Loss of Nkd1 is dominant over loss of Axin2 in regulating Wnt signaling.

Ian Bell[1*], Haider Khan[1], Nathan Stutt[2], Matthew Horn[1], Teesha Hydzik[1], Whitney Lum[1], Victoria Rea[1], Emma Clapham[1], Lisa Hoeg[3], and Terence J. Van Raay[1*].

1. Department of Molecular and Cellular Biology, University of Guelph, Ontario, Canada.
2. Department of Molecular Genetics, University of Toronto, Ontario, Canada.
3. Department of Bioinformatics, University of Guelph, Ontario, Canada.

* Corresponding Authors: ijbell@outlook.com, tvanraay@uoguelph.ca.

Abstract

Wnt signaling is a crucial developmental pathway involved in early development as well as stem cell maintenance in adults and its misregulation leads to numerous diseases. Thus, understanding the regulation of this pathway becomes vitally important. Axin2 and Nkd1 are widely utilized negative feedback regulators in Wnt signaling where Axin2 functions to destabilize cytoplasmic β-catenin, and Nkd1 functions to inhibit the nuclear localization of β-catenin. Here, we set out to further understand how Axin2 and Nkd1 regulate Wnt signaling by creating *axin2*$^{-/-}$, *nkd1*$^{-/-}$ single mutants and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant zebrafish using sgRNA/Cas9. All three Wnt regulator mutants were viable and had impaired heart looping, neuromast migration defects, and behavior abnormalities in common, but there were no signs of synergy in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants. Further, Wnt target gene expression by qRT-PCR, and RNA-seq analysis and protein expression by mass spec demonstrated that the double *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant resembled the *nkd1*$^{-/-}$ phenotype demonstrating that Axin functions upstream of Nkd1 and that loss of Nkd1 is dominant over the loss of Axin2. In support of this, the data further demonstrates that Axin2 uniquely alters the properties of β-catenin-dependent transcription having novel readouts of Wnt activity compared to *nkd1*$^{-/-}$ or the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant. We also tested the sensitivity of the Wnt regulator mutants to exacerbated Wnt signaling, where the single mutants displayed characteristic heightened Wnt sensitivity, resulting in an eyeless phenotype. Surprisingly, this phenotype was rescued in the double mutant, where we speculate that cross-talk between Wnt/β-catenin and Wnt/Planar Cell Polarity pathways could lead to altered Wnt signaling in some scenarios. Collectively, the data emphasizes both the commonality and the complexity in the feedback regulation of Wnt signaling.

Introduction

Canonical Wnt signaling is important in regulating many events during early development such as initiating the dorsal-ventral axis, maintaining the midbrain-hindbrain barrier, and regulating stem cell differentiation and maintenance[1–3]. Active canonical Wnt signaling has been found to play a crucial role in the development of a majority of the major organs, including the kidneys, liver, and colon[2,4,5]. Dysregulation of Wnt signaling, primarily due to mutations causing the pathway to become hyperactive, have been linked to many diseases including cancer, diabetes, bone density defects and neurological disorders to name a few[6–11].

When Wnt signaling is off, cytoplasmic β-catenin is unstable due to a multi-protein complex termed the destruction complex[12,13]. The destruction complex is composed of Glycogen Synthase Kinase-3β (Gsk3β), Adenomatous Polyposis Coli (Apc), Axin, and Casein Kinase 1α (Ck1α), and functions to phosphorylate and ultimately destabilize cytoplasmic β-catenin[12,13]. Apc and Axin act as scaffolding proteins in the assembly of the destruction complex, whereas Gsk3β and Ck1α are kinases that phosphorylate β-catenin leading to its polyubiquitination and degradation by the proteasome[12,13]. At the transcriptional level, Wnt target genes remain silent due to transcriptional repressors, such as Groucho, binding to Tcf/Lef transcription factors[14].

When Wnt signaling is on, a Wnt ligand binds its receptors Frizzled and Lipoprotein Receptor-Related Protein 5 and 6 (Lrp5/6) which recruits the cytoplasmic protein Dishevelled (Dvl) and the destruction complex to the membrane, resulting in the inactivation of the destruction complex[13]. Once this complex is deactivated, cytoplasmic levels of β-catenin stabilize allowing for the translocalization of β-catenin to the nucleus where it interacts with various coactivators to transcriptionally activate Wnt target genes[15]. Two of these Wnt target genes are *axin2* and *nkd1*, which function as negative feedback regulators for the pathway[16,17].

Nkd1 is an antagonist in both the canonical and non-canonical Wnt signaling pathways[18–20]. At the N-terminal end of Nkd1 is an N-terminal myristoylation sequence which acts as a reversible membrane localization switch. The Nkd1 myristoylation and its localization to the membrane is necessary for its activity in both canonical and non-canonical Wnt signaling[21,22]. There is also an EF-hand in Nkd1 which is most similar to the Recoverin protein EF-Hand[23]. Recoverin is a myristoyl switch protein that changes its membrane localization upon calcium binding to the EF-Hand[24]. While calcium was found not to bind to the Drosophila EF-Hand of Nkd, it is not clear if calcium is required for vertebrate Nkd1 function[25]. Importantly, the EF-Hand of Drosophila is unique from other arthropods and vertebrate Nkd EF-Hands suggesting it has derived a novel function[19]. Nonetheless, the EF-Hand of Nkd1 is important for its ability to antagonize Wnt signaling as the EF-hand has been found to interact with several proteins including Dvl, β-catenin, and p62[22,23,25,26]. Furthermore, Nkd1 interaction with Dvl, via the EF-hand, is also important for its function in non-canonical Wnt signaling as Nkd1 constructs containing mutations in conserved residues in the EF-hand, are still able to rescue a Wnt8 induced eyeless phenotype in zebrafish but have altered non-canonical Wnt signaling[19]. There are other conserved domains including a homology domain that is also speculated to bind Dvl and a C-terminal poly histidine tail that is involved with its interaction with Axin and Axin2[22,26,27]. The model put forth by Gammons et al suggests that the poly-histidine tail is required for Nkd1's aggregation that is dependent on its interaction with Dvl[26]. The aggregation of Nkd1 ensures that it only functions after sustained Wnt signaling[26]. Importantly, Nkd1's function and interaction with β-catenin is Wnt ligand dependent, as Nkd1 was unable to inhibit Wnt signaling induced by ligand-independent mechanisms or in the absence of its myristoylation sequence[20,22,28]. Further, our model suggests that Nkd1 functions downstream of Dvl to inhibit the nuclear accumulation of β-catenin[22].

Loss of Nkd1 in Drosophila results in a naked cuticle phenotype in the larva and in wing patterning defects later in development, both of which demonstrate that Nkd functions to inhibit canonical Wnt signaling[29,30]. In mice, loss of Nkd1 and/or Nkd2 has subtle defects in cranial bone morphology, slightly reduced litter sizes and effects on spermatogenesis, but are otherwise viable[31,32]. In zebrafish, knockdown of Nkd1 results in exacerbated Wnt signaling[20]. Finally, misexpression of Nkd's in multiple systems further demonstrates that Nkd's act as Wnt signaling antagonists[19,20,22,23,28,33]. Curiously, loss of Nkd1 in cell culture experiments has been shown to attenuate canonical Wnt signaling, suggesting that Nkd1 may function to promote Wnt signaling under certain conditions[26,27].

Axin1 and Axin2 serve a similar function acting as scaffolding proteins in the assembly of the destruction complex. They have unique expression patterns with *axin1* being more ubiquitously expressed whereas *axin2* expression is more restricted [34–36]. Furthermore, Axin1 appears to be a more

potent inhibitor of canonical Wnt signaling as overexpression of Axin1 reduces β-catenin protein levels more than overexpression of Axin2 as seen by immunocytochemistry in SW480 cells[17]. However, when Wnt signaling is activated, Axin2 is a more potent antagonist for the pathway as knocking down Axin2, but not Axin1, increased dephosphorylated levels of β-catenin in the presence of the Wnt ligand revealing that Axin1 cannot compensate for the loss of Axin2[17]. Furthermore, both Axin1 and Axin2 have RGS and DIX domains which are important in facilitating polymerization, however, the domains are not identical to one another, allowing the proteins to have unique activity[37,38]. Mutating the DIX domain of both Axin1 and Axin2 inhibits their ability to self polymerize and reduces their ability to antagonize Wnt signaling[38]. Interestingly, the DIX domain of Axin1 is more sensitive to Dvl2 inhibition as chimeric Axin1 proteins, swapping in the DIX domain of Axin2, are no longer inhibited by Dvl2[17]. Also, an independent study found that Axin1, but not Axin2, binds to Dvls[27]. The unique response to Dvl proteins could allow Axin1 and Axin2 to have separate activity when the Wnt pathway is active. The differential importance for Axin1 and Axin2 during development is highlighted in knockout mouse models as loss of Axin1 is lethal whereas loss of Axin2 has no obvious phenotype during early embryonic development[39,40]. Later in development, Axin2 knockout mice have a small head phenotype, delayed heart valve maturation, and thickened aortic valve but are viable and fertile[41,42].

In vitro, it has been shown that Nkd1 interacts with both Axin1 and Axin2[27]. Furthermore, Nkd1 forms a ternary complex with Dvl2 and Axin1. Axin1 only interacted with a high molecular weight aggregate form of Nkd1 with the interaction being abolished by mutating the C-terminal histidine cluster of Nkd1[26]. Nkd1 interaction with Axin1 could be important for its function as knocking down Axin1 in HEK293T cells reduced Nkd1's ability to antagonize Wnt3a activity[27]. Furthermore, HEK293T cells with the C-terminal histidine cluster mutated in Nkd1 had reduced Wnt3a topflash activity revealing a possible positive regulatory function for Nkd1[26]. Collectively, these results suggest that Axins and Nkd1 proteins function at the level of β-catenin to antagonize Wnt signaling.

Current research has revealed that Wnt target gene activation is dynamic depending on the coactivators that interact with β-catenin[43]. In the development of colorectal cancer, there is the "just right" hypothesis for Wnt signaling to provide the "right" conditions for tumour growth[44]. This hypothesis was established by analyzing second hit *APC* mutations found in colorectal adenomas from familial adenomatous polyposis patients with a majority of second hits being point mutations which would disrupt β-catenin binding rather than completely abolishing the proteins activity[44]. Through these experiments we are beginning to understand that different levels of Wnt signaling have different consequences, however, investigation into understanding this concept can be challenging.

Nkd1 and Axin2 are both Wnt target genes and both interact with each other as well β-catenin to inhibit Wnt signaling. Axin2 function is ligand independent and acts to degrade β-catenin while Nkd1 activity is ligand dependent and acts to prevent β-catenin's nuclear accumulation. The interaction between Axins and Nkds is curious given their unique modes of action and it is unclear how they would function together to inhibit Wnt signaling. Alternatively, it could be that the interaction is merely a mechanism to aggregate Wnt signaling components together to nucleate inhibitors[45]. Based on available information, we speculate that these two regulators function independently of one another to provide differential regulation of the pathway, not just on or off as we now know that different levels of β-catenin have different consequences on gene transcription and ultimately cell function. Thus, we hypothesized that compared to both single mutants, the combined loss of Axin2 and Nkd1 would result in a synergistic phenotype, resulting in hyperactivated or hypersensitized Wnt signaling. Counter to our hypothesis, we found that Axin2 and Nkd1 function in a classic dominant/recessive scenario, where Nkd1 functions downstream of Axin2 and loss of Nkd1 is dominant over loss of Axin2.

Results

Knocking out Axin2 and Nkd1.

The Axin2 and Nkd1 knockout lines were created by injecting sgRNA/Cas9 with their respective sgRNAs at the one cell stage and grown to adulthood (P0). These were back crossed into their respective genetic lines and F1 founders were tail clipped and genotyped via indel detection by amplicon analysis followed by sequencing. We evaluated several mutants for each gene, all of which showed similar phenotypes (not shown) and thus we only focused on one genotype for each of *axin2* and *nkd1*. The *axin2*$^{-/-}$ zebrafish were made in the tail long background (TL) whereas the *nkd1*$^{-/-}$ zebrafish were generated in the Tubingen (TU) background. The *axin2*$^{-/-}$ mutant has a 12-nucleotide deletion with a 1 nucleotide insertion in exon 2 leading to a frameshift mutation and premature stop codon (Supplemental Fig 1). The *nkd1*$^{-/-}$ mutation is a 16-nucleotide deletion in exon 3 leading to a frameshift mutation and premature stop codon (Supplemental Fig 2). The *axin2*$^{-/-}$;*nkd1*$^{-/-}$ zebrafish were created by crossing individual *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants generating double heterozygous *axin2*$^{+/-}$; *nkd1*$^{+/-}$ mutants that were then incrossed and genotyped as above to identify homozygous *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants.

For reference, in the figure legends, capital "N" refers to the number of biological replicates, whereas lowercase "n" refers to the total number of observations. In addition, all mutants discussed below are maternal-zygotic mutants unless otherwise specified.

Wnt regulator mutants have developmental defects.

Our first objective was to determine if the different mutants had developmental defects and to report on those that we consistently observe. While some of the experiments reported below have only 1 biological replicate, all have been observed multiple times and in different genetic backgrounds. We report only on the ones where we have included all the relevant controls.

The Wnt regulator mutants have no overt phenotype, appear phenotypically wild type until early adult hood (see below), and are viable. Upon close examination it was demonstrated that the mutants have heart looping defects (Fig. 1A). Specifically, at 2 days post fertilization (dpf), wild type larvae had D looped hearts 90.6% of the time, whereas, the *axin2*$^{-/-}$, *nkd1*$^{-/-}$, and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ had D looped hearts 41.3%, 53.8%, and 26.9% of the time, respectively. In each of the Wnt regulator mutants there was an increase in proportion of D incomplete looped, L looped, L incomplete looped, and unlooped hearts when compared to wild type. Since heart looping phenotypes can be variable based on the genetic background, we injected wild type embryos with sgRNA/Cas9 targeting both Axin2 and Nkd1 and observed a similar proportion of heart looping phenotypes compared to the knockout lines (Fig. 1B). These results support research on Wnt signaling in heart development where *apc* mutant zebrafish also develop with unlooped hearts, although at a much greater proportion[46]. Furthermore, past research has demonstrated that knock down of Nkd1 with morpholinos impairs heart development by inducing a heart jogging defect[33].

We also examined other known Wnt dependent events for signs of activated Wnt signaling. Wnt signaling is necessary for neuromast formation and migration where it works alongside FGF signaling for proper patterning[47–49]. Hyperactivation of Wnt signaling has previously been shown to reduce neuromast migration in zebrafish which lead us to investigate if neuromast migration is impaired in the Wnt regulator mutants (Fig. 1C)[47]. At 5dpf, wild type zebrafish had 5-7 terminal neuromast, whereas, *axin2*$^{-/-}$, *nkd1*$^{-/-}$, and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants had 4-6, 4-7, and 0-6 terminal neuromast, respectively. To confirm that hyperactivation of Wnt causes reduced neuromast migration, wild type embryos were treated with a small molecule Wnt activator, 6-bromoindirubin-3'-oxime (BIO), which caused larvae to have 0-4 terminal neuromasts at 5dpf (Fig. 1D). Total number of neuromasts for all Wnt regulator

mutants did not change (data not shown) suggesting that the migration of the neuromasts are perturbed, consistent with published reports[47].

In the adults, the *axin2*$^{-/-}$ zebrafish occasionally (~20-50%) develop a curved spine phenotype, which is not surprising as Wnt signaling is important in bone formation where both hyperactivation and hypoactivation of the pathway leads to skeletal deformities[40,50,51]. Furthermore, Axin2 knockout mice have axial skeletal defects and a small head phenotype[40]. To determine if there was a maternal effect, we performed an incross using *axin2*$^{+/-}$ zebrafish followed by phenotyping and genotyping. The incross resulted in 42 adults, of which, 2/6 *axin2*$^{-/-}$, 0/25 *axin2*$^{+/-}$, and 0/11 *axin2*$^{+/+}$ developed with a curved spine phenotype (Fig. 1E-F). The frequency was similar to the maternal-zygotic mutants suggesting that there is no maternal contribution to this phenotype. The *axin2*$^{-/-}$;*nkd1*$^{-/-}$ zebrafish also develop a curved spine, albeit with less frequency, whereas *nkd1*$^{-/-}$ single mutants do not (data not shown).

Further, the *axin2*$^{-/-}$ adult zebrafish often develop with receding opercula (Fig. 1G-H). Like above, *axin2*$^{-/-}$ mutants from an incross of *axin2*$^{+/-}$ had complete penetrance (6/6) of receding opercula in the one cross that we performed whereas *axin2*$^{+/-}$ and axin2$^{+/+}$ had 9/25 and 2/11 with receding opercula, respectively, suggesting that there may be a dosage effect (Fig 1H). Operculum malformation is common in stressed or old age zebrafish, however, *axin2*$^{-/-}$ mutants consistently develop with a receding operculum at an early age (~5 months). The *axin2*$^{-/-}$;*nkd1*$^{-/-}$ often have receding opercula at a frequency seen in the *axin2*$^{-/-}$ mutants, whereas, *nkd1*$^{-/-}$ mutants do not (data not shown).

The varying degree in the number of terminal neuromasts suggests that the Wnt regulator mutant larva might have altered responses to stimulation. Therefore, we assessed 5dpf Wnt regulator mutants for swimming behavior and tap response. The larvae were recorded for 1.5 hours under dark conditions using a 24 well plate and assayed for thigmotaxis. We discovered that the wild type larvae had a greater thigmotactic behavior when compared to the Wnt regulator mutants. Furthermore, while wild type larvae spent 82.6% of their time along the edge of the well, *axin*2$^{-/-}$, *nkd1*$^{-/-}$, and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants spent 70.6%, 70.9%, and 75.4%, of their time along the edge, respectively. Startle response was measured for 30 taps over a period of 30 seconds with the first three taps in the analysis showing the greatest difference in response when comparing the Wnt regulator mutants to wild type larvae. The *axin2*$^{-/-}$ larvae had a mild reduction in startle response whereas the *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ larvae had a severe reduction in startle response when compared to wild type (Supplemental Fig. 3).

Wnt regulator mutants have differential sensitivities to exogenous Wnt8.

While Axin2 and Nkd1 are both negative feedback regulators of Wnt signaling acting at the level of β-catenin, we have previously demonstrated that exogenous Nkd1 is only active in the presence of ligand activated Wnt signaling[28]. In contrast, Axin2 does not require the Wnt ligand for its activation[28]. Therefore, we wanted to assess the response of the regulator mutants to overactive Wnt signaling. Overexpression of *wnt8* leads to an eyeless phenotype that is easily visible at 1dpf (Fig. 2A-B). Given that both Axin2 and Nkd1 are negative feedback regulators, we hypothesized that loss of these regulators would exacerbate the eyeless phenotype induced by Wnt8 overexpression. Injection of a low dose of *wnt8* where only 4.7% of embryos are eyeless in wild type, increased to 33.8% and 31.6% in *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants respectively, supporting our hypothesis (Fig. 2C). We further expected the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants to be even more sensitive than either of the single mutants but surprisingly, only 4.6% of *axin2*$^{-/-}$;*nkd1*$^{-/-}$ larvae were eyeless, suggesting the loss of one regulator rescues the effect of the loss of the other (Fig. 2C). Furthermore, *nkd1*$^{-/-}$ injected mutants had a large proportion of embryos develop with a short-twisted axis when *wnt8* was overexpressed, which was rarely seen in either *axin2*$^{-/-}$ single or *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants (Fig. 2D-F). This suggests that loss of Axin2 can rescue loss of Nkd1 in the short- twisted axis phenotype, but it is unknown if this is the case in the eyeless phenotype.

To further explore the effect of knocking out Axin2 and/or Nkd1 on Wnt activity we utilized the Wnt reporter zebrafish line (7XTCF:GFP)[52]. We injected *axin2* and/or *nkd1* sgRNAs along with Cas9 protein into one-cell stage transgenic embryos derived from the Wnt reporter line (Crispants). Wnt activity via GFP expression in the brain was then analyzed at 1dpf. Uninjected wild type embryos predominantly have GFP expression at the midbrain-hindbrain boundary with some activity being present in the forebrain (Fig. 2G). Knock down of Axin2 caused an anterior-ward expansion in GFP expression in the forebrain, whereas knock down of Nkd1 only caused a mild increase in GFP expression in the forebrain. When both Axin2 and Nkd1 were knocked down there was a similar expansion in GFP expression as seen in the *axin2* crispants alone, suggesting the double crispant is more similar to the *axin2* crispant but the variability in the data precludes statistical testing. To further explore this, we overexpressed *wnt8,* which caused *axin2* crispants to have a major anterior expansion in GFP expression which was milder in the *nkd1* or *axin2*;*nkd1* crispant embryos (Fig. 2G). Importantly, the eyeless phenotype induced in this transgenic line mimicked the knockouts where the single crispants had increased sensitivity while the double crispants showed the same insensitivity to exogenous Wnt8 (data not shown).

Thus far, loss of either Axin2 or Nkd1 produces results that are consistent with their roles as negative feedback regulators; however, things get confusing when both Axin2 and Nkd1 are simultaneously knocked out, especially in response to exogenous Wnt8. Therefore, we wanted to test how these mutants responded when Wnt signaling was overactivated in a ligand independent manner. To accomplish this, we used the small molecule BIO, which inhibits Gsk3β activity (Fig. 3A,B)[53]. Wild type embryos treated with 0.5µM of BIO reduces the size of the eye approximately 45.7% (±19.9%) (Fig. 3C). Using the same dose of BIO, *axin2*$^{-/-}$, *nkd1*$^{-/-}$, and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant eyes were reduced by 69.5% (±21.6%), 96.1% (±3.9%), and 89.4% (±0.42%), respectively (Fig. 3C, supplemental Fig. 4). To ensure that the sensitivity to BIO was not a consequence of the different genetic backgrounds we created *axin2*$^{-/-}$ mutants from an *axin2*$^{+/-}$ incross. Here, *axin2*$^{-/-}$ embryos treated with BIO had an increase in the eyeless phenotype when compared to embryos with at least one copy of wild type Axin2 from the same cross demonstrating that the sensitivity is specific for loss of Axin2 (Supplemental Fig. 5). In stark contrast to the effect of ectopic Wnt8, the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant was highly sensitive to BIO. This suggests that a ligand, or receptor proximal, dependent event is somehow inhibiting Wnt/β-catenin signaling in the absence of two intracellular negative feedback regulators during early gastrulation when canonical Wnt signaling is patterning the hindbrain[54].

Wnt regulator mutants have differential effects on Wnt target genes.

The data so far suggests that regulation of Wnt by negative feedback regulators is a complex process. To further explore the effect of Wnt8 and determine if we could separate out how these two regulators function to restrict Wnt signaling, we analyzed the expression of several well characterized Wnt target genes. At 30% and 50% epiboly *axin2* expression is restricted to the ventral lateral domain of the developing zebrafish embryo. Overexpression of Axin1 abolished *axin2* expression at 30% and 50% epiboly, whereas hyperactivating Wnt signaling using a stable form of β-catenin (ΔN-β-catenin) causes ectopic *axin2* expression (Supplemental Fig. 6). Combined with published results, this confirms that Axin2, along with Nkd1 and Sp5a are excellent markers for active Wnt signaling during pre-gastrulation development[55–58].

