Links between behaviour and acute hyperthermia and hypoxia resistance in rainbow trout using isogenic lines

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Abstract

In the context of adaptation to climate change, acute hyperthermia and hypoxia resistance are

traits of growing interest in aquaculture. Although the feasibility of genetic improvement of

these resistance traits through selection has been demonstrated in rainbow trout

(*Oncorhynchus mykiss*), little is known about the mechanisms underlying such traits. This

poses a problem, as genetic selection of poorly understood traits can be inefficient and even

harmful. To better understand these mechanisms, the present study investigated the link

between several behaviour variables, and the resistance to acute hyperthermia and hypoxia.

Six rainbow trout isogenic lines were phenotyped for behaviour variables and for acute

hyperthermia and hypoxia resistance, using systematically different individuals. The

behaviour variables of the fish were phenotyped using an individual test in a new environment

and a group test during a moderate thermal challenge. Experimental design was robust, with

150 fish phenotyped per isogenic line for each resistance trait and 18 fish per isogenic line for

behavioural traits, distributed in triplicates. Acute hyperthermia and hypoxia resistance

phenotypes were related to behaviour phenotypes at the level of isogenic lines.

Significant differences in behaviour between isogenic lines were found, with certain

behaviour variables being highly correlated with acute hyperthermia and hypoxia resistance.

Isogenic lines with a higher activity level appeared to be more sensitive to acute hyperthermia

and more resistant to acute hypoxia than lines with a lower activity level. This previously

unknown link between behaviour and resistance phenotypes also suggests that some

behaviour variables could be used as predictors for acute hyperthermia and hypoxia resistance

in fish. This result could also lead to more ethical acute hyperthermia and hypoxia resistance

phenotyping protocols, as the current protocols in use are classified as severe by French ethics

committees.

Keywords: selective breeding, behaviour, indirect phenotyping, temperature, oxygen, climate

change, isogenic lines

Introduction

Resistance to acute hyperthermia and hypoxia are phenotypes of interest in fish for numerous

fields of research, such as evolutionary biology (Mandic, Todgham & Richards, 2009; Comte

& Olden, 2017), stress-coping physiology (Rytkönen et al., 2007; Saravia et al., 2021),

forecasts for climate change effects (Messmer et al., 2017; Potts, Mandrak & Chapman,

2021), and genetics for breeding robust animals (Baer & Travis, 2000; Perry et al., 2005;

Debes et al., 2021; Lagarde et al., 2023a; Prchal et al., 2023). The methods in use for

measuring these resistance phenotypes consist of increasing the temperature of the water for

hyperthermia phenotyping or decreasing the O2 saturation of the water for hypoxia

phenotyping until the fish lose equilibrium. The resistance phenotypes include the resistance

time and the temperature/O2 saturation at the equilibrium loss (Beitinger, Bennett &

Mccauley, 2000; Roze et al., 2013).

Previously, genetic determinism for acute hyperthermia and hypoxia resistance have been

demonstrated in rainbow trout (Perry et al., 2005; Lagarde et al., 2023a,b; Prchal et al., 2023).

In particular, Lagarde et al. (2023b) showed that some genotypes were more resistant to acute

hyperthermia or hypoxia using rainbow trout isogenic lines. However, little is known about

the physiological mechanisms underlying the breaking point leading to loss of equilibrium

during acute hyperthermia or hypoxia stress and hence about the origin of this observed

genetic variability. For acute hyperthermia resistance, the main theories about these

mechanisms are the failure of short-term essential organs such as the brain, the failure of

mitochondria processes, the thermal inactivation of enzymes, the desynchronization of

interdependent metabolic processes because of different response to temperature, the

alteration of membrane structures, and a lack of oxygen (Fry, 1971; Schmidt-Nielsen, 1997;

Pörtner, 2010; Chung & Schulte, 2020; Lefevre, Wang & McKenzie, 2021; Ern, Andreassen

& Jutfelt, 2023). For acute hypoxia resistance, loss of equilibrium may be caused by an

inability to produce adenosine triphosphate (ATP) fast enough with anaerobic substrate to

compensate the failure of the aerobic metabolism, or by damage induced by the harmful

metabolites produced by anaerobic metabolism, such as lactate or H+ (Nilsson & Östlund-

Nilsson, 2008; Mandic, Todgham & Richards, 2009; Bergstedt, Pfalzgraff & Skov, 2021).

There is no consensus within the scientific community on the reality of these mechanisms and

their relative importance, whether for resistance to acute hyperthermia or resistance to acute

hypoxia (Nilsson & Östlund-Nilsson, 2008; Ern, Andreassen & Jutfelt, 2023). It seems

therefore premature to breed for acute hyperthermia or hypoxia resistance, as selecting for

poorly understood traits could be ineffective or harmful in the long term. Thus, it is now

essential to understand why genotypes are resistant or not. Previous studies, aiming at the

evaluation of the resistance to acute hyperthermia and hypoxia of rainbow trout (Dupont-

Nivet et al., 2014; Lagarde et al., 2023b), have observed expression of an extensive

behavioural repertoire, suggesting that behaviour may be linked to resistance phenotypes. The

present paper aims to test whether behaviour may be linked with acute hyperthermia and

hypoxia resistance.

In addition, a strong interest in identifying mechanisms underlying resistance to acute

hyperthermia and hypoxia and whose phenotyping would be reasonably intrusive, such as

behavioural ones, would be to use them as predictors to replace the current classical

phenotyping methods. Indeed, although the classical phenotyping methods of acute

hyperthermia and hypoxia resistance have the advantage of directly measuring the phenotypes

of interest, they have several drawbacks. Firstly, acute hyperthermia and hypoxia challenges

push fish to their limits, with the loss of equilibrium as an endpoint generating animal

suffering. Within the 3R framework (Replacement, Reduction, Refinement), active research

must be conducted to reduce animal suffering in scientific experiments, and society has a

growing pressure on the scientific community to reduce animal suffering (Sloman et al.,

2019). Secondly, they affect the physical integrity of the fish. For example, mortality can be

as high as 25%-55% the week following an acute hyperthermia challenge in some strains of

rainbow trout (Strowbridge et al., 2021). Yet this result must be tempered as, in other strains

of rainbow trout, mortality was less than 5% (Strowbridge et al., 2021) and in zebrafish and

European sea bass, fish can recover reasonably quickly from acute hyperthermia or hypoxia

challenges with less than 1% of mortality (Claireaux et al., 2013; Morgan, Finnøen & Jutfelt,

2018).

In the present article, we investigate whether resistance to acute hyperthermia and hypoxia

may be linked to behaviour and discuss the possibility of using behaviour variables as a novel

way of phenotyping these resistances.

Our methodology used the rainbow trout isogenic lines, which have been previously shown to

have different resistance to either hyperthermia or hypoxia (Lagarde et al., 2023b). The

behaviour of fish was monitored using an individual test and a group test, the latter being

associated with moderate thermal stress. Measured behaviour phenotypes were related to

activity (acceleration, speed, distance travelled, movement), risk-taking (emergence, time

spent in a risky zone), and schooling behaviour (inter-individual distance, contacts between

fish). The resistance and behaviour phenotypes, measured on different individuals, were

related at the level of lines, thanks to the complete genetic homogeneity among fish within an

isogenic line. The advantage of measuring phenotypes on different individuals is to avoid a

previous experience to interfere with a later one.

1   Material and methods

1.1 Experimental fish

Experimental fish were produced at the INRAE PEIMA experimental fish farm (doi: 10.15454/1.5572329612068406E12, Sizun, France) with broodstock issued from the INRAE homozygous isogenic rainbow trout lines (Quillet et al., 2007). Homozygous isogenic lines are a powerful genetic resource, as fish are homozygous and genetically identical within a line. Therefore, fish from a given line can be considered as replicates of a unique genotype. The fish used in the present study are from the same rearing batch as those described in Lagarde et al. (2023b). Figure 1 summarises the rearing process of the fish and the experiments conducted. Briefly, six heterozygous isogenic lines were produced by fertilising eggs from same-day naturally spawning females of the homozygous isogenic line B57 with the sperm of six neomales (sex-reversed XX males) issued from the homozygous isogenic lines A02, A22, A32, B45, N38 and R23. At this stage, the only source of variation between heterozygous lines was expected to be the paternal genetic effect. Fertilised eggs were incubated in one hatching tray per line, supplied with spring water of little thermal amplitude over the year (11.5°C - 12.4°C). At 21 dpf (days post-fertilisation), hatching trays were each randomly divided into three hatching trays with no mixing between isogenic lines (>500 eyed eggs per hatching tray, 3x6 hatching trays). At 43 dpf, each hatching tray was transferred into a 0.25 m^3 indoor tank with no mixing between hatching trays (~500 larvae per tank, 3x6 tanks) supplied with the same spring water. At 145 dpf, the number of fries was reduced to 230 in each of the 18 tanks by randomly removing exceeding fries. At 150 dpf, the number of individuals was randomly reduced to 160 in each of the eighteen tanks, decreasing the density from 7.0 to 4.5 kg/m^3. The same day, among the approximately 160 kept individuals per tank,

100 were individually anaesthetised, PIT-tagged (Biolog-id, pit-tag dimensions: 12*9*2 mm,

weight: 0.1g) in the dorsal muscle just behind the head, and put back into their tanks.

