Mutant mice lacking alternatively spliced p53 isoforms unveil *Ackr4* as a male-specific prognostic factor in Myc-driven B-cell lymphomas

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Abstract

The gene encoding p53, a major tumor suppressor protein, encodes several alternative isoforms of elusive biological significance. Here we show that mice lacking the *Trp53* Alternatively Spliced (AS) exon, thereby expressing the canonical p53 protein but not isoforms with the AS C-terminus, have unexpectedly lost a male-specific protection against Myc-induced B-cell lymphomas. Lymphomagenesis was delayed in $p53^{+/+}$ Eμ-Myc males compared to $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males, but also compared to $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc females. Pre-tumoral splenocytes from $p53^{+/+}$ Eμ-Myc males exhibited a higher expression of *Ackr4,* encoding an atypical chemokine receptor with tumor suppressive effects. We show that *Ackr4* is a p53 target gene, but that its p53-mediated transactivation is inhibited by estrogens. We identify *Ackr4* as a male-specific factor of good prognosis, relevant for murine Eμ-Myc-induced and human Burkitt lymphomas. These data demonstrate the functional relevance of alternatively spliced p53 isoforms and reveal sex disparities in Myc-driven B-cell lymphomagenesis.

Keywords

p53, Ackr4, Myc, Burkitt lymphoma, sex disparity in cancer, p53 isoforms, alternative splicing

Introduction

TP53, the human gene for tumor suppressor p53, encodes several isoforms owing to distinct promoters, alternative splicing and multiple translation initiation sites (Bourdon et al., 2005; Courtois et al., 2002; Flaman et al., 1996; Yin et al., 2002). p53 alternative isoforms can be abnormally expressed in cancer cells and some may regulate the canonical p53 protein (Anbarasan and Bourdon, 2019; Bourdon et al., 2005; Mondal et al., 2013; Senturk et al., 2014). However, aberrant RNA splicing is a common feature of cancer cells (Graubert et al., 2012; Martin et al., 2013; Pajares et al., 2007; Sette and Paronetto, 2022) and to which extent alternative splicing generates functionally relevant proteins is controversial (Abascal et al., 2015; Blencowe, 2017; Tress et al., 2017a, 2017b; Ule and Blencowe, 2019; Weatheritt et al., 2016). Thus, the biological importance of many p53 isoforms remains elusive.

Like its human *TP53* homolog, the murine *Trp53* gene encodes multiple isoforms differing in their N- or C-termini (Arai et al., 1986; Marcel et al., 2011). Mouse models to evaluate the role of p53 isoforms differing in their N-terminus revealed that Δ40-p53 overexpression leads to accelerated ageing (Maier et al., 2004; Steffens Reinhardt et al., 2020). However, the potential role of p53 isoforms with an alternative C-terminus was not analyzed *in vivo*. p53 isoforms with distinct C-termini result from the splicing of two mutually exclusive final exons: exon 11, encoding the canonical "α" C-terminal domain, and the Alternatively Spliced (AS) exon, encoding another C-terminus (Arai et al., 1986). In adult mice, isoforms with the canonical C-terminus are predominant in all tissues (Figure S1A). Two models ($p53^{\Delta 31}$ and $p53^{\Delta CTD}$), designed to study the consequences of a loss of the canonical p53 C-terminus, exhibited signs of increased p53 activity leading to a rapidly lethal anemia (Hamard et al., 2013; Simeonova et al., 2013). To determine the role of p53-AS isoforms *in vivo*, we created $p53^{\Delta AS}$, a mouse model with a specific deletion of the AS exon (Figure S1B). In mouse embryonic fibroblasts (MEFs), the *Trp53*$^{\Delta AS}$ allele prevented the expression of isoforms with the AS C-terminus whereas it did not affect RNA levels for p53 isoforms with the canonical C-terminus (Figure S1C). We previously used this model to show that p53-AS isoforms had no role in the anemia affecting $p53^{\Delta 31/\Delta 31}$ mice (Simeonova et al., 2013). However, a detailed phenotyping of $p53^{\Delta AS/\Delta AS}$ mice remained to be performed. The detailed phenotyping, presented here, yielded surprising information on lymphomagenesis.

Results

Stress responses in WT and p53$^{\Delta AS/\Delta AS}$ cells

Thymocytes undergo a p53-dependent apoptosis upon irradiation (Lowe et al., 1993). We analyzed thymocytes from irradiated wild-type (WT) and p53$^{\Delta AS/\Delta AS}$ mice. In WT thymocytes, isoforms with the AS C-terminus were 5 times less abundant than isoforms with the α C-terminus at the RNA level (Figure 1A), and in western blots the p53-AS protein appeared as a faint band running just ahead of, and often hard to separate from, the band specific for p53-α, the canonical full-length p53 (Figure 1B). In p53$^{\Delta AS/\Delta AS}$ thymocytes, mRNA levels for α isoforms were slightly decreased, if at all (Figure 1A), whereas p53-α protein levels appeared markedly decreased (Figure 1B), raising the possibility that p53-AS isoforms might contribute to p53-α abundance. Nevertheless, the transactivation of classical p53 target genes (Figure 1C) and apoptotic response (Figures 1D and S1D) were not significantly altered by the loss of AS isoforms. Likewise, no significant difference was observed between WT and p53$^{\Delta AS/\Delta AS}$ fibroblasts in assays for cell cycle control (Figures 1E and S1E), expression of well-known p53 target genes (Figure 1F and S1F-G), proliferation under hyperoxic conditions (Figure1G), or the growth of tumor xenografts (Figure 1H).

Lymphomagenesis in WT and p53$^{\Delta AS/\Delta AS}$ mice

We compared spontaneous tumor onset in WT and p53$^{\Delta AS/\Delta AS}$ littermates for over 2 years and observed no significant difference in tumor-free survival (Figure 1I). Because lymphoma is a common neoplasm in C57BL6/J WT mice (Brayton et al., 2012) and our mouse cohorts resulted from >10 generations of backcrosses with C57BL6/J mice, we searched for evidence of lymphoma in the lymph nodes and spleen, by macroscopic examination at autopsy and histological analyses. B-cell lymphomas were observed in about 30% of mice of either genotype (Figure 1J). In p53$^{+/+}$ mice, a higher incidence of B-cell lymphomas was observed in females, in agreement with previous observations (Brayton et al., 2012). By contrast, no obvious sex-specific bias was observed for B-cell lymphomas in p53$^{\Delta AS/\Delta AS}$ mice (Figure 1J), raising the possibility that the loss of p53-AS isoforms affected B-cell lymphomagenesis. However, the numbers of lymphoma-bearing mice were too small to be conclusive.

We next used Eμ-Myc transgenic mice, prone to highly penetrant B-cell lymphomas (Adams et al., 1985). $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice developed B-cell lymphomas (Figure S2A) with similar survival curves when sexes were not considered (Figure S2B). Importantly however, death was accelerated, and tumor lymph nodes were larger, in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males compared to their $p53^{+/+}$ Eμ-Myc male counterparts, whereas no difference in lymphomagenesis was noticeable between $p53^{\Delta AS/\Delta AS}$ Eμ-Myc and $p53^{+/+}$ Eμ-Myc female mice (Figure 2A-B). Our data (Figures 2A-B and S2C), together with the fact that B-cell lymphomas occur with a higher incidence in WT C57Bl6/J female mice (Brayton et al., 2012), suggested that $p53^{+/+}$ Eμ-Myc male mice are more refractory to B-cell lymphomas, and that p53-AS isoforms might confer this male-specific protection against lymphomagenesis.

Cause for accelerated lymphomagenesis in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males

We next aimed to determine the mechanisms underlying the accelerated lymphomagenesis in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males. Inactivating p53 mutations were not more frequent in tumors from $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males than in those from $p53^{+/+}$ Eμ-Myc males, ruling out additional mutations at the *Trp53* locus as potential causes for accelerated lymphomagenesis in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males (Figures S2D-E). We next analyzed a subset of tumors with no detectable *Trp53* mutation in males of both genotypes, and found that Myc was expressed at similar RNA and protein levels in all tumors (Figure 2C). No difference in p53-α mRNA levels was observed in tumors from both genotypes, although a decrease at the protein level was detected in most tumors from $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males (Figure 2D). Nevertheless, similar transcript levels for classical p53 target genes were observed in tumor cells of both genotypes (Figure 2E). To test whether a higher tumor volume in $p53^{\Delta AS/\Delta AS}$ Eμ-myc males might result from lower apoptosis and/or higher cell proliferation, we next analyzed tumors by immunohistochemistry with antibodies against cleaved caspase-3 or ki-67, respectively. Similar apoptotic and proliferation indexes were observed for both genotypes (Figure 2F-G). In sum, classical assays for p53 activity in tumors failed to account for differences between the two male genotypes.

The speed of lymphomagenesis in Eμ-Myc mice correlates with the extent of B-cell expansion in the first stages of B cell differentiation (Langdon et al., 1986) and p53 was proposed to control the pool of early B cells (Slatter et al., 2010). Therefore, we determined the levels of the early pre-B / immature B cells in 6 weeks-old mice, before any sign of tumor.

We analyzed the spleen, a preferential site of B-cell expansion (Langdon et al., 1986) with a relatively high AS/α isoform ratio (Figure S1A). Flow cytometry with a combination of markers was used to discriminate the pre-B, immature, transitional and mature B subpopulations. As expected (Langdon et al., 1986), we observed high numbers of pre-B and immature B cells in Eμ-Myc mice. In males, pre-B and immature B cells were more abundant in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc animals, while no difference was observed for transitional and mature B cells (Figures 3A and S3A). By contrast, in the spleen of $p53^{+/+}$ and $p53^{\Delta AS/\Delta AS}$ 6 weeks-old male mice without the Eμ-Myc transgene, most B cells were mature B cells (Figure S3B). In Eμ-Myc females, the numbers of pre-B and immature B cells were similar between genotypes, as were the numbers of transitional and mature B cells (Figure 3A). Interestingly, $p53^{+/+}$ Eμ-myc males, which develop lymphomas less rapidly, exhibited the lowest number of immature B cells (Figure 3A), suggesting a direct correlation between the level of immature B cell expansion and the speed of lymphomagenesis. Together, these data suggested that p53-AS isoforms may not be required to control the pool of early B cells under normal conditions, but that in an Eμ-Myc context they would limit the expansion of pre-tumor early B cells, specifically in males.

