An interface-based molecular generative framework for protein-protein interaction inhibitors

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Abstract

Protein-protein interactions (PPIs) play a crucial role in many biochemical processes and biological processes. Recently, many structure-based molecular generative models have been proposed. However, PPI sites and compounds targeting PPIs have distinguished physicochemical properties compared to traditional binding pockets and drugs, it is still a challenging task to generate compounds targeting PPIs by considering PPI complexes or interface hotspot residues. In this work, we propose a specifically molecular generative framework based on PPI interfaces, named GENiPPI. We evaluated the framework and found it can capture the implicit relationship between the PPI interface and the active molecules, and can generate novel compounds that target the PPI interface. Furthermore, the framework is able to generate diverse novel compounds with limited PPI interface inhibitors. The results show that PPI interface-based molecular generative model enriches structure-based molecular generative models and facilitates the design of inhibitors based on PPI structures.

Main

A vast network of genes is inter-linked through protein-protein interactions and is critical component of almost every biological process under physiological conditions, and can be ubiquitous in many living organisms and biological pathways[1-4]. Modulation of PPIs expands the drug target space and has enormous potential in drug discovery. In homo sapiens, it is estimated that the entire interactome comprises between 130,000 to 930,000 binary PPIs[5-7]. Despite significant efforts in developing modulators of PPIs, drug design and development for PPI targets, especially targeting the PPI interfaces, remains challenging[6,8-12]. Structure-based rational design serves as an important tool for the discovery of lead compounds in drug discovery[13-17]. Traditional drug targets and PPIs targets have different bio-chemical features (**Table 1**)[11,18-22], so conventional drugs and PPIs inhibitors have different physicochemical properties and drug-like properties (**Table 1**)[11,23-30]. Given their differences, developing molecular generative models of different paradigms are essential for the drug design of different target types[11,19,31].

Generative artificial intelligence(AI) is enable to model the distribution of training samples and generate novel samples[32,33]. In drug discovery, generative AI can accelerate drug discovery by generating novel molecules with desired properties. Numerous excellent review articles have summarized the development in this field[16,17,34-41]. Molecular generative models in drug design can be roughly divided into three categories: ligand-based molecular generative (LBMG) models, structure-based(pockets or binding sites) molecular generative models (SBMG), and fragment-based molecular generative models (FBMG), among which SBMG models have received much attention. [17,39,42]. Currently, some significant methods in structure-based molecular generative models can be found in [43-51], molecular generative models for PPI structures or PPI interfaces have been rarely reported in the literature. In recent years, classical machine learning[52-54], active learning[55], and deep learning-assisted methods [56] is better screening and design of PPIs inhibitors have been explored, and ligand-based molecular generative models of PPI inhibitors have been reported[57]. There are few

structure-based molecular generative models for PPIs targets have not been sufficiently

explored.

In this study, we developed a conditional molecular generative framework based on

protein-protein interaction interfaces (named GENiPPI) for the design of PPI interface

inhibitors. The framework was developed by a conditional Wasserstein generative

adversarial network (cWGAN) with convolutional neural networks (CNNs), integrated

graph attention networks (GATs) and long short-term memory (LSTM). It was designed

to efficiently capture the relationship between PPI interface with active/inactive

compounds to train conditional molecular generative models (**Fig. 1**). As demonstrated

by the conditional evaluation, GENiPPI is an effective architecture for capturing the

implicit relationships between the PPI interface and active compounds. In summary,

GENiPPI represents a potent deep learning framework for structure-based design of

PPI inhibitors.

Results

Generation of molecules targeting the PPI interface

Here, we introduce GENiPPI a modular deep learning framework for the design of

structure-based PPIs inhibitors (**Fig. 1**). GENiPPI is composed of four main modules:

GATs module [58-60] for representation learning of the protein complex interface, CNNs

module for molecular representation learning, cWGAN module [61] for conditional

molecular generation, and molecular captioning network module for SMILES strings

decoding (**as shown in Supplementary Figs.1, Figs.2, Figs.3 and Figs.4,**

respectively).

Our framework undergoes four steps to accomplish the generation of molecules

targeting the PPI interface. In the first step, we use GATs module designed for the

protein complex interface is to effectively capture the nuanced atomic-level interaction

characteristics inherent to the protein complex interface region. Next, we use CNN

module to provide a representation of the compound that contains voxel and electronic density information in three-dimension space [62]. And, the cGAN module is designed to generate compounds that target PPI interfaces using features from the protein complex interface region to regulate the inputs [63]. The cGAN module consists of a generator, a discriminator, and a conditional network. The generator takes a Gaussian random noise vector, and the protein complex interface features to generate a vector in the molecular embedding space, the discriminator evaluates whether the generated molecule embedding corresponds to a real or generated molecule, and the conditional network evaluates whether the molecule embedding matches the protein complex interface features. Finally, we use the molecular captioning network, which is made by a 3D CNNs and a recurrent LSTM [64] to decode molecular representations. The molecular representation generated by the generator is fed as input to the 3D convolutional network with the LSTM subsequently decoding the SMILES strings.