At 175 dpf, PIT-tagged fish were removed from rearing tanks to form the three replicates for

phenotyping the resistance of isogenic lines to acute hyperthermia and the three replicates for

phenotyping the resistance of isogenic lines to acute hypoxia. The resistance to acute

hyperthermia and hypoxia were phenotyped between 182 and 187 dpf as already presented in

Lagarde et al. (2023b).

Among the 60 untagged fish kept in the rearing tanks, six were randomly collected to evaluate

individual and group behaviour between 177 and 179 dpf according to experimental setups

presented below.

During the rearing process, the maximum reached rearing density was 8.7 kg/m^3, and fish

were fed to satiation with a commercial diet from Le Gouessant's company.

1.2 Conduct of acute hyperthermia and hypoxia resistance experiments

The methods and devices used to determine the mean acute hyperthermia and hypoxia

resistance of isogenic lines were previously presented in Lagarde et al. (2023b) and will be

briefly summarized here.

Acute hyperthermia and hypoxia challenges were performed between 182 dpf and 187 dpf.

Fish mean body weight was 15.2 ± 3.5 g. Isogenic lines were phenotyped in triplicate for

acute hyperthermia and hypoxia resistance (Fig. 1). Each replicate (named A, B and C in Fig.

1) contained 50 fish from each of the six isogenic lines, *i.e.*, a total of 300 fish per replicate.

For the acute hyperthermia challenge, the temperature in the challenge tank was first

increased at a rate of 3.6°C/hour (12.4°C to 23.2°C in three hours). The temperature increase

rate was then slowed down to 0.6°C/hour and maintained at this rate until all fish lost

equilibrium. For the acute hypoxia challenge, dissolved oxygen saturation was decreased from

nearly 90% to 15% in 4.5 hours. Saturation was then stabilised around 15% for the rest of the

challenge.

As temperature increased in acute hyperthermia challenge or O2 saturation decreased in acute

hypoxia challenge, fish gradually lost equilibrium. Immediately after a fish lost equilibrium, it

was caught, its isogenic line was identified reading the PIT tag, and the time was recorded.

The fish was then anaesthetised, weighed, and euthanised.

Acute hyperthermia resistance was quantified as the cumulative thermal exposure (CTE) in

degree-minutes (deg-min), and acute hypoxia resistance was quantified as the cumulative

hypoxia exposure (CHE) in saturation minutes (sat-min), as described in Lagarde et al.

(2023b). CTE is the sum of the differences between the temperature in the challenge tank and

the temperature at the start of the challenge, calculated at each minute from the start of the

challenge to the time of loss of equilibrium. CHE is the sum of the differences between a

reference O2 saturation of 90%, the O2 saturation before the start of the challenge, and the O2

saturation measured in the challenge tank, calculated at each minute from the start of the

challenge to the time of loss of equilibrium. A high CTE or CHE indicates a high resistance to

acute hyperthermia or hypoxia, respectively.

As explained in Lagarde et al. (2023b), the effect of isogenic line on acute hyperthermia and

hypoxia resistance was analysed by fitting a model with *line* as a fixed effect, fish *body*

weight as a covariate, and their interaction. *line*, *body weight* and their interaction were all

significant in hyperthermia and hypoxia challenges (Lagarde et al. 2023b). Because the effect

of *body weight* on resistance was different between lines, the predicted lsmeans of *lines* on

acute hyperthermia and hypoxia resistance were estimated at 15.2 g, the mean body weight,

using lsmeans 2.30–0 R package (Lagarde et al. 2023b).

1.3 General conduct of behavioural experiments

Behavioural experiments were carried out in a quiet room with no traffic during the experiments. Figure 2 presents the diagram of the experimental devices.

Behaviour was simultaneously evaluated on six isolated individuals from the same isogenic line for 25 minutes (called individual test after that), and immediately after, the same six individuals were grouped in a tank and exposed to moderate heat stress for 150 minutes (called group test after that). Devices and duration of individual test were derived from previously published protocol used on zebrafish and seabass (Vignet et al., 2015; Alfonso et al., 2020; Alfonso, Peyrafort & Cousin, 2020; Alfonso, Gesto & Sadoul, 2021). Temperatures during the individual test and at the start of the group test were set similarly to the ones in home tanks. The same water source was used in home tanks and during the behaviour tests. Fish transfer between the two experiments was performed in buckets containing temperature-equilibrated water and lasted approximately 1 minute (Fig. 2). These behavioural assessment sessions were conducted in triplicates for each of the six lines, totalling 18 sessions. Each session lasted approximately 186 minutes: 5 minutes of acclimation, 25 minutes of individual test, 1 minute of transfer, 5 minutes of acclimation and 150 minutes of group test. Sessions were spread over three days at a rate of 6 sessions per day. Time slots of sessions were: 9h00-12h10, 10h00-13h10, 11h00-14h10, 13h00-16h10, 14h00-17h10, 15h-18h10. We arranged sessions for the three replicates of each isogenic line during morning, midday and afternoon slots so that there was no confusion between line and times slot.

1.4 Conduct of individual behaviour test

The purpose of the individual behaviour test was to evaluate behaviour in a novel environment. Six fish from the same isogenic line were caught in one of the rearing tanks according to the pre-determined session order and immediately placed randomly in six individual rectangular aquariums (20 × 40 cm, Europrix) filled with 12 L of system water corresponding to 15 cm depth of water (Fig. 2.a). The six aquariums were placed in a homemade rectangular tent lit by spots on the outside top of the tent to ensure homogeneous semi-darkness conditions. Aquariums were isolated from neighbouring aquariums by opaque white walls and backlit by an infrared device (IR floor 1 × 1 m, Noldus, The Netherlands) as infrared radiation is invisible for trout (Rader et al., 2007).

Aquariums were divided into two zones: on one side, a zone of size 20 x 8 cm covered by a plate opaque to visible light but transparent to infrared light called the safe zone, and on the other side, an uncovered zone of size 20 x 32 cm called the risky zone (Fig. 2.a). Fish were first placed in the safe zone, separated from the risky zone by a closed door. Fish were left for 5 minutes in the safe zone, and the door was then opened by a remote-controlled device, allowing fish to freely move between the safe and risky zones. Once the door opened, fish activity was video recorded for 25 minutes with a USB camera (DMK 31AU03, The Imaging Source) equipped with a 2.7–13.5-mm lens (Fujinon) and an infrared filter (The Imaging Source), using ICapture (The Imaging Source). After 25 minutes, the fish were immediately caught and placed in a bucket filled with same water. At the end of each test, water of aquariums was entirely renewed before the start of the following test.

1.5 Conduct of group behaviour test

The tank for the group test was a 75 x 75 cm tank, identical to the fish rearing tanks, filled with water at a 10 cm depth (Fig. 2.b).

Fish were gently poured out of the bucket in the group tanks. Once the fish transfer was

completed, the fish were left for 5 minutes for acclimation. Then, the water temperature was

linearly increased at a rate of 5.4°C/hour, corresponding to a temperature increase from

12.2°C to 23°C in 120 minutes by adding water from a heated buffer tank. Temperature was

then stabilised at 23°C for 30 minutes (Fig. 2.b). Fish activity was recorded for 150 minutes

with a video-surveillance camera (Full HD: 1920□×□1080 px, 105°, VIZEO—Adrien

Alarme), linked to a PC with an acquisition card. The fish were lit with two spots on each

tank side for homogeneous lighting. At the end of the experiment, fish were softly fished,

anaesthetised (Tricaïne MS222, 50 mg/L), quickly wiped clean, weighed and euthanised

(Tricaïne MS222, 150 mg/L). The fish mean body weight was 11.6 ± 3.2 g, all lines included.

Detailed isogenic lines mean body weights are given in Table S1.

1.6 Videos' behaviour variables extraction

Videos were analysed with EthoVision XT15.0 software. Fish positions were tracked using a

dynamic subtraction method. The accuracy of the tracked positions was checked by viewing

the superposition of the tracked and actual positions of fish in all videos and manually editing

if required. The Lowess smoothing method was applied to eliminate background noise.