We first used qRT-PCR to quantify the expression of the Wnt target genes *axin2*, *nkd1*, and *sp5a* at 30% and 50% epiboly (when zygotic Wnt signaling is high) with or without overexpression of *wnt8* in the different genetic backgrounds (Fig 4)[20,59,60]. We did not observe a decrease in endogenous *axin2* or *nkd1* expression in any of the Wnt regulator mutants, suggesting that the mutations in *axin2* and *nkd1* are not inducing nonsense mediated decay. In the *axin2*$^{-/-}$ mutants, there was a predictable, albeit slight,

increase in all three target genes that was exacerbated in the presence of exogenous Wnt8 at both 30% and 50% epiboly. Curiously, only the expression of *axin2* in the *axin2*$^{-/-}$ mutants was significant at 30% epiboly (Fig. 4A,B). Conversely, the *nkd1* $^{-/-}$ mutants displayed a somewhat different pattern, as there was no substantive increase in either *axin2* or *nkd1* expression at 30% with or without exogenous Wnt8 when compared to controls (Fig. 4C). At 50%, the expression of *axin2* and *nkd1* in *nkd1*$^{-/-}$ was similar to the *axin2*$^{-/-}$ mutant in that there was a slight, but non-significant increase in their expression that was moderately enhanced by exogenous Wnt8 (Fig. 4D). Interestingly and curiously, *sp5a* expression in uninjected *nkd1*$^{-/-}$ mutants was not significantly impacted but skyrocketed in the presence of exogeneous Wnt8 at 30% epiboly (Fig. 4C). Furthermore, at 50% epiboly, uninjected *nkd1*$^{-/-}$ mutants had a significant increase in *sp5a* expression that was further exacerbated by the addition of exogenous Wnt8 (Fig. 4D). Importantly, the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant closely resemble the *nkd1*$^{-/-}$ mutants with respect to all three genes (Fig. 4E). This strongly suggests that each single mutant has a unique consequence on the activation of Wnt target genes and that loss of Nkd1 might be dominant over the loss of Axin2.

Wnt regulator mutants have differential effects on *gsc* expression.

To explore this differential effect further, we chose to look at the expression of *gsc* by whole mount *in situ* hybridization (WMISH) to determine how the dorsal organizer is affected by loss of Wnt feedback regulation. The dorsal organizer is under the influence of both maternal and zygotic canonical Wnt signalling[61-62]. From dome to 50% epiboly, the arc of *gsc* expression on the dorsal side of the embryo can be quantified as a read out for Wnt activity. This would also determine if there is an expansion of Wnt signaling or ectopic Wnt signaling. At dome stage and 30% epiboly there was no significant difference in the arc of *gsc* expression in the *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants when compared to wild type (Fig. 5A,B). In contrast, at 50% epiboly, the *axin2*$^{-/-}$ embryos had an 8.1° increase in *gsc* arc angle which was not seen in either the *nkd1*$^{-/-}$ or *axin2*$^{-/-}$;*nkd1*$^{-/-}$ embryos (Fig. 5C,D). This was also validated by knocking down Axin2 using morpholinos which caused an increase in *gsc* arc angle at 50% epiboly but not at 30% epiboly (Supplemental fig. 7). This is in contrast to previous research from our lab where knocking down Nkd1 using a morpholino increased *gsc* expression at 30% epiboly[20]. This may be a consequence of knocking down translation via morpholino versus knocking out maternal and zygotic transcription in the *nkd1*$^{-/-}$ or *axin2*$^{-/-}$;*nkd1*$^{-/-}$ knockouts shown here.

This suggests that Axin2, but not Nkd1, normally functions to restrict the expression of *gsc*. To determine if there was a maternal contribution in the *axin2*$^{-/-}$, we overexpressed *wnt8* in embryos that were generated from a double heterozygote incross, processed for WMISH, scored for phenotype, and then genotyped. Unlike the maternal-zygotic mutants above, these embryos would only be zygotic *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants. From the *axin2*$^{+/-}$;*nkd1*$^{+/-}$ incross, embryos with Axin2 knocked out (*axin2*$^{-/-}$;*nkd1*$^{+/-}$) had a 14.4° increase in *gsc* arc angle, whereas, embryos with Nkd1 knocked out (*axin2*$^{+/-}$;*nkd1*$^{-/-}$) had a 6.6° increase in *gsc* arc angle when compared to embryos with at least one wild type allele of both Axin2 and Nkd1 (*axin2*$^{+/-}$;*nkd1*$^{+/-}$) , both of which were significant (Fig. 5E). This suggests that the effect on *gsc* expression appears to be independent of maternal sources of Axin2 as the maternal zygotic and the zygotic mutants generated similar results.

The evaluation of *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants was hindered by the low numbers that were available, as only 1/16 of all offspring from a double heterozygous cross would be *axin2*$^{-/-}$;*nkd1*$^{-/-}$. Therefore, to increase the number of *axin2*$^{-/-}$;*nkd1*$^{-/-}$ embryos, *wnt8* overexpression was performed on embryos from a *axin2*$^{+/-}$;*nkd1*$^{-/-}$ incross. Fifty percent of these embryos would be zygotic *axin2*$^{-/-}$ and maternal-zygotic *nkd1*$^{-/-}$. Similar to the maternal-zygotic mutants experiments above (Fig. 5A-D), *wnt8* injected *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants did not have an increase in *gsc* expression when compared to both single knockout genotypes overexpressing *wnt8* (Fig. 5F). Collectively, this suggests that loss of Nkd1 rescues the loss of Axin2 or that loss of Nkd1 is dominant over loss of Axin2.

The evidence thus far suggests that there are at least 3 different responses in the loss of the negative feedback regulators: 1) Additive: terminal neuromasts are more severe in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant; 2) Counteractive: *axin2*$^{-/-}$;*nkd1*$^{-/-}$ rescues Wnt8 overexpression in single mutants and 3) Nkd1 dominant: *sp5a* expression is up in *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants but not in *axin2*$^{-/-}$; *gsc* and *axin2* expression up in *axin2*$^{-/-}$, but there is no change in *nkd1*$^{-/-}$ or *axin2*$^{-/-}$;*nkd1*$^{-/-}$. Further, we only observed the counteractive response in the eyeless phenotype, suggesting that this may be related to other signaling phenomenon occurring during patterning of the eye field.

RNA-seq analysis suggests *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants are more like *nkd1*$^{-/-}$.

To better understand the relationship between these two Wnt regulators, we performed RNA-sequence analysis on embryos at 50% epiboly, a time point when we observed significant effects on gene expression in the mutants. In addition to the overall analysis in gene expression, we also attempted to identify groups that reflect the three different responses identified above.

RNA-sequencing was performed on the Wnt regulator mutants as well as the two wild-type background genotypes: TU and TL. The two genetic backgrounds were useful in identifying and removing genetic variability unrelated to the mutations. Three biological replicates were isolated on different days, but from the same parents for each genotype. RNA-sequencing was performed using paired-end reads at a depth of 50 million reads.

A principal component analysis was performed on all genes. This revealed that all biological replicates were closely related to each other demonstrating the reproducibility of the dataset. Further, the *axin2*$^{-/-}$ mutants were more similar to the TL background from which it was derived, while the *nkd1*$^{-/-}$ mutants clustered closely with the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants (Fig. 6A). Furthermore, a heat map of the top 1000 differential regulated genes also shows a similar trend where *axin2*$^{-/-}$;*nkd1*$^{-/-}$ embryos are more similar to *nkd1*$^{-/-}$ (Fig. 6B). This data fits very well with the *sp5a* expression we observed above. To further explore the data, Venn diagrams were created to identify overlap in differentially expressed genes (DEG) found in each of the Wnt regulator mutants. Before the Venn diagrams were created, genes that were differentially expressed (FDR < 0.05) between TU and TL were removed to strengthen the reliability of our data by removing genes that were variable between the background genotypes. Using an absolute log2FC >2 and FDR < 0.05 filtering method revealed 735 DEG in the *axin2*$^{-/-}$ mutants, 869 DEG in *nkd1*$^{-/-}$ mutants and 312 DEG in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ (Fig. 6C,D). We attributed the reduced number of DEG in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants to the fact that they were filtered against both the TU and the TL backgrounds, while the single mutants were only filtered against their respective backgrounds.

From this, we conclude that the clustering of the *nkd1*$^{-/-}$ mutants with *axin2*$^{-/-}$;*nkd1*$^{-/-}$ and both of these away from the *axin2*$^{-/-}$ mutants is not due to the number of genes that are affected. Instead, it may be due to gene expression levels found in the mutants. Although not significant, a comparison of the expression values of the 34 upregulated genes shared in all mutant genotypes demonstrated that loss of Nkd1 had a greater impact on gene expression levels compared to loss of Axin2 (Supplemental fig. 8). A similar trend was observed in the 15 genes they had in common that are downregulated. Importantly, this was only observed for the shared genes, as comparison of expression levels of all the DEG for each genotype revealed nearly identical violin plots (Supplemental fig. 8C).

While both single mutants each have several hundred DEGs we were still perplexed by the lack of overlap in the number of shared genes. We wanted to determine if this was a consequence of reduced expression levels of genes in the *axin2*$^{-/-}$ that were not shared. That is, do the 735 genes (outside of the 34 shared ones) found to be differentially expressed in *axin2*$^{-/-}$ have differential expression in the *nkd1*$^{-/-}$ mutant but just don't reach the threshold of significance? To test this, we evaluated the raw expression levels of the 735 DEG identified in the *axin2*$^{-/-}$ mutants in the *nkd1*$^{-/-}$ dataset. For comparison, we also performed the reciprocal analysis (Supplemental Fig. 9). This analysis

demonstrates that the non-overlapping DEG in the *axin2*$^{-/-}$ remain relatively unchanged in the *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants (Supplemental fig. 9A,B). This suggests that modifications to the transduction of the Wnt signal in the *axin2*$^{-/-}$ mutant are very different to the modifications that occur with the loss of Nkd1 and supports the notion that these modifications are dominant over the *axin2*$^{-/-}$ modifications.

Of the 34 common genes that were upregulated and 15 common genes that were downregulated in all the Wnt regulator mutant genotypes, a large portion (28) of the 49 differentially expressed genes were uncharacterized with the vast majority of these (11/15) being downregulated (Fig. 6E).

To determine if the RNA-seq data supports the "additive" response group, we identified genes that were up (or down) in both *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants by Log2FC > 1 whose expression was exacerbated up (or down) in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants. This identified one gene, *Prkag2b*, that had a 5.5 fold change in both *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants and a 7.0 and 7.2-fold increase in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ vs TU and vs TL, respectively. There were 8 genes (Supplemental table 1) that were additive in their reduced expression.

To determine if there was evidence for the "counteractive" response, we identified genes that were up (or down) by Log2FC > 1 in both the single mutants (same direction) but were either unchanged or down (or up) by less than 1-fold in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$. This identified seven genes (Supplemental table 2), all of which had unchanged expression in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$.

To determine how many genes were in the "Nkd1 dominant" group, we selected genes that were differentially expressed by an absolute Log2FC > 1 in both the *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants, but were unchanged in the *axin2*$^{-/-}$ mutants. This identified 237 genes. Increasing the stringency to absolute Log2FC > 2 resulted in 74 genes and a further increase to absolute Log2FC > 3 resulted in 33 genes (Supplemental table 3). Thus, while there is no significant support for "additive" or "counteractive" responses in the RNA-seq data, there is clearly support for the Nkd1 dominant response. We also determined if there was an "Axin2 dominant" group that was differentially expressed in *axin2*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ by an absolute Log2FC > 1 , but were unchanged in the *nkd1*$^{-/-}$ mutant and identified only five genes (Supplemental table 4).

Wnt target gene expression in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants is more similar to *nkd1*$^{-/-}$ than *axin2*$^{-/-}$.

Analysis of the 237 "Nkd1 dominant" genes by GO and Panther analysis revealed no significant hits, regardless of the stringency we used. Curiously, only 3 of the 237 genes were related to Wnt signaling: *gsk3b*, *fzd10* and *caveolin*. We performed similar GO analyses on each of the three mutant genotypes and in each case, there were no significant hits. Analysis of the non-significant hits did not reveal any patterns; however broad neuronal systems were abundant. Surprisingly, Wnt signaling did not appear in any of the GO analyses which could be due to the analysis focusing on the proteins in the pathway rather than the expression of Wnt target genes (data not shown).

Despite the high levels of *sp5a* identified in our qRT-PCR analysis, this gene was not identified as being a significant hit in the RNA-seq data. Thus, we wanted to investigate this gene and other known Wnt target genes. Using the RNA-seq data, a list was created of 59 known Wnt target genes identified by the Wnt signaling community. It should be noted that many of these are not unique to Wnt signaling and may have a more significant role in other pathways. Of the 59 Wnt target genes, 15 of them had differential expression in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant embryos (FDR < 0.05 vs TU and FDR < 0.05 vs TL). Of these 15 differentially expressed genes, nine of them had similar expression to *sp5a* seen earlier by qRT-PCR: *fsta, lef1, nkd1, six3b, sox17, sox2, sp5a, tbx3a, and tiam1a* (Fig. 7). The inclusion of *sp5a* in the list is validation of the qRT-PCR data. Furthermore, the levels of *axin2* and *nkd1* in the RNA-seq data are also

consistent with our qRT-PCR analysis, showing modest but consistent increases in expression of all three mutant genotypes.

Collectively, the differential gene expression analysis suggests that loss of Nkd1 has a greater impact on gene expression levels compared to the loss of Axin2 and that there is no additive, synergistic or counteractive effect in the loss of both Nkd1 and Axin2. While the GO analysis did not reveal any significant hits, there was a modest, but consistent effect on Wnt target genes. Further, it seems that there are a significant number of uncharacterized genes that are targets of the Wnt pathway that have yet to be described, highlighting the complexity and depth of this pathway in multipotent progenitor cells.

The lack of overlap in target genes between *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ (only 73 genes out of a combined 1,531) is curious but does support our qRT-PCR and *gsc* expression analysis. This suggests that each contribute to modifications of β-catenin-mediated signaling resulting in an altered transcriptional response, which may include alternative transcriptional start sites in their shared genes. To explore this further, we evaluated differential transcript usage (DTU) in the different genetic backgrounds but found no obvious pattern for DTU when comparing the Wnt regulator mutants. Several of the most significant hits are shown in supplemental figure 10. Accordingly, we did not pursue this further.

Knocking out Axin2, but not Nkd1, increases cytoplasmic levels of β-catenin.

As both Axin2 and Nkd1 interact with β-catenin in the cytoplasm, we next wanted to determine if there were changes in the levels of cytoplasmic and nuclear β-catenin. As Axin2 is predicted to target β-catenin for ubiquitin mediated degradation, we predicted that loss of Axin2 would result in more cytoplasmic β-catenin. In contrast, we have previously demonstrated that Nkd1 functions to inhibit the nuclear accumulation of β-catenin and so its loss is predicted to result in more nuclear β-catenin but not necessarily a change in cytoplasmic levels[22]. To test this, we first performed Western blot analysis on cytoplasmic fractions from 30% epiboly embryos, a time point where we have previously shown cytoplasmic β-catenin levels to be sensitive to Wnt signaling hyperactivation[22]. Further, Pan-cadherin was used as a control to ensure that the cytoplasmic fractions were free of plasma membrane as β-catenin is localized to the membrane due to its interactions with cadherins[63]. Compared to uninjected controls, wild type embryos had a relative cytoplasmic β-catenin increase of 1.86 when overexpressed with *wnt8*, which is consistent with our previous reports (Fig. 8A)[22]. Analysis of β-catenin levels in the Wnt regulator mutants demonstrated a significant increase in *axin2*$^{-/-}$ mutants. In particular, *axin2*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ had a relative cytoplasmic β-catenin increase of 2.90 and 3.52, respectively, compared to their uninjected genotype control (Fig 8B). These were significantly higher than the 1.86 increase seen in wild type *wnt8* injected embryos. By comparison, the *nkd1*$^{-/-}$ mutant had a relative cytoplasmic β-catenin increase of 2.17, which was not significant when compared to wild type (Fig. 8B) but supports the higher levels seen in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant. This supports the model that Axin2 affects cytoplasmic levels of β-catenin[17,22,31]. While the statistics support our model that loss of Nkd1 does not alter cytoplasmic β-catenin levels, we are not convinced that there is no change in *nkd1*$^{-/-}$ mutants, especially when *axin2*$^{-/-}$;*nkd1*$^{-/-}$ shows a modest additive effect.

Wnt regulator mutants have no change in nuclear β-catenin levels at 30% epiboly.

With the increase in cytoplasmic β-catenin in at least the *axin2*$^{-/-}$ mutants, we next wanted to evaluate the levels of β-catenin in the nucleus. In our experience, nuclear fractions cannot be sufficiently purified from β-catenin-containing membrane fractions in zebrafish embryos for Western blot analysis, therefore we chose to perform immunohistochemistry at 30% epiboly using anti-β-catenin

antibodies[20,22]. We first evaluated nuclear β-catenin levels in the different genetic backgrounds along the ventrolateral domain which has endogenous Wnt signaling and nuclear β-catenin. In contrast, cells outside of the ventrolateral domain were assessed for potential ectopic nuclear β-catenin. While difficult to quantify levels between embryos, we did not observe any obvious differences in nuclear β-catenin levels when compared to wild type embryos (Supplemental fig. 11).

Knocking down Axin2 and/or Nkd1 does not increase nuclear β-catenin levels when Wnt signaling is hyperactive.

We next set out to evaluate nuclear β-catenin in the presence of exogenous Wnt8. To do this, we injected a cocktail of Axin2 and/or Nkd1 sgRNAs, Cas9, and *wnt8* along with a tracer molecule (Dextran or *mcherry*) into 1 cell of an 8-cell stage wild type blastocyst to generate mosaic crispants. Embryos were fixed at 30% epiboly and immunohistochemistry was performed with anti-β-catenin antibodies. Images were captured away from the ventrolateral domain to avoid any influence from endogenous Wnt signaling. Using this design, labelled cells would be Wnt8 and crispant positive, but juxtaposed cells would be crispant negative, but still receive the Wnt ligand[22]. Thus, we compared nuclear β-catenin levels in label-positive cells with juxtaposed label-negative cells. Knocking down Axin2 and/or Nkd1 did not have any significant difference in nuclear β-catenin levels when compared to uninjected juxtaposed cells (Fig. 9). This is in contrast to previous findings where knocking down Nkd1 in the dorsal forerunner cells of zebrafish, and knocking out Axin2 in mice, caused an increase in nuclear β-catenin[33,64]. It is possible that our assay is not sensitive enough to detect subtle differences considering we are knocking down regulators of the pathway and not essential components. It is also possible that the sgRNAs were ineffective, but the same sgRNA were used in the Wnt reporter analysis with success as well as their efficiency being validated by indel detection by amplicon analysis (not shown). Taken together, loss of Wnt regulation via Axin2 and Nkd1 results in an increase in cytoplasmic β-catenin (at least significantly for Axin2), but this does not translate into a measurable increase in nuclear β-catenin.

Wnt regulator mutants have lower expression of metabolism proteins.

The differences that we have observed in phenotypes, gene expression and cytoplasmic β-catenin levels suggests that Axin2 and Nkd1 have both unique and overlapping ways of controlling Wnt signaling and that this might be reflected in the protein complement in each of the mutants. We first tried to purify β-catenin to identify proteins that might be differentially interacting with it under the different genetic conditions; however, we were unable to purify sufficient levels of β-catenin from zebrafish embryos to perform this analysis. Instead, we decided to evaluate the entire protein pool using mass spectrometry with 30% blastocysts that have been rigorously purified away from yolk proteins. Using this strategy, we identified differentially expressed proteins between the Wnt regulator mutants. These results were variable between biological replicates which, despite our best efforts, was likely due to yolk contamination during isolation. Therefore, we chose samples with similar ionic chromatograms to perform our analysis (Fig. 10A). From this, we discovered 53 differentially expressed proteins (FDR < 0.1, logFC ≥ 1, peptides ≥ 2) in the Wnt regulator mutants with the majority of the proteins being either ribosomal subunits or related to metabolism (Fig. 10B). Furthermore, and similar to the RNA-seq data, we see that the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant is more similar to the *nkd1*$^{-/-}$ mutant, whereas the *axin2*$^{-/-}$ mutant is similar to wild type; however, the proteins that were differentially expressed in mass spectrometry did not always correlate to differential expression in the RNA-seq results. Furthermore, in the *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants, there is an overall trend of decreased protein expression levels. Of the 53 proteins, there were 18 proteins (Fabp3, Aldh2.2, Prdx1, ldhba, Pcxa, Tcp1, Igf2bp3, Pkma, Actb2, Cct5, Prdx6, Tpi1a, Prdx2, Ppa1b, Sod1, Cox17, Ranbp1, Stmn1a) that have a connection to Wnt signaling (Fig.

10C). Interestingly, most of the proteins related to Wnt have been demonstrated to stimulate Wnt signaling, as knocking out/down the proteins leads to either reduced β-catenin levels or Wnt target gene expression. Finally, using string analysis, we identified the top KEGG pathways to be ribosomal proteins and pyruvate metabolism related proteins (Fig. 10D-E). While this technique requires refinement to generate high quality data it does support our model where Axin2 and Nkd1 have both overlapping and unique effects in regulating Wnt signaling.

Discussion

Our initial hypothesis was that Axin2 and Nkd1 uniquely and independently affect Wnt/β-catenin signaling with Axin2 regulating the stability of cytoplasmic β-catenin, while Nkd1 prevents its nuclear accumulation[17,22]. We further anticipated a lack of a significant phenotype in each of the single mutants, based on the phenotypes in their mammalian counterparts[31,32,40,42]. Nonetheless, we speculated that with the different modes of action, there would be a convergence on the transcriptional targets and we anticipated a synergistic effect in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants resulting in more severe phenotype. Surprisingly, the phenotypic data was underwhelming. The heart looping data showed no evidence of epistasis or synergy. The axial and operculum defect in the *axin2*$^{-/-}$ single and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants suggests that Axin2 functions downstream of Nkd1, however, a detailed analysis clearly demonstrates that the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants resemble the *ndk1*$^{-/-}$ mutants. This strongly suggests that Nkd1 functions downstream of Axin2 and that loss of Nkd1 is dominant to loss of Axin2.

Evaluation of the entire transcriptome in a whole organism is a double edge sword. On one hand, we are evaluating the loss of genes in their native environment, which is very powerful. On the other hand, not every cell is actively engaged in Wnt signaling which can generate significant background. To maximize the former and minimize the later, we chose to evaluate the transcriptional readout of the mutants at 50% epiboly as there is substantial zygotic Wnt signaling occurring throughout the ventrolateral domain of the blastula. The transcriptional data supports Axin2 having a separate mode of Wnt antagonism from Nkd1. However, we don't have any evidence that loss of Axin2 is dominant over loss of Nkd1 or that Axin2 functions downstream of Nkd1. In contrast, the majority of our analysis supports the model that Nkd1 functions downstream of Axin2 and its loss is dominant over loss of Axin2 (Table 1).

The difficulty in working with negative feedback regulators is that they function to regulate the intensity, spatial distribution and/or duration of the signal and can act as a safety mechanism more than an integral component of the pathway, much like an airbag in a car[65]. For example, we see a more robust response in *sp5a* expression when we ectopically express Wnt8 in the mutants compared to wildtype. Nonetheless, we still observed significant increases in target gene transcription without the addition of exogenous Wnt8. We also speculate that the high number of DEG's identified is an overestimation of the actual targets. Increasing the absolute Log2FC to >3 resulted in 33 genes that are significantly differentially transcribed in both the *nkd1*$^{-/-}$ single and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutants but remained unchanged in the *axin2*$^{-/-}$ mutant. One, Gsk3βb, is linked to Wnt signaling and is upregulated over 60 fold; two are related to transcriptional activity; one is a histone deacetylase and nine are uncharacterized proteins with the remainder having various unrelated functions. Similarly for *axin2*$^{-/-}$ mutants with an absolute Log2FC>1 but remained unchanged in the *nkd1*$^{-/-}$ or *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants resulted in 52 genes, 1 of which is related to Wnt signaling (*sec12l8*); 9 are related to DNA binding; 4 are related to histone function; 10 are uncharacterized proteins and the remainder have various unrelated functions (Supplemental Table 5). The significant hits in DNA binding and histone modifications are in line with the model that loss of Axin2 and/or Nkd1 would alter Wnt signaling transcriptional activity, but this requires further investigation.