Conditional evaluation

First, we verified the validity of the conditions that act as conditional molecular generative models for the protein complex interfaces. For this purpose, we selected three PPI targets: MDM2(mouse double minute 2)/p53, Bcl-2(B-cell lymphoma 2)/Bax (Bcl-2 associated X), and BAZ2B(Bromodomain adjacent to zinc finger domain protein 2B)/H4(histone) for conditional evaluation. We generated 10,000 validated molecules each by the GENiPPI framework and calculated the drug-like metrics of the generated compounds: QED[27], QEPPI[28,29] and Fsp3(fraction of sp3 carbon atoms)[65]. We compared the QED, QEPPI, and Fsp3 distributions of the active compounds and the generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4 **(Fig. 2)**. As shown, the distributions of drug-like properties were similar between the generated compounds and the active compounds for the three PPI interface targets (**Fig.2a, Fig.2b, and Fig.2c**), while different distributions of drug-like properties were observed between the generated compounds based on different targets (**Fig.2d, Fig.2e, and Fig.2f**). The results demonstrate the effectiveness of the PPI interface in conditioning the molecular

generative model. The drug-like properties of the framework generated compounds

migrate relative to those of the compounds in the training dataset, indicating that the

framework captures the distributions of the training dataset and generates novel

compounds.

Model performance

In order to gain insight into the performance of the GENiPPI framework and to compare

it with other molecular generative models. We benchmarked our method by the MOSES

platform[66], a leading benchmark platform of molecular generation. We trained all

models on the full training dataset and randomly sampled 30,000 molecules. We

utilized models and hyperparameters provided by the MOSES platform, such as an

Adversarial Autoencoder(AAE)[67], character-level recurrent neural networks

(CharRNN)[68], Variational Autoencoder(VAE)[69], LatentGAN[70] and ORGAN[71]. To

validate the higher quality of the molecules generated by the conditioned model, we

compared them with molecules sampled from the GENiPPI framework and the

GENiPPI-noninterface framework without the conditioned module. We found that

molecules generated by the conditioned GENiPPI framework were superior to other

models in novelty and diversity.

As shown in **Table 2**, the GENiPPI framework has advantages in terms of uniqueness,

novelty, and diversity over the GENiPPI-noninterface. The GENiPPI framework

performs better overall in molecular generation. Compared with LatentGAN and

ORGAN, GENiPPI offers more benefits in terms of validity and diversity. While all

molecular generative models have their unique advantages in various performance

comparison. However, the molecular generative models tailored to specific tasks,

especially those based on PPI structure, have more advantages and inspirations from

the GENiPPI framework. To understand the similarities and differences between the

molecular distributions generated by the GENiPPI framework and other models. We

compared the distribution of molecular properties of the Testset, iPPI-DB inhibitor, and

the generated molecular datasets of AAE, CharRNN, VAE, LatentGAN,

GENiPPI(noninterface) and GENiPPI(**Supplementary Figs.4**). The generated

compounds have similar distributions of physicochemical properties to the compounds

from the training set. While most of the iPPI-DB inhibitors have QED values lower

than 0.5, most of them have QEPPI values higher than 0.5.

Chemical space exploration

To better obtain an estimate of the chemical space distribution of the model generated

molecules with the active compounds in the training datasets, we evaluated the

chemical drug-like space of the generated compounds by calculating t-distributed

random neighbourhood embedding (t-SNE) maps of MACCS fingerprint [72]. The t-SNE

is a dimensionality reduction method used for data points visualization in two or three-

dimensional space by mapping high-dimensional data to a lower dimension [73,74]. By

this method, similar compounds are clustered to visualize the high-dimensional

chemical space of the compounds. The distribution of the generated compounds and

active compounds in chemical drug-like space by t-SNE visualization (**Fig.3a, Fig.3b,**

and Fig.3c). The generated drug-like compounds not only share the chemical space

with the active compounds, but are also homogeneously mixed in the two-dimensional

space. The generated compounds show a similar chemical drug-like space to that of the

active compounds under 2D topological fingerprint. Adding the three dimensions of

compounds contributes to the design of promising drug-like compounds[30,75]. We

performed PMI shape analysis on the generated compounds and compared them with

drug-like compounds from DrugBank and iPPI-DB(**Fig.3d**). Many of the approved

compounds are either rod or disk shaped, and the generated drug-like compounds

library has a similar three-dimensional space. The PBF distribution of the library of