In individual behaviour experiment, eight variables were measured: the maximum

acceleration (ACC_MAX, $cm.s^{-2}$), the distance travelled (DIST_TRAV, cm), the latency to

first exit from the safe zone (EMERGENCE, s), the zone change frequency (FRQ_CHAN,

$\#.min^{-1}$), the absolute meander (MEANDER, $deg.cm^{-1}$), the moving duration (MOV%, % of

the time), the time spent in the risky zone (RISK%, % of the time) and the maximum velocity

(VEL_MAX, $cm.s^{-1}$).

In the group behaviour experiment, seven variables were measured related to the behaviour of

the group: the average inter-fish distance (DISPERSION, cm), the maximum acceleration

(ACC_MAX, cm.s^{-2}), the distance travelled (DIST_TRAV, cm), the body contact frequency

(FRQ_CONTACT, #.min^{-1}), the absolute meander (MEANDER, deg.cm^{-1}), the moving

duration (MOV%, % of the time), and the maximum velocity (VEL_MAX, cm.s^{-1}). More

details on the measured variables are given in Table 1. The software tracked individual fish

positions, but in the group test, identities swapped frequently when two fish crossed paths in

front of the camera. This is almost unavoidable, and since group variables were measured,

fish identity was not useful.

To consider the effect of time on behaviours, behaviour variables were extracted over several

time bins: every 5 minutes for the individual test and every 30 minutes for the group test. This

was done to evaluate the potential effects of acclimation of the fish along the duration of the

individual test and of temperature increase along the group test.

1.7 Behaviour data statistical analysis

Analyses were performed in R version 3.6.2 (R Development Core Team, 2019).

Behavioural variables were measured at the individual level in the individual and group tests

(6 isogenic lines x 3 replicas x 6 individuals, 108 data points per behaviour variable and time

bin). However, these individual-level data were averaged at the replica level in the statistical

analysis (6 isogenic lines x 3 replicas, 18 data points per behaviour variable and time bin). As

there were significant body weight differences between isogenic lines (Table S1), behavioural

variables needed to be corrected by the fish body weight to disentangle the genetic and the

body weight effects, as done by Millot et al. (2014). Fish body weight was only known at the

replica level, as it was measured at the end of the group test and it was not possible to reassign

it to individuals of the individual and group tests a posteriori. For the group test, a

supplementary reason for not being able to reassign individuals was that Ethovision software

could not follow each fish individually due to frequent swapping of identity when two fish

cross paths. The experimental unit was, therefore, the replica. The setup of these experiments

meant that replicas were nested within isogenic lines.

To test for an effect of the isogenic line through time bins on behaviour variables, we fitted

linear mixed effects models with *body weight* as a covariate, *time* (time bins) and *line*

(isogenic line) as categorical fixed effects, and the interactions between *time*, *line* and *body*

weight, and *replica* as a random effect. Time was set up as a categorical variable because a

visual inspection of response variables plotted against time did not reveal any linear

relationship. We only tested *line* and *body weight* effects for EMERGENCE, as this event can

only happen once in the whole test. In the group test, the *time* effect was confounded with the

temperature increase, as no control of the group test, *i.e.*, without temperature increase, was

realised in the present study.

Full and reduced linear mixed models (with vs. without *time* and/or *line* and/or *body weight*

and/or the different interactions between these variables) were fitted using the function lme in

package nlme 3.1–153 (https://CRAN.R-project.org/package=nlme). The reduced versus full

models were compared using likelihood ratio tests to determine the best model for each

response variable. To select the best model for each response variable, weighted terms

allowing variances to differ among replica and isogenic lines were considered. To account for

temporal autocorrelation, autoregressive AR1 or autoregressive moving average ARMA(1,1)

covariance structures were considered (Zuur et al., 2009). Among the compared models (with

vs. without weighted terms and/or AR1/ARMA(1,1) structures), the one with the smallest

Akaike's Information Criterion (AIC) was selected as the best one. Assumptions of normality,

homoscedasticity and independence of final linear mixed models were verified by visual

inspection of residual-fit plots.

The predicted least-squares means (lsmeans) of *line* and *time* on each response variable were

estimated using lsmeans 2.30–0 R package (https://cran.r-project.org/package=lsmeans). For

behaviour variables for which the *body weight* effect was significant, lsmeans of *time* and *line*

(when significant) were estimated considering a body weight of 11.6 g, corresponding to the

mean body weight of fish used in the behaviour experiment. Differences among *line* and *time*

lsmeans for behaviour variables were compared by Tukey's test with Type 1 error risk set at

0.05 using the same lsmeans 2.30–0 R package. For behaviour variables for which the best

model included the *line × time* interaction, *line* lsmeans were calculated at each *time* bin and

vice-versa.

To get a synthetic view of the relationships between behaviour variables, *line* lsmeans were

reduced to two principal components using PCA. Three kinds of PCA were produced: a PCA

with only the *line* lsmeans of individual test, a PCA with only the *line* lsmeans of group test

and a PCA with the *line* lsmeans of individual and group tests.

1.8 Screening for relationships between behaviour and resistance variables

The lsmeans of the same six isogenic lines for the behavioural variables and the resistance to

acute hyperthermia and hypoxia stress were calculated using the steps mentioned in the

previous sections. Behaviour variables for which lsmeans of isogenic lines were highly

correlated with lsmeans of isogenic lines on resistance phenotype were considered as

potentially genetically linked with resistance phenotypes.

Mean body weight was slightly lower in the behaviour tests (11.6 ± 3.2 g) than in resistance

challenges (15.2 ± 3.5 g) since fish had between one and two more weeks to grow in the

resistance challenges. Rankings of lines based on body weight were similar between the two

experiments (Fig. S1).

2 Results

2.1 Body weight, isogenic line and time effects influence on behaviour variables

The significance of effects (*line*, *time*, *body weight* and their interactions) on the different

behaviour variables is shown in Table 2. Figure 3 shows the raw and modelled behaviour

phenotypes obtained for each isogenic line in the individual test, and Figure 4 shows the same

in the group test. Figure S2 shows the raw and modelled behaviour phenotypes obtained for

each *time* bin in the individual test, and Figure S3 shows the same in the group test. Modelled

behaviour phenotypes data for *line* and *time* bins shown in Figures 3-4-S2-S3 are available in

Tables S2-S3-S4-S5, respectively.

2.1.1 Behaviour variables with no significant effect

Behaviour variables for which neither *time*, *line* nor *body weight* effects were significant were

EMERGENCE and MEANDER in the individual test and FRQ_CONTACT and MEANDER

in the group test (Table 2). These variables were therefore not investigated further.

2.1.2 Behaviour variables with significant *body weight* effect

The overall fish body weight was 11.6 ± 3.2 g in behaviour tests. There were significant

differences in body weight between lines (ANOVA, N = 108, df=5; F value = 17.3; P < 2.8e-

14). Details on the mean body weight of isogenic lines are given in Table S1. The lightest line

was A32h (9.6 ± 2.6 g), and the heaviest was N38h (14.9 ± 2.8 g). *body weight* had a

significant effect on ACC_MAX in individual test and had no significant effect on any of the

variables measured in group test (Table 2). *body weight* effect on ACC_MAX was an increase

in 2.5 cm.s^{-2} per gram, *i.e.*, bigger fish had a higher acceleration, all equal.

2.1.3 Behaviour variables with significant *time* effect and no significant interaction

between *time* and *line* effects

In the individual test, the *time* effect was significant for variables ACC_MAX, MOV%,

RISK% and VEL_MAX without any interaction with *line* effect (Table 2). The effect of *time*

on these behaviour variables is presented in Fig. S2 and Table S4. ACC_MAX and

VEL_MAX were significantly higher in the first time bin and then tended to decrease. In

contrast, MOV% and RISK% were significantly lower in the first time bin and then tended to

increase. These evolutions reflect the change in behaviour over time. Indeed, at the beginning

of the test, fish displayed bursts of high activity bouts (high ACC_MAX and VEL_MAX)

interspersed with freezing moments (low MOV%). In a second step, the number of activity

bursts decreased (lower ACC_MAX and VEL_MAX) and the behaviour became more

exploratory with higher moving duration and time spent in the risky zone (increase in MOV%

and RISK%).

In the group test, the *time* effect was significant for some behaviour variables but always with

a significant *time* x *line* interaction (Table 2). Results are presented in section 2.1.5.

2.1.4 Behaviour variables with significant *line* effect and no significant interaction

between *line* and *time* effects

In the individual behaviour test, the *line* effect was significant for ACC_MAX, DIST_TRAV,

FRQ_CHAN, RISK%, and VEL_MAX (Table 2). Therefore, there were significant variations

in activity levels between the lines during the individual test, presented in Fig. 3 and detailed

in Table S2. Line A32h had a significantly lower DIST_TRAVEL (0.9 m), FRQ_CHAN (0.2

#.min^{-1}), and VEL_MAX (8.7 cm.s^{-1}) than A02h (3.3 m; 0.9 #.min^{-1}; 18.1 cm.s^{-1} respectively).

