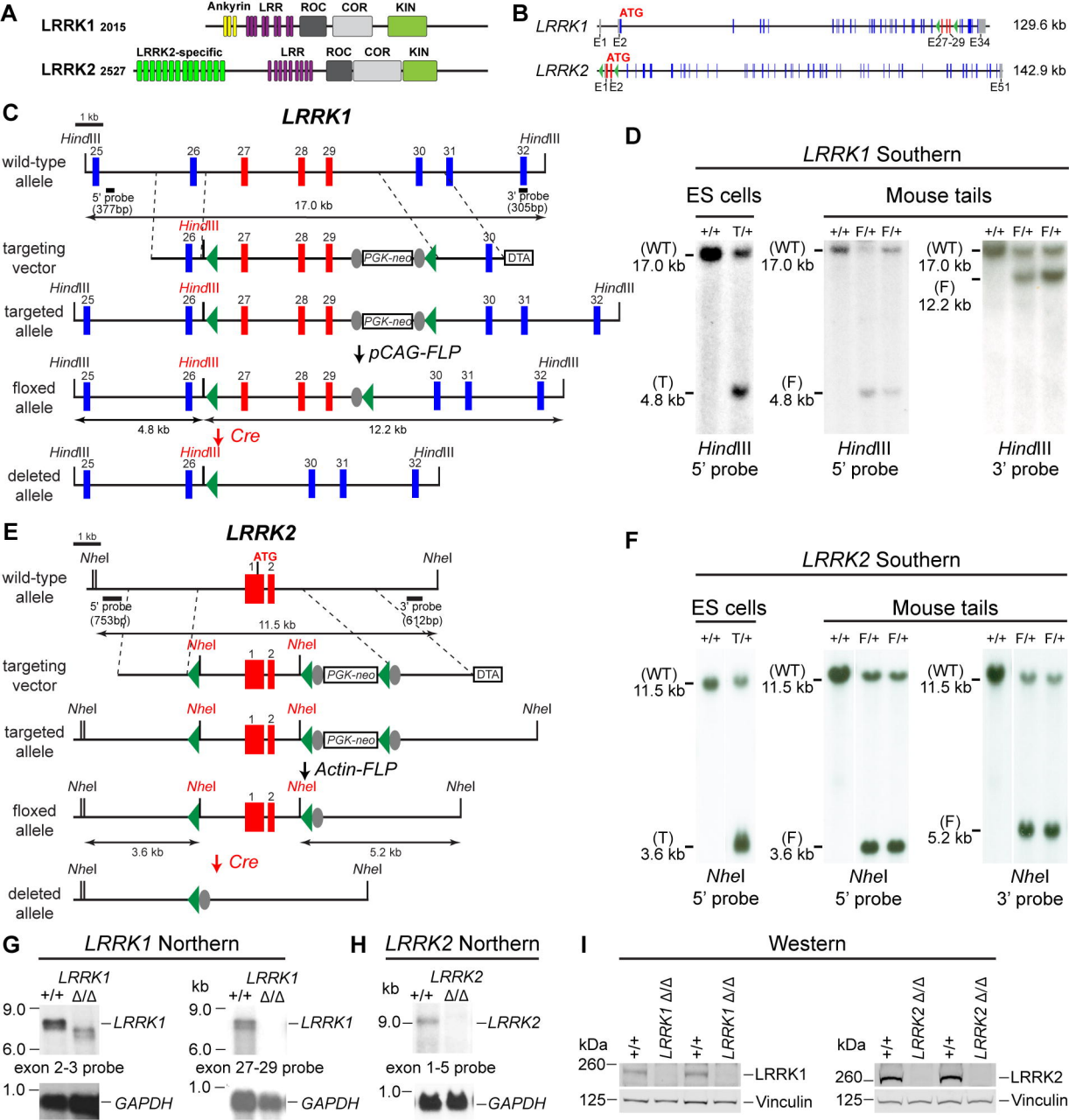
(**C**) *LRRK* cDKO (31.1 ± 1.3) and control (32.0 ± 1.5; p = 0.6410) mice at 10 months of age show similar body weight.

(**D**) *LRRK* cDKO mice and control mice at 22 months of age in the 10 mm beam walk test show similar hindlimb slips (Control: 12.0 ± 3.0; cDKO: 13.8 ± 3.3; p = 0.7022) and traversal time (Control: 15.3 ± 2.0; cDKO: 16.2 ± 2.8; p = 0.8139). In the 20 mm team walk test, there is also no difference in hindlimb slips (Control: 4.8 ± 1.6; cDKO: 4.1 ± 1.0; p = 0.7142) and traversal time (Control: 8.7 ± 1.6; cDKO: 6.4 ± 0.7; p = 0.2223) between *LRRK* cDKO and control mice.