generated drug-like compounds is about 0~2 Å(**Fig.3e**). The results show that many of

the generated drug-like compounds are derived from relatively planar molecular

scaffolds. Moreover, we evaluated the ability of the model to generate target-specific

compounds by chemical space maps. To assess the overlapping of drug-like chemical

space, we utilized Tree MAP (TMAP)[76] to create the 2D projection(**Fig.3f**). Each point

corresponds to a compound and is colored by its target label. The dark and light colors

denote the generated compounds and the active compounds in the training set. These

results suggest that our GENiPPI model can generate compounds that are similar to the

active compounds in the training set and have novel structures. The results show that

the framework enriches and expands the chemical space of PPI-targeted drug-likeness

compounds.

Few-shot molecular generation

Because of the huge consumptive costs involved in data collection, only a small amount

of labeled biomedical data are usually available. The process of drug design and

optimization often faces the problem of low data[77]. The lack of effectively labeled data

tends to diminish the practical performance of most deep learning frameworks for drug

design. To perform generalized molecular generative design with limited labeled data,

it has been a trending topic in the few-shot generative community[78,79]. The GENiPPI

model was applied to generate a virtual compound library for the heat shock protein 90

- cell division cycle 37(Hsp90-Cdc37) interaction interface. By training the model on

the PPI structure of Hsp90-Cdc37 (PDB ID: 1US7) and seven disruptors, we sampled

500 valid compounds. The similarity between active disruptors and generated

compounds of Hsp90/Cdc37 in the chemical space was visualized by t-SNE projection

maps(**Fig.4a**). After few-shot learning, the generated compounds were mostly

distributed around the active disruptor, which demonstrated the effectiveness of few-

shot learning in navigating through the targeted chemical space. We performed

pharmacophore-based matching by considering DCZ3112(a novel triazine derivative

that disrupts Hsp90-Cdc37 interactions) as a reference molecule[80]. The top 5 generated

molecules have similar pharmacophore and shape features with DCZ3112(**Fig.4b**),

demonstrating the potential of the model to be applied to low-data PPI targets. **Fig. 4c**,

shows the hot spot amino acid residues at the PPI interface of the Hsp90-Cdc37 protein

complex(PDB ID: 1US7). We performed molecular docking for prediction of the

binding poses(**Fig.4e**) of DCZ3112 with the Hsp90-Cdc37 complex by the UCSF DOCK6.9 program[81]. The structure of the Hsp90-Cdc37 complex with DCZ3112 highlights the hydrogen bond interactions with amino acid residues:Arg32A, Glu33A, Ser36A , Ser115A, Gly118A, Gln119A, and Arg167B(**Fig.4e**), which may be the major energy contributors to protein-ligand interactions. The generated compounds were performed molecular docking together with DCZ3112, and selected compounds with reasonable binding modes and higher binding affinity by visual inspection for interaction pattern analysis. The generated compounds of GENiPPI not only obtained the better docking score than the active compounds, but also reproduced the interactions with the key residues of the PPI interface. The generated compounds also formed halogen bonds, salt bridges and π-cation interactions to improve the binding affinity of the generated compounds to the target interface(**Fig.4f**). In conclusion, by analyzing the interaction patterns between the generated compounds and the PPI interface, GENiPPI learned the implicit interaction rules between the active compounds and the PPI interface.

Discussion

We developed the GENiPPI framework, which combines protein-protein interaction (PPI) interfaces features and conditional molecular generative model to generate novel modulators for PPI interfaces. We validated the ability of GENiPPI framework to learn the implicit relationship between PPI interfaces and active molecules through conditional evaluation experiments. GENiPPI used GATs to extract key features of PPI interfaces, and searched molecules by conditional wGAN with specific constraints. We compared GENiPPI with various evaluation settings and benchmarks to demonstrate its practical potential.

Despite the promising results, our framework has some limitations that can be addressed in future work. We have not tested the model on a large number of receptor-ligand pairs of PPIs, which may affect its the generalization ability. The reason is that

the current PPI has relatively little data of drug-PPI target complexes than the traditional dataset of drug-target complexes. Furthermore, the current framework does not incorporate the 3D structural information of ligand-receptor interactions of PPIs. and There are still many ways to improve representation learning, balance training speed of molecular generative models and the diversity of generated molecules. Several potential directions could further improve GENiPPI: (1) collecting and cleaning higher quality data pairs for model development and testing; (2) fusing of molecular chemical language models and pre-trained models of protein-protein structural features to fine-tune the datasets of receptor-ligand of PPIs to enhance the model generalization, novelty and diversity of the generated compounds; (3) incorporating structural information of PPIs into fragment-based molecular generative models is also a promising direction; (4) change the architecture of the model or combining it with deep reinforcement learning to generate novel compounds with better binding affinity. Therefore, we will collect more data to further develop an enhanced version of GENiPPI by combining novel representation learning methods and deep generative approaches. In summary, the GENiPPI framework brings encouraging advances in PPI structure-based molecular generative tasks and presents a tool for rational drug design in finding modulators of macromolecule-macromolecule interactions.