Line A02h had a significantly higher RISK% (68.5%) than lines N38h (30.3%) and A22h

(39.7%). Line A22h had a significantly higher ACC_MAX (47.4 cm.s^{-2}) than B45h (24.8

cm.s^{-2}) and higher VEL_MAX (18.1 cm.s^{-1}) than A32h (8.7 cm.s^{-1}).

In the group behaviour test, the *line* effect was significant and did not interact with the *time*

effect for ACC_MAX and DISPERSION (Table 2). Line A32h had a significantly lower

ACC_MAX (716 cm.s^{-2}) than line R23h (867 cm.s^{-2}) and a significantly lower group

DISPERSION (19 cm) than the other five lines (24-28 cm) (Fig. 4.a&b, Table S3 for details).

2.1.5 Behaviour variables with significant interaction between *time* and *line* effects

In the individual test, the interaction between *time* and *line* effects was not significant

regardless of the behaviour variables (Table 2).

In the group test, the interaction between *time* and *line* was significant for DIST_TRAV,

MOV% and VEL_MAX (Table 2), meaning that the effect of *time* on these behaviours was

different depending on the *line* and vice-versa. Lsmeans of *line* effect for these variables were

then estimated at each time bin (Table S3) and are presented in Fig. 4.c&d&e. For

DIST_TRAV, there were significant differences between isogenic lines at time bin 4 (B45h

higher than R23h). For MOV%, there were significant differences between isogenic lines at

time bin 4, with B45h higher than N38h. For VEL_MAX, there were significant differences

between isogenic lines at time bin 1 (A22h higher than B45h) and time bin 4 (A22h higher

than N38h). Lsmeans of *time* effect was also estimated for each isogenic line (Table S5) and

the estimates are presented in Fig. S3. Interestingly, the lines evolved differently over time

during the group challenge. The distance travelled (DIST_TRAV) tended to decrease over the

course of the challenge for some lines (A02h, A22h, A32h and R23h) before increasing again

to starting level for some of them (A22h and A32h). For other two activity remained constant

over time at high level for B45h and lower level for N38h(Fig. S3.a). MOV% was

significantly different between some time bins for B45h and R23h (Fig. S3.b). VEL_MAX

was significantly different between some time bins for A02h and A32h (Fig. S3.c).

2.2   Relationships between behaviour variables

2.2.3   PCA

The Principal Component Analyses (PCAs) of isogenic *lines* lsmeans are presented in Fig. 5.a

for the individual test, Fig. 5.b for the group test, and Fig. 5.c for the individual and group

tests together. Only the first two components were presented, as they explained more than

74% of the variance in these three PCAs. Correlations between behaviour variables in the

individual and group tests and between the individual and group tests are also presented in

Tables S6-S7-S8, respectively.

2.2.4   Individual behaviour test

The PCA of individual test variables highlights two groups of individual behaviour variables

(Fig. 5.a). The first group, constituted of ACC_MAX, DIST_TRAV, FRQ_CHAN and

VEL_MAX, contributed to PCA axis 1 for 93% and were highly positively correlated

together with a correlation coefficient (r) ranging from 0.74 to 0.98 (Table S6). These four

variables, each referring to the fish's activity level and showing strong positive and consistent

correlations between them, were named activity variables of individual test. The second

group, constituted of RISK%, only contributed to PCA axis 2 for 82% and was nearly

uncorrelated with ACC_MAX and VEL_MAX with a r ranging from 0.21 to 0.23, and

moderately correlated with DIST_TRAV and FRQ_CHAN with a r ranging from 0.51 to 0.54

(Table S6). Fish staying longer in the risky zone therefore tended to swim longer distances

than others.

2.2.5   Group behaviour test

In the group test, the interaction between *line* and *time* was significant for the variables

DIST_TRAV, MOV% and VEL_MAX (Table 2). Lsmeans were therefore calculated for

these variables at each time bin, which are all represented by different colours on the PCA

(Fig. 5.b). ACC_MAX and DISPERSION were not time bin dependant, as this effect and the

interaction with *line* were not significant (Table 2); lsmeans were therefore calculated for all

time bins merged together (Fig. 5.b).

This PCA highlights a clear separation between the DIST_TRAV, MOV%, and VEL_MAX

variables of the first and last two time bins. Indeed, within time bins 1 and 2, DIST_TRAV,

MOV% and VEL_MAX were moderately to highly positively correlated, with a r ranging

between 0.30 and 0.94 (Table S7). Within time bins 4 and 5, DIST_TRAV, MOV% and

VEL_MAX were strongly positively correlated, with a r ranging between 0.51 and 0.99

(Table S7). Correlations between VEL_MAX, DIST_TRAV, and MOV% of the time bins 1

and 2 and time bins 4 and 5 were mostly negative, close to zero, or moderate (Table S7).

Interestingly, DIST_TRAV, MOV% and VEL_MAX of time bin 3 were intermediaries

between the first and last two time bins (Fig. 5.b). Thus, correlations between VEL_MAX,

DIST_TRAV and MOV% were between -0.03 and 0.91 for time bins 1, 2 and 3 and between -

0.11 and 0.91 for time bins 3, 4 and 5.

ACC_MAX variable was generally not strongly correlated with the other activity variables

(DIST_TRAV, MOV% and VEL_MAX), contrary to what we observed in the individual test.

The DISPERSION variable was moderately to strongly positively correlated with DIST_TRAV, MOV%, and VEL_MAX at time bins 1, 2, and 3, and uncorrelated with these variables at time bins 4 and 5 (Table S7). DISPERSION was also poorly correlated with ACC_MAX (Table S7).

2.2.6 Individual and group behaviour tests

Relationships between isogenic *line* lsmeans of behaviour variables measured in the individual and group tests are presented on the PCA in Fig. 5.c, and correlation coefficients are given in Table S8. The PCA of individual and group test variables shows that the correlations between the individual and group test variables gradually turned from positive to negative with time bins in the group test as time and temperature increased (Fig. 5.c). Thus, correlations between activity variables in the individual test (ACC_MAX, DIST_TRAV, FRQ_CHAN and VEL_MAX) and DIST_TRAV, MOV% and VEL_MAX in the group test were moderately to highly correlated when considering time bins 1 and 2 of the group test, not correlated when considering time bin 3 of the group test, and negatively correlated when considering time bins 4 and 5 of the group test (Fig. 5.c and Table S8). RISK% of individual test was negatively correlated with activity variables in the group test for time bins 4 and 5, with a r between -0.53 and -0.84.

2.3 Correlations between behaviour variables and resistance phenotypes

Correlation coefficients between isogenic *line* lsmeans on behaviour variables and isogenic *line* lsmeans on resistance phenotypes are given in Table 3. Fig. 6 presents the relationship between behaviour variables and any of the two resistance phenotypes when the |Pearson correlation| between them was above an arbitrary threshold set to 0.7.

The isogenic *line* mean effect on acute hyperthermia resistance was strongly negatively

correlated ($r < -0.71$) with the isogenic *line* mean effect on DIST_TRAV and VEL_MAX in

the individual test, and with VEL_MAX in time bin 2 (VEL_MAX2) in the group test (Fig.

6.a&b). Correlation between isogenic *line* mean effect on acute hyperthermia resistance and

other activity variables in the individual test (ACC_MAX and FRQ_CHAN), and in the group

test (DIST_TRAV, MOV%, VEL_MAX; for the five time bins) were also slightly to

moderately negative, with a r between -0.11 and -0.69 (Table 3). ACC_MAX in the group test

was uncorrelated with acute hyperthermia resistance.

The isogenic *line* mean effect on acute hypoxia resistance was highly positively correlated

with the isogenic *line* mean effect on DIST_TRAV and FRQ_CHAN in the individual test (r

> 0.88; Fig. 6.c and Table 3). Again, in the individual test, acute hypoxia resistance was also

moderately positively correlated with other activity variables (ACC_MAX, VEL_MAX) and

RISK% ($0.54 < r < 0.69$; Table 3). In the group test, acute hypoxia resistance was highly

positively correlated with VEL_MAX in time bins 1 and 2 (VEL_MAX1 and VEL_MAX2)

and group DISPERSION ($r > 0.74$; Fig. 6.d and Table 3). Acute hypoxia resistance was also

moderately positively correlated with other activity variables (DIST_TRAV, MOV%) at the

beginning of the group test (*i.e.*, in the time bins 1 and 2: $0.38 < r < 0.51$). The correlations

between acute hypoxia resistance and activity variables (DIST_TRAV, MOV% and

VEL_MAX) turned moderately negative at the end of the group test (i.e. in time bins 4 and 5:

$-0.42 < r < -0.53$).