(**E**) In the pole test, *LRRK* cDKO mice at 22 months of age exhibit similar turning time (Control: 1.7 ± 0.2; cDKO: 2.8 ± 0.7; p = 0.1184) and descending time (Control: 6.6 ± 0.7; cDKO: 5.4 ± 0.3; p = 0.1413) compared to control mice.

(**F**) *LRRK* cDKO mice (34.8 ± 1.5) have similar body weight as control mice (39.6 ± 2.4; p = 0.1194).

The number in parentheses indicates the number of mice used in the study. Red-filled and open circles represent data obtained from individual male and female mice, respectively. All data are expressed as mean ± SEM. **$p < 0.01$, ***$p < 0.001$. Raw behavior data are included in Figure 9-Source Data 1.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E51
142.9 kb
C
LRRK1
1 kb
HindIII
wild-type allele
5' probe (377bp)
17.0 kb
3' probe (305bp)
targeting vector
PGK-neo
DTA
targeted allele
pCAG-FLP
floxed allele
4.8 kb
12.2 kb
Cre
deleted allele
D
LRRK1 Southern
ES cells
Mouse tails
+/+
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F/+
(WT) 17.0 kb
(T) 4.8 kb
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HindIII 3' probe
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ATG
wild-type allele
5' probe (753bp)
11.5 kb
3' probe (612bp)
targeting vector
PGK-neo
DTA
targeted allele
Actin-FLP
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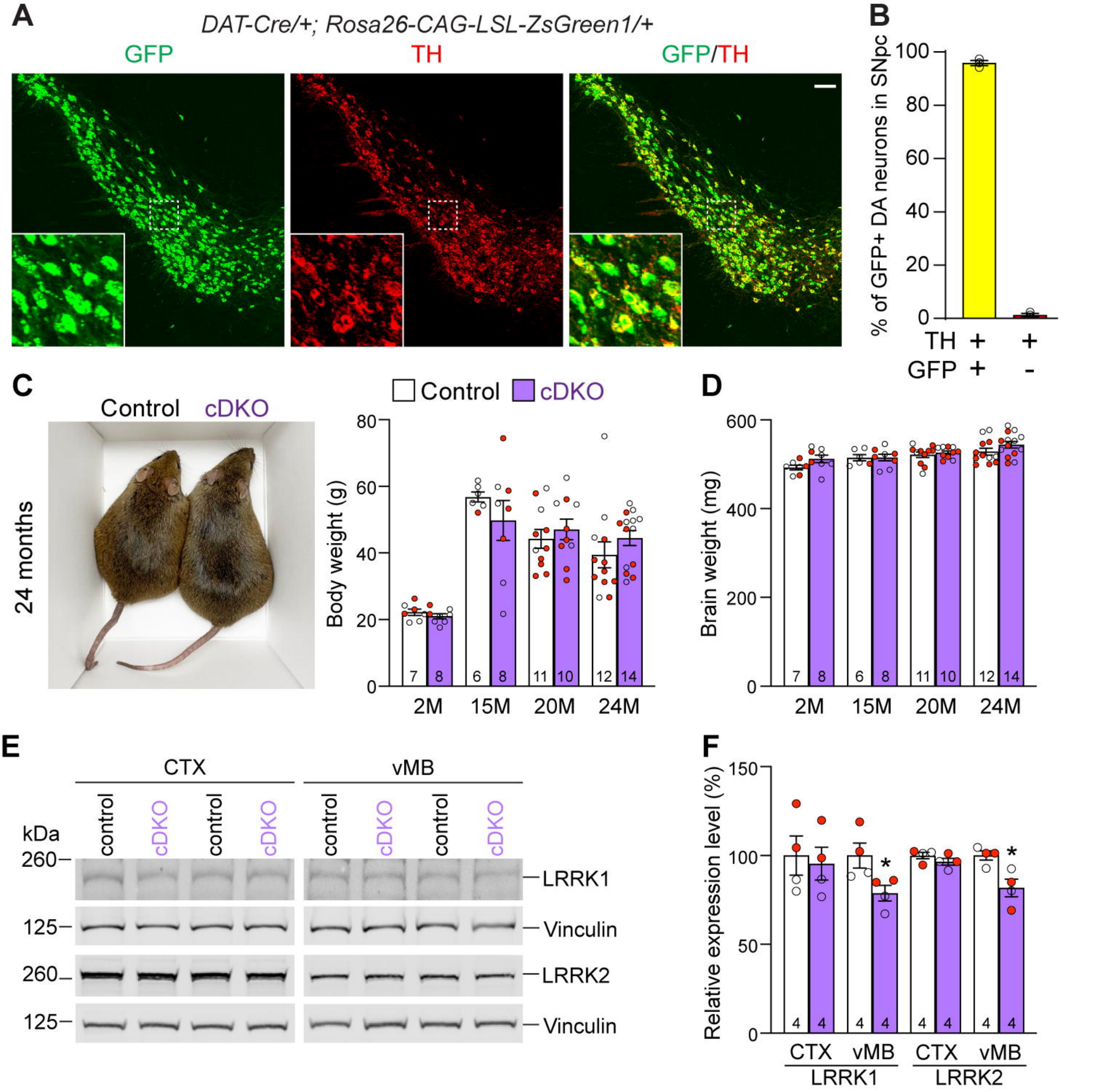
A
DAT-Cre/+; Rosa26-CAG-LSL-ZsGreen1/+
GFP
TH
GFP/TH
B
% of GFP+ DA neurons in SNpc
100
80
60
40
20
0
TH + +
GFP + -
C
Control
cDKO
24 months
Control
cDKO
Body weight (g)
80
60
40
20
0