While the data is consistent with the loss of Nkd1 being dominant over loss of Axin2, there are a few scenarios where we observe unexpected phenotypes in the presence of ectopic Wnt8. Firstly, Wnt8, but not BIO, induces a kinked tail phenotype in the *nkd1*$^{-/-}$ mutants, which is rescued in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants. We speculate that the kinked tail phenotype is a result of perturbed Wnt/planar cell polarity (PCP) signaling, but how this is rescued by loss of Axin2 is unknown.

Secondly, Axin2 and Nkd1 are both expressed along the ventrolateral margin of the gastrulating embryo when the hindbrain is being patterned[20,54,66]. Predictably, the single mutants were highly sensitive to exogenous Wnt8, resulting in the classic Wnt gain of function eyeless phenotype. Our prediction was that the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant embryos would be even more sensitive, but in fact the opposite was true; the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant was insensitive to exogenous Wnt8, resulting in a wild-type eye phenotype. To explain this, we must consider the role of Nkd1 in antagonizing the non-canonical Wnt/PCP pathway. We have previously demonstrated that Nkd1 functions to inhibit non-canonical Wnt/PCP signaling[20]. Furthermore, evidence suggests that there is cross talk between canonical Wnt and Wnt/PCP signaling where activation of Wnt/PCP can inhibit canonical Wnt/β-catenin signaling, however, this mechanism is not fully understood[67–70]. In this scenario, the presence of excess Wnt8 in combination with the loss of Nkd1 antagonism of Wnt/PCP during early gastrulation may lead to increased Wnt/PCP signaling and thus a higher inhibition of canonical Wnt signaling that could overcome the loss of Axin2 resulting in a rescue of the eyeless phenotype. In support of this model is the Wnt/PCP phenotype (kinked axis) that is induced only when Wnt8, and not BIO, is overexpressed in the *nkd1*$^{-/-}$ mutants. Rescue of the eyeless phenotype through the loss of both Axin2 and Nkd1 suggests that the Wnt/PCP-Wnt/β-catenin interaction is occurring within the cytoplasm upstream of the destruction complex and is dependent on a Wnt ligand interacting with its receptor. As Dvls are involved in both Wnt/β-catenin and Wnt/PCP signaling[71], bind Nkd1 and Axin1 but not Axin2[27], act upstream of the destruction complex and are activated by Wnt ligands[72], Dvls seem to be likely candidates to participate in this phenomenon but this requires further investigation.

Consistent with our results, there have been several studies investigating gene expression under different Wnt signaling scenarios, and Sp5 expression appears to be a highly upregulated Wnt target gene in these studies[55–57,60]. In addition, *sp5* expression is significantly higher than *axin2* and *nkd1* in these studies, which is also consistent with our study[55–57,60]. Similarly, when we compared our hits to the Wnt signaling study from Moya *et al*, 2014, we both identified *cyb5d1*, *wdr78*, *gnao1b*, and *crb2a* as Wnt target genes[56]. Further, we saw both upregulation and downregulation of Wnt target genes which is consistent with the research by Ewing *et al*, 2020 where Wnt target genes were also upregulated and downregulated when comparing HCT116 Δ45-β-catenin to HCT116 wild type cells[73]. This is also consistent with the function of β-catenin-dependent transcription being context dependent which is covered in depth in the review provided[74].

Our evaluation of the proteome by mass spectrometry is a unique approach as these studies are rarely performed on whole organisms, especially in zebrafish due to the high yolk protein content. While some replicates provided more hits, the poor reproducibility between replicates made it insufficient for any meaningful analysis and thus, we decided to focus on the samples with lower abundant proteins but with the most similar ion chromatogram profiles. This method identified 53 differentially expressed proteins with 18 of those proteins having known connections to Wnt signaling. For example, Prdx1 and Prdx2, proteins have been previously linked to enhancing β-catenin stability and were found to be downregulated in *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants in our results[73,75,76]. The transcript levels of *prdx1* were also significantly reduced, while *prdx2* transcripts were only slightly reduced. However, the majority of the proteins we identified were related to ribosomal assembly or metabolism, which may reflect the high translation rates occurring at 50% epiboly, a time when the embryo has fully transitioned into zygotic expression[77]. Interestingly, most of the proteins connected to Wnt signaling seem to have a promoting role in the pathway and are down regulated in the Wnt regulator mutants. This could suggest

that the embryos are modulating metabolism to downregulate Wnt signaling when Axin2 and/or Nkd1 is/are knocked out. Thus, our results are consistent with Wnt signaling having a known role in metabolism at both the gene and protein levels, however this is far from comprehensive and will need to be investigated further[78].

The differences between loss of Nkd1 and loss of Axin2 might suggest that Axin2 and/or Nkd1 are not negative feedback regulators of Wnt, but the overlapping results strongly suggests that they are which is consistent with the literature[17,22]. In particular, their sensitivity to Wnt8 overexpression or BIO treatment are well established indicators of excess Wnt signaling[19,21,79]. Furthermore, their common phenotypes in heart looping and neuromast migration and their effect on many Wnt target genes are also strong indicators that Axin2 and Nkd1 are involved in Wnt signaling.

Alternatively, it might be argued that the mutations in *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ are not complete nulls, which would require reading through of the premature stop codons[80]. In support of this, we do not observe nonsense mediated RNA decay in the qRT-PCR data. Without specific antibodies to Axin2 and Nkd1, it is difficult to confirm a complete null, but there is evidence to suggest they are both nulls. First, phenotypes that are common to both have similar effects; for example, we observe similar effects in heart looping and sensitivity to exogenous Wnt8 for both mutants. We also observe a significant increase in *axin2* and *sp5a* expression in *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants, respectively, with and without exogenous Wnt8. These results were also validated using RNA-seq where we saw differential expression of Wnt target genes in the Wnt regulator mutants. Furthermore, the *axin2*$^{-/-}$ mutant has a similar number of DEG compared to *nkd1*$^{-/-}$. Finally, we have previously generated *axin2* and *nkd1* crispants using at least two unique, non-overlapping sgRNAs and evaluated their phenotypes and target gene expression with or without Wnt8. All Crispants had similar eyeless phenotypes and *axin2* and *nkd1* expression patterns as observed here[81]. Thus, while we cannot be 100% certain that we have complete nulls, we are confident that at the very least the majority of the Axin2 and Nkd1 proteins are non-functional. We also injected Nkd2 sgRNA/Cas9 into the *nkd1*$^{-/-}$ mutants and found no evidence of compensation by Nkd2 (data not shown).

In conclusion, we have provided evidence that Nkd1 functions downstream of Axin2 and the destruction complex, as loss of Axin2 has minimal effect on Wnt signaling when Nkd1 is already knocked out. Further, the evidence clearly demonstrates that Axin2 and Nkd1 uniquely modify Wnt signaling which we speculate happens at the level of β-catenin. How these modifications affect gene transcription will be an active area of investigation and provide important insights into the complex regulation of a signaling pathway involved in many development processes and diseases.

Materials and methods

Knocking out Axin2 and Nkd1

The website chop chop (https://chopchop.cbu.uib.no/) was used to design sgRNAs for Axin2 and Nkd1, with sgRNAs being selected based on stringency and location[82]. sgRNAs were made using the GeneArt Precision gRNA Synthesis Kit following the protocol provided in the kit (Invitrogen A29377, Supplemental table 10). The sgRNA were mixed with InvitrogenTM TrueCut Cas9 protein (Thermofisher) and injected at the one-cell stage. Cas9 efficiency was determined by indel detection by amplicon analysis.

Neuromast staining

Embryos were treated with 0.003% PTU from 1dpf – 5dpf to inhibit pigmentation. At 5dpf larvae were fixed in 4% paraformaldehyde overnight at 4°C and then washed three times with PBST. Embryos were washed three times in alkaline tris buffer (0.5M Tris pH 9.5, 50mM MgCl2, 0.1M NaCl, 0.1% tween) and stained in alkaline tris buffer with 550nM Nitro blue tetrazolium chloride (NBT, Roche), and 400nM 5-Bromo-4-chloro-3-indolyl phosphate p-toluidine salt (BCIP, Roche).

Microinjections

Injections were performed using a PV820 pneumatic PicoPump microinjector (World Precision Instruments) with needles pulled using a Model P-1000 Flaming Brown Micropipette Puller (World Precision Instruments, TW100F-4). Needles were cut to yield an injection volume of ~1nL.

BIO

Embryos were arranged into a 6 well dish at 4 hpf containing either 0µM, 0.25µM, 0.5µM, 1µM, and 1.5µM of BIO with 1% DMSO. Embryos were incubated at 28.5°C until 30 hpf and were fixed using 4% PFA overnight at 4°C. Images were taken and measurements for the area of the eye were done using ImageJ. The Wnt regulator mutants were treated in parallel with WT embryos. For the BIO treatment on Axin2 mixed embryos, an incross of *axin2*$^{+/-}$ zebrafish was performed and embryos were treated with 0.75µM of BIO with 1% DMSO. Phenotypes for the eyes were recorded at 1dpf and each embryo was genotyped using fragment analysis.

qRT-PCR

RNA was isolated from a pool of 10 embryos using The GENEzol™ TriRNA Pure Kit (FroggaBio). RNA was then directly used in the Luna® Universal One-Step RT-qPCR Kit (New England BioLabs). Primers listed in supplemental table 10.

WMISH

Embryos were fixed at dome, 30%, and 50% epiboly using 4% paraformaldehyde overnight at 4°C. Embryos were washed three times in PBST and WMISH was carried out following the protocol provided in the Thisse and Thisse manuscript[83].

RNA sequencing preparation and analysis

RNA was isolated from a pool of 10 embryos at 50% epiboly using the GENEzol™ TriRNA Pure Kit (FroggaBio). RNA samples were DNase treated using the Invitrogen™ DNA-free™ DNA Removal Kit

(Thermo Fisher Scientific) and an RNA integrity number of more than 8.0 was confirmed for all samples using the 4200 Tapestation system (Agilent). Poly(A) mRNA library preparation was performed using NEBNext Ultra II DNA library prep kit for illumina (New England BioLabs) and 2x100bp paired-end sequencing was performed at a depth of 50 million reads on an Illumina Novaseq 6000 platform by the University of Toronto Donelly Sequencing Centre. The reads were aligned using Rsubread v2.2.6[84] to the Ensembl Genome Browser assembly ID: GRCz11. EdgeR v3.30.3[85] was used to filter reads and DESeq2 v1.28.1[86] was used to perform differential expression analysis.

Western blotting

At 30% epiboly 10 embryos were manually deyolked using forceps and lysed using a 27-gauge needle in a TKM buffer (50mM Tris-HCl pH 7.5, 25mM KCl, 5mM MgCl2, 1mM EGTA, 0.02% sodium azide, 1X SIGMAFAST protease inhibitor cocktail tablets EDTA free). An aliquot was taken for whole cell lysate and the rest of the sample was centrifuged at 100,000 x G for one hour at 4°C. The supernatant containing the cytoplasm was collected, and the pelleted plasma membrane proteins was resuspended in TKM buffer. Westerns were performed using 1:1500 anti-β-catenin (BD Biosciences Cat# 610154), 1:2000 anti-Pan-cadherin (Abcam Cat# ab16505), and 1:5000 anti-Actin (Sigma Cat# A5441)

Immunohistochemistry

At 30% epiboly embryos were fixed in 4% paraformaldehyde at 4°C overnight. Embryos were washed three times in PBST and were manually deyolked using forceps. Animal caps were blocked for 1 hour using sheep serum and bovine serum albumin and then incubated in 1:250 anti-β-catenin antibody (BD Bioscience cat# 610154) in blocking solution overnight at 4°C. Animal caps were washed in PBST + 2% DMSO and incubated in 1:200 488 anti mouse antibody (Donkey anti mouse 488, A21202, Life technology) in blocking solution for 2 hours at room temperature. Animal caps were mounted on cover slips and imaged using the Diskovery spinning disc microscope.

Mass spectrometry

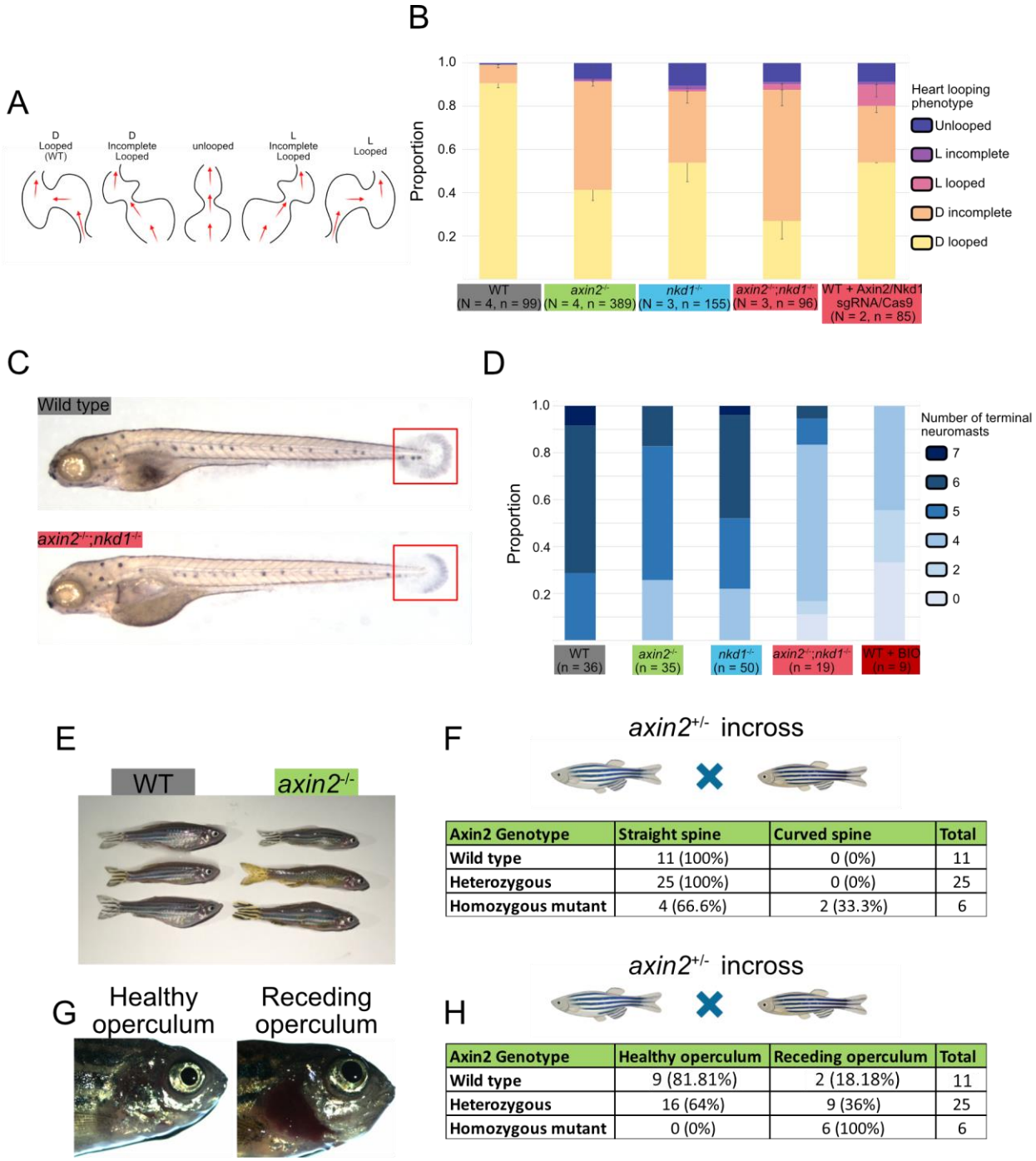
30 embryos at 30% epiboly were dechorionated and placed in a deyolking buffer (55mM NaCl, 1.8mM KCl, 1.25mM $NaHCO_3$). Embryos were vortexed on the lowest speed for 5 minutes to dissolve the yolk and centrifuged at 300 x G to pellet cells. The supernatant was removed, and the cells were washed four times using a wash buffer (110mM NaCl, 3.5mM KCl, 10mM Tris-HCl pH 8.0) followed by 300g spin to pellet cells. Cells were then washed using 1X wash buffer with added EDTA free SIGMAFAST protease inhibitor cocktail tablets. Cells were centrifuged at 300 x G and the final supernatant removed. 500µL of lysis buffer (8mM Urea, 75mM NaCl, 2mM MgCl2, 10mL 1M Tris-HCl pH 8.2, 1X SIGMAFAST protease inhibitor cocktail tablets EDTA free) was added to the cells and sonicated for 10 minutes. Lysates were incubated at 4°C for 30 minutes and centrifuged at 13,000 x G. The supernatant was moved to a clean tube and the pellet was discarded. 25uL of 200mM of iodoacetamide was added to the tubes and incubated for 30 minutes at room temperature in the dark. 1 mL of cold acetone was added, vortexed, and stored at -80°C to precipitate the protein. Protein pellets were then decanted, air dried, and resuspended in 50mM of ammonium bicarbonate. Samples were digested using trypsin and incubated at 37°C overnight and dried using vacuum centrifugation. To concentrate the samples, C18 columns were used, and protein samples were loaded into the Thermo East nLC Orbitrap Exploris 240. Analysis was performed using PEAKS 10 software. Mass spectrometry was performed by the Advanced Analysis Centre at the University of Guelph.

Authors contributions

IB and TVR conceived and planned the experiments. HK and NS made the *nkd1*$^{-/-}$ mutant. MH, EC, and IB conducted the BIO experiments. HK and IB carried out the qRT-PCR analysis. TH and IB carried out the western blotting analysis. VR and LH carried out the DTU analysis. WL performed Axin2 WMISH and *gsc* arc angle with Axin2 OE and Axin2 MO knockdown. All other experiments were carried out by IB. IB and TVR wrote the manuscript; MH, IB, and TVR edited the manuscript.

Acknowledgements

We would like to thank Matt Cornish and Mike Davies at the Hagen Aqualab facility at the University of Guelph for supporting the maintenance of the zebrafish system. We would also like to thank Amber Park and Dyanne Brewer at the Advanced Analysis Centre at the University of Guelph for their support in mass spectrometry. Lastly, we would like to thank all current and previous members in the Van Raay lab for all their support.



Axin2 Genotype	Straight spine	Curved spine	Total
Wild type	11 (100%)	0 (0%)	11
Heterozygous	25 (100%)	0 (0%)	25
Homozygous mutant	4 (66.6%)	2 (33.3%)	6

Axin2 Genotype	Healthy operculum	Receding operculum	Total
Wild type	9 (81.81%)	2 (18.18%)	11
Heterozygous	16 (64%)	9 (36%)	25
Homozygous mutant	0 (0%)	6 (100%)	6

Figure 1. Wnt regulator mutants have developmental defects.
(A) 2dpf Zebrafish embryos were observed for heart looping phenotypes and quantified in (B). (C) Zebrafish were treated with PTU from 1dpf - 5dpf and then fixed at 5dpf for neuromast staining with terminal neuromasts highlighted by the red box and quantified in (D). (E) Maternal-zygotic adult *axin2*$^{-/-}$ zebrafish occasionally develop a curved spine phenotype. (F) Adults from a heterozygous incross were phenotyped for the curved spine followed by genotyping. (G) Maternal-zygotic adult *axin2*$^{-/-}$ zebrafish often have a receding operculum phenotype. (H) Adults from a heterozygous incross were phenotyped for the receding operculum followed by genotyping. Error bars represent SEM.

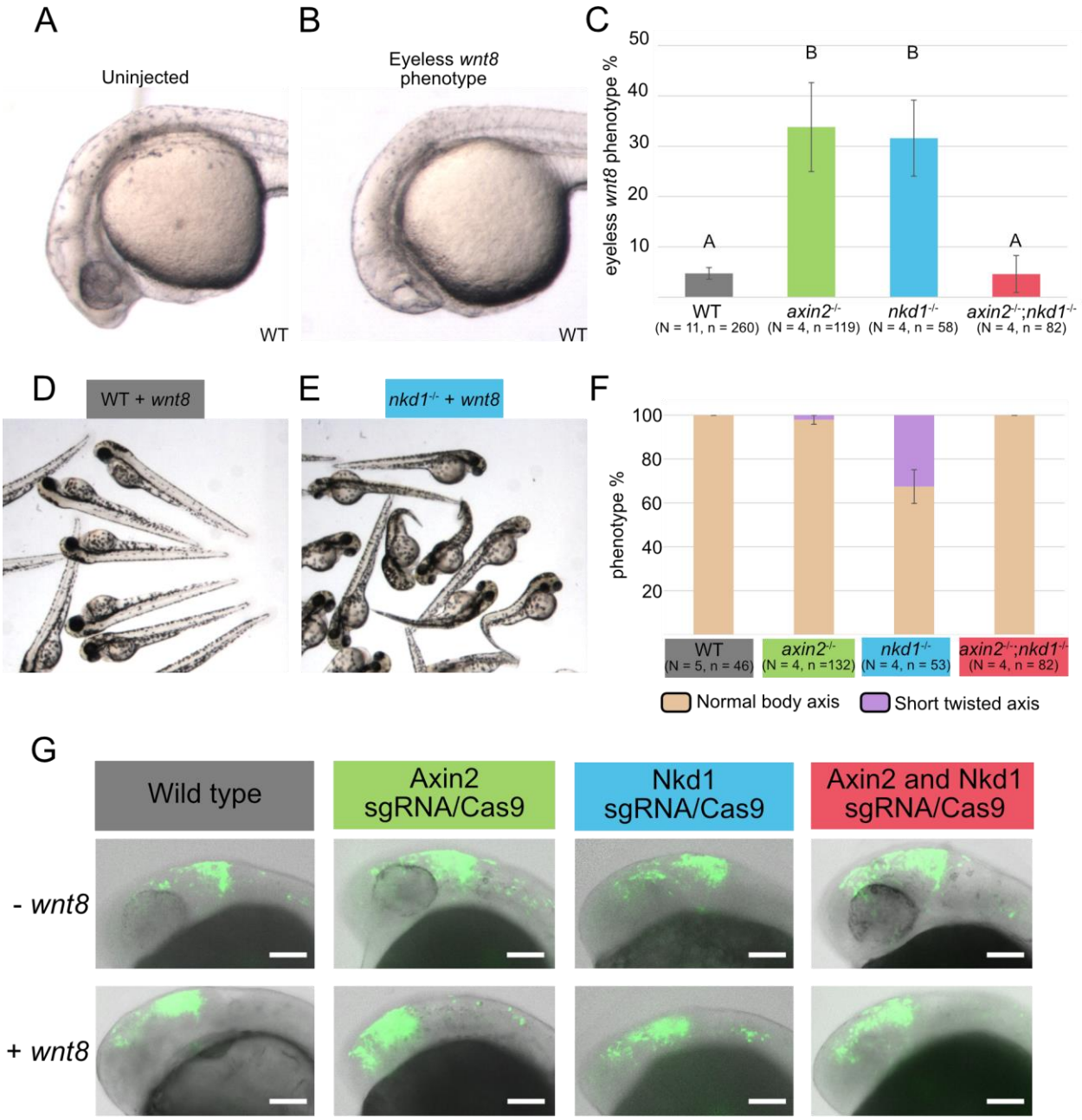


Figure 2. Wnt regulator mutants have differential sensitivities to exogenous Wnt8.
(A-C) Overexpression of 100 pg of *wnt8* in wild type embryos results in the classic Wnt gain of function eyeless phenotype in approximately 5% of embryos, but more than 30% in *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ mutants. Small eyes were considered to have eyes. In contrast, the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ resembles the wild type phenotype (C). (D-F) Analysis of body axis development revealed that only the *nkd1*$^{-/-}$ mutants develop a short-twisted axis with exogenous *wnt8* in about 30% of the embryos. (G) The Wnt reporter line was used to assess Wnt activity at the midbrain-hindbrain boundary at 1dpf when either Axin2 and/or Nkd1 was knocked down using sgRNA/Cas9. In each of the sgRNA/Cas9 injections there was an expansion of Wnt activity in the forebrain when compared to wild type embryos. Single plane brightfield images were overlaid with composite GFP images and as such the eye is not visible in all images. Error bars represent SEM, different letters above the bar signify significance with a p-value < 0.01 using a one-way ANOVA.