Transcriptomes from $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc male spleens

We next performed bulk RNA-seq and differential expression analyses comparing the spleens from 4-6 weeks-old $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males to spleens from age-matched $p53^{+/+}$ Eμ-Myc males. This revealed a limited number of significantly deregulated genes (Figure 3B), including 13 protein-coding genes and 11 pseudogenes (Figure 3C). Out of the 13 protein-coding genes, we focused on the 10 genes not encoding an immunoglobulin and analyzed the same samples by RT-qPCR (Figure 3D). For 6 of the 10 genes, expression levels were too low to be quantified (*Tcstv3*), or differences in expression were not statistically significant (*Masp2*, *Akr1c19*, *Cd300lh*, *Il5ra, Slc26a1*). Of note, *Il5ra* belonged to this group, although it is regulated by p53 (Zhu et al., 2022), which illustrates the difficulty to analyze subtle effects in our experiments. Taking this into account, we considered as potentially interesting the 4 remaining genes, exhibiting differences in mRNA levels with statistical significance ($p<0.05$) or borderline statistical significance ($p=0.057$): *Ackr4*, less expressed in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males, and *Fam132a*, *Mt2* and *Prss50*, with an increased expression in $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males (Figure 3D). Importantly, survival curves indicated that the Myc-induced lethality was delayed in $p53^{+/+}$ Eμ-Myc males compared to $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males, $p53^{+/+}$ Eμ-Myc

females and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc females (Figure S2C). Thus, we quantified transcripts for *Ackr4*, *Fam132a*, *Mt2* and *Prss50* in the spleen of 4-6 weeks-old p53$^{+/+}$ Eμ-Myc and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc females, then compared mRNA levels in p53$^{+/+}$ Eμ-Myc males versus the 3 other groups. Significantly higher expression of *Ackr4* and lower expression of *Mt2* were found in p53$^{+/+}$ Eμ-Myc males (Figure 3E).

Ackr4 encodes the atypical chemokine receptor 4, a decoy receptor promoting the degradation of chemokines that modulate cancer cell proliferation and metastasis (Chow and Luster, 2014; Marcuzzi et al., 2018; Müller et al., 2001). Our data suggested that *Ackr4* might be a gene transactivated by p53-α and/or p53-AS isoforms. Consistent with this, by extracting data from a transcriptome-wide study in MEFs (Younger et al., 2015) we found evidence for a p53-dependent transactivation of *Ackr4* in response to doxorubicin (Figure 3F). Furthermore, ChIP-Atlas, the database of chromatin immunoprecipitation experiments (Oki et al., 2018), indicated p53 binding to sequences within the intron 1 of *Ackr4* in doxorubicin-treated MEFs, and we identified a candidate p53 responsive element in this intron (Figure 3G). We next used luciferase assays to show that this p53 responsive element can be bound and regulated by both p53-α and p53-AS (Figures 3G-H and S3C). Together, these data show that *Ackr4* is indeed a p53 target gene, although RNAseq data indicated that it is expressed at much lower levels than classical p53 targets like *Cdkn1a* or *Mdm2* in the splenocytes of Eμ-Myc mice (Table S1). Furthermore, *Ackr4* was shown to be regulated by Foxl2 and estrogen signalling in ovarian cells (Georges et al., 2014) and 17-β estradiol was recently found to regulate *ACKR4* expression in meniscal cells from both sexes, albeit differentially (Knewtson et al., 2020). Accordingly, we observed, in both WT and p53$^{\Delta AS/\Delta AS}$ MEFs, that p53 activation with the Mdm2 antagonist Nutlin led to the transactivation of *Ackr4,* but that a concomitant treatment with 17-β estradiol markedly decreased, or completely abrogated, *Ackr4* transactivation (Figure 3I). By contrast, *Cdkn1a* was efficiently transactivated under both conditions in mutant and WT cells (Figure 3I). These data indicate that *Ackr4* is a p53 target gene whose p53-mediated transactivation can be inhibited by estrogens.

The *Mt2* gene, encoding the potentially oncogenic metallothionein-2 (Si and Lang, 2018), was less expressed in p53$^{+/+}$ Eμ-Myc male pre-tumoral splenocytes, which raised the possibility of its direct or indirect repression by p53, potentially through the binding of p53 or the DREAM complex at its promoter (Engeland, 2018; Peuget and Selivanova, 2021). However, ChIP-Atlas reported no binding of these proteins at the *Mt2* promoter. Alternatively, evidence that Myc may impact on *Mt2* expression was obtained previously (Qin et al., 2021),

and ChIP-Atlas reported Myc binding at the *Mt2* promoter in primary B-cells from lymph nodes of Eμ-Myc mice as well as Eμ-Myc-induced lymphoma cells (Figure S3D). This may suggest that the lower expression of *Mt2* in pre-tumoral splenic cells from $p53^{+/+}$ Eμ-Myc males (Figure 3E) might result from a subtle difference in Myc signalling.

Relevance of *ACKR4* expression in Burkitt lymphomas

Murine and human alternatively spliced p53 isoforms exhibit structural differences (Marcel et al., 2011) and the ChIP-Atlas database (Oki et al., 2018) does not report p53 binding to the human *ACKR4* intron 1. Nevertheless, we found that p53 activation in human cells also led to an increased *ACKR4* expression abrogated by 17-β estradiol, whereas 17-β estradiol had no significant effect on the p53-mediated transactivation of *CDKN1A* (Figure 4A). This led us to investigate the potential relevance of *ACKR4* expression in human B-cell lymphomas. We analyzed public databases of B-cell lymphoma patients with clinical and gene expression information. We first analyzed #GSE4475 (Hummel et al., 2006), a dataset of mature aggressive B-cell lymphomas previously used to define Burkitt lymphoma-like specific transcriptomes, comprising 159 patients (91 men, 68 women) with clinical follow-up. Overall, *ACKR4* gene expression was not significantly different between male and female patients (Figure 4B, left). However, average mRNA levels appeared higher in males when we considered the 30% patients of each sex with the highest *ACKR4* expression (Figure 4B, right). Strikingly, when we compared the survival of 30% patients with the highest ACKR4 mRNA levels to the survival of the 30% patients with the lowest ACKR4 mRNA levels, high *ACKR4* expression correlated with a better prognosis in men, but not in women (Figure 4C). By contrast, for *MT2A,* the human homolog of *Mt2*, differences in mRNA levels did not correlate with significant differences in survival for either sex (Figure S4A).

We also analyzed dataset #GSE181063 (Lacy et al., 2020), comprising mostly diffuse large B-cell lymphomas (DLBCL; 613 men, 536 women) and a few Burkitt lymphomas (65 men, 18 women). We found no difference in survival curves of DLBCL patients with low versus high *ACKR4* levels, neither in men nor in women. However, there was again an increased survival for Burkitt lymphoma male patients with high *ACKR4* expression, but not for women (Figure S4B). Finally, we analyzed #phs000235 (Morin et al., 2011), a Burkitt lymphoma-specific dataset (65 men, 37 women) comprising mostly patients diagnosed at 0-17 years of age, hence providing cohorts homogeneous for both tumor type and age of onset.

Again, *ACKR4* was expressed at higher levels in a subset of male patients (Figure 4D) and high *ACKR4* expression correlated with a better prognosis only in males (Figure 4E). We thus conclude that, as in Eμ-Myc mice, *ACKR4* is a male-specific positive prognostic factor in Burkitt lymphoma, the archetype of MYC-driven B-cell lymphomas.

Discussion

In this report we analyzed a mouse model with a specific deletion of the alternatively spliced (AS) exon of the *Trp53* gene. Despite a subtle phenotype, this model revealed that a male-specific protective effect against Eμ-Myc-induced B-cell lymphomas is lost in the absence of p53-AS isoforms. $p53^{\Delta AS/\Delta AS}$ males also appeared more prone to develop spontaneous lymphomas, suggesting that the sex-specific protective effect conferred by p53-AS isoforms might not be restricted to the Eμ-Myc model.

Our transcriptomic data from splenocytes of $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males disclosed very few differentially expressed genes, and highlighted *Ackr4* as a male-specific positive prognostic factor in Eμ-Myc-induced lymphomas. Mechanistically, we identified *Ackr4*, expressed at low levels in splenocytes, as a p53 target gene that may be transactivated by p53-α and/or p53-AS according to luciferase assays. Because the loss of p53-AS isoforms correlated with an apparent decrease in p53-α levels in both thymocytes and tumor lymph nodes, the decreased transactivation of *Ackr4* in splenocytes of $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males might result from the loss of p53-AS isoforms *per se*, but also possibly from a decrease in p53-α levels. Furthermore, we observed that 17-β estradiol can inhibit the p53-mediated transactivation of *Ackr4*. Together, our data suggest that *Ackr4* may be regulated by p53-α, p53-AS and estrogens, likely accounting for sex-specific and p53-status dependent differences in gene expression.

Our analyses reveal that in both mice and humans, *Ackr4/ACKR4* is a male-specific prognostic factor in Burkitt-like lymphomas. In fact, *Ackr4* most likely acts as a tumor suppressor of Myc-driven B-cell lymphomas because : 1) in lymph nodes, Ackr4 creates a gradient of the chemokine Ccl21, a ligand of the chemokine receptor Ccr7 (Bastow et al., 2021; Ulvmar et al., 2014); 2) the Ackr4-mediated sequestration of Ccl21 impairs the Ccr7 signalling cascade, which may lead to decreased Myc activity (Shi et al., 2015); 3) Ccr7 is also required for lymphoma cell lodging to secondary lymphoid organs, and Ccr7 deficiency was shown to delay Eμ-Myc-induced lymphomagenesis (Rehm et al., 2011). Thus, in $p53^{+/+}$ Eμ-Myc male pre-tumoral splenocytes, an increased *Ackr4* expression might delay lymphomagenesis in part by attenuating Myc signalling. Indeed, the observed lower expression of *Mt2*, known to be regulated by Myc (Qin et al., 2021), appears consistent with this hypothesis. Importantly, *Ackr4* was previously found to inhibit the growth and metastasis of breast, cervical, colorectal, hepatocellular and nasopharyngeal cancer cells (Feng et al.,

2009; Hou et al., 2013; Ju et al., 2019; Shi et al., 2015; Zhu et al., 2014), although no report mentioned any sex-specific bias for cancers occurring in both sexes. Our data provide evidence that sex-specific differences in *Ackr4* expression may have prognostic value. This suggests that measuring *ACKR4* gene expression in male patients with Burkitt lymphoma could be useful to identify the patients at higher risk, for whom specific therapeutic regimens might be required.