7 8 6 8 11 10 12 14
2M 15M 20M 24M
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Brain weight (mg)
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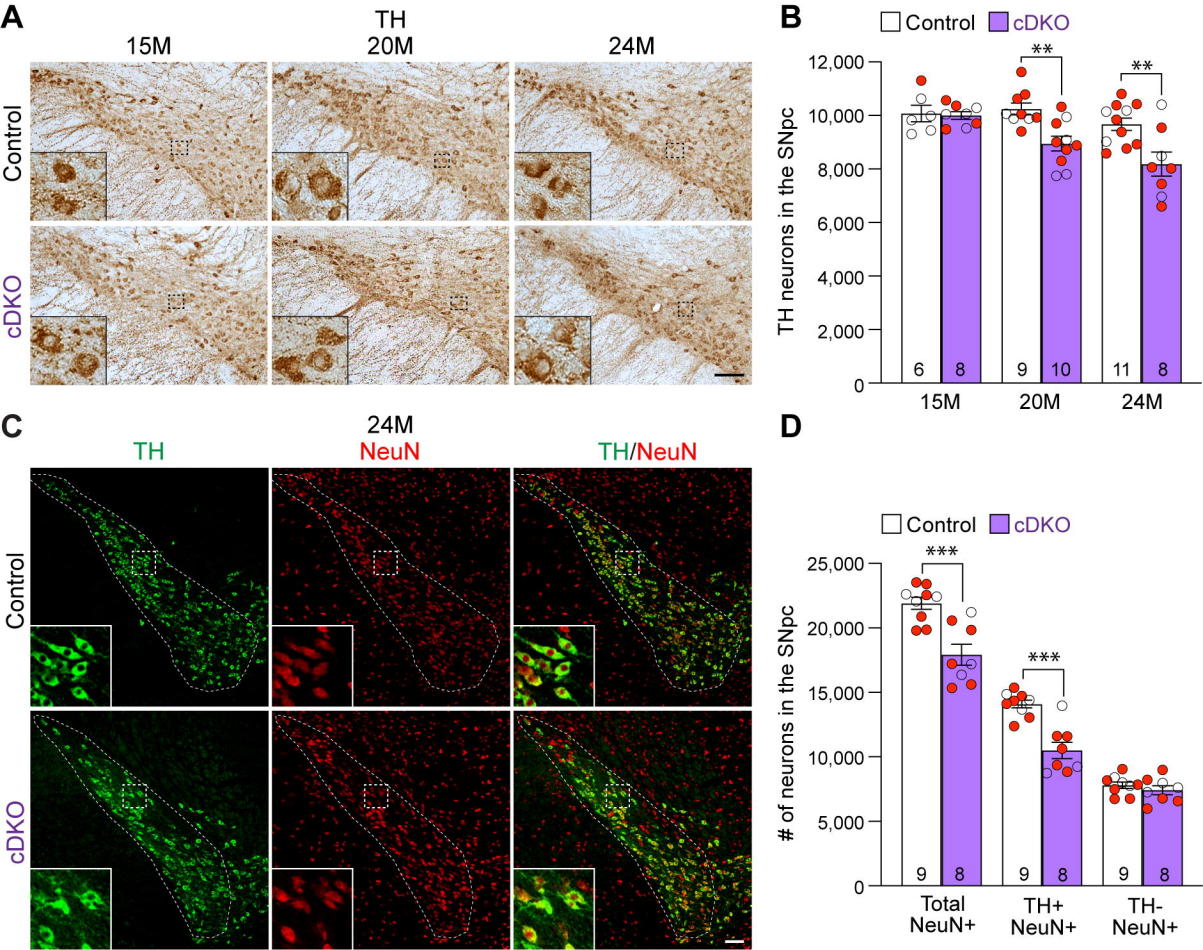
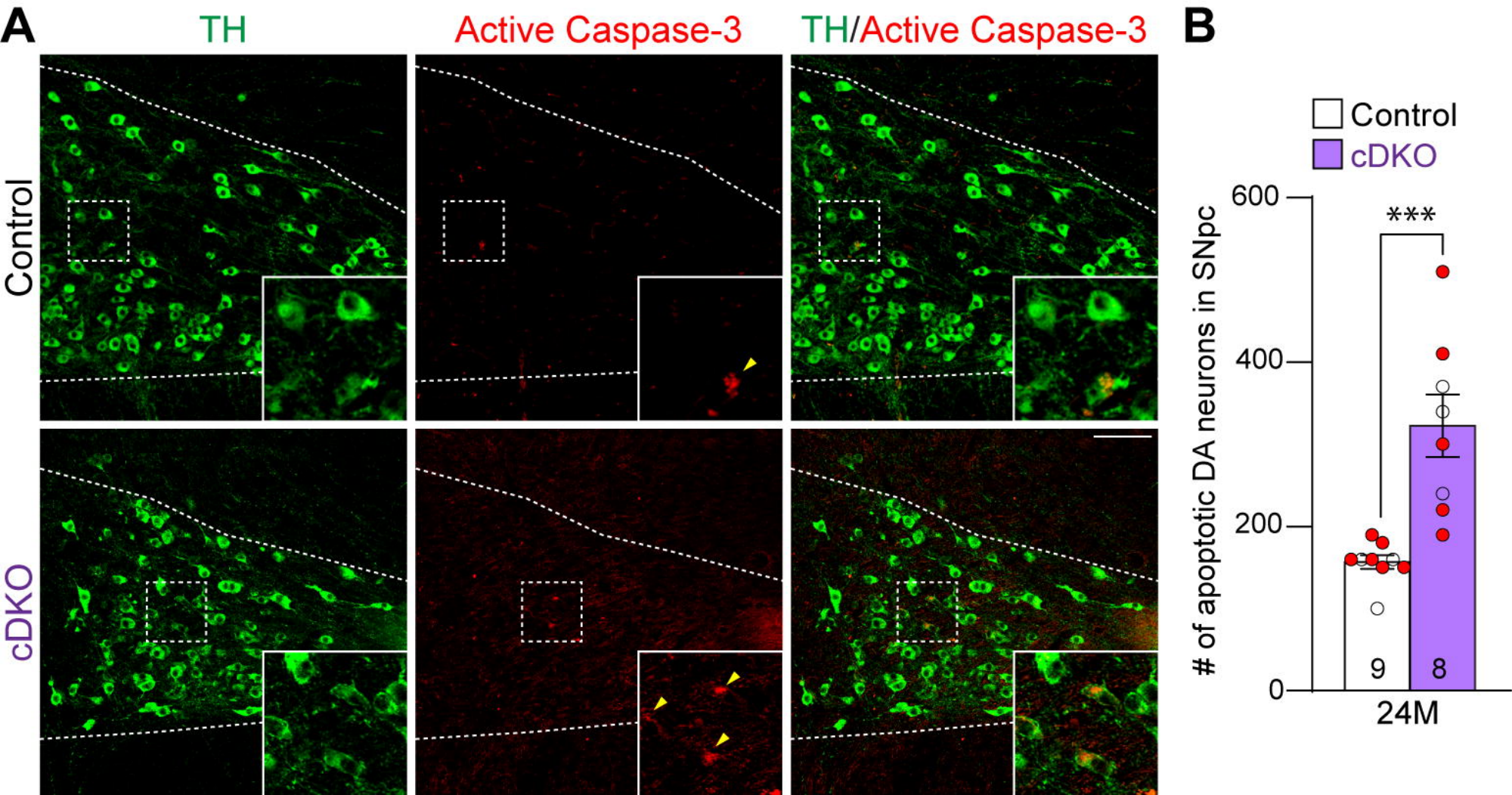
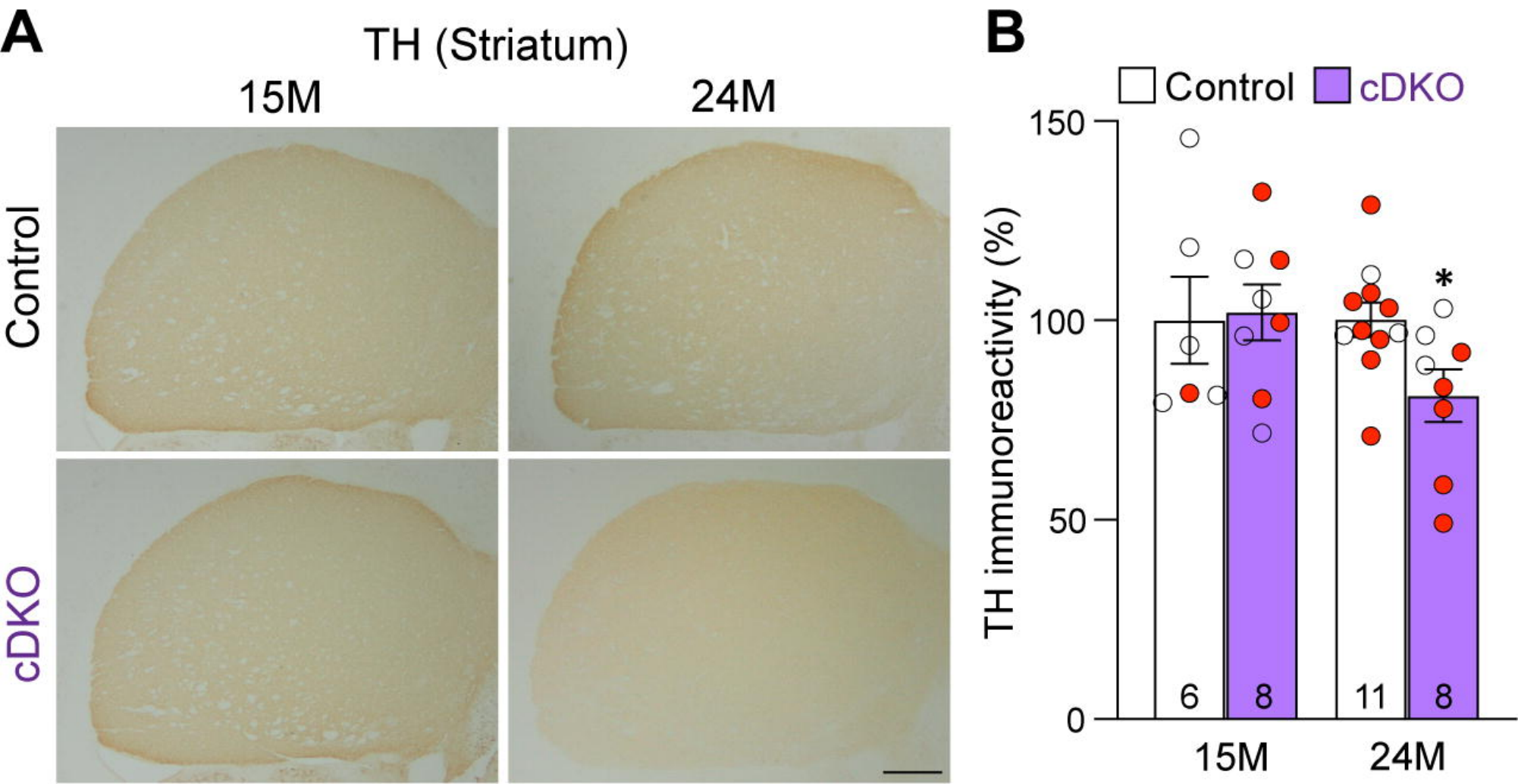
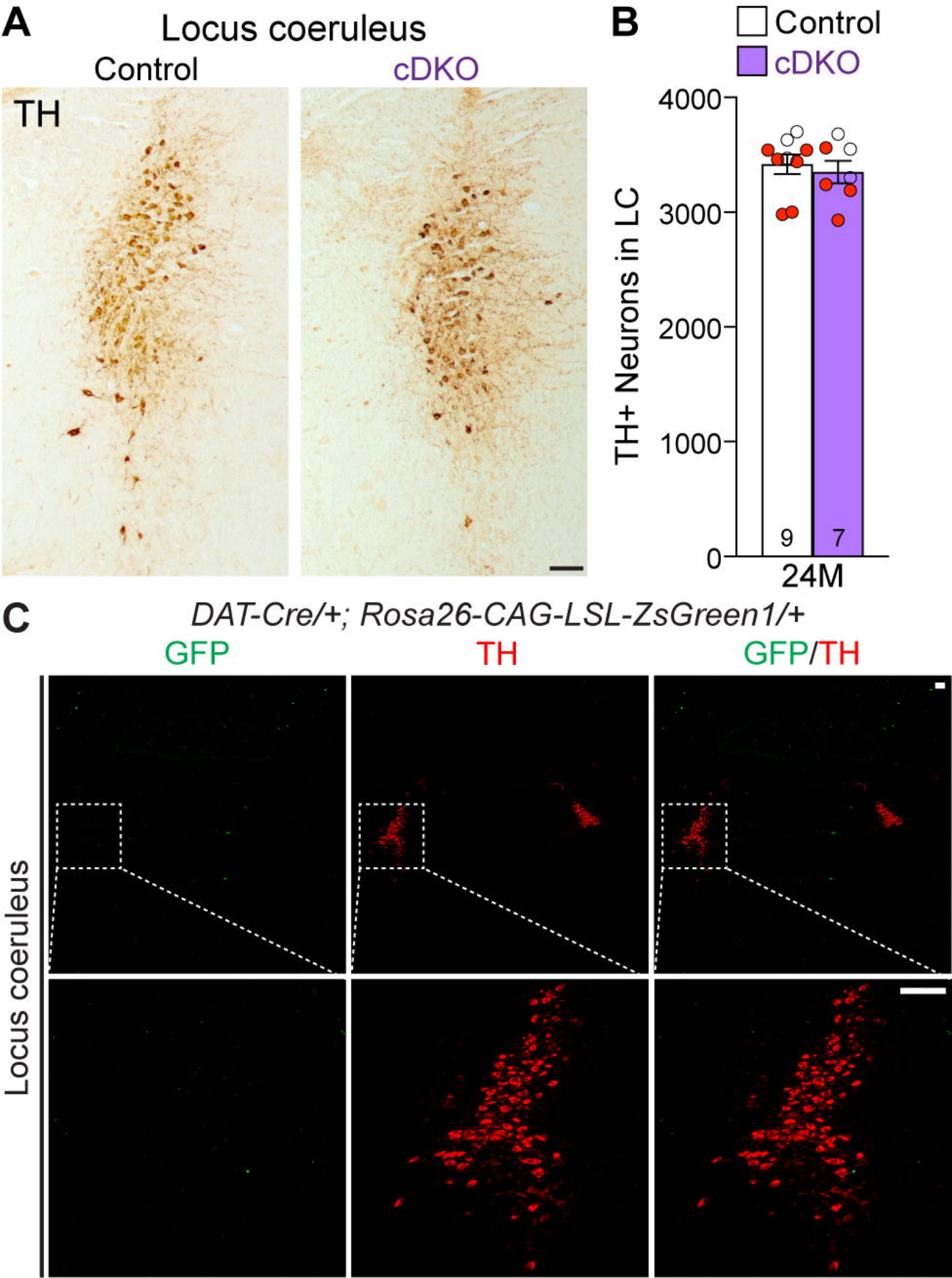
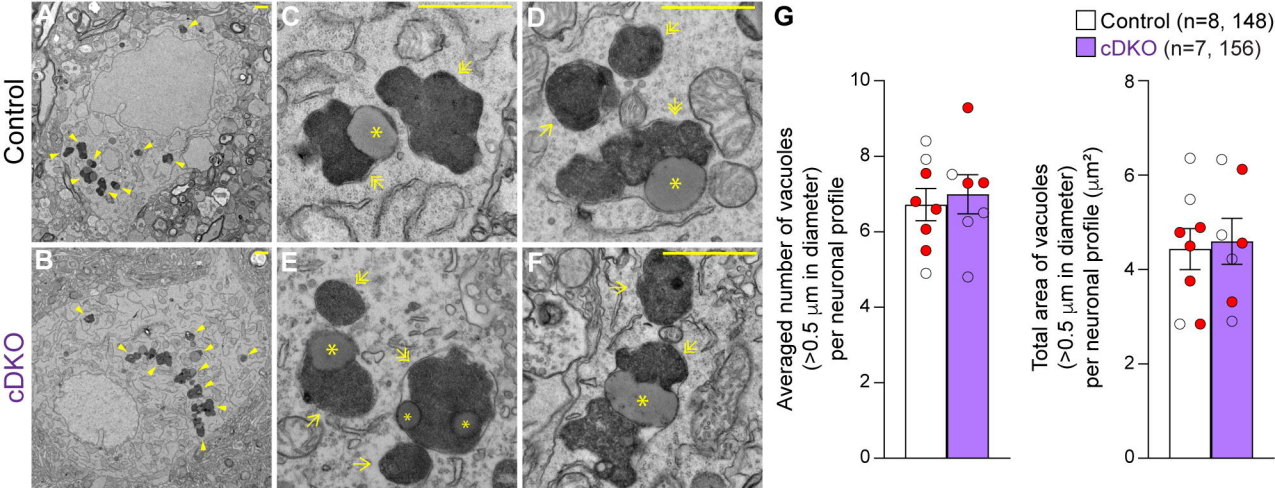
LRRK1
Vinculin
LRRK2
Vinculin
F
Relative expression level (%)
150
100
50
0
4 4 4 4 4 4 4 4
*
*
CTX vMB CTX vMB
LRRK1 LRRK2

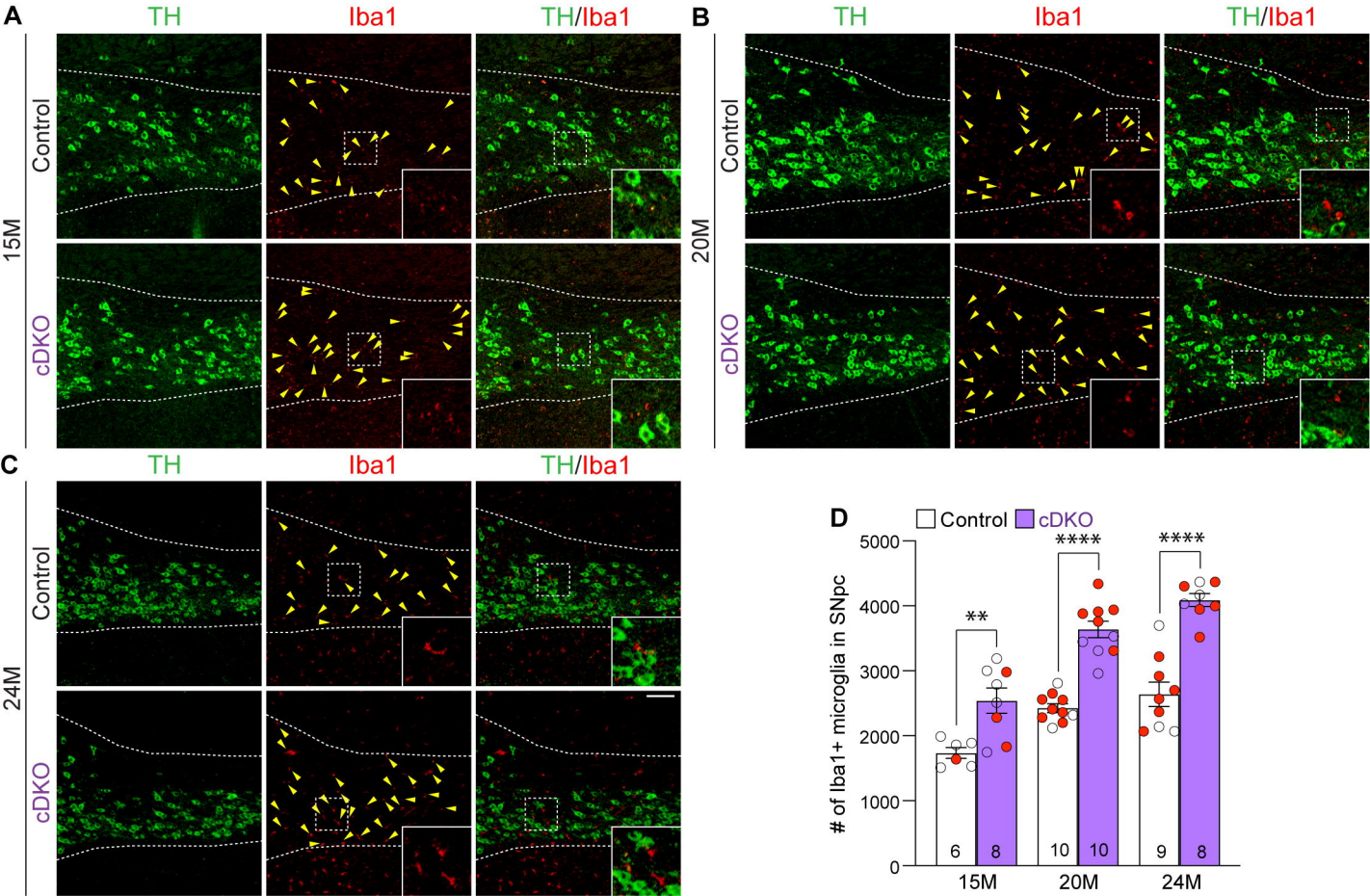