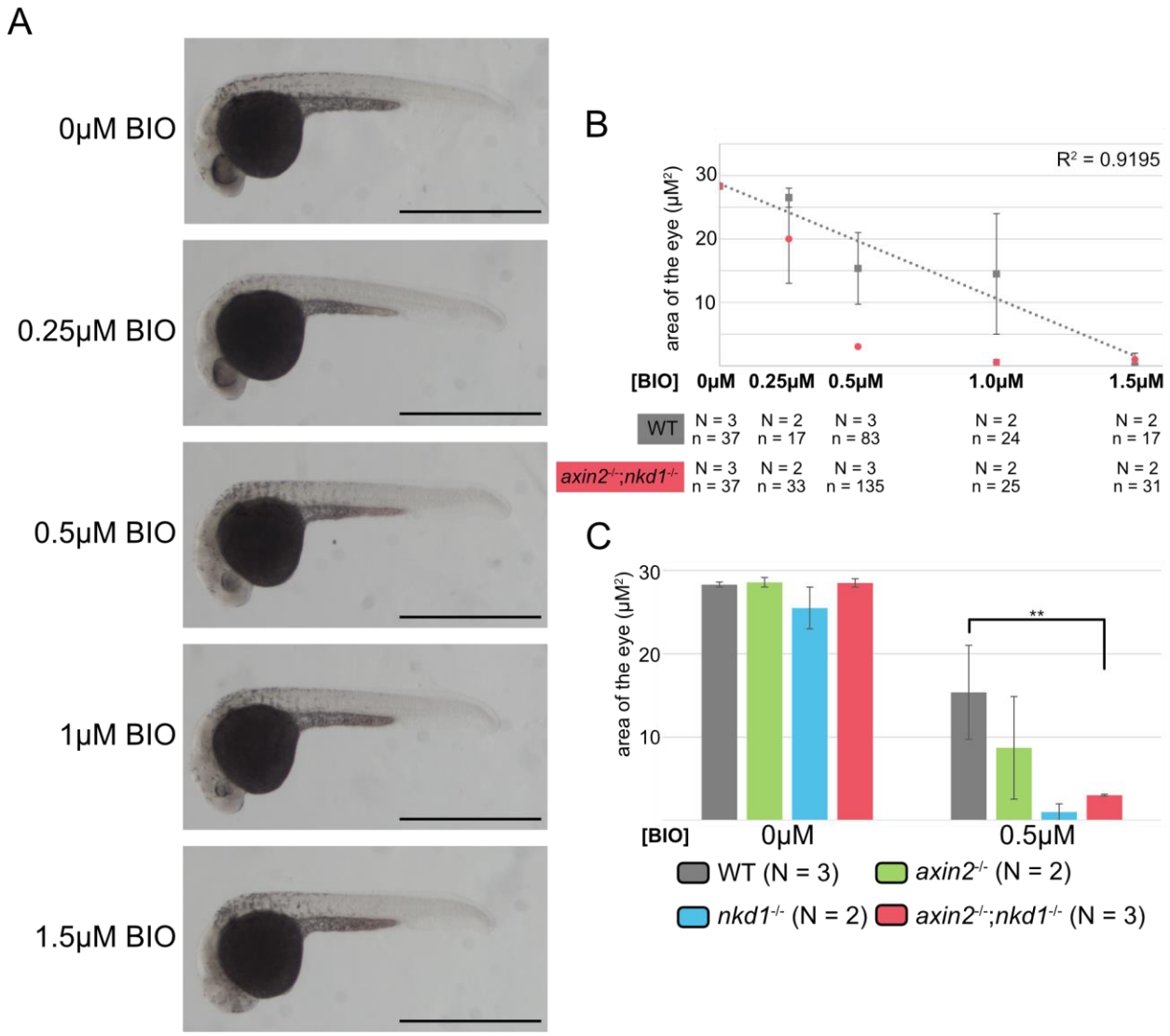


Figure 3. *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants are sensitive to activation at the destruction complex level.
(A) Embryos were treated with varying concentrations of BIO from 4 – 30 hpf with representative images taken of the wild type embryos. (B) In wild type embryos, the area of the eye has a decreasing linear association with the concentration of BIO with all embryos being eyeless at 1.5μM. In contrast, the majority of the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant embryos were eyeless at 0.5μM of BIO. (C) The *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant has no change in the size of eye in the control treatment but has significantly smaller eyes at 0.5μM of BIO when compared to wild type revealing that the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants are sensitive to BIO. Measurements were performed on imageJ and a t-test was used with ** = p-value < 0.01. All error bars represent SEM. No statistics were performed on the *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ single mutants.

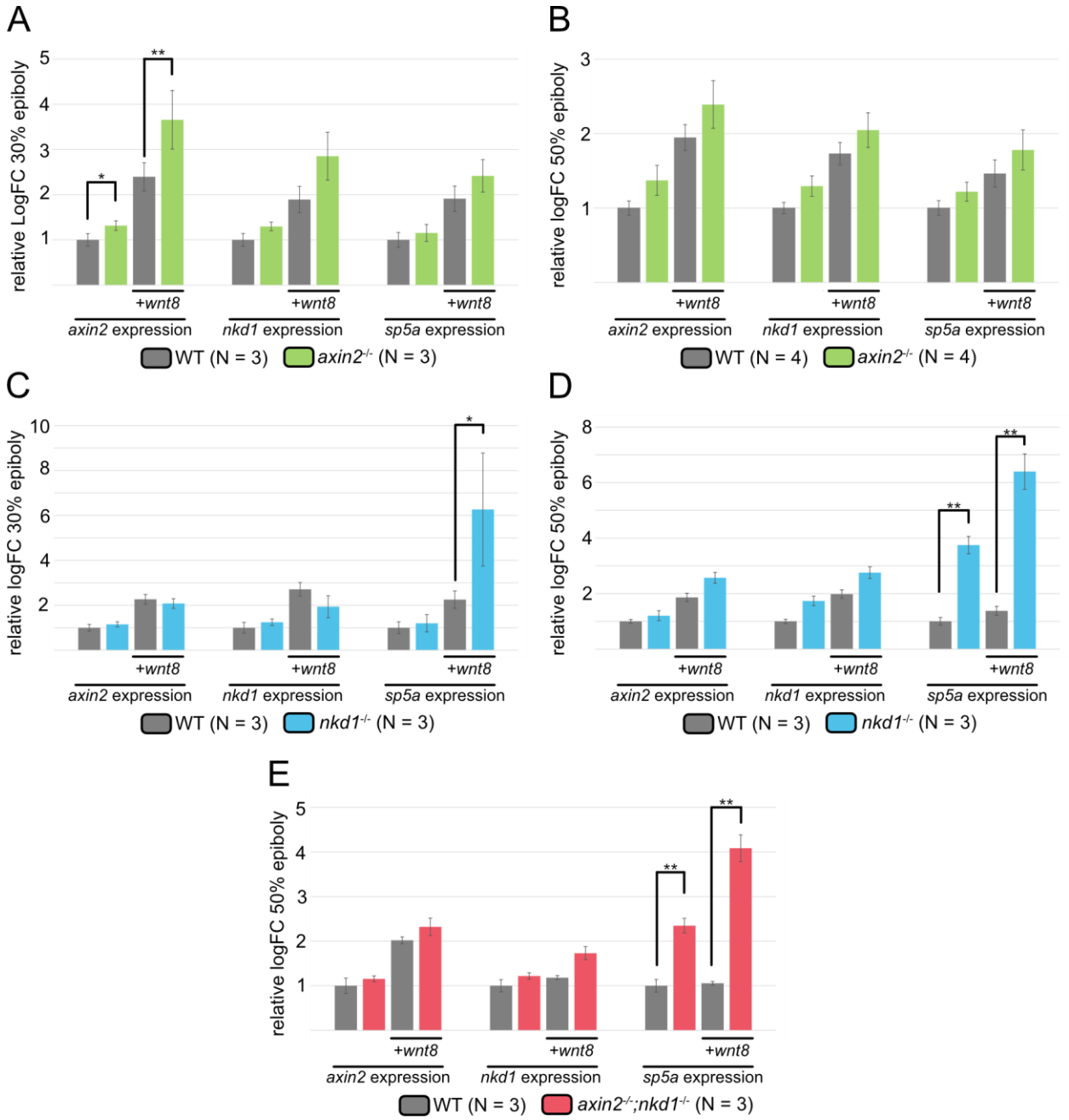


Figure 4. Wnt regulator mutants have differential effects on Wnt target genes.
qRT-PCR was performed at 30% (A,C) and 50% (B, D, E) epiboly for the Wnt target genes *axin2*, *nkd1*, and *sp5a*. Target gene expression was normalized to actin and made relative to WT uninjected expression levels. Injections were performed with 100pg of *wnt8* at the one cell stage. Expression of *axin2*, *nkd1*, and *sp5a* tended to increase in *axin2*$^{-/-}$ mutants at both 30% and 50% epiboly but only *axin2* expression increased significantly at 30% epiboly. However, in the *nkd1*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants, *axin2* and *nkd1* expression did not increase substantially compared to wild type. In contrast, *sp5a* expression showed a significant increase at 30% and 50% epiboly in the *nkd1*$^{-/-}$ mutants and at 50% in the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ (A-E, error bars represent SEM, * = p-value < 0.05, ** = p-value < 0.01, one-way ANOVA). Injection of Wnt8 into wild type (column 3) is often significant to uninjected wild type (column 1), but this is not shown for clarity.

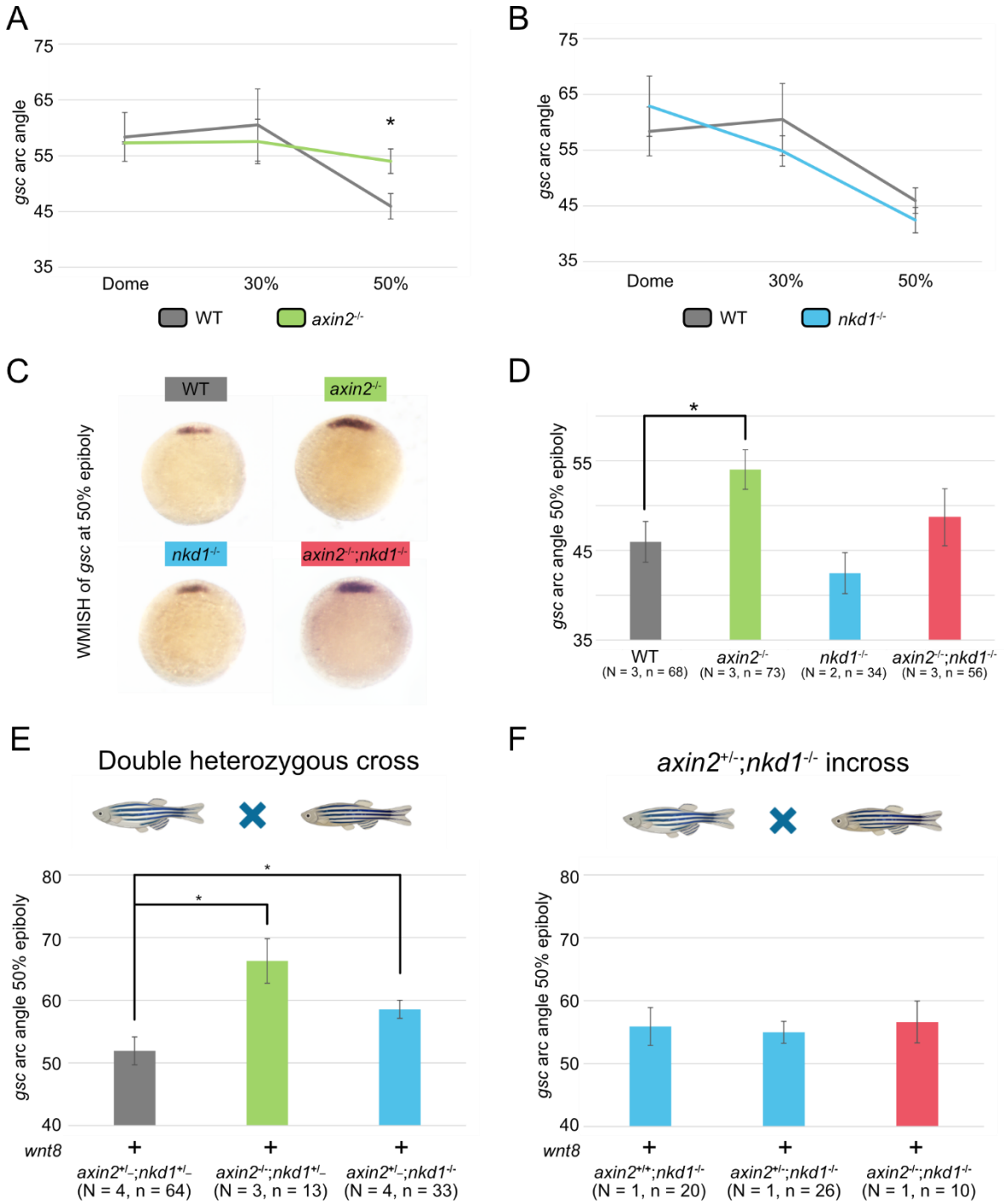
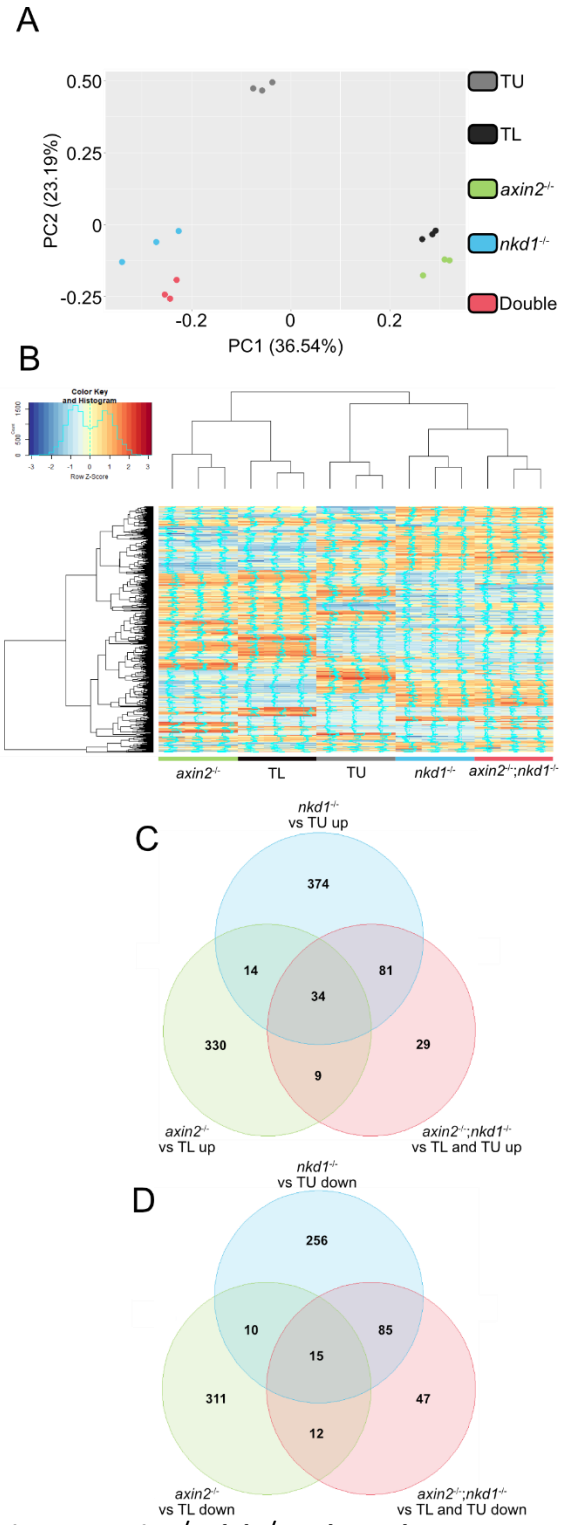


Figure 5. Wnt regulator mutants have differential effects on *gsc* expression.
(A-D) WMISH for *gsc* was performed on embryos at 3 different stages for *axin2*$^{-/-}$ and *nkd1*$^{-/-}$ single mutants (A,B) as well as 50% epiboly for all Wnt regulator mutants (C) and measured for the arc of *gsc* expression (D). At 50% epiboly, the arc angle of *gsc* expression was significantly broader in the *axin2*$^{-/-}$ embryos, which was not seen in either *nkd1*$^{-/-}$ or *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutants. (E-F) Analysis of *gsc* expression on embryos at 50% epiboly injected with 100pg of *wnt8* from either an *axin2*$^{+/-}$;*nkd1*$^{+/-}$ incross (E) or an *axin2*$^{+/-}$;*nkd1*$^{-/-}$ incross (F). (E) The *axin2*$^{-/-}$;*nkd1*$^{+/-}$ and *axin2*$^{+/-}$;*nkd1*$^{-/-}$ embryos had an increase in *gsc* arc angle when compared to *axin2*$^{+/-}$;*nkd1*$^{+/-}$. (F) In contrast, *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant embryos showed no increase in *gsc* arc angle when compared to the single knockout genotypes. Injections were performed at the one cell stage and arc angle measurements were taken on ImageJ. Embryos were genotyped using indel detection by amplicon analysis and PCR based methods. A one-way ANOVA was used for all statistics with * = p-value < 0.05, error bars represent SEM (A,B,C,E) and standard deviation (F).



E

Down regulated (FDR < 0.05) Up regulated (FDR < 0.05)

		Log fold change			
Ensembl ID	External gene name	*axin2*$^{-/-}$ vs TL	*nkd1*$^{-/-}$ vs TU	Double vs TU	Double vs TL
ENSDARG00000105403	abcc6b.1	3.142	3.905	3.791	3.252
ENSDARG00000074033	adamtsl2	3.007	4.264	4.061	2.913
ENSDARG00000097345	AL935186.3	2.377	3.272	3.623	2.915
ENSDARG00000095948	BX511100.5	2.562	2.063	2.472	3.026
ENSDARG00000097518	BX537110.1	4.136	4.465	4.613	4.782
ENSDARG00000092527	BX663522.1	2.109	3.334	3.574	3.569
ENSDARG00000078541	cers3a	2.882	5.307	5.577	5.260
ENSDARG00000076901	CR376751.1	5.909	3.816	3.833	8.175
ENSDARG00000080774	CR388189.1	2.564	3.935	4.512	3.823
ENSDARG00000060106	crb2a	2.035	2.453	3.087	3.355
ENSDARG00000097957	CU137680.2	2.313	2.021	2.401	2.681
ENSDARG00000074914	CU929346.1	2.796	7.627	7.610	2.044
ENSDARG00000056007	cyb5d1	2.547	2.912	3.465	4.663
ENSDARG00000113820	cyp46a1.4	11.041	3.524	2.495	4.281
ENSDARG00000044807	dck	2.366	2.424	2.058	3.560
ENSDARG00000044775	fut7	2.923	2.062	2.854	3.990
ENSDARG00000036058	gnao1b	3.220	2.005	2.334	2.236
ENSDARG00000112491	hist1h4l	2.279	2.170	2.177	5.733
ENSDARG00000022176	IGLON5	3.202	4.927	4.989	7.349
ENSDARG00000029874	khk	3.925	5.911	5.747	4.269
ENSDARG00000113737	LO018287.1	2.395	4.660	4.899	5.056
ENSDARG00000056690	mtmr1a	3.826	3.795	3.443	4.246
ENSDARG00000001933	prkag2b	5.483	5.526	7.049	7.245
ENSDARG00000007080	rhcgl1	5.875	3.771	3.370	5.418
ENSDARG00000103260	si:ch211-57b15.1	3.197	3.995	3.845	4.430
ENSDARG00000103237	si:ch73-299h12.8	4.668	2.320	2.813	5.269
ENSDARG00000093998	si:ch73-7i4.2	2.208	4.662	4.444	3.076
ENSDARG00000098406	si:dkey-22a18.2	4.205	3.149	3.100	3.906
ENSDARG00000089582	si:dkey-265e15.2	2.494	4.890	5.007	3.917
ENSDARG00000094385	si:dkey-85k15.6	2.624	6.890	6.790	4.392
ENSDARG00000080000	SLC25A1	5.401	5.614	5.736	6.683
ENSDARG00000088985	SPSB2	2.117	4.186	3.460	4.741
ENSDARG00000093477	zgc:123299	9.738	3.055	4.162	6.036
ENSDARG00000012281	zgc:65851	3.953	4.445	3.848	4.518
ENSDARG00000102528	AL935126.1	-3.027	-10.481	-10.249	-9.888
ENSDARG00000101966	BX664606.2	-8.552	-10.596	-10.366	-8.577
ENSDARG00000091938	BX890576.1	-4.364	-6.384	-3.833	-2.065
ENSDARG00000117806	BX901974.4	-5.521	-3.167	-6.767	-5.572
ENSDARG00000103172	BX914205.3	-3.628	-2.633	-3.673	-2.196
ENSDARG00000117107	CABZ01058647.1	-3.006	-3.495	-4.208	-5.290
ENSDARG00000116298	CR628395.1	-6.566	-2.985	-3.261	-2.326
ENSDARG00000043997	cyp2x8	-3.982	-5.865	-4.650	-4.286
ENSDARG00000009941	erc1a	-2.100	-2.080	-2.431	-3.748
ENSDARG00000097770	si:ch211-167j9.4	-3.753	-6.234	-4.871	-3.404
ENSDARG00000096522	si:ch211-176g6.1	-6.214	-6.380	-5.362	-3.397
ENSDARG00000089200	si:dkey-103g5.4	-2.692	-3.340	-4.010	-3.450
ENSDARG00000092702	si:dkeyp-20e4.8	-3.029	-5.570	-6.963	-2.274
ENSDARG00000091169	tor4ab	-2.030	-4.713	-8.616	-8.160
ENSDARG00000044400	wdr78	-2.112	-3.485	-2.963	-4.445

Figure 6. *axin2*$^{-/-}$;*nkd1*$^{-/-}$ embryos have a more similar RNA profile to *nkd1*$^{-/-}$ than to *axin2*$^{-/-}$. RNA-seq was performed at 50% epiboly for all Wnt regulator mutants as well as two wild type background genotypes: TU and TL. (A) PCA analysis and a (B) heat map analysis of the top 1000 differentially regulated genes shows that that *axin2*$^{-/-}$;*nkd1*$^{-/-}$ double mutant embryos have a similar RNA profile to the *nkd1*$^{-/-}$ single mutant embryos. (C-D) Venn diagrams were made for upregulated genes (C) (log2FC > 2 and an FDR < 0.05) and down regulated genes (D) (log2FC < -2 and an FDR < 0.05) with genes that were significantly different (FDR < 0.05) between TL and the TU backgrounds being removed. (E) A gene list for the 34 genes that were upregulated and the 15 genes that were down regulated for each of the Wnt regulator mutants was made with the log fold change values provided for comparison. For more information on DEG for each Wnt regulator mutant refer to supplemental tables 6-9.