Methods

Datasets

We first investigated PPI targets that were annotated with sufficient compound bioactivity data for training and evaluation of our model [82]. For this study, we selected 10 validated PPI drug targets that cover the binding interface(**Supplementary Table**). These targets are E3 ubiquitin-protein ligase Mdm2, apoptosis regulator Bcl-2, BAZ2B, apoptosis regulator Bcl-xL, BRD4 bromodomain 1 BRD4-1, CREB-binding protein (CREBBP), ephrin type-A receptor 4 (EphA4), induced myeloid leukemia cell differentiation protein Mcl-1, and menin. In addition, we randomly selected a subset of 250,000 compounds as additional inactive compounds from the ChEMBL [83] dataset

289 that was used as part of the training datasets. A detailed data preprocessing can be found

290 in Supplementary Note A.

Model strategy and training

Graph attention networks of protein-protein interaction interface

In this section, the representation learning of protein-protein complexes interfaces is

inspired by the work in protein docking model evaluation[60], which designed a double-

graph representation to capture the interface features and interactions of protein-protein

complexes (**Supplementary Figs.1**). The extracted interface region is constructed as

two graphs (G^1 and G^2) for representing the interfacial information and the residues

involved in the two proteins participating in the interaction. A graph G can be defined

as $G = (V, E,$ and $A)$, where V is the set of nodes, and E is the set of edges between

them, and A is the adjacency matrix for mapping the association between the nodes of

the graph, which numerically denotes the connectivity of the graph. If the graph G has

N nodes, the dimension of the adjacency matrix A of the graph is N^*N, where $A_{ij} > 0$

if the i-th node is connected to the j-th node, and $A_{ij} = 0$ otherwise. The graph G^1

describes the coding of the atomic types of all residues in the interface region, and its

adjacency matrix A^1 describes the classification of interatomic bonding types for all

residues at the interface region, which only considers the covalent bonds between atoms

of interface residues within each subunit as edges. Therefore, it is defined as follows:

$$A_{ij}^1 = \begin{cases} 1 & if \text{ atom } i \text{ and atom } j \text{ are connected by a covalent bond or } if \ i{=}j \\ 0 & \text{otherwise} \end{cases}$$

The graph G^2 links both covalent bonds (thus including G^1) and non-covalent residue

interactions as edges. The adjacency matrix A^2 for G^2 describes both covalent bonds

and non-covalent interactions between atoms within the range of 10.0 Å to each other.

The non-covalent atom pairs are defined as those which are closer than 10.0 Å to each

other. It is defined as follows:

$$315 \quad A_{ij}^2 = \begin{cases} A_{ij}^1, & \text{if } i,j \in \text{receptor } or\ i,j \in \text{ligand} \\ e^{\frac{-(d_{ij}-\mu^2)}{\sigma}}, & \text{if } d_{ij} \leq 10\text{Å } and\ i \in \text{receptor and } j \in \text{ligand}; \\ & or\ if\ d_j \leq 10\ \text{Å } and\ j \in \text{receptor } and\ i \in \text{ligand} \\ 0, & otherwise \end{cases}$$

where d_{ij} represents the distance between the i-th and the j-th atoms of all atoms of all

residues in the interaction region. μ and σ are learnable parameter with initial values of

0.0 and 1.0, respectively. The formula $e^{-(d_{ij}-\mu)^2/\sigma}$ decays with increasing distance

between atoms.

The graph representation is more flexible and natural to encode interactive information

and adjacent(local) relationships. For the node features of the graph, we considered the

physicochemical properties of the atoms. We used the same features from the previous

work[60,84,85]. Then, the feature vector of the nodes is 23 in length and was embedded into

140 features by a one-layer fully connected (FC) network.