3   Discussion

In this work, we investigated whether links could be established between behaviour variables

and resistance to acute hyperthermia or hypoxia. To this end, we monitored the behaviour of

six isogenic lines and studied links with resistance to acute hyperthermia and hypoxia

phenotypes previously determined (Lagarde et al. 2023b).

3.1 Behaviours' evolution with time and isogenic lines

The *time* effect was significant for several behaviour variables during the individual and

group tests. At the beginning of the individual tests (first time bin), low moving duration, high

maximum acceleration and velocity was observed (Fig. S2.a&b&d), which is characteristic of

a "burst and freeze" behaviour. This behaviour could be linked to a high level of stress

(Brown, Gardner & Braithwaite, 2005; Staven et al., 2019) and is consistent with the stress

induced by the opening of the door and the novel environment proposed. In the following

time bins, as stress was relieved, behaviour gradually turned into exploratory behaviour,

which was characterised by an increase in moving duration and time spent in the risky zone

(Fig. S2.b&c) (Ryer, 2004; Malavasi et al., 2008; Millot, Bégout & Chatain, 2009).

Significant behavioural differences between isogenic lines were found in the individual and

group tests. These differences are consistent with the genetic determinism in fish behaviour,

which was reviewed in Bell (2008), as well as in an earlier study that, similarly to our own,

used isogenic lines (Millot et al., 2014). In particular, Millot et al. (2014) studied four

isogenic lines that are investigated here: A02h, A22h, B45h and N38h. Establishing direct

links between their results and our own is not simple since protocols and devices were

different; specifically, Millot et al. used novel object and emergence tests. It is interesting to

note that albeit evaluated in different contexts, A02h line was more active than B45h in both

studies, carried out a decade apart, supporting the hypothesis of a genetic basis for this trait.

The interaction between *time* and *line* was significant for several behaviour variables in the

group test but not in the individual test (Table 2). This difference may be due to the longer

duration of the group test, allowing time for the behaviour of isogenic lines to evolve. A

second hypothesis could be that the temperature increase during the group test led to a

different stress response of the lines, as several studies have shown different behaviour

reactions to stress depending on fish personalities in rainbow trout (Frost et al., 2013; White et

al., 2017).

3.2   Correlated behaviours between tests

Behaviour variables reflecting the level of activity of fish, *i.e.* ACC_MAX, DIST_TRAV,

FRQ_CHAN and VEL_MAX in the individual test, were highly correlated together at the

isogenic line level (Fig. 5a, Table S6). In the group test, DIST_TRAV, MOV% and

VEL_MAX were strongly correlated within each time bin (Fig. 5b, Table S7). The only

exception was ACC_MAX in the group test, which was poorly correlated with the three

behaviour variables mentioned above (Fig. 5b, Table S7). A strong correlation between

activity variables was already found in rainbow trout for variables similar to DIST_TRAV,

MOV% and VEL_MAX (Makaras et al., 2020).

The concept of coping styles (or personalities) refers to a consistent set of behavioural and

physiological traits that co-vary across environmental contexts (Koolhaas et al., 1999; Conrad

et al., 2011). Existence of coping style have been suggested in rainbow trout isogenic lines

(Millot et al. 2014). ACC_MAX, DIST_TRAV and VEL_MAX were moderately to highly

positively correlated together between the individual tests and the two first time bins of the

group tests, at the level of the isogenic line (Fig. 5.c, Table S8). Consistent relationships

between behaviour variables in two contexts allow us to ascribe coping styles to some lines.

Thus, in the group tests we conducted, we can differentiate the lines according to their level of

activity, some lines showing a low activity level, such as A32h, others with a high activity

level, such as B45h. However, it is essential to note that a robust personality assignment

would have required more behavioural testing, such as in Cerqueira et al. (2016). As time and

temperature increased, the correlations between individual and group behaviour variables

loosened (Fig. 5.c, Table S8), although we cannot determine which of the confounded time

and temperature factors was the cause. It is highly probable, however, that heat stress played a

role, given that temperature is a well-known factor affecting fish behaviours such as

schooling, activity, and maximum swimming speed (Hurst, 2007; Colchen et al., 2017;

Kochhann et al., 2021; Pilakouta et al., 2022).

3.3   Behaviour associated with acute hyperthermia or hypoxia resistance

Interestingly, coping styles were shown to be associated with physiological traits possibly

linked with acute hyperthermia and hypoxia resistance: proactivity was shown to be

associated with thermal preference in Nile tilapia (Cerqueira et al., 2016), reactivity was

linked to faster growth in brown trout *Salmo trutta* (Adriaenssens & Johnsson, 2011), and

boldness in a refuge emergence test was related with high aerobic capacity in bluegill sunfish

Lepomis macrochirus (Binder et al., 2016). To our knowledge, no study previous to our own

has investigated the genetic relationship between behaviour and resistance to acute

hyperthermia or hypoxia traits in fish.

In the individual behaviour test, acute hyperthermia resistance was found highly negatively

correlated with DIST_TRAV and VEL_MAX, and moderately negatively correlated with

ACC_MAX and FRQ_CHAN. In contrast, acute hypoxia resistance was found to be highly

positively correlated with DIST_TRAV, FRQ_CHAN, and moderately positively correlated

with ACC_MAX, RISK% and VEL_MAX. Therefore, acute hyperthermia resistance appears

to be associated with a low level of activity and acute hypoxia resistance with a high level of

activity. In the two first bins of the group test, we observed a similar yet less clear pattern:

acute hyperthermia resistance was still associated with low activity (high negative correlation

with VEL_MAX, moderate negative correlation with DIST_TRAV and MOV%), while acute

hypoxia resistance was associated with high activity (high positive correlation with

VEL_MAX and moderate positive correlation with DIST_TRAV and MOV%).

Our results on acute hyperthermia resistance are consistent with the study of Campos, Val &

Almeida-Val (2018), although the two studies are very different. They reported that fish

species with the lowest routine activity level, indirectly measured with a proxy, had the

highest resistance to acute hyperthermia. They also hypothesised that fish with the lowest

activity have lower resting energetic demands and thus can resist longer acute hyperthermia.

For acute hypoxia resistance, our results are hardly comparable to those in the existing

literature. In rainbow trout exposed to sub-lethal levels of acute hypoxia, divergent behaviours

were identified with some fish staying quiet and not panicking while other fish actively tried

to avoid hypoxia and showed peak activity (Van Raaij et al., 1996). The survival rate of fish

during the post-exposure recovery period was significantly associated with behavioural

strategy, as fish staying quiet during the hypoxia challenge had a higher survival rate after

return to normoxia compared to fish displaying high level of activity during the hypoxia

challenge (Van Raaij et al., 1996). In contrast, we showed in this study that isogenic lines

resistant to acute hypoxia were rather highly active. However, the results from the present

study and Van Raaij et al. (1996) do not conflict since the level of activity was measured in a

hypoxia stress challenge in Van Raaij et al. (1996) which was not the case in our experiment.

3.4   Potential of behaviour as a proxy of acute hyperthermia and hypoxia resistance

traits

In the present study, some behaviour variable were highly correlated with acute hyperthermia

or hypoxia resistance at the level of the isogenic lines. These correlations were consistent

between individual and group tests, even if they weakened with the duration and temperature

increase of the group test. Together, these results suggest a potential for using behavioural

variables as novel, non-stressful proxies for acute hyperthermia and hypoxia resistance in fish,

improving experimental fish welfare.

However, as correlations calculated with only six genotypes can give misleading results, these

results still need to be validated on more genotypes and different populations. In case of

confirmation, it would also be necessary to optimise how behaviour is measured, as data

acquisition and analysis of the measurements are time-consuming compared to the classical

phenotyping method. For instance, in Lagarde et al. (2023b), it took around 700 minutes to

phenotype the resistance to acute hyperthermia or hypoxia stress of 300 fish, equivalent to 0.4

fish phenotyped/minute, while in the present study, it took between 25 minutes (individual

test) to 150 minutes (group test) to phenotype six fish, equivalent to a rate of 0.24 or 0.04

phenotyped fish/minute. In addition, while the classical resistance phenotyping method

directly gives the phenotypes of resistance, the behaviour phenotyping method implies

extracting and analysing behaviour variables from recorded videos. However, the purpose of

the behavioural phenotyping protocols in the present study did not involve optimisation.

Future studies could implement significant time savings such as determining the minimum

behaviour phenotyping time period required, increasing the number of fish in individual or

group tests, automating data extraction, etc. The accuracy of behaviour variables in

discriminating phenotypes of resistance to acute hyperthermia and hypoxia must also be

determined at the fish level to determine their value as a proxy.