Down-regulated (FDR < 0.05) Up-regulated (FDR < 0.05)

Gene name	Log fold change			
	axin2$^{-/-}$ vs TL	*nkd1*$^{-/-}$ vs TU	Double vs TU	Double vs TL
axin2	0.456	0.443	0.722	0.620
birc5a	0.055	-0.052	-0.043	0.559
bmp4	-0.851	-0.055	-0.004	0.208
ccn1	-0.638	-0.893	0.051	0.132
ccnd1	0.274	0.174	0.194	0.111
ccnd2b	-0.188	-0.196	0.000	-1.099
cdc25b	-0.203	-0.146	-0.217	-0.340
cdh1	0.182	-0.199	-0.153	0.032
cdx1a	-0.098	0.537	0.771	0.141
ctnnb1	0.029	0.196	0.208	0.148
ctnnb2	0.219	-0.072	-0.591	-0.439
cx43	-0.556	-0.304	-0.075	-2.623
elk4	-0.282	-0.193	-0.166	0.068
en1b	0.968	0.247	0.393	0.816
fgf18a	-0.357	0.568	0.635	-0.069
fgf18b	0.325	0.602	1.021	0.274
fn1a	-0.658	1.576	0.853	-0.296
fn1b	-0.115	0.007	0.224	0.383
foxn1	0.244	-0.726	-0.212	-0.517
fsta	0.498	1.031	0.995	0.924
fzd7b	-0.119	0.122	0.056	-0.109
gbx2	0.818	0.119	0.219	1.104
gsc	0.301	0.100	0.051	0.210
jag2b	-0.123	-0.310	-0.421	-0.866
jun	0.517	-0.093	-0.038	-0.310
lbh	-0.113	0.029	0.179	-0.433
lef1	0.454	1.685	1.593	1.355
lrp5	-0.280	-0.146	0.083	0.155
mcama	5.030	-1.568	5.209	6.177
met	-0.138	-1.237	-1.721	-1.770
mitfa	0.193	-0.266	-0.195	0.193
mitfb	0.187	-0.529	-1.570	-0.313
msl1a	0.536	-0.191	-0.127	0.171
myca	0.053	0.084	0.023	0.120
mycbp	0.318	0.202	0.128	0.582
mycn	0.093	0.309	0.509	-0.034
nkd1	0.401	0.764	0.863	0.600
nos2b	0.251	-0.120	-0.106	0.346
pgfb	-0.328	-0.003	-0.004	-0.306
pitx2	0.261	0.379	0.004	0.223
ppardb	0.038	0.166	0.217	0.272
ret	-0.023	-0.369	-0.485	-0.252
sall4	0.196	0.198	0.255	0.012
sfrp2	-0.463	0.419	0.841	0.726
six3b	0.035	1.164	2.974	3.280
sox17	-0.322	1.345	1.311	1.006
sox2	0.497	1.594	1.551	0.960
sox9a	-1.272	-0.024	-0.029	-0.953
sox9b	-0.839	-0.503	-0.481	0.214
sp5a	0.021	0.890	0.962	0.867
stra6	0.169	-0.033	0.119	-0.677
tbx3a	0.193	2.098	2.703	2.417
tbx3b	-0.265	0.161	0.166	1.110
TCF4	0.015	-0.099	0.407	0.425
tiam1a	0.415	0.721	1.189	1.420
tiam1b	0.134	0.361	0.427	1.136
vangl2	-0.017	0.044	0.081	0.039
vcana	-0.751	-0.173	-1.000	-1.373
wnt3a	-0.047	-0.231	-0.145	0.429

Figure 7. Wnt target gene expression in *axin2*$^{-/-}$;*nkd1*$^{-/-}$ is more similar to *nkd1*$^{-/-}$ than *axin2*$^{-/-}$.
59 Wnt target genes identified by the Wnt signaling community were used to create a list of expression comparisons between the Wnt regulator mutants compared to their respective backgrounds using the RNA-seq data.

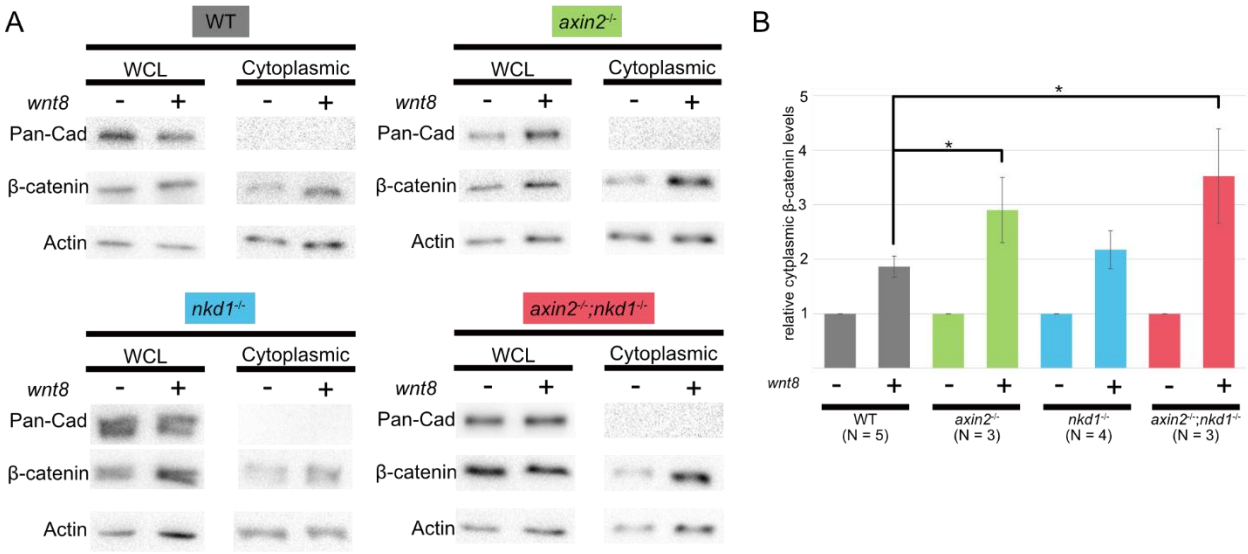


Figure 8. Knocking out Axin2, but not Nkd1, increases cytoplasmic levels of β-catenin.
(A) Whole cell lysate and cytoplasmic fractions of embryos from wild type and Wnt regulator mutants, with or without 200pg of *wnt8* injected, were collected at 30% epiboly for Western blot analysis for cytoplasmic levels of β-catenin. Pan-cadherin was used to confirm purity of the cytoplasmic fraction and Actin was used as a loading control. (B) Since each genotype was run on separate western blots, the increase in cytoplasmic β-catenin when *wnt8* was overexpressed was made relative to their uninjected counterpart. Both *axin2*$^{-/-}$ and *axin2*$^{-/-}$;*nkd1*$^{-/-}$ embryos had a greater increase in cytoplasmic β-catenin levels with *wnt8* overexpression when compared to wild type embryos. Error bars represent SEM, * = p-value < 0.05, one-way ANOVA.

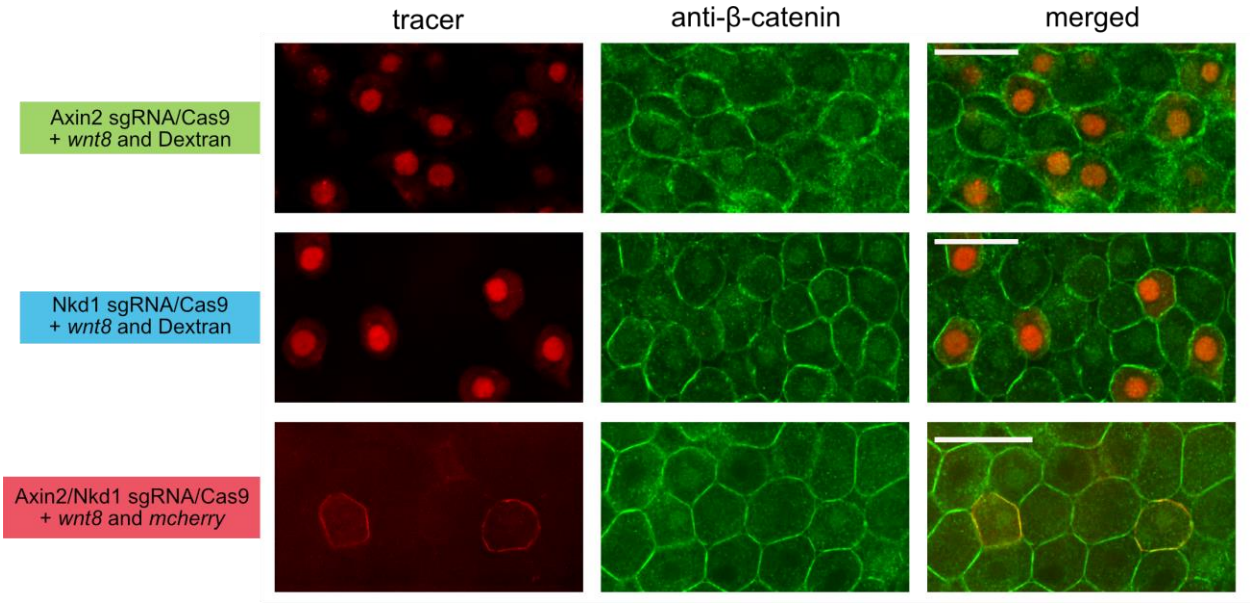
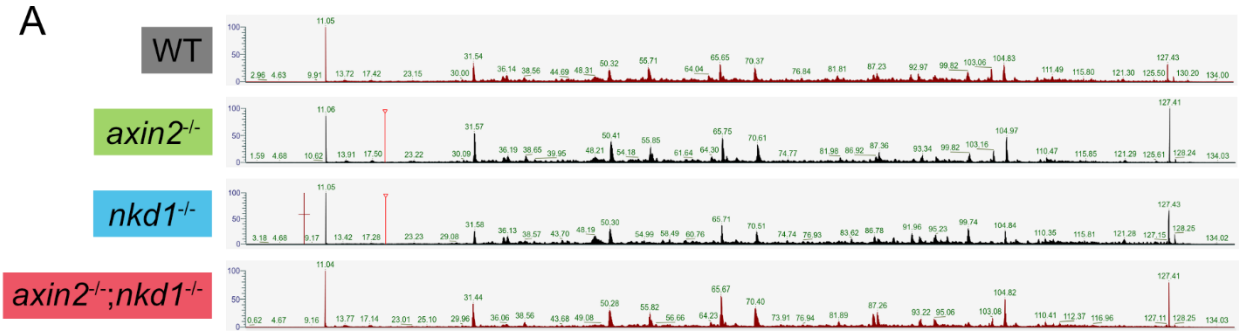


Figure 9. Knocking down Axin2 and/or Nkd1 does not increase nuclear β-catenin levels when Wnt signaling is hyperactive.

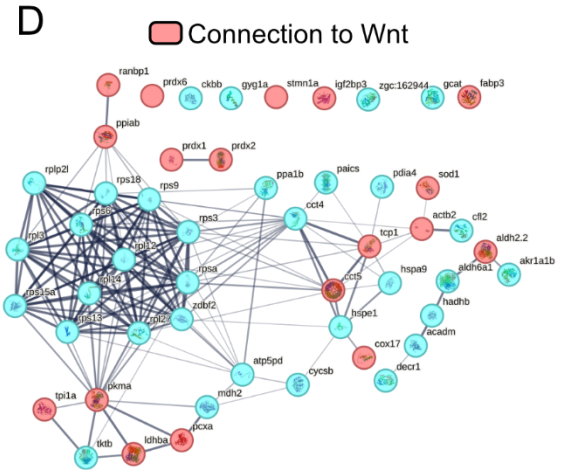
Embryos were injected with a cocktail that contained Axin2 and/or Nkd1 sgRNA/Cas9, *wnt8,* and a tracer molecule (Dextran or mcherry) into 1 cell of an 8-cell stage embryo. Embryos were fixed at 30% epiboly and immunohistochemistry was performed for β-catenin. When Axin2 and/or Nkd1 was knocked down, there was no increase in nuclear β-catenin intensity when compared to juxtaposed uninjected cells. Scale bar = 20µM.



C

Down regulated (FDR < 0.05) Up regulated (FDR < 0.05)

Protein	Mass spec LogFC vs WT: axin2-/-	nkd1-/-	DKO	Connection to Canonical Wnt signaling	RNA-seq LogFC: axin2-/- vs TL	nkd1-/- vs TU	DKO vs TU	DKO vs TL
Fabp3	-0.65	-1.90	-0.99	In zebrafish knocking down fabp3 increased Wnt1, Wnt5a and Wnt11 gene expression (Liu et al, 2013)	0.384	-0.002	-0.521	-0.236
Aldh2.2	-1.04	-1.59	-1.60	In rat heart tissue ALDH2 activation reduced β-catenin protein levels and mRNA expression (Zhao et al, 2015)	-0.511	0.212	-0.080	-0.447
Prdx1	-0.57	-2.95	-2.30	In colon cancer Caov-3 cells knocking down PRDX1 reduced β-catenin protein levels (Zheng et al, 2018)	1.147	-0.697	-1.722	-0.697
Idhba	-0.88	-1.02	-1.49	In colon cancer LS174T cells knocking out LDHA/B caused an 11 fold reduction in DKK4 gene expression (Mazzio et al, 2020)	-0.097	-0.379	-0.577	-0.249
Pcxa	-0.51	-0.97	-1.85	In breast adenocarcinoma MCF-7 cells Wnt1 and Wnt3a overexpression upregulate pyruvate carboxylate mRNA and protein levels (Lee et al, 2012)	0.201	-0.241	-0.410	-0.868
Tcp1	-0.53	-0.78	-1.58	In hepatocellular carcinoma MHCC-97H cells knocking down Tcp1 decreased β-catenin and increased p-β-catenin (Tang et al, 2020)	-0.298	-0.179	-0.342	-0.348
Igf2bp3	-0.53	-0.64	-1.41	In kidney proximal tubule HKC-8 cells Igf2bp3 binds to β-catenin mRNA and enhances its stability (Song et al, 2023)	-0.171	0.096	0.330	-0.019
Pkma	-0.43	-1.09	-1.35	In the hippocampus of mice overexpressing PKM2 increases β-catenin protein levels and increased β-catenin, c-myc, and cyclinD gene expression (Lu et al, 2021)	0.204	0.090	0.228	-0.409
Actb2	-0.30	-0.79	-0.92	In squamous cell carcinoma CAL33 cells knocking down ACTB reduced β-catenin gene expression (Gu et al, 2021)	0.172	-0.013	0.046	0.207
Cct5	-0.11	-1.03	-1.19	In gastric adenocarcinoma MKN28 cells overexpression of CCT5 increased nuclear β-catenin and decreased cytoplasmic β-catenin levels (Li et al, 2022)	-0.171	-0.252	-0.049	0.076
Prdx6	-0.31	-1.94	-1.22	In epithelial lung carcinoma H1299 and A549 knocking down PRDX6 reduced β-catenin protein levels (Xu et al, 2019)	0.274	-0.191	0.292	0.136
Tpi1a	-0.20	-1.97	-1.04	In heptocellular carcinoma HCCLM3 cell lines c-myc was shown to induce Tpi1 gene expression (Xia et al, 2023)	-0.165	0.247	0.275	-0.071
Prdx2	-0.24	-1.57	-1.13	In esophageal carcinoma Eca-109 and TE-1 cells knocking down PRDX2 reduced protein levels of Wnt3a and β-catenin (Feng et al, 2020)	-0.335	-0.063	-0.120	0.006
Ppa1b	-0.40	-1.43	-1.02	In ovarian cancer ES2 and SKOV3 cells knocking down PPA1 reduced nuclear levels of β-catenin (Niu et al, 2017)	-0.239	-0.590	-0.140	-0.054
Sod1	-0.23	-3.56	-0.67	In HEK293 cells knocking down Sod1 abolished Wnt3a-dependent nuclear localization of β-catenin (Chandrasekharan et al, 2021)	-0.049	0.455	0.502	0.366
Cox17	-0.06	-3.85	-0.15	In Cox17 mutant zebrafish there is reduced β-catenin protein levels and reduced topflash reporter activity (Li et al, 2022)	-0.075	0.584	0.097	-0.381
Ranbp1	-0.10	-2.32	-0.44	In Hematopoietic stem cells knocking out Lrp5 and 6 reduced Ranbp1 gene expression (Liu et al, 2019)	0.239	0.265	0.205	0.553
Stmn1a	-0.76	-2.70	-0.41	In human dental pulp stem cells knocking down Stmn1 reduced Wnt5a and β-catenin gene expression (Zhang et al, 2019)	-0.063	-0.056	-0.190	0.360



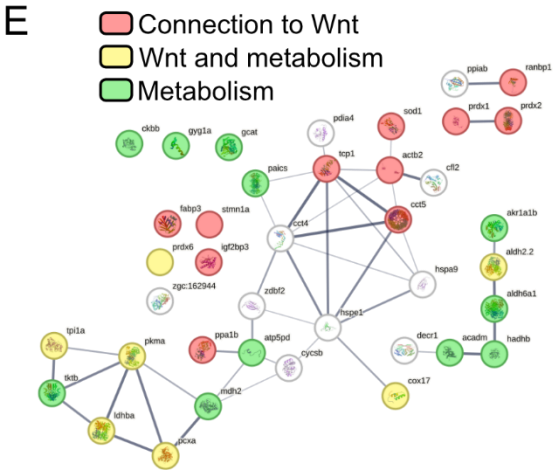


Figure 10. Wnt signaling mutants have lower expression of metabolism proteins.
Mass spectrometry was performed on embryos at 30% epiboly. (A) Only samples with similar total ionic chromatograms were used, resulting in only 1 biological replicate for each genotype. (B) Heat map of 53 differentially expressed proteins using PEAKS 10 software (FDR < 0.1, logFC ≥ 1, peptides ≥ 2). (C) Eighteen proteins were identified to have a connection to Wnt signaling in the literature: Fabp3[87], Aldh2.2[88], Prdx1[75], Idhba[89], Pcxa[90], Tcp1[91], Igf2bp3[92], Pkma[93], Actb2[94], Cct5[95], Prdx6[96], Tpi1a[97], Prdx2[76], Ppa1b[98], Sod1[99], Cox17[100], Ranbp1[101], Stmn1a[102]. (D-E) String protein networks were created using all proteins identified (D) and a network of the 39 non ribosomal proteins (E).

Experiment	*axin2*$^{-/-}$	*nkd1*$^{-/-}$	*axin2*$^{-/-}$;*nkd1*$^{-/-}$	*axin2*$^{-/-}$;*nkd1*$^{-/-}$ is like?
Heart looping	++	++	++	Both
Neuromast migration	++	++	+++	Synergistic
Spine and operculum	++	-	++	*axin2*$^{-/-}$
Thigmotaxis behavior	++	++	++	Both
Tap response	+	+++	+++	*nkd1*$^{-/-}$
Eyeless Wnt8	++	++	-	WT
Twisted axis Wnt8	-	++	-	*axin2*$^{-/-}$
Wnt activity in forebrain	++	+	++	*nkd1*$^{-/-}$
Wnt activity in forebrain Wnt8	+++	+	++	nkd1$^{-/-}$
BIO sensitivity	++	++	++	Both
axin2 expression	++	+	+	*nkd1*$^{-/-}$
axin2 expression Wnt8	++	+	+	*nkd1*$^{-/-}$
nkd1 expression	+	+	+	*nkd1*$^{-/-}$
nkd1 expression Wnt8	+	+	+	Both
sp5a expression	-	+++	+++	*nkd1*$^{-/-}$
sp5a expression Wnt8	+	+++	+++	*nkd1*$^{-/-}$
gsc expression at 50%	++	-	-	*nkd1*$^{-/-}$
gsc expression with Wnt8 OE	++	++	++	Both
RNA-seq	+	++	++	*nkd1*$^{-/-}$
RNA-seq Wnt target genes	+	++	++	*nkd1*$^{-/-}$
Cytoplasmic β-catenin	++	-	++	*axin2*$^{-/-}$
Nuclear β-catenin	-	-	-	WT
Nuclear β-catenin Wnt8	-	-	-	WT
Protein expression	+	+++	+++	*nkd1*$^{-/-}$

Table 1. Summary of results. Each experiment is listed with the effects shown for each genotype. A (-) symbol signifies no observed effect, while the (+) symbol signifies an effect. **+** = somewhat affected, **++** = affected, **+++** = very affected. Colour coding reflects genotype. In the majority of our analysis the *axin2*$^{-/-}$;*nkd1*$^{-/-}$ mutant looks similar to *nkd1*$^{-/-}$ but rarely has a more severe phenotype when compared to the single mutants.

REFERENCES

1. Kim C-H, Oda T, Itoh M, et al. Repressor activity of Headless/Tcf3 is essential for vertebrate head formation. *Nature*. 2000;407(6806):913-916. doi:10.1038/35038097

2. Kosinski C, Li VSW, Chan ASY, et al. Gene expression patterns of human colon tops and basal crypts and BMP antagonists as intestinal stem cell niche factors. *Proc Natl Acad Sci U S A*. 2007;104(39):15418-15423. doi:10.1073/pnas.0707210104

3. Lu F-I, Thisse C, Thisse B. Identification and mechanism of regulation of the zebrafish dorsal determinant. *Proc Natl Acad Sci U S A*. 2011;108(38):15876-15880. doi:10.1073/pnas.1106801108

4. Y W, CJ Z, Y L. Wnt Signaling in Kidney Development and Disease. *Prog Mol Biol Transl Sci*. 2018;153:181-207. doi:10.1016/BS.PMBTS.2017.11.019

5. Russell JO, Monga SP. Wnt/β-Catenin Signaling in Liver Development, Homeostasis, and Pathobiology. *Annu Rev Pathol*. 2018;13:351. doi:10.1146/ANNUREV-PATHOL-020117-044010

6. Yu S, Han R, Gan R. The Wnt/β-catenin signalling pathway in Haematological Neoplasms. *Biomark Res 2022 101*. 2022;10(1):1-13. doi:10.1186/S40364-022-00418-9

7. Xu X, Zhang M, Xu F, Jiang S. Wnt signaling in breast cancer: biological mechanisms, challenges and opportunities. *Mol Cancer 2020 191*. 2020;19(1):1-35. doi:10.1186/S12943-020-01276-5

8. Novellasdemunt L, Antas P, Li VSW. Targeting Wnt signaling in colorectal cancer. A review in the theme: Cell signaling: Proteins, pathways and mechanisms. *Am J Physiol - Cell Physiol*. 2015;309(8):C511-C521. doi:10.1152/ajpcell.00117.2015

9. Chen J, Ning C, Mu J, Li D, Ma Y, Meng X. Role of Wnt signaling pathways in type 2 diabetes mellitus. *Mol Cell Biochem*. 2021;476(5):2219-2232. doi:10.1007/S11010-021-04086-5

10. Gong Y, Slee RB, Fukai N, et al. LDL receptor-related protein 5 (LRP5) affects bone accrual and eye development. *Cell*. 2001;107(4):513-523. doi:10.1016/S0092-8674(01)00571-2

11. Gao J, Liao Y, Qiu M, Shen W. Wnt/β-Catenin Signaling in Neural Stem Cell Homeostasis and Neurological Diseases. *Neuroscientist*. 2021;27(1):58-72. doi:10.1177/1073858420914509/ASSET/IMAGES/LARGE/10.1177_1073858420914509-FIG3.JPEG

12. Komiya Y, Habas R. Wnt signal transduction pathways. *Organogenesis*. 2008;4(2):68-75. doi:10.4161/org.4.2.5851

13. Zeng X, Huang H, Tamai K, et al. Initiation of Wnt signaling: control of Wnt coreceptor Lrp6 phosphorylation/activation via frizzled, dishevelled and axin functions. *Development*. 2008;135(2):367-375. doi:10.1242/dev.013540

14. Flack JE, Mieszczanek J, Novcic N, Bienz M. Wnt-Dependent Inactivation of the Groucho/TLE Co-repressor by the HECT E3 Ubiquitin Ligase Hyd/UBR5. *Mol Cell*. 2017;67(2):181. doi:10.1016/J.MOLCEL.2017.06.009

15. Borrelli C, Valenta T, Handler K, et al. Differential regulation of β-catenin-mediated transcription via N- and C-terminal co-factors governs identity of murine intestinal epithelial stem cells. *Nat*

Commun 2021 121. 2021;12(1):1-16. doi:10.1038/s41467-021-21591-9

16. Angonin D, van Raay TJ. Nkd1 Functions as a Passive Antagonist of Wnt Signaling. *PLoS One*. 2013;8(8). doi:10.1371/journal.pone.0074666