The constructed graphs are used as the input for GATs. The graph consists of adjacency

matrices A^1, A^2, node matrices N_{mn}^1, N_{pq}^2, and the node features, $x^{in} =$

$\{x_1^{in}, x_2^{in}, \cdots, x_N^{in}\}$ and $x \in \mathbb{R}^F$, where F is the dimensionality of the node features. For

the input graph of x^{in}, the pure graph attention coefficients are defined as follows,

which represent the relative importance between the i-th and j-th nodes:

$$332 \qquad e_{ij} = x_i^T E x_j' + x_j^T E x_i',$$

333 where x_i' and x_j' are the transformed feature representations defined by $x_i' = W x_i^{in}$ and

334 $x_j' = W x_j^{in}.W$, $E \in \mathbb{R}^{F \times F}$ are learnable matrices in the GATs. e_{ij} and e_{ji} become

335 identical to satisfy the symmetrical property of the graph by adding $x_i^T E x_j^T$ and $x_i^T E x_i'$.

336 The coefficient will only be computed for i and j where $A_{ij} > 0$.

338 The attention coefficients will also be calculated for the elements in the adjacency

matrix. For the elements (i, j), they are defined in the following form:

$$a_{ij} = \frac{\exp(e_{ij})}{\sum_{j \in N_i} \exp(e_{ij})} A_{ij},$$

where a_{ij} represents the normalized attention coefficient between the i-th and j-th node

pairs, while e_{ij} is the computed symmetric graph attention coefficient. N_i denotes the

set of neighbors for the i-th node, which includes the interacting node j with $A_{ij} > 0$.

The purpose here is to define attention by considering both the physical structure A_{ij}

and the normalized attention coefficient e_{ij} of the interactions simultaneously.

Based on the attention mechanism, the new node features of each node are updated in

consideration of its neighboring nodes, which is a linear combination of the neighboring

node features and the final attention coefficient a_{ij} :

$$x_i'' = \sum_{j \in N_i} a_{ij} x_j',$$

Making the use of the GATs mechanism described previously, we applied four layers

of GATs to process the node embedding information of the neighboring nodes and

output the updated node embedding. For two adjacency matrices A^1 and A^2, we use a

shared GAT. the initial input to the network is the atomic feature. Working with two

matrices A^1 and A^2, we have $x_1 = GAT(x^{in}, A^1)$ and $x_2 = GAT(x^{in}, A^2)$. In order to

focus only on the intermolecular interactions at the interface of the input protein-protein

complex, we obtain the final node embedding by subtracting the embeddings of the two

graphs. By subtracting the updated embedding x_1 from x_2, we can capture aggregated

information on intermolecular interactions from only the other nodes in the protein-

protein complex interface. The output node feature is therefore defined as:

$$x^{out} = x^2 - x^1,$$

After that, the updated x^{out} became x^{in} to iteratively increase the information through

the three following GATs layers. After the four GATs layers updated the node

embeddings, the node embedding of the entire graph was summed up as the overall

intermolecular interaction representation of the protein-protein complex:

$$x_{graph} = \sum_{k \in G} x_k.$$

Finally, the FC layers were applied to the x_{graph} to obtain a [4,4,4] vector as features

of the protein-protein interface.

Molecular representation

For each SMILES string, a 3D conformer is generated using RDKit [86] and optimized

using the default settings of the MMFF94 force field. The molecular structure

information is then extracted into a 35Å grid centered at the geometric center of the

molecule using the HTMD package[87]. The atoms of the molecule are discretized into a

1 Å cubic grid, and eight channels are considered to compute voxelized information.

Finally, the electronic density of the molecules 9th channel is calculated using the

original molecule method in Multiwfn(**Supplementary Figs.2**) [88].

Conditional Wasserstein generative adverserial networks

The generator takes a conditional vector and a noise vector sampled from a Gaussian

distribution as inputs. The PPI interface features([1,4,4,4], vector shape)   are

concatenated with a noise vector of size [9, 4, 4, 4] and input to a 4-layer transposed

convolutional neural network (CNNs) with 256, 512, 1024, and 1024 filters,

respectively. The first three layers downsample the array size using concatenation

convolution (s=2). For all convolutions, we use a kernel size of 4, and the Leaky ReLU

is used as an activation function after convolution. BatchNorm3d is applied between

convolution and activation operations to normalize the values of each channel of each

sample.

The discriminator consists of a 4-layer sequential convolutional neural network (CNNs)

with 256, 512, 1024, and 1024 filters, respectively. The first three layers downsample

the array size using concatenation convolution (s=2). For all convolutions, we use a

kernel size of 4, and the Leaky ReLU (α=0.2) is used as an activation function after

convolution. InstanceNorm3d is applied between convolution and activation operations

to normalize the values of each channel of each sample.

The physical and spatial features of the compounds are derived from the molecular

representation learning module, and the PPI interface features are obtained from the

GATs module of the protein complex interface. They are used to estimate the matching

probability between molecules and PPI interface features(**Supplementary Figs.3**).