3.5   Implications for selective breeding of acute hyperthermia and hypoxia resistance

The correlations found between some behaviour variables and acute hyperthermia or hypoxia

resistances at the level of the isogenic line might reveal genetic correlations between some

behavioural variables and resistance phenotypes. Therefore, selection for resistance to acute

hyperthermia or hypoxia stress, as proposed by Perry et al. (2005), Lagarde et al. (2023a),

Prchal et al. (2023), could result in the indirect selection of certain behavioural variables. This

side effect is not surprising since, in practice, selected fish over several generations are known

to exhibit different behaviours, such as swimming, foraging, predator avoidance, social

behaviour, personality, reproduction and cognitive learning, compared to the wild type

(Pasquet, 2019). Specifically, for rainbow trout, domestication was shown to reduce predator

avoidance behaviours (Johnsson, 1993; Berejikian, 1995). Behaviour modifications of farmed

fish through direct or indirect genetic selection can positively or negatively impact fish

adaptation to farming systems (e.g. Doyle & Talbot, 1986). If they were to be selected in a

breeding program, special consideration must be paid to the effects of selecting these

resistance phenotypes on the behaviour variables. More generally, physiological mechanisms

underlying acute hyperthermia and hypoxia resistance are still not known with certainty

(Nilsson & Östlund-Nilsson, 2008; Ern, Andreassen & Jutfelt, 2023), and unintended trade-

offs with important breeding traits could therefore arise.

Conclusion

In the present study, we identified several behaviour variables strongly correlated with acute

hyperthermia and hypoxia resistance at the line level using six rainbow trout isogenic lines. In

individual test and the beginning of group test, active isogenic lines were generally found to

be sensitive to acute hyperthermia and resistant to acute hypoxia. These relationships between

behaviour and resistance to acute hyperthermia and hypoxia suggest that behaviour is a

promising approach to better understand the mechanisms underlying these resistances. These

results also support the idea that behaviour phenotyping could be a non-invasive method for

phenotyping acute hyperthermia or hypoxia resistances.

However, these conclusions, notably the correlations between behaviour and resistance traits,

must be verified in a higher number of isogenic lines and other populations of rainbow trout,

as well as at the fish level.

Ethical statement

Experiments were carried out at the INRAE experimental facilities (PEIMA, INRAE, 2021,

Fish Farming systems Experimental Facility, doi: 10.15454/1.5572329612068406E12, Sizun,

France) authorised for animal experimentation under the French regulation C29-277-02.

Experiments were carried out according to the European guidelines; the protocols were

evaluated and approved by the ethical committee CEFEA No 74 and authorised by the French

Ministry of Higher Education and Research (APAFIS#24784_2020032510486957).

References

Adriaenssens B, Johnsson JI. 2011. Shy trout grow faster: Exploring links between

personality and fitness-related traits in the wild. *Behavioral Ecology* 22:135–143. DOI:

10.1093/beheco/arq185.

Alfonso S, Gesto M, Sadoul B. 2021. Temperature increase and its effects on fish stress

physiology in the context of global warming. *Journal of Fish Biology* 98:1496–1508.

DOI: 10.1111/jfb.14599.

Alfonso S, Peyrafort M, Cousin X. 2020. Zebrafish Danio rerio shows behavioural cross-

context consistency at larval and juvenile stages but no consistency between stages.

Journal of Fish Biology 96:1411–1421. DOI: 10.1111/jfb.14310.

Alfonso S, Sadoul B, Cousin X, Bégout ML. 2020. Spatial distribution and activity patterns as

welfare indicators in response to water quality changes in European sea bass,

Dicentrarchus labrax. *Applied Animal Behaviour Science* 226:104974. DOI:

10.1016/j.applanim.2020.104974.

Baer CF, Travis J. 2000. Direct and correlated responses to artificial selection on acute

thermal stress tolerance in a livebearing fish. *Evolution* 54:238–244. DOI:

10.1111/j.0014-3820.2000.tb00024.x.

Beitinger TL, Bennett WA, Mccauley RW. 2000. Temperature tolerances of North American

freshwater fishes exposed to dynamic changes in temperature. *Environmental Biology of*

Fishes 58:237–275.

Bell AM. 2008. The Genetics of Fish Behavior. In: Magnhagen C, Braithwaite VA, Forsgren

E, Kapoor BG (eds.). 2008. *Fish Behaviour (1st ed.).* CRC Press, 151–181. DOI:

10.1201/b10757.

Berejikian BA. 1995. The effects of hatchery and wild ancestry and experience on the relative

ability of steelhead trout fry (Oncorhynchus mykiss) to avoid a benthic predator.

Canadian Journal of Fisheries and Aquatic Sciences 52:2476–2482. DOI: 10.1139/f95-

838.

Bergstedt JH, Pfalzgraff T, Skov P V. 2021. Hypoxia tolerance and metabolic coping

strategies in Oreochromis niloticus. *Comparative Biochemistry and Physiology -Part*

A□: Molecular and Integrative Physiology 257:110956. DOI:

10.1016/j.cbpa.2021.110956.

Binder TR, Wilson ADM, Wilson SM, Suski CD, Godin JGJ, Cooke SJ. 2016. Is there a

pace-of-life syndrome linking boldness and metabolic capacity for locomotion in bluegill

sunfish? *Animal Behaviour* 121:175–183. DOI: 10.1016/j.anbehav.2016.09.006.

Brown C, Gardner C, Braithwaite VA. 2005. Differential stress responses in fish from areas

of high- and low-predation pressure. *Journal of Comparative Physiology B* 175:305–312.

DOI: 10.1007/s00360-005-0486-0.

Campos DF, Val AL, Almeida-Val VMF. 2018. The influence of lifestyle and swimming

behavior on metabolic rate and thermal tolerance of twelve Amazon forest stream fish

species. *Journal of Thermal Biology* 72:148–154. DOI: 10.1016/j.jtherbio.2018.02.002.

Cerqueira M, Rey S, Silva T, Featherstone Z, Crumlish M, MacKenzie S. 2016. Thermal

preference predicts animal personality in Nile tilapia Oreochromis niloticus. *Journal of*

Animal Ecology 85:1389–1400. DOI: 10.1111/1365-2656.12555.

Chung DJ, Schulte PM. 2020. Mitochondria and the thermal limits of ectotherms. *Journal of*

Experimental Biology 223. DOI: 10.1242/jeb.227801.

Claireaux G, Théron M, Prineau M, Dussauze M, Merlin FX, Le Floch S. 2013. Effects of oil

exposure and dispersant use upon environmental adaptation performance and fitness in

the European sea bass, Dicentrarchus labrax. *Aquatic Toxicology* 130–131:160–170.

DOI: 10.1016/j.aquatox.2013.01.004.

Colchen T, Teletchea F, Fontaine P, Pasquet A. 2017. Temperature modifies activity, inter-

individual relationships and group structure in a fish. *Current Zoology* 63:175–183. DOI:

10.1093/cz/zow048.

Comte L, Olden JD. 2017. Evolutionary and environmental determinants of freshwater fish

thermal tolerance and plasticity. *Global Change Biology* 23:728–736. DOI:

10.1111/gcb.13427.

Conrad JL, Weinersmith KL, Brodin T, Saltz JB, Sih A. 2011. Behavioural syndromes in

fishes: a review with implications for ecology and fisheries management. *Journal of Fish*

Biology 78:395–435. DOI: 10.1111/j.1095-8649.2010.02874.x.

Debes P V., Solberg MF, Matre IH, Dyrhovden L, Glover KA. 2021. Genetic variation for upper thermal tolerance diminishes within and between populations with increasing acclimation temperature in Atlantic salmon. *Heredity* 127:455–466. DOI: 10.1038/s41437-021-00469-y.

Doyle RW, Talbot AJ. 1986. Artificial Selection on Growth and Correlated Selection on Competitive Behaviour in Fish. *Canadian Journal of Fisheries and Aquatic Sciences* 43:1059–1064. DOI: 10.1139/f86-132.

Dupont-Nivet M, Crusot M, Rigaudeau D, Quillet E. 2014. Genetic analysis of resistance to acute or chronic temperature stress using isogenic lines of Rainbow trout (Oncorhynchus mykiss). *Proceedings of 10th World Congress of Genetics Applied to Livestock Production*:2–4.

Ern R, Andreassen AH, Jutfelt F. 2023. Physiological mechanisms of acute upper thermal tolerance in fish. *Physiology*. DOI: 10.1152/physiol.00027.2022.

Frost AJ, Thomson JS, Smith C, Burton HC, Davis B, Watts PC, Sneddon LU. 2013. Environmental change alters personality in the rainbow trout, Oncorhynchus mykiss. *Animal Behaviour* 85:1199–1207. DOI: 10.1016/j.anbehav.2013.03.006.