Interestingly, our data suggested that *Mt2* might be a male-specific negative prognostic factor in murine Eμ-Myc induced lymphomas, but *MT2A* expression levels had no prognostic value in human lymphomas. A possible explanation for this discrepancy is suggested by the fact that *Mt2* is regulated by Myc. A translocation leading to MYC overexpression drives oncogenesis in all Burkitt lymphomas, but half of them exhibit additional missense MYC mutations enhancing its tumorigenicity (Chakraborty et al., 2015). The transcriptional program of a WT and two lymphoma-associated Myc mutants were recently compared, and we noticed that one of the mutants led to an alteration in *Mt2* expression (Mahani et al., 2021), which would abrogate any potential prognostic value.

Finally, a polymorphism in the *MDM2* gene promoter provided evidence that sex-specific hormones may affect p53 signalling and tumorigenesis (Bond and Levine, 2007). More recently, a higher frequency of *TP53* mutations in men, together with an increased vulnerability to alterations of X-linked genes encoding p53 regulators, were proposed to explain a higher cancer incidence and death in male patients (Haupt et al., 2019). Here, on the contrary, male mice and a subset of male patients were more efficiently protected against Burkitt-like lymphomas, which adds another layer of complexity to sex-specific differences in tumorigenesis. The p53 pathway thus underlies cancer sex-disparities through multiple mechanisms, that may notably include variations in p53 isoforms or Ackr4 expression.

Materials and Methods

Mice

Design and construction of the $p53^{\Delta AS}$ mouse model was previously described (Simeonova et al., 2013). A minimum of 10 backcrosses with C57Bl6/J mice of both sexes (Charles River Laboratories) were performed before establishing the cohorts of $p53^{+/+}$ and $p53^{\Delta AS/\Delta AS}$ littermate mice used in this study. Mouse genotyping with multiple primer sets confirmed >99% C57Bl6/J genetic background after 10 backcrosses (primer sequences available upon request). Cohorts of $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice were established with identical parental origin of the Eμ-Myc transgene. For all experiments, mice housing and treatment were conducted according to Institutional Animal Care and Use Committee of the Institut Curie.

Cells and cell culture reagents

Mouse embryonic fibroblasts (MEFs) were isolated from 13.5 days post-coitum embryos and cultured in a 5% CO_2 and 3% O_2 incubator, in DMEM Glutamax (GIBCO), with 15% FBS (PAN Biotech), 100 μM 2-mercaptoethanol (Millipore), 0.1 mM Non-Essential Amino-Acids and Penicillin/Streptomycin (GIBCO) for less than 5 passages, except for 3T3 experiments, performed in a 5% CO_2 incubator for 9 passages. Cells were treated for 24h with 0.5 μg/ml Doxorubicin (Sigma-Aldrich), 15 μM Etoposide (Sigma-Aldrich), 10 μM Nutlin 3a (Vassilev et al., 2004) (Sigma-Aldrich) and/or 5 μg/ml 17β-Estradiol (Merck). At least 3 independent experiments with at least 2 independent littermate MEF clones of each genotype and each sex were performed to measure DNA damage responses. For estradiol assays, four independent experiments with three independent MEF male clones of each genotype were performed. Human lung fibroblast MRC5 and its SV40-transformed derivatives were cultured in a 5% CO2 and 3% O2-regulated incubator in MEM medium without Phenol Red (Gibco), completed with 10% FBS, 2 mM L-glutamine (Gibco), 1 mM pyruvate, 0.1 mM Non-Essential Amino-Acids, and penicillin/streptomycin, and treated for 24h with 10 μM Nutlin 3a (Sigma-Aldrich) and/or 5 μg/ml 17β-Estradiol (Merck). Four independent experiments were performed.

Quantitative RT-PCR

Total RNAs were extracted using nucleospin RNA II (Macherey-Nagel), reverse-transcribed using superscript IV (Invitrogen), and real-time quantitative PCRs were performed on an ABI

PRISM 7500 using Power SYBR Green (Applied Biosystems) as previously described (Simeonova et al., 2012). For quantification of p53 isoforms in healthy tissues, a forward primer in exon 10 and a reverse primer encompassing the boundary between exons 10 and 11 were used for p53-α amplification, whereas the same forward primer and a reverse primer located in exon AS were used for p53-AS amplification (see Table S2 for primer sequences). To determine the AS/α mRNA ratios, expression levels were compared with a standard curve generated by serial dilutions of a plasmid containing both p53-AS and p53-α cDNAs.

Western blots

Thymocytes were lysed in RIPA buffer (50 mM Tris-HCl pH 8, 150 mM NaCl, 5 mM EDTA, 0.5% deoxycholic acid, 0.1% SDS, 1% NP-40) with a cocktail of protease inhibitors (Roche) and 1 mM PMSF (Sigma). Whole-cell extracts were sonicated three times for 10 s and centrifuged at 13 000 r.p.m. for 30 min to remove cell debris. MEFs or B-cell lymphomas were lysed in Giordano's buffer (50 mM Tris-HCl pH 7.4, 250 mM NaCl, 5 mM EDTA, 0.1% Triton X-100) with a cocktail of protease inhibitors (Roche) and 1 mM PMSF (Sigma). Protein lysate concentration was determined by BCA assay (Thermo Scientific) and 30 µg of each lysate was fractionated by SDS–PAGE on a 4-12% polyacrylamide gel and transferred onto PDVF membrane (Amersham). Membranes were incubated with antibodies against p53 (CM5, Novocastra), myc (9E-10, Santa Cruz), p21 (F-5, Santa Cruz) and actin (actin-HRP sc47778, Santa Cruz) and revealed with SuperSignal West femto detection reagent (Thermo Scientific).

Apoptosis Assays

Six weeks-old $p53^{+/+}$ and $p53^{\Delta AS/\Delta AS}$ male mice were whole-body irradiated with 5 Gy of γ-irradiation. Mice were sacrificed 4h later and thymocytes were recovered, stained with AnnexinV-FITC Apoptosis detection kit (Abcam), then analyzed by flow cytometry using FlowJo.

Cell-Cycle assays

Log phase MEFs were irradiated at room temperature with a CS γ-irradiator at doses of 3 or 10 Gy, incubated for 24 h, then pulse-labeled for 1h with 10 µM BrdU, fixed in 70% ethanol, double-stained with FITC anti BrdU and propidium iodide, and sorted by flow cytometry using a BD FACSort. Data were analyzed using FlowJo.

Oncogene-induced tumor xenografts

MEFs with the indicated genotypes were sequentially infected with pWZL-E1A12S and pBABE-Hrasv12 viruses as previously described (Toledo et al., 2006). In total, $5x10^6$ E1A- and Ras- (E1A+Ras) expressing MEFs of each genotype were injected subcutaneously into the flanks of 7-weeks-old female athymic nude mice (at least 4 mice per genotype) and tumor volumes were determined 1, 8, 15, 21 and 25 days after injection. Importantly, populations of (E1A+Ras)-expressing cells were used to minimize potential differences in expression levels that could result from independent viral insertion sites.

Cell sorting of B-cell sub-populations

Splenocytes were recovered from 6 weeks-old asymptomatic mice and incubated with DAPI and the following antibodies: APC rat anti-mouse CD45R/B220, FITC rat anti-mouse CD43, PE rat anti-mouse IgM and BV605 rat anti-mouse IgD (BD Pharmingen). First, the B220+CD43- cells were selected by flow cytometry from DAPI negative living cells, yielding subsequently 4 different B subpopulations based on IgM and IgD labeling: IgM-/IgD- preB lymphocytes, IgM low/IgD- immature B lymphocytes, IgM high/IgD- transitional B lymphocytes and IgM+/IgD+ mature B lymphocytes.

RNA-Seq analysis

Total RNA was extracted from the spleen of 4-6 weeks-old asymptomatic mice using nucleospin RNA II (Macherey-Nagel). The quality of RNA was checked with Bioanalyzer Agilent 2100 and RNAs with a RIN (RNA integrity number) $\geq$ 6 were retained for further analysis. RNA was depleted from ribosomal RNA, then converted into cDNA librairies using a TruSeq Stranded Total Library preparation kit (Illumina). Paired-end sequencing was performed on an Illumina MiSeq platform. Reads were mapped to the mouse genome version GRCm38 and counted on gene annotation gencode.vM18 with featureCounts (Liao et al., 2014). Differentially expressed genes of C57Bl6/J genetic background with an adjusted p-value < 0.05 were identified using the DESeq2 R package (Love et al., 2014).

Luciferase assays

The candidate p53 responsive element (p53 RE) in the *Ackr4* promoter was identified by using the JASPAR database of binding profiles (Fornes et al., 2020) with the position weight matrix scanner PWMscan (Ambrosini et al., 2018). A 1.5 kb fragment from *Ackr4* intron 1, containing a WT or mutant p53 RE at its center, was cloned upstream a SV40 minimal promoter and a luciferase reporter gene in the backbone of a PGL3 plasmid (Promega). We used lipofectamine 2000 to transfect $p53^{-/-}$ MEFs with 2 µg of either luciferase expression vector, 2 µg of an expression vector for $p53^{WT}$, $p53^{AS}$ or the DNA-binding mutant $p53^{R270H}$, and 30 ng of a renilla luciferase expression plasmid (pGL4.73, Promega) for normalization. Transfected cells were incubated for 24 h then trypsinized, resuspended in 75 µl culture medium with 7.5% FBS and transferred into a well of an optical 96-well plate (Nunc). The dual-glo luciferase assay system (Promega) was used according to the manufacturer's protocol to lyse the cells and read firefly and renilla luciferase signals. Results were normalized, then the average luciferase activity in cells transfected with the WT p53RE luciferase reporter and the $p53^{R270H}$ expression plasmid were assigned a value of 1.

Statistical Analyses

Student's unpaired *t* tests were used in most Figures to analyze differences between WT vs ΔAS values. Log-Rank Mantel-Cox were used to analyze Kaplan-Meier tumor-free survival curves. Analyses were performed using Graphpad Prism 5, and values of $p<0.05$ were considered significant.

Data availability

RNA sequencing data have been deposited in the Gene Expression Omnibus (GEO) under the accession code GSE209708.