Molecular captioning network

In this section, we will describe how to decode the generated molecular representation

into a SMILES strings. Our work is inspired by shape-based molecular generation[89,90],

which designs a combination network of convolutional neural networks (CNNs) and

Long Short-Term Memory (LSTM) [64] to generate SMILES strings. In brief, the

molecular captioning network consists of a 3D CNNs and a recurrent LSTM. The

molecular representation generated by the generator is fed as input to the 3D CNNs,

and the output of the 3D CNN is fed into the LSTM to decode the SMILES strings

(**Supplementary Figs.4**).

Model training

The conditional generative adversarial network is trained with Wasserstein loss. The

loss functions for the generator $\left(G_{(0(z,c))}\right)$ and discriminator $\left(D_0(x)\right)$ are：

$$L_{x_0} = E_{iy_{xx}}[-D_y(x)] + E_{z_{xx},iy_{yx}}[D_{yy_{yx}}(G_{zy}(z,c))] + \lambda E_{iy_1}[(\parallel \nabla_{zx}D_y(\hat{x}) \parallel_z - 1)^2],$$
$$I_{x_{xx}} = E_{z_{xx},ixx_{yx}}\left[-D_z\left(G_{zy}(z,c)\right) - \alpha lo\,g\big(f_u(G_u(z,c),c)\big)\right]$$

418 where x and c are molecular representations and PPI interface features, respectively,

419 sampled from the true data distribution p_{real}, z is a random noise vector sampled from

420 a Gaussian distribution (p_z), and f_0 is a function that evaluates the probability that a

421 PPI interface feature corresponds to a molecular representation. λ and α terms are

422 regularization parameters, both empirically set to 10. λ term weighs the effect of the

423 gradient penalty on discriminator loss. α term weighs the effect of the effect of f_0 on

424 the loss of the generator.

426 The model was trained for 50,000 calendar hours with a batch size of 8 (65 steps per

427 calendar hour). The discriminators were updated after each step, while the generators

428 were updated every 30 steps. The network was trained using the RMSprop optimizer

429 with a learning rate of $1 \times 10\text{-}4$ for the generator and discriminator. during training, we

430 monitored the similarity between real and generated molecular representations using

431 Fréchet distances. The weights of the conditional networks were pre-trained on a binary

432 cross-entropy loss and frozen during GAN training. Training was performed on a single

433 NVIDIA A40 GPU, and all neural networks were built and trained using Pytorch 1.7.1

434 [91] and Tensorflow 2.5 [92].

436 **Molecular generation**

437 After the model has been trained, the embedding information of the protein-protein

438 complex interface is used to guide the model to generate novel molecules from the

439 latent space. The maximum sampling strategy was used in the LSTM, meaning that the

440 SMILES strings are generated by selecting the next token based on the highest

441 prediction probability[89].

443 **Evaluation settings**

444 **Conditional evaluation metrics**

445 In this study, the key is to evaluate the effectiveness of the proposed framework of

446 protein-protein interaction interface-based conditional molecular generation. Therefore,

447 we sampled the same number of valid molecules for the three PPI targets. Then we

448 calculated the QED values and Fsp3 by RDKit[86] and calculated the QEPPI values by

449 the QEPPI package (https://github.com/ohuelab/QEPPI) for the generated compounds

450 and others，and plotted the density distribution for comparing the differences of drug-

451 likeness.

MOSES evaluation metrics

454 To evaluate the performance of our proposed conditional molecule generation

455 framework, we used the evaluation metrics of validity, uniqueness, novelty and

456 diversity provided by the MOSES platform[66], which are defined as follows:

457 Validity: Molecules defined as valid in the generated molecules.

$$458 \quad \text{Validity} = \frac{N_{\text{valid}}}{N_{\text{generalated}}}$$

459 Uniqueness: The proportion of unique molecules found among the generated valid

460 molecules.

$$461 \quad \text{Uniqueness} = \frac{N_{\text{unique}}}{N_{\text{valid}}}$$

462 Novelty: The generated molecules are not to be covered in the training set.

$$463 \quad \text{Novelty} = \frac{N_{\text{novel}}}{N_{\text{unique}}}$$

464 FCD(Fréchet ChemNet Distance)：To detect whether the generated molecules are

465 diverse and whether they have chemical and biological properties that are similar with

466 the real molecules[93].