17. Bernkopf DB, Hadjihannas M V., Behrens J. Negative-feedback regulation of the Wnt pathway by conductin/axin2 involves insensitivity to upstream signalling. *J Cell Sci*. 2015;128(1):33-39. doi:10.1242/jcs.159145

18. Wang Y, Yang C, Li W, et al. Identification of colon tumor marker NKD1 via integrated bioinformatics analysis and experimental validation. *Cancer Med*. 2021;10(20):7383. doi:10.1002/CAM4.4224

19. Marsden AN, Derry SW, Schneider I, et al. The Nkd EF-hand domain modulates divergent wnt signaling outputs in zebrafish. *Dev Biol*. 2018;434(1):63-73. doi:10.1016/j.ydbio.2017.11.012

20. Van Raay TJ, Coffey RJ, Solnica-Krezel L. Zebrafish Naked1 and Naked2 antagonize both canonical and non-canonical Wnt signaling. *Dev Biol*. 2007;309(2):151-168. doi:10.1016/j.ydbio.2007.04.018

21. Hu T, Li C, Cao Z, et al. Myristoylated Naked2 Antagonizes Wnt-β-Catenin Activity by Degrading Dishevelled-1 at the Plasma Membrane. *J Biol Chem*. 2010;285(18):13561. doi:10.1074/JBC.M109.075945

22. van Raay TJ, Fortino NJ, Miller BW, et al. Naked1 antagonizes wnt signaling by preventing nuclear accumulation of β-catenin. *PLoS One*. 2011;6(4). doi:10.1371/journal.pone.0018650

23. Wharton KA, Zimmermann G, Rousset R, Scott MP. Vertebrate proteins related to Drosophila Naked Cuticle bind Dishevelled and antagonize Wnt signaling. *Dev Biol*. 2001;234(1):93-106. doi:10.1006/DBIO.2001.0238

24. Timr Š, Pleskot R, Kadlec J, Kohagen M, Magarkar A, Jungwirth P. Membrane Binding of Recoverin: From Mechanistic Understanding to Biological Functionality. *ACS Cent Sci*. 2017;3(8):868-874. doi:10.1021/ACSCENTSCI.7B00210/ASSET/IMAGES/LARGE/OC-2017-00210W_0004.JPEG

25. Rousset R, Wharton KA, Zimmermann G, Scott MP. Zinc-dependent interaction between dishevelled and the Drosophila Wnt antagonist naked cuticle. *J Biol Chem*. 2002;277(50):49019-49026. doi:10.1074/jbc.M203246200

26. Gammons M V., Renko M, Flack JE, Mieszczanek J, Bienz M. Feedback control of wnt signaling based on ultrastable histidine cluster co-aggregation between Naked/NKD and axin. *Elife*. 2020;9:1-22. doi:10.7554/ELIFE.59879

27. Miller BW, Lau G, Grouios C, et al. Application of an integrated physical and functional screening approach to identify inhibitors of the Wnt pathway. *Mol Syst Biol*. 2009;5:315. doi:10.1038/msb.2009.72

28. Larraguibel J, Weiss ARE, Pasula DJ, Dhaliwal RS, Kondra R, Van Raay TJ. Wnt ligand–dependent activation of the negative feedback regulator Nkd1. Bronner M, ed. *Mol Biol Cell*. 2015;26(12):2375-2384. doi:10.1091/mbc.E14-12-1648

29. Zeng W, Wharton KA, Mack JA, et al. naked cuticle encodes an inducible antagonist of Wnt

signalling. *Nat 2000 4036771*. 2000;403(6771):789-795. doi:10.1038/35001615

30. Wang R, Xie H, Yang L, et al. Naked cuticle inhibits wingless signaling in Drosophila wing development. *Biochem Biophys Res Commun*. 2021;576:1-6. doi:10.1016/J.BBRC.2021.08.077

31. Zhang S, Çagatay T, Amanai M, et al. Viable Mice with Compound Mutations in the Wnt/Dvl Pathway Antagonists nkd1 and nkd2. *Mol Cell Biol*. 2007;27(12):4454-4464. doi:10.1128/mcb.00133-07

32. Li Q, Ishikawa TO, Miyoshi H, Oshima M, Taketo MM. A targeted mutation of Nkd1 impairs mouse spermatogenesis. *J Biol Chem*. 2005;280(4):2831-2839. doi:10.1074/JBC.M405680200

33. Schneider I, Schneider PN, Derry SW, et al. Zebrafish Nkd1 promotes Dvl degradation and is required for left-right patterning. *Dev Biol*. 2010;348(1):22. doi:10.1016/J.YDBIO.2010.08.040

34. Lange M, Granados A, VijayKumar S, et al. Zebrahub – Multimodal Zebrafish Developmental Atlas Reveals the State-Transition Dynamics of Late-Vertebrate Pluripotent Axial Progenitors. *bioRxiv*. June 2023:2023.03.06.531398. doi:10.1101/2023.03.06.531398

35. Yamamoto H, Kishida S, Uochi T, et al. Axil, a Member of the Axin Family, Interacts with Both Glycogen Synthase Kinase 3β and β-Catenin and Inhibits Axis Formation of Xenopus Embryos. *Mol Cell Biol*. 1998;18(5):2867. doi:10.1128/MCB.18.5.2867

36. Zeng L, Fagotto F, Zhang T, et al. The Mouse Fused Locus Encodes Axin, an Inhibitor of the Wnt Signaling Pathway That Regulates Embryonic Axis Formation. *Cell*. 1997;90(1):181-192. doi:10.1016/S0092-8674(00)80324-4

37. Bernkopf DB, Brückner M, Hadjihannas M V., Behrens J. An aggregon in conductin/axin2 regulates Wnt/β-catenin signaling and holds potential for cancer therapy. *Nat Commun*. 2019;10(1):1-14. doi:10.1038/s41467-019-12203-8

38. Fiedler M, Mendoza-Topaz C, Rutherford TJ, Mieszczanek J, Bienz M. Dishevelled interacts with the DIX domain polymerization interface of Axin to interfere with its function in down-regulating β-catenin. *Proc Natl Acad Sci U S A*. 2011;108(5):1937-1942. doi:10.1073/pnas.1017063108

39. Xie R, Jiang R, Chen D. Generation of Axin1 conditional mutant mice. *genesis*. 2011;49(2):98-102. doi:10.1002/dvg.20703

40. Yu HMI, Jerchow B, Sheu TJ, et al. The role of Axin2 in calvarial morphogenesis and craniosynostosis. *Development*. 2005;132(8):1995-2005. doi:10.1242/dev.01786

41. Yu HMI, Jerchow B, Sheu TJ, et al. The role of Axin2 in calvarial morphogenesis and craniosynostosis. *Development*. 2005;132(8):1995-2005. doi:10.1242/DEV.01786

42. Hulin A, Moore V, James JM, Yutzey KE. Loss of Axin2 results in impaired heart valve maturation and subsequent myxomatous valve disease. *Cardiovasc Res*. 2017;113(1):40-51. doi:10.1093/cvr/cvw229

43. Rieger ME, Zhou B, Solomon N, et al. p300/β-Catenin Interactions Regulate Adult Progenitor Cell Differentiation Downstream of WNT5a/Protein Kinase C (PKC). *J Biol Chem*. 2016;291(12):6569. doi:10.1074/JBC.M115.706416

44. Albuquerque C, Breukel C, van der Luijt R, et al. The 'just-right' signaling model: APC somatic mutations are selected based on a specific level of activation of the β-catenin signaling cascade.

Hum Mol Genet. 2002;11(13):1549-1560. doi:10.1093/HMG/11.13.1549

45. Colozza G, Koo BK. Wnt/β-catenin signaling: Structure, assembly and endocytosis of the signalosome. *Dev Growth Differ*. 2021;63(3):199. doi:10.1111/DGD.12718

46. Lin X, Xu X. Distinct functions of Wnt/β-catenin signaling in KV development and cardiac asymmetry. *Development*. 2009;136(2):207-217. doi:10.1242/DEV.029561

47. Head JR, Gacioch L, Pennisi M, Meyers JR. Activation of canonical Wnt/β-catenin signaling stimulates proliferation in neuromasts in the zebrafish posterior lateral line. *Dev Dyn*. 2013;242(7):832-846. doi:10.1002/DVDY.23973

48. Valdivia LE, Young RM, Hawkins TA, et al. Lef1-dependent Wnt/β-catenin signalling drives the proliferative engine that maintains tissue homeostasis during lateral line development. *Development*. 2011;138(18):3931-3941. doi:10.1242/DEV.062695

49. Ma EY, Raible DW. Signaling Pathways Regulating Zebrafish Lateral Line Development. *Curr Biol*. 2009;19(9):R381-R386. doi:10.1016/j.cub.2009.03.057

50. Qu X, Liao M, Liu W, et al. Loss of Wnt16 Leads to Skeletal Deformities and Downregulation of Bone Developmental Pathway in Zebrafish. *Int J Mol Sci Artic Int J Mol Sci*. 2021;22:6673. doi:10.3390/ijms22136673

51. Hayes M, Gao X, Yu LX, et al. ptk7 mutant zebrafish models of congenital and idiopathic scoliosis implicate dysregulated Wnt signalling in disease. *Nat Commun*. 2014;5. doi:10.1038/NCOMMS5777

52. Moro E, Ozhan-Kizil G, Mongera A, et al. In vivo Wnt signaling tracing through a transgenic biosensor fish reveals novel activity domains. *Dev Biol*. 2012;366(2):327-340. doi:10.1016/J.YDBIO.2012.03.023

53. Meijer L, Skaltsounis AL, Magiatis P, et al. GSK-3-Selective Inhibitors Derived from Tyrian Purple Indirubins. *Chem Biol*. 2003;10(12):1255-1266. doi:10.1016/J.CHEMBIOL.2003.11.010

54. Moens CB, Prince VE. Constructing the hindbrain: Insights from the zebrafish. *Dev Dyn*. 2002;224(1):1-17. doi:10.1002/DVDY.10086

55. Mukherjee S, Luedeke DM, McCoy L, Iwafuchi M, Zorn AM. SOX transcription factors direct TCF-independent WNT/β-catenin responsive transcription to govern cell fate in human pluripotent stem cells. *Cell Rep*. 2022;40(8):111247. doi:10.1016/J.CELREP.2022.111247

56. Moya N, Cutts J, Gaasterland T, Willert K, Brafman DA. Endogenous WNT Signaling Regulates hPSC-Derived Neural Progenitor Cell Heterogeneity and Specifies Their Regional Identity. *Stem Cell Reports*. 2014;3(6):1015-1028. doi:10.1016/J.STEMCR.2014.10.004

57. Dunty WC, Kennedy MWL, Chalamalasetty RB, Campbell K, Yamaguchi TP. Transcriptional Profiling of Wnt3a Mutants Identifies Sp Transcription Factors as Essential Effectors of the Wnt/β-catenin Pathway in Neuromesodermal Stem Cells. *PLoS One*. 2014;9(1):e87018. doi:10.1371/JOURNAL.PONE.0087018

58. Weidinger G, Thorpe CJ, Wuennenberg-Stapleton K, Ngai J, Moon RT. The Sp1-Related Transcription Factors sp5 and sp5-like Act Downstream of Wnt/β-Catenin Signaling in Mesoderm and Neuroectoderm Patterning. *Curr Biol*. 2005;15(6):489-500. doi:10.1016/J.CUB.2005.01.041

59. Jho E, Zhang T, Domon C, Joo C-K, Freund J-N, Costantini F. Wnt/β-Catenin/Tcf Signaling Induces the Transcription of Axin2, a Negative Regulator of the Signaling Pathway. *Mol Cell Biol*. 2002;22(4):1172. doi:10.1128/MCB.22.4.1172-1183.2002

60. Huggins IJ, Bos T, Gaylord O, et al. The WNT target SP5 negatively regulates WNT transcriptional programs in human pluripotent stem cells. *Nat Commun*. 2017;8(1). doi:10.1038/S41467-017-01203-1

61. Kelly C, Chin AJ, Leatherman JL, Kozlowski DJ, Weinberg ES. Maternally controlled (beta)-catenin-mediated signaling is required for organizer formation in the zebrafish. *Development*. 2000;127(18):3899-3911. doi:10.1242/DEV.127.18.3899

62. Bellipanni G, Varga M, Maegawa S, et al. Essential and opposing roles of zebrafish β-catenins in the formation of dorsal axial structures and neurectoderm. *Development*. 2006;133(7):1299-1309. doi:10.1242/DEV.02295

63. Tian X, Liu Z, Niu B, et al. E-Cadherin/β-Catenin Complex and the Epithelial Barrier. *J Biomed Biotechnol*. 2011;2011. doi:10.1155/2011/567305

64. Yan Y, Tang D, Chen M, et al. Axin2 controls bone remodeling through the β-catenin-BMP signaling pathway in adult mice. *J Cell Sci*. 2009;122(19):3566-3578. doi:10.1242/jcs.051904

65. Freeman M. Feedback control of intercellular signalling in development. *Nature*. 2000;408(6810):313-319. doi:10.1038/35042500

66. Gao R, Ma LQ, Du X, et al. Rnf25/AO7 positively regulates wnt signaling via disrupting Nkd1-Axin inhibitory complex independent of its ubiquitin ligase activity. *Oncotarget*. 2016;7(17):23850-23859. doi:10.18632/oncotarget.8126

67. Nemeth MJ, Topol L, Anderson SM, Yang Y, Bodine DM. Wnt5a inhibits canonical Wnt signaling in hematopoietic stem cells and enhances repopulation. *Proc Natl Acad Sci U S A*. 2007;104(39):15436-15441. doi:10.1073/pnas.0704747104

68. Tortes MA, Yang-Snyder JA, Purcell SM, Demarais AA, Mcgrew LL, Moon RT. *Activities of the Wnt-1 Class of Secreted Signaling Factors Are Antagonized by the Wnt-5A Class and by a Dominant Negative Cadherin in Early Xenopus Development*. https://rupress.org/jcb/article-pdf/133/5/1123/389130/1123.pdf. Accessed April 30, 2020.

69. Bisson JA, Mills B, Helt JCP, Zwaka TP, Cohen ED. Wnt5a and Wnt11 inhibit the canonical Wnt pathway and promote cardiac progenitor development via the Caspase-dependent degradation of AKT. *Dev Biol*. 2015;398(1):80-96. doi:10.1016/j.ydbio.2014.11.015

70. Bell IJ, Horn MS, Van Raay TJ. Bridging the gap between non-canonical and canonical Wnt signaling through Vangl2. *Semin Cell Dev Biol*. 2022;125:37-44. doi:10.1016/J.SEMCDB.2021.10.004

71. Sharma M, Castro-Piedras I, Simmons GE, Pruitt K. Dishevelled: a masterful conductor of complex Wnt signals. *Cell Signal*. 2018;47:52. doi:10.1016/J.CELLSIG.2018.03.004

72. Ma W, Chen M, Kang H, et al. Single-molecule dynamics of dishevelled at the plasma membrane and Wnt pathway activation. *Proc Natl Acad Sci U S A*. 2020;117(28):16690-16701. doi:10.1073/PNAS.1910547117/SUPPL_FILE/PNAS.1910547117.SM02.AVI

73. Ewing RM, Song J, Gokulrangan G, et al. Multiproteomic and Transcriptomic Analysis of Oncogenic β-Catenin Molecular Networks. *J Proteome Res*. 2018;17(6):2216. doi:10.1021/ACS.JPROTEOME.8B00180

74. Valenta T, Hausmann G, Basler K. The many faces and functions of Î 2-catenin. *EMBO J*. 2012;31(12):2714-2736. doi:10.1038/emboj.2012.150

75. Zheng MJ, Wang J, Wang HM, et al. Decreased expression of peroxiredoxin1 inhibits proliferation, invasion, and metastasis of ovarian cancer cell. *Onco Targets Ther*. 2018;11:7745-7761. doi:10.2147/OTT.S175009

76. Feng AL, Han X, Meng X, et al. PRDX2 plays an oncogenic role in esophageal squamous cell carcinoma via Wnt/β-catenin and AKT pathways. *Clin Transl Oncol*. 2020;22(10):1838-1848. doi:10.1007/S12094-020-02323-9/METRICS

77. Vastenhouw NL, Cao WX, Lipshitz HD. The maternal-to-zygotic transition revisited. 2019. doi:10.1242/dev.161471

78. Sethi JK, Vidal-Puig A. Wnt signalling and the control of cellular metabolism. *Biochem J*. 2010;427(1):1. doi:10.1042/BJ20091866

79. Robertson JK, Danzmann K, Charles S, et al. Targeting the Wnt pathway in zebrafish as a screening method to identify novel therapeutic compounds: *https://doi.org/101177/1535370213514322*. 2014;239(2):169-176. doi:10.1177/1535370213514322

80. Li C, Zhang J. Stop-codon read-through arises largely from molecular errors and is generally nonadaptive. *PLoS Genet*. 2019;15(5). doi:10.1371/JOURNAL.PGEN.1008141

81. Pasula D, Guelph C©. Investigating the Role of Axin2 in the Regulation of the Wnt Signaling Pathway in Early Zebrafish Development. 2016.

82. Labun K, Montague TG, Krause M, Torres Cleuren YN, Tjeldnes H, Valen E. CHOPCHOP v3: expanding the CRISPR web toolbox beyond genome editing. *Nucleic Acids Res*. 2019;47(W1):W171-W174. doi:10.1093/NAR/GKZ365

83. C T, B T. High-resolution in situ hybridization to whole-mount zebrafish embryos. *Nat Protoc*. 2008;3(1):59-69. doi:10.1038/NPROT.2007.514

84. Liao Y, Smyth GK, Shi W. The R package Rsubread is easier, faster, cheaper and better for alignment and quantification of RNA sequencing reads. *Nucleic Acids Res*. 2019;47(8):e47-e47. doi:10.1093/NAR/GKZ114

85. Robinson MD, McCarthy DJ, Smyth GK. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics*. 2010;26(1):139-140. doi:10.1093/BIOINFORMATICS/BTP616

86. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol*. 2014;15(12):1-21. doi:10.1186/S13059-014-0550-8/FIGURES/9

87. Liu YQ, Song GX, Liu HL, et al. Silencing of FABP3 leads to apoptosis-induced mitochondrial dysfunction and stimulates Wnt signaling in zebrafish. *Mol Med Rep*. 2013;8(3):806-812. doi:10.3892/MMR.2013.1586/HTML

88. Zhao X, Hua Y, Chen H, et al. Aldehyde dehydrogenase-2 protects against myocardial infarction-related cardiac fibrosis through modulation of the Wnt/β-catenin signaling pathway. *Ther Clin Risk Manag*. 2015;11:1371. doi:10.2147/TCRM.S88297

89. Mazzio E, Badisa R, Mack N, et al. Whole-transcriptome Analysis of Fully Viable Energy Efficient Glycolytic-null Cancer Cells Established by Double Genetic Knockout of Lactate Dehydrogenase A/B or Glucose-6-Phosphate Isomerase. *Cancer Genomics Proteomics*. 2020;17(5):469. doi:10.21873/CGP.20205

90. Lee SY, Jeon HM, Ju MK, et al. Wnt/snail signaling regulates cytochrome c oxidase and glucose metabolism. *Cancer Res*. 2012;72(14):3607-3617. doi:10.1158/0008-5472.CAN-12-0006/650294/AM/WNT-SNAIL-SIGNALING-REGULATES-CYTOCHROME-C-OXIDASE

91. Tang N, Cai X, Peng L, Liu H, Chen Y. TCP1 regulates Wnt7b/β-catenin pathway through P53 to influence the proliferation and migration of hepatocellular carcinoma cells. *Signal Transduct Target Ther*. 2020;5(1). doi:10.1038/S41392-020-00278-5

92. Song D, Shang J, Long Y, et al. Insulin-like growth factor 2 mRNA-binding protein 3 promotes kidney injury by regulating β-catenin signaling. *JCI Insight*. 2023;8(2). doi:10.1172/JCI.INSIGHT.162060

93. Lu L, Wang H, Liu X, et al. Pyruvate kinase isoform M2 impairs cognition in systemic lupus erythematosus by promoting microglial synaptic pruning via the β-catenin signaling pathway. *J Neuroinflammation*. 2021;18(1). doi:10.1186/S12974-021-02279-9

94. Gu Y, Tang S, Wang Z, et al. A pan-cancer analysis of the prognostic and immunological role of β-actin (ACTB) in human cancers. *Bioengineered*. 2021;12(1):6166. doi:10.1080/21655979.2021.1973220

95. Li Y, Liu C, Zhang X, et al. CCT5 induces epithelial-mesenchymal transition to promote gastric cancer lymph node metastasis by activating the Wnt/β-catenin signalling pathway. *Br J Cancer*. 2022;126(12):1684. doi:10.1038/S41416-022-01747-0

96. Xu J, Su Q, Gao M, Liang Q, Li J, Chen X. Differential Expression And Effects Of Peroxiredoxin-6 On Drug Resistance And Cancer Stem Cell-Like Properties In Non-Small Cell Lung Cancer. *Onco Targets Ther*. 2019;12:10477. doi:10.2147/OTT.S211125

97. Xia P, Zhang H, Lu H, et al. METTL5 stabilizes c-Myc by facilitating USP5 translation to reprogram glucose metabolism and promote hepatocellular carcinoma progression. *Cancer Commun*. 2023;43(3):338. doi:10.1002/CAC2.12403

98. Niu H, Zhou W, Xu Y, et al. Silencing PPA1 inhibits human epithelial ovarian cancer metastasis by suppressing the Wnt/β-catenin signaling pathway. *Oncotarget*. 2017;8(44):76266. doi:10.18632/ONCOTARGET.19346

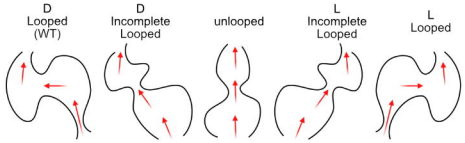
99. Chandrasekharan B, Montllor-Albalate C, Colin AE, Andersen JL, Jang YC, Reddi AR. Cu/Zn Superoxide Dismutase (Sod1) regulates the canonical Wnt signaling pathway. *Biochem Biophys Res Commun*. 2021;534:720. doi:10.1016/J.BBRC.2020.11.011

100. Li LY, Chen MY, Liu WY, Tai PZ, Liu X, Liu JX. Zebrafish cox17 modulates primitive erythropoiesis via regulation of mitochondrial metabolism to facilitate hypoxia tolerance. *FASEB J*. 2022;36(11):e22596. doi:10.1096/FJ.202200829R

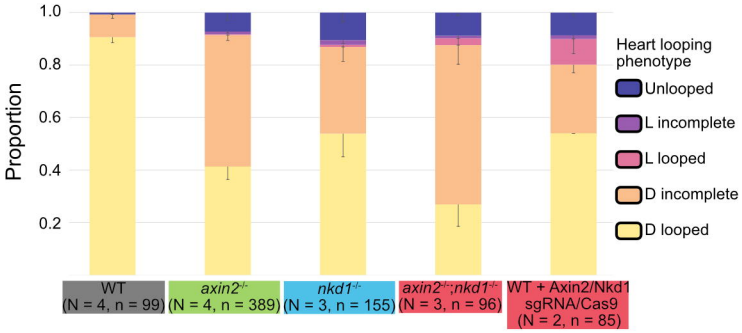
101. Liu J, Cui Z, Wang F, et al. Lrp5 and Lrp6 are required for maintaining self-renewal and differentiation of hematopoietic stem cells. *FASEB J*. 2019;33(4):5615. doi:10.1096/FJ.201802072R

102. Zhang X, Ning T, Wang H, et al. Stathmin regulates the proliferation and odontoblastic/osteogenic differentiation of human dental pulp stem cells through Wnt/β-catenin signaling pathway. *J Proteomics*. 2019;202:103364. doi:10.1016/J.JPROT.2019.04.014