A
TH
15M
20M
24M
Control
cDKO
B
Control
cDKO
TH neurons in the SNpc
12,000
10,000
8,000
6,000
4,000
2,000
0
**
**
6
8
9
10
11
8
15M
20M
24M
C
24M
TH
NeuN
TH/NeuN
Control
cDKO
D
Control
cDKO
# of neurons in the SNpc
25,000
20,000
15,000
10,000
5,000
0
***
***
9
8
9
8
9
8
Total NeuN+
TH+ NeuN+
TH- NeuN+

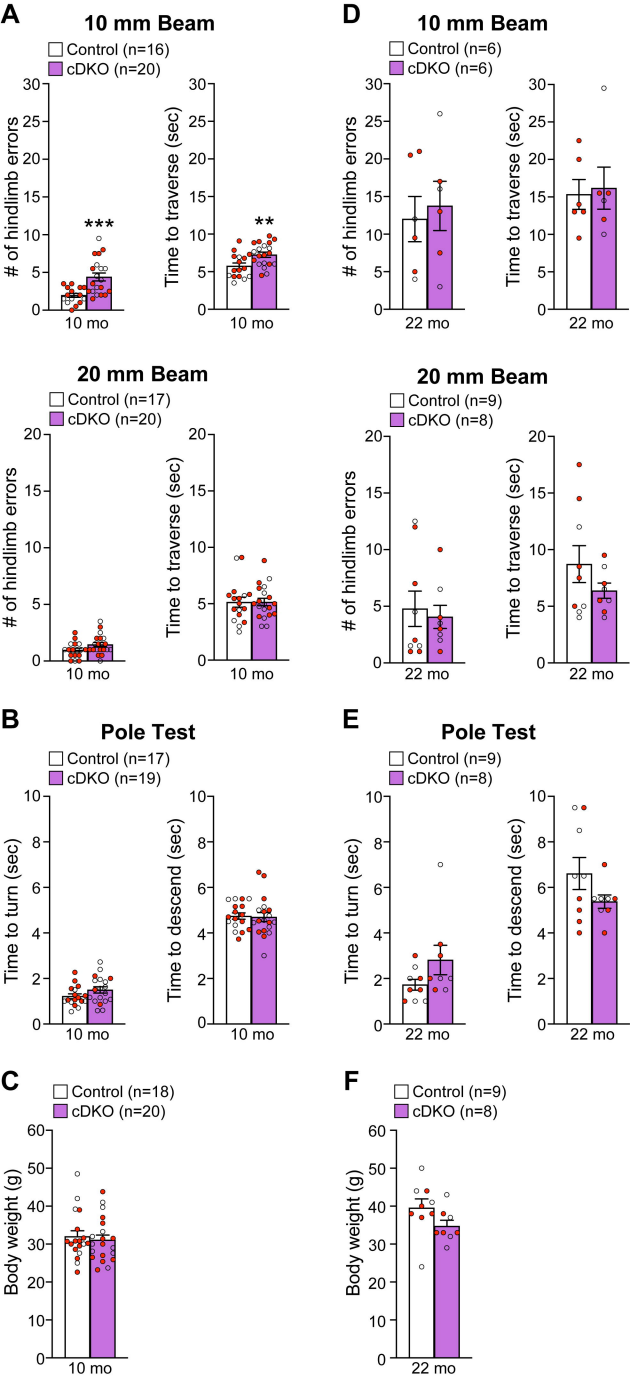

A
TH
Active Caspase-3
TH/Active Caspase-3
Control
cDKO
B
Control
cDKO
# of apoptotic DA neurons in SNpc
600
400
200
0
***
9
8
24M


A
TH (Striatum)
15M
24M
Control