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Author Contributions

F.T, J.C.B. and I.S. initiated the project; F.T., B.B. and A.F. designed the research and analyzed data; B.B., A.F., I.S., J.L., A.M., V.L., F.T., A.H., W.V.--Z., E.E. and J.R. performed the research; M.G., M.P. and A.M. analyzed RNAseq data; F.T. acquired funding; F.T., A.F. and B.B. wrote the paper.

Declaration of interests

The authors declare no competing interest.

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Figures

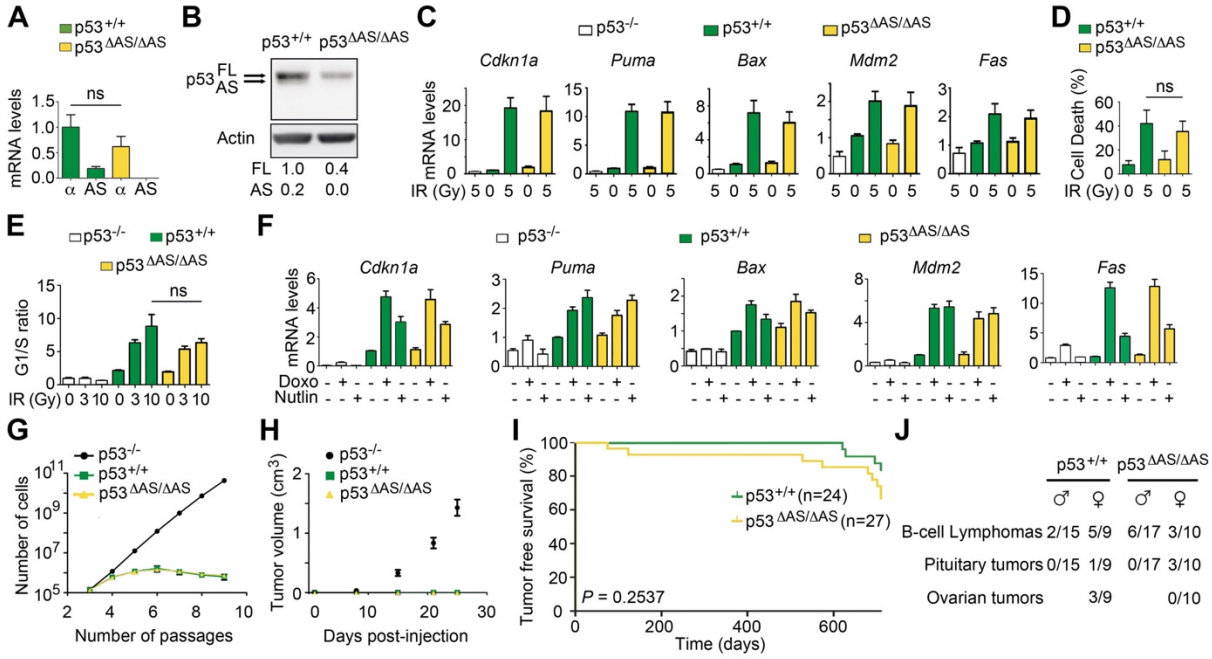


Figure 1. The loss of p53-AS isoforms does not alter cellular stress responses or survival to spontaneous tumors.

(A) mRNAs for p53-α and p53-AS isoforms from thymocytes of irradiated mice were quantified by RT-qPCR, and p53-α levels in $p53^{+/+}$ mice were assigned a value of 1. Means ± SEM (n=3). (B) Protein extracts from thymocytes of irradiated mice were immunoblotted with p53 or actin antibodies. After normalisation to actin, full-length (FL) p53-α levels in $p53^{+/+}$ thymocytes were assigned a value of 1. (C) mRNA levels of p53 target genes in thymocytes, before or after γ-irradiation. Means ± SEM (n=3). (D) Thymocyte apoptotic response to γ-irradiation. Means ± SEM (n=6). (E) Cell cycle control in mouse embryonic fibroblasts (MEFs) after γ-irradiation. Asynchronous MEFs were exposed to 0-10 Gy γ-irradiation, and after 24 hr, cells were labelled with BrdU for 1 hr and analyzed by FACS. Means ± SEM from >3 independent experiments with at least 2 independent MEF clones per genotype. (F) mRNA levels of p53 target genes in MEFs untreated or treated with 0.5 µg/ml of clastogenic Doxorubicin (Doxo) or 10 µM of the Mdm2 antagonist Nutlin. Means ± SEM from > 3 experiments with ≥ 2 independent MEF clones. (G) MEFs proliferation under hyperoxic conditions. Cells were grown according to a 3T3 protocol. Each point is the mean from 4 independent MEF clones, the value for each clone resulting from triplicates. (H) Growth of tumor xenografts. E1A+Ras-expressing MEFs were injected into the flanks of nude mice and tumor volumes were determined after 1-25 days. Means ± SD (n=4 per timepoint and genotype). (I) Tumor-free survival of $p53^{+/+}$ and $p53^{\Delta AS/\Delta AS}$ mice (n=cohort size). (J) Incidence of the indicated tumor types, determined at death after macroscopic examination and histological analysis. In A, D, E, ns=non-significant in Student's t-test.

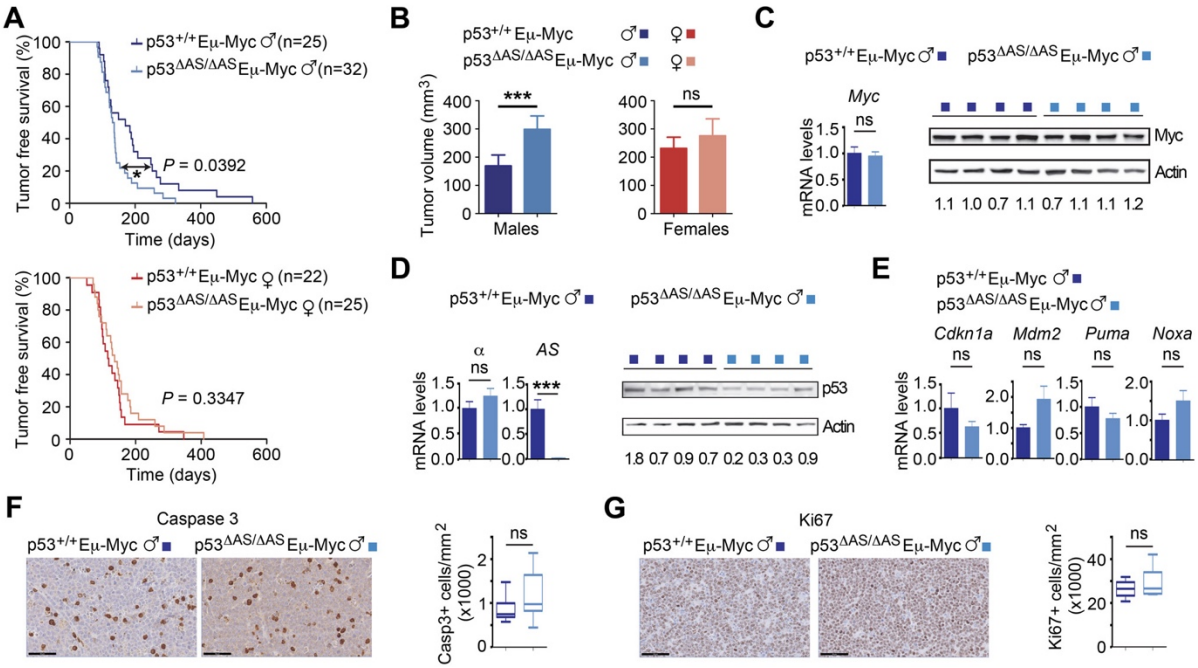


Figure 2. Male-specific acceleration of Myc-induced B-cell lymphomagenesis in mice lacking p53-AS isoforms.

(A) Tumor-free survival of $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice, classified according to sex. (n=cohort size). (B) Tumor volumes upon dissection of $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice, classified according to sex. Means ± SEM from 100 lymph nodes from $p53^{+/+}$ Eμ-Myc males, 96 from $p53^{+/+}$ Eμ-Myc females, 148 from $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males and 124 from $p53^{\Delta AS/\Delta AS}$ Eμ-Myc females. (C) Myc mRNA and protein levels in lymph node tumors. (D) Levels of p53-α and p53-AS transcripts and p53 protein levels in lymph node tumors. (E) Transcript levels of the indicated p53 target genes. Means ± SEM (n=6 per genotype). (F-G) Apoptosis (F) and cell proliferation (G) in tumor lymph nodes from Eμ-Myc males were determined by immunohistochemistry with antibodies against cleaved caspase-3 and ki67, respectively. Positive cells were counted and normalized to the analyzed areas. Means ± SEM (n=6 mice per assay and genotype). In F, G scale bars=50 μm. Statistical analyses with Mantel-Cox (A) and Student's t (B-G) tests. ***P<0.001, *P<0.05, ns: non-significant.