Molecular shape

To evaluate the shape space of molecules, we used two widely adopted molecular

descriptors to represent the three dimensions of molecular structure: principal moment

of inertia (PMI)[94] and the best-fit plane (PBF)[95]. The PMI descriptor classifies the

geometric shape of molecules into the degree of rod-shaped (linear shape, such as

acetylene), disk-shaped (planar shape, such as benzene), and sphere (spherical shape,

such as adamantane). The normalized PMI ratios (NPRs) are plotted in two-

dimensional triangle and then used to compare the shape space covered by different sets

of molecules, evaluating and visualizing the diversity of the molecular shape associated

with a given set of molecules[30]. PBF is a three-dimensional descriptor that represents

the deviation of a molecule from a plane. The PBF descriptor is the mean distance of

each heavy atom from the best-fit plane passing via all heavy atoms[95].

Tree MAP

To explore and explain the chemical space by unsupervised visualization of high-

dimensional data[76], we calculated MinHash fingerprint[96] vectors for active compounds

and generated compounds. Then tmap[76] and faerun[97] were utilized to construct two-

dimensional projections of Tree MAP (TMAP).

Protocol for few-shot generation

Targeting the Hsp90-Cdc37 PPI interface is recognized as an important option for

cancer therapy. The crystal structure of the Hsp90-Cdc37 protein complex (PDB ID:

1US7) is available for molecular docking[98]. In addition, known Hsp90-Cdc37 PPI

disruptors were collected for training of few-shot generative. They are DCZ3112,

Celastrol , FW-04-804, Sulforaphane, Withaferin A, Platycodin D, Kongensin A [99].

OpenPharmacophore(https://github.com/uibcdf/OpenPharmacophore) was utilized to

create pharmacophore models and virtual screening. The protein structures were

processed by using UCSF Chimera[100], the program DOCK6.9 was used for semiflexible

docking[81], and PyMOL[101] was used to create the figures. A detailed docking protocol

can be found in Supplementary Note B.

Data Availability

The datasets are available at Github (https://github.com/AspirinCode/GENiPPI). The

data implementation will be provided upon acceptance of the manuscript for publication.

Code Availability

All the codes are freely available at Github (https://github.com/AspirinCode/GENiPPI).

The code implementation will be provided upon acceptance of the manuscript for

publication.

Acknowledgments

Ethics declarations

The authors declare no competing interests.

Competing Interests Statement

The authors have declared no competing interests.

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Figure. legends

Fig. 1. Generation of molecules targeting PPI. 3D structural information of the protein-protein complex interface is represented as a graph. Feature representation of the interface region is captured by using a graph attention neural networks. The representation of the voxel and electron density of the compound is encoded by a 3D convolutional neural networks (CNNs). A conditional Wasserstein generative adversarial networks is trained to generate molecular embeddings with interface features as conditions. Generator: takes interface features and random noise vectors to generate molecular embeddings for the input features. Discriminator: calculates the probability that a molecule is from a real or a fake molecule. Condition: controls or regulates the generation of molecules constrained by a specific protein-protein interface. Finally, a long short-term memory (LSTM) networks parse SMILES strings from molecular representation.

Fig. 2: Results of conditional evaluation. (a) The QED, QEPPI and Fsp3 distribution of active compounds and compounds generated by the GENiPPI framework for MDM2/p53; (b) The QED, QEPPI and Fsp3 distribution of active compounds and compounds generated by the GENiPPI framework for Bcl-2/Bax; (c) The QED, QEPPI and Fsp3 distribution of active compounds and compounds generated by the GENiPPI framework for BAZ2B/H4; (d) The QED distribution of generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4; (e) The QEPPI distribution of generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4; (f) The Fsp3 distribution of generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4;

Fig. 3: Chemical space exploration. (a) The t-SNE visualization of active compounds and generated compounds for MDM2/p53; (b) The t-SNE visualization of active compounds and generated compounds for Bcl-2/Bax; (c) The t-SNE visualization of active compounds and generated compounds for BAZ2B/H4; (d) The PMI ternary density plots of generated compounds, small molecule drugs of DrugBank, and iPPI-DB inhibitors. Top left: propyne, bottom: benzene, and the top right: adamantane; (e) The molecular three-dimensionality distribution of the generated molecules was visualized with NPR descriptors and PBF descriptors. (f) TMAP visualization of active compounds and generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4.

Fig. 4: Few shot molecular generation analysis. (a) The t-SNE visualization of the distribution of active compounds and generated compounds for Hsp90/Cdc37; (b) Comparison of the pharmacophore of the generated molecules with the reference molecule(DCZ3112); (c) PPI interface region(in green) of the Hsp90(in palecyan)/Cdc37(in lightpink) complex; (d) The complex structure of DCZ3112(in green) and Hsp90(in palecyan)-CDC37(in lightpink) modeled by molecular docking (PDB ID: 1US7); (e) The binding poses of generated compounds(in green) and Hsp90(in palecyan)-CDC37(in lightpink) modeled by molecular docking (PDB ID: 1US7). Hydrogen bonds are displayed as blue dotted lines. π-cation Interactions are displayed as orange dotted lines.