A

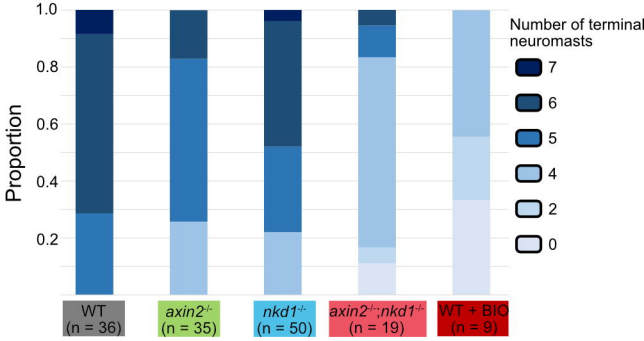


B

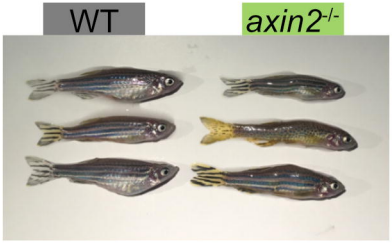


C

D



E

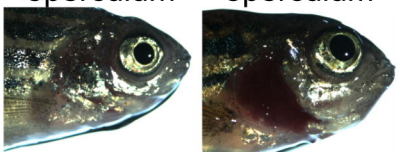


F

axin2$^{+/-}$ incross

Axin2 Genotype	Straight spine	Curved spine	Total
Wild type	11 (100%)	0 (0%)	11
Heterozygous	25 (100%)	0 (0%)	25
Homozygous mutant	4 (66.6%)	2 (33.3%)	6

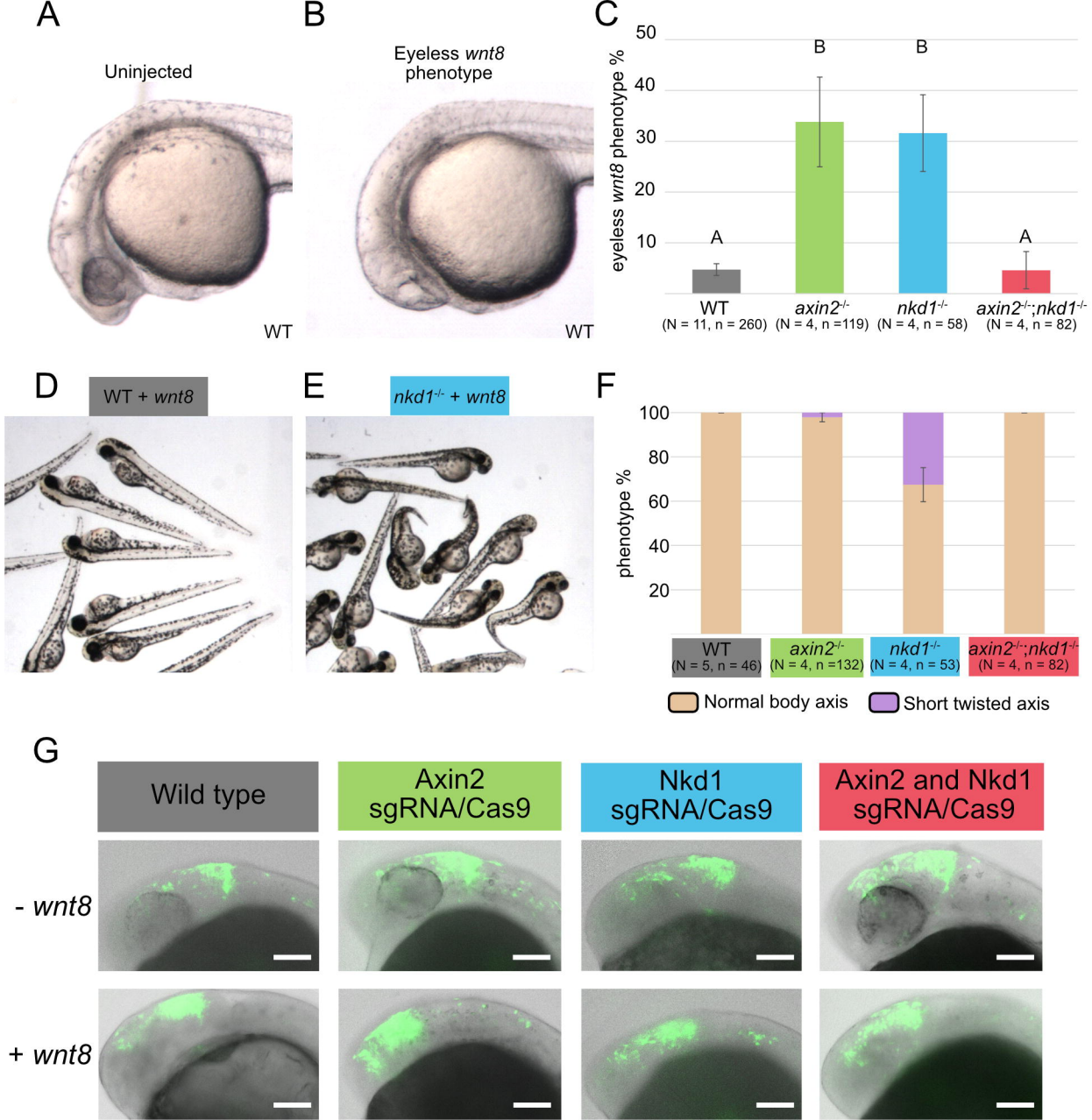
G



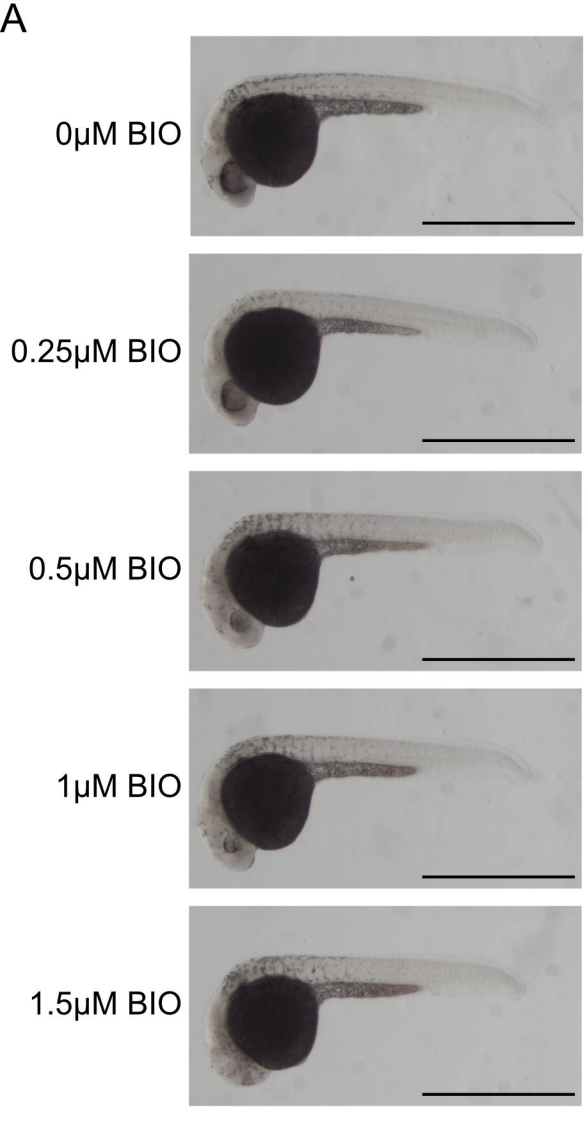
H

axin2$^{+/-}$ incross

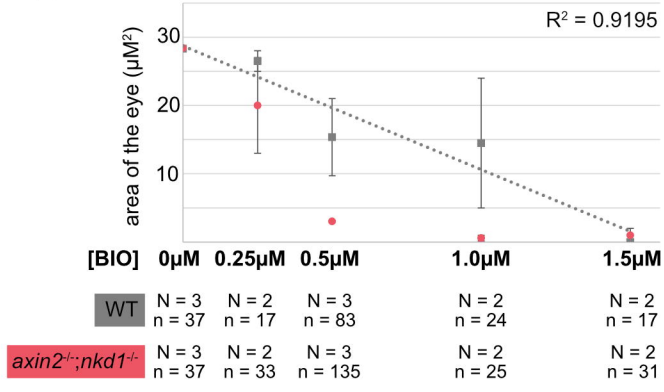
Axin2 Genotype	Healthy operculum	Receding operculum	Total
Wild type	9 (81.81%)	2 (18.18%)	11
Heterozygous	16 (64%)	9 (36%)	25
Homozygous mutant	0 (0%)	6 (100%)	6


A
Uninjected
WT
B
Eyeless wnt8 phenotype
WT
C
eyeless wnt8 phenotype %
50
40
30
20
10
A
B
B
A
WT (N = 11, n = 260)
axin2-/- (N = 4, n =119)
nkd1-/- (N = 4, n = 58)
axin2-/-;nkd1-/- (N = 4, n = 82)
D
WT + wnt8
E
nkd1-/- + wnt8
F
phenotype %
100
80
60
40
20
WT (N = 5, n = 46)
axin2-/- (N = 4, n =132)
nkd1-/- (N = 4, n = 53)
axin2-/-;nkd1-/- (N = 4, n = 82)
Normal body axis
Short twisted axis
G
Wild type
Axin2 sgRNA/Cas9
Nkd1 sgRNA/Cas9
Axin2 and Nkd1 sgRNA/Cas9
- wnt8
+ wnt8

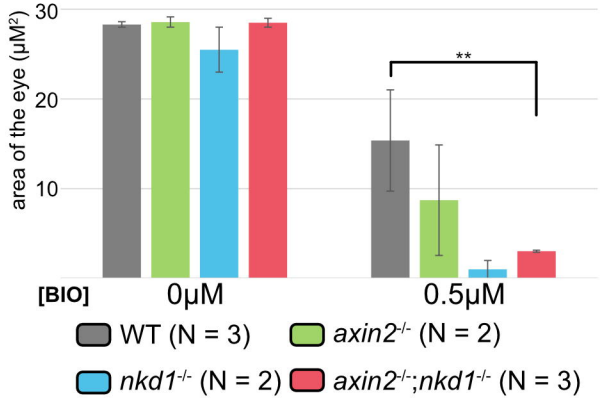
A

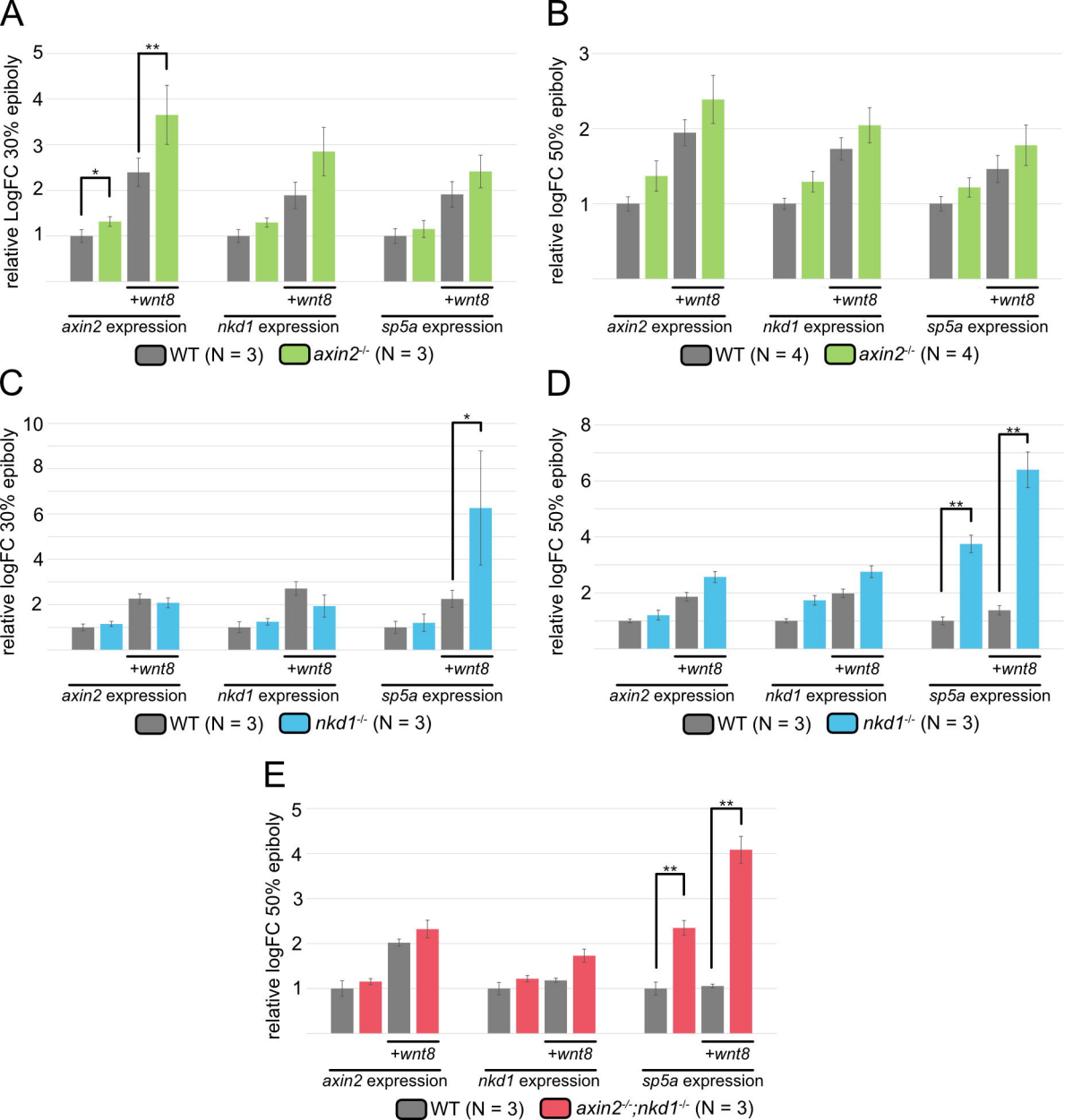

0μM BIO
0.25μM BIO
0.5μM BIO
1μM BIO
1.5μM BIO

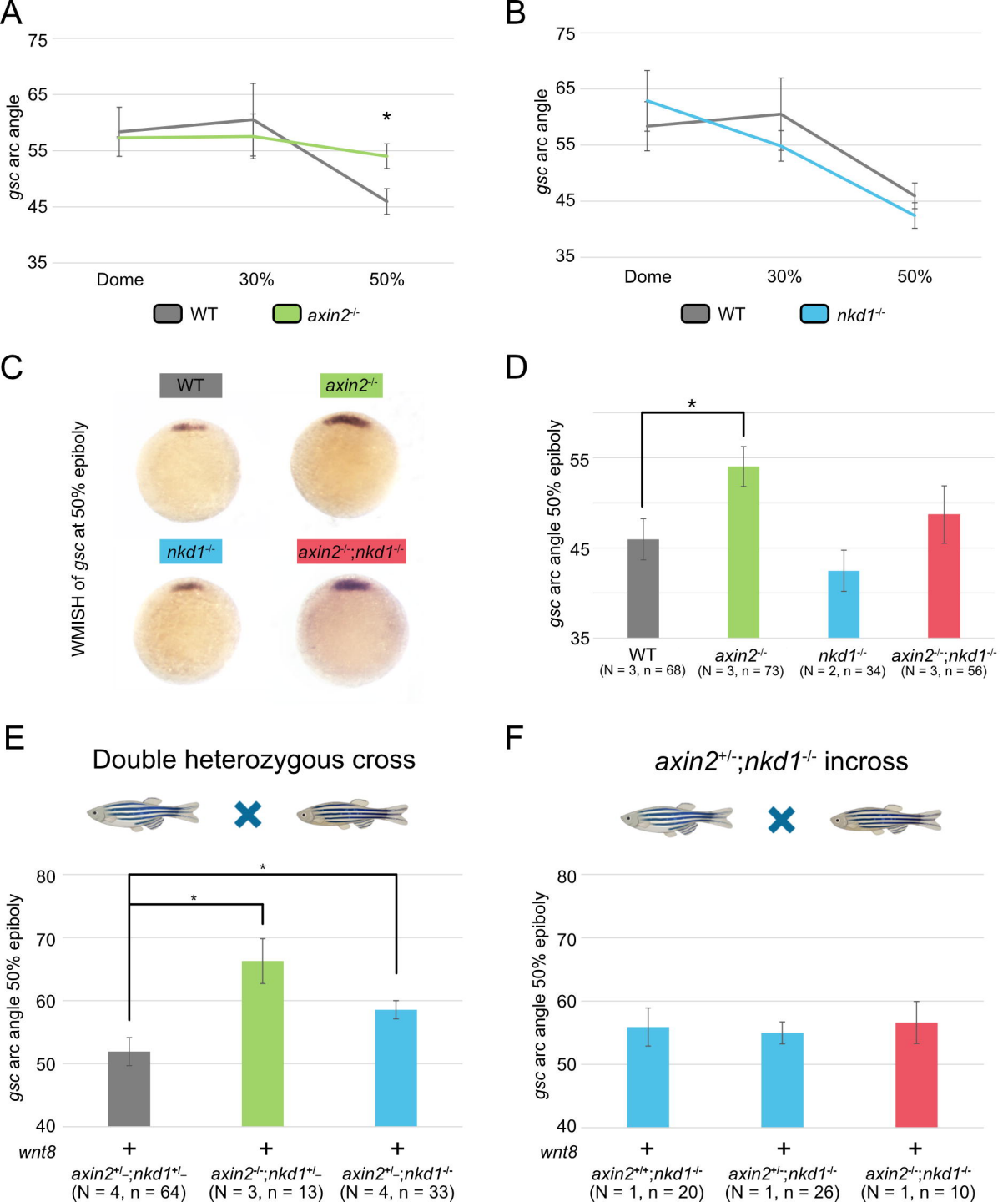
B


R² = 0.9195
area of the eye (μM²)
30
20
10
[BIO] 0μM 0.25μM 0.5μM 1.0μM 1.5μM
WT N = 3 n = 37 N = 2 n = 17 N = 3 n = 83 N = 2 n = 24 N = 2 n = 17
axin2-/-;nkd1-/- N = 3 n = 37 N = 2 n = 33 N = 3 n = 135 N = 2 n = 25 N = 2 n = 31

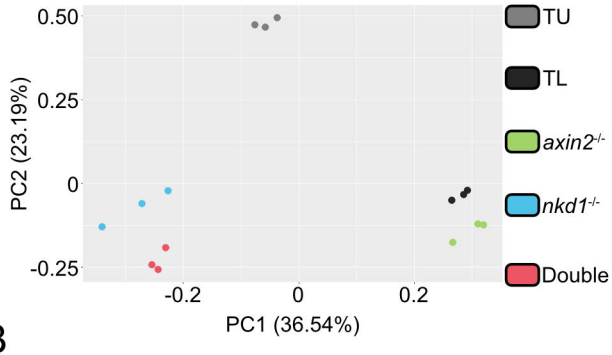
C


area of the eye (μM²)
30
20
10
**
[BIO] 0μM 0.5μM
WT (N = 3)
axin2-/- (N = 2)
nkd1-/- (N = 2)
axin2-/-;nkd1-/- (N = 3)

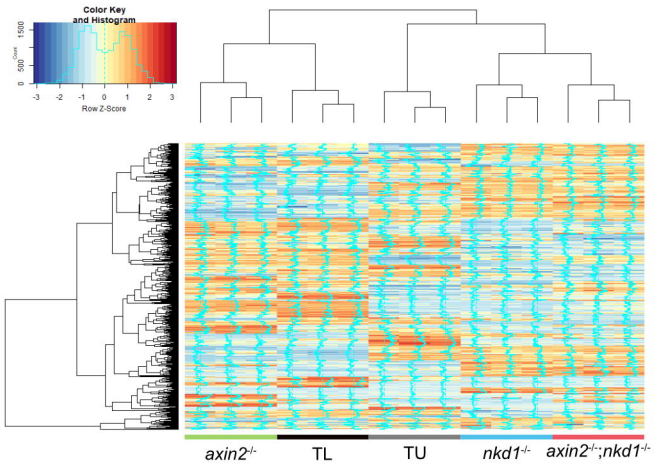

A
relative LogFC 30% epiboly
5
4
3
2
1
*
**
+wnt8
+wnt8
+wnt8
axin2 expression
nkd1 expression
sp5a expression
WT (N = 3)
axin2-/- (N = 3)
B
relative logFC 50% epiboly
3
2
1
+wnt8
+wnt8
+wnt8
axin2 expression
nkd1 expression
sp5a expression
WT (N = 4)
axin2-/- (N = 4)
C
relative logFC 30% epiboly
10
8
6
4
2
*
+wnt8
+wnt8
+wnt8
axin2 expression
nkd1 expression
sp5a expression
WT (N = 3)
nkd1-/- (N = 3)
D
relative logFC 50% epiboly
8
6
4
2
**
**
+wnt8
+wnt8
+wnt8
axin2 expression
nkd1 expression
sp5a expression
WT (N = 3)
nkd1-/- (N = 3)
E
relative logFC 50% epiboly
5
4
3
2
1
**
**
+wnt8
+wnt8
+wnt8
axin2 expression
nkd1 expression
sp5a expression
WT (N = 3)
axin2-/-;nkd1-/- (N = 3)


A
75
65
55
45
35
gsc arc angle
*
Dome
30%
50%
WT
axin2-/-
B
75
65
55
45
35
gsc arc angle
Dome
30%
50%
WT
nkd1-/-
C
WT
axin2-/-
nkd1-/-
axin2-/-;nkd1-/-
WMISH of gsc at 50% epiboly
D
*
55
45
35
gsc arc angle 50% epiboly
WT
(N = 3, n = 68)
axin2-/-
(N = 3, n = 73)
nkd1-/-
(N = 2, n = 34)
axin2-/-;nkd1-/-
(N = 3, n = 56)
E
Double heterozygous cross
80
70
60
50
40
gsc arc angle 50% epiboly
*
*
wnt8
+
+
+
axin2+/-;nkd1+/-
(N = 4, n = 64)
axin2-/-;nkd1+/-
(N = 3, n = 13)
axin2+/-;nkd1-/-
(N = 4, n = 33)
F
axin2+/-;nkd1-/- incross
80
70
60
50
40
gsc arc angle 50% epiboly
wnt8
+
+
+
axin2+/+;nkd1-/-
(N = 1, n = 20)
axin2+/-;nkd1-/-
(N = 1, n = 26)
axin2-/-;nkd1-/-
(N = 1, n = 10)