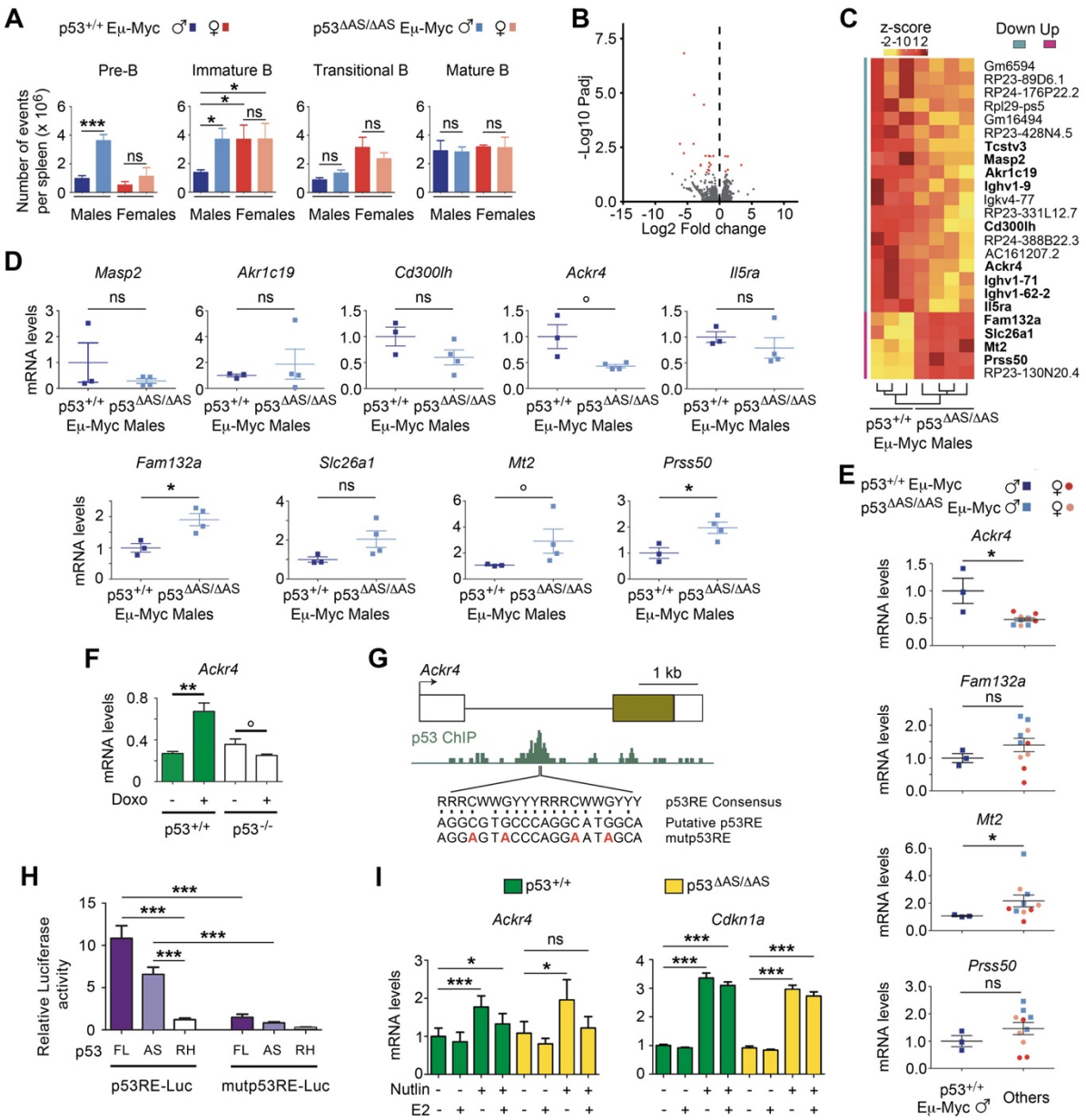


Figure 3. The loss of p53-AS isoforms affects *Ackr4* expression in Eμ-Myc male mice.

(A) B-cell subpopulations in spleens of 6 weeks-old p53$^{+/+}$ Eμ-Myc and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc mice. Means ± SEM (n=6 per genotype). (B-C) RNAseq analysis of spleens from p53$^{+/+}$ Eμ-Myc (n=3) and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc (n=4) 4-6 weeks-old male mice. Volcano plot (B), with differentially expressed genes (DEGs) in red. Unsupervised clustering heat-map plot (C), with DEGs ranked according to mean fold changes, and protein-coding genes in bold. (D) RT-qPCR analysis of candidate DEGs from spleens of p53$^{+/+}$ Eμ-Myc males and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc males. Means ± SEM (n=3-4 per genotype). (E) RT-qPCR analysis of indicated DEGs from spleens of 4-6 weeks-old p53$^{+/+}$ Eμ-Myc males, p53$^{\Delta AS/\Delta AS}$ Eμ-Myc males, p53$^{+/+}$ Eμ-Myc females and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc females. Means ± SEM (n=3-4 per sex and genotype). (F) *Ackr4* is transactivated by p53 in response to stress. Ackr4 mRNAs in untreated or doxorubicin-treated WT and p53$^{-/-}$ MEFs. Data from 2-3 MEFs per genotype (Younger et al., 2015). (G) A putative p53 response element in *Ackr4* intron 1. Top: Map of the *Ackr4* gene. (boxes: exons (brown box: translated region), black line: intron 1); middle: p53 ChIP in doxorubicin-treated MEFs according to ChIP-Atlas (SRX270554) (Oki et al., 2018); bottom: p53 Response Element (p53RE) consensus sequence (R=G or A, W=A or T, Y=C or T), the putative p53RE and its

mutated counterpart. (H) Luciferase assays of the candidate p53RE. A 1.5 kb fragment containing the WT or mutant p53RE was cloned upstream a luciferase reporter, then transfected into $p53^{-/-}$ MEFs together with an expression plasmid for full length p53 (FL), p53-AS or the DNA-binding mutant $p53^{R270H}$ (RH). Means ± SEM (n=4-6). (I) In MEFs, p53 activation leads to an increased *Ackr4* expression attenuated by estradiol. *Ackr4* and *Cdkn1a* mRNAs were quantified by RT-qPCR from $p53^{+/+}$ and $p53^{\Delta AS/\Delta AS}$ MEFs, untreated or treated with 10 µM Nutlin and/or 5 µg/ml 17-β estradiol (E2). Means ± SEM from 4 independent experiments. In A, D, E, F, H, I ***P<0.001, **P<0.01, *P<0.05, °P≤0.057, ns: non-significant in Student's t or Mann-Whitney tests.

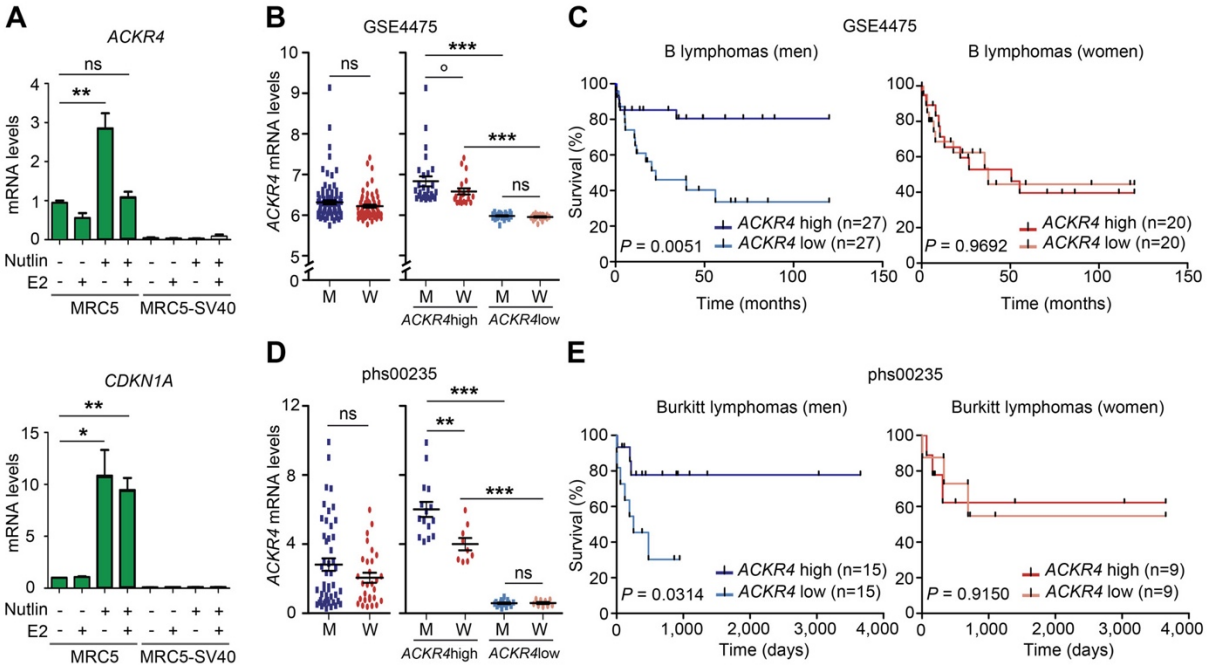


Figure 4. *ACKR4* is a male-specific prognostic factor in Burkitt lymphoma.

(A) In human cells, p53 activation leads to an increased *ACKR4* expression abrogated by estradiol. *ACKR4* and *CDKN1A* mRNAs were quantified by RT-qPCR from p53-proficient (MRC5) and p53-deficient (MRC5-SV40) human fibroblasts, untreated or treated with Nutlin and/or estradiol (E2). Means ± SEM from 4 independent experiments. (B-C) Analysis of lymphoma dataset #GSE4475. *ACKR4* gene expression was plotted for all lymphoma patients with clinical follow-up (91 men [M], 68 women [W]), classified according to sex (B, left). Gene expression (B, right) or survival curves (C) were plotted for the 30% patients (27 men, 20 women) with the highest or lowest *ACKR4* expression, classified according to sex. (D-E) Analysis of Burkitt lymphoma-specific dataset #phs00235. *ACKR4* gene expression was plotted for all patients with a Burkitt lymphoma diagnosed at age 0-17 (48 males, 29 females), classified according to sex (D, left). Gene expression (D, right) or survival curves (E) were plotted for the 30% patients (15 men, 9 women) with the highest or lowest *ACKR4* expression, classified according to sex. Statistical analyses by Student's t or Mann-Whitney tests (A, B, D) and Mantel-Cox (C, E) test. ***P<0.001, **P<0.01, * P<0.01, °P=0.054, ns: non-significant.