Fry FEJ. 1971. The effect of environmental factors on the physiology of fish. In *Fish physiology* (Vol. 6, pp. 1-98). Academic press.

Hurst TP. 2007. Thermal effects on behavior of juvenile walleye pollock (Theragra chalcogramma): Implications for energetics and food web models. *Canadian Journal of Fisheries and Aquatic Sciences* 64:449–457. DOI: 10.1139/F07-025.

Johnsson JI. 1993. Big and brave: size selection affects foraging under risk of predation in juvenile rainbow trout, Oncorhynchus mykiss. *Animal Behaviour* 45:1219–1225. DOI: 10.1006/anbe.1993.1143.

Kochhann D, Sarmento CG, de Oliveira JC, Queiroz HL, Val AL, Chapman LJ. 2021. Take

time to look at the fish: Behavioral response to acute thermal challenge in two

Amazonian cichlids. *Journal of Experimental Zoology Part A: Ecological and*

Integrative Physiology 335:735–744. DOI: 10.1002/jez.2541.

Koolhaas JM, Korte SM, De Boer SF, Van Der Vegt BJ, Van Reenen CG, Hopster H, De

Jong IC, Ruis MAW, Blokhuis HJ. 1999. Coping styles in animals: Current status in

behavior and stress- physiology. *Neuroscience and Biobehavioral Reviews* 23:925–935.

DOI: 10.1016/S0149-7634(99)00026-3.

Lagarde H, Lallias D, Patrice P, Dehaullon A, Prchal M, François Y, D'Ambrosio J, Segret E,

Acin-Perez A, Cachelou F, Haffray P, Dupont-Nivet M, Phocas F. 2023a. Genetic

architecture of acute hyperthermia resistance in juvenile rainbow trout (Oncorhynchus

mykiss) and genetic correlations with production traits. *Genetics Selection Evolution*

55:39. DOI: 10.1186/s12711-023-00811-4.

Lagarde H, Phocas F, Pouil S, Goardon L, Bideau M, Guyvarc'h F, Labbé L, Dechamp N,

Prchal M, Dupont-Nivet M, Lallias D. 2023b. Are resistances to acute hyperthermia or

hypoxia stress similar and consistent between early and late ages in rainbow trout using

isogenic lines? *Aquaculture* 562:738800. DOI: 10.1016/j.aquaculture.2022.738800.

Lefevre S, Wang T, McKenzie DJ. 2021. The role of mechanistic physiology in investigating

impacts of global warming on fishes. *Journal of Experimental Biology* 224. DOI:

10.1242/jeb.238840.

Makaras T, Montvydienė D, Kazlauskienė N, Stankevičiūtė M, Raudonytė-Svirbutavičienė E.

2020. Juvenile fish responses to sublethal leachate concentrations: comparison of

sensitivity of different behavioral endpoints. *Environmental Science and Pollution*

Research 27:4876–4890. DOI: 10.1007/s11356-019-07211-6.

Malavasi S, Georgalas V, Mainardi D, Torricelli P. 2008. Antipredator responses to overhead

fright stimuli in hatchery-reared and wild European sea bass (Dicentrarchus labrax L.)

juveniles. *Aquaculture Research* 39:276–282. DOI: 10.1111/j.1365-2109.2007.01894.x.

Mandic M, Todgham AE, Richards JG. 2009. Mechanisms and evolution of hypoxia tolerance

in fish. *Proceedings of the Royal Society B: Biological Sciences* 276:735–744. DOI:

10.1098/rspb.2008.1235.

Messmer V, Pratchett MS, Hoey AS, Tobin AJ, Coker DJ, Cooke SJ, Clark TD. 2017. Global

warming may disproportionately affect larger adults in a predatory coral reef fish. *Global*

Change Biology 23:2230–2240. DOI: 10.1111/gcb.13552.

Millot S, Bégout M-L, Chatain B. 2009. Exploration behaviour and flight response toward a

stimulus in three sea bass strains (Dicentrarchus labrax L.). *Applied Animal Behaviour*

Science 119:108–114. DOI: 10.1016/j.applanim.2009.03.009.

Millot S, Péan S, Labbe L, Kerneis T, Quillet E. 2014. Assessment of Genetic Variability of

Fish Personality Traits using Rainbow Trout Isogenic Lines. :383–393. DOI:

10.1007/s10519-014-9652-z.

Morgan R, Finnøen MH, Jutfelt F. 2018. CTmax is repeatable and doesn't reduce growth in

zebrafish. *Scientific Reports* 8:1–8. DOI: 10.1038/s41598-018-25593-4.

Nilsson GE, Östlund-Nilsson S. 2008. Does size matter for hypoxia tolerance in fish?

Biological Reviews 83:173–189. DOI: 10.1111/j.1469-185X.2008.00038.x.

Pasquet A. 2019. Effects of Domestication on Fish Behaviour. In: *Animal Domestication*.

IntechOpen, 13. DOI: 10.5772/intechopen.78752.

Perry GML, Martyniuk CM, Ferguson MM, Danzmann RG. 2005. Genetic parameters for

upper thermal tolerance and growth-related traits in rainbow trout (Oncorhynchus

mykiss). *Aquaculture* 250:120–128. DOI: 10.1016/j.aquaculture.2005.04.042.

Pilakouta N, O'Donnell PJ, Crespel A, Levet M, Claireaux M, Humble JL, Kristjánsson BK,

Skúlason S, Lindström J, Metcalfe NB, Killen SS, Parsons KJ. 2022. A warmer

environment can reduce sociability in an ectotherm. *Global Change Biology*:1–9. DOI:

10.1111/gcb.16451.

Pörtner HO. 2010. Oxygen- And capacity-limitation of thermal tolerance: A matrix for

integrating climate-related stressor effects in marine ecosystems. *Journal of*

Experimental Biology 213:881–893. DOI: 10.1242/jeb.037523.

Potts LB, Mandrak NE, Chapman LJ. 2021. Coping with climate change: Phenotypic

plasticity in an imperilled freshwater fish in response to elevated water temperature.

Aquatic Conservation: Marine and Freshwater Ecosystems 31:2726–2736. DOI:

10.1002/aqc.3620.

Prchal M, D'Ambrosio J, Lagarde H, Lallias D, Patrice P, François Y, Poncet C, Desgranges

A, Haffray P, Dupont-Nivet M, Phocas F. 2023. Genome-wide association study and

genomic prediction of tolerance to acute hypoxia in rainbow trout. *Aquaculture* 565.

DOI: 10.1016/j.aquaculture.2022.739068.

Quillet E, Dorson M, Le Guillou S, Benmansour A, Boudinot P. 2007. Wide range of

susceptibility to rhabdoviruses in homozygous clones of rainbow trout. *Fish and*

Shellfish Immunology 22:510–519. DOI: 10.1016/j.fsi.2006.07.002.

Van Raaij MTM, Pit DSS, Balm PHM, Steffens AB, Van Den Thillart GEEJM. 1996.

Behavioral strategy and the physiological stress response in rainbow trout exposed to

severe hypoxia. *Hormones and Behavior* 30:85–92. DOI: 10.1006/hbeh.1996.0012.

Rader RB, Belish T, Young MK, Rothlisberger J. 2007. The scotopic visual sensitivity of four

species of trout: A comparative study. *Western North American Naturalist* 67:524–537.

DOI: 10.3398/1527-0904(2007)67[524:TSVSOF]2.0.CO;2.

Roze T, Christen F, Amerand A, Claireaux G. 2013. Trade-off between thermal sensitivity,

hypoxia tolerance and growth in fish. *Journal of Thermal Biology* 38:98–106. DOI:

10.1016/j.jtherbio.2012.12.001.

Ryer C. 2004. Comparison of genetically based antipredator behaviour in hatchery and wild

coho fry from a coastal Oregon river: interpreting a negative result. *Journal of Fish Biology* 64:1413–1418. DOI: 10.1111/j.0022-1112.2004.00348.x.

Rytkönen KT, Vuori KAM, Primmer CR, Nikinmaa M. 2007. Comparison of hypoxia-inducible factor-1 alpha in hypoxia-sensitive and hypoxia-tolerant fish species. *Comparative Biochemistry and Physiology - Part D: Genomics and Proteomics* 2:177–186. DOI: 10.1016/j.cbd.2007.03.001.

Saravia J, Paschke K, Oyarzún-Salazar R, Cheng C-HC, Navarro JM, Vargas-Chacoff L. 2021. Effects of warming rates on physiological and molecular components of response to CTMax heat stress in the Antarctic fish Harpagifer antarcticus. *Journal of Thermal Biology* 99:103021. DOI: 10.1016/j.jtherbio.2021.103021.

Schmidt-Nielsen K. 1997. *Animal Physiology. Adaptation and Environment, 4th edn.* Cambridge. DOI: 10.5962/bhl.title.50838.