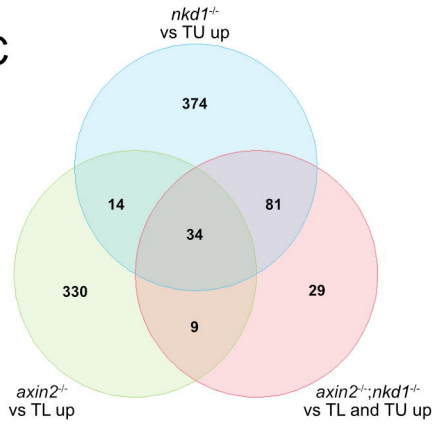
A



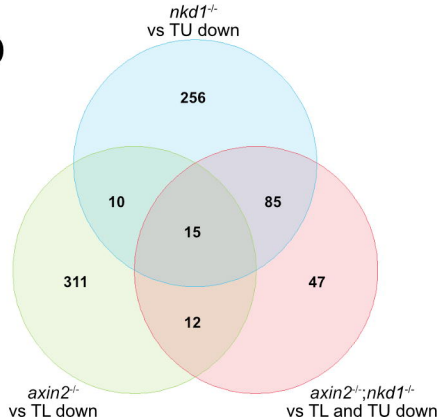
B



C



D



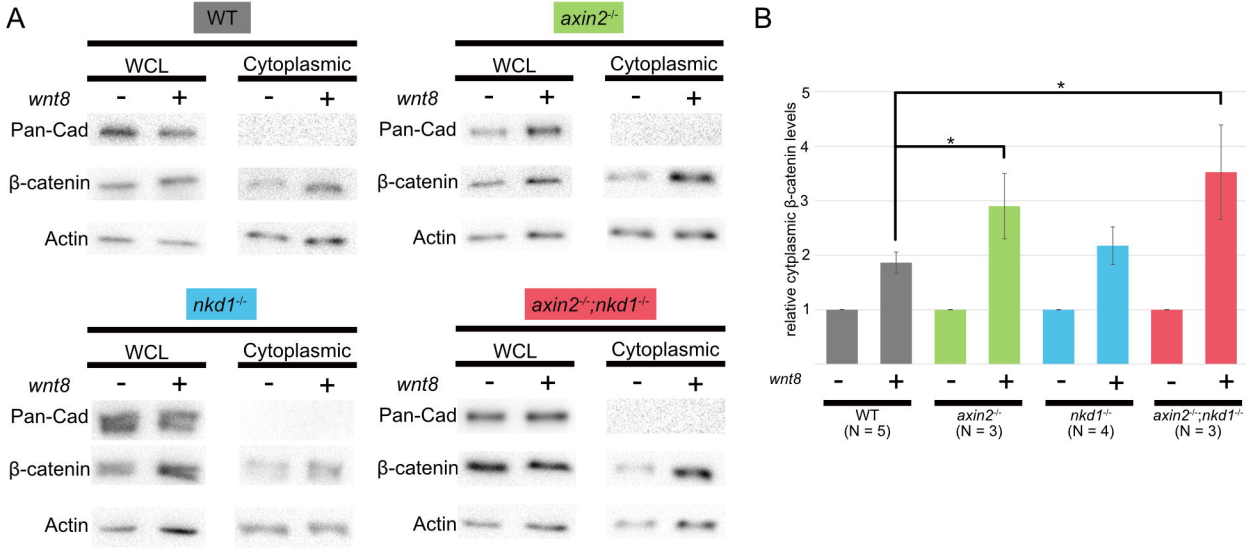
E

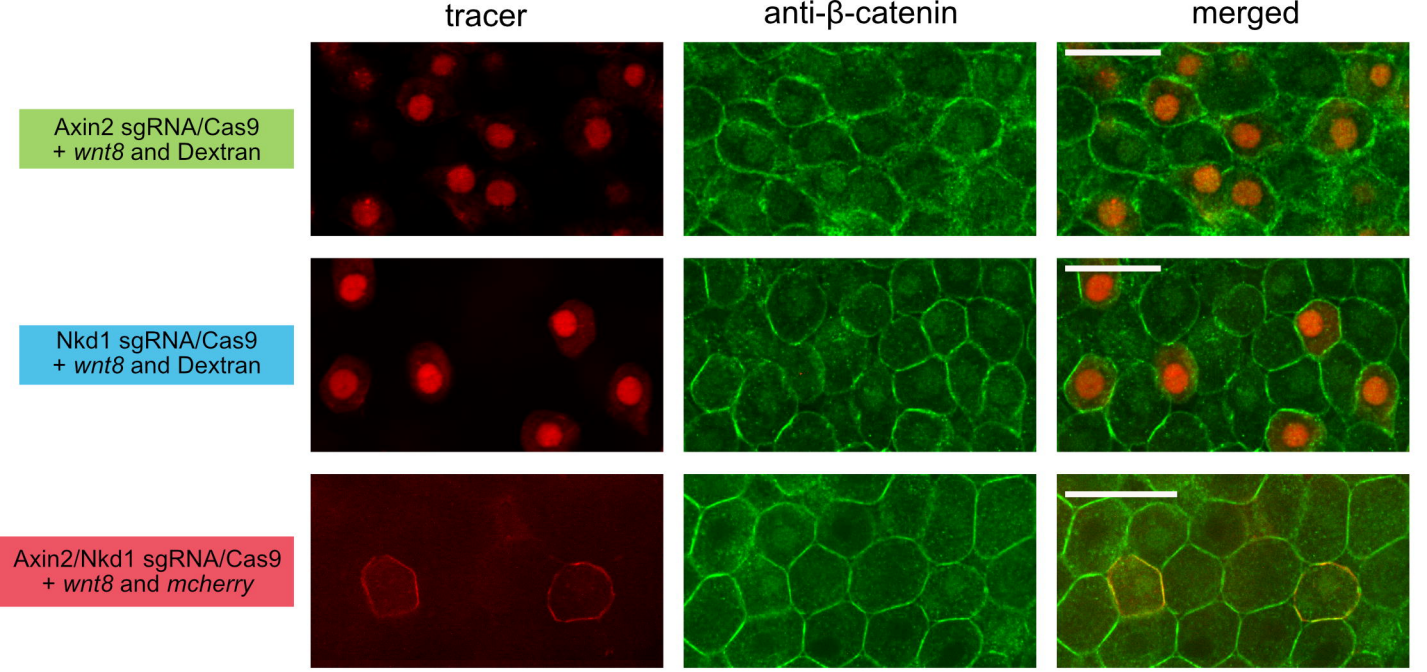
Down regulated (FDR < 0.05)
Up regulated (FDR < 0.05)

Ensembl ID	External gene name	Log fold change			
		axin2$^{-/-}$ vs TL	*nkd1*$^{-/-}$ vs TU	Double vs TU	Double vs TL
ENSDARG00000105403	abcc6b.1	3.142	3.905	3.791	3.252
ENSDARG00000074033	adamtsl2	3.007	4.264	4.061	2.913
ENSDARG00000097345	AL935186.3	2.377	3.272	3.623	2.915
ENSDARG00000095948	BX511100.5	2.562	2.063	2.472	3.026
ENSDARG00000097518	BX537110.1	4.136	4.465	4.613	4.782
ENSDARG00000092527	BX663522.1	2.109	3.334	3.574	3.569
ENSDARG00000078541	cers3a	2.882	5.307	5.577	5.260
ENSDARG00000076901	CR376751.1	5.909	3.816	3.833	8.175
ENSDARG00000080774	CR388189.1	2.564	3.935	4.512	3.823
ENSDARG00000060106	crb2a	2.035	2.453	3.087	3.355
ENSDARG00000097957	CU137680.2	2.313	2.021	2.401	2.681
ENSDARG00000074914	CU929346.1	2.796	7.627	7.610	2.044
ENSDARG00000056007	cyb5d1	2.547	2.912	3.465	4.663
ENSDARG00000113820	cyp46a1.4	11.041	3.524	2.495	4.281
ENSDARG00000044807	dck	2.366	2.424	2.058	3.560
ENSDARG00000044775	fut7	2.923	2.062	2.854	3.990
ENSDARG00000036058	gnao1b	3.220	2.005	2.334	2.236
ENSDARG00000112491	hist1h4l	2.279	2.170	2.177	5.733
ENSDARG00000022176	IGLON5	3.202	4.927	4.989	7.349
ENSDARG00000029874	khk	3.925	5.911	5.747	4.269
ENSDARG00000113737	LO018287.1	2.395	4.660	4.899	5.056
ENSDARG00000056690	mtmr1a	3.826	3.795	3.443	4.246
ENSDARG00000001933	prkag2b	5.483	5.526	7.049	7.245
ENSDARG00000007080	rhcgl1	5.875	3.771	3.370	5.418
ENSDARG00000103260	si:ch211-57b15.1	3.197	3.995	3.845	4.430
ENSDARG00000103237	si:ch73-299h12.8	4.668	2.320	2.813	5.269
ENSDARG00000093998	si:ch73-7i4.2	2.208	4.662	4.444	3.076
ENSDARG00000098406	si:dkey-22a18.2	4.205	3.149	3.100	3.906
ENSDARG00000089582	si:dkey-265e15.2	2.494	4.890	5.007	3.917
ENSDARG00000094385	si:dkey-85k15.6	2.624	6.890	6.790	4.392
ENSDARG00000080000	SLC25A1	5.401	5.614	5.736	6.683
ENSDARG00000088985	SPSB2	2.117	4.186	3.460	4.741
ENSDARG00000093477	zgc:123299	9.738	3.055	4.162	6.036
ENSDARG00000012281	zgc:65851	3.953	4.445	3.848	4.518
ENSDARG00000102528	AL935126.1	-3.027	-10.481	-10.249	-9.888
ENSDARG00000101966	BX664606.2	-8.552	-10.596	-10.366	-8.577
ENSDARG00000091938	BX890576.1	-4.364	-6.384	-3.833	-2.065
ENSDARG00000117806	BX901974.4	-5.521	-3.167	-6.767	-5.572
ENSDARG00000103172	BX914205.3	-3.628	-2.633	-3.673	-2.196
ENSDARG00000117107	CABZ01058647.1	-3.006	-3.495	-4.208	-5.290
ENSDARG00000116298	CR628395.1	-6.566	-2.985	-3.261	-2.326
ENSDARG00000043997	cyp2x8	-3.982	-5.865	-4.650	-4.286
ENSDARG00000009941	erc1a	-2.100	-2.080	-2.431	-3.748
ENSDARG00000097770	si:ch211-167j9.4	-3.753	-6.234	-4.871	-3.404
ENSDARG00000096522	si:ch211-176g6.1	-6.214	-6.380	-5.362	-3.397
ENSDARG00000089200	si:dkey-103g5.4	-2.692	-3.340	-4.010	-3.450
ENSDARG00000092702	si:dkeyp-20e4.8	-3.029	-5.570	-6.963	-2.274
ENSDARG00000091169	tor4ab	-2.030	-4.713	-8.616	-8.160
ENSDARG00000044400	wdr78	-2.112	-3.485	-2.963	-4.445

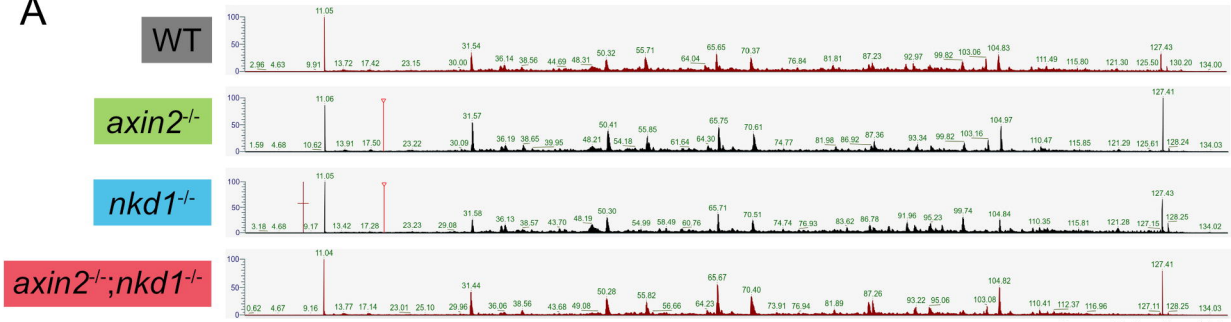
Down-regulated (FDR < 0.05) Up-regulated (FDR < 0.05)

Gene name	Log fold change			
	axin2$^{-/-}$ vs TL	*nkd1*$^{-/-}$ vs TU	Double vs TU	Double vs TL
axin2	0.456	0.443	0.722	0.620
birc5a	0.055	-0.052	-0.043	0.559
bmp4	-0.851	-0.055	-0.004	0.208
ccn1	-0.638	-0.893	0.051	0.132
ccnd1	0.274	0.174	0.194	0.111
ccnd2b	-0.188	-0.196	0.000	-1.099
cdc25b	-0.203	-0.146	-0.217	-0.340
cdh1	0.182	-0.199	-0.153	0.032
cdx1a	-0.098	0.537	0.771	0.141
ctnnb1	0.029	0.196	0.208	0.148
ctnnb2	0.219	-0.072	-0.591	-0.439
cx43	-0.556	-0.304	-0.075	-2.623
elk4	-0.282	-0.193	-0.166	0.068
en1b	0.968	0.247	0.393	0.816
fgf18a	-0.357	0.568	0.635	-0.069
fgf18b	0.325	0.602	1.021	0.274
fn1a	-0.658	1.576	0.853	-0.296
fn1b	-0.115	0.007	0.224	0.383
foxn1	0.244	-0.726	-0.212	-0.517
fsta	0.498	1.031	0.995	0.924
fzd7b	-0.119	0.122	0.056	-0.109
gbx2	0.818	0.119	0.219	1.104
gsc	0.301	0.100	0.051	0.210
jag2b	-0.123	-0.310	-0.421	-0.866
jun	0.517	-0.093	-0.038	-0.310
lbh	-0.113	0.029	0.179	-0.433
lef1	0.454	1.685	1.593	1.355
lrp5	-0.280	-0.146	0.083	0.155
mcama	5.030	-1.568	5.209	6.177
met	-0.138	-1.237	-1.721	-1.770
mitfa	0.193	-0.266	-0.195	0.193
mitfb	0.187	-0.529	-1.570	-0.313
msl1a	0.536	-0.191	-0.127	0.171
myca	0.053	0.084	0.023	0.120
mycbp	0.318	0.202	0.128	0.582
mycn	0.093	0.309	0.509	-0.034
nkd1	0.401	0.764	0.863	0.600
nos2b	0.251	-0.120	-0.106	0.346
pgfb	-0.328	-0.003	-0.004	-0.306
pitx2	0.261	0.379	0.004	0.223
ppardb	0.038	0.166	0.217	0.272
ret	-0.023	-0.369	-0.485	-0.252
sall4	0.196	0.198	0.255	0.012
sfrp2	-0.463	0.419	0.841	0.726
six3b	0.035	1.164	2.974	3.280
sox17	-0.322	1.345	1.311	1.006
sox2	0.497	1.594	1.551	0.960
sox9a	-1.272	-0.024	-0.029	-0.953
sox9b	-0.839	-0.503	-0.481	0.214
sp5a	0.021	0.890	0.962	0.867
stra6	0.169	-0.033	0.119	-0.677
tbx3a	0.193	2.098	2.703	2.417
tbx3b	-0.265	0.161	0.166	1.110
TCF4	0.015	-0.099	0.407	0.425
tiam1a	0.415	0.721	1.189	1.420
tiam1b	0.134	0.361	0.427	1.136
vangl2	-0.017	0.044	0.081	0.039
vcana	-0.751	-0.173	-1.000	-1.373
wnt3a	-0.047	-0.231	-0.145	0.429

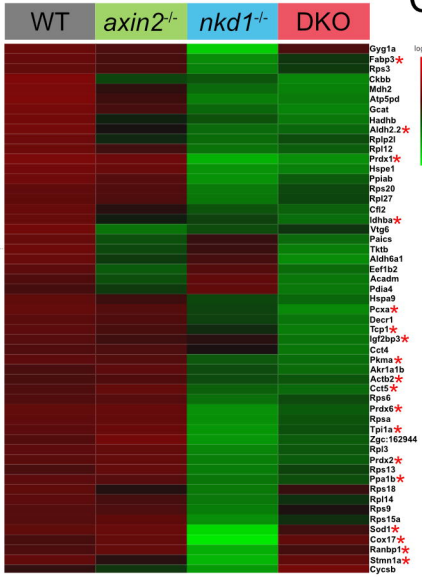

A
WT
WCL
Cytoplasmic
wnt8
-
+
Pan-Cad
β-catenin
Actin
axin2-/-
nkd1-/-
axin2-/-;nkd1-/-
B
relative cytplasmic β-catenin levels
5
4
3
2
1
*
*
WT
(N = 5)
axin2-/-
(N = 3)
nkd1-/-
(N = 4)
axin2-/-;nkd1-/-
(N = 3)


tracer
anti-β-catenin
merged
Axin2 sgRNA/Cas9
+ wnt8 and Dextran
Nkd1 sgRNA/Cas9
+ wnt8 and Dextran
Axin2/Nkd1 sgRNA/Cas9
+ wnt8 and mcherry

A



B

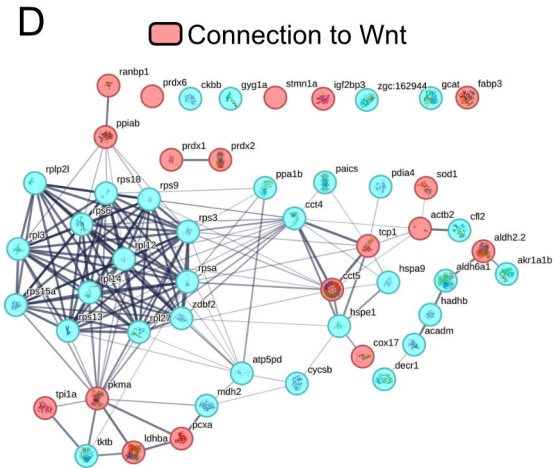


C

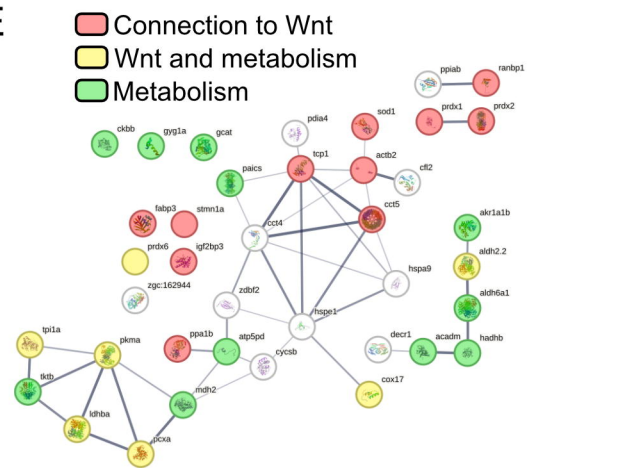
Down regulated (FDR < 0.05)
Up regulated (FDR < 0.05)

Protein	Mass spec LogFC vs WT: axin2-/-	Mass spec LogFC vs WT: nkd1-/-	Mass spec LogFC vs WT: DKO	Connection to Canonical Wnt signaling	RNA-seq LogFC: axin2-/- vs TL	RNA-seq LogFC: nkd1-/- vs TU	RNA-seq LogFC: DKO vs TU	RNA-seq LogFC: DKO vs TL
Fabp3	-0.65	-1.90	-0.99	In zebrafish knocking down fabp3 increased Wnt1, Wnt5a and Wnt11 gene expression (Liu et al, 2013)	0.384	-0.002	-0.521	-0.236
Aldh2.2	-1.04	-1.59	-1.60	In rat heart tissue ALDH2 activation reduced β-catenin protein levels and mRNA expression (Zhao et al, 2015)	-0.511	0.212	-0.080	-0.447
Prdx1	-0.57	-2.95	-2.30	In colon cancer Caov-3 cells knocking down PRDX1 reduced β-catenin protein levels (Zheng et al, 2018)	1.147	-0.697	-1.722	-0.697
Idhba	-0.88	-1.02	-1.49	In colon cancer LS174T cells knocking out LDHA/B caused an 11 fold reduction in DKK4 gene expression (Mazzio et al, 2020)	-0.097	-0.379	-0.577	-0.249
Pcxa	-0.51	-0.97	-1.85	In breast adenocarcinoma MCF-7 cells Wnt1 and Wnt3a overexpression upregulate pyruvate carboxylate mRNA and protein levels (Lee et al, 2012)	0.201	-0.241	-0.410	-0.868
Tcp1	-0.53	-0.78	-1.58	In hepatocellular carcinoma MHCC-97H cells knocking down Tcp1 decreased β-catenin and increased p-β-catenin (Tang et al, 2020)	-0.298	-0.179	-0.342	-0.348
Igf2bp3	-0.53	-0.64	-1.41	In kidney proximal tubule HKC-8 cells Igf2bp3 binds to β-catenin mRNA and enhances its stability (Song et al, 2023)	-0.171	0.096	0.330	-0.019
Pkma	-0.43	-1.09	-1.35	In the hippocampus of mice overexpressing PKM2 increases β-catenin protein levels and increased β-catenin, c-myc, and cyclinD gene expression (Lu et al, 2021)	0.204	0.090	0.228	-0.409
Actb2	-0.30	-0.79	-0.92	In squamous cell carcinoma CAL33 cells knocking down ACTB reduced β-catenin gene expression (Gu et al, 2021)	0.172	-0.013	0.046	0.207
Cct5	-0.11	-1.03	-1.19	In gastric adenocarcinoma MKN28 cells overexpression of CCT5 increased nuclear β-catenin and decreased cytoplasmic β-catenin levels (Li et al, 2022)	-0.171	-0.252	-0.049	0.076
Prdx6	-0.31	-1.94	-1.22	In epithelial lung carcinoma H1299 and A549 knocking down PRDX6 reduced β-catenin protein levels (Xu et al, 2019)	0.274	-0.191	0.292	0.136
Tpi1a	-0.20	-1.97	-1.04	In heptocellular carcinoma HCCLM3 cell lines c-myc was shown to induce Tpi1 gene expression (Xia et al, 2023)	-0.165	0.247	0.275	-0.071
Prdx2	-0.24	-1.57	-1.13	In esophageal carcinoma Eca-109 and TE-1 cells knocking down PRDX2 reduced protein levels of Wnt3a and β-catenin (Feng et al, 2020)	-0.335	-0.063	-0.120	0.006
Ppa1b	-0.40	-1.43	-1.02	In ovarian cancer ES2 and SKOV3 cells knocking down PPA1 reduced nuclear levels of β-catenin (Niu et al, 2017)	-0.239	-0.590	-0.140	-0.054
Sod1	-0.23	-3.56	-0.67	In HEK293 cells knocking down Sod1 abolished Wnt3a-dependent nuclear localization of β-catenin (Chandrasekharan et al, 2021)	-0.049	0.455	0.502	0.366
Cox17	-0.06	-3.85	-0.15	In Cox17 mutant zebrafish there is reduced β-catenin protein levels and reduced topflash reporter activity (Li et al, 2022)	-0.075	0.584	0.097	-0.381
Ranbp1	-0.10	-2.32	-0.44	In Hematopoietic stem cells knocking out Lrp5 and 6 reduced Ranbp1 gene expression (Liu et al, 2019)	0.239	0.265	0.205	0.553
Stmn1a	-0.76	-2.70	-0.41	In human dental pulp stem cells knocking down Stmn1 reduced Wnt5a and β-catenin gene expression (Zhang et al, 2019)	-0.063	-0.056	-0.190	0.360

D



E



Experiment	*axin2*$^{-/-}$	*nkd1*$^{-/-}$	*axin2*$^{-/-}$;*nkd1*$^{-/-}$	*axin2*$^{-/-}$;*nkd1*$^{-/-}$ is like?
Heart looping	++	++	++	Both
Neuromast migration	++	++	+++	Synergistic
Spine and operculum	++	-	++	*axin2*$^{-/-}$
Thigmotaxis behavior	++	++	++	Both
Tap response	+	+++	+++	*nkd1*$^{-/-}$
Eyeless Wnt8	++	++	-	WT
Twisted axis Wnt8	-	++	-	*axin2*$^{-/-}$
Wnt activity in forebrain	++	+	++	*nkd1*$^{-/-}$
Wnt activity in forebrain Wnt8	+++	+	++	nkd1$^{-/-}$
BIO sensitivity	++	++	++	Both
axin2 expression	++	+	+	*nkd1*$^{-/-}$
axin2 expression Wnt8	++	+	+	*nkd1*$^{-/-}$
nkd1 expression	+	+	+	*nkd1*$^{-/-}$
nkd1 expression Wnt8	+	+	+	Both
sp5a expression	-	+++	+++	*nkd1*$^{-/-}$
sp5a expression Wnt8	+	+++	+++	*nkd1*$^{-/-}$
gsc expression at 50%	++	-	-	*nkd1*$^{-/-}$
gsc expression with Wnt8 OE	++	++	++	Both
RNA-seq	+	++	++	*nkd1*$^{-/-}$
RNA-seq Wnt target genes	+	++	++	*nkd1*$^{-/-}$
Cytoplasmic β-catenin	++	-	++	*axin2*$^{-/-}$
Nuclear β-catenin	-	-	-	WT
Nuclear β-catenin Wnt8	-	-	-	WT
Protein expression	+	+++	+++	*nkd1*$^{-/-}$