Supplemental Information

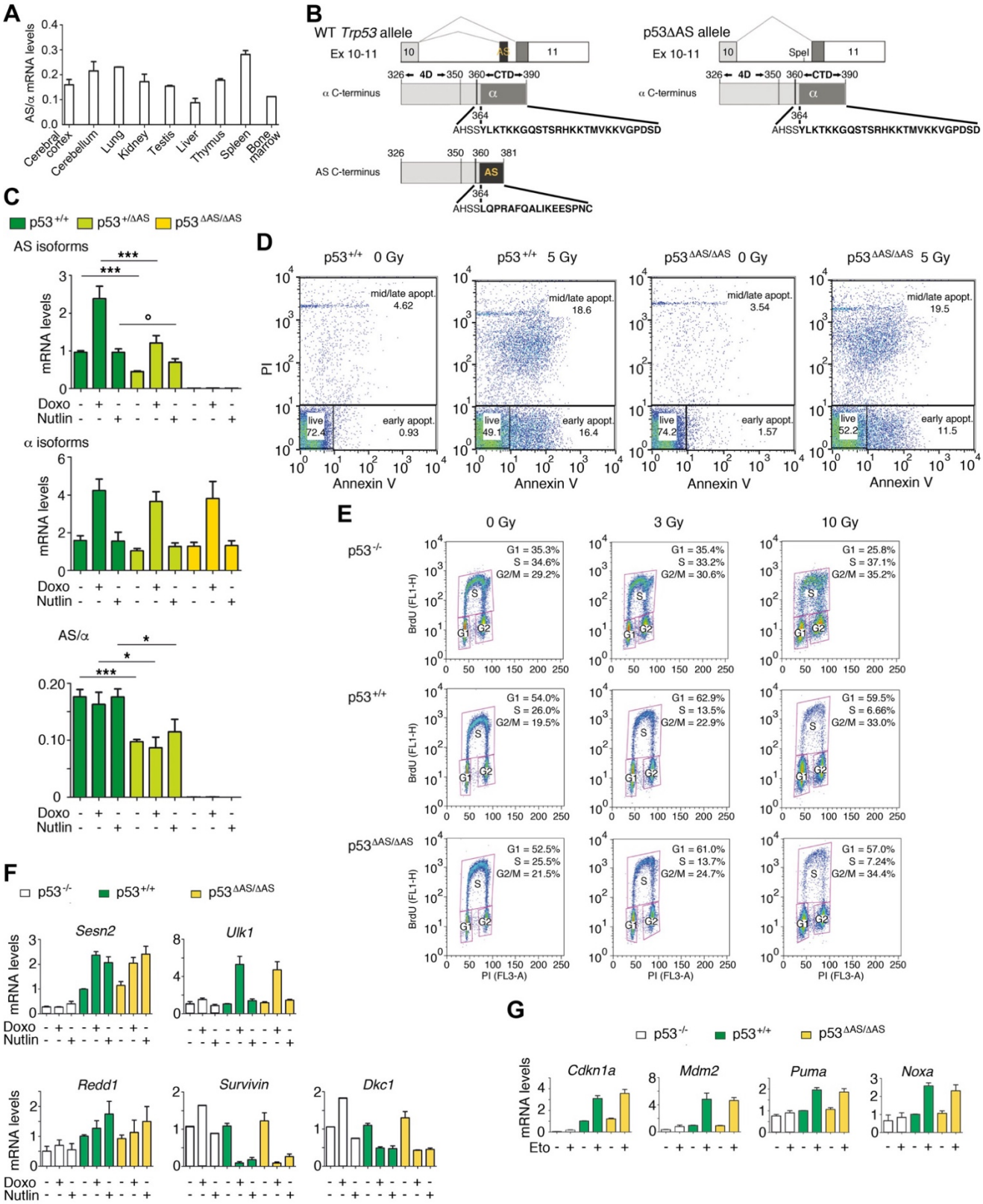
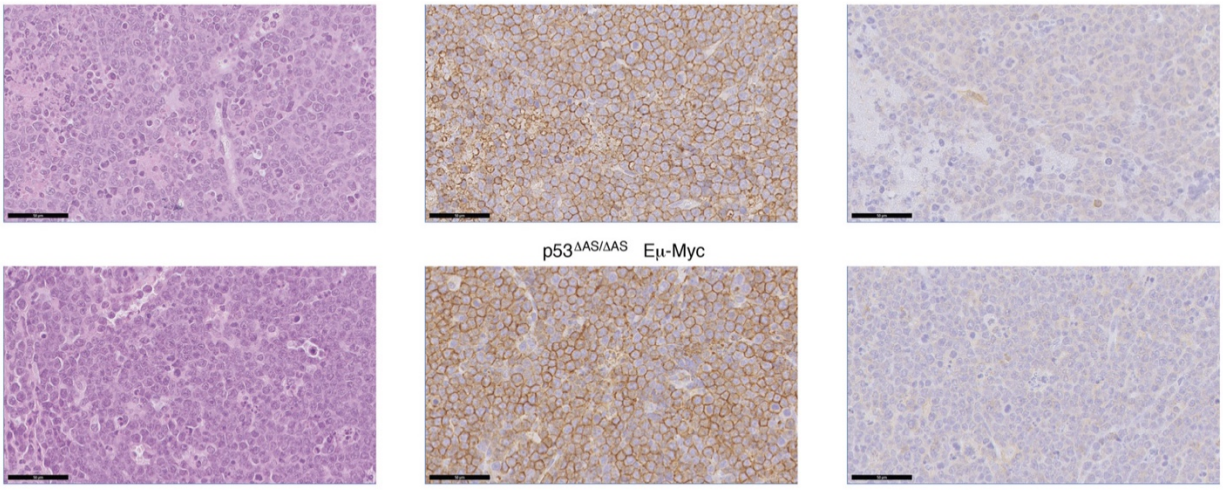


Figure S1

Figure S1. Description of the p53$^{\Delta AS}$ mouse model and analysis of p53$^{\Delta AS/\Delta AS}$ thymocytes and fibroblasts.

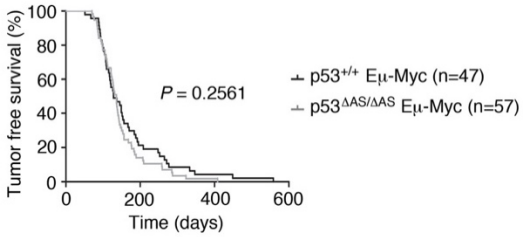
(A) Relative expression of p53 isoforms with an α or AS C-terminus in tissues of wild-type mice. RNAs were prepared from the indicated tissues of wild-type mice, then p53-AS and p53-α mRNAs were quantified by RT-qPCR. Results are expressed as mean AS/a ratios ± SD, from 3 independent experiments. (B) Comparative maps of the wild-type (WT) and ΔAS *Trp53* alleles. Left, top: the 3' end of the WT *Trp53* gene is shown - black line: intron 10; boxes are for exons, with greytones for translated regions from exon 10 (light gray), exon 11 (dark gray), and exon AS (black). Left, below: the WT *Trp53* allele encodes proteins with two different C-termini. (4D: tetramerization domain, CTD: C-terminal domain). Right, top: the 3' end of the ΔAS *Trp53* allele is shown; the AS exon was deleted and replaced by a SpeI restriction site. Right, below: the ΔAS *Trp53* allele only enables the synthesis of proteins with the 'canonical' α C-terminus. (C) Quantifications of p53 isoforms with an AS or an α C-terminus, in mouse embryonic fibroblasts (MEFs) of the indicated genotypes, that were either untreated, or treated for 24hr with 0.5 µg/ml doxorubicin (Doxo) or 10 µM Nutlin. Means ± SEM from >3 experiments with ≥ 2 independent MEF clones of each genotype are shown. ***P<0.001, *P<0.05, °P=0.07 by Student's t-test. (D) Apoptotic responses of p53$^{+/+}$ and p53$^{\Delta AS/\Delta AS}$ thymocytes. p53$^{+/+}$ and p53$^{\Delta AS/\Delta AS}$ mice were irradiated and their thymocytes were recovered and analyzed by FACS after annexin V-FITC staining. A typical experiment for cells of each genotype and condition is shown. Numbers indicate % cells, apopt.: apoptotic cells. (E) Cell cycle control of p53$^{-/-}$, p53$^{+/+}$ and p53$^{\Delta AS/\Delta AS}$ fibroblasts. Asynchronous MEFs were exposed to 0-10 Gy γ-irradiation, then after 24 hr cells were labelled with BrdU for 1 hr and analyzed by FACS. A typical experiment for cells of each genotype and condition is shown, with % of cells in G1, S and G2/M mentioned in each panel. (F) mRNA levels of the indicated genes were quantified in MEFs of the indicated genotypes left untreated or treated for 24 hr with 0.5 µg/ml Doxorubicin (Doxo), 10 µM Nutlin-3 (Nutlin). Means ± SEM from at least 3 experiments with 2 independent MEF clones of p53$^{+/+}$ and p53$^{\Delta AS/\Delta AS}$ genotypes are shown. (G) mRNA levels of the indicated genes were quantified in MEFs of the indicated genotypes left untreated or treated for 24 hr with 15 µM etoposide (Eto). Means ± SEM from at least 3 experiments with 2 independent MEF clones per genotype.

A



H & E B220 CD3

B



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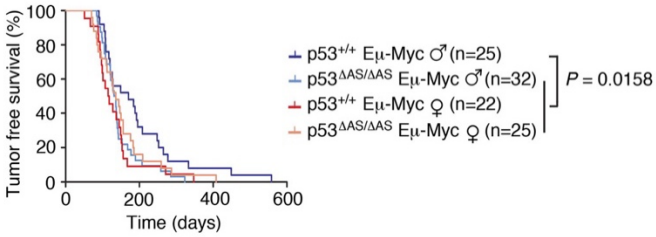


D

WT ♂	#10	#11	#B	#31	#45	#46	#50	#C	#28	#34	#36	#47	#51	#52
Trp53 DBD mutation	Yes	Yes	Yes	No	No	No	No	No	No	No	No	No	No	No

ΔAS ♂	#2	#14	#F	#17	#19	#24	#D	#16	#E	#15	#43	#4	#5	#7	#20	#21	#23	#41
Trp53 DBD mutation	Yes	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No	No

E

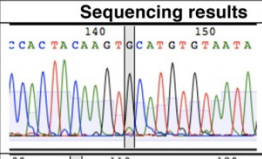
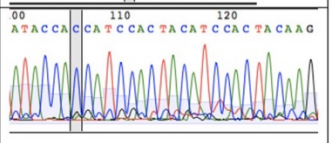
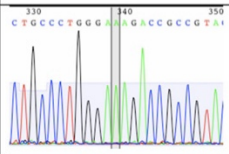
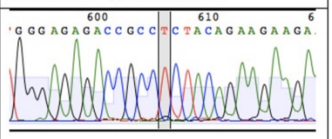
Tumor	DBD Mutation	Reference Sequence	Mutated Sequence	Sequencing results
WT #10	**DNA** : Exon 7 Substitution : 1235 :A>G **Protein** : Y233C	**DNA** : Exon 7 5'-accatccactacaagtacatgtgtaat-3' **Protein** : 228-TIHYKYMCN-236	**DNA :** Exon 7 5'-accatccactacaagt**G**catgtgtaat-3' **Protein** : 228-TIHYK**C**MCN-236	
WT #11	**DNA** : Exon 7 Duplication : 1230 :+catccacta **Protein** : Y231-IHY-K235	**DNA** : Exon 7 5'-accatccactacaagtacatgtgtaat-3' **Protein** : 228-TIHYKYMCN-236	**DNA** : Exon 7 5'-accatccacta**CATCCACTA**caagtacatgtgtaat-3' **Protein** : 228-TIHY**IHY**KYMCN-239	
WT #B	**DNA** : Exon 8 Substitution : 1367 :G>A **Protein** : R277K	**DNA** : Exon 8 5'-tgccctgggagagaccgccgt-3' **Protein** : 274-CPGRDRR-280	**DNA** : Exon 8 5'-tgccctggga**A**agaccgccgt-3' **Protein** : 274-CPG**K**DRR-280	
ΔAS #2	**DNA** : Exon 8 Insertion : 1376 : +T Substitution : 1377 : G>C **Protein** : R281fs	**DNA** : Exon 8 5'-gggagagaccgccgtacagaagaagaa aatttccgcaaaaaggaagtcctttgccctga-3' **Protein** : 276-GRDRRTEEE-284	**DNA** : Exon 8 5'-gggagagaccgcc**TC**tacagaagaagaa aatttccgcaaaaaggaagtcctttgccctga-3' **Protein** : 276- GRDR**LYRRRKFPQKGSPLP***	

Figure S2

Figure S2. Analysis of tumors from $p53^{+/+}$ Eμ-myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice.