Sloman KA, Bouyoucos IA, Brooks EJ, Sneddon LU. 2019. Ethical considerations in fish research. *Journal of Fish Biology* 94:556–577. DOI: 10.1111/jfb.13946.

Staven FR, Nordeide JT, Imsland AK, Andersen P, Iversen NS, Kristensen T. 2019. Is Habituation Measurable in Lumpfish Cyclopterus lumpus When Used as Cleaner Fish in Atlantic Salmon Salmo salar Aquaculture? *Frontiers in Veterinary Science* 6:1–9. DOI: 10.3389/fvets.2019.00227.

Strowbridge N, Northrup SL, Earhart ML, Blanchard TS, Schulte PM. 2021. Acute measures of upper thermal and hypoxia tolerance are not reliable predictors of mortality following environmental challenges in rainbow trout (Oncorhynchus mykiss). *Conservation Physiology* 9:1–16. DOI: 10.1093/conphys/coab095.

Vignet C, Joassard L, Lyphout L, Guionnet T, Goubeau M, Le Menach K, Brion F, Kah O, Chung BC, Budzinski H, Bégout ML, Cousin X. 2015. Exposures of zebrafish through diet to three environmentally relevant mixtures of PAHs produce behavioral disruptions

in unexposed F1 and F2 descendant. *Environmental Science and Pollution Research*

22:16371–16383. DOI: 10.1007/s11356-015-4157-8.

White SL, Wagner T, Gowan C, Braithwaite VA. 2017. Can personality predict individual

differences in brook trout spatial learning ability? *Behavioural Processes* 141:220–228.

DOI: 10.1016/j.beproc.2016.08.009.

Zuur AF, Ieno EN, Walker N, Saveliev AA, Smith GM. 2009. *Mixed effects models and*

extensions in ecology with R. New York, NY: Springer New York. DOI: 10.1007/978-0-

387-87458-6.

Figures :

Fig. 1 Rearing history of fish. Each isogenic line is represented by a different colour. The entire rearing history and experimental use were only represented for lines A22h and N38h. The four other lines were not represented to not overload the figure, but they had a similar life history. Rounded rectangles represent rearing tanks. Replicate rearing tanks also corresponding to replicate behaviour and resistance phenotyping are named A, B and C. The theoretical number of fish is indicated inside each rectangle. The number of fish and lines used in each experiment is indicated in each experiment rectangular, with the first number indicating the number of isogenic lines and the second the number of fish per isogenic line. The lines represent fish movements, solid for fish used in behaviour tests and dotted for fish used in acute hyperthermia and hypoxia challenges. "Hype" stands for acute hyperthermia phenotyping, and "Hypo" for acute hypoxia phenotyping. Figure adapted from Lagarde et al. (2023b).

Fig. 2 Diagram of behaviour tests. a) Tanks used for monitoring individual behaviour. Grey bands represent the safe zone, covered with a lid opaque to visible light. The six fish monitored in parallel were from the same isogenic line. Temperature was kept constant during the 5 minutes of acclimation and 25 minutes of the behaviour test. b) Fish were then transferred to a unique tank for 150 minutes for a group test under moderate hyperthermia stress. Temperature was linearly increased from 12.2°C to 23.0 °C in 2 hours after five minutes acclimation.

Fig. 3 Behaviour variables from individual test, for which *line* effect was statistically significant: a) ACC_MAX – maximal acceleration, b) DIST_TRAV – travelled distance, c) FRQ_CHAN – change of zone frequency, d) RISK% - percentage of time in the risky zone, e) VEL_MAX – maximum velocity. Dark lines represent fits of models. Error bars are standard errors of the model predictions. Means with same letters are not significantly different. Points represent the replicates mean behaviour variables, all time bins considered.

Fig. 4 Behaviour variables from group test, for which *line* was statistically significant: a) ACC_MAX – maximal acceleration, b) DISPERSION – average inter-fish distance, c) DIST_TRAV – travelled distance, d) MOV% – percentage of the time moving, e) VEL_MAX – maximum velocity. Dark lines represent fits of models. Error bars are standard errors of the model predictions. Means with identical letters are not significantly different. For ACC_MAX and DISPERSION, in the absence of interaction between *line* and *time*, points represent the replicates mean behaviour variables, all-time bins considered. For DIST_TRAV, MOV%, and VEL_MAX, behaviour variables for which the best model had a significant interaction between *line* and *time* effects, points represent the replicates' mean behaviour variables at each time bin. ns means there was no significant difference between lines.

Fig. 5 PCA of isogenic lines lsmeans in behaviour tests. A) Individual test. B) Group test. "All bins" means that there was no significant interaction between *line* and *time* effects for the given variable; it was, therefore, possible to estimate *line* lsmeans on the whole test. "Bin 1", "Bin 2", "Bin 3", "Bin 4", and "Bin 5" are the lsmeans estimated at each time bin when there was a significant interaction between *line* and *time* effects. C) Individual and group tests together.

Fig. 6 Correlations between mean isogenic line effects on behaviour traits and mean isogenic line effects

on acute hyperthermia or hypoxia resistances when correlations are higher than |0.7|.

Table 1 Definition of behaviour variables. Test 1 stands for individual behaviour test, and test 2 stands for group test.

Test	Abbreviation	Full name	Unit	Details
1 and 2	ACC_MAX	Maximum acceleration	cm.s^{-2}	The maximum acceleration. Acceleration is the difference in velocity divided by the time difference between the current sample and the previous one.
1 and 2	DIST_TRAV	Distance moved	m	The cumulated distance travelled by the body centre point of the subject from the previous sample to the current one.
1 and 2	MEANDER	Meander	deg.cm^{-1}	Meander is the change in direction of movement of a subject relative to the distance moved by that subject
1 and 2	MOV%	Percentage of the time moving	%	The fraction of time the subject was in moving state, *i.e.* when the subject velocity was above 1.5 cm/s.
1 and 2	VEL_MAX	Maximum velocity	cm.s^{-1}	The maximum velocity. Velocity is the distance moved by the body centre point of the subject per unit of time. The velocity was averaged over 2 frames.
1	EMERGENCE	Latency to exit from the safe zone	s	The time until fish first exit from the safe zone
1	FRQ_CHAN	Zone change frequency	#.min^{-1}	The number of time per minute the subject cross the border between the risky and the safe zone.
1	RISK%	Percentage of the time in the risk zone	%	The fraction of time the subject remains in the risky zone.
2	DISPERSION	Average inter-fish distance	cm	The mean distance between the body centre point of a subject and the body points of all other individuals.
2	FRQ_CONTACT	Body contact frequency	#.min^{-1}	The number of time per minute the subject is in the state "In contact", *i.e.* the subject body contour is in contact with the body contour of any of the other individuals.

Table 2 Best model for behaviour variables in individual and group tests

Behaviour variables	Fixed structure best model
Individual test	
ACC_MAX	*line, time, body weight*
DIST_TRAV	*line*
EMERGENCE	no significant effect
FRQ_CHAN	*line*
MEANDER	no significant effect
MOV%	*time*
RISK%	*line, time*
VEL_MAX	*line, time*
Group test	
ACC_MAX	*line*
DISPERSION	*line*
DIST_TRAV	*line, time, time × line*
FRQ_CONTACT	no significant effect
MEANDER	no significant effect
MOV%	*line, time, time × line*
VEL_MAX	*line, time, time × line*

Table 3 Correlations between isogenic line mean effect (lsmeans) on resistance phenotypes and isogenic line mean effect on behaviour variables. Pearson correlation coefficients above an arbitrary threshold set to |0.7| are highlighted in bold.

Test type	Behaviour variables	Time bin (if interaction)	Acute hyperthermia resistance (min)	Acute hypoxia resistance (min)
Individual test	ACC_MAX		-0.55	0.54
	DIST_TRAV		**-0.71**	**0.90**
	FRQ_CHAN		-0.69	**0.88**
	RISK%		0.19	0.56
	VEL_MAX		**-0.73**	0.69
Group test	ACC_MAX		-0.01	0.40
	DISPERSION		-0.67	**0.74**
	DIST_TRAV	bin 1	-0.20	0.51
		bin 2	-0.34	0.41
		bin 3	-0.30	0.05
		bin 4	-0.17	-0.48
		bin 5	-0.21	-0.51
	MOV%	bin 1	-0.35	0.38
		bin 2	-0.55	0.45
		bin 3	-0.44	0.05
		bin 4	-0.22	-0.42
		bin 5	-0.22	-0.44
	VEL_MAX	bin 1	-0.21	**0.85**
		bin 2	**-0.81**	**0.78**
		bin 3	-0.49	0.46
		bin 4	-0.11	-0.53
		bin 5	-0.27	-0.49