Figure 1

Targeting protein-protein interaction interface

Fig. 1. Generation of molecules targeting the PPI interface. 3D structural information of the protein-protein complex interface is represented as a graph. Feature representation of the interface region is captured by using a graph attention neural networks. The representation of the voxel and electron density of the compound is encoded by a 3D convolutional neural networks (CNNs). A conditional Wasserstein generative adversarial networks is trained to generate molecular embeddings with interface features as conditions. Generator: takes interface features and random noise vectors to generate molecular embeddings for the input features. Discriminator: calculates the probability that a molecule is from a real or a fake molecule. Condition: controls or regulates the generation of molecules constrained by a specific protein-protein interface.

Figure 2

Fig. 2. Results of conditional evaluation. (a) The QED, QEPPI and Fsp3 distribution of active compounds and compounds generated by the GENiPPI framework for MDM2/p53; **(b)** The QED, QEPPI and Fsp3 distribution of active compounds and compounds generated by the GENiPPI framework for Bcl-2/Bax; **(c)** The QED, QEPPI and Fsp3 distribution of active compounds and compounds generated by the GENiPPI framework for BAZ2B/H4; **(d)** The QED distribution of generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4; **(e)** The QEPPI distribution of generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4; **(f)** The Fsp3 distribution of generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4.

Figure 3

Fig. 3. Chemical space exploration. (a) The t-SNE visualization of active compounds and generated compounds for MDM2/p53; (b) The t-SNE visualization of active compounds and generated compounds for Bcl-2/Bax; (c) The t-SNE visualization of active compounds and generated compounds for BAZ2B/H4; (d) The PMI ternary density plots of generated compounds, small molecule drugs of DrugBank, and iPPI-DB inhibitors. Top left: propyne, bottom: benzene, and the top right: adamantane; (e) The molecular three-dimensionality distribution of the generated molecules was visualized with NPR descriptors and PBF descriptors. (f) TMAP visualization of active compounds and generated compounds for MDM2/p53, Bcl-2/Bax and BAZ2B/H4.

Figure 4

Fig. 4. Few shot molecular generation analysis. (a)The t-SNE visualization of the distribution of active compounds and generated compounds for Hsp90/Cdc37; (b) Comparison of the pharmacophore of the generated molecules with the reference molecule(DCZ3112); (c)PPI interface region(in green) of the Hsp90(in palecyan)/Cdc37(in lightpink) complex; (d)The complex structure of DCZ3112(in green) and Hsp90(in palecyan)-CDC37(in lightpink) modeled by molecular docking (PDB ID: 1US7); (e)The binding poses of generated compounds(in green) and Hsp90(in palecyan)-CDC37(in lightpink) modeled by molecular docking (PDB ID: 1US7). Hydrogen bonds are displayed as blue dotted lines. π-cation Interactions are displayed as orange dotted lines.

Table. 1.

Table 1. Comparisons between PPI interfaces and binding sites

PPI interfaces	Binding sites
Target properties	
Large surface area (1000-6000 $Å^2$)	Small surface (300-1000 $Å^2$) Hydrophobic
Preference for Trp (W), Tyr (Y), and Arg (R) as PPI hotspot residues; subpockets	Large volume (~260 $Å^3$)
Shallow, flat, flexible	Pocket, cliff
Hydrophobic, featureless, undruggability	Diverse properties
Chemical space	
MW$\geq$ 400	MW$\leq$ 500
LogP $\geq$ 4	LogP$\leq$ 500
HBA $\geq$ 4	HBA $\leq$ 10
number of rings: $\geq$ 4	HBD $\leq$ 5
Ro4 Morelli's rules	Lipinski's Rule of 5 (Ro5)
Quantitative estimate of drug-likeness scores	
QEPPI	QED

Table. 2.

Table 2. Valid, unique, novelty and FCD of sampling SMILES after training. We sampled 30,000 SMILES each time.

Model	valid	Unique@1k	Unique@10k	novelty	FCD	
					Test	TestSF
AAE	0.881	1.000	0.995	0.995	8.573	9.117
CharRNN	0.985	0.999	0.988	0.994	8.7564	8.952
VAE	0.834	1.000	0.996	0.994	7.703	8.141
LatentGAN	0.724	1.000	0.999	0.998	7.595	8.160
ORGAN	0.609	0.996	0.994	0.999	39.800	41.158
GENiPPI(noninterface)	0.999	0.997	0.975	0.997	7.653	8.132
GENiPPI	0.999	0.998	0.977	0.998	7.450	7.884