(A) Histological analyses of Eμ-Myc-induced tumor lymph nodes. Tumor lymph nodes were analyzed by Hematoxilin-Eosin staining (H & E), or antibodies against B220 (a B-cell specific marker) or CD3 (a T-cell specific marker). $p53^{+/+}$ Eμ-Myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice developed similar B-cell lymphomas, characterized by massive tissue homogenization of the lymph node by B-cells. Scale bars=50 μM. (B) Tumor-free survival of $p53^{+/+}$ Eμ-myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc mice are similar when sexes are not considered (n=cohort size). (C) Increased tumor-free survival of $p53^{+/+}$ Eμ-myc males compared to $p53^{\Delta AS/\Delta AS}$ Eμ-Myc males, $p53^{+/+}$ Eμ-Myc females and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc females (n=cohort size). Statistical analysis with Mantel-Cox test. (D) *Trp53* DNA sequencing of $p53^{+/+}$ Eμ-myc and $p53^{\Delta AS/\Delta AS}$ Eμ-Myc tumors. The portion of the *Trp53* gene encoding the DNA Binding domain (DBD), most frequently mutated in cancers, was sequenced for 14 tumors from $p53^{+/+}$ (WT) Eμ-myc males and 18 tumors from $p53^{\Delta AS/\Delta AS}$ (ΔAS) Eμ-myc males, and *Trp53* mutations were found in 3/14 and 1/18 tumors, respectively. (E) Details on the 4 p53 mutations identified in (D).

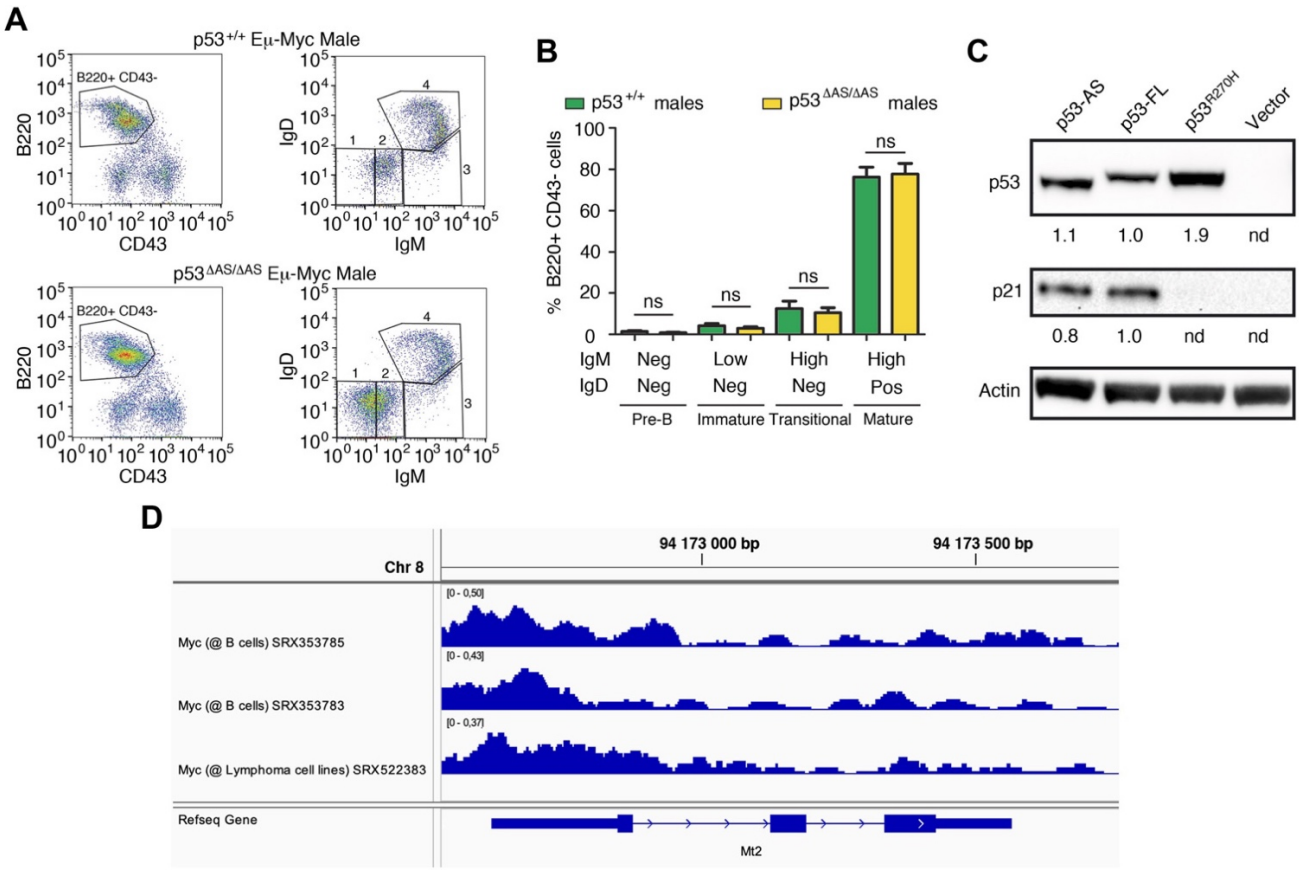


Figure S3. Analysis of pre-tumoral spleens.

(A) Cell sorting of B-cell subpopulations by FACS. Splenocytes were incubated with DAPI and the following antibodies: APC rat anti-mouse CD45R/B220, FITC rat anti-mouse CD43, PE rat anti-mouse IgM and BV605 rat anti-mouse IgD. First, the B220+CD43- cells were selected from DAPI negative living cells, subsequently yielding 4 different B subpopulations based on IgM and IgD labeling: IgM-/IgD- pre-B cells (box 1), IgM low/IgD- immature B cells (box 2), IgM high/IgD- transitional B cells (box 3) and IgM+/IgD+ mature B cells (box 4). Typical results with a $p53^{+/+}$ Eμ-Myc male mouse and a $p53^{\Delta AS/\Delta AS}$ Eμ-Myc male mouse are shown. (B) The loss of p53-AS isoforms does not impair B-cell differentiation in 6 weeks-old non-transgenic male mice. Means ± SEM from 4 mice per genotype. ns: non-significant by Student's t-test. (C) Control for expression of p53-FL, p53-AS and $p53^{R270H}$ in luciferase assays. As a control to luciferase experiments reported in Fig. 3H, protein extracts from $p53^{-/-}$ MEFs transfected with expression plasmids for p53-FL, p53-AS or $p53^{R270H}$ were immunoblotted with antibodies against p53, p21 and actin. Quantifications are relative to actin. p53-FL and p53-AS were expressed at similar levels and both transactivated p21, whereas the DNA-binding mutant $p53^{R270H}$ was expressed at higher amounts but failed to transactivate p21, as expected. nd: not determined. (D) Evidence for Myc binding at the *Mt2* promoter in B-cells. ChIP-Atlas reports Myc binding to *Mt2* promoter sequences in primary B-cells from the lymph nodes of Eμ-Myc mice (SRX353785, SRX353783) and in Eμ-Myc-induced lymphoma cells (SRX522383). Chr: chromosome.