cDKO
B
Control
cDKO
TH immunoreactivity (%)
150
100
50
0
*
6
8
11
8
15M
24M


A
Locus coeruleus
Control
cDKO
TH
B
Control
cDKO
TH+ Neurons in LC
4000
3000
2000
1000
0
9
7
24M
C
DAT-Cre/+; Rosa26-CAG-LSL-ZsGreen1/+
GFP
TH
GFP/TH
Locus coeruleus


Control
cDKO
A
B
C
D
E
F
G
Control (n=8, 148)
cDKO (n=7, 156)
Averaged number of vacuoles (>0.5 μm in diameter) per neuronal profile
Total area of vacuoles (>0.5 μm in diameter) per neuronal profile (μm²)
0
2
4
6
8
10


A
TH
Iba1
TH/Iba1
15M
Control
cDKO
B
20M
C
24M
D
Control
cDKO
# of Iba1+ microglia in SNpc
5000
4000
3000
2000
1000
0
**
****
****
6
8
10
10
9
8
15M
20M
24M


A
10 mm Beam
Control (n=16)
cDKO (n=20)
# of hindlimb errors
Time to traverse (sec)
***
**
10 mo
20 mm Beam
Control (n=17)
cDKO (n=20)
D
10 mm Beam
Control (n=6)
cDKO (n=6)
22 mo
20 mm Beam
Control (n=9)
cDKO (n=8)
B
Pole Test
Control (n=17)
cDKO (n=19)
Time to turn (sec)
Time to descend (sec)
E
Pole Test
Control (n=9)
cDKO (n=8)
C
Control (n=18)
cDKO (n=20)
Body weight (g)
F
Control (n=9)
cDKO (n=8)