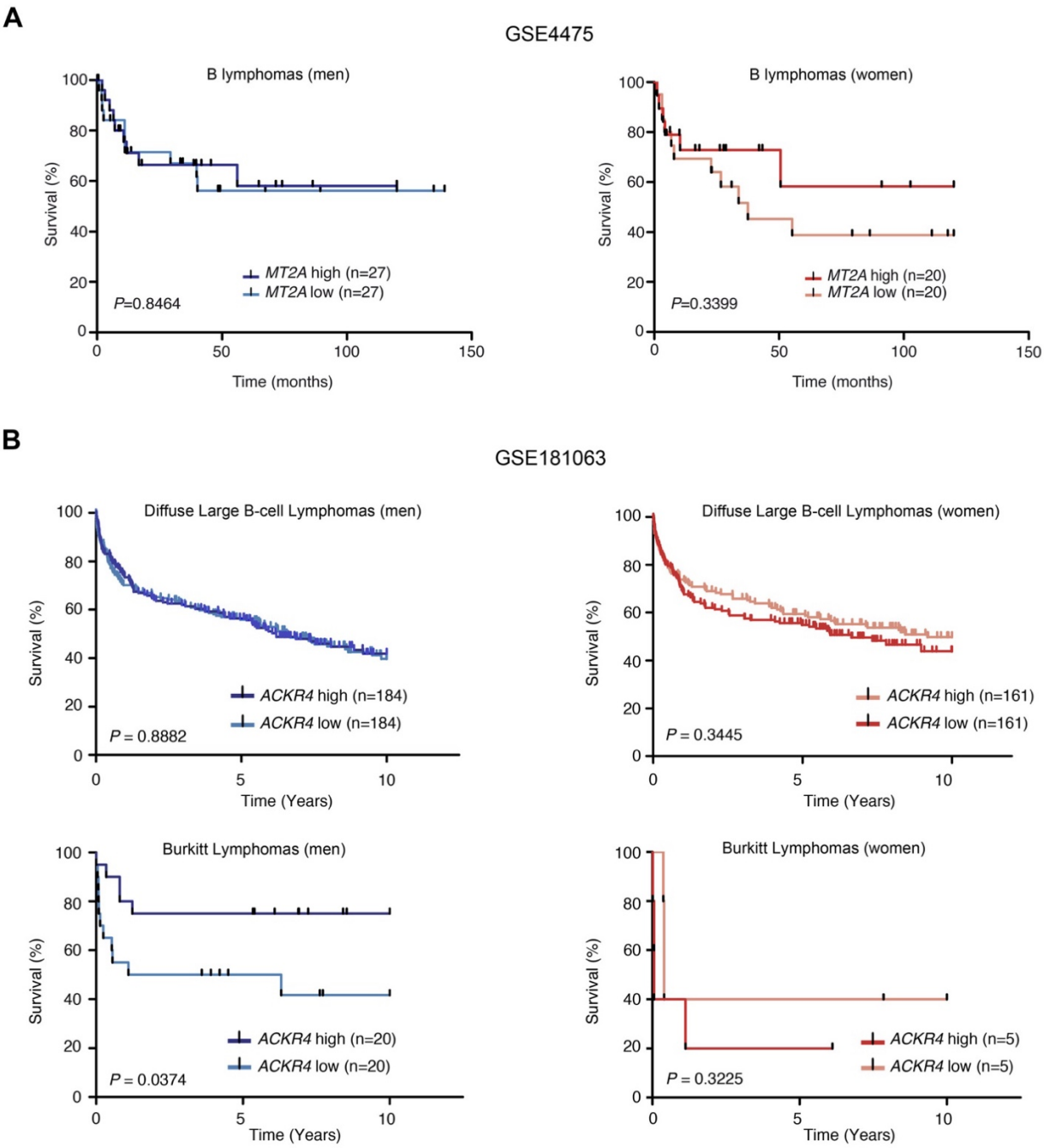


Figure S4. Further analyses of human B-cell lymphoma datasets.

(A) *MT2A* gene expression is not a prognostic marker in human B-cell lymphomas. Survival curves for the 30% patients with the highest *MT2A* mRNA levels and the 30% patients with the lowest *MT2A* mRNA levels according to sex (n=cohort sizes), for patients from dataset #GSE4475. Statistical analyses by Mantel-Cox tests. (B) *ACKR4* is a prognostic factor in Burkitt lymphomas but not in diffuse large B-cell lymphomas. Survival curves of patients from dataset #GSE181063, for the 30% patients with the highest *ACKR4* mRNA levels and the 30% patients with the lowest *ACKR4* mRNA levels, classified according to sex (n=cohort sizes), and diagnosed with either a diffuse large B-cell (top) or a Burkitt (bottom) lymphoma. Statistical analyses by Mantel-Cox tests.

Gene	WT_Myc_1	WT_Myc_2	WT_Myc_3	ΔAS_Myc_1	ΔAS_Myc_2	ΔAS Myc_3	ΔAS _Myc_4
Ackr4	267.610698766789	187.913317967838	103.698438238889	80.2839670298248	58.3264519788058	44.2019721676723	58.626719832054
Cdkn1a	1172.3897279307	1179.72136964871	1227.565295909	1248.51861547663	1018.37985154995	800.288338193645	1079.90417930643
Mdm2	4419.00743604647	4820.24950903391	5133.53980290714	4756.31040570283	4240.33305885918	4132.88439767736	5521.46447378284

Table S1. Expression of *Ackr4*, *Cdkn1a* and *Mdm2* in p53$^{+/+}$ Eμ-Myc and p53$^{\Delta AS/\Delta AS}$ Eμ-Myc male splenocytes.

Read numbers for the indicated genes, obtained by Bulk RNA-seq from the spleens of three p53$^{+/+}$ Eμ-Myc (WT_Myc) and four p53$^{\Delta AS/\Delta AS}$ Eμ-Myc (ΔAS_Myc) male mice.

Locus	Species	Usage	Forward	Reverse
Trp53	Mouse	Genotyping	5' AAGGGGTATGAGGGACAAGG 3'	5' GAAGACAGAAAAGGGGAGGG 3'
Eμ-Myc	Mouse	Genotyping	5' ACCCAGGCTAAGAAGGCAAT 3'	5' CGCTCACTCCCTCTGTCTCT 3'
Trp53 exon4	Mouse	Sequencing	5' CAGAGCAGAAAGGGACTTGG 3'	5' GCTAAAAAGGTTCAGGGCAAA 3'
Trp53 exon5-6	Mouse	Sequencing	5'TGGTGCTTGGACAATGTGTT 3'	5'TAGCACTCAGGAGGGTGAGG 3'
Trp53 exon7-9	Mouse	Sequencing	5'TGCCGAACAGGTGGAATATC 3'	5'CCTTGGTACCTTGAGGGTGA 3'
Ackr4	Mouse	RT-qPCR	5' GCACCTCTCCCAGCTTAAACA 3'	5' AATAGTATTCCGCTGACTGGTTCAG 3'
Akr1c19	Mouse	RT-qPCR	5' AGTTGCCTACTGTGCTCTTGGAT 3'	5' GAGAACTGGAGAGCTTGGGTCTA 3'
Bax	Mouse	RT-qPCR	5' CGGCGAATTGGAGATGAACT 3'	5' CCGTGTCCACGTCAGCAA 3'
Cdkn1a	Mouse	RT-qPCR	5' GCAGACCAGCCTGACAGATTTC3'	5' TTCAGGGTTTTCTCTTGCAGAAG 3'
Cd300lh	Mouse	RT-qPCR	5' GAACATTGGCCAGGTGACTCA 3'	5' AAGAAGGAGATGCTGCTCAACAG 3'
Fam132a	Mouse	RT-qPCR	5' GACAAGAAGACTCTGGTGGAACTG 3'	5' AGGAAGGCGCCCTGAGTAGT 3'
Fas	Mouse	RT-qPCR	5' ATGCACACTCTGCGATGAAG 3'	5' CAGTGTTCACAGCCAGGAGA 3'
Il5ra	Mouse	RT-qPCR	5' CCGCCTGCTTCGTCTTGTTA 3'	5' CAACCTGGTCCATAGATGACACA 3'
Masp2	Mouse	RT-qPCR	5' CCAATGAGAAGCCGTTCACA 3'	5' GAGACACTCTGCATTCATCCACAT 3'
Mdm2	Mouse	RT-qPCR	5' GTCTACCGAGGGTGCTGCAA 3'	5' AAGCAATGGTTTTGGTCTAACCA 3'
Mt2	Mouse	RT-qPCR	5' ACAATGCAAATGTACTTCCTGC 3'	5' CACTTCGCACAGCCCACG 3'
Myc	Mouse	RT-qPCR	5' CCACCAGCAGCGACTCTGA 3'	5' TCCACAGACACCACATCAATTTC 3'
Noxa	Mouse	RT-qPCR	5' CCTGGGAAGTCGCAAAAGAG 3'	5' GAGCACACTCGTCCTTCAAGTCT 3'
Prss50	Mouse	RT-qPCR	5' CGTGGCCCATTGCTTGA 3'	5' GCTCCCCGCCCTCACT 3'
Puma	Mouse	RT-qPCR	5' GAGCGGCGGAGACAAGAA 3'	5' GAGTCCCATGAAGAGATTGTACATGA 3'
Redd1	Mouse	RT-qPCR	5' GAGTCCCTGGACAGCAGCAA 3'	5' CATCCAGGTATGAGGAGTCTTCCT 3'
Sesn2	Mouse	RT-qPCR	5' CTTCCGCCACTCAGAGAAGGT 3'	5' CTTGCATGCGGGCTTCA 3'
Slc26a1	Mouse	RT-qPCR	5' GCCACTGCCCTTACTCTGATG 3'	5' GCCGGAGGATACCCATGAG 3'
Survivin	Mouse	RT-qPCR	5' TGTTTTTTCTGCTTTAAGGAATTGG 3'	5' TCTATGCTCCTCTATCGGGTTGTC 3'
Tcstv3	Mouse	RT-qPCR	5' CTCCAGCTGTTGTGGAATAAGTTC 3'	5' CCATGGATCCCTGAAGGTAAATC 3'
Trp53-α	Mouse	RT-qPCR	5' AAAGGATGCCCATGCTACAGA 3'	5' TCTTGGTCTTCAGGTAGCTGGAG 3'
Trp53-AS	Mouse	RT-qPCR	5' AAAGGATGCCCATGCTACAGA 3'	5' TGAAGTGATGGGAGCTAGCAGTT 3'
Ulk1	Mouse	RT-qPCR	5' TGTACATGGCTCCTGAGGTCAT 3'	5' CTCCACAGGTCAGCCTTTCC 3'
Ppia	Mouse	RT-qPCR control (MEFs)	5' TCTCCTTCGAGCTGTTTGCA 3'	5' CAGTGCTCAGAGCTCGAAAGTTT 3'
Rplp0	Mouse	RT-qPCR control (MEFs)	5' CGACCTGGAAGTCCAACTAC 3'	5' ATCTGCTGCATCTGCTTG 3'
Il2rg	Mouse	RT-qPCR control (spleen)	5' GGAGCTCCAAGGTCCTCATG 3'	5' TGTAGAAGTCAGGATCAAATCAGCTT 3'
Polr2a	Mouse	RT-qPCR control (spleen, thymus)	5' CTTTGAGGAAACGGTGGATGTC 3'	5' TCCCTTCATCGGGTCACTCT 3'
Vps4a	Mouse	RT-qPCR control (thymus)	5' GACAACGTCAACCCTCCAGAAA 3'	5' TCTGTGGCTTTTGTCACCAGAT 3'
ACKR4	Human	RT-qPCR	5' ACTGCTCCTCTCTGCCGACTAC 3'	5' GCCATTCATTTCATTTTCCTCAT 3'
CDKN1A	Human	RT-qPCR	5' ACCATGTGGACCTGTCACTGTCTT 3'	5' AGAAGATGTAGAGCGGGCCTTTGA 3'
PPIA	Human	RT-qPCR control	5' CATCTGCACTGCCAAGACTGA 3'	5' TTCATGCCTTCTTTCACTTTGC 3'
RPLP0	Human	RT-qPCR control	5' CTTGTCTGTGGAGACGGATTACAC 3'	5' TCAGCCAAGAAGGCCTTGA 3'

Table S2. Primer Sequences.